

Supporting Information

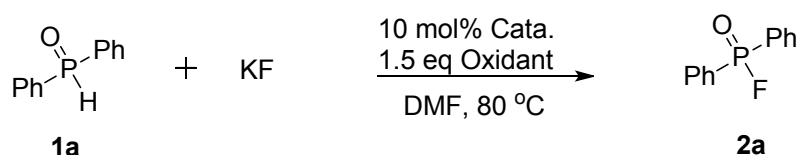
Copper-promoted Oxidative –Fluorination of Arylphosphine under Mild Condition

Na Liu,^a Liu-Liang Mao,^a Bin Yang,^a Shang-Dong Yang^{*a,b}

1. General information and materials

NMR spectra were obtained using a Bruker advance III 400 spectrometer. Chemical shifts are reported in units of parts per million (ppm) downfield from tetramethylsilane (TMS) and CDCl₃, and all coupling constants are reported in hertz. The infrared spectra were obtained with KBr plates using Nexus 670 FT-IR spectrometer and only major peaks are reported in cm⁻¹. Mass spectra were measured using Bruker microTOF-Q II MS and measured in EI or ESI mode. The starting materials were purchased from Aldrich, Acros Organics, J&K Chemicals or TCI and used without further purification. DMF was purified and dried according to the procedure from "Purification of Laboratory Chemicals book". Column chromatography was carried out on silica gel (particle size 200-400 mesh ASTM). Substrates **1** were prepared according to the literature procedure,^{1,2} **2n** was described in previous report.³

2. Table S1 Optimization different Catalysts and Oxidant^{ab}



Entry	Cata.(mol%)	Fsource(equive)	Oxidant(equive)	Yield(%)
1	CuI (10)	KF (1.5)	DDQ (1.5)	69
2	CuCl ₂ (10)	KF (1.5)	DDQ (1.5)	75
3	CuO (10)	KF (1.5)	DDQ (1.5)	59
4	CuBr ₂ (10)	KF (1.5)	DDQ (1.5)	80
5	Cu(OAc) ₂ (10)	KF (1.5)	DDQ (1.5)	64
6	PdCl ₂ (10)	KF (1.5)	DDQ (1.5)	72
7	Pd(TFA) ₂ (10)	KF (1.5)	DDQ (1.5)	69
8	Pd(OAc) ₂ (10)	KF (1.5)	DDQ (1.5)	65
9	Pd(acac) ₂ (10)	KF (1.5)	DDQ (1.5)	56
10	FeCl ₃ (10)	KF (1.5)	DDQ (1.5)	52
11	Ag ₂ CO ₃ (10)	KF (1.5)	DDQ (1.5)	49
12	AgTFA (10)	KF (1.5)	DDQ (1.5)	58
13	AgF (10)	KF (1.5)	DDQ (1.5)	55
14	AgOAc (10)	KF (1.5)	DDQ (1.5)	58
15	AgOTf (10)	KF (1.5)	DDQ (1.5)	60
16	CuBr ₂ (10)	KF (1.5)	K ₂ S ₂ O ₈ (1.5)	33
17	CuBr ₂ (10)	KF (1.5)	Cu(OAc) ₂ (1.5)	n.d.
18	CuBr ₂ (10)	KF (1.5)	AgOAc (1.5)	17
18	CuBr ₂ (10)	KF (1.5)	^t BuOOH (1.5)	trace
20	CuBr ₂ (10)	KF (1.5)	CNA (1.5)	19
21	CuBr ₂ (10)	KF (1.5)	PhI(OAc) ₂ (1.5)	38
22	CuBr ₂ (10)	KF (1.5)	Oxone (1.5)	n.d.
23	CuBr ₂ (10)	KF (1.5)	BQ (1.5)	n.d.
24		KF (1.5)	DDQ (1.5)	47
25 ^c	ZnBr ₂ (5.0)	NaF (1.5)	DDQ (1.5)	57

26 ^c	Zn(OTf) ₂ (7.5)	NaF (1.5)	DDQ 1.5)	70
27 ^c	ZnCl ₂ (5.0)	NaF (1.5)	DDQ 1.5)	69

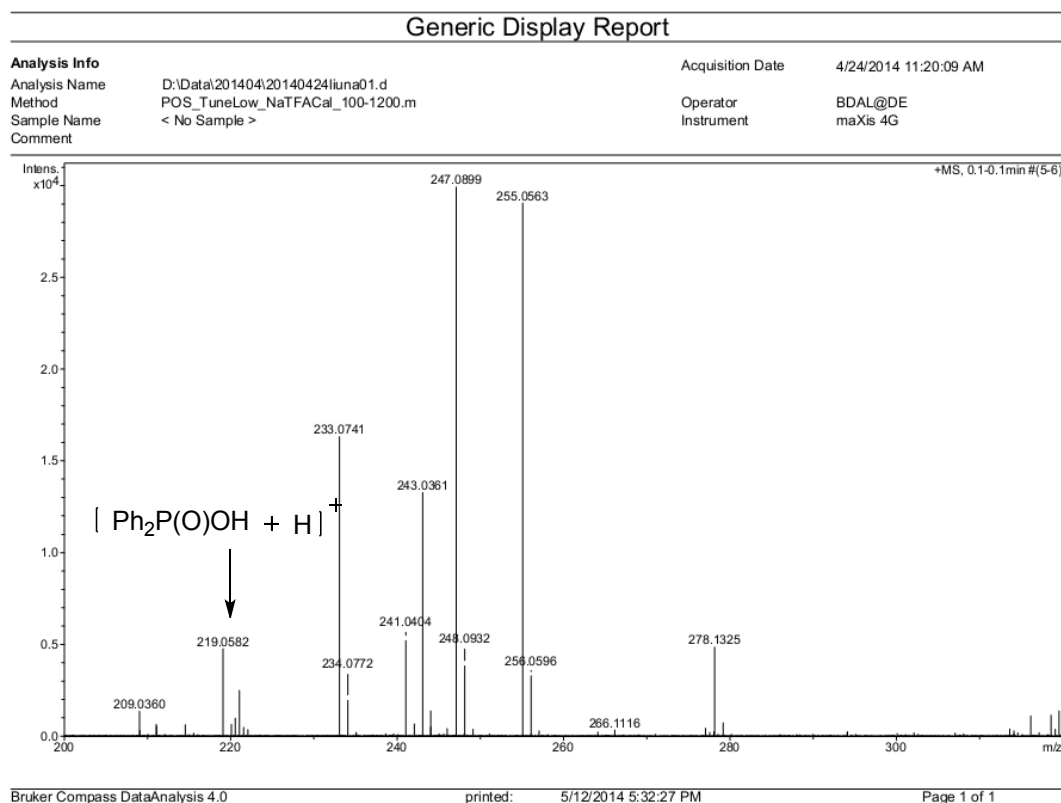
^a All the reaction were carried out with Catalysts (10 mol%) in the presence of **1a** (0.3 mmol), KF (1.5 eq) and DDQ (1.5 eq) in 2.0 mL DMF at 80 °C under argon for 4 h. ^b Isolated yield. ^c R.T.

3. Preliminary mechanistic studies.

3.1 Analytical data of ESI-MS:

In a reaction tube, diphosphine oxide **1a** (0.30 mmol), NaF (0.45 mmol), DDQ (0.45 mmol), and CuBr₂ (0.015 mmol) were added and charged with Ar three times. Then, anhydrous DMF (1 mL) were added. The reaction mixture was stirred at RT for 10 min with vigorous stirring. The reaction was diluted with DMF(1/100) prior to the injection into the mass spectrometer.

The positive-ion mode of ESI-MS spectrum showed the signals corresponding to the A (m/z) 427.0441, 428.0471, 429.0481. The signal might mean that DDQ can easily react with diphenylphosphine oxide to form the intermediate A. On the other hand, a signal of m/z 219.0582 may be the Diphenylphosphinic acid. The signal of m/z 419.0986 illustrated that intermediate A easily react with Diphenylphosphinic acid to generate the molecular B, which can react with NaF to the final products.



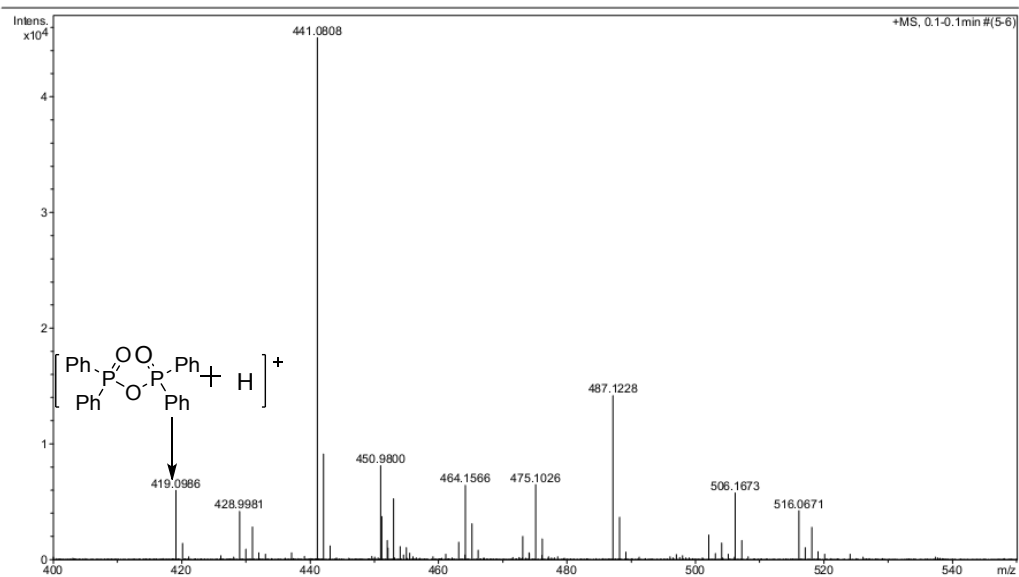
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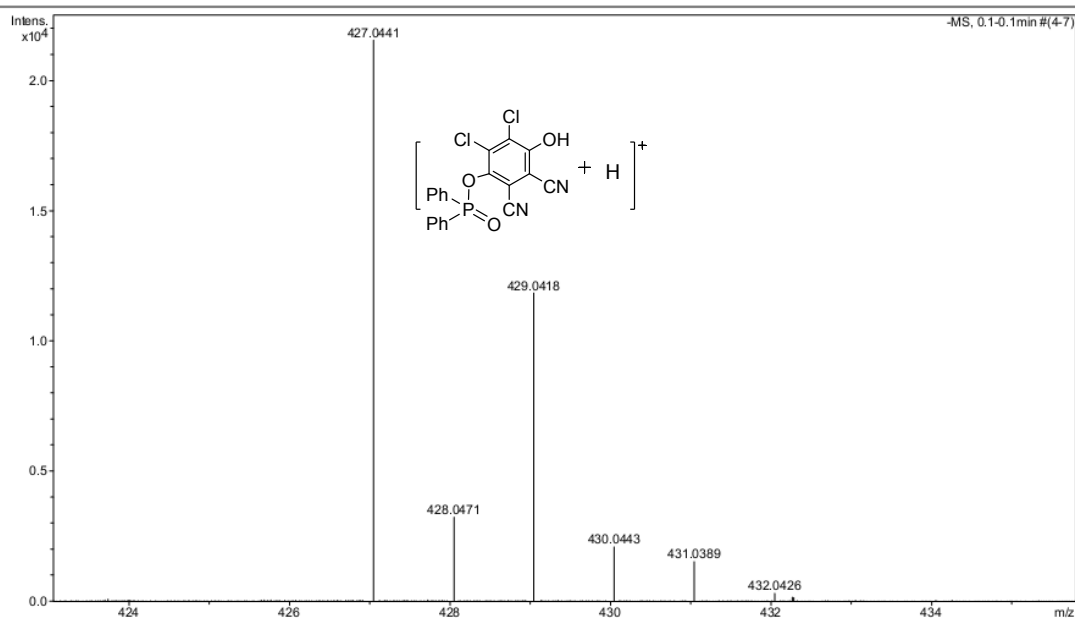
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Figure S1. ESI-MS spectrum of the reaction solution.

We used $[\text{Ph}_2\text{P}(\text{O})]_2\text{O}$ as the substrate under the reaction conditions resulted in the yield of 95% at 60 °C, but in the absence of the DDQ, only obtained 11% yield at 60 °C [Scheme S1, Eq. (1), (2)]. When the reaction was degassed under conditions that the product was obtained in the yield of 85% [Scheme S1, Eq. (3)]. Chemical trapping of radicals using TEMPO (a well-known radical-trapping reagent) and AIBN under the reaction conditions [Scheme S1, Eq. (4), (5)]. As illustrated in Scheme S1, the addition of 3.0 equivalents of TEMPO and AIBN were performed under the standard condition. When TEMPO was added, the yield was decreased. Above all the reactions were carried out, the results were illustrated that the reaction was not go through the radical processure.

4. General Procedure for the Copper-promoted Oxidative-Fluorination of Arylphosphine.

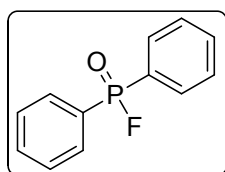
Method A: In a reaction tube, arylphosphine oxides **1** (0.30 mmol), NaF (0.45 mmol), DDQ (0.45 mmol), and CuBr_2 (0.015 mmol, 5.0 mol %) were added and charged with Ar three times. Then, anhydrous DMF (1 mL) were added. The mixture was stirred at room temperature for 4 h, until the reaction was completed. DMF was distilled under reduced pressure. The crude product was purified by flash chromatography on silica gel to give the desired products (PE : EA = 4:1).

Method B: In a reaction tube, arylphosphine oxides **1** (0.30 mmol), DDQ (0.45 mmol), and CuBr_2 (0.015 mmol, 5.0 mol %) were added and charged with Ar three times. Then, anhydrous DMF (1 mL) were added. The mixture was stirred at room temperature for 20 min, the nucleophilic was added slowly, the mixture was stirred until the reaction was completed. DMF was distilled under reduced pressure, The crude product was purified by flash chromatography on silica gel to give the desired products (PE : EA = 2:1).

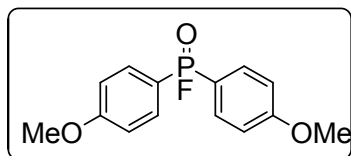
References:

1. S. Montel, T. Jia, P. J. Walsh. *Org. Lett.* 2014, **16**, 130-133.
2. Q. Xu, C.-Q. Zhao, L.-B. Han. *J. Am. Chem. Soc.* 2008, **130**, 12648-12655.
3. H. D. Durst, D. K. Rohrbaugh, P. Smith, J. M. Nilles, T. Connell, J. A. Laramee, S. Munavalli. *Phosphorus, Sulfur and Silicon.* 2008, **183**, 2655-2668.

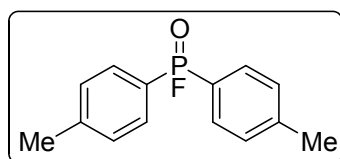
5. Characterization of new compounds.



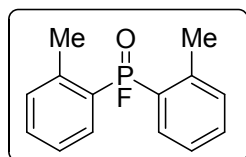
2a Yield: 90%, pale yellow oil. ^1H NMR (400 MHz, CDCl_3) δ 7.90 – 7.77 (m, 4H), 7.69 – 7.57 (m, 4H), 7.51 (m, 4H). ^{31}P NMR (400 MHz, CDCl_3) δ 40.94 (d, $J = 1019.7$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -75.17 (d, $J = 1019.8$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 133.35, 133.33, 131.45, 131.43, 131.34, 131.32, 129.38 (d, $J = 22.3$ Hz), 128.87, 128.73, 127.97 (d, $J = 22.3$ Hz). MS : m/z (M+H): 221.



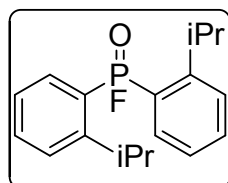
2b Yield: 95%, faint orange solid. ^1H NMR (400 MHz, CDCl_3) δ 7.74 (m, 4H), 6.99 (m, 4H), 3.85 (s, 6H). ^{31}P NMR (400 MHz, CDCl_3) δ 42.41 (d, $J = 1008.3$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -72.64 (d, $J = 1008.4$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 163.34, 163.31, 133.34, 133.32, 133.21, 133.19, 120.89 (d, $J = 23.9$ Hz), 119.40 (d, $J = 23.9$ Hz), 114.31, 114.16, 55.31. IR (film/ cm^{-1}) ν_{max} : 3393, 2935, 2842, 1599, 1505, 1299, 1255, 1135, 1183. MS : m/z (M+H): 281.



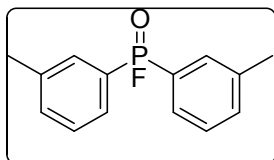
2c Yield: 93%, white solid. ^1H NMR (400 MHz, CDCl_3) δ 7.70 (m, 4H), 7.30 (m, 4H), 2.40 (s, 6H). ^{31}P NMR (400 MHz, CDCl_3) δ 42.11 (d, $J = 1014.1$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -74.33 (d, $J = 1014.3$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 143.97, 143.94, 131.37, 131.35, 131.25, 131.23, 129.48, 129.34, 126.47 (d, $J = 22.8$ Hz), 125.04 (d, $J = 22.8$ Hz), 21.63. IR (film/ cm^{-1}) ν_{max} : 3392, 2923, 2866, 1604, 1449, 1261, 1214, 1180, 1132, 1113. MS : m/z (M+H): 249.



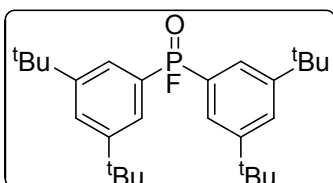
2e Yield: 75%, pale yellow solid. ^1H NMR (400 MHz, CDCl_3) δ 7.80 (m, 2H), 7.50 (t, $J = 7.6$ Hz, 2H), 7.30 (m, 4H), 2.44 (s, 6H). ^{31}P NMR (400 MHz, CDCl_3) δ 43.10 (d, $J = 1018.4$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -74.57 (d, $J = 1018.6$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 141.75, 141.63, 133.30, 133.27, 132.94, 132.91, 132.83, 132.79, 131.58, 131.45, 128.26 (d, $J = 19.1$ Hz), 126.91 (d, $J = 19.1$ Hz), 125.83, 125.69, 21.11, 21.07, 21.06. IR (film/ cm^{-1}) ν_{max} : 3555, 3488, 3061, 2968, 2928, 1595, 1476, 1454, 1284, 1256, 1147, 758. MS : m/z (M+H): 249.



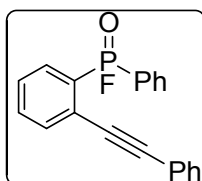
2f Yield: 18%, white solid. ^1H NMR (400 MHz, CDCl_3) δ 7.79 (dt, $J = 16.0, 8.2$ Hz, 2H), 7.57 (t, $J = 7.6$ Hz, 2H), 7.49 – 7.41 (m, 2H), 7.31 (tt, $J = 8.9, 4.4$ Hz, 2H), 3.44 (dq, $J = 13.4, 6.7$ Hz, 2H), 1.08 (dd, $J = 12.3, 6.7$ Hz, 12H). ^{31}P NMR (162 MHz, CDCl_3) δ 43.50 (d, $J = 1017.2$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -71.19 (d, $J = 1017.5$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 153.20, 153.07, 133.57, 133.54, 132.61, 132.57, 132.49, 132.45, 128.15 (d, $J = 18.5$ Hz), 126.95, 126.88, 126.82, 126.79 (d, $J = 18.5$ Hz), 125.95, 125.81, 31.61, 31.56, 23.83, 23.77. IR (film/ cm^{-1}) ν_{max} : 3404, 2967, 2929, 1594, 1442, 1251, 1134, 830, 769. MS : m/z (M+H): 305.



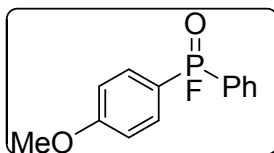
2g Yeld:80%, yellow oil. ^1H NMR (400 MHz, CDCl_3) δ 7.70 – 7.56 (m, 4H), 7.44 – 7.35 (m, 4H), 2.40 (s, 6H). ^{31}P NMR (162 MHz, CDCl_3) δ 41.71 (d, $J = 1019.7$ Hz). ^{19}F NMR (376 MHz, CDCl_3) δ -75.51 (d, $J = 1022.2$ Hz). ^{13}C NMR (101 MHz, CDCl_3) δ 138.81, 138.67, 134.09, 134.06, 131.81, 131.79, 131.70, 131.68, 129.31 (d, $J = 22.1$ Hz), 128.73, 128.58, 128.49, 128.47, 128.38, 128.36, 127.91 (d, $J = 22.0$ Hz), 21.29. IR(film/ cm^{-1}) ν_{max} 3052, 2924, 2857, 1599, 1480, 1262, 1228, 1124.705, 567. MS : m/z (M+H): 249.



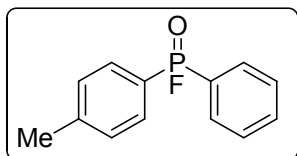
2h Yeld:55%, white solid. ^1H NMR (400 MHz, CDCl_3) δ 7.70 (t, $J = 5.2$ Hz, 2), 7.67 (s, 4), 1.33 (s, 36) ^{31}P NMR (400 MHz, CDCl_3) δ 43.64 (d, $J = 1017.0$ Hz). ^{19}F NMR (400MHz Cl_3) δ -72.41 (d, $J = 1017.2$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 151.45, 151.32, 128.76 (d, $J = 21.8$ Hz), 127.43, 127.40, 127.38 (d, $J = 21.8$ Hz) 125.61, 125.60, 125.49, 125.48, 35.02, 31.22. IR(film/ cm^{-1}) ν_{max} 3057, 2959, 905, 2870, 1594, 1477, 1427, 1364, 1255, 1154, 916, 878, 603. MS : m/z (M+H): 445.



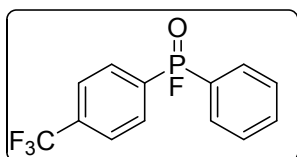
2i Yeld:73%, yellow oil ^1H NMR (400 MHz, CDCl_3) δ 8.11 (m, 1H), 7.91 – 7.79 (m, 2H), 7.73 – 7.47 (m, 4H), 7.43 (m, 2H), 7.38 – 7.28 (m, 5H). ^{31}P NMR (400 MHz, CDCl_3) δ 39.56 (d, $J = 1022.3$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -76.31 (d, $J = 1022.3$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 133.65, 133.54, 133.14, 133.11, 133.06, 133.02, 131.49, 131.44, 131.43, 131.33, 131.31, 130.56 (d, $J = 19.4$ Hz), 129.68 (d, $J = 21.0$ Hz), 128.89, 128.56, 128.42, 128.24, 128.10, 127.97, 126.25, 126.15, 122.15, 96.49, 86.82, 86.76. IR(film/ cm^{-1}) ν_{max} 3402, 3060, 2925, 2855, 1588, 1492, 1440, 1257, 1140, 757, 692, 532. MS : m/z (M+H): 321.



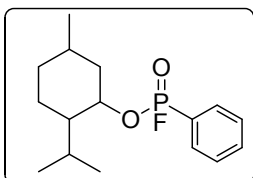
2j Yeld:84%, pale yellow oil. ^1H NMR (400 MHz, CDCl_3) δ 7.85 – 7.68 (m, 5H), 7.63 – 7.54 (m, 1H), 7.49 (td, $J = 7.6, 3.9$ Hz, 2H), 7.00 (dd, $J = 8.7, 2.8$ Hz, 2H), 3.85 (s, 3H). ^{31}P NMR (400 MHz, CDCl_3) δ 41.64 (d, $J = 1014.3$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -73.90 (d, $J = 1014.2$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 163.57, 163.54, 133.58, 133.56, 133.45, 133.43, 133.12, 133.10, 131.29, 131.27, 128.79, 128.65, 120.30 (d, $J = 22.9$ Hz), 118.82 (d, $J = 22.9$ Hz) 114.44, 114.29, 55.39. IR(film/ cm^{-1}) ν_{max} : 3396, 2925, 1597, 1253, 1133, 534. MS : m/z (M+H): 251.



