

Chemical Communications

Electronic Supporting Information

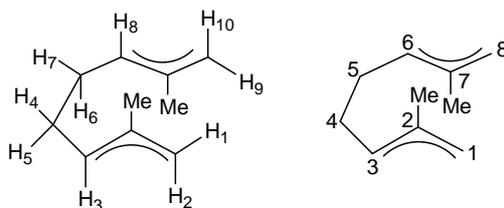
**An efficient ruthenium(IV) catalyst for the
selective hydration of nitriles to amides in water
under mild conditions**

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EXPERIMENTAL SECTION

General methods: Synthetic procedures were performed under an atmosphere of dry nitrogen using vacuum-line and standard Schlenk techniques. Organic solvents were dried by standard methods and distilled under nitrogen before use. All reagents were obtained from commercial suppliers and used without any further purification, with the exception of complex $[\{\text{RuCl}(\mu\text{-Cl})(\eta^3:\eta^3\text{-C}_{10}\text{H}_{16})\}_2]$ (**1**),¹ the secondary phosphine oxides $\text{R}_2\text{P}(=\text{O})\text{H}$ ($\text{R} = \text{Ph}, \text{Me}$)² and 4-cyano-1-(2,6-difluorobenzyl)-1*H*-1,2,3-triazole (**3aa**),³ which were prepared by following the methods reported in the literature. Infrared spectra were recorded on a Perkin-Elmer 1720-XFT spectrometer. GC measurements were made on a Hewlett-Packard HP6890 equipment using a Supelco Beta-DexTM 120 column (30 m length; 250 μm diameter). NMR spectra were recorded on a Bruker DPX-300 or AV400 instruments. The chemical shift values (δ) are given in parts per million and are referred to the residual peak of the deuterated solvent employed (^1H and ^{13}C) or an external 85% aqueous H_3PO_4 solution (^{31}P). DEPT experiments have been carried out for all the compounds reported. Elemental analyses were provided by the Analytical Service of the Instituto de Investigaciones Químicas (IIQ-CSIC) of Seville. The numbering for protons and carbons of the 2,7-dimethylocta-2,6-diene-1,8-diyl skeleton is as follows:



Synthesis of $[\text{RuCl}_2(\eta^3:\eta^3\text{-C}_{10}\text{H}_{16})\{\text{PPh}_2(\text{OH})\}]$ (2a**):** A solution of the dimeric precursor $[\{\text{RuCl}(\mu\text{-Cl})(\eta^3:\eta^3\text{-C}_{10}\text{H}_{16})\}_2]$ (**1**)

(0.200 g, 0.32 mmol) and $\text{Ph}_2\text{P}(=\text{O})\text{H}$ (0.131 g, 0.64 mmol) in dichloromethane (50 mL) was stirred at room temperature for 48 h. The solvent was then removed under reduced pressure. The resulting yellow solid washed with hexanes (2 x 10 mL) and diethyl ether (2 x 10 mL), and dried under vacuum. Yield: 0.269 g (82%). IR (KBr): 3035 (s), 2909 (w), 2854 (w), 1437 (s), 1380 (m), 1216 (w), 1146 (s), 1100 (m), 1027 (w), 878 (s), 742 (m), 690 (s), 609 (w), 538 (s), 489 (s) cm^{-1} . $^{31}\text{P}\{^1\text{H}\}$ NMR (CDCl_3) δ = 80.2 (s) ppm. ^1H NMR (CDCl_3) δ = 2.14 (s, 6H, Me), 2.70–2.75 (m, 2H, H_4 and H_6), 3.15 (d, 2H, $^3J_{\text{PH}}$ = 3.3 Hz, H_2 and H_{10}), 3.52–3.58 (m, 2H, H_5 and H_7), 3.85 (d, 2H, $^3J_{\text{PH}}$ = 10.4 Hz, H_1 and H_9), 5.14–5.18 (m, 2H, H_3 and H_8), 7.39–7.44 (m, 6H, Ph), 7.75 (s, 1H, OH), 8.00–8.06 (m, 2H, Ph), 8.11–8.17 (m, 2H, Ph) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3) δ = 21.1 (s, Me), 37.2 (s, C_4 and C_5), 67.1 (s, C_1 and C_8), 107.7 (br, C_3 and C_6), 125.2 (s, C_2 and C_7), 128.0 (d, $^2J_{\text{CP}}$ = 10.5 Hz, Ph), 128.2 (d, $^2J_{\text{CP}}$ = 10.5 Hz, Ph), 129.6–129.8 (m, Ph), 139.1 (d, $^1J_{\text{CP}}$ = 54.3 Hz, Ph), 140.8 (d, $^1J_{\text{CP}}$ = 53.6 Hz, Ph) ppm. Elemental analysis calcd. (%) for $\text{C}_{22}\text{H}_{27}\text{Cl}_2\text{OPRu}$: C 51.77, H 5.33; found: C 51.59, H 5.38.

Synthesis of $[\text{RuCl}_2(\eta^3:\eta^3\text{-C}_{10}\text{H}_{16})\{\text{PMe}_2(\text{OH})\}]$ (2b): A solution of the dimeric precursor $[\{\text{RuCl}(\mu\text{-Cl})(\eta^3:\eta^3\text{-C}_{10}\text{H}_{16})\}_2]$ (**1**) (0.150 g, 0.24 mmol) and $\text{Me}_2\text{P}(=\text{O})\text{H}$ (0.037 g, 0.48 mmol) in dichloromethane (50 mL) was stirred at room temperature for 48 h. The solvent was then removed under reduced pressure and the resulting oily residue dissolved in diethyl ether (30 mL) and filtered. The filtrate was evaporated to dryness and the yellow syrup formed washed with hexanes (10 mL) to afford a dark yellow solid, which was dried under vacuum. Yield: 0.156 g (84%). IR (KBr): 3188 (m), 2962 (w), 2906 (m), 2852 (w), 1457(w), 1413 (m), 1383 (m), 1283 (m), 1137 (s), 1024 (s), 948 (s), 912 (s), 849 (s), 736 (m), 699 (s), 556 (m) cm^{-1} . $^{31}\text{P}\{^1\text{H}\}$ NMR (CDCl_3) δ = 108.5 (s) ppm. ^1H

NMR (CDCl₃) δ = 1.91 (d, 3H, $^2J_{HP}$ = 9.6 Hz, PMe), 2.03 (d, 3H, $^2J_{HP}$ = 9.5 Hz, PMe), 2.24 (s, 6H, Me), 2.84-2.88 (m, 2H, H₄ and H₆), 3.06 (d, 2H, $^3J_{PH}$ = 4.8 Hz, H₂ and H₁₀), 3.42-3.47 (m, 2H, H₅ and H₇), 4.25 (d, 2H, $^3J_{PH}$ = 8.2 Hz, H₁ and H₉), 4.91-4.95 (m, 2H, H₃ and H₈) ppm; OH signal not observed. $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl₃) δ = 19.8 (d, $^1J_{CP}$ = 41.1 Hz, PMe), 20.6 (s, Me), 22.2 (d, $^1J_{CP}$ = 35.7 Hz, PMe), 36.5 (C₄ and C₅), 63.4 (d, $^2J_{CP}$ = 6.2 Hz, C₁ and C₈), 108.2 (d, $^2J_{CP}$ = 9.7 Hz, C₃ and C₆), 121.0 (s, C₂ and C₇) ppm. Elemental analysis calcd. (%) for C₁₂H₂₃Cl₂OPRu: C 37.31, H 6.00; found: C 37.17, H 6.38.

General procedure for the catalytic hydration of nitriles with complex [RuCl₂(η^3 : η^3 -C₁₀H₁₆){PMe₂(OH)}] (2b): Under a nitrogen atmosphere, the corresponding nitrile **3a-ac** (1 mmol), water (3 mL), and the ruthenium complex **2b** (0.0038 g, 0.01 mmol; except for **3h** and **3ab,ac** which required 0.03 and 0.05 mmol of **2b**, respectively) were introduced into a Teflon-capped sealed tube, and the reaction mixture stirred at 60 °C for the indicated time (see Table 2 and Schemes 2-3). The course of the reaction was monitored regularly taking samples of ca. 20 μL , which, after extraction with CH₂Cl₂ (3 mL), were analyzed by GC (in the case of **3ab,ac** the reactions were monitored by ^1H NMR spectroscopy). Once the reaction is completed, two different procedures were applied to isolate the primary amide product depending on its solubility. For amides which partially precipitate in the medium, the reaction mixture was cooled at r.t., or 0 °C in an ice bath, until complete precipitation/crystallization of the product. The suspension was then filtered, the solid washed with water (2 x 3 mL) and hexanes (3 mL) and dried in vacuo. For those amides that do not precipitate, isolation was carried out as follows: After elimination of the water solvent under

reduced pressure, the crude reaction mixture was pulverized in diethyl ether and the resulting solid filtered, washed and dried in vacuo. In some cases additional purification by column chromatography was required (silica gel with CH₂Cl₂ as the eluent). The identity of the amide products was assessed by comparison of their NMR spectroscopic data with those reported in the literature (copies of the NMR spectra are included below).

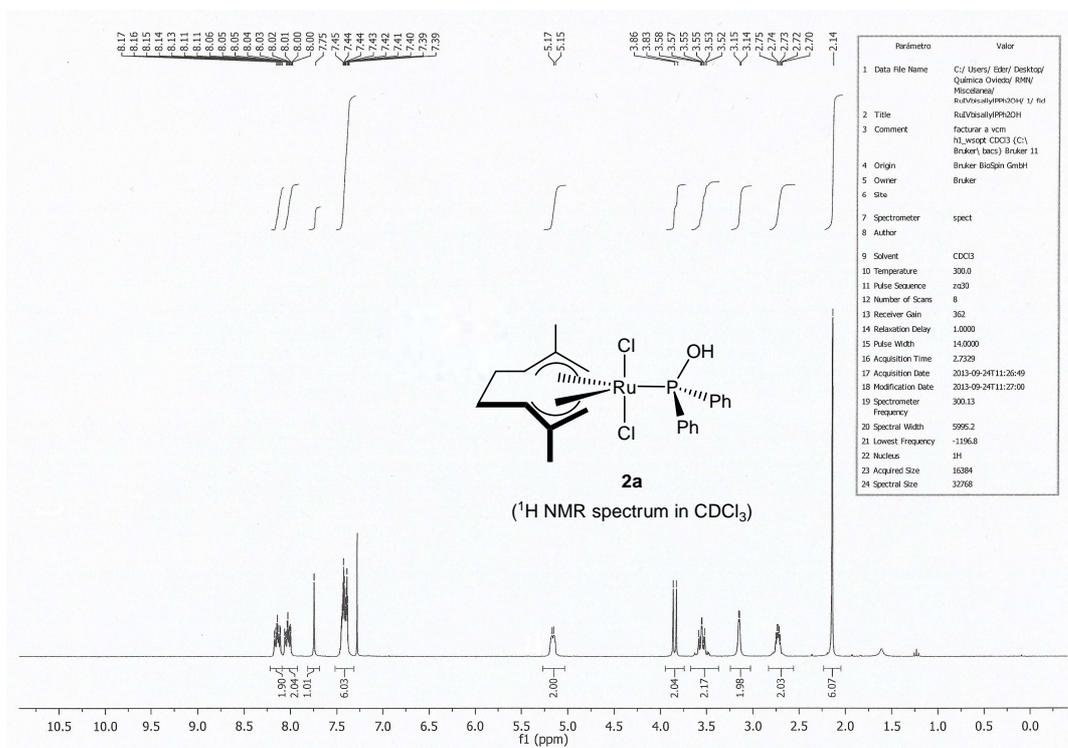
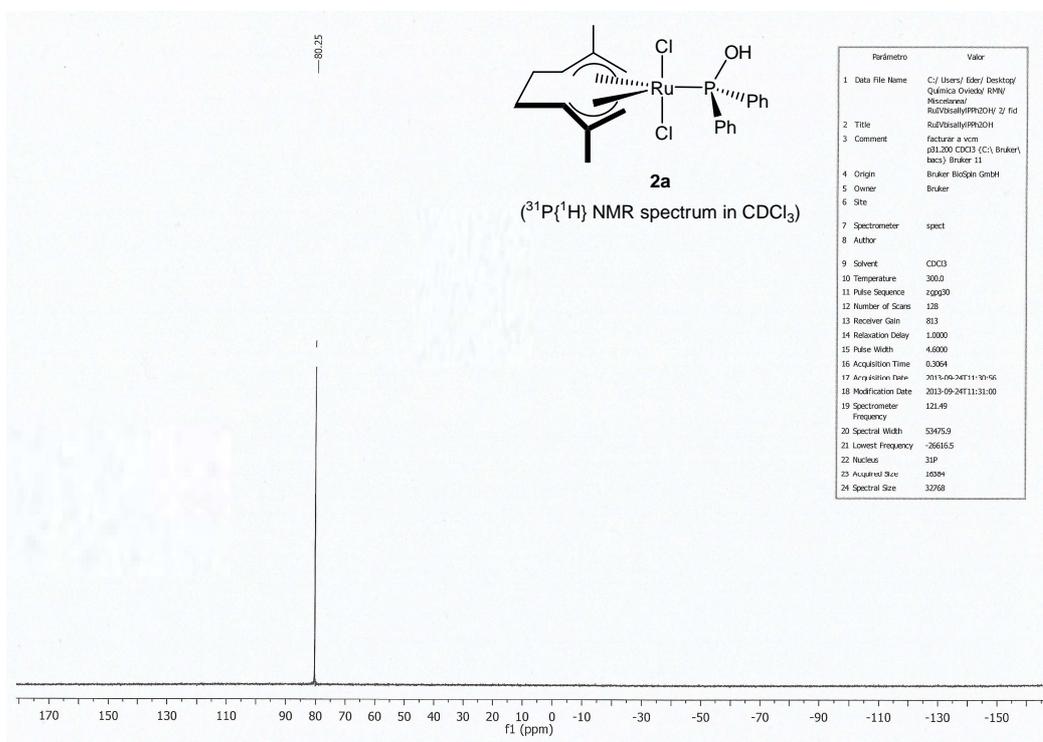
Recycling of complex [RuCl₂(η³:η³-C₁₀H₁₆){PMe₂(OH)}] (2b):

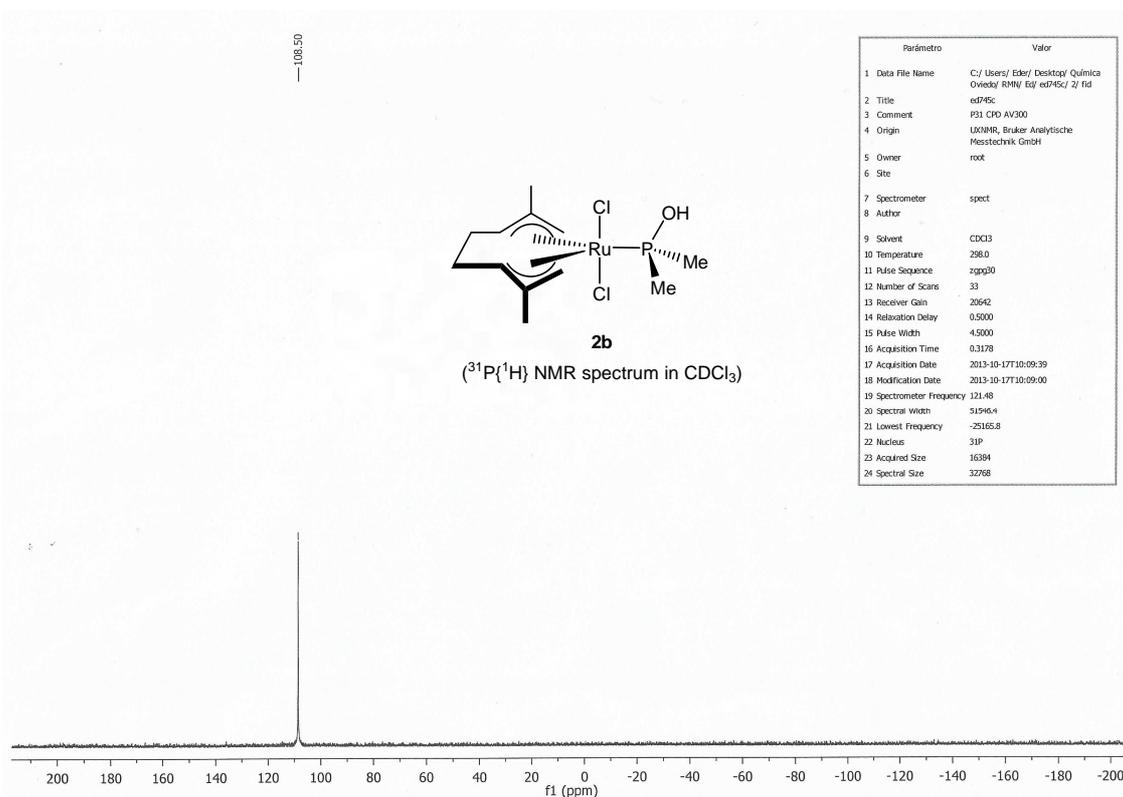
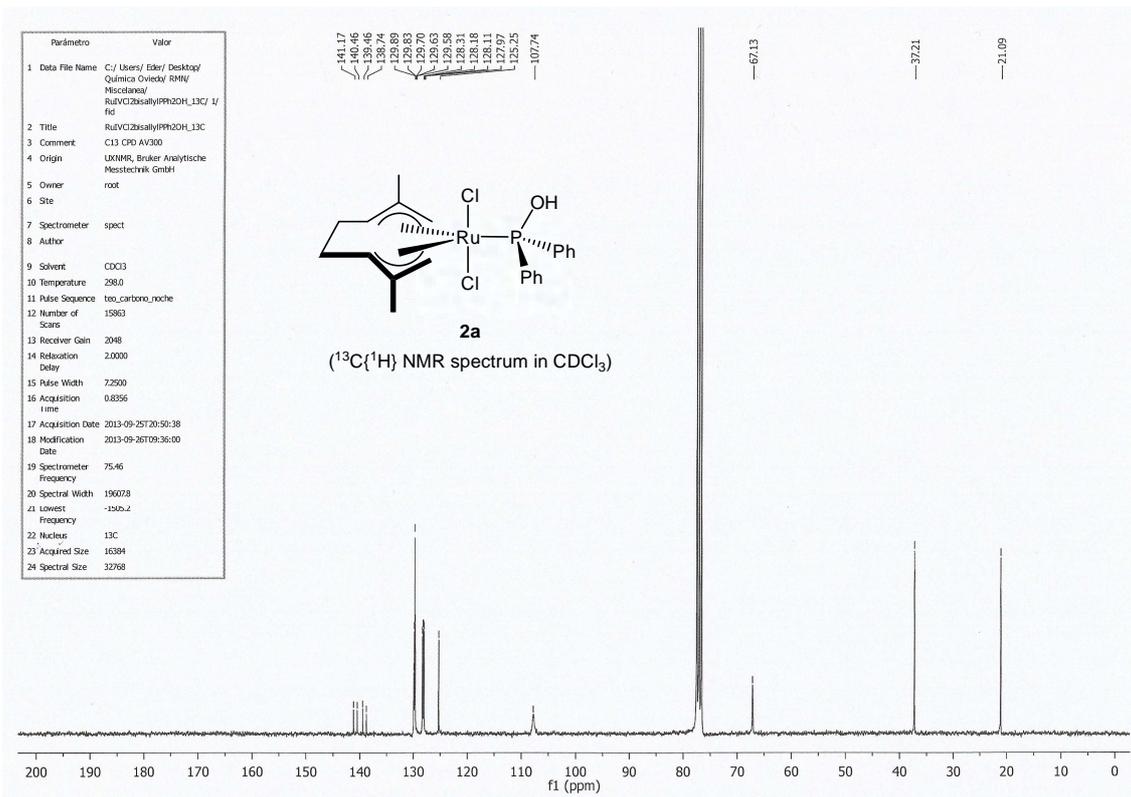
Under a nitrogen atmosphere, pentafluorobenzonitrile **3b** (0.193 g, 1 mmol), water (3 mL), and the ruthenium complex **2b** (0.0038 g, 0.01 mmol) were introduced into a Teflon-capped sealed tube, and the reaction mixture was stirred at 60 °C for 30 min. After this time, a sample of ca. 20 μL was taken and, after extraction with CH₂Cl₂ (3 mL), analyzed by GC to determine the conversion of **3b** into **4b**. The reaction mixture was then cooled at 0 °C in an ice bath until the complete crystallization of **4b**. The suspension was filtered and pentafluorobenzonitrile **3b** (0.193 g, 1 mmol) added to the aqueous filtrate containing **2b**. The resulting mixture was introduced into a Teflon-capped sealed tube, and stirred at 60 °C for 30 min. The same procedure was repeated for 10 times.

