

## Supplementary Information

### The Stereoselective Synthesis of $\alpha$ -Amino Aldols Starting from Terminal Alkynes

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#### Table of Contents:

S2	General Methods and Materials		
S3	Typical procedure for the synthesis of 1-sulfonyl-1,2,3-triazole		
S4	Spectroscopic Data ( <b>3i</b> , <b>3j</b> , <b>3l</b> )		
S4	Typical procedure for the one-pot reaction of terminal alkynes with silanol [Table1, left column]		
S4	Typical procedure for the synthesis of silyl enol ethers [Table2]		
S5	Spectroscopic Data ( <b>5a</b> , <b>5b</b> , <b>5c</b> )		
S5	Spectroscopic Data ( <b>5d</b> , <b>5e</b> , <b>5g</b> , <b>5h</b> , <b>5i</b> )		
S6	Spectroscopic Data ( <b>5j</b> , <b>5k</b> , <b>5l</b> , <b>5m</b> , <b>5f</b> )		
S7	Spectroscopic Data ( <b>5n</b> , <b>5o</b> , <b>5p</b> , <b>5q</b> , <b>5r</b> )		
S8	Typical procedure for the Mukaiyama-aldol reaction of silyl enol ether with aldehydes [Table 1, right column]		
S8	Spectroscopic Data ( <b>7aa</b> , <b>7bb</b> , <b>7cb</b> , <b>7da</b> )		
S9	Spectroscopic Data ( <b>7eb</b> , <b>7fc</b> , <b>7ad</b> , <b>7ae</b> )		
S10	Determination of Stereochemistries		
S11	Typical procedure for the addition reaction of PhMgBr onto the aldol products <b>7</b>		
S11	Spectroscopic Data ( <b>8fc</b> , <b>8aa</b> , <b>8ae</b> )		
S12	Typical procedure for the deprotection of <i>t</i> -butyldimethylsilyl group		
S12	Spectroscopic Data ( <b>9fc</b> , <b>9aa</b> , <b>9ae</b> )		
S13	Typical procedure for the acetonide protection of 1,3-diol <b>9</b>		
S13	Spectroscopic Data ( <b>10fc</b> , <b>10aa</b> , <b>10ae</b> )		
S14	Experiments for the synthesis of the silyl enol ether from $\alpha$ -tosylamino acetophenone		
S15–22	Detail of the Single-Crystal X-Ray Analysis ( <b>7fc</b> )		
S23–24	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>3i</b>	S65–66	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>7aa</b>
S25–26	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>3j</b>	S67–68	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>7bb</b>
S27–28	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>3l</b>	S69–70	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>7cb</b>
S29–30	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5a</b>	S71–72	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>7da</b>
S31–32	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5b</b>	S73–74	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>7eb</b>
S33–34	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5c</b>	S75–76	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>7fc</b>
S35–36	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5d</b>	S77–78	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>7ad</b>
S37–38	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5e</b>	S79–80	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>7ae</b>
S39–40	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5g</b>	S81–82	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>8aa</b>
S41–42	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5h</b>	S83–84	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>9aa</b>
S43–44	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5i</b>	S85–86	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>10aa</b>
S45–46	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5j</b>	S87–88	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>8fc</b>
S47–48	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5k</b>	S89–90	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>9fc</b>
S49–50	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5l</b>	S91–92	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>10fc</b>
S51–52	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5m</b>	S93–94	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>8ae</b>
S53–54	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5f</b>	S95–96	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>9ae</b>
S55–56	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5n</b>	S97–98	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>10ae</b>
S57–58	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5o</b>		
S59–60	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5p</b>		
S61–62	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5q</b>		
S63–64	$^1\text{H}$ and $^{13}\text{C}$ NMR Spectra of <b>5r</b>		

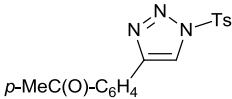
**General Methods:** All reactions were carried out under a nitrogen atmosphere unless otherwise noted. The microwave irradiation was carried out with a Biotage® Initiator 2.5 microwave synthesizer. IR measurements were performed on a FTIR SHIMADZU DR-8000 spectrometer fitted with a Pike Technologies MIRacle Single Reflection ATR adapter. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Varian Mercury-vx400 (<sup>1</sup>H at 400.44 MHz and <sup>13</sup>C at 100.69 MHz) and a JEOL JNM-ECA600 (<sup>1</sup>H at 600.17 MHz and <sup>13</sup>C at 150.92 MHz) spectrometer. NMR data were obtained in CDCl<sub>3</sub> unless otherwise noted. Proton chemical shifts were referenced to the residual proton signal of the solvent at 7.26 ppm (CHCl<sub>3</sub>) and 7.16 ppm (C<sub>6</sub>D<sub>6</sub>). Carbon chemical shifts were referenced to the carbon signal of the solvent at 77.0 ppm (CDCl<sub>3</sub>) and 128.0 ppm (C<sub>6</sub>D<sub>6</sub>). High-resolution mass spectra were recorded on a Thermo Scientific Exactive (ESI) spectrometer or a JEOL JMS-T100CS (NSI) spectrometer. Preparative thin-layer chromatography was performed on silica gel plates with PF254 indicator (Merck). Flash column chromatography was performed with silica gel 60N (Kanto) and diol-silica gel DIOL MB 100–40/75 (Fuji Silysia Chemical Ltd.). Gel permeation chromatography (GPC) was carried out with a Japan Analytical Industry LC-9210 NEXT.

**Materials:** Rh<sub>2</sub>(OCOC<sub>7</sub>H<sub>15</sub>)<sub>4</sub> (Aldrich) was used as received from the commercial sources. Rh<sub>2</sub>(OCO1-Ad)<sub>4</sub> was prepared according to the literature procedure.<sup>1</sup> Tetrahydrofuran and dichloromethane (Kanto, dehydrate) was used as received from commercial sources. Chloroform (Wako, super dehydrated, amylene as stabilizer) was distilled from phosphorus oxide(V). *tert*-Butyldimethylsilanol (Aldrich) was distilled from CaO. Terminal alkynes **1**, [phenylethyne (**1a**, Aldrich), (4-tolyl)ethyne (**1b**, TCI), (4-methoxyphenyl)ethyne (**1c**, Wako), (4-trifluoromethylphenyl)ethyne (**1d**, Aldrich), (3-thienyl)ethyne (**1e**, Aldrich), (*o*-tolyl)ethyne (**1j**, Aldrich), 1-pentyne (**1k**, Wako), and 4-methylpent-1-yne (**1l**, Aldrich)] were distilled at reduced pressure prior to use. (4-ethoxycarbonylphenyl)ethyne (**1h**) and (4-Acetylphenyl)ethyne (**1i**)<sup>2</sup> was prepared by Sonogashira coupling reaction between the corresponding iodo-compounds and 1-trimethylsilylethyne. Aldehydes **6**, [*p*-chlorobenzaldehyde (**6a**, Nacalai), benzaldehyde (**6b**, Wako), *p*-bromobenzaldehyde (**6c**, Aldrich), *p*-nitrobenzaldehyde (**6d**, Aldrich), and enanthaldehyde (**6e**, Nacalai)] were distilled at reduced pressure prior to use. Titanium(IV) chloride (Wako) was used as receiver from commercial sources. 1,4-Disubstituted 1-sulfonyl-1,2,3-triazoles **3a–r** were prepared according to the literature procedure.<sup>3</sup> The analytical data of compounds **3a**,<sup>3</sup> **3g**,<sup>4</sup> **3h**,<sup>5</sup> **3k**,<sup>6</sup> **3m**,<sup>7</sup> **3n**,<sup>3</sup> **3o**,<sup>3</sup> **3p**,<sup>3</sup> **3q**,<sup>8</sup> and **3r**<sup>9</sup> have been already reported.

**Typical procedure for the synthesis of 1-sulfonyl-1,2,3-triazole:** To a solution of CuTC (95.2 mg, 0.5 mmol, 10 mol %) in toluene (10 mL) was added a solution of **1j** (600.2 mg, 5.2 mmol, 1.0 equiv) in toluene (5 mL) and tosyl azide (982.1 mg, 5.0 mmol, 1.0 equiv) in toluene (5 mL). After 15 h, the reaction mixture was diluted with saturated NH<sub>4</sub>Cl aq (20 mL) and extracted with ethyl acetate (3 x 15 mL). The combined organic phase was washed with brine (10 mL), dried over MgSO<sub>4</sub>, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (hexane/ethyl acetate = 5:1), then reprecipitation (hexane/dichloromethane = 20:1) at –20 °C to give **3j** (1.01 g, 3.3 mmol, 65%) as a white powder.

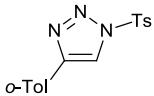
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- 1 T. D. Nelson, Z. J. Song, A. S. Thompson, M. Zhao, A. DeMarco, R. A. Reamer, M. F. Huntington, E. J. J. Grabowski, P. J. Reider, *Tetrahedron Lett.* **2000**, *41*, 1877.
  - 2 X. Shao, X. Wang, T. Yang, L. Lu, Q. Shen, *Angew. Chem. Int. Ed.* **2013**, *52*, 3457.
  - 3 J. Raushel, V. V. Fokin, *Org. Lett.* **2010**, *12*, 4952.
  - 4 T. Miura, M. Yamauchi, M. Murakami, *Chem. Commun.* **2009**, 1470.
  - 5 B. Chattopadhyay, V. Gevorgyan, *Org. Lett.* **2011**, *13*, 3746.
  - 6 T. Miura, K. Hiraga, T. Biyajima, T. Nakamuro, M. Murakami, *Org. Lett.* **2013**, *15*, 3298.
  - 7 T. Miura, T. Biyajima, T. Fujii, M. Murakami, *J. Am. Chem. Soc.* **2012**, *134*, 194.
  - 8 E. J. Yoo, M. Ahlquist, S. H. Kim, I. Bae, V. V. Fokin, K. B. Sharpless, S. Chang, *Angew. Chem. Int. Ed.* **2007**, *46*, 1730.
  - 9 T. Miura, Y. Funakoshi, T. Tanaka, M. Murakami, *Org. Lett.* **2014**, *16*, 2760.

**3i:**



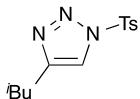
IR (ATR): 1680, 1394, 1190, 1169, 989 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = 2.45 (s, 3H), 2.62 (s, 3H), 7.40 (d, *J* = 8.4 Hz, 2H), 7.93 (d, *J* = 8.8 Hz, 2H), 8.02 (d, *J* = 8.4 Hz, 2H), 8.03 (d, *J* = 8.4 Hz, 2H), 8.42 (s, 1H); <sup>13</sup>C NMR: δ = 21.8, 26.7, 119.9, 126.0, 128.7, 129.0, 130.5, 132.7, 133.2, 137.1, 146.1, 147.6, 197.4; HRMS (ESI<sup>+</sup>): Calcd for C<sub>17</sub>H<sub>16</sub>N<sub>3</sub>O<sub>3</sub>S, M+H<sup>+</sup> 342.0907. Found m/z 342.0902.

**3j:**



IR (ATR): 1591, 1389, 1348, 1196, 1173, 986 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = 2.448 (s, 3H), 2.453 (s, 3H), 7.23–7.33 (m, 3H), 7.40 (d, *J* = 8.0 Hz, 2H), 7.73–7.77 (m, 1H), 8.04 (d, *J* = 8.4 Hz, 2H), 8.21 (s, 1H); <sup>13</sup>C NMR: δ = 21.3, 21.8, 120.8, 126.2, 128.1, 128.7, 128.9, 129.0, 130.4, 131.0, 133.0, 135.8, 146.6, 147.3; HRMS (ESI<sup>+</sup>): Calcd for C<sub>16</sub>H<sub>16</sub>N<sub>3</sub>O<sub>2</sub>S, M+H<sup>+</sup> 314.0958. Found m/z 314.0950.

**3l:**



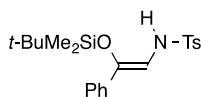
IR (ATR): 2957, 1595, 1387, 1192, 1175 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = 0.89 (d, *J* = 6.8 Hz, 6H), 1.88–2.01 (m, 1H), 2.43 (s, 3H), 2.56 (d, *J* = 7.2 Hz, 2H), 7.36 (d, *J* = 8.0 Hz, 2H), 7.84 (s, 1H), 7.96 (d, *J* = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = 21.8, 22.1, 28.3, 34.3, 120.8, 128.5, 130.3, 133.2, 147.0, 147.1; HRMS (ESI<sup>+</sup>): Calcd for C<sub>13</sub>H<sub>18</sub>N<sub>3</sub>O<sub>2</sub>S, M+H<sup>+</sup> 280.1114. Found m/z 280.1110.

**Typical procedure for the one-pot reaction of terminal alkynes with silanol [Table 1, left column]:**

Phenylethyne **1a** (41.7 mg, 0.4 mmol), TsN<sub>3</sub> **2a** (79.4 mg, 0.4 mmol), CuTC (7.6 mg, 40 µmol), and CHCl<sub>3</sub> (2 mL) were added to an oven-dried 2–5 mL Biotage® microwave vial equipped with a stir bar. The vial was sealed with a cap containing an inner Teflon film. The reaction mixture was stirred at room temperature for 6 h. Then, the vial was taken inside the glove box. To the resulting mixtures were added silanol **4** (84.3 mg, 0.6 mmol), Rh<sub>2</sub>(OCOC<sub>7</sub>H<sub>15</sub>)<sub>4</sub> (3.2 mg, 4 µmol), 4 A MS (40 mg), and CHCl<sub>3</sub> (3 mL). The reaction mixture was heated to 100 °C for 15 min under microwave irradiation. After the reaction mixture was cooled and passed through a cotton stopper with CHCl<sub>3</sub>, the filtrate was concentrated under reduced pressure. The residue was purified by gel permeation chromatography (GPC) to give the product **5a** as a white solid (121.6 mg, 0.3 mmol, 76%).

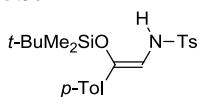
**Typical procedure for the synthesis of silyl enol ethers [Table 2]:** In a nitrogen-filled glove-box, Rh<sub>2</sub>(OCOC<sub>7</sub>H<sub>15</sub>)<sub>4</sub> (0.8 mg, 1 µmol), **3a** (61.0 mg, 0.20 mmol), 4 A MS (20 mg), silanol **4** (40.1 mg, 0.30 mmol), and CHCl<sub>3</sub> (4 mL) were added to an oven-dried 2-5 mL Biotage® microwave vial. The vial was sealed with a cap containing an inner Teflon film. The reaction mixture was heated to 100 °C for 15 min under microwave irradiation. After the reaction mixture was cooled and passed through a cotton stopper with CHCl<sub>3</sub>, the filtrate was concentrated under reduced pressure. The residue was purified by silica gel column chromatography (CHCl<sub>3</sub>) to give the product **5a** as a white solid (78.8 mg, 0.195 mmol, 96%).

**5a:**



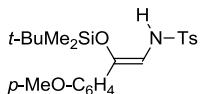
Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3371, 2930, 1335, 1153 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.15 (s, 6H), 0.93 (s, 9H), 2.41 (s, 3H), 6.12 (d, *J* = 10.4 Hz, 1H), 6.21 (d, *J* = 10.8 Hz, 1H), 7.22–7.32 (m, 7H), 7.76 (d, *J* = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = -4.3, 18.2, 21.5, 25.7, 107.5, 125.3, 126.8, 127.8, 128.2, 129.8, 136.4, 137.0, 138.4, 143.7; HRMS (ESI<sup>+</sup>): Calcd for C<sub>21</sub>H<sub>30</sub>NO<sub>3</sub>SSi, M+H<sup>+</sup> 404.1710. Found m/z 404.1700.

**5b:**



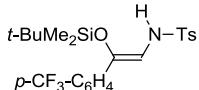
Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3375, 2953, 1335, 1155 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.14 (s, 6H), 0.94 (s, 9H), 2.32 (s, 3H), 2.41 (s, 3H), 6.08 (d, *J* = 10.8 Hz, 1H), 6.21 (d, *J* = 10.8 Hz, 1H), 7.09 (d, *J* = 8.4 Hz, 2H), 7.20 (d, *J* = 8.0 Hz, 2H), 7.29 (d, *J* = 8.4 Hz, 2H), 7.76 (d, *J* = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = -4.3, 18.2, 21.1, 21.5, 25.7, 106.9, 125.3, 126.8, 128.8, 129.7, 133.5, 137.0, 137.7, 138.6, 143.6; HRMS (ESI<sup>+</sup>): Calcd for C<sub>22</sub>H<sub>32</sub>NO<sub>3</sub>SSi, M+H<sup>+</sup> 418.1867. Found m/z 418.1854.

**5c:**



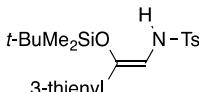
Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3285, 2930, 1601, 1248, 1155 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.16 (s, 6H), 0.92 (s, 9H), 2.41 (s, 3H), 3.79 (s, 3H), 5.99 (d, *J* = 10.4 Hz, 1H), 6.14 (d, *J* = 10.8 Hz, 1H), 6.81 (d, *J* = 8.0 Hz, 2H), 7.22 (d, *J* = 8.4 Hz, 2H), 7.29 (d, *J* = 8.1 Hz, 2H), 7.75 (d, *J* = 8.2 Hz, 2H); <sup>13</sup>C NMR: δ = -4.3, 18.1, 21.4, 25.7, 55.2, 106.2, 113.5, 126.7, 126.8, 128.8, 129.7, 136.9, 138.6, 143.6, 159.4; HRMS (ESI<sup>+</sup>): Calcd for C<sub>22</sub>H<sub>32</sub>NO<sub>4</sub>SSi, M+H<sup>+</sup> 434.1816. Found m/z 434.1821.

**5d** (reaction time, 20 h):



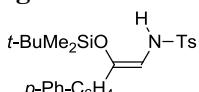
Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3306, 2930, 1614, 1321, 1157 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.12 (s, 6H), 0.95 (s, 9H), 2.41 (s, 3H), 6.26 (d, *J* = 10.4 Hz, 1H), 6.30 (d, *J* = 10.8 Hz, 1H), 7.31 (d, *J* = 8.0 Hz, 2H), 7.42 (d, *J* = 8.0 Hz, 2H), 7.54 (d, *J* = 8.0 Hz, 2H), 7.76 (d, *J* = 8.0 Hz, 2H); <sup>13</sup>C NMR: δ = -4.2, 18.2, 21.5, 25.7, 109.5, 124.0 (q, *J* = 270.1 Hz), 125.0, 125.3 (q, *J* = 3.7 Hz), 126.8, 129.5 (q, *J* = 32.2 Hz), 129.9, 136.4, 136.9, 140.0, 144.0; HRMS (ESI): Calcd for C<sub>22</sub>H<sub>27</sub>F<sub>3</sub>NO<sub>4</sub>SSi, M-H<sup>-</sup> 470.1438. Found m/z 470.1428.

5e:



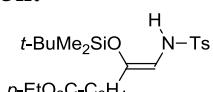
Purified by silica gel column chromatography ( $\text{CHCl}_3$ ); IR (ATR): 3292, 2928, 1663, 1331, 1153  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta = -0.09$  (s, 6H), 0.94 (s, 9H), 2.41 (s, 3H), 6.12 (d,  $J = 10.4$  Hz, 1H), 6.17 (d,  $J = 10.8$  Hz, 1H), 7.01 (dd,  $J = 5.2, 1.2$  Hz, 1H), 7.11 (dd,  $J = 2.4, 1.2$  Hz, 1H), 7.23 (dd,  $J = 5.2, 2.8$  Hz, 1H), 7.30 (d,  $J = 8.0$  Hz, 2H), 7.75 (d,  $J = 8.4$  Hz, 2H);  $^{13}\text{C}$  NMR:  $\delta = -4.3, 18.2, 21.5, 25.7, 107.1, 120.5, 125.0, 125.8, 126.7, 129.7, 134.5, 136.9, 137.8, 143.7$ ; HRMS (APCI $^+$ ): Calcd for  $\text{C}_{19}\text{H}_{28}\text{NO}_3\text{S}_2\text{Si}$ ,  $\text{M}+\text{H}^+$  410.1274. Found m/z 410.1268.

5g:



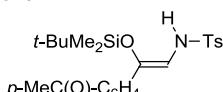
Purified by silica gel column chromatography ( $\text{CHCl}_3$ ); IR (ATR): 3381, 2930, 1659, 1335, 1155  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta = -0.07$  (s, 6H), 0.98 (s, 9H), 2.42 (s, 3H), 6.23 (d,  $J = 10.8$  Hz, 1H), 6.30 (d,  $J = 10.8$  Hz, 1H), 7.32 (d,  $J = 8.0$  Hz, 2H), 7.34–7.38 (m, 1H), 7.41 (d,  $J = 8.4$  Hz, 2H), 7.45 (t,  $J = 7.6$  Hz, 2H), 7.55 (d,  $J = 8.4$  Hz, 2H), 7.60 (d,  $J = 7.2$  Hz, 2H), 7.80 (d,  $J = 8.4$  Hz, 2H);  $^{13}\text{C}$  NMR:  $\delta = -4.2$ , 18.2, 21.5, 25.7, 107.6, 125.6, 126.7, 126.79, 126.80, 127.4, 128.7, 129.8, 135.2, 136.9, 137.9, 140.3, 140.5, 143.7; HRMS (ESI $^+$ ): Calcd for  $\text{C}_{27}\text{H}_{34}\text{NO}_3\text{SSi}$ ,  $\text{M}+\text{H}^+$  480.2023. Found m/z 480.2031.

5h:



Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3277, 2932, 1713, 1339, 1273, 1151 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.13 (s, 6H), 0.94 (s, 9H), 1.37 (t, *J* = 7.2 Hz, 3H), 2.40 (s, 3H), 4.35 (q, *J* = 7.2 Hz, 2H), 6.28 (d, *J* = 10.8 Hz, 1H), 6.32 (d, *J* = 10.8 Hz, 1H), 7.30 (d, *J* = 8.4 Hz, 2H), 7.37 (d, *J* = 8.4 Hz, 2H), 7.75 (d, *J* = 8.4 Hz, 2H), 7.95 (d, *J* = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = -4.2, 14.3, 18.1, 21.5, 25.7, 60.9, 109.4, 124.6, 126.7, 129.4, 129.5, 129.8, 136.8, 136.9, 140.7, 143.9, 166.1; HRMS (ESI<sup>+</sup>): Calcd for C<sub>24</sub>H<sub>34</sub>NO<sub>5</sub>SSi, M+H<sup>+</sup> 476.1921. Found m/z 476.1930.

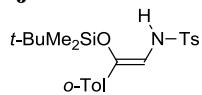
5i:



Purified by diol-silica gel column chromatography ( $\text{CHCl}_3$ ); IR (ATR): 3290, 2930, 1680, 1601, 1339, 1256, 1157  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta = -0.13$  (s, 6H), 0.95 (s, 9H), 2.41 (s, 3H), 2.58 (s, 3H), 6.32 (s, 2H), 7.31 (d,  $J = 8.0$  Hz, 2H), 7.39 (t,  $J = 8.8$  Hz, 2H), 7.75 (d,  $J = 8.4$  Hz, 2H), 7.87 (d,  $J = 8.8$  Hz, 2H);  $^{13}\text{C}$  NMR:  $\delta = -4.2$ , 18.2, 21.5, 25.7, 26.5, 109.7, 124.7, 126.7, 128.4, 129.9, 136.0, 136.6, 136.8, 141.0, 144.0, 197.3; HRMS (ESI $^+$ ):

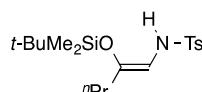
Calcd for C<sub>23</sub>H<sub>32</sub>NO<sub>4</sub>SSi, M+H<sup>+</sup> 446.1816. Found m/z 446.1807.

**5j:**



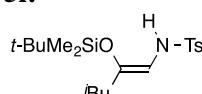
Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3344, 2928, 1661, 1595, 1333, 1163 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.35 (s, 6H), 0.85 (s, 9H), 2.15 (s, 3H), 2.43 (s, 3H), 5.74 (d, J = 10.8 Hz, 1H), 6.21 (d, J = 10.8 Hz, 1H), 7.08–7.14 (m, 3H), 7.18–7.23 (m, 1H), 7.32 (d, J = 8.0 Hz, 2H), 7.77 (d, J = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = -5.0, 18.0, 19.7, 21.5, 25.5, 108.5, 125.5, 126.9, 128.6, 129.2, 129.6, 130.2, 135.5, 136.7, 137.1, 139.7, 143.6; HRMS (ESI<sup>+</sup>): Calcd for C<sub>22</sub>H<sub>32</sub>NO<sub>3</sub>SSi, M+H<sup>+</sup> 418.1867. Found m/z 418.1861.

**5k:**



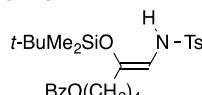
Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3364, 2930, 1684, 1339, 1165 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.04 (s, 6H), 0.81 (t, J = 7.6 Hz, 3H), 0.85 (s, 9H), 1.38 (sext, J = 7.2 Hz, 2H), 1.91 (t, J = 7.6 Hz, 2H), 2.40 (s, 3H), 5.48 (d, J = 10.4 Hz, 1H), 5.89 (d, J = 10.4 Hz, 1H), 7.27 (d, J = 8.0 Hz, 2H), 7.70 (d, J = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = -4.3, 13.3, 18.1, 19.8, 21.5, 25.6, 35.5, 104.7, 126.8, 129.5, 136.8, 140.5, 143.4; HRMS (ESI<sup>+</sup>): Calcd for C<sub>18</sub>H<sub>32</sub>NO<sub>3</sub>SSi, M+H<sup>+</sup> 370.1867. Found m/z 370.1860.

**5l:**



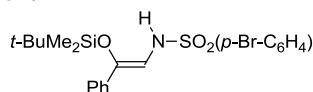
Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3356, 2955, 1342, 1165, 781 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.05 (s, 6H), 0.76 (d, J = 6.4 Hz, 6H), 0.85 (s, 9H), 1.61–1.73 (m, 1H), 1.76 (d, J = 6.8 Hz, 2H), 2.41 (s, 3H), 5.48 (d, J = 10.4 Hz, 1H), 5.89 (d, J = 10.4 Hz, 1H), 7.27 (d, J = 8.0 Hz, 2H), 7.70 (d, J = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = -4.3, 18.1, 21.5, 22.0, 25.4, 25.6, 42.9, 105.6, 126.9, 129.5, 136.6, 139.7, 143.5; HRMS (ESI<sup>+</sup>): Calcd for C<sub>19</sub>H<sub>34</sub>NO<sub>3</sub>SSi, M+H<sup>+</sup> 384.2023. Found m/z 384.2024.

**5m:**

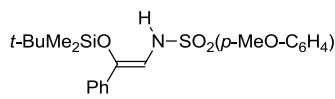


Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3285, 2955, 1717, 1273, 1165 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.04 (s, 6H), 0.85 (s, 9H), 1.49–1.57 (m, 2H), 1.60–1.70 (m, 2H), 2.01 (t, J = 7.2 Hz, 2H), 2.37 (s, 3H), 4.26 (t, J = 6.4 Hz, 2H), 5.53 (d, J = 10.4 Hz, 1H), 5.91 (d, J = 10.8 Hz, 1H), 7.26 (d, J = 8.0 Hz, 2H), 7.44 (t, J = 8.0 Hz, 2H), 7.56 (t, J = 7.6 Hz, 1H), 7.70 (d, J = 8.4 Hz, 2H), 8.03 (d, J = 8.0 Hz, 2H); <sup>13</sup>C NMR: δ = -4.3, 18.1, 21.5, 23.0, 25.6, 27.8, 33.1, 64.5, 105.1, 126.8, 128.3, 129.5, 129.6, 130.2, 132.9, 136.6, 139.9, 143.5, 166.6; HRMS (ESI<sup>+</sup>): Calcd for C<sub>26</sub>H<sub>38</sub>NO<sub>5</sub>SSi, M+H<sup>+</sup> 504.2234. Found m/z 504.2234.

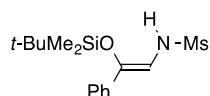
**5f:**



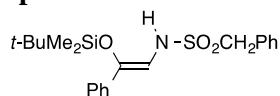
Purified by silica gel column chromatography (CHCl<sub>3</sub>); IR (ATR): 3341, 2928, 1333, 1169, 1067 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.13 (s, 6H), 0.94 (s, 9H), 6.10 (d, J = 10.8 Hz, 1H), 6.27 (d, J = 10.4 Hz, 1H), 7.26–7.33 (m, 5H), 7.65 (d, J = 8.8 Hz, 2H), 7.74 (d, J = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = -4.3, 18.2, 25.7, 106.9, 125.4, 127.9, 128.1, 128.2, 128.3, 132.4, 136.1, 138.8, 139.2; HRMS (ESI<sup>+</sup>): Calcd for C<sub>20</sub>H<sub>27</sub>BrNO<sub>3</sub>SSi, M+H<sup>+</sup> 468.0659. Found m/z 468.0670.

**5n:**

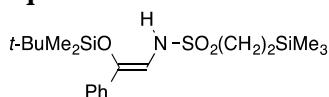
Purified by silica gel column chromatography ( $\text{CHCl}_3$ ); IR (ATR): 3302, 2928, 1661, 1595, 1335, 1258, 1151  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta = -0.14$  (s, 6H), 0.94 (s, 9H), 3.84 (s, 3H), 6.12 (d,  $J = 10.4$  Hz, 1H), 6.22 (d,  $J = 10.8$  Hz, 1H), 6.97 (d,  $J = 8.8$  Hz, 2H), 7.23–7.32 (m, 5H), 7.81 (d,  $J = 8.8$  Hz, 2H);  $^{13}\text{C}$  NMR:  $\delta = -4.3$ , 18.2, 25.7, 55.6, 107.6, 114.3, 125.3, 127.8, 128.2, 128.9, 131.5, 136.3, 138.3, 163.0; HRMS (ESI $^+$ ): Calcd for  $\text{C}_{21}\text{H}_{30}\text{NO}_4\text{SSi}$ ,  $\text{M}+\text{H}^+$  420.1659. Found m/z 420.1657.

**5o:**

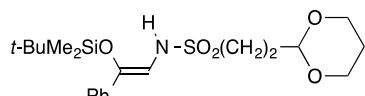
Purified by silica gel column chromatography ( $\text{CHCl}_3$ ); IR (ATR): 3310, 2930, 1659, 1331, 1148  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta = 0.00$  (s, 6H), 1.01 (s, 9H), 3.04 (s, 3H), 6.13 (s, 2H), 7.25–7.40 (m, 5H);  $^{13}\text{C}$  NMR:  $\delta = -4.0$ , 18.2, 25.8, 40.6, 107.4, 125.2, 128.0, 128.3, 136.2, 138.3; HRMS (ESI $^+$ ): Calcd for  $\text{C}_{15}\text{H}_{26}\text{NO}_3\text{SSi}$ ,  $\text{M}+\text{H}^+$  328.1397. Found m/z 328.1395.

**5p:**

Purified by silica gel column chromatography ( $\text{CHCl}_3$ ); IR (ATR): 3346, 3287, 2930, 1661, 1323, 1146  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta = -0.06$  (s, 6H), 0.88 (s, 9H), 4.31 (s, 2H), 6.00 (d,  $J = 11.2$  Hz, 1H), 6.10 (d,  $J = 10.8$  Hz, 1H), 7.27–7.43 (m, 10H);  $^{13}\text{C}$  NMR:  $\delta = -4.1$ , 18.1, 25.6, 58.4, 107.9, 125.1, 127.9, 128.3, 128.6, 128.8, 129.0, 130.7, 136.3, 137.7; HRMS (ESI $^+$ ): Calcd for  $\text{C}_{21}\text{H}_{30}\text{NO}_3\text{SSi}$ ,  $\text{M}+\text{H}^+$  404.1710. Found m/z 404.1702.

**5q:**

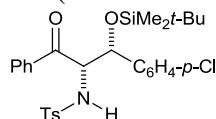
Purified by silica gel column chromatography ( $\text{CHCl}_3$ ); IR (ATR): 3285, 2951, 1661, 1327, 1252, 1142  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta = 0.00$  (s, 6H), 0.04 (s, 9H), 1.02 (s, 9H), 1.04–1.09 (m, 2H), 2.98–3.04 (m, 2H), 6.10 (s, 2H), 7.24–7.38 (m, 5H);  $^{13}\text{C}$  NMR:  $\delta = -4.0$ , -2.1, 10.6, 18.2, 25.7, 49.0, 108.0, 125.2, 127.9, 128.3, 136.3, 138.0; HRMS (ESI $^+$ ): Calcd for  $\text{C}_{19}\text{H}_{36}\text{NO}_3\text{SSi}_2$ ,  $\text{M}+\text{H}^+$  414.1949. Found m/z 414.1954.

**5r:**

Purified by silica gel column chromatography ( $\text{CHCl}_3$ ); IR (ATR): 3298, 2930, 2856, 1664, 1331, 1142  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta = 0.00$  (s, 6H), 1.00 (s, 9H), 1.28–1.37 (m, 1H), 1.95–2.15 (m, 3H), 3.23–3.30 (m, 2H), 3.74 (dt,  $J = 12.2$ , 2.4 Hz, 2H), 4.03–4.10 (m, 2H), 4.69 (t,  $J = 4.4$  Hz, 1H), 6.10 (d,  $J = 10.4$  Hz, 1H), 6.14 (d,  $J = 10.4$  Hz, 1H), 7.25–7.39 (m, 5H);  $^{13}\text{C}$  NMR:  $\delta = -4.0$ , 18.2, 25.4, 25.8, 29.2, 47.9, 66.7, 99.1, 107.7, 125.1, 127.7, 128.2, 136.3, 137.5; HRMS (ESI $^+$ ): Calcd for  $\text{C}_{20}\text{H}_{34}\text{NO}_5\text{SSi}$ ,  $\text{M}+\text{H}^+$  428.1921. Found m/z 428.1917.

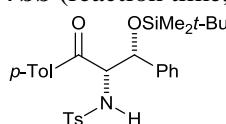
**Typical procedure for the Mukaiyama-aldo reaction of silyl enol ether with aldehydes (Table 1, right column):** Aldehyde **6a** (39.9 mg, 0.28 mmol), silyl enol ether **5a** (99.2 mg, 0.25 mmol), and dry CH<sub>2</sub>Cl<sub>2</sub> (8 mL) were added to an oven-dried two-necked round bottom flask equipped with a stir bar. The reaction mixture was cooled to -78 °C. Then, TiCl<sub>4</sub> (31 µL, 0.28 mmol) was added dropwise via syringe. The resulting mixture was stirred at -78 °C for 13 h, and quenched with water (1 mL). The aqueous layer was repeatedly extracted with dichloromethane. The combined organic layers were washed with brine and dried over MgSO<sub>4</sub>. The solvent was removed under reduced pressure and the residue was purified by preparative thin-layer chromatography (CHCl<sub>3</sub>) to give the product **7aa** as a white solid (119.7 mg, 0.22 mmol, 88%).

**7aa** (reaction time, 13 h):



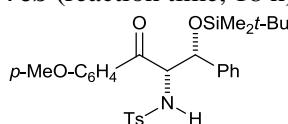
Purified by preparative thin-layer chromatography (CHCl<sub>3</sub>); IR (ATR): 3290, 2928, 1692, 1340, 1165, 1084 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.35 (s, 3H), -0.29 (s, 3H), 0.75 (s, 9H), 2.30 (s, 3H), 4.94 (dd, J = 9.6, 2.8 Hz, 1H), 5.04 (d, J = 2.8 Hz, 1H), 5.69 (d, J = 9.6Hz, 1H), 7.05 (d, J = 8.4 Hz, 2H), 7.13-7.21 (m, 4H), 7.44–7.51 (m, 4H), 7.61 (t, J = 7.2 Hz, 1H), 7.81 (d, J = 8.4 Hz, 2H); <sup>13</sup>C NMR: δ = -5.6, -4.9, 17.9, 21.4, 25.5, 63.4, 74.8, 126.8, 127.9, 128.2, 128.6, 128.8, 129.4, 133.8, 133.9, 134.7, 136.7, 138.3, 143.3, 195.2; HRMS (ESI<sup>+</sup>): Calcd for C<sub>28</sub>H<sub>35</sub>ClNO<sub>4</sub>SSi, M+H<sup>+</sup> 544.1739. Found m/z 544.1736.

**7bb** (reaction time, 18 h):



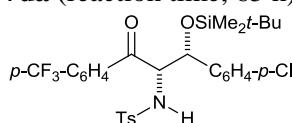
Purified by preparative thin-layer chromatography (hexane/ethyl acetate 4:1 then CHCl<sub>3</sub>); IR (ATR): 3285, 2926, 1692, 1333, 1161, 1096 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.33 (s, 3H), -0.26 (s, 3H), 0.77 (s, 9H), 2.26 (s, 3H), 2.43 (s, 3H), 4.95 (dd, J = 9.8, 2.2 Hz, 1H), 5.04 (s, 1H), 5.67 (d, J = 9.2Hz, 1H), 7.03 (d, J = 8.0 Hz, 2H), 7.22-7.30 (m, 7H), 7.47 (d, J = 8.0 Hz, 2H), 7.70 (d, J = 8.0 Hz, 2H); <sup>13</sup>C NMR(150 MHz): δ = -5.5, -4.9, 18.0, 21.3, 21.7, 25.6, 63.5, 75.5, 126.6, 126.8, 127.9, 128.0, 128.7, 129.316, 129.324, 132.3, 136.9, 139.8, 143.0, 144.7, 194.9; HRMS (ESI<sup>+</sup>): Calcd for C<sub>29</sub>H<sub>38</sub>NO<sub>4</sub>SSi, M+H<sup>+</sup> 524.2285. Found m/z 524.2272.

**7cb** (reaction time, 18 h):



Purified by preparative thin-layer chromatography (hexane/ethyl acetate 4:1 then CHCl<sub>3</sub>); IR (ATR): 3236, 2928, 1674, 1593, 1165, 1067 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.31 (s, 3H), -0.24 (s, 3H), 0.78 (s, 9H), 2.26 (s, 3H), 3.88 (s, 3H), 4.92 (dd, J = 9.6, 2.8 Hz, 1H), 5.05 (d, J = 2.8 Hz, 1H), 5.71 (d, J = 9.6Hz, 1H), 6.92 (d, J = 8.8 Hz, 2H), 7.02 (d, J = 8.0 Hz, 2H), 7.22-7.30 (m, 5H), 7.47 (d, J = 8.4 Hz, 2H), 7.79 (d, J = 8.8 Hz, 2H); <sup>13</sup>C NMR(150 MHz): δ = -5.5, -4.9, 18.0, 21.3, 25.6, 55.5, 63.2, 75.8, 113.8, 126.6, 126.8, 127.7, 127.92, 127.94, 129.3, 130.9, 136.9, 139.9, 143.0, 163.9, 193.7; HRMS (ESI<sup>+</sup>): Calcd for C<sub>29</sub>H<sub>38</sub>NO<sub>5</sub>SSi, M+H<sup>+</sup> 540.2234. Found m/z 540.2222.

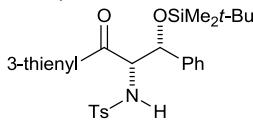
**7da** (reaction time, 65 h):



Purified by preparative thin-layer chromatography (hexane/ethyl acetate 4:1) then GPC(CHCl<sub>3</sub>); IR (ATR): 3302, 2953, 1707, 1315, 1153, 1067 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.33 (s, 3H), -0.25 (s, 3H), 0.74 (s, 9H), 2.31 (s, 3H), 4.94 (dd, J = 9.4, 3.4 Hz, 1H), 5.01 (d, J = 3.2 Hz, 1H), 5.62 (d, J = 9.6Hz, 1H), 7.07 (d, J = 8.0 Hz, 2H), 7.11 (d, J = 8.4 Hz, 2H), 7.18 (d, J = 8.4 Hz, 2H), 7.46 (d, J = 8.4 Hz, 2H), 7.73 (d, J = 8.0 Hz, 2H), 7.90 (d, J = 8.4 Hz, 2H); <sup>13</sup>C NMR(150 MHz): δ = -5.4, -4.9, 17.9, 21.4, 25.5, 63.6, 74.8, 123.3 (q, J = 270.7 Hz),

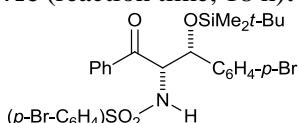
125.7 (q,  $J = 3.6$  Hz) 126.8, 127.9, 128.3, 128.9, 129.5, 134.1, 134.9 (q,  $J = 32.3$  Hz), 136.5, 137.7, 137.8, 143.6; HRMS (ESI $^+$ ): Calcd for C<sub>29</sub>H<sub>34</sub>NO<sub>4</sub>SSiF<sub>3</sub>Cl, M+H $^+$  612.1613. Found m/z 612.1599.

**7eb** (reaction time, 18 h):



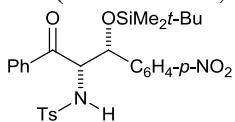
Purified by preparative thin-layer chromatography (hexane/ethyl acetate 3:1 then CHCl<sub>3</sub>); IR (ATR): 3265, 2926, 1682, 1339, 1165, 1094 cm $^{-1}$ ; <sup>1</sup>H NMR:  $\delta = -0.29$  (s, 3H),  $-0.20$  (s, 3H),  $0.78$  (s, 9H),  $2.28$  (s, 3H),  $4.71$  (dd,  $J = 9.2, 3.2$  Hz, 1H),  $5.08$  (d,  $J = 3.2$  Hz, 1H),  $5.63$  (d,  $J = 9.6$  Hz, 1H),  $7.03$  (d,  $J = 8.0$  Hz, 2H),  $7.24$  (s, 5H),  $7.30$  (dd,  $J = 4.8, 3.2$  Hz, 1H),  $7.40$  (dd,  $J = 5.2, 1.2$  Hz, 1H),  $7.45$  (d,  $J = 8.4$  Hz, 2H),  $8.02$  (dd,  $J = 2.8, 1.2$  Hz, 1H); <sup>13</sup>C NMR:  $\delta = -5.5, -4.9, 18.0, 21.4, 25.6, 65.2, 75.7, 126.5, 126.6, 126.9, 127.0, 128.01, 128.04, 129.3, 133.3, 136.7, 139.6, 139.7, 143.1, 189.6$ ; HRMS (ESI $^+$ ): Calcd for C<sub>26</sub>H<sub>34</sub>NO<sub>4</sub>S<sub>2</sub>Si, M+H $^+$  516.1693. Found m/z 516.1683.

**7fc** (reaction time, 18 h):



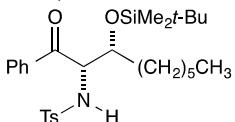
Purified by preparative thin-layer chromatography (dichloromethane/hexane 20:1 then 7:1); IR (ATR): 3292, 2926, 1690, 1342, 1167, 1084 cm $^{-1}$ ; <sup>1</sup>H NMR:  $\delta = -0.35$  (s, 3H),  $-0.31$  (s, 3H),  $0.76$  (s, 9H),  $4.95$  (dd,  $J = 9.6, 2.8$  Hz, 1H),  $5.05$  (d,  $J = 2.4$  Hz, 1H),  $5.75$  (d,  $J = 9.6$  Hz, 1H),  $7.12$  (d,  $J = 8.4$  Hz, 2H),  $7.36$  (d,  $J = 8.4$  Hz, 2H),  $7.41$  (s, 4H),  $7.50$  (t,  $J = 7.8$  Hz, 2H),  $7.63$  (t,  $J = 7.2$  Hz, 1H),  $7.84$  (d,  $J = 8.2$  Hz, 2H); <sup>13</sup>C NMR:  $\delta = -5.6, -4.9, 17.9, 25.5, 63.7, 74.7, 122.2, 127.5, 128.2, 128.3, 128.6, 128.9, 131.2, 132.0, 134.1, 134.3, 138.8, 138.9, 194.8$ ; HRMS (ESI $^+$ ): Calcd for C<sub>27</sub>H<sub>35</sub>Br<sub>2</sub>N<sub>2</sub>O<sub>4</sub>SSi, M+NH $^+$  669.0448. Found m/z 669.0435.

**7ad** (reaction time, 13 h):



Purified by preparative thin-layer chromatography (CHCl<sub>3</sub>); IR (ATR): 3275, 2928, 1697, 1528, 1340, 1165 cm $^{-1}$ ; <sup>1</sup>H NMR:  $\delta = -0.36$  (s, 3H),  $-0.31$  (s, 3H),  $0.76$  (s, 9H),  $2.25$  (s, 3H),  $4.99$  (dd,  $J = 9.4, 2.4$  Hz, 1H),  $5.18$  (d,  $J = 2.8$  Hz, 1H),  $5.77$  (d,  $J = 9.2$  Hz, 1H),  $7.03$  (d,  $J = 8.8$  Hz, 2H),  $7.42-7.52$  (m, 6H),  $7.63$  (t,  $J = 7.6$  Hz, 1H),  $7.84$  (d,  $J = 7.6$  Hz, 2H),  $8.08$  (d,  $J = 8.8$  Hz, 2H); <sup>13</sup>C NMR:  $\delta = -5.7, -4.9, 17.9, 21.3, 25.5, 63.2, 74.7, 123.2, 126.7, 127.5, 128.6, 128.9, 129.4, 134.17, 134.18, 136.5, 143.6, 147.2, 147.5, 194.5$ ; HRMS (ESI $^+$ ): Calcd for C<sub>28</sub>H<sub>35</sub>N<sub>2</sub>O<sub>6</sub>SSi, M+H $^+$  555.1980. Found m/z 555.1974.

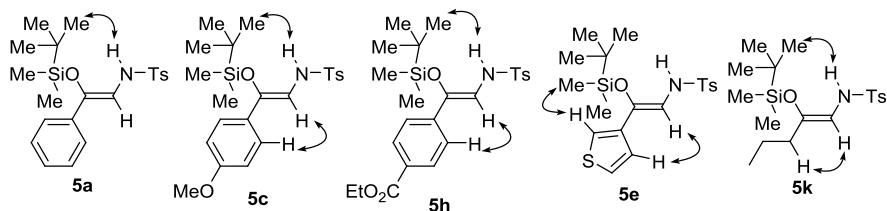
**7ae** (reaction time, 19 h):



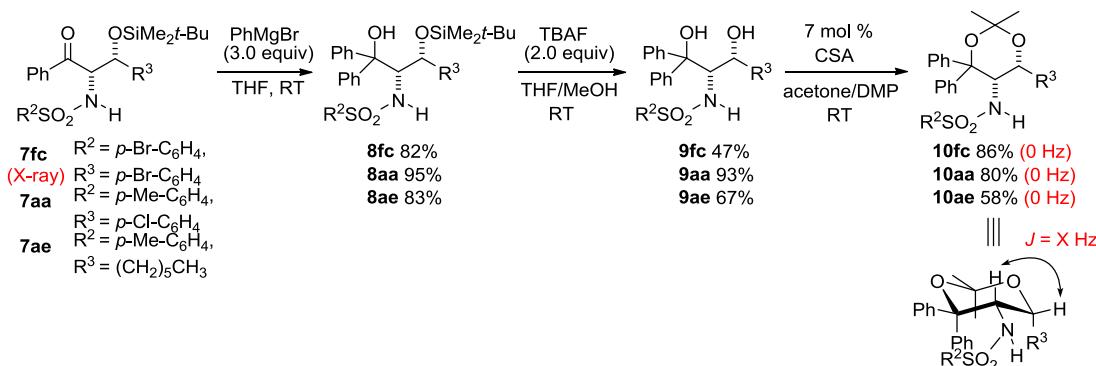
Purified by preparative thin-layer chromatography (CHCl<sub>3</sub>); IR (ATR): 3287, 2926, 1697, 1333, 1163, 1080 cm $^{-1}$ ; <sup>1</sup>H NMR:  $\delta = -0.36$  (s, 3H),  $-0.16$  (s, 3H),  $0.76$  (s, 9H),  $0.91$  (t,  $J = 3.2$  Hz, 3H),  $1.24-1.48$  (m, 9H),  $1.85-1.96$  (m, 1H),  $2.28$  (s, 3H),  $3.91-3.96$  (m, 1H),  $4.91$  (d,  $J = 9.8$  Hz, 1H),  $5.63$  (d,  $J = 10.0$  Hz, 1H),  $7.12$  (d,  $J = 8.4$  Hz, 2H),  $7.43$  (t,  $J = 7.6$  Hz, 2H),  $7.57$  (t,  $J = 7.6$  Hz, 1H),  $7.67$  (d,  $J = 8.4$  Hz, 2H),  $7.70$  (d,  $J = 8.0$  Hz, 2H); <sup>13</sup>C NMR:  $\delta = -5.1, -4.5, 14.0, 17.8, 21.4, 22.6, 25.3, 25.6, 29.3, 31.7, 34.5, 60.3, 73.4, 127.0, 128.2, 128.7, 129.4, 133.6, 134.2, 137.0, 143.2, 195.4$ ; HRMS (ESI $^+$ ): Calcd for C<sub>28</sub>H<sub>44</sub>NO<sub>4</sub>SSi, M+H $^+$  518.2755. Found m/z 518.2753.

