

Supporting Information

Aerobic Oxygenative Cleavage of Electron-Deficient Triple Bonds in the Gold-Catalyzed Cyclization of 1,6-Enynes

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1. General Information

All solvents were dried and distilled according to the standard methods before use.¹ Au(SPhos)Cl, Au(JohnPhos)Cl, Au(*t*Bu-XPhos)Cl and Au(*t*Bu-DavePhos)Cl were prepared from Au(SMe₂)Cl and the corresponding phosphine ligand following the standard protocol.² Au(PPh₃)Cl, AuCl₃, AgOTf, AgSbF₆ and AgNTf₂ were purchased from Strem Chemicals or Aldrich Chemicals and was stored in a dry-keeper before use. Propiolamide substrates were prepared via standard DCC coupling³ followed by their *N*-allylation. Propiolamides with aryl substituted alkynes (**1a**, **1c**, **1d**, **1g** and **1h**) were prepared by coupling of the propiolamide precursors with boronic acids.⁴ (2-(bromomethyl)allyloxy)(*tert*-butyl)dimethyl silane,⁵ (3-bromoprop-1-en-2-yl)benzene⁶ for the preparation of substrates **1k**, **1l** and **1m**, respectively, were prepared according to the literature.

TLC (Thin-layer chromatography) analysis was carried out on Merck silica gel 60 F254 TLC plate and was visualized with UV lamp and potassium permanganate solution. Flash column chromatography was performed on Kieselgel 60 (230-400 mesh). ¹H, ¹³C NMR and COSY spectra were recorded on a Varian (Mercury, 300 MHz and 400 MHz) spectrometer with TMS as an internal standard. High resolution mass spectra (HRMS) were obtained from Korea Basic Science Institute (KBSI) at Seoul and Daegu. X-ray crystallographic analysis was conducted at Gyeongsang National University.

¹ Armarego, W. L. F.; Chai, C. L. L. *Purification of Laboratory Chemicals*; Elsevier: Oxford, 2009.

² (a) Mézailles, N.; Ricard, L.; Gagosz, F. *Org. Lett.* **2005**, *7*, 4133; (b) de Frémont, P.; Scott, N. M.; Stevens, E. D.; Nolan, S. P. *Organometallics*, **2005**, *24*, 2411.

³ Srinivasachari, S.; Fichter, K. M.; Reineke, T. M. *J. Am. Chem. Soc.* **2008**, *130*, 4618.

⁴ M. -B. Zhou, W. -T. Wei, Y. -X. Xie, Y. Lei, and J. -H. Li, *J. Org. Chem.* **2010**, *75*, 5635

⁵ Couladouros, E. A.; Dakanali, M.; Demadis, K. D.; Vidali, V. P. *Org. Lett.* **2009**, *11*, 4430.

⁶ Geoghegan K.; Evans, P.; Rozas, I.; Alkorta, I. *Chem. Eur. J.* **2012**, *18*, 13379.

2. Screening of Reaction Conditions

The following reactions were conducted using the substrate **1a** inside a flask closed under air, *i.e.* without rigorous exclusion of air. The catalyst were generated *in-situ*: Au(L)Cl (5 mol %) and AgX (5 mol %) was added to a solution of **1a** (0.1 M) in a vial closed under air.

Table S1. Screening of reaction conditions employing **1a**^a

Reaction scheme: Substrate **1a** reacts with Au(L)Cl (5 mol%) and AgX (5 mol%) in a solvent at room temperature (rt) for 4 hours in a flask closed under air to yield products **2a** and **3a**.

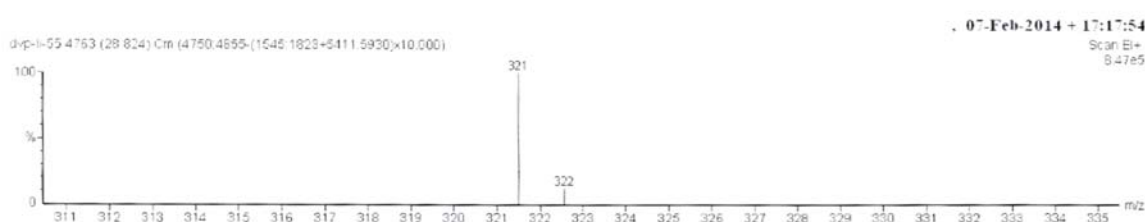
Entry	Ligand (L)	Ag salt (X)	Solvent	2a (%) ^b	3a (%) ^b
1	SPhos	AgSbF ₆	1,4 dioxane	-	75
2	JohnPhos	AgSbF ₆	1,4 dioxane	-	66
3	<i>t</i> Bu-XPhos	AgSbF ₆	1,4 dioxane	-	36
4	<i>t</i> Bu-DavePhos	AgSbF ₆	1,4 dioxane	-	55
5	PPh ₃	AgSbF ₆	1,4 dioxane	-	37
6	IPr	AgSbF ₆	1,4 dioxane	-	74
7	AuCl ₃	-	1,4 dioxane	-	-
8	SPhos	AgOTf	1,4 dioxane	-	51
9	SPhos	AgNTf ₂	1,4 dioxane	-	45
10	SPhos	AgBF ₄	1,4 dioxane	24	52
11	SPhos	AgSbF ₆	CHCl ₃	74	<5
12	SPhos	AgSbF ₆	toluene	25	45
13	SPhos	AgSbF ₆	DCE	66	<5
14	SPhos	AgSbF ₆	THF	-	54
15	SPhos	AgSbF ₆	diethyl ether	7	63
16	SPhos	AgSbF₆	CF₃CH₂OH	-	86

^a Au(I)-complex were generated *in-situ*. ^b Crude yield (¹H-NMR, 1,3,5-trimethoxy-benzene as an internal standard).

3. Incorporation of ^{18}O into **3b**

3.1. Exchange with $^{18}\text{OH}_2$

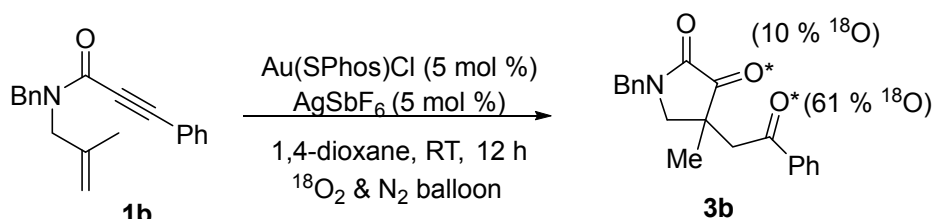
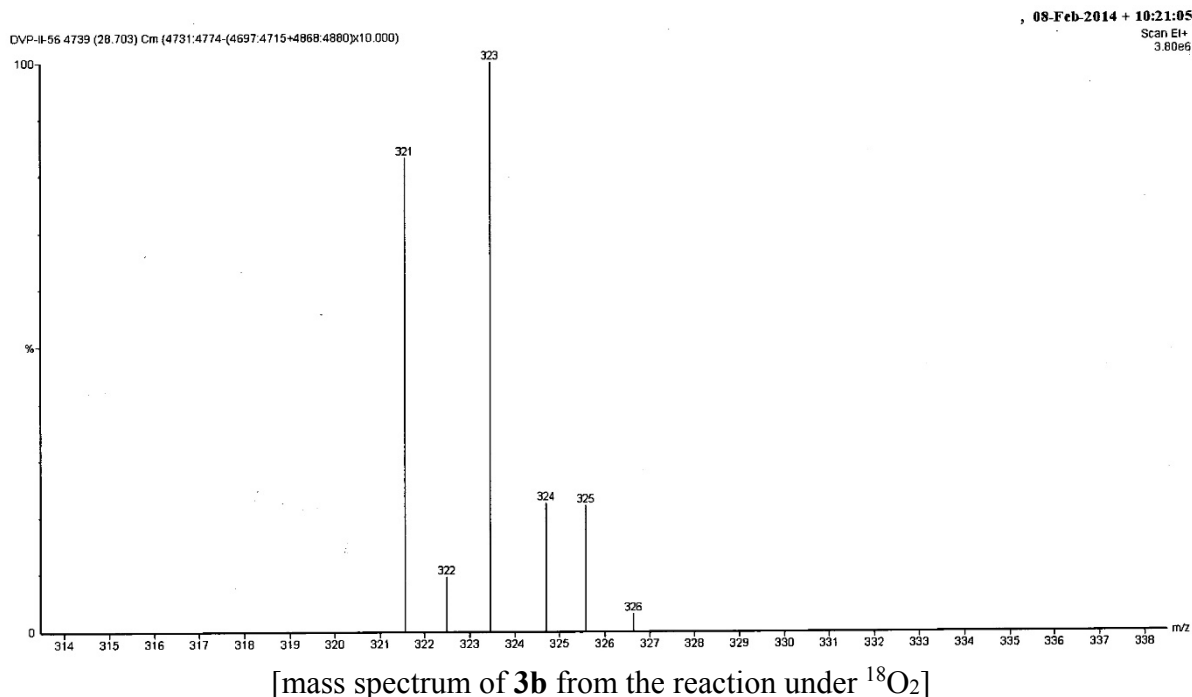
To test whether oxygen from $^{18}\text{OH}_2$ (99 atom %) is incorporated into the product (**3b**), the reaction of **1b** was conducted in the presence of 15 equiv. of $^{18}\text{OH}_2$ in 1,4-dioxane under otherwise standard conditions. To remove the possibility of carbonyl oxygen exchange with adventitious water via a hydrate after the formation of 1,2-dicarbonyl **3b**, the reaction mixture was treated with aqueous 1 N HCl before usual workup. An aliquot was taken from organic layer for GC-MS analysis, which indicated no ^{18}O incorporation in the product **3b**.



[mass spectrum of normal isotopomer **3b**]

3.2. Exchange with $^{18}\text{O}_2$

For $^{18}\text{O}_2$ incorporation experiment, a gas mixture of $^{18}\text{O}_2$ (90 atom %) and N_2 (ratio of partial pressure = 1:4) in a balloon was used. A mixture of $\text{Au}(\text{SPhos})\text{Cl}$ (3.2 mg, 0.005 mmol) and AgSbF_6 (1.7 mg, 0.005 mmol) was dissolved in anhydrous 1,4-dioxane and the mixture was stirred for 10 min at RT. Substrate **1b** (29.8 mg, 0.10 mmol) was placed in a separate test tube and capped with a septum. The tube was evacuated and then filled with N_2 gas. This evacuation and re-filling was repeated twice before a balloon of $^{18}\text{O}_2$ and N_2 mixture (1:4) was connected through the septum. To the test tube was added above solution of pre-catalyst in dioxane via a syringe. The reaction mixture was allowed to stir at rt for 12 h. An aliquot was taken directly from the crude reaction mixture for GC-MS analysis to reduce the oxygen exchange of 1,2-dicarbonyl group with trace amount of water.



[Calculation]

For normal oxygen isotopomer **3b** (no ^{18}O), only m/z 321 and 322 was observed for molecular peaks (please see the mass spectrum in **3.1**).

Total relative peak heights of all **3b** species (m/z 321, 322, 323, 324, 325 and 326) from above mass spectrum = 83 + 10 + 100 + 23 + 22 + 3 = 241. Therefore,

3b (%): peak height (m/z 321 & 322)/241 x 100 = 38.6 %

^{18}O -**3b** (%): peak height (m/z 323 & 324)/241 x 100 = 51.0 %

$^{18}\text{O}_2$ -**3b** (%): peak height (m/z 325 & 326)/241 x 100 = 10.4 %

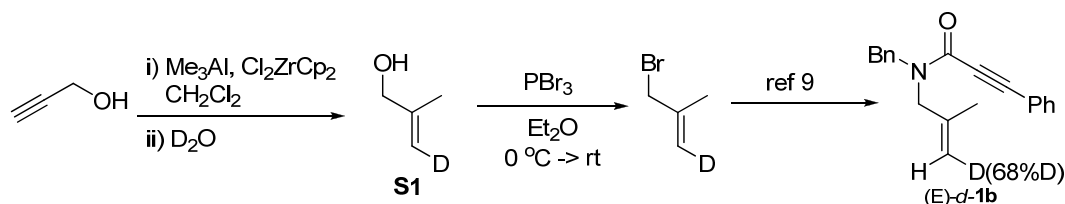
Assuming that all ^{18}O -**3b** resulted from oxygen exchange of $^{18}\text{O}_2$ -**3b** with extraneous water (H_2O) via a hydrate, mono ^{18}O -**3b** should have ^{18}O atom in the phenyl ketone, not α -ketocarbonyl. Therefore,

^{18}O content at α -ketocarbonyl (%): 10.4 % (only from $^{18}\text{O}_2$ -**3b**)

^{18}O content at phenylketone carbonyl (%): 10.4 % (from $^{18}\text{O}_2$ -**3b**) + 51.0 % (from ^{18}O -**3b**) = 61.4 %

4. Preparation of *d*-labeled substrates

Preparation of **S1** for (*E*)-**d-1b**⁷

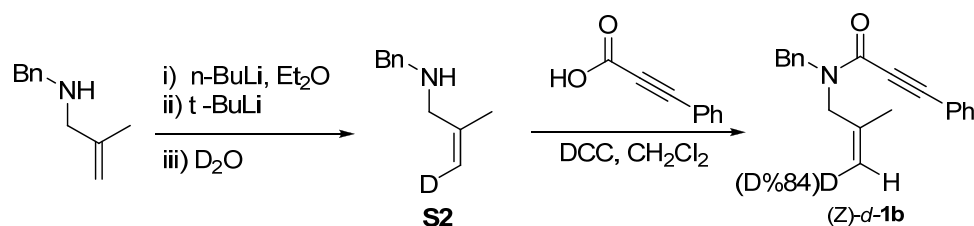


To a solution of Me_3Al (pyrophoric!) (1.5 mL, 3.0 mmol) and Cp_2ZrCl_2 (292.3 mg, 1.0 mmol) in 1,2-dichloroethane (10 mL) was added dropwise 3-butyne-1-ol (58 μL , 1.0 mmol) in 1,2-dichloroethane (10 mL) at 0 °C. A rapid evolution of methane occurs. After the reaction mixture is stirred for 12 h at room temperature, it was treated with D_2O (1.0 mL) and then with saturated aqueous K_2CO_3 at 0 °C. The heterogeneous mixture was extracted with hexane and ether. The organic layers were combined, dried over MgSO_4 , and concentrated *in vacuo*. Flash column chromatography (EtOAc:HEX=30:70) provided the corresponding allylic alcohol **S1** (24.9 mg, 34 %) as a colorless oil.

The allylic alcohol **S1** was brominated with PBr_3 and coupled to the amides as described before⁸ to provide (*E*)-**d-1b**: 89 % yield (68 % D incorporation).

(*E*)-**d-1b**: ^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.2:1): δ 7.58-7.46 (m, 2H, major/minor), 7.44-7.24 (m, 8H, major/minor), 5.01 (s, H(D), major), 4.96 (s, H(D), minor), 4.90 (s, H, major), 4.81 (s, 2H, minor), 4.80 (s, H, minor), 4.63 (s, 2H, major), 4.11 (s, 2H, major), 3.95 (s, 2H, minor), 1.78 (s, 3H, major), 1.72 (s, 3H, minor); ^{13}C NMR (100 MHz, CDCl_3): δ 155.3, 154.9, 139.9, 139.6, 136.5, 136.4, 132.6, 132.5, 130.24, 130.19, 128.9, 128.8, 128.7, 128.64, 128.58, 128.0, 127.8, 127.7, 120.6, 120.5, 114.1, 114.0, 113.8, 113.5, 113.2, 90.7, 90.6, 81.8, 81.7, 53.9, 51.4, 48.7, 46.4, 20.1, 19.9.

Preparation of **S2** for (*Z*)-**d-1b**⁹



To a solution of corresponding amine (161.2 mg, 1.0 mmol) in diethyl ether (5 mL) was added *n*-BuLi (2.5M in hexane, 0.4 mL, 1.0 mmol) at -50 °C under Ar and the mixture was stirred for 20 min at -50 °C. To the resulting mixture was added *t*-BuLi (1.7M in pentane,

⁷ C. L. Rand, D. E. Van Horn, M. W. Moore, E. -i. Negishi, *J. Org. Chem.* **1981**, 46, 4093.

⁸ Koo, J.; Park, H. -S.; Shin, S. *Tetrahedron Lett.* **2013**, 54, 834.

⁹ J. Baluenga, R. Gonz  les, F. J. Fa  n  s, *Tetrahedron Lett.* **1992**, 33, 7573

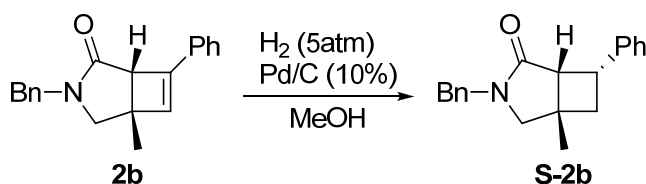
0.71 mL, 1.2 mmol) at -30 °C and stirred 2 h. The mixture was treated with D₂O (22 μL, 1.2 mmol) at -50 °C and stirred overnight. The mixture was quenched with saturated NH₄Cl and extracted with diethyl ether. The combined organic layers were washed with brine, dried over anhydrous MgSO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc:Hex = 20:80) to give corresponding amine **S2** (100.1 mg, 62 %).

The secondary amine was then coupled with phenylpropionic acid by DCC,³ to give (*Z*)-*d*-**1b**: 76 % yield (84 % D incorporation).

(*Z*)-*d*-**1b**: ¹H NMR (400 MHz, CDCl₃, rotameric mixture, 1.2:1): δ 7.58-7.46 (m, 2H, major/minor), 7.44-7.24 (m, 8H, major/minor), 5.00 (s, H, major), 4.94 (s, H, minor), 4.90 (s, H(D), major), 4.81 (s, 2H, minor), 4.63 (s, 2H, major), 4.11 (s, 2H, major), 3.95 (s, 2H, minor), 1.78 (s, 3H, major), 1.72 (s, 3H, minor); ¹³C NMR (100 MHz, CDCl₃): δ 155.3, 154.9, 139.9, 139.6, 136.5, 136.4, 132.6, 132.5, 130.23, 130.20, 128.9, 128.8, 128.7, 128.65, 128.63, 128.56, 128.0, 127.8, 127.7, 120.6, 120.5, 114.1, 114.0, 113.8, 113.6, 113.4, 113.1, 112.9, 90.7, 90.6, 81.8, 81.7, 53.9, 51.4, 48.6, 46.4, 20.2, 20.1, 20.0, 19.9.

5. Structural Determination of **2b**

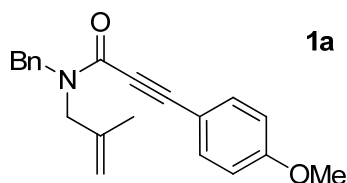
A confirmation of the structure of **2b** was conducted by the following hydrogenation reaction.



To a solution of **2b** (13.9 mg, 0.065 mmol) in MeOH (2 mL) was added Pd/C (6.9 mg of 10 % Pd/C). The mixture was hydrogenated at 5 atm of H₂ for 48 h at rt. The mixture was filtered off through a Celite and separated by silica gel column chromatography.

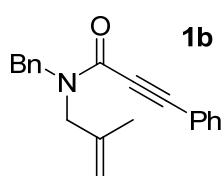
S-2b : ¹H NMR (400 MHz, CDCl₃): δ 7.40-7.18 (m, 10H), 4.65 (d of ABq, *J* = 14.4 Hz, H), 4.13 (d of ABq, *J* = 14.5 Hz, H), 4.01 (q, *J* = 9.8 Hz, H), 3.15 (d of ABq, *J* = 9.8 Hz, H), 3.09-3.01 (m, 2H), 2.33 (t, *J* = 11.0 Hz, H), 2.25 (ddd, *J* = 2.8, 9.4, 12.1 Hz, H), 1.38 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.8, 139.6, 136.9, 128.7, 128.6, 128.1, 127.7, 127.6, 59.6, 51.9, 46.8, 37.9, 37.3, 33.7, 22.9.

6. Characterization of Substrates



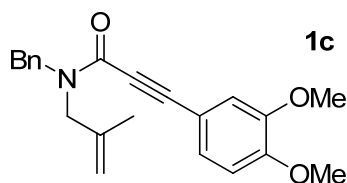
1a

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.1:1): δ 7.46 (app t, $J = 9.0$ Hz, 2H, major/minor), 7.44-7.22 (m, 5H, major/minor), 6.86 (app t, $J = 9.0$ Hz, 2H, major/minor), 5.01 (s, H, major), 4.95 (s, H, minor), 4.89 (s, H, major), 4.81 (s, 2H, minor), 4.79 (s, H, minor), 4.62 (s, 2H, major), 4.10 (s, 2H, major), 3.94 (s, 2H, minor), 3.83 (s, 3H, major), 3.82 (s, 3H, minor), 1.77 (s, 3H, major), 1.72 (s, 3H, minor); ^{13}C NMR (100 MHz, CDCl_3): δ 161.05, 161.03, 155.5, 155.1, 140.0, 139.7, 136.5, 136.4, 134.3, 134.2, 128.8, 128.6, 128.4, 127.8, 127.7, 127.5, 114.22, 114.19, 113.9, 113.2, 112.4, 112.3, 91.19, 91.14, 80.99, 80.85, 55.4, 53.8, 51.3, 48.5, 46.3, 20.0, 19.9.



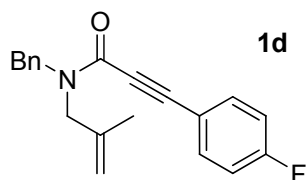
1b

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.2:1): δ 7.58-7.46 (m, 2H, major/minor), 7.44-7.24 (m, 8H, major/minor), 5.01 (s, H, major), 4.96 (s, H, minor), 4.90 (s, H, major), 4.81 (s, 2H, minor), 4.80 (s, H, minor), 4.63 (s, 2H, major), 4.11 (s, 2H, major), 3.95 (s, 2H, minor), 1.78 (s, 3H, major), 1.72 (s, 3H, minor); ^{13}C NMR (100 MHz, CDCl_3): δ 155.2, 154.8, 139.9, 139.6, 136.4, 136.2, 132.5, 132.4, 130.13, 130.10, 128.8, 128.7, 128.55, 128.52, 128.4, 127.9, 127.7, 127.6, 120.5, 120.4, 114.0, 113.3, 90.6, 90.5, 81.7, 81.6, 53.8, 51.3, 48.6, 46.4, 20.1, 19.9.



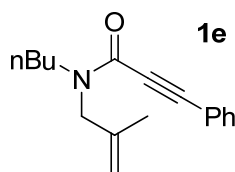
1c

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.1:1): δ 7.42-7.24 (m, 5H, major/minor), 7.20-7.10 (m, H, major/minor), 7.06-6.96 (m, H, major/minor), 6.86-6.78 (m, H, major/minor), 5.02 (s, H, major), 4.95 (s, H, minor), 4.91 (s, H, major), 4.82 (s, 2H, minor), 4.80 (s, H, minor), 4.63 (s, 2H, major), 4.11 (s, 2H, major), 3.95 (s, 2H, minor), 3.93-3.80 (m, 6H, major/minor), 1.78 (s, 3H, major), 1.72 (s, 3H, minor); ^{13}C NMR (100 MHz, CDCl_3): δ 155.4, 155.0, 151.0, 150.9, 148.72, 148.68, 139.9, 139.6, 136.5, 136.4, 128.8, 128.6, 128.4, 127.8, 127.7, 127.5, 126.4, 126.3, 114.83, 114.79, 113.9, 113.2, 112.3, 112.2, 110.97, 110.94, 91.3, 91.2, 80.7, 80.6, 55.96, 55.93, 53.8, 51.3, 48.6, 46.3, 20.0, 19.9.



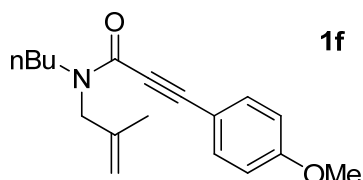
1d

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.2:1): δ 7.58-7.44 (m, 2H, major/minor), 7.42-7.20 (m, 5H, major/minor), 7.04 (app q, $J = 9.0$ Hz, 2H, major/minor), 5.02 (s, H, major), 4.96 (s, H, minor), 4.90 (s, H, major), 4.80 (s, 3H, major/minor), 4.62 (s, 2H, major), 4.09 (s, 2H, major), 3.95 (s, 2H, minor), 1.77 (s, 3H, major), 1.72 (s, 3H, minor); ^{13}C NMR (100 MHz, CDCl_3): δ 163.5 (d, $J = 254.3$ Hz), 155.0, 154.7, 139.8, 139.5, 136.4, 136.2, 134.6 (d, $J = 8.1$ Hz), 134.5 (d, $J = 8.8$ Hz), 128.8, 128.7, 128.4, 127.9, 127.6, 116.6 (d, $J = 3.7$ Hz), 116.5 (d, $J = 3.7$ Hz), 116.1 (d, $J = 22.8$ Hz), 116.0 (d, $J = 22.1$ Hz), 114.0, 113.3, 89.6, 89.5, 81.6, 81.5, 81.4, 53.8, 51.3, 48.6, 46.4, 20.0, 19.9.



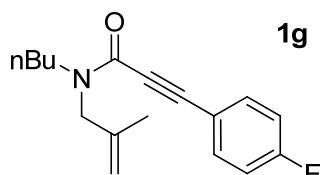
1e

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1:1): δ 7.60-7.48 (m, 2H), 7.48-7.32 (m, 3H), 4.96 (s, H), 4.91 (s, 2H), 4.83 (s, H), 4.18 (s, 2H), 4.04 (s, 2H), 3.57 (t, $J = 7.4$ Hz, 2H), 3.37 (t, $J = 7.4$ Hz, 2H), 1.77 (s, 3H), 1.72 (s, 2H), 1.70-1.48 (m, 2H), 1.46-1.28 (m, 2H), 0.97 (t, $J = 7.4$ Hz, 3H), 0.93 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 154.8, 154.6, 140.4, 140.1, 132.33, 132.29, 129.97, 129.91, 128.53, 128.48, 120.66, 120.65, 113.4, 112.7, 89.8, 89.6, 81.9, 81.8, 55.0, 49.5, 47.9, 43.9, 30.5, 29.1, 20.17, 20.02, 19.99, 19.86, 13.8.



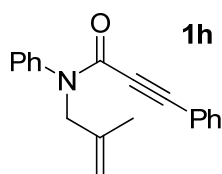
1f

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1:1): δ 7.49 (d, $J = 8.9$ Hz, 2H), 7.46 (d, $J = 9.0$ Hz, 2H), 6.88 (d, $J = 9.0$ Hz, 2H), 6.87 (d, $J = 8.6$ Hz, 2H), 4.96 (s, H), 4.90 (s, 2H), 4.82 (s, H), 4.18 (s, 2H), 4.03 (s, 2H), 3.83 (s, 3H), 3.82 (s, 3H), 3.56 (t, $J = 7.4$ Hz, 2H), 3.37 (t, $J = 7.5$ Hz, 2H), 1.77 (s, 3H), 1.71 (s, 3H), 1.64 (quintet, $J = 7.4$ Hz, 2H), 1.54 (quintet, $J = 7.4$ Hz, 2H), 1.46-1.28 (m, 2H), 0.97 (t, $J = 7.5$ Hz, 3H), 0.93 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 160.95, 160.90, 155.1, 154.9, 140.6, 140.2, 134.11, 134.06, 114.22, 114.16, 113.3, 112.61, 112.56, 112.54, 90.3, 90.2, 81.2, 81.1, 55.3, 55.0, 49.5, 47.9, 43.9, 30.5, 29.1, 20.2, 20.03, 19.96, 19.8, 13.8.



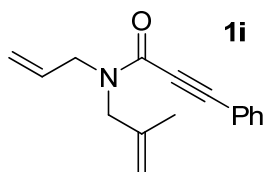
1g

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1:1): δ 7.60-7.44 (m, 2H), 7.14-6.98 (m, 2H), 4.97 (s, H), 4.90 (s, 2H), 4.83 (s, H), 4.17 (s, 2H), 4.04 (s, 2H), 3.55 (t, $J = 7.4$ Hz, 2H), 3.37 (t, $J = 7.4$ Hz, 2H), 1.76 (s, 3H), 1.71 (s, 3H), 1.70-1.46 (m, 2H), 1.44-1.24 (m, 2H), 0.97 (t, $J = 7.4$ Hz, 3H), 0.93 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 163.5 (d, $J = 252.9$ Hz), 163.4 (d, $J = 252.8$ Hz), 154.6, 154.5, 140.3, 140.0, 134.48 (d, $J = 8.8$ Hz), 134.44 (d, $J = 8.8$ Hz), 116.76 (d, $J = 3.7$ Hz), 116.73 (d, $J = 3.7$ Hz), 115.98 (d, $J = 22.9$ Hz), 115.96 (d, $J = 22.8$ Hz), 113.4, 112.7, 88.8, 88.6, 81.7, 81.6, 54.9, 49.5, 47.8, 43.9, 30.5, 29.1, 20.1, 20.01, 19.97, 19.86.



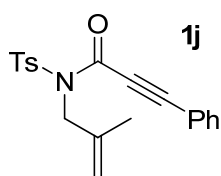
1h

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 5.5:1): δ 7.57 (d, $J = 7.1$ Hz, 2H, minor), 7.50-7.18 (m, 8H, major/minor), 7.12 (d, $J = 7.4$ Hz, 2H, major), 4.95 (s, H, minor), 4.92 (s, H, minor), 4.88 (s, H, major), 4.80 (s, H, major), 4.59 (s, 2H, minor), 4.40 (s, 2H, major), 1.83 (s, 3H, minor), 1.70 (s, 3H, major); ^{13}C NMR (100 MHz, CDCl_3): δ 154.6, 142.1, 140.2, 132.7, 130.5, 130.2, 129.3, 129.2, 128.8, 128.6, 128.4, 128.3, 127.3, 126.4, 120.6, 113.9, 91.5, 82.8, 54.6, 20.6, 20.4.



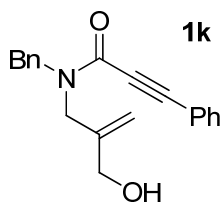
1i

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.2:1): δ 7.60-7.48 (m, 2H, major/minor), 7.48-7.30 (m, 3H, major/minor), 5.92-5.69 (m, H, major/minor), 5.30-5.12 (m, 2H, major/minor), 4.98 (s, H, major), 4.93 (s, H, minor), 4.90 (s, H, major), 4.82 (s, H, minor), 4.20 (d, $J = 5.9$ Hz, 2H, major/minor), 4.17 (s, 2H, major/minor), 4.03 (s, 2H, major/minor), 1.77 (s, 3H, major), 1.72 (s, 3H, minor); ^{13}C NMR (100 MHz, CDCl_3): δ 154.71, 154.67, 140.0, 139.8, 132.6, 132.4, 132.3, 131.9, 130.06, 130.05, 128.5, 120.5, 118.2, 118.1, 113.6, 113.0, 90.06, 89.99, 81.6, 81.5, 54.0, 50.4, 49.0, 46.0, 20.0, 19.9.



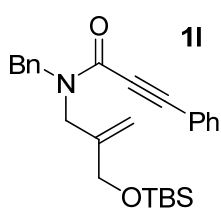
1j

^1H NMR (400 MHz, CDCl_3): δ 7.93 (d, $J = 8.2$ Hz, 2H), 7.49 (d, $J = 7.1$ Hz, 2H), 7.44 (d, $J = 7.4$ Hz, H), 7.37 (t, $J = 7.8$ Hz, 2H), 7.30 (d, $J = 7.8$ Hz, 2H), 5.09 (s, H), 4.99 (s, 1H), 4.63 (s, 2H), 2.42 (s, 3H), 1.81 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 153.0, 145.2, 139.9, 135.8, 132.8, 131.0, 129.4, 129.0, 128.8, 119.5, 112.9, 92.7, 81.6, 52.5, 21.7, 20.3.



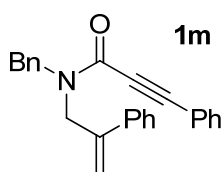
1k

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 2.2:1): δ 7.58-7.48 (m, 2H, major/minor), 7.46-7.26 (m, 8H, major/minor), 5.32 (s, H, minor), 5.18 (s, H, major), 5.10 (s, H, minor), 4.81 (s, 2H, major), 4.66 (s, 2H, minor), 4.25 (s, 2H, minor), 4.16 (s, 2H, minor), 4.04 (s, 4H, major); ^{13}C NMR (100 MHz, CDCl_3): δ 155.6, 155.3, 143.2, 143.0, 136.3, 135.7, 132.53, 132.49, 130.4, 130.2, 129.0, 128.7, 128.6, 128.5, 128.4, 128.1, 127.8, 127.7, 120.3, 120.0, 115.0, 113.6, 91.5, 90.8, 81.4, 81.2, 63.6, 63.5, 51.8, 50.2, 46.7, 45.7.



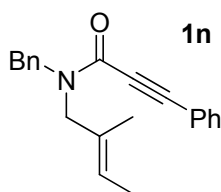
1l

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1:1): δ 7.56-7.48 (m, 2H), 7.44-7.26 (m, 8H), 5.30 (s, 1H), 5.29 (s, H), 5.03 (s, H), 4.95 (s, H), 4.84 (s, 2H), 4.65 (s, 2H, minor), 4.19 (s, 2H), 4.15 (s, 2H), 4.11 (s, 2H), 4.00 (s, 2H), 0.90 (s, 9H), 0.88 (s, 9H), 0.07 (s, 3H), 0.05 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 155.2, 154.8, 142.9, 142.6, 136.4, 136.2, 132.5, 132.4, 130.2, 130.1, 128.8, 128.7, 128.6, 128.52, 128.51, 128.4, 127.9, 127.7, 127.6, 120.5, 120.4, 112.4, 112.2, 90.6, 90.5, 81.6, 81.5.



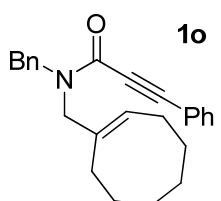
1m

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.2:1): δ 7.60-7.20 (m, 15H, major/minor), 5.54 (s, H, major), 5.52 (s, H, minor), 5.21 (s, H, minor), 5.12 (s, H, major), 4.75 (s, 2H, major), 4.65 (s, 2H, minor), 4.59 (s, 2H, minor), 4.48 (s, 2H, major); ^{13}C NMR (100 MHz, CDCl_3): δ 155.5, 154.9, 143.1, 142.6, 138.9, 138.2, 136.5, 136.2, 132.6, 130.2, 129.0, 128.8, 128.7, 128.67, 128.64, 128.61, 128.5, 128.34, 128.29, 128.0, 127.8, 127.7, 126.5, 126.4, 120.6, 120.5, 115.7, 115.0, 90.8, 90.6, 81.8, 51.7, 51.3, 46.9, 46.3.



1n

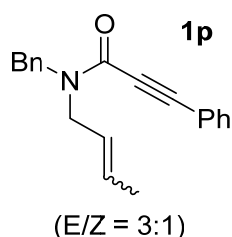
^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.2:1): δ 7.60-7.42 (m, 2H, major/minor), 7.42-7.20 (m, 8H, major/minor), 5.46-5.36 (m, H, major), 5.36-5.26 (m, H, minor), 4.76 (s, 2H, minor), 4.57 (s, 2H, major), 4.10 (s, 2H, major), 3.93 (s, 2H, minor), 1.70-1.56 (m, 6H, major/minor); ^{13}C NMR (100 MHz, CDCl_3): δ 155.6, 155.59, 155.5, 137.2, 137.0, 136.8, 136.6, 133.05, 133.02, 132.9, 131.0, 130.8, 130.7, 130.6, 129.5, 129.34, 129.3, 129.2, 129.13, 129.1, 129.0, 128.6, 128.4, 128.3, 128.2, 128.1, 124.5, 123.7, 121.1, 121.0, 120.9, 91.4, 91.3, 91.0, 82.4, 82.3, 82.2, 56.3, 52.0, 51.5, 50.7, 46.9, 46.4, 14.4, 14.2, 14.0, 13.95.



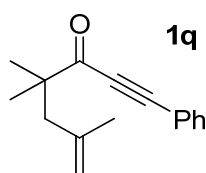
1o

^1H NMR (400 MHz, CDCl_3 , rotameric mixture, 1.5:1): δ 7.60-7.20 (m, 10H), 5.53 (t, $J = 8.2$ Hz H, major), 5.44 (t, $J = 8.2$ Hz H, minor), 4.80 (s, 2H, minor), 4.62 (s, 2H, major), 4.09 (s, 2H, major), 3.94 (s, 2H, minor), 2.19-2.09 (m, 4H, major/minor), 1.54-1.49 (m,

8H, major/minor); ^{13}C NMR (100 MHz, CDCl_3): δ 155.1, 154.9, 136.7, 136.5, 135.2, 134.9, 132.5, 132.3, 130.1, 130.0, 128.8, 128.6, 128.58, 128.53, 128.3, 127.9, 127.8, 127.7, 127.5, 120.6, 120.5, 90.4, 90.3, 81.9, 81.8, 53.4, 51.2, 48.2, 46.3, 29.7, 29.6, 28.58, 28.55, 26.77, 26.57, 26.50, 26.44, 26.33, 26.25, 26.22.

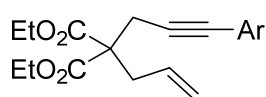


^1H NMR (400 MHz, CDCl_3 rotameric mixture, 1:1): δ 7.61-7.20 (m, 10H, major/minor), 5.77-5.52 (m, H, major/minor), 5.52-5.33 (m, H, major/minor), 4.82 (s, 2H, minor), 4.64 (s, 2H, major), 4.21 (d, J = 6.6 Hz, H, cis), 4.10 (d, J = 5.8 Hz, H, trans), 4.00 (d, J = 7.0 Hz, H, cis), 3.91 (d, J = 5.8 Hz, H, trans), 1.71, (dd J = 3.5, 12.9 Hz, 3H, trans), 1.58, (dd J = 6.3, 18.3 Hz, 3H, cis); ^{13}C NMR (100 MHz, CDCl_3): δ 154.5, 154.3, 136.5, 136.3, 132.3, 132.2, 130.1, 129.97, 129.91, 128.78, 128.70, 128.6, 128.5, 128.45, 128.41, 128.21, 127.79, 127.74, 127.50, 127.4, 125.2, 124.9, 124.5, 124.1, 120.4, 120.3, 90.4, 90.2, 90.1, 90.0, 81.8, 81.7, 81.6, 51.7, 51.4, 49.9, 46.7, 46.5, 45.3, 44.6, 39.8, 17.6.

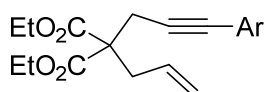


^1H NMR (400 MHz, CDCl_3): δ 7.58 (d, J = 7.1 Hz, 2H), δ 7.45 (t, J = 7.4 Hz, H), δ 7.38 (t, J = 7.8 Hz, 2H), 4.80 (s, H), 4.72 (s, H), 2.49 (s, 2H), 1.71 (s, 3H), 1.25 (s, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 194.1, 142.2, 133.1, 130.7, 128.7, 120.3, 114.9, 92.5, 86.4, 48.5, 47.8, 24.6, 24.1.

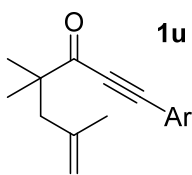
1r is a known compound and our material matched the ^1H and ^{13}C spectra in the literature (D. A. Candito, M. Lautens, *Synlett*, **2011**, 1987-1992).



^1H NMR (400 MHz, CDCl_3): δ 6.99 (s, 2H), δ 6.91 (s, H), 5.73-5.63 (m, H), 5.20 (d, J = 17.2 Hz, H), 5.13 (d, J = 10.2 Hz, H), 4.20 (q, J = 7.0 Hz, 4H), 2.99 (s, 2H), 2.86 (d, J = 7.5 Hz, 2H), 2.26 (s, 6H), 1.26 (t, J = 7.0 Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 169.9, 137.7, 132.0, 129.8, 129.3, 122.8, 119.7, 83.7, 83.4, 61.5, 57.0, 36.5, 23.4, 21.1, 20.9, 14.1.

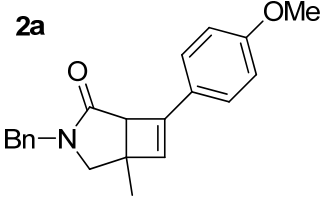


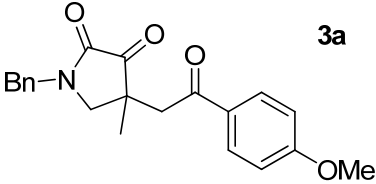
^1H NMR (400 MHz, CDCl_3): δ 7.33 (dt, J = 1.9, 3.5 Hz, 2H), δ 6.96 (t, J = 8.6 Hz, 2H), 5.74-5.63 (m, H), 5.20 (d, J = 16.8 Hz, H), 5.15 (d, J = 9.8 Hz, H), 4.22 (q, J = 7.0 Hz, 4H), 2.99 (s, 2H), 2.85 (d, J = 7.4 Hz, 2H), 1.26 (t, J = 7.0 Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 169.7, 163.6 (d, J = 247.8 Hz), 133.5 (d, J = 8.8 Hz), 131.9, 119.8, 119.4 (d, J = 3.3 Hz), 115.6 (d, J = 22.0 Hz), 84.2, 82.5, 61.7, 57.0, 36.7, 23.5, 14.1.

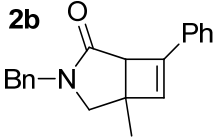


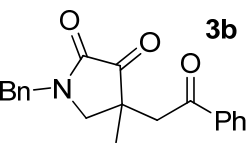
^1H NMR (400 MHz, CDCl_3): δ 7.53 (d, J = 9.0 Hz, 2H), 6.90 (d, J = 8.6 Hz, 2H), 4.83 (s, H), 4.70 (s, 1H), 3.84 (s, 3H), 2.48 (s, 2H), 1.70 (s, 3H), 1.25 (s, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 194.0, 161.5, 142.2, 134.9, 114.6, 114.3, 112.0, 93.5, 86.1, 55.4, 48.1, 47.7, 24.5, 24.0.

7. Characterization of Products

2a  ¹H NMR (400 MHz, CDCl₃): δ 7.58 (d, *J* = 8.5 Hz, 2H), 7.38-7.22 (m, 3H), 7.17 (d, *J* = 7.1 Hz, 2H), 6.89 (d, *J* = 8.6 Hz, 2H), 6.28 (s, H), 4.45 (d of ABq, *J* = 14.8 Hz, H), 4.38 (d of ABq, *J* = 14.9 Hz, H), 3.81 (s, 3H), 3.47 (s, H), 3.23 (d of ABq, *J* = 9.8 Hz, H), 3.07 (d of ABq, 10.6 Hz, H), 1.37 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.7, 159.8, 147.3, 136.5, 130.8, 128.6, 128.0, 127.4, 127.0, 125.9, 113.9, 55.3, 55.0, 54.4, 46.8, 40.2, 22.1; HRMS(EI⁺) Calcd for C₂₁H₂₁O₂N [M]⁺ 319.1567, found 319.1568.