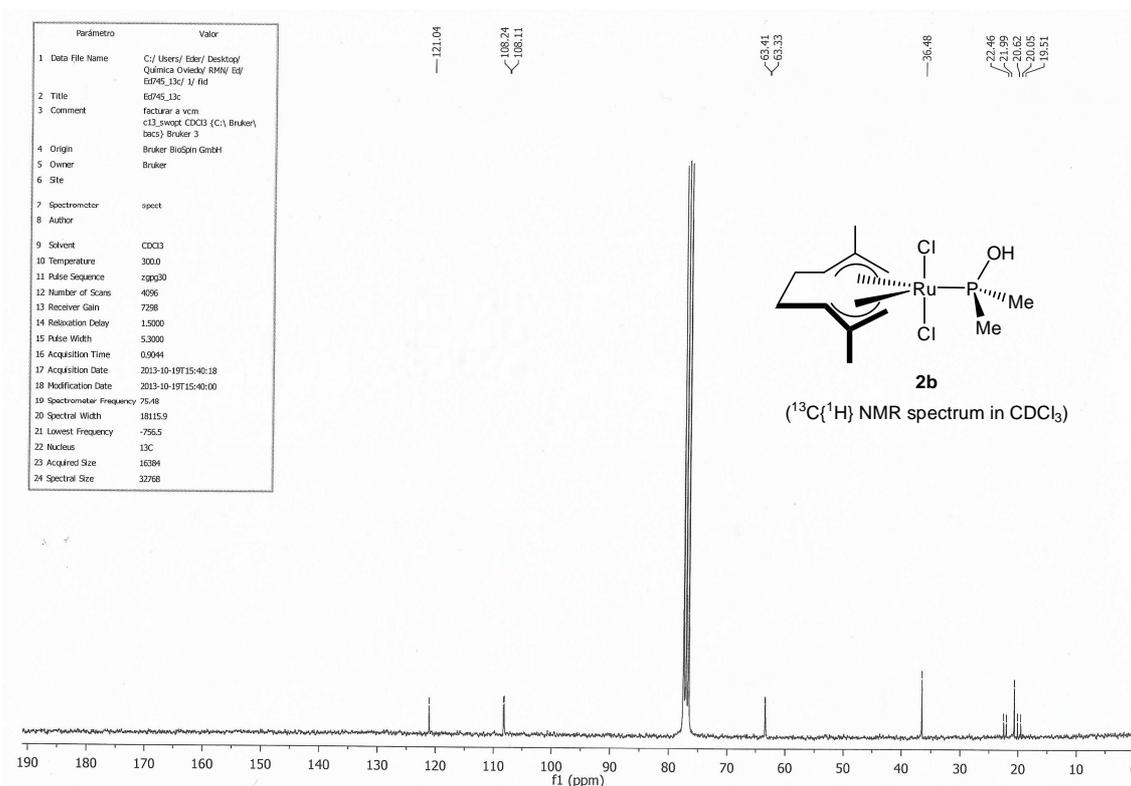
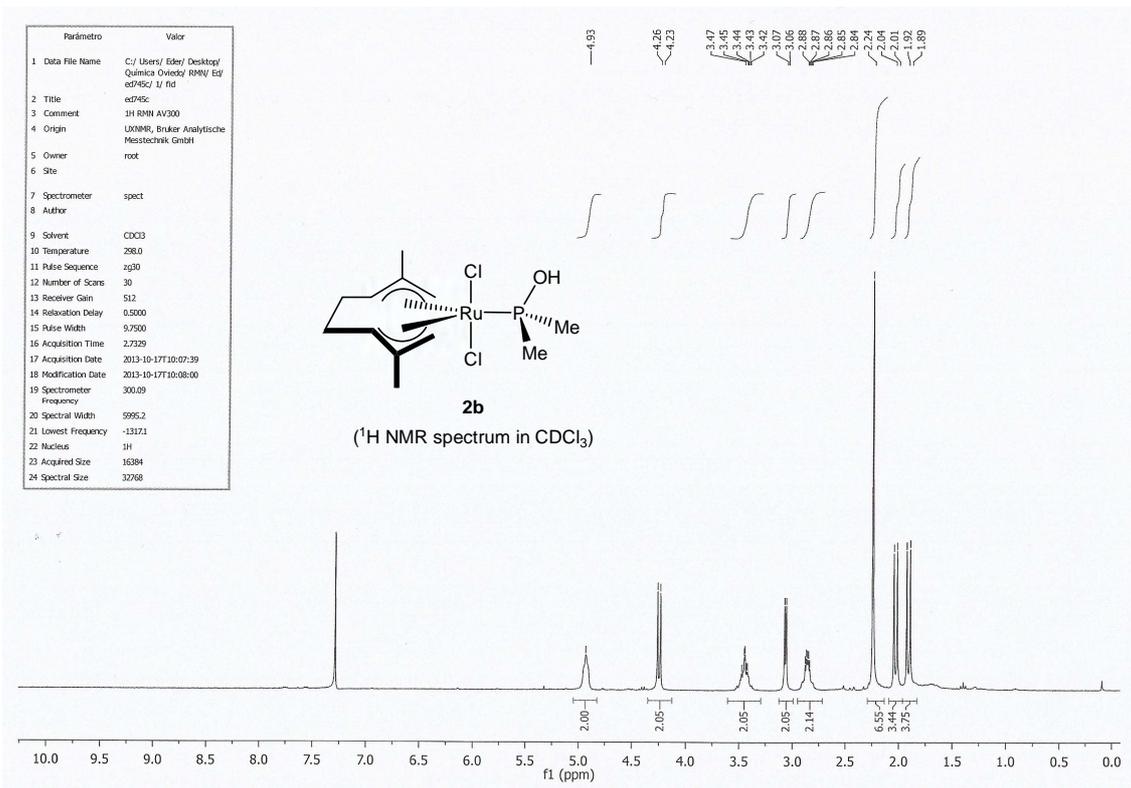
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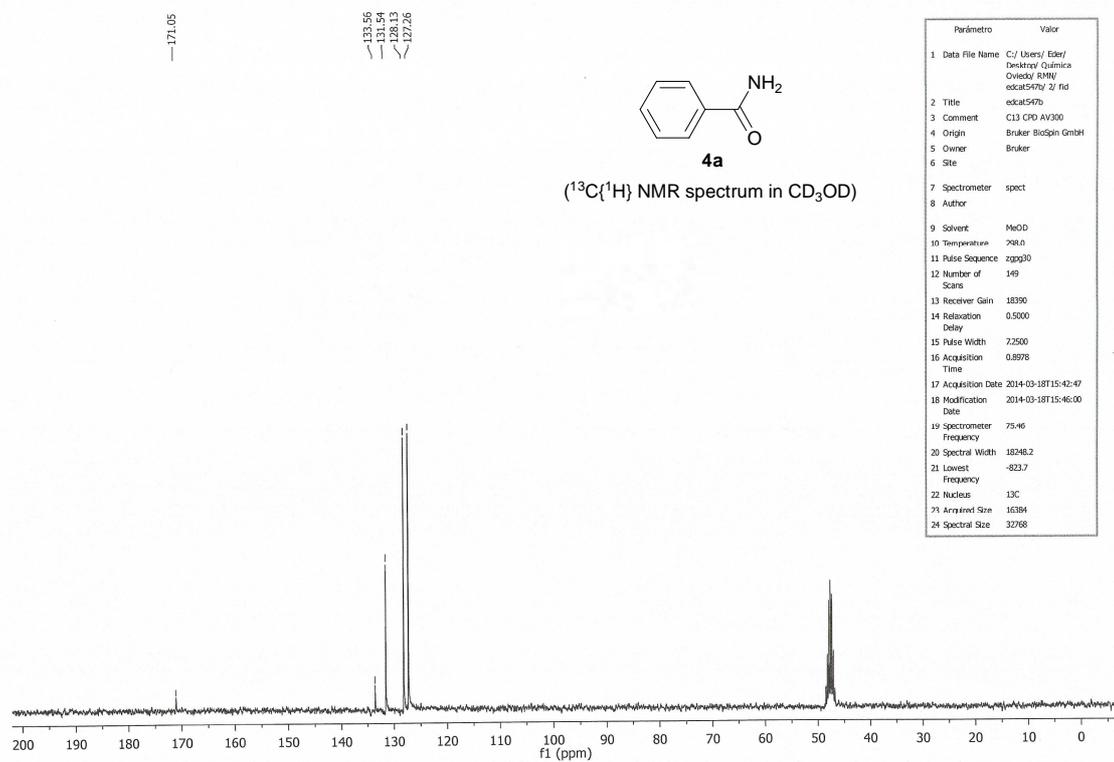
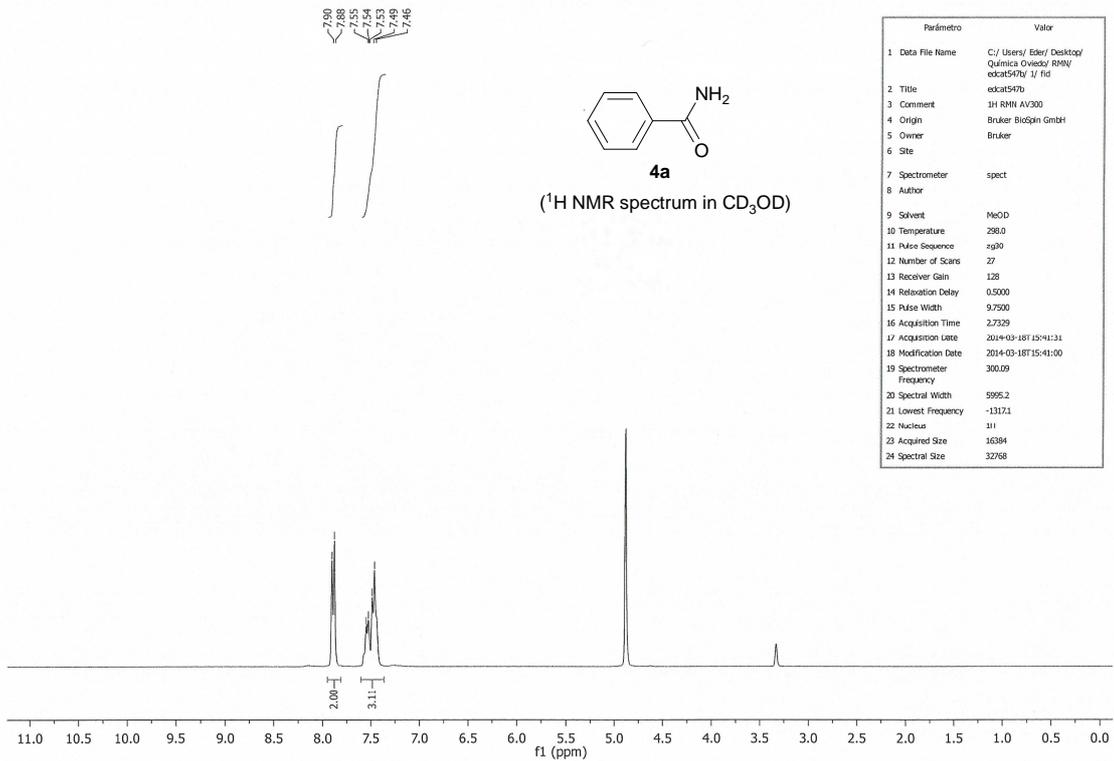
- 1 (a) L. Porri, M. C. Gallazzi, A. Colombo and G. Allegra, *Tetrahedron Lett.*, 1965, **47**, 4187; (b) A. Salzer, A. Bauer, S. Geysler and F. Podewils, *Inorg. Synth.*, 2004, **34**, 59.
- 2 (a) A. Camp, C. P. Healy, I. D. Jenkins, B. W. Skelton and A. H. White, *J. Chem. Soc., Perkin Trans. 1*, 1991, 1323; (b) H. R. Hays, *J. Org. Chem.*, 1968, **33**, 3690.

COPIES OF THE NMR SPECTRA OF COMPLEXES 2a,b AND AMIDES 4a-ac

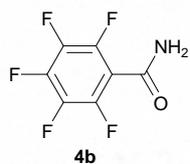




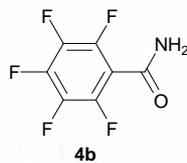
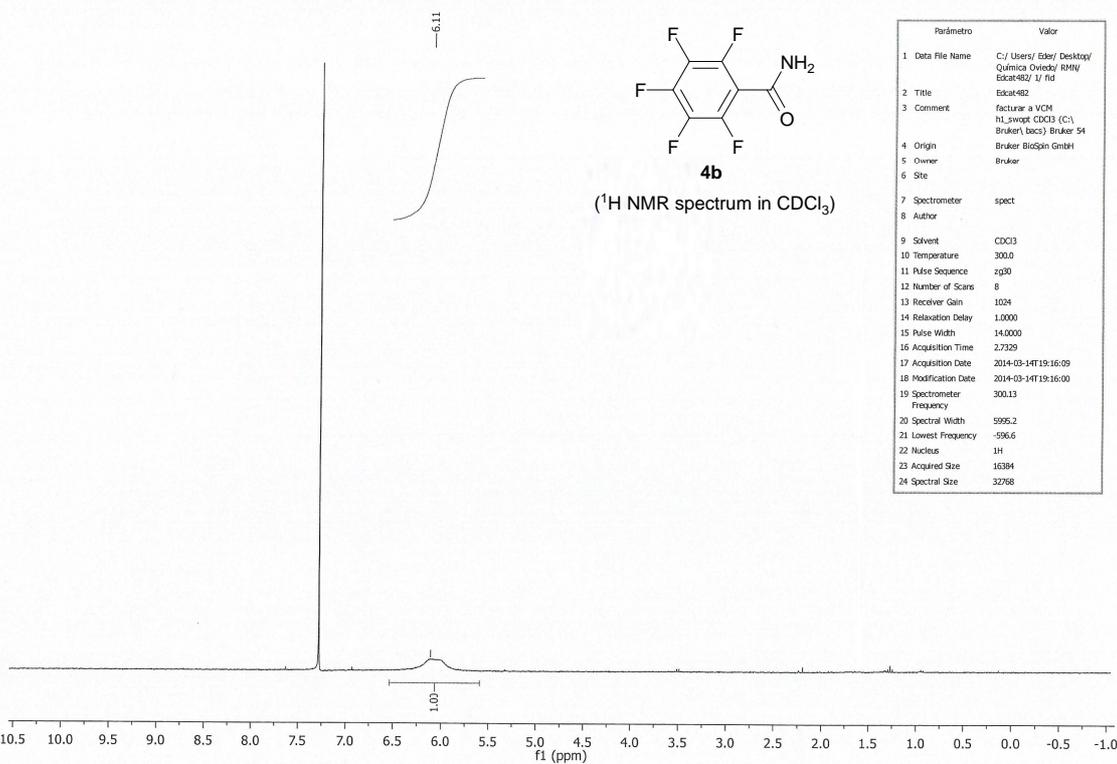
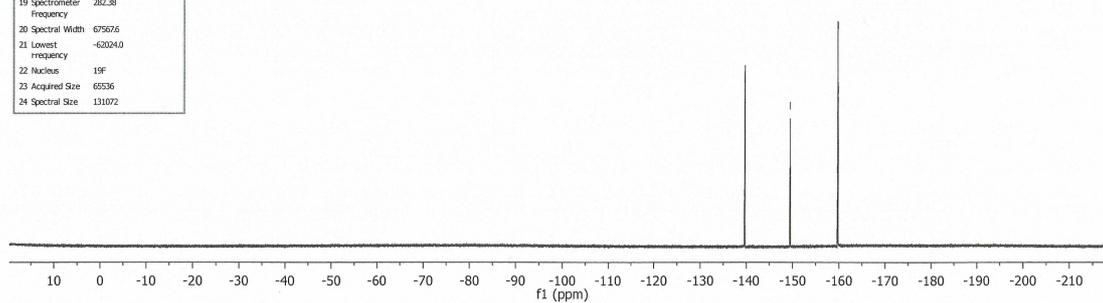




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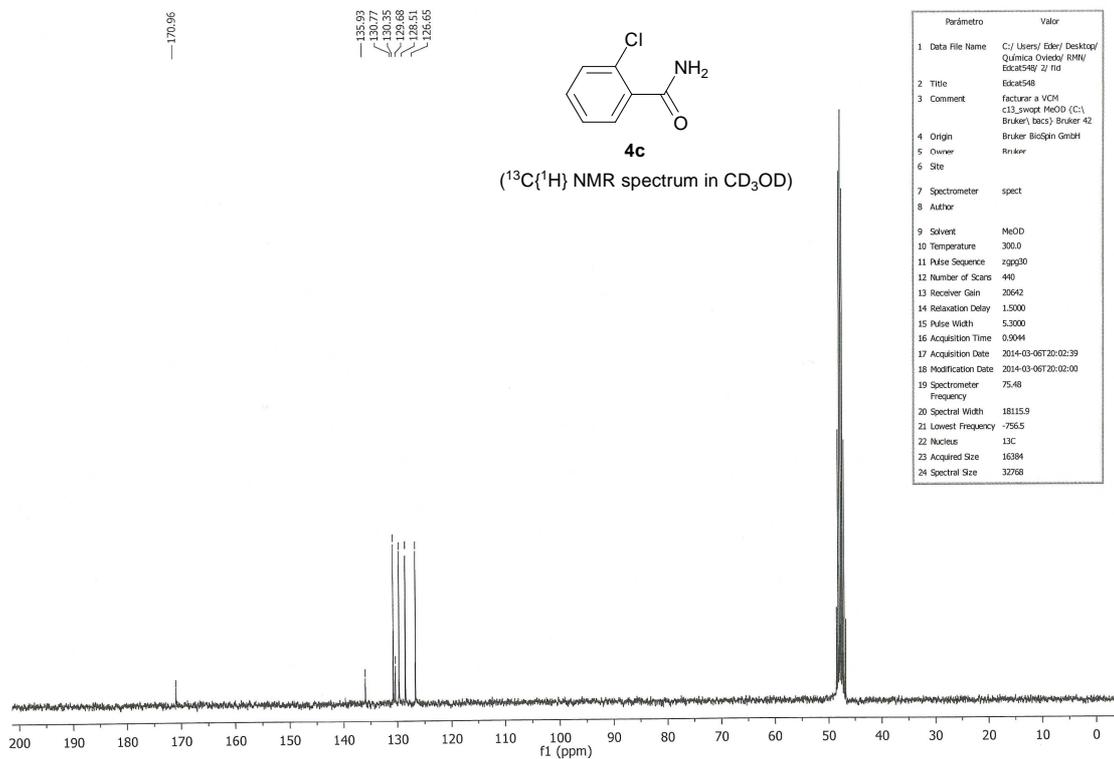
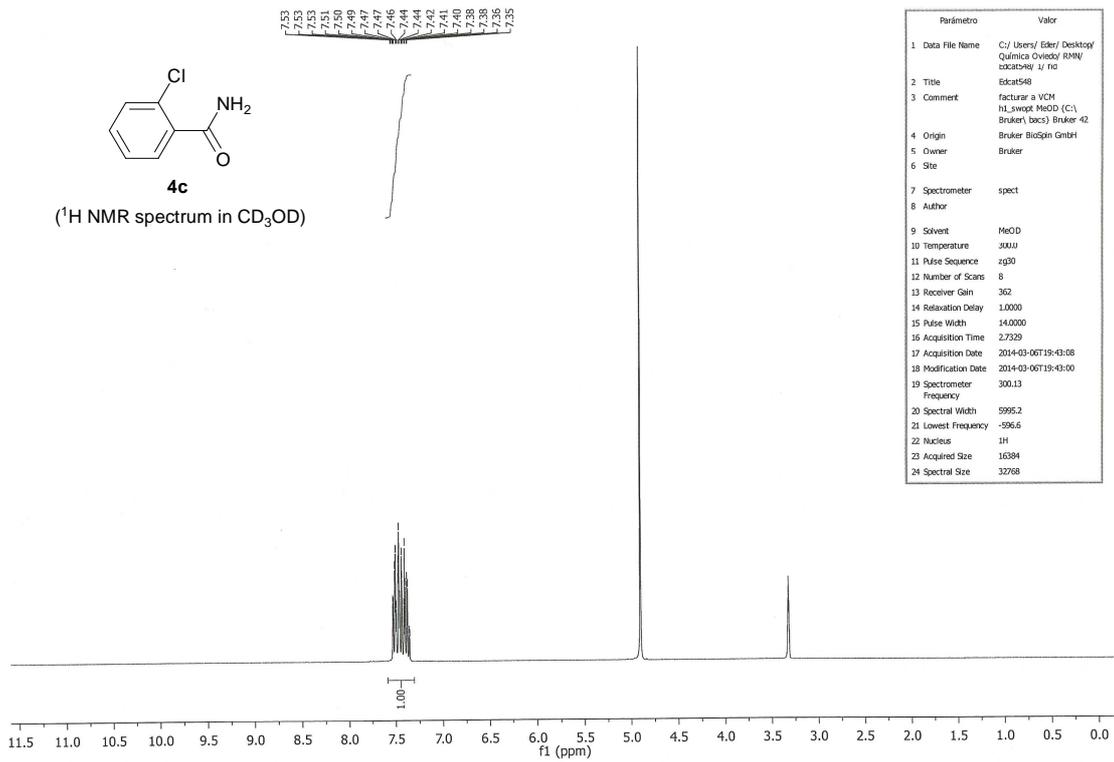


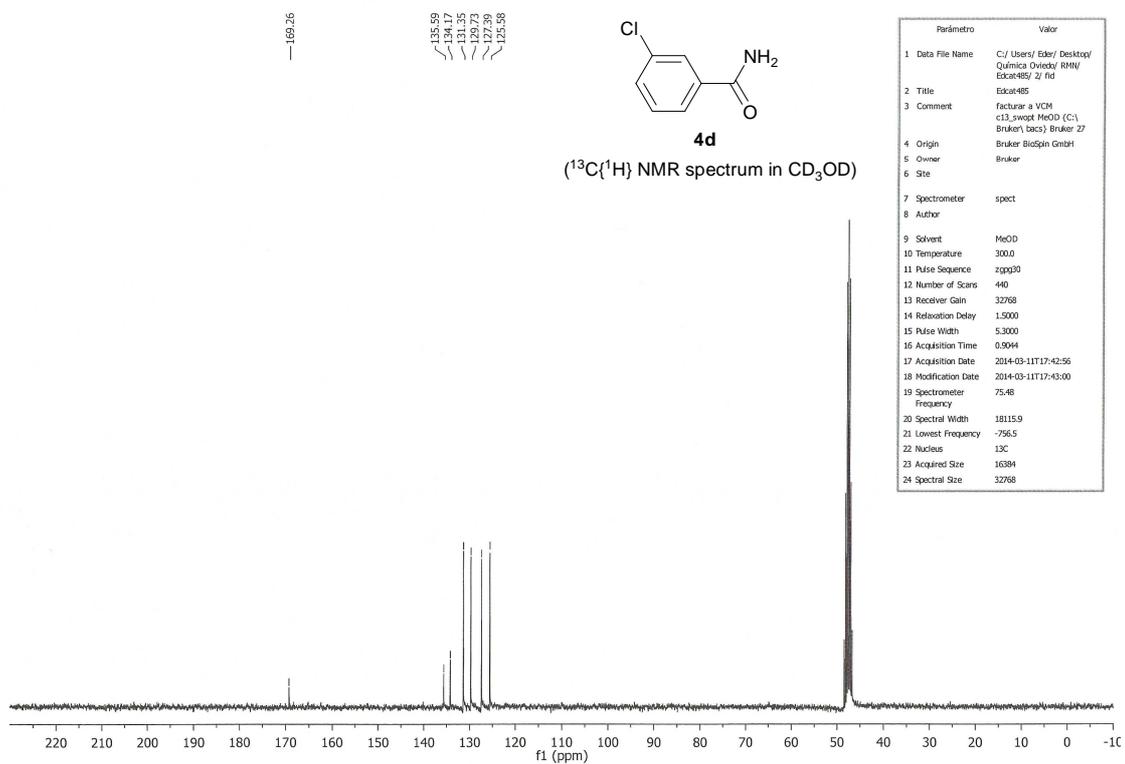
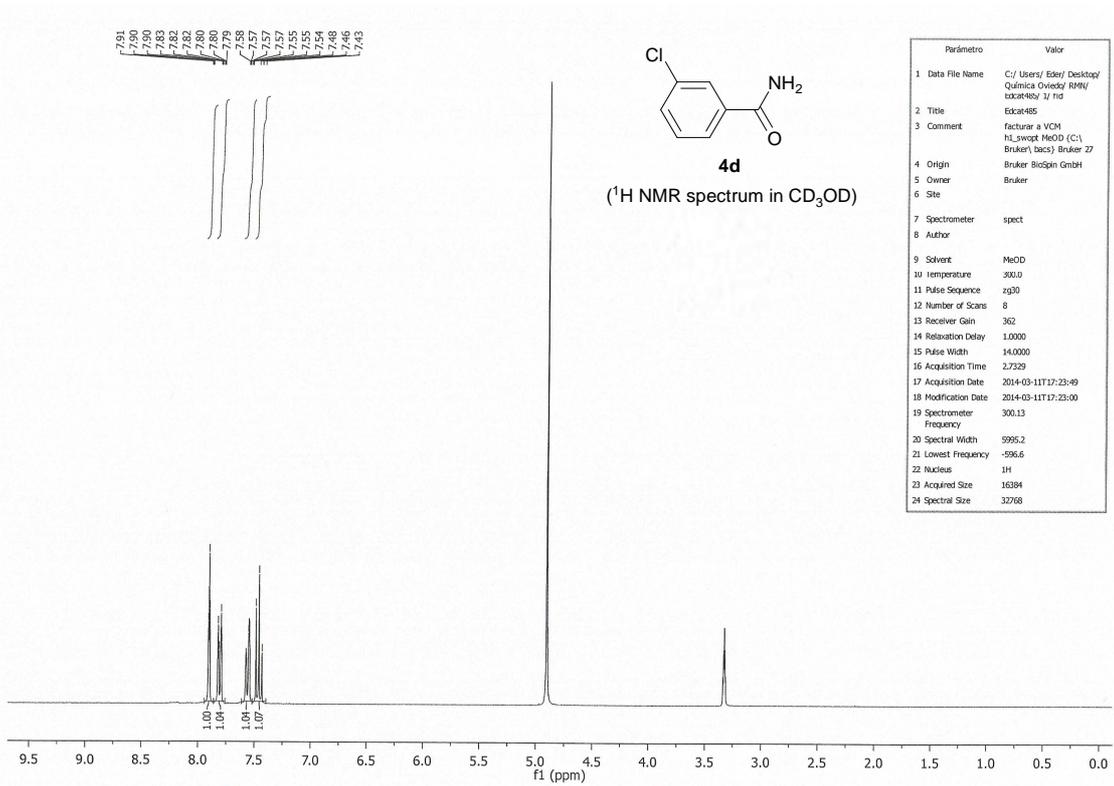
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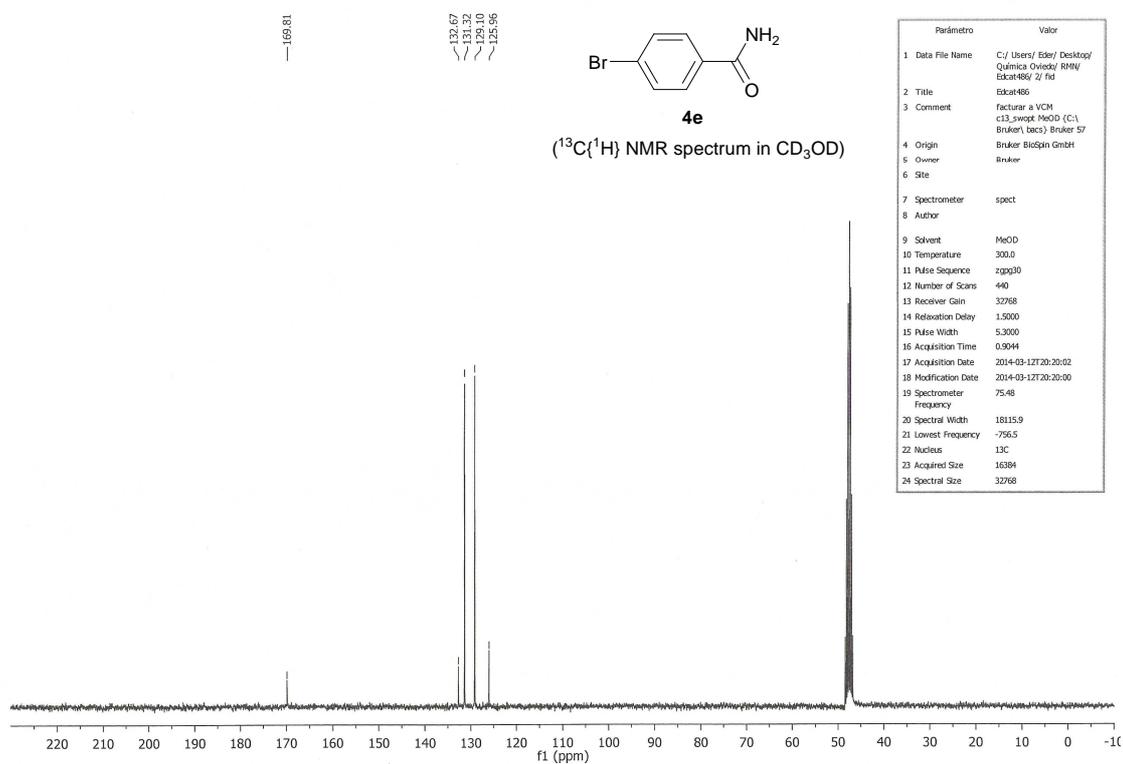
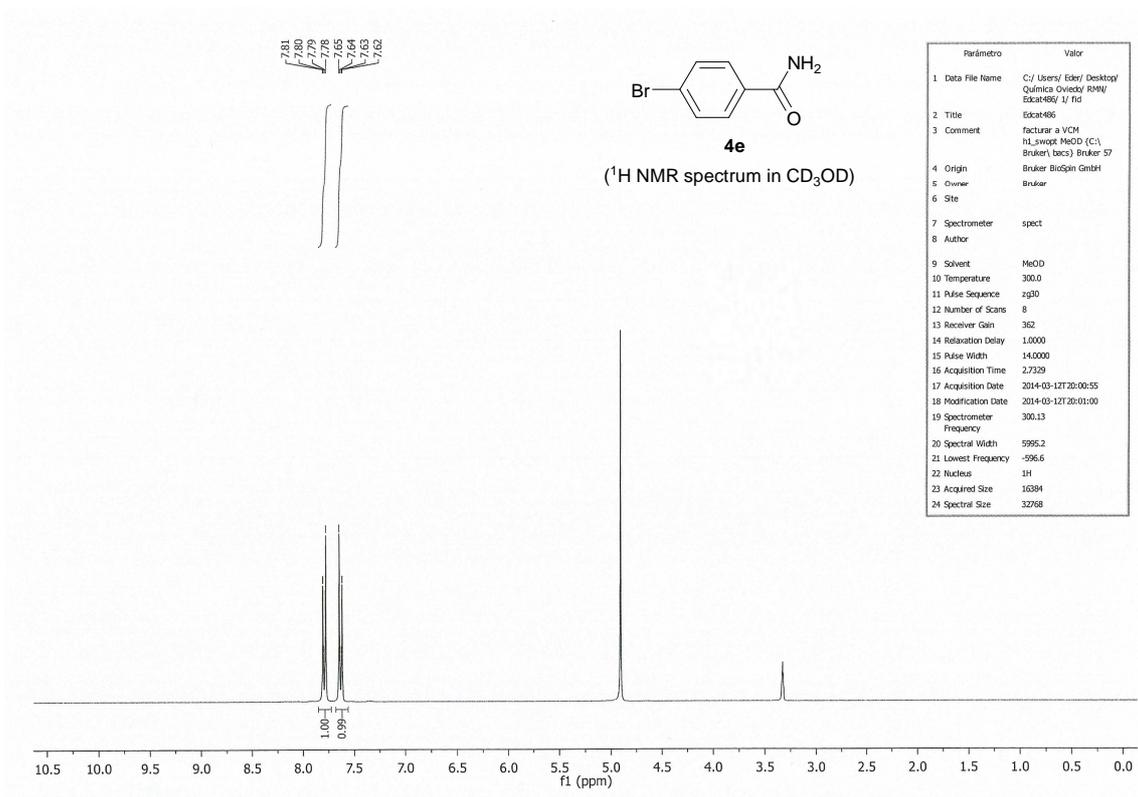


