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Quantifying chloride binding and salt extraction with poly(methyl methacrylate) copolymers bearing aryl-triazoles as anion receptor side chains

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S1. General Methods

All reagents were obtained from commercial suppliers and used as received unless otherwise noted. 3,5-Diethynylbenzenemethanol,^{S1} 1-azido-4-*tert*-butylbenzene,^{S2} 4-azidobenzyl alcohol^{S3} and tris-[(1-benzyl-1H-1,2,3-triazol-4-yl)methyl]amine (TBTA)^{S4} were prepared according to literature procedures. Methyl methacrylate (contains \leq 30 ppm MEHQ as inhibitor, 99%, Aldrich) and styrene (ReagentPlus, contains 4-*tert*-butylcatechol as stabilizer, \geq 99%, Aldrich) were eluted over a column of basic Al₂O₃ prior to use. 2,2'-Azobis(2-methylpropionitrile) (AIBN) was recrystallized from methanol. Column chromatography was performed on silica gel plates (0.25 mm thick, #1615126, Sorbent Technologies, USA) and observed under UV light. Nuclear magnetic resonance (NMR) spectra were recorded on Varian Inova (500, 400 MHz), Varian VXR (400 MHz) and Varian Gemini (300 MHz) spectrometers at room temperature (298 K). Chemical shifts were referenced on tetramethylsilane (TMS) or residual solvent peaks. High-resolution electrospray ionization (ESI), electron impact (EI), or chemical ionization (CI) mass spectrometry was performed on a Thermo Electron Corporation

MAT 95XP-Trap mass spectrometer. Thermal gravimetric analysis (TGA) was performed on a TA Instruments TGA Q5000 IR Thermogravimetric Analyzer under a N₂ atmosphere at a ramp rate of 10° C / minute. Gel permeation chromatography (GPC) was performed using an Agilent HPLC equipped with two ResiPore (300×7.5 mm) columns in series and stabilized tetrahydrofuran (THF) as the solvent unless otherwise noted. Relative molecular weights were determined based on polymethyl(methacrylate) (PMMA) standards (EasiVial, Agilent). Dynamic light scattering (DLS) measurements were made on Zetasizer Nano-ZS using a quartz cuvette.

List of Abbreviations

AIBN	2,2'-Azobis(2-methylpropionitrile)
DIPA	Diisopropylamine
Na ₂ EDTA	Ethylenediaminetetracetic acid, disodium salt
PMMA	Polymethyl(methacrylate)
TBA	Tetra- <i>n</i> -butylammonium
TBTA	Tris-[(1-benzyl-1H-1,2,3-triazol-4-yl)methyl]amine
THF	Tetrahydrofuran
TMS	Tetramethylsilane
TMSA	Trimethylsilylacetylene

S2. Synthesis and Compound Characterization



Scheme S1. Two-step preparation of PMMA-unorg from symmetric methacrylate 1



3,5-Diethynylbenzyl methacrylate (3): 3,5-Diethynylbenzenemethanol^S1 (1.23 g, 7.88 mmol) was dissolved in dry CH_2Cl_2 (50 mL), triethylamine (2.25 mL) and cooled to 0 °C. Methacryloyl chloride (860 mg, 8.27 mmol) was then added and the solution was allowed to warm slowly to room temperature over 90 minutes. After the starting material was consumed, the solution was washed with NaHCO₃, dried over MgSO₄, and concentrated in vacuo. The crude product was chromatographed over SiO₂ (2% ethyl

acetate/hexanes) to yield **3** (1.15 g, 64%) in addition to an unidentified byproduct that could be converted back to starting material upon hydrolysis in K₂CO₃/MeOH. Based on recovered starting material (185 mg), the product was obtained in a 77% yield as a white solid. ¹H NMR (400 MHz, CDCl₃) δ = 7.56 (s, 1H), 7.47 (s, 2H), 6.17 (s, 1H), 5.62 (s, 1H), 5.14 (s, 2H), 3.11 (s,

2H), 1.98 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ = 166.89, 136.82, 135.87, 135.18, 131.68, 126.21, 122.81, 82.20, 78.30, 65.12, 18.29. HR-ESI-MS: C₁₅H₁₃O₂ [M + H⁺], Calculated: 225.0914, Found: 225.0910.