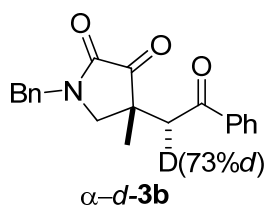
3a  IR (neat): 1763, 1710, 1674 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.84 (d, *J* = 9.0 Hz, 2H), 7.42-7.20 (m, 5H), 6.90 (d, *J* = 8.9 Hz, 2H), 4.81 (d of ABq, *J* = 14.5 Hz, H), 4.68 (d of ABq, *J* = 14.4 Hz, H), 3.86 (s, 3H), 3.57 (d of ABq, *J* = 18.4 Hz, H), 3.44 (d of ABq, *J* = 18.4 Hz, H), 3.39 (d of ABq, *J* = 10.5 Hz, H), 3.17 (d of ABq, *J* = 10.5 Hz, H), 1.23 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 202.3, 194.8, 164.1, 159.7, 134.8, 130.5, 128.9, 128.36, 128.33, 128.1, 113.9, 55.5, 53.6, 48.3, 47.4, 40.0, 23.5; HRMS(FAB⁺) Calcd for C₂₁H₂₂O₄N [M+1]⁺ 352.1543, found 352.1544; white solid; mp 178-182 °C.

2b  ¹H NMR (400 MHz, CDCl₃): δ 7.64 (d, *J* = 7.1 Hz, 2H), 7.37 (t, *J* = 7.0 Hz, 2H), 7.34-7.22 (m, 4H), 7.17 (d, *J* = 6.6 Hz, 2H), 6.44 (s, H), 4.45 (d of ABq, *J* = 14.8 Hz, H), 4.39 (d of ABq, *J* = 14.5 Hz, H), 3.51 (s, H), 3.25 (dd, *J* = 1.6, 10.5 Hz, H), 3.08 (d, 10.2 Hz, H), 1.39 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.7, 148.0, 136.6, 133.6, 133.0, 128.8, 128.6, 128.2, 127.6, 125.6, 55.2, 54.3, 47.0, 40.6, 22.2; HRMS(FAB⁺) Calcd for C₂₀H₂₀ON [M+1]⁺ 290.1539, found 290.1543.

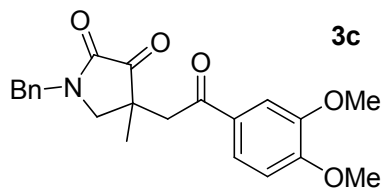
3b  ¹H NMR (400 MHz, CDCl₃): δ 7.87 (d, *J* = 7.4 Hz, 2H), 7.58 (t, *J* = 7.8 Hz, H), 7.45 (t, *J* = 7.8 Hz, 2H), 7.40-7.28 (m, 5H), 4.80 (d of ABq, *J* = 14.4 Hz, H), 4.72 (d of ABq, *J* = 14.4 Hz, H), 3.62 (d of ABq, *J* = 18.4 Hz, H), 3.48 (d of ABq, *J* = 18.4 Hz, H), 3.40 (d of ABq, *J* = 10.5 Hz, H), 3.19 (d of ABq, *J* = 10.5 Hz, H), 1.23 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 202.4, 196.5, 159.6, 135.2, 134.7, 133.9, 128.9, 128.8, 128.3, 128.2, 128.1, 53.5, 48.2, 47.6, 40.0, 23.5; HRMS(FAB⁺) Calcd for C₂₀H₂₀O₃N [M+1]⁺ 322.1438, found 322.1438; white solid; mp 148-152 °C.

(Obtained from (*E*)-**1b**) ¹H NMR (400 MHz, CDCl₃): δ 7.87 (d, *J* = 7.4 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, H), 7.45 (t, *J* = 7.5 Hz, 2H), 7.40-7.28 (m, 5H), 4.80 (d of ABq, *J* = 14.8 Hz, H), 4.72 (d of ABq, *J* = 14.8 Hz, H), 3.62 (d of ABq, *J* = 18.4 Hz, 0.32H of **3b**), 3.48 (d of ABq, *J* = 18.4 Hz, 0.32H of **3b**), 3.47 (s, 0.68H of *d*-**3b**), 3.39 (d of ABq, *J* = 10.5 Hz, H), 3.19 (d of ABq, *J* = 10.5 Hz, H), 1.23 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 202.4, 196.6, 159.7, 135.4, 134.8, 134.1, 129.0, 128.9, 128.5, 128.3, 128.2, 53.6, 48.4, 47.5(**3b**), 47.3

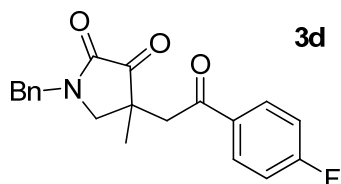
(t, $J_{CD} = 19.1$ Hz, **d-3b**), 40.0, 23.6.



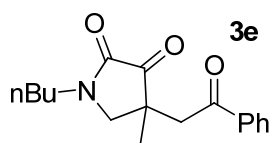
(Obtained from (Z)-**1b**) ^1H NMR (400 MHz, CDCl_3): δ 7.86 (d, $J = 7.4$ Hz, 2H), 7.58 (t, $J = 7.5$ Hz, H), 7.45 (t, $J = 7.4$ Hz, 2H), 7.40-7.28 (m, 5H), 4.80 (d of ABq, $J = 14.5$ Hz, H), 4.72 (d of ABq, $J = 14.5$ Hz, H), 3.62 (d of ABq, $J = 18.3$ Hz, 0.26H of **3b**), 3.59 (s, 0.69H of **d-3b**), 3.48 (d of ABq, $J = 18.4$ Hz, 0.26H of **3b**), 3.39 (d of ABq, $J = 10.5$ Hz, H), 3.19 (d of ABq, $J = 10.5$ Hz, H), 1.23 (s, 3H).



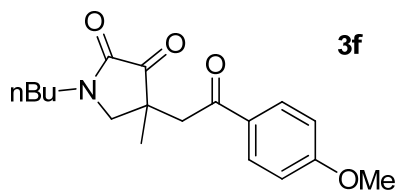
^1H NMR (400 MHz, CDCl_3): δ 7.54-7.46 (m, H), 7.44-7.28 (m, 6H), 6.85 (d, $J = 8.6$ Hz, H), 4.86 (d of ABq, $J = 14.4$ Hz, H), 4.65 (d of ABq, $J = 14.8$ Hz, H), 3.95 (s, 3H), 3.91 (s, 3H), 3.58 (d of ABq, $J = 18.3$ Hz, H), 3.46 (d of ABq, $J = 18.4$ Hz, H), 3.40 (d of ABq, $J = 10.5$ Hz, H), 3.19 (d of ABq, $J = 10.5$ Hz, H), 1.24 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 202.3, 195.1, 159.6, 153.9, 149.1, 134.8, 128.9, 128.5, 128.3, 128.1, 123.1, 110.02, 110.00, 56.1, 56.0, 53.6, 48.3, 47.3, 40.0, 23.5; HRMS(FAB+) Calcd for $\text{C}_{22}\text{H}_{24}\text{O}_5\text{N}$ $[\text{M}+1]^+$ 382.1649, found 382.1654; sticky oil.



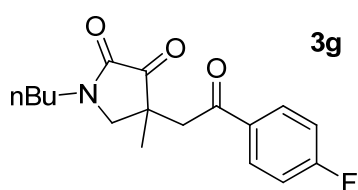
^1H NMR (400 MHz, CDCl_3): δ 7.90 (dd, $J = 5.5, 8.6$ Hz, 2H), 7.44-7.22 (m, 5H), 7.12 (t, $J = 8.6$ Hz, 2H), 4.79 (d of ABq, $J = 14.4$ Hz, H), 4.73 (d of ABq, $J = 14.8$ Hz, H), 3.58 (d of ABq, $J = 18.3$ Hz, H), 3.44 (d of ABq, $J = 18.8$ Hz, H), 3.39 (d of ABq, $J = 10.9$ Hz, H), 3.19 (d of ABq, $J = 10.5$ Hz, H), 1.25 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 202.2, 194.8, 166.2 (d, $J = 255.8$ Hz), 159.5, 134.7, 131.7, (d, $J = 2.9$ Hz), 130.9 (d, $J = 9.5$ Hz), 128.9, 128.3, 128.1, 115.9 (d, $J = 22.1$ Hz), 54.5, 48.3, 47.3, 40.0, 23.6; HRMS(FAB+) Calcd for $\text{C}_{20}\text{H}_{19}\text{O}_3\text{NF}$ $[\text{M}+1]^+$ 340.1343, found 340.1342; white solid; mp 186-190 $^\circ\text{C}$.



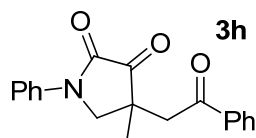
^1H NMR (400 MHz, CDCl_3): δ 7.90 (d, $J = 7.4$ Hz, 2H), 7.59 (t, $J = 7.4$ Hz, H), 7.46 (t, $J = 7.8$ Hz, 2H), 3.37-3.47 (m, 5H), 3.35 (d of ABq, $J = 10.5$ Hz, H), 1.74-1.54 (m, 2H), 1.40 (sextet, $J = 7.4$ Hz, 2H), 1.31 (s, 3H), 0.96 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 202.5, 196.5, 159.5, 135.4, 133.9, 128.8, 128.2, 54.1, 47.1, 44.2, 40.1, 28.8, 23.7, 19.9, 13.7; HRMS(FAB+) Calcd for $\text{C}_{17}\text{H}_{22}\text{O}_3\text{N}$ $[\text{M}+1]^+$ 288.1594, found 288.1594; white solid; mp 123-125 $^\circ\text{C}$.



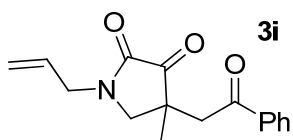
^1H NMR (400 MHz, CDCl_3): δ 7.87 (d, $J = 8.6$ Hz, 2H), 6.94 (d, $J = 8.6$ Hz, 2H), 3.87 (s, 3H), 3.62-3.47 (m, 5H), 3.33 (d, $J = 10.5$ Hz, H), 1.74-1.54 (m, 2H), 1.48-1.34 (m, 2H), 1.30 (s, 3H), 0.96 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 202.6, 194.9, 164.1, 159.6, 130.5, 128.5, 113.9, 55.5, 54.2, 46.9, 44.2, 40.1, 28.8, 23.7, 19.9, 13.7; HRMS(FAB+) Calcd for $\text{C}_{18}\text{H}_{24}\text{O}_4\text{N}$ $[\text{M}+1]^+$ 318.1700, found 318.1699; white solid; mp 158-162 $^\circ\text{C}$.



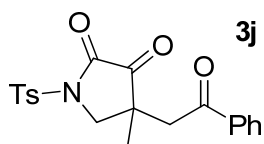
3g ^1H NMR (400 MHz, CDCl_3): δ 7.94 (dd, J = 5.1, 8.6 Hz, 2H), 7.13 (t, J = 8.6 Hz, 2H), 3.66-3.46 (m, 5H) 3.35 (d of ABq, J = 10.6 Hz, H), 1.74-1.54 (m, 2H), 1.41 (sextet, J = 7.0 Hz, 2H), 1.31 (s, 3H), 0.96 (t, J = 7.4 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) : δ 202.5, 194.9, 166.2 (d, J = 255.8 Hz), 159.4, 131.8 (d, J = 3.0 Hz), 130.9 (d, J = 9.6 Hz), 115.9 (d, J = 22.1 Hz), 54.1, 47.0, 44.2, 40.1, 28.8, 23.7, 19.9, 13.7; HRMS(FAB+) Calcd for $\text{C}_{17}\text{H}_{21}\text{O}_3\text{NF}$ $[\text{M}+1]^+$ 306.1500, found 306.1501; white solid; mp 164-165 $^\circ\text{C}$.



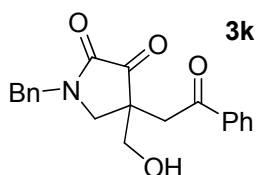
3h ^1H NMR (400 MHz, CDCl_3): δ 7.92 (d, J = 7.8 Hz, 2H), 7.86 (d, J = 7.8 Hz, 2H), 7.60 (t, J = 7.5 Hz, H), 7.47 (t, J = 7.9 Hz, 2H), 7.46 (t, J = 8.2 Hz, 2H), 7.28 (t, J = 7.4 Hz, H), 4.03 (d of ABq, J = 10.1 Hz, H), 3.83 (d of ABq, J = 10.1 Hz, H), 3.74 (d of ABq, J = 18.7 Hz, H), 3.67 (d of ABq, J = 18.8 Hz, H), 1.43 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 201.6, 196.5, 158.1, 138.9, 135.2, 134.1, 129.2, 128.8, 128.3, 119.5, 126.5, 55.0, 47.7, 39.8, 23.8; HRMS(FAB+) Calcd for $\text{C}_{19}\text{H}_{18}\text{O}_3\text{N}$ $[\text{M}+1]^+$ 308.1281, found 308.1283; white solid; mp 162-164 $^\circ\text{C}$.



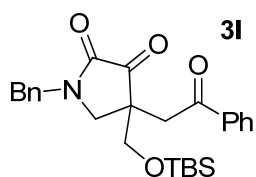
3i ^1H NMR (400 MHz, CDCl_3): δ 7.90 (d, J = 7.4 Hz, 2H), 7.60 (t, J = 7.4 Hz, H), 7.46 (t, J = 7.4 Hz, 2H), 5.81 (ddd, J = 5.8, 10.6, 17.2 Hz, H), 5.33 (d, J = 17.5 Hz, H), 5.29 (d, J = 10.6 Hz, H), 4.19 (d, J = 6.2 Hz, 2H), 3.65 (d of ABq, J = 18.8 Hz, H), 3.55 (d of ABq, J = 18.7 Hz, H), 3.53 (d of ABq, J = 10.5 Hz, H), 3.32 (d of ABq, J = 10.5 Hz, H), 1.32 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) : δ 202.4, 196.7, 159.4, 135.4, 134.1, 130.9, 128.9, 128.3, 119.6, 53.9, 47.6, 47.1, 40.2, 23.8; HRMS(FAB+) Calcd for $\text{C}_{16}\text{H}_{18}\text{O}_3\text{N}$ $[\text{M}+1]^+$ 272.1281, found 272.1281; sticky oil.



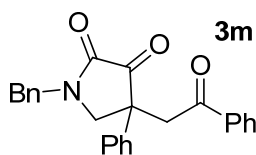
3j ^1H NMR (400 MHz, CDCl_3): δ 7.99 (d, J = 8.2 Hz, 2H), 7.77 (d, J = 7.0 Hz, 2H), 7.59 (t, J = 7.4 Hz, H), 7.44 (t, J = 7.8 Hz, 2H), 7.37 (d, J = 8.2 Hz, 2H), 3.92 (d of ABq, J = 10.6 Hz, H), 3.72 (d of ABq, J = 10.2 Hz, H), 3.68 (d of ABq, J = 18.7 Hz, H), 3.60 (d of ABq, J = 18.7 Hz, H), 2.47 (s, 3H), 1.20 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 199.0, 196.5, 156.7, 146.1, 134.7, 134.4, 133.9, 130.0, 128.9, 128.6, 128.4, 53.5, 49.8, 40.7, 23.2, 21.9; HRMS(FAB+) Calcd for $\text{C}_{20}\text{H}_{20}\text{NO}_5\text{S}$ $[\text{M}+1]^+$ 386.1057, found 386.1062; sticky oil.



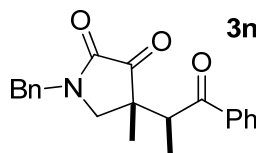
3k ^1H NMR (400 MHz, CDCl_3): δ 7.85 (d, J = 7.4 Hz, 2H), 7.56 (t, J = 7.4 Hz, H), 7.42 (t, J = 7.4 Hz, 2H), 7.38-7.28 (m, 5H), 4.82 (d of ABq, J = 14.8 Hz, H), 4.69 (d of ABq, J = 14.9 Hz, H), 3.88-3.62 (m, 3H), 3.52 (d of ABq, J = 19.1 Hz, 1H), 3.47 (d of ABq, J = 11.3 Hz, H), 3.28 (d of ABq, J = 10.9 Hz, H); ^{13}C NMR (100 MHz, CDCl_3): δ 202.2, 196.7, 160.4, 135.3, 134.7, 134.1, 129.0, 128.9, 128.6, 128.4, 128.2, 66.0, 49.8, 48.5, 46.1, 44.4; HRMS(FAB+) Calcd for $\text{C}_{20}\text{H}_{20}\text{O}_4\text{N}$ $[\text{M}+1]^+$ 338.1387, found 338.1392; colorless oil.



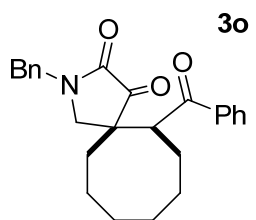
3l ^1H NMR (400 MHz, CDCl_3): δ 7.85 (d, $J = 7.4$ Hz, 2H), 7.58 (t, $J = 7.4$ Hz, H), 7.45 (t, $J = 7.8$ Hz, 2H), 7.40-7.26 (m, 5H), 4.79 (d of ABq, $J = 14.8$ Hz, H), 4.74 (d of ABq, $J = 14.8$ Hz, H), 3.73 (d of ABq, $J = 9.0$ Hz, H), 3.60 (d of ABq, $J = 9.0$ Hz, 1H), 3.58 (d of ABq, $J = 9.3$ Hz, H), 3.51 (s, 2H), 3.28 (d of ABq, $J = 10.6$ Hz, H), 0.81 (s, 9H), 0.00 (s, 3H), -0.04 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 202.2, 196.2, 160.1, 135.4, 134.9, 134.1, 128.95, 128.90, 128.6, 128.3, 128.1, 67.4, 49.8, 48.5, 46.1, 44.0, 25.8, 18.3, -5.4; HRMS(FAB+) Calcd for $\text{C}_{26}\text{H}_{34}\text{O}_4\text{NSi}$ $[\text{M}+1]^+$ 452.2252, found 452.2257; colorless oil.



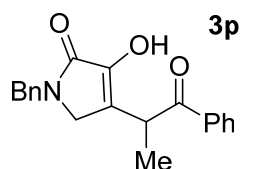
3m ^1H NMR (400 MHz, CDCl_3): δ 7.88 (d, $J = 7.4$ Hz, 2H), 7.59 (t, $J = 7.4$ Hz, H), 7.45 (t, $J = 7.4$ Hz, 2H), 7.40-7.24 (m, 8H), 7.24-7.18 (m, 2H), 4.82 (d of ABq, $J = 14.5$ Hz, H), 4.77 (d of ABq, $J = 14.5$ Hz, H), 3.97 (d of ABq, $J = 18.3$ Hz, H), 3.93 (d of ABq, $J = 10.9$ Hz, H), 3.90 (d of ABq, $J = 18.7$ Hz, H), 3.71 (d of ABq, $J = 10.9$ Hz, H); ^{13}C NMR (100 MHz, CDCl_3): δ 199.2, 196.4, 159.7, 138.1, 135.5, 134.6, 134.1, 129.4, 129.0, 128.9, 128.7, 128.3, 128.2, 126.2, 53.6, 48.6, 48.3, 48.1; HRMS(EI+) Calcd for $\text{C}_{25}\text{H}_{21}\text{O}_3\text{N}$ $[\text{M}+1]^+$ 383.1516, found 383.1518; sticky oil.



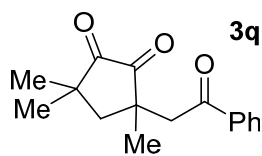
3n ^1H NMR (400 MHz, CDCl_3): δ 7.90 (d, $J = 7.4$ Hz, 2H), 7.60 (t, $J = 7.4$ Hz, H), 7.48 (t, $J = 7.8$ Hz, 2H), 7.40-7.24 (m, 5H), 4.73 (d of ABq, $J = 14.9$ Hz, H), 4.69 (d of ABq, $J = 14.9$ Hz, H), 3.93 (q, $J = 7.4$ Hz, 1H), 3.64 (d of ABq, $J = 11.7$ Hz, H), 3.31 (d of ABq, $J = 11.7$ Hz, H), 1.23 (d, $J = 7.4$ Hz, 3H), 1.22 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 202.7, 201.6, 159.4, 135.0, 134.6, 133.9, 128.9, 128.6, 128.4, 128.3, 128.2, 52.5, 48.8, 48.4, 43.8, 22.7, 14.0; HRMS(FAB+) Calcd for $\text{C}_{21}\text{H}_{22}\text{O}_3\text{N}$ $[\text{M}+1]^+$ 336.1594, found 336.1596; sticky oil.



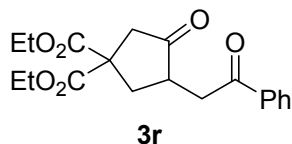
3o ^1H NMR (400 MHz, CDCl_3): δ 7.82 (d, $J = 7.4$ Hz, 2H), 7.58 (t, $J = 7.4$ Hz, H), 7.45 (t, $J = 7.4$ Hz, 2H), 7.28-7.21 (m, 5H), 4.73 (d of ABq, $J = 14.9$ Hz, H), 4.59 (d of ABq, $J = 14.9$ Hz, H), 3.90 (d, $J = 5.4$ Hz, H), 3.25 (d of ABq, $J = 11.0$ Hz, H), 2.81 (d of ABq, $J = 11.0$ Hz, H), 2.65-2.57 (m, H), 2.00 (t, $J = 13.1$ Hz, H), 1.86-1.67 (m, 8H), 1.60-1.52 (m, 2H), 1.47-1.43 (m, H); ^{13}C NMR (100 MHz, CDCl_3): δ 201.1, 200.9, 160.6, 135.0, 134.8, 133.8, 128.97, 128.93, 128.90, 128.2, 55.3, 48.1, 44.9, 31.2, 28.1, 27.6, 25.7, 24.9, 23.2; HRMS(FAB+) Calcd for $\text{C}_{25}\text{H}_{28}\text{O}_3\text{N}$ $[\text{M}+1]^+$ 390.2064, found 390.2069; sticky oil.



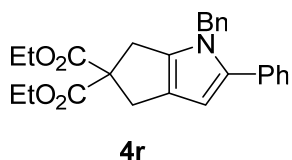
3p ^1H NMR (400 MHz, CDCl_3): δ 8.05 (d, $J = 7.8$ Hz, 2H), 7.56 (t, $J = 7.4$ Hz, H), 7.45 (t, $J = 7.4$ Hz, 2H), 7.35-7.27 (m, 3H), 7.17 (d, $J = 6.6$ Hz, 2H), 4.88 (q, $J = 7.0$ Hz, H), 4.69 (d of ABq, $J = 14.9$ Hz, H), 4.46 (d of ABq, $J = 14.9$ Hz, H), 3.72 (d of ABq, $J = 18.4$ Hz, H), 3.55 (d of ABq, $J = 18.4$ Hz, H), 1.35 (d, $J = 6.7$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 199.9, 167.3, 142.0, 136.4, 135.7, 133.6, 128.8, 128.7, 128.6, 128.0, 127.7, 119.9, 47.08, 37.9, 30.1, 16.2; HRMS(FAB+) Calcd for $\text{C}_{20}\text{H}_{20}\text{NO}_3$ $[\text{M}+1]^+$ 322.1438, found 322.1443; thick oil.



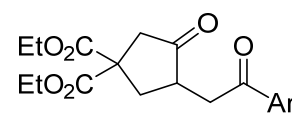
3q ^1H NMR (400 MHz, CDCl_3): δ 7.88 (d, $J = 7.4$ Hz, 2H), 7.58 (t, $J = 7.4$ Hz, H), 7.44 (d, $J = 7.8$ Hz, 2H), 3.75 (d of ABq, $J = 18.7$ Hz, H), 3.64 (d of ABq, $J = 18.4$ Hz, H), 2.00 (d, $J = 13.3$ Hz, H), 1.91 (d, $J = 13.3$ Hz, H), 1.32 (s, 3H), 1.29 (s, 3H), 1.20 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 208.5, 206.5, 198.0, 134.9, 134.0, 128.7, 128.4, 54.8, 46.0, 41.2, 40.4, 27.1, 25.24, 25.23; HRMS(FAB+) Calcd for $\text{C}_{16}\text{H}_{19}\text{O}_3$ $[\text{M}+1]^+$ 259.1329, found 259.1334; yellow solid, mp 135-137 $^\circ\text{C}$.



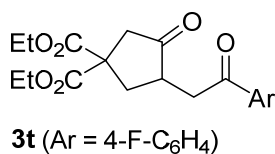
3r ^1H NMR (400 MHz, CDCl_3): δ 7.93 (d, $J = 7.4$ Hz, 2H), 7.57 (t, $J = 7.4$ Hz, H), 7.46 (t, $J = 7.4$ Hz, 2H), 4.28-4.19 (m, 4H), 3.51 (dd, $J = 2.8, 18.3$ Hz, H), 3.24 (dd, $J = 6.6, 18.7$ Hz, H), 3.05 (d of ABq, $J = 18.7$ Hz, H), 2.98 (d of ABq, $J = 18.7$ Hz, H), 2.92-2.84 (m, 2H), 2.24 (t, $J = 15.6$ Hz, H), 1.27 (t, $J = 7.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 215.1, 197.1, 171.3, 170.7, 136.3, 133.5, 128.7, 128.1, 62.2, 62.1, 55.2, 44.7, 43.3, 38.6, 35.7, 14.1. HRMS (EI+) Calcd for $\text{C}_{19}\text{H}_{22}\text{O}_6$ $[\text{M}]^+$ 346.1411, found 346.1414; colorless oil.



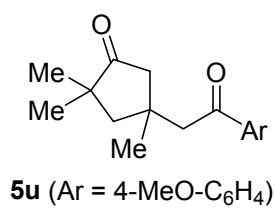
4r ^1H NMR (400 MHz, CDCl_3): δ 7.31-7.21 (m, 8H), 6.99 (d, $J = 7.0$ Hz, 2H), 6.04 (s, H), 5.04 (s, 2H), 4.17 (q, $J = 7.0$ Hz, 4H), 3.37 (s, 2H), 3.20 (s, 2H), 1.22 (t, $J = 7.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 171.8, 138.8, 138.4, 135.6, 133.6, 128.8, 128.6, 128.5, 127.3, 126.8, 126.3, 122.6, 103.8, 64.8, 61.7, 49.2, 34.7, 33.8, 14.1. HRMS (EI+) Calcd for $\text{C}_{26}\text{H}_{27}\text{NO}_4$ $[\text{M}]^+$ 417.1935, found 417.1939; sticky oil.



3s (Ar = 3,5-di-Me- C_6H_3) ^1H NMR (400 MHz, CDCl_3): δ 7.53 (s, 2H), 7.20 (s, H), 6.84 (d, $J = 20.7$ Hz, H), 4.27-4.18 (m, 4H), 3.48 (dd, $J = 2.7, 18.3$ Hz, H), 3.19 (dd, $J = 6.6, 18.3$ Hz, H), 3.05 (d of ABq, $J = 18.7$ Hz, H), 2.97 (d of ABq, $J = 18.7$ Hz, H), 2.92-2.82 (m, 2H), 2.68 (d, $J = 16.0$ Hz, H), 2.35 (s, 3H), 2.27 (s, 3H), 1.36 (t, $J = 7.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 215.1, 204.4, 197.6, 171.3, 170.7, 170.3, 170.1, 139.2, 138.4, 137.7, 136.5, 135.1, 129.2, 128.0, 125.9, 62.2, 62.15, 62.10, 57.8, 55.3, 44.7, 43.4, 41.6, 38.7, 38.3, 35.8, 30.1, 25.7, 24.9, 21.39, 21.33, 14.1, 14.0. HRMS (EI+) Calcd for $\text{C}_{21}\text{H}_{26}\text{O}_6$ $[\text{M}]^+$ 374.1724, found 374.1728; colorless oil.



3t (Ar = 4-F- C_6H_4) ^1H NMR (400 MHz, CDCl_3): δ 7.98-7.94 (m, 2H), 7.13 (t, $J = 8.6$ Hz, 2H), 4.28-4.21 (m, 4H), 3.46 (dd, $J = 3.2, 18.3$ Hz, H), 3.20 (dd, $J = 7.0, 18.7$ Hz, H), 3.05 (d of ABq, $J = 18.7$ Hz, H), 2.97 (d of ABq, $J = 18.7$ Hz, H), 2.92-2.84 (m, 2H), 2.23 (t, $J = 15.6$ Hz, H), 1.27 (t, $J = 7.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 214.9, 195.6, 171.2, 170.7, 166.0 (d, $J = 260$ Hz), 132.9 (d, $J = 8.8$ Hz), 130.8 (d, $J = 3.7$ Hz), 116.0 (d, $J = 22.0$ Hz), 62.2, 62.1, 55.3, 44.6, 43.3, 38.5, 35.7, 29.8, 14.1; HRMS (EI+) Calcd for $\text{C}_{19}\text{H}_{21}\text{FO}_6$ $[\text{M}]^+$ 364.1317, found 364.1321; colorless oil.



¹H NMR (400 MHz, CDCl₃): δ 7.91 (d, *J* = 9.0 Hz, 2H), 6.91 (d, *J* = 8.2 Hz, 2H), 3.87 (s, 3H), 3.05 (s, 2H), 2.60 (d of ABq, *J* = 17.5 Hz, H), 2.33 (d of ABq, *J* = 17.5 Hz, H), 2.06 (d of ABq, *J* = 13.3 Hz, H), 1.93 (d of ABq, *J* = 13.7 Hz, H), 1.22 (s, 3H), 1.19 (s, 3H), 1.15 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 223.3, 197.4, 163.6, 131.0, 130.4, 113.8, 55.6, 51.2, 51.0, 49.2, 45.0, 35.4, 28.6, 27.8, 27.3. HRMS (EI⁺)

Calcd for C₁₇H₂₂O₃ [M]⁺ 274.1563, found 274.1568; colorless oil.

8. X-ray crystallographic analysis data for compound 3e

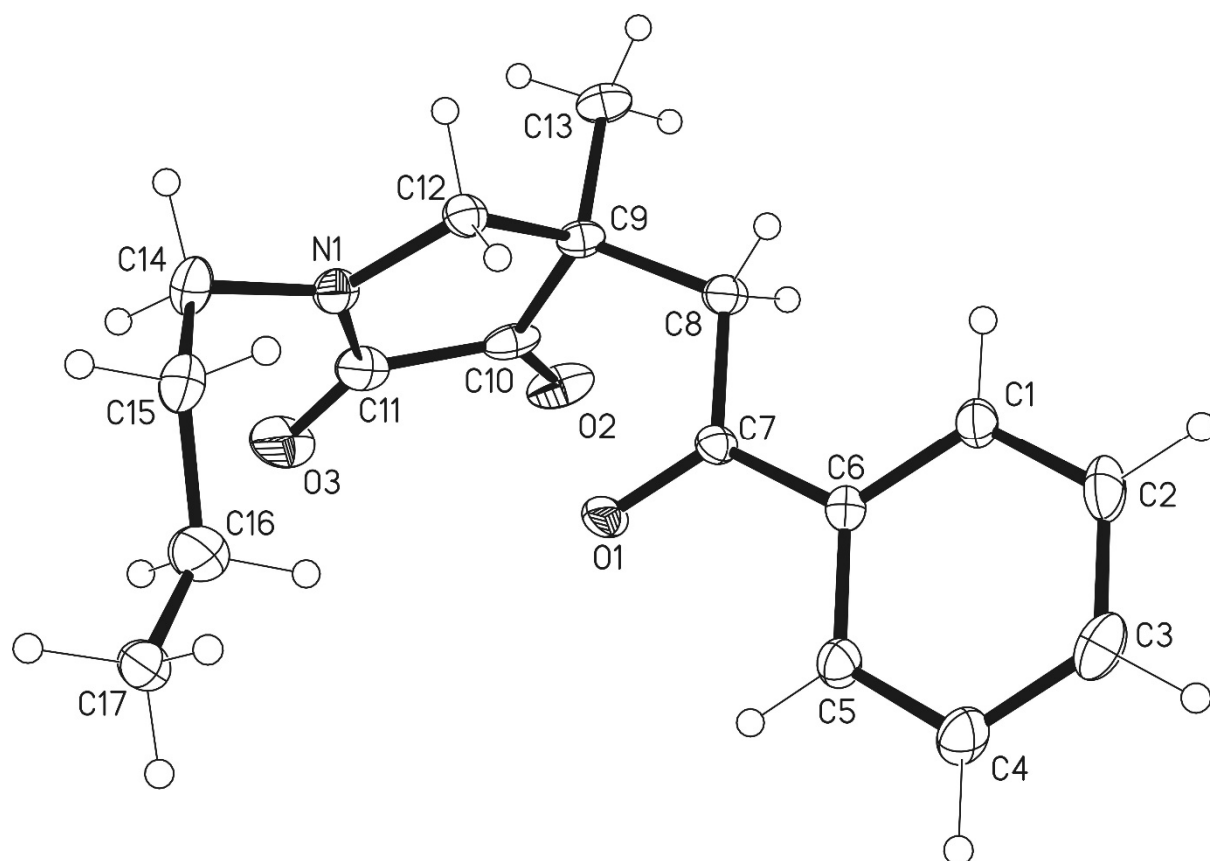


Table S2. Crystal data and structure refinement for **3e**

Identification code	3e	
Empirical formula	C ₁₇ H ₂₁ N O ₃	
Formula weight	287.35	
Temperature	173(2) K	
Wavelength	0.71073 Å	
Crystal system	Hexagonal	
Space group	P6(5)	
Unit cell dimensions	a = 11.2648(8) Å	α = 90°.
	b = 11.2648(8) Å	β = 90°.
	c = 21.2304(15) Å	γ = 120°.
Volume	2333.1(3) Å ³	
Z	6	
Density (calculated)	1.227 Mg/m ³	
Absorption coefficient	0.084 mm ⁻¹	
F(000)	924	
Crystal size	0.32 x 0.09 x 0.08 mm ³	
Theta range for data collection	2.09 to 27.58°.	
Index ranges	-14 ≤ h ≤ 7, -13 ≤ k ≤ 13, -27 ≤ l ≤ 27	
Reflections collected	14031	
Independent reflections	3606 [R(int) = 0.0604]	
Completeness to theta = 27.58°	99.9 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.9933 and 0.9737	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	3606 / 1 / 199	
Goodness-of-fit on F ²	1.032	
Final R indices [I > 2σ(I)]	R1 = 0.0563, wR2 = 0.1160	
R indices (all data)	R1 = 0.1063, wR2 = 0.1336	
Absolute structure parameter	-1.6(17)	
Largest diff. peak and hole	0.159 and -0.203 e.Å ⁻³	

Table S3. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for **3e**. U(eq) is defined as one third of the trace of the orthogonalized U_{ij} tensor.

	x	y	z	U(eq)
O(1)	9040(2)	7767(2)	1488(1)	48(1)
O(2)	8741(3)	5159(2)	2171(1)	66(1)
O(3)	6414(2)	4074(2)	1317(2)	83(1)
N(1)	8133(2)	5151(2)	586(1)	46(1)
C(1)	12487(3)	10650(3)	1556(1)	43(1)
C(2)	13153(3)	12061(3)	1598(1)	56(1)
C(3)	12434(4)	12722(3)	1668(1)	58(1)
C(4)	11017(3)	11986(3)	1703(2)	53(1)
C(5)	10347(3)	10586(2)	1660(1)	42(1)
C(6)	11066(2)	9893(2)	1582(1)	32(1)
C(7)	10290(3)	8376(2)	1517(1)	34(1)
C(8)	11029(3)	7581(3)	1476(1)	40(1)
C(9)	10130(3)	6126(3)	1236(1)	37(1)
C(10)	8828(3)	5358(3)	1609(1)	46(1)
C(11)	7621(3)	4774(3)	1161(2)	55(1)
C(12)	9609(2)	6082(3)	562(1)	40(1)
C(13)	10918(3)	5339(3)	1277(2)	53(1)
C(14)	7317(3)	4811(3)	13(2)	66(1)
C(15)	7165(4)	5993(4)	-240(2)	71(1)
C(16)	6353(5)	6391(5)	181(2)	94(1)
C(17)	5866(7)	7223(7)	-125(3)	50(2)
C(17')	5131(13)	5940(20)	139(7)	199(9)

Table S4. Bond lengths [Å] and angles [°] for **3e**.

O(1)-C(7)	1.221(3)
O(2)-C(10)	1.209(3)
O(3)-C(11)	1.228(3)
N(1)-C(11)	1.326(4)
N(1)-C(14)	1.456(4)
N(1)-C(12)	1.457(3)
C(1)-C(2)	1.380(4)
C(1)-C(6)	1.388(3)
C(2)-C(3)	1.355(5)
C(3)-C(4)	1.384(5)
C(4)-C(5)	1.370(4)
C(5)-C(6)	1.388(4)
C(6)-C(7)	1.486(3)
C(7)-C(8)	1.499(3)
C(8)-C(9)	1.520(4)
C(9)-C(10)	1.502(4)
C(9)-C(12)	1.539(4)
C(9)-C(13)	1.539(4)
C(10)-C(11)	1.514(5)
C(14)-C(15)	1.522(5)
C(15)-C(16)	1.498(5)
C(16)-C(17')	1.210(12)
C(16)-C(17)	1.455(7)
C(11)-N(1)-C(14)	124.6(3)
C(11)-N(1)-C(12)	114.8(2)
C(14)-N(1)-C(12)	120.2(2)
C(2)-C(1)-C(6)	120.0(3)
C(3)-C(2)-C(1)	120.7(3)
C(2)-C(3)-C(4)	120.3(3)
C(5)-C(4)-C(3)	119.5(3)
C(4)-C(5)-C(6)	121.0(3)
C(5)-C(6)-C(1)	118.5(2)
C(5)-C(6)-C(7)	119.0(2)
C(1)-C(6)-C(7)	122.5(2)
O(1)-C(7)-C(6)	120.1(2)

O(1)-C(7)-C(8)	119.3(2)
C(6)-C(7)-C(8)	120.6(2)
C(7)-C(8)-C(9)	113.2(2)
C(10)-C(9)-C(8)	112.1(2)
C(10)-C(9)-C(12)	103.0(2)
C(8)-C(9)-C(12)	112.6(2)
C(10)-C(9)-C(13)	108.5(2)
C(8)-C(9)-C(13)	109.7(2)
C(12)-C(9)-C(13)	110.7(2)
O(2)-C(10)-C(9)	126.3(3)
O(2)-C(10)-C(11)	124.6(3)
C(9)-C(10)-C(11)	109.0(2)
O(3)-C(11)-N(1)	128.3(3)
O(3)-C(11)-C(10)	125.0(3)
N(1)-C(11)-C(10)	106.7(2)
N(1)-C(12)-C(9)	105.6(2)
N(1)-C(14)-C(15)	113.4(3)
C(16)-C(15)-C(14)	113.8(4)
C(17')-C(16)-C(17)	61.2(10)
C(17')-C(16)-C(15)	124.8(7)
C(17)-C(16)-C(15)	114.2(4)

Symmetry transformations used to generate equivalent atoms:

Table S5. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for **3e**. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

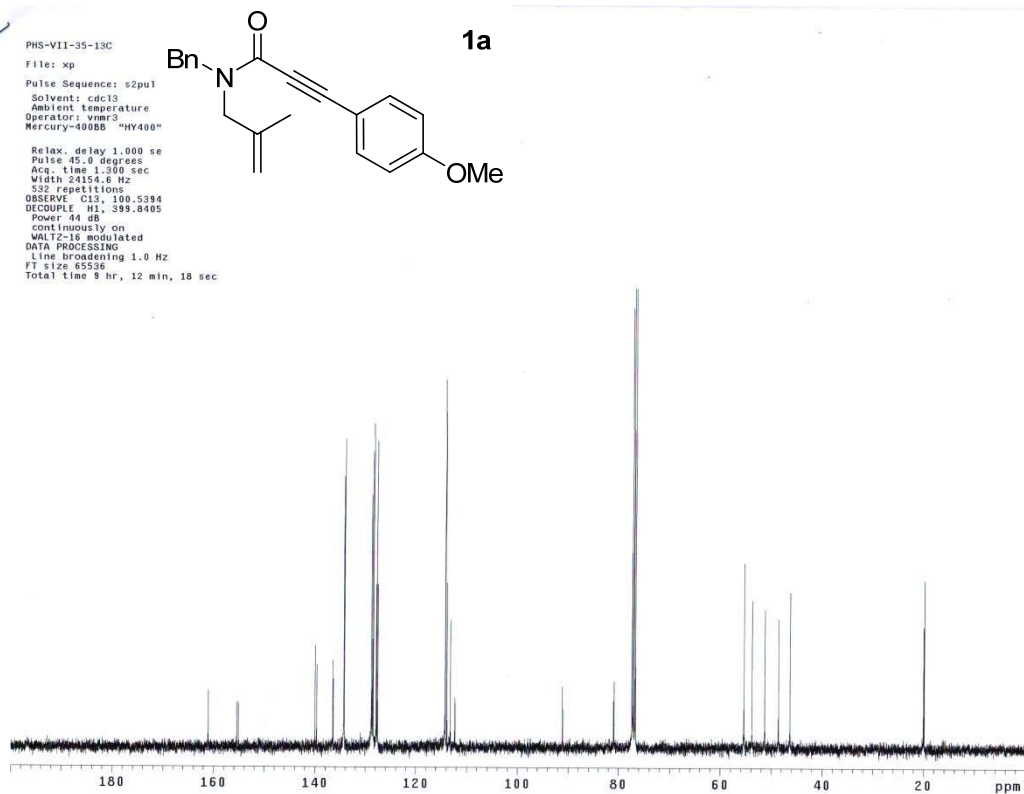
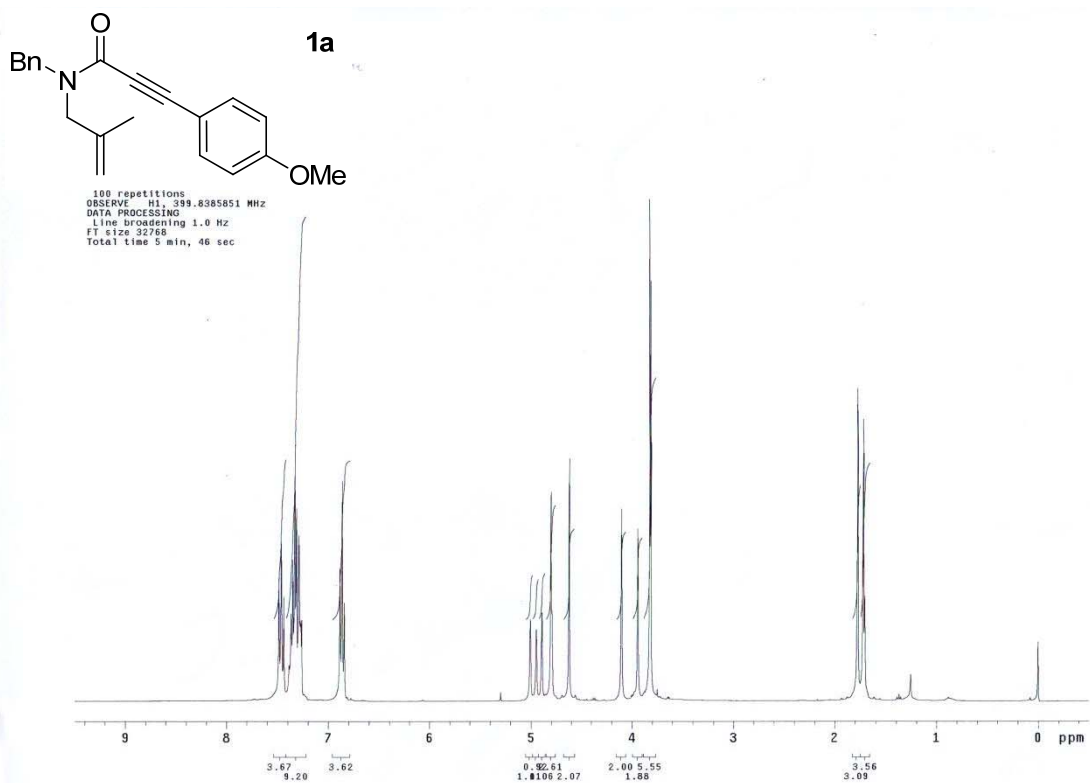
	U^{11}	U^{22}	U^{33}	U^{23}	U^{13}	U^{12}
O(1)	28(1)	32(1)	78(1)	-3(1)	9(1)	12(1)
O(2)	104(2)	54(1)	53(1)	19(1)	35(1)	48(1)
O(3)	47(1)	46(1)	131(2)	9(1)	40(1)	6(1)
N(1)	37(1)	29(1)	63(2)	-4(1)	8(1)	9(1)
C(1)	35(2)	41(2)	44(2)	-6(1)	0(1)	12(1)
C(2)	40(2)	45(2)	53(2)	-4(1)	3(1)	-2(1)
C(3)	73(2)	31(2)	56(2)	-1(1)	-4(2)	16(2)
C(4)	60(2)	34(2)	67(2)	0(1)	-11(2)	24(2)
C(5)	41(2)	33(1)	51(2)	0(1)	-5(1)	16(1)
C(6)	33(1)	32(1)	28(1)	2(1)	0(1)	12(1)
C(7)	30(1)	34(1)	36(1)	3(1)	5(1)	15(1)
C(8)	39(2)	40(2)	43(2)	0(1)	4(1)	22(1)
C(9)	46(2)	34(1)	37(1)	6(1)	9(1)	25(1)
C(10)	66(2)	30(1)	49(2)	12(1)	27(2)	29(1)
C(11)	44(2)	25(1)	87(2)	5(2)	20(2)	10(1)
C(12)	33(1)	33(1)	48(2)	2(1)	8(1)	12(1)
C(13)	67(2)	52(2)	57(2)	0(1)	7(2)	43(2)
C(14)	52(2)	55(2)	75(2)	-29(2)	-14(2)	14(2)
C(15)	67(2)	84(3)	68(2)	-36(2)	-28(2)	42(2)
C(16)	89(3)	143(4)	83(3)	-45(3)	-30(2)	83(3)
C(17)	44(4)	62(4)	48(3)	-5(3)	-2(3)	31(3)
C(17')	112(10)	350(20)	159(13)	-157(15)	-79(10)	132(14)

