

ELECTRONIC SUPPLEMENTARY INFORMATION

Protein Assembly Mediated by Sulfonatocalix[4]arene

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Methods

Co-crystallization of lysozyme and sclx4. Hen white egg lysozyme (62971 Fluka) was purchased from Sigma-Aldrich and used without further purification. The hanging drop vapour diffusion method was used for crystallization at 20° C. Co-crystals of lysozyme and sclx4 were grown from identical conditions to those reported for cytochrome *c*¹ except that an oil barrier was not used. Drops were prepared by combining 1 µL volumes of lysozyme (1.4 mM), sclx4 (17 mM) and the reservoir solution (20-30 % polyethylene glycol (PEG) 8000, 50 mM NaCl, 100 mM MgCl₂ and 50 mM sodium cacodylate pH 6.3). Premixing the protein and ligand was not possible as this resulted in complete precipitation.

X-ray Diffraction and Structure Determination. Crystals were transferred to reservoir solution supplemented with 25 % glycerol and flash-frozen under a stream of nitrogen gas at 100 K (X-stream 2000). Diffraction data were collected from a single crystal of the lysozyme:sclx4 complex at Soleil (PROXIMA 1, Pilatus 6M detector, φ scans of 0.1° over 180° to a resolution of 1.72 Å). Data processing and scaling were performed in MOSFLM² and SCALA,³ respectively. The data collection and refinement statistics are given in Table S1. The structure was solved by molecular replacement in PHASER.⁴ Refinement and manual rebuilding were performed in REFMAC5 as implemented in CCP4⁵ and COOT,⁶ respectively. Solvent molecules were placed automatically using ARP/wARP⁷ and refinement was continued until no features remained in the F_o – F_c difference maps. Molprobity⁸ was used to check the structure quality. Coordinates and structure factors were deposited in the Protein Data Bank with the accession code 4PRQ. The protein-ligand and protein-protein interfaces were analysed in COOT and PISA.⁹ Interface areas [the inaccessible surface (Å²) of the protein or ligand in the complex] were calculated as described previously.¹

Arginine solvent accessibility in lysozyme. 15 high resolution (0.9-2.1 Å) crystal structures of hen egg white lysozyme were analyzed using the Accessible Surface Areas calculation in CCP4.⁴ Each file (PDB codes: 194L, 1GWD, 1JIS, 1LPI, 1YIK, 2CDS, 2D4K, 2FBB, 2I25, 2ZQ3, 3A67, 3AGH, 3AW7, 3LZT, 4J1A) was manually edited to contain a single copy of lysozyme. Other proteins, water molecules, ions and alternate conformations were removed prior to the calculations.

Table S1. Summary of crystallization conditions, data collection and refinement statistics

<i>Crystallization Conditions^a</i>	
[protein], [sclx ₄] (mM)	1.4, 17
PEG 8,000 (%)	24
Buffer, (CH ₃) ₂ AsO ₂ Na	0.05 M, pH 6.3
Salts	0.05 M NaCl, 0.1 M MgCl ₂
<i>Data Collection^b</i>	
Space group	<i>P</i> 2 ₁
	<i>a</i> = 44.01 Å
	<i>b</i> = 81.75 Å
Cell constants	<i>c</i> = 72.10 Å
	$\alpha = \gamma = 90^\circ$
	$\beta = 105.37^\circ$
Resolution (Å)	81.75-1.72 (1.77-1.72)
Wavelength (Å)	1.00792
Unique reflections	1386759 (51142)
Multiplicity	3.0 (2.6)
<i>I</i> / σ	9.4 (1.8)
Completeness (%)	98.7 (94.9)
<i>R</i> _{merge} ^c (%)	7.3 (55.6)
Solvent content (%)	43.86
<i>Refinement</i>	
<i>R</i> _{factor} (%)	18.48
<i>R</i> _{free} (%)	22.42
rmsd ^d bonds (Å)	0.01
rmsd angles (°)	1.12
# molecules in asymmetric unit	
Protein	4
sclx ₄	5
PEG	5
Mg ²⁺	3
Solvent	322
Average <i>B</i> factors ^e (Å ²)	
Protein	21.51
sclx ₄	19.70
PEG	45.46
Mg ²⁺	28.88
Solvent	28.44
Ramachandran analysis ^f	
% residues (favoured regions)	98.8
% residues (allowed regions)	100.0

^aThe crystallization drops comprised 1 μL each of the protein, ligand and reservoir solutions; ^bValues in parentheses correspond to the highest resolution shell; ^c $R_{\text{merge}} = \sum_{hkl} \sum_i |I_i(hkl) - \langle I(hkl) \rangle| / \sum_{hkl} \sum_i I_i(hkl)$; ^droot mean square deviation; ^ecalculated from the B values of all non-hydrogen atoms; ^fcalculated with Molprobity.

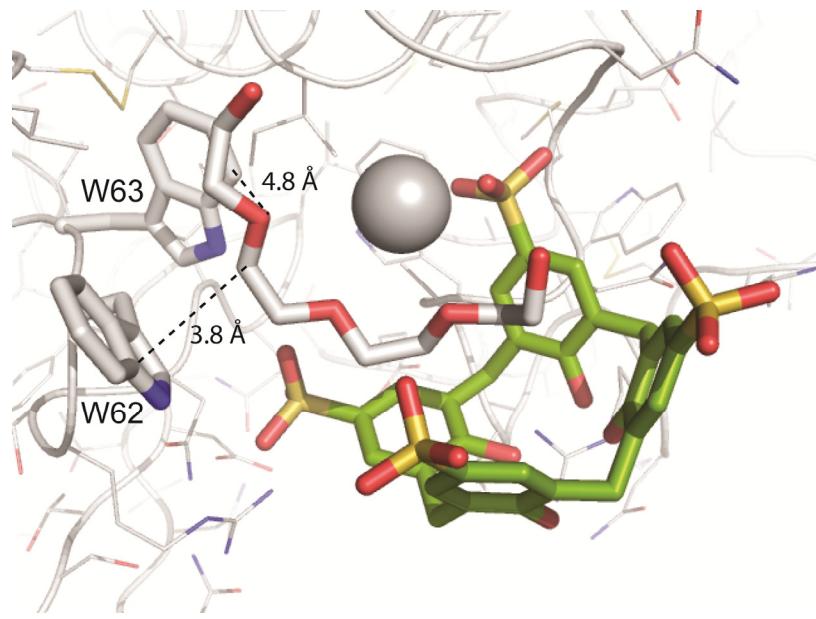


Fig S1. A calixarene bound near the active site in chain B. This calixarene is complexed with Mg^{2+} and a fragment of PEG (in a crown ether like conformation). The PEG also makes van der Waals contacts with the indole rings of the active site residues Trp62 and Trp63.

References

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