### Determination of Stereochemistries

The (*Z*)-stereochemistries of the silyl enol ethers (**5a**, **5c**, **5h**, **5e**, and **5k**) were determined by NOE experiments. Curved arrows shown below indicate the observed NOE. Other silyl enol ethers **5** were determined by analogy.

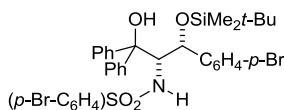


The relative stereochemistry of the aldol product **7fc** was unambiguously determined as *syn* by a single-crystal X-ray analysis. Other aldol products **7** were determined by NMR studies of its derivative.



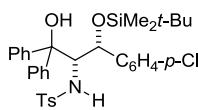
**Typical procedure for the addition reaction of PhMgBr onto the aldol products 7:** PhMgBr (0.3M in THF, 2.5 mL) and dry THF (3 mL) were added to an oven-dried two-necked round bottom flask equipped with a stir bar. The reaction mixture was cooled to 0 °C. Then, a solution of **7fc** (143.2 mg, 0.22 mmol) in THF (5 mL) was added dropwise via syringe. The resulting mixture was stirred at room temperature for 3 h, and quenched with sat. NH<sub>4</sub>Cl (8 mL). The aqueous layer was repeatedly extracted with ethyl acetate. The combined organic layers were washed with brine and dried over MgSO<sub>4</sub>. The solvent was removed under reduced pressure and the residue was purified by preparative thin-layer chromatography (hexane/ethyl acetate 5:1) to give the product **8fc** (130.8 mg, 0.18 mmol, 82%).

**8fc:**



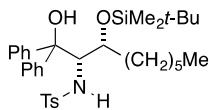
Purified by preparative thin-layer chromatography (hexane/ethyl acetate 5:1); IR (ATR): 3443, 2932, 2856, 1576, 1487, 1339, 1157, 1065 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz): δ = -0.50 (s, 3H), -0.46 (s, 3H), 0.91 (s, 9H), 4.64 (dd, J = 9.3, 1.5 Hz, 1H), 4.70 (s, 1H), 5.31 (s, 1H), 5.47 (d, J = 9.6 Hz, 1H), 6.97–7.03 (m, 5H), 7.14 (d, J = 8.4 Hz, 2H), 7.23–7.30 (m, 3H), 7.37 (d, J = 6.6 Hz, 2H), 7.38–7.43 (m, 4H), 7.70 (d, J = 7.8 Hz, 2H); <sup>13</sup>C NMR (150 MHz): δ = -6.0, -3.8, 17.8, 25.8, 65.3, 75.2, 81.2, 122.3, 125.2, 125.6, 126.1, 126.8, 127.3, 127.4, 128.0, 128.70, 128.72, 131.1, 131.7, 139.4, 140.9, 143.4, 145.1; HRMS (APCI): Calcd for C<sub>33</sub>H<sub>36</sub>Br<sub>2</sub>NO<sub>4</sub>SSi, M-H<sup>-</sup> 728.0507. Found m/z 728.0522.

**8aa:**



Purified by silica gel column chromatography (hexane/ethyl acetate 3:1); IR (ATR): 3447, 2928, 1597, 1491, 1333, 1155 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.51 (s, 3H), -0.44 (s, 3H), 0.93 (s, 9H), 2.35 (s, 3H), 4.70 (dd, J = 9.0, 1.4 Hz, 1H), 4.79 (s, 1H), 5.33 (s, 1H), 5.46 (d, J = 9.2 Hz, 1H), 6.95 (d, J = 8.0 Hz, 2H), 6.98–7.18 (m, 9H), 7.29 (t, J = 7.2 Hz, 1H), 7.43 (t, J = 8.0 Hz, 2H), 7.51 (d, J = 8.2 Hz, 2H), 7.76 (d, J = 7.6 Hz, 2H); <sup>13</sup>C NMR: δ = -6.1, -4.0, 17.7, 21.2, 25.8, 65.1, 75.0, 81.3, 125.4, 125.65, 125.71, 126.6, 127.2, 127.7, 127.9, 128.3, 128.6, 129.0, 133.6, 138.9, 139.0, 141.7, 143.3, 145.2; HRMS (ESI<sup>+</sup>): Calcd for C<sub>34</sub>H<sub>40</sub>ClNO<sub>4</sub>SSiNa, M+Na<sup>+</sup> 644.2028. Found m/z 644.2022.

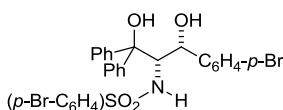
**8ae:**



Purified by preparative thin-layer chromatography (hexane/ethyl acetate 7:1); IR (ATR): 3437, 3306, 2924, 1431, 1321, 1157 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = -0.08 (s, 3H), 0.03 (s, 3H), 0.89–1.00 (m, 12H), 1.18–1.42 (m, 8H), 1.58–1.70 (m, 1H), 1.88–2.00 (m, 1H), 2.33 (s, 3H), 4.11 (dd, J = 11.2, 4.0 Hz, 1H), 4.75 (s, 1H), 4.83 (d, J = 9.2 Hz, 1H), 5.10 (d, J = 8.8 Hz, 1H), 6.99 (d, J = 8.0 Hz, 2H), 7.02–7.14 (m, 3H), 7.18–7.26 (m, 3H), 7.33 (t, J = 7.2 Hz, 2H), 7.55 (d, J = 8.4 Hz, 2H), 7.59 (d, J = 8.0 Hz, 2H); <sup>13</sup>C NMR: δ = -4.5, -3.3, 14.0, 17.9, 21.2, 22.6, 25.1, 25.9, 29.1, 31.7, 34.4, 59.7, 74.4, 81.3, 125.2, 125.3, 126.2, 126.3, 126.8, 127.9, 128.3, 129.0, 139.0, 142.0, 144.5, 145.9; HRMS (ESI<sup>+</sup>): Calcd for C<sub>34</sub>H<sub>48</sub>NO<sub>4</sub>SSi, M-H<sup>-</sup> 594.3079. Found m/z 594.3081.

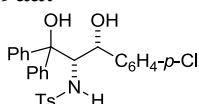
**Typical procedure for the deprotection of *t*-butyldimethylsilyl group:** Alcohol **8fc** (142.0 mg, 0.194 mmol), dry THF (6 mL), and dry MeOH (2 mL) were added to an oven-dried two-necked round bottom flask equipped with a stir bar. Then, TBAF solution (0.1 M in THF, 0.7 mL) was added dropwise via syringe. The resulting mixture was stirred at room temperature for 9 h, and diluted with water (8 mL). The aqueous layer was repeatedly extracted with ethyl acetate. The combined organic layers were washed with brine and dried over MgSO<sub>4</sub>. The solvent was removed under reduced pressure and the residue was purified by preparative thin-layer chromatography (chloroform/ethyl acetate/acetic acid 10:1:0.1) to give the product **9fc** (55.7 mg, 0.09 mmol, 47%).

**9fc:**



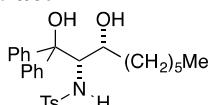
Purified by preparative thin-layer chromatography (chloroform/ethyl acetate/acetic acid 10:1:0.1); IR (ATR): 3360, 3024, 1576, 1448, 1327, 1153, 1067 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = 3.18 (s, 1H), 4.746 (d, *J* = 8.8 Hz, 1H), 4.753 (s, 1H), 5.02 (s, 1H), 5.68 (d, *J* = 9.2 Hz, 1H), 6.94 (dt, *J* = 8.4, 2.8 Hz, 2H), 7.02 (d, *J* = 8.4 Hz, 2H), 7.06–7.15 (m, 3H), 7.21–7.32 (m, 5H), 7.42 (t, *J* = 7.8 Hz, 2H), 7.47–7.54 (m, 2H), 7.67 (d, *J* = 7.8 Hz, 2H); <sup>13</sup>C NMR: δ = 64.8, 72.8, 82.1, 121.9, 125.1, 125.5, 126.2, 127.14, 127.15, 127.3, 127.4, 128.1, 128.9, 131.3, 131.6, 139.4, 140.8, 143.1, 144.8; HRMS (APCI): Calcd for C<sub>27</sub>H<sub>22</sub>Br<sub>2</sub>NO<sub>4</sub>S, M-H<sup>-</sup> 613.9642. Found m/z 613.9649.

**9aa:**



Purified by GPC; IR (ATR): 3366, 3026, 1597, 1491, 1448, 1323, 1151 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = 2.36 (s, 3H), 3.44 (d, *J* = 2.4 Hz, 1H), 4.72 (d, *J* = 8.8 Hz, 1H), 4.86 (s, 1H), 4.98 (d, *J* = 1.6 Hz, 1H), 5.56 (d, *J* = 9.2 Hz, 1H), 6.89 (d, *J* = 8.0 Hz, 2H), 6.93 (d, *J* = 8.4 Hz, 2H), 6.98–7.17 (m, 7H), 7.30 (t, *J* = 7.2 Hz, 1H), 7.43 (t, *J* = 8.0 Hz, 2H), 7.55 (d, *J* = 7.2 Hz, 2H), 7.69 (d, *J* = 7.2 Hz, 2H); <sup>13</sup>C NMR: δ = 21.3, 64.6, 72.6, 82.2, 125.2, 125.6, 125.7, 126.9, 127.0, 127.3, 127.9, 128.1, 128.8, 128.9, 133.3, 138.8, 139.0, 142.0, 143.1, 144.9; HRMS (ESI): Calcd for C<sub>28</sub>H<sub>25</sub>ClNO<sub>4</sub>S, M-H<sup>-</sup> 506.1198. Found m/z 506.1200.

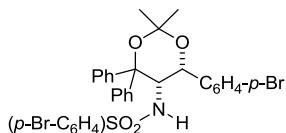
**9ae:**



Purified by preparative thin-layer chromatography (chloroform/ethyl acetate/acetic acid 10:1:0.1 then 10:1:0.05); IR (ATR): 3456, 3354, 2926, 1599, 1450, 1333, 1155 cm<sup>-1</sup>; <sup>1</sup>H NMR: δ = 0.88 (t, *J* = 6.8 Hz, 3H), 1.14–1.40 (m, 8H), 1.48–1.72 (m, 2H), 2.33 (s, 3H), 2.61 (d, *J* = 5.2 Hz, 1H), 3.91–3.97 (m, 1H), 4.62 (d, *J* = 8.4 Hz, 1H), 4.74 (s, 1H), 5.51 (d, *J* = 8.8 Hz, 1H), 6.92–7.10 (m, 5H), 7.20 (t, *J* = 7.6 Hz, 1H), 7.24 (d, *J* = 8.4 Hz, 2H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.43 (d, *J* = 7.6 Hz, 2H), 7.52 (d, *J* = 7.2 Hz, 2H); <sup>13</sup>C NMR: δ = 14.0, 21.3, 22.6, 25.3, 29.0, 31.6, 35.0, 61.4, 72.4, 81.8, 125.1, 125.2, 126.3, 126.5, 127.0, 128.0, 128.5, 129.2, 138.5, 142.3, 144.1, 145.4; HRMS (ESI): Calcd for C<sub>28</sub>H<sub>34</sub>NO<sub>4</sub>S, M-H<sup>-</sup> 480.2214. Found m/z 480.2219.

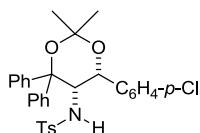
**Typical procedure for the acetonide protection of 1,3-diol 9:** 1,3-Diol **9fc** (142.0 mg, 0.194 mmol), 10-camphorsulfonic acid (CSA; 1.7 mg, 7.32  $\mu$ mol), dry acetone (0.8 mL), and dry 2,2-dimethoxypropane (DMP; 0.2 mL) were added to an oven-dried two-necked round bottom flask equipped with a stir bar. The resulting mixture was stirred at room temperature for 6 h, and quenched with Et<sub>3</sub>N (2 drops). Then, volatile materials were removed under reduced pressure. The residue was purified by preparative thin-layer chromatography (hexane/ethyl acetate 5:1) to give the product **10fc** (40.4 mg, 0.06 mmol, 86%).

**10fc:**



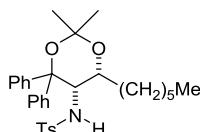
Purified by preparative thin-layer chromatography (hexane/ethyl acetate 5:1); IR (ATR): 3387, 2993, 1489, 1340, 1159, 1070  $\text{cm}^{-1}$ ; <sup>1</sup>H NMR:  $\delta$  = 1.01 (s, 3H), 1.72 (s, 3H), 4.83 (d,  $J$  = 10.0 Hz, 1H), 4.89 (d,  $J$  = 9.6 Hz, 1H), 5.48 (s, 1H), 6.92 (dt,  $J$  = 8.8, 2.4 Hz, 2H), 7.05–7.32 (m, 10H), 7.35 (dt,  $J$  = 8.4, 2.2 Hz, 2H), 7.40 (d,  $J$  = 7.2 Hz, 2H), 7.53 (d,  $J$  = 7.2 Hz, 2H); <sup>13</sup>C NMR (55 °C):  $\delta$  = 24.2, 31.2, 57.6, 70.3, 81.9, 101.5, 122.0, 125.6, 126.2, 126.8, 127.2, 127.4, 127.6, 127.7, 127.9, 128.5, 131.3, 131.7, 137.4, 141.0, 144.1, 144.8; HRMS (APCI): Calcd for C<sub>30</sub>H<sub>26</sub>Br<sub>2</sub>NO<sub>4</sub>S, M-H<sup>-</sup> 653.9955. Found m/z 653.9967.

**10aa:**



Purified by preparative thin-layer chromatography (hexane/ethyl acetate 4:1); IR (ATR): 3391, 2989, 1599, 1491, 1339, 1155  $\text{cm}^{-1}$ ; <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>):  $\delta$  = 0.80 (s, 3H), 1.50 (s, 3H), 2.00 (s, 3H), 4.90 (d,  $J$  = 9.6 Hz, 1H), 5.02 (d,  $J$  = 9.6 Hz, 1H), 5.36 (s, 1H), 6.57 (d,  $J$  = 8.0 Hz, 2H), 6.94–7.20 (m, 12H), 7.39 (d,  $J$  = 7.2 Hz, 2H), 7.52 (d,  $J$  = 7.6 Hz, 2H); <sup>13</sup>C NMR:  $\delta$  = 21.2, 24.2, 31.0, 57.5, 70.5, 82.4, 101.3, 126.3, 127.2, 127.6, 127.7, 127.9, 128.1, 128.4, 128.5, 128.6, 129.2, 133.7, 137.5, 140.4, 141.5, 144.9, 145.3; HRMS (ESI): Calcd for C<sub>31</sub>H<sub>29</sub>ClNO<sub>4</sub>S, M-H<sup>-</sup> 546.1511. Found m/z 546.1524.

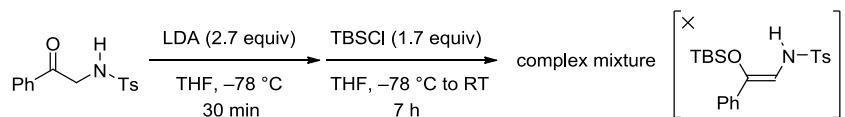
**10ae:**



Purified by preparative thin-layer chromatography (hexane/ethyl acetate 5:1); IR (ATR): 3287, 2926, 1429, 1327, 1157  $\text{cm}^{-1}$ ; <sup>1</sup>H NMR (600 MHz):  $\delta$  = 0.91 (t,  $J$  = 6.6 Hz, 3H), 0.95 (s, 3H), 1.25–1.46 (m, 8H), 1.54–1.63 (m, 5H), 2.34 (s, 3H), 4.31 (dd,  $J$  = 7.2, 6.0 Hz, 1H), 4.57 (d,  $J$  = 9.6 Hz, 1H), 4.83 (d,  $J$  = 9.0 Hz, 1H), 6.93 (tt,  $J$  = 7.2, 1.2 Hz, 1H), 6.97–7.02 (m, 4H), 7.13 (d,  $J$  = 7.2 Hz, 2H), 7.18 (dt,  $J$  = 7.8, 1.2 Hz, 2H), 7.21 (tt,  $J$  = 7.2, 1.2 Hz, 1H), 7.29 (t,  $J$  = 7.2 Hz, 2H), 7.34 (d,  $J$  = 8.4 Hz, 2H); <sup>13</sup>C NMR (150 MHz):  $\delta$  = 14.0, 21.3, 22.6, 24.2, 25.1, 29.1, 31.1, 31.7, 32.4, 55.2, 69.7, 81.8, 100.9, 125.6, 126.2, 126.6, 127.2, 127.3, 128.2, 129.1, 139.1, 142.1, 144.9, 145.8; HRMS (ESI<sup>+</sup>): Calcd for C<sub>31</sub>H<sub>43</sub>N<sub>2</sub>O<sub>4</sub>S, M+NH<sub>4</sub><sup>+</sup> 539.2938. Found m/z 539.2924.

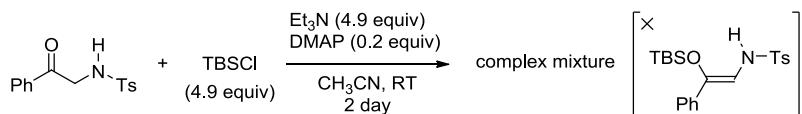
### Experiments for the synthesis of the silyl enol ether from $\alpha$ -tosylamino acetophenone

i) Kinetic conditions using LDA as base according to the literature procedure.<sup>4</sup>



A solution of  $\alpha$ -tosylamino acetophenone (62.1 mg, 0.215 mmol) in THF (1.0 mL) was added dropwise via syringe to a solution of LDA (0.39 M in THF, 1.5 mL) at -78°C. The resulting solution was further stirred for 30 min at -78°C, then a solution of *tert*-butyldimethylsilylchloride (55.2 mg, 0.366 mmol) in THF (1.0 mL) was added. The reaction mixture allowed to slowly warm to room temperature. After stirring for 7 h, the solution was filtrated through the patch of celite and removed under reduced pressure. The residue was checked by <sup>1</sup>H NMR, and a complex mixture was observed.

ii) Thermodynamic conditions using Et<sub>3</sub>N as base according to the literature procedure.<sup>5</sup>



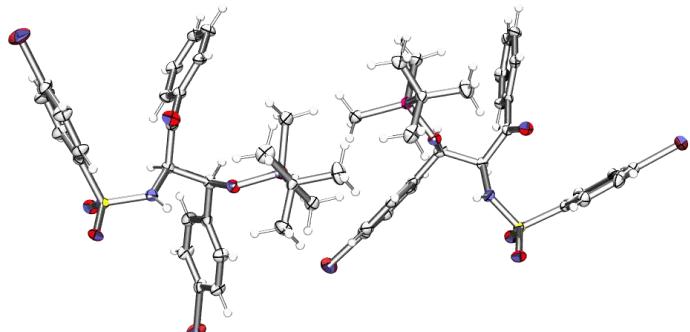
Et<sub>3</sub>N (94.5 mg, 0.93 mmol) and DMAP (4.7 mg, 0.038 mmol) were added to a solution of  $\alpha$ -tosylamino acetophenone (53.9 mg, 0.19 mmol) and *t*-butyldimethylsilylchloride (140.2 mg, 0.93 mmol) in dry acetonitrile (0.9 mL). The resulting solution was stirred at room temperature for 2 days. Then, volatile materials were removed under reduced pressure. The residue was checked by <sup>1</sup>H NMR, and a complex mixture was observed.

4 I. Fleming, I. Paterson, *Synthesis* **1979**, 736.

5 M. J. Crimmin, P. J. O' Hanlon, N. H. Rogers, F. M. Sime, G. Walker, *J. Chem. Soc. Perkin. Trans. I.* **1989**, 2059.

**Detail of the Single-Crystal X-Ray Analysis (7fc).**

The single crystal was mounted on a glass capillary. X-ray diffractions were collected on a Rigaku R-AXIS RAPID-F graphite-monochromated Mo K $\alpha$  radiation diffractometer with imaging plate. A symmetry-related absorption correction was carried out by using the program ABSCOR.<sup>1</sup> The analysis was carried out with direct methods (SHELX-97<sup>2</sup> or SIR92<sup>3</sup>) using Yadokari-XG.<sup>4</sup> Details of crystal and data collection parameters are shown in *Table S1–S5*.



ORTEP structure of **7fc**

**Table S1.** Crystal data and structure refinement.

Empirical formula	C <sub>27</sub> H <sub>31</sub> Br <sub>2</sub> N O <sub>4</sub> S Si		
Formula weight	653.50		
Temperature	296(2) K		
Wavelength	0.71075 Å		
Crystal system	Monoclinic		
Space group	p_21/a		
Unit cell dimensions	a = 18.9903(5) Å	$\alpha = 90^\circ$ . $\beta = 108.1468(7)^\circ$ . $\gamma = 90^\circ$ .	
	b = 12.1460(3) Å		
	c = 26.2415(6) Å		
Volume	5751.7(2) Å <sup>3</sup>		
Z	8		
Density (calculated)	1.509 Mg/m <sup>3</sup>		
Absorption coefficient	2.966 mm <sup>-1</sup>		
F(000)	2656		
Crystal size	0.80 x 0.80 x 0.50 mm <sup>3</sup>		
Theta range for data collection	3.12 to 27.49°.		
Index ranges	-24 <= h <= 24, -15 <= k <= 15, -33 <= l <= 28		
Reflections collected	53458		
Independent reflections	13126 [R(int) = 0.0794]		
Completeness to theta = 27.49°	99.4 %		
Absorption correction	Semi-empirical from equivalents		
Max. and min. transmission	0.3187 and 0.2001		
Refinement method	Full-matrix least-squares on F <sup>2</sup>		
Data / restraints / parameters	13126 / 0 / 659		
Goodness-of-fit on F <sup>2</sup>	1.162		
Final R indices [I>2sigma(I)]	R1 = 0.0552, wR2 = 0.1320		
R indices (all data)	R1 = 0.0875, wR2 = 0.1860		
Largest diff. peak and hole	0.877 and -2.055 e.Å <sup>-3</sup>		

**Table S2.** Atomic coordinates (x 10<sup>4</sup>) and equivalent isotropic displacement parameters (Å<sup>2</sup>x 10<sup>3</sup>). U(eq) is defined as one third of the trace of the orthogonalized U<sup>ij</sup> tensor.

	x	y	z	U(eq)
Br(1)	-747(1)	5550(1)	3367(1)	34(1)

Br(2)	1508(1)	-1359(1)	-2123(1)	42(1)
Br(3)	617(1)	5544(1)	1673(1)	41(1)
Br(4)	3166(1)	-1479(1)	7142(1)	45(1)
S(5)	785(1)	1366(1)	5376(1)	19(1)
S(6)	407(1)	1365(1)	-426(1)	20(1)
Si(7)	1034(1)	-163(1)	3271(1)	21(1)
Si(8)	2626(1)	140(1)	1716(1)	21(1)
O(9)	505(2)	2499(2)	-518(1)	24(1)
O(17)	2029(2)	627(2)	1154(1)	20(1)
O(18)	918(2)	479(2)	3801(1)	21(1)
O(19)	1958(2)	-523(2)	4912(1)	27(1)
O(13)	-296(2)	955(2)	-418(1)	24(1)
O(14)	66(1)	1027(2)	5377(1)	22(1)
O(15)	1001(2)	2495(2)	5451(1)	24(1)
N(20)	984(2)	1043(3)	154(1)	20(1)
O(16)	1991(2)	-439(2)	28(1)	25(1)
C(83)	1165(2)	-550(4)	-1628(2)	29(1)
N(19)	850(2)	989(3)	4799(1)	20(1)
C(20)	1446(2)	597(3)	5876(1)	21(1)
C(21)	851(2)	3280(4)	1514(2)	27(1)
C(22)	1496(2)	1292(3)	4653(1)	18(1)
C(23)	4274(2)	192(4)	341(2)	25(1)
C(24)	2471(2)	-622(4)	6619(2)	30(1)
C(25)	2325(2)	-1328(3)	1727(2)	25(1)
C(26)	1029(2)	4329(4)	1407(2)	29(1)
C(27)	527(2)	-498(4)	-976(2)	26(1)
C(28)	3548(2)	-50(3)	298(1)	21(1)
C(29)	3008(2)	776(3)	172(1)	18(1)
C(30)	3324(2)	-76(3)	4706(1)	22(1)
C(31)	-428(2)	3268(4)	3419(2)	27(1)
C(32)	607(2)	4426(4)	3879(2)	28(1)
C(33)	-130(2)	4291(4)	3577(2)	26(1)
C(34)	1050(2)	1154(4)	-1216(2)	29(1)
C(35)	482(3)	-1461(3)	3221(2)	27(1)
C(36)	1811(2)	-1114(4)	6324(2)	30(1)
C(37)	671(2)	624(3)	-915(1)	20(1)
C(38)	763(2)	2456(3)	3870(1)	21(1)
C(39)	2240(2)	486(3)	141(1)	19(1)
C(40)	4011(2)	196(4)	4672(2)	25(1)
C(41)	2034(2)	-463(4)	3401(2)	31(1)
C(42)	-319(2)	-1173(4)	3182(2)	29(1)
C(43)	1755(2)	1387(3)	283(1)	18(1)
C(44)	1681(2)	2568(3)	1063(1)	19(1)
C(45)	1267(2)	1463(3)	4037(2)	20(1)
C(46)	2077(2)	1628(3)	893(1)	22(1)
C(47)	780(3)	-1097(4)	-1334(2)	31(1)
C(48)	1841(2)	3638(3)	948(2)	26(1)
C(49)	4468(2)	1257(4)	249(2)	26(1)
C(50)	488(3)	-2143(4)	2729(2)	38(1)
C(51)	1513(2)	4528(4)	1115(2)	30(1)
C(52)	2842(3)	-1949(4)	2209(2)	34(1)
C(53)	3113(2)	1813(4)	4880(2)	25(1)
C(54)	1533(3)	-1364(4)	1762(2)	34(1)
C(55)	1292(2)	-494(4)	5948(2)	26(1)
C(56)	2543(2)	927(4)	2301(2)	25(1)
C(57)	3938(2)	2081(4)	115(2)	28(1)
C(58)	1180(2)	2389(4)	1340(1)	24(1)
C(80)	2106(2)	410(3)	4812(1)	19(1)
C(60)	829(3)	-2148(4)	3732(2)	33(1)
C(61)	3808(2)	2076(4)	4862(2)	27(1)

C(62)	21(2)	2347(4)	3572(2)	26(1)
C(63)	3212(2)	1844(4)	87(1)	23(1)
C(64)	679(2)	728(4)	2668(2)	27(1)
C(65)	2858(2)	731(3)	4800(1)	21(1)
C(66)	1049(2)	3508(3)	4022(2)	26(1)
C(67)	2104(2)	1080(4)	6175(2)	31(1)
C(68)	4253(2)	1271(4)	4750(2)	28(1)
C(69)	1307(3)	551(4)	-1577(2)	34(1)
C(70)	3592(2)	265(4)	1701(2)	34(1)
C(72)	2623(3)	454(4)	6548(2)	38(1)
C(78)	2329(3)	-1905(4)	1205(2)	38(1)

**Table S3.** Bond lengths [Å] and angles [°].

Br(1)-C(33)	1.900(4)
Br(2)-C(83)	1.899(4)
Br(3)-C(26)	1.903(4)
Br(4)-C(24)	1.892(4)
S(5)-O(14)	1.426(3)
S(5)-O(15)	1.427(3)
S(5)-N(19)	1.622(3)
S(5)-C(20)	1.773(4)
S(6)-O(9)	1.421(3)
S(6)-O(13)	1.431(3)
S(6)-N(20)	1.622(3)
S(6)-C(37)	1.763(4)
Si(7)-O(18)	1.667(3)
Si(7)-C(41)	1.857(5)
Si(7)-C(64)	1.861(4)
Si(7)-C(35)	1.876(4)
Si(8)-O(17)	1.664(3)
Si(8)-C(70)	1.853(5)
Si(8)-C(56)	1.857(4)
Si(8)-C(25)	1.875(4)
O(17)-C(46)	1.412(5)
O(18)-C(45)	1.413(5)
O(19)-C(80)	1.215(5)
N(20)-C(43)	1.457(5)
O(16)-C(39)	1.219(5)
C(83)-C(69)	1.363(6)
C(83)-C(47)	1.385(7)
N(19)-C(22)	1.443(5)
C(20)-C(67)	1.382(5)
C(20)-C(55)	1.383(6)
C(21)-C(26)	1.371(6)
C(21)-C(58)	1.395(6)
C(22)-C(80)	1.538(5)
C(22)-C(45)	1.551(5)
C(23)-C(28)	1.379(6)
C(23)-C(49)	1.386(6)
C(24)-C(72)	1.364(6)
C(24)-C(36)	1.387(6)
C(25)-C(52)	1.535(5)
C(25)-C(54)	1.535(6)
C(25)-C(78)	1.541(5)
C(26)-C(51)	1.389(6)
C(27)-C(47)	1.388(6)
C(27)-C(37)	1.389(6)
C(28)-C(29)	1.400(5)

C(29)-C(63)	1.391(6)
C(29)-C(39)	1.476(5)
C(30)-C(40)	1.375(6)
C(30)-C(65)	1.394(6)
C(31)-C(33)	1.374(6)
C(31)-C(62)	1.388(6)
C(32)-C(66)	1.376(6)
C(32)-C(33)	1.388(6)
C(34)-C(37)	1.381(6)
C(34)-C(69)	1.401(6)
C(35)-C(42)	1.533(6)
C(35)-C(50)	1.536(5)
C(35)-C(60)	1.543(5)
C(36)-C(55)	1.380(6)
C(38)-C(62)	1.389(5)
C(38)-C(66)	1.396(6)
C(38)-C(45)	1.516(5)
C(39)-C(43)	1.549(5)
C(40)-C(68)	1.378(6)
C(43)-C(46)	1.553(5)
C(44)-C(58)	1.384(5)
C(44)-C(48)	1.389(6)
C(44)-C(46)	1.508(6)
C(48)-C(51)	1.383(6)
C(49)-C(57)	1.384(6)
C(53)-C(61)	1.375(6)
C(53)-C(65)	1.393(6)
C(57)-C(63)	1.390(6)
C(80)-C(65)	1.491(5)
C(61)-C(68)	1.382(6)
C(67)-C(72)	1.381(6)

O(14)-S(5)-O(15)	120.79(18)
O(14)-S(5)-N(19)	105.69(16)
O(15)-S(5)-N(19)	107.33(17)
O(14)-S(5)-C(20)	107.72(18)
O(15)-S(5)-C(20)	107.26(17)
N(19)-S(5)-C(20)	107.43(17)
O(9)-S(6)-O(13)	120.84(18)
O(9)-S(6)-N(20)	107.91(17)
O(13)-S(6)-N(20)	105.22(17)
O(9)-S(6)-C(37)	106.77(18)
O(13)-S(6)-C(37)	108.12(18)
N(20)-S(6)-C(37)	107.35(17)
O(18)-Si(7)-C(41)	109.41(17)
O(18)-Si(7)-C(64)	109.26(18)
C(41)-Si(7)-C(64)	110.5(2)
O(18)-Si(7)-C(35)	103.65(17)
C(41)-Si(7)-C(35)	111.4(2)
C(64)-Si(7)-C(35)	112.41(19)
O(17)-Si(8)-C(70)	110.99(17)
O(17)-Si(8)-C(56)	109.81(17)
C(70)-Si(8)-C(56)	108.3(2)
O(17)-Si(8)-C(25)	103.04(16)
C(70)-Si(8)-C(25)	112.8(2)
C(56)-Si(8)-C(25)	111.84(19)
C(46)-O(17)-Si(8)	127.4(2)
C(45)-O(18)-Si(7)	126.6(2)
C(43)-N(20)-S(6)	119.0(3)
C(69)-C(83)-C(47)	122.6(4)

C(69)-C(83)-Br(2)	118.4(4)
C(47)-C(83)-Br(2)	119.0(3)
C(22)-N(19)-S(5)	119.6(3)
C(67)-C(20)-C(55)	121.4(4)
C(67)-C(20)-S(5)	120.2(3)
C(55)-C(20)-S(5)	118.4(3)
C(26)-C(21)-C(58)	119.3(4)
N(19)-C(22)-C(80)	112.4(3)
N(19)-C(22)-C(45)	109.0(3)
C(80)-C(22)-C(45)	108.6(3)
C(28)-C(23)-C(49)	119.8(4)
C(72)-C(24)-C(36)	122.0(4)
C(72)-C(24)-Br(4)	119.8(3)
C(36)-C(24)-Br(4)	118.2(3)
C(52)-C(25)-C(54)	108.9(4)
C(52)-C(25)-C(78)	109.3(4)
C(54)-C(25)-C(78)	108.4(4)
C(52)-C(25)-Si(8)	111.1(3)
C(54)-C(25)-Si(8)	109.7(3)
C(78)-C(25)-Si(8)	109.4(3)
C(21)-C(26)-C(51)	121.6(4)
C(21)-C(26)-Br(3)	119.2(3)
C(51)-C(26)-Br(3)	119.2(4)
C(47)-C(27)-C(37)	119.7(4)
C(23)-C(28)-C(29)	120.5(4)
C(63)-C(29)-C(28)	119.1(4)
C(63)-C(29)-C(39)	122.2(3)
C(28)-C(29)-C(39)	118.7(4)
C(40)-C(30)-C(65)	120.7(4)
C(33)-C(31)-C(62)	118.9(4)
C(66)-C(32)-C(33)	118.7(4)
C(31)-C(33)-C(32)	121.8(4)
C(31)-C(33)-Br(1)	118.8(3)
C(32)-C(33)-Br(1)	119.4(3)
C(37)-C(34)-C(69)	119.8(4)
C(42)-C(35)-C(50)	109.5(4)
C(42)-C(35)-C(60)	109.1(4)
C(50)-C(35)-C(60)	109.0(4)
C(42)-C(35)-Si(7)	109.6(3)
C(50)-C(35)-Si(7)	111.3(3)
C(60)-C(35)-Si(7)	108.4(3)
C(55)-C(36)-C(24)	118.9(4)
C(34)-C(37)-C(27)	120.7(4)
C(34)-C(37)-S(6)	119.6(3)
C(27)-C(37)-S(6)	119.6(3)
C(62)-C(38)-C(66)	118.9(4)
C(62)-C(38)-C(45)	121.7(4)
C(66)-C(38)-C(45)	119.4(3)
O(16)-C(39)-C(29)	122.4(4)
O(16)-C(39)-C(43)	119.6(4)
C(29)-C(39)-C(43)	117.9(3)
C(30)-C(40)-C(68)	119.8(4)
N(20)-C(43)-C(39)	111.8(3)
N(20)-C(43)-C(46)	109.7(3)
C(39)-C(43)-C(46)	108.0(3)
C(58)-C(44)-C(48)	119.5(4)
C(58)-C(44)-C(46)	121.7(4)
C(48)-C(44)-C(46)	118.8(4)
O(18)-C(45)-C(38)	112.3(3)
O(18)-C(45)-C(22)	106.6(3)

C(38)-C(45)-C(22)	110.8(3)
O(17)-C(46)-C(44)	113.2(3)
O(17)-C(46)-C(43)	105.9(3)
C(44)-C(46)-C(43)	112.1(3)
C(83)-C(47)-C(27)	118.6(4)
C(51)-C(48)-C(44)	120.8(4)
C(57)-C(49)-C(23)	120.5(4)
C(48)-C(51)-C(26)	118.6(4)
C(61)-C(53)-C(65)	120.4(4)
C(36)-C(55)-C(20)	119.1(4)
C(49)-C(57)-C(63)	119.6(4)
C(44)-C(58)-C(21)	120.1(4)
O(19)-C(80)-C(65)	122.5(4)
O(19)-C(80)-C(22)	120.2(3)
C(65)-C(80)-C(22)	117.2(3)
C(53)-C(61)-C(68)	120.0(4)
C(31)-C(62)-C(38)	120.7(4)
C(57)-C(63)-C(29)	120.4(4)
C(53)-C(65)-C(30)	118.7(4)
C(53)-C(65)-C(80)	122.2(4)
C(30)-C(65)-C(80)	119.0(4)
C(32)-C(66)-C(38)	120.9(4)
C(72)-C(67)-C(20)	119.3(4)
C(40)-C(68)-C(61)	120.3(4)
C(83)-C(69)-C(34)	118.6(4)
C(24)-C(72)-C(67)	119.3(4)

Symmetry transformations used to generate equivalent atoms:

**Table S4.** Anisotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ).

The anisotropic displacement factor exponent takes the form:  $-2p^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^{*} b^{*} U^{12}]$

	U <sup>11</sup>	U <sup>22</sup>	U <sup>33</sup>	U <sup>23</sup>	U <sup>13</sup>	U <sup>12</sup>
Br(1)	32(1)	20(1)	43(1)	5(1)	1(1)	5(1)
Br(2)	38(1)	55(1)	31(1)	-13(1)	6(1)	17(1)
Br(3)	42(1)	29(1)	51(1)	-13(1)	12(1)	7(1)
Br(4)	41(1)	48(1)	38(1)	19(1)	0(1)	12(1)
S(5)	19(1)	17(1)	22(1)	2(1)	6(1)	2(1)
S(6)	16(1)	19(1)	22(1)	-1(1)	4(1)	1(1)
Si(7)	23(1)	18(1)	22(1)	0(1)	6(1)	1(1)
Si(8)	22(1)	20(1)	20(1)	0(1)	5(1)	2(1)
O(9)	26(2)	17(2)	27(1)	1(1)	7(1)	2(1)
O(17)	20(1)	16(2)	23(1)	2(1)	4(1)	0(1)
O(18)	23(1)	17(2)	25(1)	-2(1)	9(1)	0(1)
O(19)	27(2)	15(2)	40(2)	3(1)	14(1)	-1(1)
O(13)	17(1)	24(2)	31(1)	1(1)	5(1)	-1(1)
O(14)	15(1)	26(2)	30(1)	1(1)	13(1)	-1(1)
O(15)	27(2)	13(2)	32(1)	-1(1)	10(1)	0(1)
N(20)	17(2)	28(2)	15(1)	1(1)	4(1)	-3(1)
O(16)	23(2)	20(2)	31(1)	-3(1)	7(1)	-3(1)
C(83)	25(2)	36(3)	21(2)	-4(2)	3(2)	6(2)
N(19)	18(2)	20(2)	22(2)	-2(1)	5(1)	1(1)
C(20)	22(2)	21(2)	19(2)	1(1)	8(2)	0(2)
C(21)	24(2)	24(2)	30(2)	-4(2)	5(2)	1(2)
C(22)	16(2)	17(2)	22(2)	0(1)	7(1)	-3(2)
C(23)	22(2)	27(2)	29(2)	0(2)	10(2)	5(2)
C(24)	32(2)	33(3)	21(2)	9(2)	4(2)	4(2)
C(25)	32(2)	22(2)	22(2)	-1(2)	8(2)	4(2)
C(26)	26(2)	29(3)	29(2)	-6(2)	4(2)	4(2)
C(27)	28(2)	27(2)	25(2)	-2(2)	8(2)	-4(2)

C(28)	23(2)	23(2)	18(2)	-2(1)	7(1)	-2(2)
C(29)	20(2)	19(2)	16(2)	2(1)	9(1)	2(2)
C(30)	19(2)	19(2)	25(2)	-1(2)	4(2)	1(2)
C(31)	25(2)	24(2)	27(2)	1(2)	2(2)	0(2)
C(32)	32(2)	16(2)	30(2)	2(2)	1(2)	-2(2)
C(33)	31(2)	22(2)	22(2)	2(2)	5(2)	3(2)
C(34)	35(2)	24(2)	30(2)	-1(2)	11(2)	0(2)
C(35)	34(2)	20(2)	22(2)	-3(2)	3(2)	-2(2)
C(36)	32(2)	21(2)	35(2)	8(2)	8(2)	-1(2)
C(37)	16(2)	23(2)	19(2)	1(1)	3(1)	1(2)
C(38)	26(2)	22(2)	16(2)	2(1)	8(1)	1(2)
C(39)	24(2)	15(2)	18(2)	1(1)	4(1)	2(2)
C(40)	21(2)	23(2)	32(2)	-1(2)	10(2)	5(2)
C(41)	31(2)	31(3)	27(2)	-2(2)	5(2)	4(2)
C(42)	24(2)	28(3)	40(2)	-3(2)	17(2)	-4(2)
C(43)	20(2)	13(2)	21(2)	-2(1)	5(1)	-2(2)
C(44)	13(2)	19(2)	22(2)	-2(1)	1(1)	-1(2)
C(45)	23(2)	15(2)	24(2)	1(1)	10(2)	-3(2)
C(46)	23(2)	15(2)	24(2)	2(2)	3(2)	-2(2)
C(47)	39(3)	22(2)	25(2)	-7(2)	0(2)	-2(2)
C(48)	27(2)	23(2)	25(2)	0(2)	4(2)	-3(2)
C(49)	24(2)	29(3)	27(2)	2(2)	11(2)	-2(2)
C(50)	58(3)	24(3)	37(2)	-10(2)	22(2)	-8(2)
C(51)	32(2)	23(2)	29(2)	0(2)	3(2)	0(2)
C(52)	46(3)	22(2)	31(2)	2(2)	9(2)	5(2)
C(53)	23(2)	21(2)	30(2)	-3(2)	6(2)	1(2)
C(54)	40(3)	27(3)	31(2)	2(2)	5(2)	-7(2)
C(55)	21(2)	26(2)	29(2)	3(2)	4(2)	-3(2)
C(56)	24(2)	25(2)	24(2)	-1(2)	6(2)	-2(2)
C(57)	26(2)	27(3)	34(2)	5(2)	15(2)	-6(2)
C(58)	19(2)	26(2)	24(2)	-2(2)	0(2)	-3(2)
C(80)	18(2)	20(2)	20(2)	-1(1)	6(1)	-1(2)
C(60)	40(3)	23(3)	37(2)	7(2)	12(2)	-4(2)
C(61)	23(2)	19(2)	35(2)	0(2)	3(2)	-5(2)
C(62)	23(2)	19(2)	32(2)	-3(2)	5(2)	-4(2)
C(63)	20(2)	21(2)	28(2)	1(2)	8(2)	2(2)
C(64)	26(2)	28(3)	27(2)	1(2)	9(2)	-2(2)
C(65)	20(2)	20(2)	22(2)	1(2)	6(2)	2(2)
C(66)	23(2)	23(2)	29(2)	0(2)	5(2)	-5(2)
C(67)	26(2)	29(3)	34(2)	6(2)	1(2)	-8(2)
C(68)	21(2)	29(3)	32(2)	1(2)	6(2)	-3(2)
C(69)	36(3)	40(3)	29(2)	-2(2)	14(2)	-1(2)
C(70)	31(2)	48(3)	23(2)	4(2)	7(2)	5(2)
C(72)	30(2)	38(3)	36(2)	4(2)	-4(2)	-2(2)
C(78)	54(3)	26(3)	34(2)	-3(2)	15(2)	5(2)

**Table S5.** Hydrogen coordinates ( $x \times 10^4$ ) and isotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ).

	x	y	z	U(eq)
H(61)	837	675	382	24
H(62)	501	610	4582	24
H(1)	515	3163	1702	32
H(57)	1694	1986	4831	22
H(9)	4633	-357	431	30
H(10)	263	-846	-777	32
H(11)	3417	-768	352	25
H(12)	3169	-806	4665	26
H(2)	-923	3194	3214	32
H(3)	799	5125	3983	33

H(13)	1133	1908	-1178	35
H(14)	1719	-1850	6378	36
H(15)	4312	-344	4596	30
H(27)	2308	214	3460	46
H(28)	2101	-839	3098	46
H(29)	2209	-920	3714	46
H(30)	-536	-740	2865	43
H(31)	-323	-761	3493	43
H(32)	-599	-1839	3162	43
H(58)	1780	2059	82	22
H(59)	1713	1579	3932	24
H(60)	2601	1824	974	26
H(16)	693	-1850	-1377	37
H(4)	2172	3758	757	31
H(17)	4957	1419	278	31
H(33)	990	-2306	2748	57
H(34)	254	-1732	2408	57
H(35)	222	-2818	2723	57
H(5)	1615	5243	1033	36
H(36)	2668	-2691	2211	50
H(37)	2846	-1586	2534	50
H(38)	3334	-1959	2182	50
H(18)	2810	2360	4947	30
H(39)	1208	-961	1468	51
H(40)	1526	-1039	2094	51
H(41)	1370	-2115	1747	51
H(19)	846	-806	5745	31
H(42)	2624	1695	2252	38
H(43)	2906	668	2621	38
H(44)	2056	826	2330	38
H(20)	4068	2789	43	33
H(6)	1062	1674	1411	29
H(45)	524	-2781	3729	50
H(46)	862	-1707	4042	50
H(47)	1316	-2384	3743	50
H(21)	3979	2797	4924	33
H(7)	-177	1650	3473	31
H(22)	2859	2402	11	28
H(48)	171	915	2620	41
H(49)	713	339	2358	41
H(50)	970	1388	2716	41
H(8)	1545	3589	4222	31
H(23)	2195	1818	6126	38
H(24)	4718	1456	4729	33
H(25)	1570	895	-1778	41
H(51)	3653	-176	1414	51
H(52)	3927	15	2036	51
H(53)	3696	1021	1645	51
H(26)	3072	763	6748	45
H(54)	2814	-1852	1167	57
H(55)	1976	-1555	905	57
H(56)	2199	-2665	1218	57

- (1) Higashi, T. *ABSCOR. Program for Absorption Correction.*; Rigaku Corporation: Japan, 1995.  
 (2) Sheldrick, G. M. *SHELX-97. Programs for Crystal Structure Analysis.*; University of Göttingen: Germany, 1997.  
 (3) Altomare, A.; Burla, M. C.; Camalli, M.; Cascarano, G. L.; Giacovazzo, C.; Guagliardi, A.; Moliterni, A. G. G.; Polidori, G.; Spagna, R. *J. Appl. Cryst.* **1999**, 32, 115–119.  
 (4) Wakita, K. *Yadokari-XG. Program for Crystal Structure Analysis.*; 2000.