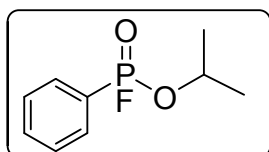
2k Yeld:83%, pale yellow oil ^1H NMR (400 MHz, CDCl_3) δ 7.87 – 7.78 (m, 2), 7.72 (dd, $J = 12.9, 8.0$ Hz, 2), 7.66 – 7.56 (m, 2), 7.51 (td, $J = 7.6, 3.9$ Hz, 1H), 7.32 (dd, $J = 7.8, 3.5$ Hz, 2H). 2.42 (s, 3H). ^{31}P NMR (400 MHz, CDCl_3) δ 41.45 (d, $J = 1016.8$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -74.76 (d, $J = 1017.1$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 144.21, 133.19, 133.17, 131.54, 131.52, 131.43, 131.40, 131.38, 131.29, 131.27, 129.82 (d, $J = 22.8$ Hz), 129.61, 129.47, 128.81, 128.67, 128.42 (d, $J = 22.7$ Hz), 21.73. IR(film/ cm^{-1}) ν_{max} 3408, 2924, 1604, 1439, 1258, 1135, 536. MS : m/z (M+H): 235



2l Yeld:47%, pale yellow oil ^1H NMR (400 MHz, CDCl_3) δ 7.97 (m, 2H), 7.89 – 7.81 (m, 2H), 7.78 (m, 2H), 7.78 (m, 2H), 7.67 (m, 1H), 7.55 (mz, 2H). ^{31}P NMR (400 MHz, CDCl_3) δ 38.72 (d, $J = 1025.3$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -63.44, -75.17 (d, $J = 1025.6$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 135.14, 134.84, 133.89, 133.87, 133.62, 132.48, 132.24, 132.02, 132.00, 131.90, 131.88, 131.66, 131.54, 131.52, 131.42, 131.40, 131.22, 131.12, 129.14, 129.00, 128.59, 128.48, 128.26, 127.05, 126.84, 125.86, 125.82, 125.78, 125.75, 125.72, 125.68, 125.64, 125.61, 124.60, 121.89. IR(film/ cm^{-1}) ν_{max} 3373, 3061, 2927, 2857, 2588, 1678, 1439, 1399, 1324, 1170, 1131, 965, 739, 567. MS : m/z (M+H): 289.

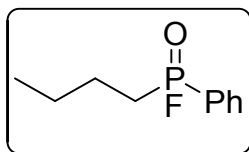


2mj Yeld:78%, pale yellow oil ^1H NMR (400 MHz, CDCl_3) δ 7.83 (m, 2H), 7.61 (m, 1H), 7.49 (m, 2H), 4.60 – 4.47 (m, 1H), 2.30 (d, $J = 11.8$ Hz, 1H), 2.24 – 2.13 (m, 1H), 1.94 (m, 1H), 1.69 (m, 2H), 1.53 – 1.14 (m, 4H), 1.04 (m, 1H), 0.95 (t, $J = 7.3$ Hz, 3H), 0.90 – 0.81 (m, 6H), 0.70 (d, $J = 6.9$ Hz, 2H). ^{31}P NMR (400 MHz, CDCl_3) δ 19.18 (d, $J = 106.1$ Hz), 12.74 (d, $J = 103.6$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -59.22 (d, $J = 919.0$ Hz), -62.00 (d, $J = 916.9$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 133.30, 131.48, 131.38, 128.65, 128.64, 128.49, 128.48, 126.60 (dd, $J = 45.9, 31.2$ Hz), 124.58 (dd, $J = 46.4, 31.3$ Hz), 80.43, 80.36, 80.05, 79.98, 48.28, 48.22, 48.15, 42.92, 42.83, 33.82, 31.52, 31.48, 25.81, 25.75, 22.84, 21.82, 21.73, 20.78, 20.67, 15.47. IR(film/ cm^{-1}) ν_{max} 3365, 2956, 2930, 2871, 1595, 1457, 1442, 1292, 1136, 1014, 996, 847. MS : m/z (M+H): 299

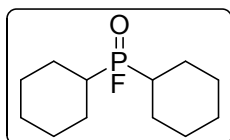


2n Yeld:44%, pale yellow oil. ^1H NMR (400 MHz, CDCl_3) δ 7.91 – 7.80 (m, 2H), 7.63 (m, 1H), 7.51 (dd, $J = 12.6, 7.6$ Hz, 2H), 5.07 – 4.92 (m, 1H), 1.47 (d, $J = 6.2$ Hz, 3H), 1.38 (d, $J = 6.2$ Hz,

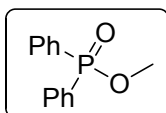
3H). ^{31}P NMR (400 MHz, CDCl_3) δ 15.92 (d, $J = 1041.3$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -61.69 (d, $J = 1041.0$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 133.43, 133.40, 131.62, 131.52, 128.69, 128.53, 126.31 (d, $J = 30.7$ Hz), 124.31 (d, $J = 30.7$ Hz), 73.47, 73.41, 30.88, 23.89, 23.85, 23.77, 23.73. IR (film/ cm^{-1}) ν_{max} 3494, 3063, 2984, 2932, 1595, 1441, 1287, 1137, 1011, 995, 848. MS (M+H): 203



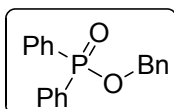
2o Yield: 65%, pale yellow oil ^1H NMR (400 MHz, CDCl_3) δ 7.78 (m, 2H), 7.67 – 7.56 (m, 1H), 7.51 (m, 2H), 2.16 – 1.94 (m, 2H), 1.69 – 1.49 (m, 2H), 1.47 – 1.32 (m, 2H), 0.87 (t, $J = 7.3$ Hz, 3H). ^{31}P NMR (400 MHz, CDCl_3) δ 55.62 (d, $J = 1021.4$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -75.55 (d, $J = 1021.5$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 133.31, 133.29, 131.01, 130.99, 129.07 (d, $J = 19.4$ Hz), 127.77 (d, $J = 19.2$ Hz), 28.93 (d, $J = 17.2$ Hz), 27.97 (d, $J = 17.2$ Hz), 23.59, 23.43, 23.29, 23.25, 13.35. IR (film/ cm^{-1}) ν_{max} 3404, 2958, 2931, 2870, 1594, 1439, 1165, 1128, 965, 741. MS : m/z (M+H): 201.



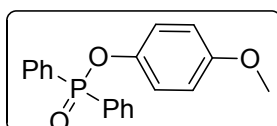
2p Yield: 75%, pale yellow solid ^1H NMR (400 MHz, CDCl_3) δ 2.00 – 1.89 (m, 2H), 1.89 – 1.73 (m, 8H), 1.69 (d, $J = 1.5$ Hz, 2H), 1.55 – 1.33 (m, 4H), 1.31 – 1.11 (m, 6H). ^{31}P NMR (400 MHz, CDCl_3) δ 70.75 (d, $J = 1036.4$ Hz). ^{19}F NMR (400 MHz, CDCl_3) δ -94.88 (d, $J = 1036.7$ Hz). ^{13}C NMR (400 MHz, CDCl_3) δ 35.29 (d, $J = 13.3$ Hz), 34.43 (d, $J = 13.3$ Hz), 25.90, 25.87, 25.76, 25.74, 25.53, 25.51, 24.61, 24.58, 24.54. IR (film/ cm^{-1}) ν_{max} 3453, 2930, 2854, 1448, 1279, 1245, 1202, 858. MS : m/z (M+H): 233.



3a Yield: 88%, pale yellow solid ^1H NMR (400 MHz, CDCl_3) δ 7.86 – 7.77 (m, 4H), 7.52 (m, 2H), 7.44 (m, 4H), 3.76 (3H). ^{31}P NMR (400 MHz, CDCl_3) δ 33.22. ^{13}C NMR (400 MHz, CDCl_3) δ 132.16, 132.13, 131.63, 131.53, 130.26, 128.56, 128.43, 51.49, 51.43. MS : m/z (M+H): 233

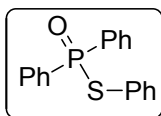


3b Yield: 81%, pale yellow solid ^1H NMR (400 MHz, CDCl_3) δ 7.90 – 7.80 (m, 2H), 7.52 (t, $J = 7.3$ Hz, 2H), 7.44 (dm, 4H), 7.39 – 7.28 (m, 4H), 5.08 (d, $J = 6.8$ Hz, 2H). ^{31}P NMR (400 MHz, CDCl_3) δ 32.32. ^{13}C NMR (400 MHz, CDCl_3) δ 136.29, 136.22, 132.13, 132.10, 131.65, 131.55, 131.26 (d, $J = 136.7$ Hz), 128.51, 128.43, 128.38, 128.16, 127.74, 66.20, 66.15. MS : m/z (M+H): 309



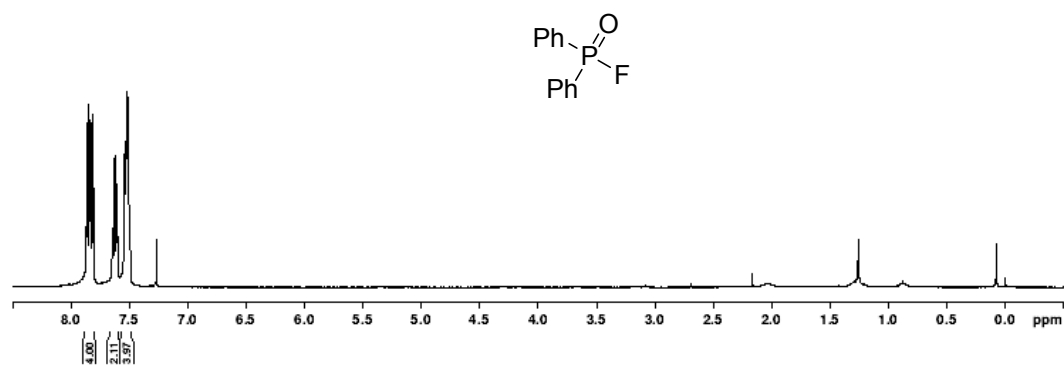
3d Yield: 46%, pale yellow solid. ^1H NMR (400 MHz, CDCl_3) δ 7.92 – 7.84 (m, 4H), 7.54 (m, 2H), 7.46 (m, 4H), 7.12 – 7.07 (m, 2H), 6.78 – 6.71 (m, 2H), 3.72 (s, 3H). ^{31}P NMR (400 MHz, CDCl_3) δ

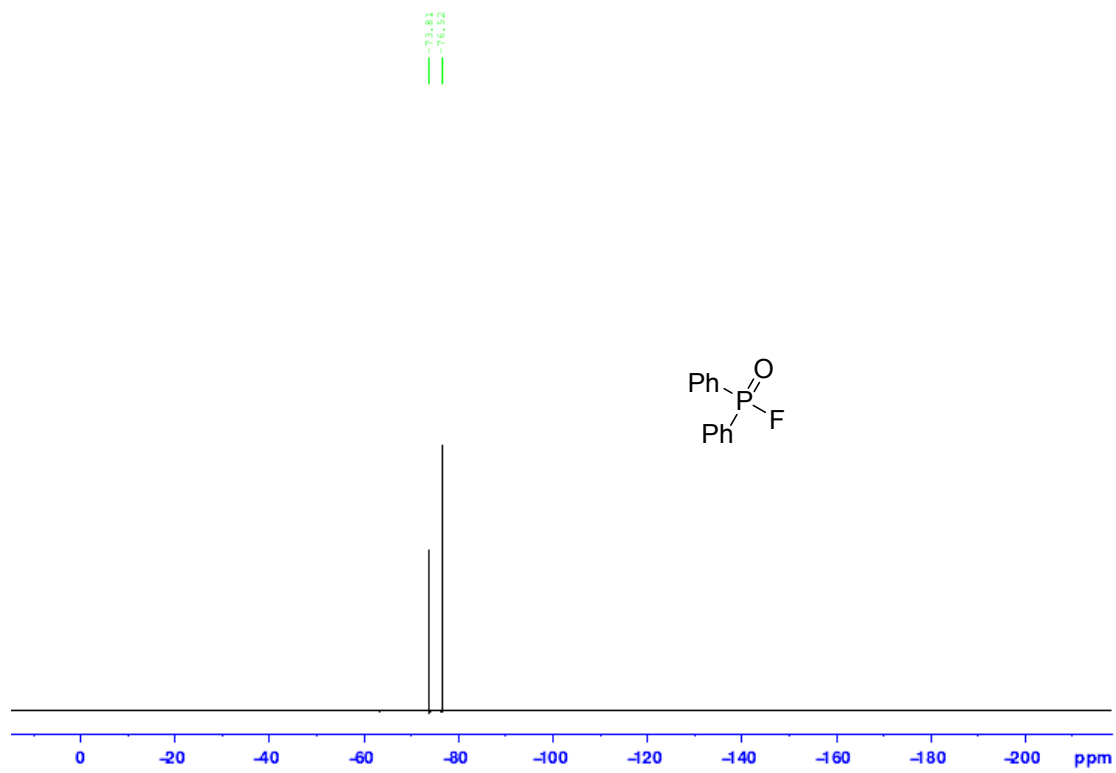
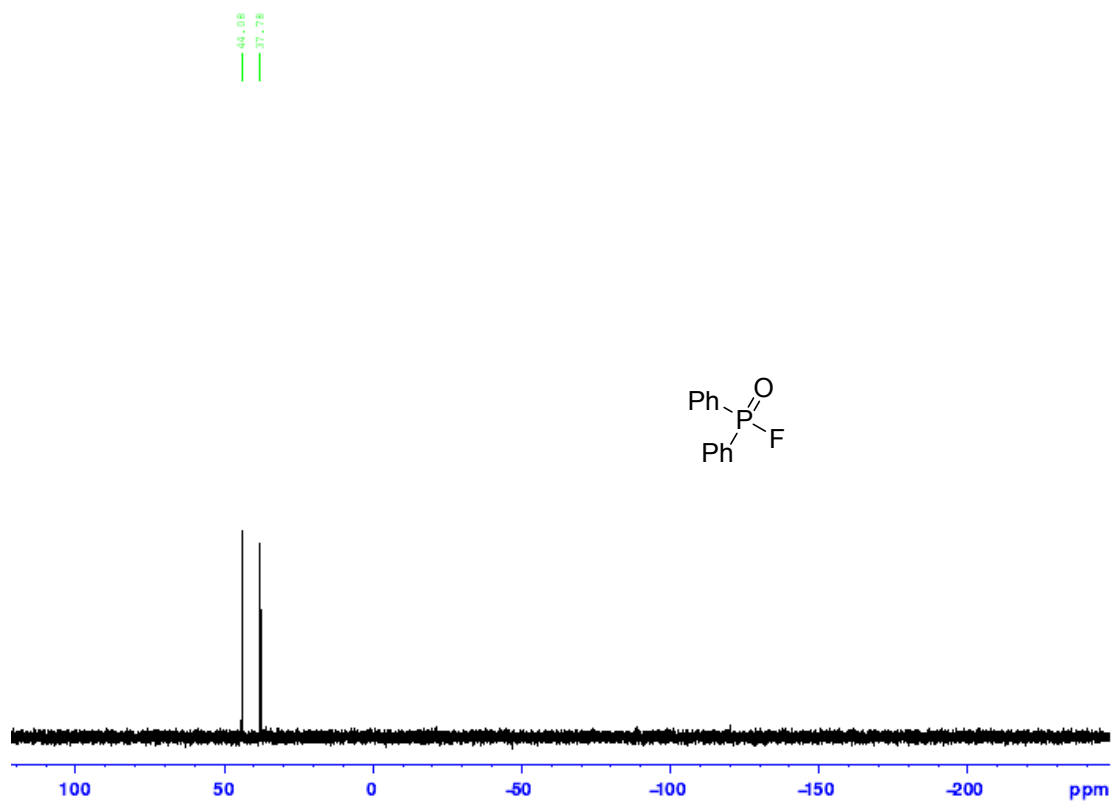
30.72. ^{13}C NMR (400 MHz, CDCl_3) δ 156.34, 144.14, 144.05, 132.47, 132.44, 131.82, 131.71, 130.59 (d, $J = 138.5$ Hz), 128.61, 128.48, 121.58, 121.54, 114.53, 55.45. MS : m/z (M+H): 325.

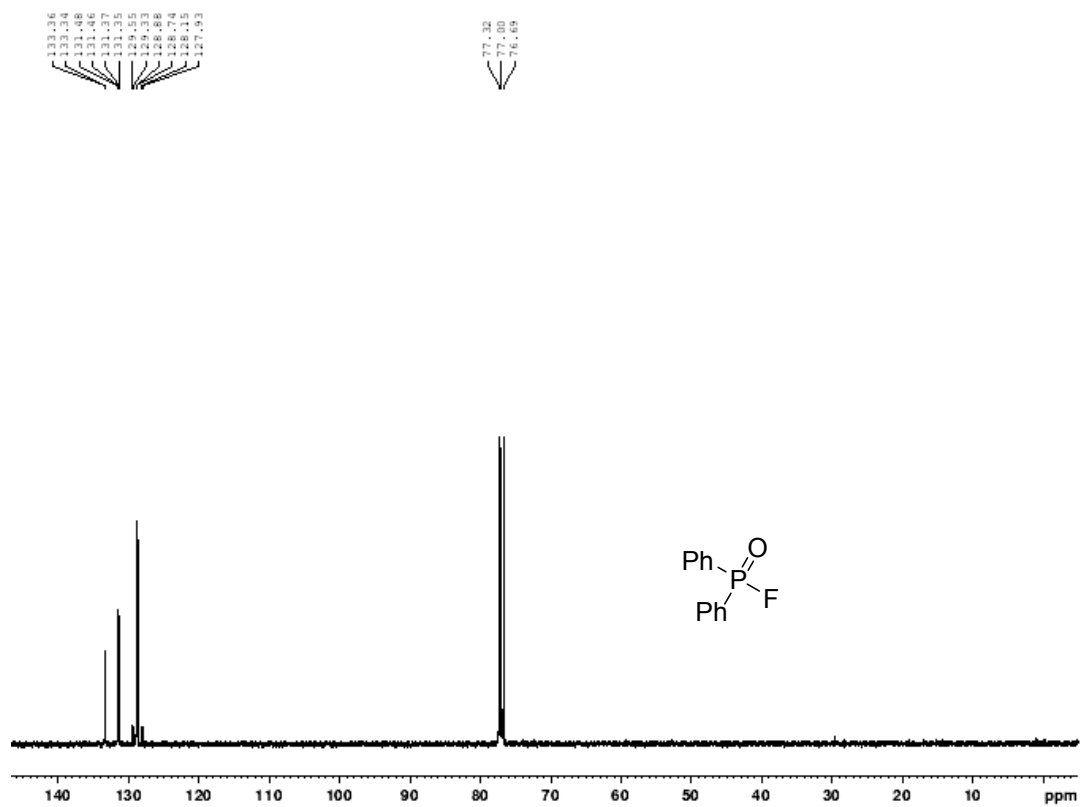


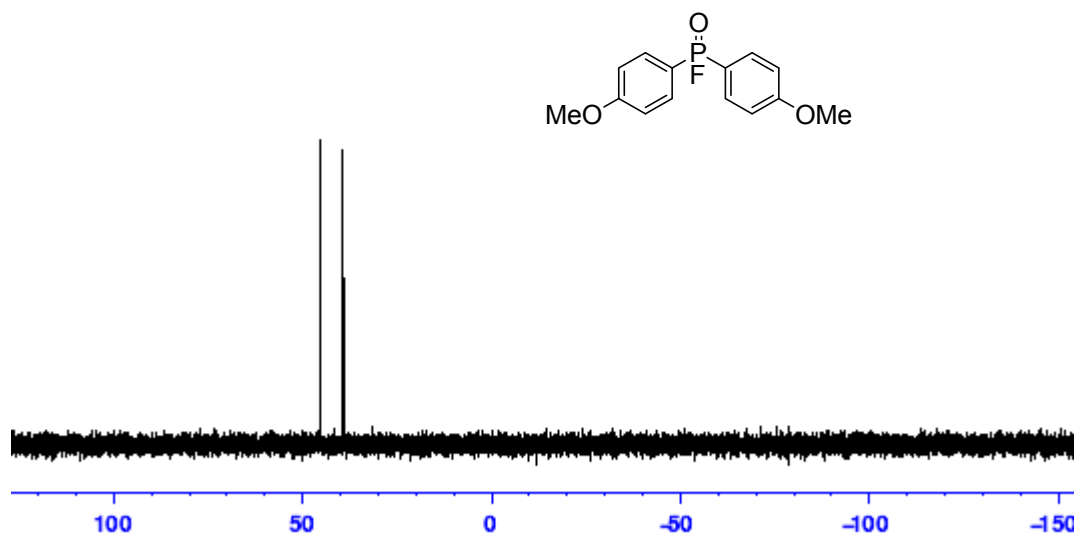
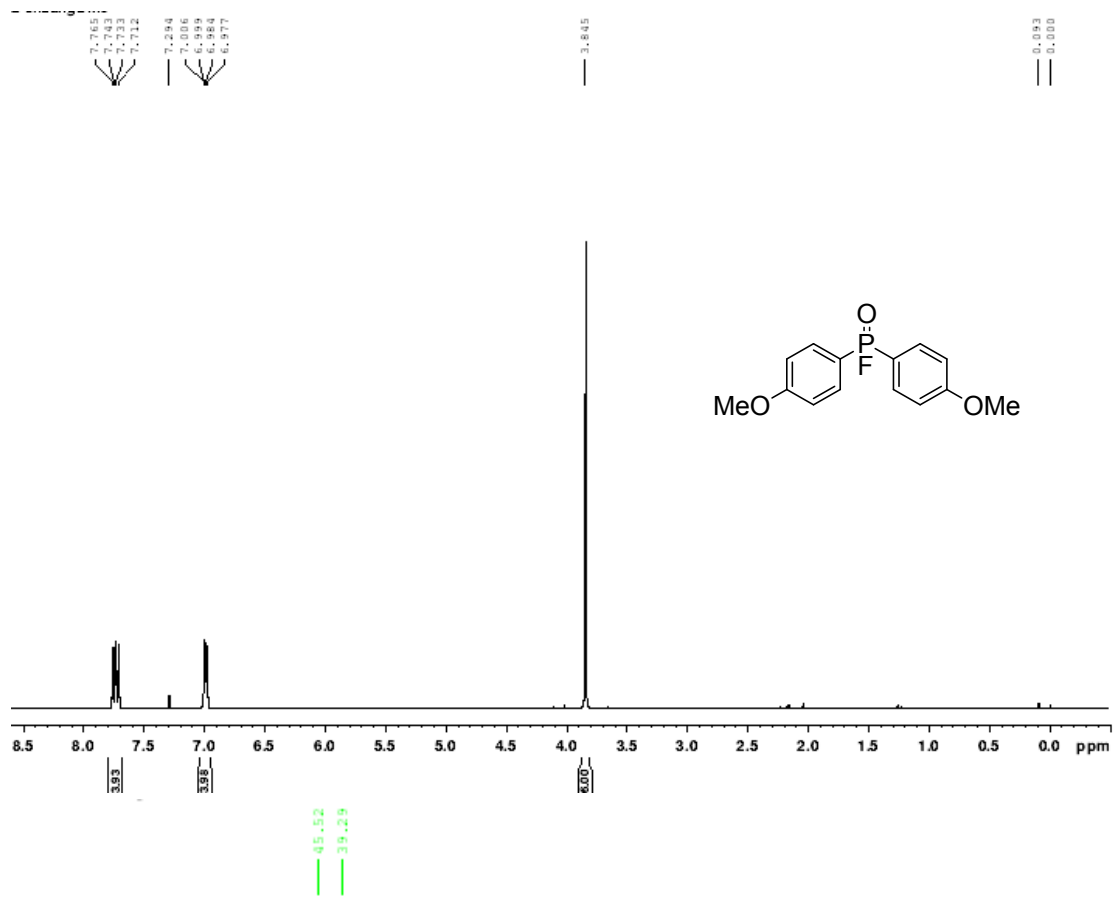
3e Yield:81%, white solid. ^1H NMR (400 MHz, DMSO) δ 7.86 – 7.78 (m, 4H), 7.63 – 7.57 (m, 2H), 7.57 – 7.50 (m, 4H), 7.46 – 7.41 (m, 2H), 7.33 – 7.23 (m, 3H). ^{31}P NMR (400 MHz, DMSO) δ 39.58. ^{13}C NMR (400 MHz, DMSO) δ 134.66, 134.62, 132.41, 132.39, 132.34 (d, $J = 106.0$ Hz), 131.04, 130.94, 129.19, 129.03, 129.02, 128.85, 128.84, 128.74, 128.61, 127.04, 125.96, 125.91. MS: m/z (M+H): 311.