Unorganized pentad (1): **3** (78 mg, 0.35 mmol), 1-azido-4*tert*-butylbenzene^S2 (135 mg, 0.77 mmol), and TBTA (18 mg) was dissolved in THF (7 mL), *t*-butanol (3 mL), and H₂O (2 mL). The solution was degassed with argon for 15 minutes followed by the addition of CuSO₄•5H₂O (9 mg dissolved in 500 μ L H₂O) and sodium ascorbate (14 mg dissolved in 500 μ L H₂O). The solution was stirred at room temperature for 4 hours. After removing the solvents in vacuo, the crude product was dissolved in CH₂Cl₂ (50 mL), washed with Na₂EDTA, brine, and then dried over MgSO₄. The product was then purified over SiO₂ (15% ethyl acetate/hexanes) to yield **1** (167 mg, 84%) as a

white solid. ¹H NMR (400 MHz, CDCl₃) $\delta = 8.37$ (m, 3H), 7.93 (s, 1H), 7.68 (d, J = 8.3 Hz, 2H), 7.47 (d, J = 8.3 Hz, 2H), 1.96 (s, 3H), 1.32 (s, 18H). ¹³C NMR (100 MHz, CDCl₃) $\delta = 167.09$, 152.01, 147.39, 137.66, 136.01, 13.44, 131.32, 126.58, 126.10, 125.01, 122.73, 119.93, 118.26, 65.98, 34.70, 31.20, 18.34. HR-ESI-MS: C₃₅H₃₈N₆O₂Cl [M + Cl⁻], Calculated: 609.2745, Found: 609.2746.



PMMA-unorganized pentad copolymer (PMMA-unorg): 1 (200 mg, 0.35 mmol) and methyl methacrylate (350 mg) were dissolved in THF (10 mL) and degassed with argon. AIBN (6 mg, 0.03 mmol) was then added and the solution was warmed to 70 °C and stirred for 48 hours. The resulting solution was cooled and precipitated in CH₃OH (150 mL) to yield **PMMA-unorg** as a white solid (320 mg) in a 58% yield. ¹H NMR (400 MHz, CDCl₃) δ = 8.67 – 8.23 (br, 3H), 8.00-7.85 (br, 2H), 7.84-7.65 (br, 4H), 7.60-7.45 (br, 4H), 5.14 (br, 2H), 3.58 (br, 28H), 1.85 (br, 19.6 H), 1.37 (br, 27.4H), 0.99, 0.82 (br, 29.9 H).



Scheme S2. Preparation of alkyne 4 through a stepwise desymmetrization of the central bisamidophenylene



N,N'-(**4-Iodo-1,3-phenylene)bis(2,2-dimethylpropanamide)** (5): *N,N'-*(1,3-Phenylene)bis(2,2-dimethylpropanamide)⁵ (1.00 g, 3.61 mmol) was dissolved in CHCl₃ (50 mL) and cooled to 0 °C in an ice-water bath. Iodine monochloride (ICl, 700 mg) dissolved in CHCl₃ (25 mL) and added dropwise over 30 minutes. After addition was

complete, the solution was slowly warmed to 40 °C. Once the staring material was consumed, the solution was neutralized with Na₂CO₃, washed with Na₂S₂O₃, and dried over MgSO₄. The volatiles were removed in vacuo and the crude material was chromatographed over SiO₂ (10% ethyl acetate/hexanes) to yield **5** (1.28 g, 88%) as a white solid. mp = 201-203 °C. ¹H NMR (400 MHz, CDCl₃) δ = 8.20 (d, *J* = 2.4 Hz, 1H), 7.83 (br s), 7.67 (d, *J* = 8.8 Hz, 1H), 7.61 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.47 (br s), 1.37 (s, 9H), 1.29 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ = 177.04, 176.73, 139.20, 138.72, 138.20, 117.73, 112.57, 82.58, 40.16, 39.67, 27.60, 27.48. HR-ESI-MS: C₁₆H₂₄N₂O₂I [M + H⁺], Calculated: 403.0882, Found: 403.0894.



N,*N*'-(4-((Trimethylsilyl)ethynyl)-1,3-phenylene)bis(2,2-di-methylpropanamide) (5-TMS): 5 (3.0 g, 7.46 mmol) was dissolved in THF/DIPA (5:1, 60 mL) and degassed with argon for 15 minutes. To the solution was then added $PdCl_2(PPh_3)_2$ (160 mg, 0.22 mmol), CuI (42 mg, 0.22 mmol) and TMSA (1.6 mL, 11 mmol). The solution was

stirred for 16 hours at rt. The resulting suspension was filtered over Celite, washed with THF, and concentrated in vacuo. The crude material was chromatographed over SiO₂ (5% ethyl acetate/hexanes) to yield **5-TMS** (2.25, 81%) as a light brown, waxy, solid. ¹H NMR (400 MHz, CDCl₃) δ = 8.41 (br s, 1H), 8.31 (d, *J* = 1.9 Hz, 1H), 7.80 (dd, *J* = 8.6, 2.0 Hz, 1H), 7.60 (br s, 1H), 7.37 (d, *J* = 8.6 Hz, 1H), 1.34 (s, 9H), 1.28 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ = 176.69, 176.61, 139.79, 139.59, 132.34, 114.61, 109.47, 107.09, 101.18, 100.20, 40.09, 39.64, 27.48, 27.41, -0.08. HR-CI-MS: C₂₁H₃₃N₂O₂Si [M^{•+}], Calculated: 372.2225, Found: 372.2228.