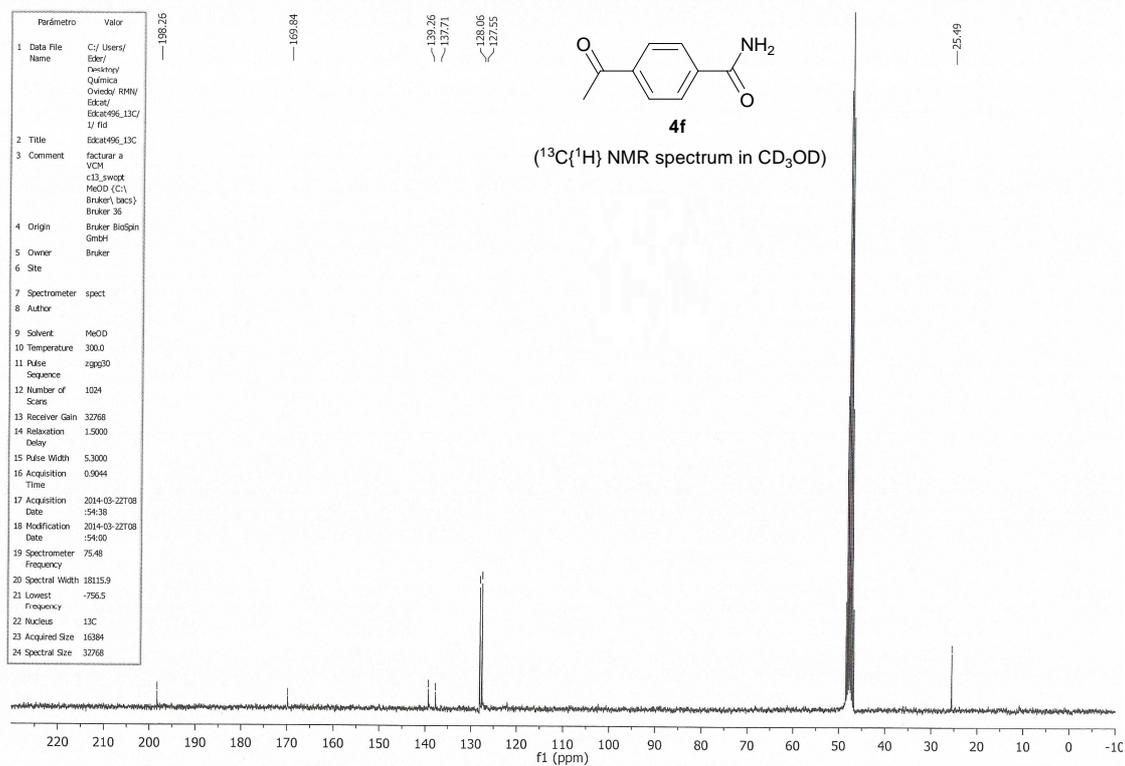
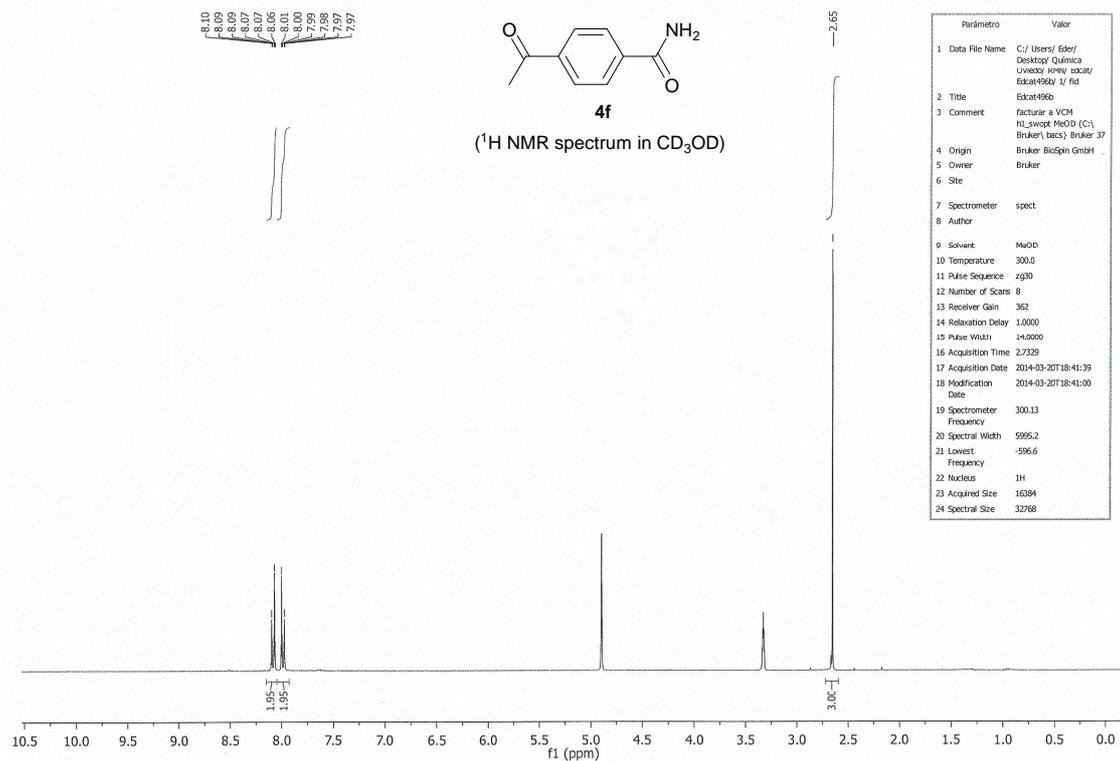
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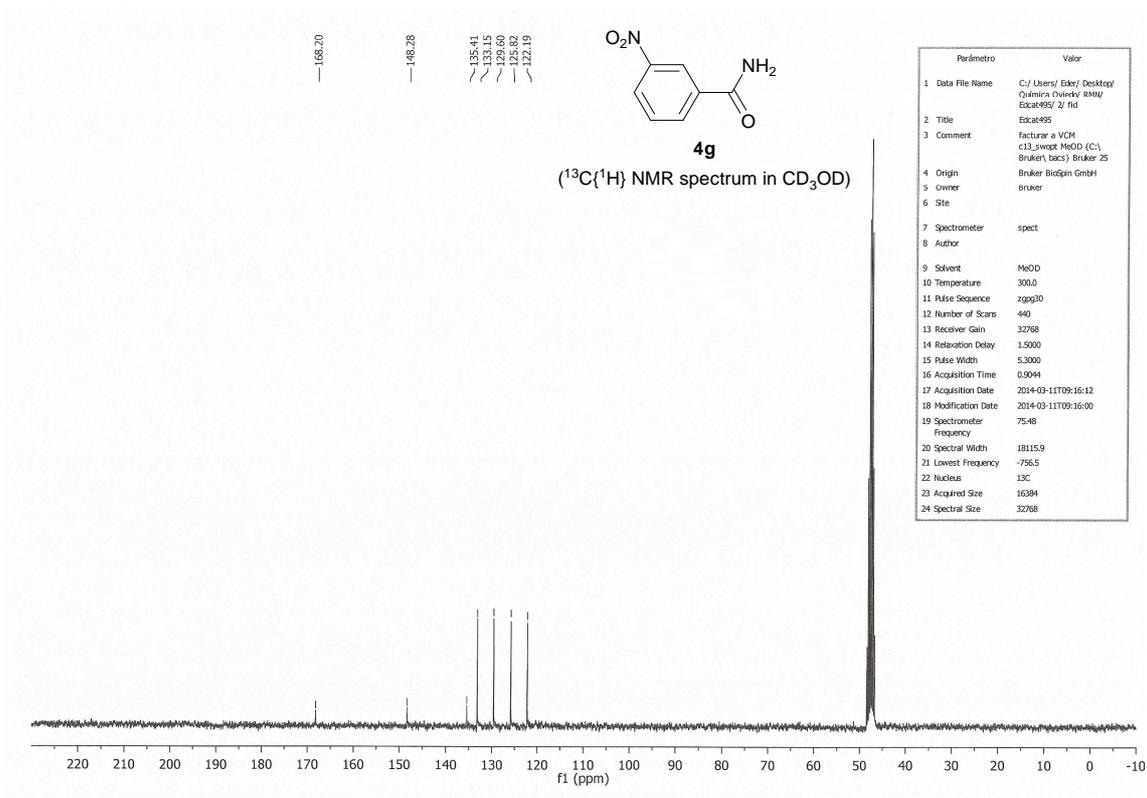
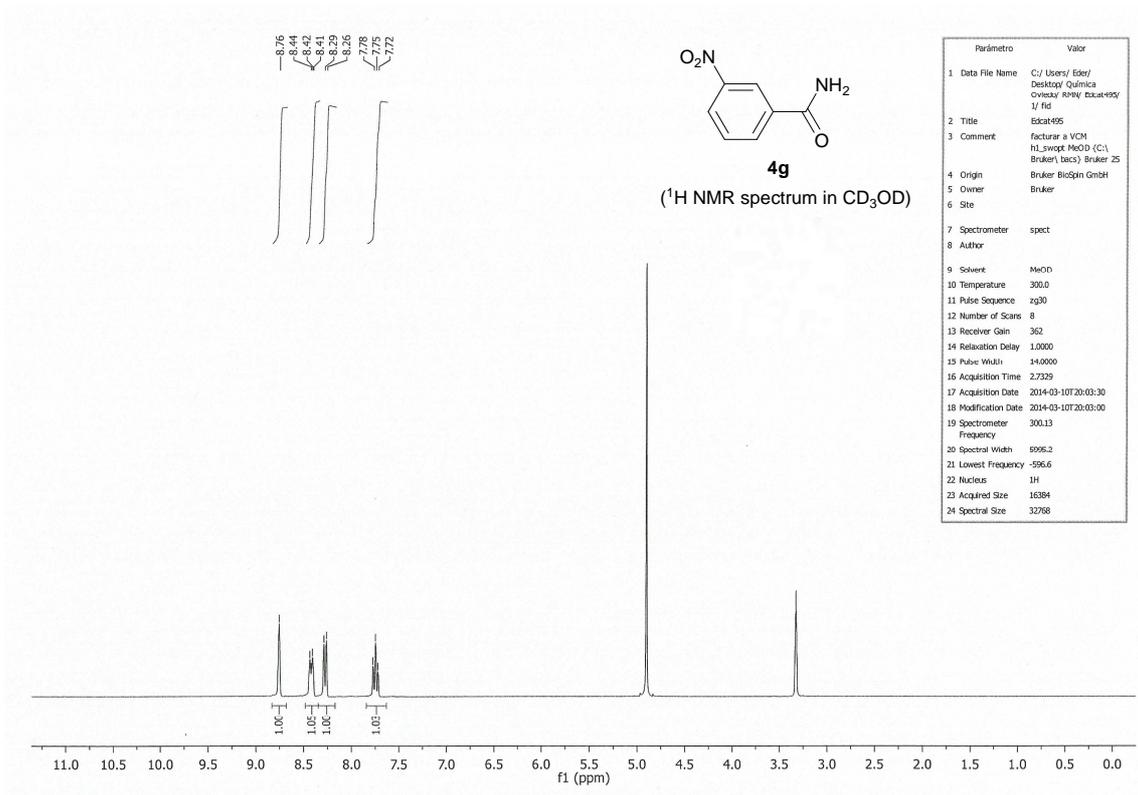
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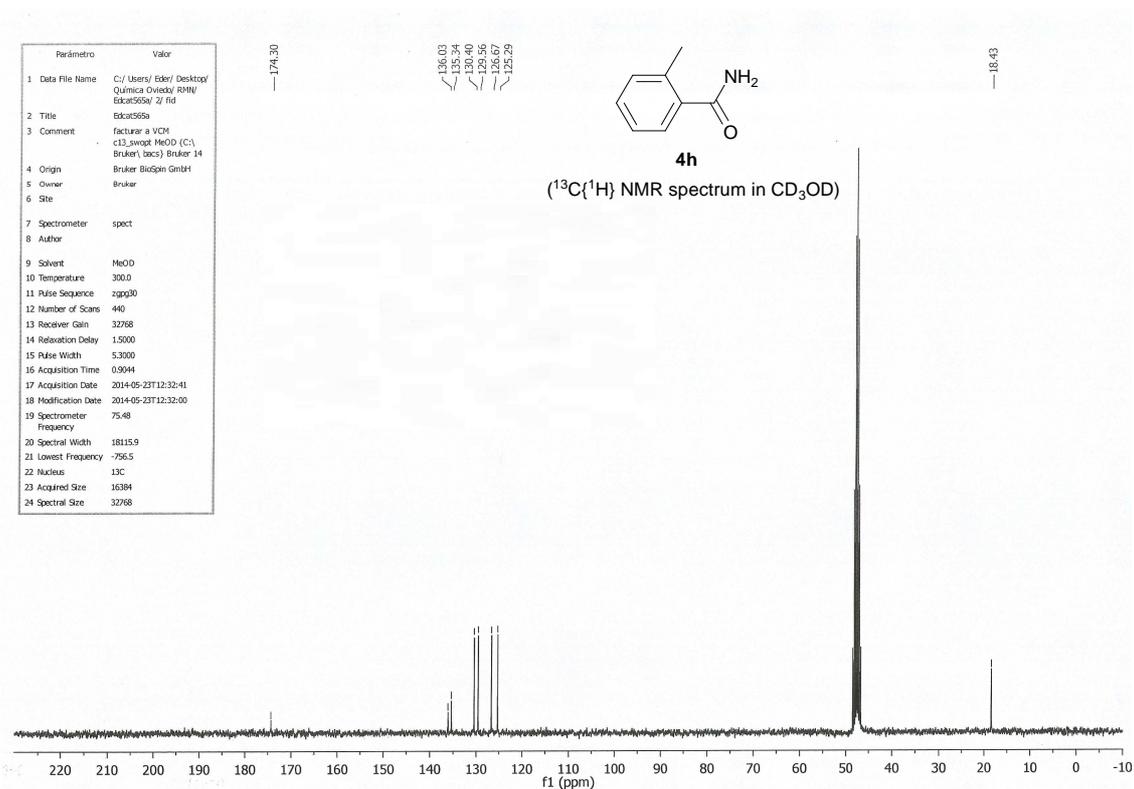
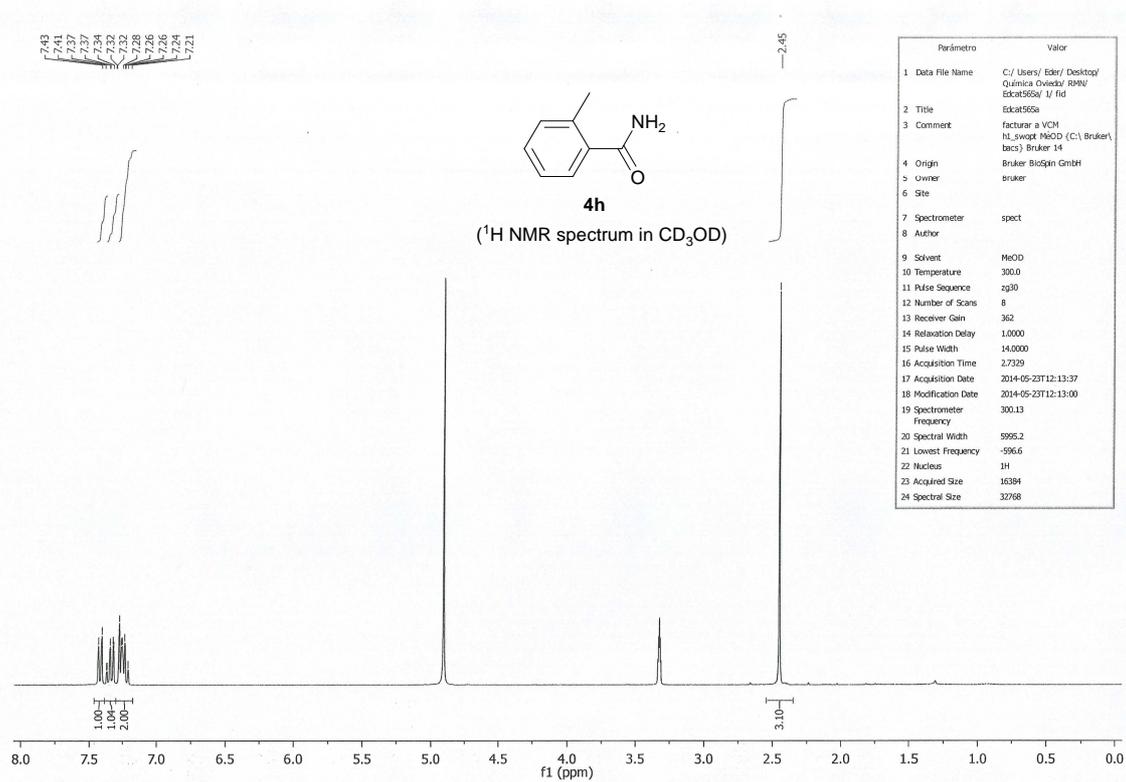


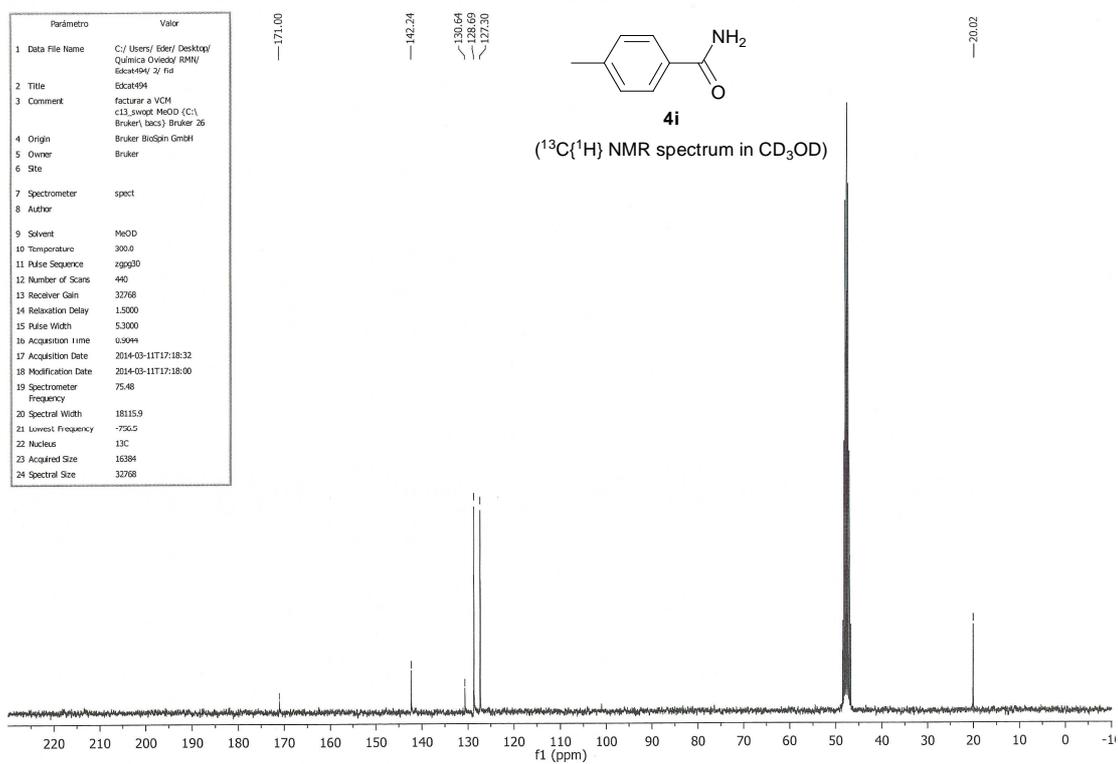
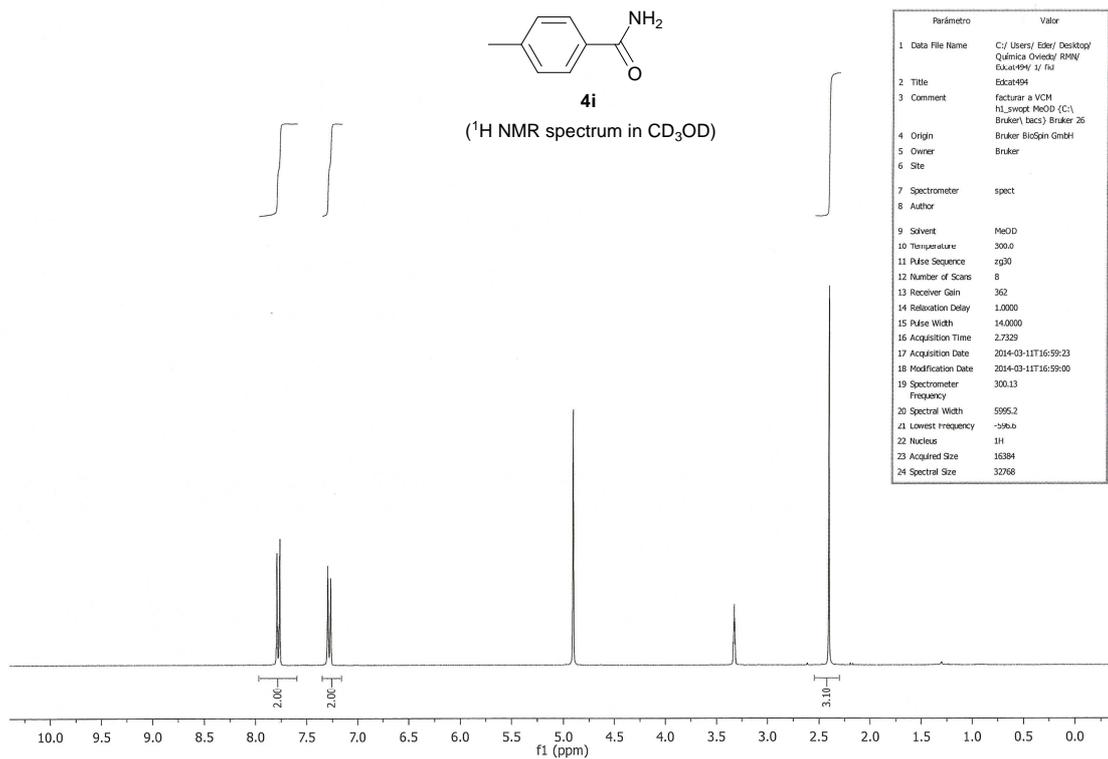