## triazole Ac

File: exp

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vmlml

Mercury-400B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acc time 3.500 sec

width 64.0 Hz

16 repetitions

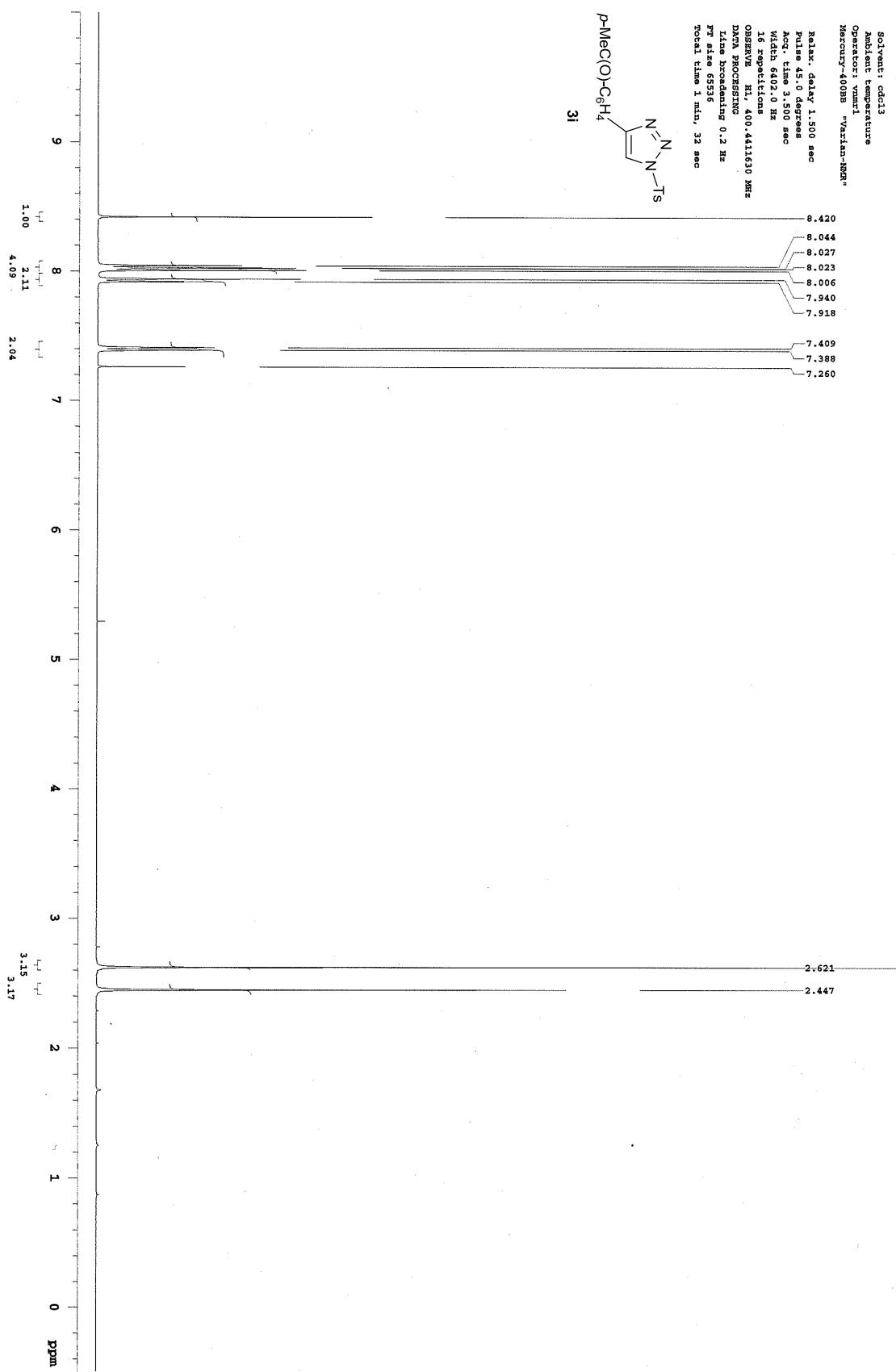
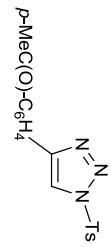
OBSERVE H1, 400.4411630 MHz

DATA PROCESSING

Line broadening 0.2 Hz

FT size 65536

Total time 1 min, 32 sec



triazole-Nc

File: exp

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vnmrl

Mercury-400B "Varian-NMR"

376

Relax. delay 0.700 sec

Pulse 15.0 degrees

Acc. time 1.300 sec

Width 24154.5 Hz

10560 repetitions

OBSERVE C13, 100.6910157 MHz

DECOUPLE H1, 400.4431956 MHz

Power 41 dB

continuously on

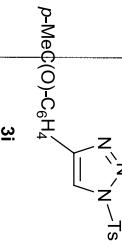
WALTZ-16 modulated

DATA PROCESSING

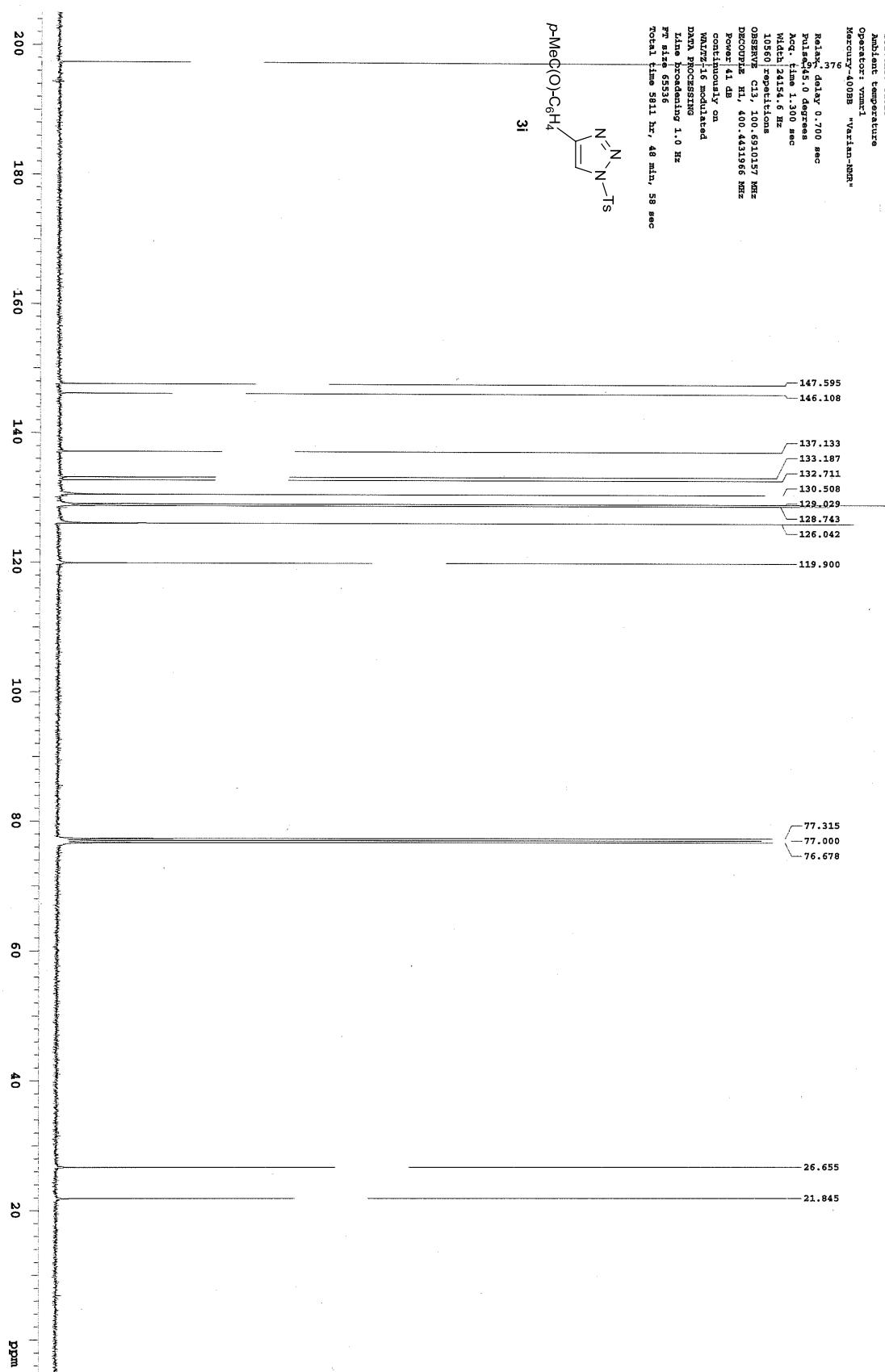
Line broadening 1.0 Hz

FT size 65536

Total time 5811 hr, 48 min, 58 sec



3i





1038-pu

File: exp

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Temp. 24.0 C / 297.1 K

Operator: vmlml

Mercury-400B "Varian-NMR"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acc. time 1.300 sec

width 24154.6 Hz

102 repetitions

OBSERVE C13, 100.6510157 MHz

DECOUPLE H1, 400.4431986 MHz

Power 41 dB

continuously on

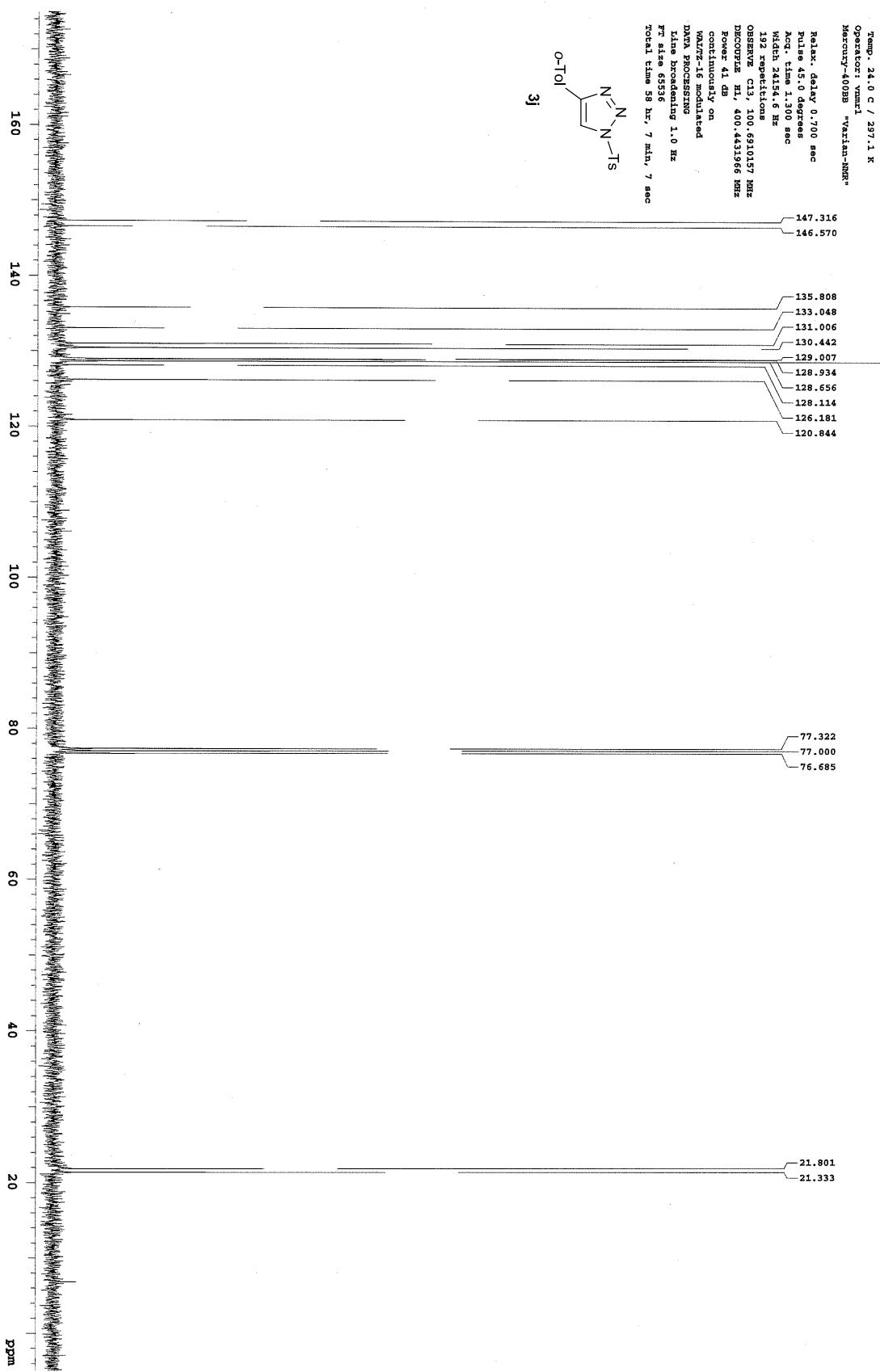
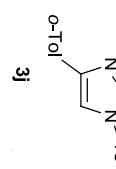
WAVEZ-16 modulated

DATA PROCESSING

line broadening 1.0 Hz

FT size 65536

Total time 58 hr, 7 min, 7 sec



tirazole i-Bu

File: exp

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Mercury-400B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

width 642.0 Hz

16 repetitions

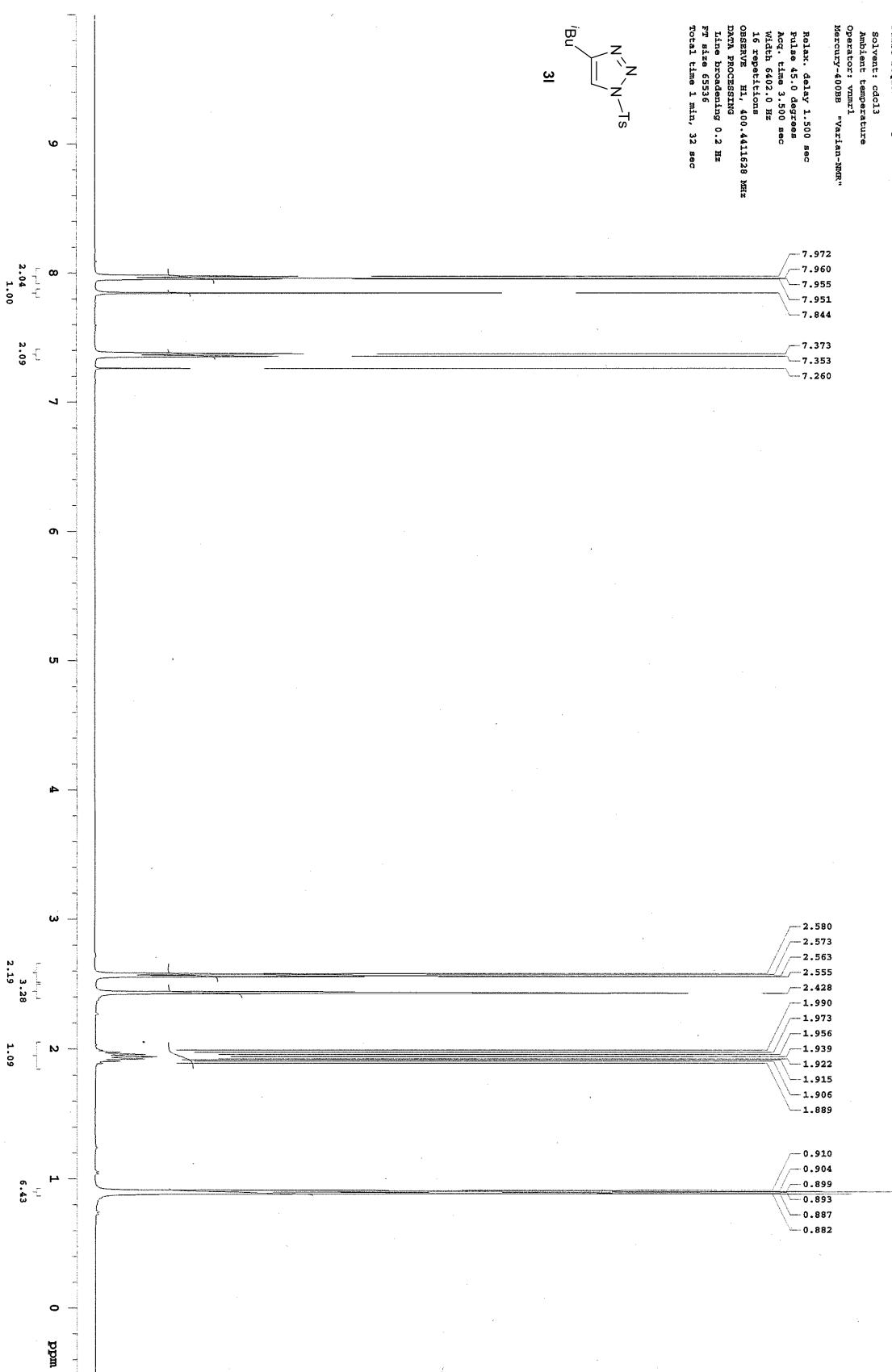
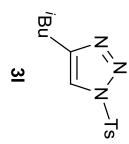
OBSERVE H, 400.4411628 MHz

DATA PROCESSING

Line broadening 0.2 Hz

FT size 65536

Total time 1 min, 32 sec



tetraole i-Bu

File: exp

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vml1

Mercury-400BB "Varian-NMR"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 2414.6 Hz

128 repetitions

OBSERVE CL1, 100.6910157 MHz

DECOUPLE HI, 400.4431986 MHz

Power 41 dB

continuously on

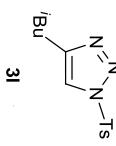
WAVEM-16 modulated

DATA PROCESSING

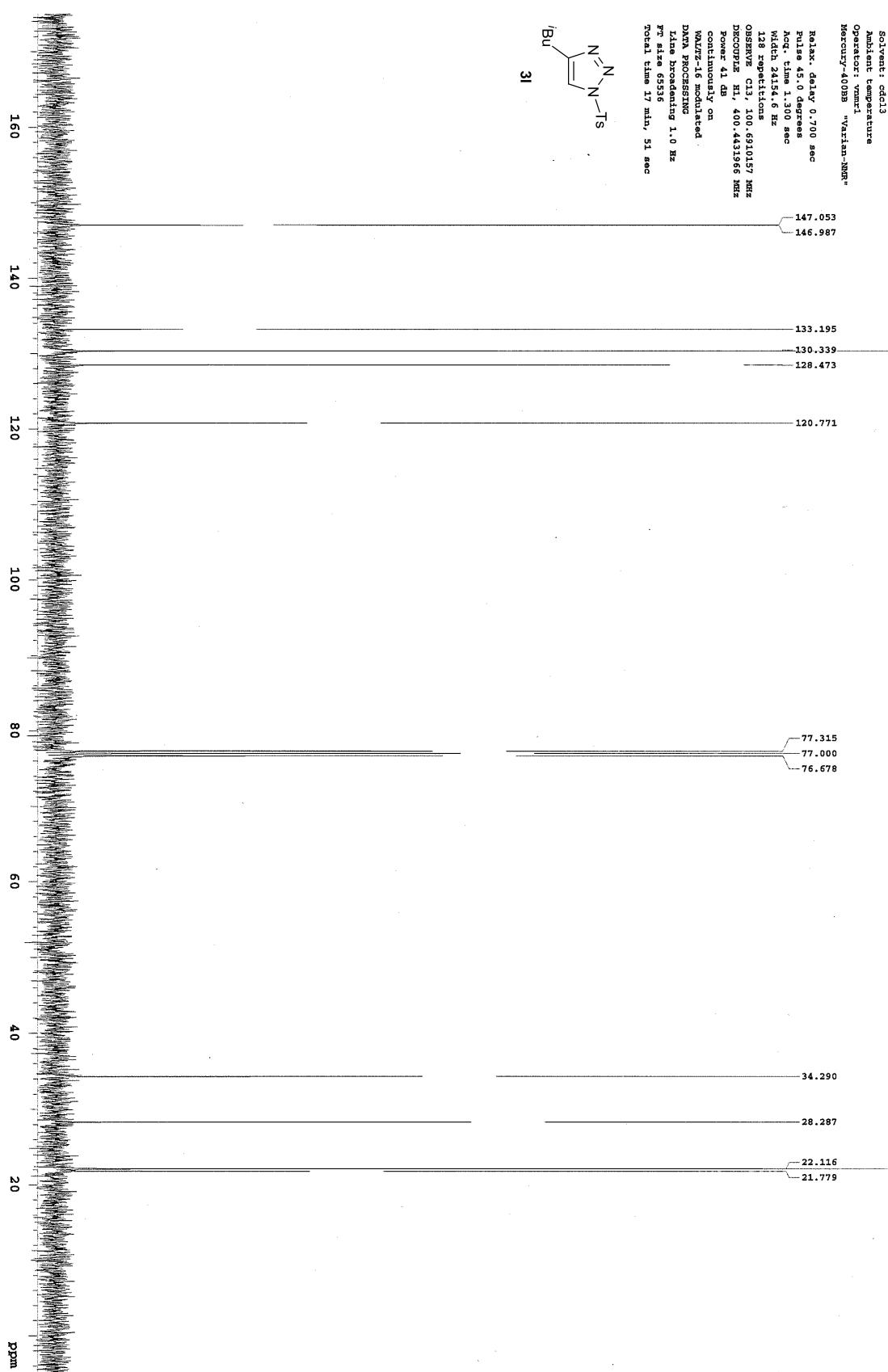
Line broadening 1.0 Hz

FT size 65536

Total time 17 min, 51 sec



31



## SelectFlux F

File: /home/vmrc1/vmrcsys/data/murakami\_1ab/Nakamura/TBSCl phenyl\_product2.fid

Pulse Sequence: s2p1

Solvent: cdcl3

Ambient temperature

Operator: vmlc1

File: TBSO phenyl\_product2

MERCURY-400NB "Varian-MP"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Ave. time 3.500 sec

Width 600.0 Hz

16 repetitions

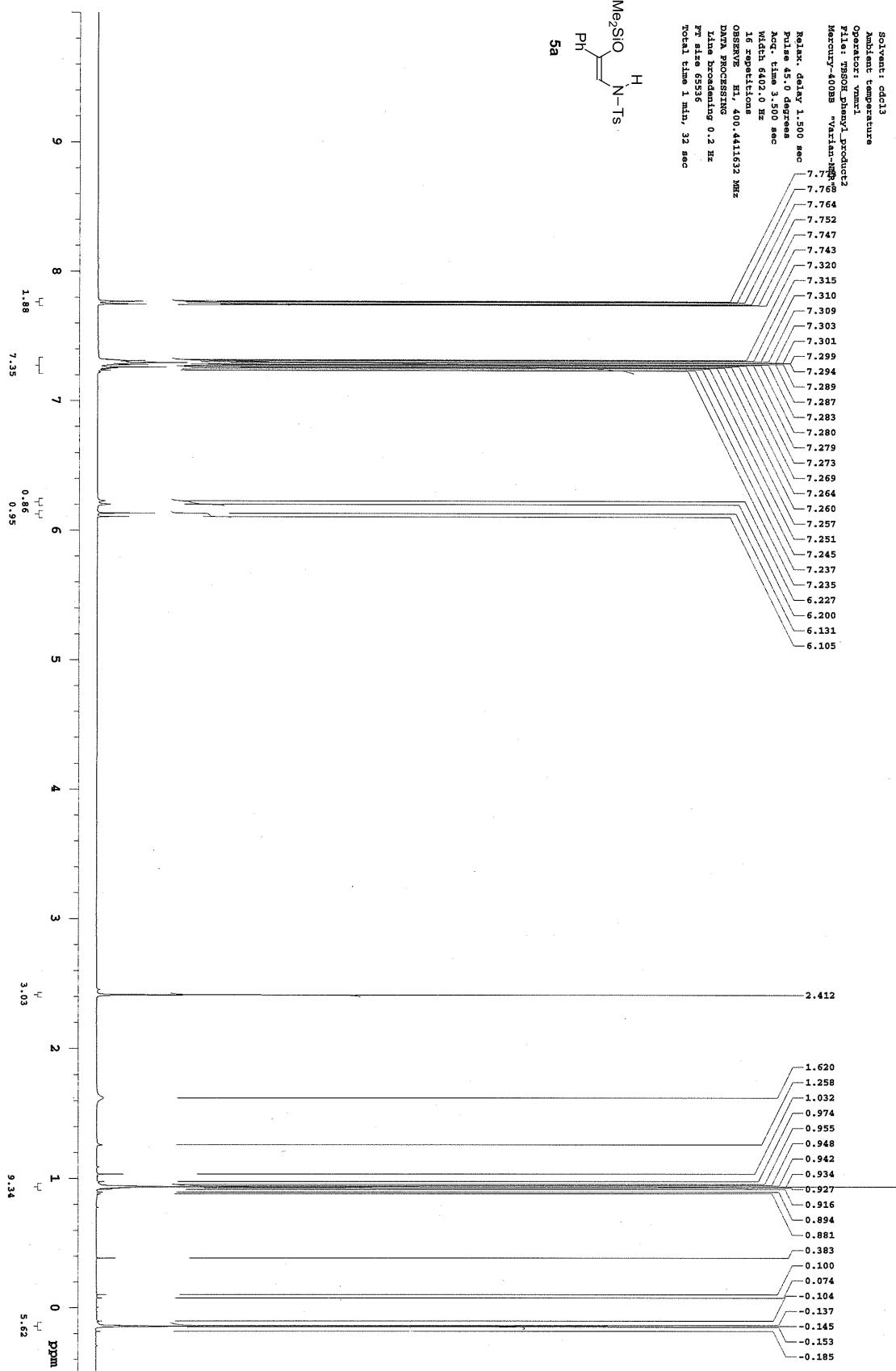
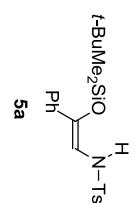
OBSERVE H1, 400.4411632 MHz

DATA PROCESSING

Line broadening 0.2 Hz

FID size 65536

Total time 1 min, 32 sec



File: /Home/vnmrl1/vnmrsys/data/murakami\_1ab/Rakamuro/Triazole\_Silanol/MBSOH\_phenyl\_product\_13C.fid

Pulse Sequence: s2p1

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vnmrl1

File: MBSOH\_phenyl\_product\_13C

Mercury-400MHz "Varian-NMR"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 2145.6 Hz

118 repetitions

OBSERVE CH, 100.6910137 MHz

DECOUPLE PH, 400.4431956 MHz

Power 40 dB

continuously on

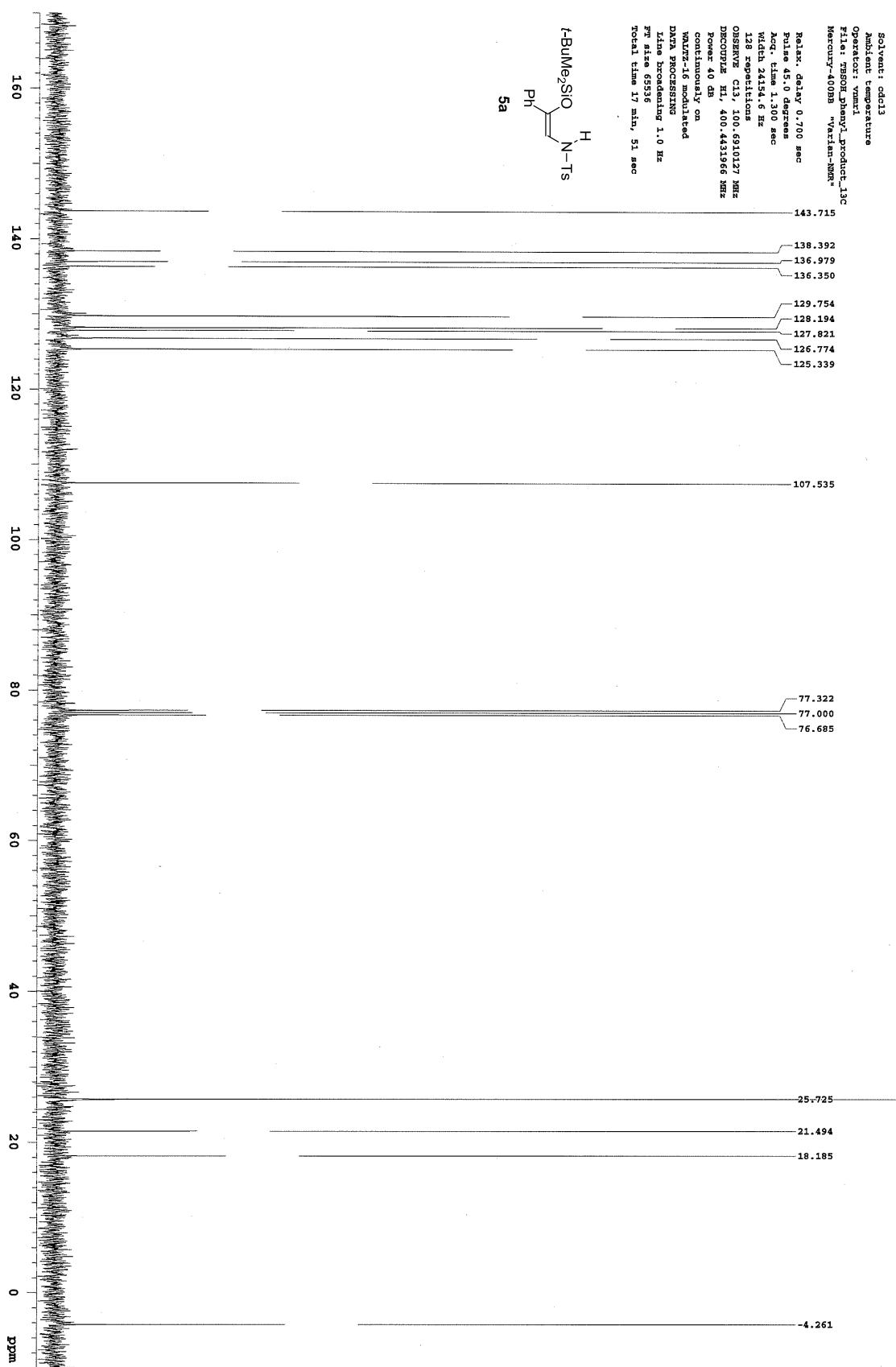
WAVZ-16 undelayed

DATA PROCESSING

Lam decimating 1.0 Hz

FT size 65536

Total time 17 min, 51 sec



File: /home/vmml1/vmmlsys/data/murakami.lab/Nakamuro/triazole.Silanol/mbor\_4-tol\_product.fid

Pulse Sequence: s2p11

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vmml

File: mbor\_4-tol\_product

Mercury-400B Varian-NMR<sup>a</sup>

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.000 sec

Width 64.0 2.0 Hz

16 repetitions

OBSERVE H1, 400.4411630 MHz

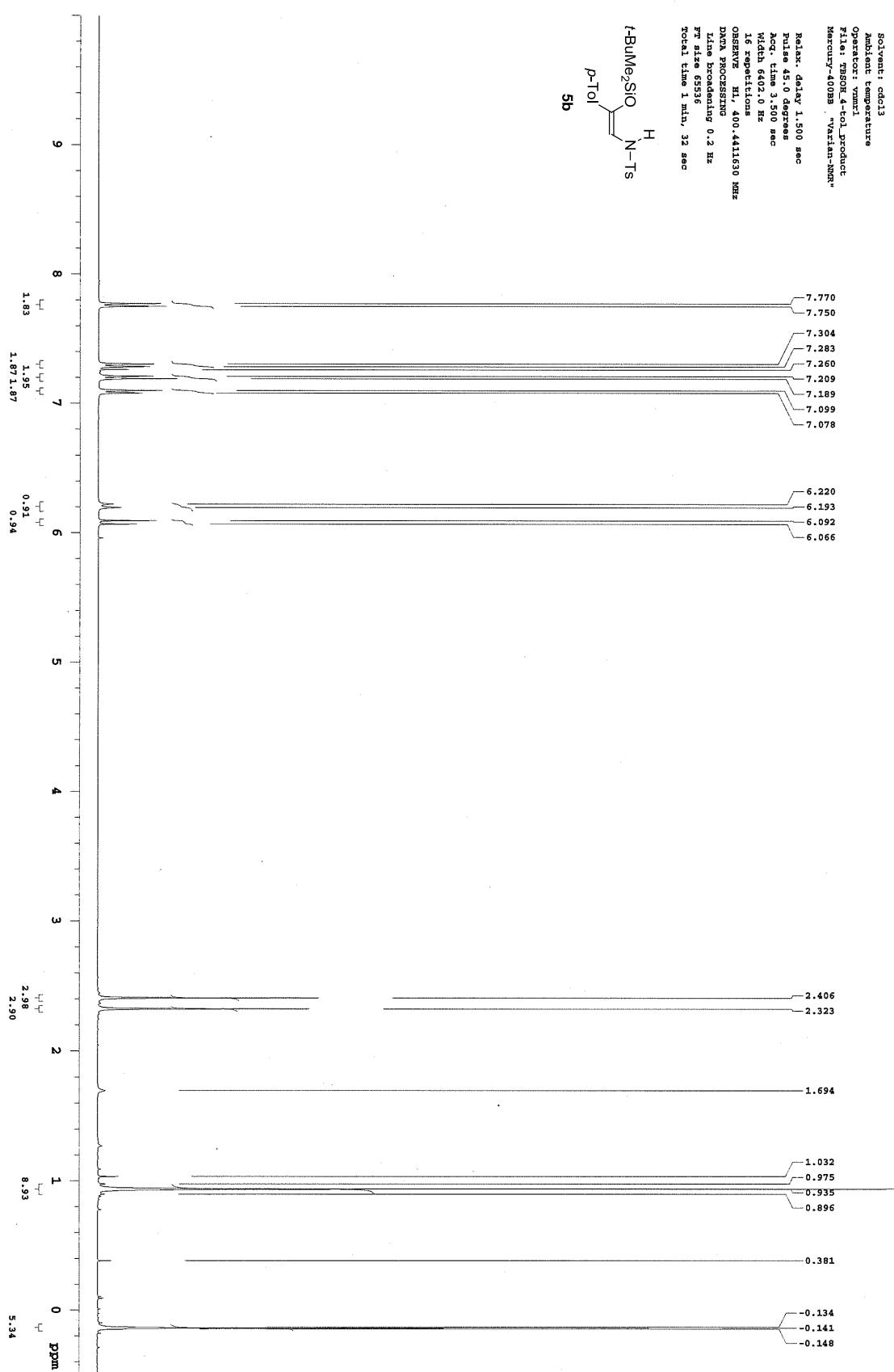
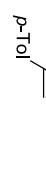
DATA PROCESSING

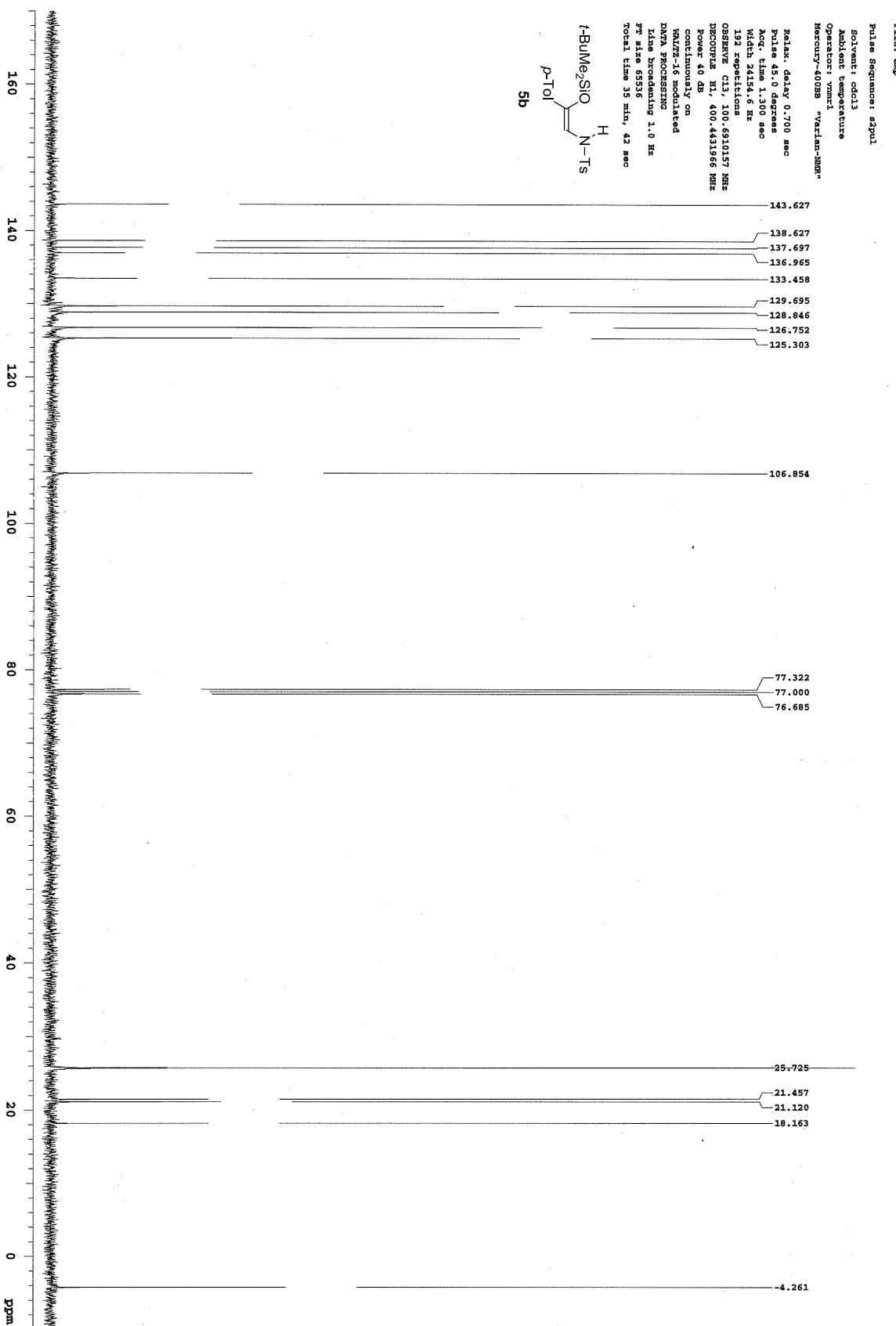
LINE BROADENING 0.2 Hz

WT size 65536

total time 1 min, 32 sec

t-BuMe<sub>2</sub>SiO<sup>H</sup>





new experiment

File: /home/vmml1/vmml1/murakami/data/murakami\_lab/Nakamuro/triazole\_Silanol/mBSOH\_methoxy\_product.fid

Pulse Sequence: s2pul

Ambient temperature

Solvent: cdcl<sub>3</sub>

Operator: vmml1

File: mBSOHi\_methoxy\_product

Mercury-400B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 642.0 Hz

16 repetitions

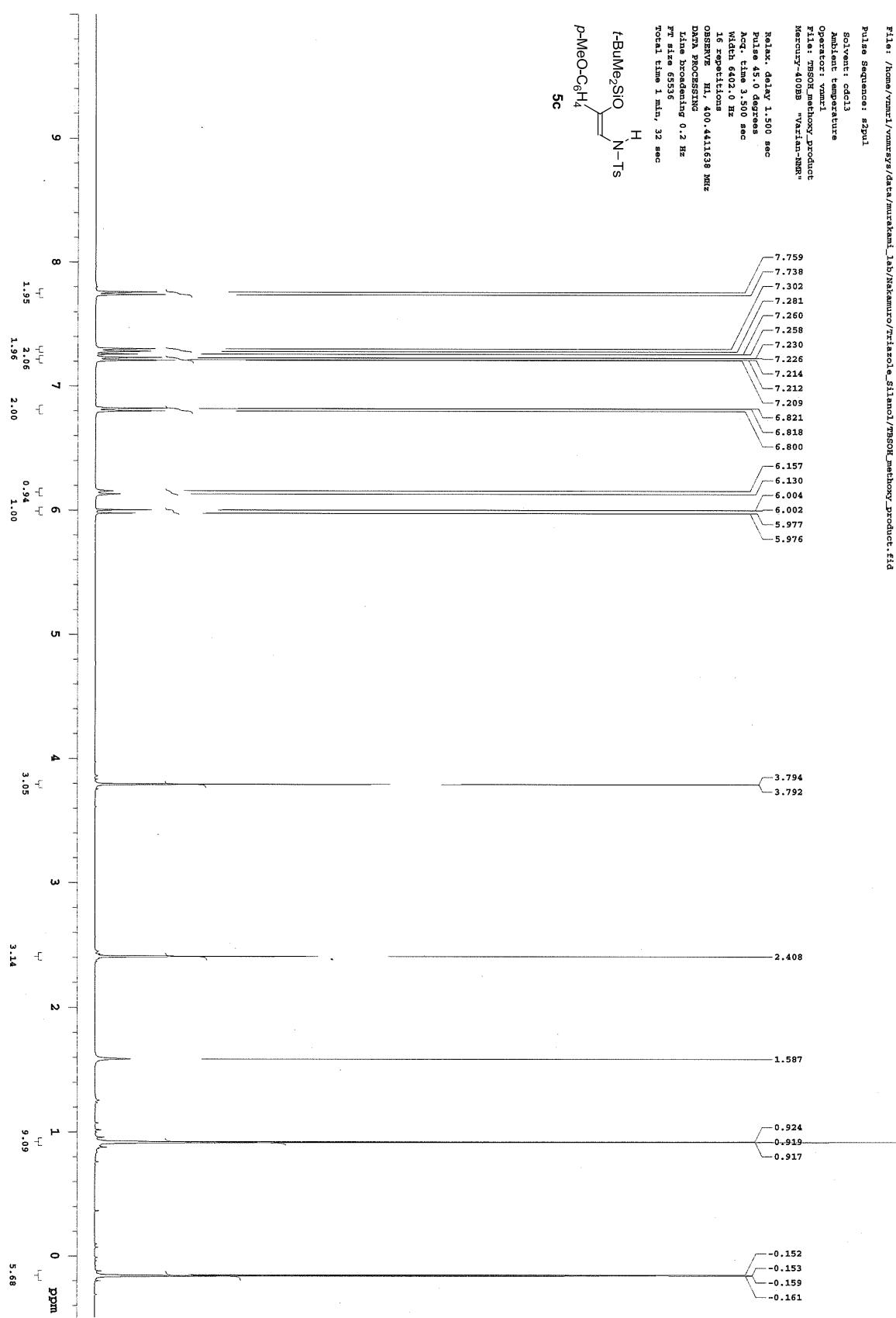
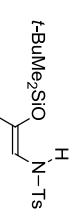
OBSERVE HI, 400.4411638 MHz

DATA PROCESSING

LINE BROADENING 0.2 Hz

FT SIZE 65536

TOTAL TIME 1 min, 32 sec



07 column

File: /home/vmncl1/vmnclsys/data/murakami\_lab/Nakamura/Triazole\_Silanol/TMSOL.methoxy\_product\_13C\_3.fid

Pulse Sequence: a2p1

Solvent:

ambient temperature

Operator: vmncl1

File: TMSOL.methoxy\_product\_13C\_3

MERCHRY-400P "VARIAN-NMR"

Relax. deg Np 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Watch 2414.6 Hz

128 repetitions

OBSERVE C13, 100.6910179 MHz

DECOUPLE H1, 400.4431986 MHz

Power 40 dB

continuously on

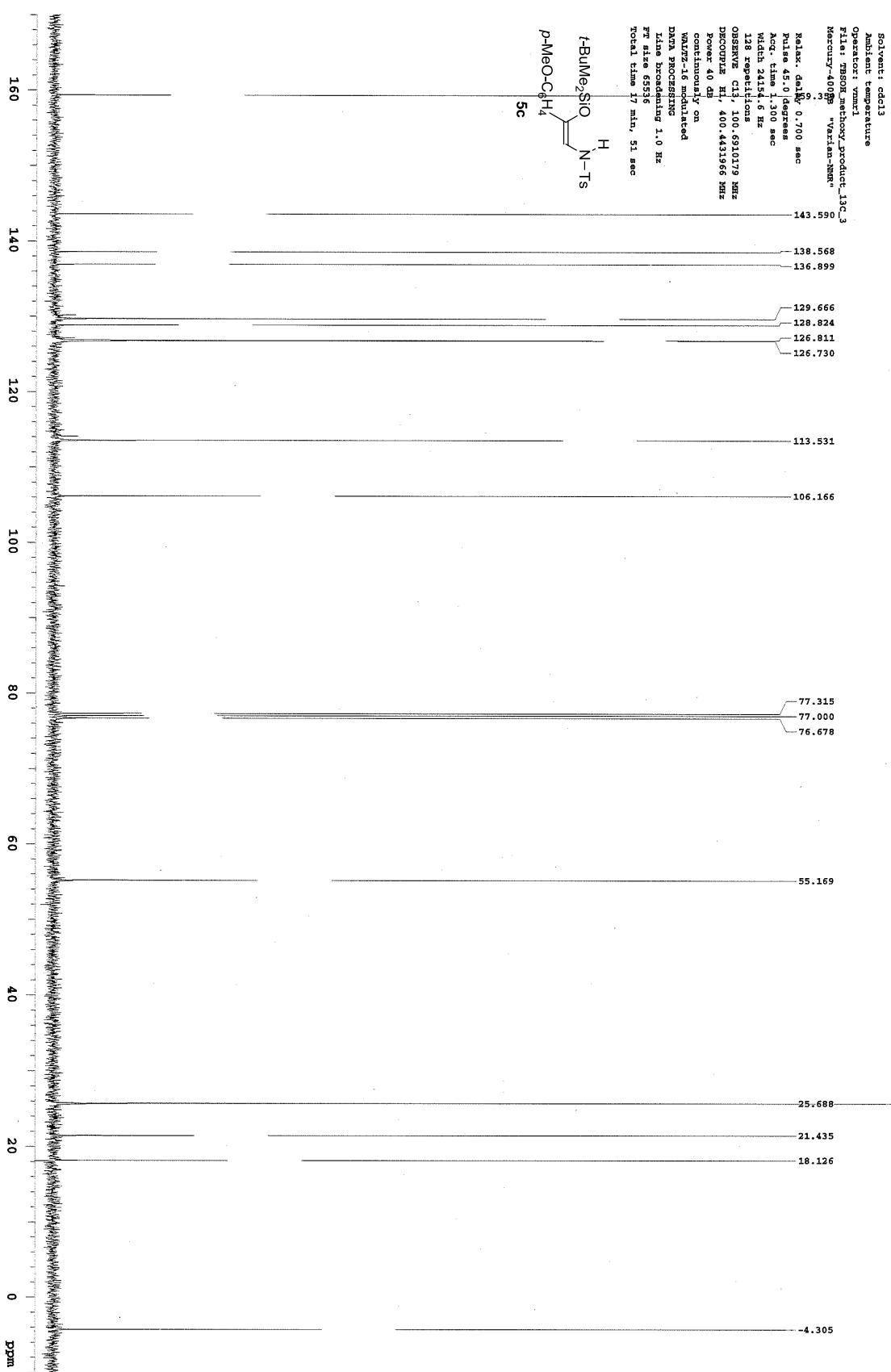
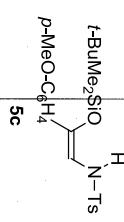
WAVEZ-16 uncalibrated

DATA PROCESSING

Lam. broadening 1.0 Hz

FT size 65536

Total time 17 min, 51 sec



File: /home/vmrc1/vmrcsys/data/murakami\_lab/nakamura/triazole\_Silanol/TBSOH\_trifluoro\_product.fid

Pulse Sequence: s2pul

Solvent:

Scout1: edc13

Ambient temperature

Operator: vmlci

File: TBSOH\_trifluoro\_product

Mercury-400BB "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 6402.0 Hz

16 repetitions

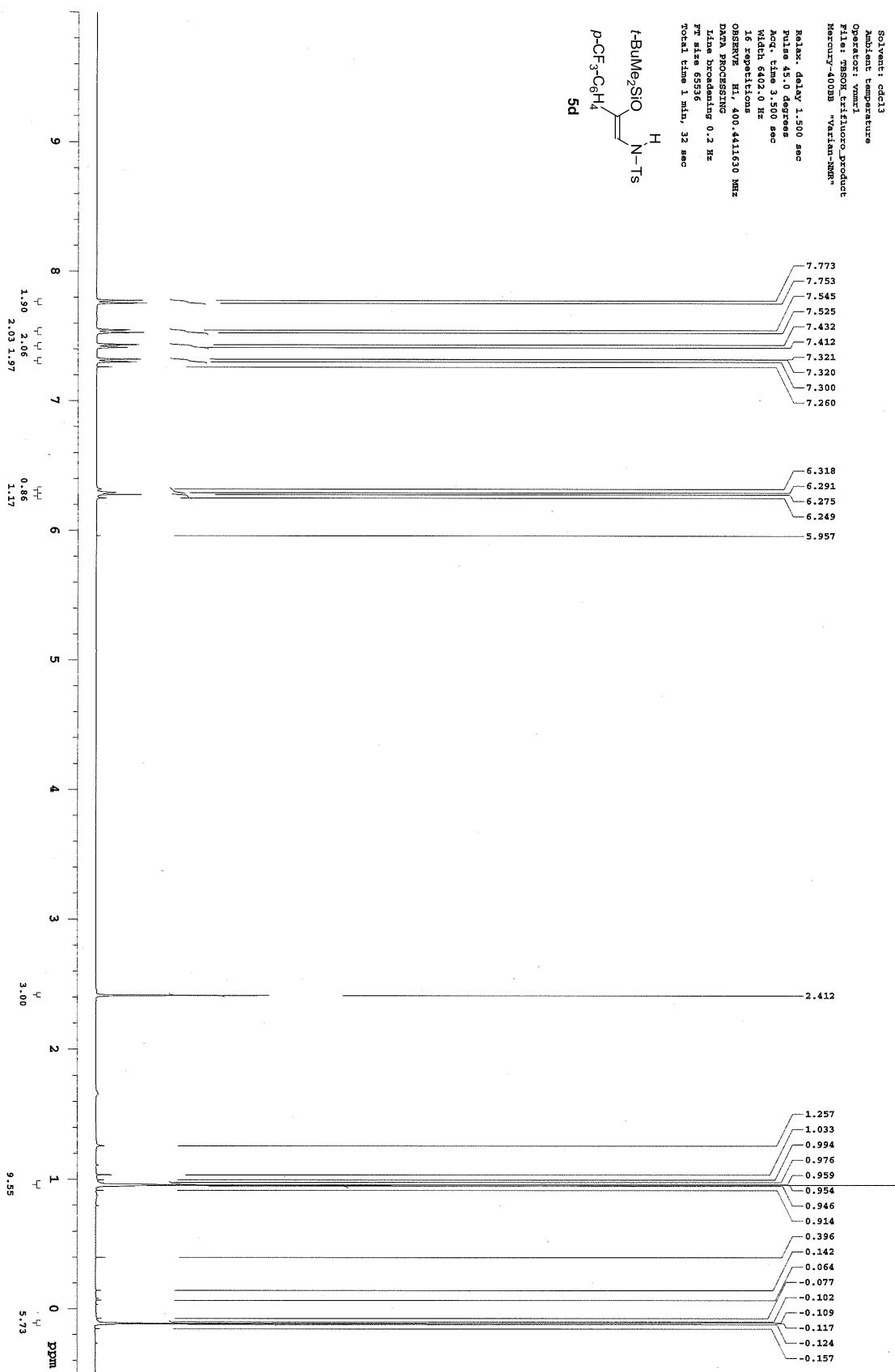
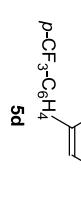
Observe: H1 400.4411610 MHz

