

## Supporting Information

### Regulated Assemblies and Anion Responsive Vesicles Based on 1,3-Alternate Oxacalix[2]arene[2]triazene Amphiphiles

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## 1. General information

<sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on 300 MHz NMR spectrometer. Chemical shifts are reported in ppm versus tetramethylsilane or the residual solvent resonance used as an internal standard. Melting points are uncorrected. All chemicals, except if stated otherwise, were obtained from commercial sources and used without further purification. Elemental analyses, mass spectrometry, SEM, TEM, DLS, LSCM and X-ray diffraction analysis were performed at the Institute of Chemistry. The fluorescence spectra were measured with a Hitachi F-4500 fluorescence spectrophotometer. Synthesis of macrocyclic compound **1** and **4**, intermediates **2c'**, **2d''** and **2d** followed the reported methods.<sup>1-4</sup>

## 2. Experimental details

**Preparation of vesicles or micelle:** A small volume (100 ul) of the stock solution of **3** (**3a-d**) in THF ( $5 \times 10^{-4}$  M) was transferred to a sample bottle. Double distilled water (1 ml) was injected quickly, giving a mixture solution of  $5 \times 10^{-5}$  M. The mixture solution was put in the ultrasonic bath for 30 minutes, incubated at 60 °C for 30 minutes to remove the organic solvent. Solutions of **3a** and **3b** hence yielded as opalescent, whereas **3c** and **3d** as clear dispersions.

**Laser scanning confocal microscopy (LSCM):** Laser scanning confocal microscopic experiments were carried out by preparing the vesicles in a solution containing lucigenin. The fluorescence agent outside the vesicles was then removed through repeated dialysis. The resulting vesicle solution were deposited on a glass surface, covered with a glass slide, and then visualized using a confocal laser scanning microscope (LSCM, Olympus FV1000, Olympus FV1000 with a 100×oil-immersion objective and a numerical aperture of 1.4. The excitation wavelength was set at 405nm).

**Dynamic light scattering (DLS):** The DLS measurements were carried out at 25°C using an LLS spectrometer (ALV/SP-125) with a multi- $\tau$  digital time correlator (ALV-5000). Light of  $\lambda=632.8\text{nm}$  from a solid-state He-Ne laser (22 mW) was used as the incident beam. The measurement was performed at a scattering angle of 90°. The correlation function was analyzed from the scattering data via the CONTIN method to obtain the distribution of diffusion coefficients (D) of the solutes. The apparent hydrodynamic radius  $R_h$  was deduced from D by the Stokes-Einstein equation  $R_h=k_B T/(6\pi\eta D)$  for spherical particles, where  $k_B$  is the Boltzmann constant, T is the Kelvin temperature, and  $\eta$  is viscosity of solvent. For the effect of salts on

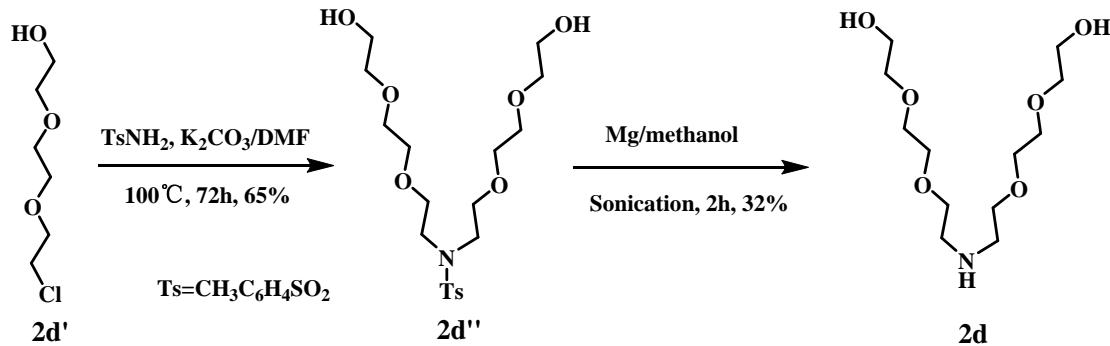
vesicle sizes, small volume of salt solutions (2.5 M) was added to the vesicle solutions (2 mL) assembled from **3a** or **3b**, the mixture was then balanced for 1 minute and the size distribution was recorded by DLS equipment.

**Zeta potential measurement:** The zeta potential measurements were conducted at 25 °C, using a Malvern Zetasizer Nano-ZS instrument (ZEN3600, Malvern Instruments, Worcestershire, UK) equipped with a 4 mW He–Ne laser at a wavelength of 633 nm. A clear disposable capillary cell (DTS1060C) was used. All the measurements were performed at  $\theta=173^\circ$ . The zeta potential was calculated using the Helmholtz-Smoluchowski relationship from the mobility measured in an electrophoretic light-scattering (ELS) experiment.

**X-ray diffraction (XRD) analysis:** Samples for XRD measurement were prepared through directly casting vesicle solution on the glass slide for 4 times and the solvent was evaporated at room temperature. XRD analysis was performed on an X-ray diffractometer (SW, X'PERT) with nickel filtered Cu K $\alpha$  radiation ( $\lambda=1.54060\text{ \AA}$ ) operated at 40 kV and 10 mA.

### 3. Synthesis

#### Synthesis of **2d**



**Scheme S1.** Synthesis of **2d**.

1.32 g (3.03 mmol) of **2d''** was dissolved in methanol (50 ml) and magnesium powders (0.42 g, 30 mmol) were added. The resulting suspension was sonicated for 2 h when TLC on silica (ethyl acetate–methanol, 6 : 1) indicated that all starting material had been consumed. After filtration, the filtrate was evaporated under reduced pressure to afford oily residue, which was further purified by column chromatography using silica gel with acetone/methanol (v/v, 20/1) as eluent to give **2d** as oil (0.27 g, 32%):  $^1\text{H}$  NMR (300 MHz,  $\text{CD}_3\text{Cl}$ , ppm):  $\delta$  4.46 (br. s, 3H), 3.57–3.72 (m, 20H), 2.89 (t,  $J=5.3\text{ Hz}$ , 4H); MS (ESI):  $m/z$  282.16 [ $\text{M}+\text{H}]^+$ , 304.06 [ $\text{M}+\text{Na}]^+$ .

### General procedures for the synthesis of **3a**, **3b**, **3c'** and **3d**

A mixture of **1** (0.46 g, 0.5 mmol for **3a**, 0.30 g, 0.3 mol for **3b**, 0.13 g, 0.14 mmol for **3c'**, 0.18 g, 0.2 mmol for **3d**), **2** (**2a**: 0.16 g, 1.5 mmol, **2b**: 0.10 g, 1.0 mmol, **2c'**: 0.13 g, 0.4 mmol, **2d**: 0.23 g, 0.8 mmol) and DIPEA (0.52 g, 4.0 mmol for **3a**, 0.34 g, 2.6 mmol for **3b**, 0.14 g, 1.1 mmol for **3c'**, 0.21 g, 1.6 mmol for **3d**) in THF (30 ml) was stirring at room temperature for 5 h. After removal of the solvent, the residue was subjected to silica gel chromatography to get pure products of **3** as white solids (0.42 g, 81% for **3a**, 0.32 g, 92% for **3b**, 0.22 g, 92% for **3c'** and 0.12 g, 46% for **3d**).

**3a**: mp 159~160°C; IR (KBr) v 3335, 3081, 2923, 2853, 1645, 1603, 1583, 1525, 1392, 1304, 1172; <sup>1</sup>H NMR (300 MHz, DMSO-d<sub>6</sub>, ppm): δ 8.36 (t, J=5.4Hz, 2H), 7.34 (d, J=2.0Hz, 4H), 7.20 (t, J=2.0Hz, 2H), 4.85 (t, J=5.3Hz, 4H), 3.70~3.72 (m, 8H), 3.63~3.66 (m, 8H), 3.13~3.19 (m, 4H), 1.41~1.46 (m, 4H), 1.20~1.24 (m, 44H), 0.84 (t, J=6.5Hz, 6H). <sup>13</sup>C NMR (300 MHz, DMSO-d<sub>6</sub>, ppm): δ 170.92, 167.47, 163.81, 151.78, 136.53, 119.29, 117.02, 58.32, 50.52, 31.29, 29.05, 29.02, 28.96, 28.84, 28.71, 26.45, 22.09, 13.92. MS (MALDI-TOF): m/z 1081.3 [M+Na]<sup>+</sup>. Calcd for C<sub>56</sub>H<sub>86</sub>N<sub>10</sub>O<sub>10</sub>: C, 63.49; H, 8.18; N, 13.22. Found: C, 63.55; H, 8.37; N, 13.19.

**3b**: mp 86~87°C; IR (KBr) v 3078, 2964, 2934, 2874, 1589, 1523, 1391, 1265, 1201, 1141; <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>Cl, ppm): δ 7.13 (d, J=2.1Hz, 4H), 6.78 (t, J=2.1Hz, 2H), 6.06 (t, J=5.6Hz, 2H), 3.57 (t, J=7.7Hz, 8H), 3.34 (q, J=6.8Hz, 4H), 1.64~1.74 (m, 8H), 1.52~1.59 (m, 4H), 1.23~1.32 (m, 44H), 0.96 (t, J=7.35Hz, 12H), 0.88 (J=6.6Hz, 6H). <sup>13</sup>C NMR (300 MHz, CD<sub>3</sub>Cl, ppm): δ 171.88, 167.83, 166.22, 152.58, 137.36, 120.03, 118.01, 49.47, 40.43, 32.07, 29.81, 29.78, 29.73, 29.66, 29.50, 27.20, 22.83, 20.97, 14.25, 11.44. MS (MALDI-TOF): m/z 1073.7 [M+Na]<sup>+</sup>. Calcd for C<sub>60</sub>H<sub>94</sub>N<sub>10</sub>O<sub>6</sub>: C, 68.54; H, 9.01; N, 13.32. Found: C, 68.64; H, 9.06; N, 13.32.

**3c'**: mp 116~117°C; IR (KBr) v 3305, 3066, 3034, 2924, 2853, 1750, 1645, 1579, 1538, 1386, 1300, 1168, 1030; <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>Cl, ppm): δ 7.32-7.37 (m, 20H), 7.13 (d, J=2.0 Hz, 4H), 6.71 (t, J=2.0 Hz, 2H), 6.08 (t, J=2.3 Hz, 2H), 5.22 (s, 8H), 4.58 (s, 8H), 3.36 (q, J=5.0 Hz, 4H), 1.59 (m, 4H), 1.21-1.32 (m, 44H), 0.88 (t, J=6.9 Hz, 6H). <sup>13</sup>C NMR (300 MHz, CD<sub>3</sub>Cl, ppm): δ 172.06, 169.18, 168.98, 165.95, 152.37, 137.76, 135.34, 128.80, 128.64, 128.42, 119.67, 118.20, 67.37, 49.65, 40.48, 32.07, 29.84, 29.80, 29.73, 29.64, 29.50, 27.20, 22.83, 14.26. MS (MALDI-TOF): m/z 1497.5 [M+Na]<sup>+</sup>. Calcd for C<sub>84</sub>H<sub>102</sub>N<sub>10</sub>O<sub>14</sub>: C, 68.36; H, 6.97; N, 9.49.