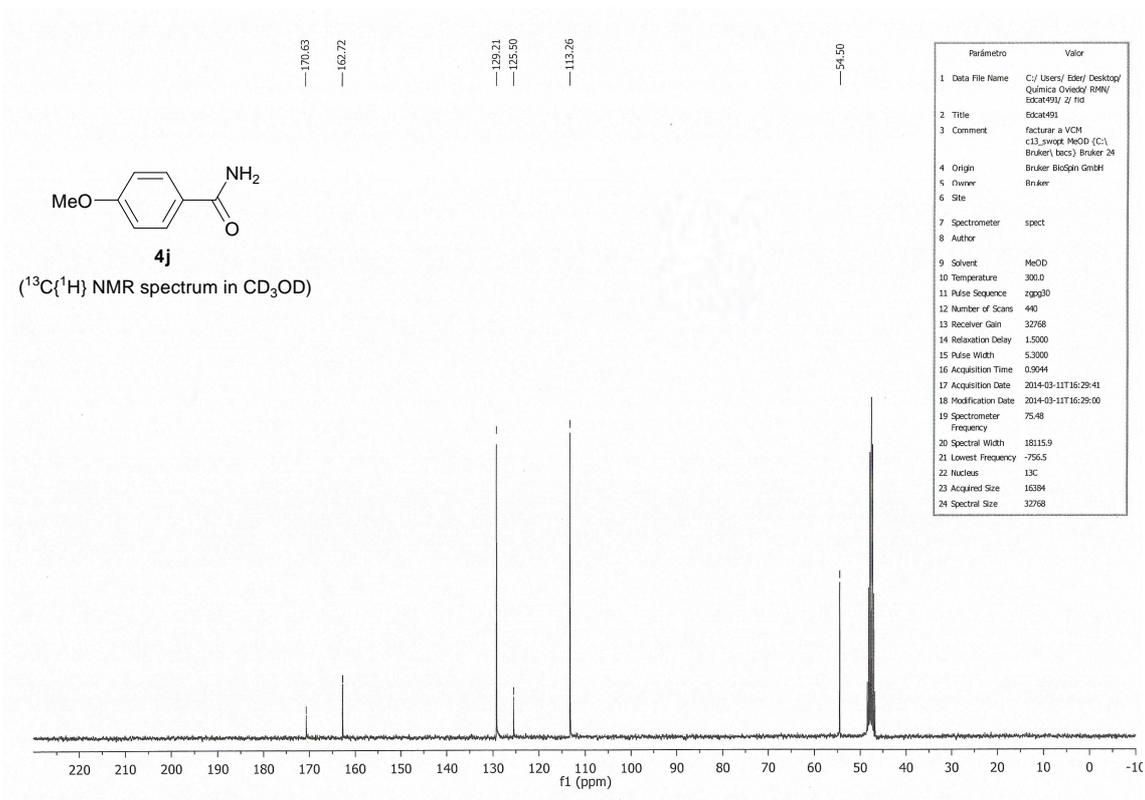
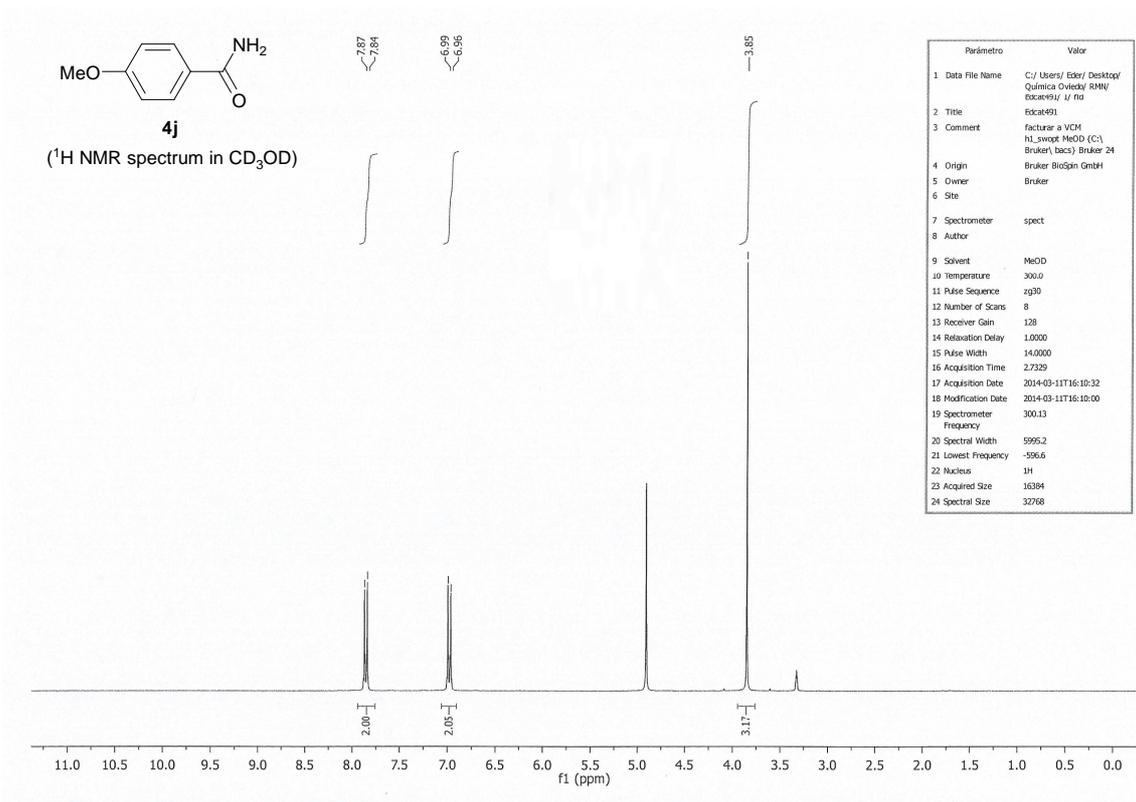


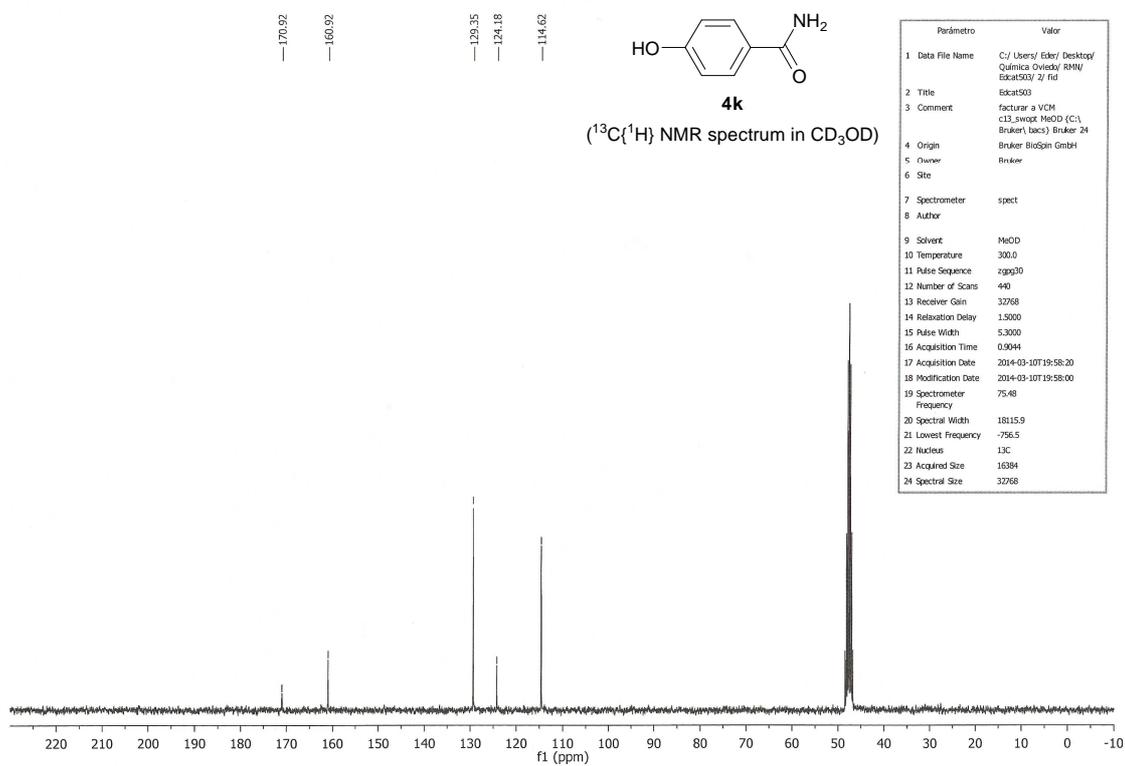
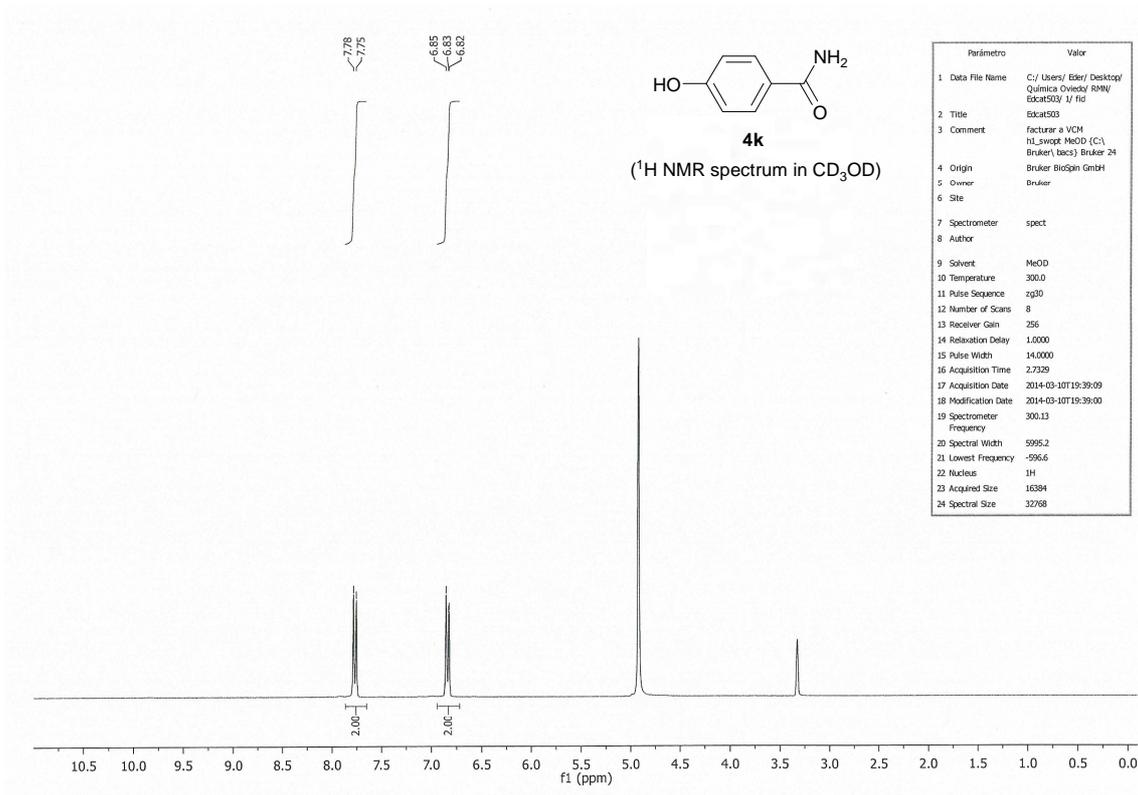




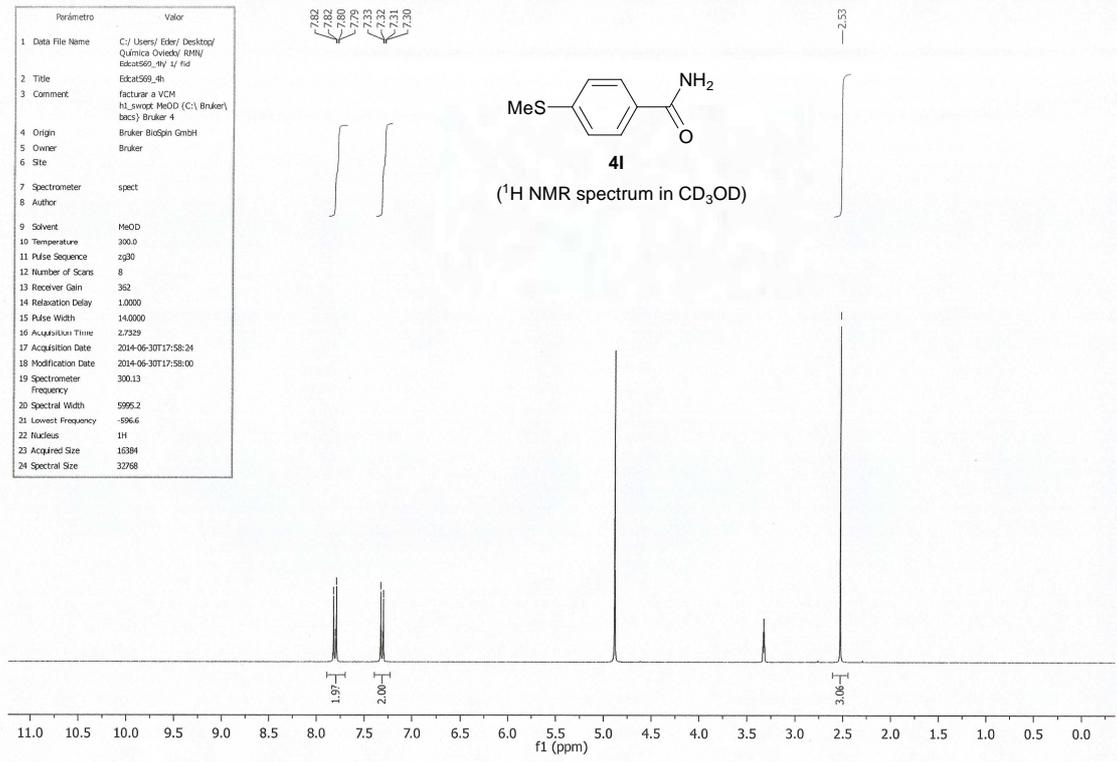




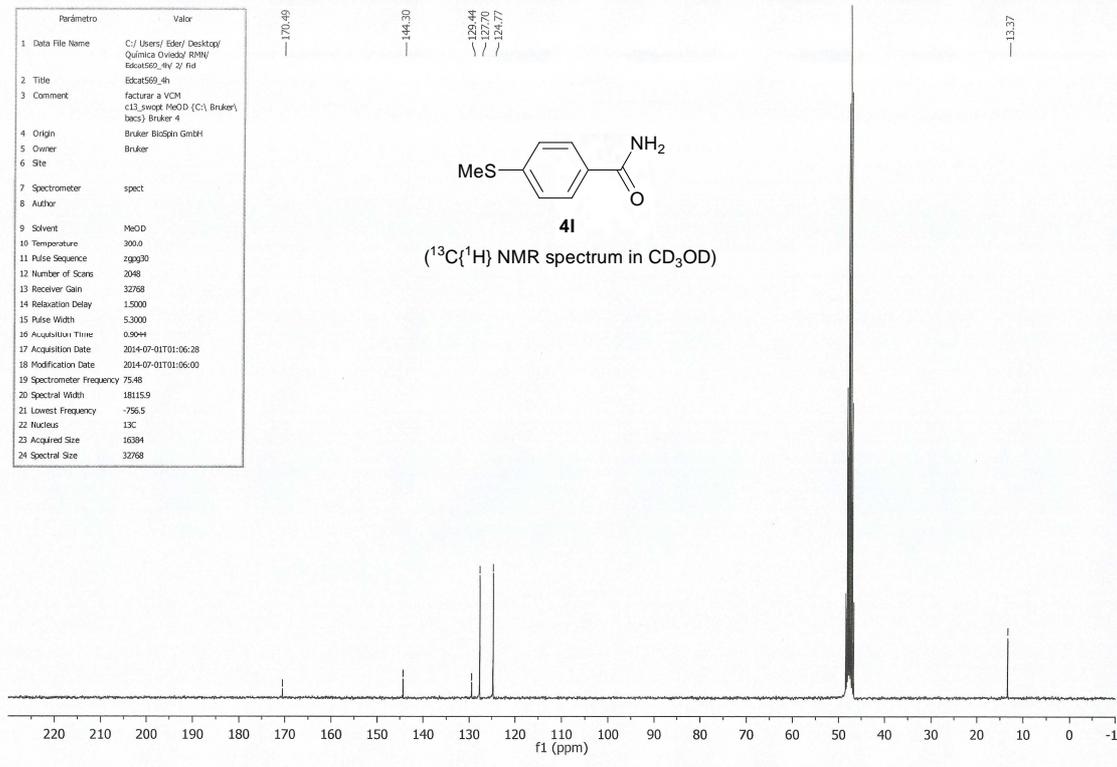


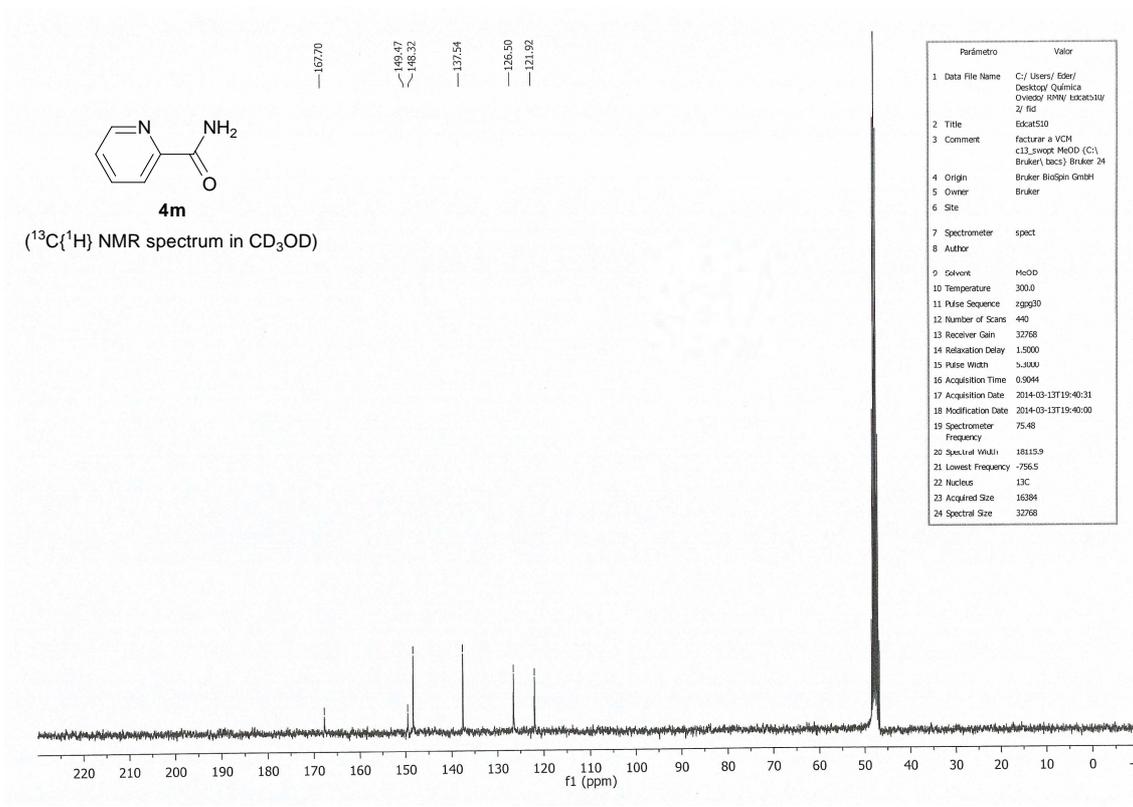
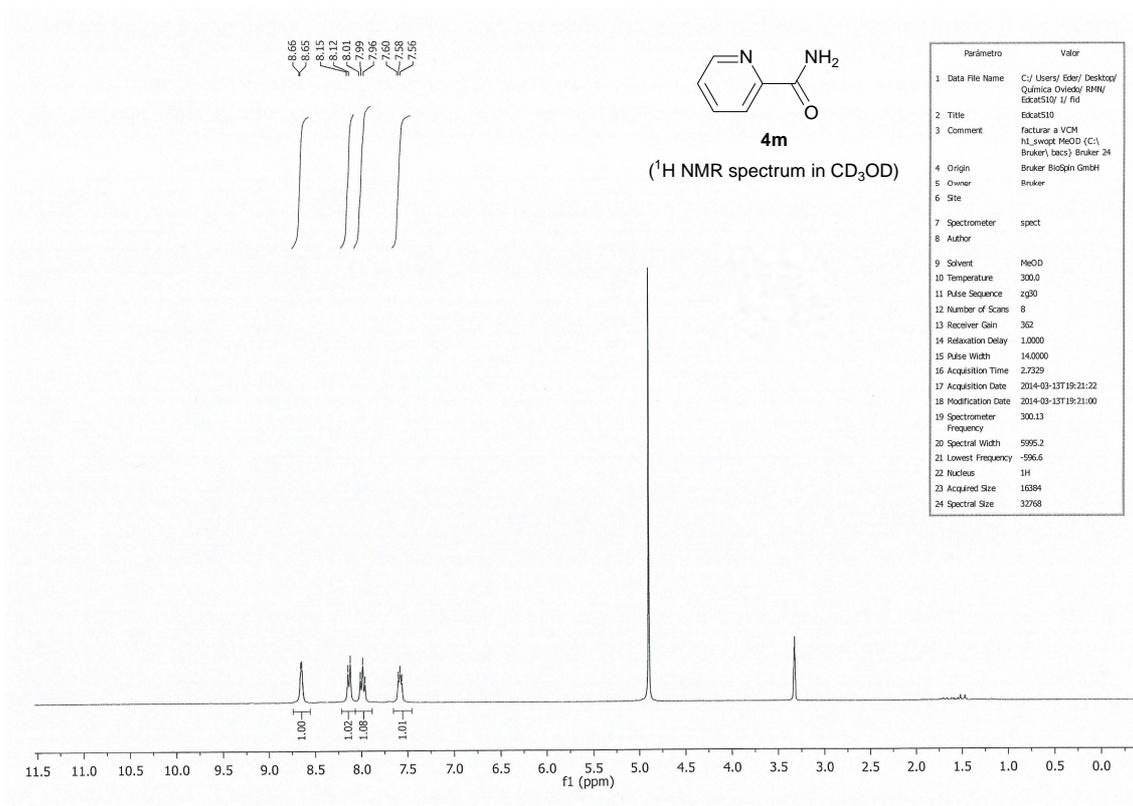