N,N'-(4-Ethynyl-1,3-phenylene)bis(2,2-dimethylpropanamide) (6): 5-TMS (2.24 g, 6.01 mmol) was dissolved in THF (70 mL) and saturated K₂CO₃/MeOH (10 mL) and stirred for 10 minutes. After consumption of the starting material, NH₄Cl (150 mL) was added and

extracted with dichloromethane. After drying over MgSO₄, the crude material was purified over SiO₂ (10-15% acetone/hexanes) to yield **6** (1.68 g, 93%) as an off-white solid. mp = 147-149 °C. ¹H NMR (400 MHz, CDCl₃) δ = 8.38 (br s, 1H), 8.27 (d, *J* = 1.6 Hz, 1H), 7.88 (dd, *J* = 8.6, 2.0 Hz, 1H), 7.43 (d, *J* = 8.6 Hz, 2H), 7.40 (br s, 1H), 3.52 (s, 1H), 1.34 (s, 9H), 1.29 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ = 177.13, 176.66, 140.13, 139.78, 114.63, 109.43, 105.95, 83.79, 79.24, 40.24, 39.75, 27.51, 27.49. HR-ESI-MS: C₁₈H₂₅N₂O₂ [M + H⁺], Calculated: 301.1916, Found: 301.1918.



Triad-H (7): 6 (100 mg, 0.33 mmol) and 4-azido-*tert*-butylbenzene (58 mg, 0.33 mmol) were dissolved in THF (10 mL), *t*-BuOH (3 mL), and H₂O (3 mL). After degassing the solution for 15 minutes, TBTA (9 mg), CuSO₄•5H₂O (4 mg), and sodium ascorbate (13 mg) were added sequentially. The solution was warmed to 60 °C and stirred for 2 hours. After consumption of the starting materials, the solution was concentrated, diluted with dichloromethane, washed with Na₂EDTA(aq), brine, and dried over MgSO₄. After removal of the solvent in vacuo, the crude material was purified over SiO₂ (10-25%)

ethyl acetate/hexanes) to yield 7 (149 mg, 94%) as a light yellow solid. ¹H NMR (400 MHz, CDCl₃) δ = 11.59 (br s, 1H), 8.53 (d, *J* = 2.0 Hz, 1H), 8.26 (s, 1H), 7.99 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.73 (d, *J* = 8.6 Hz, 2H), 7.55 (d, *J* = 8.9 Hz, 2H), 7.52 (br s, 1H), 1.40 (s, 9H), 1.37 (s, 9H), 1.29 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ = 178.56, 176.82, 152.66, 147.87, 138.91, 137.29, 134.18, 127.88, 126.72, 120.36, 118.45, 114.89, 113.11, 111.54, 40.43, 39.68, 34.81, 31.21, 27.70, 27.54. HR-ESI-MS: C₂₈H₃₈N₅O₂ [M + H⁺], Calculated: 476.3026, Found: 476.3044.



Triad-I (8): 7 (430 mg, 0.91 mmol) and ICl (177 mg, 1.1 mmol) was dissolved in CHCl₃, warmed to 70 °C and stirred for 16 hours. After cooling, the dark red solution was neutralized with Na₂CO₃ (aq), washed with Na₂S₂O₃ (aq) and concentrated in vacuo. The crude material was chromatographed over SiO₂ (10-20% ethyl acetate/hexanes) to yield 7 (480 mg, 88%) as a white solid. ¹H NMR (400 MHz, CDCl₃) δ = 11.33 (s, 1H), 9.61 (s, 1H), 8.38 (s, 1H), 7.85 (s, 1H), 7.75 (d, *J* = 8.4 Hz, 2H), 7.66 (s, 1H), 7.53 (d, *J* = 8.6 Hz, 2H),

1.35 (s, 9H), 1.34 (s, 9H), 1.31 (s, 9H). ¹³C NMR (400 MHz, CDCl₃) δ = 177.51, 176.15, 152.56, 146.36, 138.54, 137.99, 136.10, 134.12, 126.66, 120.29, 118.93, 115.54, 114.55, 82.16, 40.42, 40.04, 34.79, 31.23, 27.69, 27.66. HR-ESI-MS: C₂₈H₃₇N₅O₂I, [M + H⁺], Calculated: 602.1992, Found: 602.1985.