Table S6. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^{-3}$) for **3e**.

	x	y	z	U(eq)
H(1A)	13002	10196	1510	52
H(2A)	14125	12574	1577	68
H(3A)	12904	13694	1694	69
H(4A)	10514	12450	1757	64
H(5A)	9375	10081	1684	51
H(8A)	11380	7545	1899	48
H(8B)	11826	8069	1192	48
H(12A)	9817	7007	425	47
H(12B)	10046	5739	264	47
H(13A)	10335	4402	1123	79
H(13B)	11172	5312	1716	79
H(13C)	11748	5802	1018	79
H(14A)	6394	4022	99	79
H(14B)	7749	4527	-315	79
H(15A)	8089	6800	-301	86
H(15B)	6714	5733	-657	86
H(16A)	5554	5547	345	113
H(16B)	6930	6905	545	113
H(16C)	6519	6194	616	113
H(16D)	6787	7400	148	113
H(17A)	5355	7450	179	60
H(17B)	5264	6711	-477	60
H(17C)	6650	8070	-283	60
H(17D)	4843	6376	459	238
H(17E)	4645	4951	205	238
H(17F)	4914	6146	-280	238

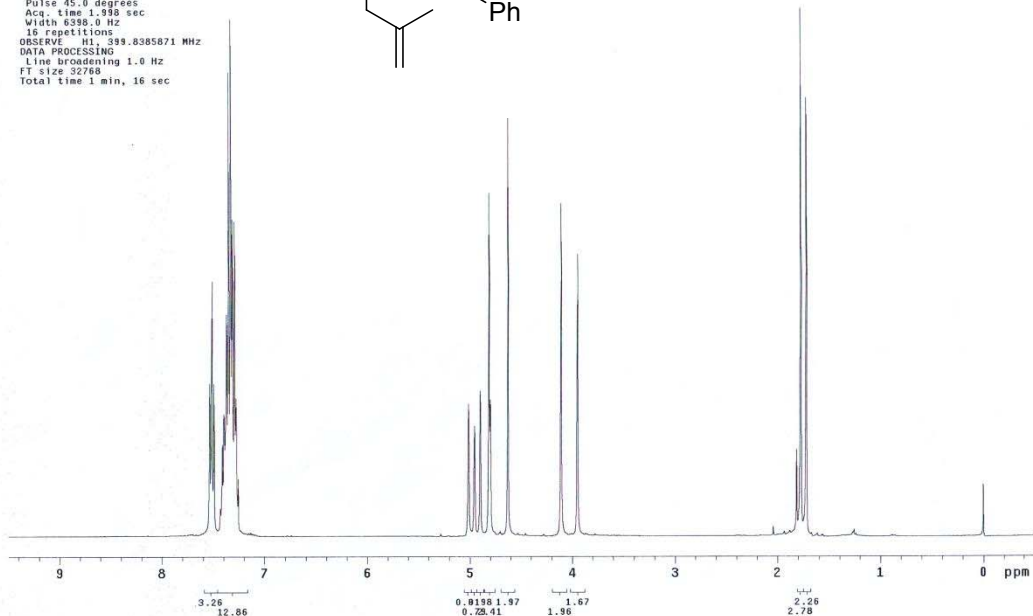
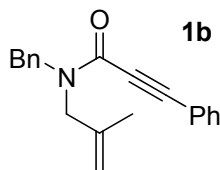
Appendix

Copy of ^1H and ^{13}C NMR Spectra of
Substrates and Products



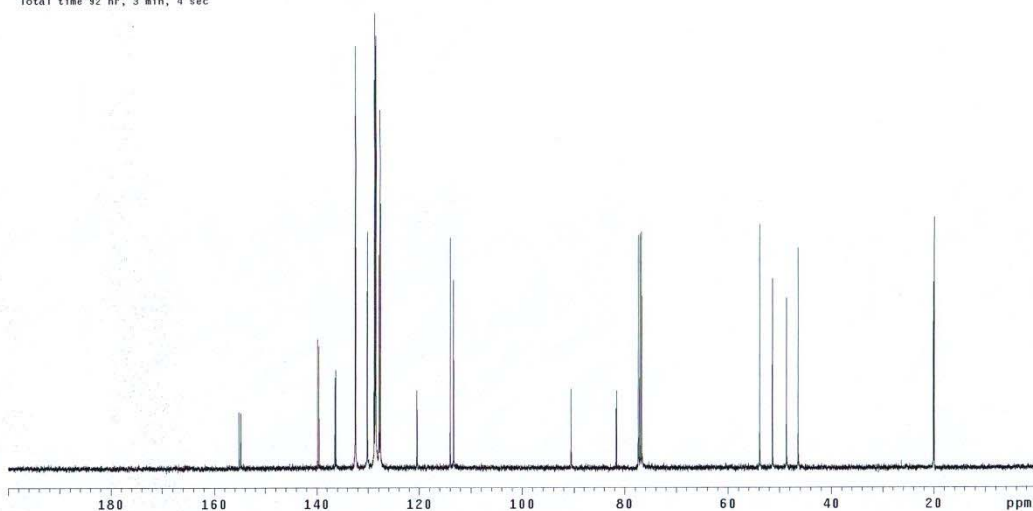
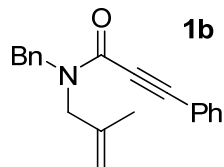
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 Solvent: cdc13
 Ambient temperature
 Operator: vnmr3
 File: PHS-VII-18-2-2
 Mercury-400BB "HY400"

Relax. delay 1.000 sec
 Pulse 45.0 degrees
 Acq. time 1.998 sec
 Width 6398.0 Hz
 16 repetitions
 OBSERVE H1, 399.8385871 MHz
 DATA PROCESSING
 Line broadening 1.0 Hz
 FT size 32768
 Total time 1 min, 16 sec

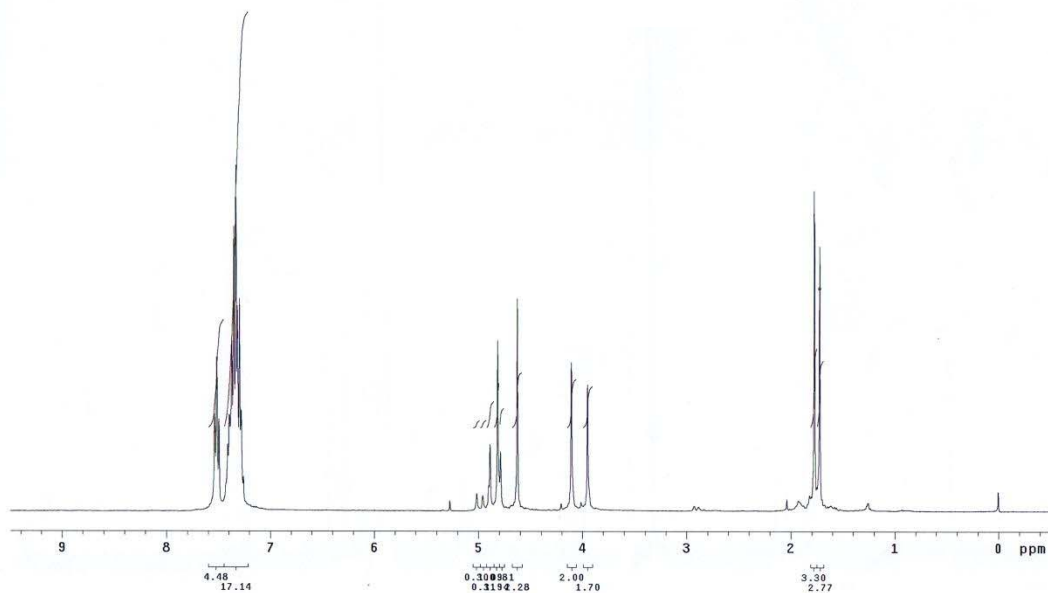
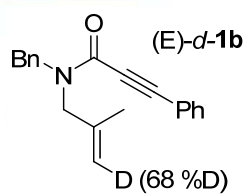


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 Solvent: cdc13
 Ambient temperature
 Operator: vnmr3
 File: PHS-VII-18-13C
 Mercury-400BB "HY400"

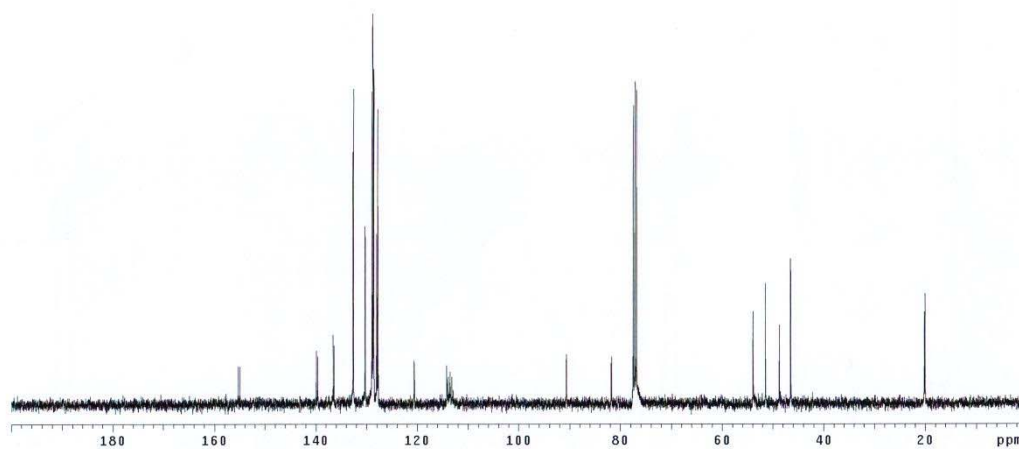
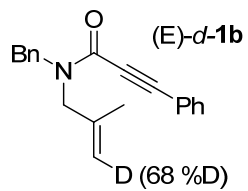
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 532 repetitions
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 DECOUPLE H1, 399.8405742 MHz
 Power 44 dB
 continuously on
 WALTZ-16 modulated
 DATA PROCESSING
 Line broadening 1.0 Hz
 FT size 65536
 Total time 32 hr, 3 min, 4 sec

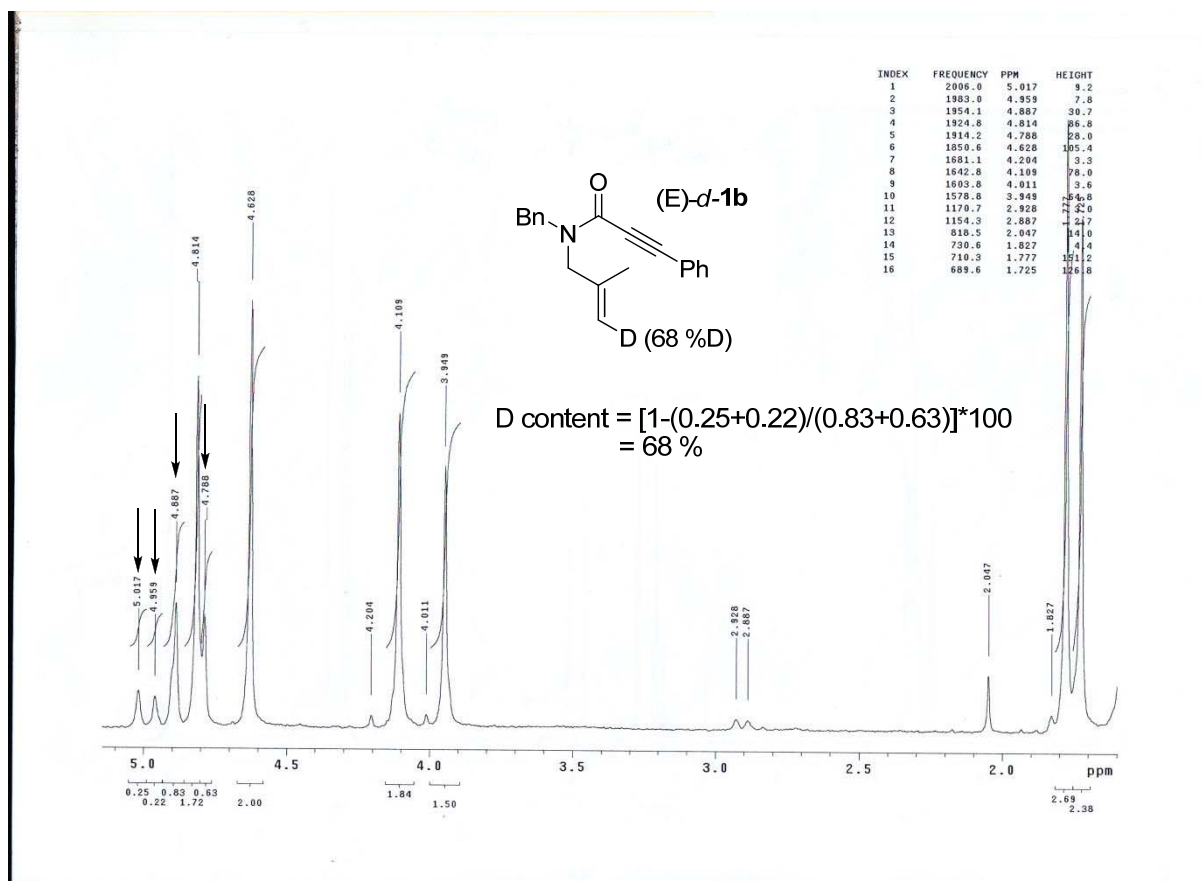


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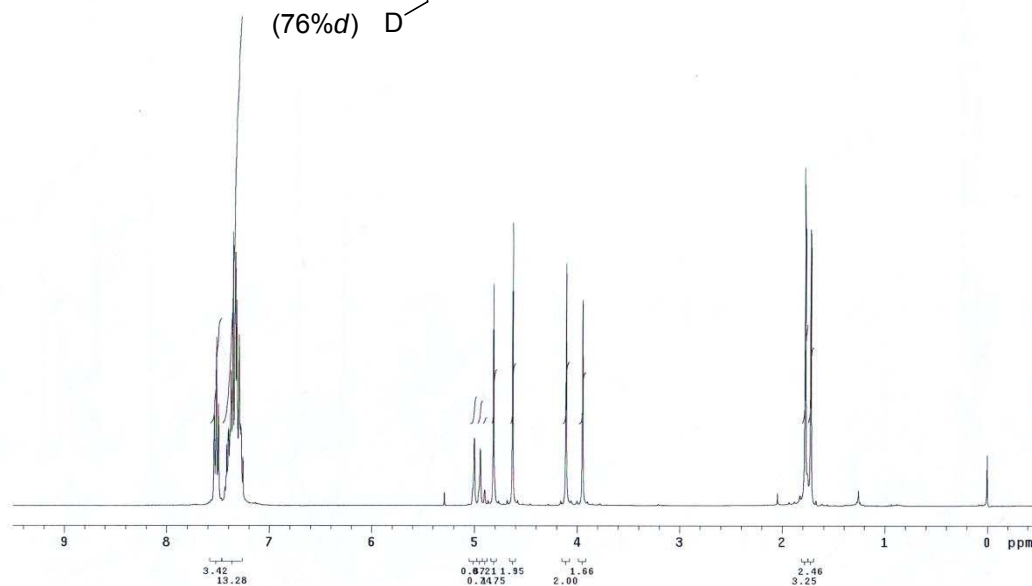
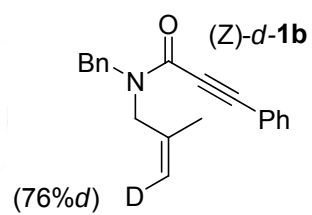


KJY-VI-32-13C
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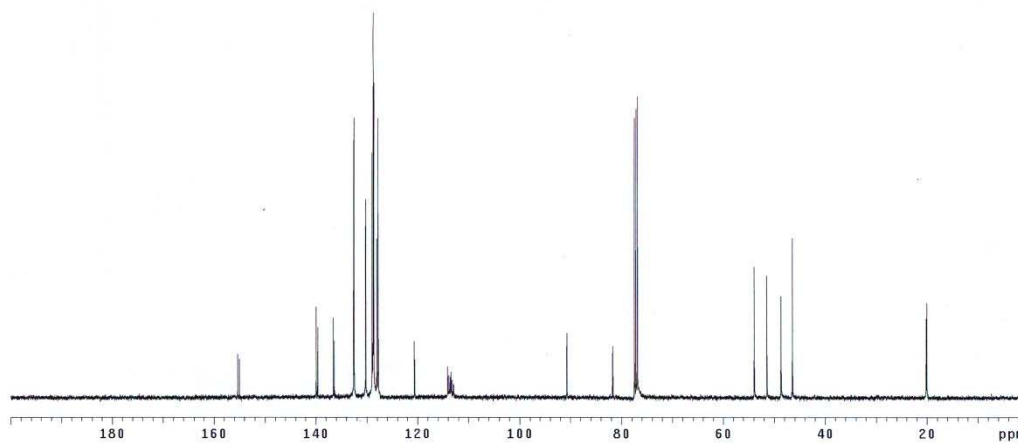
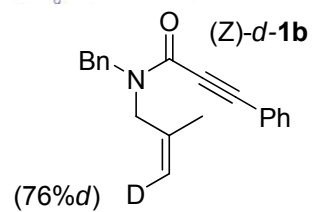


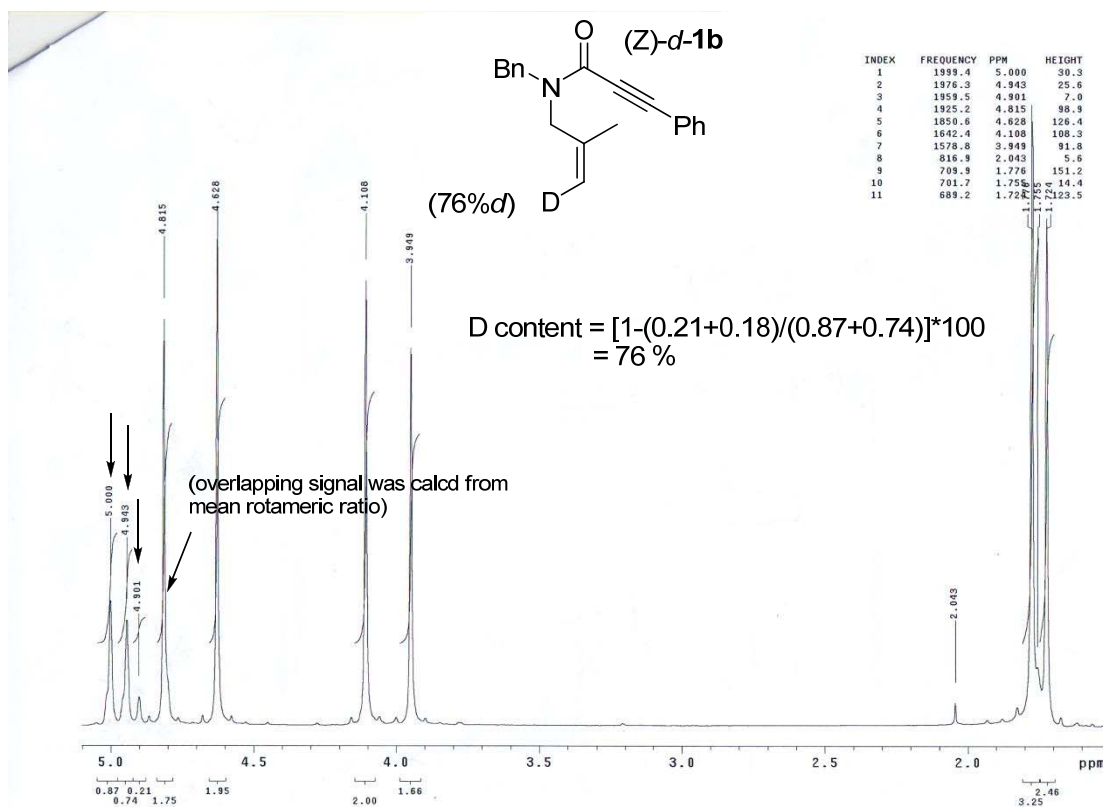


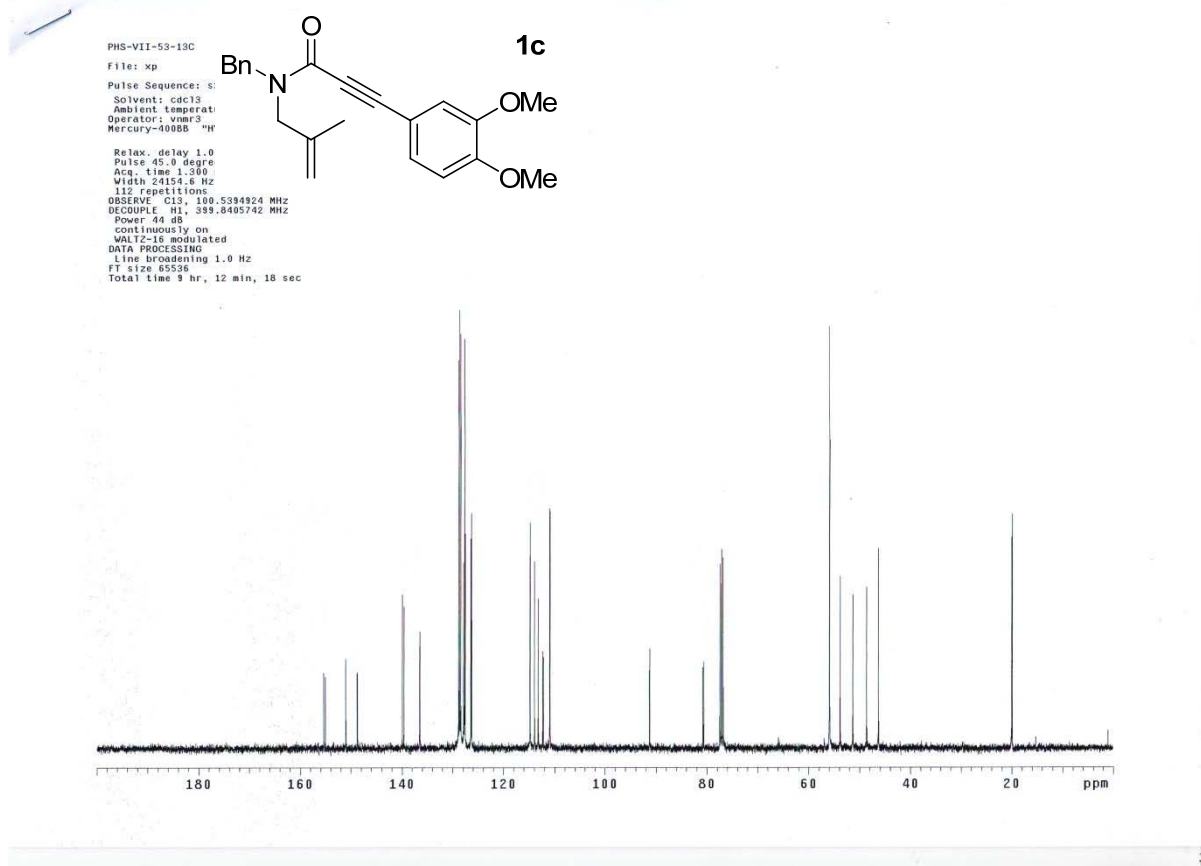
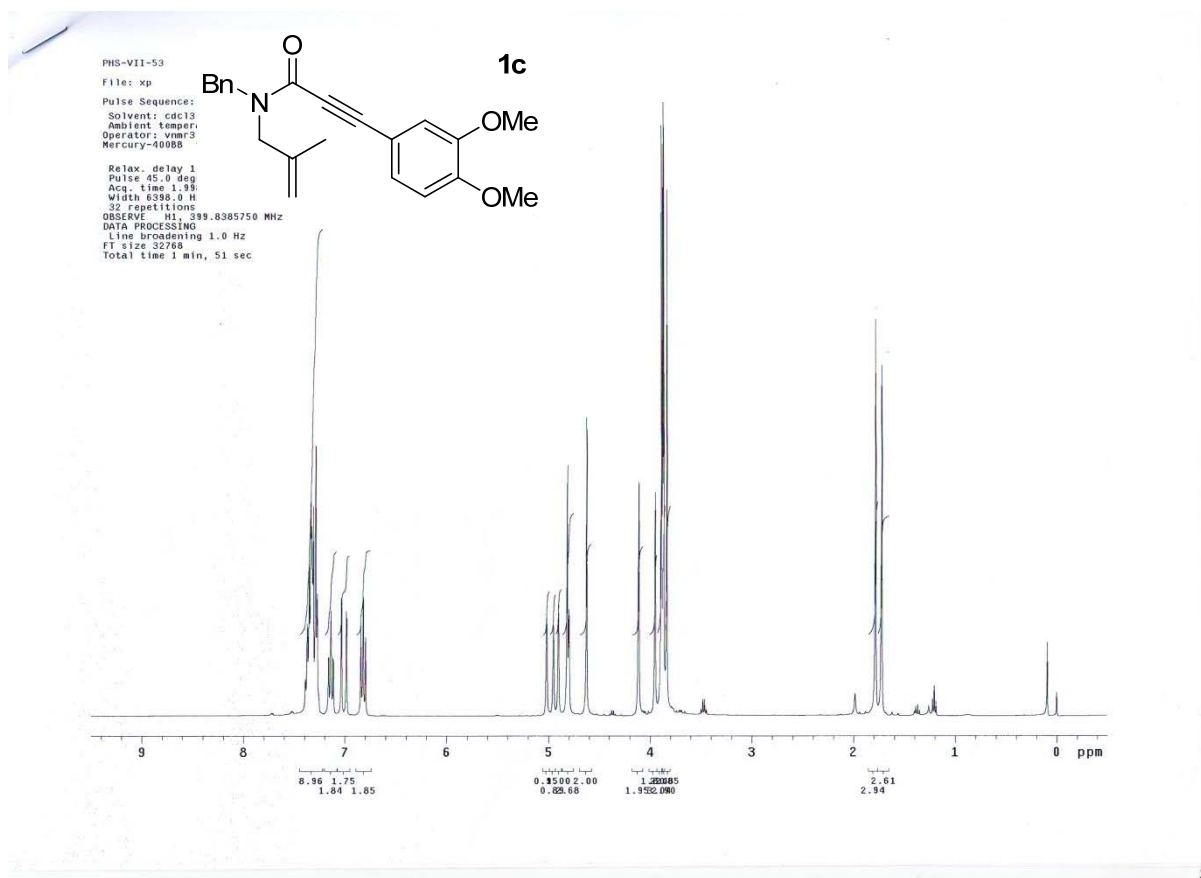
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Pulse Sequence: s2pu1

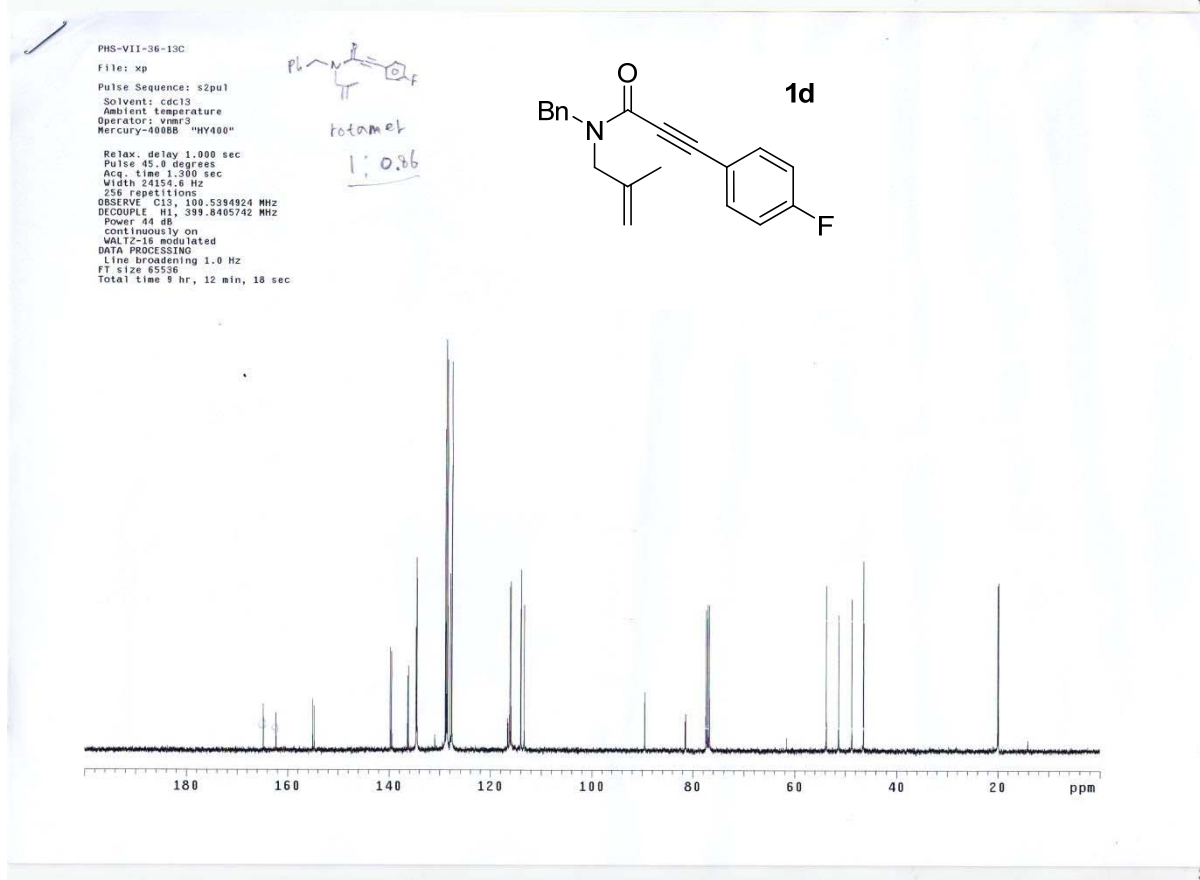
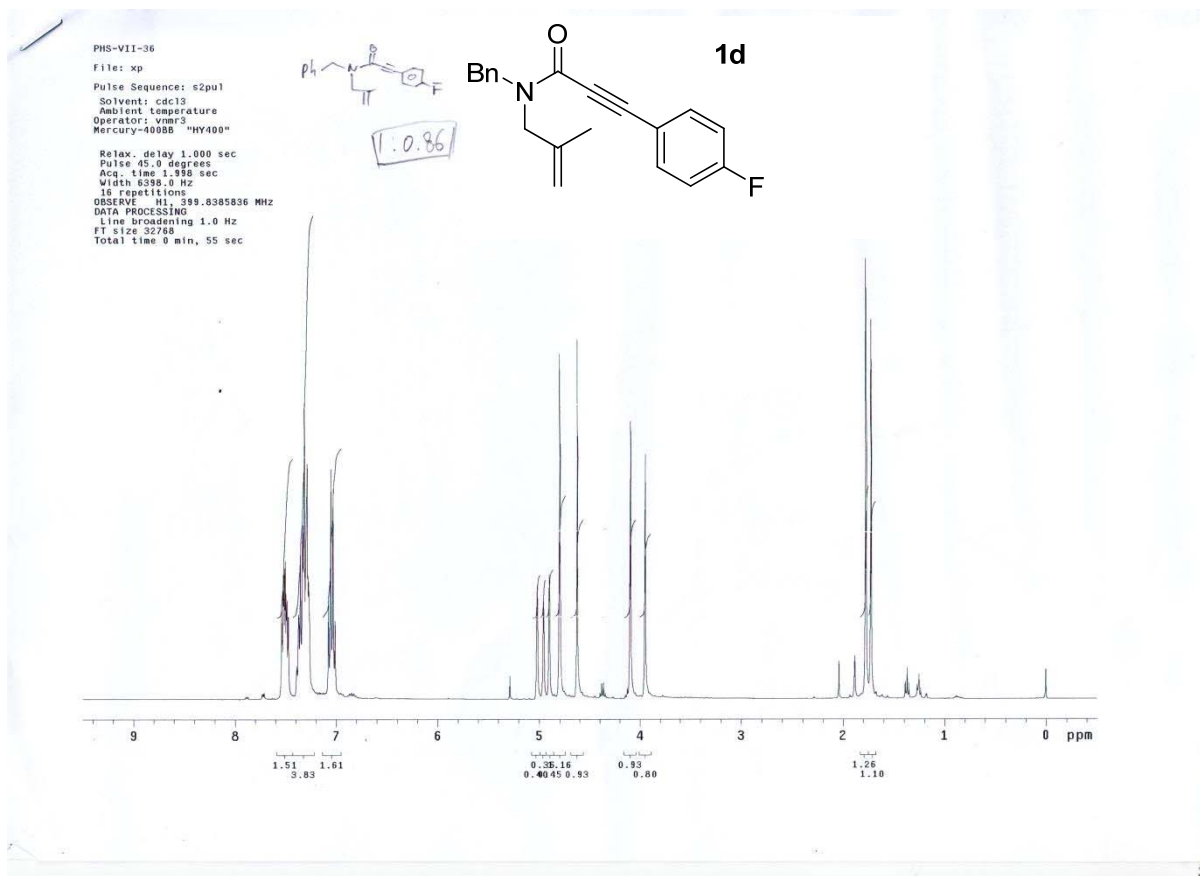


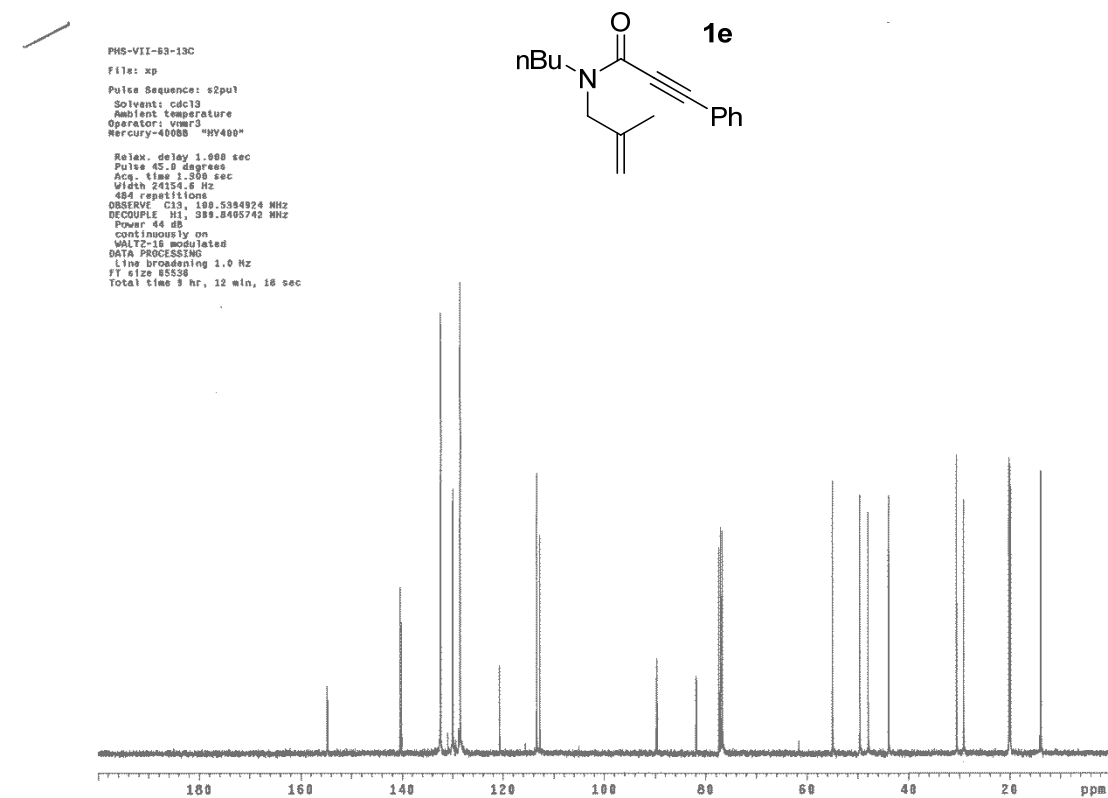
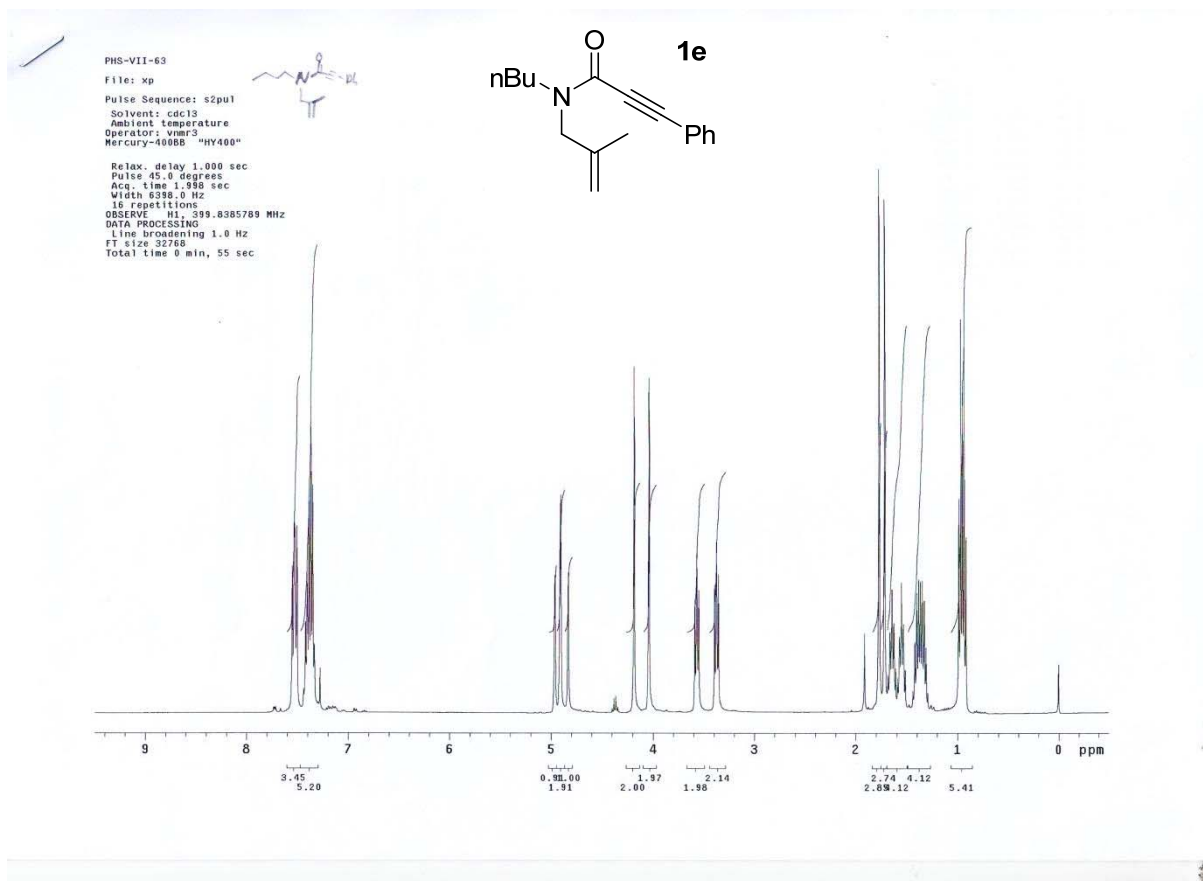
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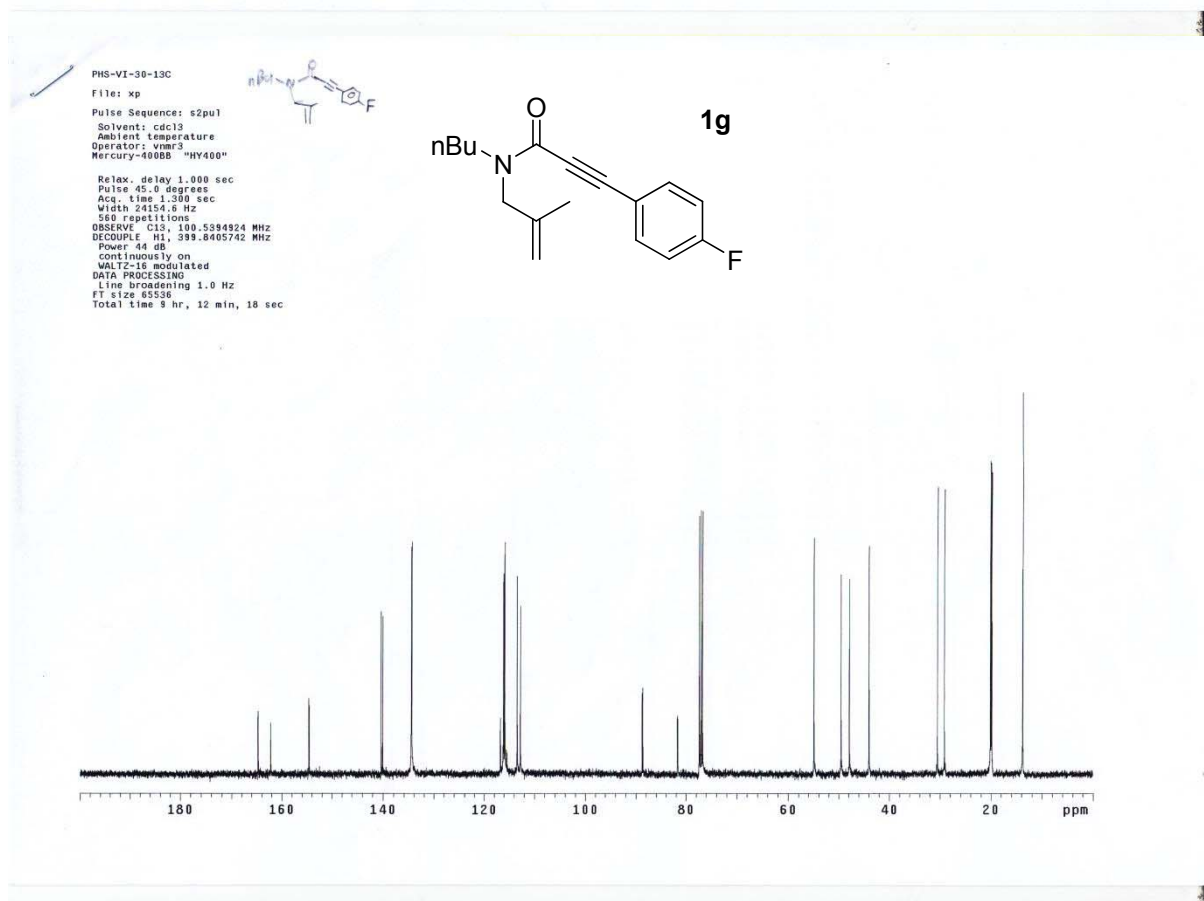
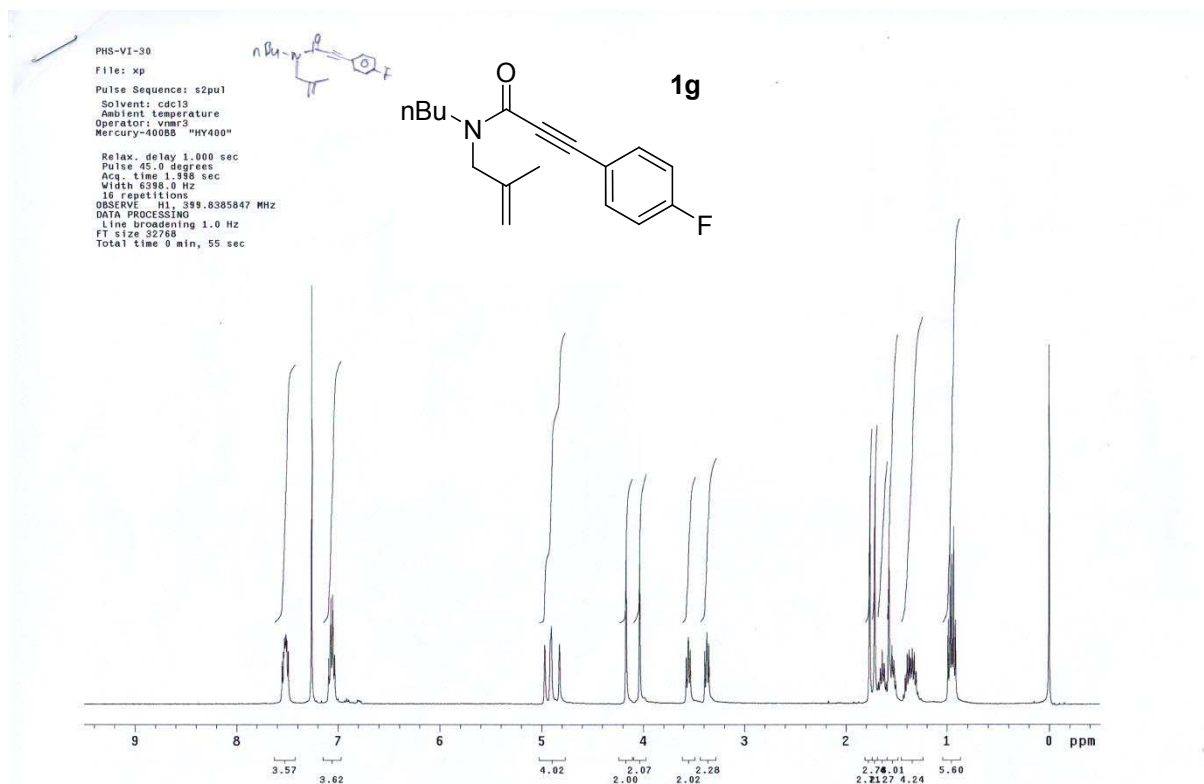


