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Electronic Supplementary Information (ESI)

for

A low dose, highly selective and sensitive colorimetric and fluorescent probe for biothiols and its application for bioimaging

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1. Experimental general

All reagents were purchased from commercial suppliers and used without further purification. All aqueous solutions and buffers were prepared with using distilled water that had been passed through a Millipore-Q ultrapurification system. TLC analysis was performed using precoated plates. Melting points were determined using an X-4 apparatus and are not corrected. NMR spectra were measured on a Varian Mercury 400 instrument, operating at 400 MHz for ¹H NMR and 100 MHz for ¹³C NMR. IR spectra were recorded on a Perkin-Elmer Spectrum BX FT-IR spectrophotometer as KBr pellets and were reported in cm⁻¹. Electrospray mass spectra (ESI-MS) were acquired on Agilent 1100 Series LC/MS ion trap mass spectrometers and 6530 Accurate-Mass QTOF spectrometer coupled to an Agilent HPLC 1200 series. UV-vis spectra and fluorescent spectra were recorded on an Agilent Cary 100 UV-vis spectrophotometer and an Agilent Cary Eclipse fluorescence spectrophotometer, respectively. Both spectrophotometers are equipped with a temperature controller. Standard quartz cuvettes with a 10 mm lightpath were used for all optical measurements. Cell imaging was performed in an Olypus IX71 inverted fluorescence microscopy with a 20× objective lens.

Optical measurements: Stock solutions of probe **1** (1 mM) were prepared in DMSO (HPLC grade). Stock solutions (0.01-10 M) of the analytes were prepared in ultrapure water. For optical measurements, a solution of probe **1** (10 μ M or as stated) was prepared in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v). Then 3.0 mL of the probe **1** solution was placed in a quartz cell until the temperature reached at 37 °C over a few minutes. The UV-vis or fluorescent spectra were then recorded upon addition of various analytes.

Determination of the fluorescence quantum yield: In our system, the fluorescence quantum yields of probe 1 ($\Phi = 0.09$) and compound 2 ($\Phi = 0.56$) were determined in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v), using rhodamine B ($\Phi_f = 0.89$ in ethanol) as standard. The quantum yield was calculated using the following equation:

$$\Phi_{\rm x} = \Phi_{\rm s}(A_{\rm s}F_{\rm x} / A_{\rm x}F_{\rm s}) (n_{\rm x}^2/n_{\rm s}^2)$$

where, A_x and A_s are the absorbance of the sample and the reference, respectively, at

the same excitation wavelength, F_x and F_s are the corresponding relative integrated fluorescence intensities, and n is the refractive index of the solvent. Absorbance of sample and reference at their respective excitation wavelengths was controlled to be lower than 0.05.

Cell imaging experiments: HeLa cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% FBS (Fetal Bovine Serum), 100mg/mL penicillin and 100 μ g/mL streptomycin in a 5% CO₂, water saturated incubator at 37 °C, and then were seeded in a 12-well culture plate for one night before cell imaging experiments. For living cells imaging experiment of probe 1, cells were incubated with 1 μ M (or 10 μ M) of probe 1 (with 0.2% DMSO, v/v) for 60 min at 37 °C and washed three times with prewarmed PBS, and then imaged immediately. For N-ethylmaleimide (NEM) treated experiments, HeLa cells were pretreated with 0.5 mM NEM for 60 min at 37 °C, washed three times with prewarmed PBS, and then incubated with 10 μ M probe 1 for 60 min at 37 °C. Cell imaging was then carried out after washing cells with prewarmed PBS buffer.

2. Synthesis of compound 2 and probe 1



Scheme S1. Synthesis of probe 1.

Synthesis of 3-benzothiazolyl-7-hydroxycoumarin (2). Compound **2** was synthesized in 75 % yield according to a previously reported method (Ref. W. Lin, L. Long and W. Tan, *Chem. Commun.*, 2010, **46**, 1503–1505). M.p. 304-306 °C. TLC (silica plate): R_f 0.3 (petroleum ether : ethyl acetate 2:1, v/v); ¹H NMR (400 MHz, d_6 -DMSO, Me₄Si): δ (ppm) 9.14 (d, J = 1.7 Hz, 1H), 8.15 (d, J = 7.9 Hz, 1H), 8.04 (d, J = 8.2 Hz, 1H), 7.92 – 7.89 (m, 1H), 7.55 (t, J = 7.4 Hz, 1H), 7.45 (t, J = 7.1 Hz, 1H), 6.95 (d, J = 8.6 Hz, 1H), 6.90 (s, 1H). ¹³C NMR (100 MHz, d_6 -DMSO, Me₄Si): δ (ppm) 188.3, 164.2, 160.9, 160.2, 156.2, 152.2, 143.0, 136.0, 132.3, 126.9, 125.4, 122.5, 115.0, 114.7, 111.8, 102.5. IR (KBr) v_{max} (cm⁻¹): 2965, 2737, 1721 (s), 1707, 1615, 1599, 1585, 1558, 1525, 1452 (s), 1344, 1298, 1257 (s), 858, 755, 508. EI-MS: m/z found 295.15 (M⁺, 100%), 267.14 (M⁺ - C₂H₄, 35%).