Triad-TMS (9): 8 (480 mg, 0.80 mmol) was dissolved in THF/DIPA(4:1, 25 mL) and degassed for 15 minutes. PdCl₂(PPh₃)₂ (30 mg. 0.04 mmol), CuI (8) mg, 0.04 mmol) and (trimethylsilyl)acetylene (250 µL, 1.60 mmol) were then added and the solution was stirred for 16 h. The resulting suspension was filtered over Celite, washed with THF, concentrated in vacuo and purified over SiO₂ (0-3% acetone/dichloromethane) to yield 9 (446 mg, 98%) as a light brown solid. ¹H NMR (400 MHz, CDCl₃) δ =

11.46 (s, 1H), 9.91 (s, 1H), 8.34 (br s, 1H), 8.24 (s, 1H), 7.72 (d, J = 8.5 Hz, 2H), 7.64 (s, 1H), 7.57 (d, J = 8.6 Hz, 2H), 1.39 (s, 9H), 1.38 (s, 9H), 1.34 (s, 9H), 0.30 (s, 9H). ¹³C NMR (400 MHz, CDCl₃) $\delta = 177.61$, 175.97, 152.72, 147.35, 140.34, 138.64, 134.16, 129.94, 126.74, 120.31, 118.28, 112.17, 111.22, 106.23, 101.17, 100.10, 40.50, 40.23, 34.83, 31.22, 27.69, 27.62, -0.01. HR-ESI-MS: C₃₃H₄₆N₅O₂Si [M + H⁺], Calculated: 572.3421, Found: 572.3428.





Scheme S3. Two-step preparation of PMMA-org from alkyne 4 and azido-methacrylate 10

10

4-Azido-benzylmethacrylate (10): 4-Azidobenzyl alcohol (1.13 g, 7.57 mmol) and 4-*N*,*N*[°]-dimethylaminopyridine (DMAP, 93 mg, 0.75 mmol) were dissolved in CH₂Cl₂/Et₃N (10:1, 20 mL) and cooled to 0 °C. Methacryloyl chloride (940 mg, 8.99 mmol) was then added and the solution was slowly warmed to rt over 4 hours. After the starting material had been consumed, NaHCO₃(aq) was added, extracted with dichloromethane, washed with brine, and dried over MgSO₄. After removal of the solvent, the crude material was purified over SiO₂ (1% ethyl acetate/hexanes) to

yield **10** (1.23 g, 75%) as a yellow oil. **Note:** This compound should be stored in the dark at -20 °C to hinder self-polymerization. ¹H NMR (400 MHz, CDCl₃) δ = 7.38 (d, J = 8.3 Hz, 2H), 7.03 (d, J = 8.6 Hz, 2H), 6.15 (s, 1H), 5.80 (s, 1H), 5.17 (s, 2H), 1.97 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ = 167.14, 139.97, 136.13, 132.84, 129.75, 125.89, 119.11, 65.77, 18.31. HR-CI-MS: C₁₁H₁₂C₂N₃ [M + H⁺], Calculated: 218.0924, Found: 218.0932.



Methacrylate-preorganized pentad (2): 4 (106 mg, 0.21 mmol) and 10 (51 mg, 0.23 mmol) were dissolved in THF (10 mL), *t*-BuOH (3 mL), and H₂O (2 mL) and degassed with argon. TBTA (6 mg, 0.01 mmol), CuSO₄•5H₂O (3 mg, 0.01 mmol), and sodium ascorbate (4 mg, 0.02 mmol) were then added sequentially, and the solution was stirred for 16 hours at rt. After consumption of the starting material, NH₄Cl (75 mL) was added and extracted with dichloromethane. After washing with Na₂EDTA (aq), the organic layer was dried over MgSO₄ and concentrated in vacuo. The crude material was purified over SiO₂ (15% acetone/hexanes) to yield **2** (128 mg, 84%) as an offwhite solid. ¹H NMR (400 MHz, CDCl₃) δ = 11.09 (s, 1H),

11.08 (s, 1H), 9.88 (s, 1H), 8.46 (s, 1H), 8.27 (s, 1H), 7.89 (d, J = 8.6 Hz, 2H), 7.72 (d, J = 8.5 Hz, 2H), 7.61 (s, 1H), 7.57 (d, J = 6.2 Hz, 2H), 7.55 (d, J = 7.0 Hz, 2H), 6.21 (s, 1H), 5.65 (s, 1H), 5.28 (s, 2H), 2.01 (s, 3H), 1.38 (s, 9H), 1.32 (s, 9H), 1.31 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) $\delta = 177.60$, 177.48, 167.06, 152.60, 147.60, 147.38, 137.44, 137.40, 137.25, 136.45, 136.00, 134.24, 129.27, 126.64, 126.20, 125.71, 120.85, 120.56, 118.83, 118.68, 114.18, 113.10, 112.91, 65.43, 40.30, 40.29, 34.83, 31.24, 27.68, 18.33. HR-ESI-MS: C₄₁H₄₈N₆O₄Cl [M + Cl⁻], Calculated: 751.3487, Found: 751.3458.