Data Processing

Line broadening 0.2 Hz

FT size 65536

Total time 1 min, 32 sec



new experiment

File: /home/vmuzl/vmnsysa/data/murakami\_lab/Sakamuro/Triazole\_Silanol/BSO-trifluoro-product\_13C\_3.fida

Pulse Sequence: s2pul

Solvent: CDCl<sub>3</sub>

Ambient temperature

Operator: vmuzl

File: BSO-trifluoro-product\_13C\_3

Mercury-4010B "Varian"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 2414.6 Hz

15168 repetitions

OBSERVE C13, 100.691012 MHz

DECOPOL. H1, 400.4431986 MHz

Power 40 dB

continuously on

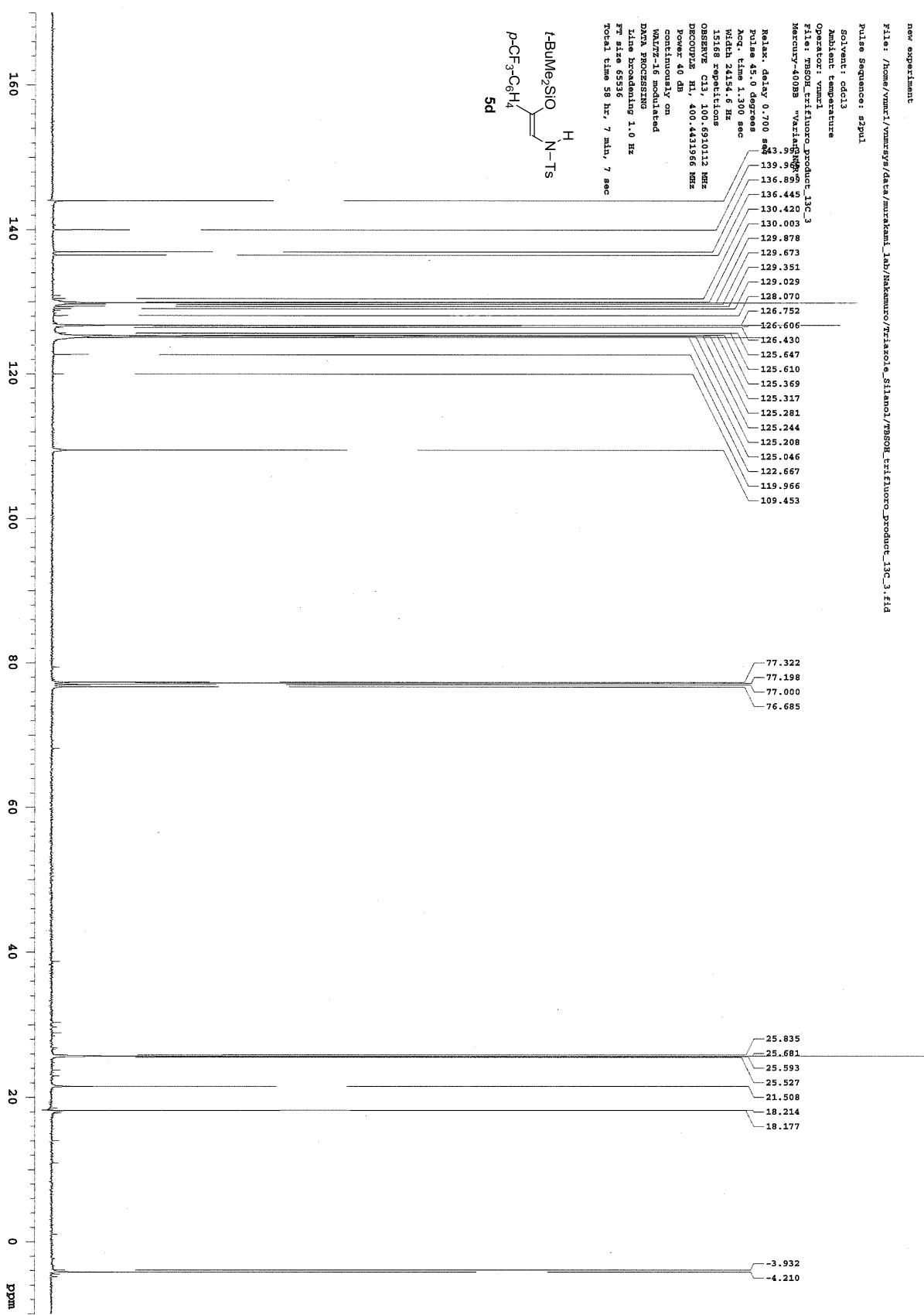
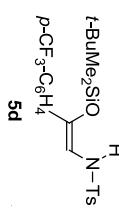
WAVEZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

FT size 65336

Total time 58 hr, 7 min, 7 sec



455 column

File: /home/vmrc1/vmcrcsys/data/murakami\_lab/Nakamoto/triazole\_Silanol/MSOR\_thienvl\_product\_1H\_exp455.fid

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

temp. 21.0 C / 294.1 K

Operator: vmcrc

File: MSOR\_thienvl\_product\_1H\_exp455

Mercury-400B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 640.0 Hz

16 repetitions

observe H, 400.4411632 MHz

DATA PROCESSING

Linn decoupling 0.2 Hz

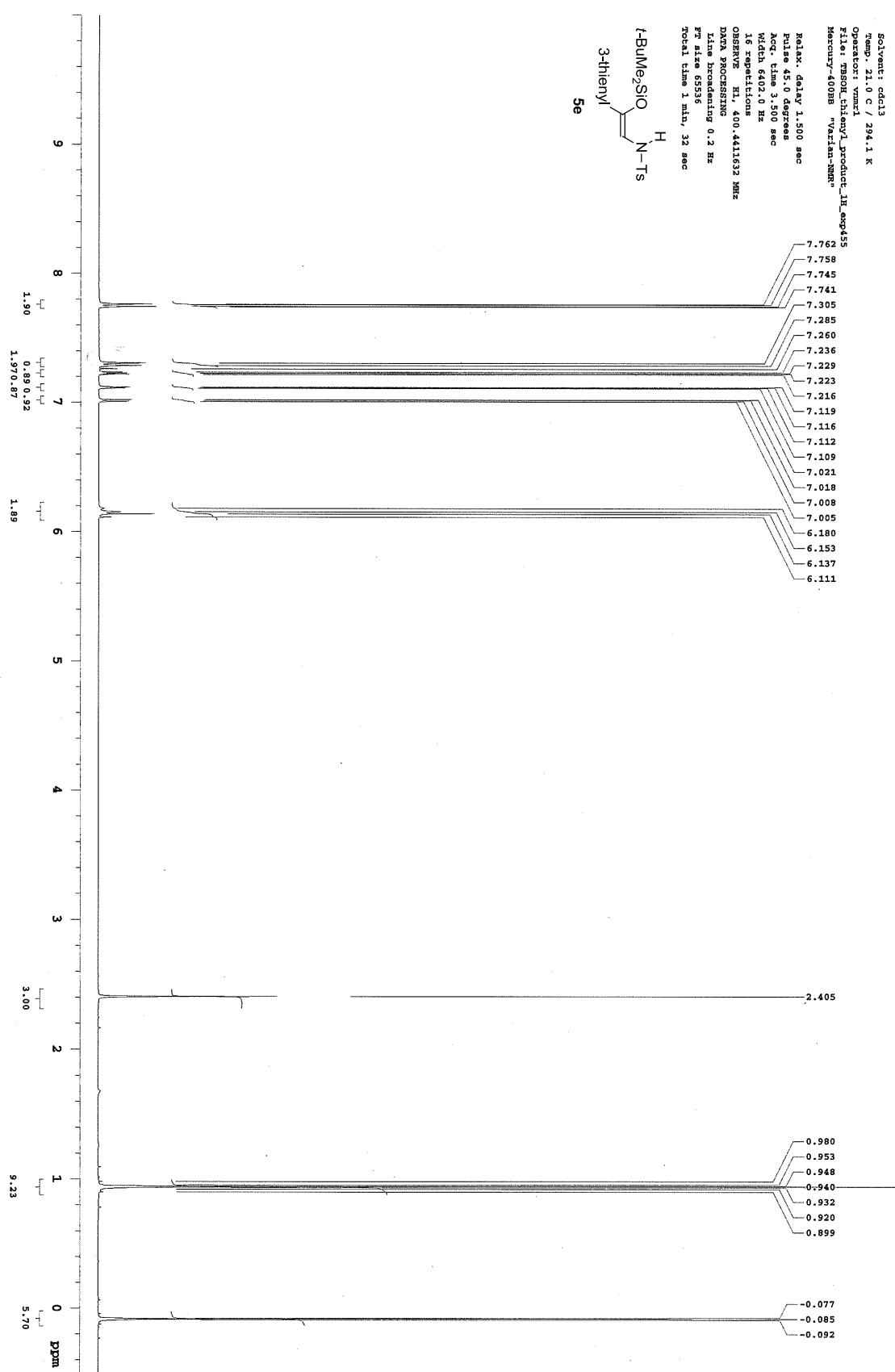
FT size 5556

Total time 1 min, 32 sec

Line broadening 0.2 Hz

Chemical shift 0.00 ppm

5e



Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vrml

Mercury-400BB "Varian-BBR"

Relax. delay 0.700 sec

Pulse 45.0 degrees sec

Acc. time 1.300 sec

Width 2414.6 Hz

256 acquisitions

OBSERVE C13 100.6910154 MHz

DECOPLE B1 400.4431966 MHz

Power 40 dB

continuously on

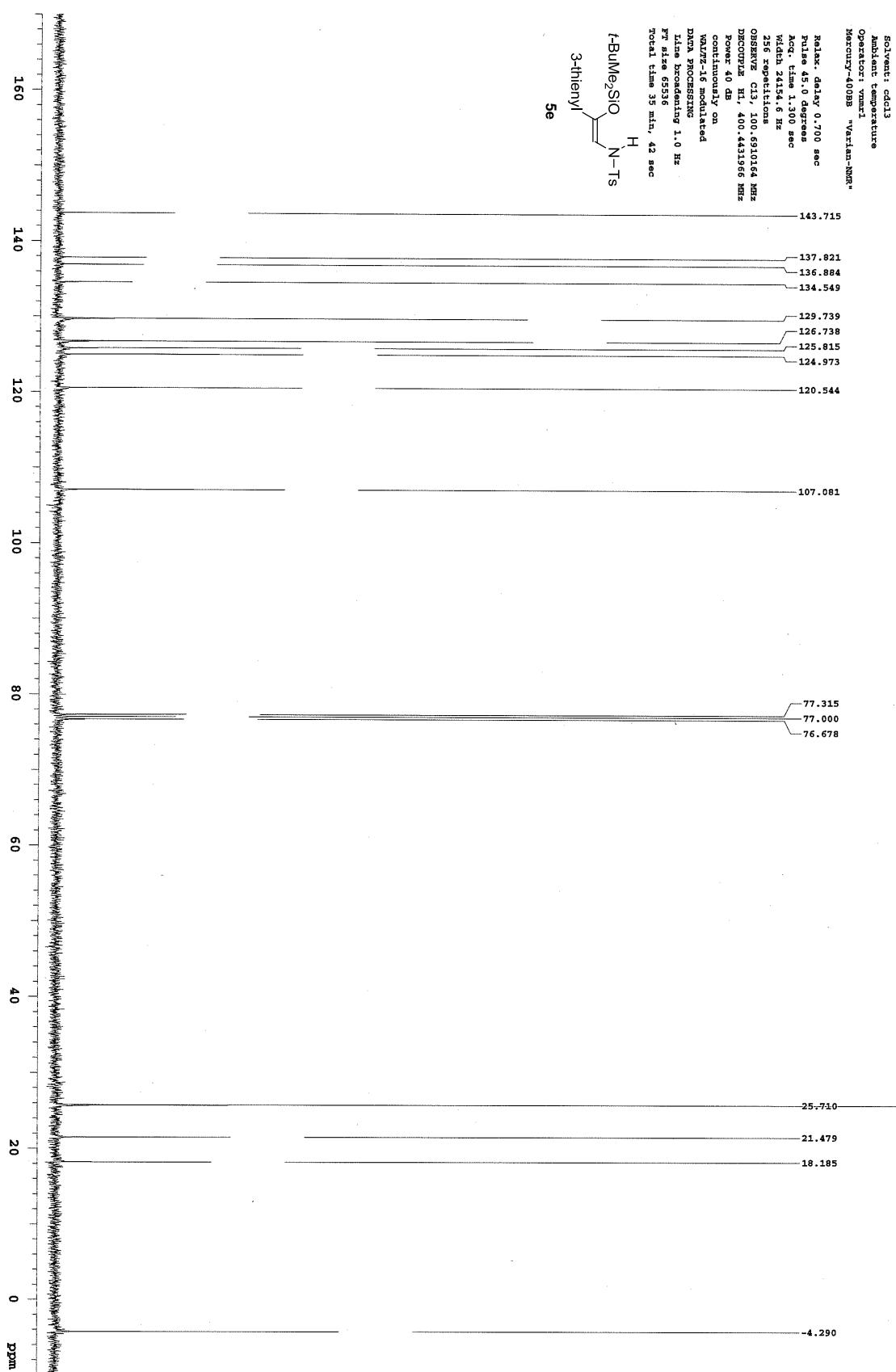
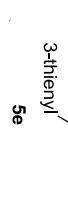
WAVE2:16 modulated

DATA PROCESSING

line broadening 1.0 Hz

FT size 65536

Total time 35 min, 42 sec





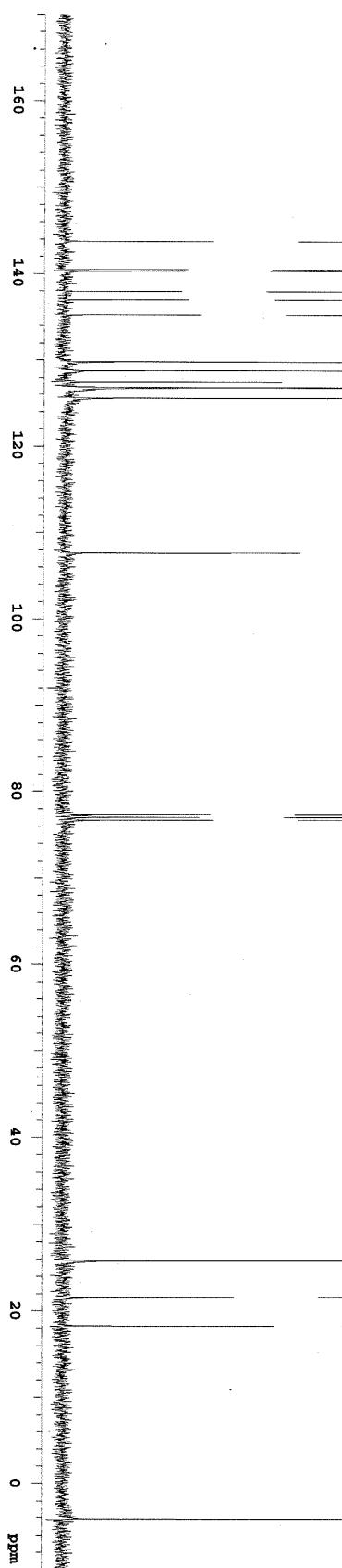
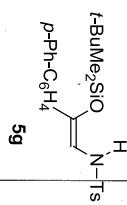
File: exp  
 Pulse Sequence: s2pul  
 Solvent: cdcl<sub>3</sub>  
 Ambient temperature  
 Operator: ymri  
 Marcell-400BB "Varian-NMR"

Relax. delay 0.700 sec  
 pulse 45.0 degrees  
 acc. time 1.300 sec  
 width 24154.6 Hz  
 128 experiments  
 observe cl3, 100.6910193 MHz  
 decouple h1, 400.4431966 mhz  
 power 40 dB  
 continuously on  
 waltz-16 modulated  
 data processing  
 line broadening 1.0 Hz  
 pw size 65536  
 total time 17 min, 51 sec  
 107.623

77.315  
 77.000  
 76.678

25.732  
 21.472  
 18.185

-4.202



File: /home/vmusr1/vmnrsys/data/murakami\_lab/Nakamura/Triazole\_Silanol/TBSOH\_CO2Et\_Product.fid

Pulse Sequences: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vml

File: MSOR-CORE-product

Mercury-400BB "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acc. time 3.500 sec

Width 6402.0 Hz

16 repetitions

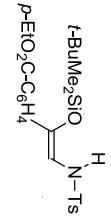
OBSERVE H1, 400.4411626 MHz

DATA PROCESSING

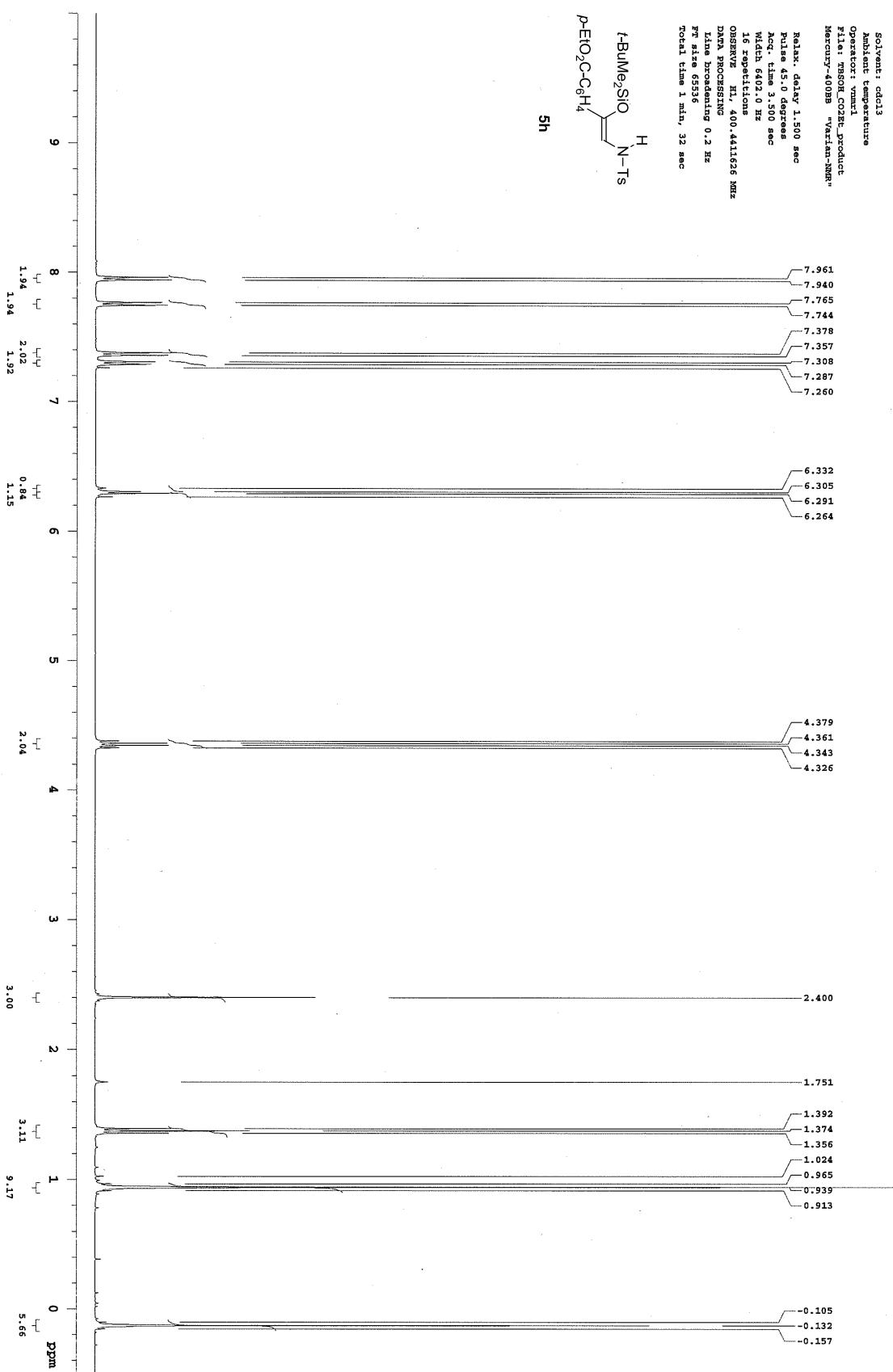
Line broadening 0.2 Hz

RT size 65536

Total time 1 min., 32 sec



5



File: /home/vmlc1/vmlcasya/data/murakami\_lab/nakamura/Triazole\_Silanol/TBSON\_CO2EC\_product\_13C.fid

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vmlc1

File: T200L.C02EC\_product\_13C

Memory=400KB "Varian-NMR"

1.1

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 2414.6 Hz

448 repetitions

observe C13 100.6910157 MHz

decouple H1 400.4433956 MHz

Power 40 dB

continuously on

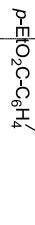
WAVE=16 modulated

DATA PROCESSING

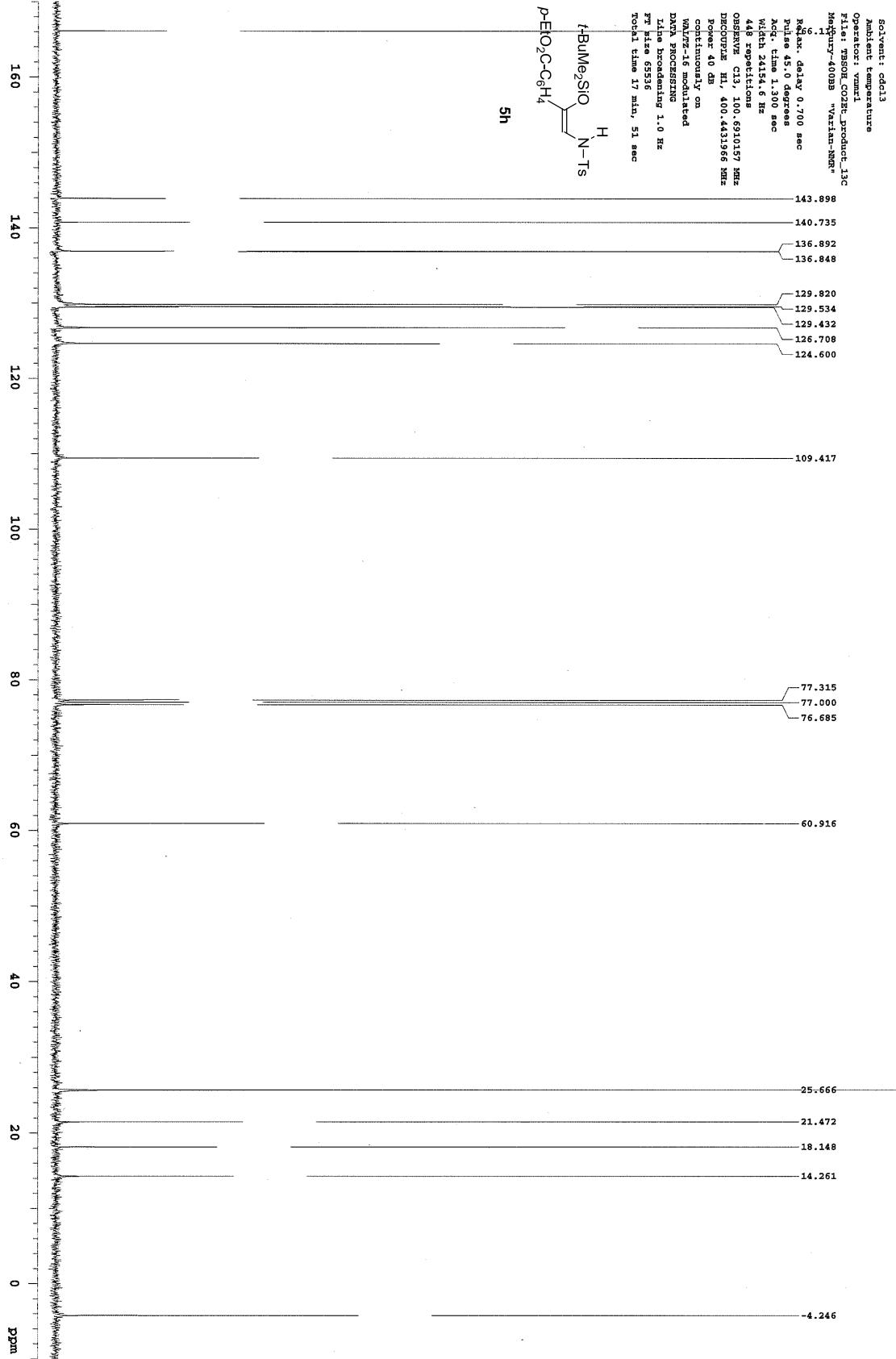
line broadening 1.0 Hz

FT size 65135

Total time 17 min, 51 sec



5h



1433 column

File: /home/vmnrl/vmnrsys/data/murakami\_lab/Nakamura/1433\_1H.fid

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vmnrl

File: 1433\_1H

Mercury-40 DB "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

width 640.0 Hz

16 repetitions

OBSERVE: H1, 400.441628 MHz

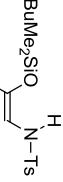
DATA PROCESSING

Line broadening 0.2 Hz

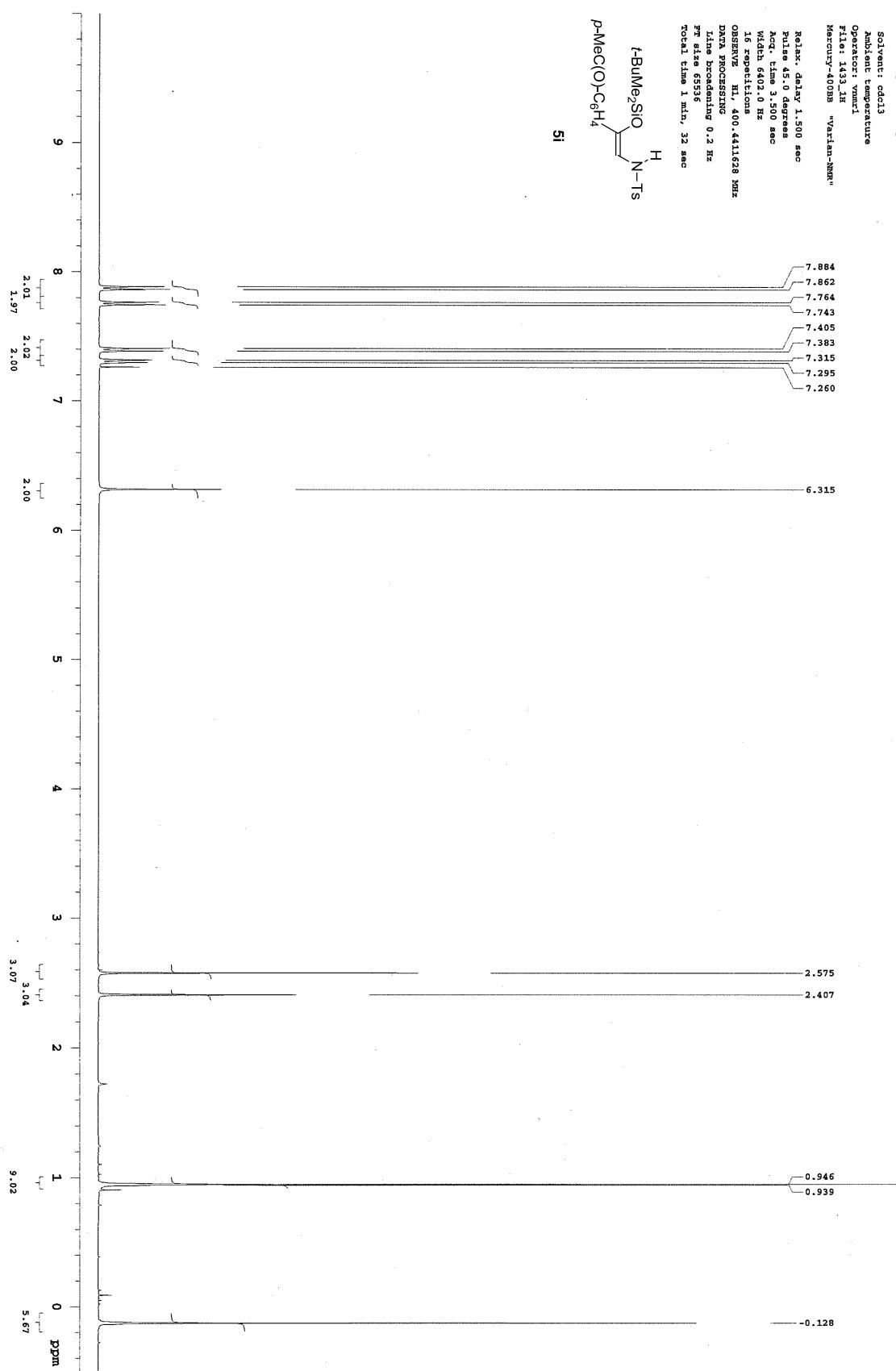
PP size 65536

Total time 1 min, 32 sec

*t*-BuMe<sub>2</sub>SiO



5i



File: /home/vmml1/vmmlsys/data/murakami\_1ab/Nakamura/1433-13C-4.fid

Pulse sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vml1

File: 1433-13C-4

Mercury-409BB "Varian-NMR"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 24154.6 Hz

7232 repetitions

OBSERVE C13, 100.651014 MHz

DECOPLEAR H1, 400.4431986 MHz

Power 41 dB

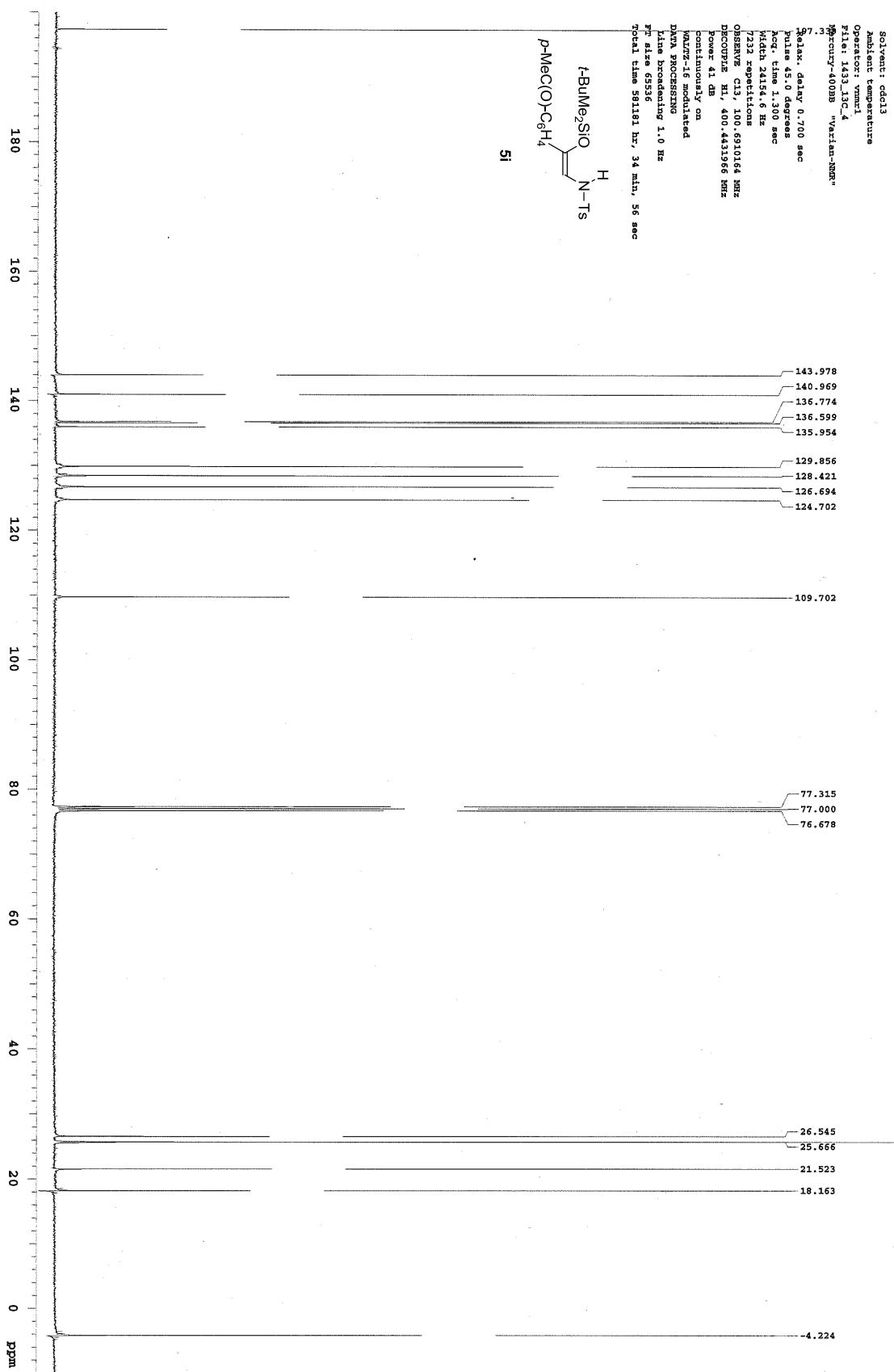
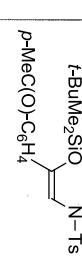
Continuously on

WAVEZ-16 modulated

LINE broadening 1.0 Hz

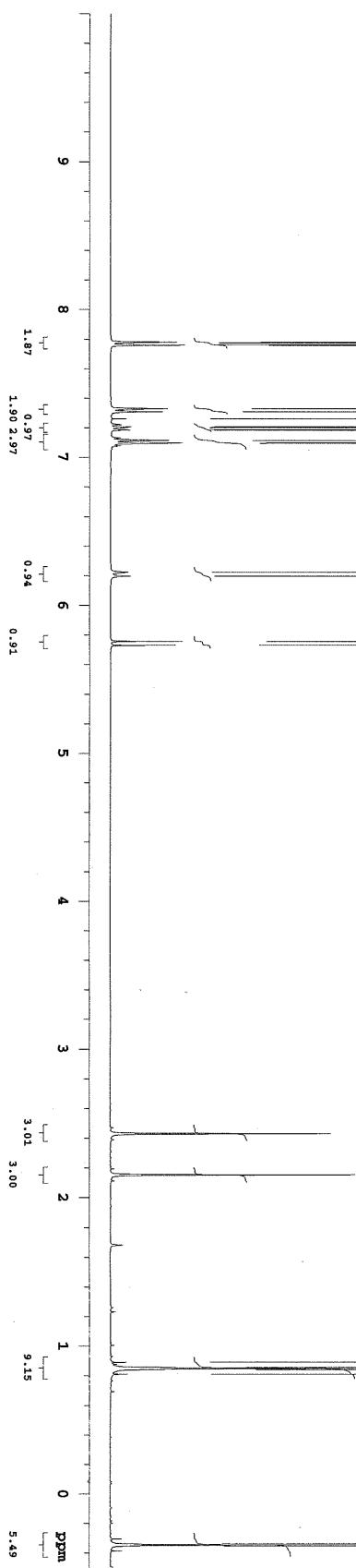
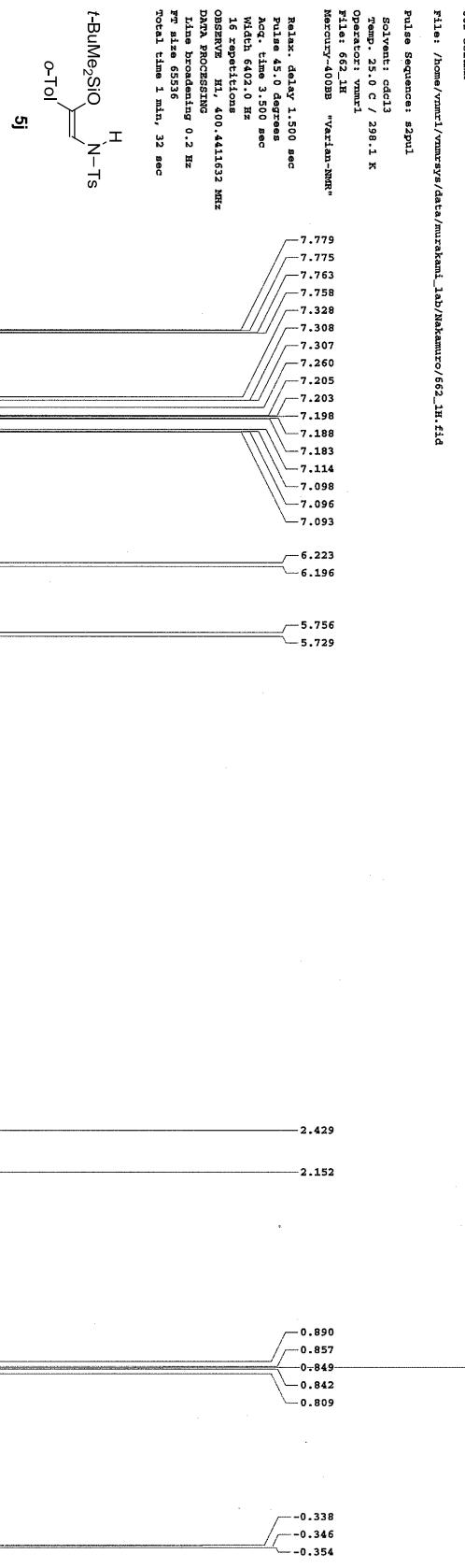
RF size 55336

Total time 581.161 sec, 34 min, 56 sec



**File:** /home/vmuru1/vmurusys/data/murakami\_lab/Nakamuro/662\_1H.fid

Pulse Sequence: s2pul  
 Solvent: cdd13  
 Temp.: 25.0 C 298.1 K  
 Operator: manual  
 File#: 002-1R "Varian-NMR"  
 Mercury-400BPP



662 column

file: /Home/vmuz1/vmuzsys/data/muzakami\_lab/Nakamoto/662\_13C.fid

Pulse Sequence: x2pul

Solvent: ccl4,3

Temp: 25.0 C, 298.1 K

Operator: vmuz1

File: 662\_13C

Mercury-400BB "Varian-MER"

Relax. delay: 0.700 sec  
Pulse 45.0 degrees  
Acc. time: 1.300 sec  
Width: 2414.6 Hz  
128 acquisitions

OBSERVE: C13, 100.6910171 MHz

DECODES: EL, 400.4431966 MHz

Power: 40 dB

continuously on

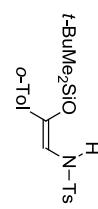
WAVZ-1K modulated

DATA PROCESSING

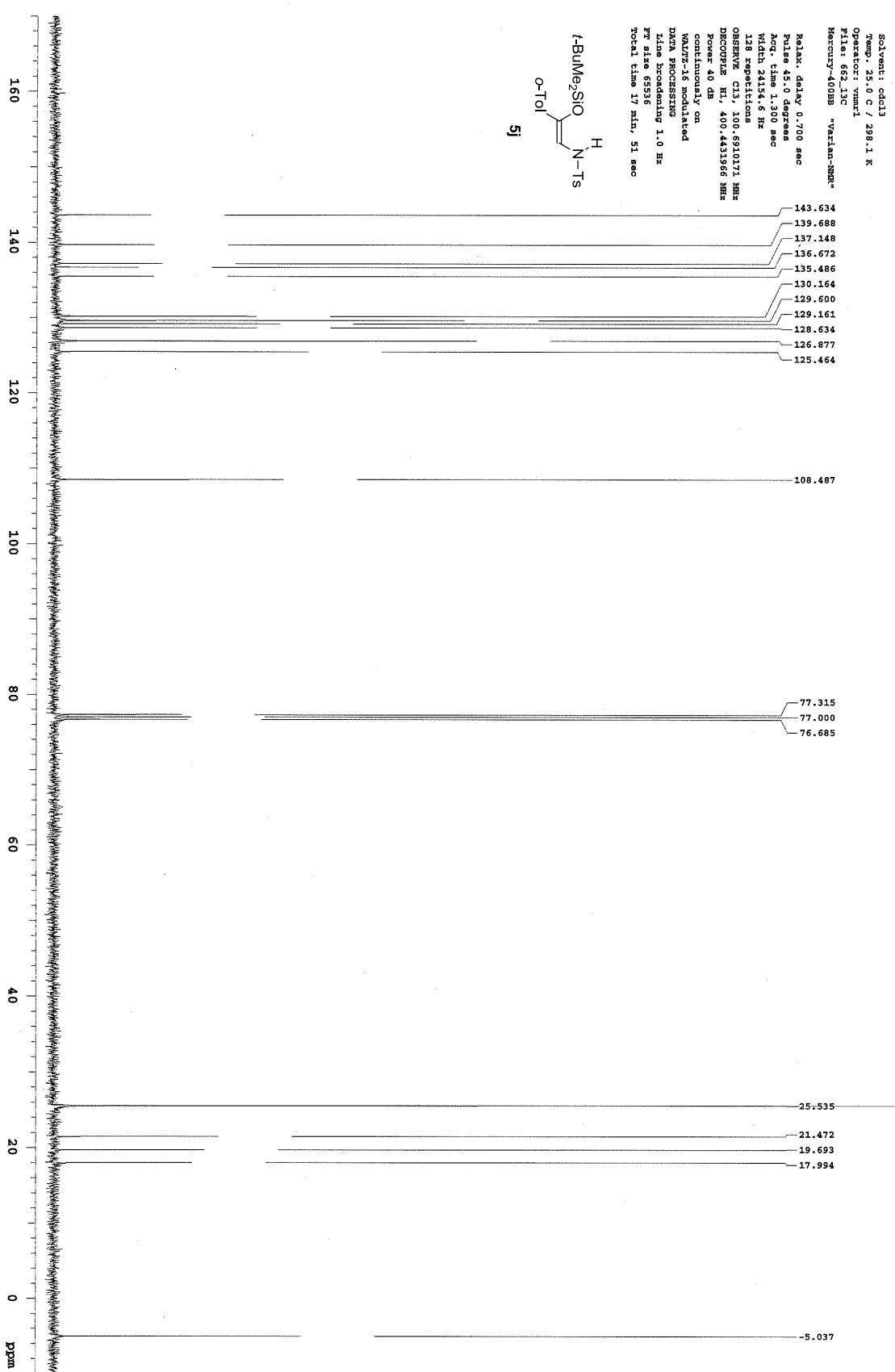
line broadening: 1.0 Hz

RT size: 65336

Total time: 17 min, 51 sec



5j



851 column  
File: /Home/vnmrl1/vnmrsys/data/murakami\_lab/Nakamura/851.fid

Pulse Sequence: s2pnl

Solvent: cdcl<sub>3</sub>

Temp: 24.0 C / 297.1 K

Operator: vnmrl1

File: 851  
Mercury-400B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 642.0 Hz

16 repetitions

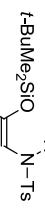
OBSERVE Hz, 400.441630 MHz

DATA PROCESSING

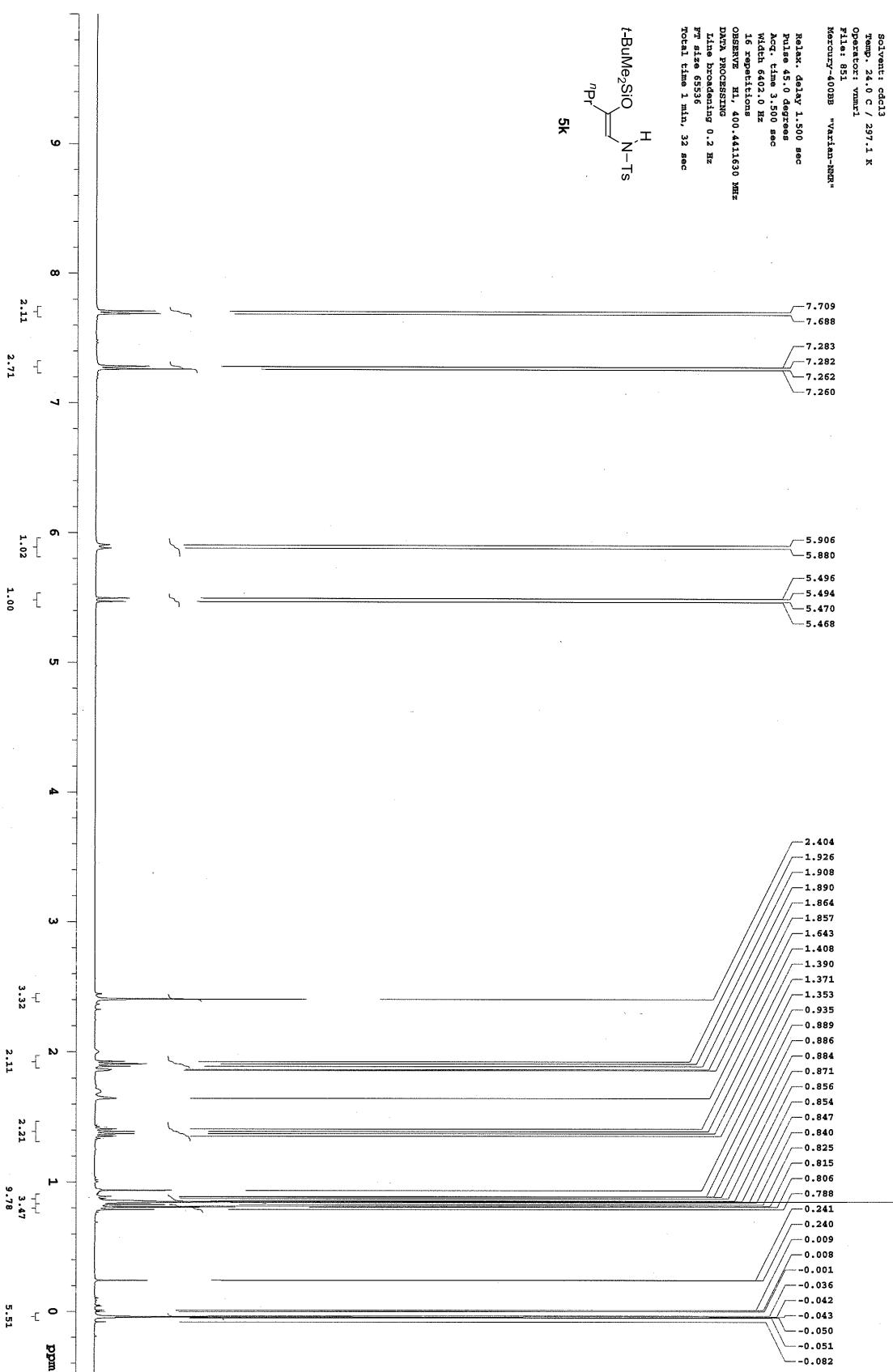
Line broadening 0.2 Hz

RT size 65536

Total time 1 min, 32 sec



5k



851 column

file: exp

Pulse Sequence: *s2pul*

Solvent: *cdcl3*

Temp. 24.0 C / 247.1 K

Operator: *vnmrl*

Mercury-400B "Varian-MER"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 24154.6 Hz