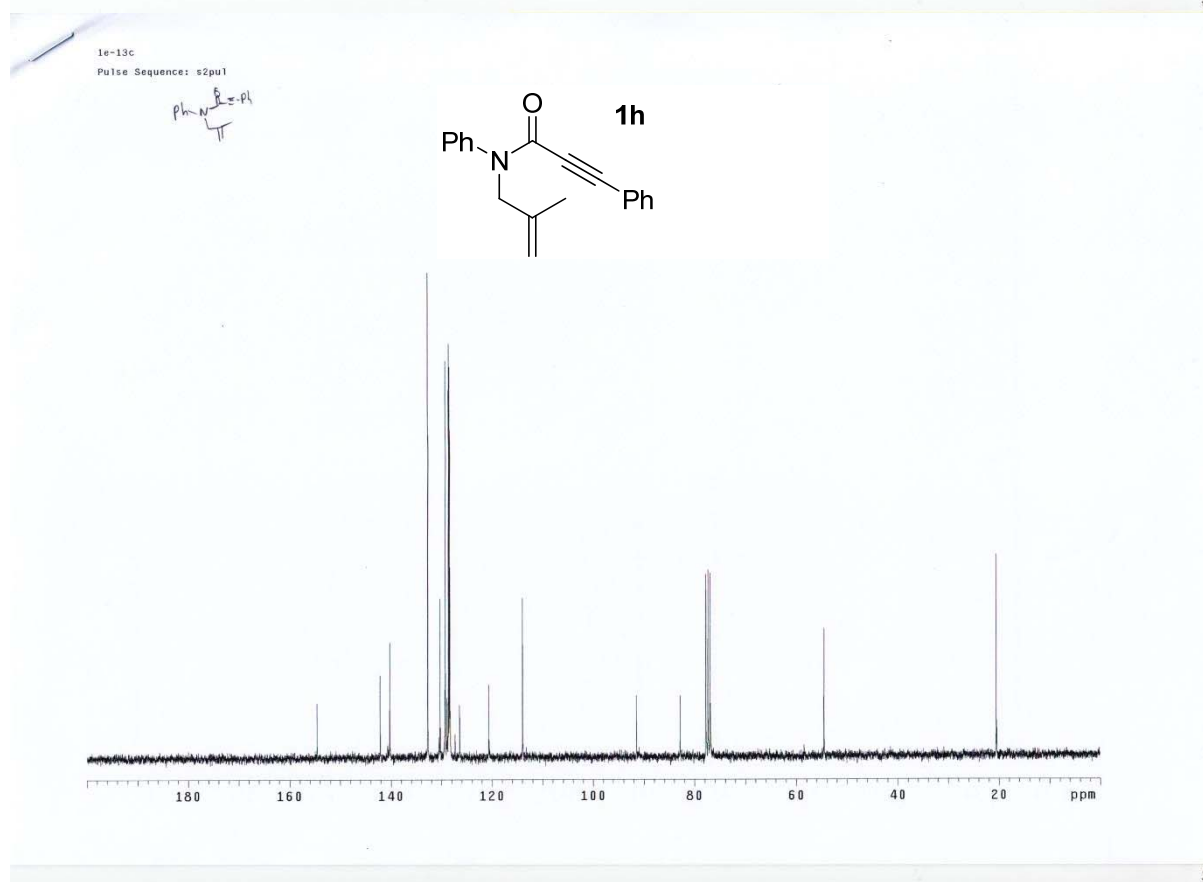
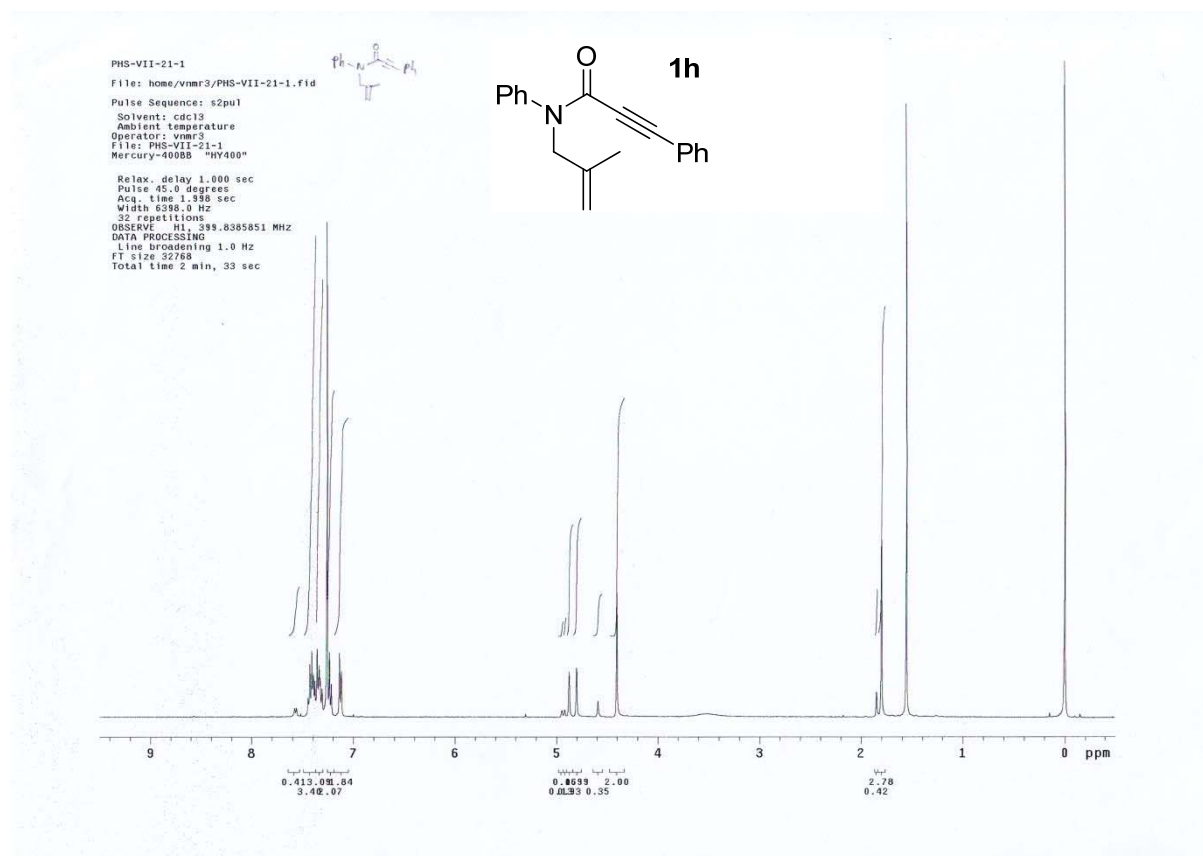


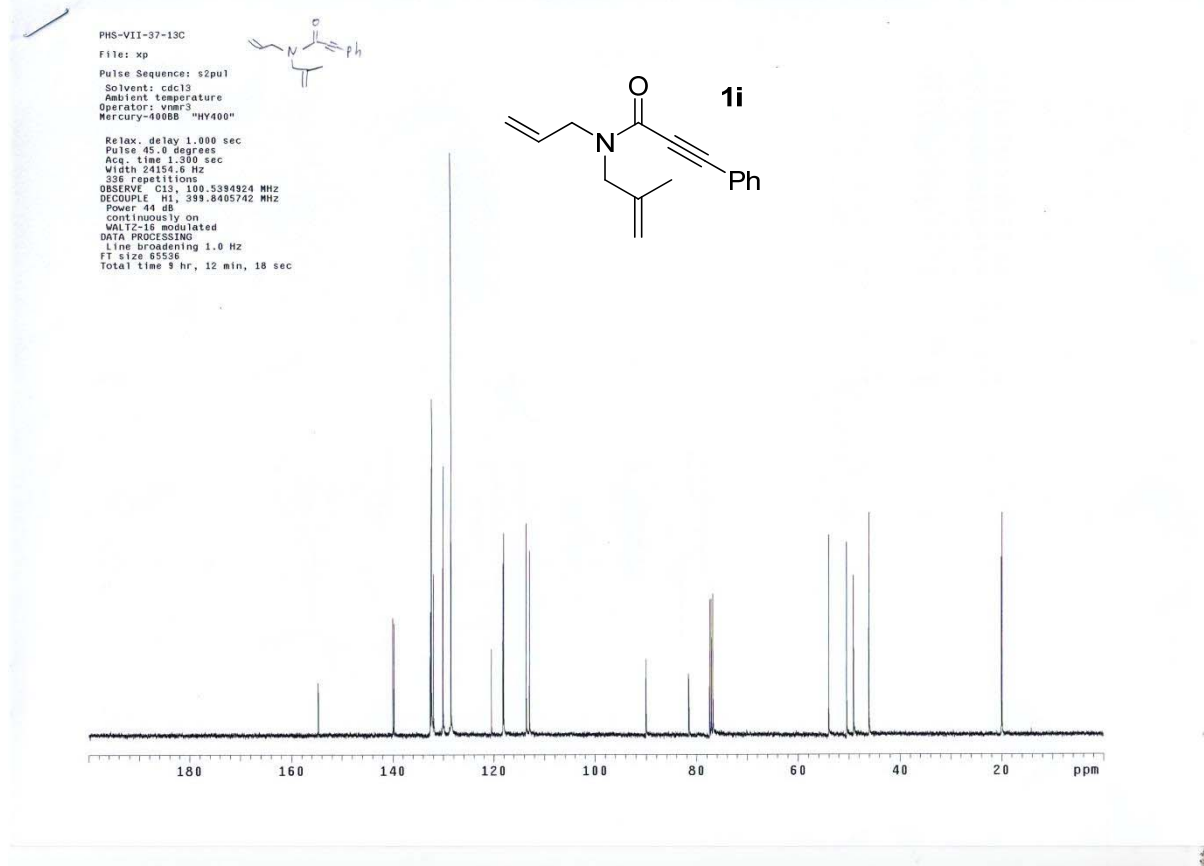
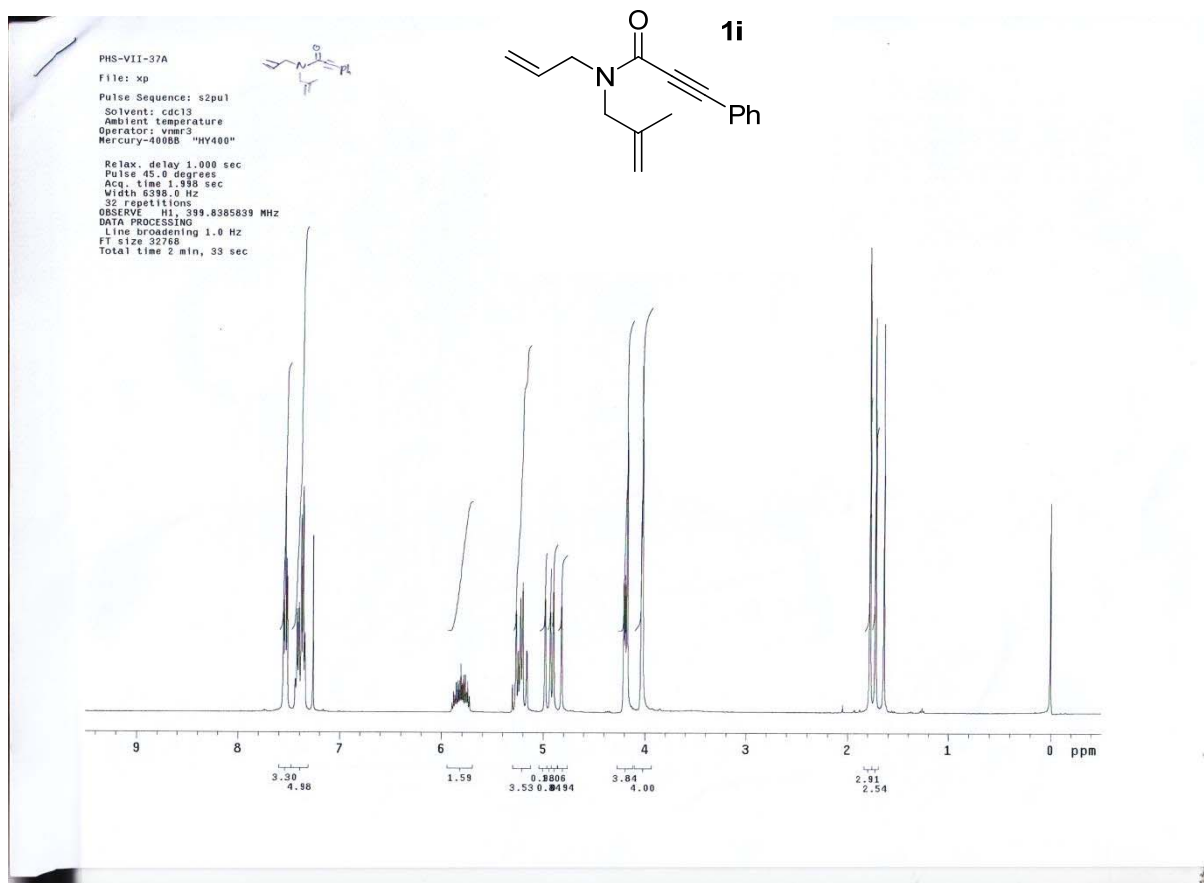




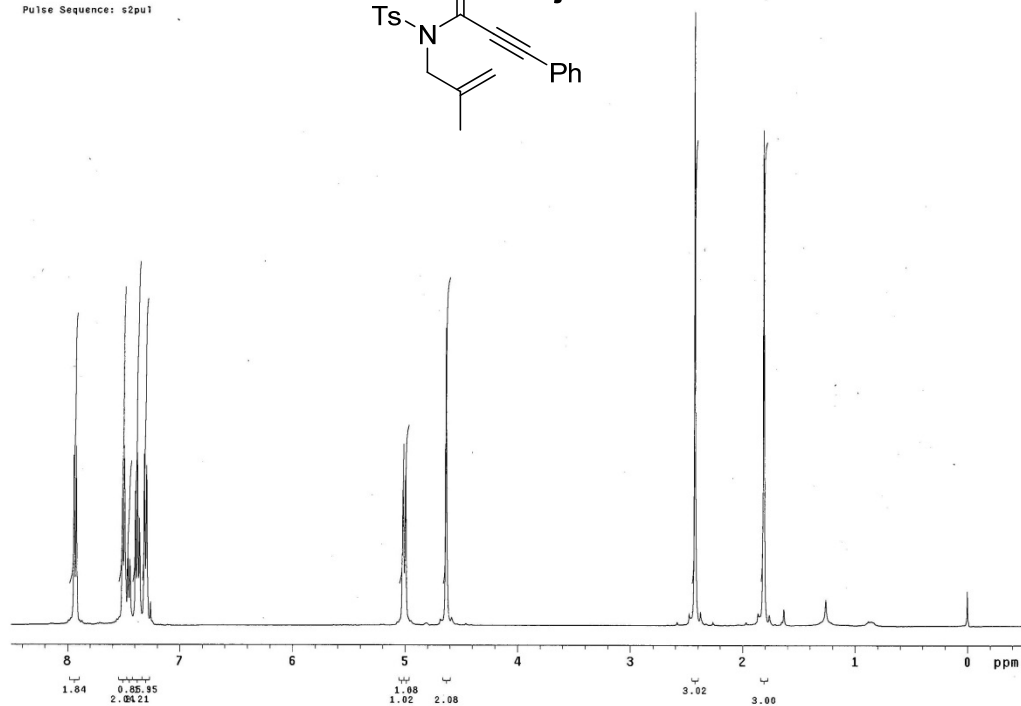
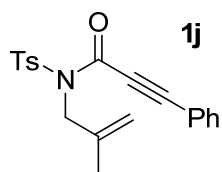




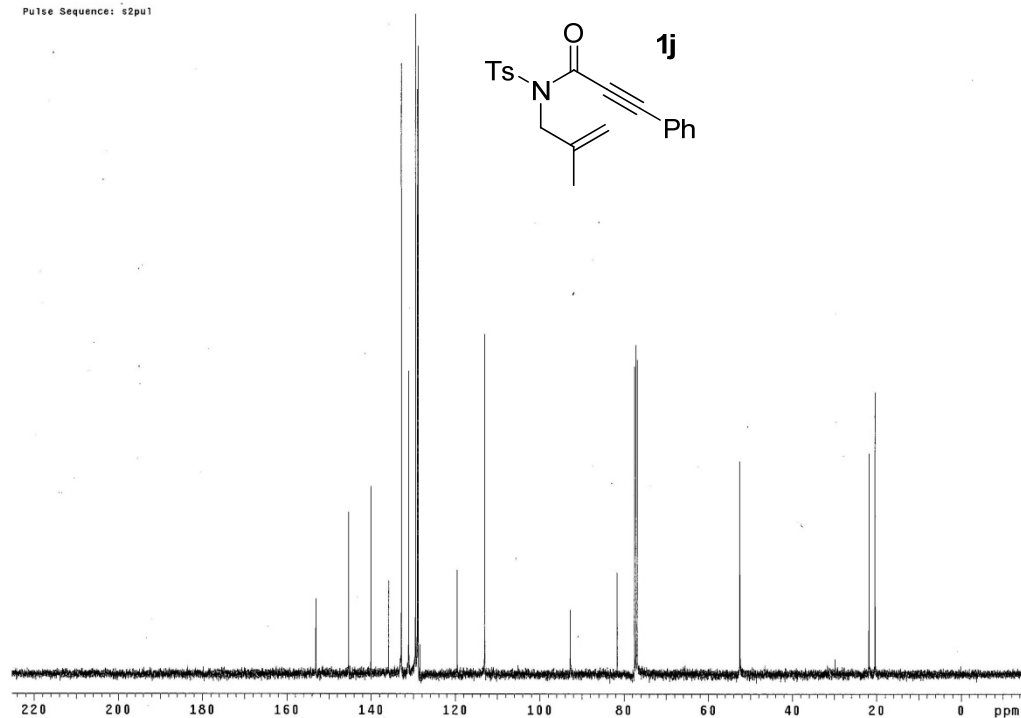
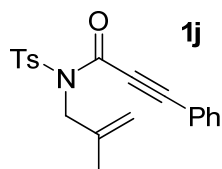




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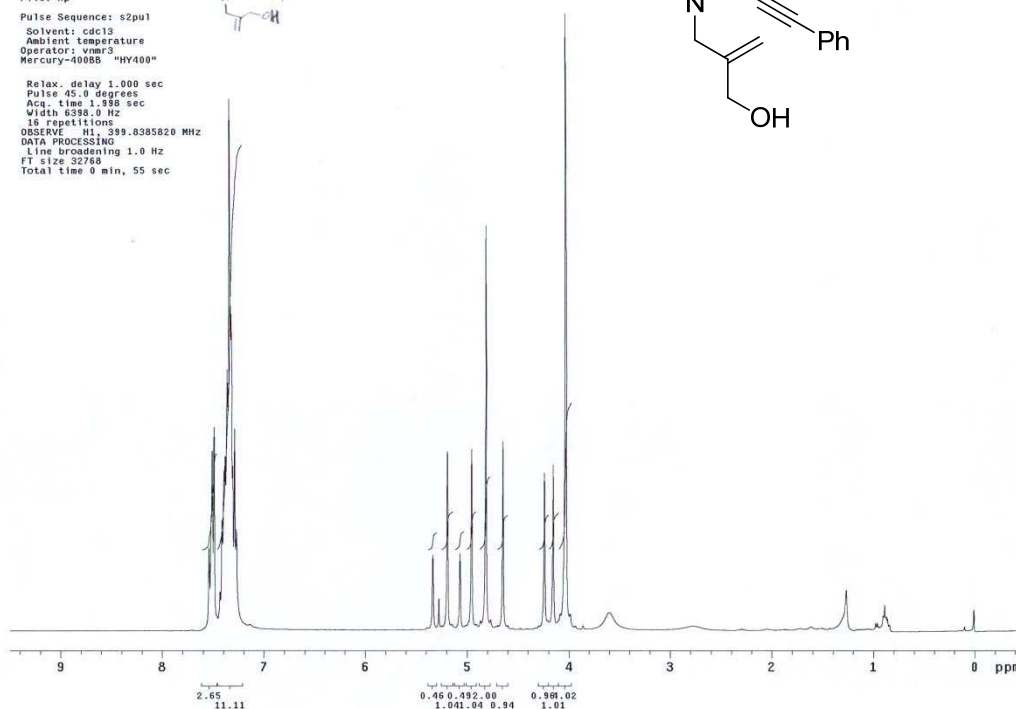
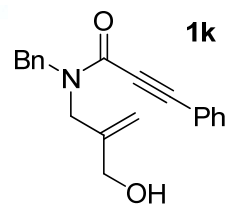


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Pulse Sequence: s2pu1



PHS-VIII-27
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Ambient temperature
Operator: vnmr3
Mercury-400BB "HY400"

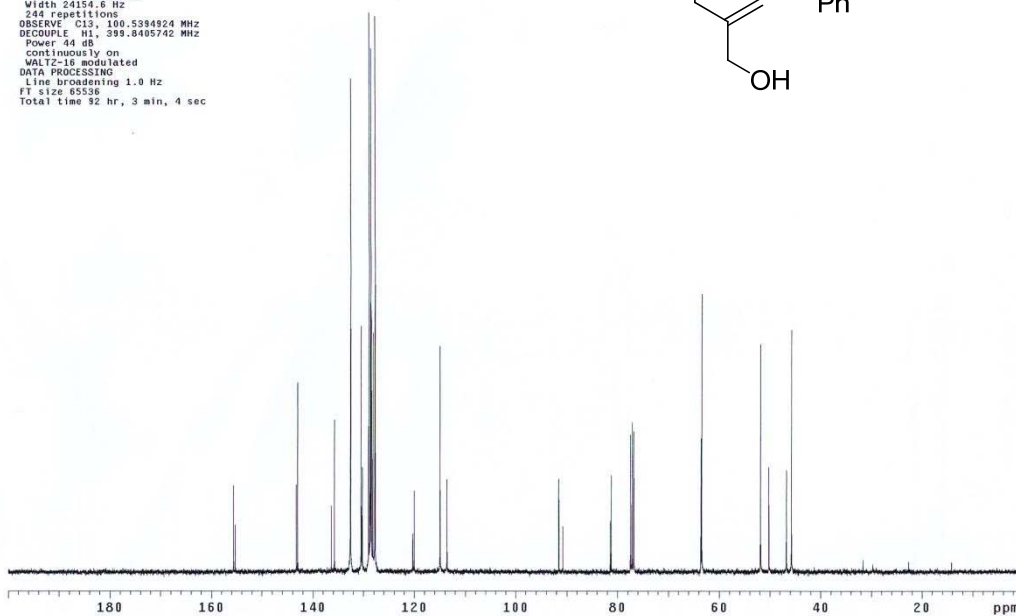
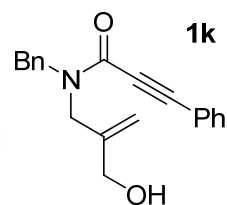
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DATA PROCESSING
Line broadening 1.0 Hz
FT size 32768
Total time 0 min, 55 sec



PHS-VIII-27-13C

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Pulse Sequence: s2pul
Solvent: cdcl3
Ambient temperature
Operator: vnmr3
Mercury-400BB "HY400"

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Acq. time 1.300 sec
Width 24154.0 Hz
244 repetitions
OBSERVE C13, 100.5394924 MHz
DECOUPLE H1, 399.8405742 MHz
Power 44 dB
continuously on
WALTZ-16 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 65536
Total time 52 hr, 3 min, 4 sec



PHS-VIII-16

File: xp

Pulse Sequence: s2pul

Solvent: cdc13

Ambient temperature

Operator: vnmr3

Mercury-400BB "HY400"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.958 sec

Width 6398.0 Hz

32 repetitions

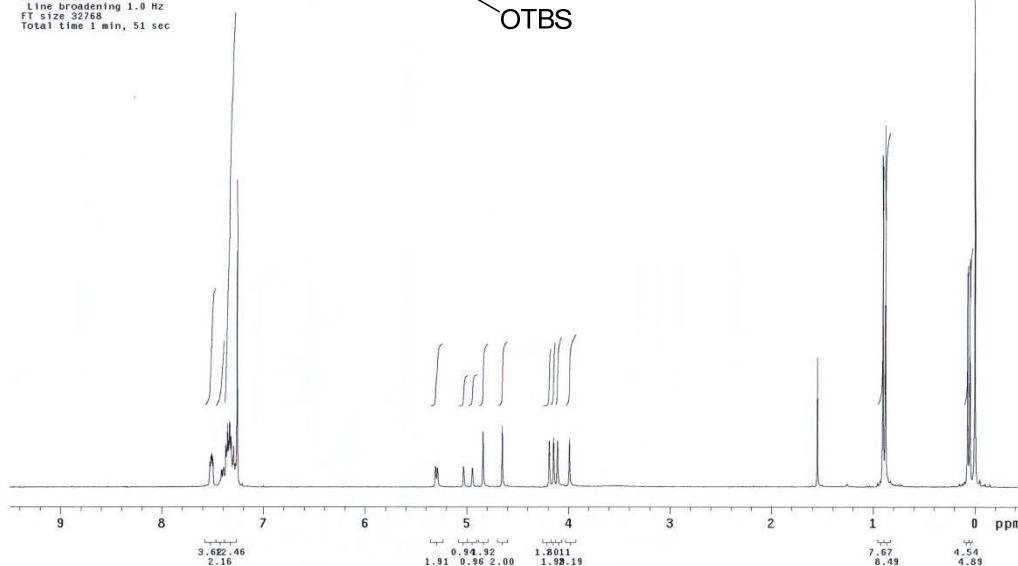
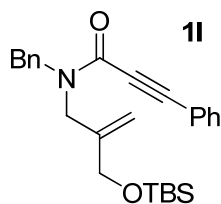
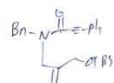
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DATA PROCESSING

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FT size 32768

Total time 1 min, 51 sec



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File: xp

Pulse Sequence: s2pul

Solvent: cdc13

Ambient temperature

Operator: vnmr3

Mercury-400BB "HY400"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 24154.6 Hz

604 repetitions

OBSERVE C13, 100.5394924 MHz

DECOUPLE H1, 399.8495742 MHz

Power 44 dB

continuously on

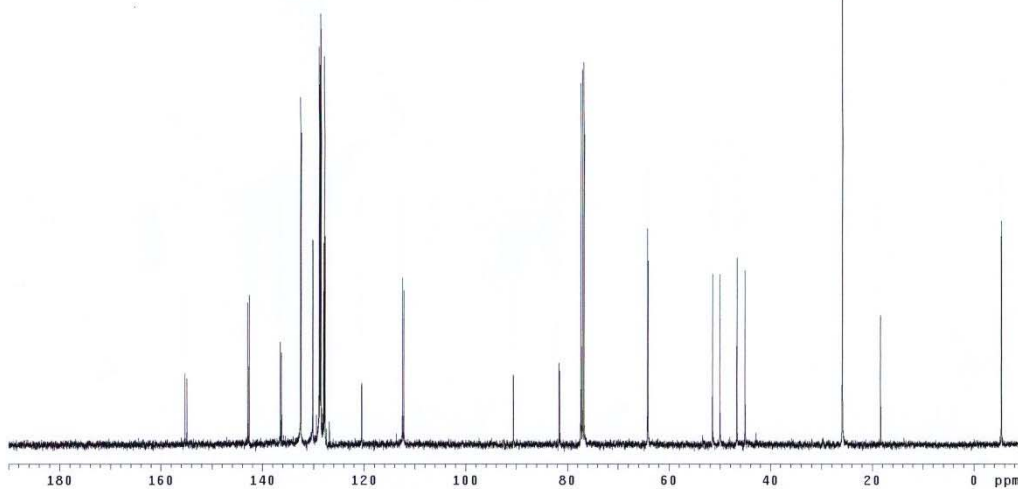
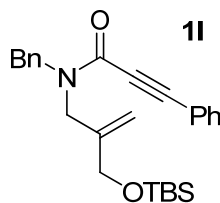
WALTZ-16 modulated

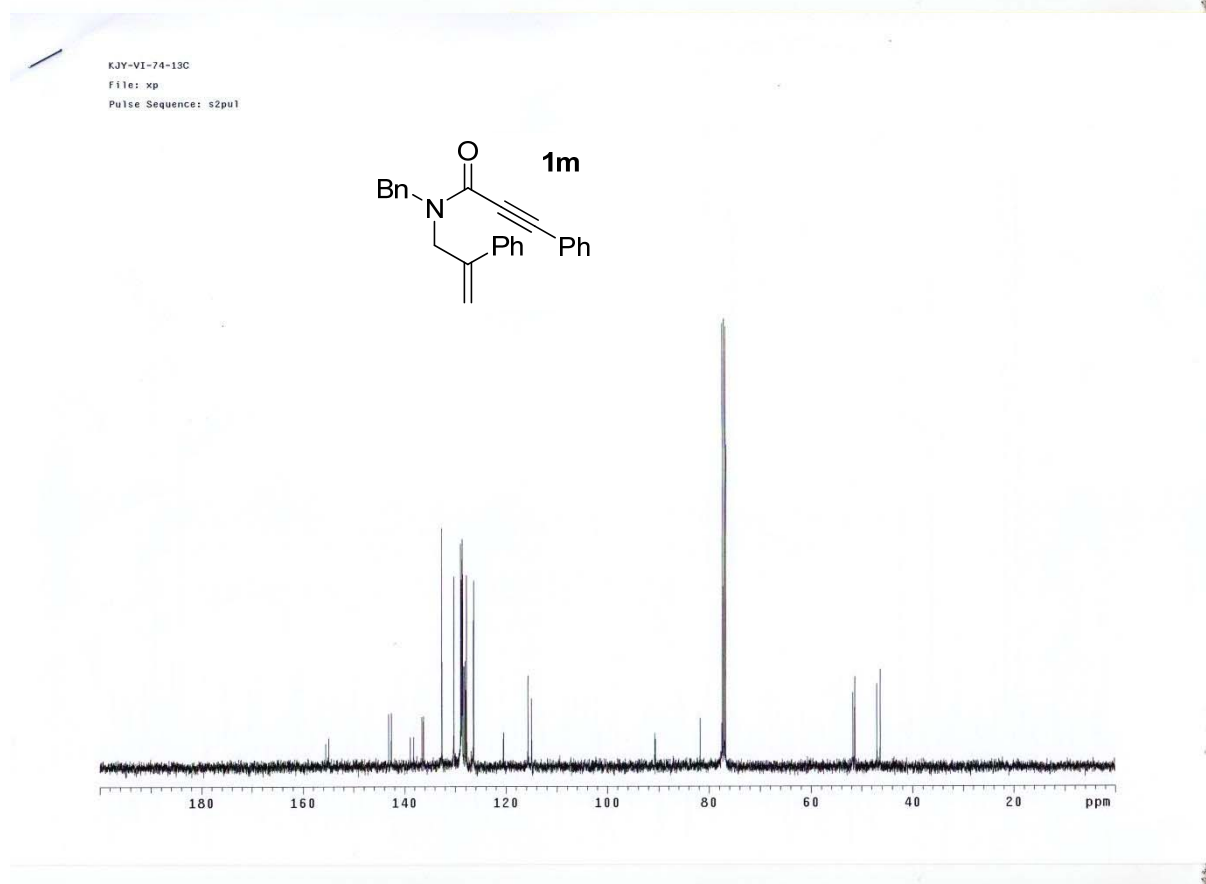
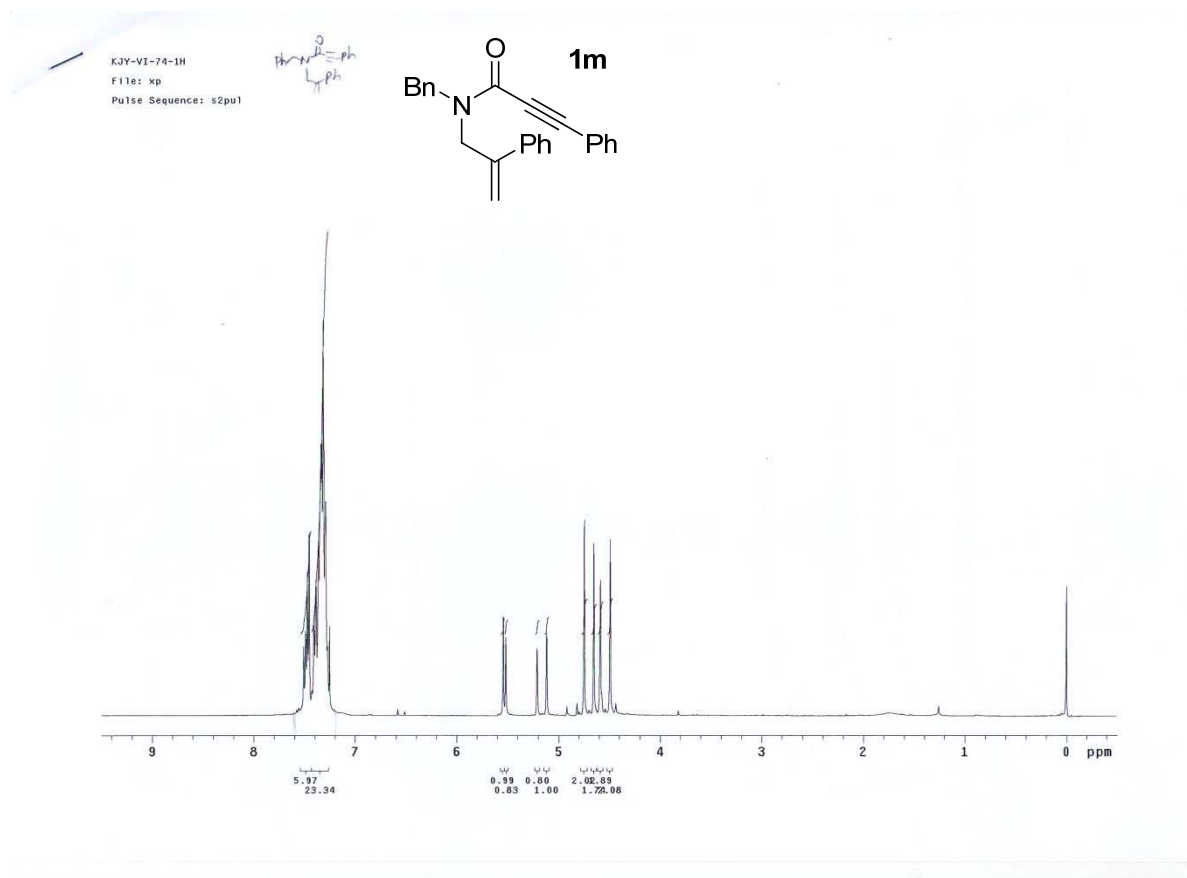
DATA PROCESSING

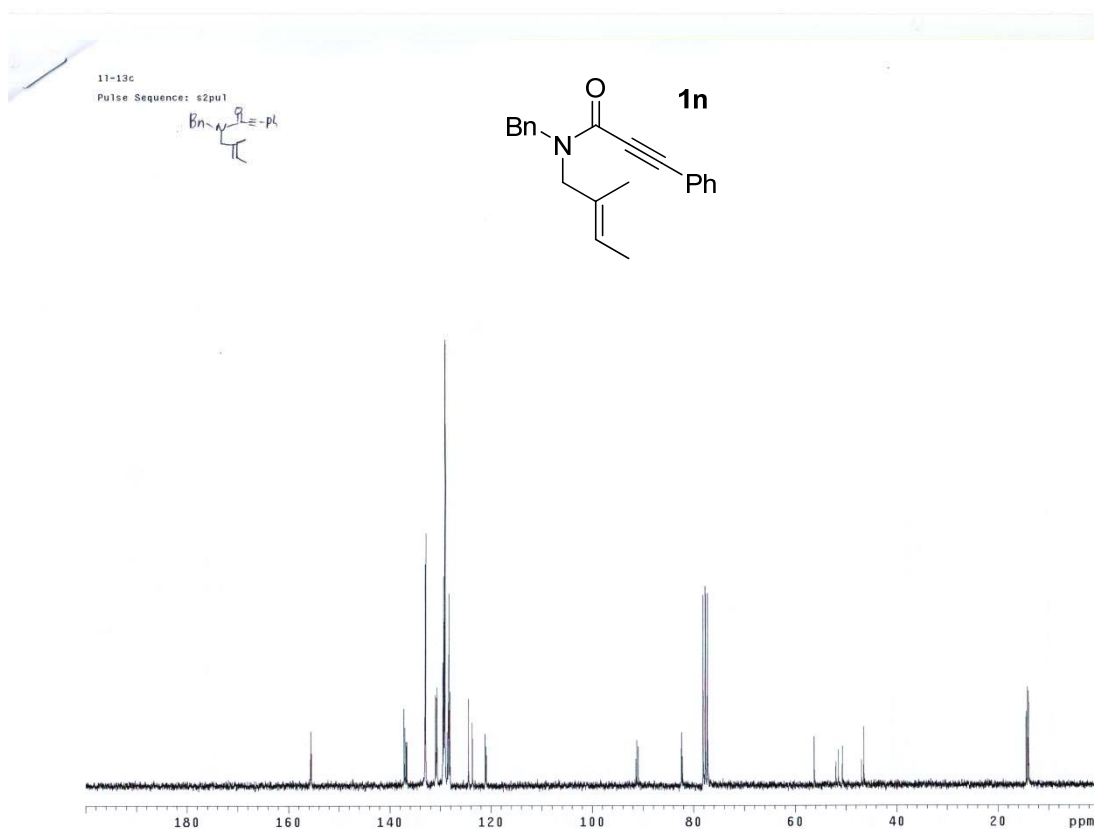
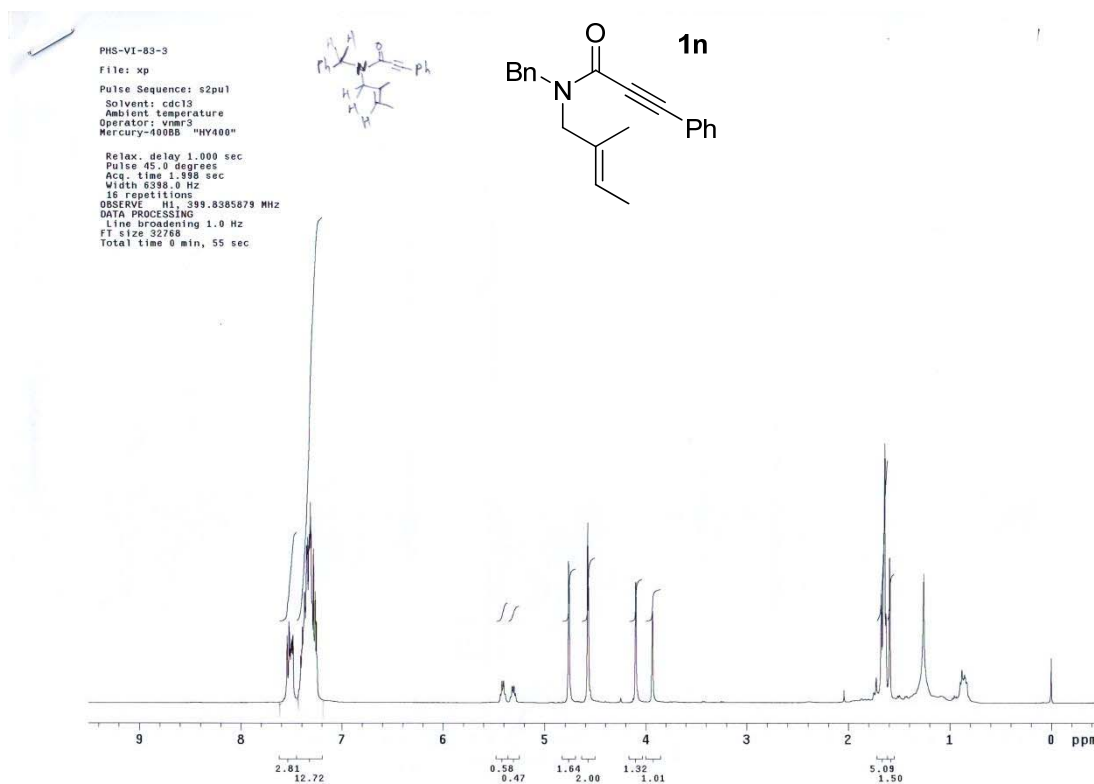
Line broadening 1.0 Hz