PMMA-organized pentad copolymer (PMMA-org): PMMA-org was prepared following a similar procedure as **PMMA-unorg** using **2** as the comonomer (1:10 feed ratio) with methyl methacrylate. 83% yield. ¹H NMR (400 MHz, CD₂Cl₂) δ = 11.15 (br, 2H), 9.96 (br, 1H), 8.44 (br, 2H), 7.87-7.50 (br, 9H), 5.08 (br, 2H), 3.60 (br, 22H), 2.05-1.7 (br, 13.9H), 1.51-1.14 (br, 36.7H), 1.02-0.84 (br, 23.5H).

S3. 2D ROESY NMR – Monomer 2



Figure S1. 2D ROESY NMR spectrum for monomer 2 (CD₂Cl₂, 298 K, 500 MHz)

S4. Anion Titrations

Dried polymer samples were dissolved in CD_2Cl_2 (400 μ L) in a septum-lined screw capped NMR tube to achieve the effective concentration of the receptor. An initial ¹H NMR spectrum was recorded and additional spectra were obtained after aliquots of a TBA⁺ halide solution (in CD_2Cl_2) was injected sequentially using a microsyringe. The ¹H NMR peak data was fit to a binding model including K_1 , K_2 , and K_{ion} using HypNMR.⁶ For **PMMA-org**, changes in the triazole (H_{d,e}), terminal phenylene (H_{i,j}) and α -TBA⁺ resonances were used in the data fitting while the triazole (H_a), and central phenylene (H_b) resonances were used in determining K_{eff} for **PMMA-unorg**. The effective concentration of the receptor was determined based on the average weight percentage of the receptor relative to methyl methacrylate as determined by ¹H NMR integration.

Figure S2. (a) ¹H NMR titration of **PMMA-org** (36 mg mL⁻¹, CD₂Cl₂, [receptor]_{eff} = ~24 mM, CD₂Cl₂, 298 K) with TBACl. (b) Resonances assigned for **PMMA-org** based upon the corresponding monomer. (c) Plot of $\delta(H_{i,j})$ vs. equivalents TBACl.

Figure S3. (a) ¹H NMR titration of **PMMA-org** (6 mg mL⁻¹, [receptor]_{eff} = ~4 mM, CD₂Cl₂, 298 K) with TBACl. (b) Resonances assigned for **PMMA-org** based upon the corresponding monomer. (c) Plot of triazole chemical shift vs. equivalents TBACl compared to **PMMA-unorg**.

Figure S4. Comparison of the terminal phenylene $(H_{i,j})$ signals for **PMMA-org** at two different polymer concentrations ([receptor]_{eff} = ~4 and ~24 mM; 6 and 36 mg mL⁻¹) during ¹H NMR titrations. The earlier inflection point and higher upfield change in chemical shift for the ~24 mM titration is consistent with a higher concentration of 2:1 cross-links formed.

Figure S5. (a) ¹H NMR titration of **PMMA-unorg** (~4 mM receptor, CD₂Cl₂, 298 K) with TBACl. (b) Resonances assigned for **PMMA-unorg** based upon the corresponding monomer. (c) Plot of δ (H_a) vs. equivalents TBACl and the corresponding binding affinity obtained from fitting to an approximate 1:1 binding model with ion-pairing (TBACl) and ion-pair complexes included.^{S7} Relative to known binding models for aryl-triazoles, *K*₂ was not included as no inflection point was observed in the signal @ 7.5 ppm stemming from the terminal phenylenes.

Figure S6. (a) Chloride binding model for unorganized aryl-triazole pentad including K_1 (1:1 binding), K_{ion} (ion pairing between the TBA⁺ and chloride), and K_{ipc} (ion pairing between the chloride complex and TBA⁺). (b) Fitting obtained from the ¹H NMR data of **PMMA-unorg**. The known value for K_{ion} was fixed while K_1 was changed systematically. The range of K_{ipc} values were obtained from this scan of K_1 . (c) Plot of observed and calculated chemical shift positions for the triazole (H_a) resonance in **PMMA-unorg**.

Figure S7. (a) Chloride binding model for **PMMA-org** and (b) values obtained from fitting ¹H NMR data using the triazole ($H_{d,e}$), terminal pheneylene ($H_{i,j}$) and the α -CH₂ resonance from the TBA⁺ cation. (c) Plot of observed and calculated triazole ($H_{d,e}$) chemical shifts in **PMMA-org**.

Figure S8. ¹H NMR titration of **PMMA-org** (100 mg mL⁻¹, [receptor]_{eff} = ~70 mM, CD₂Cl₂, 298 K) with TBACl and plots of chemical shift vs. equivalents. The H_{d,e} signals saturate upon addition of 1-2 eq. TBACl while Hi,j shows an inflection point at 0.5 eq. of added TBACl. The α proton of the TBA⁺ cation also shows an inflection point at 0.5 eq. that indicates a free TBA⁺ that is not associating with the Cl⁻ anion. Its shift back downfield is associated with pairing to form TBACl.

