

*ELECTRONIC SUPPLEMENTARY INFORMATION*

**Protein Assembly Mediated by Sulfonatocalix[4]arene**

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## Methods

**Co-crystallization of lysozyme and sclx<sub>4</sub>.** Hen white egg lysozyme (62971 Fluka) was purchased from Sigma-Aldrich and used without further purification. The hanging drop vapour diffusion method was used for crystallization at 20° C. Co-crystals of lysozyme and sclx<sub>4</sub> were grown from identical conditions to those reported for cytochrome *c*<sup>1</sup> except that an oil barrier was not used. Drops were prepared by combining 1 µL volumes of lysozyme (1.4 mM), sclx<sub>4</sub> (17 mM) and the reservoir solution (20-30 % polyethylene glycol (PEG) 8000, 50 mM NaCl, 100 mM MgCl<sub>2</sub> and 50 mM sodium cacodylate pH 6.3). Premixing the protein and ligand was not possible as this resulted in complete precipitation.

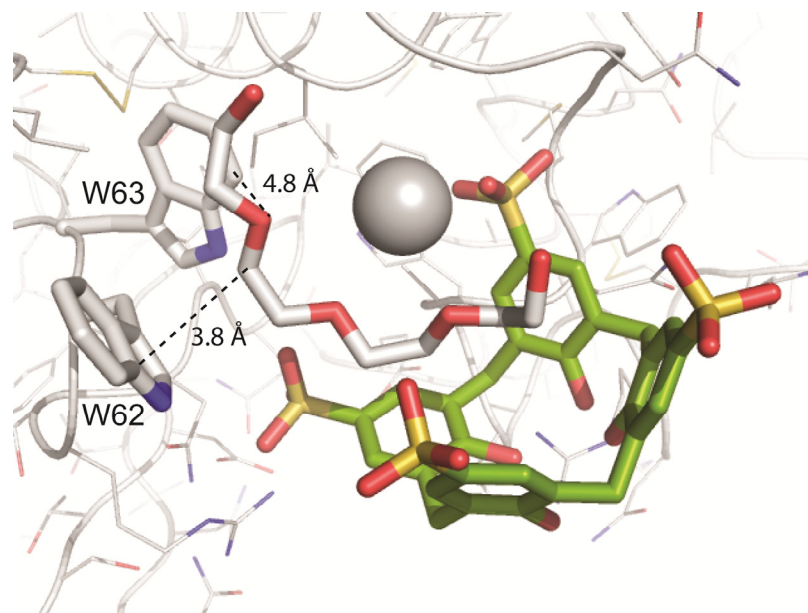
**X-ray Diffraction and Structure Determination.** Crystals were transferred to reservoir solution supplemented with 25 % glycerol and flash-frozen under a stream of nitrogen gas at 100 K (X-stream 2000). Diffraction data were collected from a single crystal of the lysozyme:sclx<sub>4</sub> complex at Soleil (PROXIMA 1, Pilatus 6M detector,  $\phi$  scans of 0.1° over 180° to a resolution of 1.72 Å). Data processing and scaling were performed in MOSFLM<sup>2</sup> and SCALA,<sup>3</sup> respectively. The data collection and refinement statistics are given in Table S1. The structure was solved by molecular replacement in PHASER.<sup>4</sup> Refinement and manual rebuilding were performed in REFMAC5 as implemented in CCP4<sup>5</sup> and COOT,<sup>6</sup> respectively. Solvent molecules were placed automatically using ARP/wARP<sup>7</sup> and refinement was continued until no features remained in the  $F_o - F_c$  difference maps. Molprobity<sup>8</sup> was used to check the structure quality. Coordinates and structure factors were deposited in the Protein Data Bank with the accession code 4PRQ. The protein-ligand and protein-protein interfaces were analysed in COOT and PISA.<sup>9</sup> Interface areas [the inaccessible surface (Å<sup>2</sup>) of the protein or ligand in the complex] were calculated as described previously.<sup>1</sup>

**Arginine solvent accessibility in lysozyme.** 15 high resolution (0.9-2.1 Å) crystal structures of hen egg white lysozyme were analyzed using the Accessible Surface Areas calculation in CCP4.<sup>4</sup> Each file (PDB codes: 194L, 1GWD, 1JIS, 1LPI, 1YIK, 2CDS, 2D4K, 2FBB, 2I25, 2ZQ3, 3A67, 3AGH, 3AW7, 3LZT, 4J1A) was manually edited to contain a single copy of lysozyme. Other proteins, water molecules, ions and alternate conformations were removed prior to the calculations.

**Table S1.** Summary of crystallization conditions, data collection and refinement statistics

<i>Crystallization Conditions<sup>a</sup></i>	
[protein], [sclx <sub>4</sub> ] (mM)	1.4, 17
PEG 8,000 (%)	24
Buffer, (CH <sub>3</sub> ) <sub>2</sub> AsO <sub>2</sub> Na	0.05 M, pH 6.3
Salts	0.05 M NaCl, 0.1 M MgCl <sub>2</sub>
<i>Data Collection<sup>b</sup></i>	
Space group	<i>P</i> 2 <sub>1</sub>
	<i>a</i> = 44.01 Å
	<i>b</i> = 81.75 Å
Cell constants	<i>c</i> = 72.10 Å
	<i>α</i> = <i>γ</i> = 90°
	<i>β</i> = 105.37°
Resolution (Å)	81.75-1.72 (1.77-1.72)
Wavelength (Å)	1.00792
Unique reflections	1386759 (51142)
Multiplicity	3.0 (2.6)
<i>I</i> / <i>σ</i>	9.4 (1.8)
Completeness (%)	98.7 (94.9)
<i>R</i> <sub>merge</sub> <sup>c</sup> (%)	7.3 (55.6)
Solvent content (%)	43.86
<i>Refinement</i>	
<i>R</i> <sub>factor</sub> (%)	18.48
<i>R</i> <sub>free</sub> (%)	22.42
rmsd <sup>d</sup> bonds (Å)	0.01
rmsd angles (°)	1.12
# molecules in asymmetric unit	
Protein	4
sclx <sub>4</sub>	5
PEG	5
Mg <sup>2+</sup>	3
Solvent	322
Average <i>B</i> factors <sup>e</sup> (Å <sup>2</sup> )	
Protein	21.51
sclx <sub>4</sub>	19.70
PEG	45.46
Mg <sup>2+</sup>	28.88
Solvent	28.44
Ramachandran analysis <sup>f</sup>	
% residues (favoured regions)	98.8
% residues (allowed regions)	100.0

<sup>a</sup>The crystallization drops comprised 1 μL each of the protein, ligand and reservoir solutions; <sup>b</sup>Values in parentheses correspond to the highest resolution shell; <sup>c</sup> $R_{\text{merge}} = \frac{\sum_{hkl} \sum_i |I_i(hkl) - \langle I(hkl) \rangle|}{\sum_{hkl} \sum_i I_i(hkl)}$ ; <sup>d</sup>root mean square deviation; <sup>e</sup>calculated from the *B* values of all non-hydrogen atoms; <sup>f</sup>calculated with Molprobity.



**Fig S1.** A calixarene bound near the active site in chain B. This calixarene is complexed with  $\text{Mg}^{2+}$  and a fragment of PEG (in a crown ether like conformation). The PEG also makes van der Waals contacts with the indole rings of the active site residues Trp62 and Trp63.

## References

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