**3d**: mp 72~73°C; IR (KBr) v 3342, 3079, 2924, 2853, 1644, 1581, 1525, 1386, 1305, 1138, 1078,

811;  $^1\text{H}$  NMR (300 MHz,  $\text{CD}_3\text{Cl}$ , ppm):  $\delta$  7.16 (d,  $J=2.1\text{Hz}$ , 4H), 6.76 (t,  $J=2.1\text{Hz}$ , 2H), 6.37 (s, 2H), 3.96 (t,  $J=4.7\text{Hz}$ , 8H), 3.80 (t,  $J=5.0\text{Hz}$ , 8H), 3.67-3.71 (m, 24H), 3.59 (t,  $J=4.7\text{Hz}$ , 8H) 0-3.37 (m, 8H), 1.21-1.57 (m, 4H), 1.22-1.32 (m, 44H), 0.88 (t,  $J=6.6\text{Hz}$ , 6H).  $^{13}\text{C}$  NMR (300 MHz,  $\text{CD}_3\text{Cl}$ , ppm):  $\delta$  171.69, 168.07, 166.23, 152.18, 137.30, 119.67, 118.11, 72.71, 70.47, 69.11, 61.71, 48.32, 40.36, 31.99, 29.77, 29.73, 29.69, 29.49, 29.43, 27.13, 22.76, 14.21. MS (MALDI-TOF): m/z 1434.0 [ $\text{M}+\text{Na}$ ] $^+$ . Calcd for  $\text{C}_{72}\text{H}_{118}\text{N}_{10}\text{O}_{18}$ : C, 61.25; H, 8.42; N, 9.92. Found: C, 60.95; H, 8.42; N, 9.80.

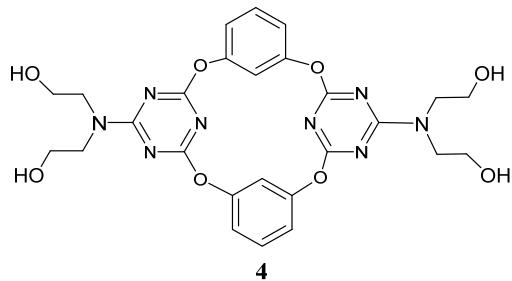
#### Procedures for synthesis of 3c

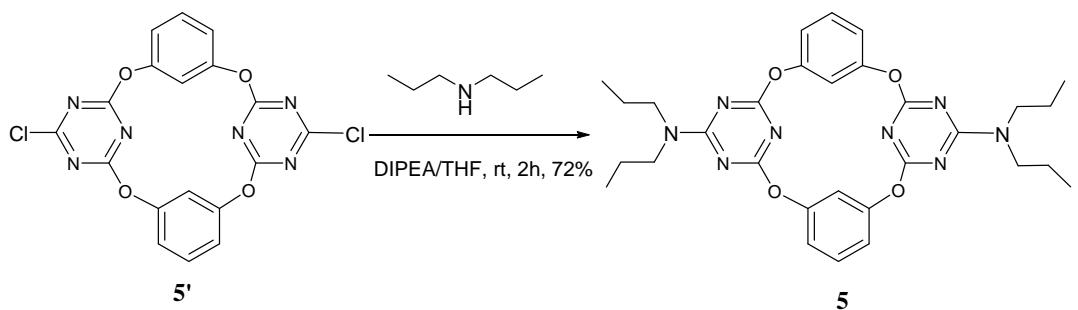
0.22 g of **3c'** was dissolved in THF, to which Pd/C (10%, 0.04 g) was added. The resulting mixture was stirred at room temperature overnight under hydrogen atmosphere. The resulting solution was filtrated through celite. After removal of organic solvent, the residues were recrystallized with a mixture of chloroform and acetonitrile to afford **3c** as white solid (0.15 g, 88%).

**3c:** mp 212~213°C; IR (KBr) v 3325, 3084, 2924, 2853, 2543, 1732, 1578, 1541, 1387, 1300, 1170;  $^1\text{H}$  NMR (300 MHz,  $\text{DMSO-d}_6$ , ppm):  $\delta$  12.88 (br, 4H), 8.32 (t,  $J=5.4\text{Hz}$ , 2H), 7.37 (d,  $J=2.1\text{Hz}$ , 4H), 7.27 (t,  $J=2.1\text{Hz}$ , 2H), 4.37 (s, 8H), 3.12-3.18 (m, 4H), 1.42-1.46 (m, 4H), 1.20-1.25 (m, 44H), 0.84 (t,  $J=6.6\text{Hz}$ , 6H).  $^{13}\text{C}$  NMR (300 MHz,  $\text{DMSO-d}_6$ , ppm):  $\delta$  170.98, 170.25, 168.22, 163.61, 151.59, 136.56, 119.31, 117.29, 49.86, 31.27, 29.03, 28.99, 28.82, 28.74, 28.69, 26.47, 22.06, 13.85. MS (MALDI-TOF): m/z 1137.4 [ $\text{M}+\text{Na}$ ] $^+$ . HRMS (APCI): Calcd for  $\text{C}_{56}\text{H}_{78}\text{N}_{10}\text{O}_{14}$  [ $\text{M}+\text{H}$ ] $^+$ : 1115.5772. Found: 1115.5770.