128 repetitions

OBSERVE C13, 100.6910127 MHz

DECOUPLE HI, 40.4431986 MHz

Power 4.1 dB

continuously on

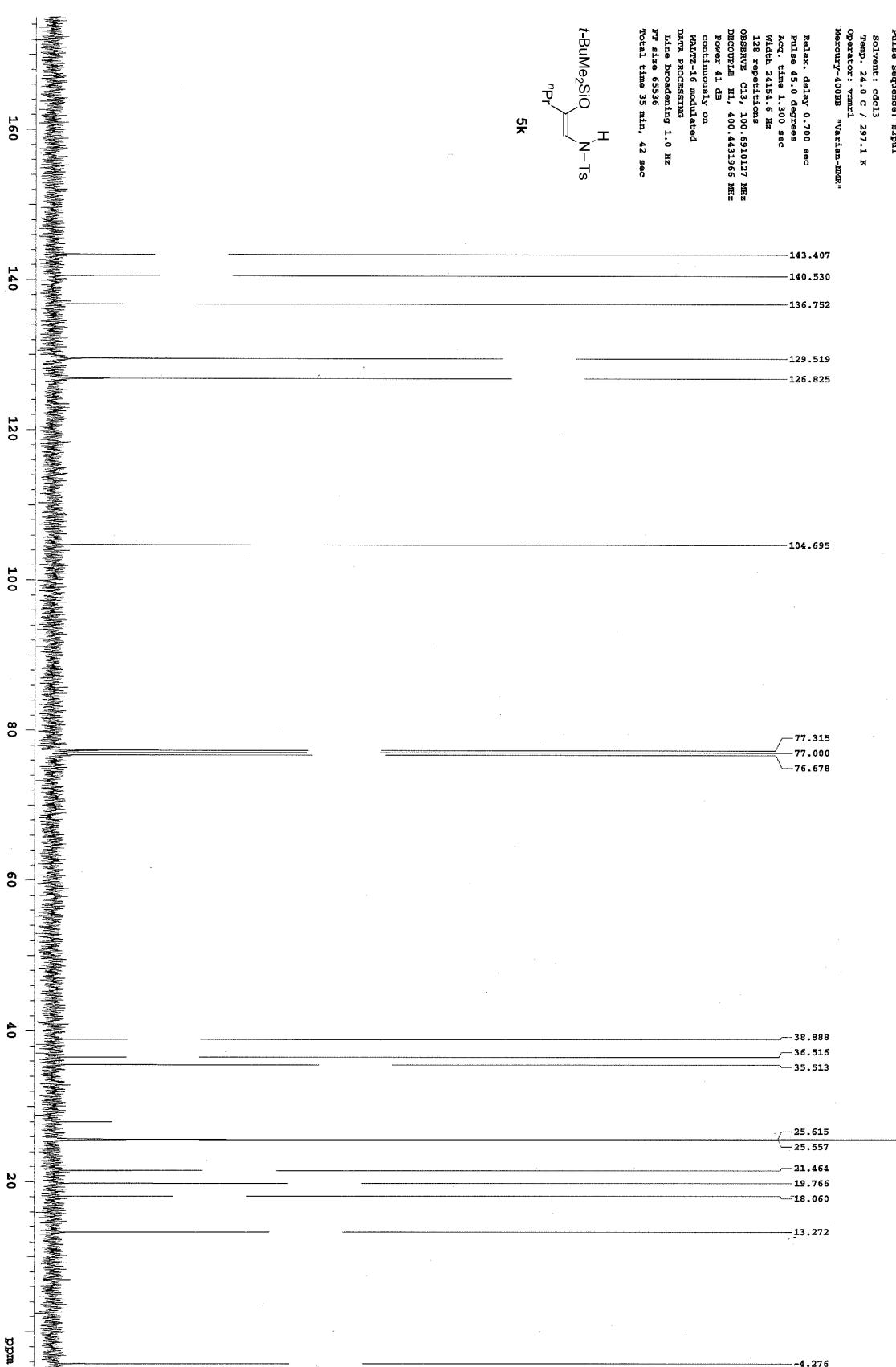
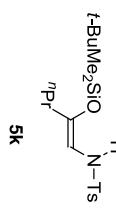
WAVEZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

FT size 65536

Total time 35 min., 42 sec



1411 column major

file: /home/vmml1/vmmls/sdata/murakami\_1b/Nakamuro/1411Lmajor\_1H.fid

Pulse sequence: 2-pul

Solvent: ccl4:13

Ambient temperature

Operator: vmml

file: 1411 major\_1H

Mercury-400BB "Varian-MER"

Relax. delay 1.500 sec

Pulse 45° degrees

Acq. time 3.500 sec

Width 600.0 Hz

16 replications

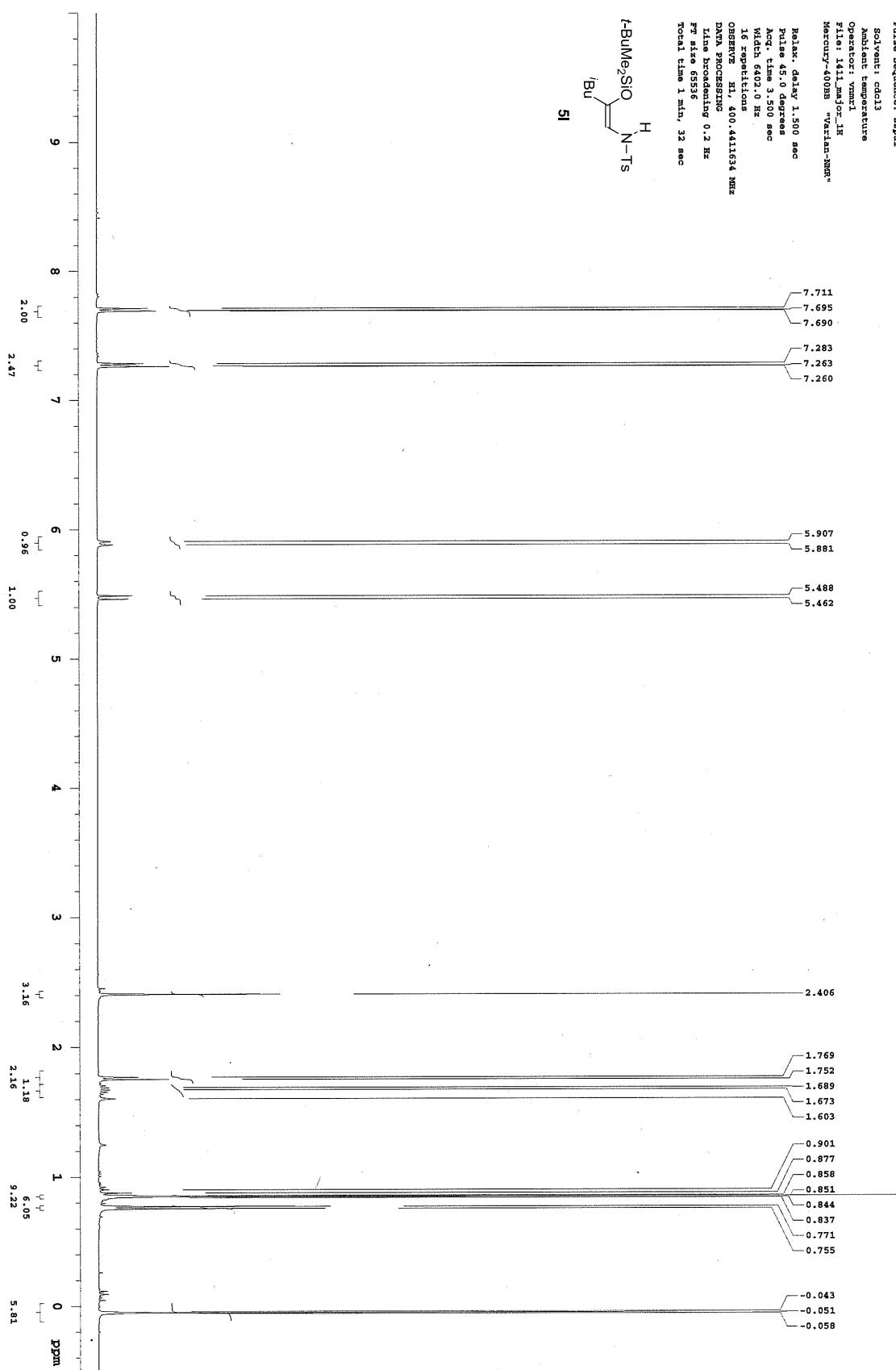
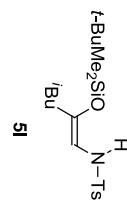
OBSERVE H1 400.4411634 MHz

DATA PROCESSING

line broadening 0.2 Hz

PPM size 65136

Total time 1 min, 32 sec



1411 column major

File: /Home/vmrx1/vmrxsys/data/murakami\_lab/Nakamoto/1411\_major\_13C.fid

Pulse Sequence: s2pul

Solvent: cdcl3

Ambient temperature

Operator: vml1

Date: 1411-08-15C

Mercury-400BB "Varian-NMR"

Relax. delay 0.700 sec  
Pulse 45.0 degrees  
Acq. time 1.300 sec  
Width 24.154.6 Hz  
192 repetitions  
Observe c13, 100.6910134 MHz  
Decouple H1, 400.4431956 MHz  
Power 41 dB  
continuously on

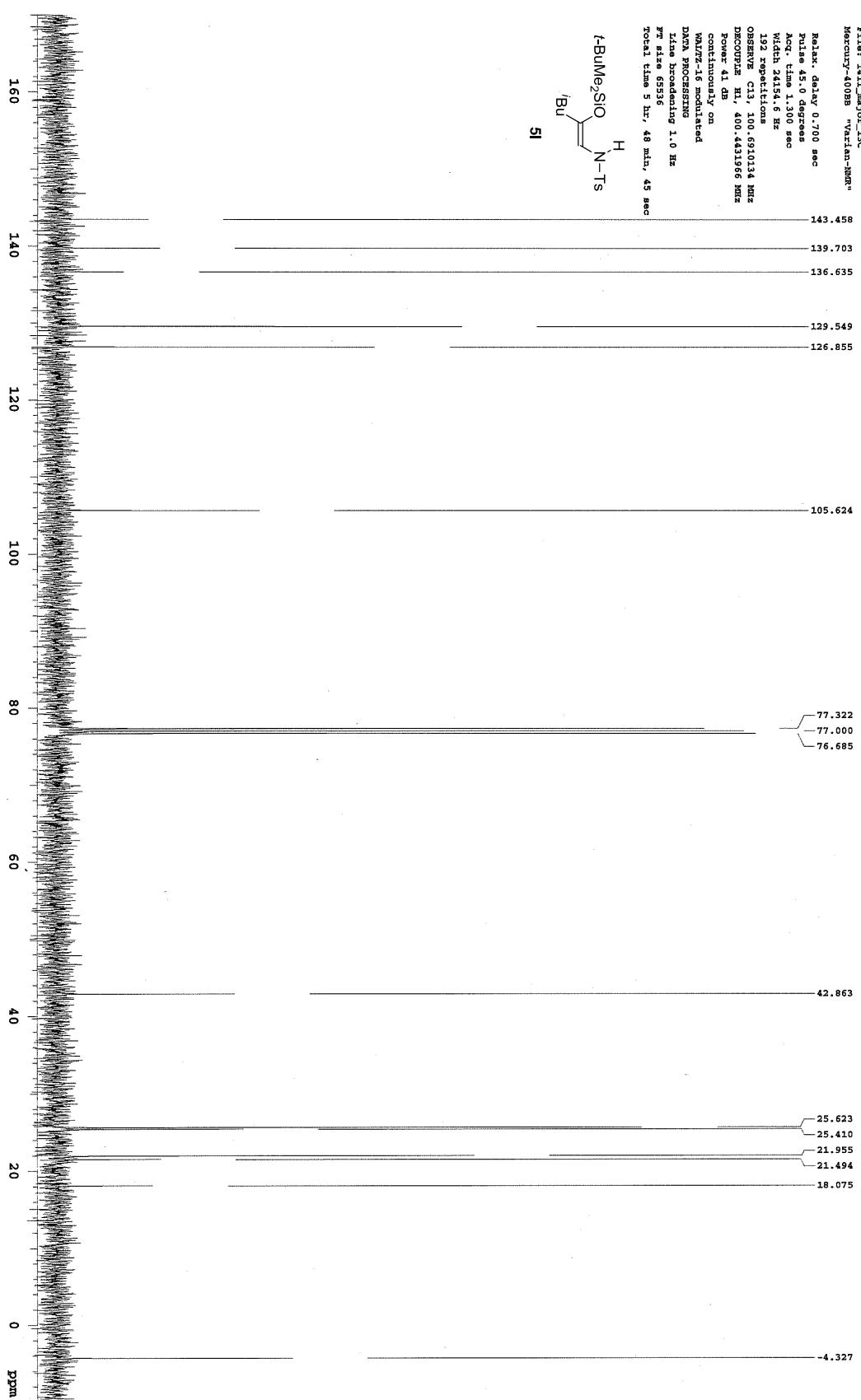
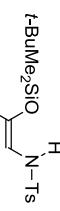
WIDFT=16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

FFT size 65536

Total time 5 hr, 48 min, 45 sec



1412 column

File: /home/vmxml/vmrssys/data/murakami\_lab/Nakamuro/1412\_1H.fid

Pulse Sequence: s2pul

Solvent: cdcl3

Operator: vimrl

Mercury-400BB "Varifan"

8.

Pulse 45.0 degrees

Width 6402.0 Hz

18 Repetitions

DATA PROCESSING

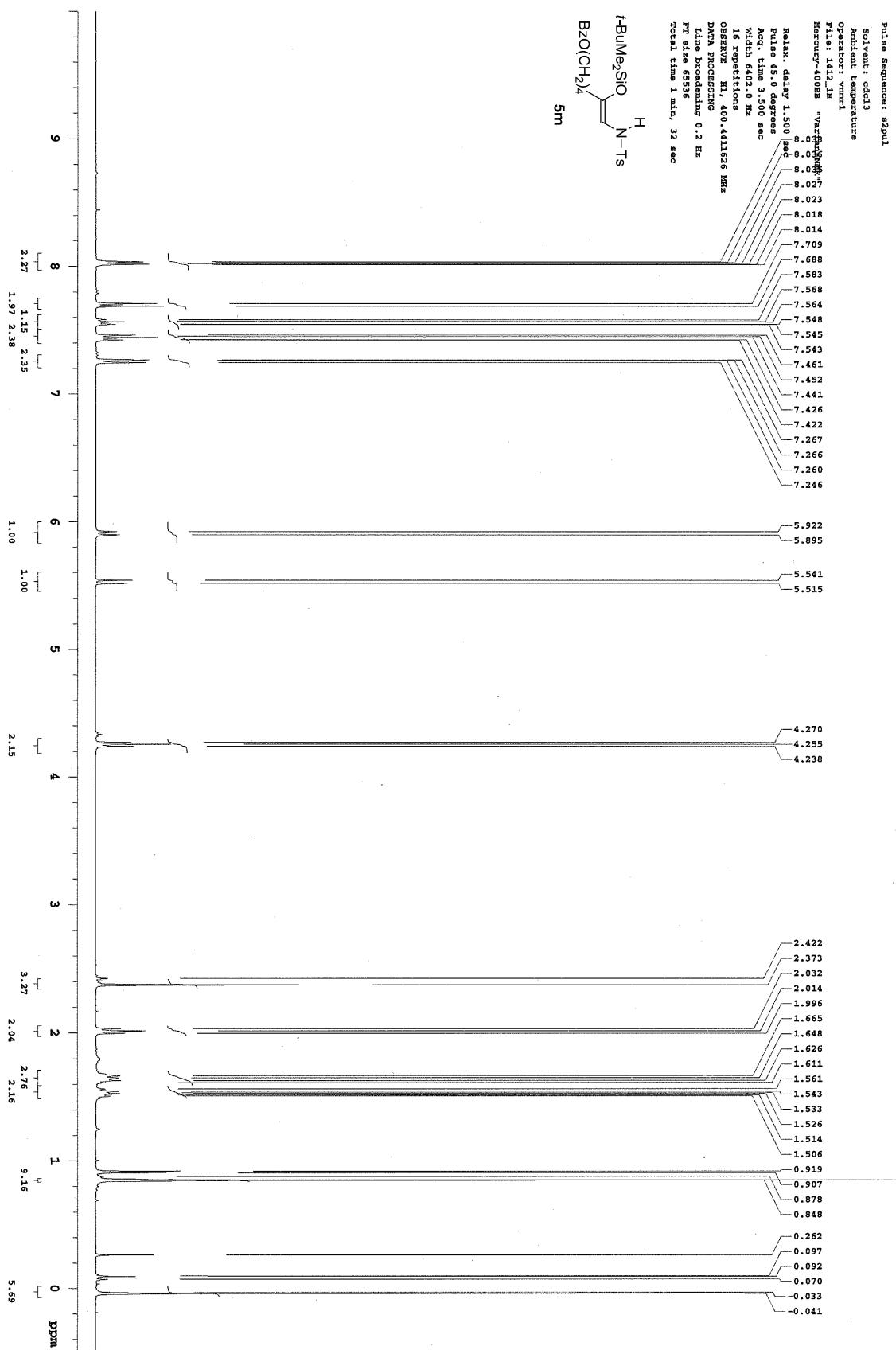
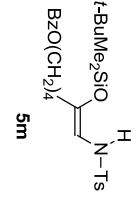
FT size 65536

工

i-BuMe<sub>2</sub>SiO

BORGH

$$\text{BzO}(\text{CH}_2)_4-\text{C}=\text{CH}-\text{N}-\text{Ts}$$



1412 column

File: /home/vmrc1/vmrcsys/data/murakami1.lab/Nakamuro/1412-13C-2.fid

Pulse Sequence: a2pul

Solvent: cde13

Ambient temperature

Operator: vmarl

file: 1412-13C-2

McQuay-400BBS "Varian-MERK"

5

Relax. delay 0.700 sec

Pulse 45.0 degrees

Ring time 1.300 sec

Whch 21134.5 Hz

512 repetitions

OBSERVE C13, 100.6910142 MHz

DECOUPLE H1, 400.4431986 MHz

Power 41 dB

continuously on

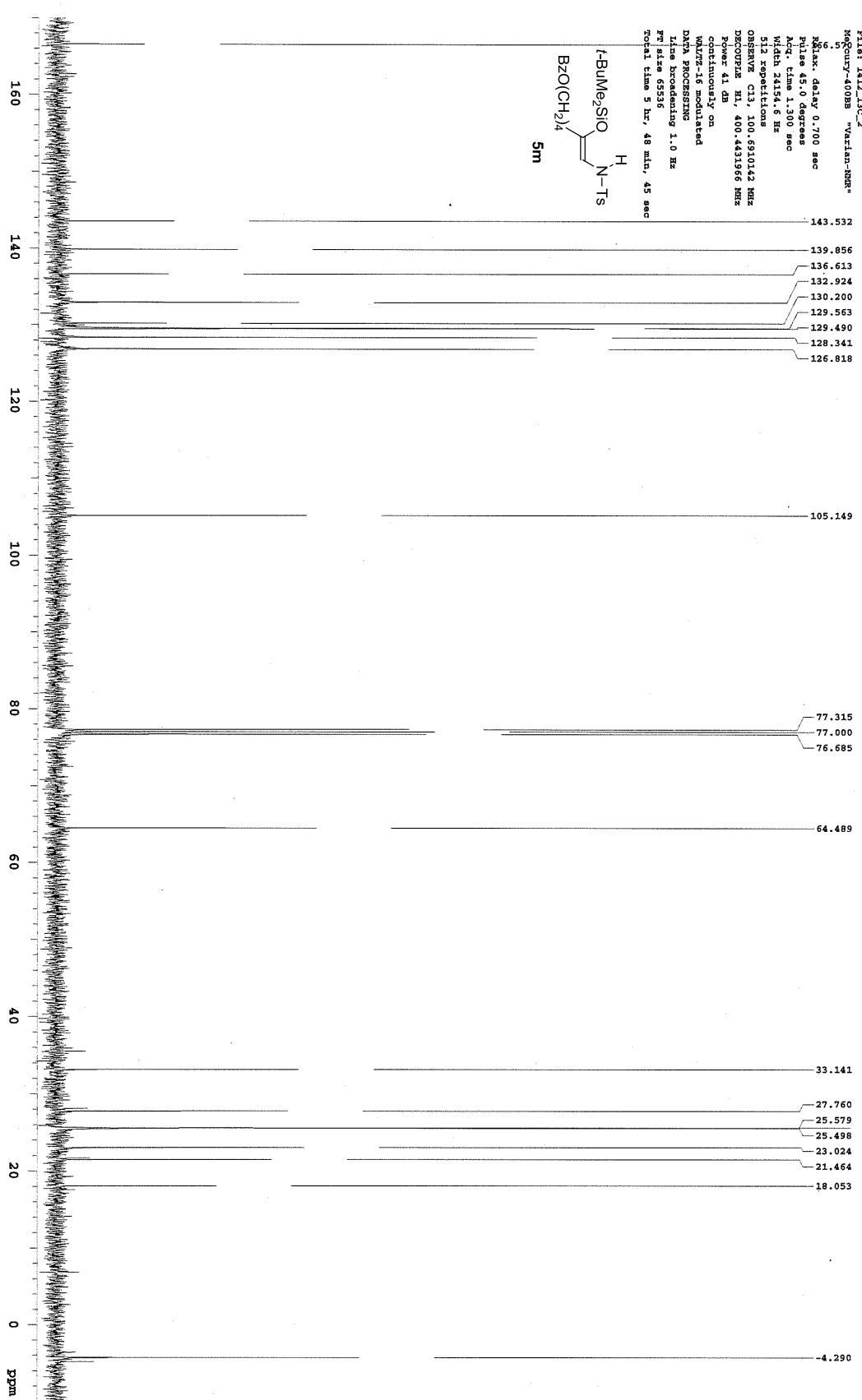
VOLUME=1 modulated

DATA PROCESSING

Line decoupling 1.0 Hz

FT size 65536

Total time 5 hr., 48 min., 45 sec



654 column  
file: /home/vmnrt1/vmnrtsys/data/murakami\_lab/Nakamuro/654\_1H.fid  
Pulse Sequences: a2spin

Solvent: dcd13

Temp. 23.0 C / 298.1 K

Operator: vmnrt1

File: 654\_1H

Mercury-4098B "Varian-MER"

Relax. delay 1.500 sec  
Pulse 45.0 degrees  
Acq. time 3.500 sec  
Width 6.02.0 Hz  
16 replications

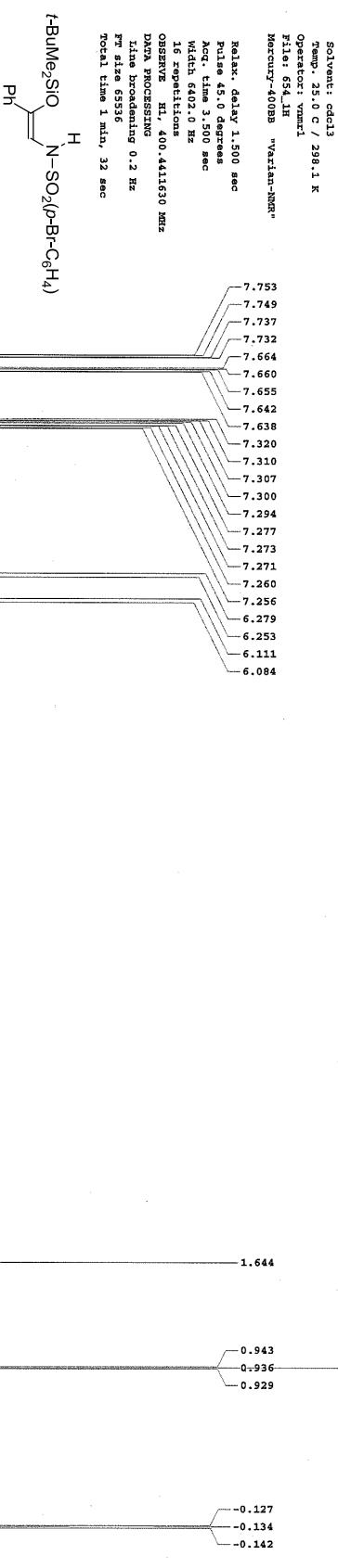
OBSERVE H, 400.4411630 MHz

DATA PROCESSING

Line broadening 0.2 Hz

FFT size 65336

Total time 1 min, 32 sec



5f

654 column  
File: /home/vnmrl1/vnmrsys/data/nakamaki\_lab/nakamuro/654-13c.zfd

Pulse Sequence: r2gppd1

Solvent: cdc13

Temp: 25.0 C - 298.1 K

Operator: vnmrl1

file: 654-13C

Mercury=000003 "Varian-NMR"

Relax: delay 0.700 sec

pulse 45.0 degrees

Acq. time 1.300 sec

Width 2414.6 Hz

128 repetitions

OBSERVE C13, 100.6910164 MHz

DECOUPLER H1, 400.4431966 MHz

Power 40 dB

continuously on

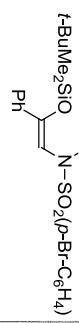
WALTZ-16 modulated

DATA PROCESSING

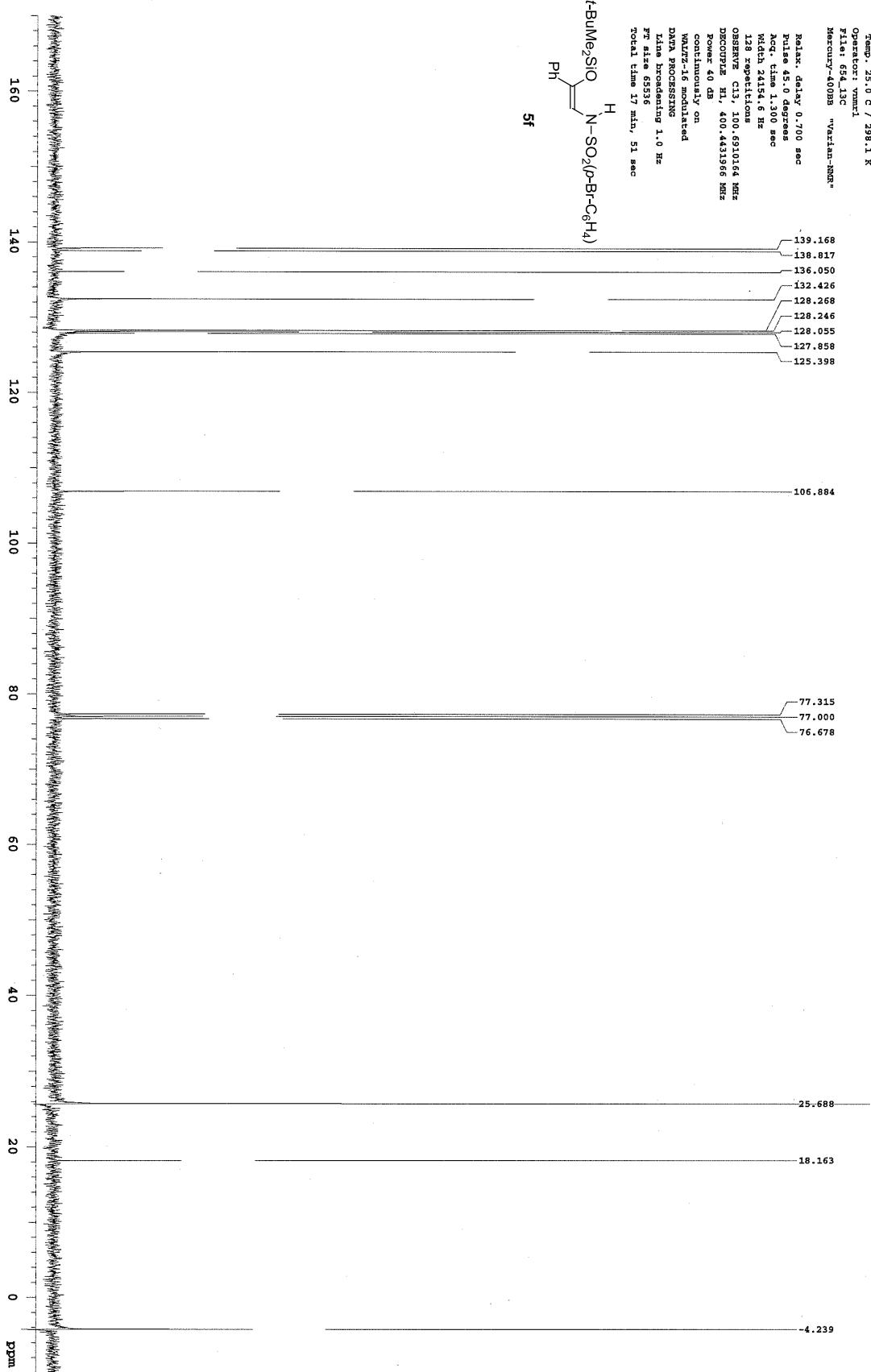
LINE BROADENING 1.0 Hz

FT size 65536

Total time 17 min., 51 sec



5f





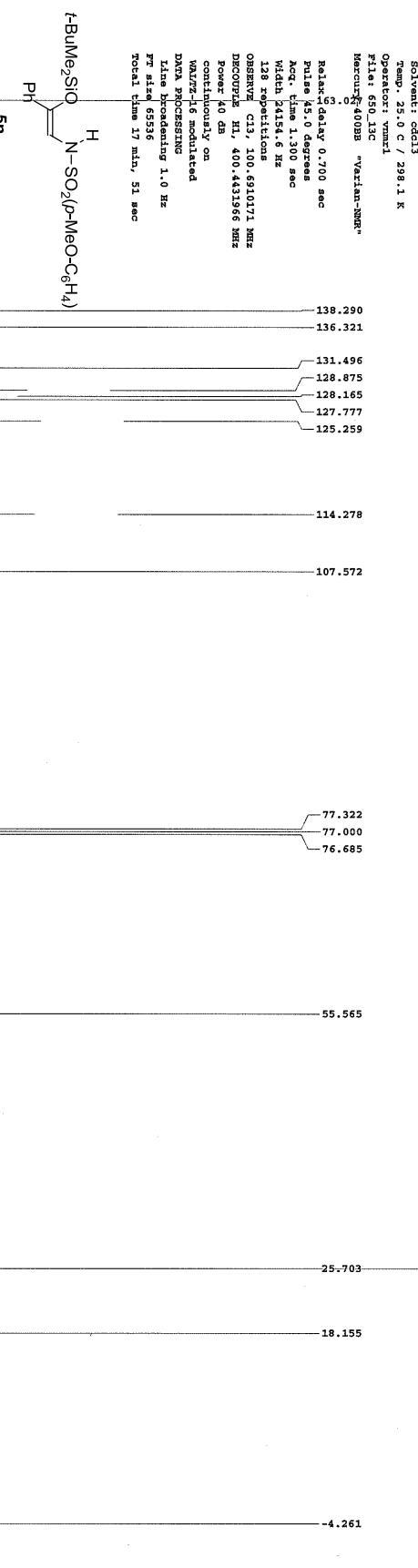
650 column  
File: /home/vmmt1/vmmr/sys/data/murakami\_lab/Nakamura/650\_13C.fid  
Pulse Sequence: ap2p1

Solvent: cdcl<sub>3</sub>  
Temp: 22.0 C / 298.1 K  
Operator: vmmr1  
File: 650\_13C  
Mercury: 300BB "Varian-MER"

Relax1: 1.0 sec  
Relax2: 0.700 sec  
Pulse: 45.0 degrees  
Acq. time: 1.300 sec  
Width: 2414.6 Hz  
128 repetitions  
OBSERVE: C13, 100.6910171 MHz  
DECOUPLE: H1, 400.4431966 MHz  
Power: 40 dB  
continuously on  
WALTZ-16 modulated

DATA PROCESSING:  
Line broadening 1.0 Hz

FT size: 65536  
Total time: 17 min, 51 sec



6.49 column

File: /home/vnmri/vnmr1/vnmr1sys/data/murakami.lab/murakami/6.49\_1H.fid

Pulse Sequence: a2pul

Solvent: cdcl<sub>3</sub>

Temp: 21.0 C / 298.1 K

Operator: vnmr1

File: 64\_1H "Varian-NMR"

Mercury-4010B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 640.0 Hz

16 repetitions

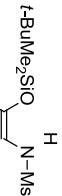
OBSERVE H1, 400.4411634 MHz

DATA PROCESSING

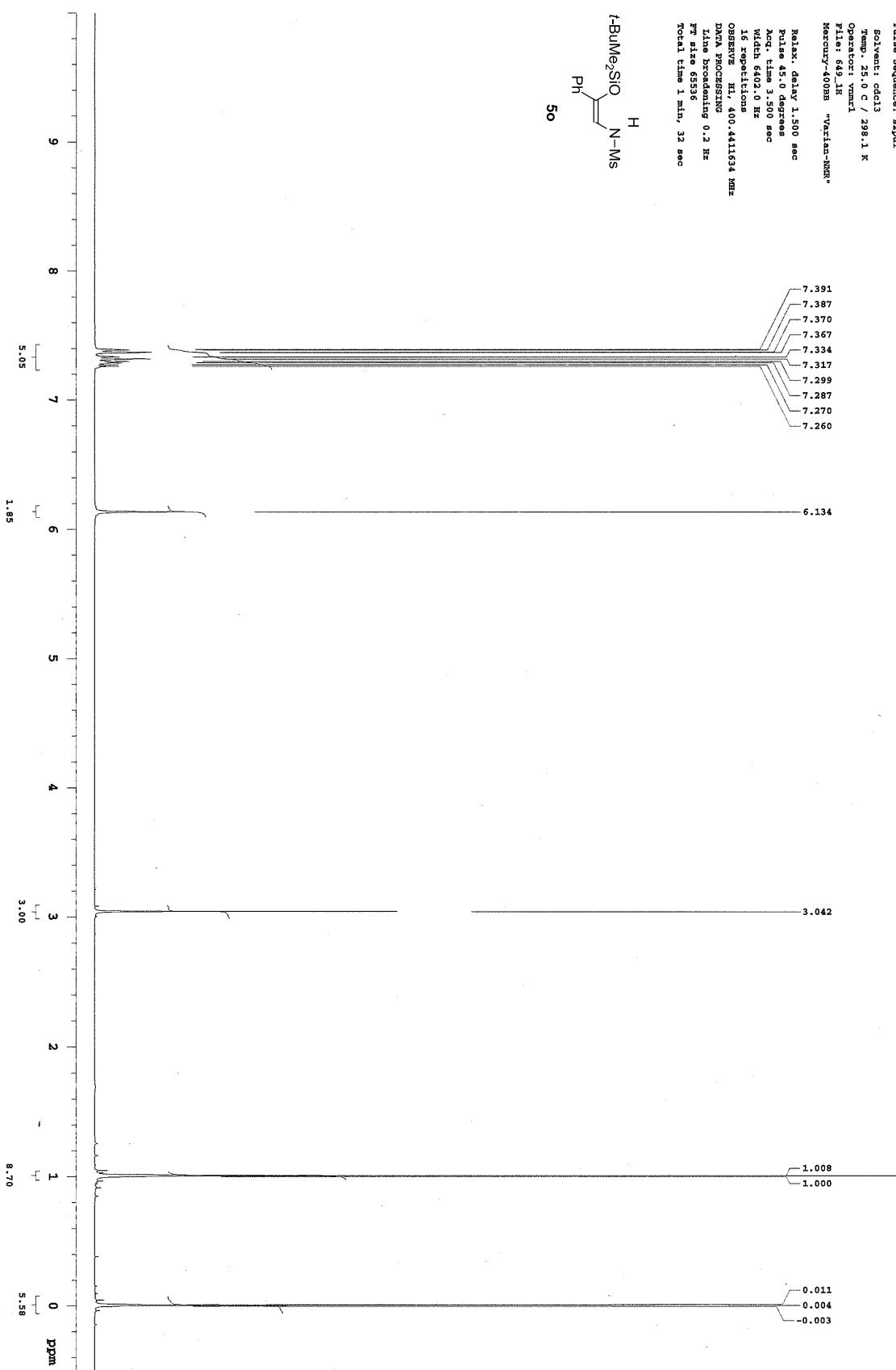
Line broadening 0.2 Hz

RT size 65136

Total time 1 min, 32 sec



50



649 column

File: /home/vmnri1/vmnrisys/data/murakami\_lab/Nakamura/649\_13C.fid

Pulse Sequences: s2pul

Solvent: ddc13

Temp. 22.0 C / 298.1 K

Operator: vmnri

File: 649\_13C

Mercury-40mB "Varian-MER"

Relax: delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 2114.6 Hz

128 repetitions

OBSERVE C13, 100.6910157 MHz

DECOUPLE H1, 400.4431986 MHz

Power 40 dB

continuously on

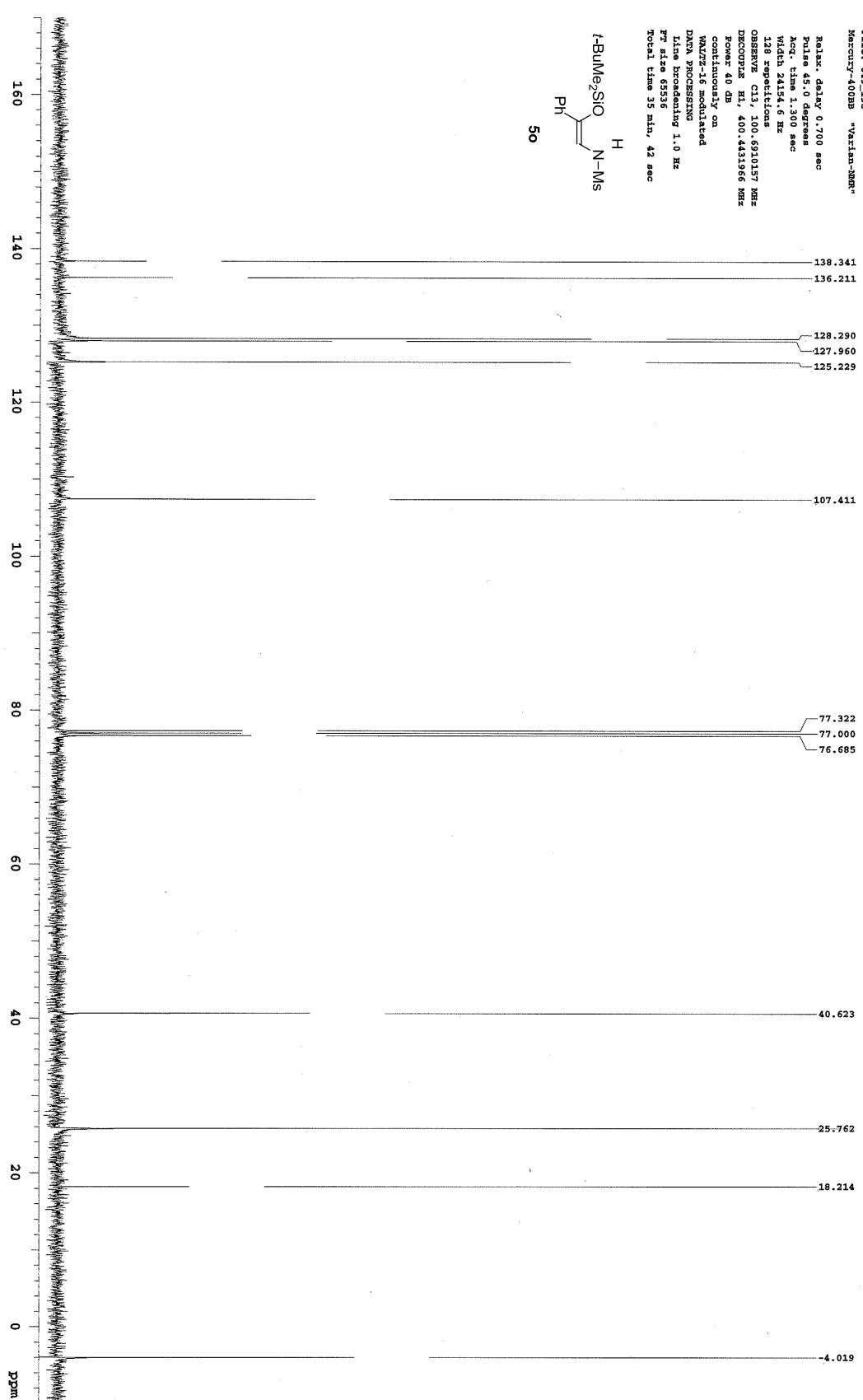
WAVEZ-15 modulated

DATA PROCESSING

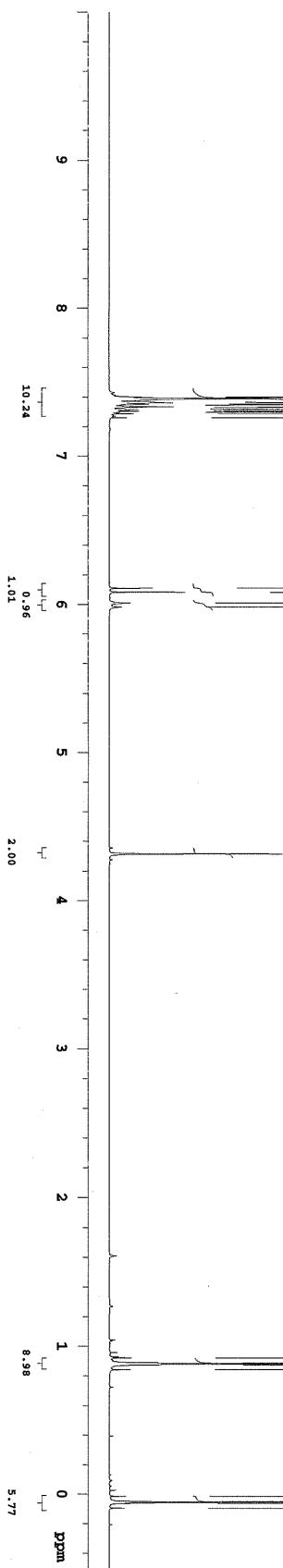
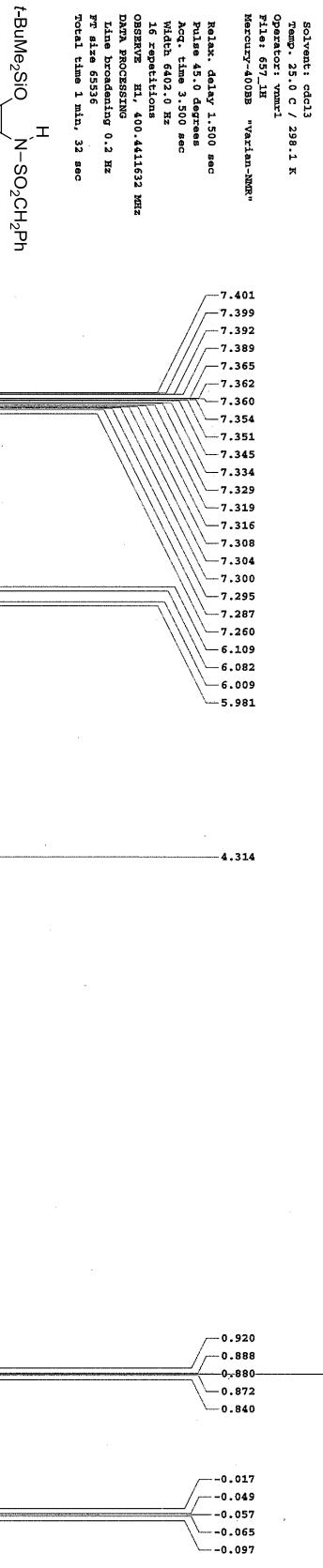
Line broadening 1.0 Hz

FFT size 65536

Total time 35 min., 42 sec



65<sup>7</sup> column  
 File: /home/vmrci/vmrcisys/data/murakami.lab/Nakamura/65<sup>7</sup>.1H.fid  
 Pulse sequence: a2pul1  
 Solvent:  $\text{CDCl}_3$   
 Temp: 22.0 C / 298.1 K  
 Operator: vmlml  
 File: 65<sup>7</sup>.1H  
 Mercury-400B3 "Varian-MER"  
 Relax: delay 1.500 sec  
 Pulse 45.0 degrees  
 ACG time 3.500 sec  
 Width 6.02, 0 Hz  
 16 replications  
 OBSERVE H: 400.4411632 MHz  
 DATA PROCESSING  
 Line broadening 0.2 Hz  
 FT size 65336  
 Total time 1 min, 32 sec



65<sup>7</sup> column  
File: /home/vmml1/vmmlsys/data/murakami/lab/Nakamura/65<sup>7</sup>-13C.fid  
Pulse sequence: m2pul

Solvent: ccd13

Temp. 22.0 C / 298.1 K

Operator: vmlr1

File: 65<sup>7</sup>-13C

MERCURY-400BB "Varian-NMR"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Watch 2.114.6 Hz

128 repetitions

OBSERVE C13, 100.6910171 MHz

DECOUPLE H1, 400.4431986 MHz

Power 40 dB

continuously on

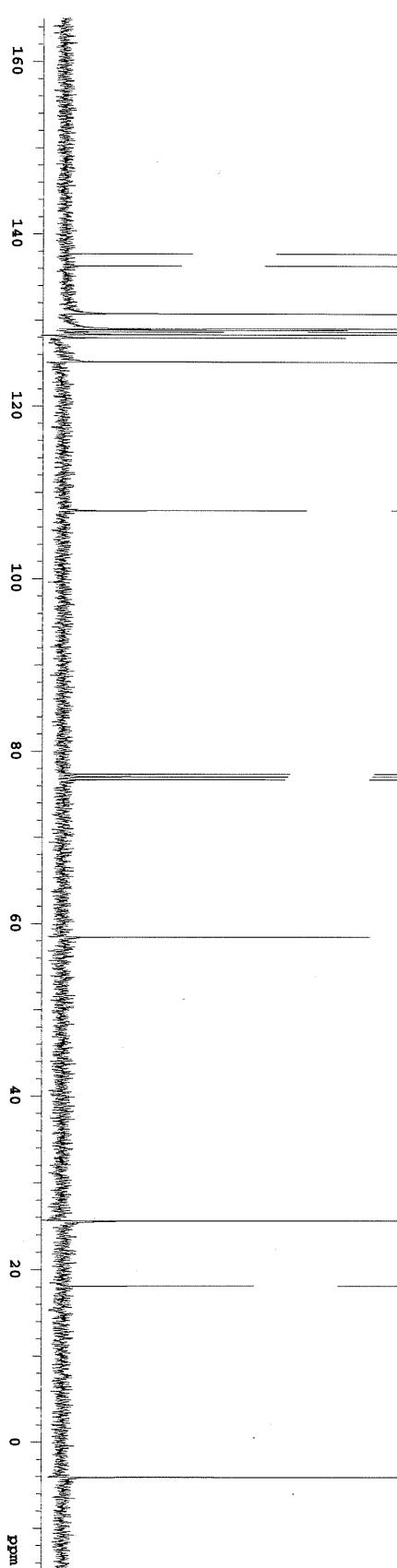
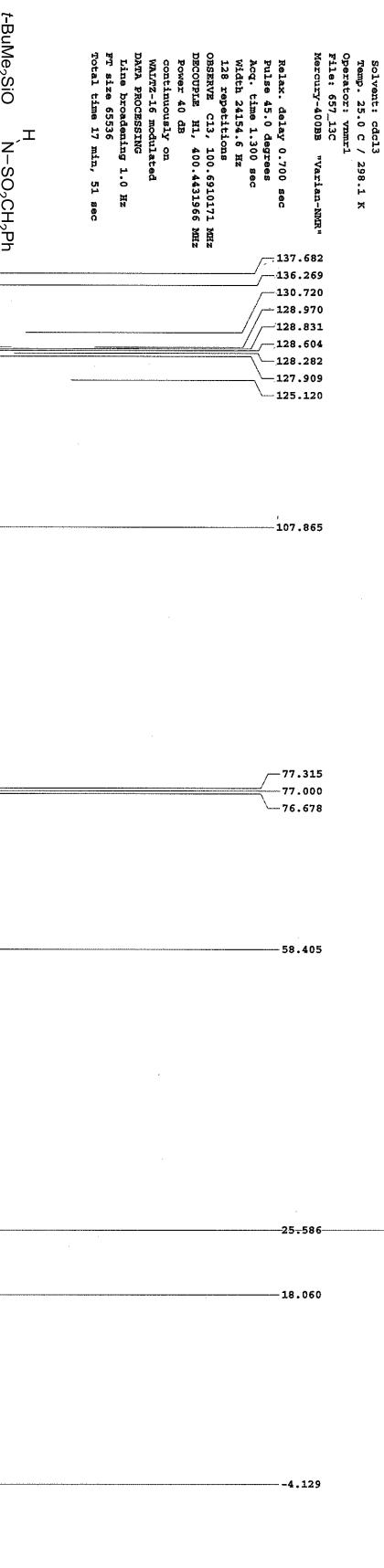
WATER-1H modulated