S5. Monomer Binding Comparison

Figure S9. Comparison of titration data for the (a) triazole and (b) terminal phenylene ¹H NMR peak shifts observed for **PMMA**-org (~4 mM, CD₂Cl₂) and for a monomeric analogue (**control**, 5 mM, CD₂Cl₂). The similar trends in triazole peak migration shows that the binding affinity of the aryl-triazole receptor is unchanged after incorporation into the polymer ($K_{eff} \sim 5 \times 10^4 \text{ M}^{-1}$. The trends in chemical shift change observed for H_{i,j} show the same inflection point which correlates to similar stabilities of the 2:1 complexes formed.

S6. Diffusion NMR

Figure S10. (a) Representative plots of peak intensity, $\ln (I/I_0)$, against the gradient squared (G^2) for **PMMA-org** with 10 eq. TBACl ([receptor]_{eff} = ~70 mM, CD₂Cl₂, 298 K). (b) *D* values for the TBACl titration with **PMMA-org** at ~4 mM (squares, 600 MHz), ~24 mM (triangles, 500 MHz) and ~70 mM (circles, 600 MHz) (CD₂Cl₂, 298 K). *D* values are based on the OMe proton from MMA. (c) The same plot as part (b) but redrawn vs. TBACl concentration. ¹H NMR spectra upon addition of TBACl that correspond to the diffusion data in Tables S1 and S2 for the (d) PMMA homopolymer (25 mg mL⁻¹) and (e) neat CD₂Cl₂.

$C1^{-}$	$D (10^{-6} \text{ cm}^2 \text{ s}^{-1})$				
concentration (mM)		PMMA-org		PM	MA
	OMe ^a	receptor ^b	α -H TBA ⁺	OMe ^a	α -H TBA ⁺
0	$1.94\pm\!\!0.03$	1.66 ± 0.03	-	$4.18\pm\!\!0.08$	-
18.5	1.96 ± 0.04	1.66 ± 0.04	$6.27\pm\!\!0.03$	$4.13\pm\!\!0.08$	14.5 ± 0.4
36.8	1.96 ± 0.05	1.68 ± 0.06	6.2 ± 0.1	$4.07\pm\!\!0.07$	15.8 ± 0.8
72.1	1.99 ± 0.05	1.69 ± 0.04	7.97 ± 0.07	$3.98\pm\!\!0.07$	14.4 ± 0.1
312	$1.69\pm\!\!0.03$	1.3 ± 0.1	$8.92\pm\!\!0.03$	$3.45\pm\!\!0.06$	12.3 ± 0.1
535	1.53 ± 0.04	1.2 ± 0.01	7.99 ± 0.01	$2.94\pm\!\!0.03$	10.3 ± 0.1
703	-	-	-	$2.55\pm\!\!0.02$	8.9 ± 0.1

Table S1. Summary of diffusion coefficients for selected peaks in the **PMMA-org** (\sim 70 mM) and the PMMA homopolymer (25 mg mL⁻¹) upon addition of TBACl salt (600 MHz)

^a Methoxy protons on PMMA. ^bAmide protons on the receptor.

Table S2. Summary of diffusion coefficients associated with the addition of TBACl salt to neat
 CD₂Cl₂ (600 MHz) _

Cl ⁻	$D (10^{-6} \text{ cm}^2 \text{ s}^{-1})$	
concentration (mM)	CHDCl ₂	ΤΒΑ-α
0.0	46 ±1	-
5.1	48 ±2	17 ± 1
25.1	45 ±2	15.9 ± 0.3
116	47 ±2	14.4 ± 0.3
213	41 ±2	13.3 ± 0.2
366	42 ±2	11.6 ± 0.2

S7. Gel Permeation Chromatography (GPC)

Figure S11. GPC elugrams for (a) methacrylate-based copolymers. Relative molecular weights (b) were determined from calibration curves obtained from PMMA standards. The degree of polymerization (DP) was calculated from the average M_n values and the molar ratios (m, n) of the two monomers.

S8. Thermal Gravimetric Analysis

Figure S12. Thermogram of PMMA-based homo- and copolymers taken under an atmosphere of nitrogen at a scan rate of 10 $^{\circ}$ C / min. The decrease in % weight loss for the copolymers provides indirect evidence for incorporation of the aryl-triazole oligomer into the polymer.