#### Procedures for syntheses of compound 4 and 5

Compound **4** was prepared according to the reported methods.<sup>5</sup>



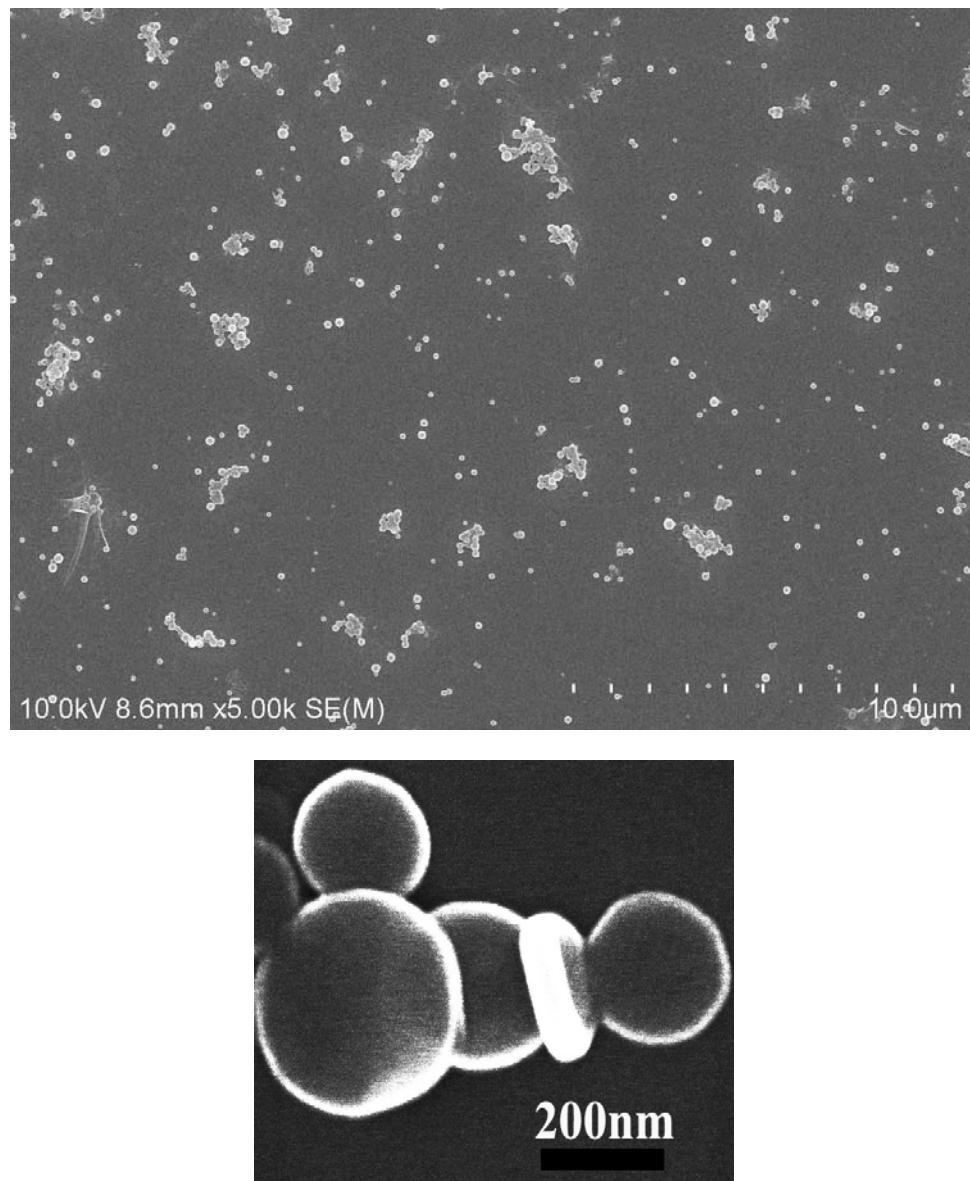


**Scheme S2.** Synthesis of compound **5**.

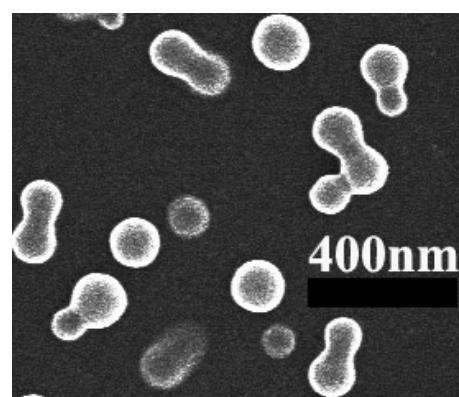
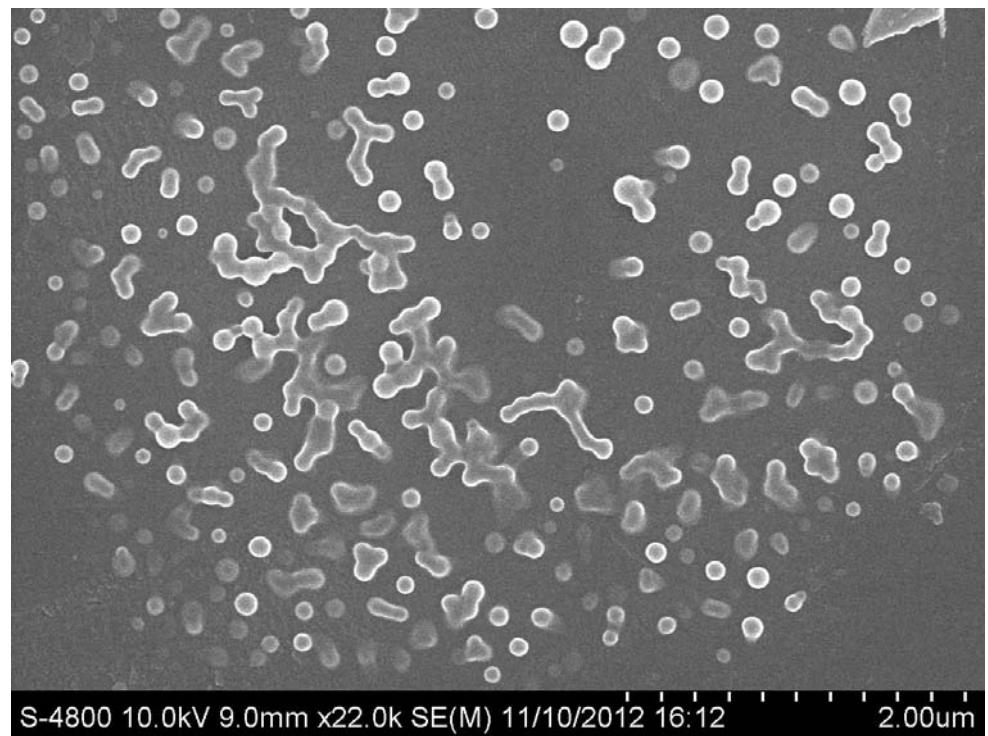
#### Synthesis of **5**

Tetraoxacalix[2]arene[2]triazene (**5'**) (1 mmol, 442 mg) was dissolved in THF (30 ml), to which n-dipropylamine (3 mmol, 303 mg) and DIPEA (8 mmol, 1.032 g) were added. The resulting mixture was stirring for 2 hours at room temperature. After removal of the organic solvent, the residue was subjected to silica gel chromatography with a mixture of dichloride methane and n-hexane as an eluent, giving pure **5** as white solids (0.411 g, 72%): mp 167-168°C; IR (KBr)  $\nu$  3084, 2964, 2934, 2874, 1589, 1523, 1464, 1391, 1322, 1266, 1201, 1141;  $^1\text{H}$  NMR (300 MHz,  $\text{CD}_3\text{Cl}$ , ppm):  $\delta$  7.14 (t,  $J=8.1$  Hz, 2H), 6.78 (dd,  $J=2.1$  Hz,  $J=8.1$  Hz, 4H), 6.67 (t,  $J=2.1$  Hz, 2H), 3.57 (t,  $J=7.7$  Hz, 8 H), 1.64-1.76 (m, 8H), 0.96 (t,  $J=7.4$  Hz, 12H);  $^{13}\text{C}$  NMR (300 MHz,  $\text{CD}_3\text{Cl}$ , ppm):  $\delta$  172.14, 167.88, 152.52, 129.47, 118.96, 117.42, 49.32, 20.98, 11.44; HRMS: m/z 573.2932 [ $\text{M}+\text{H}]^+$  (Calcd.: 573.2938); Anal. Calcd for  $\text{C}_{30}\text{H}_{36}\text{N}_8\text{O}_4$ : C, 62.92; H, 6.34; N, 19.57. Found: C, 63.08; H, 6.50; N, 19.13.