DATA PROCESSING

Line broadening 1.0 Hz

FT size 65536

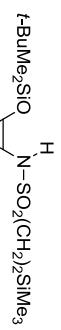
Total time 17 min, 51 sec



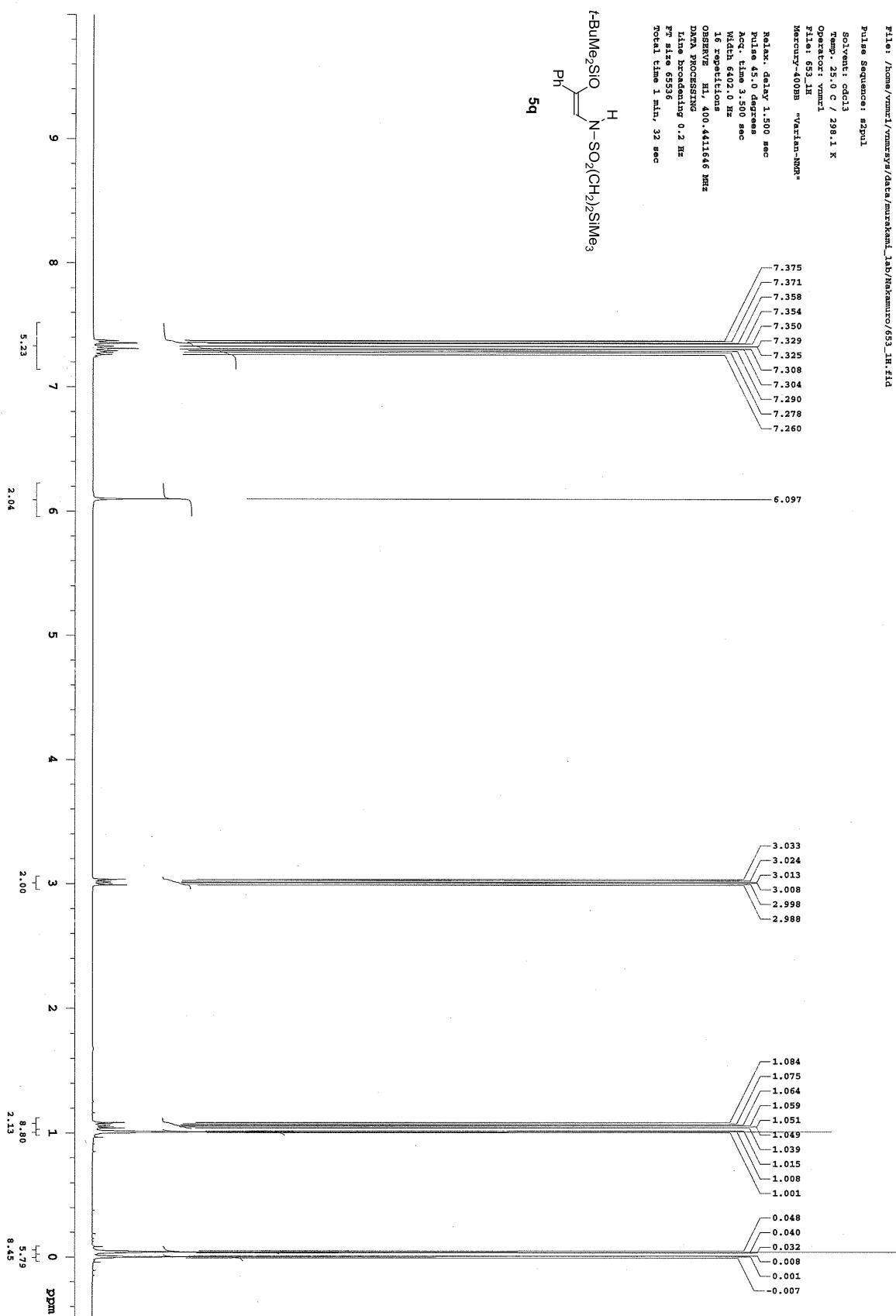
653 column  
File: /home/rmml1/rmmlsys/data/murakami\_lab/Nakamura/653\_1H.fid  
Pulse Sequence: s2p11

Solvent: CDCl<sub>3</sub>  
Temp: 25.0 C / 298.1 K  
Operator: rmml1  
File: 653\_1H "varian-NMR"  
Mercury-400B "varian-NMR"

Relax. delay 1.500 sec  
Pulse 45.0 degrees  
Acc. time 3.500 sec  
Width 6422.0 Hz  
16 repetitions  
Observe H1, 400.4411646 MHz  
Data Processing  
Line broadening 0.2 Hz  
PPM size 65536  
Total time 1 min, 32 sec



5q



653 column

File: /home/vnmrl1/vnmrsys/data/murakami1.lab/Nakamura/653\_13C.fid

Pulse Sequence: apsp1

Solvent: cdcl3

Temp: 25.0 C / 298.1 K

Operator: vnmrl1

File: 653\_13C

MERCURY-400B "Varian-MER"

Relax delay 0.700 sec

Pulse 45.0 degrees

Avg. time 1.300 sec

Width 2414.6 Hz

128 repetitions

OBSERVE C13, 100.6910171 MHz

DECOMPR. HI, 400.4431966 MHz

Power 40 dB

continuously on

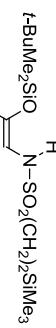
WALFZ-16 modulated

DATA PROCESSING

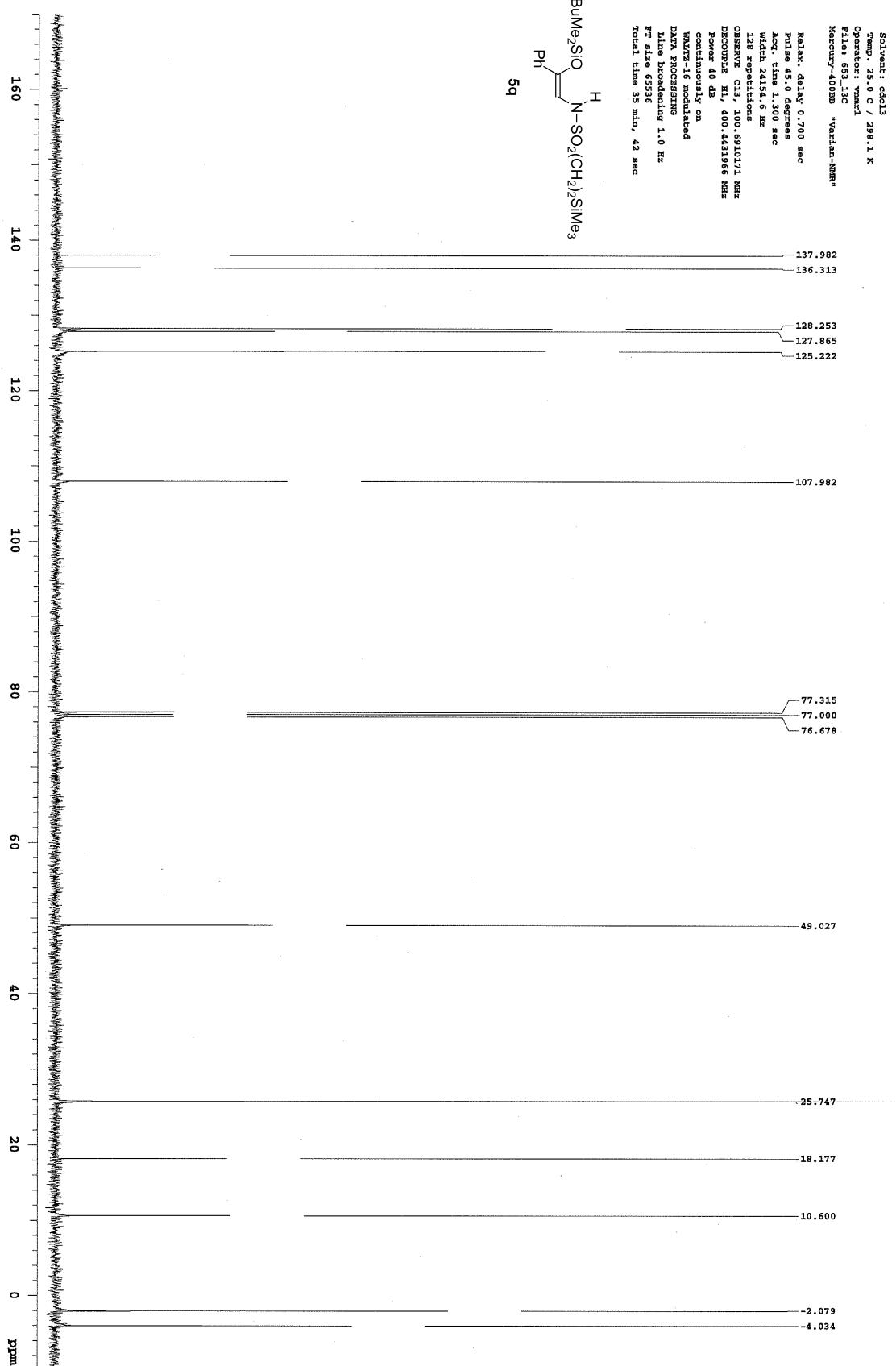
Line broadening 1.0 Hz

FFT size 65536

Total time 35 min., 42 sec



5q



1384 column

File: /Home/vnmrl/vnmrsys/data/murakami\_lab/Nakamura/1384\_1H.fid

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vnmrl

File: 1384\_1H  
Mercury-400B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 640.0 Hz

16 repetitions

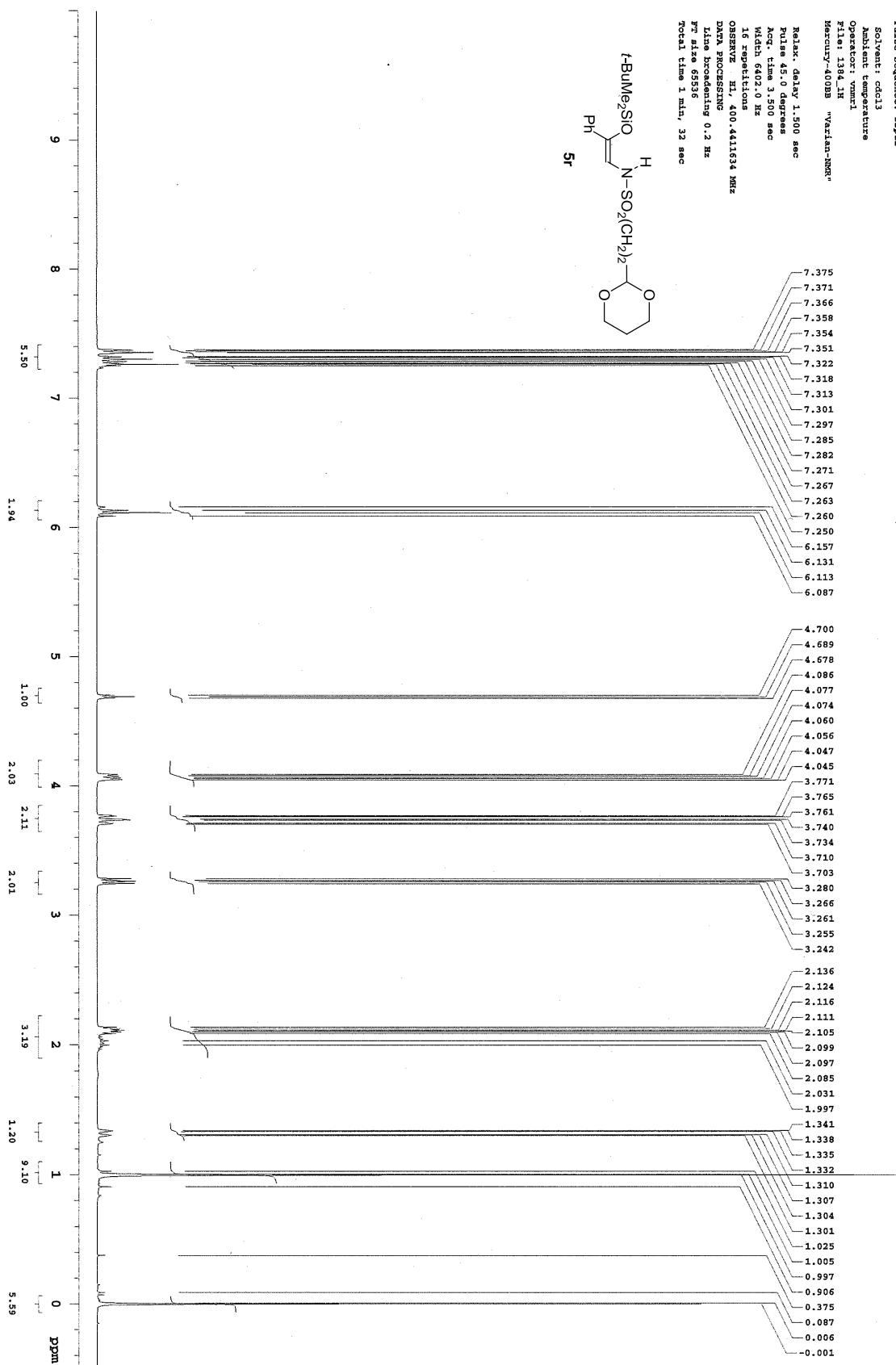
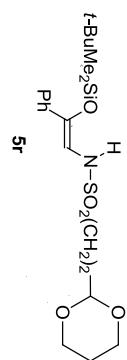
OBSERVE Hz, 400.4411634 MHz

DATA PROCESSING

Line broadening 0.2 Hz

FT size 65536

Total time 1 min, 32 sec



1384 column:

File: /home/rnakami/vnmr1/vnmrsys/data/nakamai\_1ab/Nakamairo/1384\_13C.fid

Pulse Sequence: sp2pul

Solvent: cdcl<sub>3</sub>

ambient temperature

Operator: vnmri

Model: 1384.13C "Varian-NMR"

Mercury-400B "Varian-NMR"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acc. time 1.300 sec

Width 24.154.6 Hz

128 repetitions

OBSERVE CL3, 100.6910179 kHz

RECDIM 1M, 400.4431966 kHz

Power 41 dB

continuously on

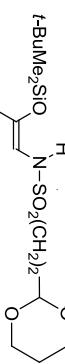
WAVE=16 modulated

DATA PROCESSING

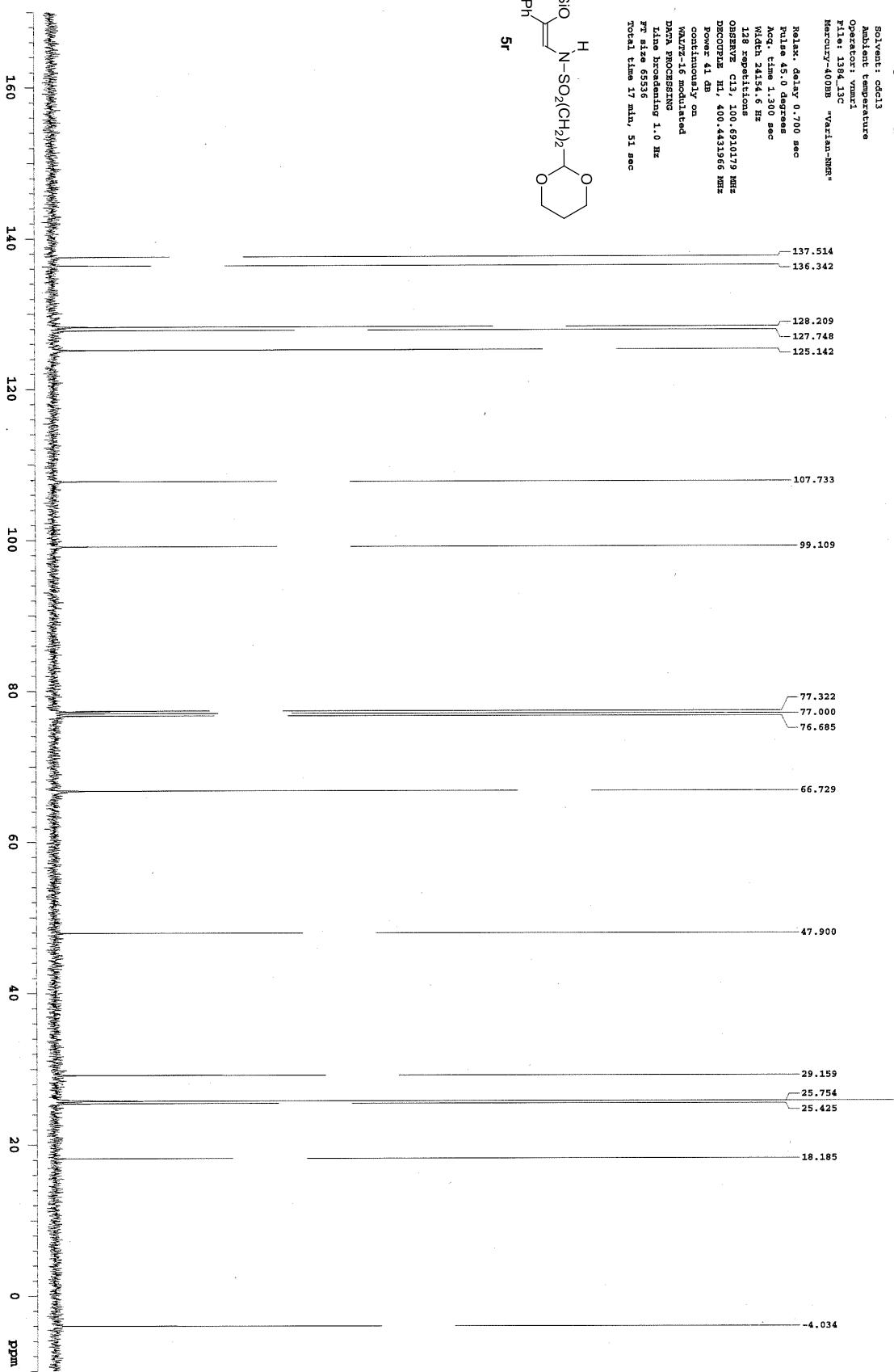
line broadening 1.0 Hz

RT size 65336

Total time 17 min, 51 sec



5r



20-2

File: /home/vmml1/vmmlsys/data/murakami\_lab/Nakamuro/Mukaiyama\_aldol/NaIodol\_2x13.2.fid

Pulse Sequence: 42pul

Solvent: ccl4.3

Temp: 21.0 C / 294.1 K

Operator: vmml

File: m\_s10d\_2x13.2

Mercury-400MHz "Varian-NMR"

Relax: delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Watch: 600.1 Hz

16 replications

OBSERVE: H1 400.4411628 MHz

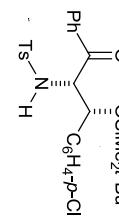
DATA PROCESSING

Line broadening 0.2 Hz

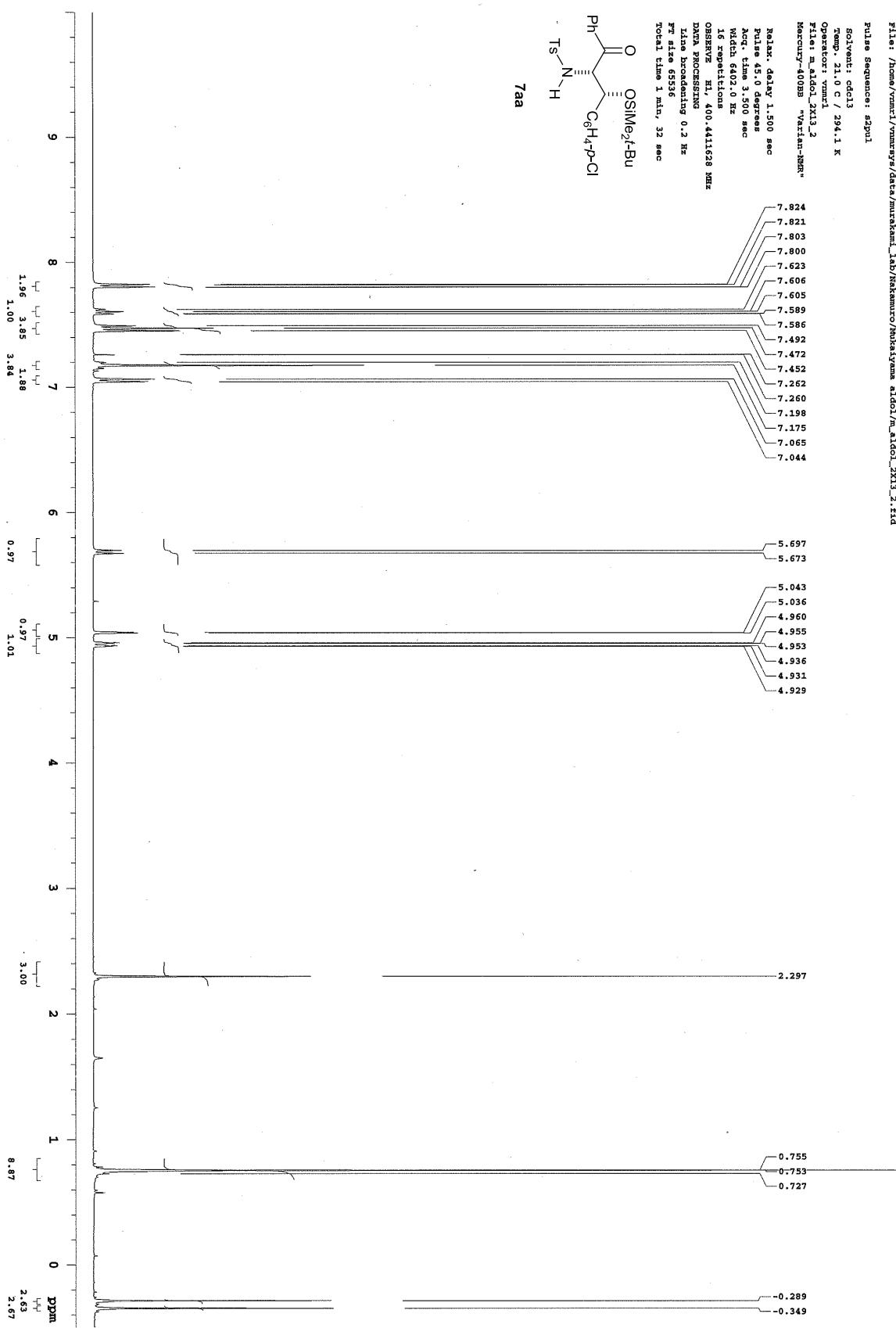
FF size 65336

Total time 1 min, 32 sec

Chemical structure of 7aa:



7aa



File: /home/vmml/vmml/vnresys/data/murakami\_lab/Nakamura/Makiyama\_zld1/m\_zld1\_2x13\_13C.fid

Pulse Sequence: s2pul1

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vml1

File: m\_zld1\_2x13\_13C

Marconi-400BB "Varian-NMR"

Relax: delay 0.700 sec

Pulse 45.0 degrees

Acc. time 1.300 sec

Width 2414.6 Hz

576 repetitions

OBSERVE C13 100.691042 MHz

DECOURSE H1 400.443196 MHz

Power 40 dB

continuously on

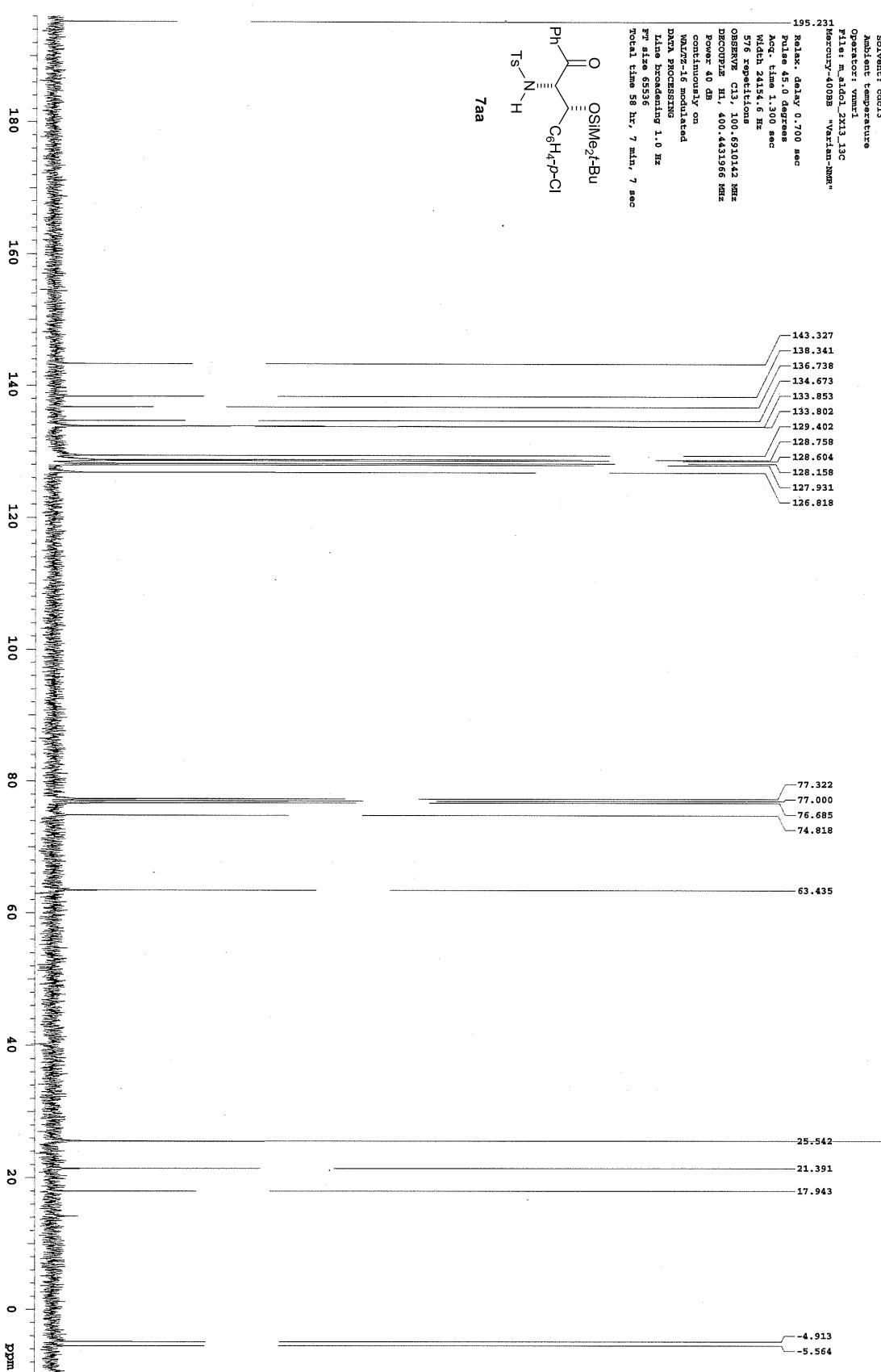
WIDENING modulated

DATA PROCESSING

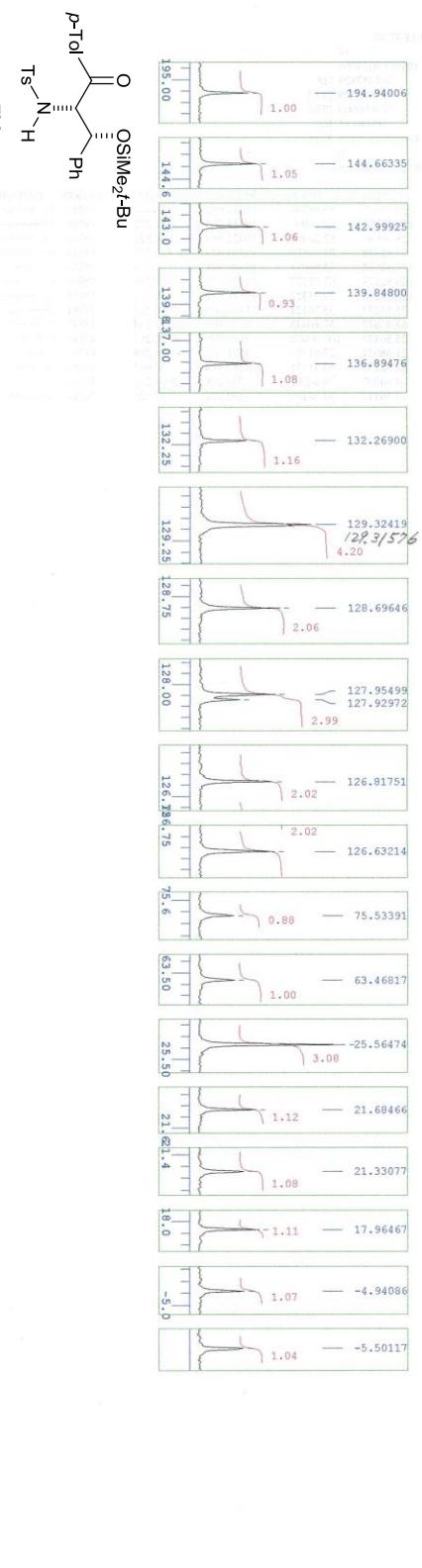
LINE broadening 1.0 Hz

FT size 65536

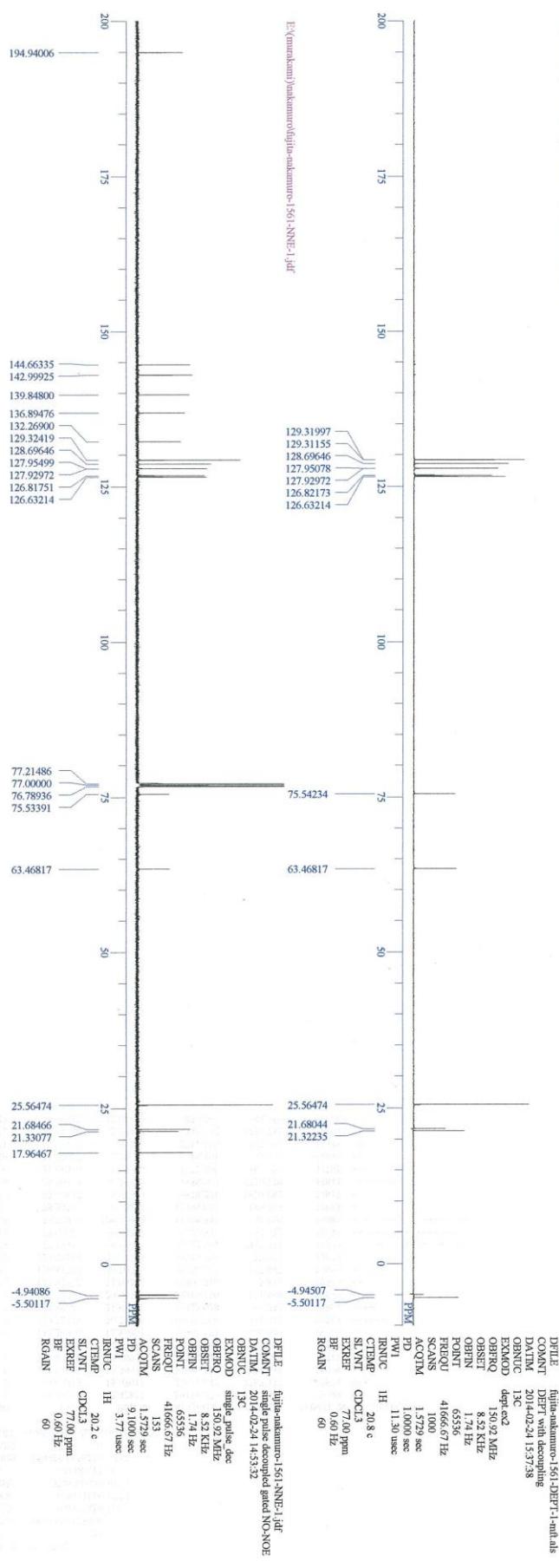
total time 58 hr, 7 min, 7 sec







130448



1564 pt1c

file: /home/vnmr1/vnmrsys/data/murakami\_lab/Nakamuro/1562\_1H.fid

Pulse Sequence: s2pul

Solvate: gaseous

Operator: vimrl

Mercury-400BB "Varian-NMR"

File: 1562.JH "Varian-NMR"  
Mercury-400MHz

Relax. delay 1.500 sec  
Pulse 45.0 degrees  
Acq. time 3.500 sec  
Width 6402.0 Hz  
16 repetitions  
OBSERVE HI, 400.4411615 MHz  
DATA PROCESSING 0.2 Hz  
FT size 65536  
Total time 1 min, 32 sec

7.796  
7.774  
7.479  
7.458  
7.273  
7.260  
7.257  
7.246  
7.236  
7.032  
7.012  
6.926  
6.904  
  
5.722  
5.698  
  
5.049  
5.042  
4.942  
4.934  
4.918  
4.910

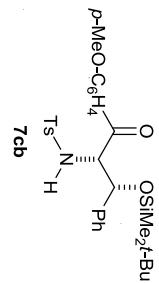
3.879

2.258

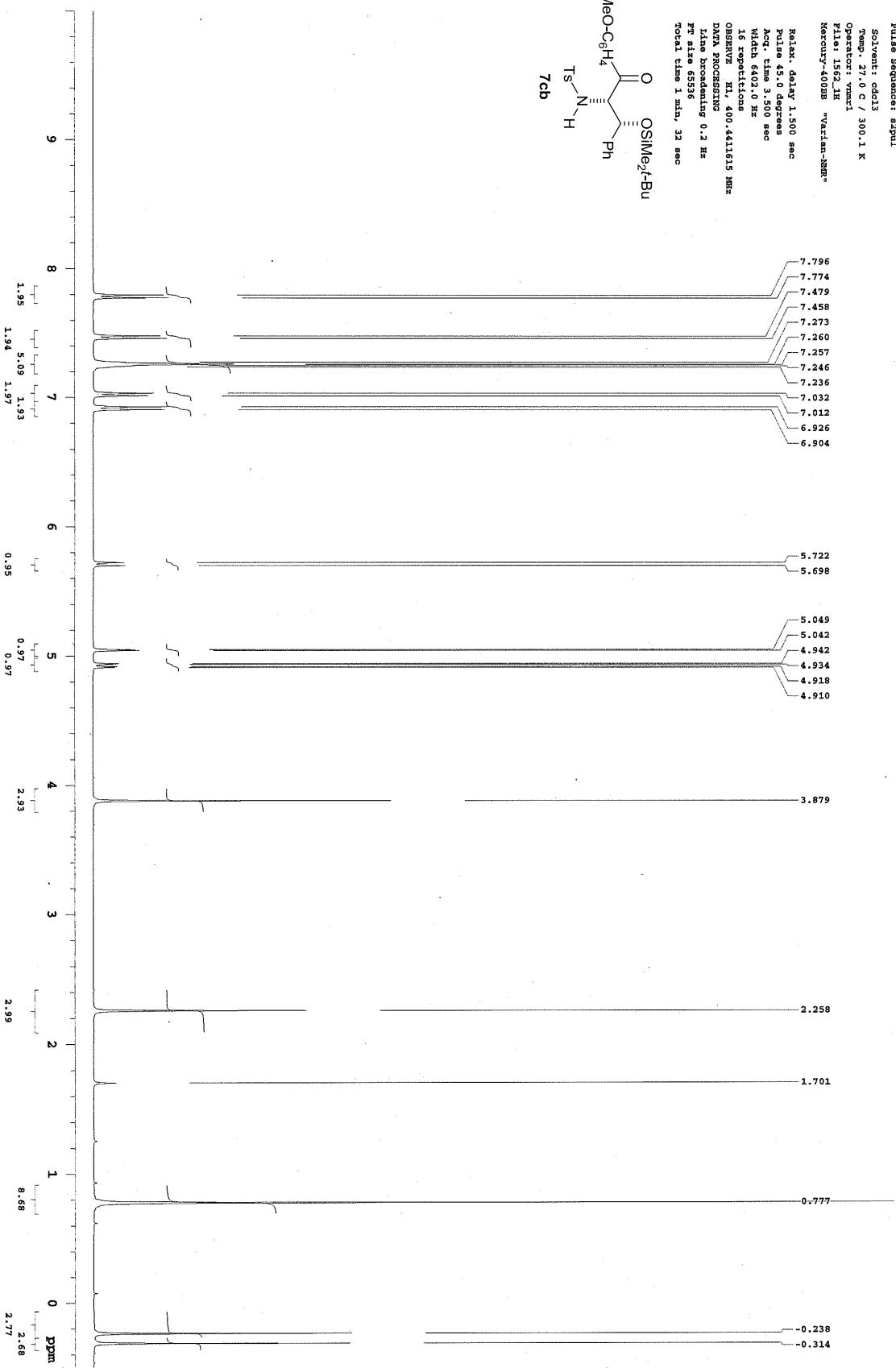
1.701

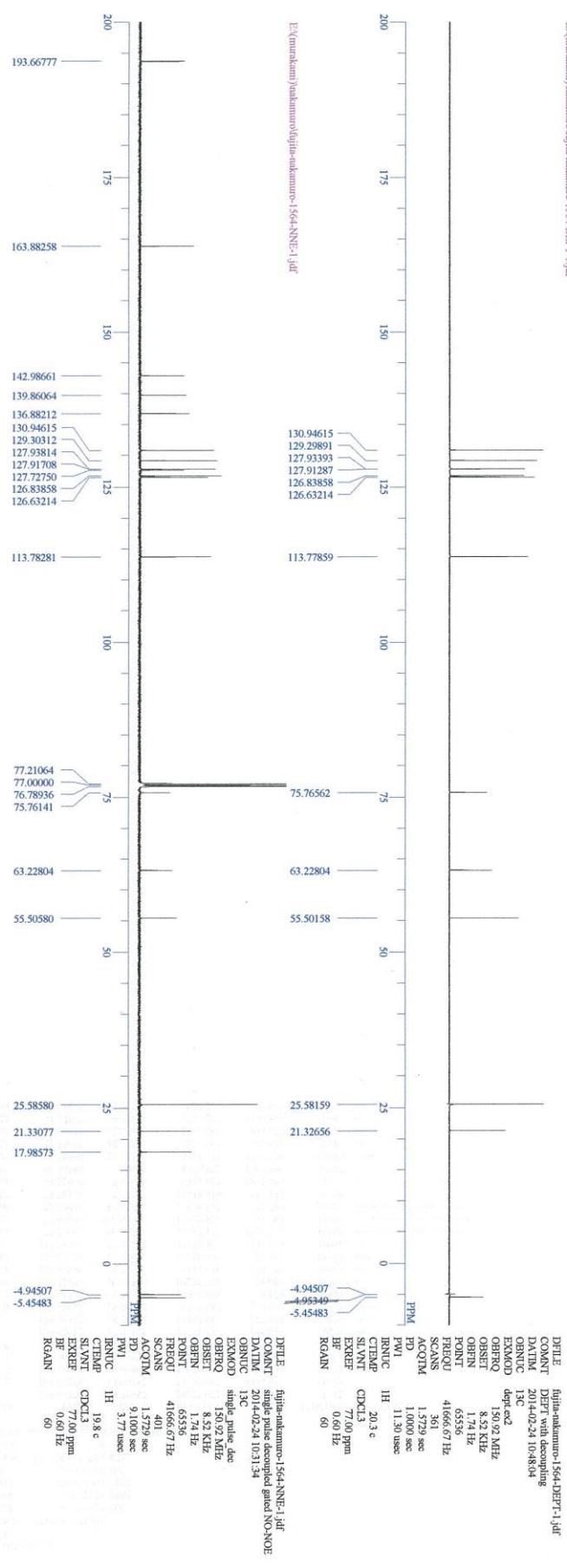
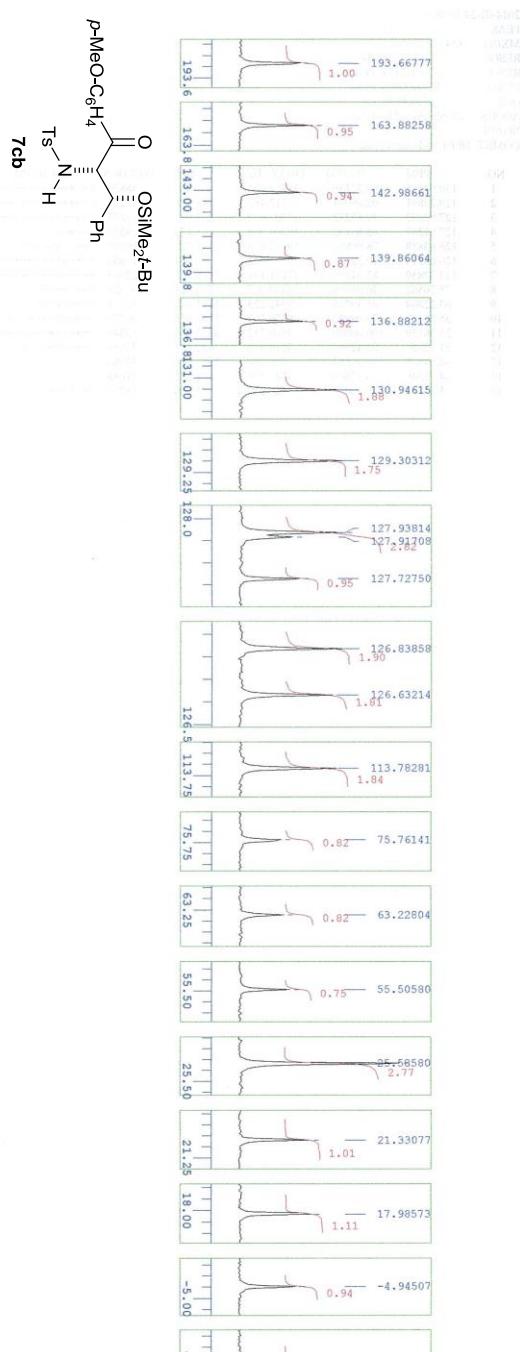
0.777

-0.238  
-0.314



13





new experiment

File: /Name/vnmr1/vnmrsys/data/nakamuro/1556\_1H.fid

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Temp. 27.0 °C / 300.1 K

Operator: vnmr1

File: 1556\_1H

Mercury-400MHz "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 6403.0 Hz

4 repetitions

OBSERVE H1 400.4411630 MHz

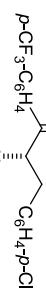
DATA PROCESSING

L100 breaking 0.2 Hz

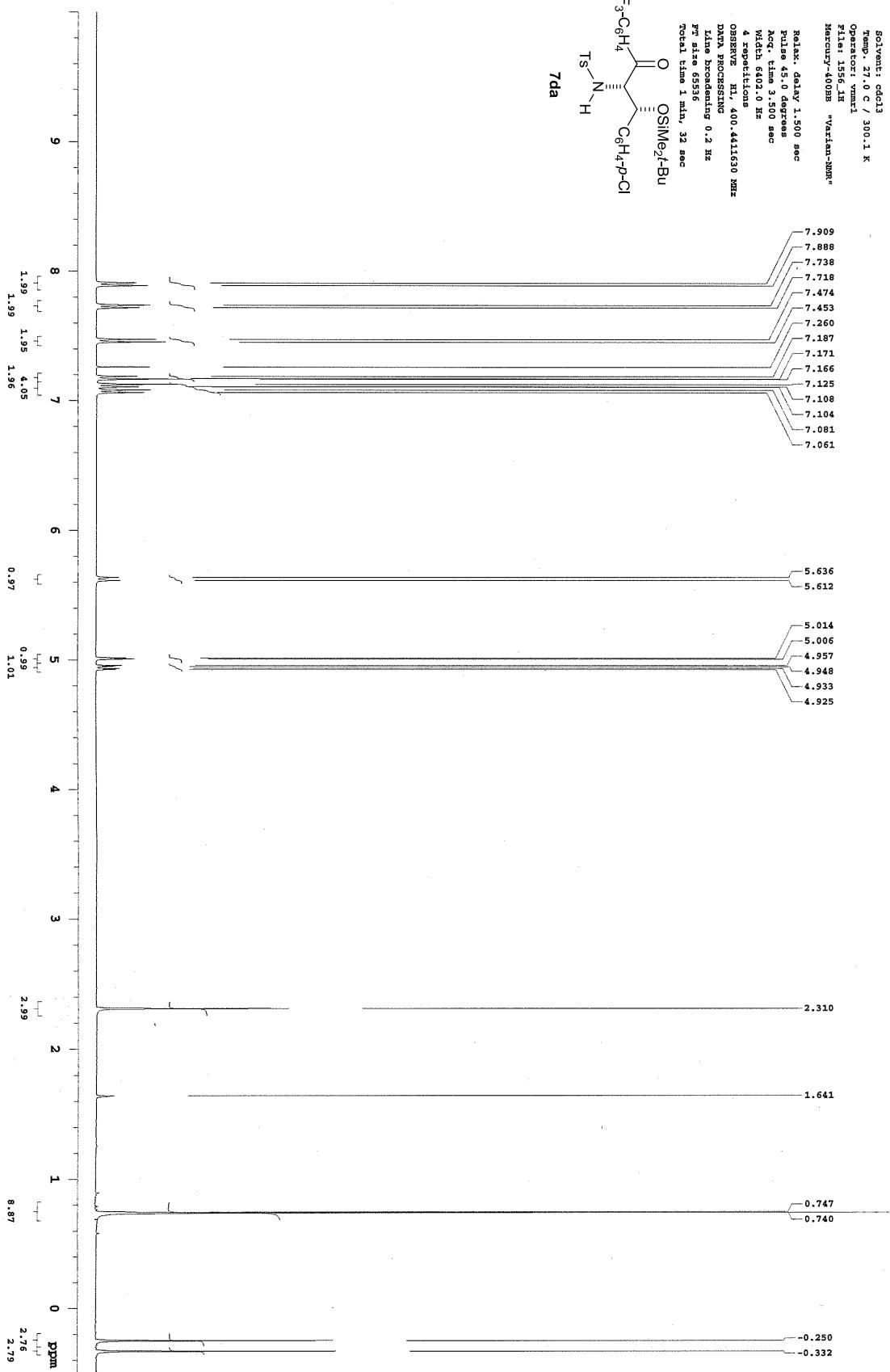
FT size 65536

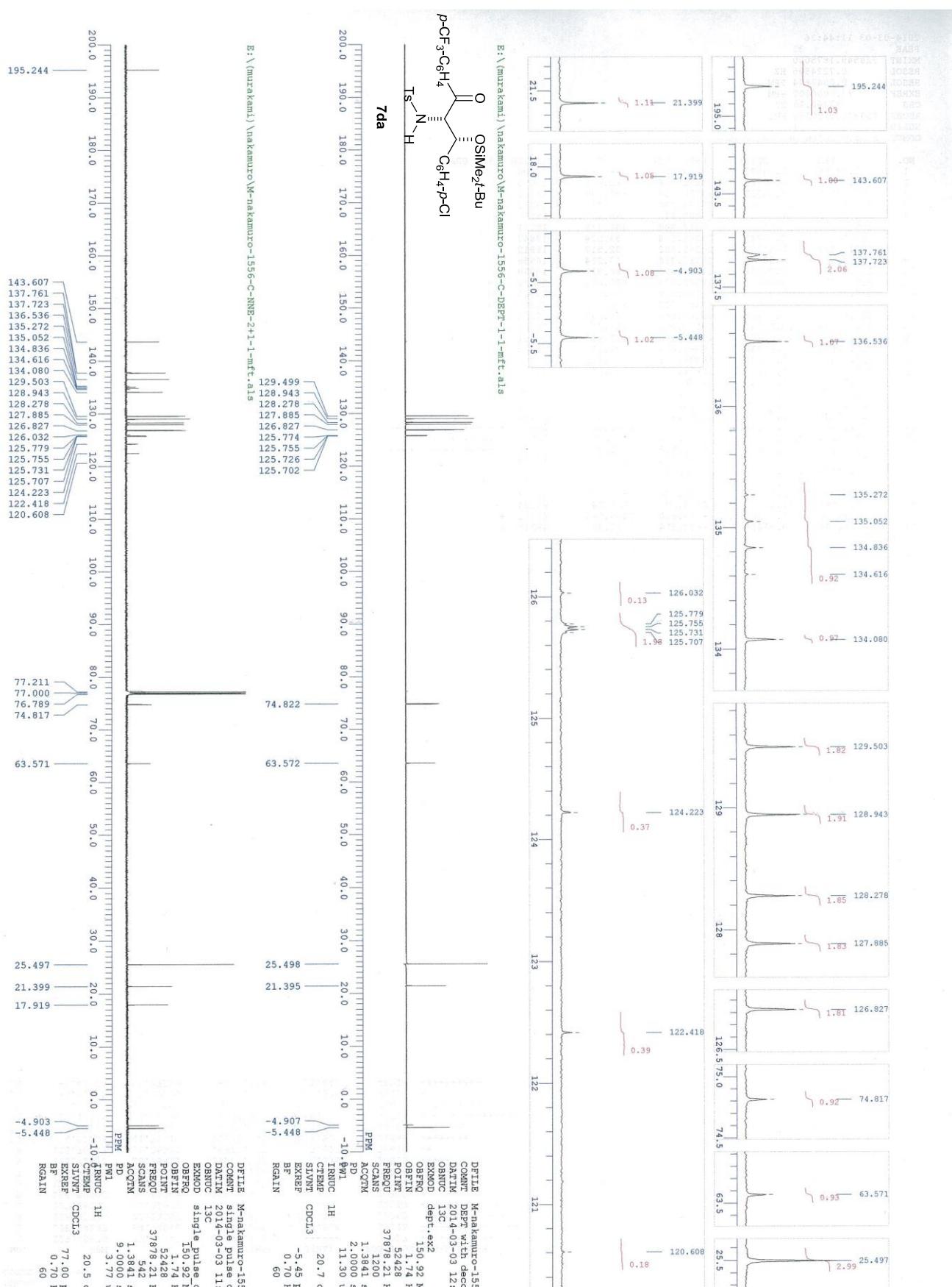
L100 total 1 min, 32 sec

Total time 1 min, 32 sec



7da





1552.p1c

File: /home/vmusr1/muraysu/data/murakami\_lab/Nakamuro/1552\_1H.fid

Pulse Sequence: s2pul

Solvent: cdcl<sub>3</sub>

Temp. 27.0 °C / 300.1 K

File: 1552\_1H

Operator: vmsr1

Mercury-400BMS "Varian-NMR"