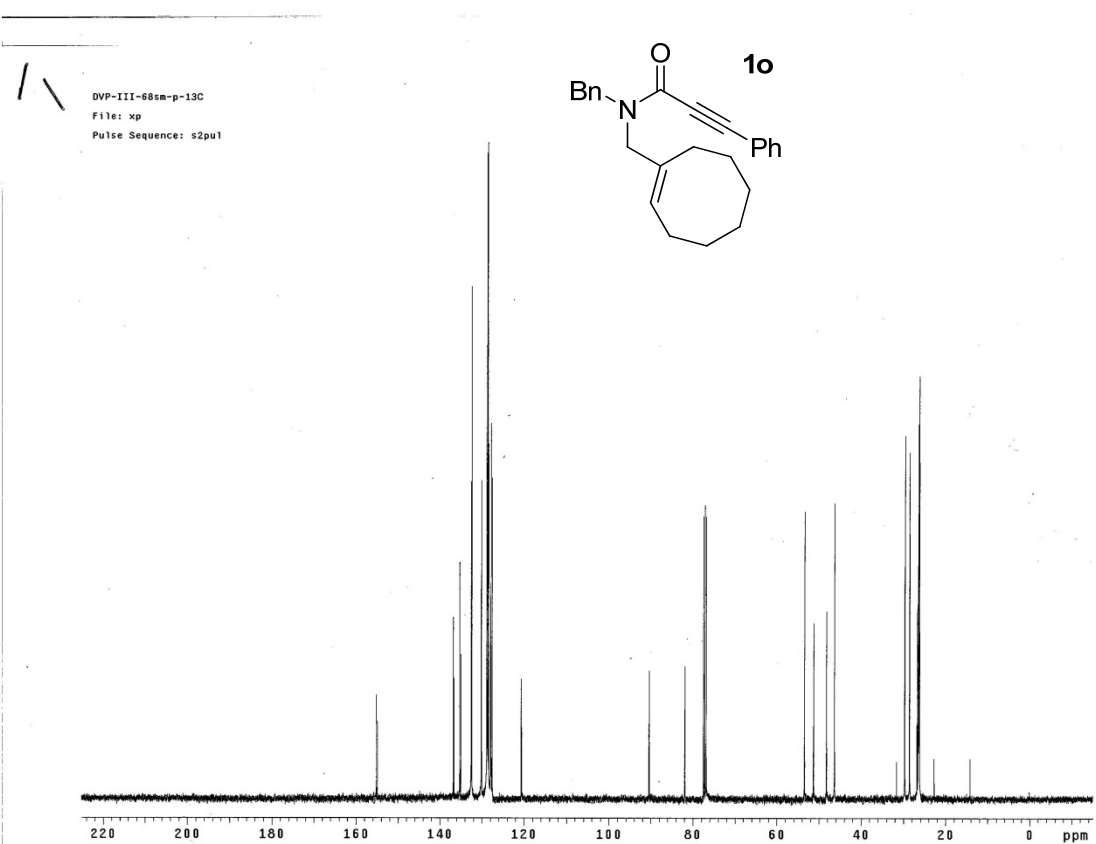
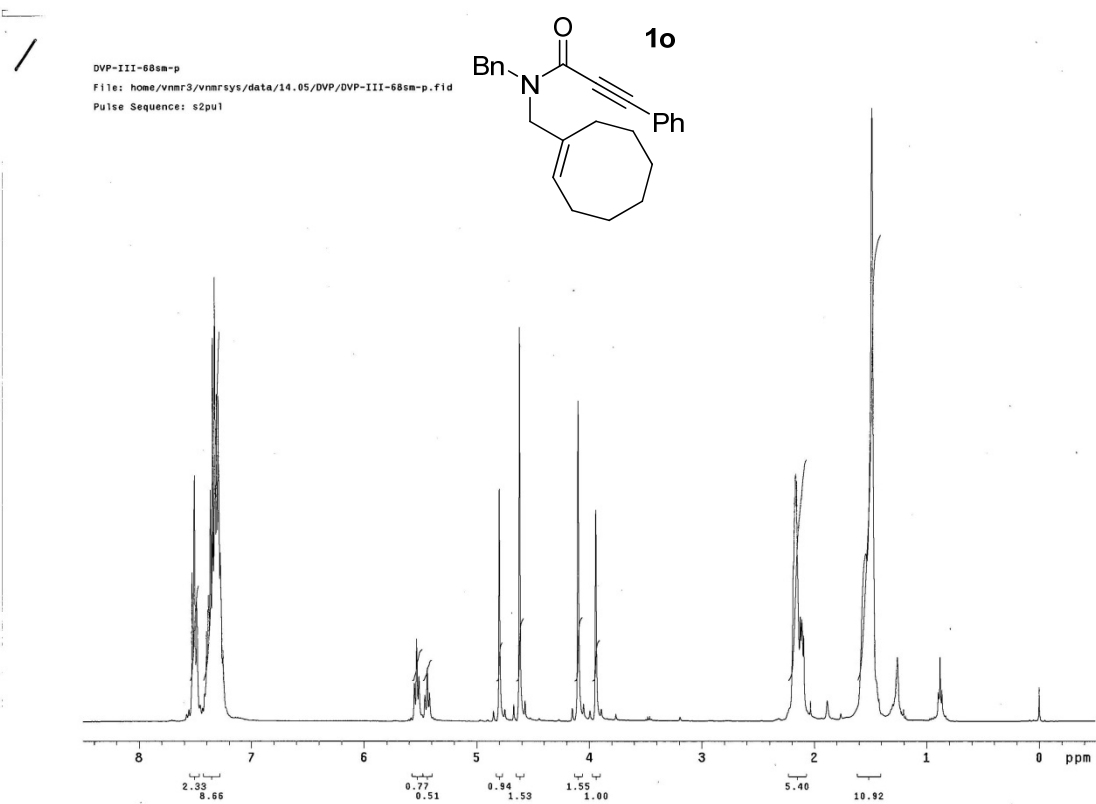
FT size 65536

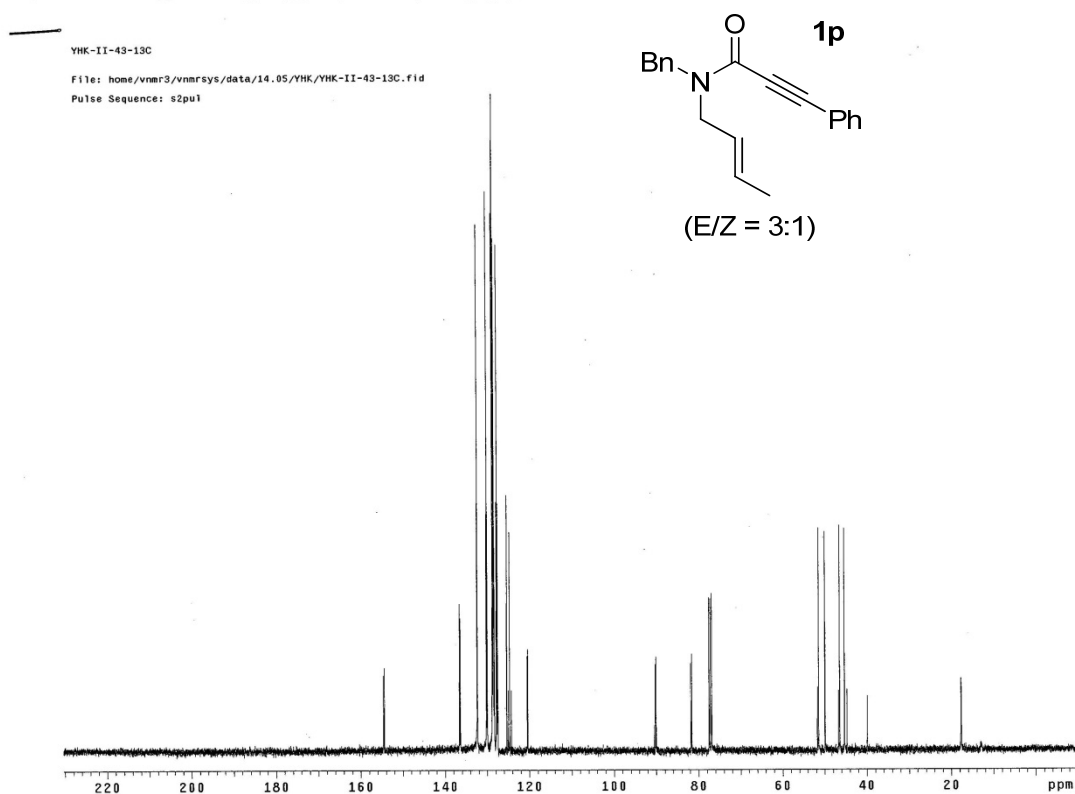
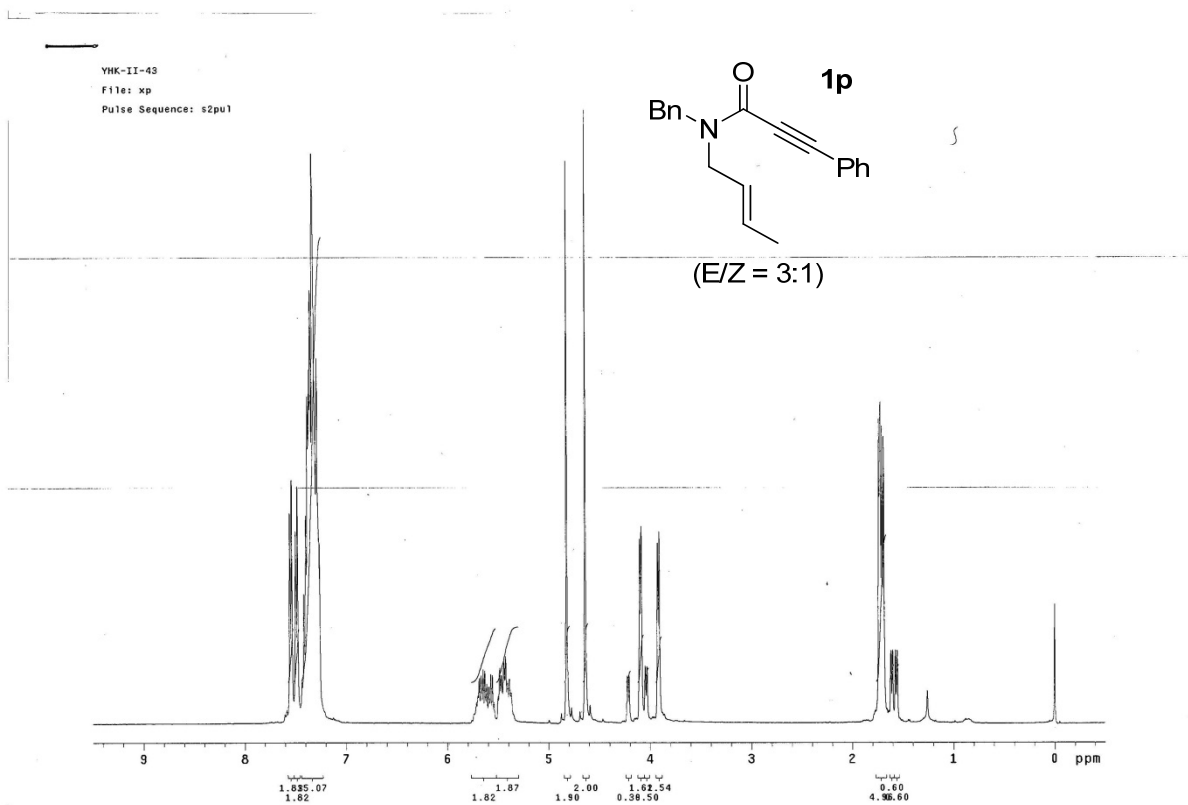
Total time 92 hr, 3 min, 4 sec



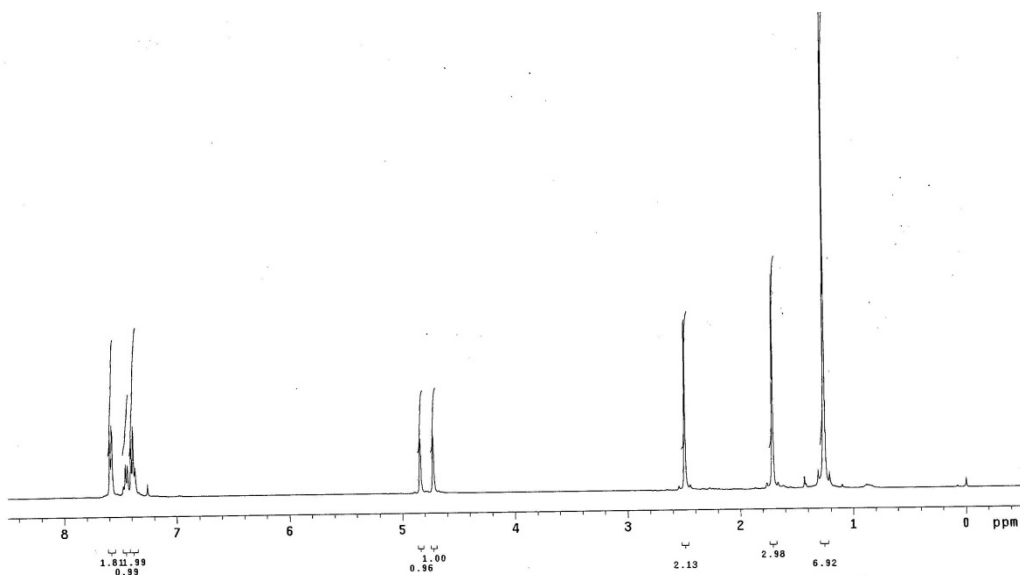
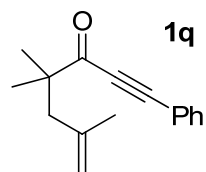




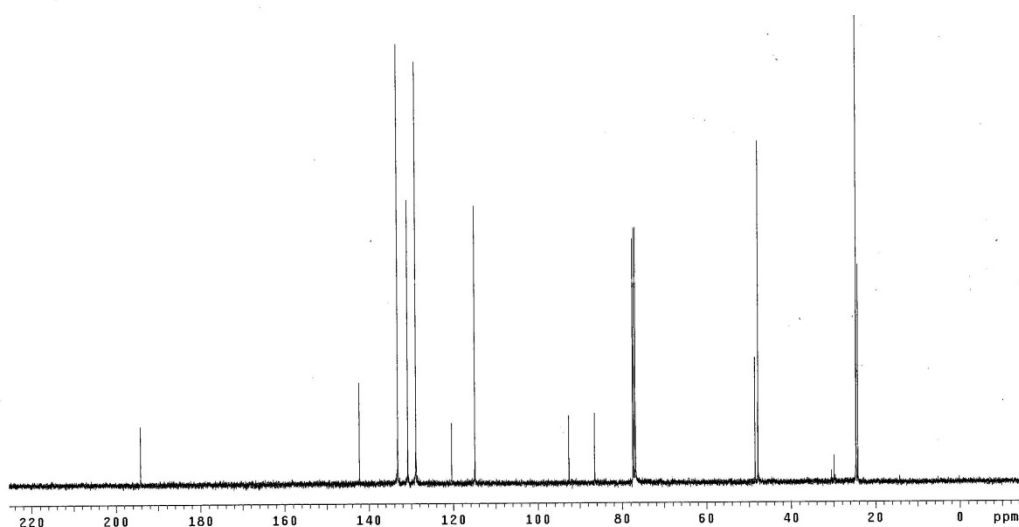
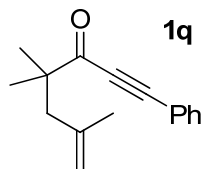




DVP-II-90fr1
 File: home/vnmr3/vnmr3sys/data/DVP/DVP-II-90fr1.fid
 Pulse Sequence: s2pul

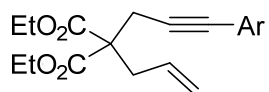


DVP-II-90fr1-13C
 File: xp
 Pulse Sequence: s2pul

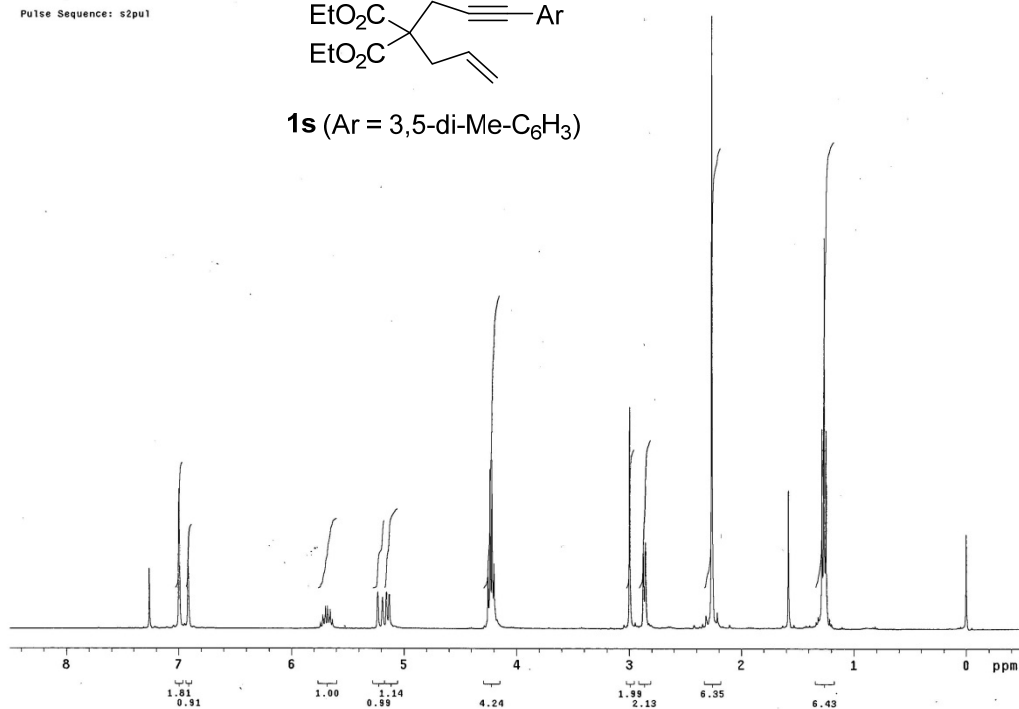


/

DVP-IV-41
File: xp
Pulse Sequence: s2pu1

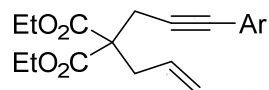


1s (Ar = 3,5-di-Me-C₆H₃)

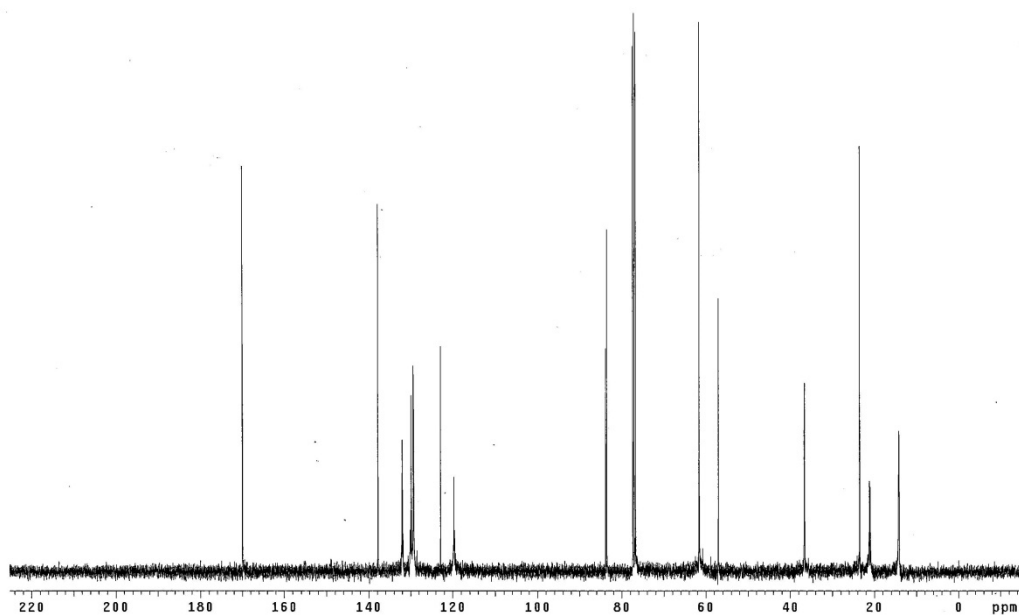


/

DVP-IV-41-13C
File: home/vnmr3/vnmr3sys/data/14.07/DVP/DVP-IV-41-13C.fid
Pulse Sequence: s2pu1

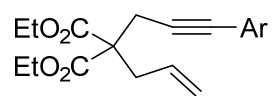


1s (Ar = 3,5-di-Me-C₆H₃)

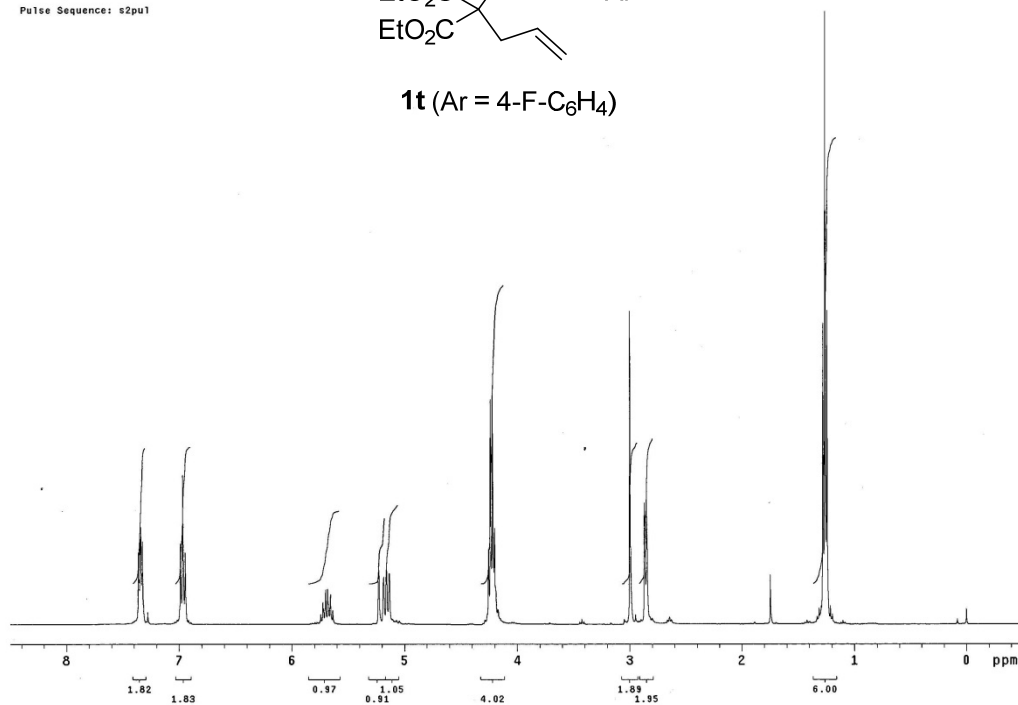


/

DVP-IV-44
File: xp
Pulse Sequence: s2pu1

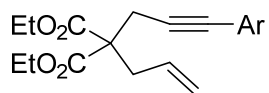


1t (Ar = 4-F-C₆H₄)

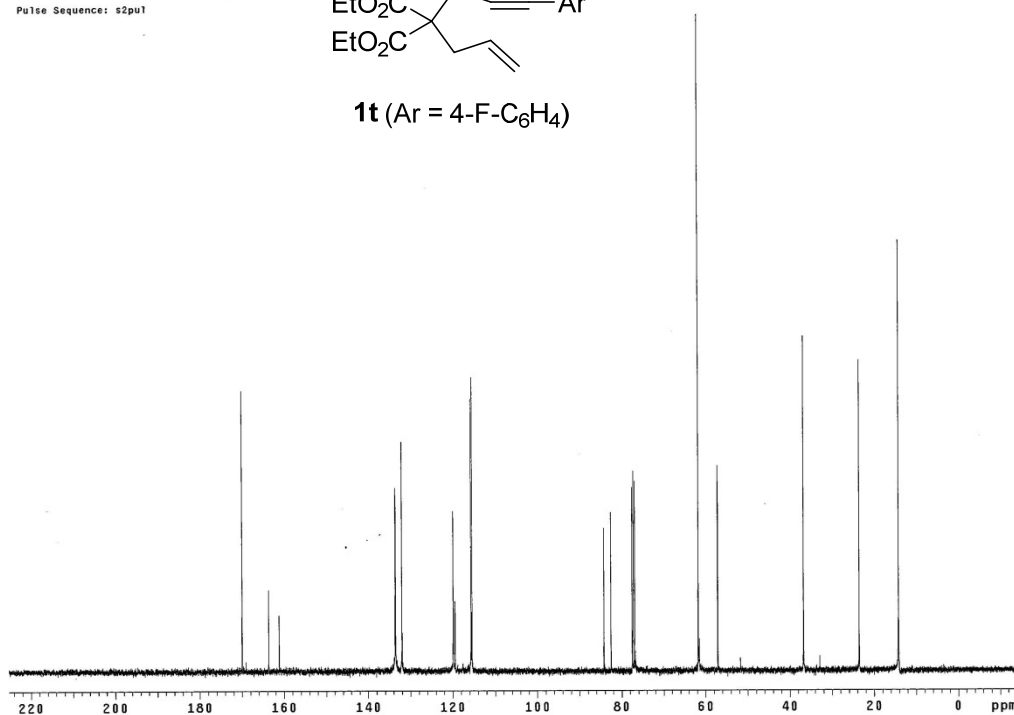


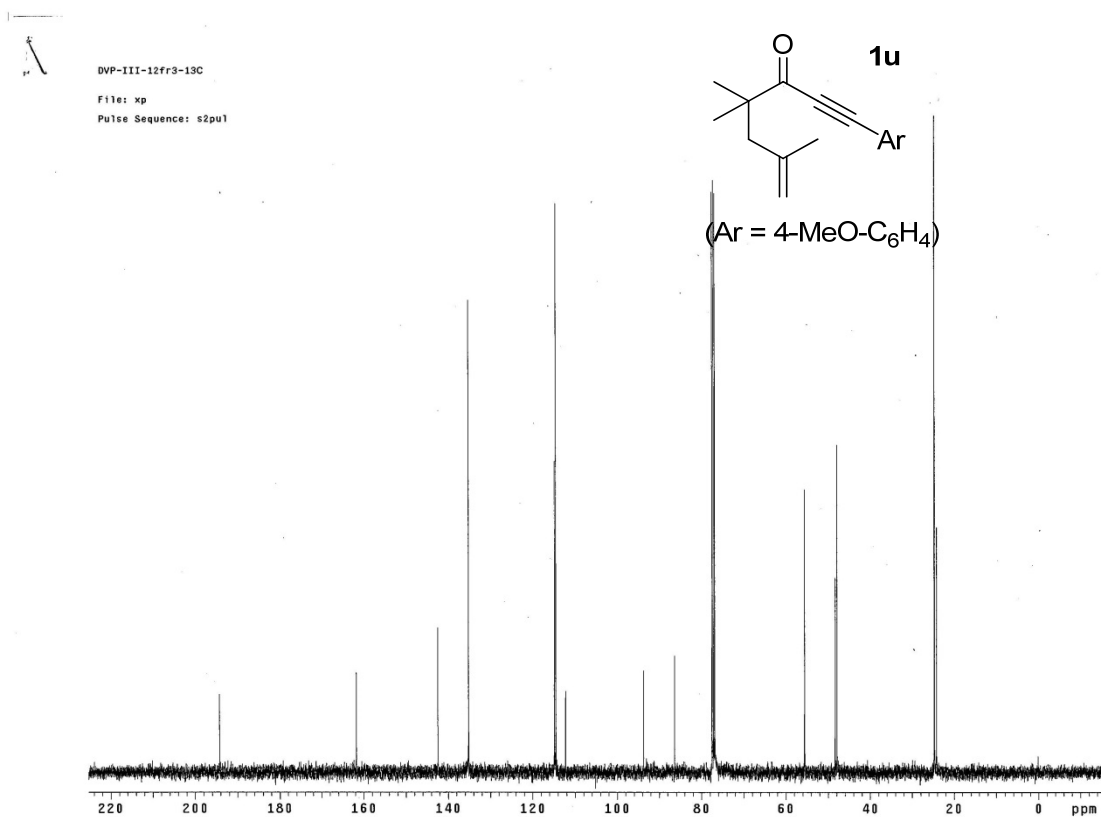
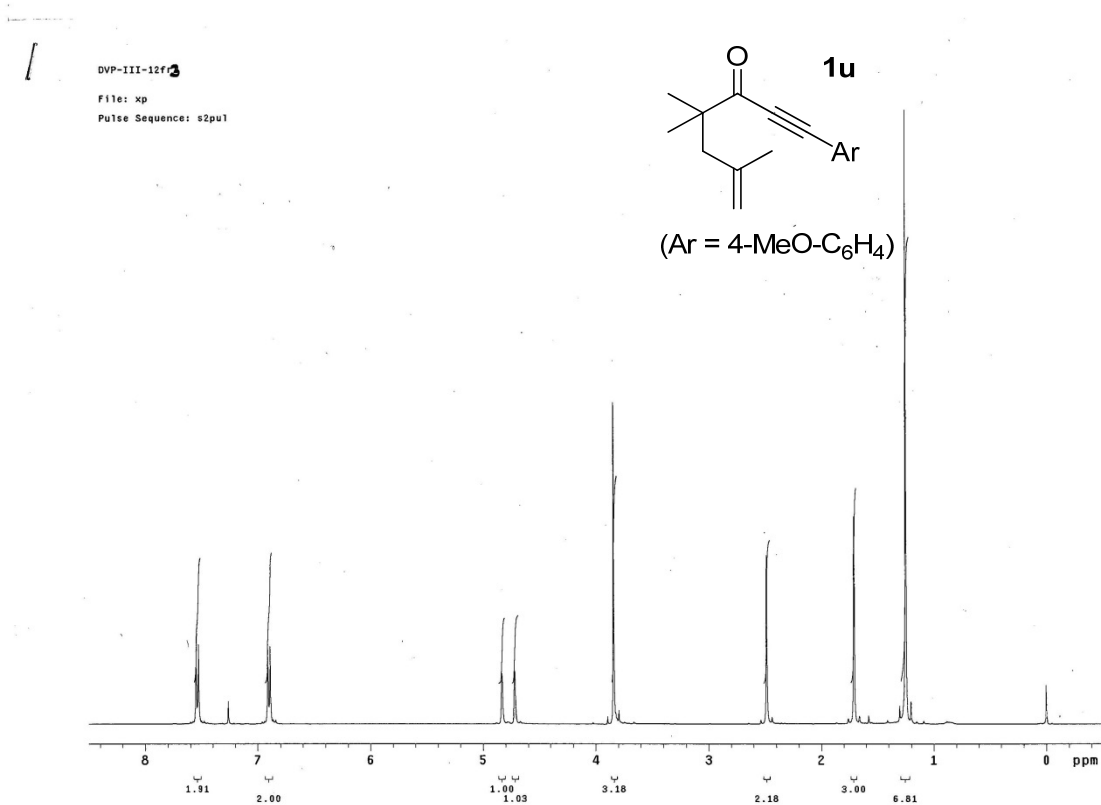
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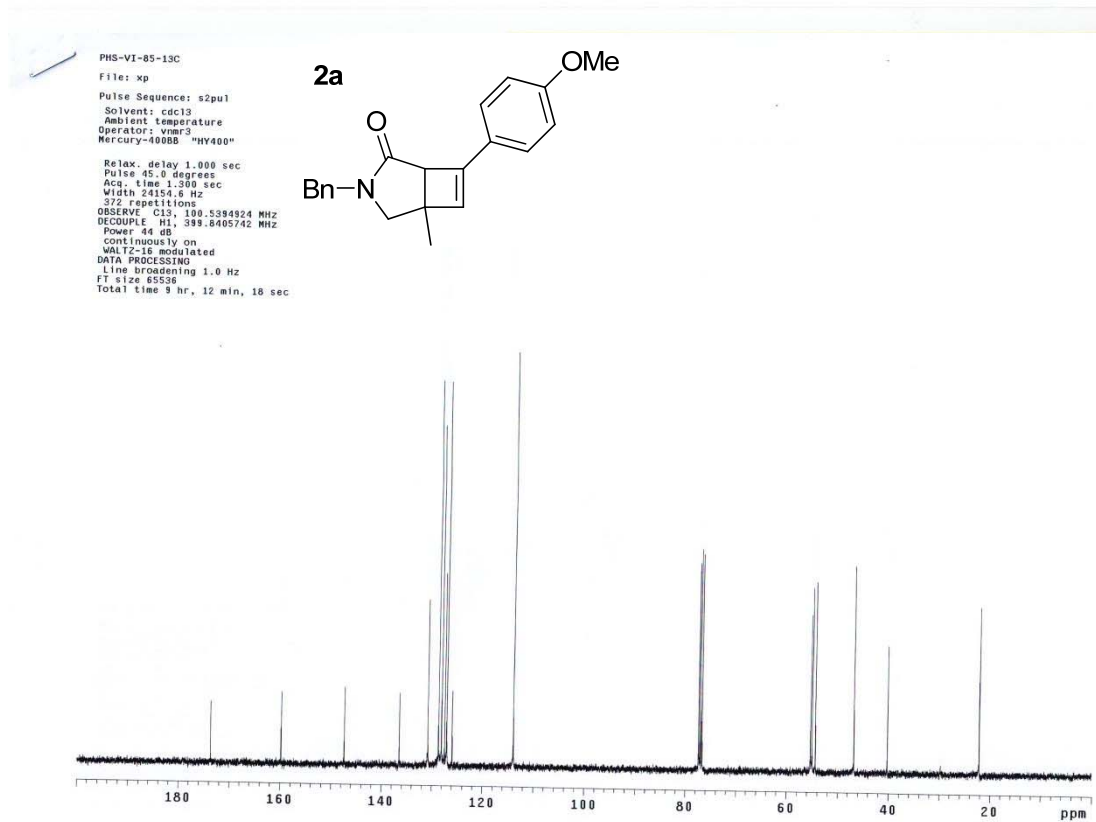
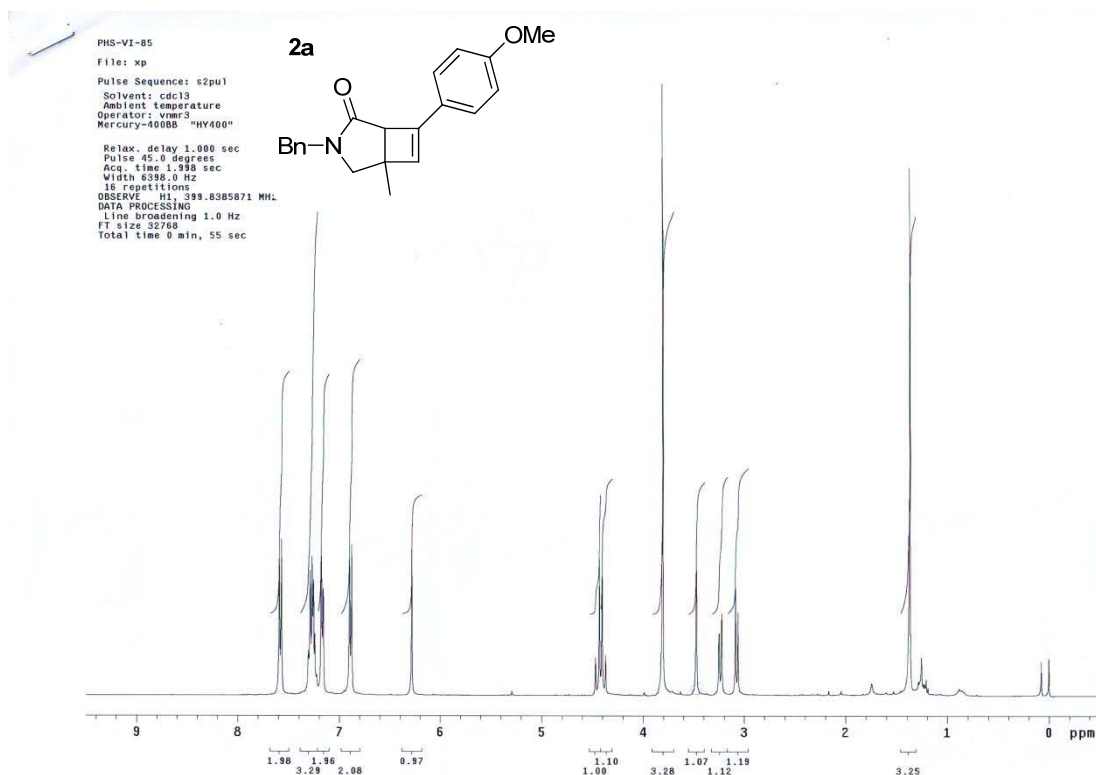
DVP-IV-44-13C
File: xp
Pulse Sequence: s2pu1

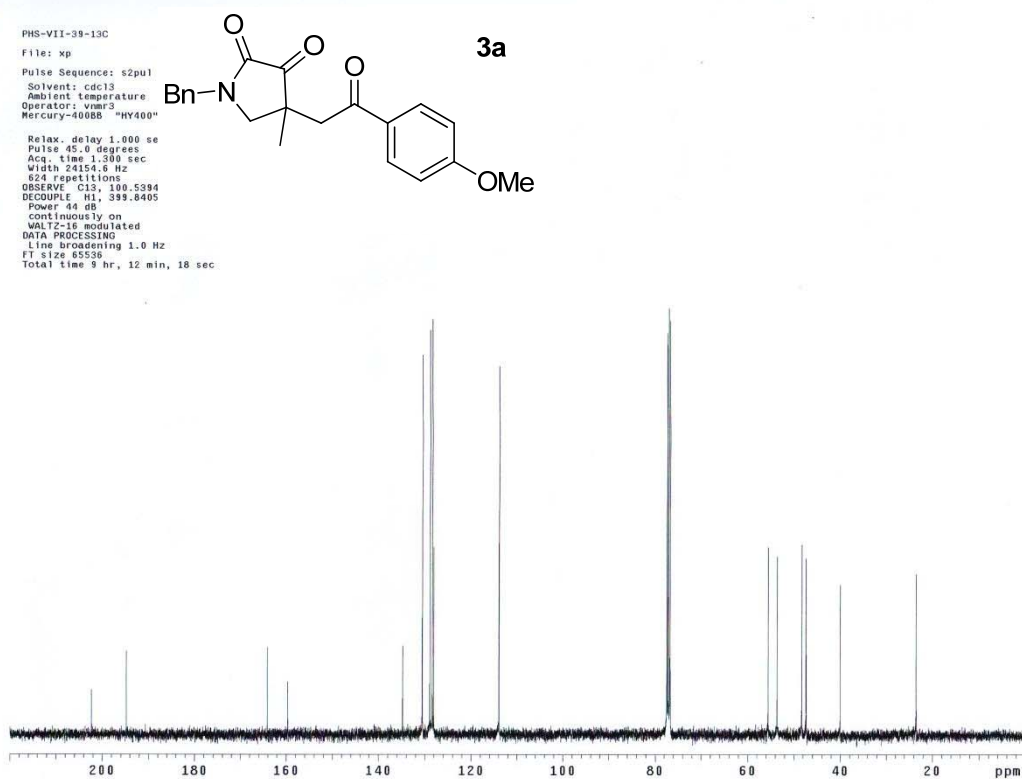
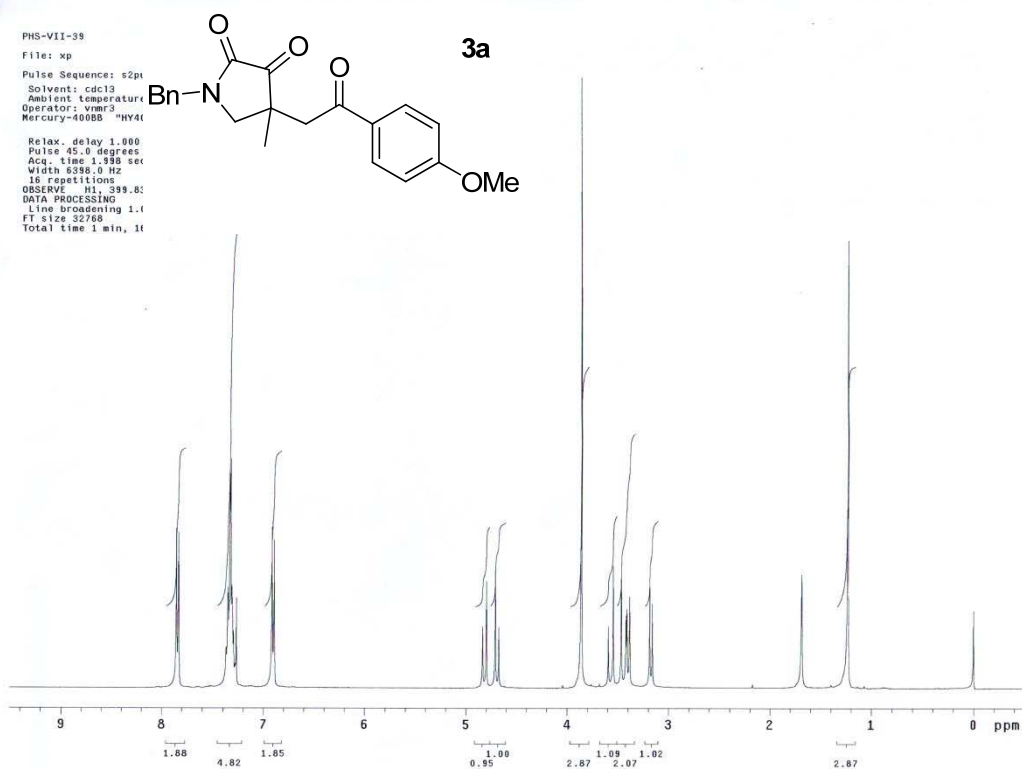