**4. SEM images of 3a and 3b**

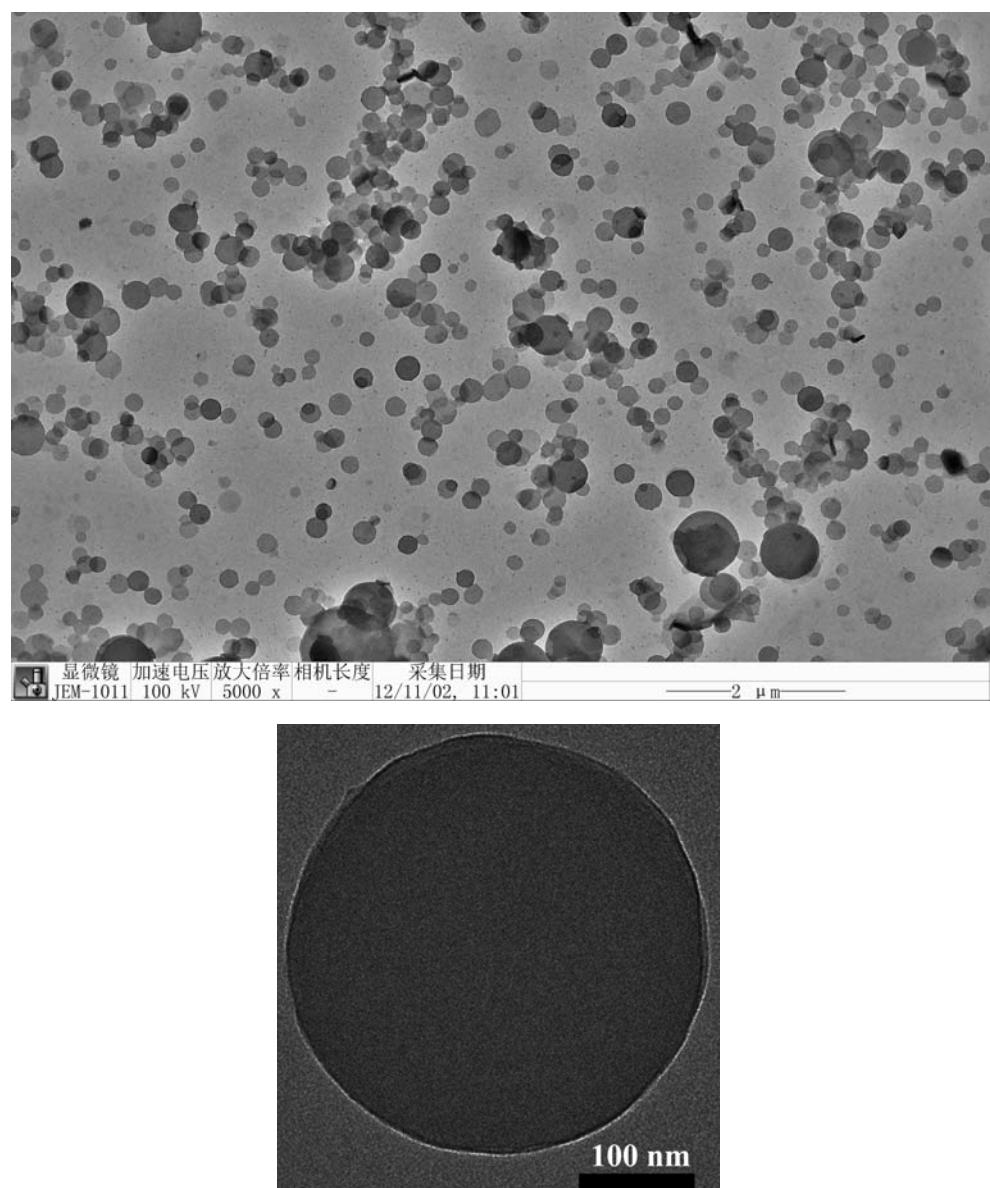


**Fig. S1** SEM images of vesicles formed with **3a** in water.

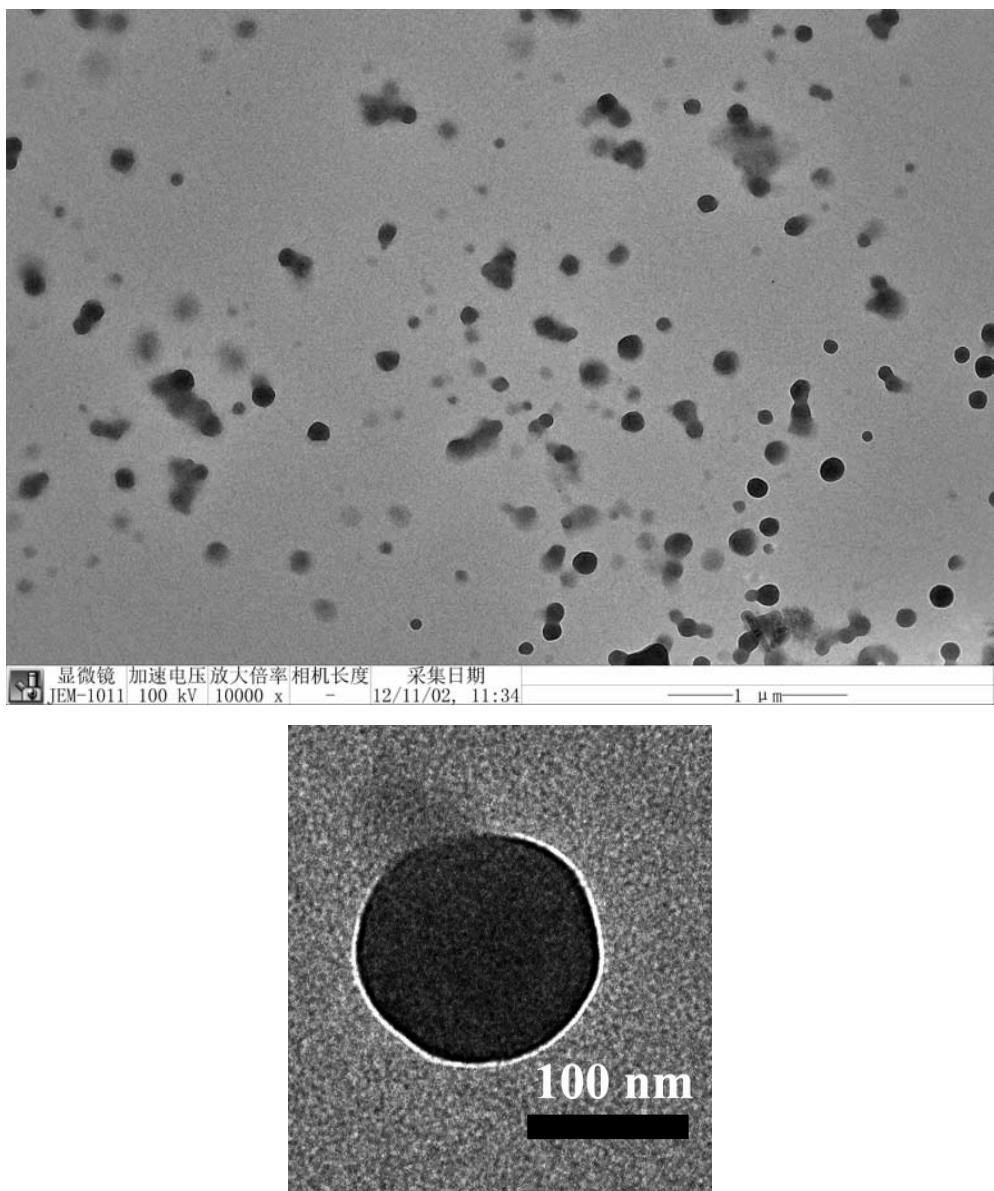


**Fig. S2** SEM images of vesicles formed with **3b** in water.

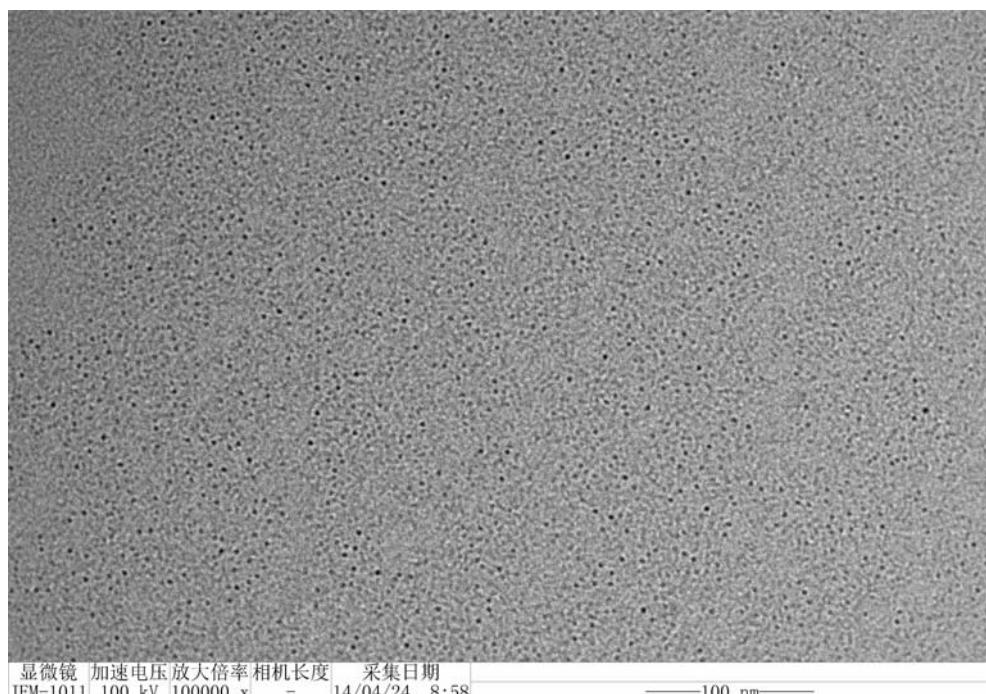
5. TEM images of 3a-d



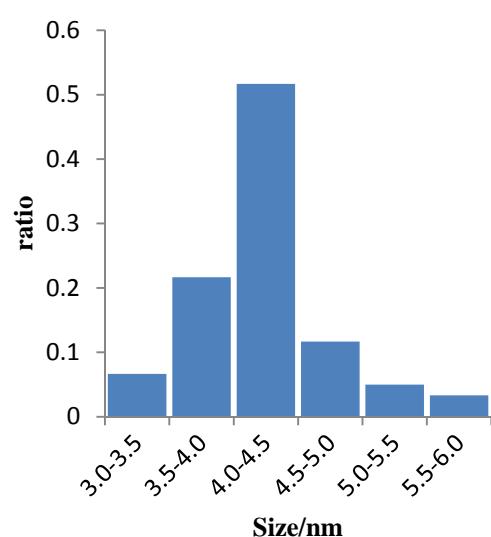
**Fig. S3** TEM images of assemblies formed with **3a** in water.



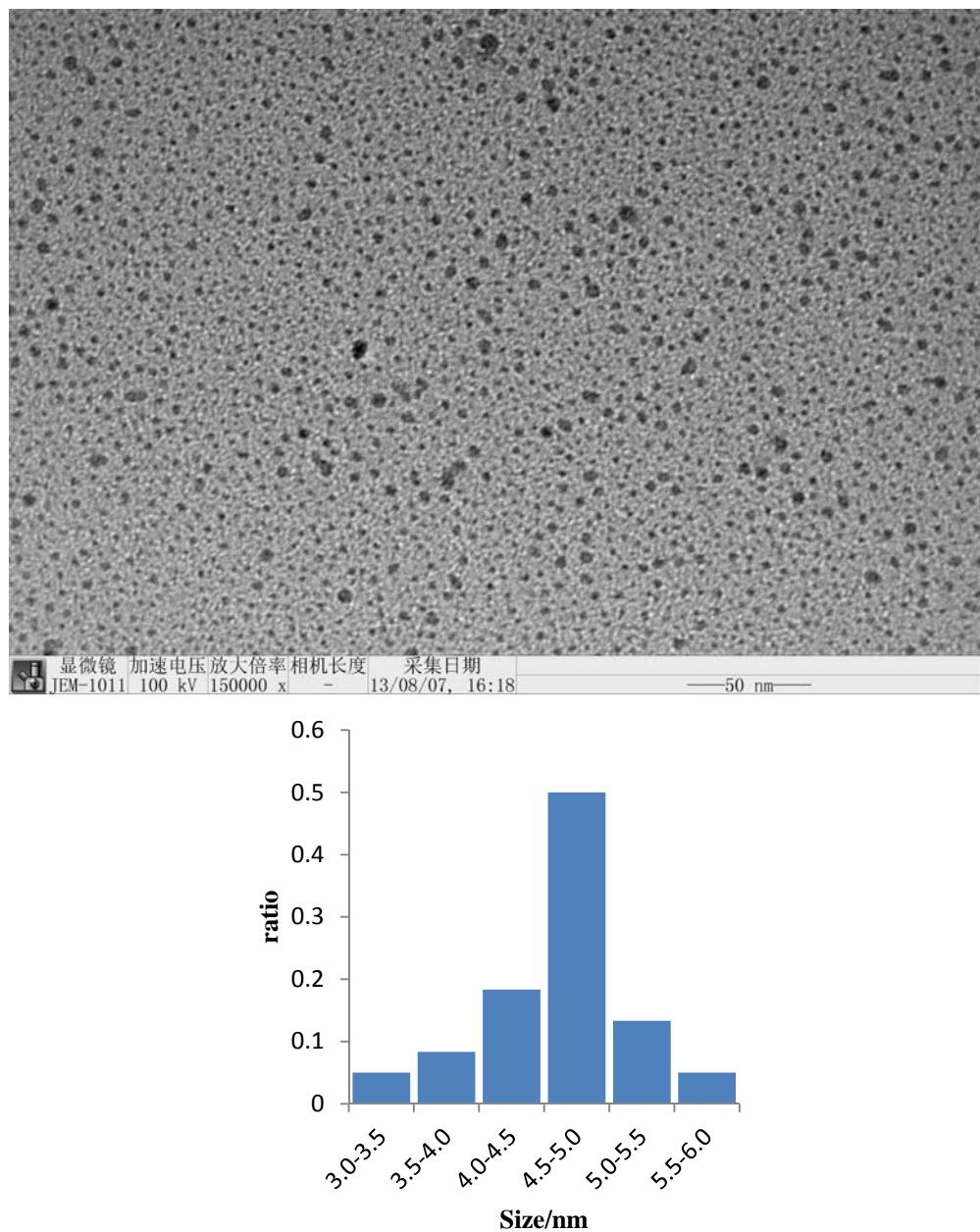
**Fig. S4** TEM images of assemblies formed with **3b** in water.



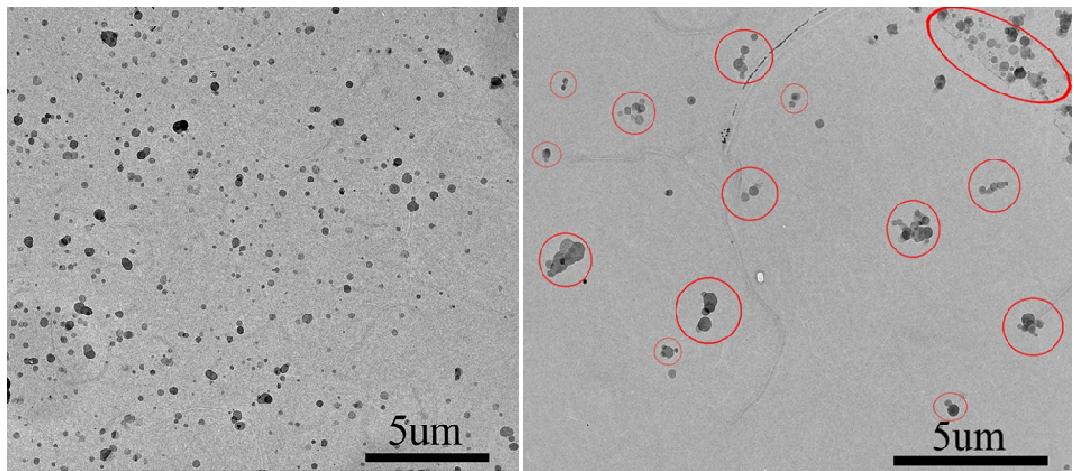
显微镜 加速电压 放大倍率 相机长度  
JEM-1011 100 kV 100000 x - 14/04/24, 8:58  
—100 nm—



**Fig. S5** TEM images of assemblies formed with **3c** in water (top) and the statistic size distribution from TEM images (bottom).