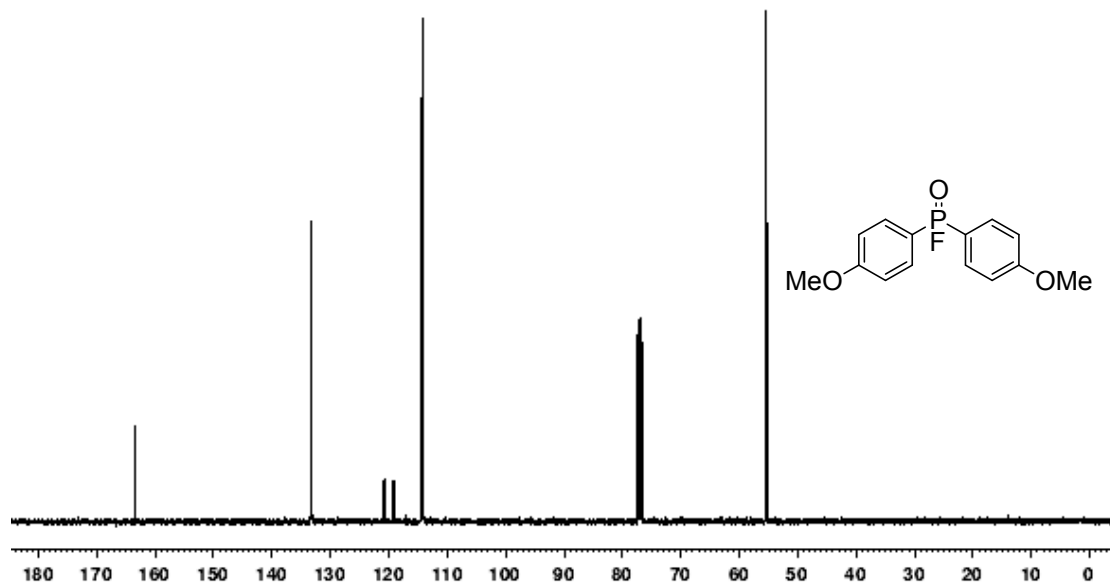
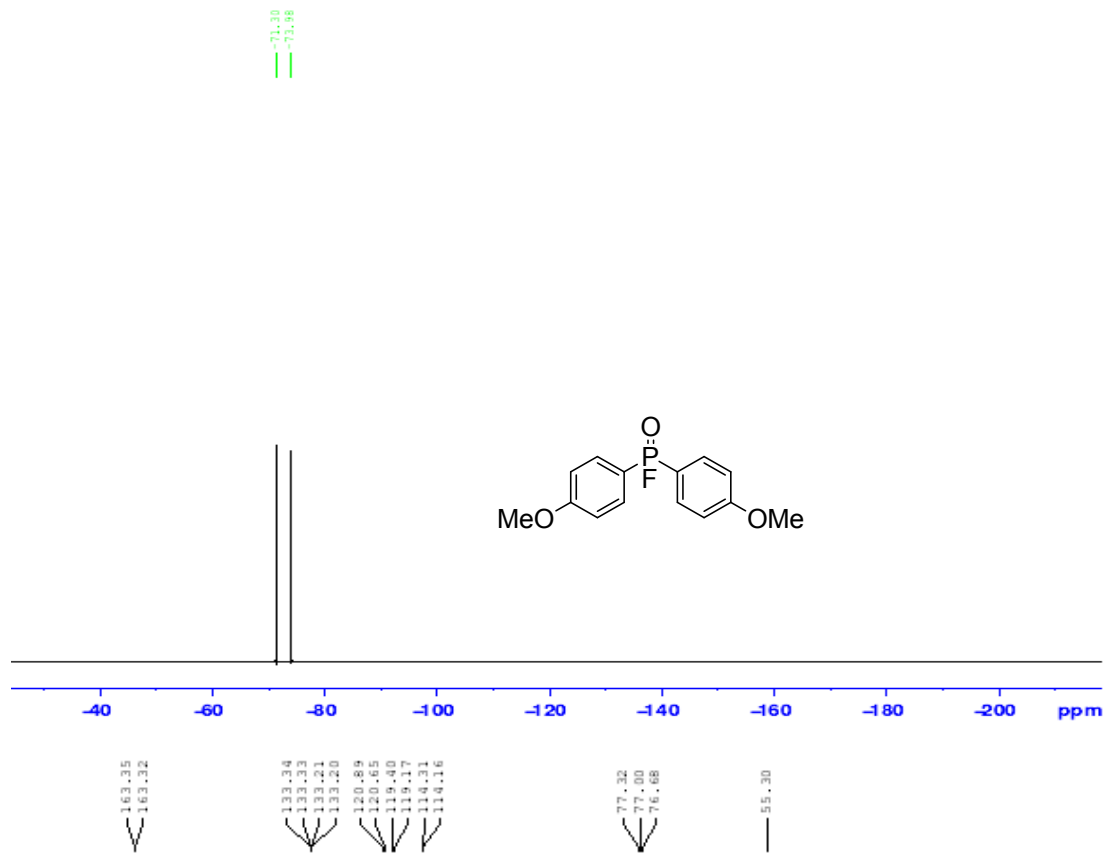
6. Charts

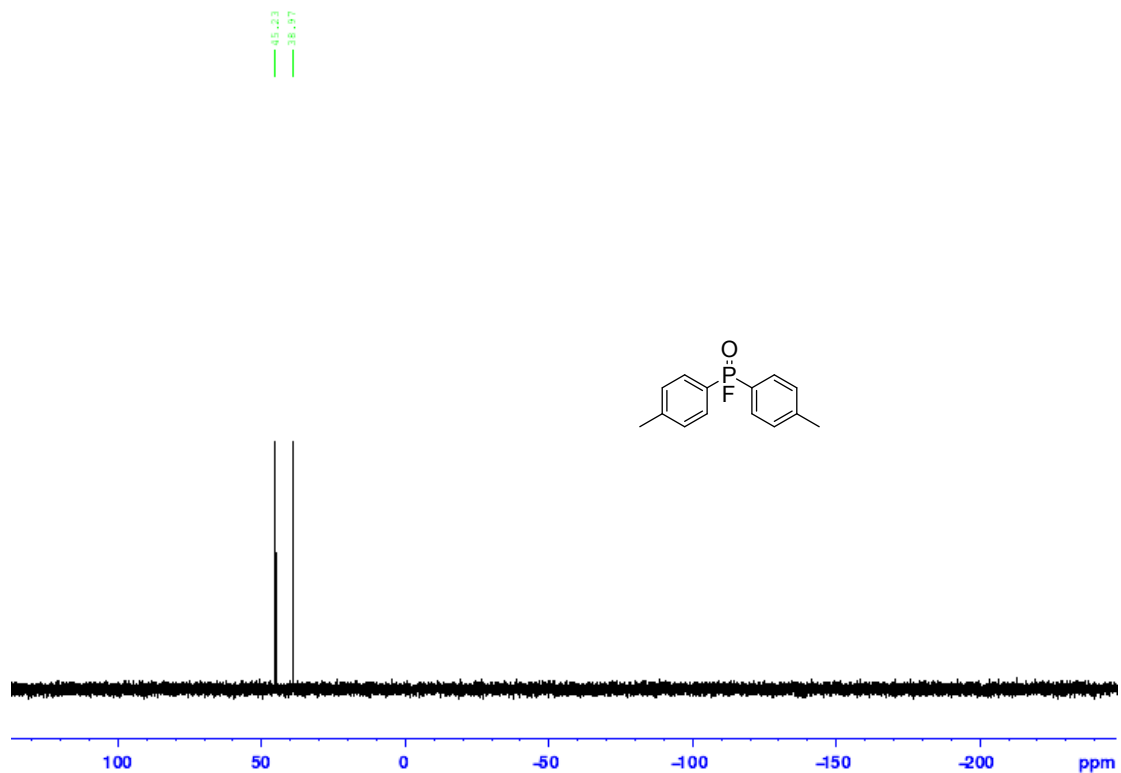
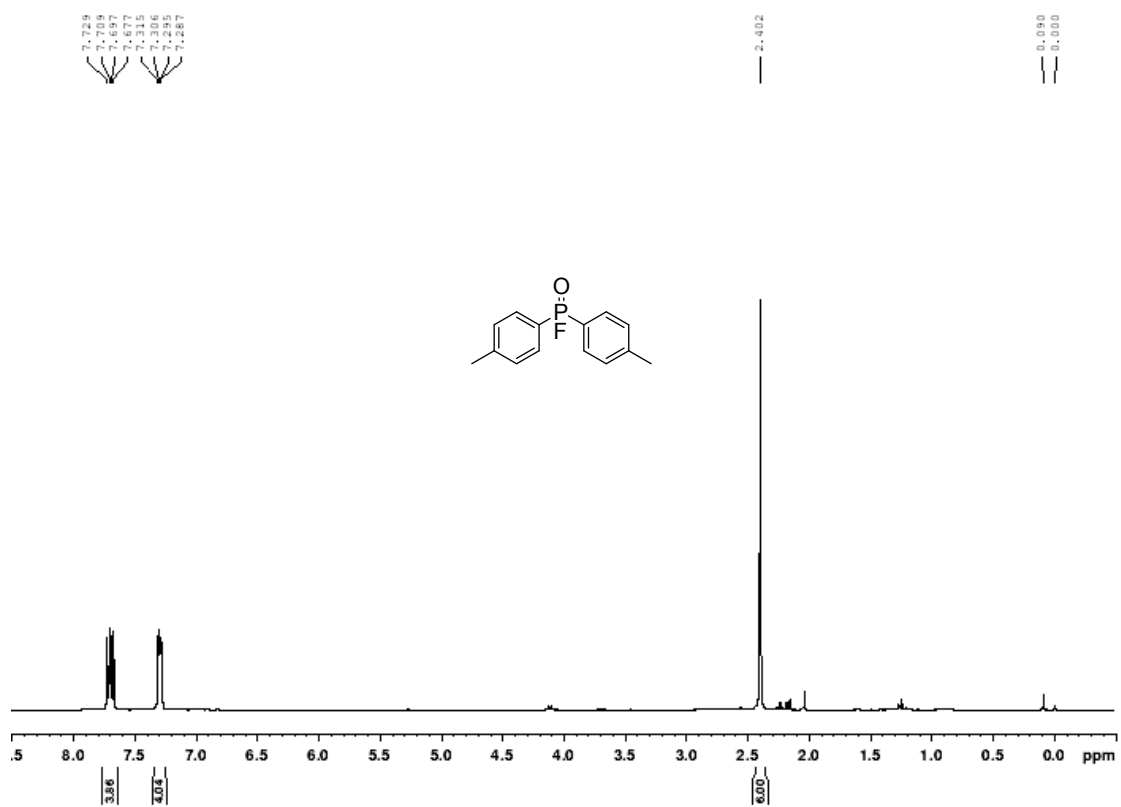


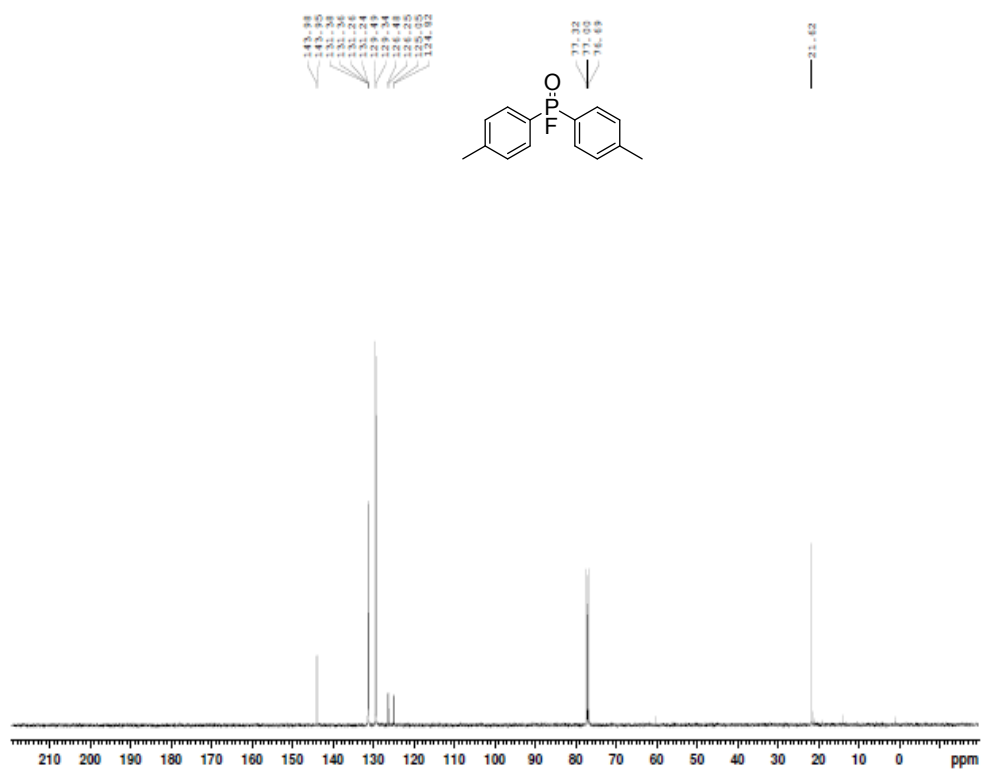
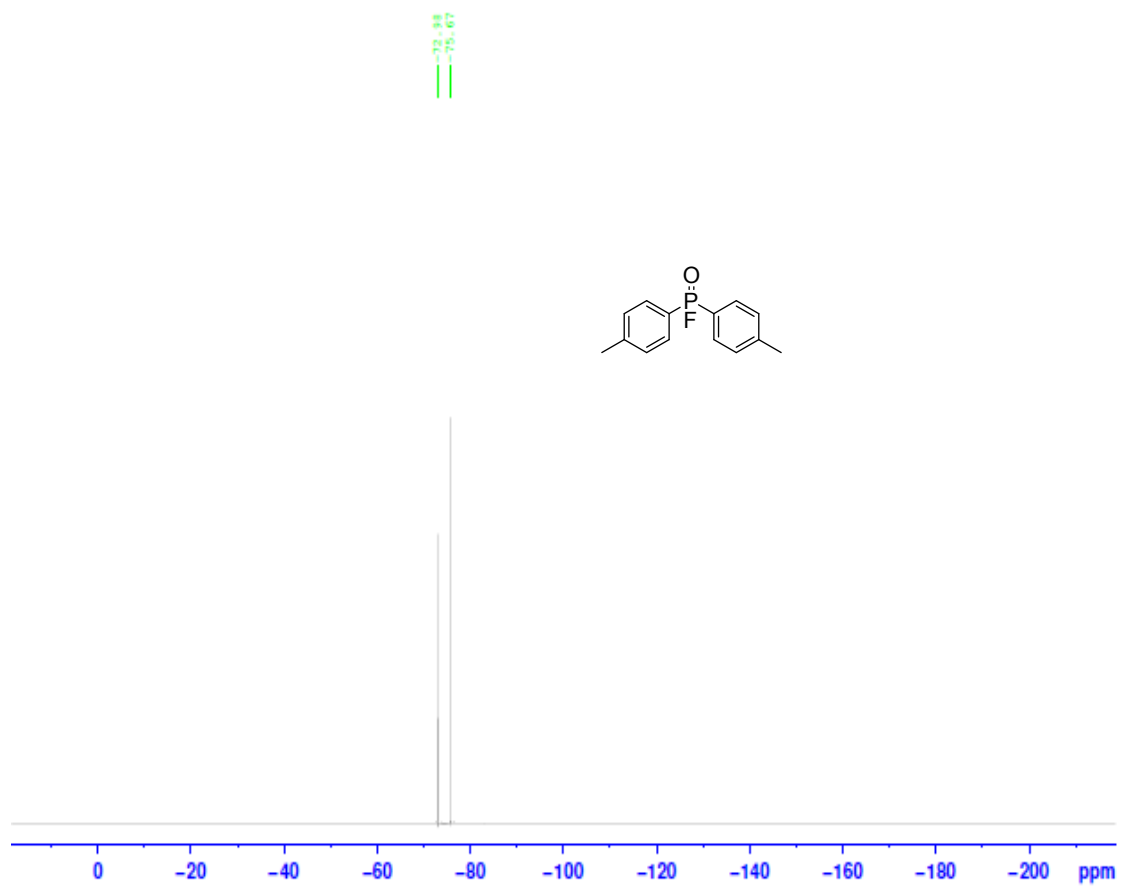


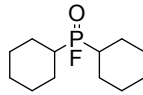
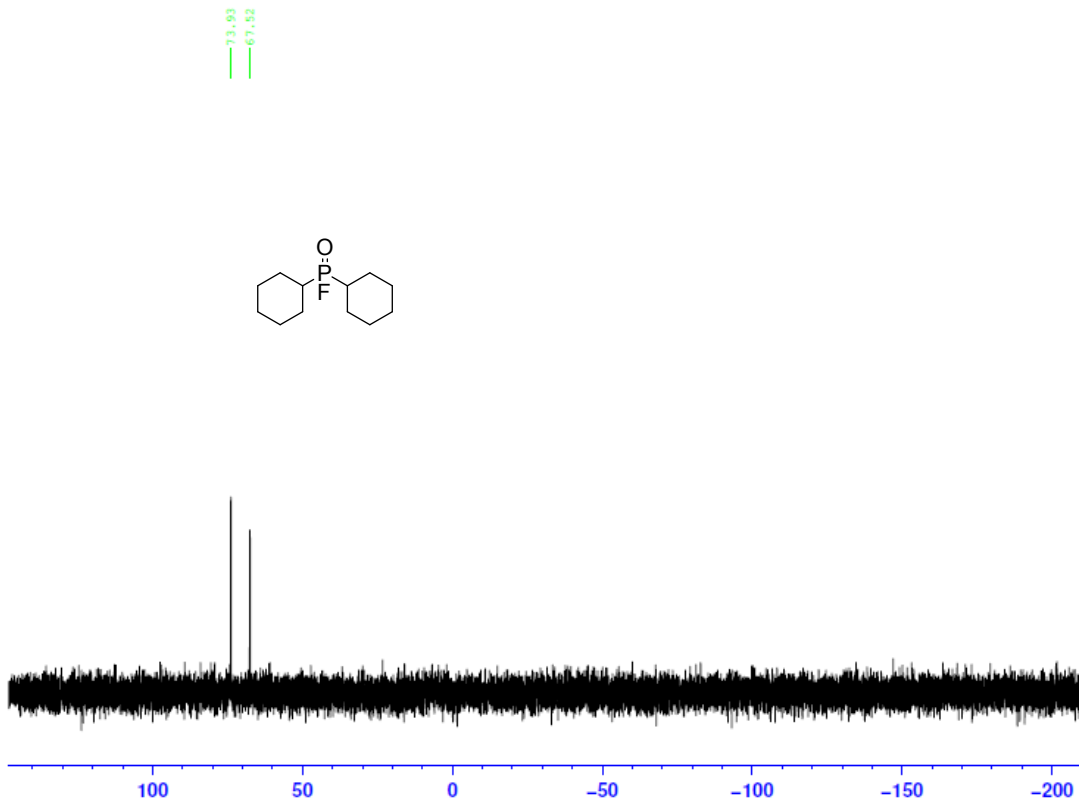
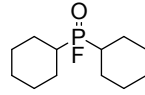
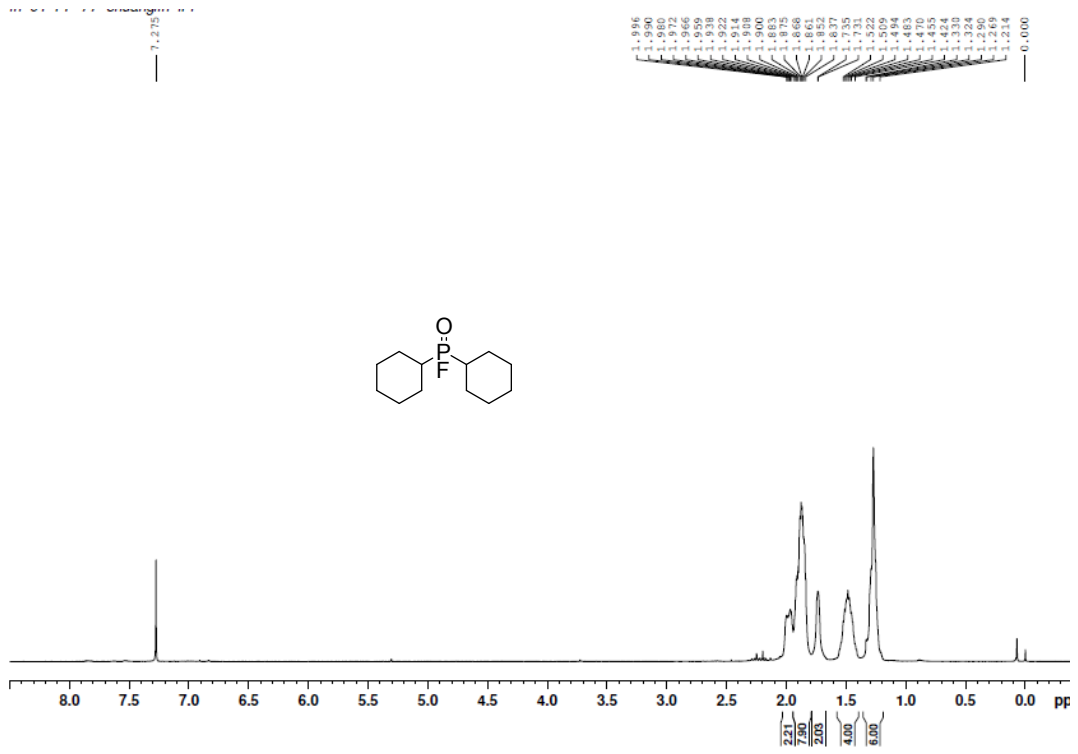