Synthesis of probe 1. To a solution of compound 2 (147 mg, 0.5 mmol) in dry dichloromethane (5 mL) was added acryloyl chloride (90 μ L) and Et₃N (150 μ L). The resulting mixture was stirred at room temperature until the reaction was complete (monitored by TLC in a silica plate, the Rf of the starting material 2 is 0.3 using petroleum ether : ethyl acetate 1:1 (v/v) as mobile phase. Water (10 mL) was used to wash the resulting solution three times, and the dichloromethane phase was dried over Na₂SO₄. After filtered and removal of the organic solvent, a yellow solid product was formed, which can be further purified by recrystallization from ethanol to afford the pure product (140 mg, 80%). M.p. 227-229 °C; TLC (silica plate): Rf 0.7 (petroleum ether : ethyl acetate 2:1, v/v); ¹H NMR (400 MHz, CDCl₃, Me₄Si) : δ (ppm) 9.06 (s, 1H), 8.08 (d, J = 8.2 Hz, 1H), 7.98 (d, J = 7.9 Hz, 1H), 7.75 (d, J = 8.5 Hz, 1H), 7.54 (t, J = 7.6 Hz, 1H), 7.43 (t, J = 7.5 Hz, 1H), 7.30 (s, 1H), 7.21 (dd, J = 8.4, 1.6 Hz, 1.6 Hz)1H), 6.68 (d, J = 17.3 Hz, 1H), 6.36 (dd, J = 17.2, 10.4 Hz, 1H), 6.12 (d, J = 10.4 Hz, 1H). ¹³C NMR (100 MHz, *d*₆-DMSO) δ 182.1, 164.2, 160.3, 159.8, 154.6, 154.5, 152.5, 142.2, 136.5, 135.1, 131.9, 127.8, 127.3, 126.1, 123.1, 120.0, 119.5, 117.5, 110.7, 55.49. IR (KBr) v_{max} (cm⁻¹): 3440 (br s), 2978, 2942, 2739, 2677, 2604, 2494, 1732 (s), 1610, 1477, 1398, 1156, 1037, 767; EI-MS: m/z found 349.17 (M⁺, 40%), 295.19 (M⁺ - C₂H₃CO, 100%). HR-MS Calc. for C₁₉H₁₂NO₄S⁺ (M + H⁺) 350.04815, found 350.04805.



¹H-NMR spectrum of compound **2** in d_6 -DMSO



IR spectrum of compound 2





¹H-NMR spectrum of probe 1 in CDCl₃









MS (EI) spectrum of probe 1



IR spectrum of probe 1

3. Additional studies



Fig. S1 Fluorescent kinetics of probe **1** (10 μ M) in the absence and presence of 50 μ M of Cys in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v) at 37 °C. The spectra were monitored at 498 nm and collected at10 s intervals, respectively, with $\lambda_{ex} = 458$ nm; $d_{ex} = d_{em} = 2.5$ nm.

Sensing mechanism of probe 1 for Cys



Scheme S2. A proposed mechanism for sensing of Cys by probe 1.

To a 150 mL flask, probe **1** (0.14 g, 0.4 mmol) and Cys (1.25 eq) were combined in 100 mL of MeOH : H₂O (90 : 10, v/v) solution, and the mixture stirred at room temperature for 1 h. Then, Et₃N (30 μ L) was added and the solution stirred ca. 2 h. The solvents was removed under reduced pressure and the crude product was subjected to column chromatography (eluted with CH₂Cl₂: MeOH, 5 : 1, v/v) to afford (58 mg, 49.2%) of product **A** and 18 mg of product **B** as an off-white solid. Product **A** was checked by TLC, ¹H NMR and Mass spectra, which was proved to be **2** by comparison with those of reference sample of compound **2**. Product **B** was proved to

be **3** by its ¹H NMR spectrum with comparison to that of previously reported spectrum of **3**. See below.



Fig. S2 TLC analysis of the isolated product **A** from the reaction of probe **1** and Cys. plate under different light used to compare probe **1**, the reference sample of compound **2** and the isolated reaction product of probe **1** and Cys. (A) under room light, (B) under light of 254 nm, (C) under light of 365 nm. Spots on the TLC plate are: a. probe **1**, b. the reaction product **A**, c. mixture of product **A** and the reference sample of compound **2**, d. the reference sample of compound **2**. The eluent for TLC: petroleum ether : ethyl acetate = 2 : 1 (v/v).