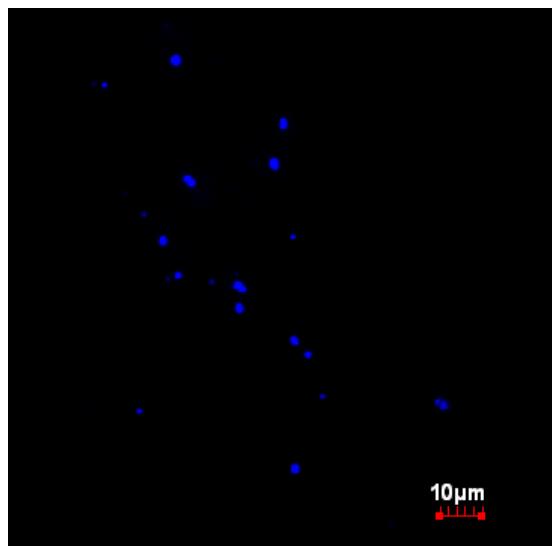


**Fig. S6** TEM images of assemblies formed with **3d** in water (top) and statistic size distribution from TEM images (bottom)

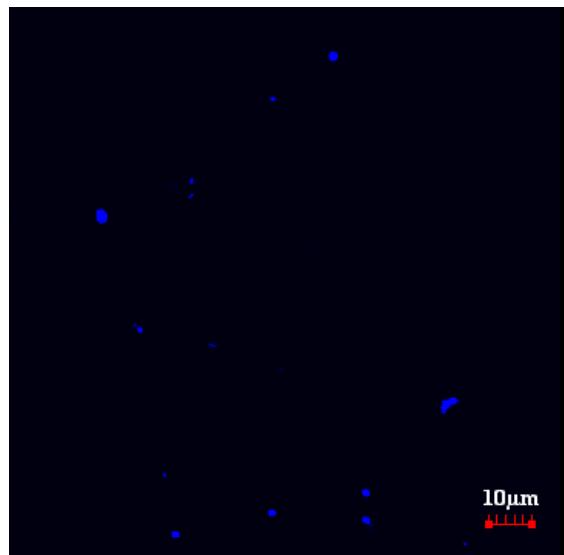


**Fig. S7** TEM images of vesicles of **3a** before (left) and after (right) treated with NaCl.

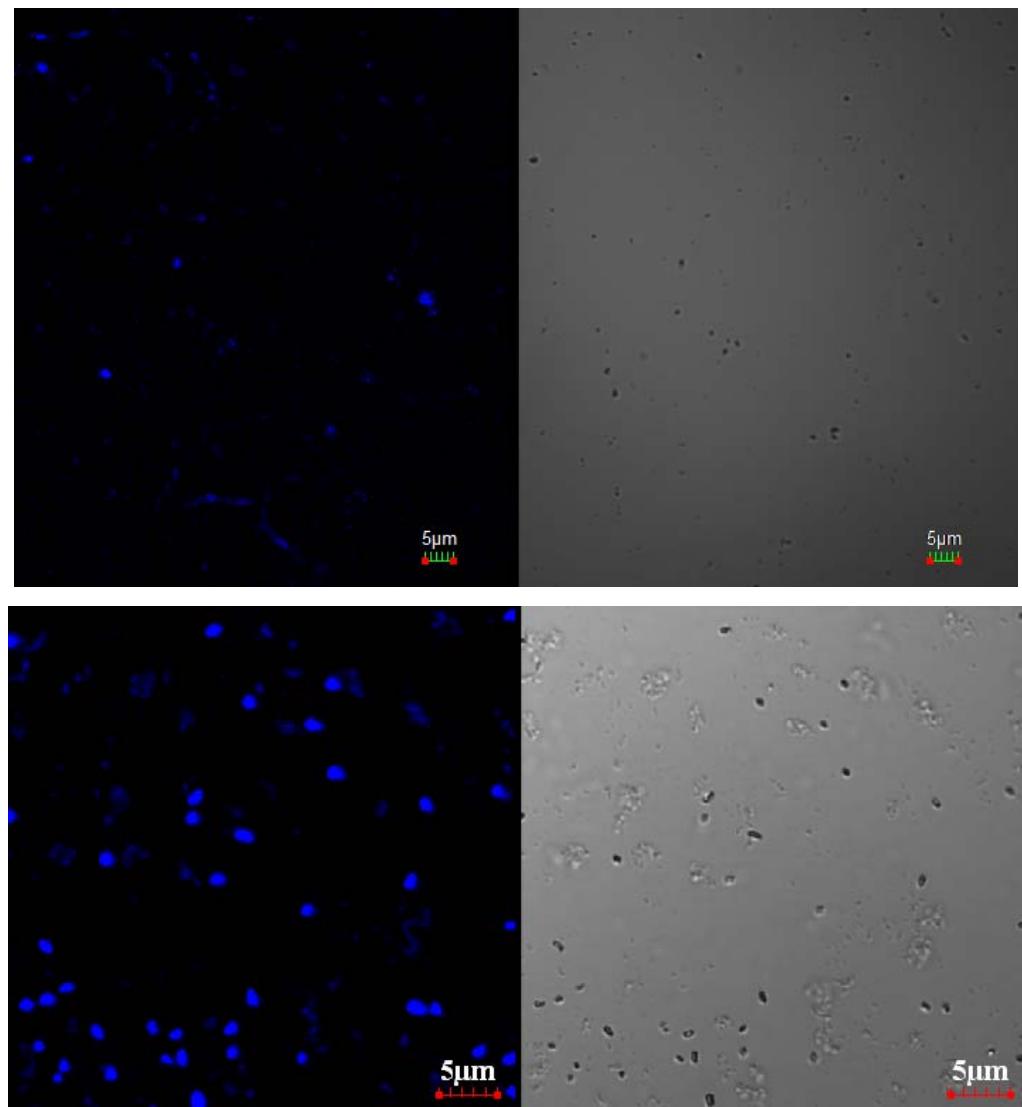
#### 6. LSCM images of **3a** and **3b**