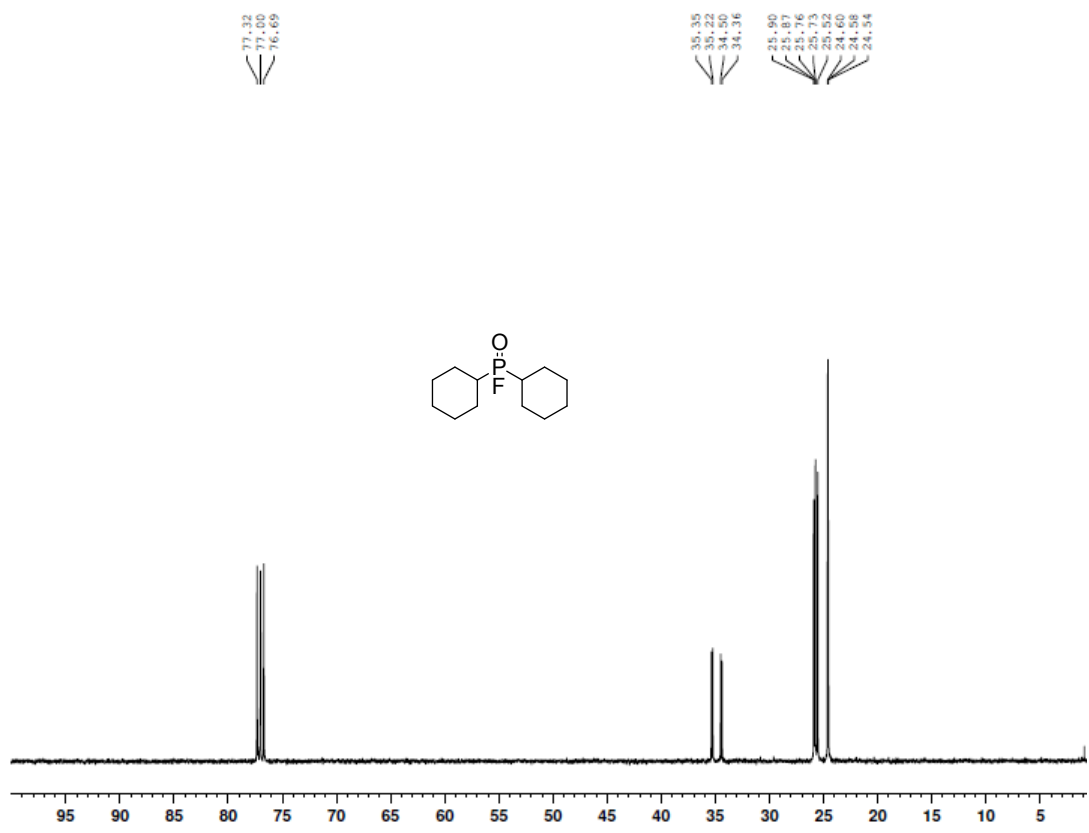
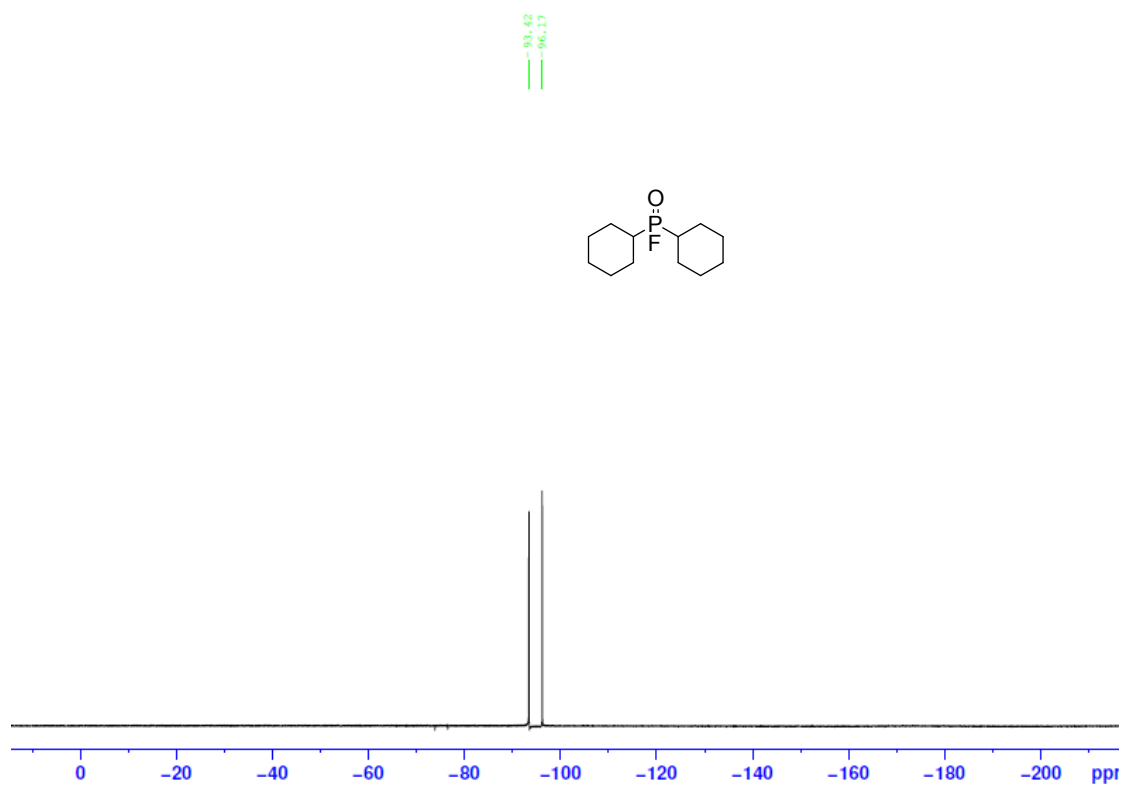


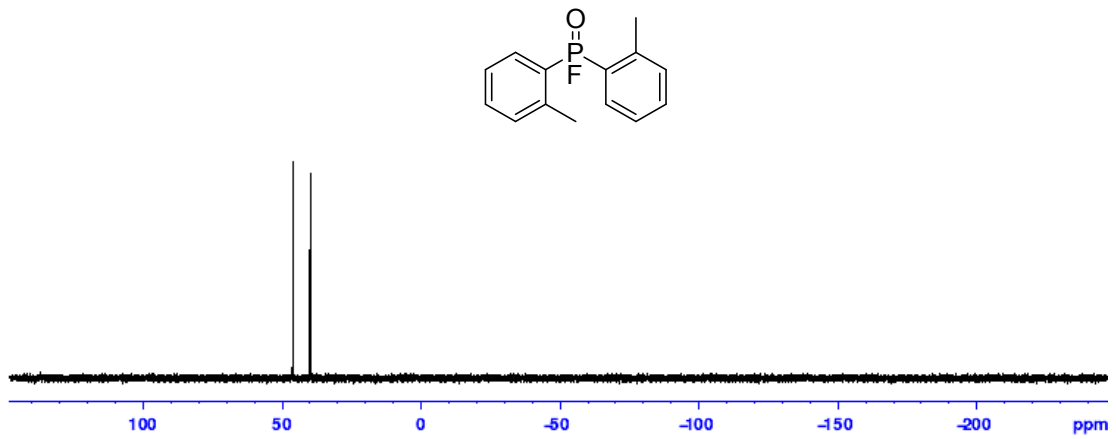
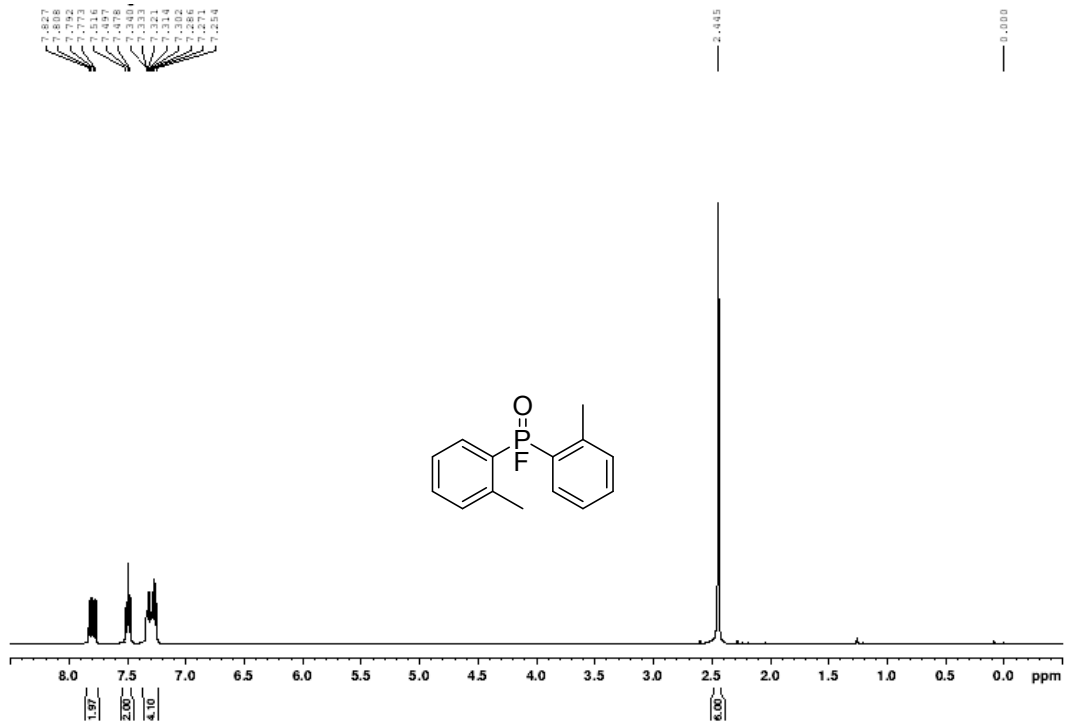


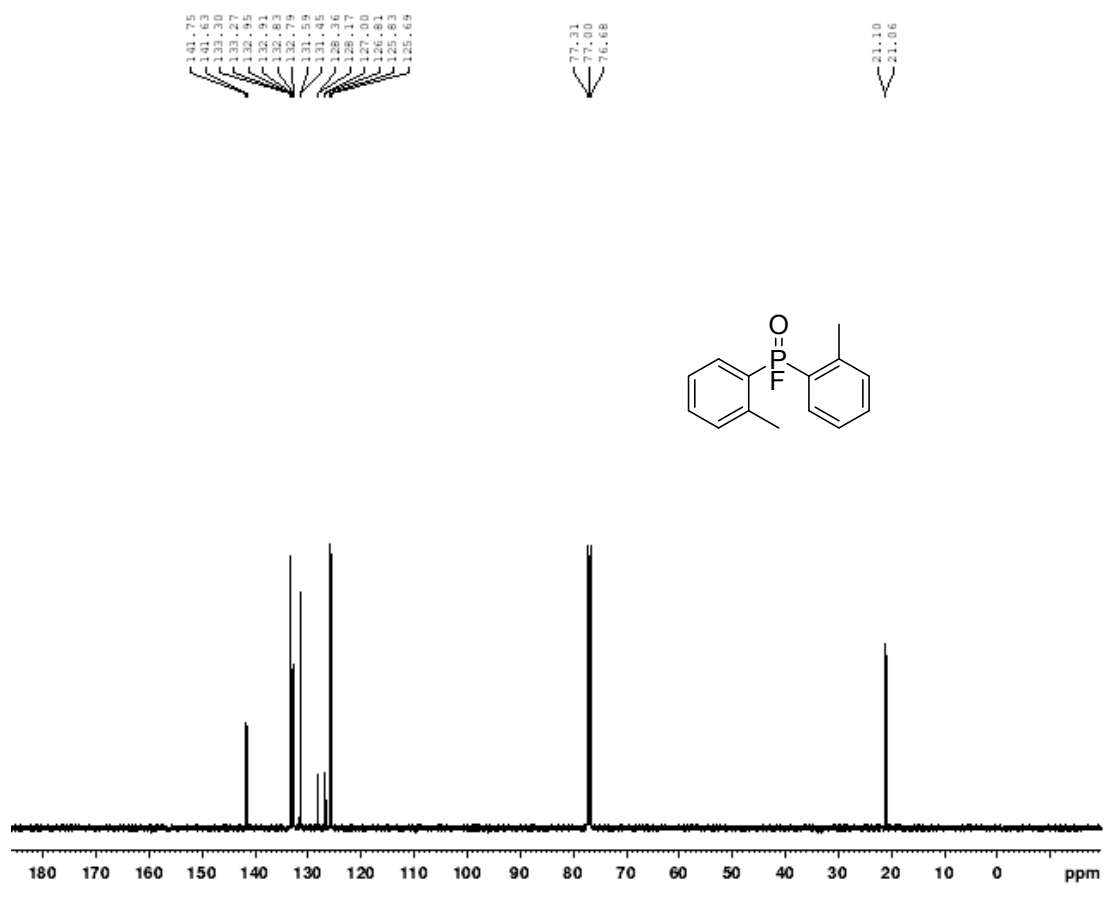
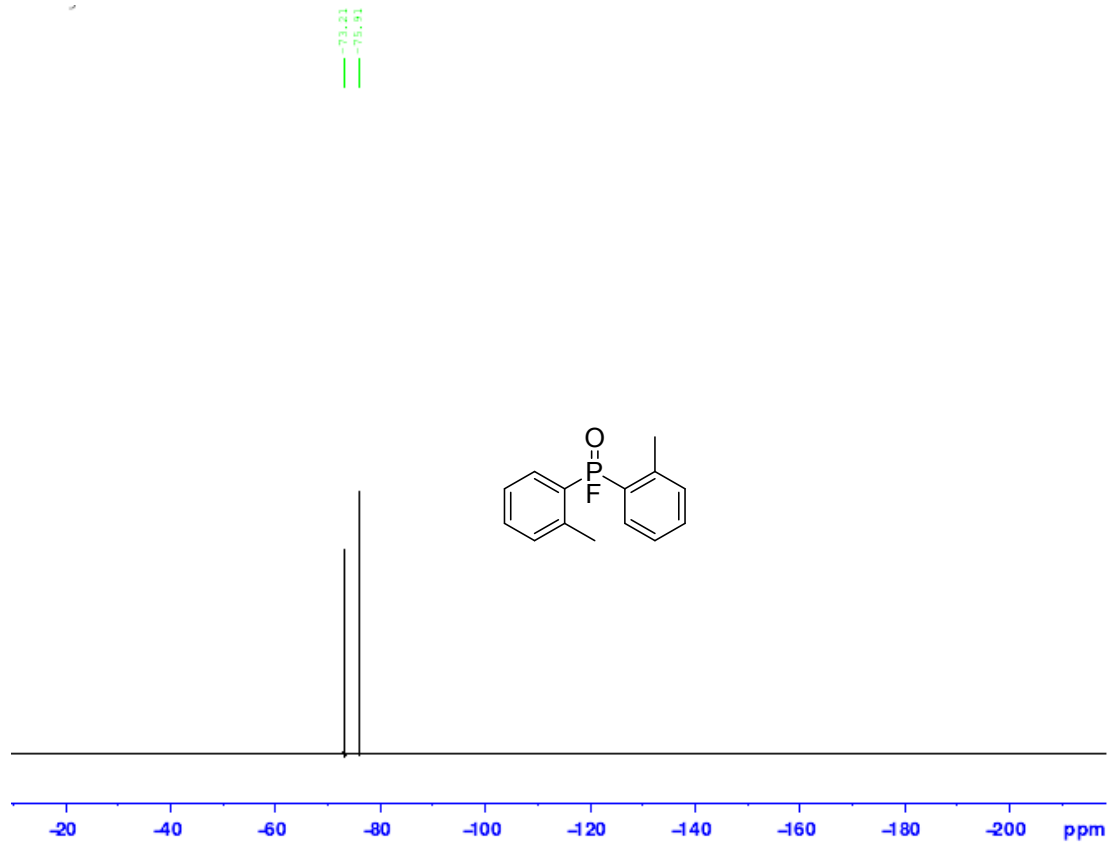


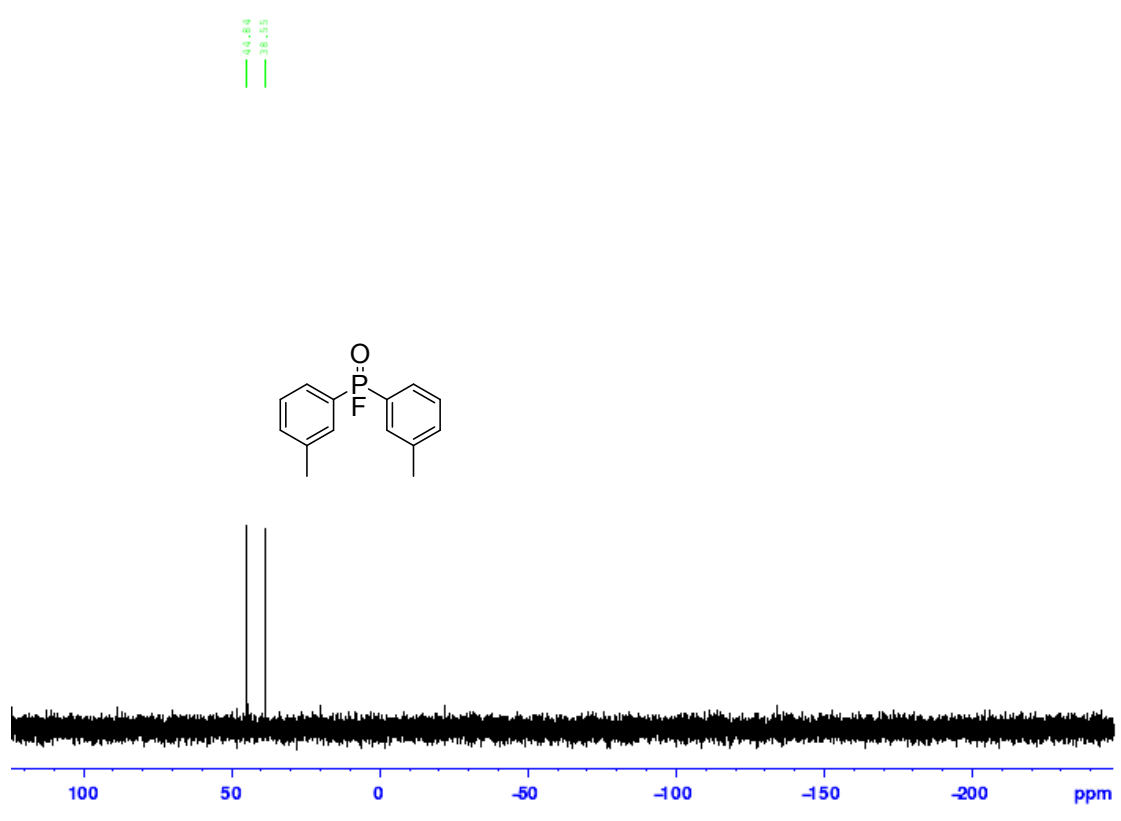
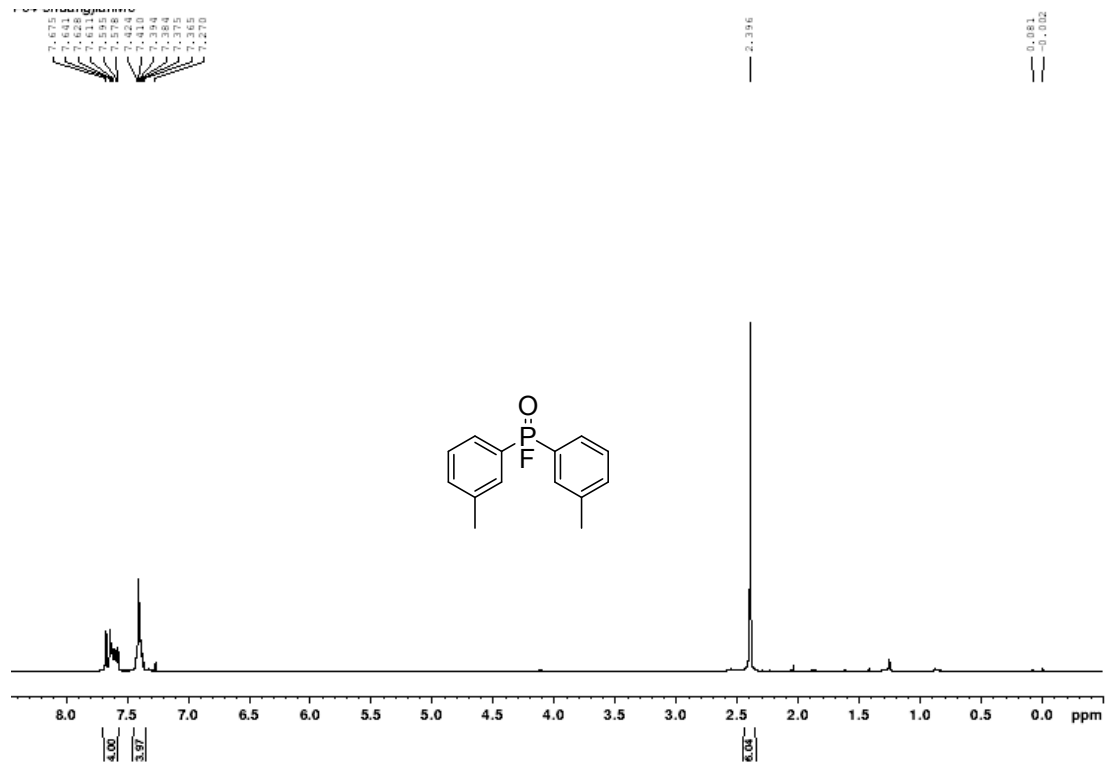


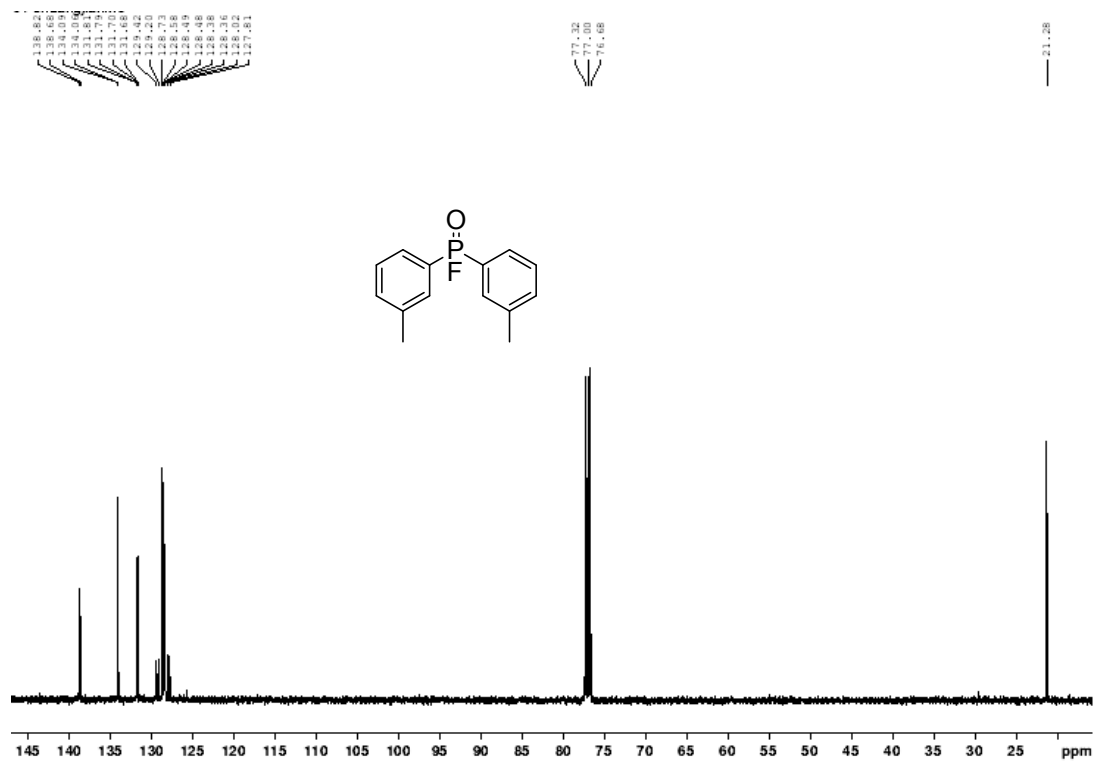
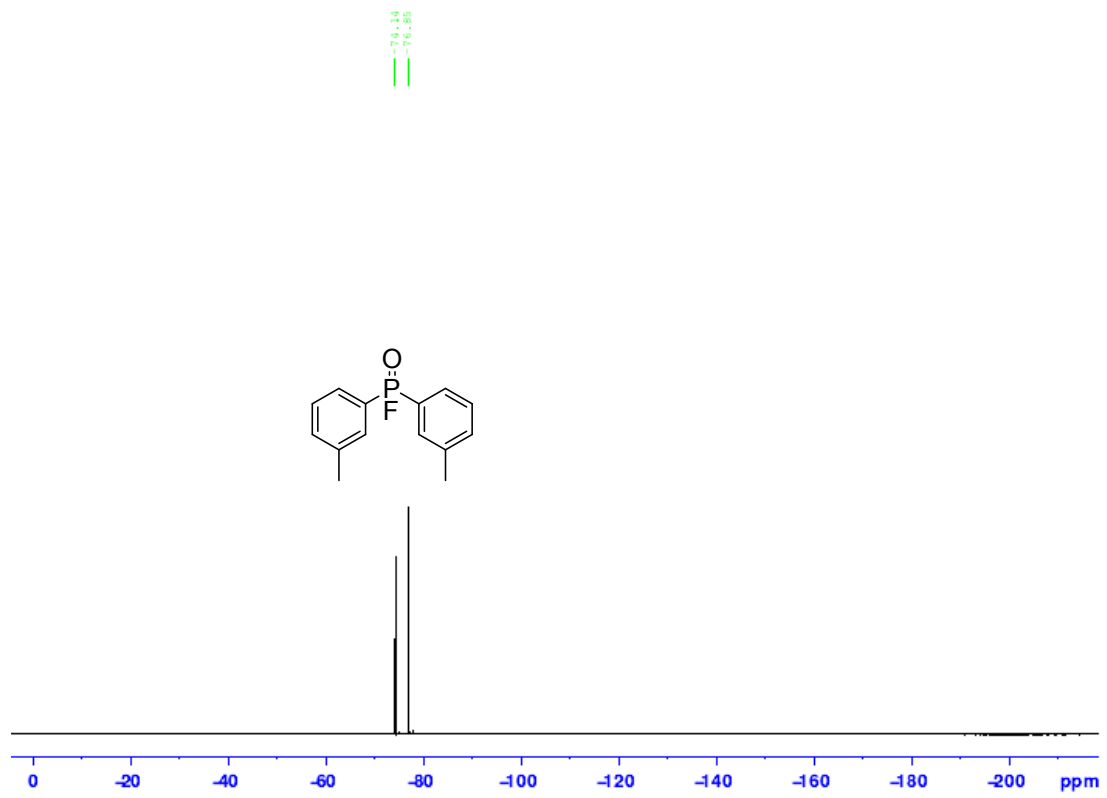