Relax. delay 1.500 sec

pulse 45.0 degrees

Actr. time 3.500 sec

Width 64.02.0 Hz

16 repetitions

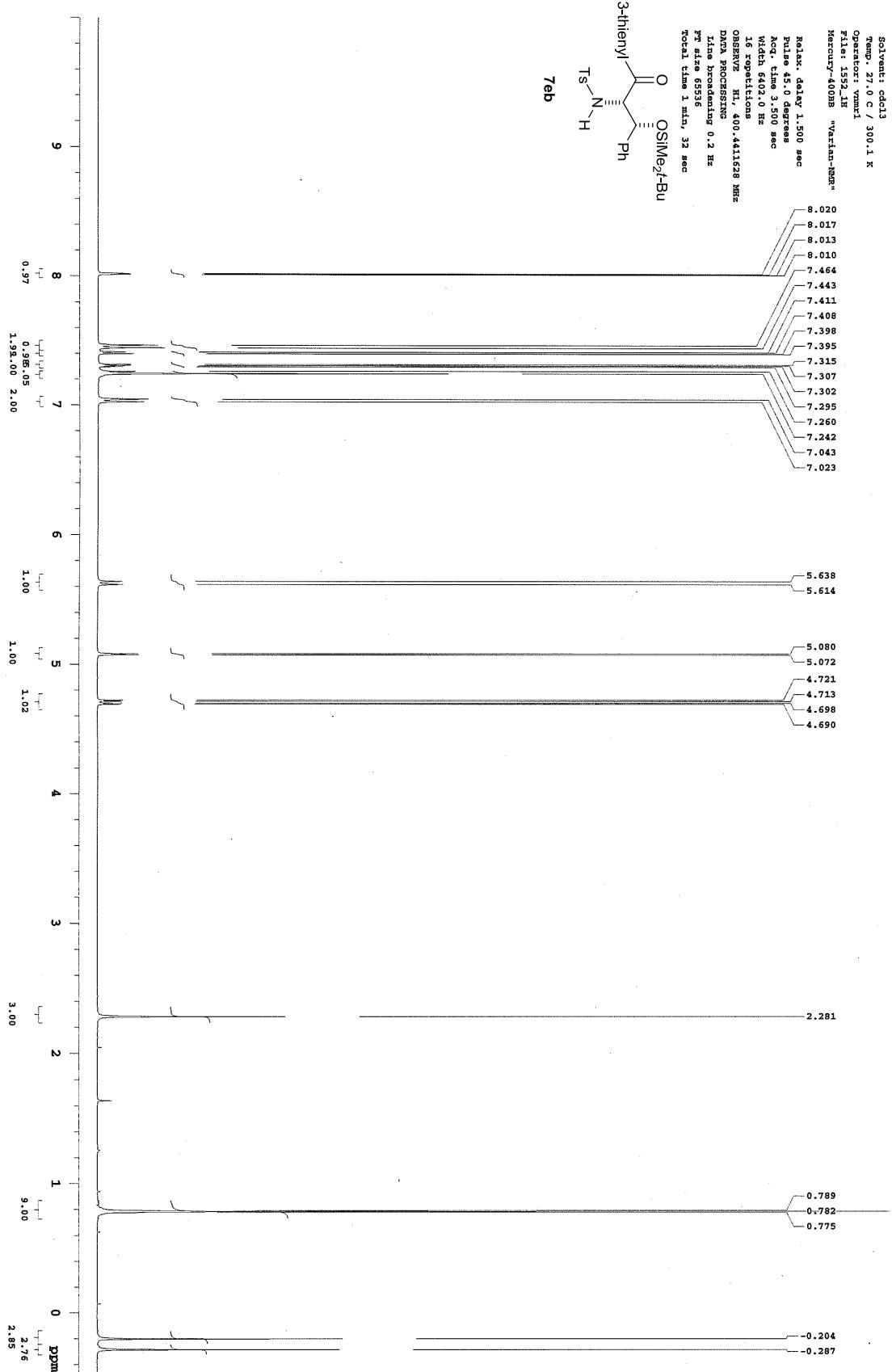
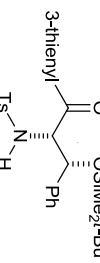
Observe H1, 400.441123 MHz

Data Processing

Liss broadening 0.2 Hz

RT size 65536

Total time 1 min., 32 sec



1552.p1c

File: /home/vmnml1/vmnml1s9/data/murakami\_1ab/Nakamura/1552\_13C.fid

Pulse sequence: s2pul

Solvent: cdcl3

Temp: 27.0 C / 300.1 K

Operator: vmnml

File: 1552\_13C

Mercury-400B "Varian-NMR"

Relax: 90°

Pulse: 45.0 degrees

Acq. time: 1.300 sec

Width: 24154.6 Hz

1472 repetitions

OBSERVE: 0.13, 100.6910164 MHz

DECOPPLE: H1, 400.4433.966 kHz

Power: 41 dB

continuously on

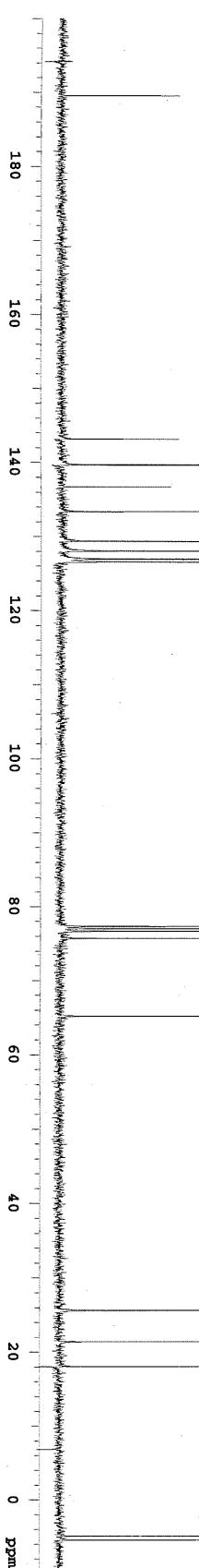
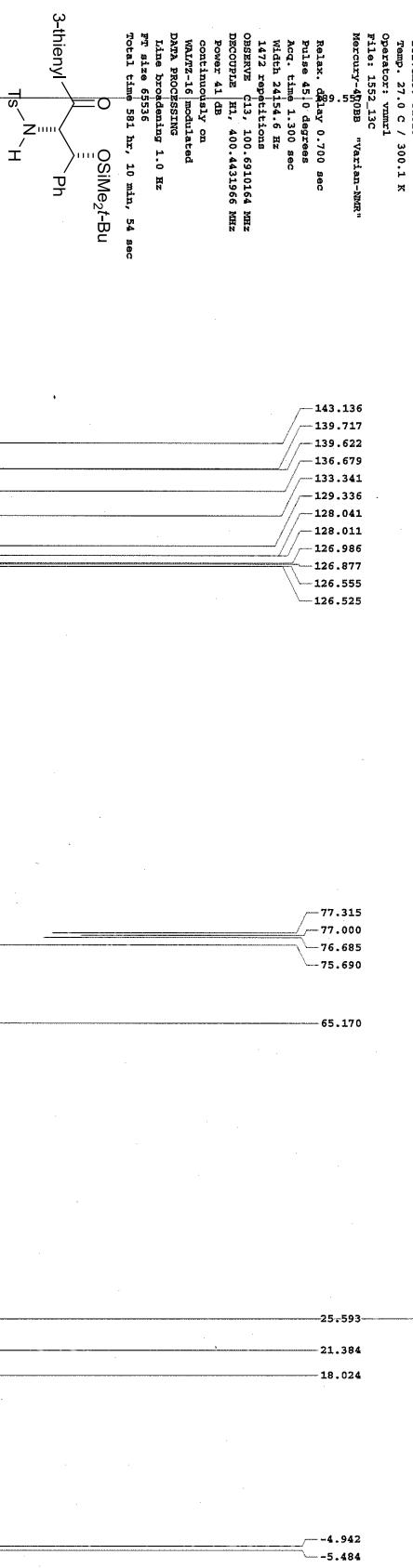
WALZ-16 modulated

DATA PROCESSING

Line broadening: 1.0 Hz

RT size: 65536

Total time: 58.1 hr, 10 min, 54 sec



818.p1c

File: /home/vnmr1/vnmrsys/data/murakami\_lab/Nakamura/818.z1d

Pulse Sequence: sspul

Solvent: cdcl3

Ambient temperature

Operator: vnmr1

File: 818

Mercury-400B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 602.0 Hz

16 repetitions

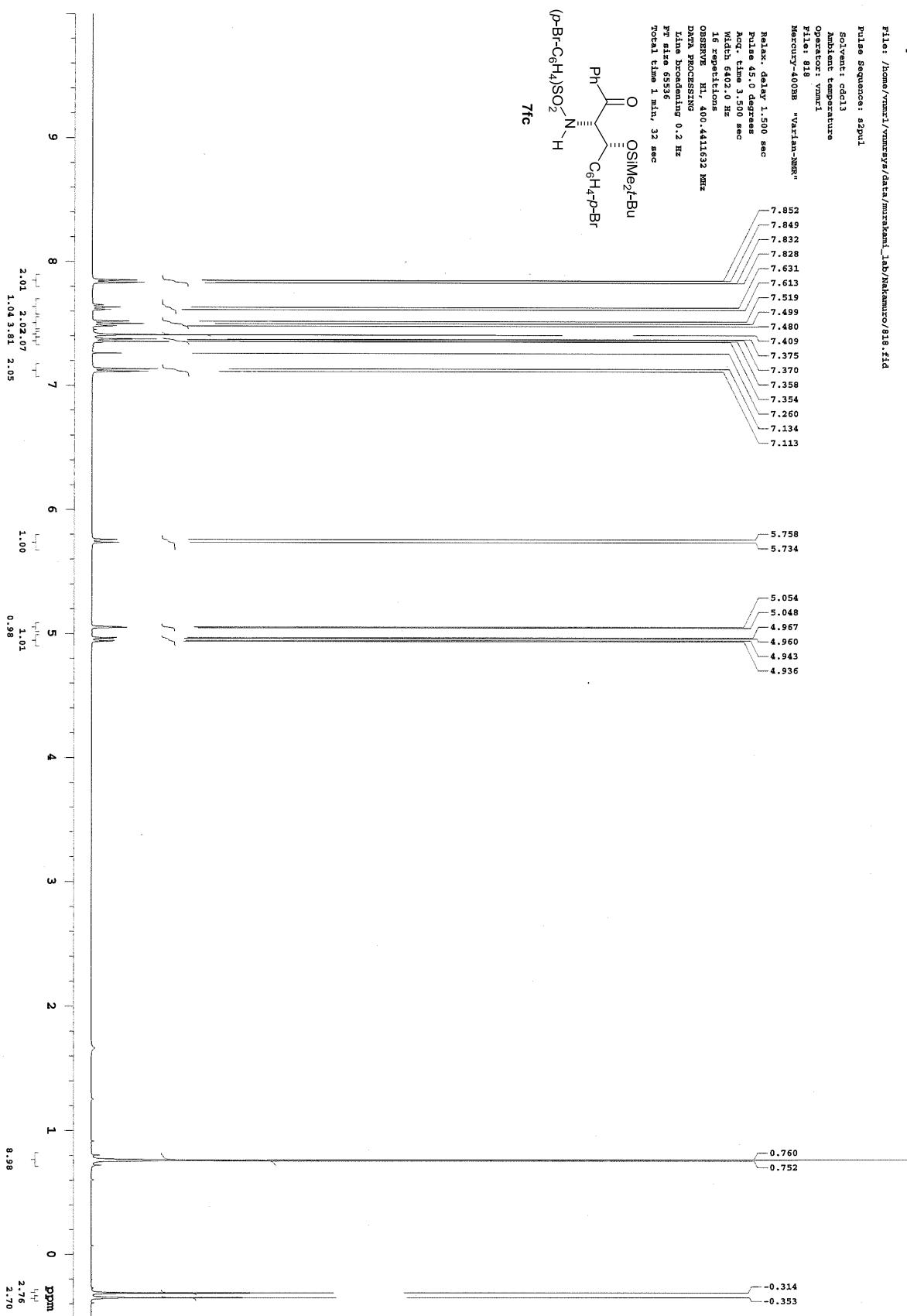
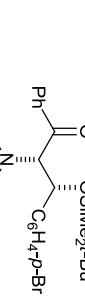
OBSERVE H1, 400.4411632 MHz

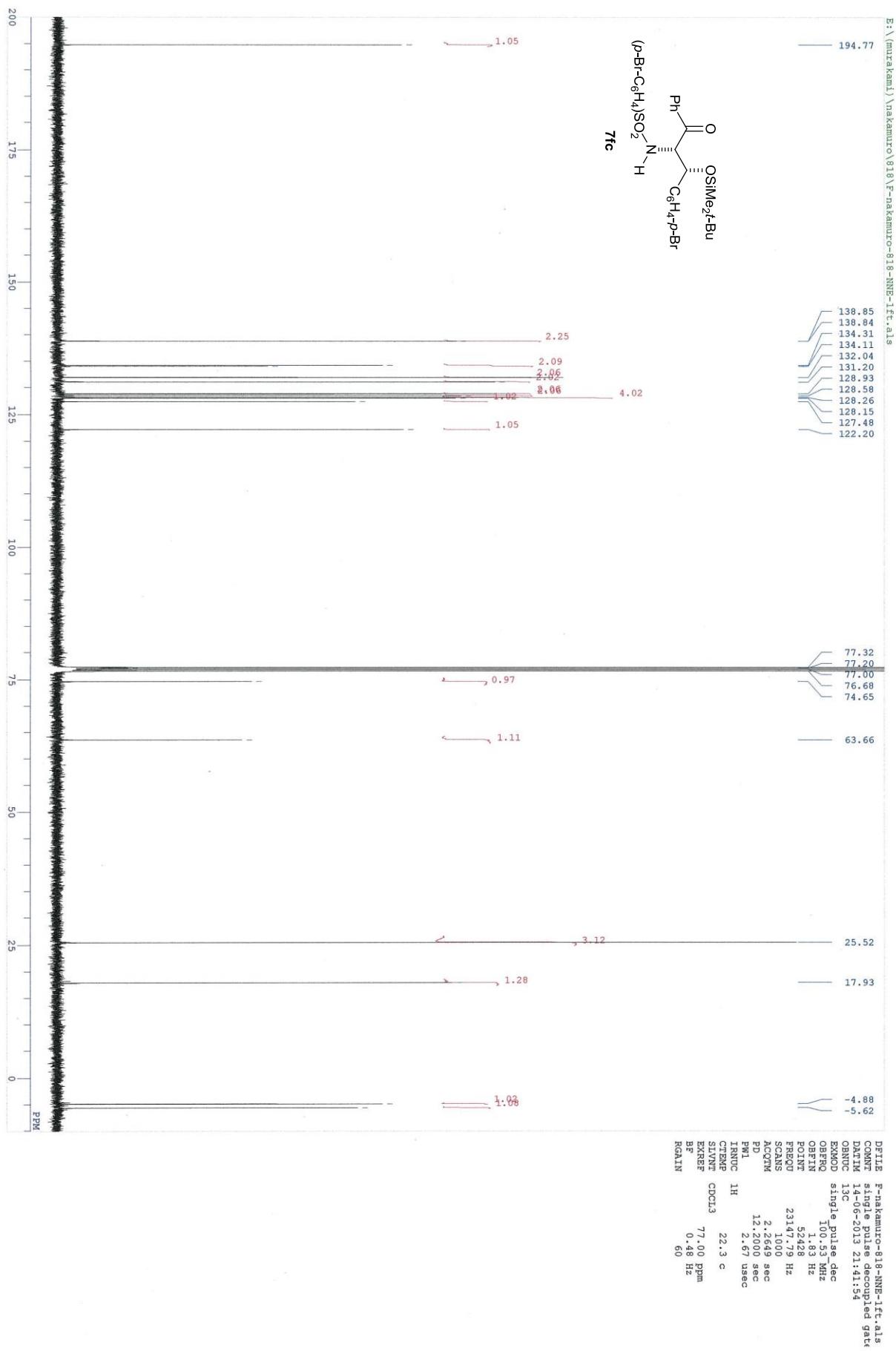
DATA PROCESSING

Line broadening 0.2 Hz

FT size 65536

Total time 1 min, 32 sec





File: /home/vmrci/vmrci/vmrcsys/data/murakami\_1ab/Nakamura/Mukaiyama\_sido1/m.sido1\_381\_1R\_2.fid

Pulse Sequence: 2spul

Solvent: cdcl<sub>3</sub>

Temp: 21.0 C 294.1 K

Operator: vmrci

Mercury-400BB "Varian" 2

File: m.sido1\_381\_1R\_2

Mercury-400BB "Varian" 2

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acc. time 3.500 sec

Width 6402.0 Hz

16 repetitions

Pulse 45.0 degrees

Acc. time 3.500 sec

Width 6402.0 Hz

OBSERVE H1 400.4411630 MHz

DATA PROCESSING

Line broadening 0.2 Hz

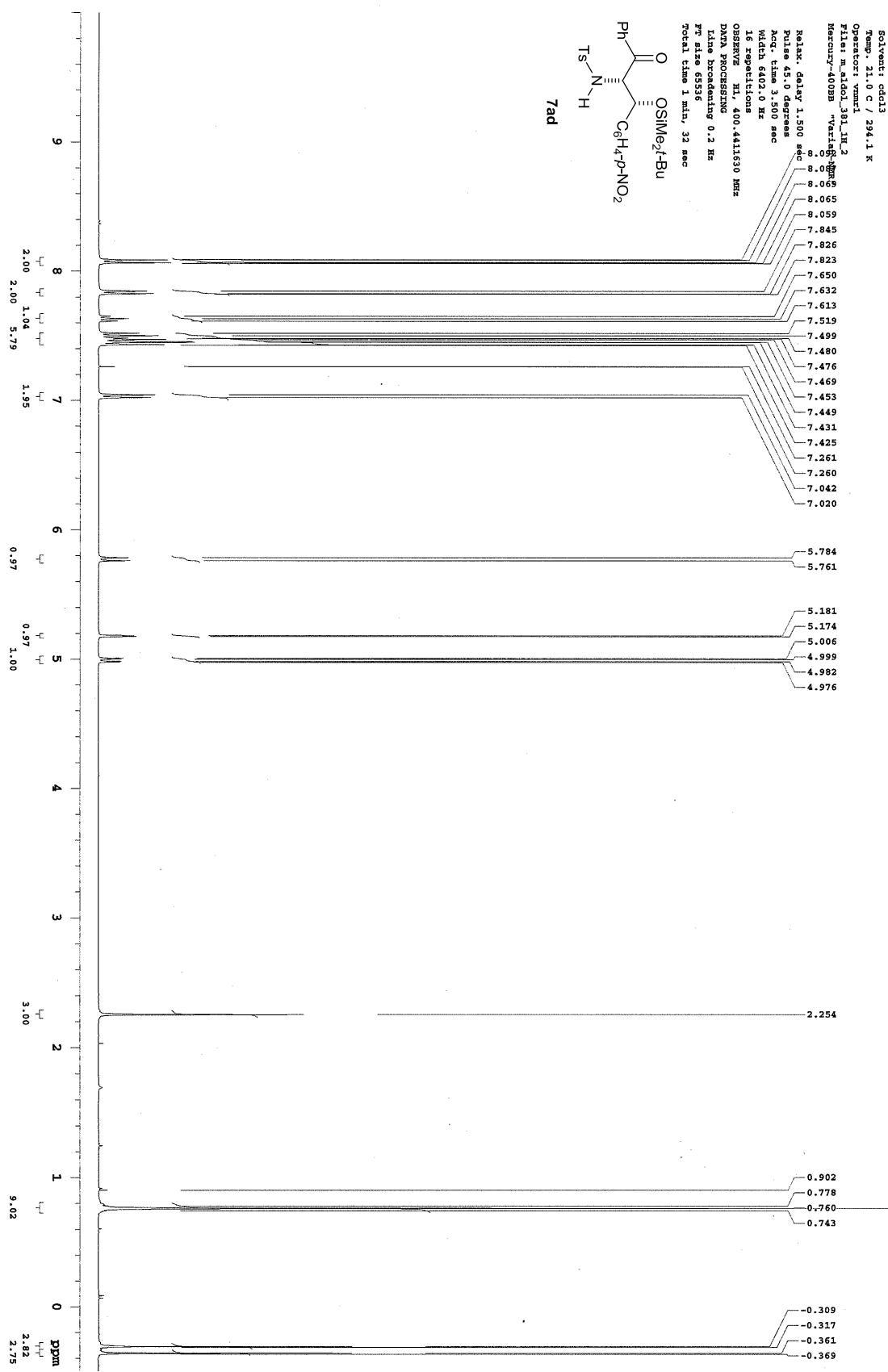
FT size 65536

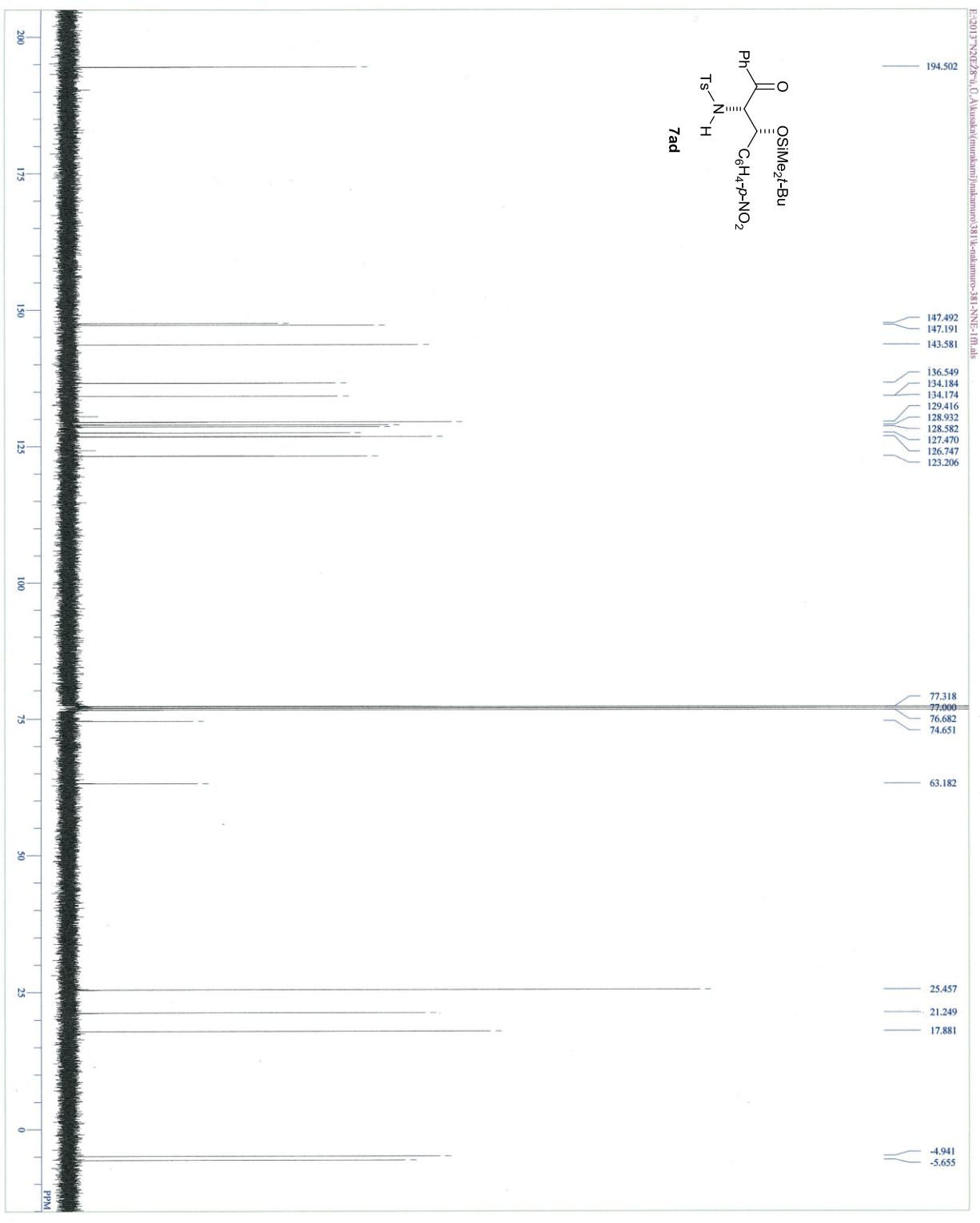
Total time 1 min. 32 sec

Ph  
C(=O)  
|  
C=C  
|  
OSiMe<sub>2</sub>t-Bu  
|  
C<sub>6</sub>H<sub>4</sub>-p-NO<sub>2</sub>

Ts-N-H

7ad





File: /Name/vnmr1/vnmrsys/data/murakami\_lab/Rakamuro/mulayama\_aido/maidol\_2707\_323\_1H.fid

Pulse Sequence: s2p11

Solvent: cdcl<sub>3</sub>

Temp: 21.0 C / 294.1 K

Operator: vnmr1

File: maidol\_2707\_323\_1H

MAGNET: 400B Varian-NMR

Relax. delay 1.500 sec

Pulse 45.0 degrees

Aqc. time 3.500 sec

Width 642.0 Hz

16 repetitions

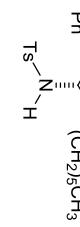
OBSERVE: H1, 400.4411642 MHz

DATA PROCESSING

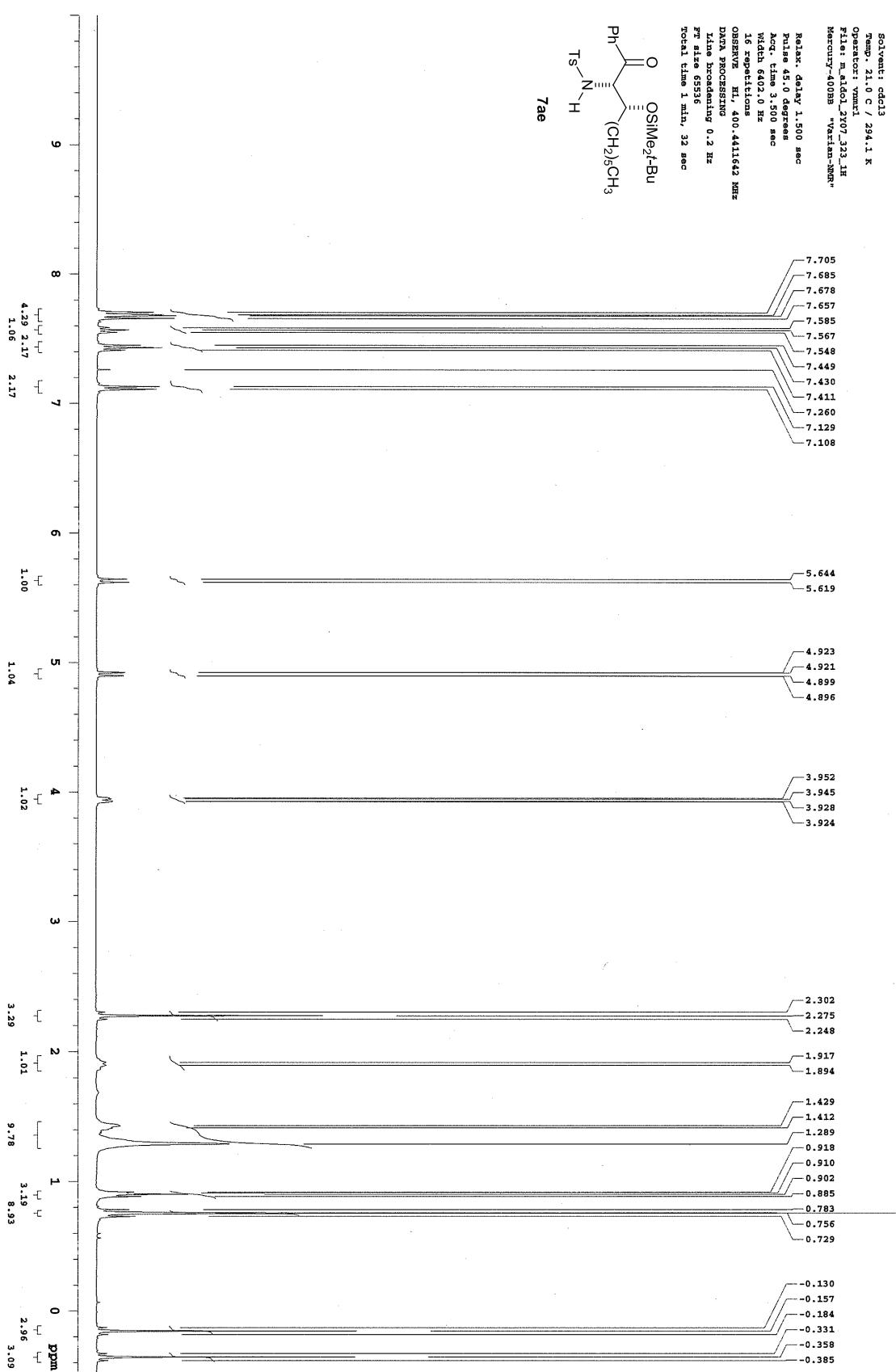
LINE BROADENING 0.2 Hz

RT size 65536

Total time 1 min, 32 sec



7ae



new experiment

file: exp

Pulse Sequence: s2pul

Solvent: CDCl<sub>3</sub>

Temp.: 21.0 C / 294.1 K

Operator: vnmr1

Mercury-400BB "Varian-NMR"

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File: /home/vmmt1/vmmtsys/data/murakami\_lab/Nakamura/474.3.fid

Pulse Sequence: s2spul

Solvent: cdd13

Temp. 19.0 C / 292.1 K

Operator: vmtmt1

File: 474.3

Mercury-40<sup>13</sup>C

7.74 7.74

7.52 7.52

7.51 7.51

7.49 7.49

7.44 7.44

7.43 7.43

7.40 7.40

7.36 7.36

7.28 7.28

7.26 7.26

7.20 7.20

7.12 7.12

7.11 7.11

7.10 7.10

7.09 7.09

7.08 7.08

7.07 7.07

7.07 7.07

7.06 7.06

7.05 7.05

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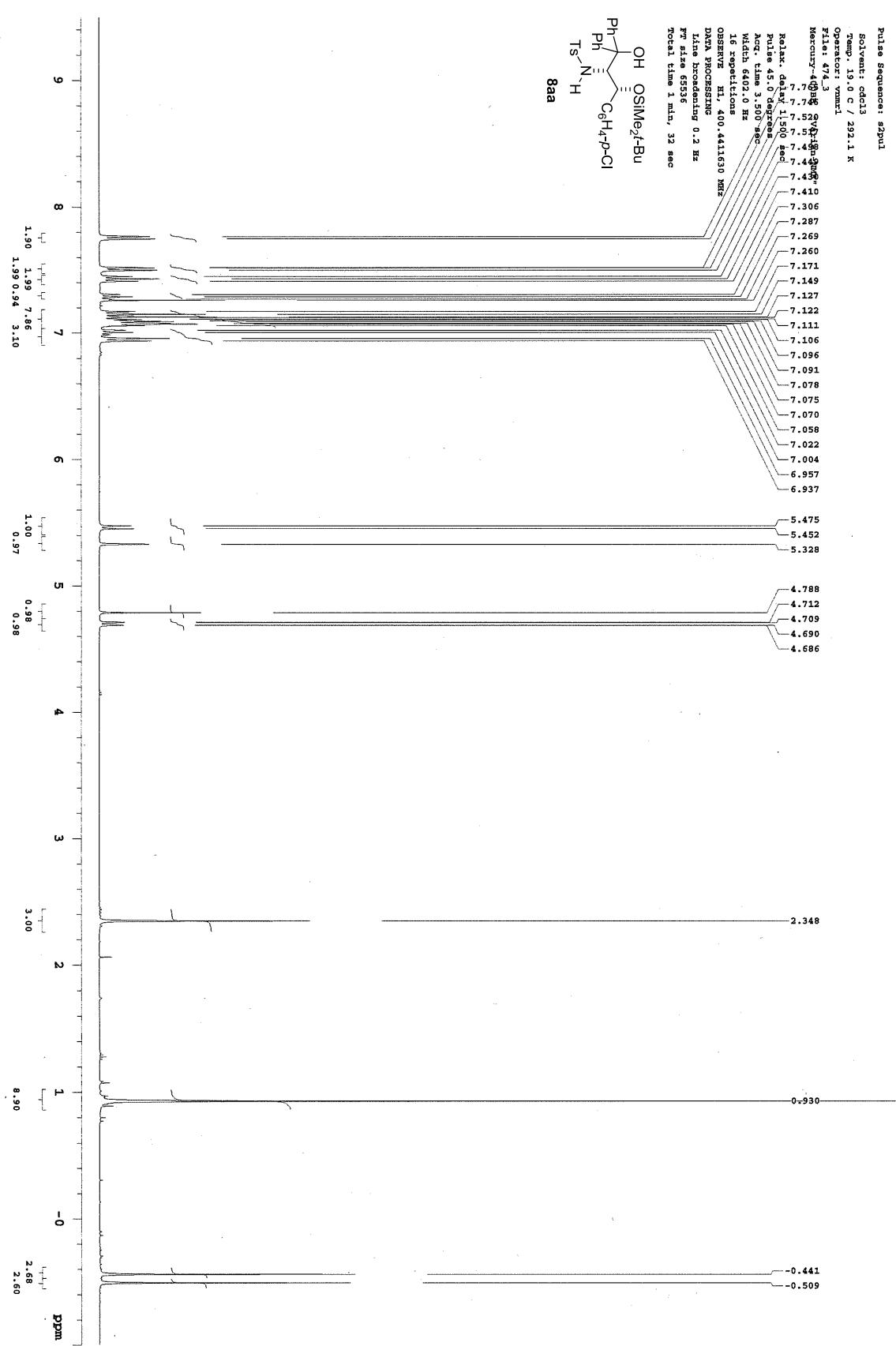
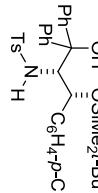
7.05 7.05

7.05 7.05

7.05 7.05

7.05 7.05

7.05 7.05



File: /Name/vnmr1/vnmrsvr/data/murakami\_1ab/Nakamura/474\_3\_13C.fid

Pulse sequence: s2pul

Solvent: cdcl<sub>3</sub>

Operator: vnmr1

Temp: 19.0 C / 292.1 K

File: 474\_3\_13C

Mercury-400MHz "VarianPro"

Relax. delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.300 sec

Width 24154.6 Hz

128 repetitions

OBSERVE C13, 100.631299 MHz

DECOUPLE CH, 400.4431986 MHz

Power 40 dB

continuously on

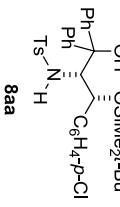
WAVEZ-16 modulated

DATA PROCESSING

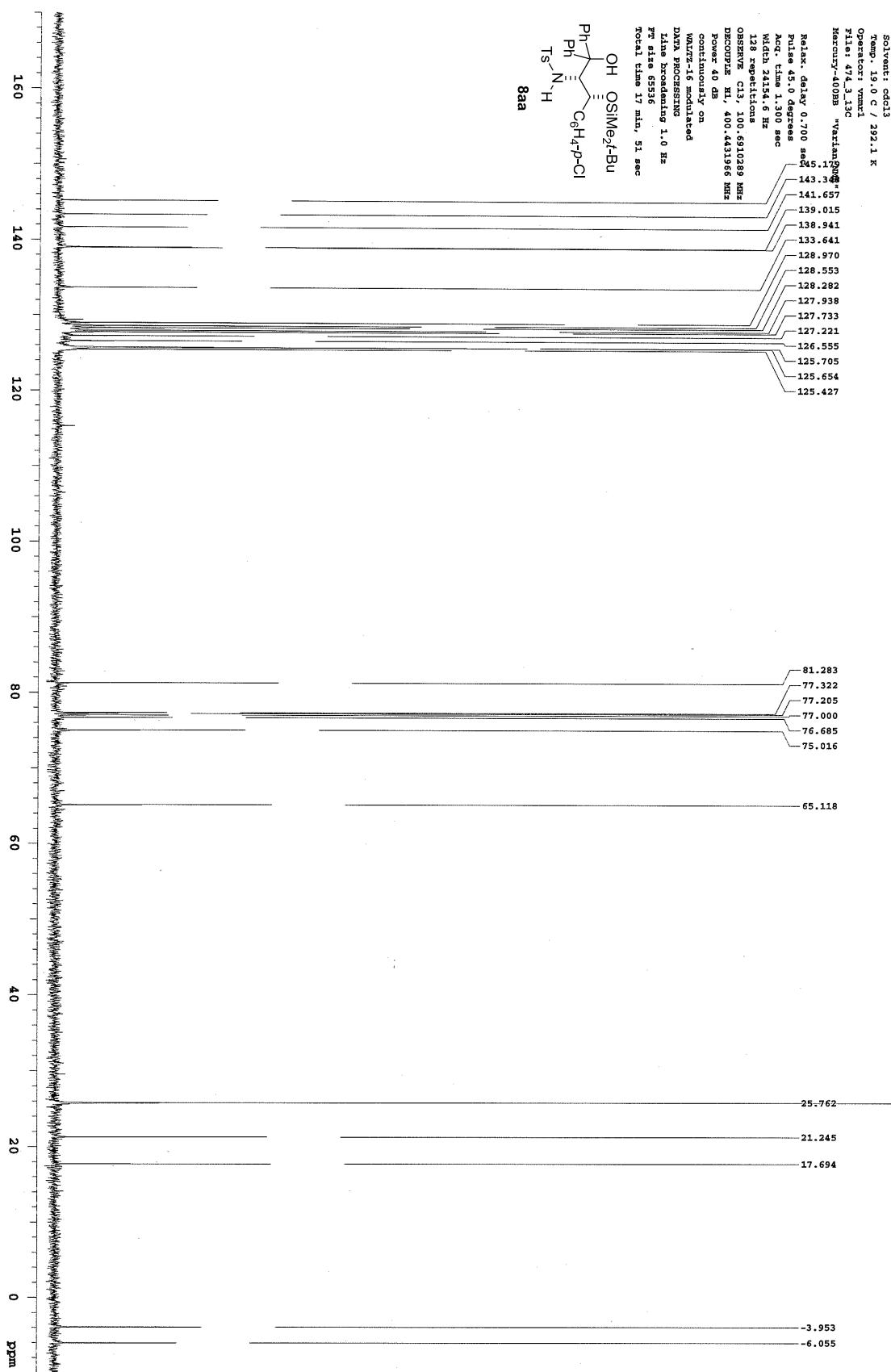
line broadening 1.0 Hz

RT size 65536

Total time 17 min, 51 sec



8aa



file: /home/vnmrl1/vnmrcys/data/murakami1\_lab/Nakamura/522\_1H\_2.fid

pulse sequence: s2pul

Solvent: cdcl<sub>3</sub>

Temp: 25.0 C / 288.1 K

Operator: vnmrl

File: 522\_1H\_2

Mercury-400B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acq. time 3.500 sec

Width 6402.0 Hz

16 repetitions

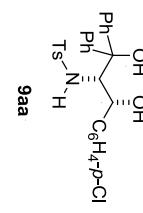
OBSERVE H1, 400.4411625 MHz

DATA PROCESSING

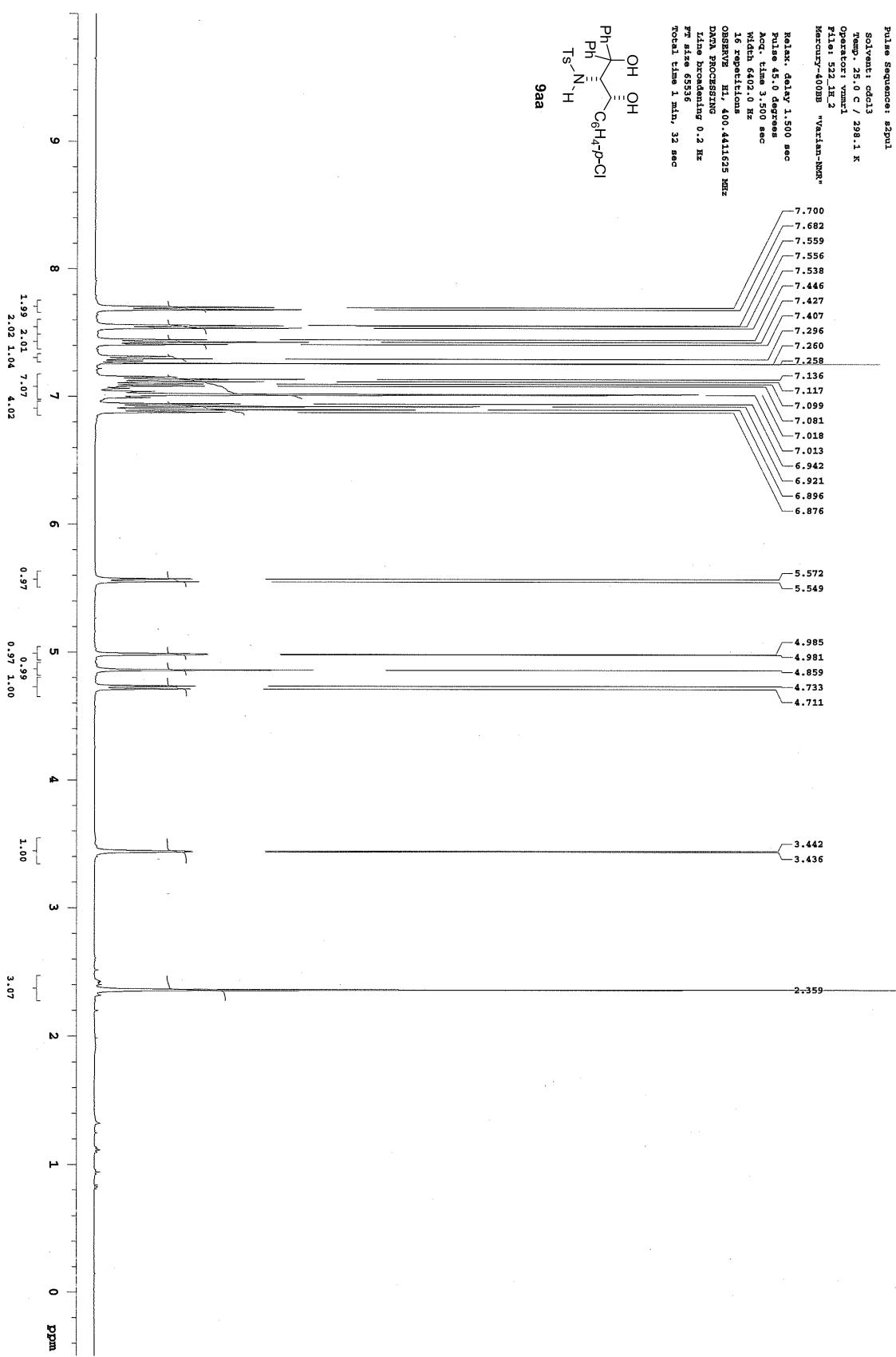
LINewidth 0.2 Hz

RT size 65536

Total time 1 min, 32 sec



9aa



File: /home/rmrt1/unixsys/data/murakami\_lab/Nakamura/522-13C.fid

Pulse Sequence: s2pul

Solvent: cdc13

Temp: 25.0 C / 238.1 K

Operator: "nmr1"

File: 522-13C

Mercury-400MHz "varian-NMR"