**Fig. S8** LSCM images of vesicles formed with **3a**.

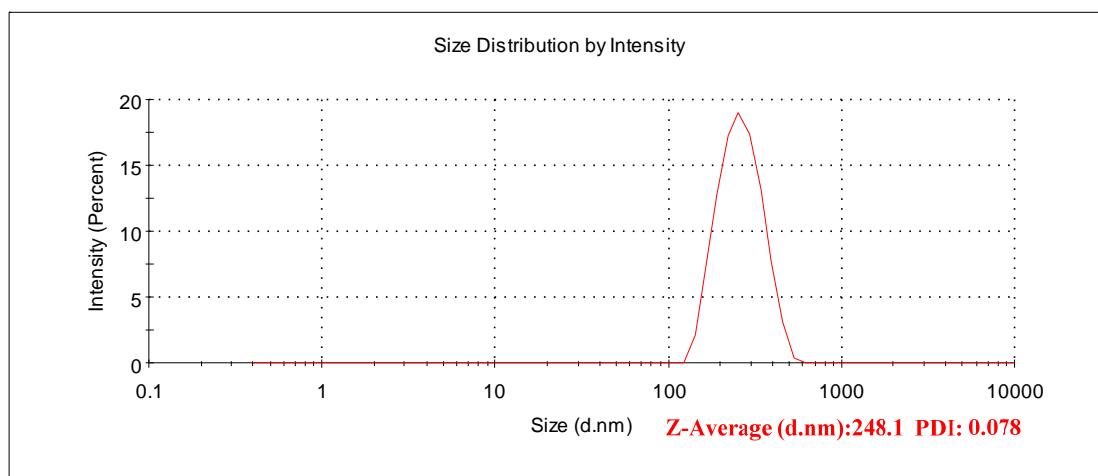


**Fig. S9** LSCM images of vesicles formed with **3b**.

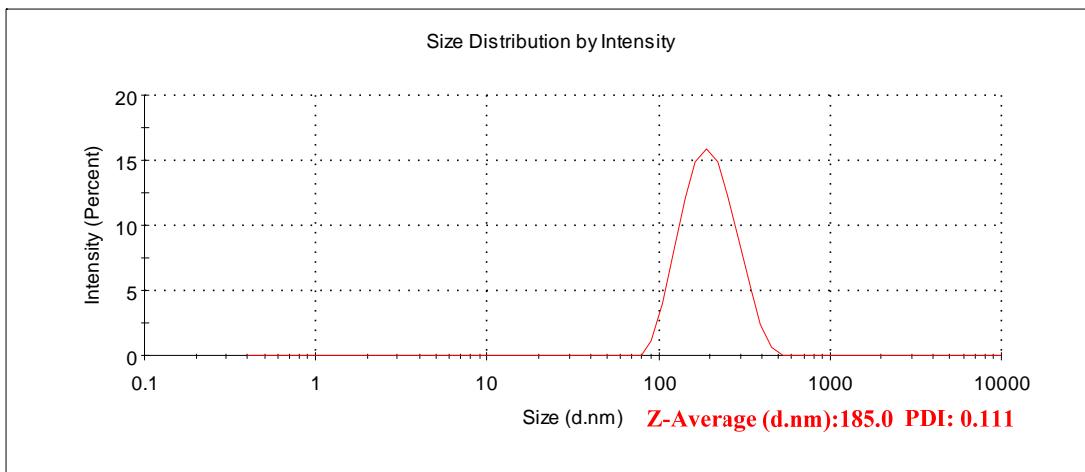


**Fig. S10** LSCM images of vesicles of **3a** before (top) and after (bottom) treatment with NaCl.

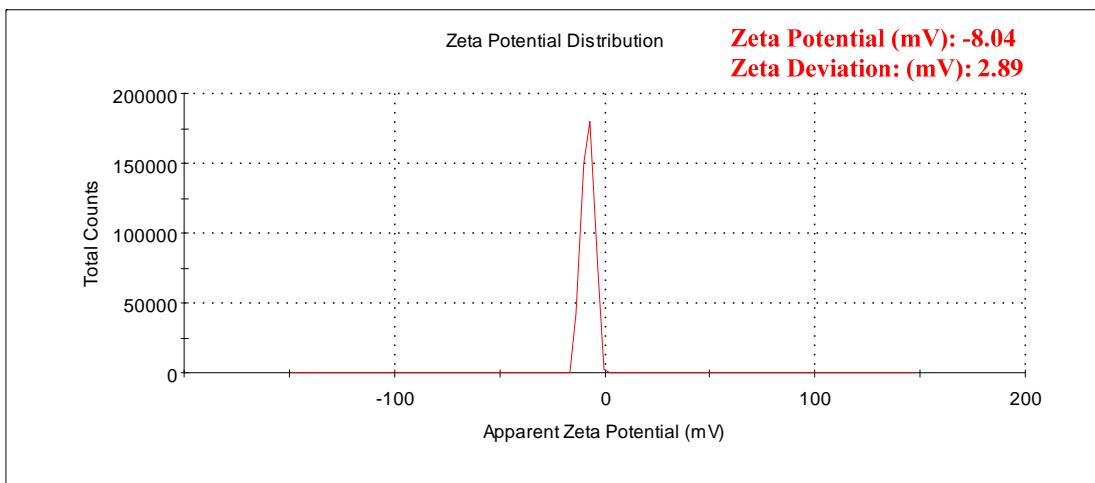
## 7. DLS results of 3a-d



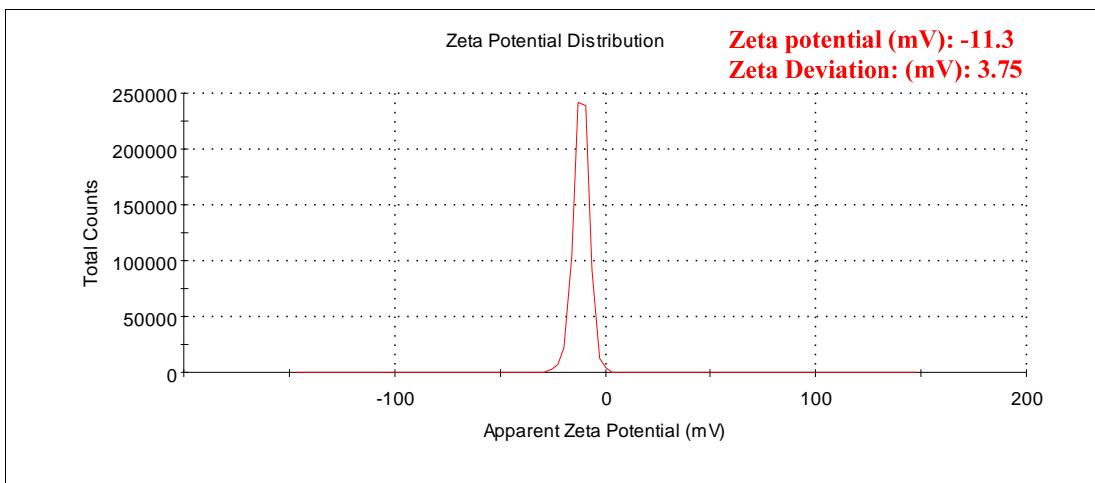
**Fig. S11** Size distribution of assemblies formed with **3a**.



**Fig. S12** Size distribution of assemblies formed with **3b**.

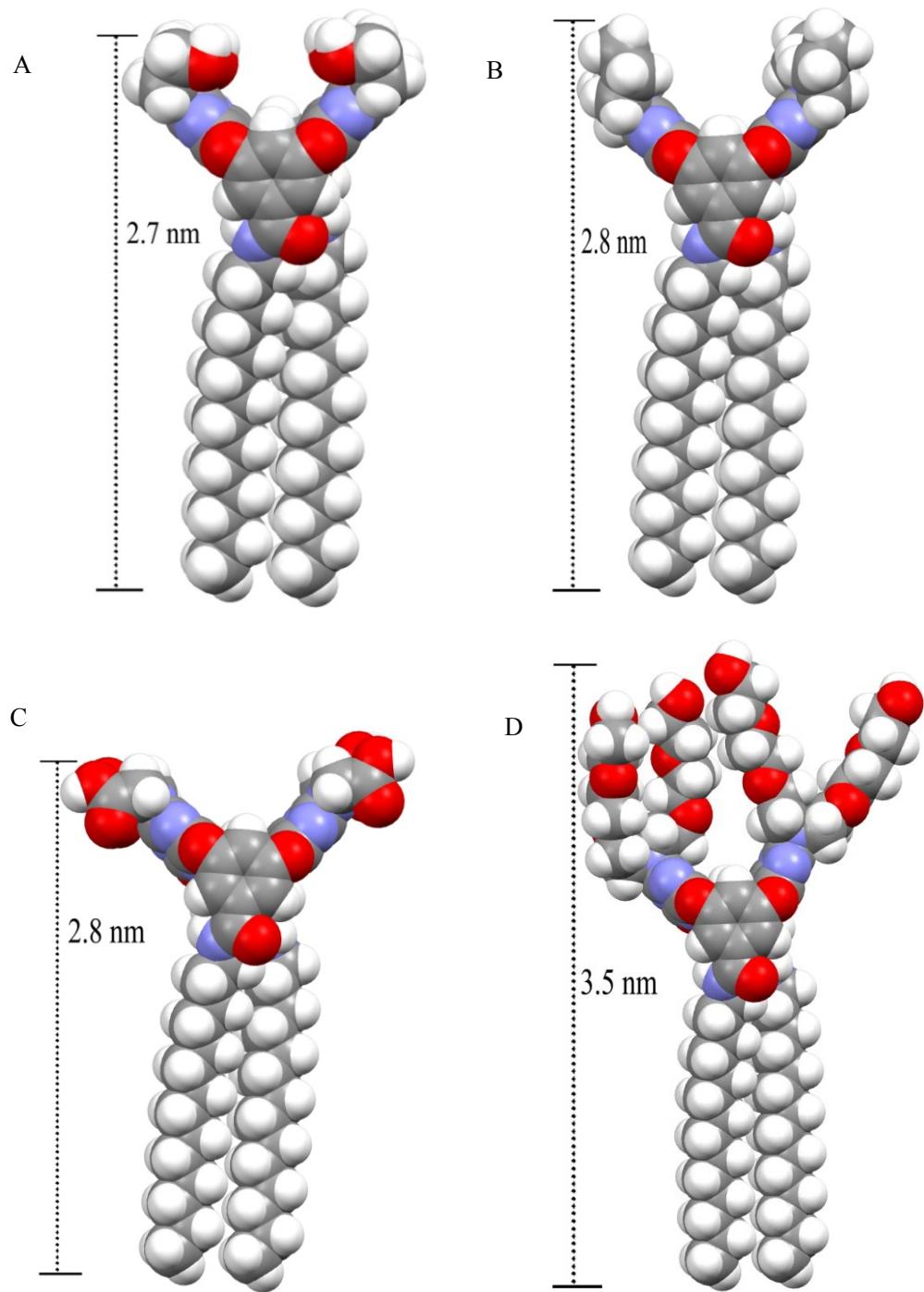


**Fig. S13** Zeta potential of vesicles formed with **3a** in the presence of NaCl (0.63 mM)



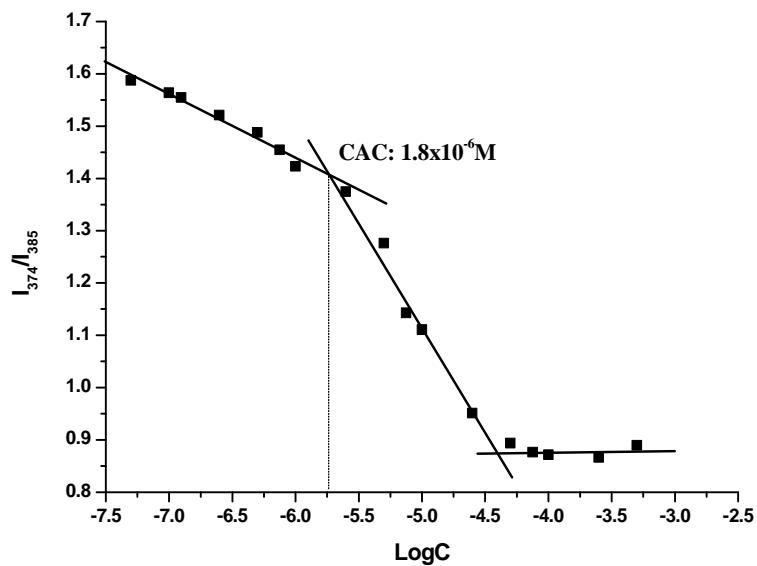
**Fig. S14** Zeta potential of vesicles formed with **3b** in the presence of NaCl (6.3 mM)

**8. Calculated molecular models of 3a-d**

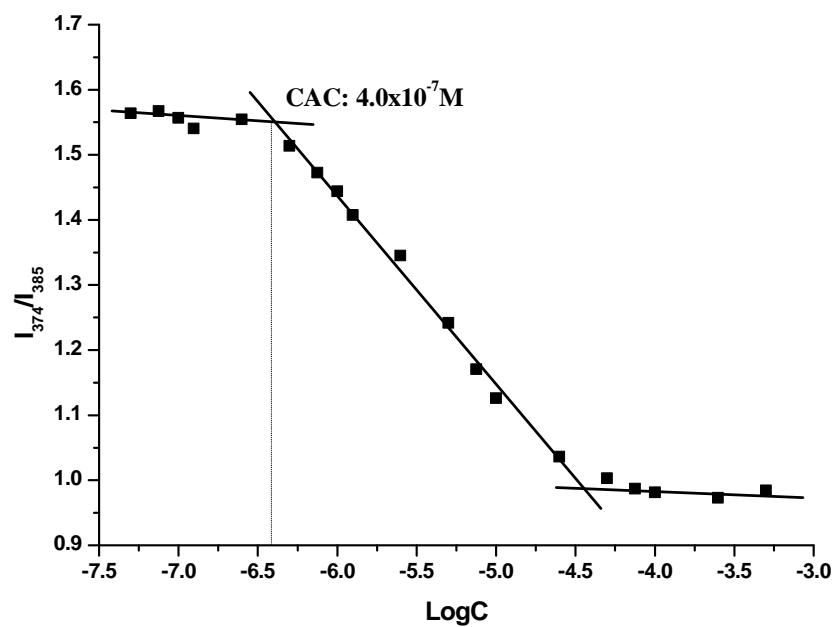


**Fig. S15** Molecular model of **3a** (A), **3b** (B), **3c** (C) and **3d** (D). Optimized at b3lyp/6-31g\* level

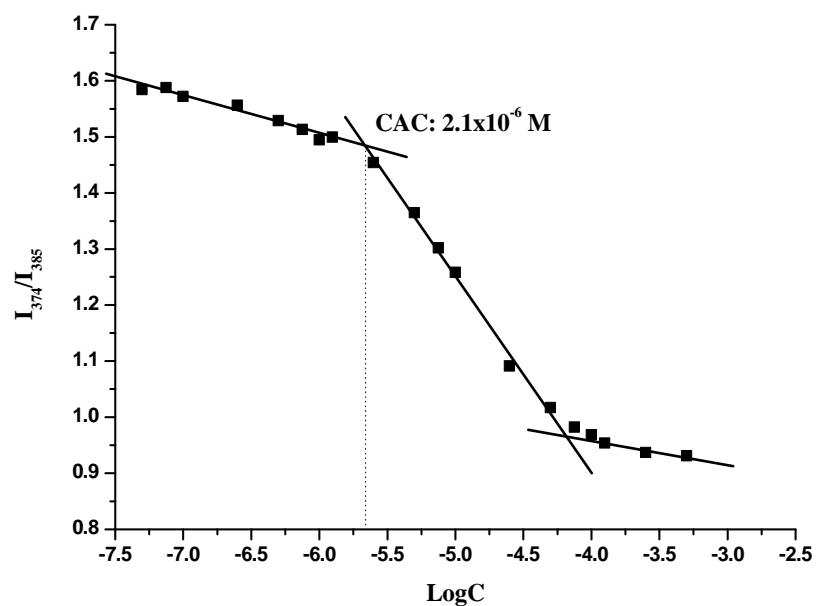
**9. Determination of CAC values of 3a-d**