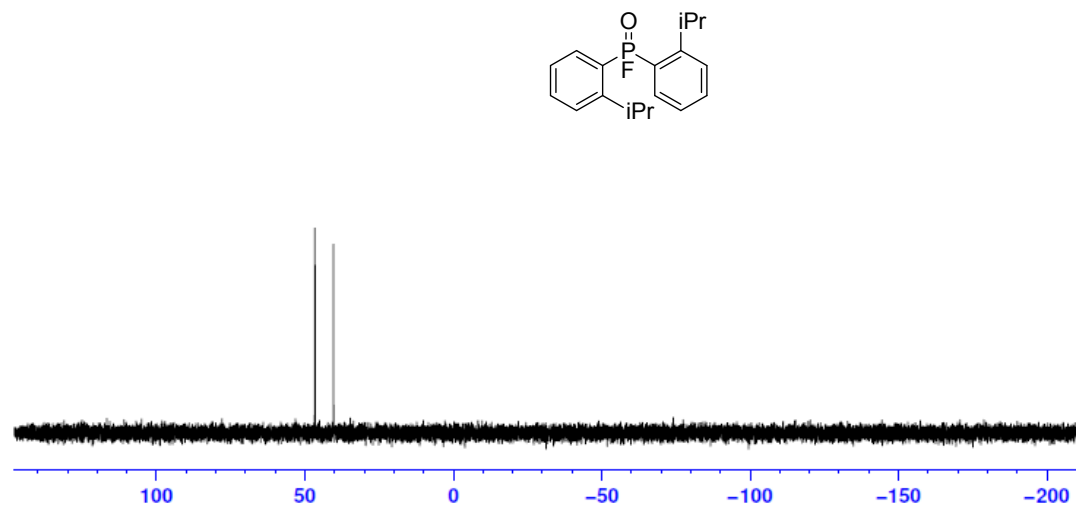
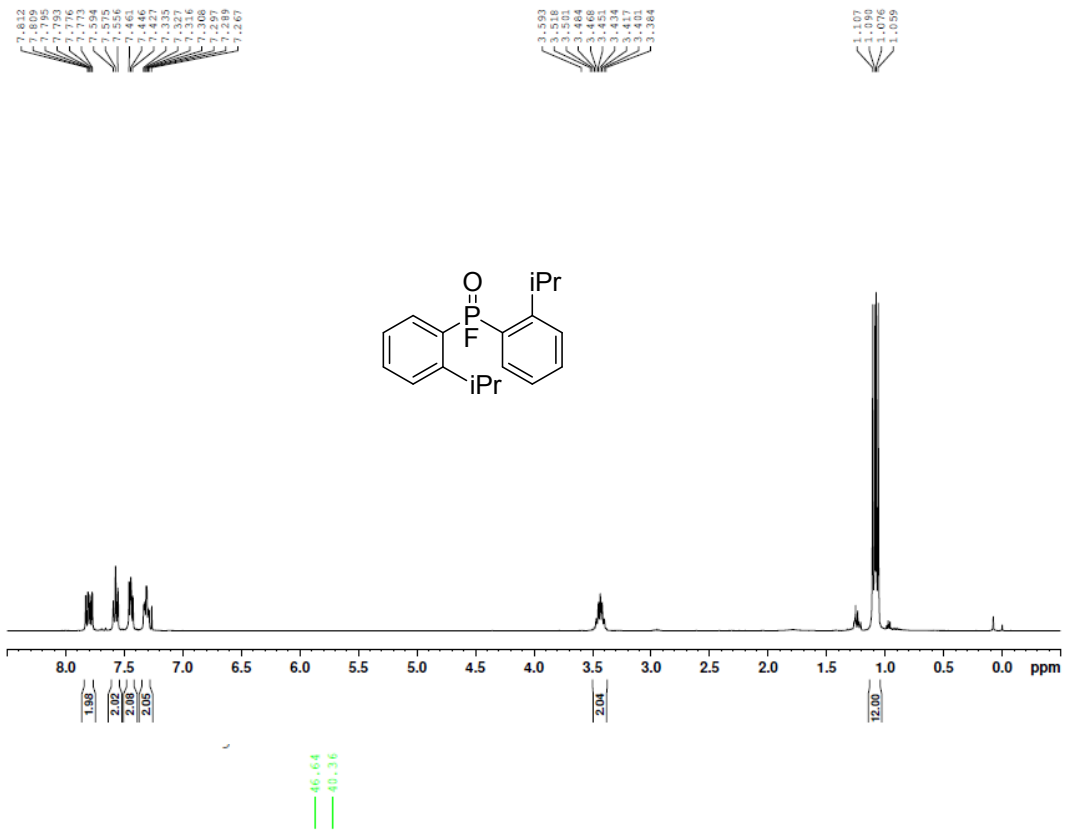


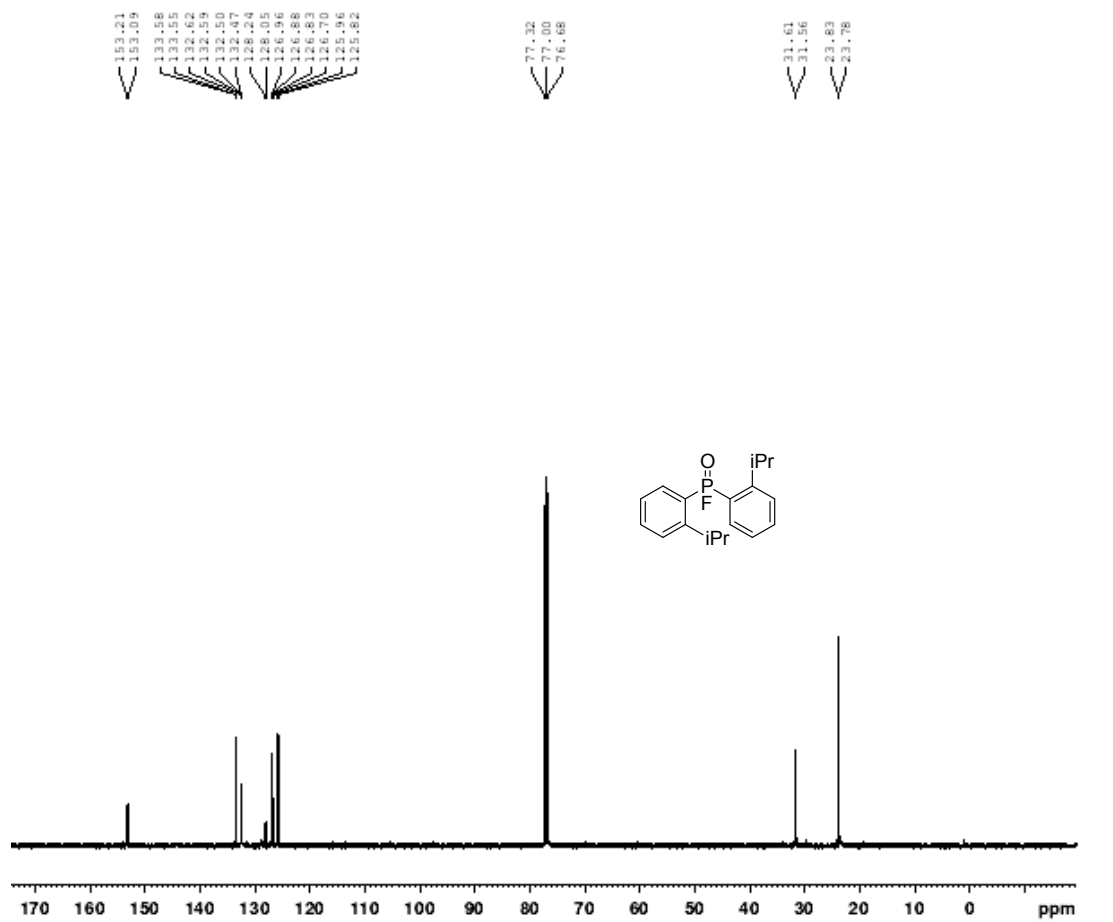
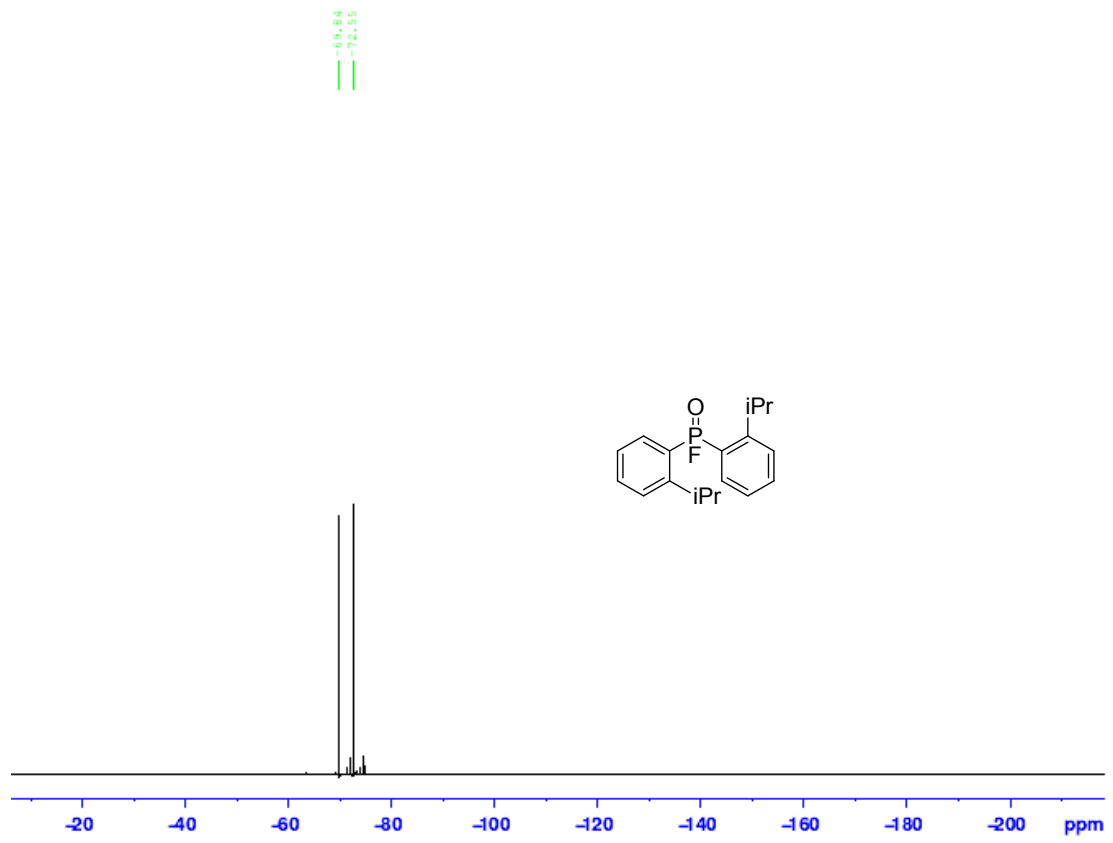


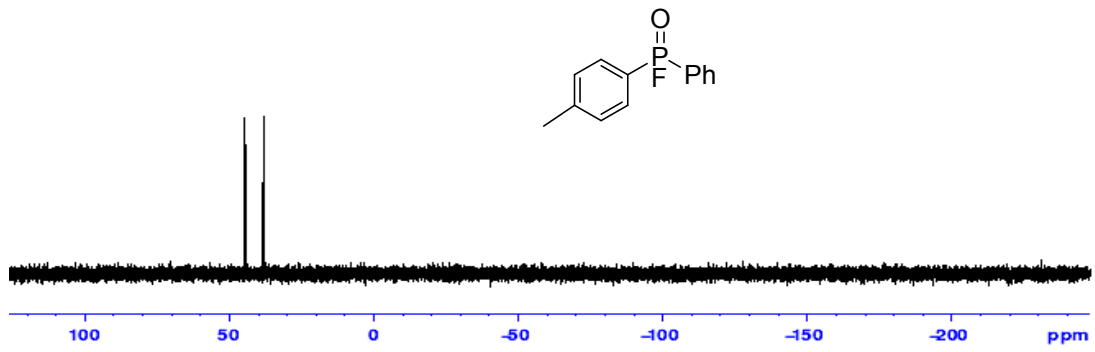
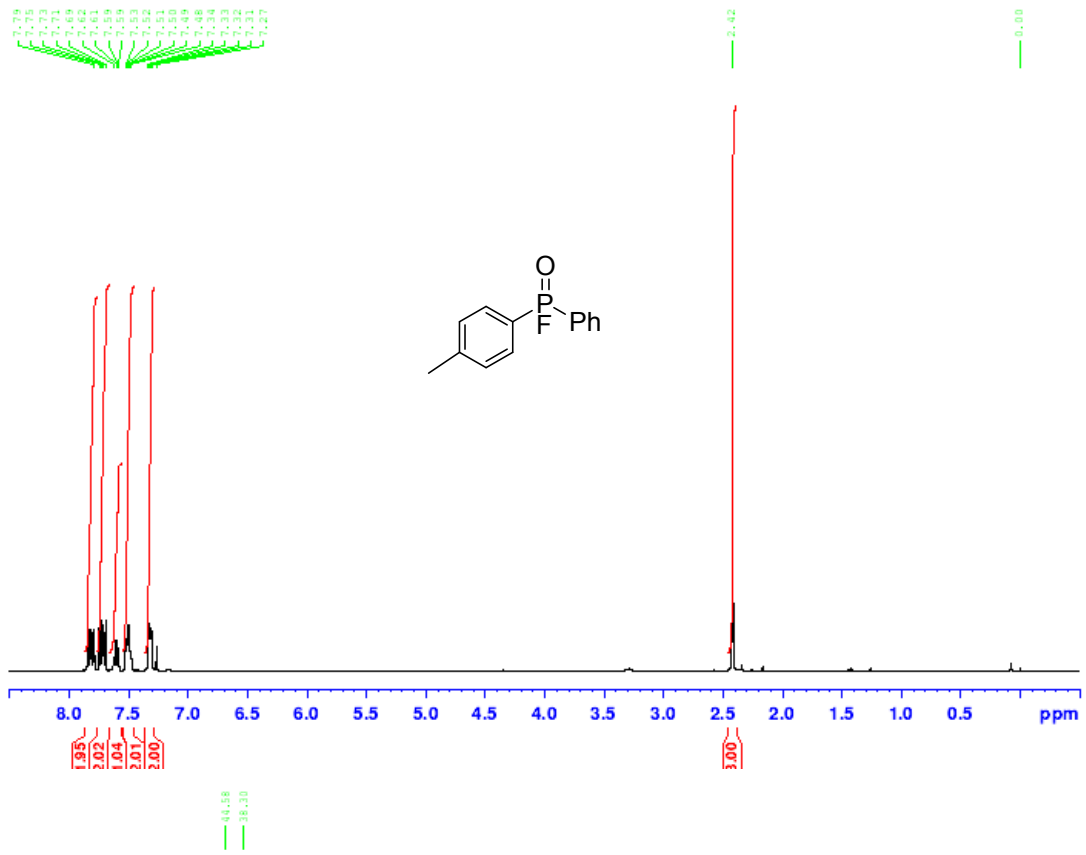


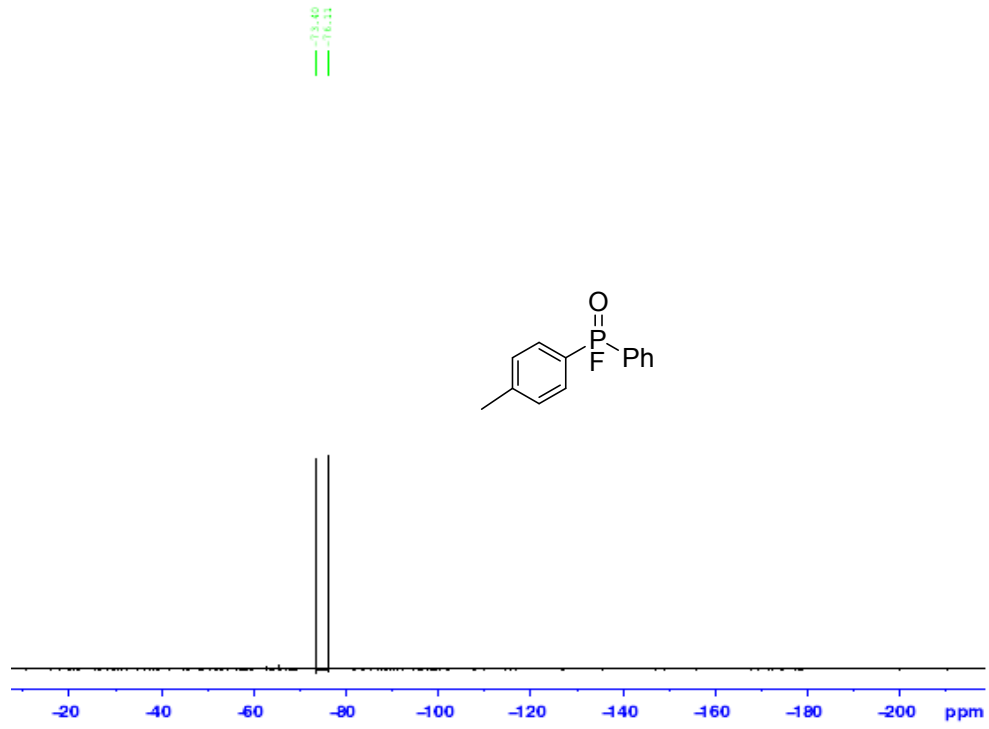








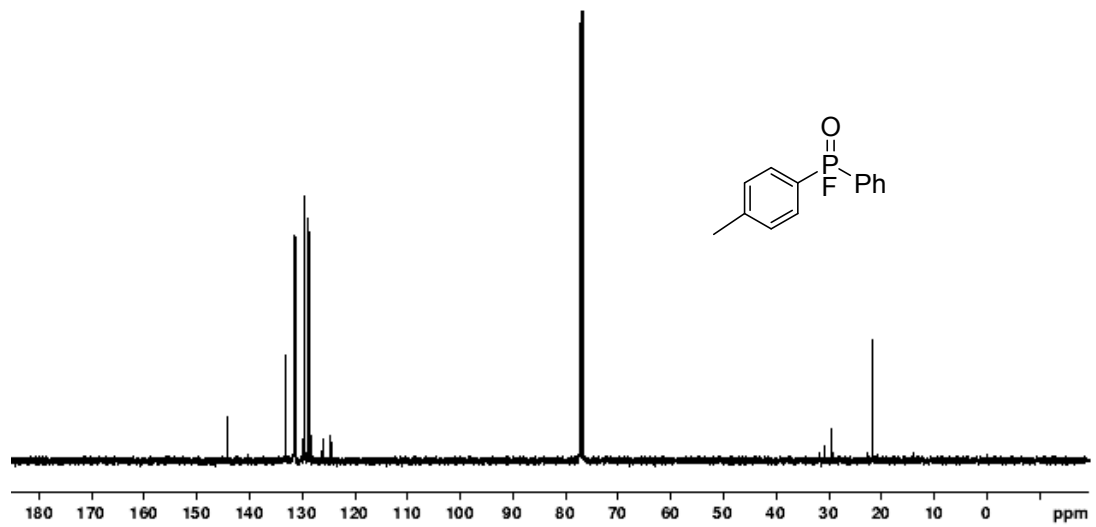




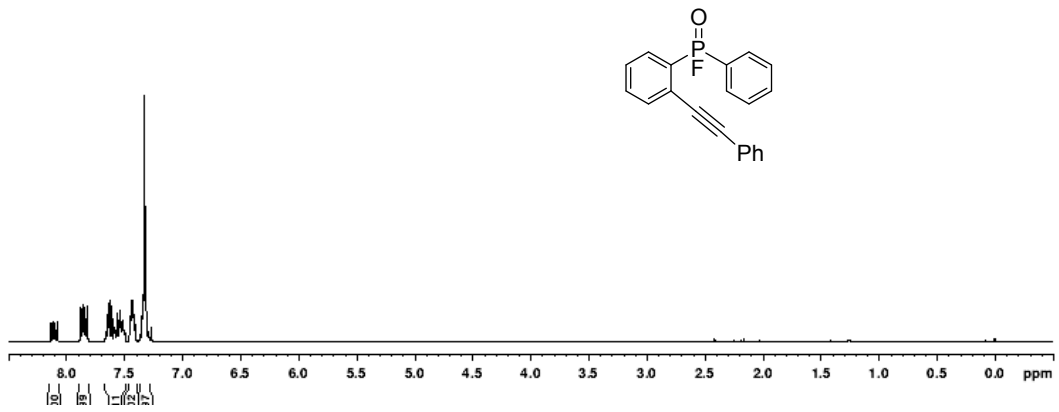
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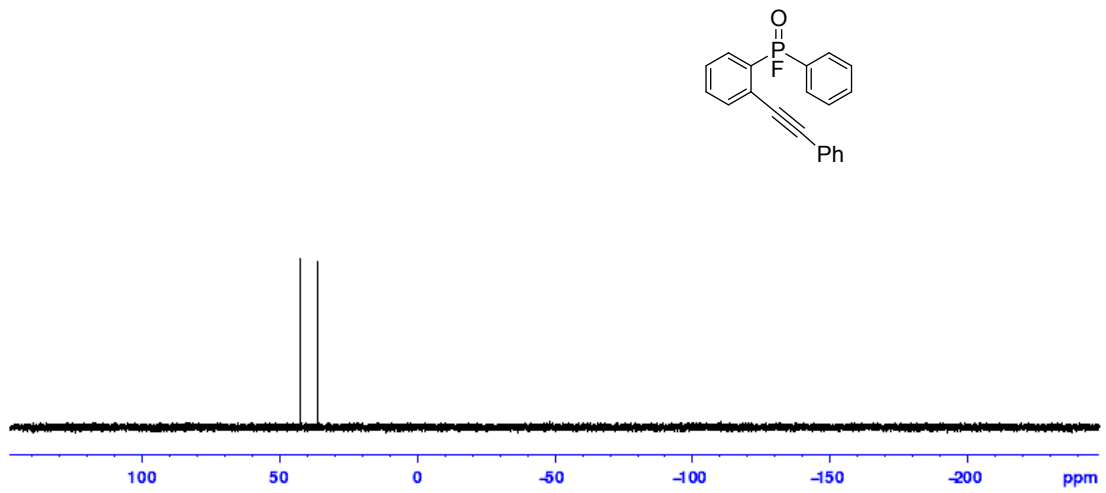
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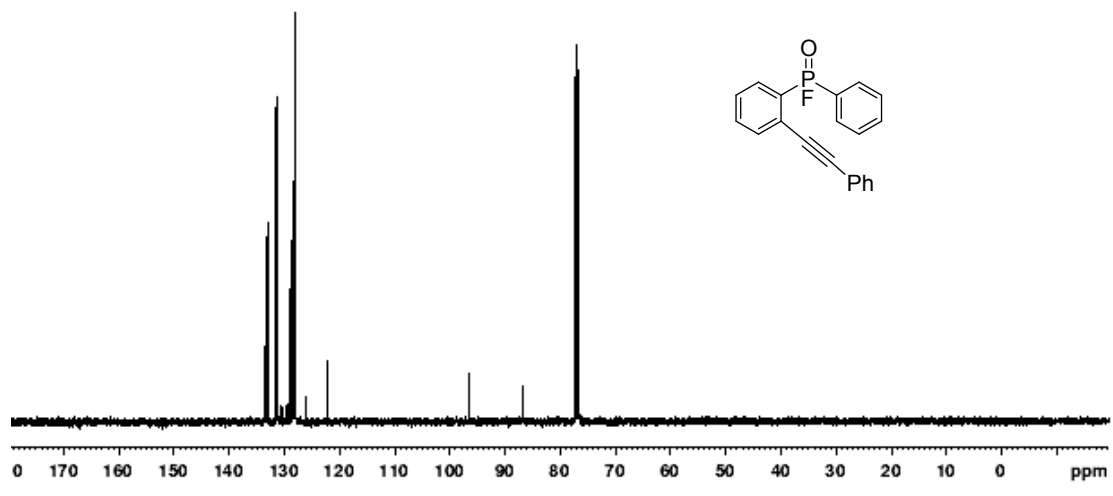