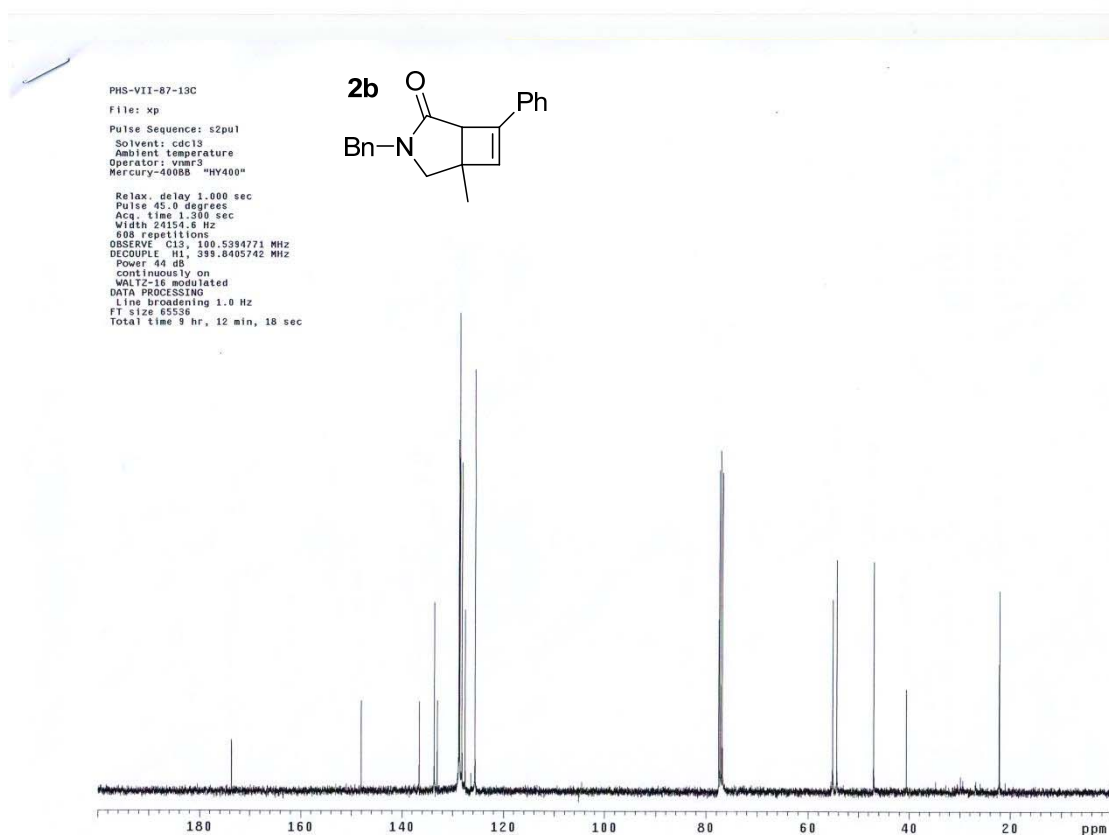
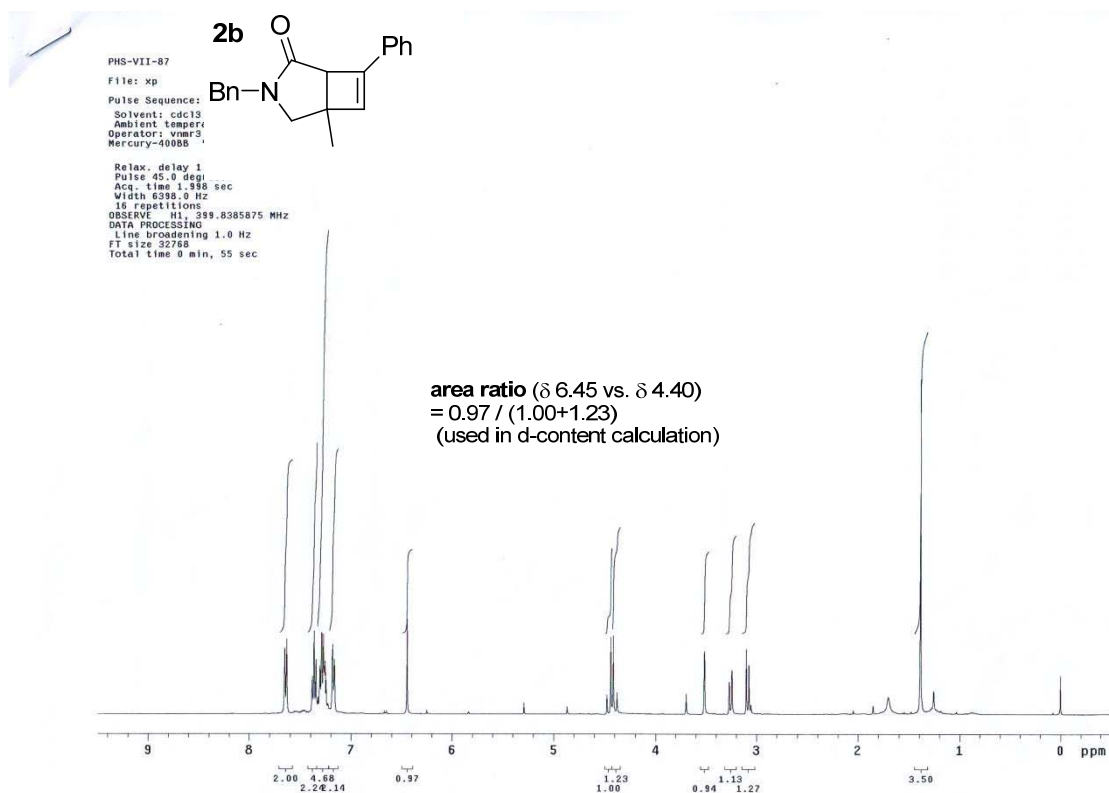
1t (Ar = 4-F-C₆H₄)

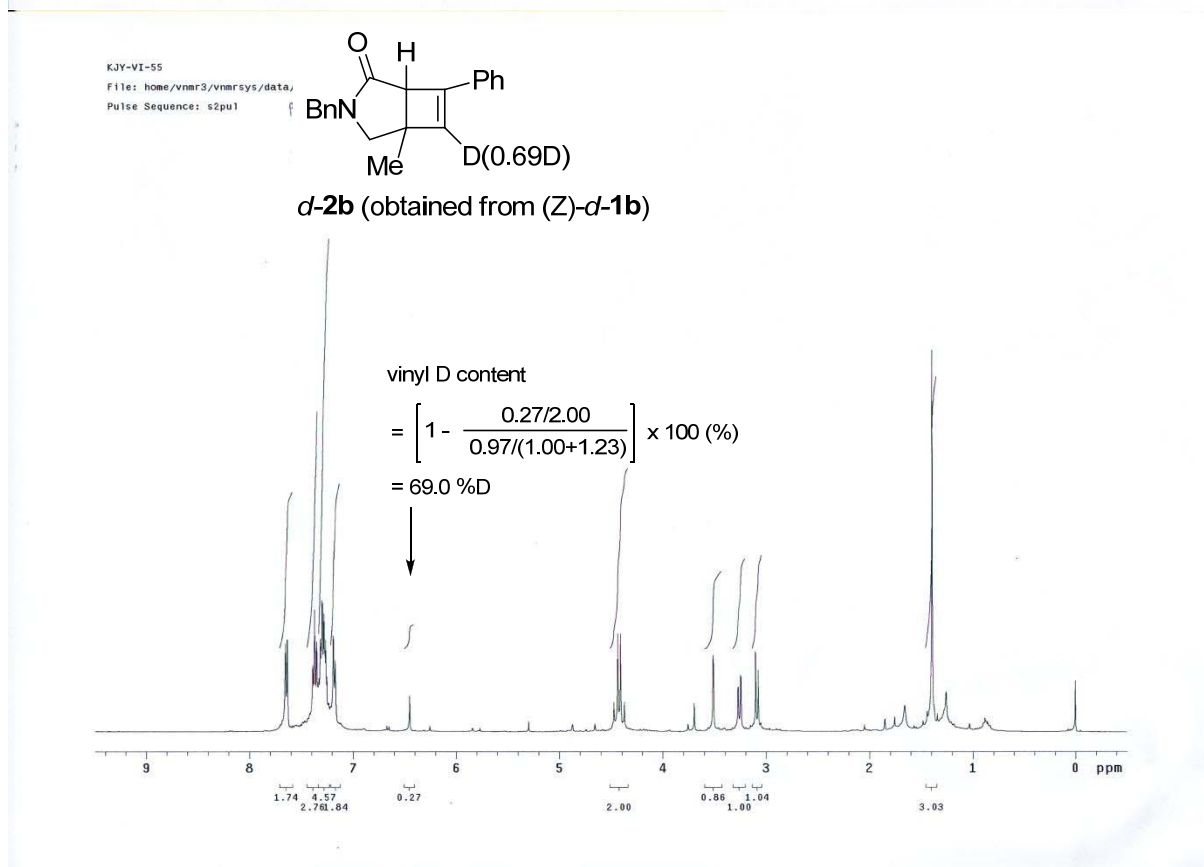
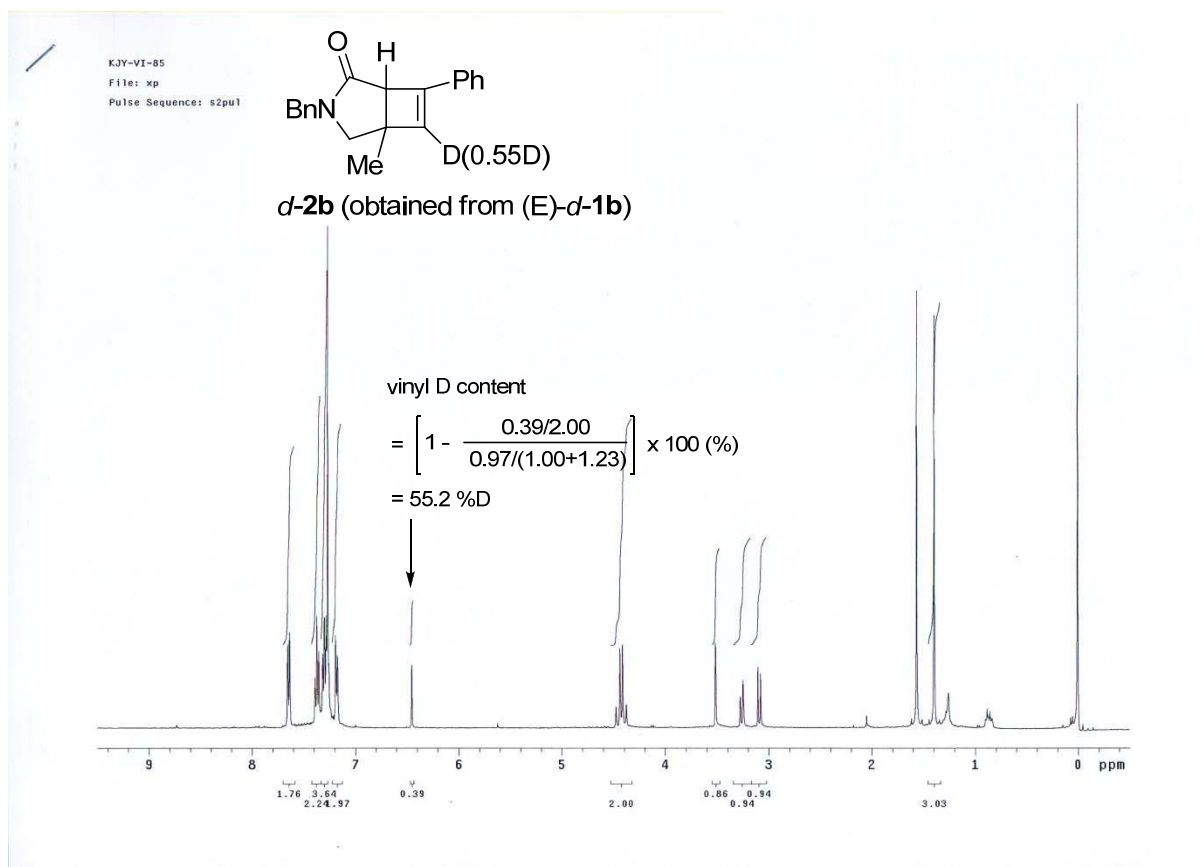




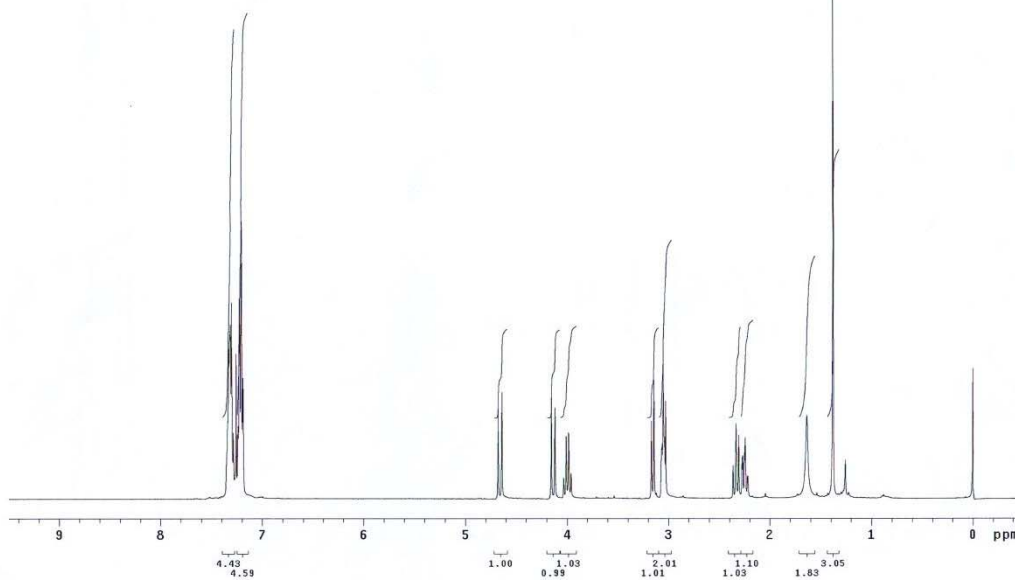
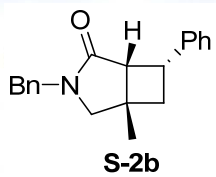




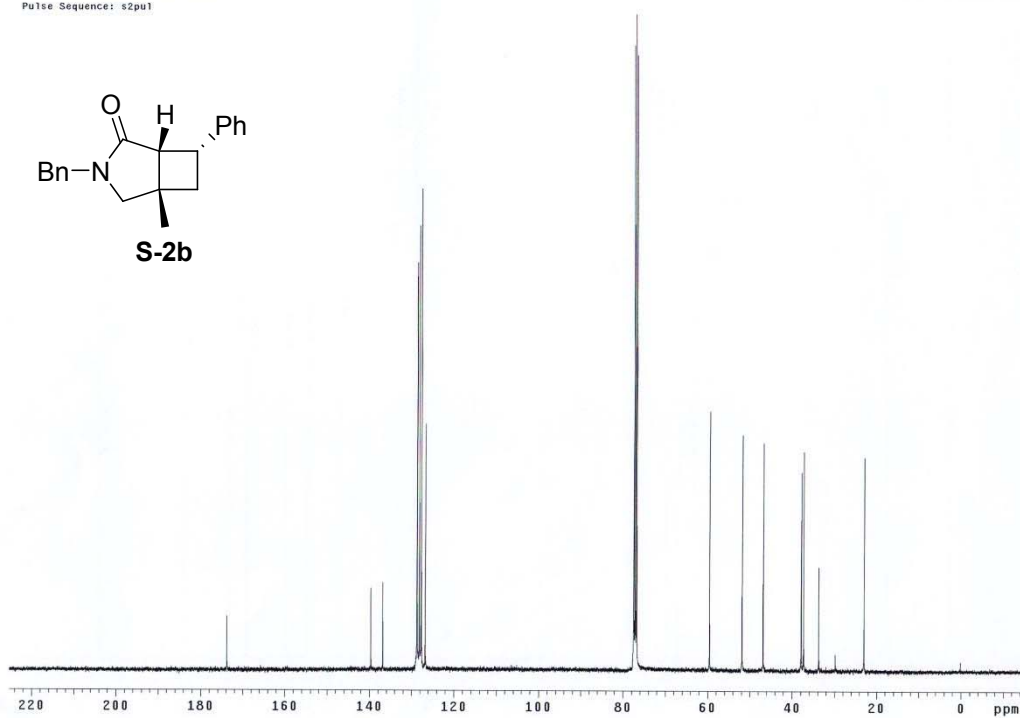
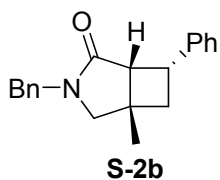




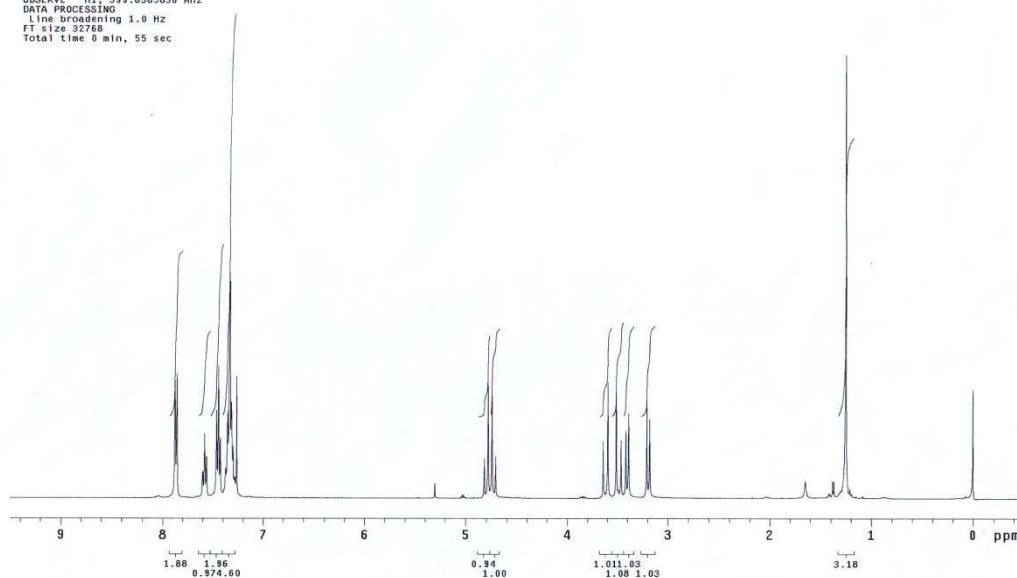
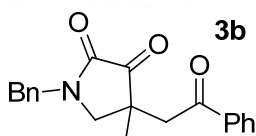
KJY-V-140-hydrogenation
File: xp
Pulse Sequence: s2pu1



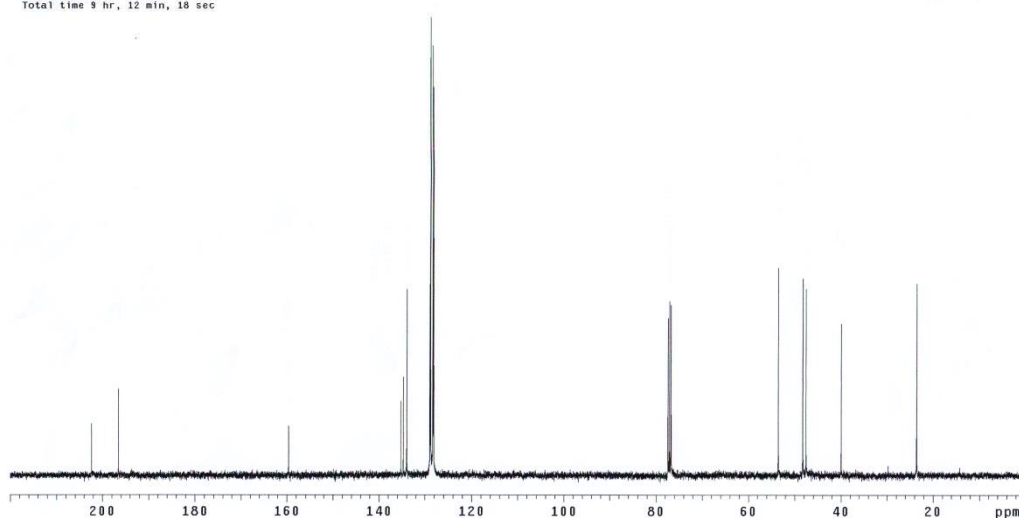
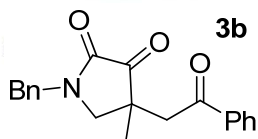
KJY-V-140-13C
File: home/vnmr3/KJY-V-140-13C.fid
Pulse Sequence: s2pu1



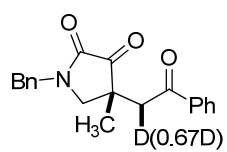
PHS-VI-54
 File: xp
 Pulse Sequence: s2pul
 Solvent: cdc13
 Ambient temperature
 Operator: vnmr3
 Mercury-400B "HY400"
 Relax. delay 1.000 sec
 Pulse 45.0 degrees
 Acq. time 1.998 sec
 Width 6398.0 Hz
 16 repetitions
 OBSERVE H1, 399.8385836 MHz
 DATA PROCESSING
 Line broadening 1.0 Hz
 FT size 32768
 Total time 0 min, 55 sec



PHS-VII-93-13C
 File: xp
 Pulse Sequence: s2pul
 Solvent: cdc13
 Ambient temperature
 Operator: vnmr3
 Mercury-400B "HY400"
 Relax. delay 1.000 sec
 Pulse 45.0 degrees
 Acq. time 1.300 sec
 Width 24154.6 Hz
 182 repetitions
 OBSERVE C13, 100.5334924 MHz
 DECOUPLE H1, 399.8405742 MHz
 Power 44 dB
 continuously on
 WALTZ-16 modulated
 DATA PROCESSING
 Line broadening 1.0 Hz
 FT size 65536
 Total time 9 hr, 12 min, 18 sec

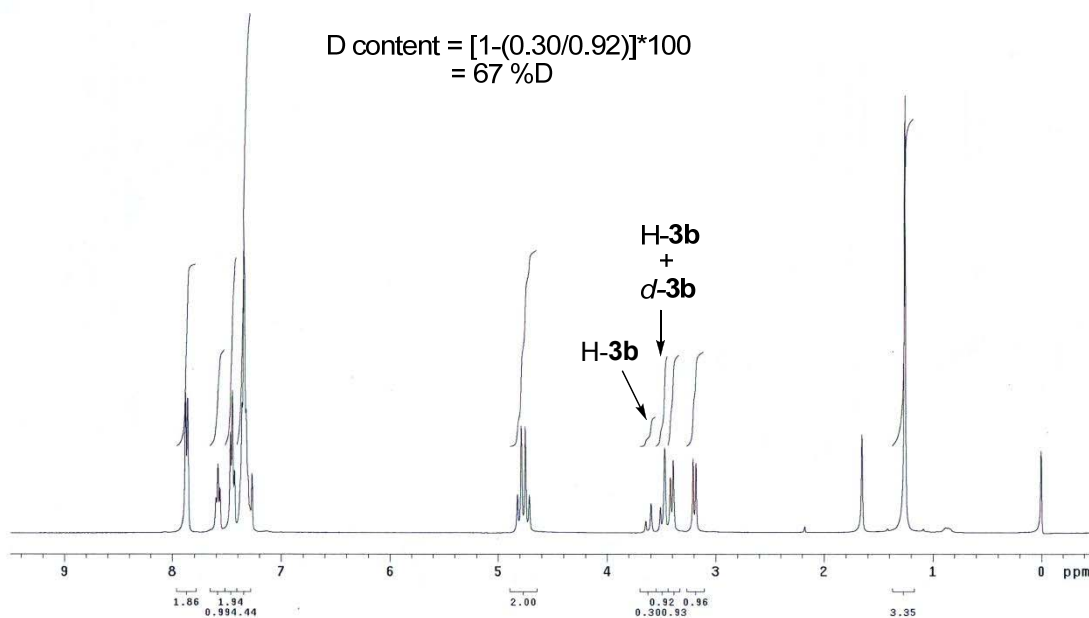


KJY-VI-33-recryst
File: xp
Pulse Sequence: s2pu1

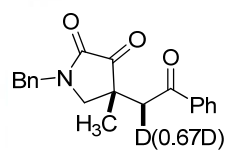


d-**3b** (Eq 3)

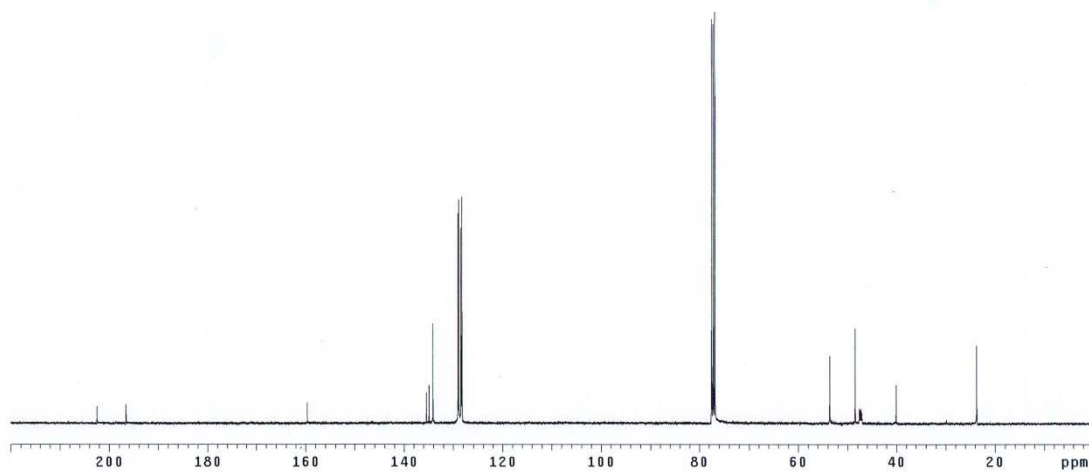
$$\text{D content} = [1 - (0.30/0.92)] \times 100 = 67\% \text{D}$$



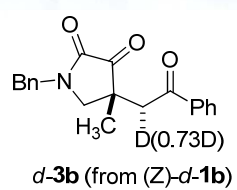
Std Carbon experiment
File: home/vnmr3/vnmrsys/data/KJY/KJY-VI-33-13C.fid
Pulse Sequence: s2pu1



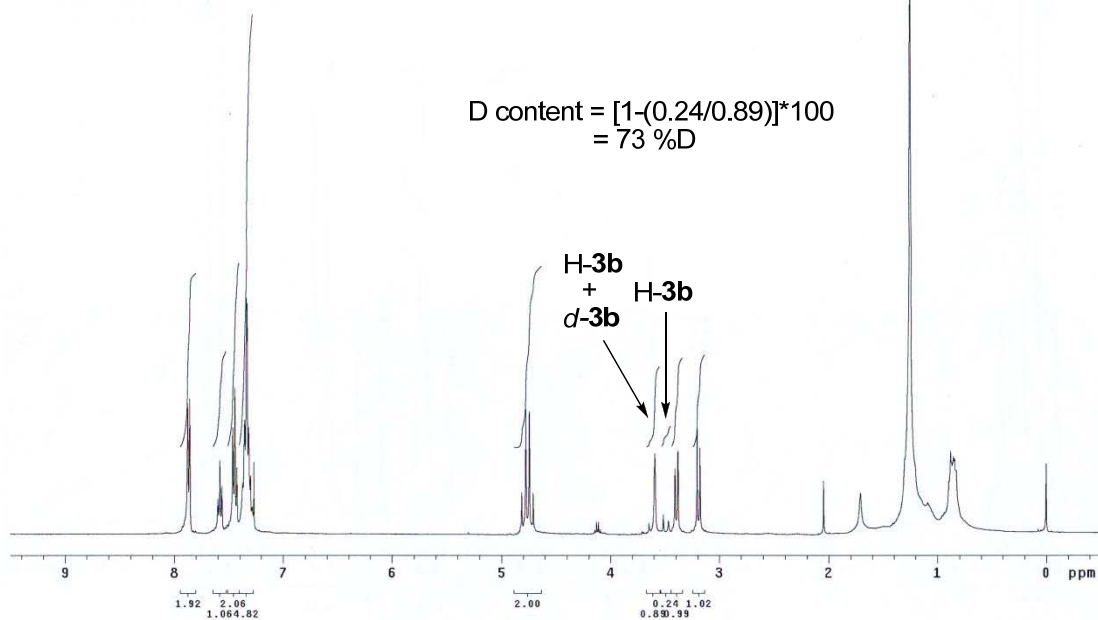
d-**3b** (Eq 3)

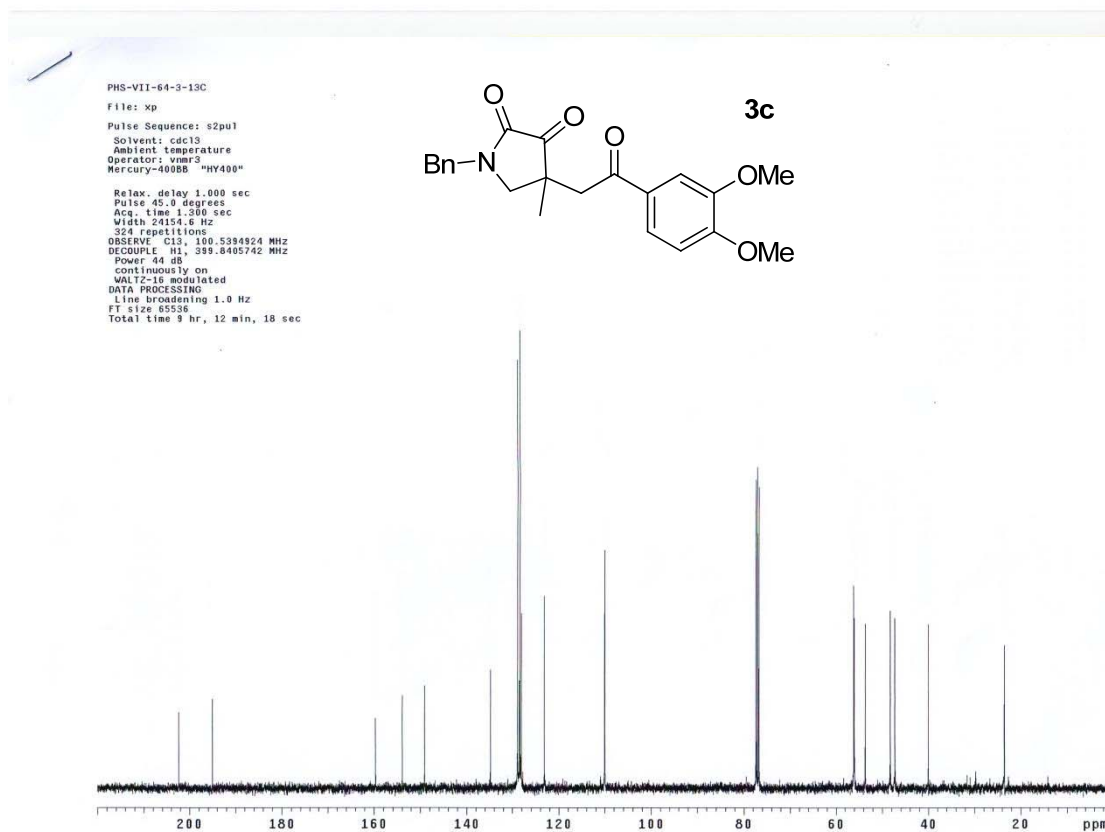
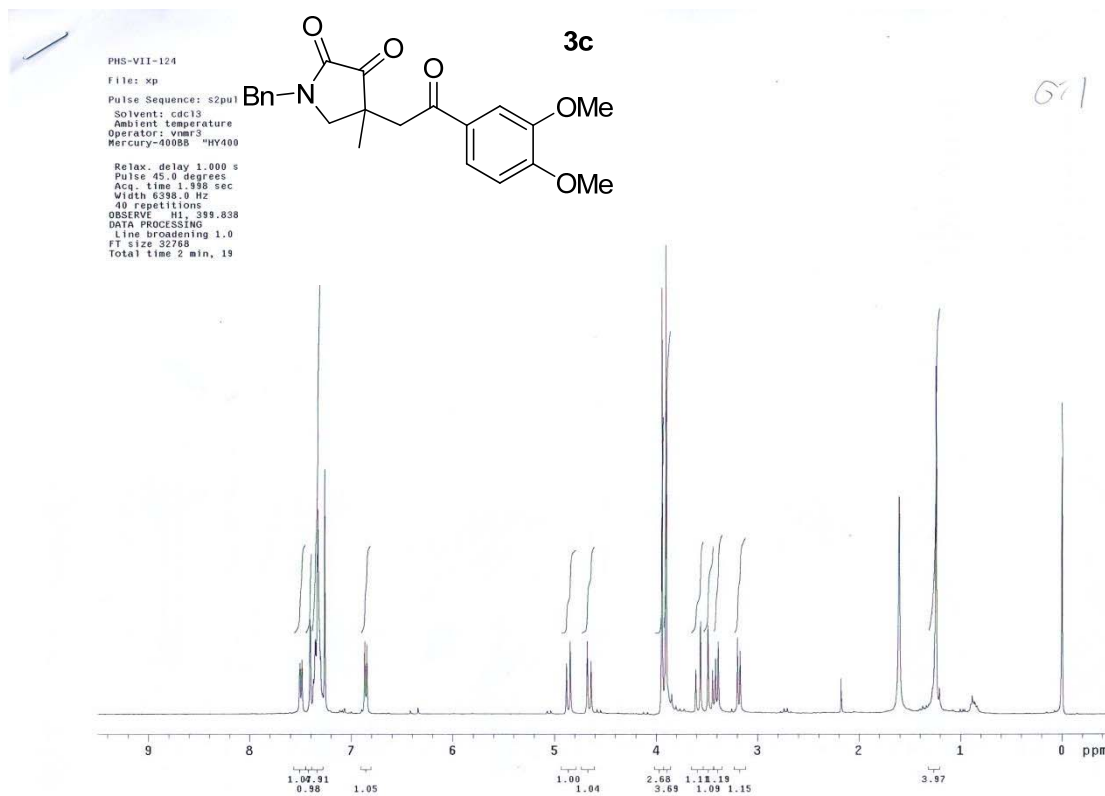


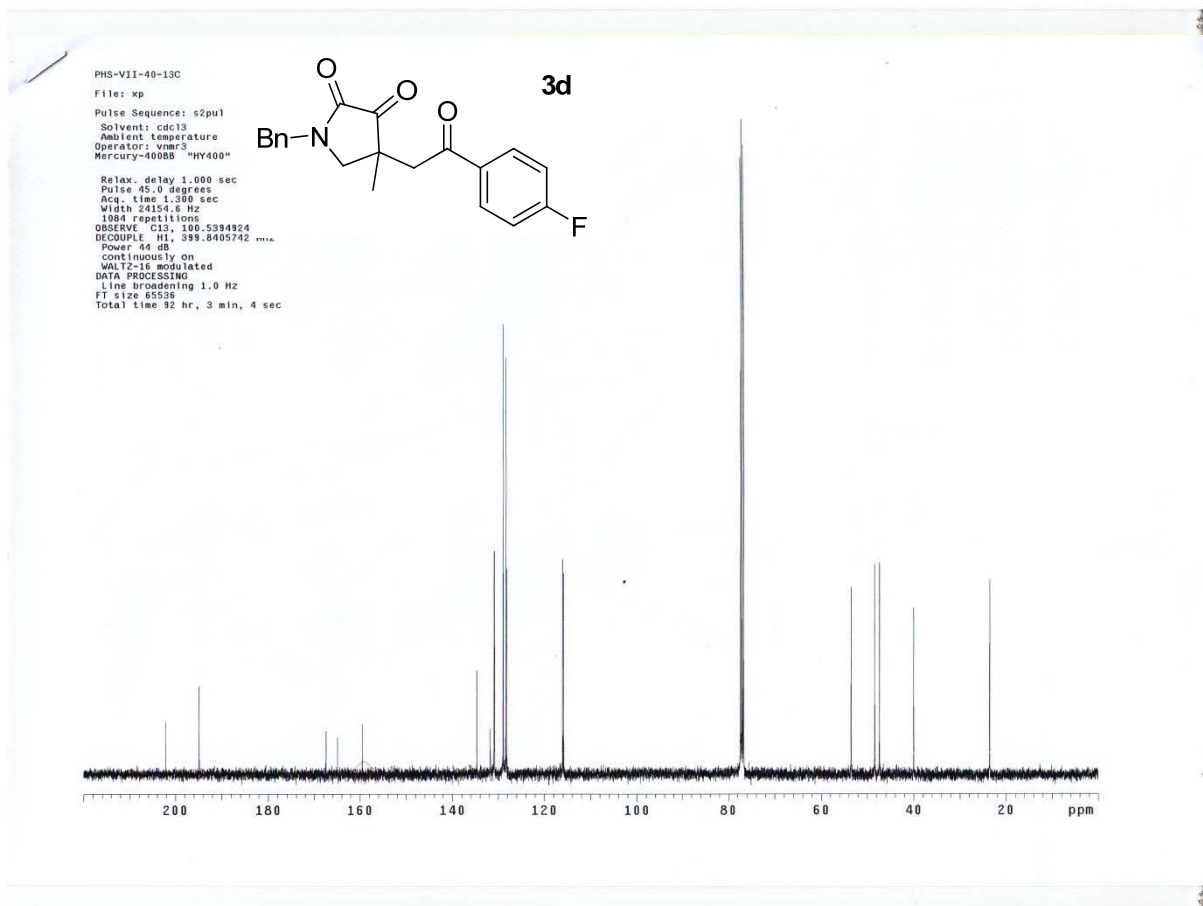
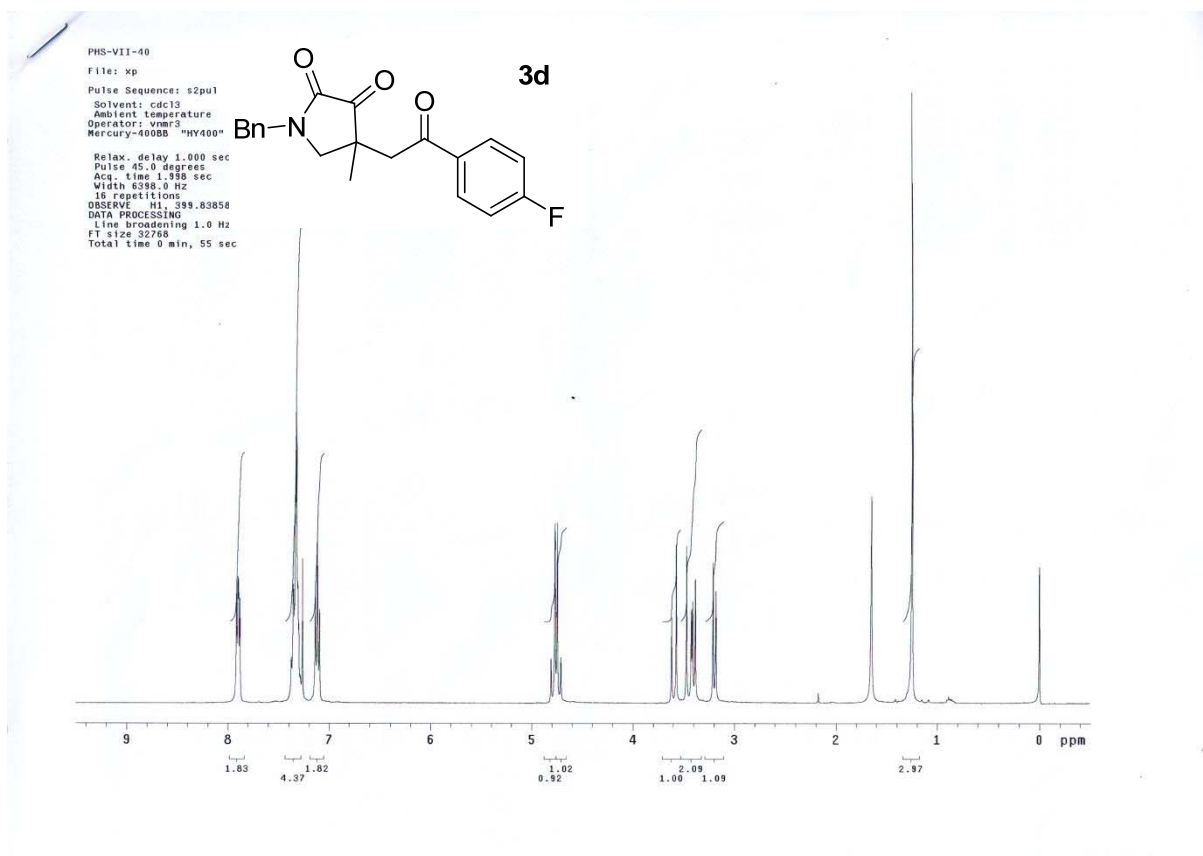
KJY-VI-50
 File: xp
 Pulse Sequence: s2pu1