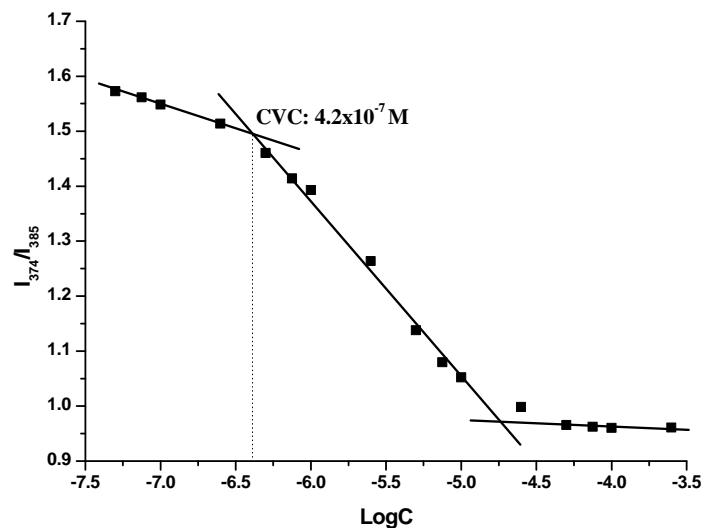
**Fig. S16** Plot of  $I_1/I_3$  ratio of pyrene vs concentration of **3a**. [pyrene]= $5 \times 10^{-7}$  M, excitation 335 nm.



**Fig. S17** Plot of  $I_1/I_3$  ratio of pyrene vs concentration of **3b**. [pyrene]= $5 \times 10^{-7}$  M, excitation 335 nm.

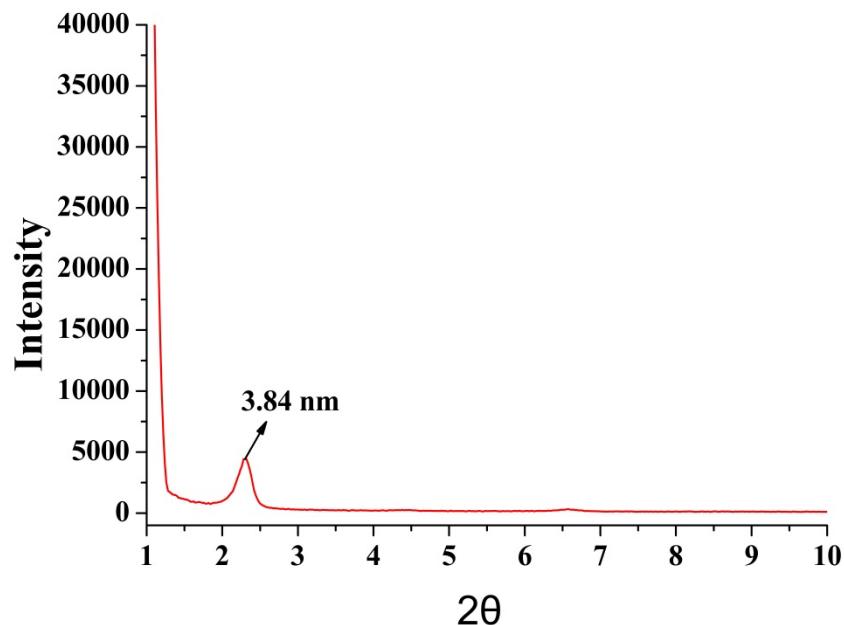


**Fig. S18** Plot of  $I_1/I_3$  ratio of pyrene vs concentration of **3c**.  $[\text{pyrene}] = 5 \times 10^{-7} \text{ M}$ , excitation 335 nm.

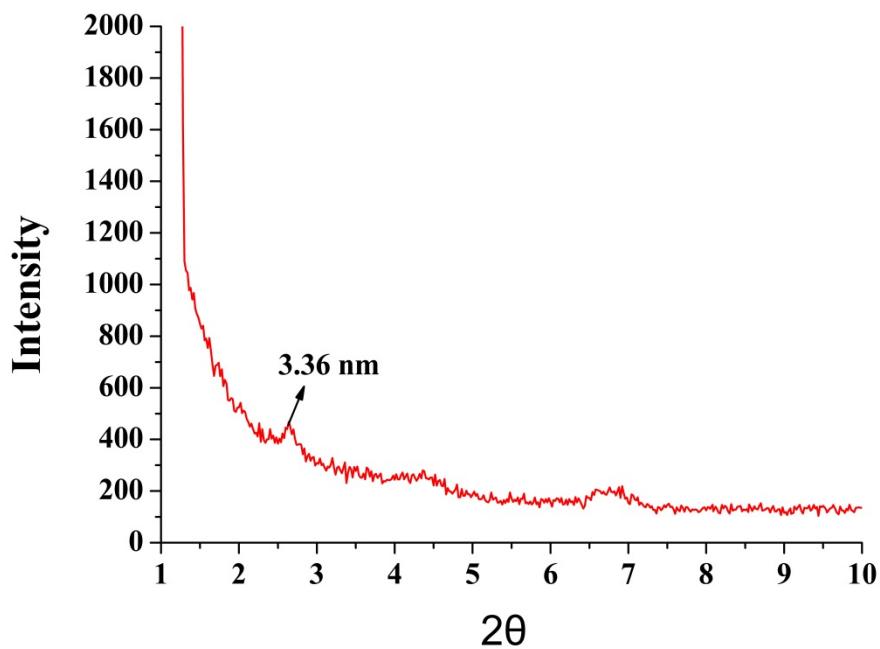


**Fig. S19** Plot of  $I_1/I_3$  ratio of pyrene vs concentration of **3d**.  $[\text{pyrene}] = 5 \times 10^{-7} \text{ M}$ , excitation 335 nm.