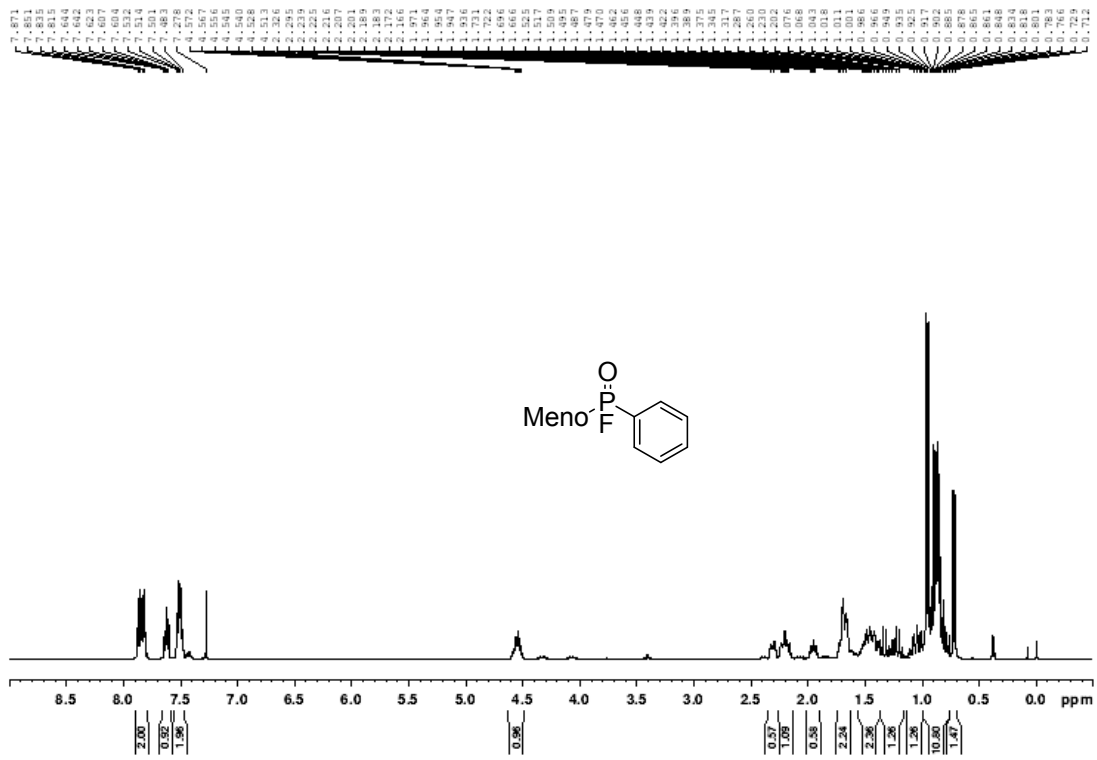
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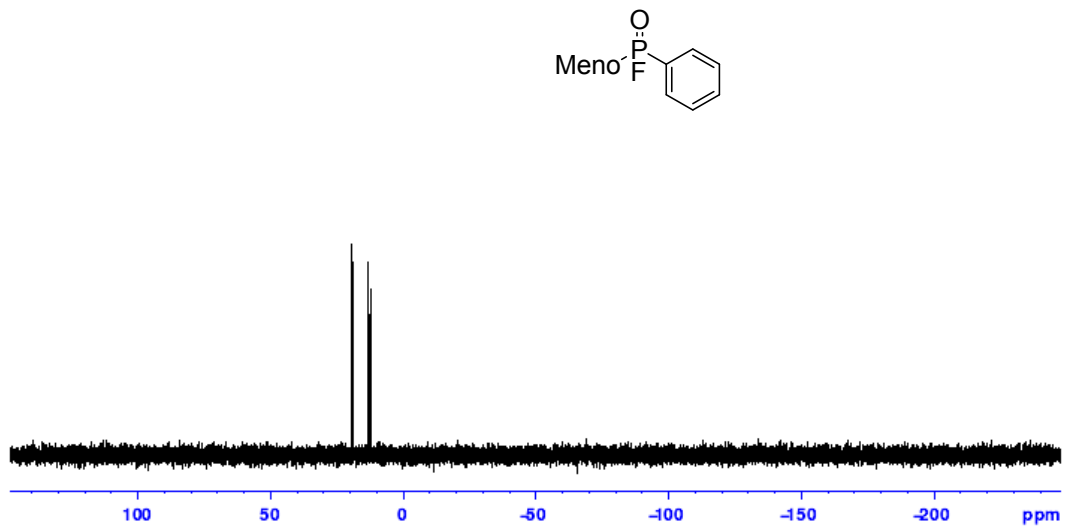


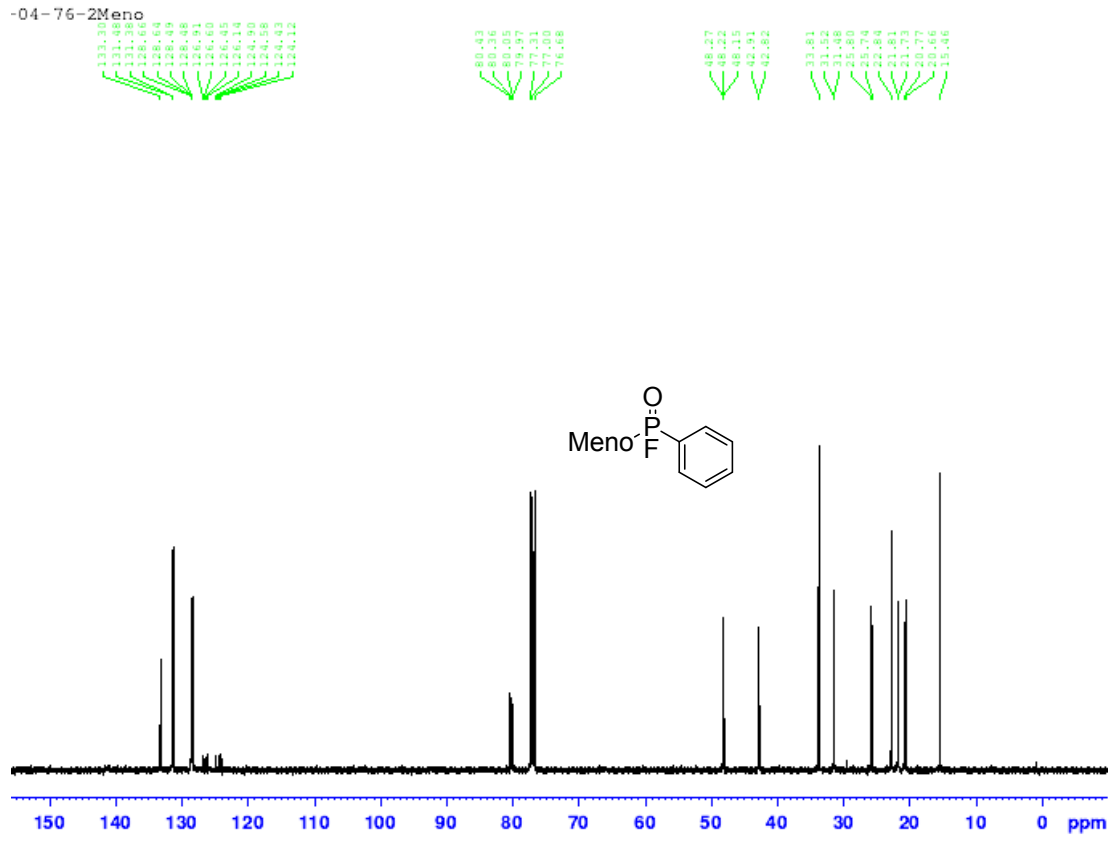
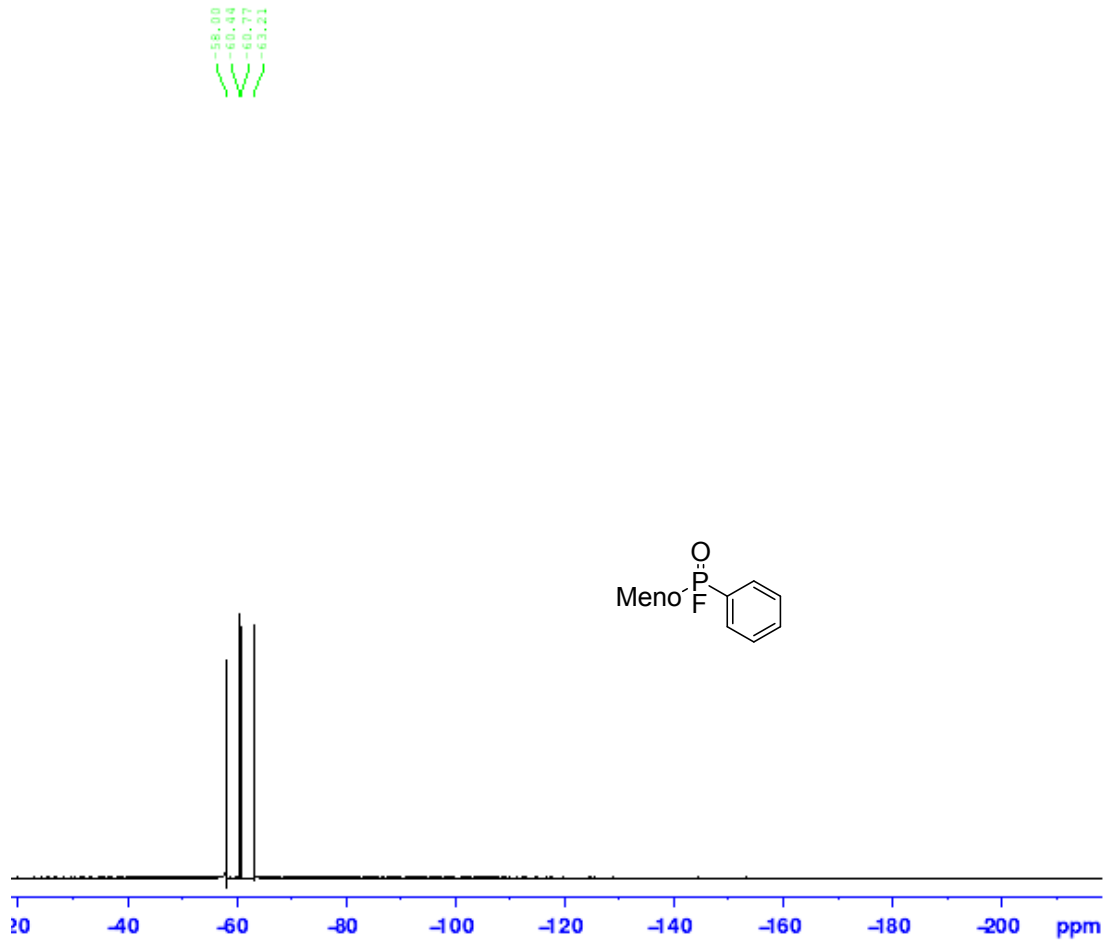
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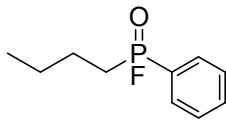
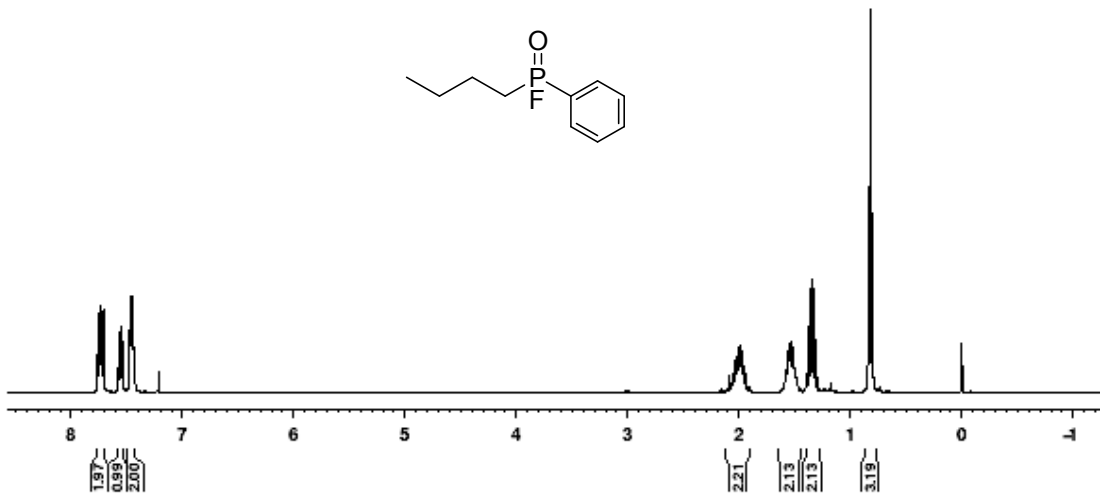
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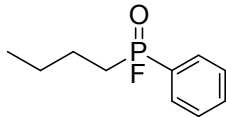
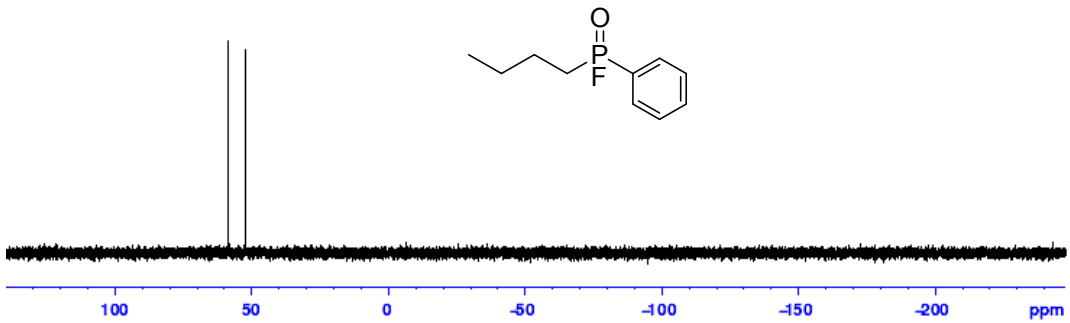
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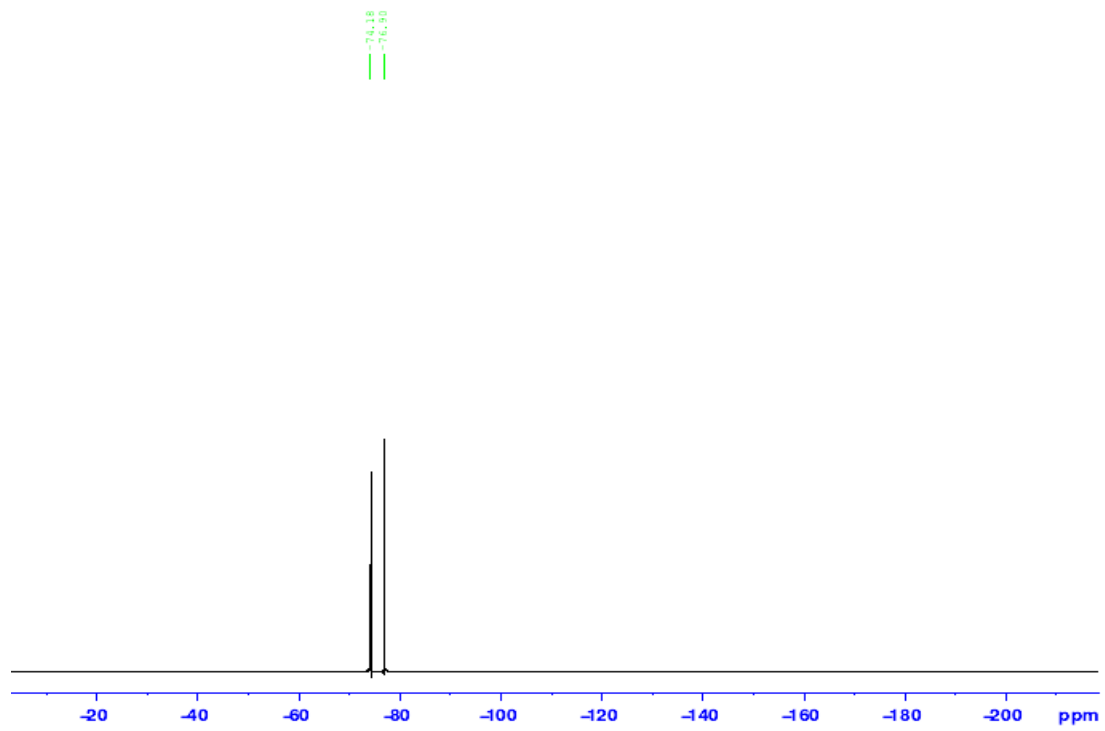
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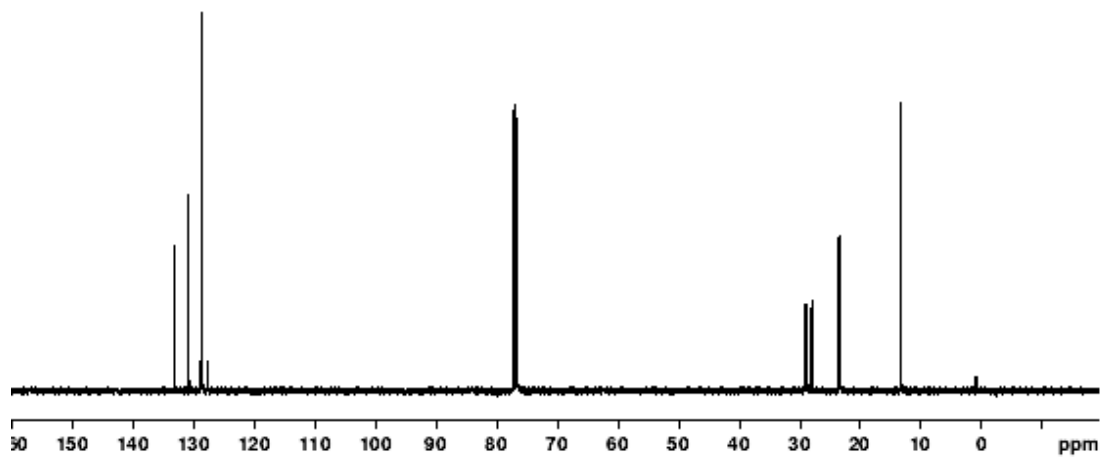
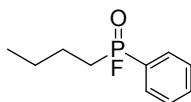




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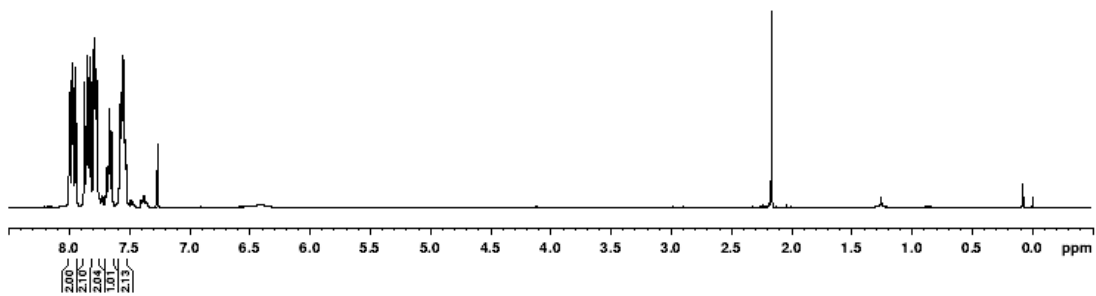
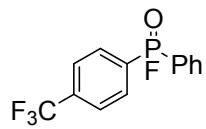
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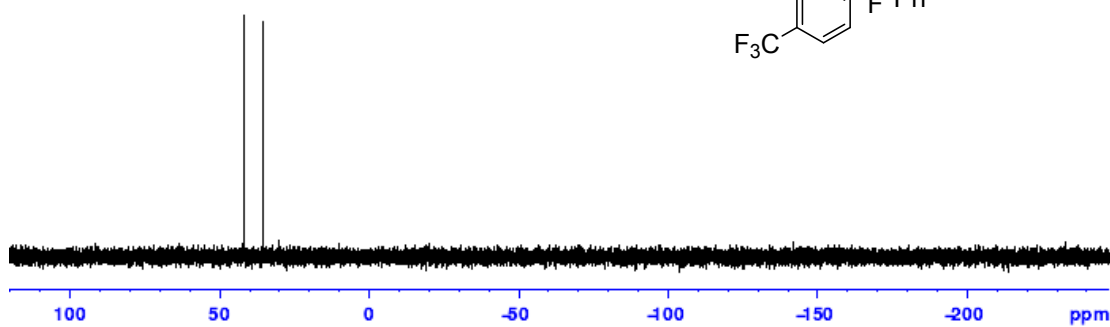
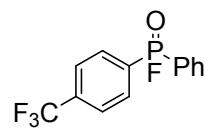