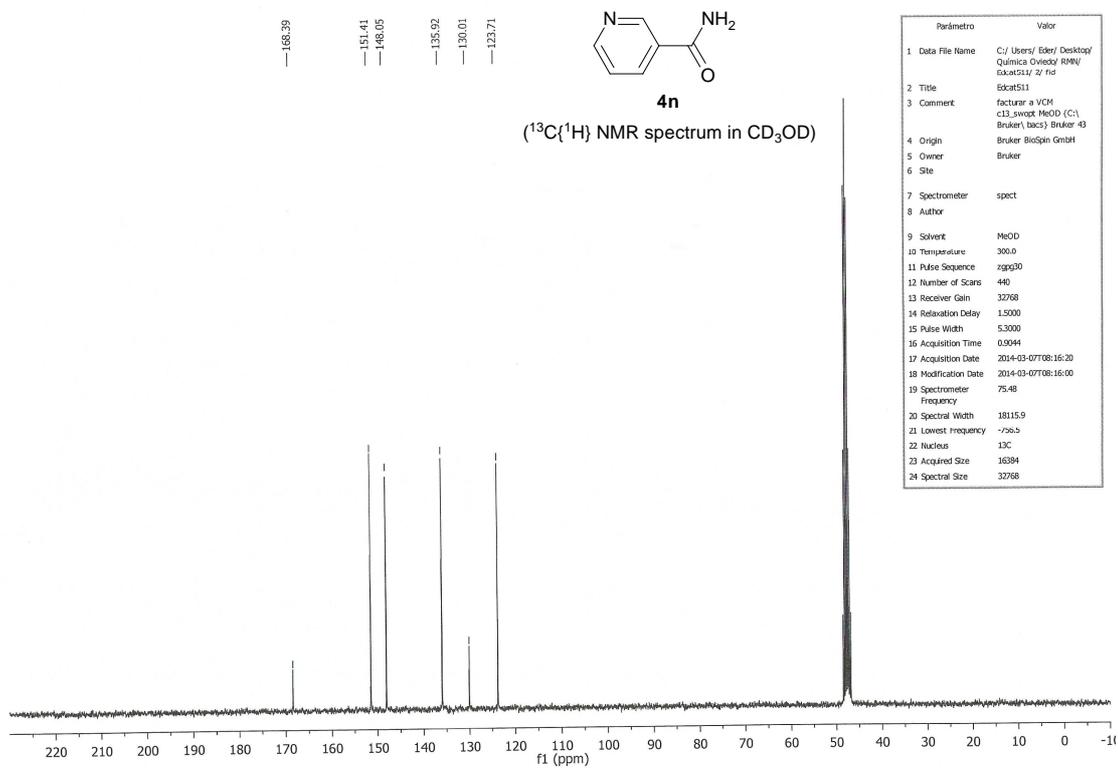
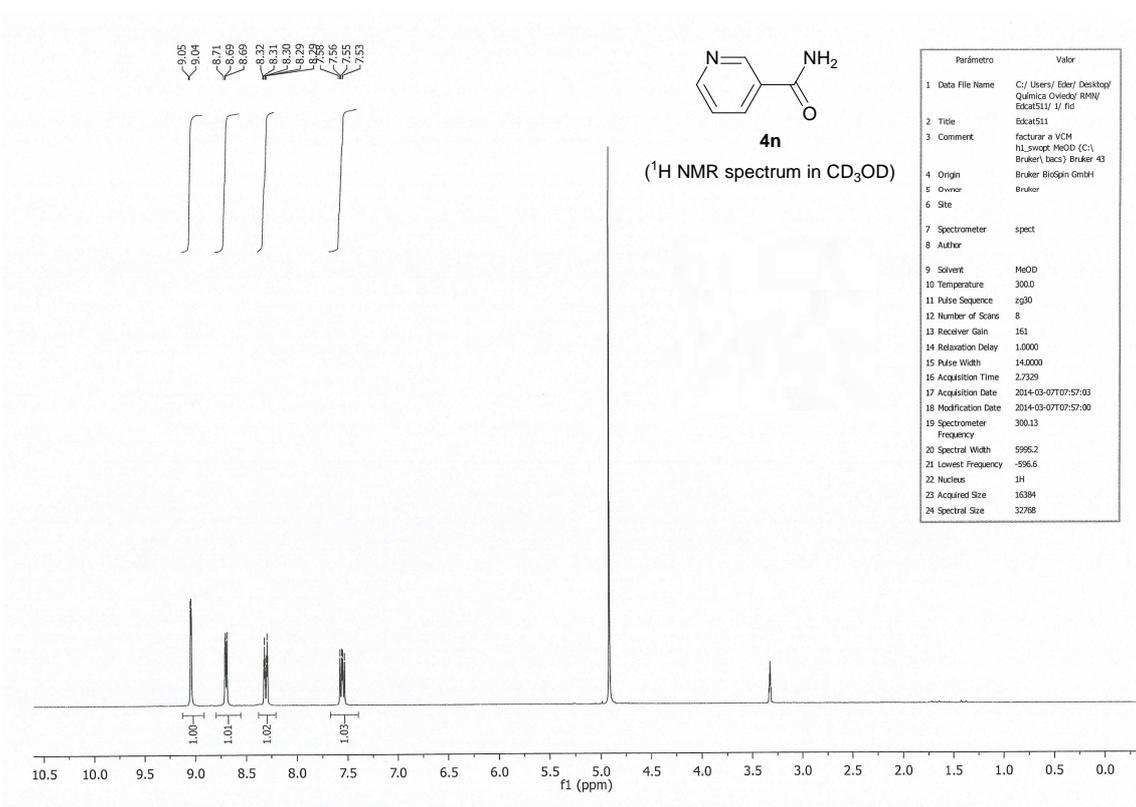
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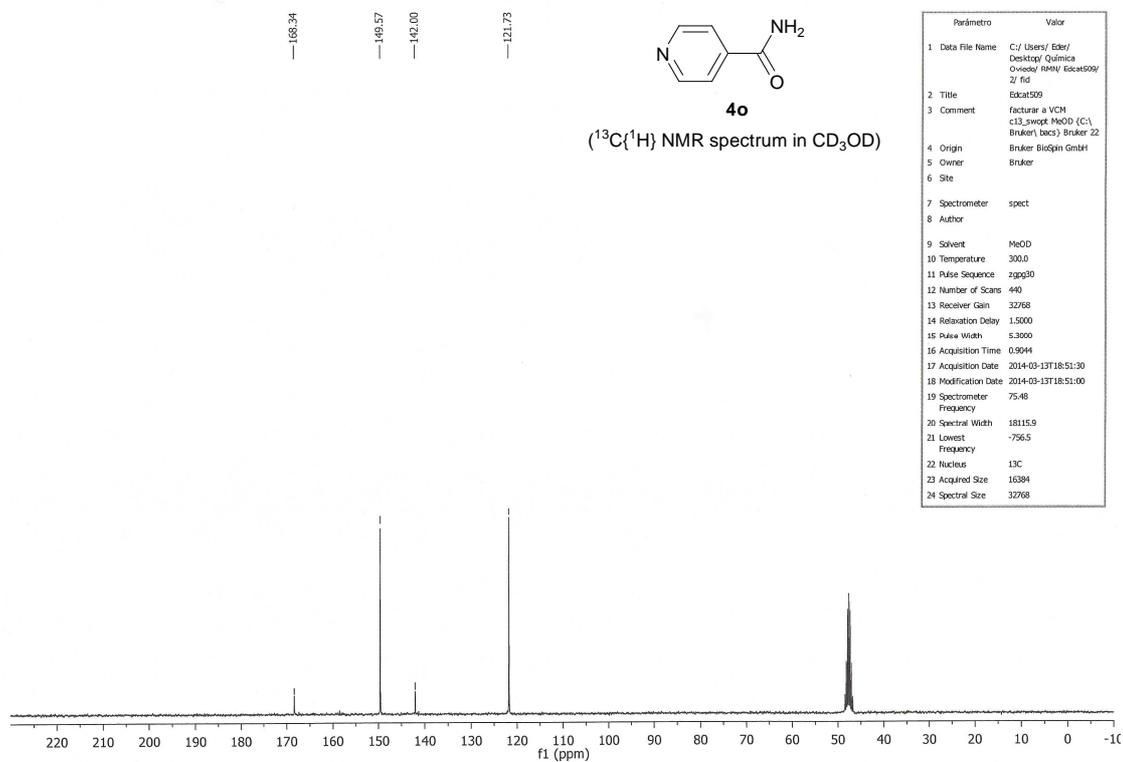
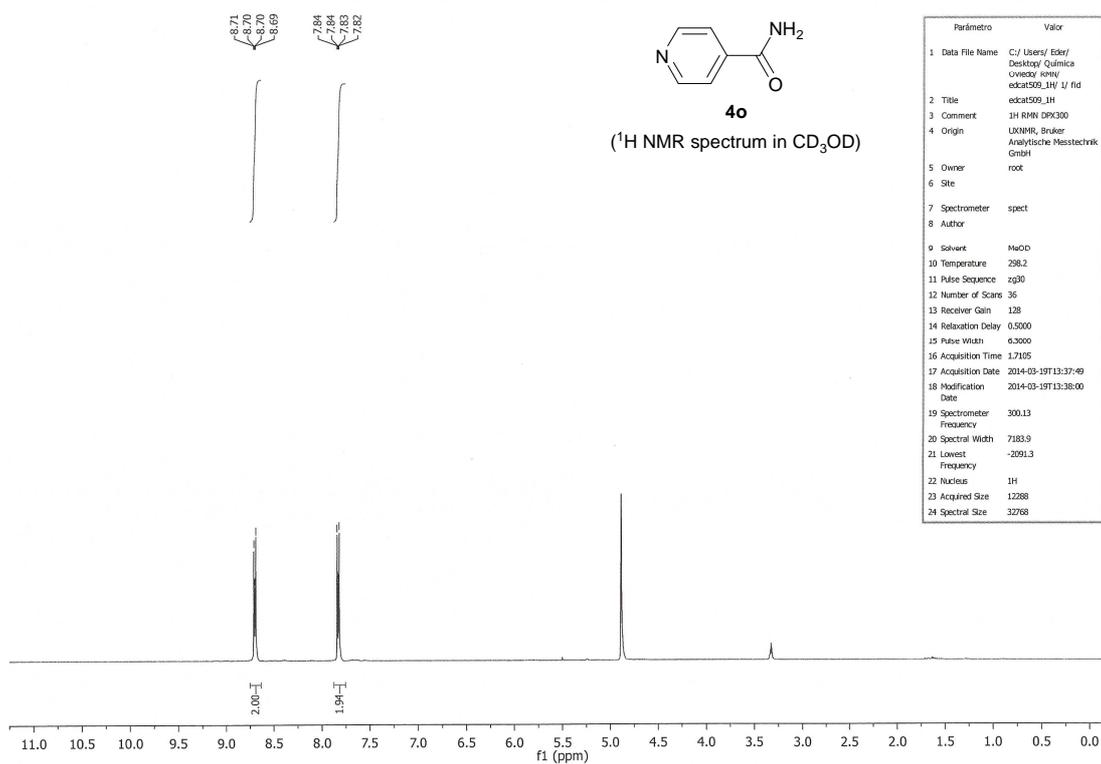


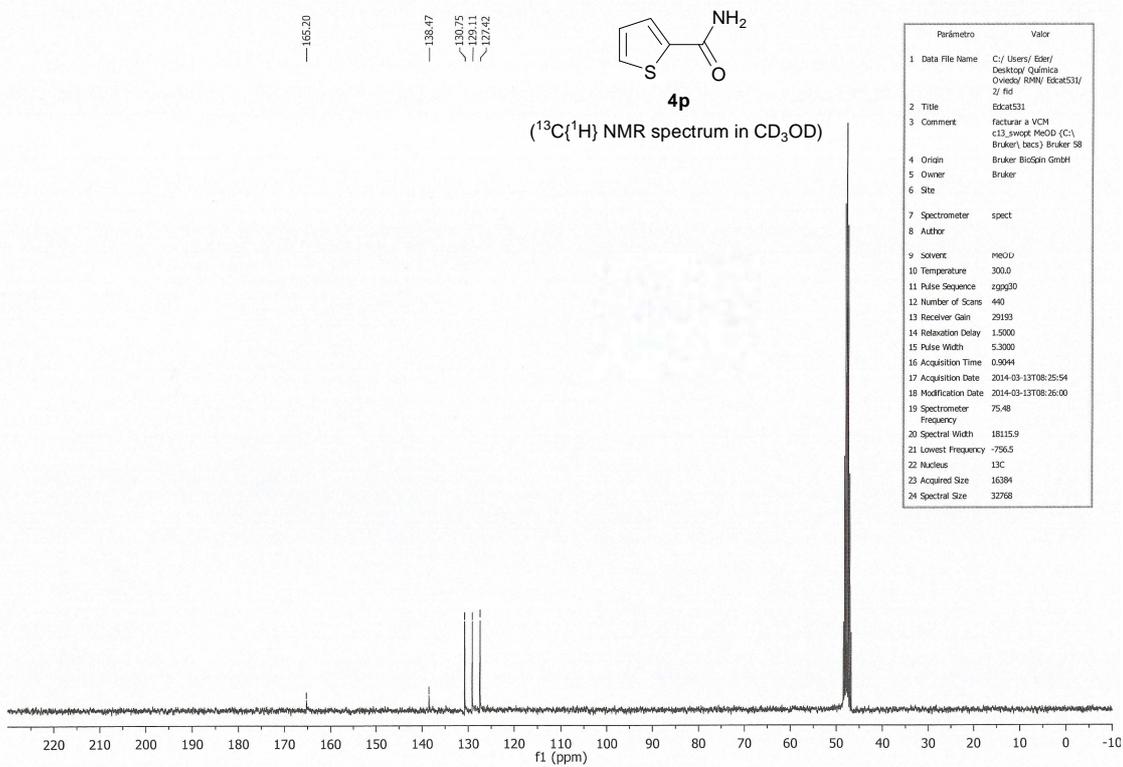
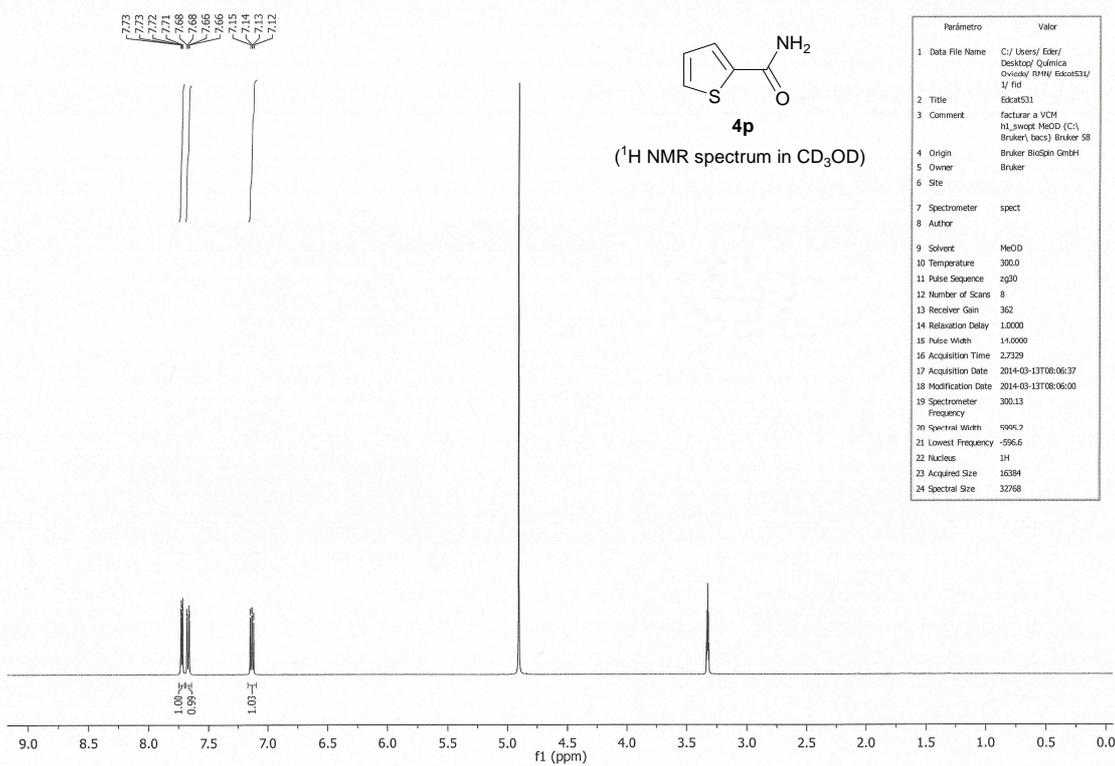
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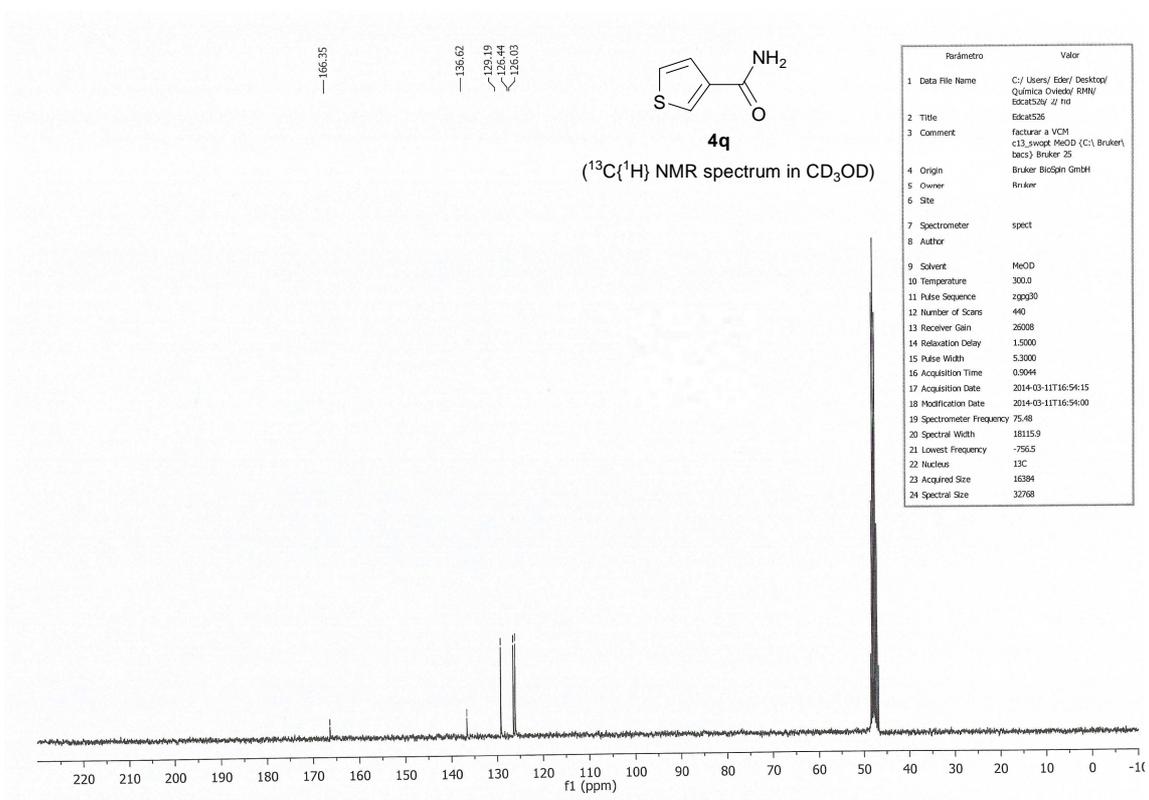
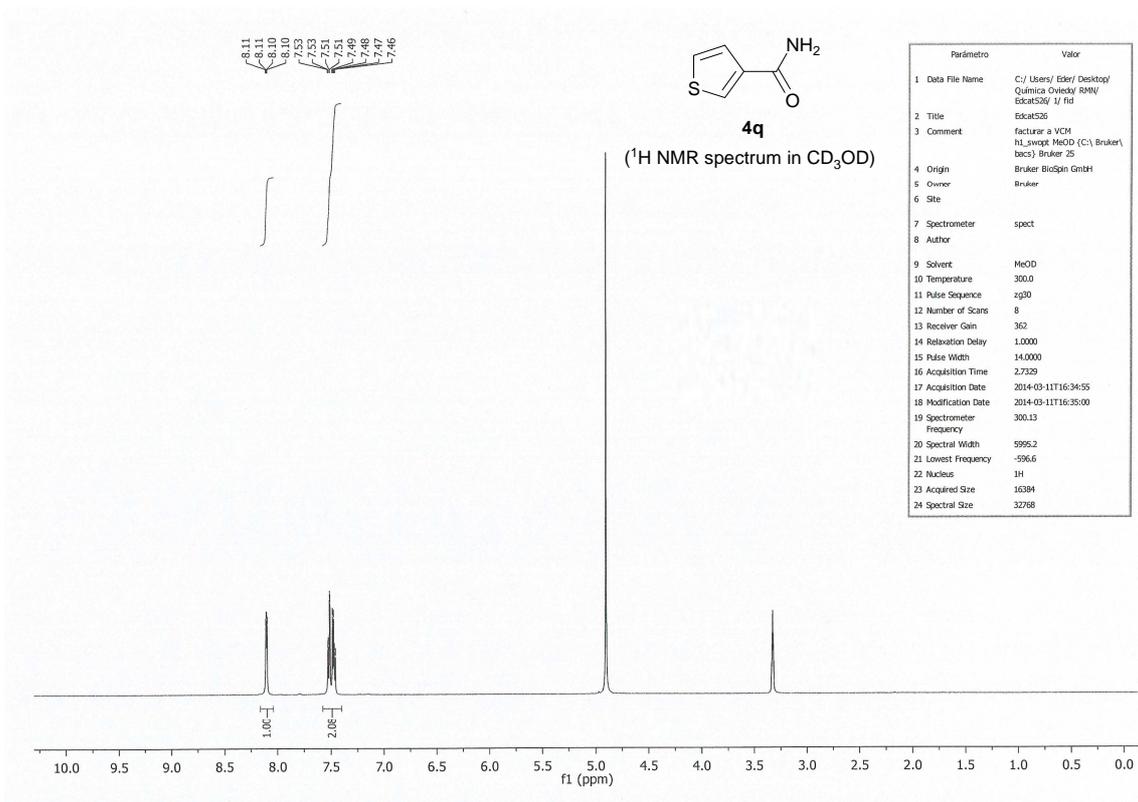