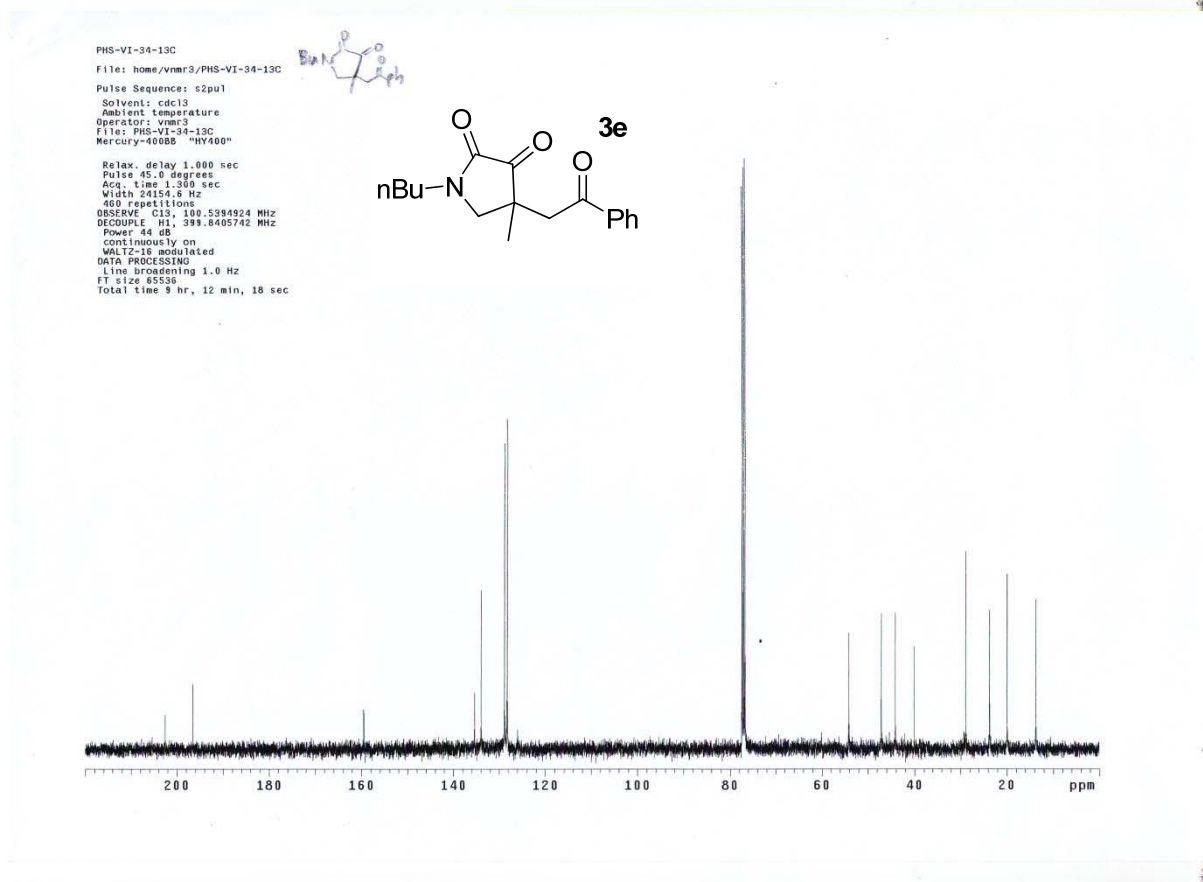
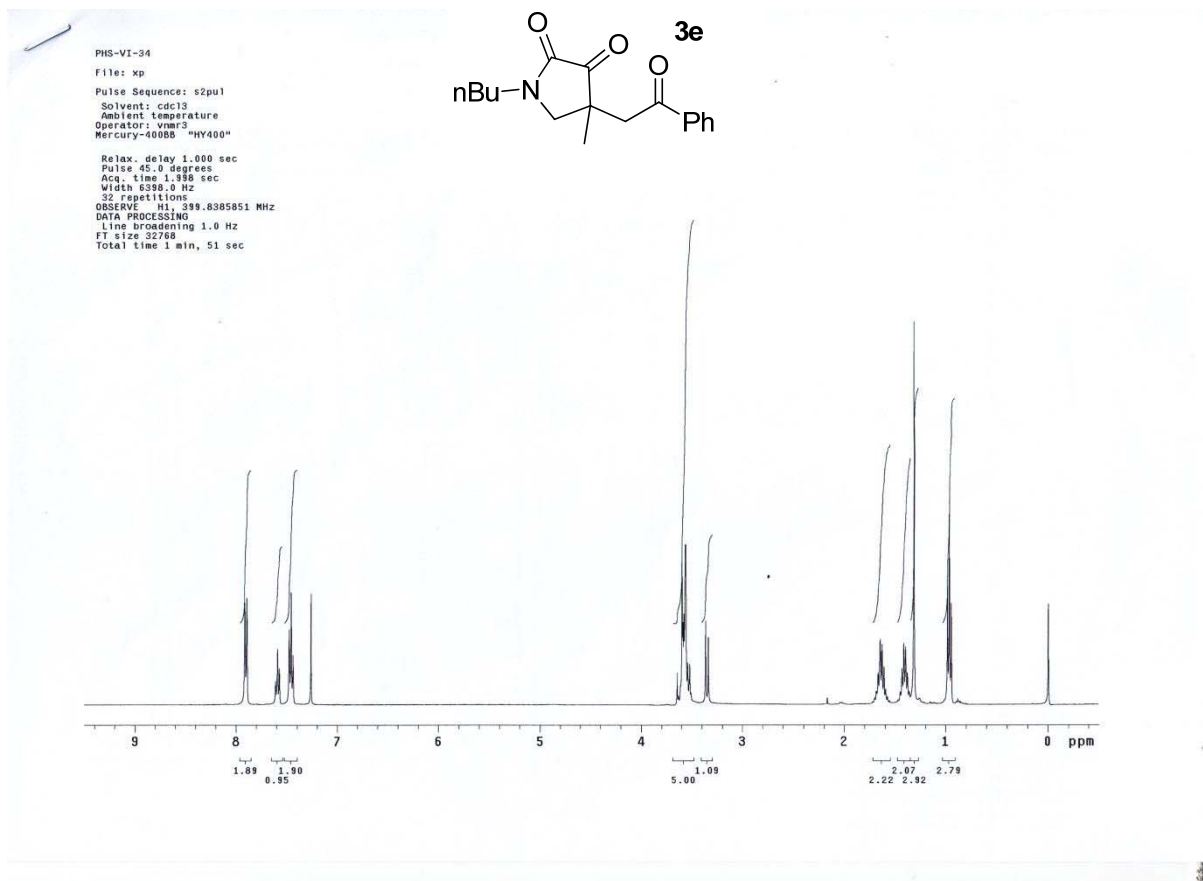


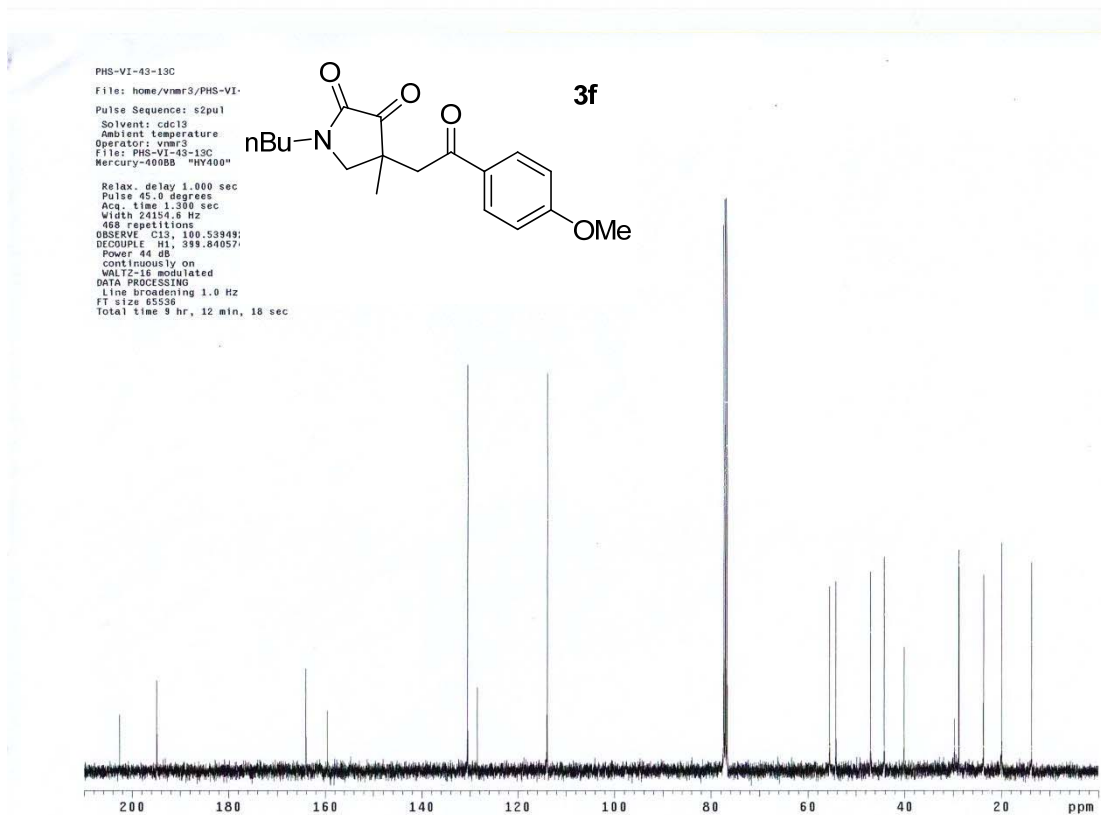
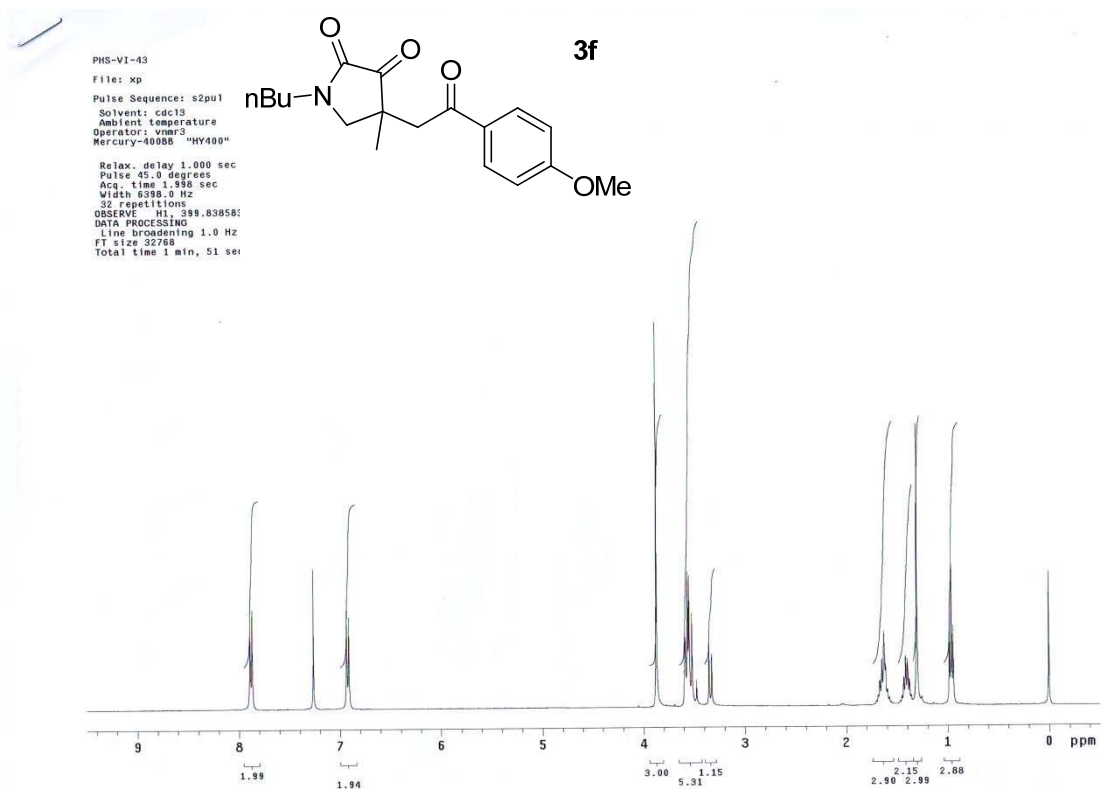
$$\text{D content} = [1 - (0.24/0.89)] * 100 = 73 \% \text{D}$$

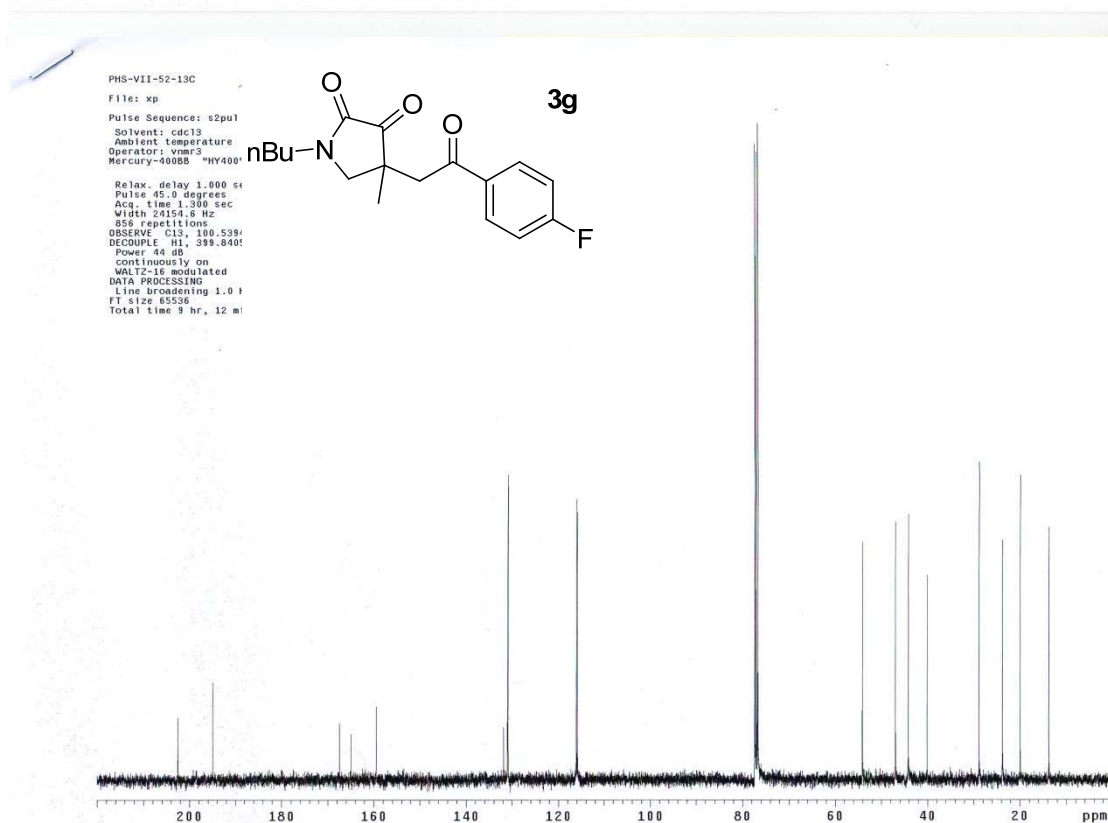
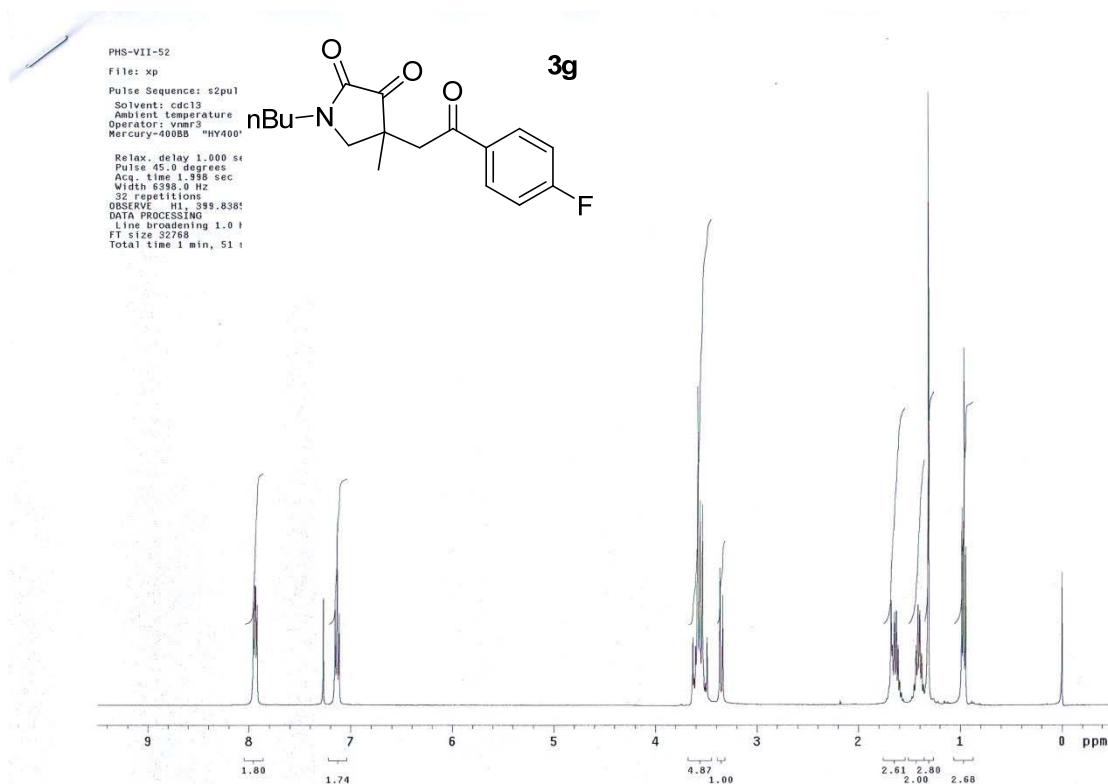


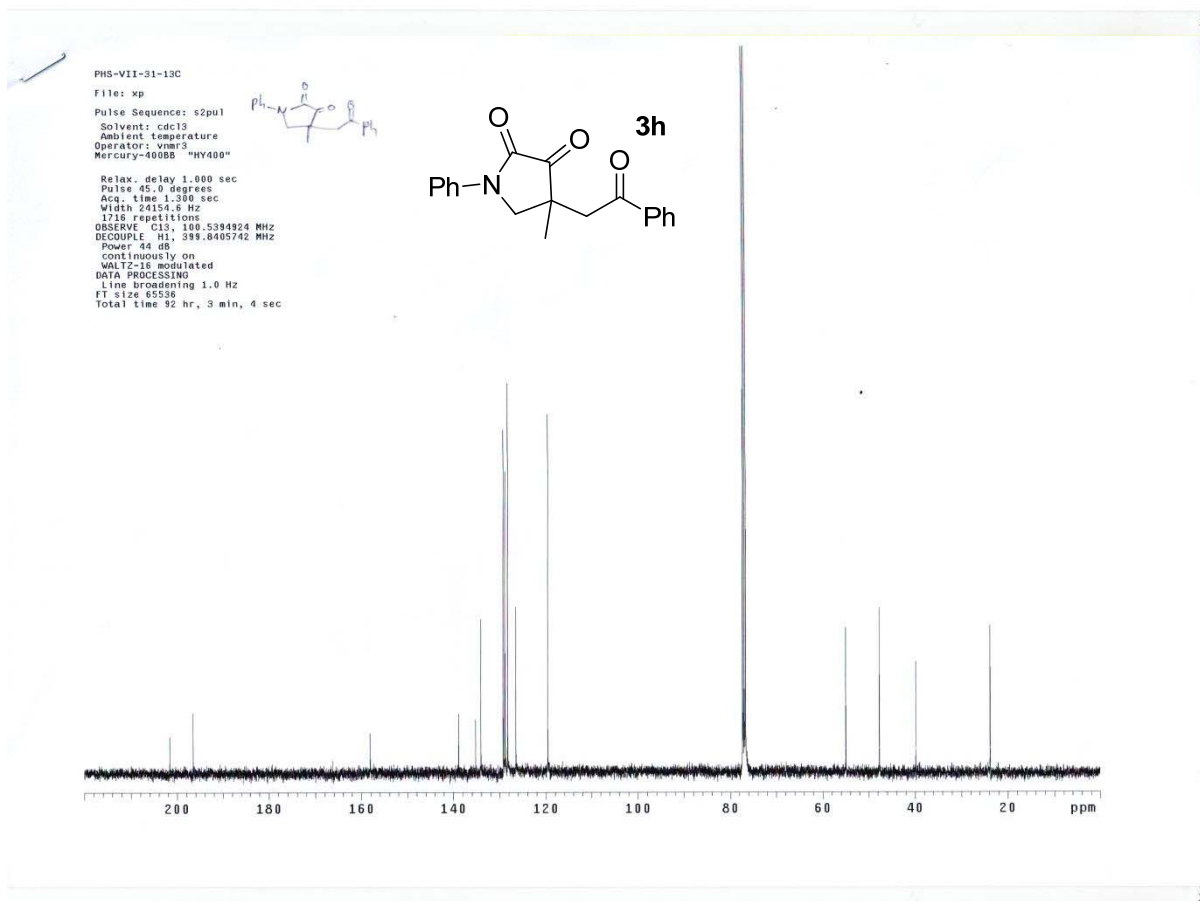
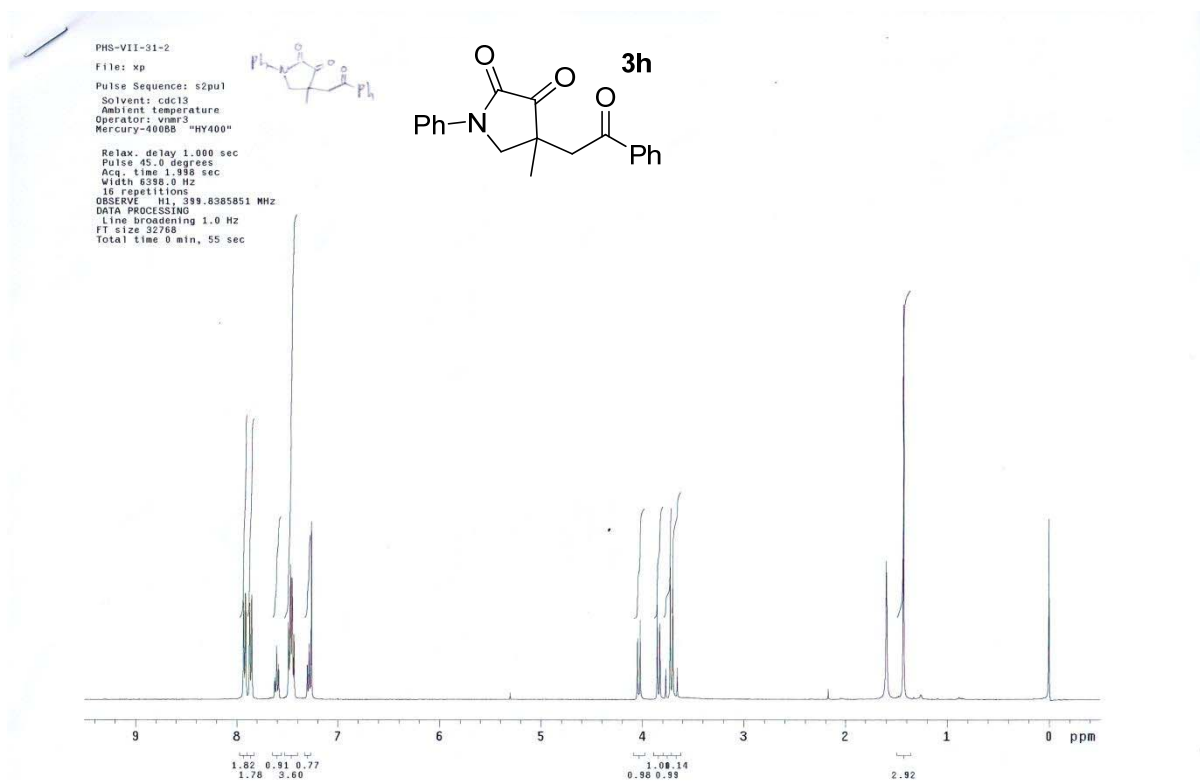












PHS-VII-29-2

File: xp

Pulse Sequence: s2pul

Solvent: cdcl3

Ambient temperature

Operator: vnmr3

Mercury-400BB "HY400"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.988 sec

Width 6389.0 Hz

68 repetitions

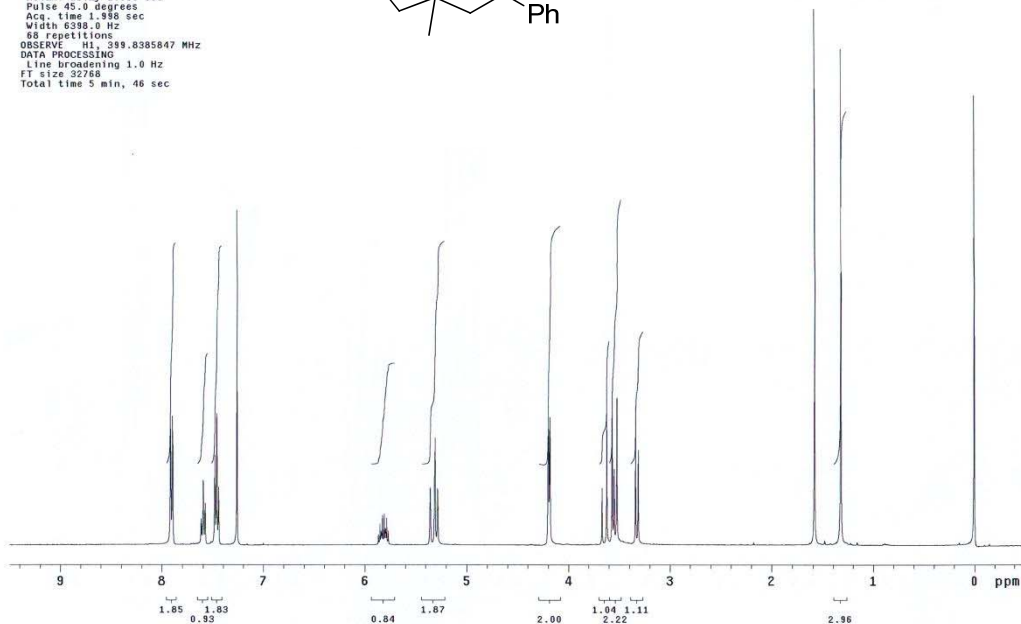
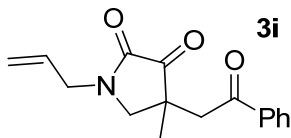
OBSERVE H1, 399.8385847 MHz

DATA PROCESSING

Line broadening 1.0 Hz

FT size 32768

Total time 5 min, 46 sec



PHS-VII-29-2-13C

File: home/vnmr3/PHS-VII-29-2-13C.fid

Pulse Sequence: s2pul

Solvent: cdcl3

Ambient temperature

Operator: vnmr3

File: PHS-VII-29-2-13C

Mercury-400BB "HY400"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.980 sec

Width 24154.6 Hz

1748 repetitions

OBSERVE C13, 100.5394925 MHz

DECOUPLE H1, 399.8405742 MHz

Power 44 dB

continuously on

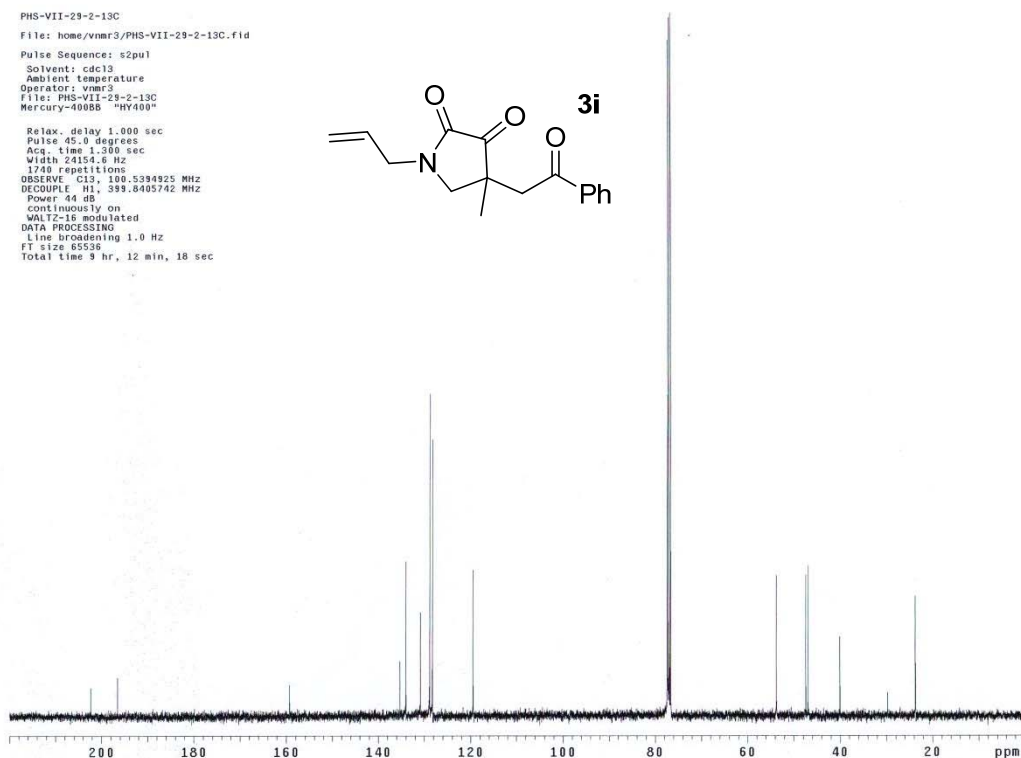
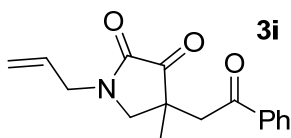
WALTZ-16 modulated

DATA PROCESSING

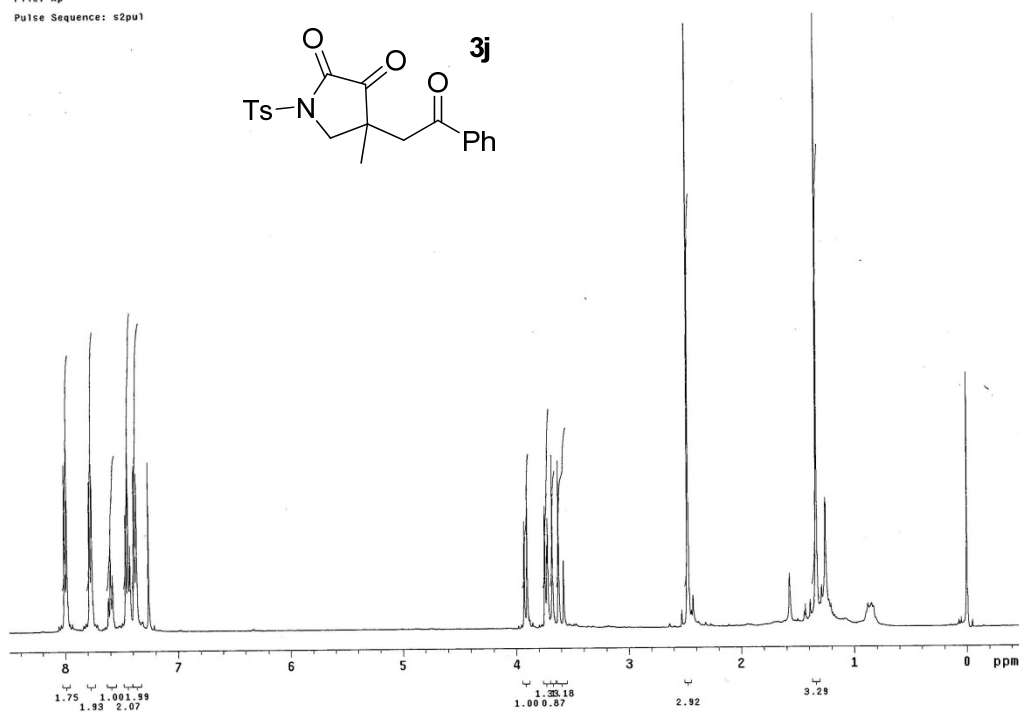
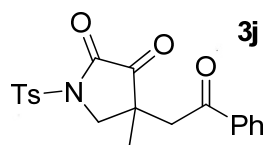
Line broadening 1.0 Hz

FT size 65536

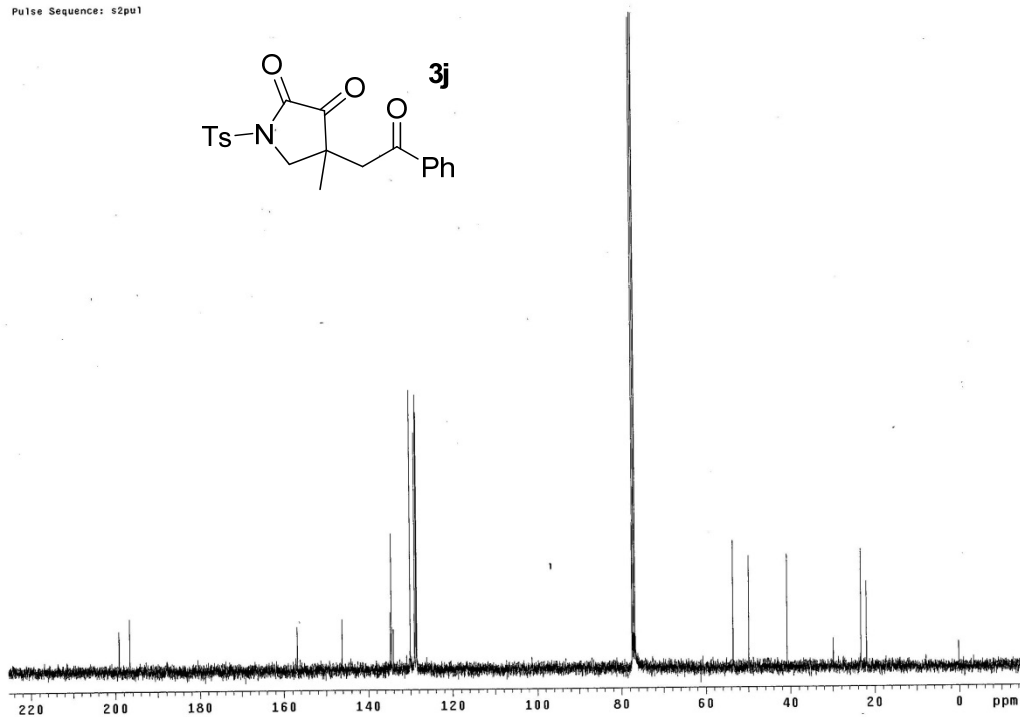
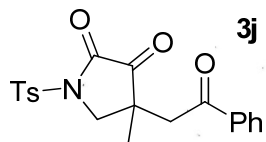
Total time 9 hr, 12 min, 18 sec

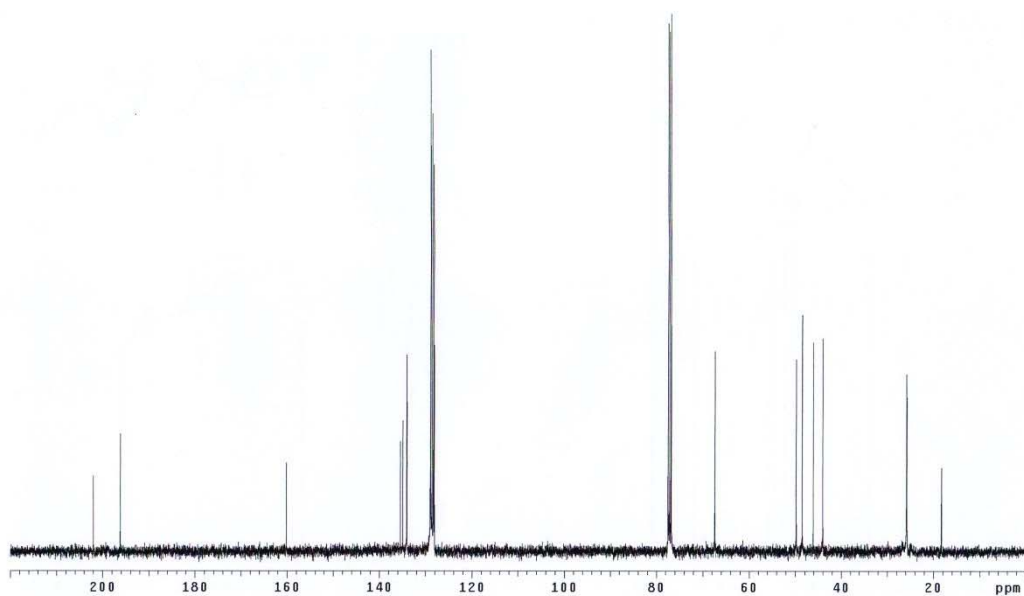
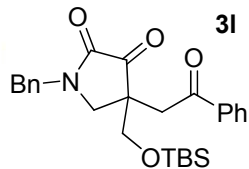
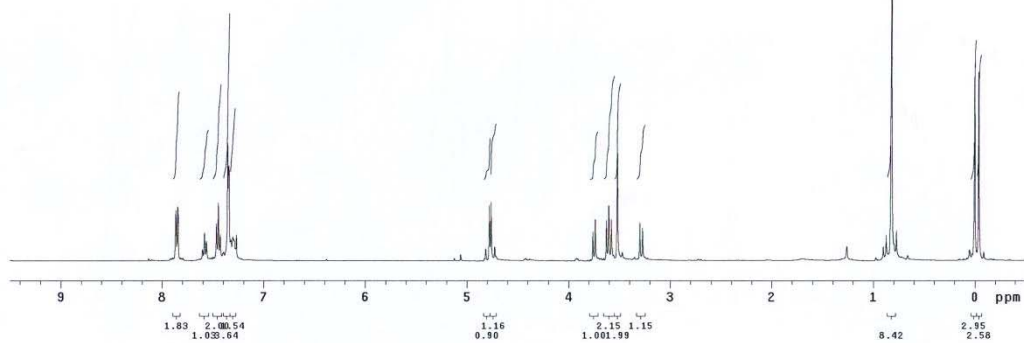
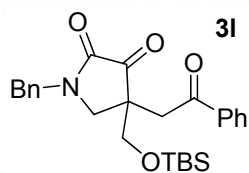


DVP-III-44fr2-p
File: xp
Pulse Sequence: s2pu1

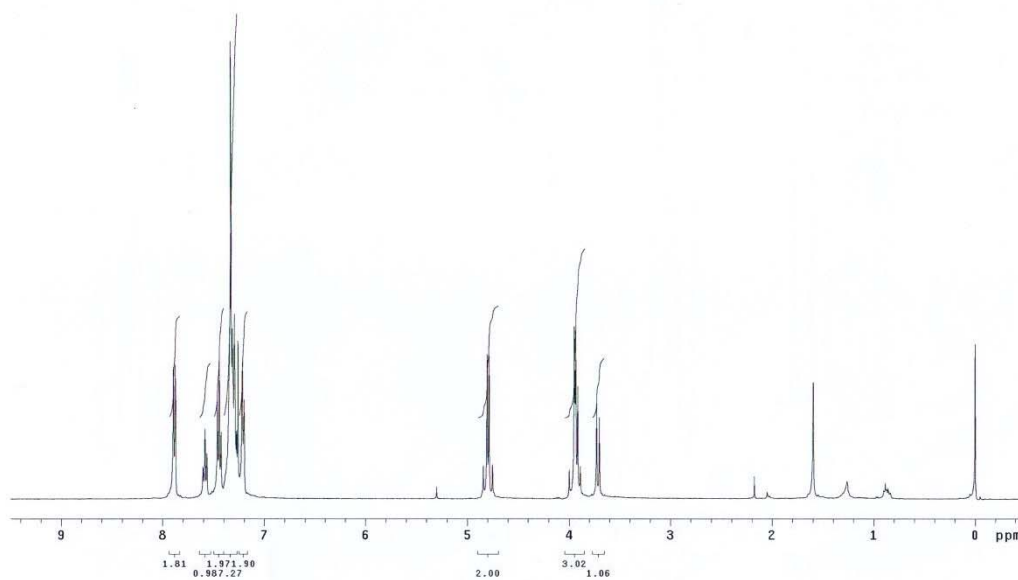
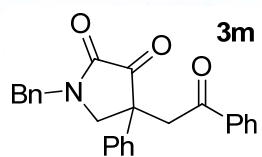


DVP-III-44fr2-p-13C
File: xp
Pulse Sequence: s2pu1

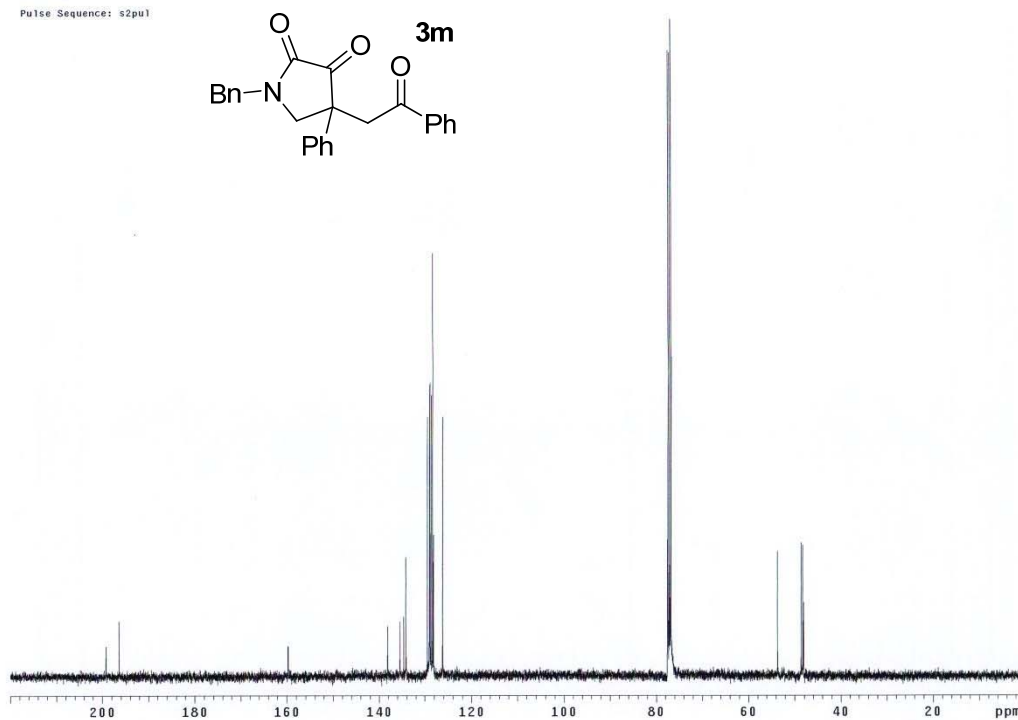
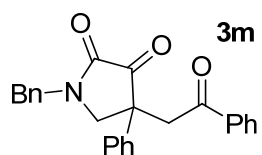