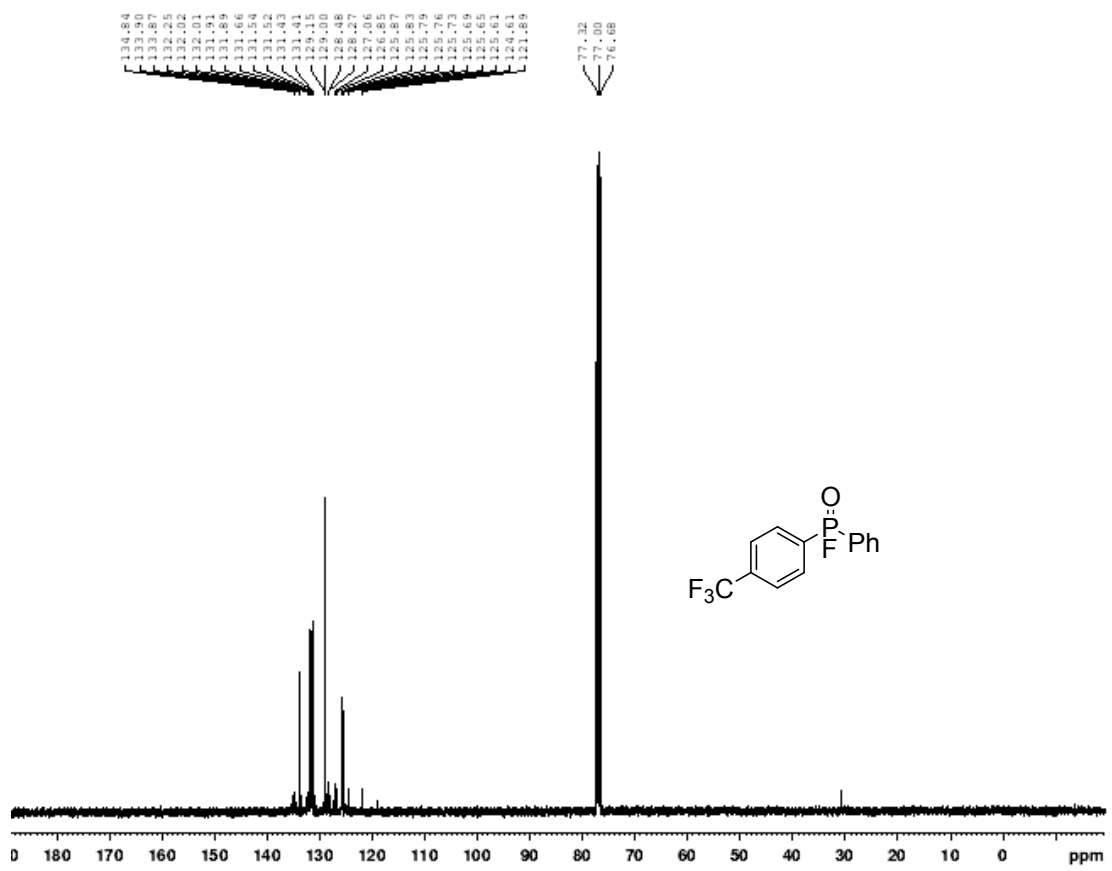
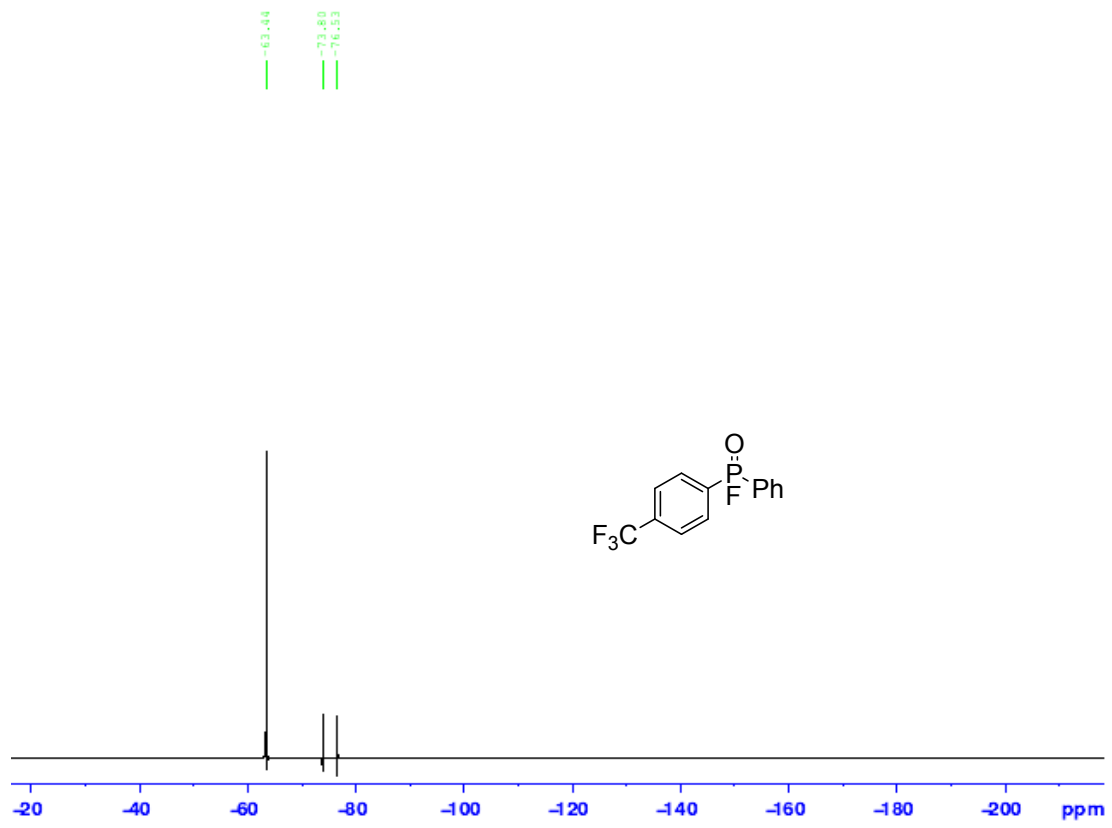
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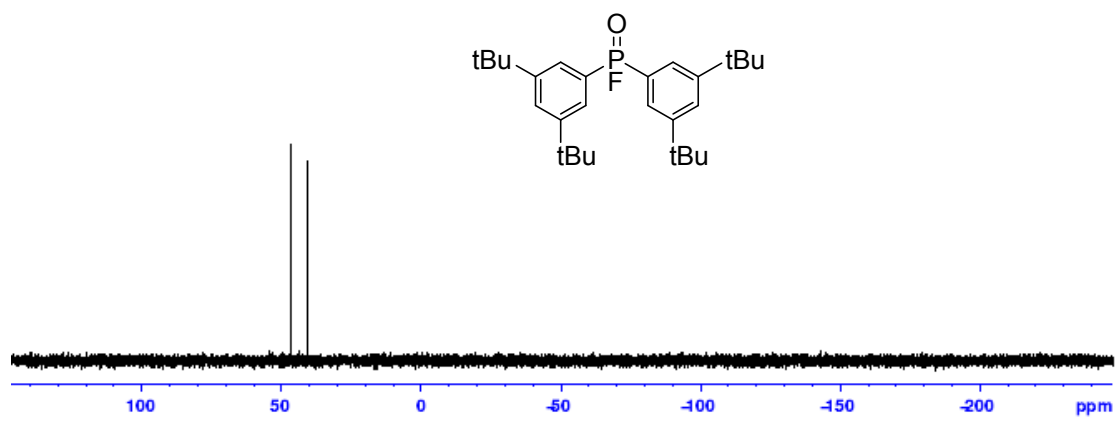
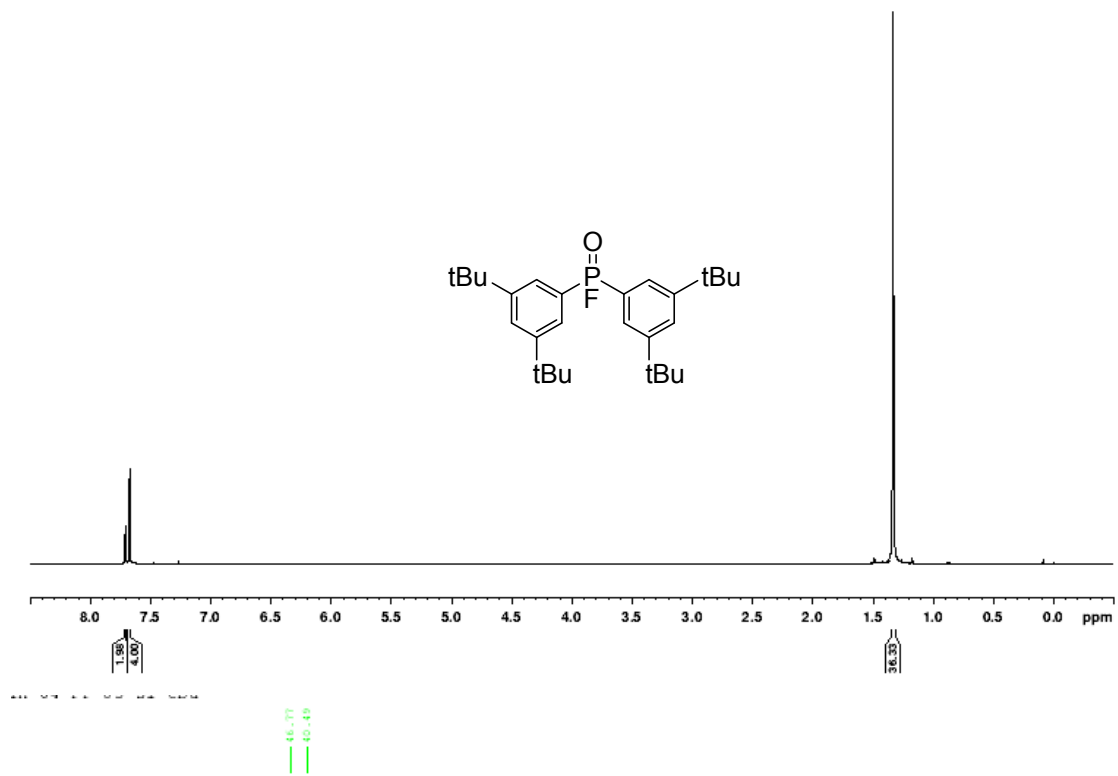
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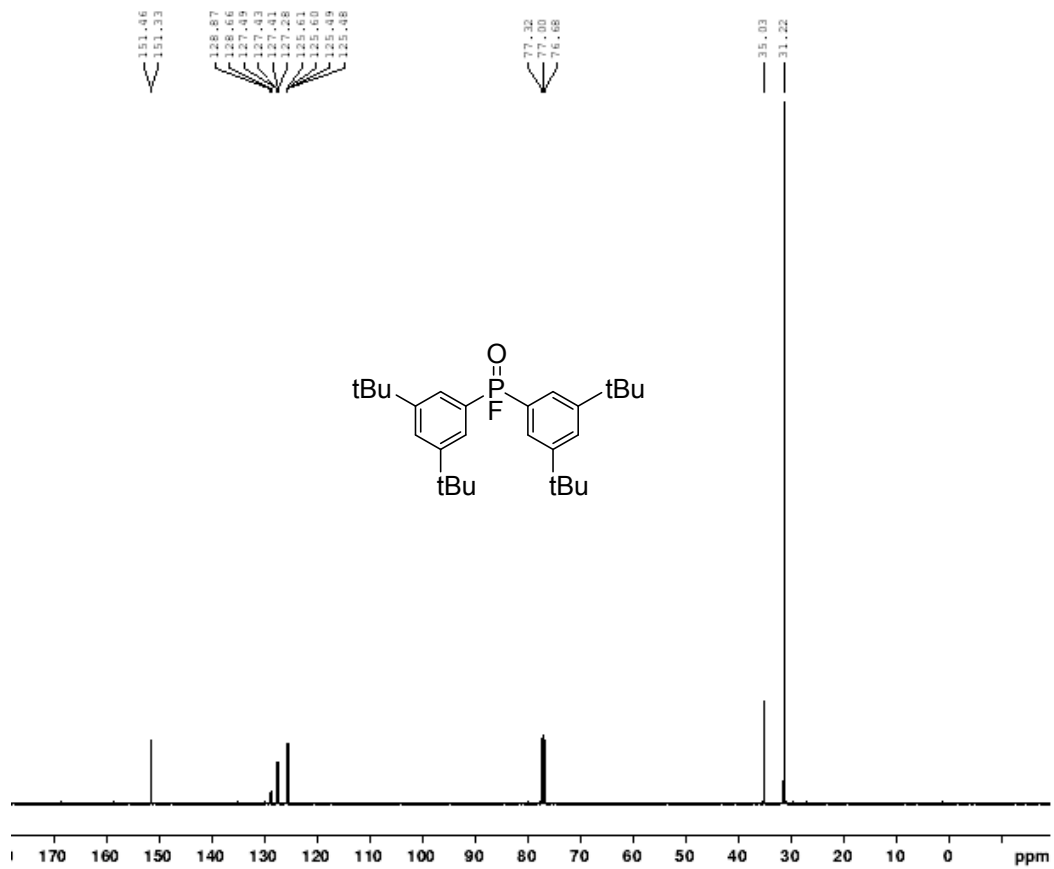
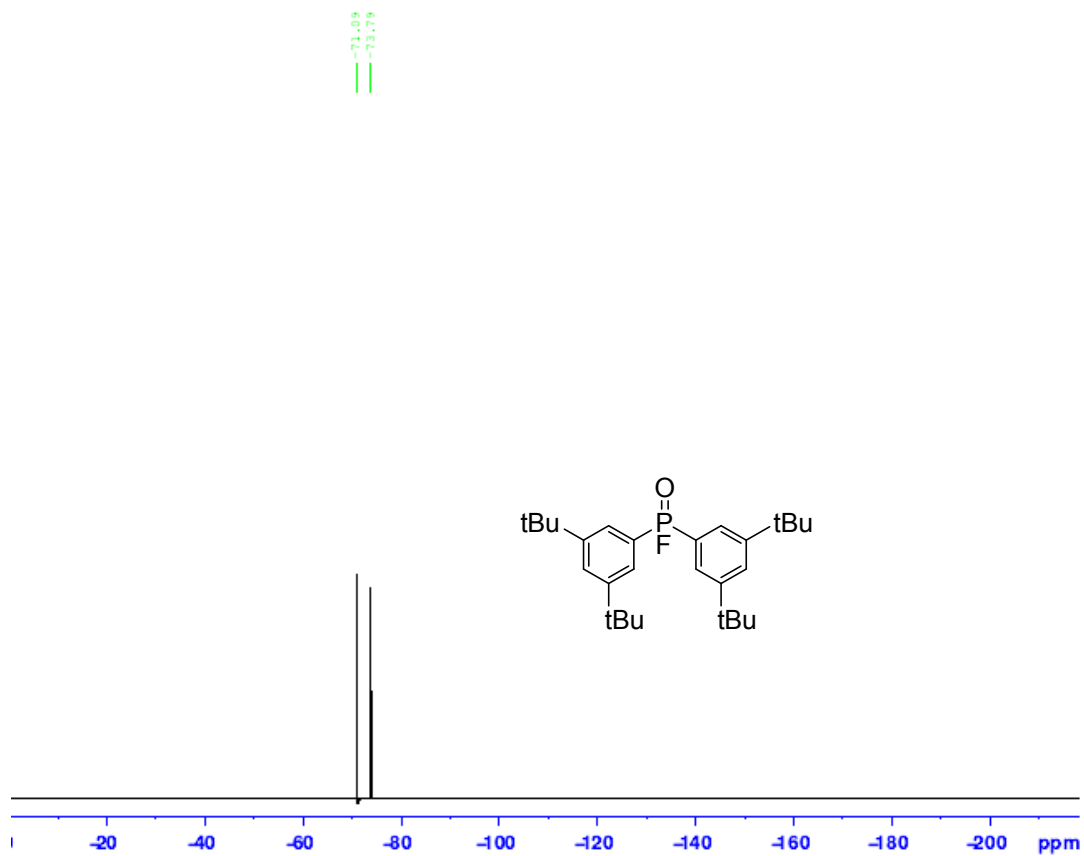


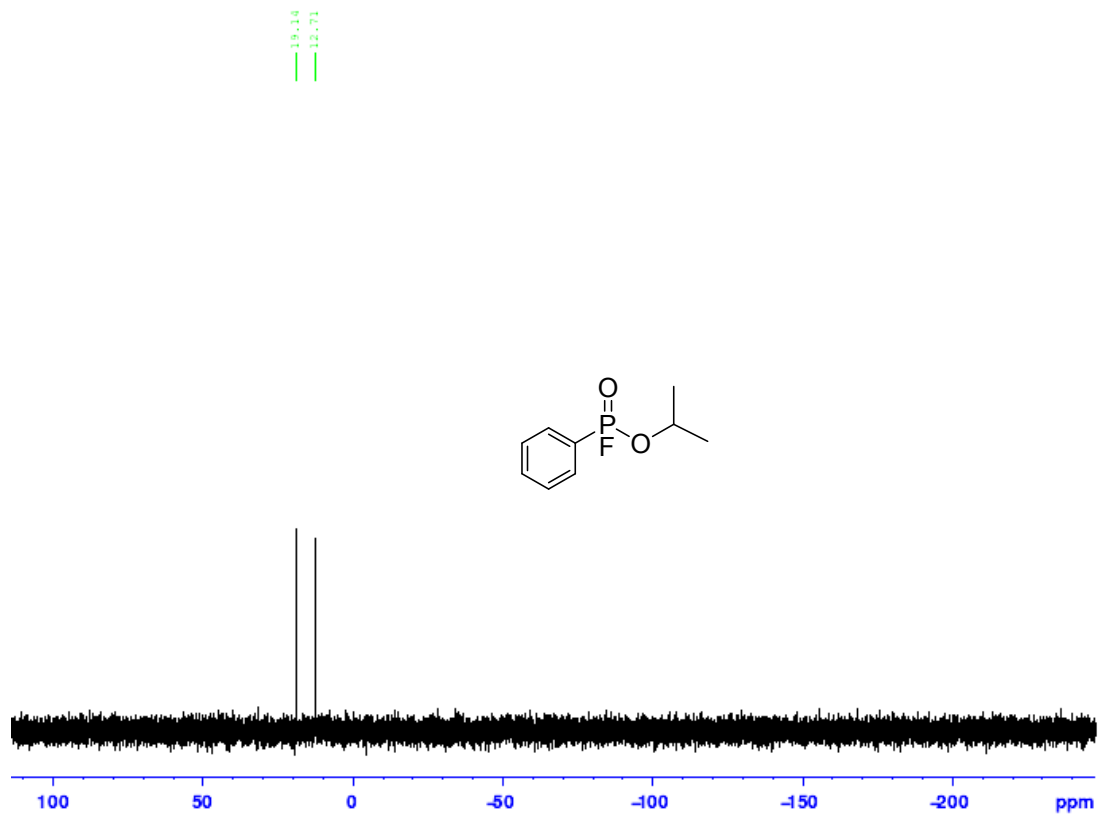
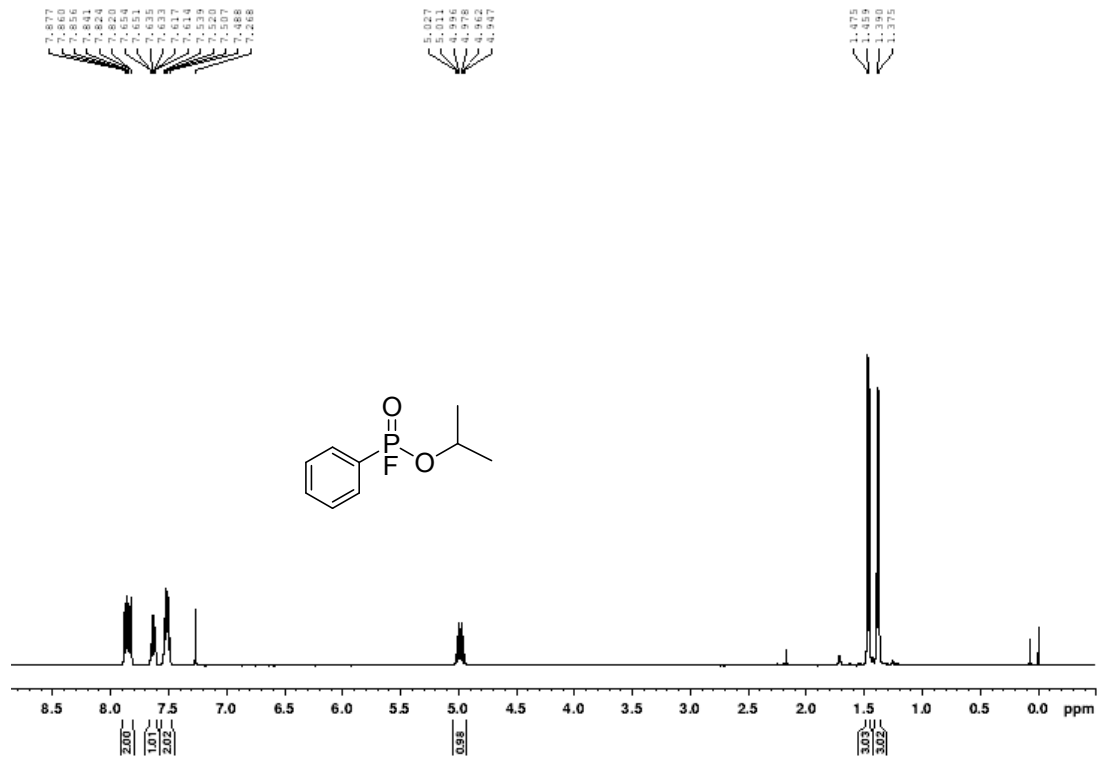
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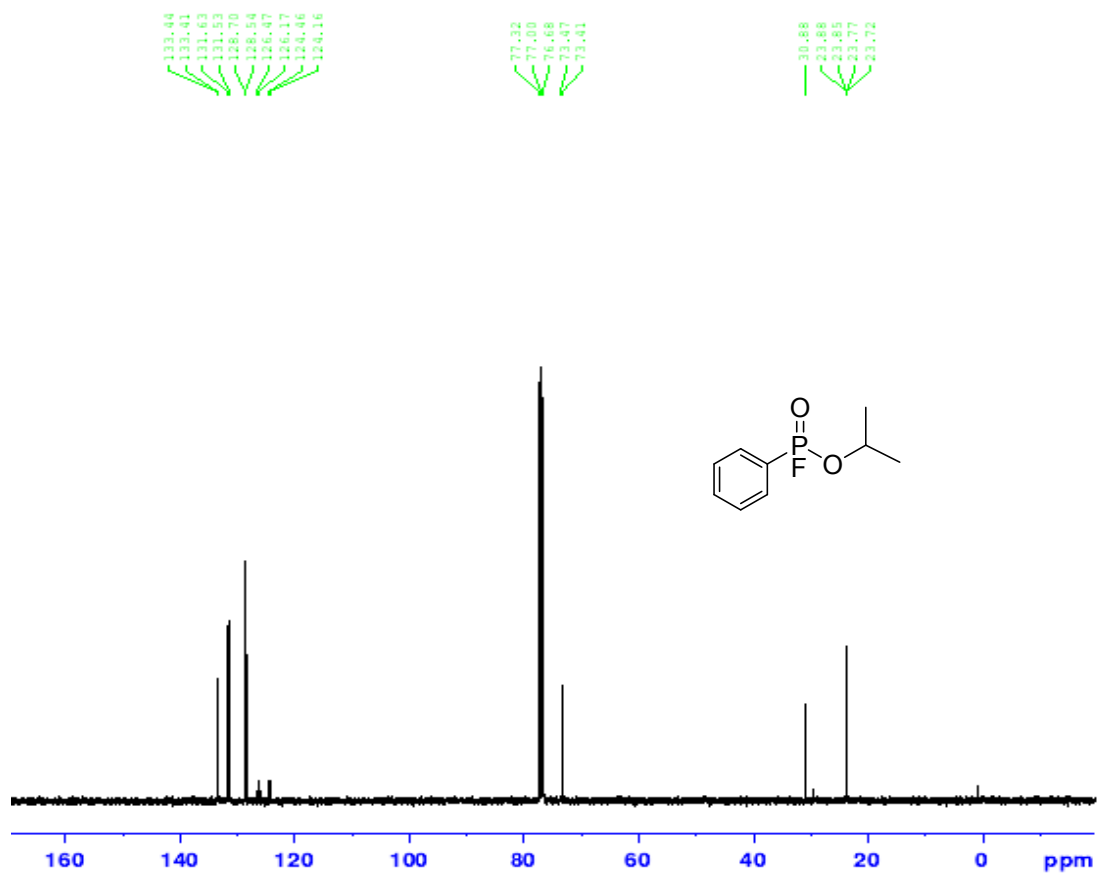
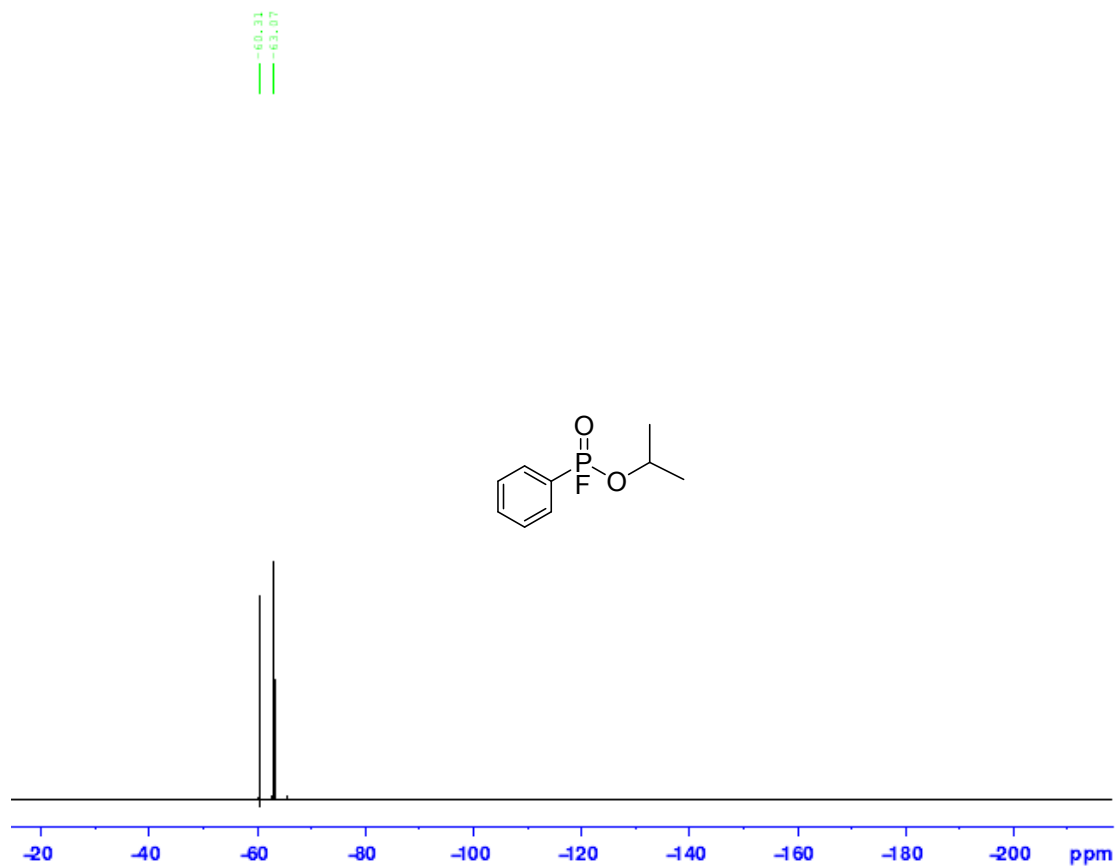