S9. Extraction

Figure S13. ¹H NMR spectra (CD₂Cl₂, 298 K) collected of **PMMA-org** ([receptor]_{eff} = 8.8 mM) after extraction with D₂O solutions (100 mM) of (a) TMACl, (b) TEACl, (c) TPACl, and (d) TBACl. The extent of extraction was determined by comparison of the peak intensities of the tetraalkylcation (R_4N^+) and the methoxy signal (–OCH₃) of methyl methacrylate.

The ability of **PMMA-org** to extract salt was determined using liquid-liquid extractions experiments and subsequent analysis was conducted using ¹H NMR. Samples of a PMMA homopolymer (5 mg, $M_n = 3,520$) and **PMMA-org** (10 mg, $M_n = 12,600$) were dissolved in CD₂Cl₂ (750 μ L, [receptor]_{eff} = 10 mM). These concentrations were selected so that the amount of PMMA in solution would be equal. Tetrapropylammonium chloride (TPACl, 16.5 mg) was dissolved in D₂O (750 μ L, [TPACl] = 100 mM). The salt and polymer solutions were added to a glass vial, sealed, and stirred for 12 h (25 °C, 1000 rpm). The mixture was then allowed to phase separate for 30 min before the organic phase was transferred into an NMR tube for analysis. Determination of extraction efficacy was done by comparing integration of the PMMA methyl group (~3.6 ppm) to the α -proton of the tetraalkylammonium cation (~3.2 ppm).

A countercation effect was demonstrated. The extraction of salts using neutral organic receptors is related to the corresponding partition coefficients (Figure 5, main text). Control experiments with the PMMA homopolymer showed this effect to be related to increased partitioning of the larger, hydrophobic cation (Figure S14). Minimal partitioning was observed for TMA, TEA, or TPACI. However, integration of the TBA⁺ resonance following extraction with the PMMA homopolymer was found to be identical to that seen in extraction experiments with **PMMA-org**.

Figure S14. ¹H NMR spectra (CD₂Cl₂, 298 K) collected after extraction experiments involving various chloride salts with the PMMA homopolymer (10 mg / mL, $M_n = 3520$). The role of the cation in effecting extraction was explored by comparing the relative integration of methyl methacrylate (MMA) to the cation (a) TMACl, (b) TEACl, (c), TPACl, and (d) TBACl.

 Table S3.
 Chloride concentrations and enhancement factors observed in the extraction of chloride salts by PMMA-org and the PMMA homopolymer

	Relative Integration (MMA/R₄N⁺)		Enhancement Factor
	PMMA-org	РММА	[CI [_]] _{PMMA-org} [CI [_]] _{PMMA}
TBACI	3:2 (20 mM)	3:2 (20 mM)	1.0
TPACI	20:1 (1.6 mM)	40:1 (0.8 mM)	2.0
TEACI	40:1 (0.8 mM)	40:1 (0.8 mM)	1.0
TMACI	47:1 (0.6 mM)	40:1 (0.8 mM)	1.2

S10. X-Ray Crystallographic Analysis of 2:1 Chloride Complex with Control

Data collection

Single crystals of a 2:1 complex of **control**^{S5} and TEACl was grown by slow diffusion of diethyl ether into a solution of 1,2-dichloroethane. A colorless crystal (approximate dimensions $0.425 \times 0.121 \times 0.074 \text{ mm}^3$) was placed onto the tip of MiTeGen and mounted on an Apex Kappa Duo diffractometer and measured at 100 K. A preliminary set of cell constants was calculated from reflections harvested from three sets of 12 frames. These initial sets of frames were oriented such that orthogonal wedges of reciprocal space were surveyed. This produced initial orientation matrices determined from 339 reflections. The data collection was carried out using Mo Ka radiation (graphite monochromator) with a frame time of 60 seconds and a detector distance of 5.0 cm. A randomly oriented region of reciprocal space was surveyed to achieve complete data with a redundancy of 4. Sections of frames were collected with 0.50° steps in ω and ϕ scans. Data to a resolution of 0.84 Å were considered in the reduction. Final cell constants were calculated from the xyz centroids of 7334 strong reflections from the actual data collection after integration (SAINT).^{S8} The intensity data were corrected for absorption (SADABS).^{S9} Please refer to Table S4 for additional crystal and refinement information.

Structure solution and refinement

The space group P–1 was determined based on intensity statistics and systematic absences. The structure was solved using SIR-92^{S10} and refined (full-matrix-least squares) using the Oxford University Crystals for Windows system.^{S11} A direct-methods solution was calculated, which provided most non-hydrogen atoms from the E-map. Full-matrix least squares / difference Fourier cycles were performed, which located the remaining non-hydrogen atoms. All non-hydrogen atoms were refined with anisotropic displacement parameters.