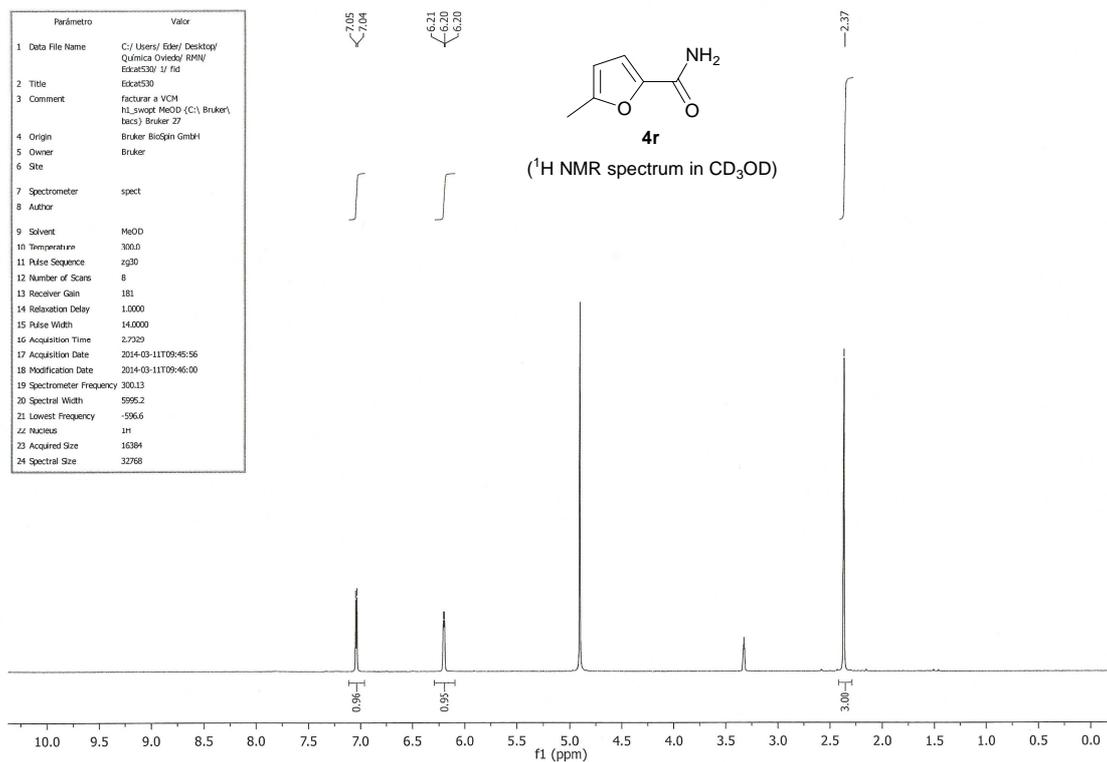




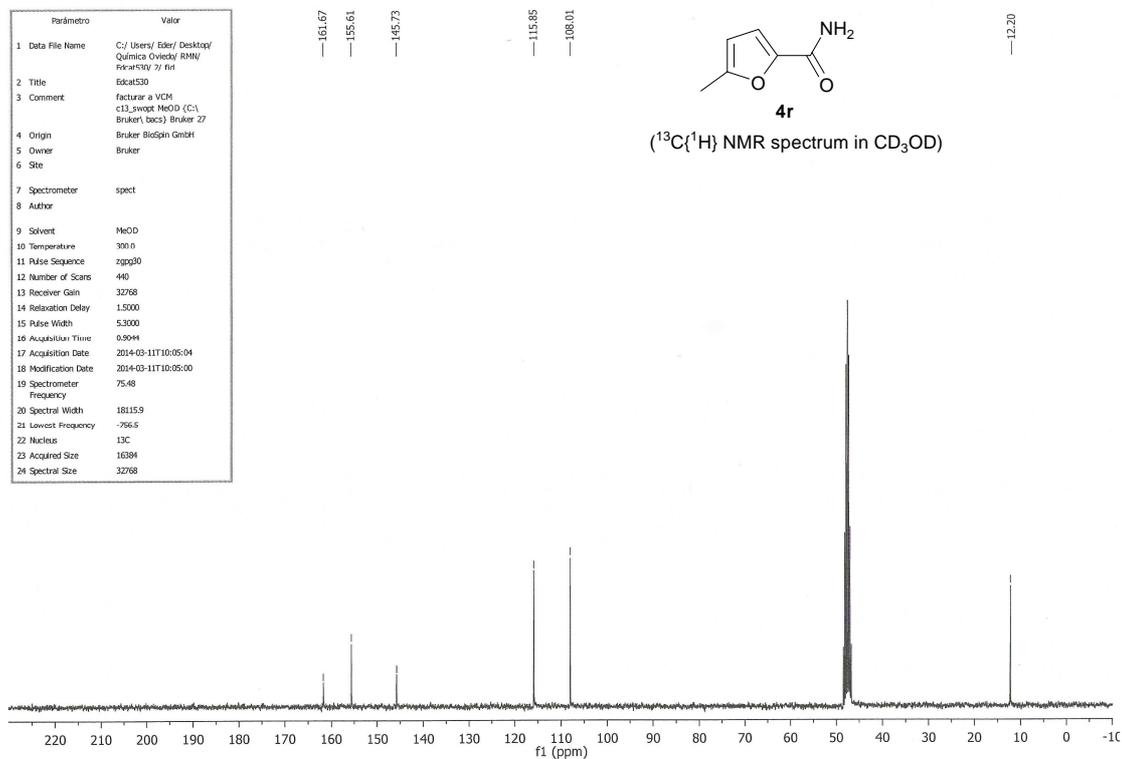


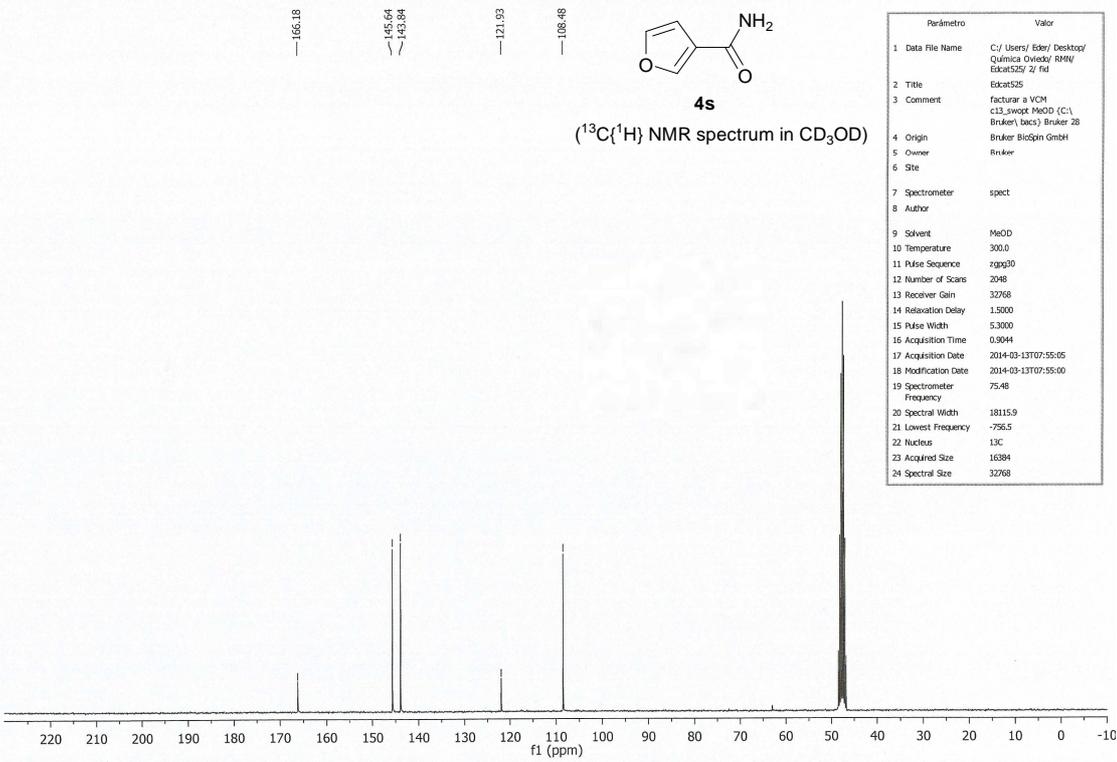
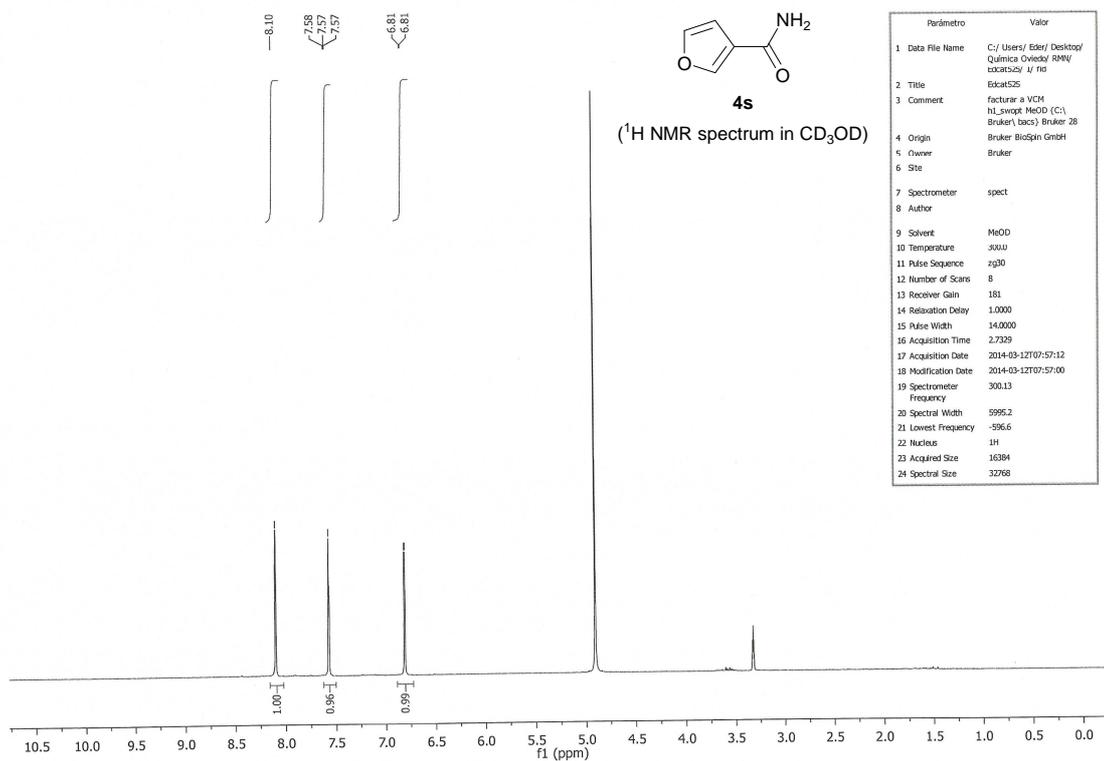


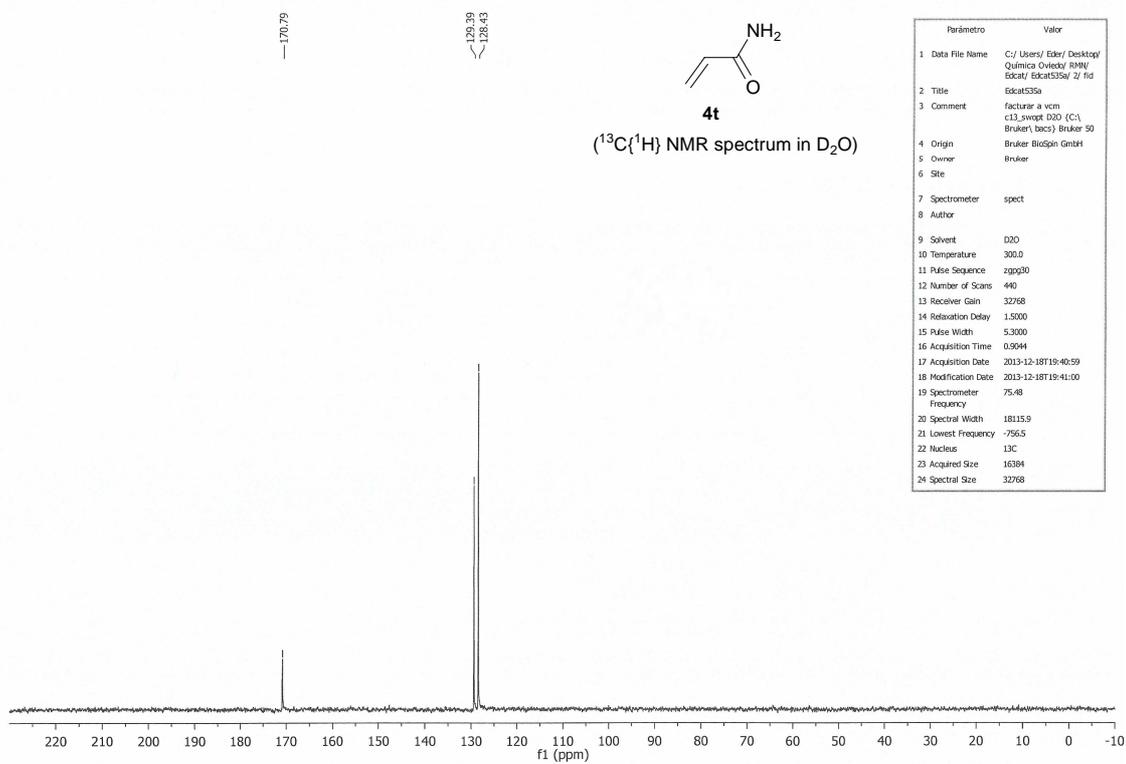
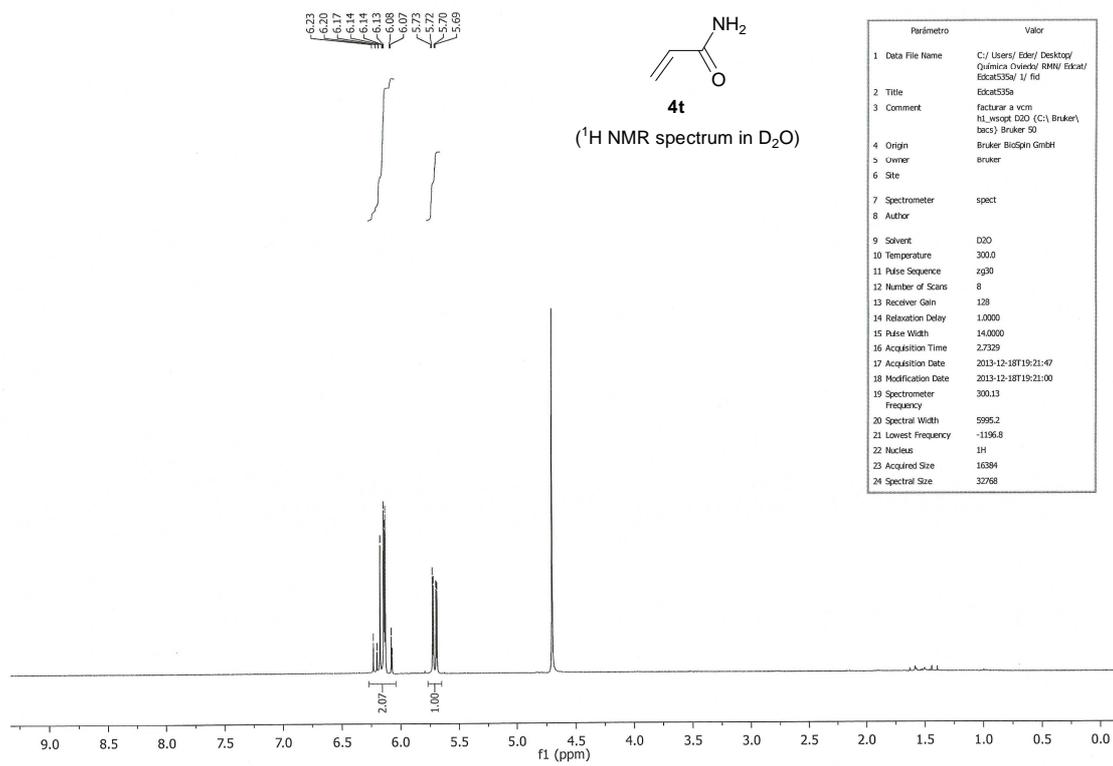
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1 Data File Name	C:/Users/Eder/Desktop/Química Ovecky RMN/Edcat530_1.fid
2 Title	Edcat530
3 Comment	facturar a VCM HL_swapt MeOD (C:\Bruker\hacs) Bruker Z7
4 Origin	Bruker Biospin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	MeOD
10 Temperature	300.0
11 Pulse Sequence	zg30
12 Number of Scans	8
13 Receiver Gain	181
14 Relaxation Delay	1.0000
15 Pulse Width	14.0000
16 Acquisition Time	2.7320
17 Acquisition Date	2014-03-11T09:45:56
18 Modification Date	2014-03-11T09:46:00
19 Spectrometer Frequency	300.13
20 Spectral Width	5996.2
21 Lowest Frequency	-596.6
22 Nucleus	1H
23 Acquired Size	16384
24 Spectral Size	32768



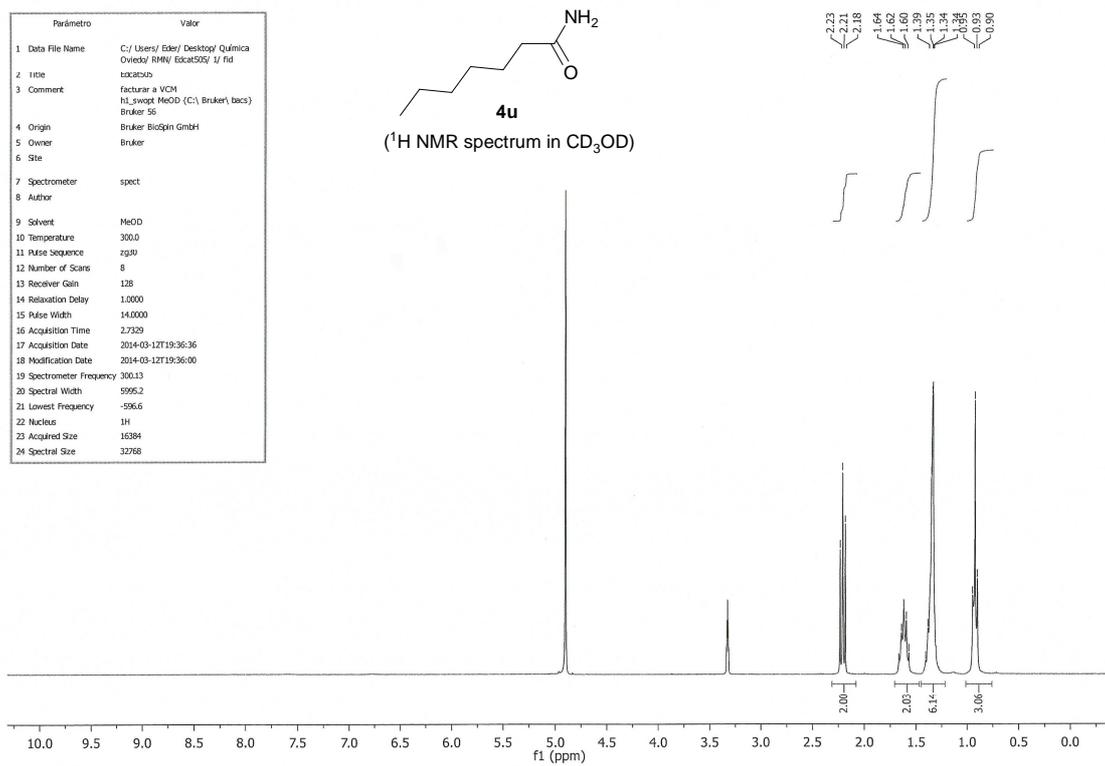
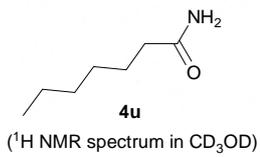
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1 Data File Name	C:/Users/Eder/Desktop/Química Ovecky RMN/Edcat530_2.fid
2 Title	Edcat530
3 Comment	facturar a VCM c13_swapt MeOD (C:\Bruker\hacs) Bruker Z7
4 Origin	Bruker Biospin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	MeOD
10 Temperature	300.0
11 Pulse Sequence	zgpg30
12 Number of Scans	400
13 Receiver Gain	32768
14 Relaxation Delay	1.5000
15 Pulse Width	5.3000
16 Acquisition Time	0.9094
17 Acquisition Date	2014-03-11T10:05:04
18 Modification Date	2014-03-11T10:05:00
19 Spectrometer Frequency	75.48
20 Spectral Width	18115.9
21 Lowest Frequency	-786.5
22 Nucleus	13C
23 Acquired Size	16384
24 Spectral Size	32768



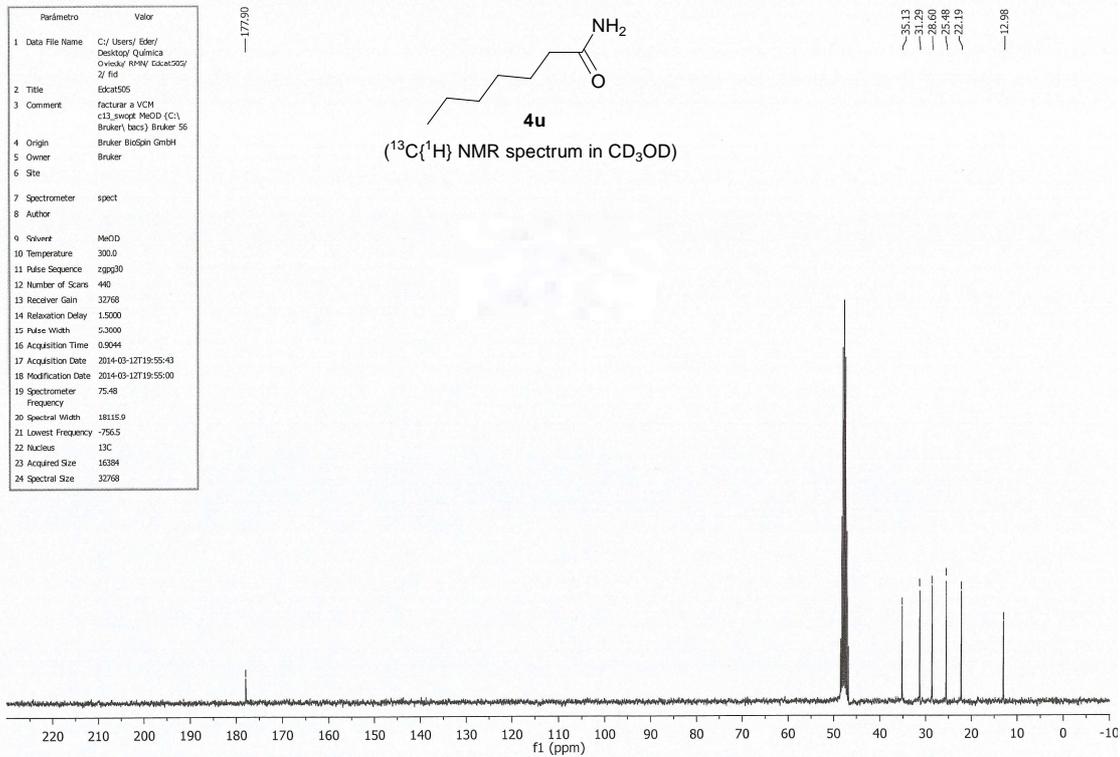
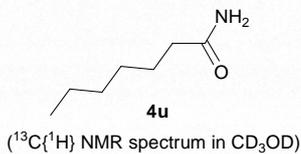




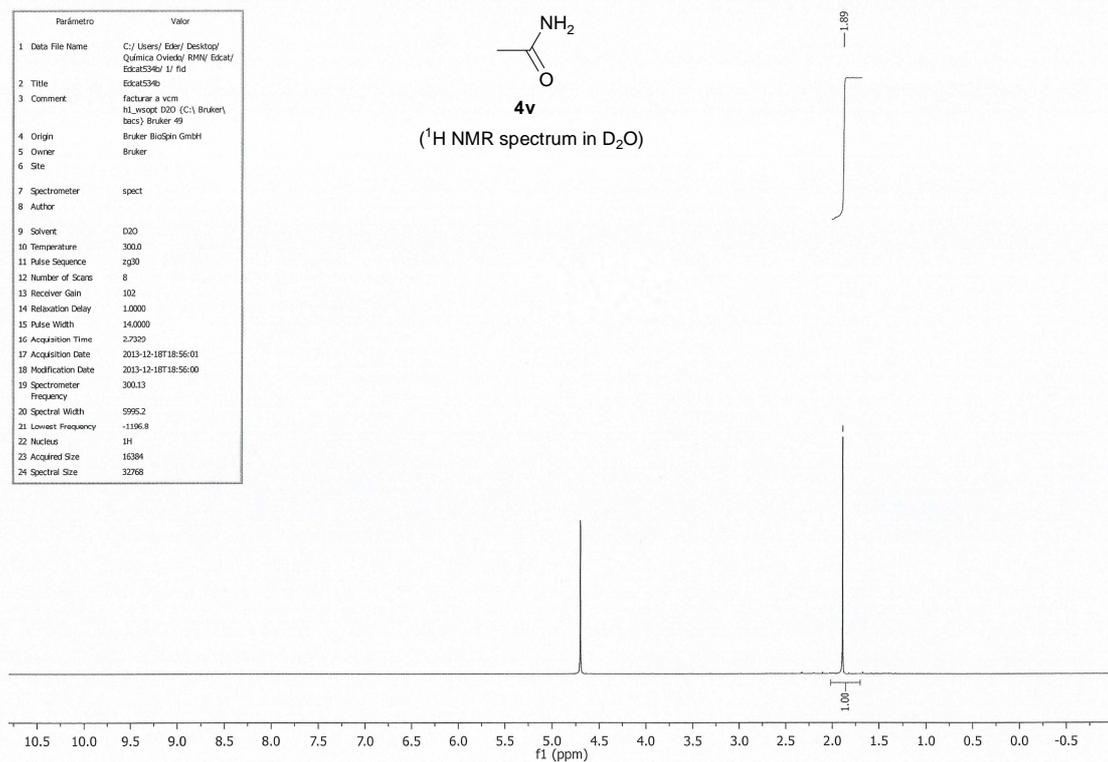
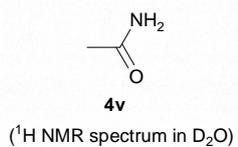
Parámetro	Valor
1 Data File Name	C:/Users/Eder/Desktop/Química/Oviedo/RMW/Escat505/1/ f1d
2 Title	Escat505
3 Comment	facturar a VCM h1_swopt MeOD (C), Bruker(bacs) Bruker 56
4 Origin	Bruker Biospin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	MeOD
10 Temperature	300.0
11 Pulse Sequence	zgpg30
12 Number of Scans	8
13 Receiver Gain	128
14 Relaxation Delay	1.0000
15 Pulse Width	14.0000
16 Acquisition Time	2.7329
17 Acquisition Date	2014-03-12T19:36:36
18 Modification Date	2014-03-12T19:36:00
19 Spectrometer Frequency	300.13
20 Spectral Width	5995.2
21 Lowest Frequency	-596.6
22 Nucleus	¹ H
23 Acquired Size	16384
24 Spectral Size	32768