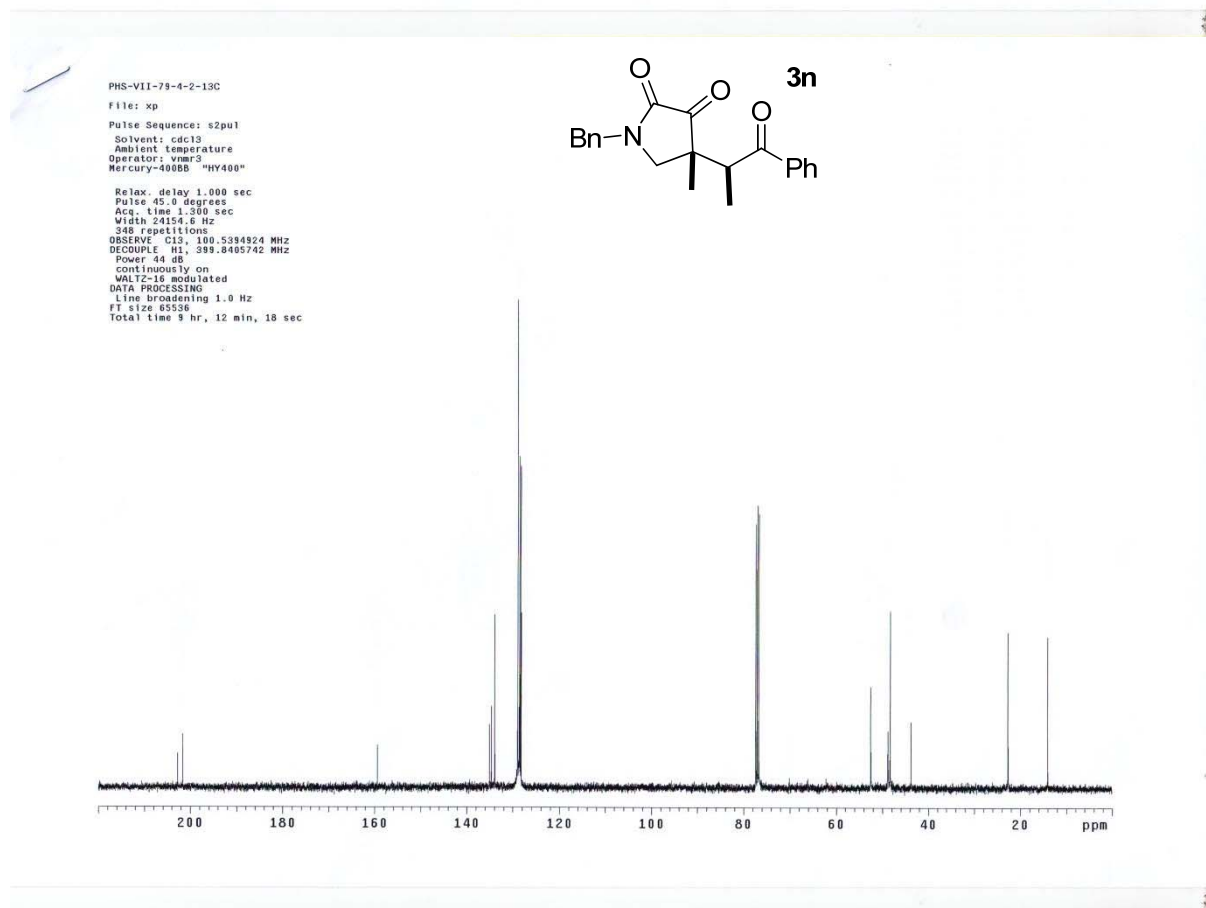
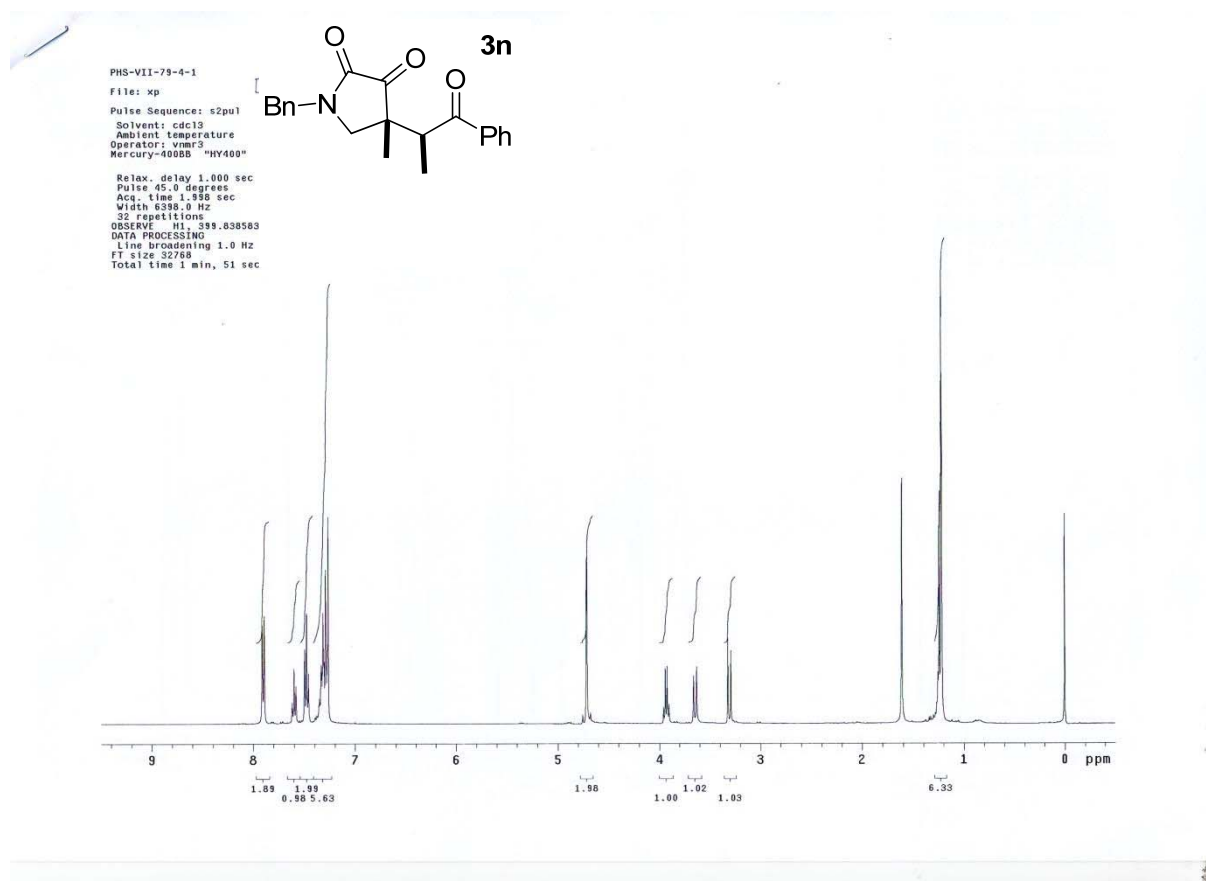


KJY-VI-81-recrystalliz-1H
File: xp
Pulse Sequence: s2pu1



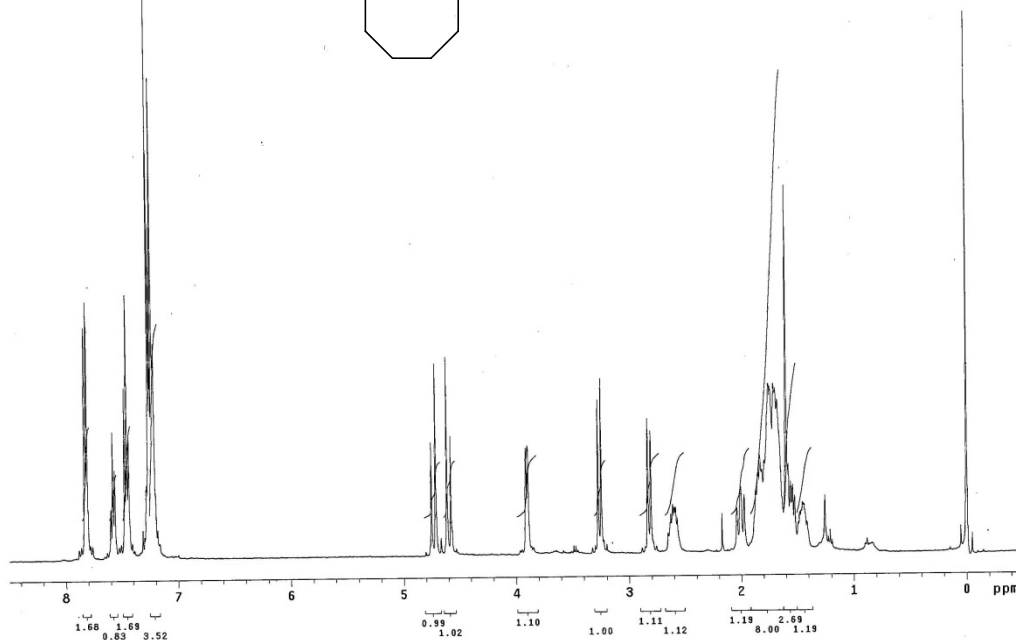
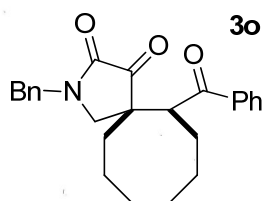
KJY-VI-81-recrystalliz-13C
File: xp
Pulse Sequence: s2pu1



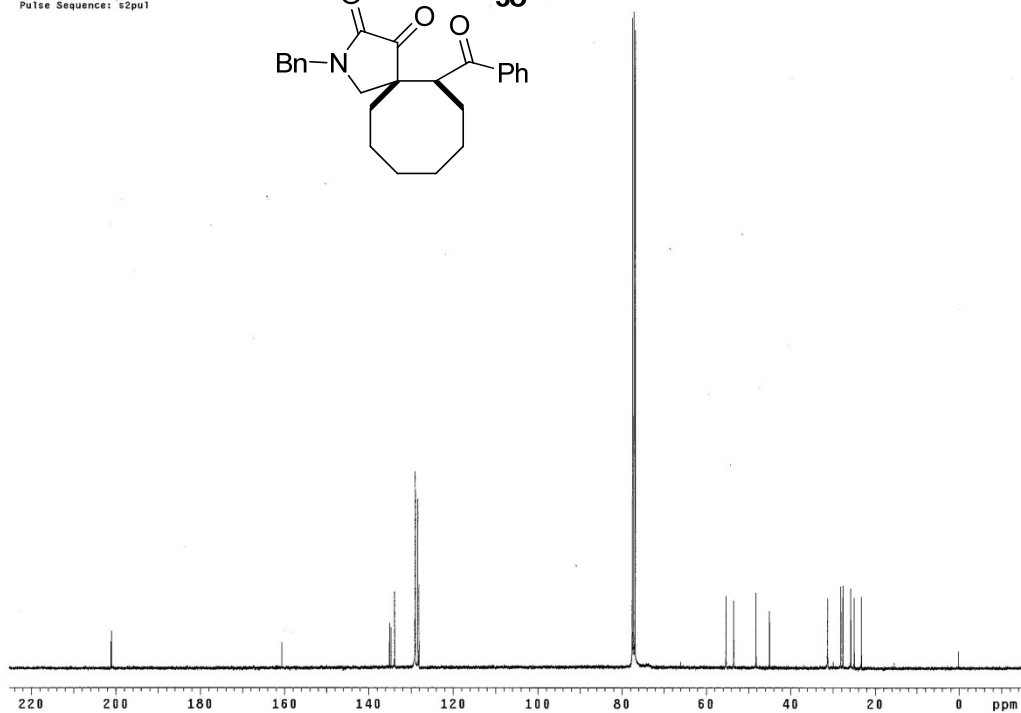
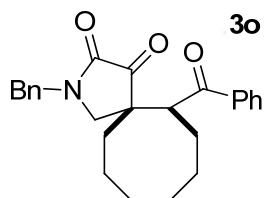


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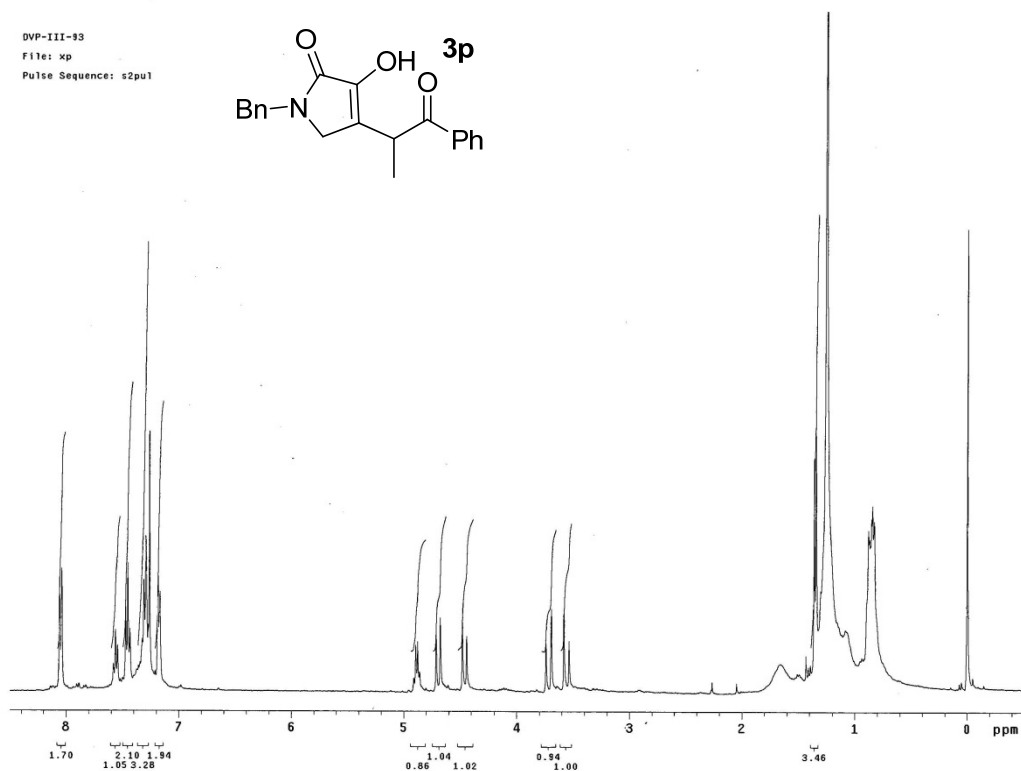
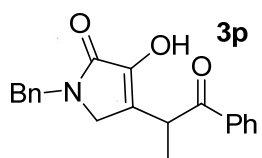
DVP-III-70
File: xp
Pulse Sequence: s2pu1



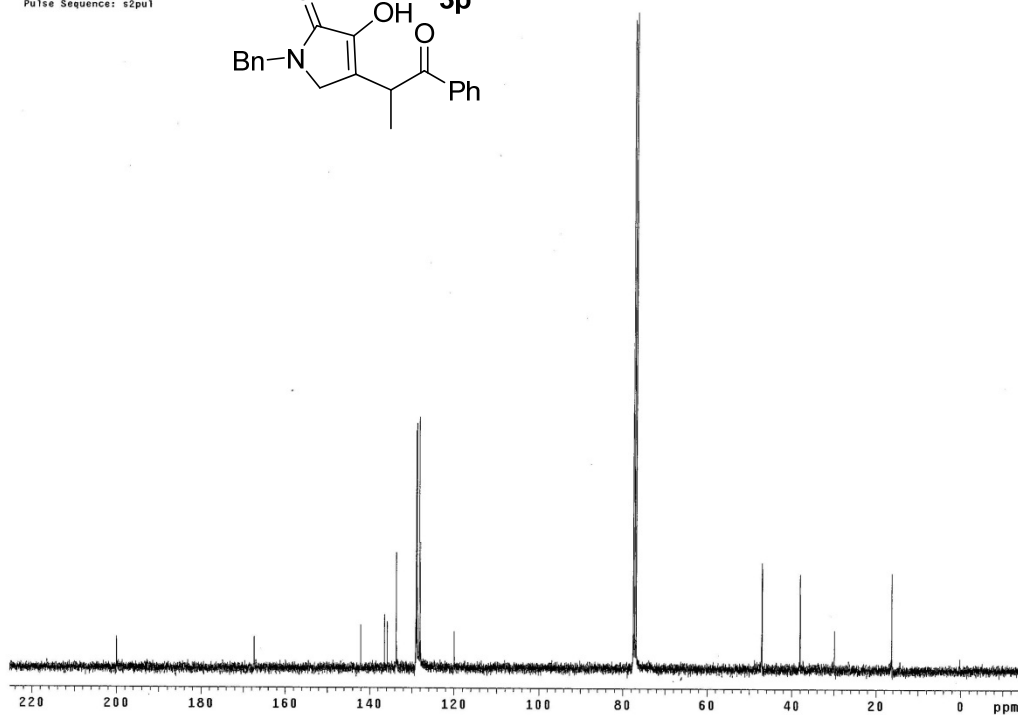
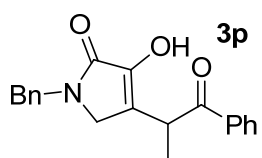
DVP-III-70-13C
File: xp
Pulse Sequence: s2pu1



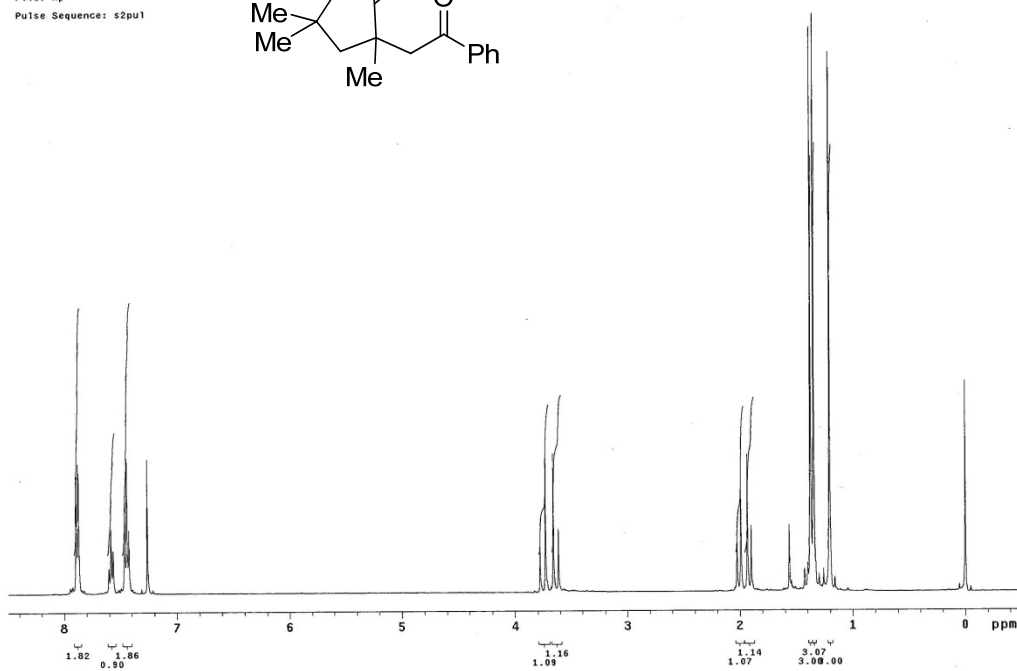
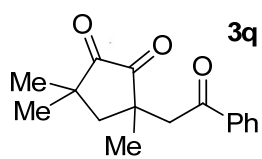
DVP-III-93
File: xp
Pulse Sequence: s2pu1



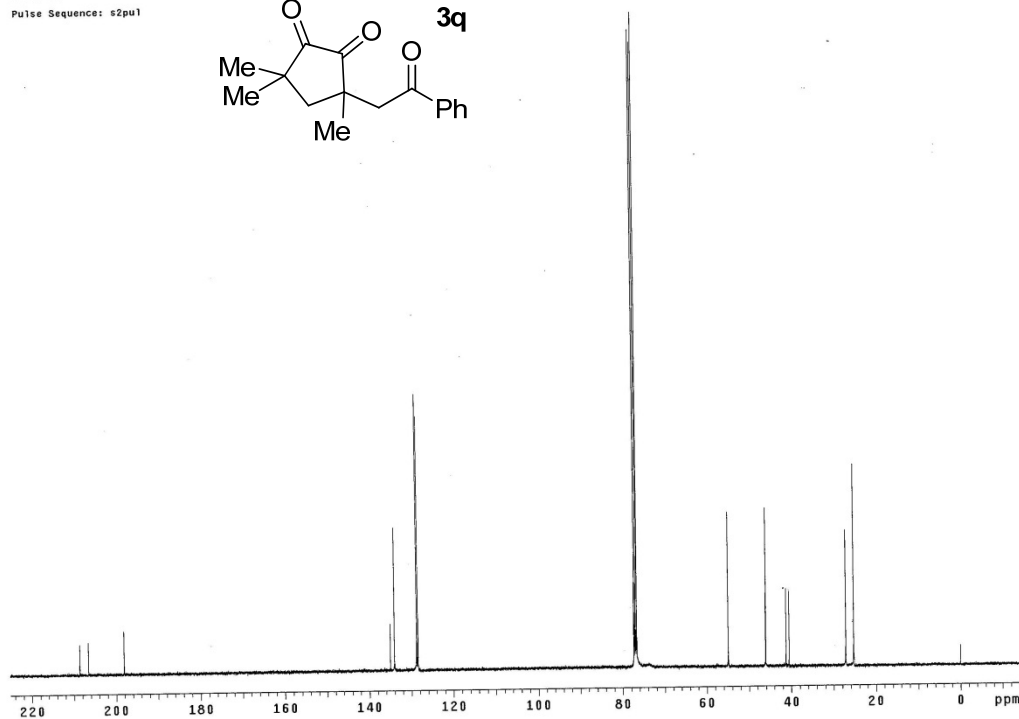
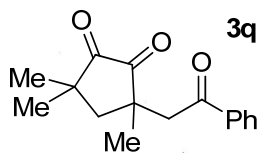
DVP-III-93-13C
File: xp
Pulse Sequence: s2pu1



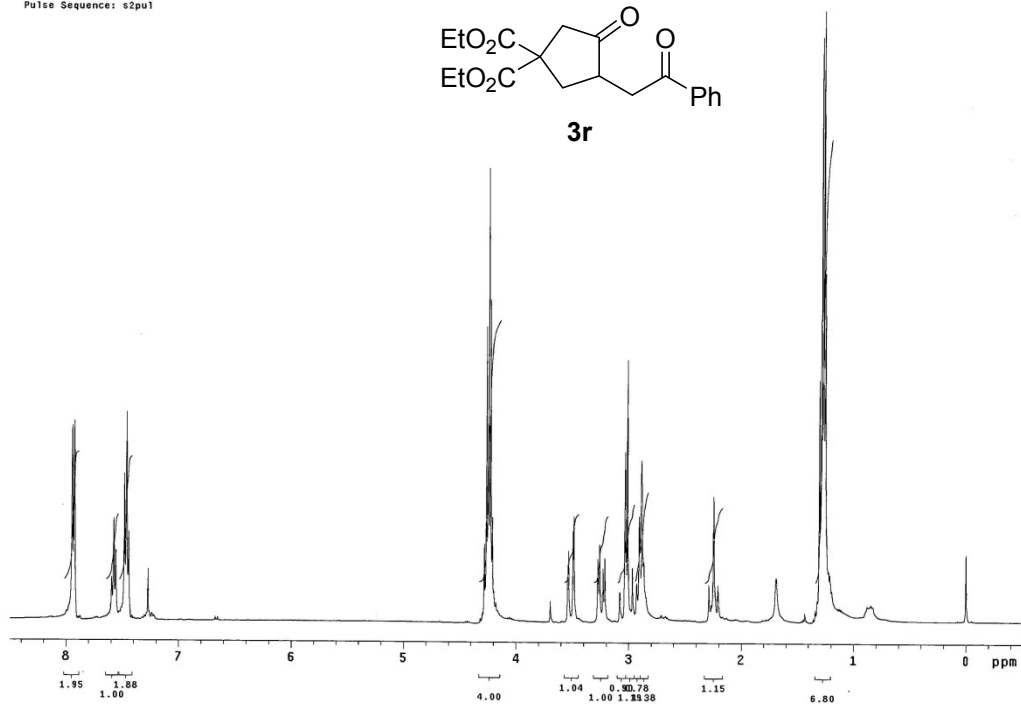
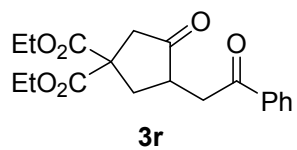
DVP-III-49
File: xp
Pulse Sequence: s2pul



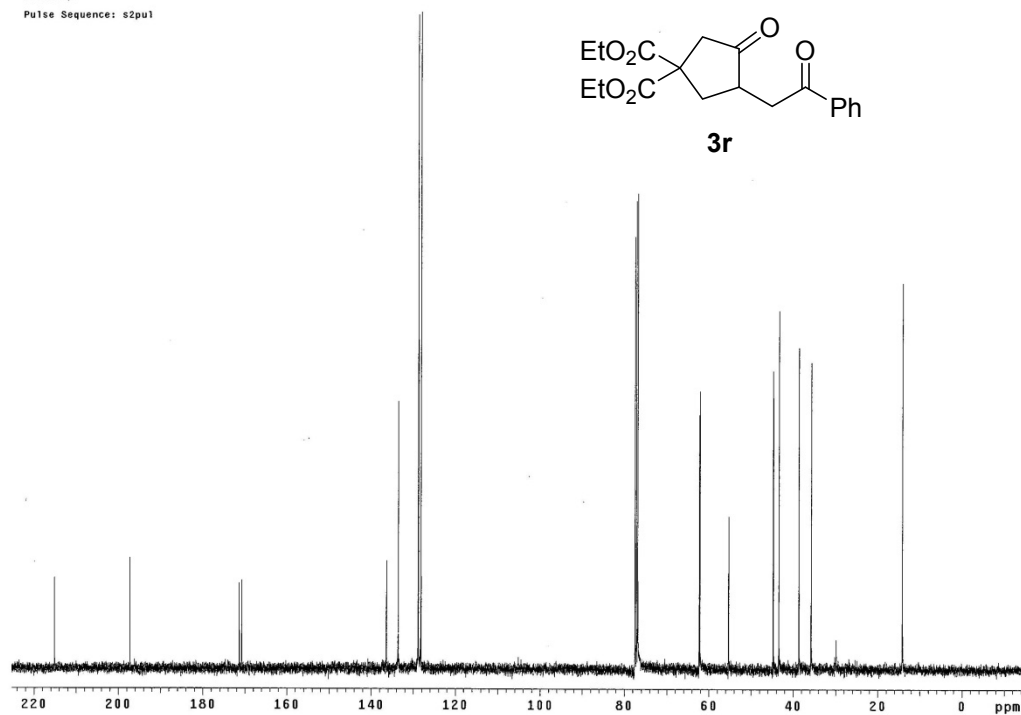
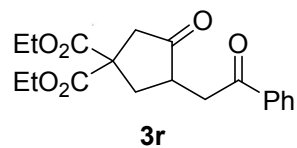
DVP-III-49-13C
File: xp
Pulse Sequence: s2pul



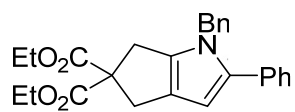
DVP-IV-30p
 File: home/vnmr3/vnmr3/data/14.07/DVP/DVP-IV-30p.fid
 Pulse Sequence: s2pu1



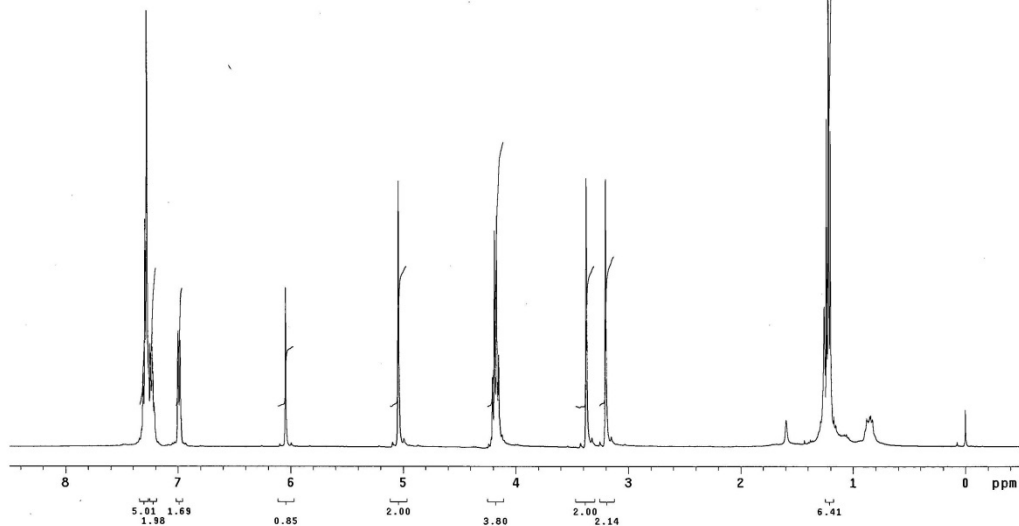
DVP-IV-30p-13C
 File: xp
 Pulse Sequence: s2pu1



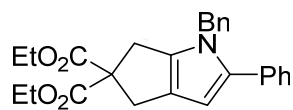
DVP-IV-39
 File: xp
 Pulse Sequence: s2pu1



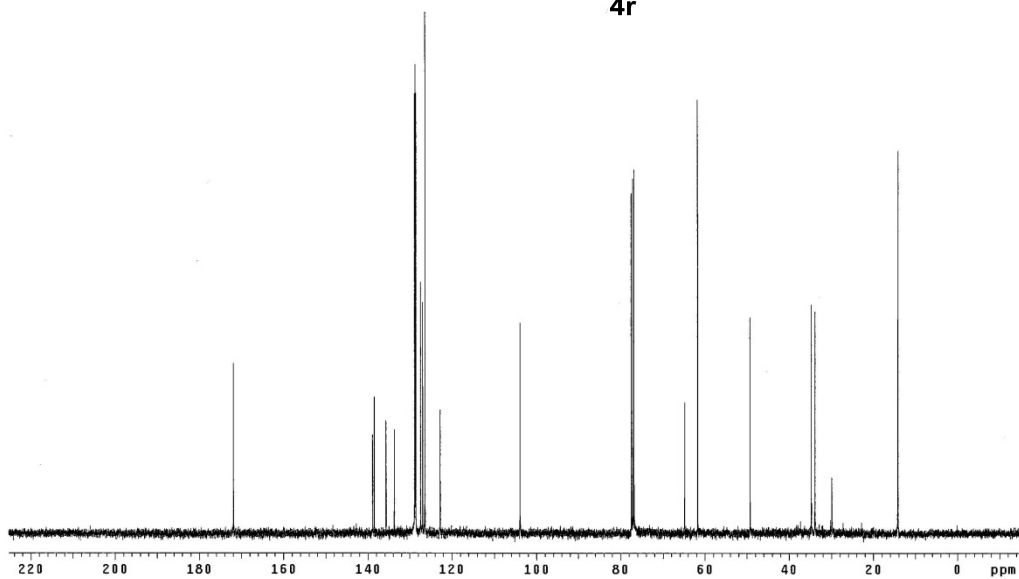
4r



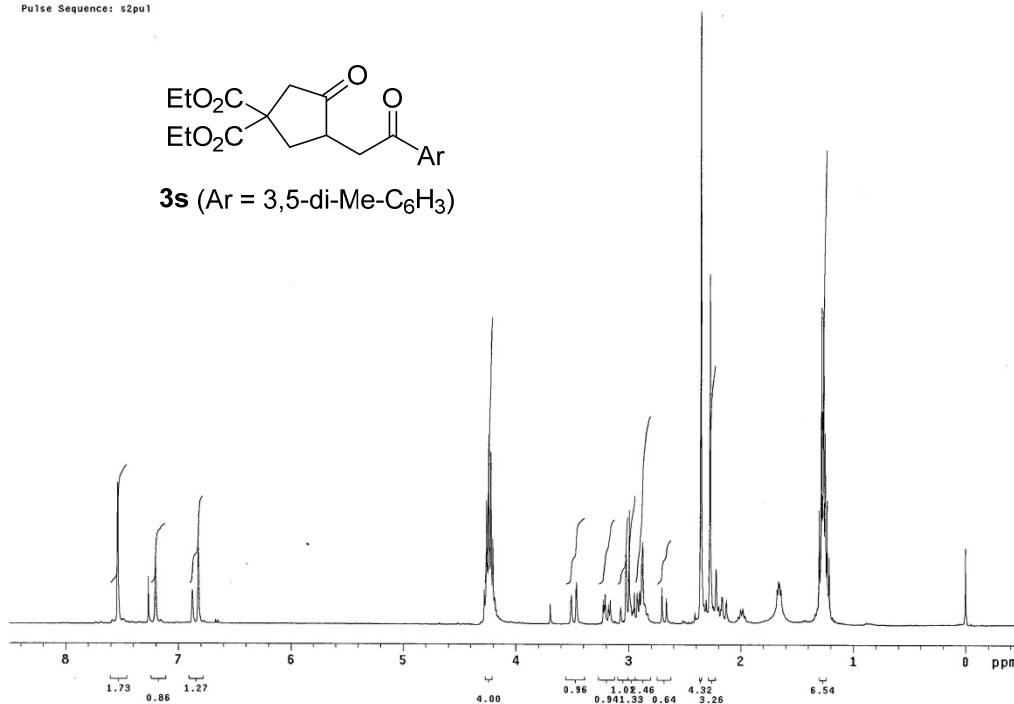
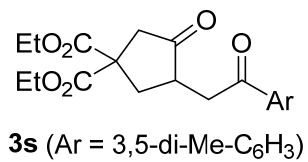
DVP-IV-39-13C
 File: xp
 Pulse Sequence: s2pu1



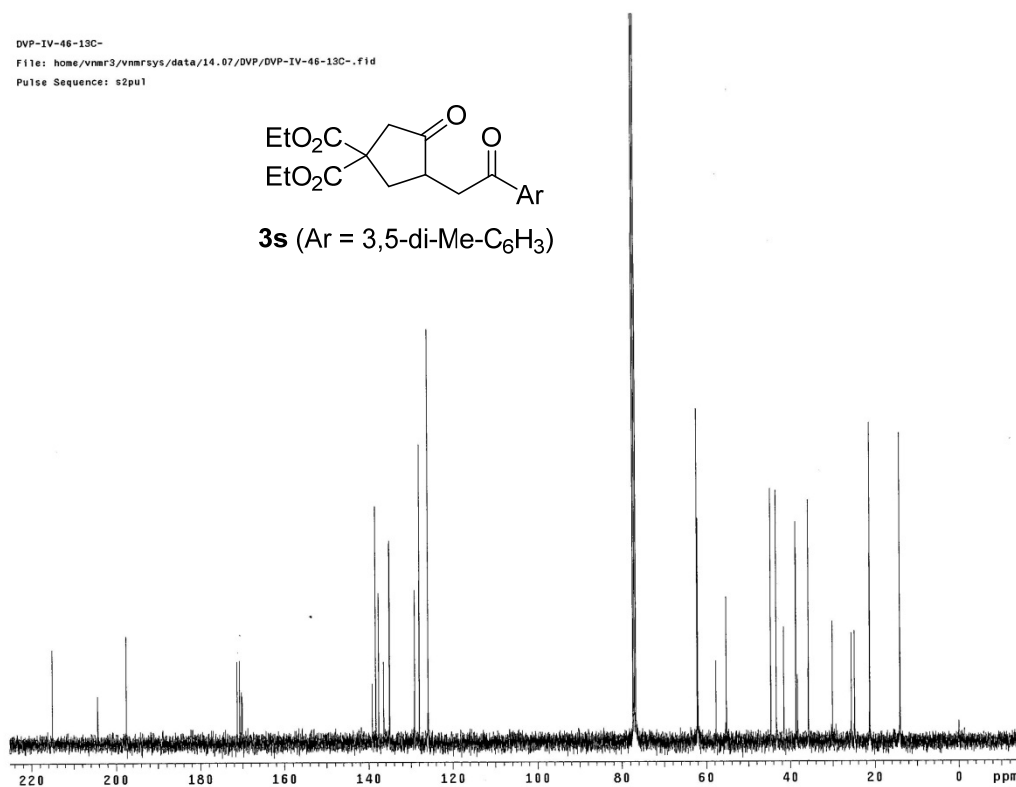
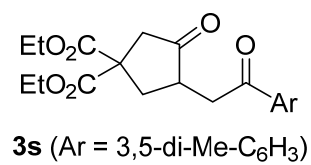
4r



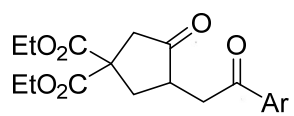
DVP-IV-46
 File: home/vnmr3/vnmr3sys/data/14.07/DVP/DVP-IV-46.fid
 Pulse Sequence: s2pu1



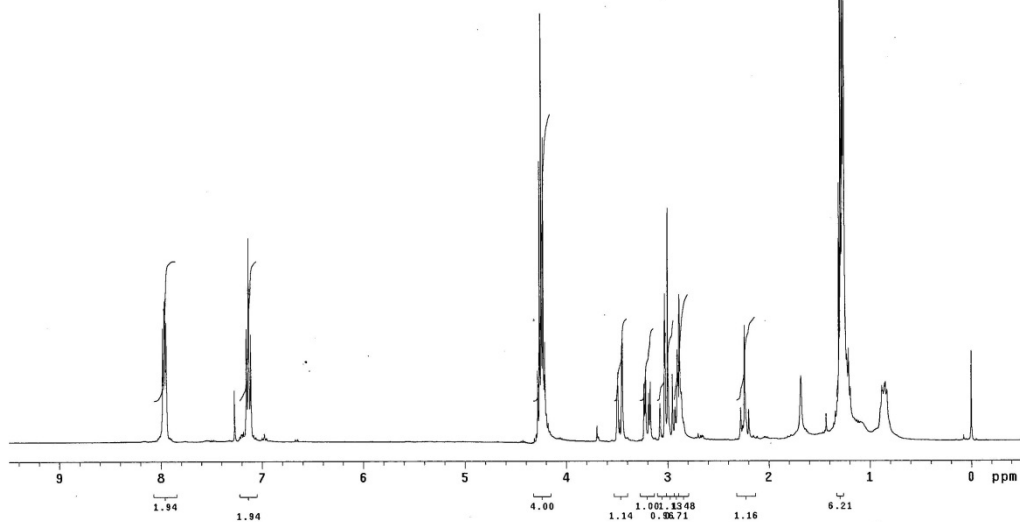
DVP-IV-46-13C-
 File: home/vnmr3/vnmr3sys/data/14.07/DVP/DVP-IV-46-13C-.fid
 Pulse Sequence: s2pu1



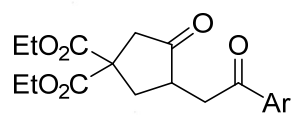
DVP-IV-47
File: xp
Pulse Sequence: s2pu1



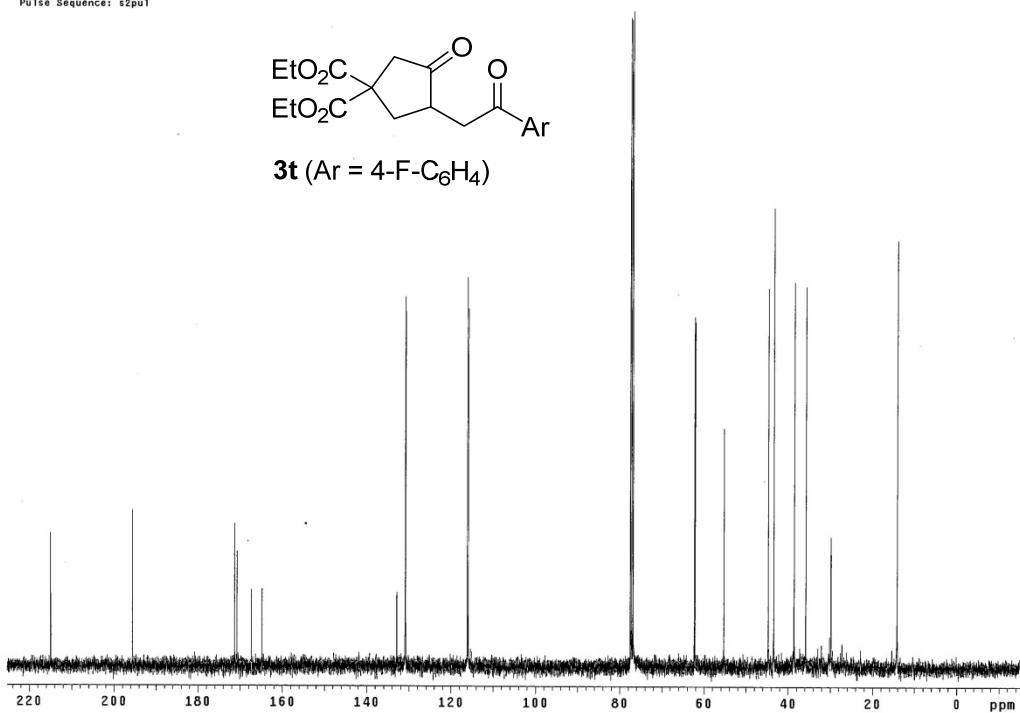
3t (Ar = 4-F-C₆H₄)

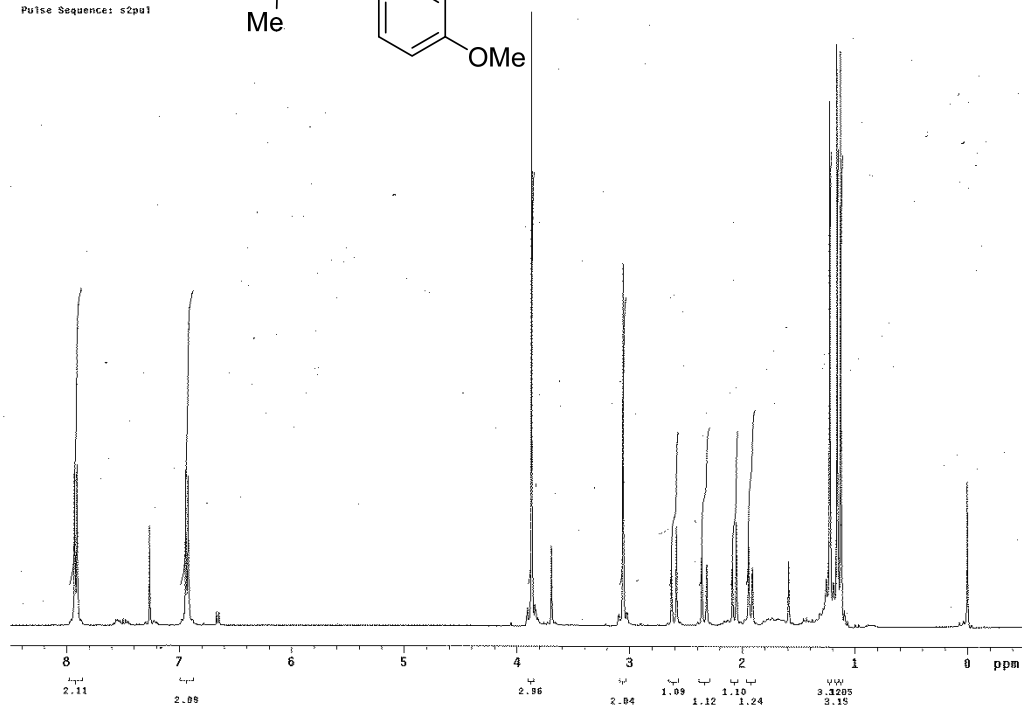
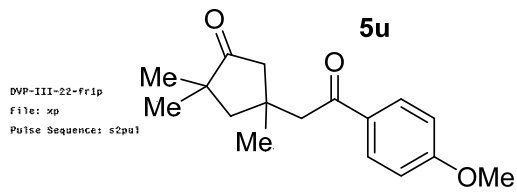


DVP-IV-47P-13C
File: xp
Pulse Sequence: s2pu1



3t (Ar = 4-F-C₆H₄)





DVP-III-22fr1-p-13C
 File: xp
 Pulse Sequence: s2pu1