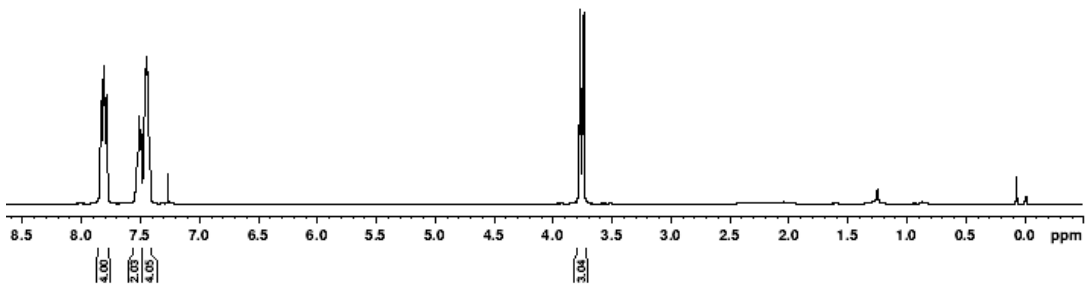
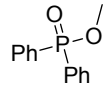




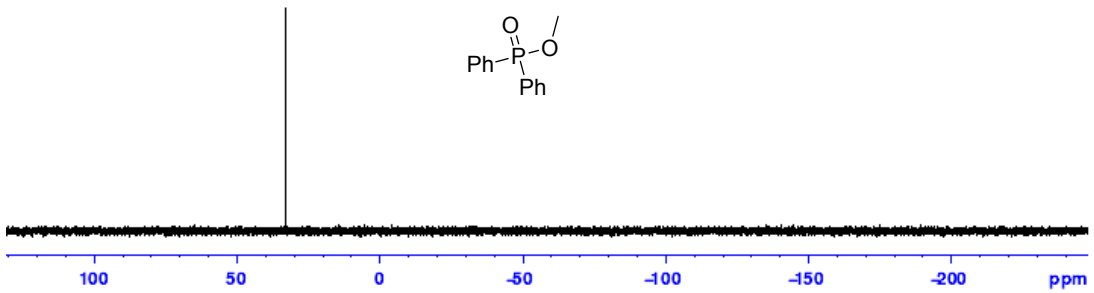
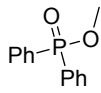
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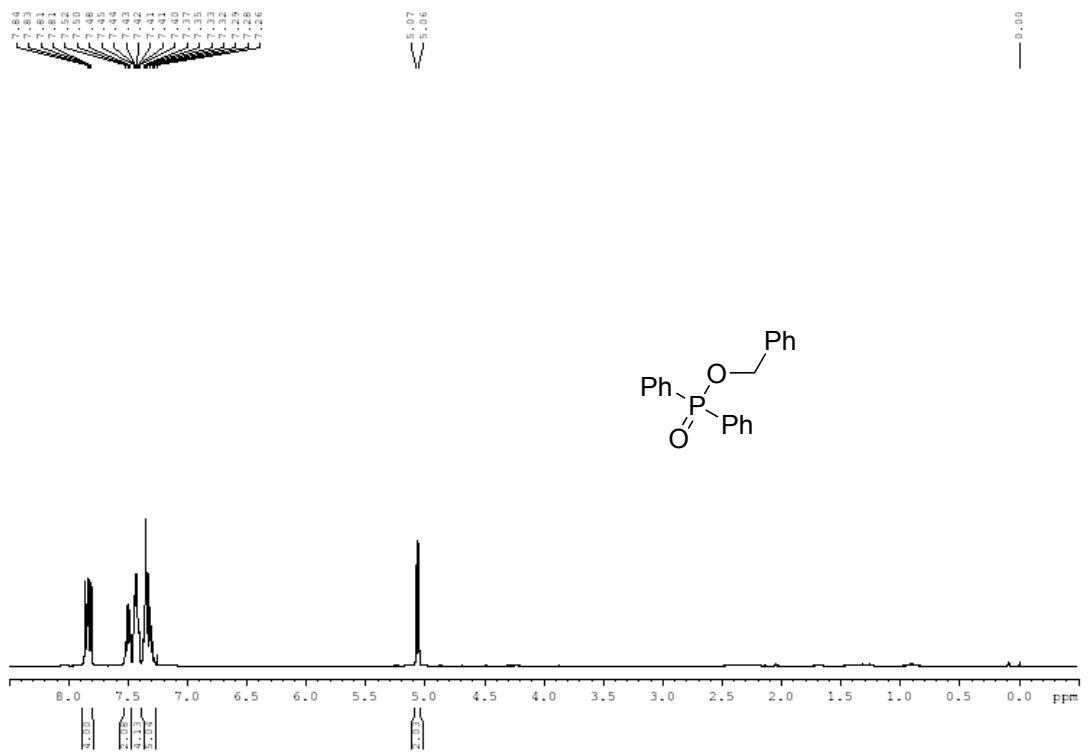
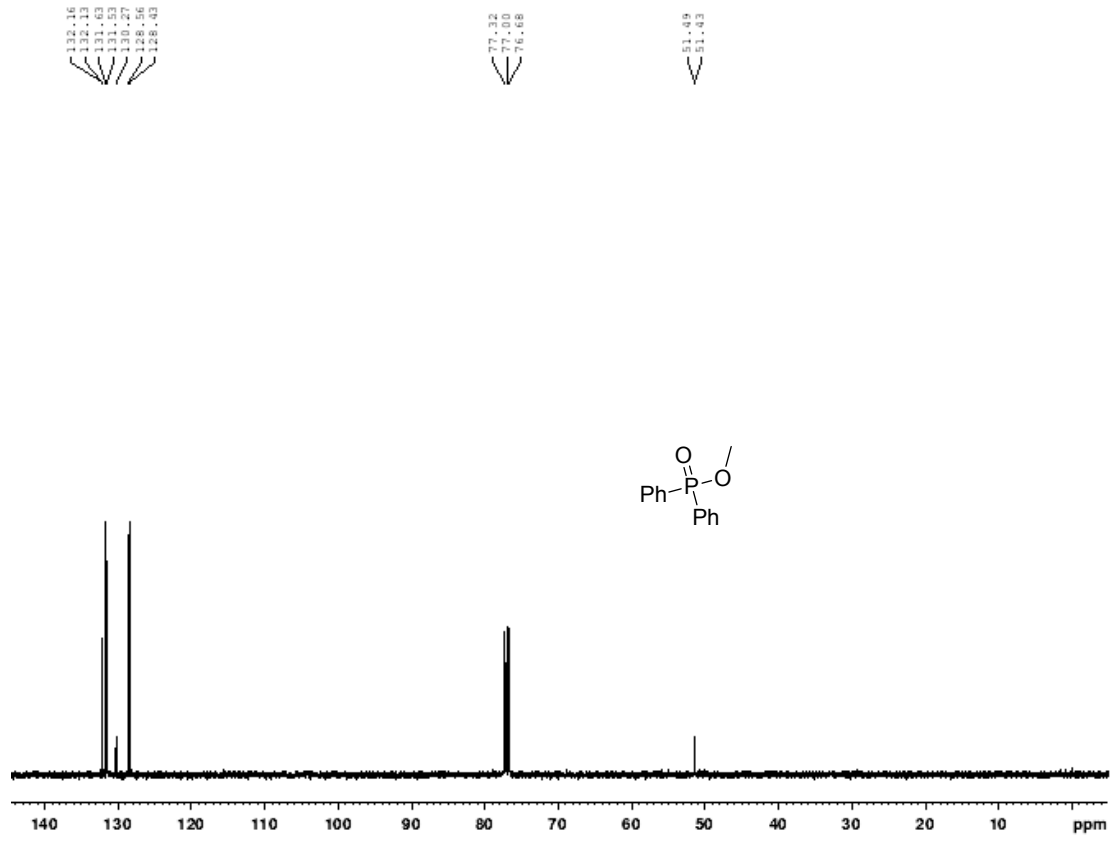
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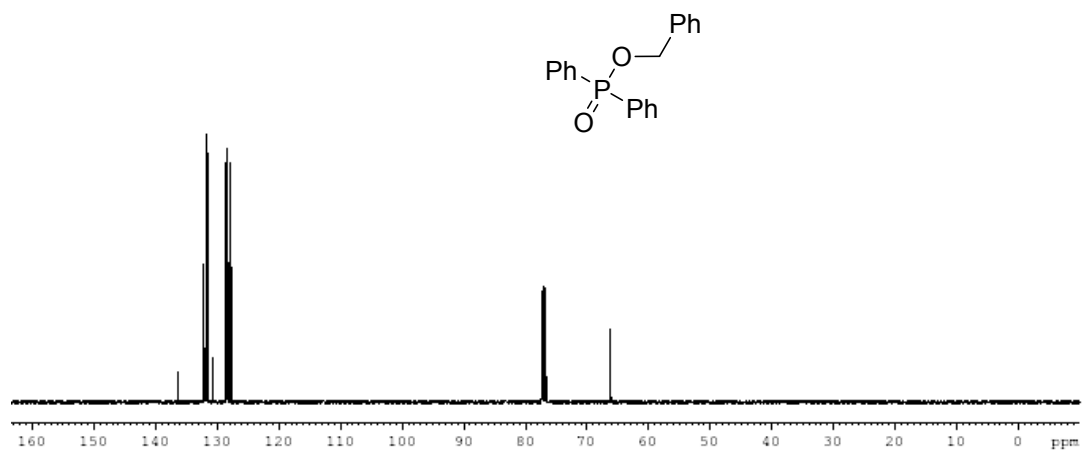
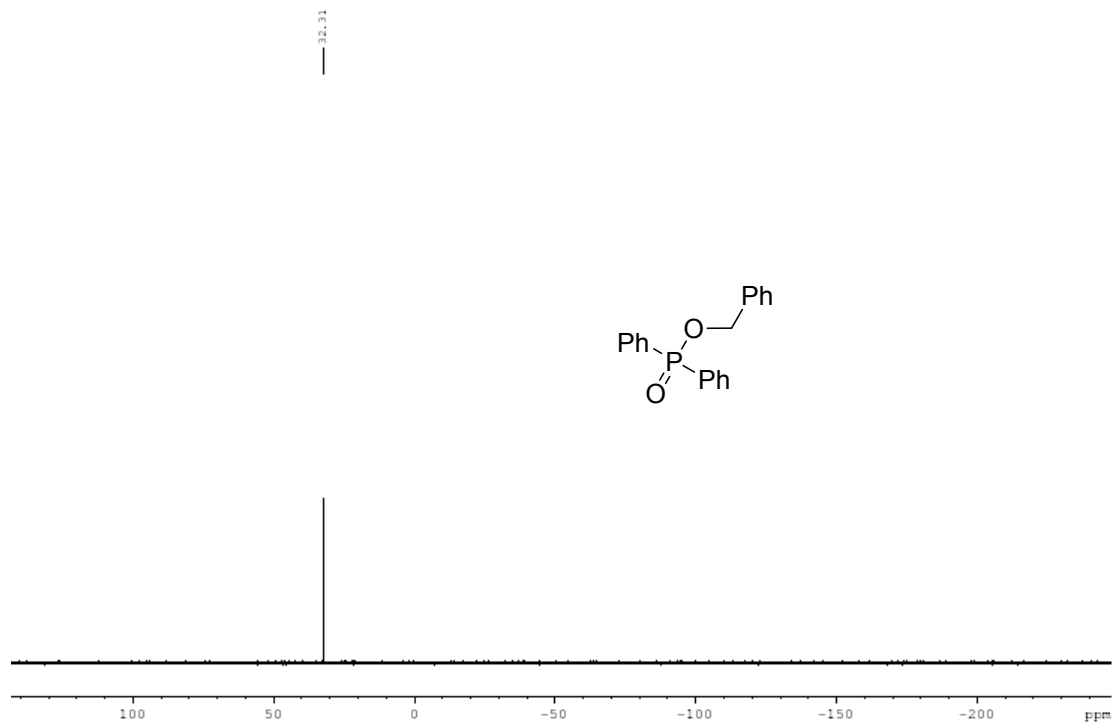
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-0.010



33.22

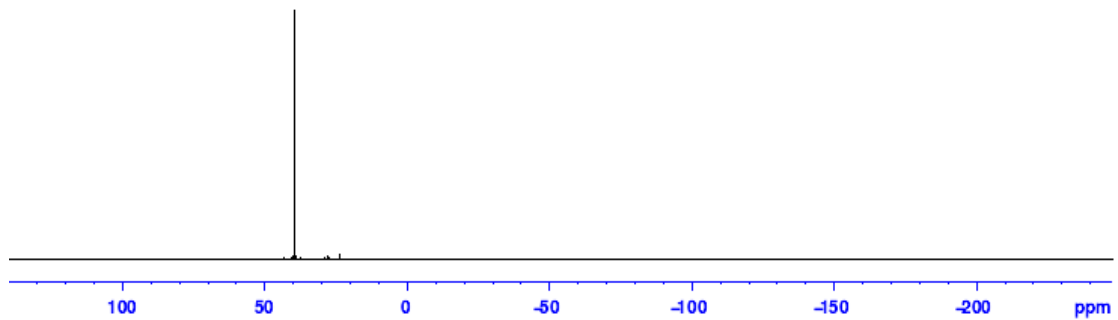
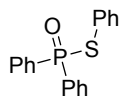
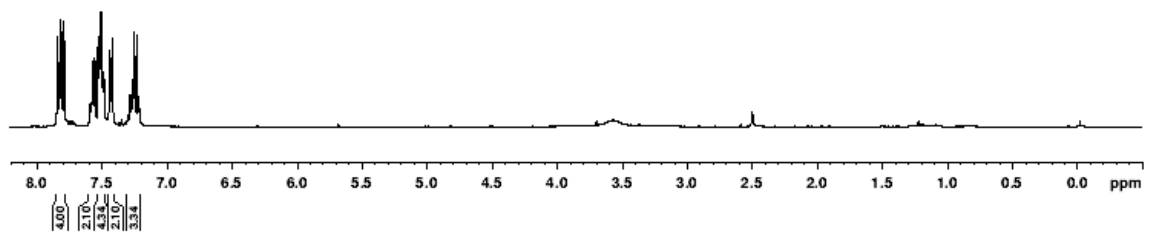
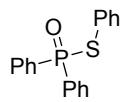


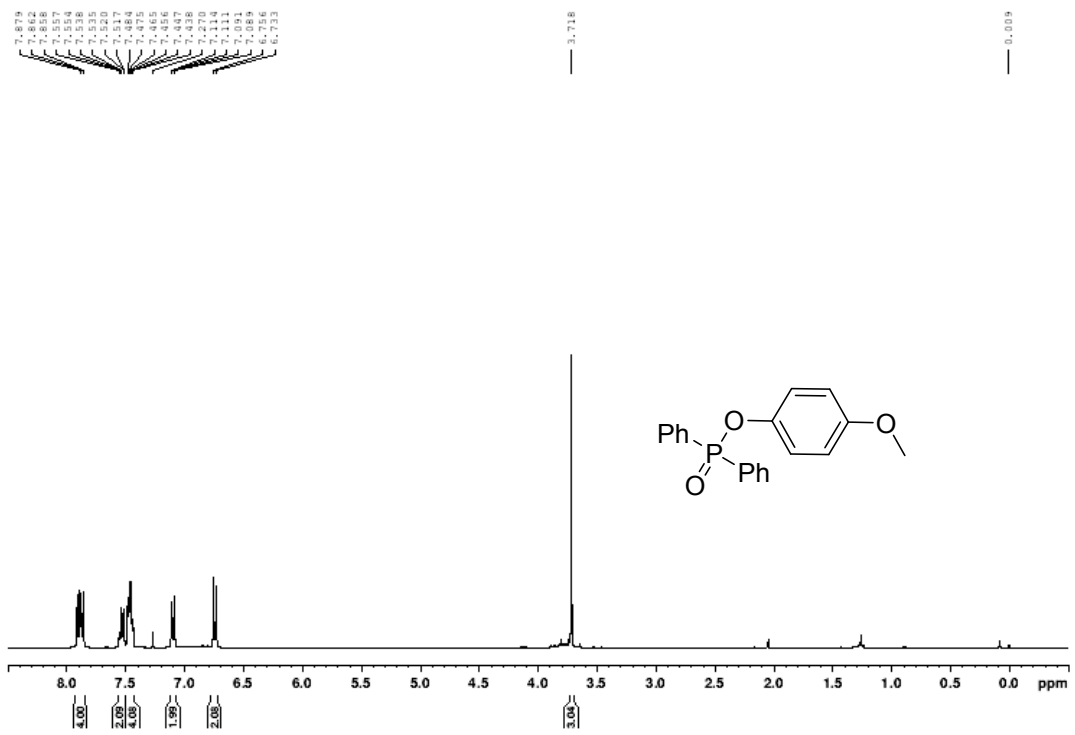
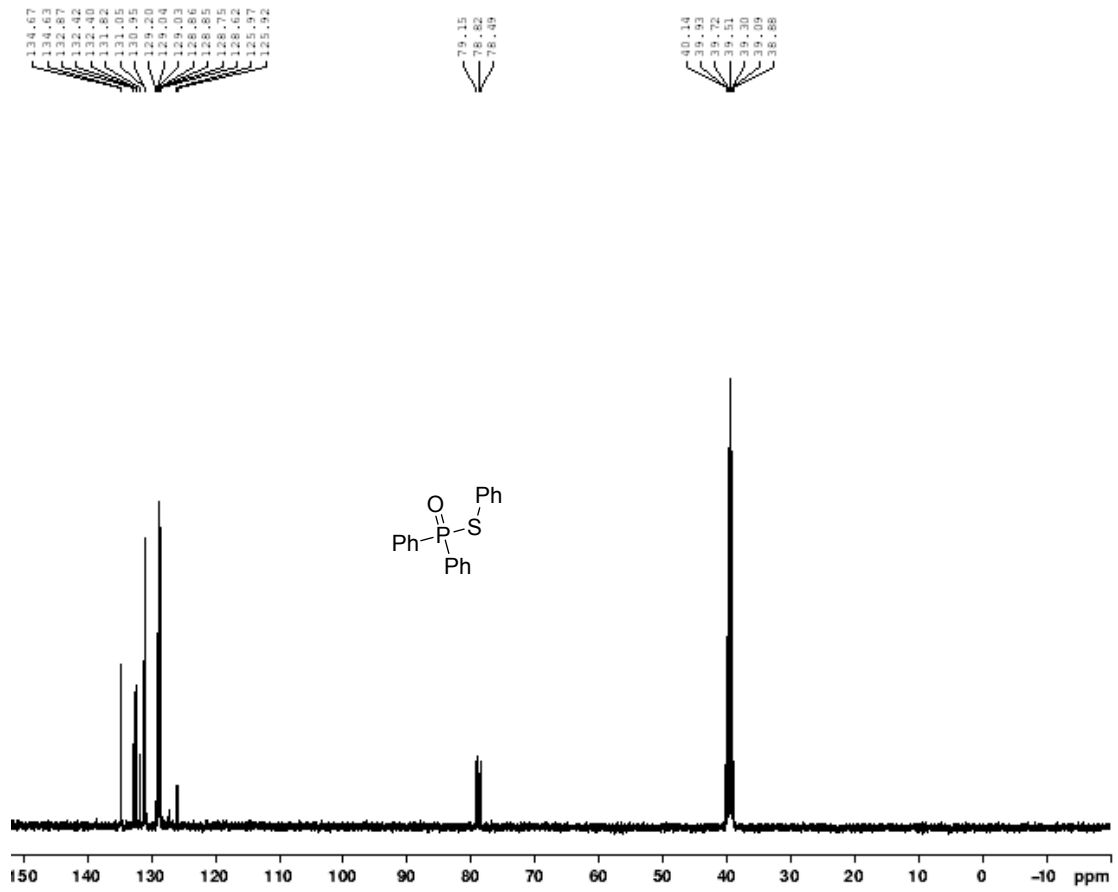


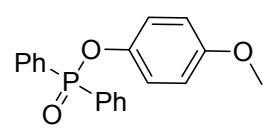
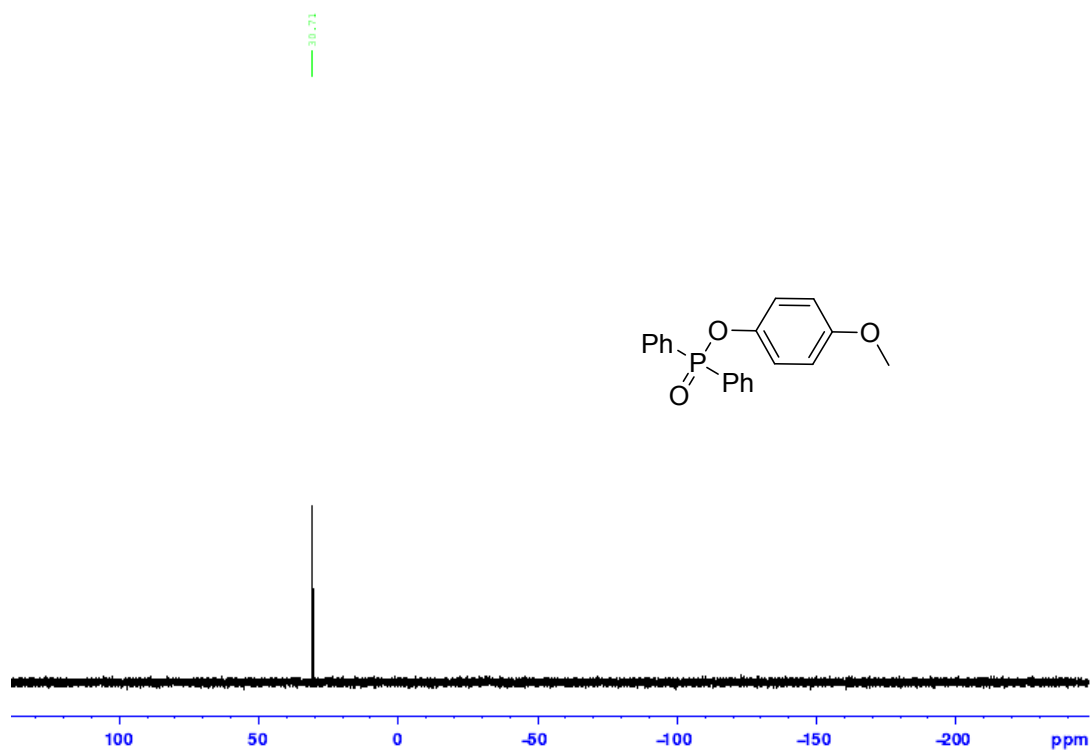


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2.460







- 156.34
- 144.44
- 144.16
- 132.91
- 132.38
- 131.84
- 131.73
- 131.49
- 130.12
- 128.46
- 121.60
- 121.56
- 114.54
- 77.32
- 77.00
- 76.68
- 55.46