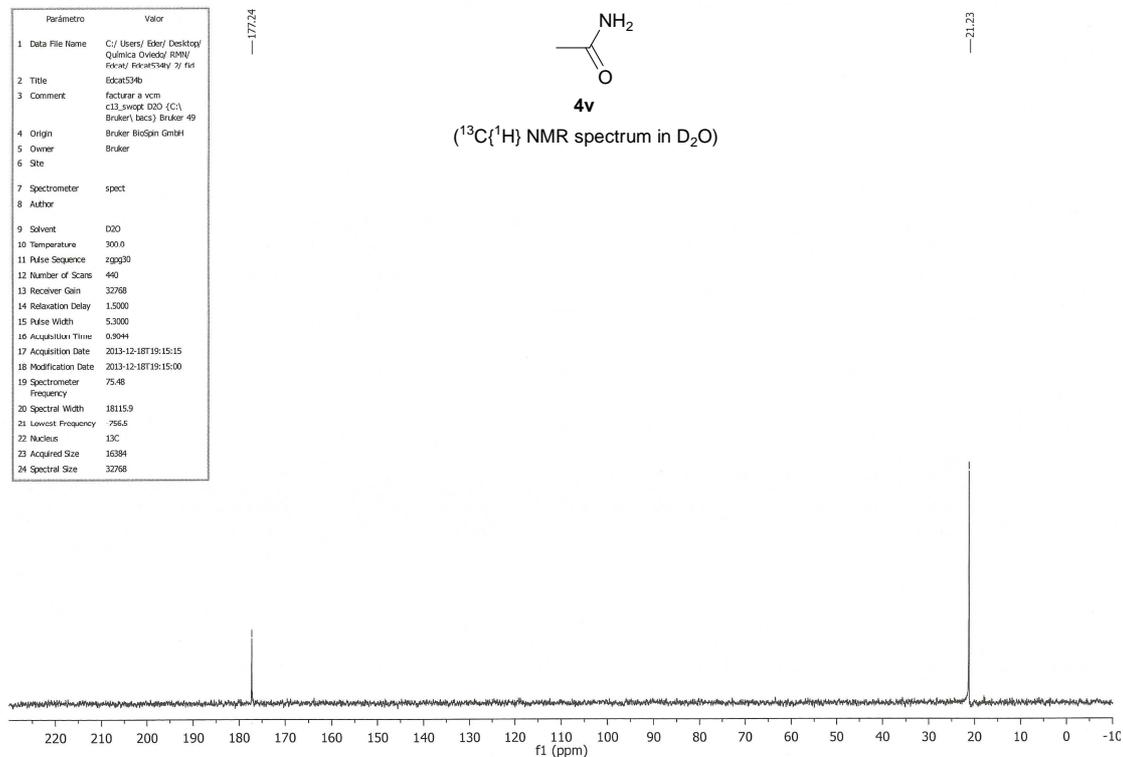
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1 Data File Name	C:/Users/Eder/Desktop/Química/Oviedo/RMW/Escat505/2/ f1d
2 Title	Escat505
3 Comment	facturar a VCM c13_swopt MeOD (C), Bruker(bacs) Bruker 56
4 Origin	Bruker Biospin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	MeOD
10 Temperature	300.0
11 Pulse Sequence	zgpg30
12 Number of Scans	490
13 Receiver Gain	32768
14 Relaxation Delay	1.5000
15 Pulse Width	5.3000
16 Acquisition Time	0.9044
17 Acquisition Date	2014-03-12T19:55:43
18 Modification Date	2014-03-12T19:55:00
19 Spectrometer Frequency	75.48
20 Spectral Width	18115.9
21 Lowest Frequency	-756.5
22 Nucleus	¹³ C
23 Acquired Size	16384
24 Spectral Size	32768

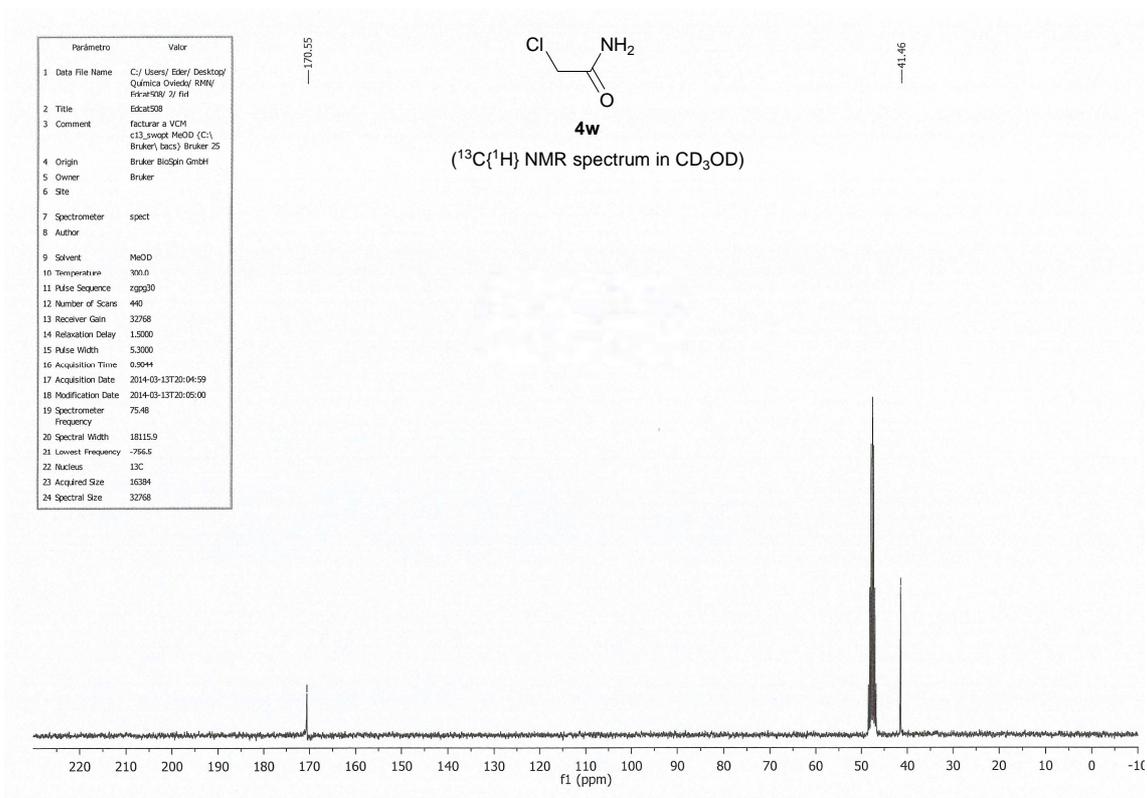
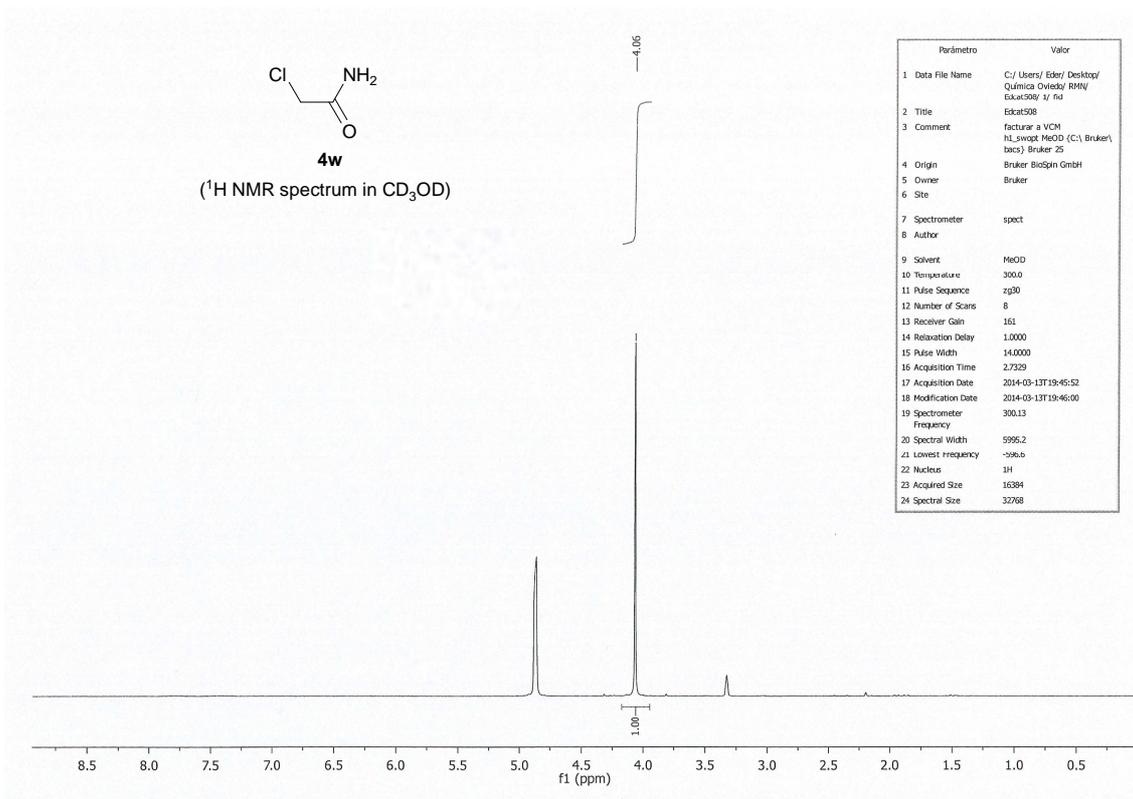


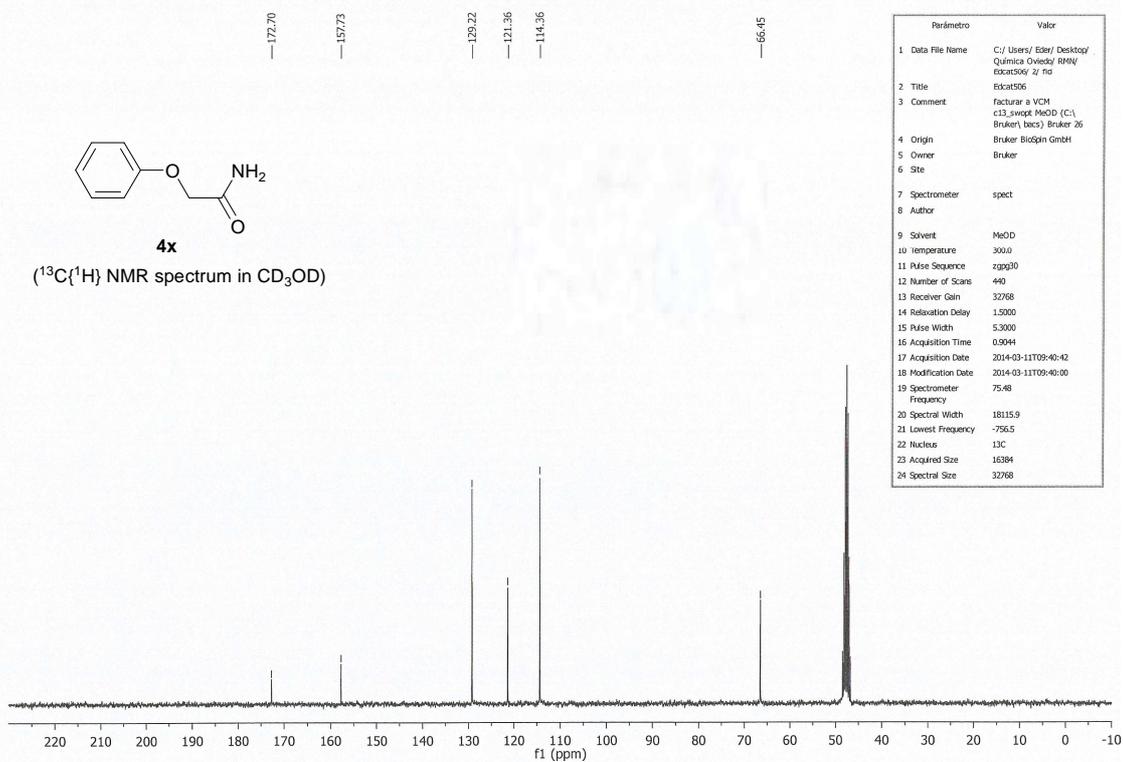
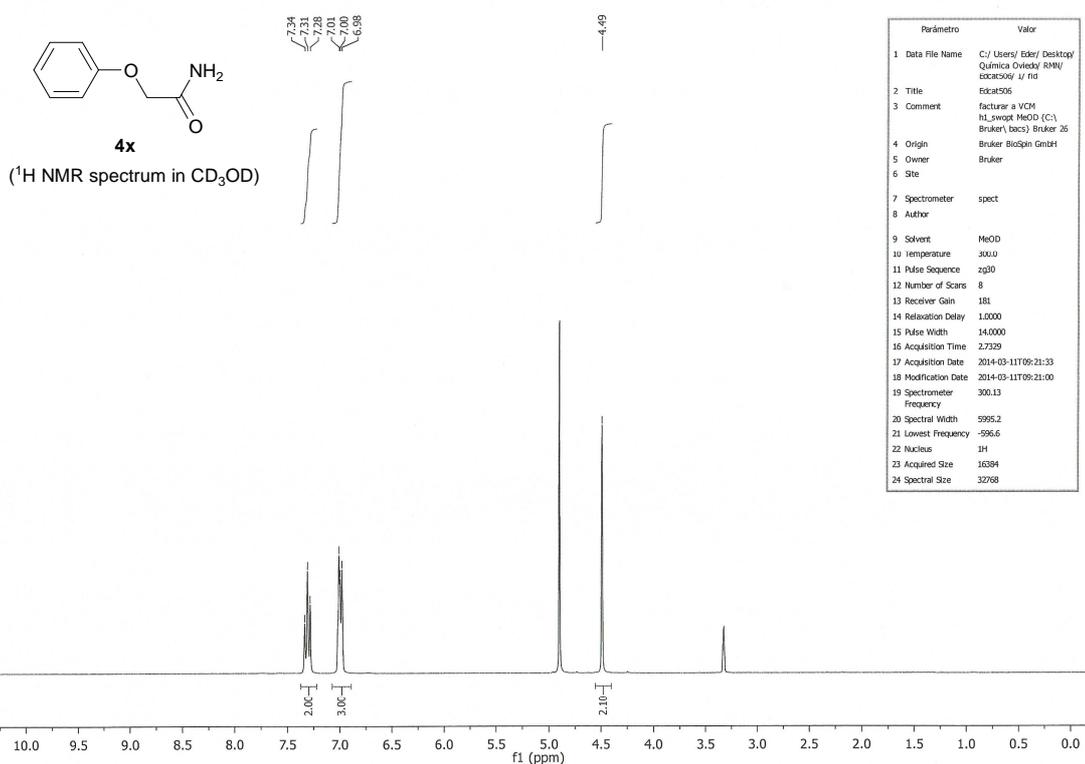
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1 Data File Name	C:/Users/Eder/Desktop/Química Oviedo/ RMN/ Edcat/ Edcat534b/ 1/ fid
2 Title	Edcat534b
3 Comment	facturar a vcm ML_Novak D2O (C-1; Bruker) bacs) Bruker 49
4 Origin	Bruker BioSpin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	D2O
10 Temperature	300.0
11 Pulse Sequence	zg30
12 Number of Scans	8
13 Receiver Gain	102
14 Relaxation Delay	1.0000
15 Pulse Width	14.0000
16 Acquisition Time	2.7325
17 Acquisition Date	2013-12-18T18:56:01
18 Modification Date	2013-12-18T18:56:00
19 Spectrometer Frequency	300.13
20 Spectral Width	5995.2
21 Lowest Frequency	-1196.8
22 Nucleus	¹ H
23 Acquired Size	16384
24 Spectral Size	32768



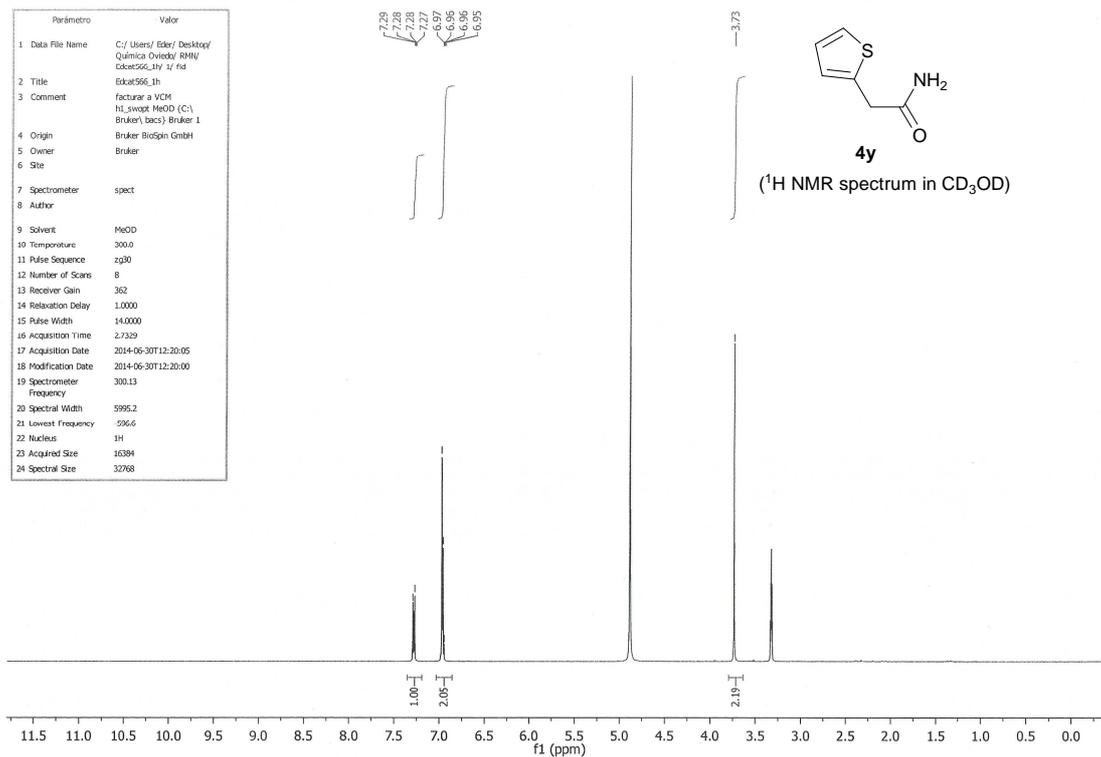
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1 Data File Name	C:/Users/Eder/Desktop/Química Oviedo/ RMN/ Novak/ Edcat534b/ 2/ fid
2 Title	Edcat534b
3 Comment	facturar a vcm c13_swapt D2O (C-1; Bruker) bacs) Bruker 49
4 Origin	Bruker BioSpin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	D2O
10 Temperature	300.0
11 Pulse Sequence	zgpg30
12 Number of Scans	460
13 Receiver Gain	32768
14 Relaxation Delay	1.5000
15 Pulse Width	5.3000
16 Acquisition Time	0.8944
17 Acquisition Date	2013-12-18T19:15:15
18 Modification Date	2013-12-18T19:15:00
19 Spectrometer Frequency	75.48
20 Spectral Width	18115.9
21 Lowest Frequency	756.5
22 Nucleus	¹³ C
23 Acquired Size	16384
24 Spectral Size	32768



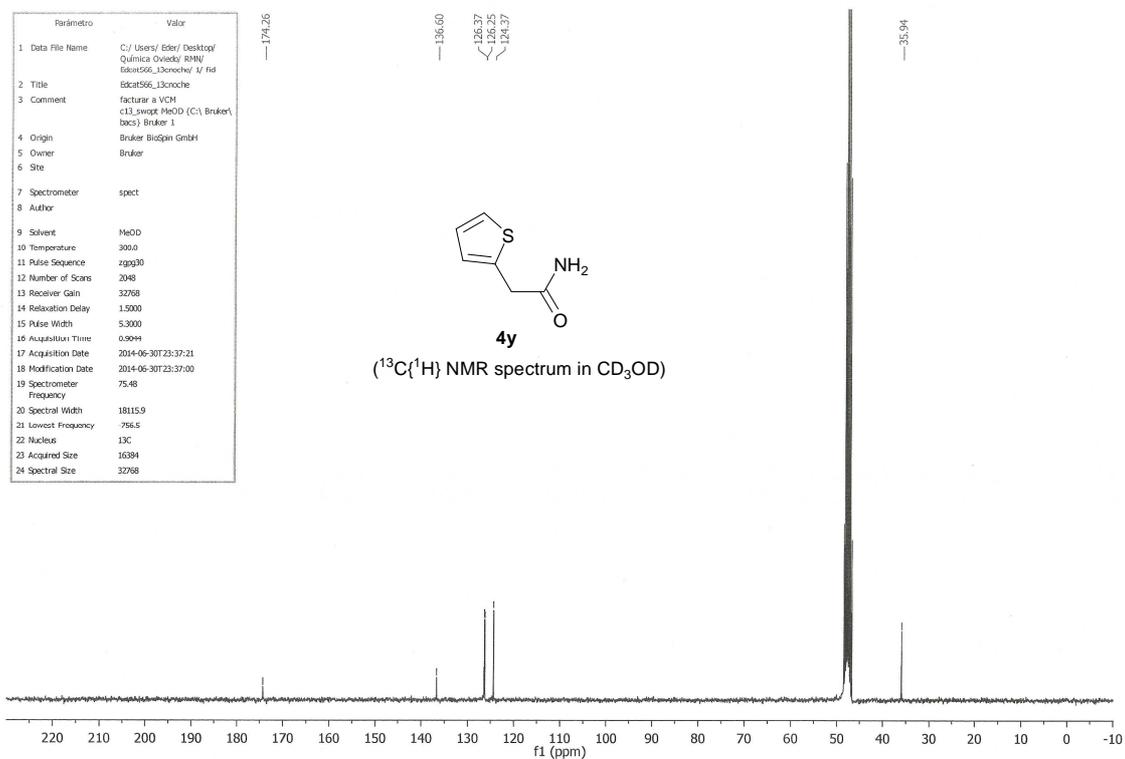


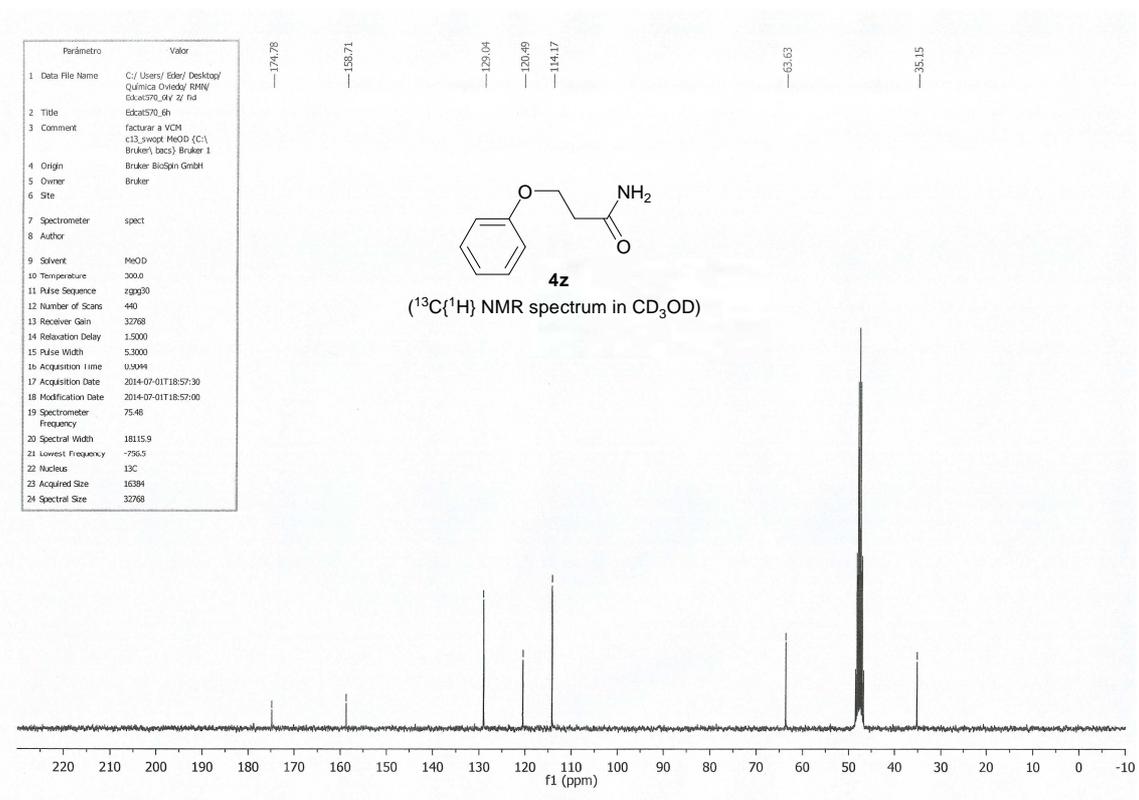
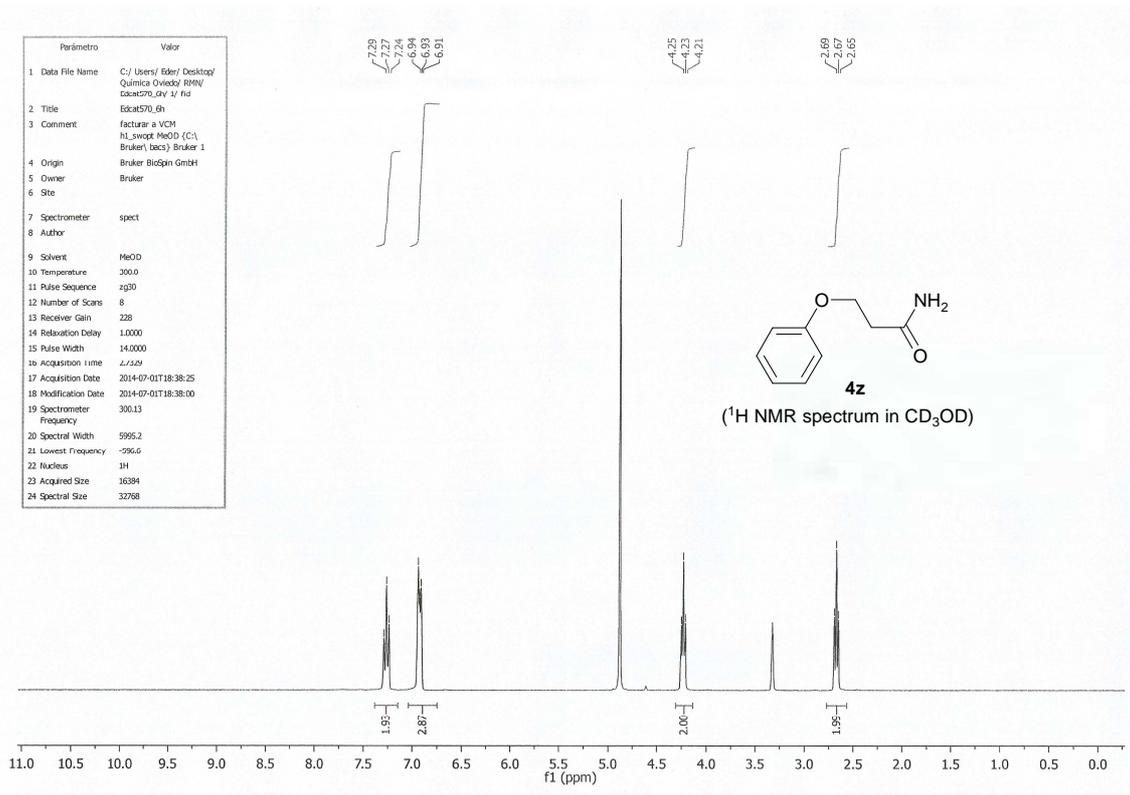