Relax: delay 0.700 sec

Pulse 45.0 degrees

Avg: time 1.300 sec

Width 24154.6 Hz

128 repetitions

Observe: C13, 100.6910230 MHz

Decouple: H1, 400.4431966 MHz

Power: 40 dB

continuously on

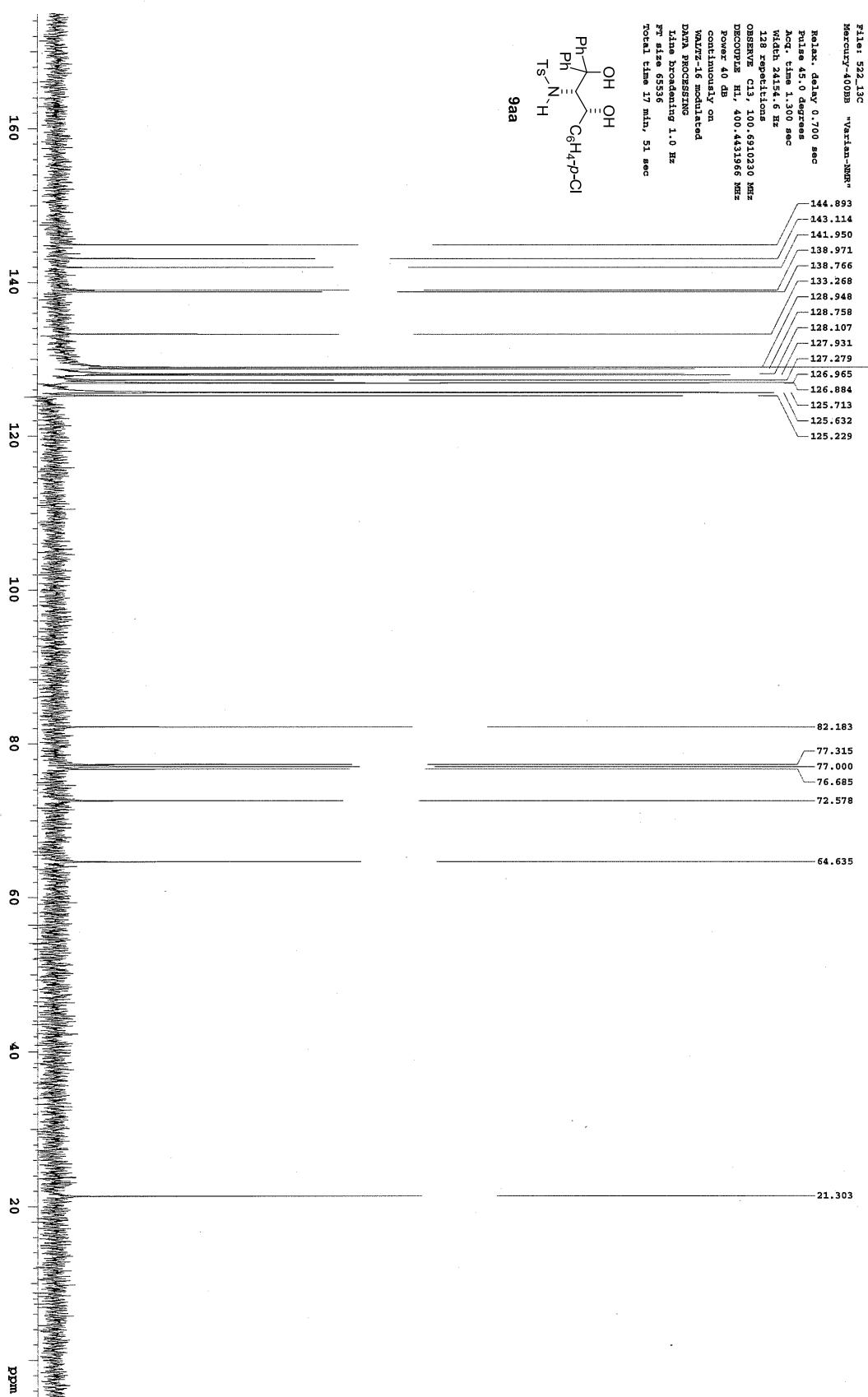
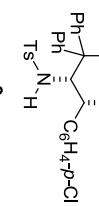
WALTZ-15 modulated

Data Processing

Line broadening 1.0 Hz

PP size 65536

Total time 17 min, 51 sec



file: /Home/vmncl1/vmnclsy/data/murakami\_lab/Nakamura/545\_benzene\_1H\_3.fid

Pulse Sequence: apsp1

Solvent: CDCl<sub>3</sub>

Temp: 23.0 C / 298.1 K

Operator: vmncl1

file: 545\_benzene\_1H\_3

MERCURY-00DBB "Varian-MER"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Avg. time 3.500 sec

Width 600.0 Hz

16 repetitions

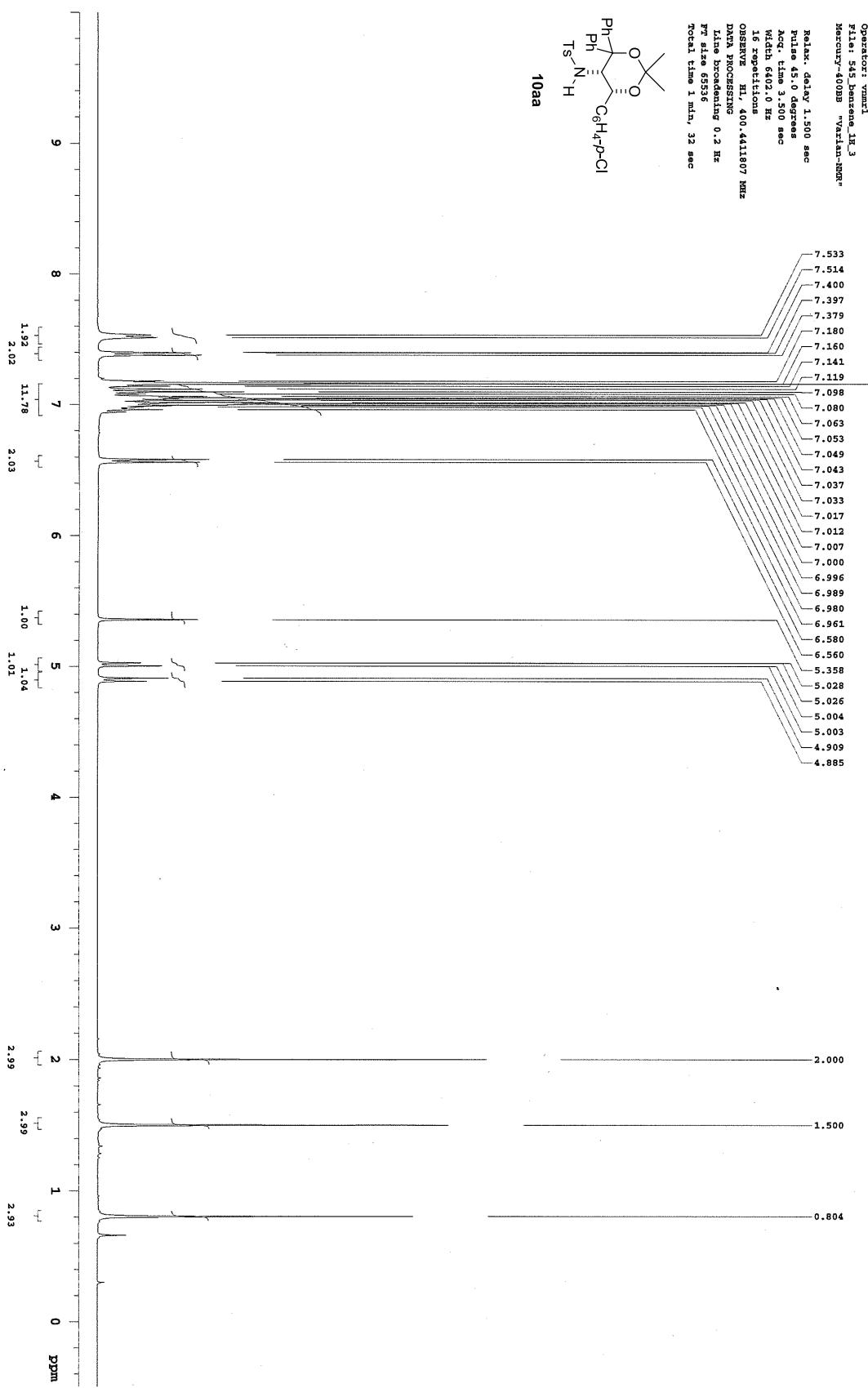
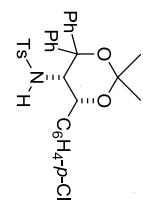
OBSERVE H, 400.4411807 MHz

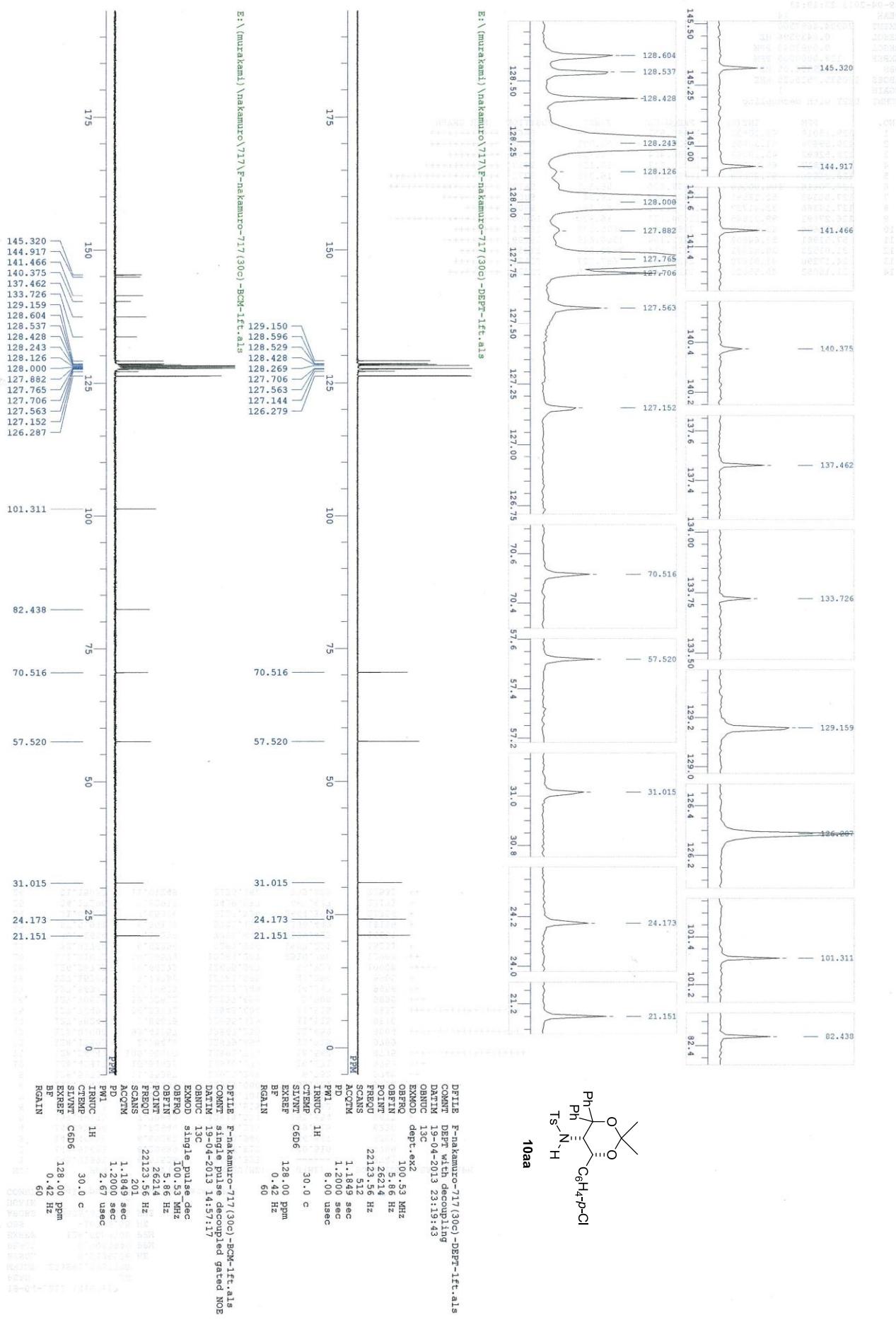
DATA PROCESSING

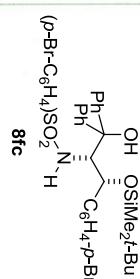
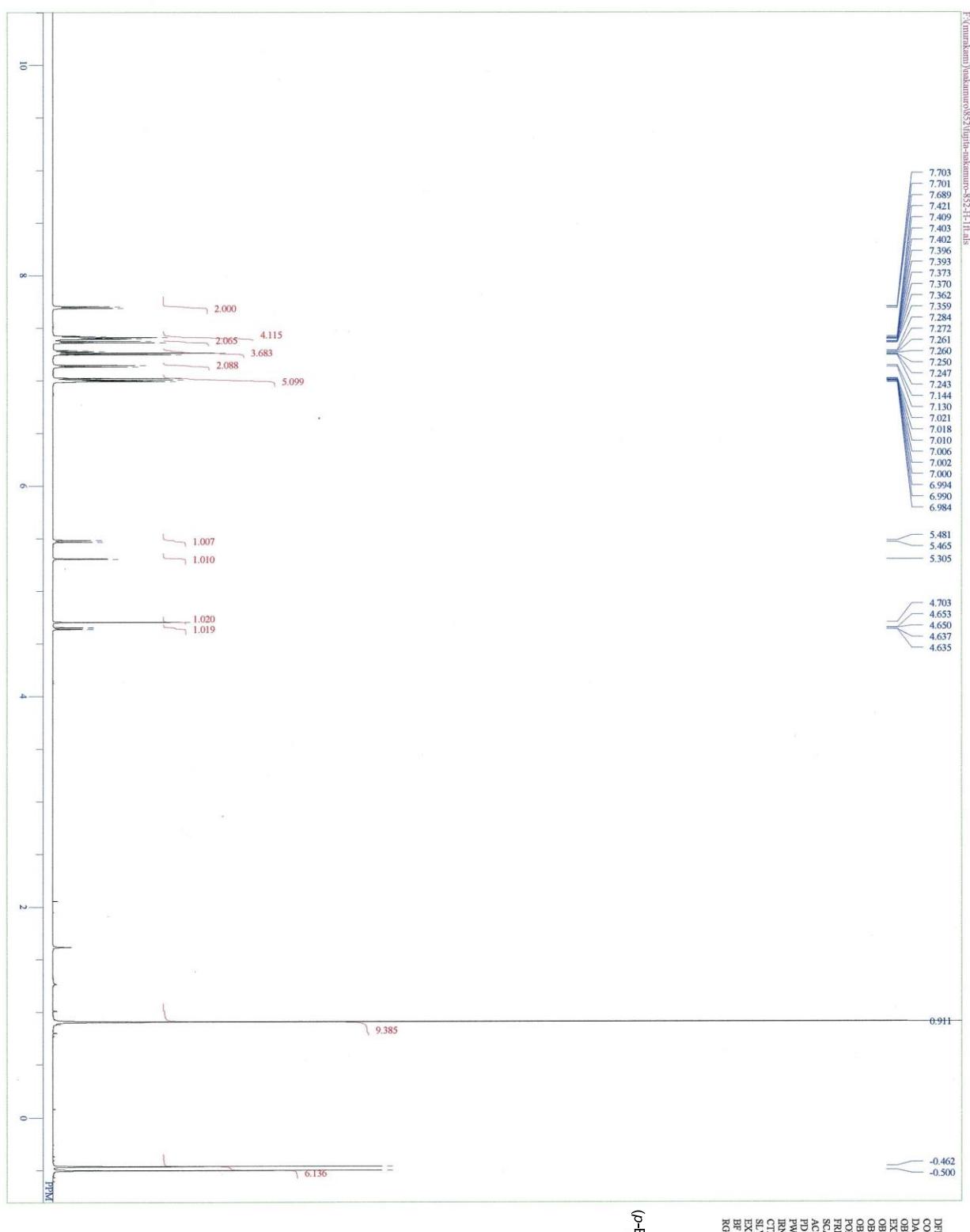
Line broadening 0.2 Hz

RT size 65336

Total time 1 min, 32 sec







E:\murakami\nakamuro\852\fujita-nakamuro-852-NNBB-1ft.als

1.02

1.18

1.14

0.92

1.02

1.02



DFT1E fujita-nakamuro-852-DEPT-1ft.als

DEPT with decoupling

DATIM 25-06-2013 17:41:15  
OBNUC 13C  
EXMO dept-ex2

OBPRO 150.92 MHz  
OPTIN 2.63 Hz  
POINT 131072

FREQ 33602.15 Hz  
SCANS 600  
ACQTIME 1.984 sec  
PWL 11.70 usec

CTAMP 1H  
IRUC 24.1 c  
SLVNT CDCL3

EXPEF 25.85 ppm  
BPP 0.75 Hz  
RGAIN 60

DFT1E fujita-nakamuro-852-NNBB-1ft.als  
CONT single pulse decoupled gated NO-

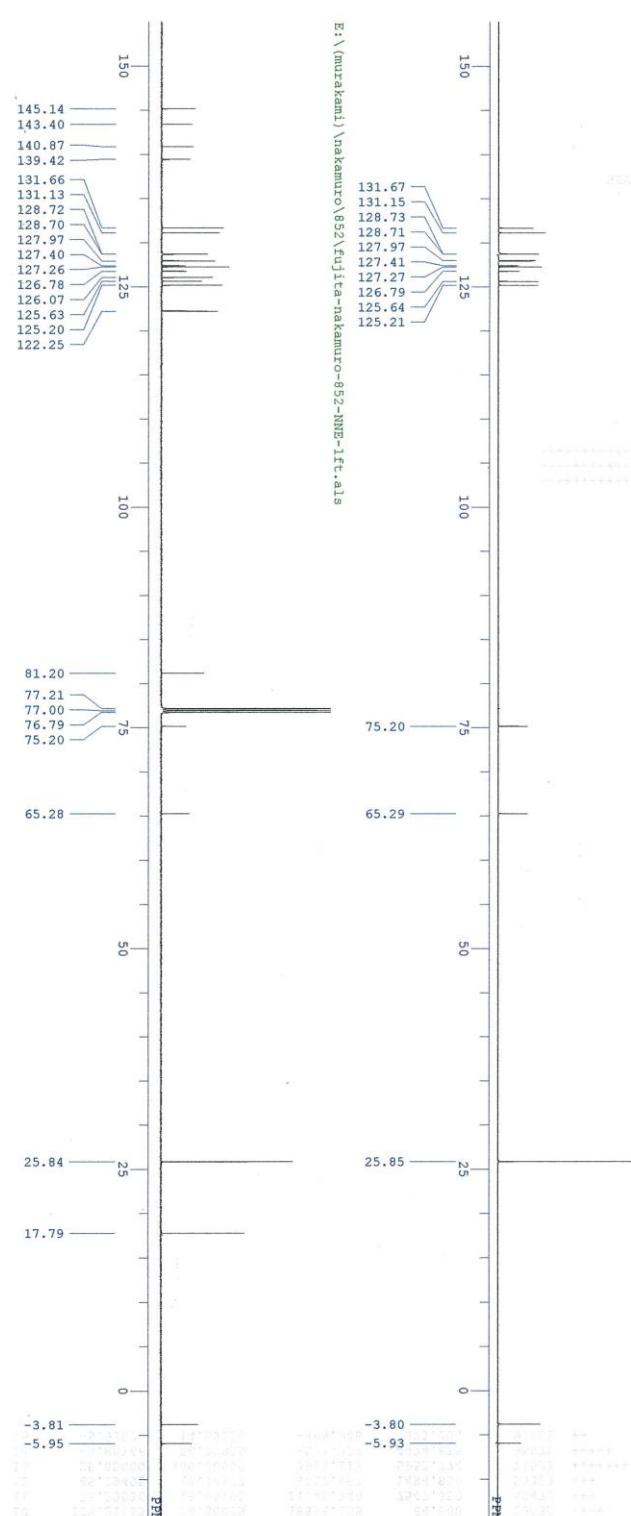
DATIM 26-06-2013 00:01:14  
OBNUC 13C  
EXMO single pulse dec

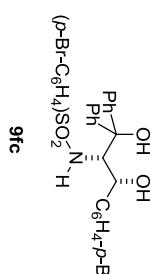
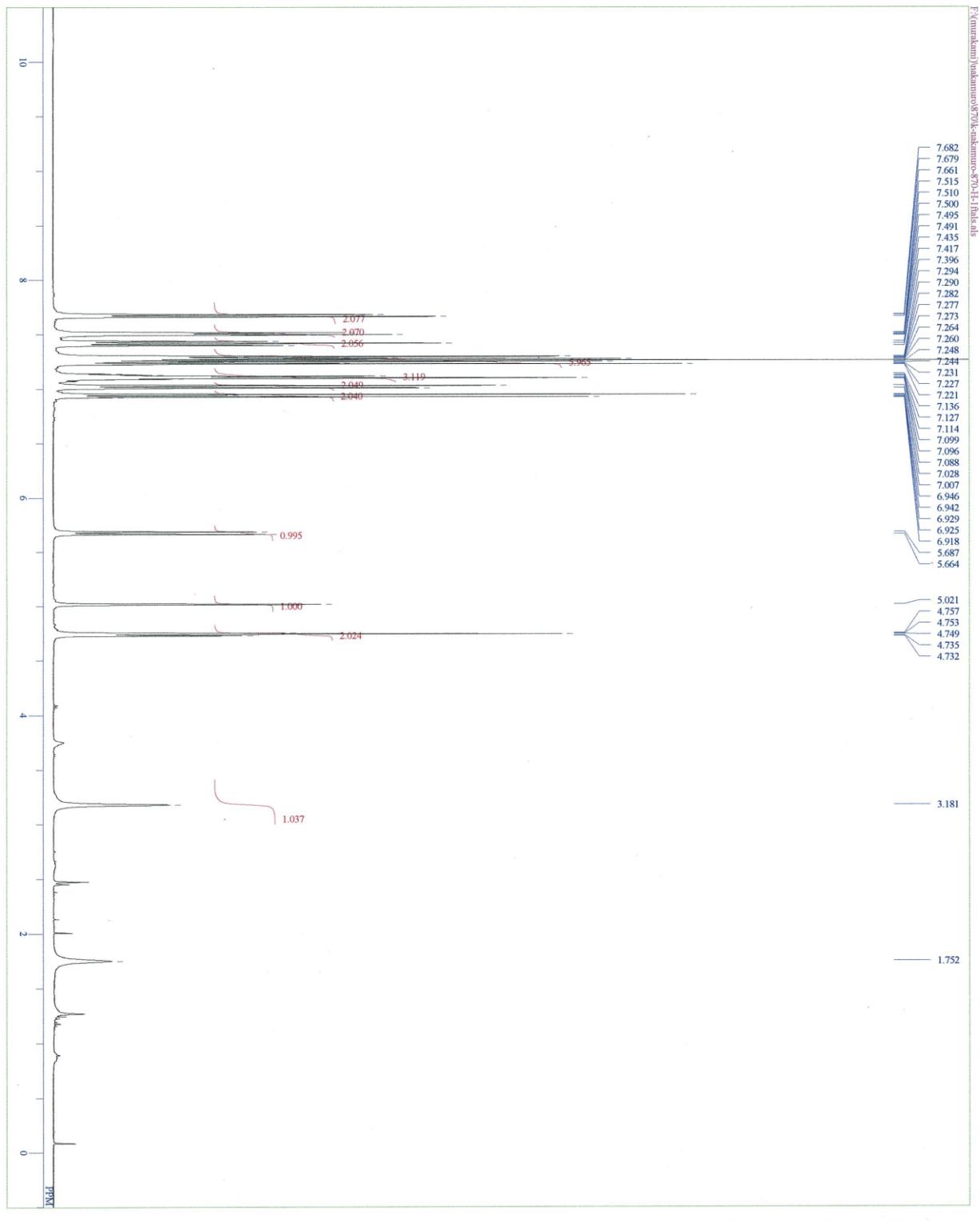
OBPRO 150.92 MHz  
OPTIN 2.63 Hz  
POINT 52428

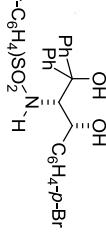
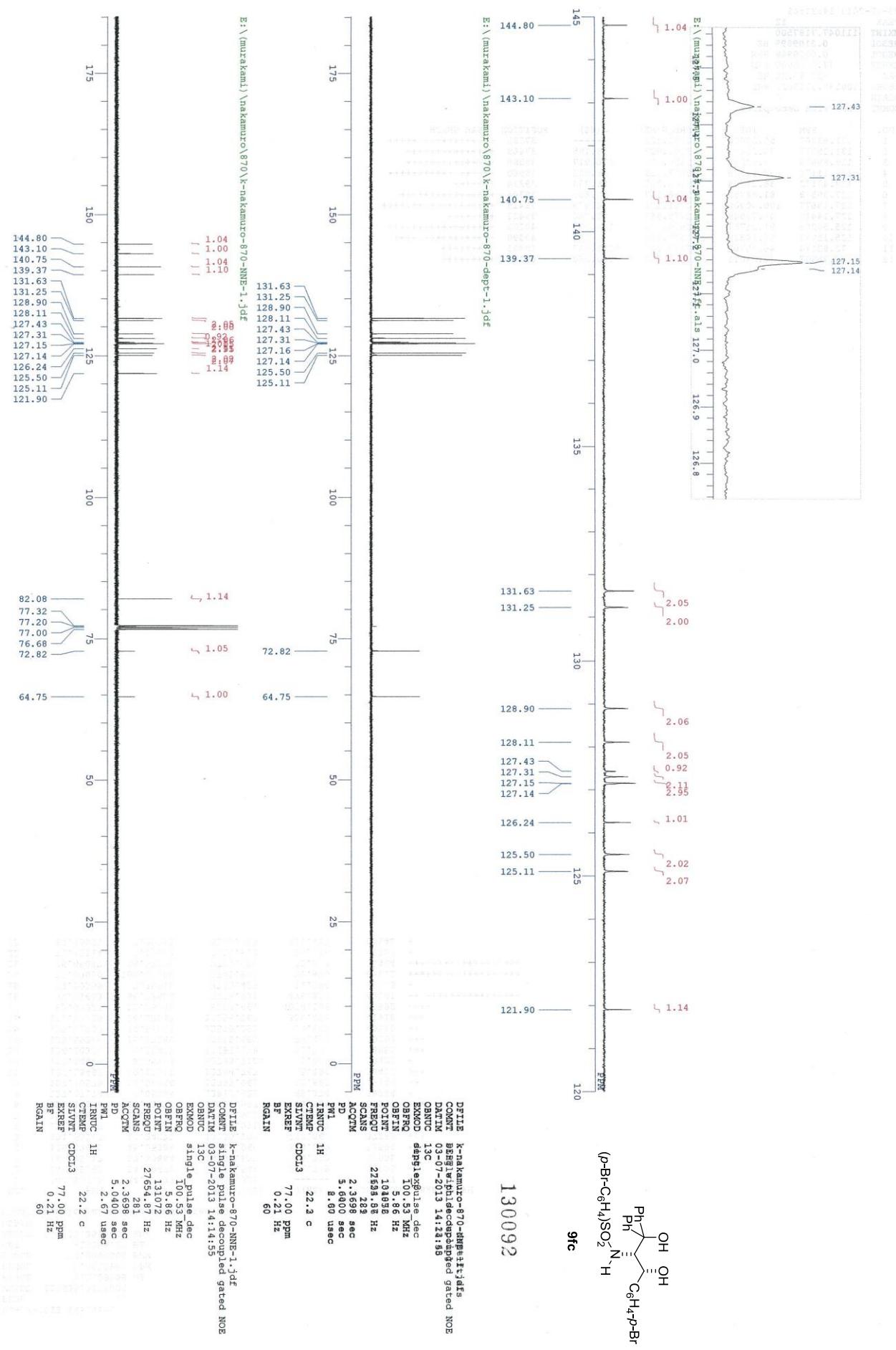
FREQ 26881.31 Hz  
SCANS 4000  
ACQTIME 1.9504 sec  
PWL 9.100 sec  
IRUC 3.90 usec

CTAMP 22.3 c  
SLVNT CDCL3

EXPEF 77.00 ppm  
BPP 0.26 Hz  
RGAIN 60

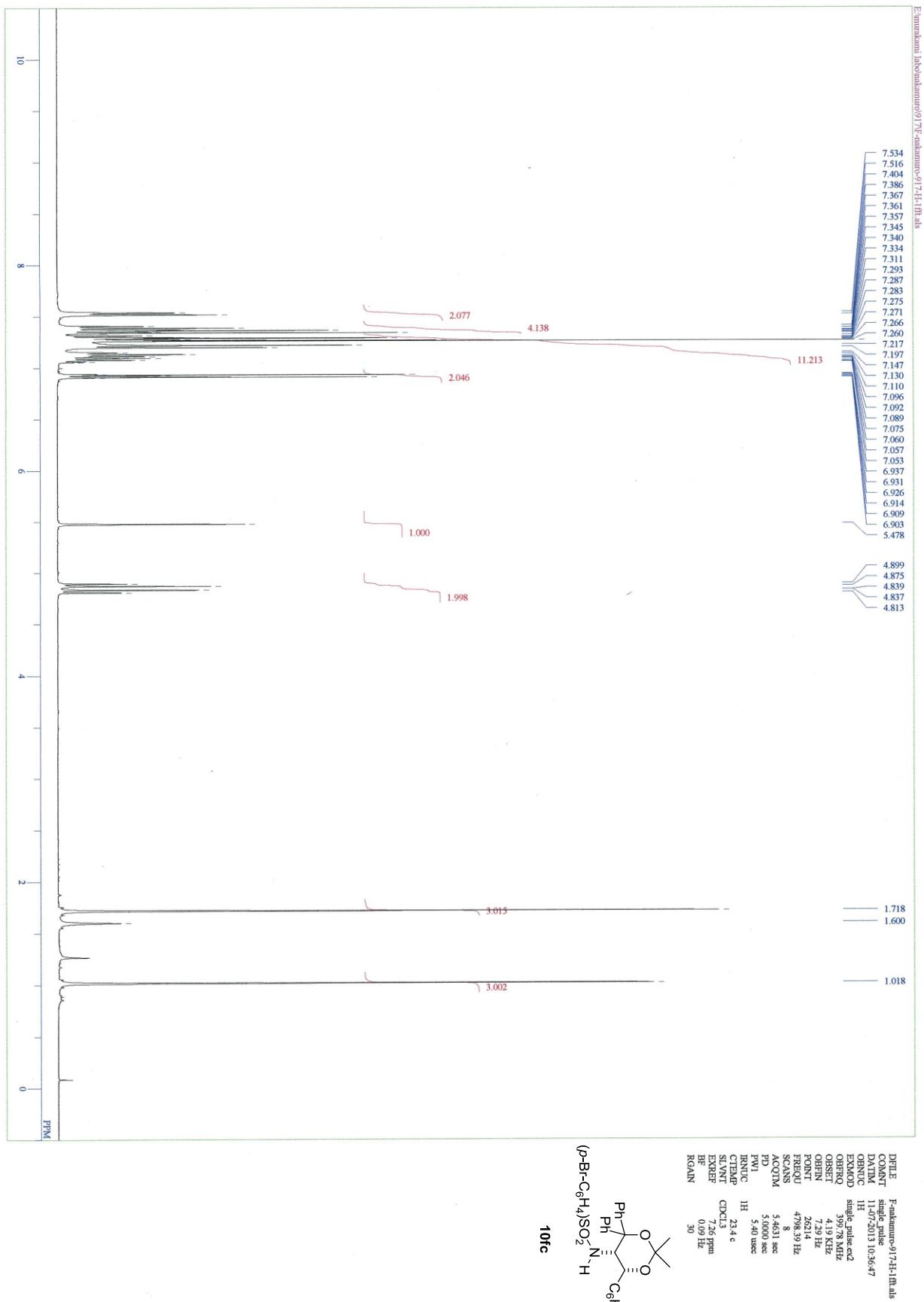


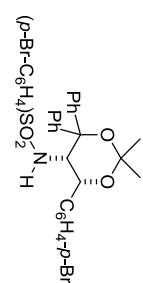
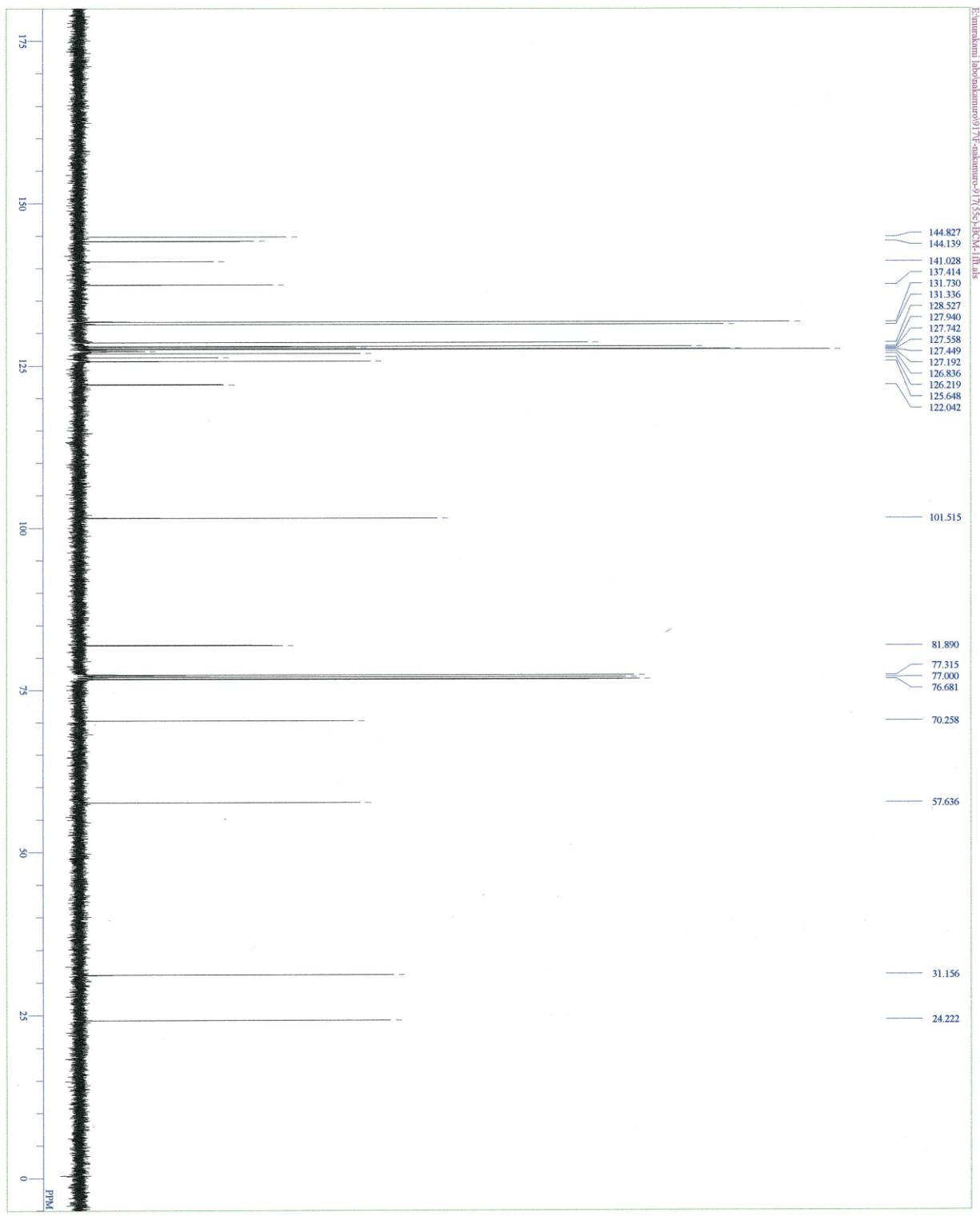




13092

S90





667 column

File: /home/vmarr1/vmarrsys/data/murakami\_lab/Nakamura/877\_1H.fid

Pulse Sequence: PREPAT

Solvent: cdcl<sub>3</sub>

Ambient temperature

Operator: vmar1

File: 677\_1H

Relax: 0.020 sec

Relax: 0.020 degrees

Relax: 1.998 sec

width 6410.3 Hz

16 repetitions

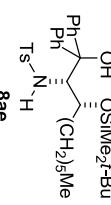
OBSERVE H1, 400.4411633 MHz

DATA PROCESSING

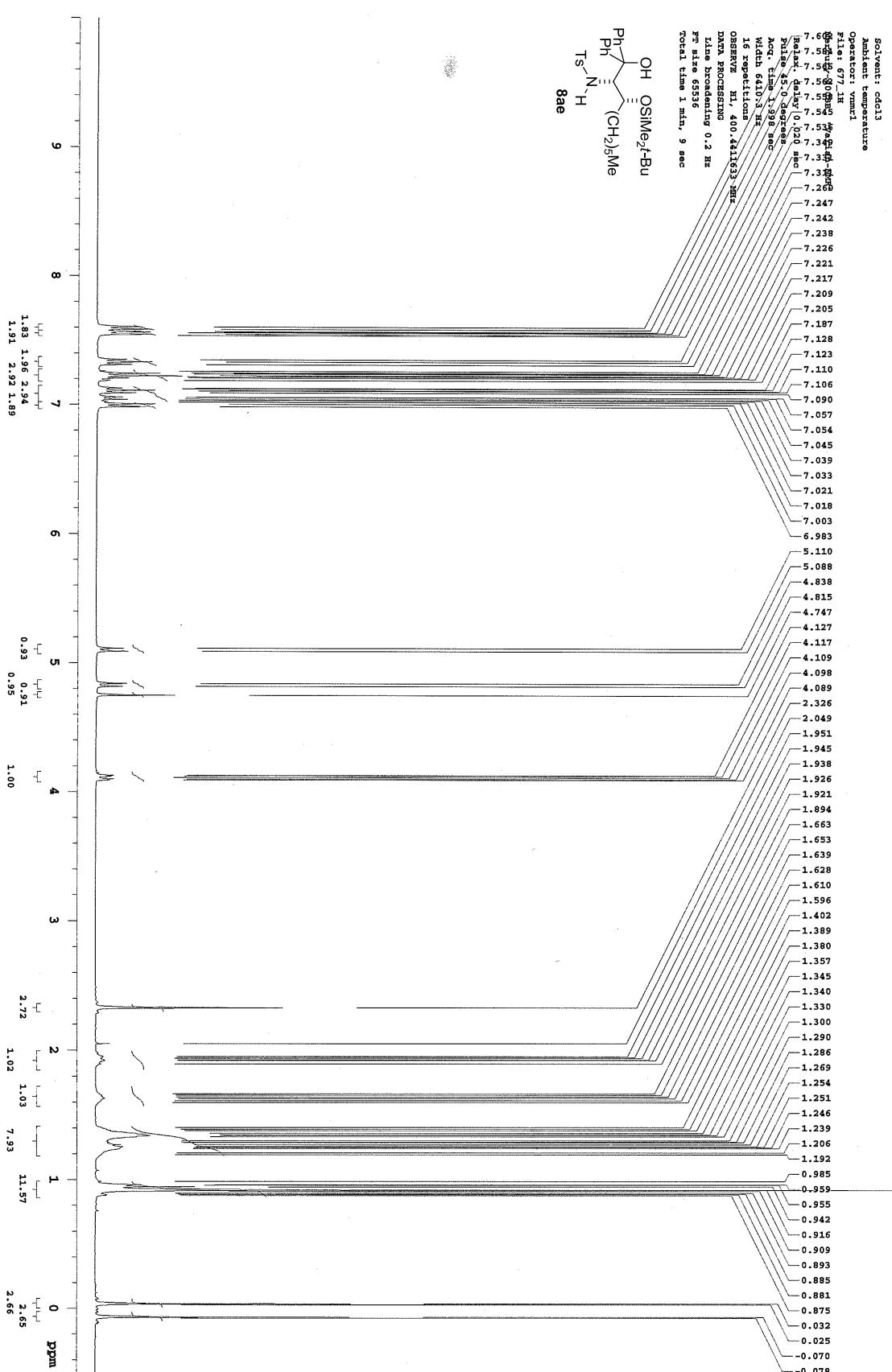
Line broadening 0.2 Hz

PP size 65536

Total time 1 min, 9 sec



8ae



667 column

File: /Home/vmrc1/vmrcsys/Data/murakami\_1ab/Nakamura/677\_13C.fid

Pulse Sequence: s2pul

Solvent: cdc13

Ambient temperature

Operator: vnmrc1

File: 677\_13C "Varian-NMR"

Mercury-400WB

Relax. delay 0.700 sec

Pulse 45.2 degrees

Acq. time 1.300 sec

Width 2415.6 Hz

64 repetitions

OBSERVE C13, 100.691238 MHz

DECOUPLE CH, 400.4431956 MHz

Power 40 dB

continuously on

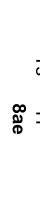
WAVELENGTH modulated

DATN PROCESSING

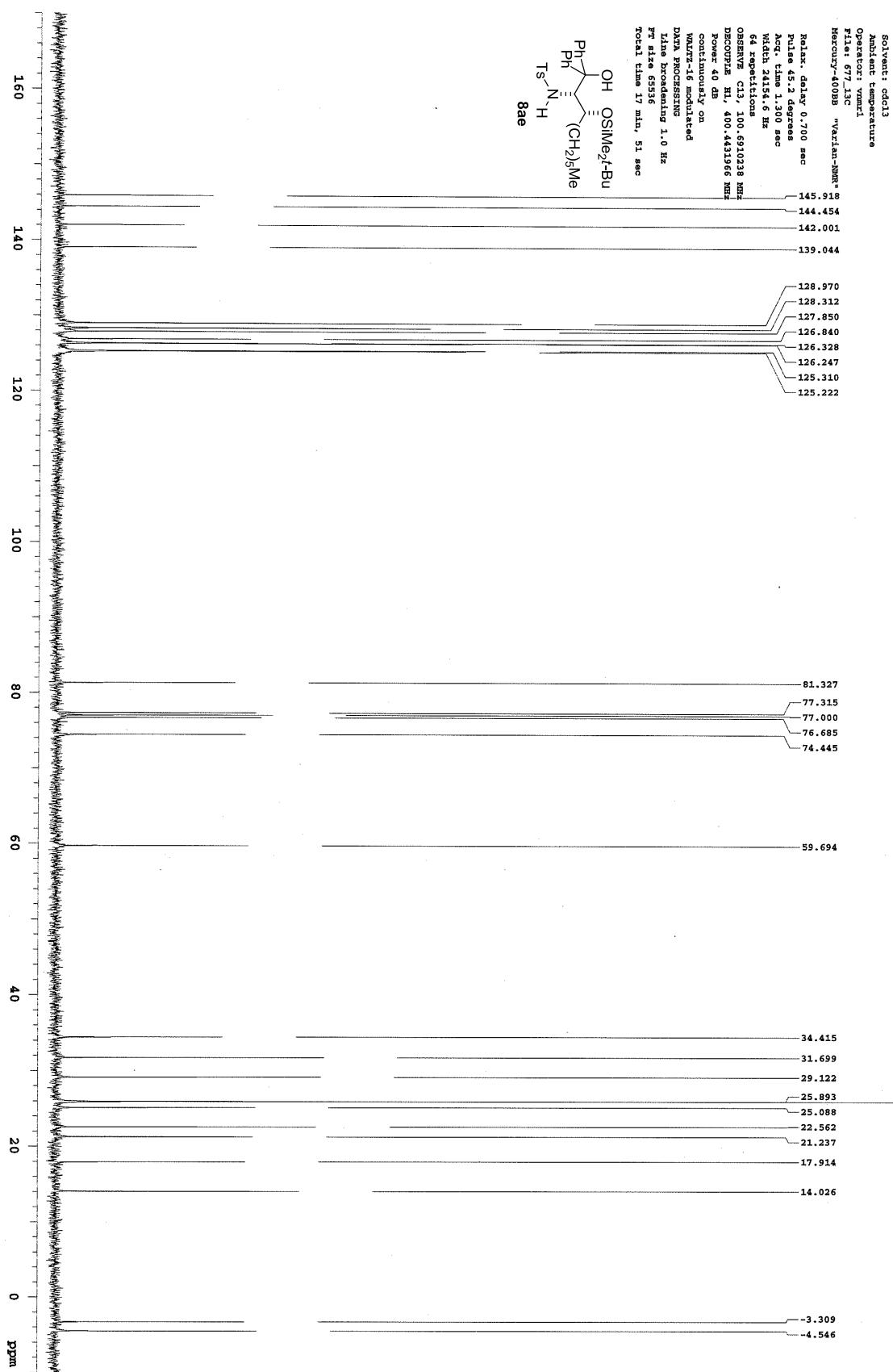
Line broadening 1.0 Hz

RT size 65536

Total time 17 min, 51 sec



8ae



682 ptic

File: /home/vmml1/vmmlsys/data/nakamuro/682\_1H.tid

Pulse Sequence: a2pul

Solvent: cdcl3

Ambient temperature

Operator: vml1

File: 682\_1H Mercury-4012B "Varian-NMR"

Relax. delay 1.500 sec

Pulse 45.0 degrees

Acc. time 3.500 sec

Width 6402.0 Hz

16 repetitions

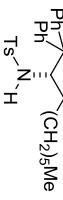
OBSERVE H1 400.4411638 MHz

DATA PROCESSING

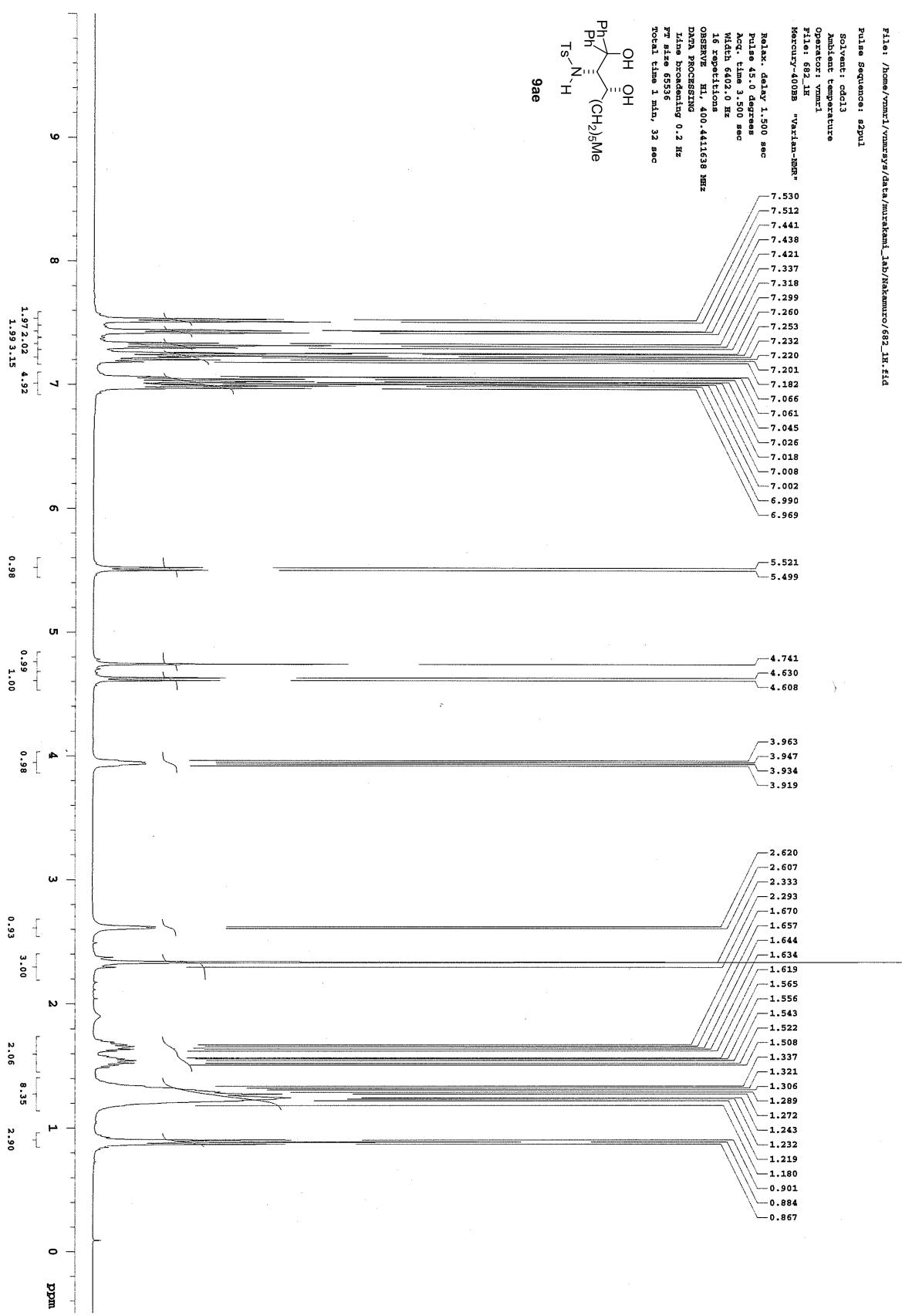
line broadening 0.2 Hz

PPM size 65536

Total time 1 min., 32 sec



9ae



682.pt1c

File: /home/vmml1/vmmcrsys/data/nakamani\_1ab/Nakamizo/682-13C.fid

Pulse Sequence: s2pul

Solvent: cdcl3

Ambient temperature

Operator: vmmrl

File: 682-13C

Mercury-4000B "Varian-NMR"

Relax: delay 0.700 sec

Pulse 45.2 degrees

Acq. time 1.300 sec

Width 24.14.6 Hz

128 repetitions

OBSERVE C13, 100.6910171 MHz

DICOUPLE H1, 400.4431966 MHz

Power 40 dB

continuously on

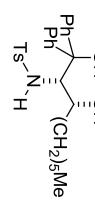
WATER-1H modulated

DATA PROCESSING

Line broadening 1.0 Hz

FFT size 65536

Total time 17 min, 51 sec



9ae