Fig. S3 ¹H NMR spectrum of product **A** in DMSO- d_6 , which is identical to that of the reference sample of compound **2** (see above).



Fig. S4 Mass spectrum of product A, which shows the right mass of compound 2



Fig. S5 ¹H-NMR spectrum of compound **B** in D_2O , which is identical to the previously reported spectrum of **3**, see Ref: X. Yang, Y. Guo and R. Strongin, *Angew. Chem., Int. Ed.*, 2011, **50**, 10690–10693, Figure S28 in the ESI of this paper.



Fig. S6 (a) UV-vis spectra changes of probe **1** (10 μ M) upon addition of various analytes (100 μ M, Cys, Hcy, and GSH were used 50 μ M). (b) Absorption changes of probe **1** (10 μ M) at 460 nm for Cys (50 μ M) in the presence of various analytes (100 μ M). Black bars represent the addition of a single analyte. Red bars represent the subsequent addition of Cys to the mixture. Analytes 1-31: 1. none, 2. F⁻, 3. Cl⁻, 4. Br⁻, 5. I⁻ 6. NO₃⁻, 7. NO₂⁻, 8. AcO⁻, 9. SCN⁻, 10. CO₃²⁻, 11. SO₄²⁻, 12. CN⁻, 13. SO₃²⁻, 14. S₂O₃²⁻, 15. Ala, 16. Glu, 17. Thr , 18. Trp, 19. Phe, 20. Gln, 21. Gly, 22. Lys, 23. Arg, 24. Ile, 25. Asp, 26. Leu, 27. Ser, 28. Met, 29. His, 30. GSH, 31. Hcy. Each spectrum was collected after 15 min of mixing each analyte with probe **1** in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v) at 37 °C.



Fig. S7 Color changes and fluorescence changes (under a 365 nm UV lamp) of probe **1** (10 μ M) upon addition of various analytes (from left to right: none, Ala, Glu, Arg, Asp, Gln, Gly, His, Leu, Lys, Met, Phe, Trp, Ser, Ile, Thr, Cys, Hcy, and GSH. Except Hcy, GSH and Cys were used 50 μ M, others were use 100 μ M.).



Fig. S8 Fluorescent kinetics of probe **1** (10 μ M) with 50 μ M of Cys, Hcy and GSH in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v) at 37 °C. The reactions are monitored at 498 nm with $\lambda_{ex} = 458$ nm, slit: $d_{ex} = d_{em} = 2.5$ nm.



Fig. S9 (a) UV/Vis absorption spectra of probe 1 (10 μ M) upon addition of different concentrations of Cys in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v) at 37 °C. Final concentration of Cys: 0, 1, 5, 10, 15, 20, 25, 30, 35 40, 45 and 50 μ M, respectively. (b) The changes of the absorbance intensity at 460 nm of probe 1 (10 μ M) against concentration of Cys. Each spectrum was obtained 15 min after Cys addition.



Fig. S10 (a) Fluorescent spectra changes of probe 1 (10 μ M) upon addition of different concentrations of Cys in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v) at 37 °C. Final concentration of Cys: 0, 1, 5, 10, 15, 20, 25, 30, 35, 40, 45 and 50 μ M, respectively. (b) Fluorescent intensity changes of probe 1 (10 μ M) at 498 nm against concentration of Cys. Each spectrum was obtained 15 min after Cys addition.



Fig. S11 (a) Fluorescent spectra changes of probe **1** (1 μ M) upon addition of 5 μ M Cys after 10 min. (b) Fluorescence kinetics of probe **1** (1 μ M) monitored at 498 nm upon addition of Cys (1 μ M and 5 μ M) in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v) at 37 °C. The kinetics data are fitted (solid line) by a first-order reaction scheme as shown in the figure, and the observed pseudo-first-order rate constant k_{obs} was also shown. $\lambda_{ex} = 458$ nm, $d_{ex} = 2.5$ nm, $d_{em} = 5$ nm.



Fig. S12 Detection of Cys using very low concentrations of probe **1** in DMSO-PBS buffer (20 mM, pH 7.4, 1:1, v/v) at 37 °C. $\lambda_{ex} = 458$ nm, slit for (a) and (b): $d_{ex} = 5$ nm, $d_{em} = 10$ nm, for (c) and (d): $d_{ex} = d_{em} = 10$ nm, respectively. Insert in (d): fluorescent intensity changes at 498 nm as a function of time.



Fig. S13 The effect of pH on the fluorescence intensity changes of probe 1 (1 μ M) at 498 nm in absence and presence of Cys (5 μ M) in DMSO-PBS buffer (20 mM, 1:1, v/v) at 37 °C. All the data was obtained 15 min after mixing. $\lambda_{ex} = 458$ nm, $d_{ex} = 2.5$ nm, $d_{em} = 5$ nm. Green color area indicates the best working pH range for probe 1.