The structure of **control**₂•Cl⁻ exhibits structural disorder on one of the tetraethylammonium cations as well as solvent 1,2-dichloroethane. On account of a low reflection-to-parameter ratio as well as severe solvent disorder that could not be modeled successfully, Platon SQUEEZE¹² was implemented to treat the solvents with neutral charge, with tetraethylammonium being refined isotropically. Hydrogen atoms were placed in ideal positions and refined as riding atoms. The final full matrix least squares refinement converged to R1 = 0.0717 and wR2 = 0.2423 (F², all data).

Table S4. Crystal data and structure refinement for control₂•TEACl

Empirical formula	$C_{176} \ H_{240} \ Cl_2 \ N_{34} \ O_8$		
Formula weight	3030.99		
Crystal color, shape, size	colorless block, $0.425 \times 0.121 \times 0.074 \text{ mm}^3$		
Temperature	100 K		
Wavelength	0.71073 Å		
Crystal system, space group	Triclinic, P–1		
Unit cell dimensions	$a = 20.868(4) \text{ Å}$ $\alpha = 79.546(3)^{\circ}$		
	$b = 22.184(4) \text{ Å}$ $\beta = 81.047(4)^{\circ}.$		
	$c = 22.207(5) \text{ Å}$ $\gamma = 77.042(3)^{\circ}$		
Volume	9781(3) Å ³		

Z Density (calculated) Absorption coefficient F(000)

Data collection Diffractometer Theta range for data collection Index ranges Reflections collected Independent reflections Observed Reflections Completeness to theta = 25.153°

Solution and Refinement Absorption correction Max. and min. transmission Solution Refinement method Weighting scheme

Data / restraints / parameters Goodness-of-fit on F² Final R indices [I>2sigma(I)] R indices (all data) Largest diff. peak and hole 2 1.026 Mg/m³ 0.091 mm⁻¹ 3264

Bruker Apex Kappa Duo, Bruker 1.009 to 25.153° . $-24 \le h \le 24, -25 \le k \le 26, 0 \le l \le 26$ 34411 34411 [R(int) = 0.000] 1300998.2 %

Numerical 0.99 and 0.99 Direct methods Full-matrix least-squares on F² $w = [\sigma^2 Fo^{2+} AP^{2+} BP]^{-1}$, with $P = (Fo^{2+} 2 Fc^2)/3$, A = 0.136, B = 0.00034411 / 142 / 1912 0.8877 R1 = 0.0717, wR2 = 0.1864 R1 = 0.1685, wR2 = 0.2423 1.48 and -1.43 e.Å^{-3}

Figure S15. Asymmetric unit of **control**₂•TEACl crystal structure containing four pentads and two TEACl salt molecules (50% ellipsoids, hydrogen atoms omitted for clarity).

Figure S16. Two pairs of **control**₂•TEACl complexes stacked on top of one another (hydrogen atoms omitted for clarity).

Figure S17. Columnar stacking of **control**₂•TEACl complexes in the crystal structure (hydrogen atoms omitted for clarity).

Figure S18. C–H•••Cl⁻ distances within **control**₂•Cl⁻ pairs (a) AB and (b) CD.

References

- S1 Fukushima, K.; Vandenbos, A. J.; Fujiwara, T. Chem. Mater. 2007, 19, 644-646.
- S2 Lee, S.; Hua, Y.; Park, H.; Flood, A. H. Org. Lett. 2010, 12, 2100-2102.
- S3 Lamara, K.; Smalley, R. K. *Tetrahedron* **1991**, *47*, 2277-2290.
- S4 Chan, T. R.; Hilgraf, R.; Sharpless, K. B.; Fokin, V. V. Org. Lett. 2004, 6, 2853-2855.
- S5 McDonald, K. P.; Ramabhadran, R. O.; Lee, S.; Raghavachari, K.; Flood, A. H. Org. Lett. 2011, 13, 6260-6263.
- S6 Frassineti, C.; Ghelli, S.; Gans, P.; Sabatini, A.; Moruzzi, M. S.; Vacca, A. Analytical Biochemistry 1995, 231, 374-382
- S7 Y. Hua, R. O. Ramabhadran, E. O. Uduehi, J. A. Karty, K. Raghavachari and A. H. Flood, *Chem. Eur. J.*, 2011, 17, 312-321.
- S8 SAINT, Bruker Analytical X-Ray Systems, Madison, WI, current version.
- S9 Blessing, R. Acta Cryst. A. 1995, 51, 33-38.
- S10 Altomare, A; Cascarano, G; Giacovazzo, G.; Guagliardi, A.; Burla, M. C.; Polidori, G.; Camalli, M. J. Appl. Cryst. 1994, 27, 435.
- S11 Betteridge, P. W.; Carruthers, J. R.; Cooper, R. I.; Prout, K.; Watkin, D. J. J. Appl. Cryst.
 2003, 36, 1487.
- S12 Spek, A, L. Acta Cryst. D, 2009, 65, 148-155.