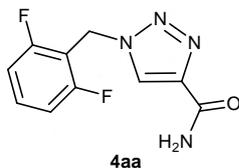
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1 Data File Name	C:/Users/Eder/Desktop/Química Ovedes/RMN/Excat566_1H_1/fid
2 Title	Excat566_1h
3 Comment	factor a VCM h1_swept MeOD (C-), Bruker(bacs) Bruker 1
4 Origin	Bruker Biospin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	MeOD
10 Temperature	300.0
11 Pulse Sequence	zg30
12 Number of Scans	8
13 Receiver Gain	362
14 Relaxation Delay	1.0000
15 Pulse Width	14.0000
16 Acquisition Time	2.7329
17 Acquisition Date	2014-06-30T12:20:05
18 Modification Date	2014-06-30T12:20:00
19 Spectrometer Frequency	300.13
20 Spectral Width	5955.2
21 Lowest Frequency	596.6
22 Nucleus	1H
23 Acquired Size	16384
24 Spectral Size	32768



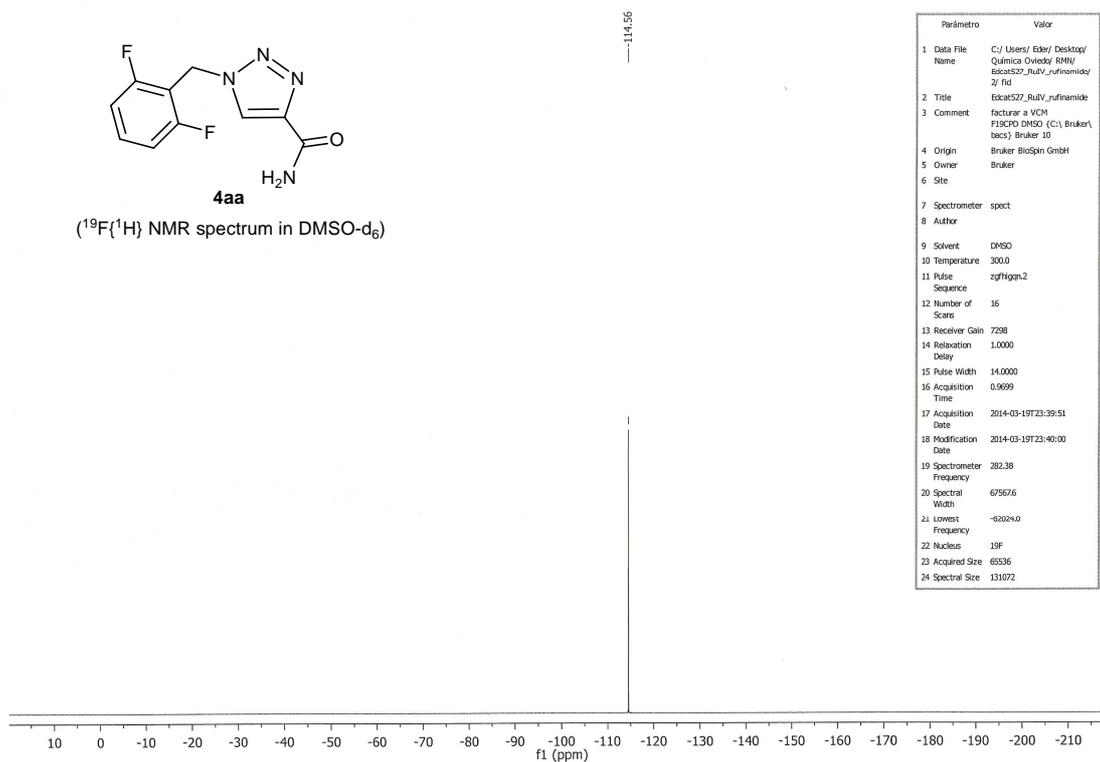
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1 Data File Name	C:/Users/Eder/Desktop/Química Ovedes/RMN/Excat566_13choche/1/fid
2 Title	Excat566_13choche
3 Comment	factor a VCM c13_swept MeOD (C-), Bruker(bacs) Bruker 1
4 Origin	Bruker Biospin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	MeOD
10 Temperature	300.0
11 Pulse Sequence	zgpg30
12 Number of Scans	2048
13 Receiver Gain	32768
14 Relaxation Delay	1.5000
15 Pulse Width	5.3000
16 Acquisition Time	0.9994
17 Acquisition Date	2014-06-30T23:37:21
18 Modification Date	2014-06-30T23:37:00
19 Spectrometer Frequency	75.48
20 Spectral Width	18115.9
21 Lowest Frequency	-756.5
22 Nucleus	13C
23 Acquired Size	16384
24 Spectral Size	32768



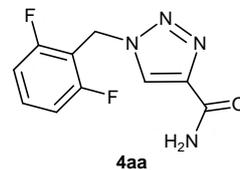
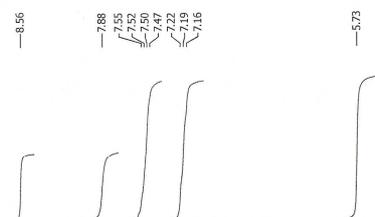




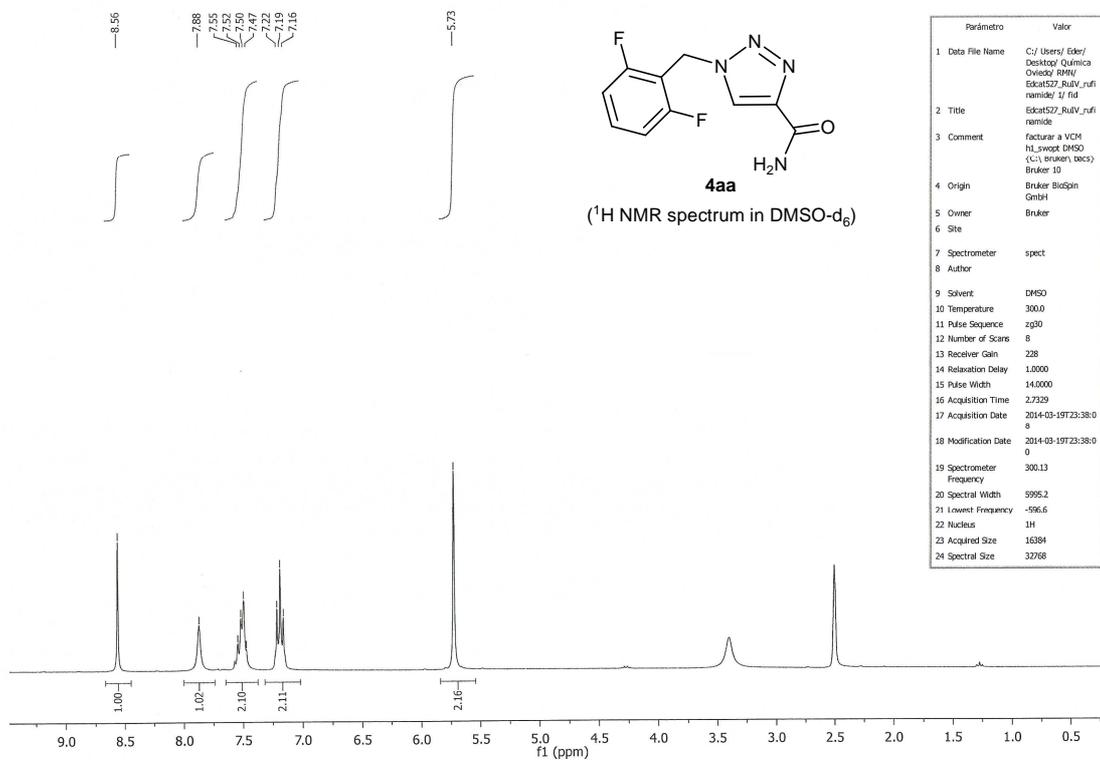
(¹⁹F{¹H} NMR spectrum in DMSO-d₆)



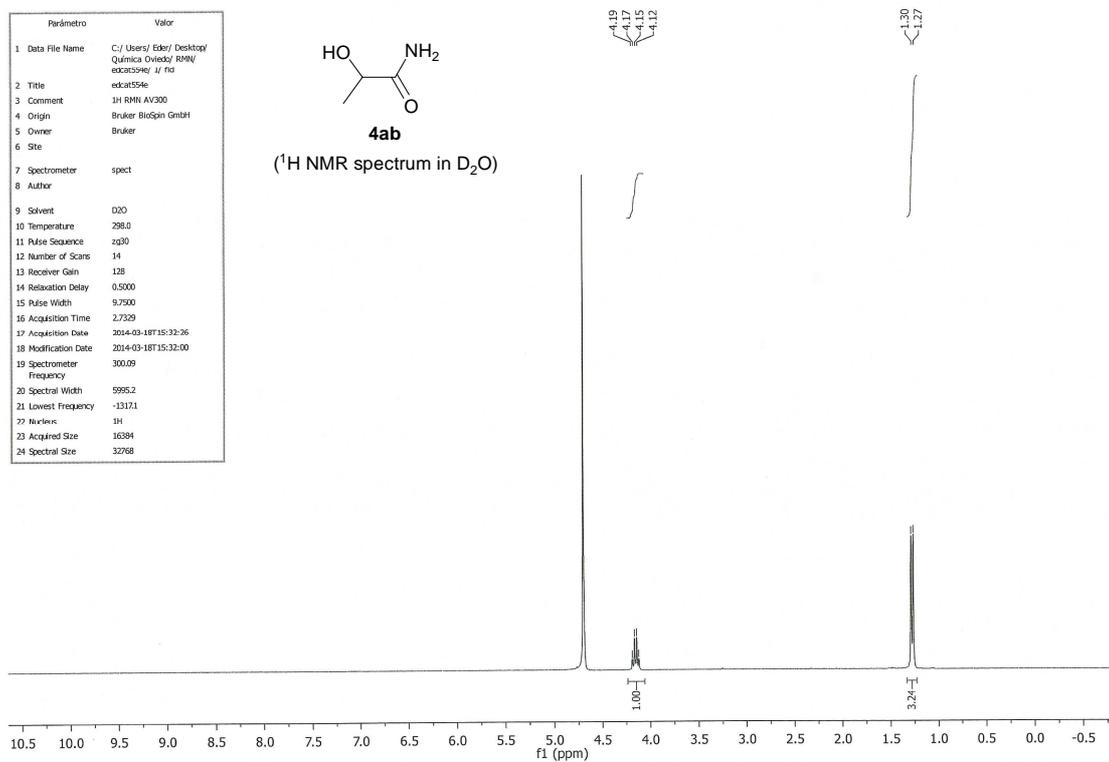
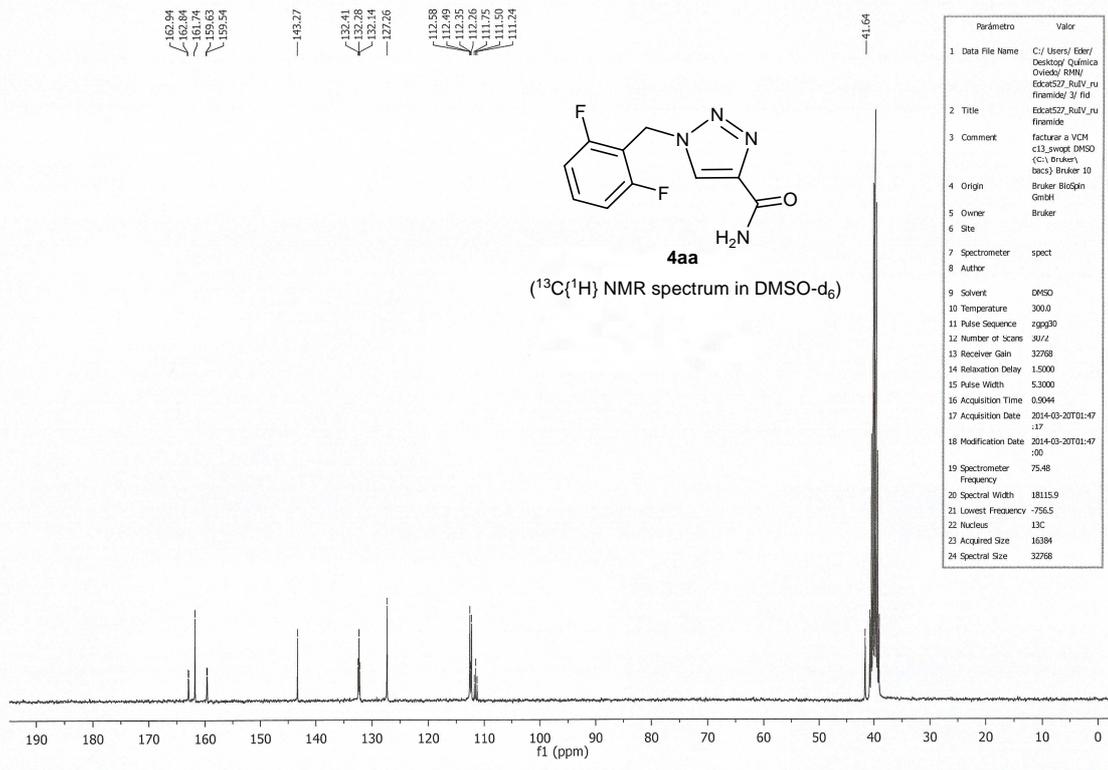
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1 Data File Name	C:/Users/Eder/Desktop/Química Oviesty/ RMN/ Ekat527_RuIV_rufinamide/ 2/ f1d
2 Title	Ekat527_RuIV_rufinamide
3 Comment	factorar a VCM F3200 DMSO (C1) Bruker/ bacs) Bruker 10
4 Origin	Bruker BioSpin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	DMSO
10 Temperature	300.0
11 Pulse Sequence	zgpgqz2
12 Number of Scans	16
13 Receiver Gain	7298
14 Relaxation Delay	1.0000
15 Pulse Width	14.0000
16 Acquisition Time	0.9699
17 Acquisition Date	2014-03-19T23:39:51
18 Modification Date	2014-03-19T23:40:00
19 Spectrometer Frequency	282.38
20 Spectral Width	67567.6
21 Lowest Frequency	-62024.0
22 Nucleus	19F
23 Acquired Size	6536
24 Spectral Size	131072

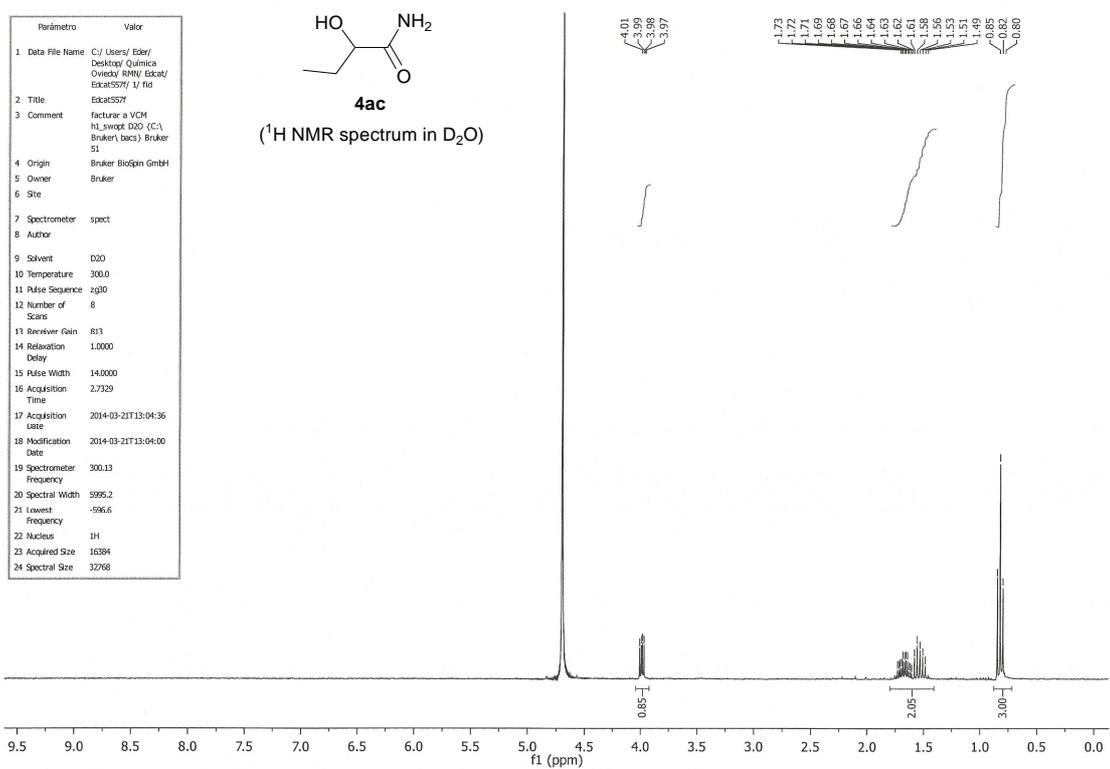
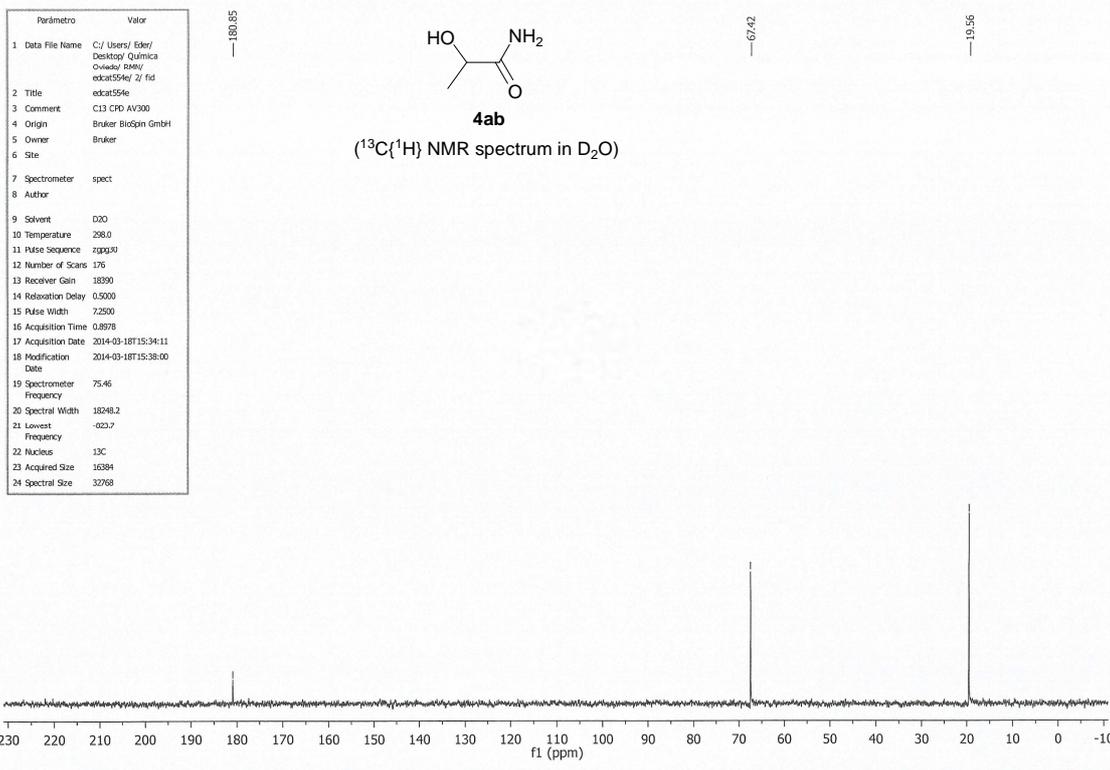


(¹H NMR spectrum in DMSO-d₆)

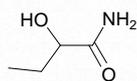


Parámetro	Valor
1 Data File Name	C:/Users/Eder/Desktop/Química Oviesty/ RMN/ Ekat527_RuIV_rufinamide/ 2/ f1d
2 Title	Ekat527_RuIV_rufinamide
3 Comment	factorar a VCM H1_swopt DMSO (C1) Bruker/ bacs) Bruker 10
4 Origin	Bruker BioSpin GmbH
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	DMSO
10 Temperature	300.0
11 Pulse Sequence	zg30
12 Number of Scans	8
13 Receiver Gain	228
14 Relaxation Delay	1.0000
15 Pulse Width	14.0000
16 Acquisition Time	2.7329
17 Acquisition Date	2014-03-19T23:38:00
18 Modification Date	2014-03-19T23:38:00
19 Spectrometer Frequency	300.13
20 Spectral Width	5995.2
21 Lowest Frequency	-596.6
22 Nucleus	1H
23 Acquired Size	16384
24 Spectral Size	32768





Parámetro	Valor
1 Data File Name	C:/Users/Eden/Desktop/Química/Ovickay_RMW/Eden557_1/ fid
2 Title	Eden557
3 Comment	facturar a VCM c13_swapt D2O (C-1; Bruker), bacs) Bruker 7
4 Origin	bruker biospin center
5 Owner	Bruker
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	D2O
10 Temperature	300.0
11 Pulse Sequence	zgpg30
12 Number of Scans	480
13 Receiver Gain	32768
14 Relaxation Delay	15000
15 Pulse Width	5.3000
16 Acquisition Time	0.9944
17 Acquisition Date	2014-03-21T15:13:30
18 Modification Date	2014-03-21T15:32:00
19 Spectrometer Frequency	75.48
20 Spectral Width	18115.9
21 Lowest Frequency	-756.5
22 Nucleus	13C
23 Acquired Size	16384
24 Spectral Size	32768



4ac

(¹³C{¹H} NMR spectrum in D₂O)