**10. XRD results for determination of the membrane thicknesses of 3a and 3b**

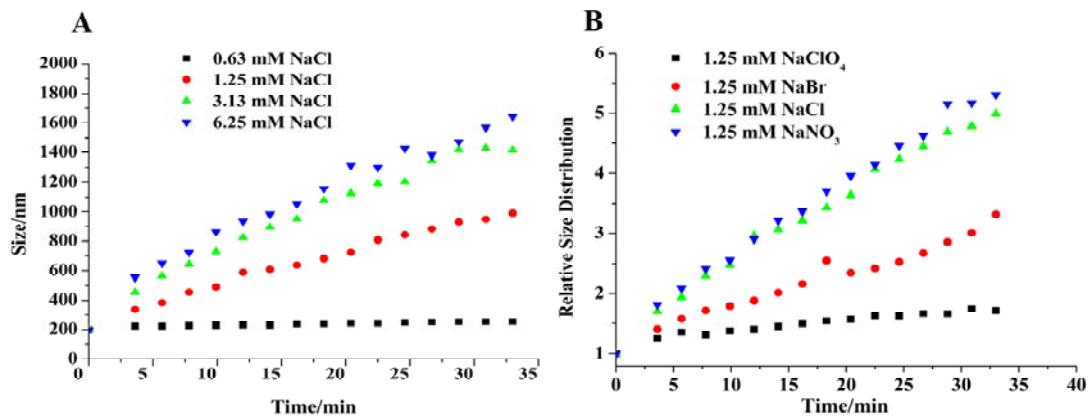


**Fig. S20** XRD result of 3a.

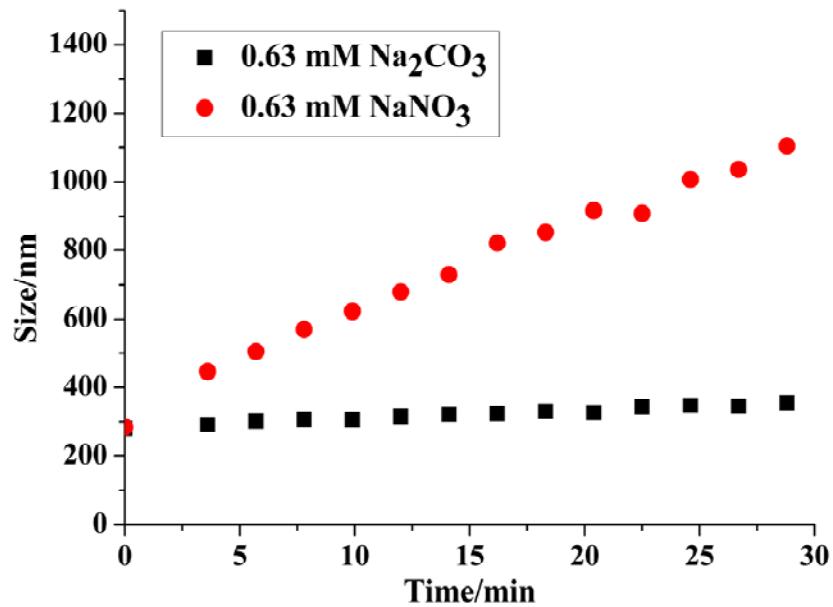


**Fig. S21** XRD result of 3b.

**11. Vesicular size responses of **3b** to anions**

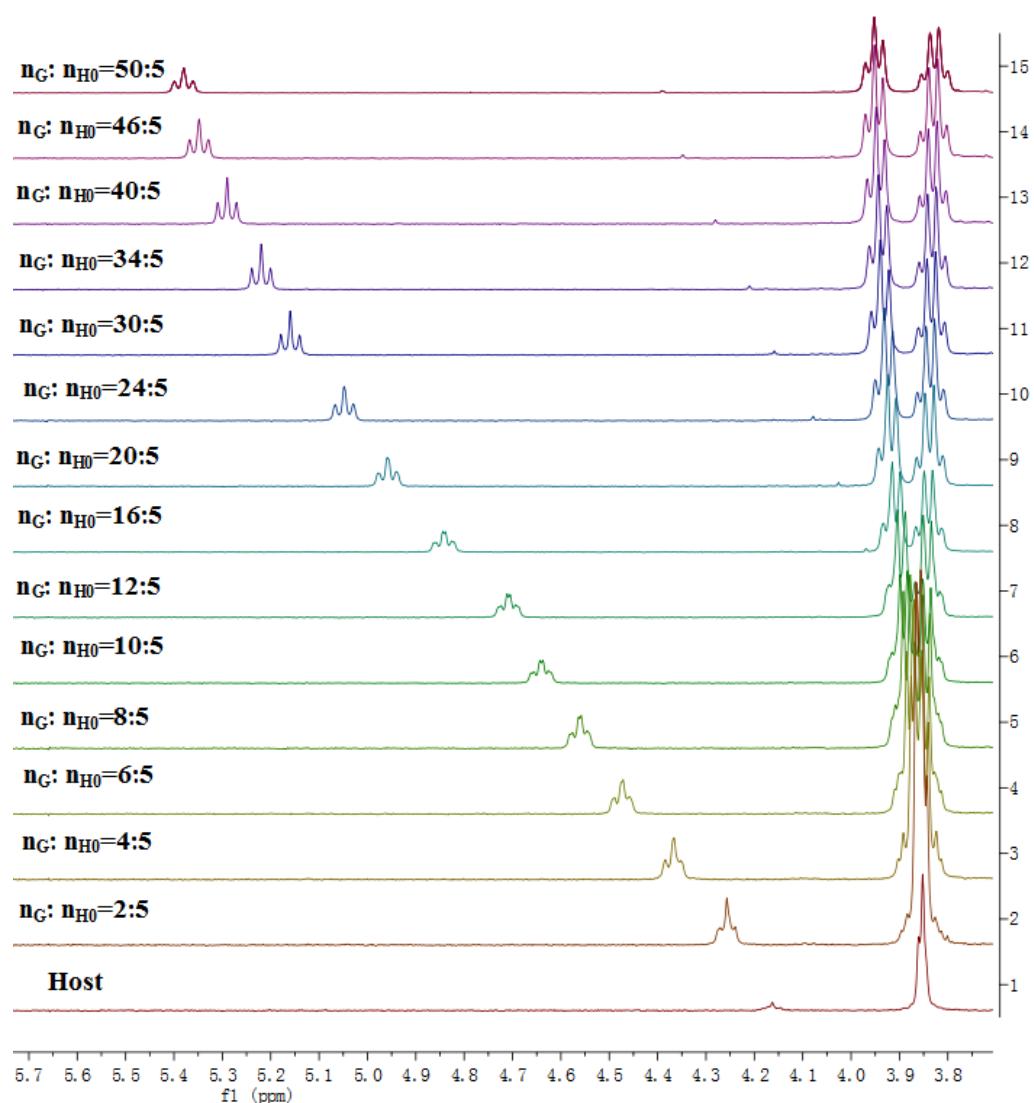


**Fig. S22** Vesicular size response of **3b** to (A) NaCl of different concentrations and various anions.

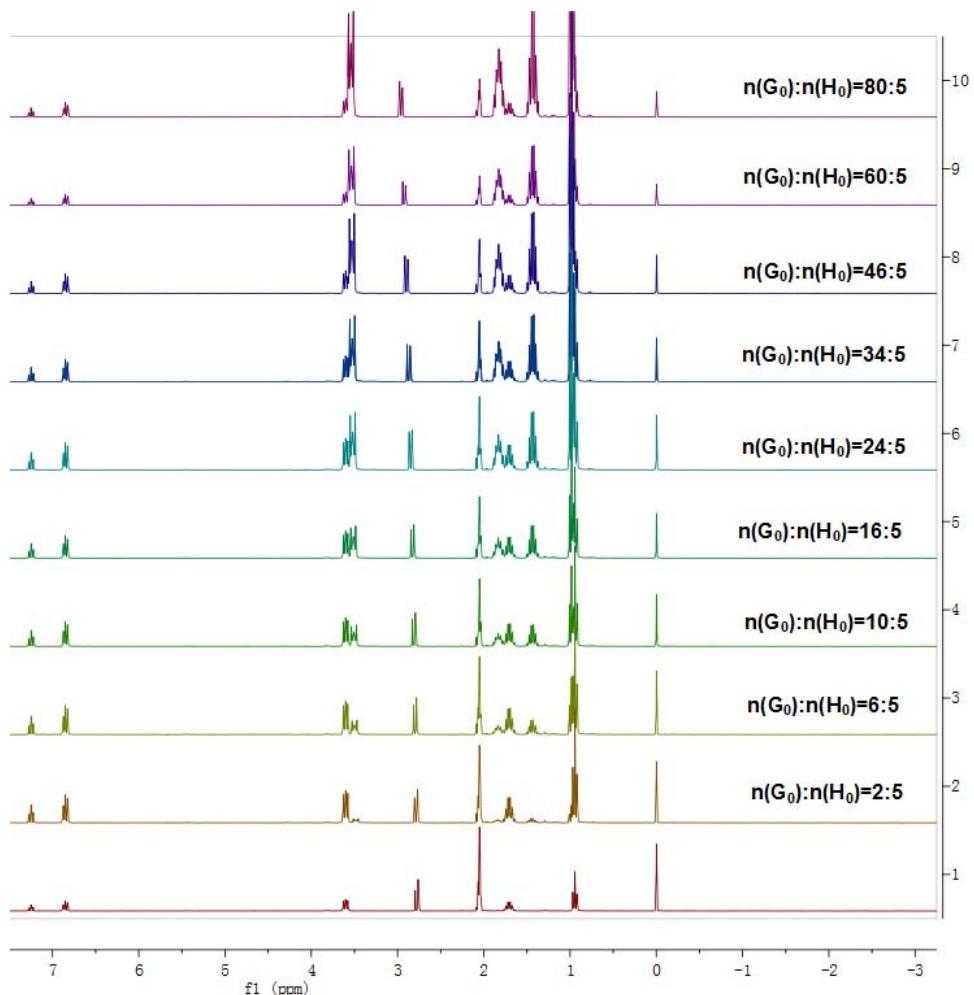


**Fig. S23** Comparison of effect nitrate and carbonate on the size of **3a**.

## 12. NMR titrations



**Fig. S24** <sup>1</sup>H NMR titrations of **4** in presence of n-Bu<sub>4</sub>NCl in acetone-d<sub>6</sub>.

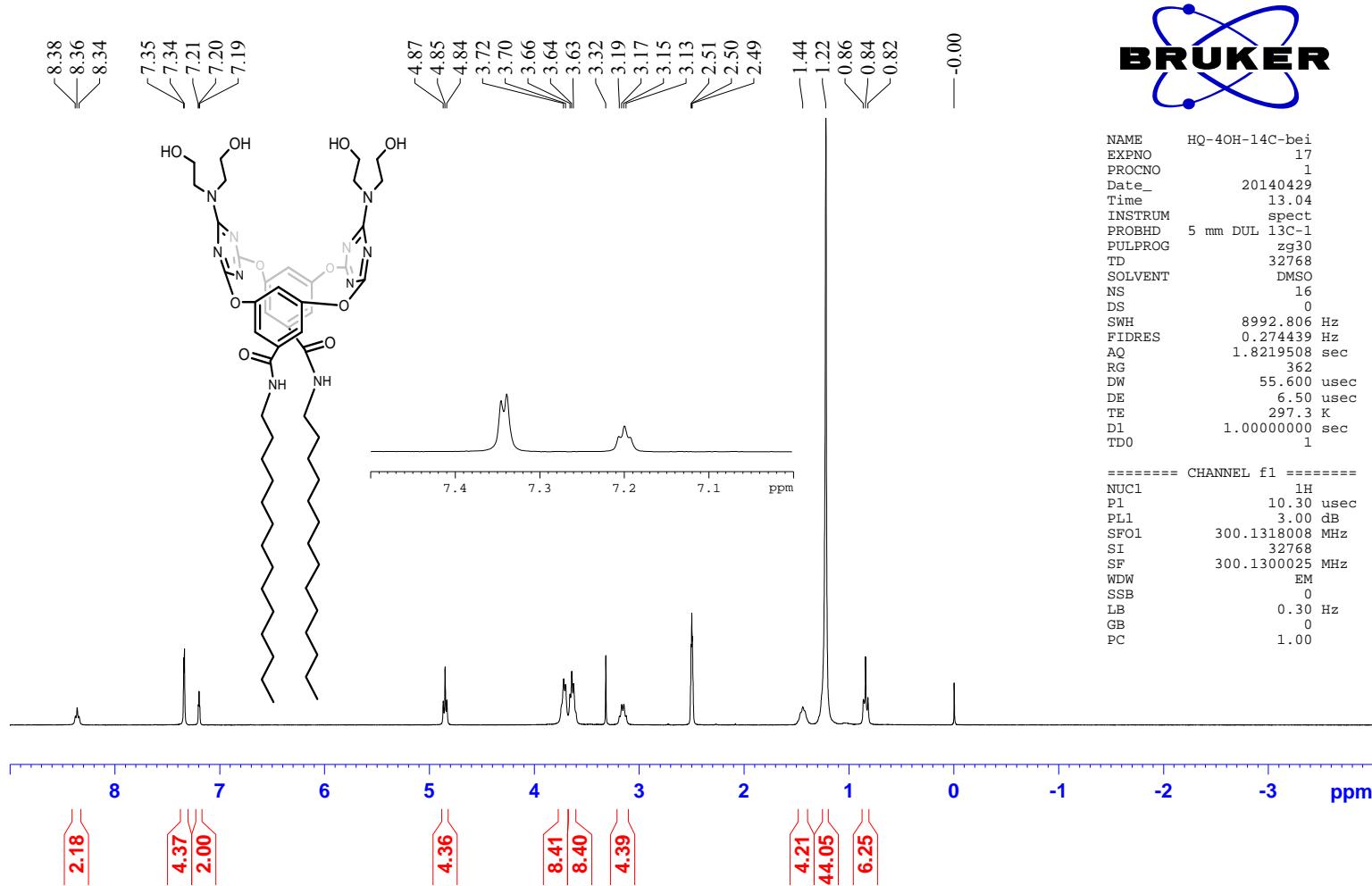


**Fig. S25**  $^1\text{H}$  NMR titrations of **5** in presence of  $\text{n-Bu}_4\text{NCl}$  in acetone- $\text{d}_6$ .

#### References:

1. J. P. Hagemann and P. T. Kaye, *Synthetic Commun.*, 1997, **27**, 2539-2546.
2. M. X. Wang and H. B. Yang, *J. Am. Chem. Soc.*, 2004, **126**, 15412-15422.
3. S. Quici, A. Manfredi, G. Pozzi, M. Cavazzini and A. Rozzoni, *Tetrahedron*, 1999, **55**, 10487-10496.
4. M. Iqbal, J. Huskens, M. Sypula, G. Modolo and W. Verboom, *New. J. Chem.*, 2011, **35**, 2591-2600.
5. H. -B. Yang, D. -X. Wang, Q. -Q. Wang and M. -X. Wang, *J. Org. Chem.*, 2007, **72**, 3757-3763.

**13  $^1\text{H}$  NMR spectra and  $^{13}\text{C}$  NMR spectra of the compounds**



**Figure S26**  $^1\text{H}$  NMR spectrum of 3a.

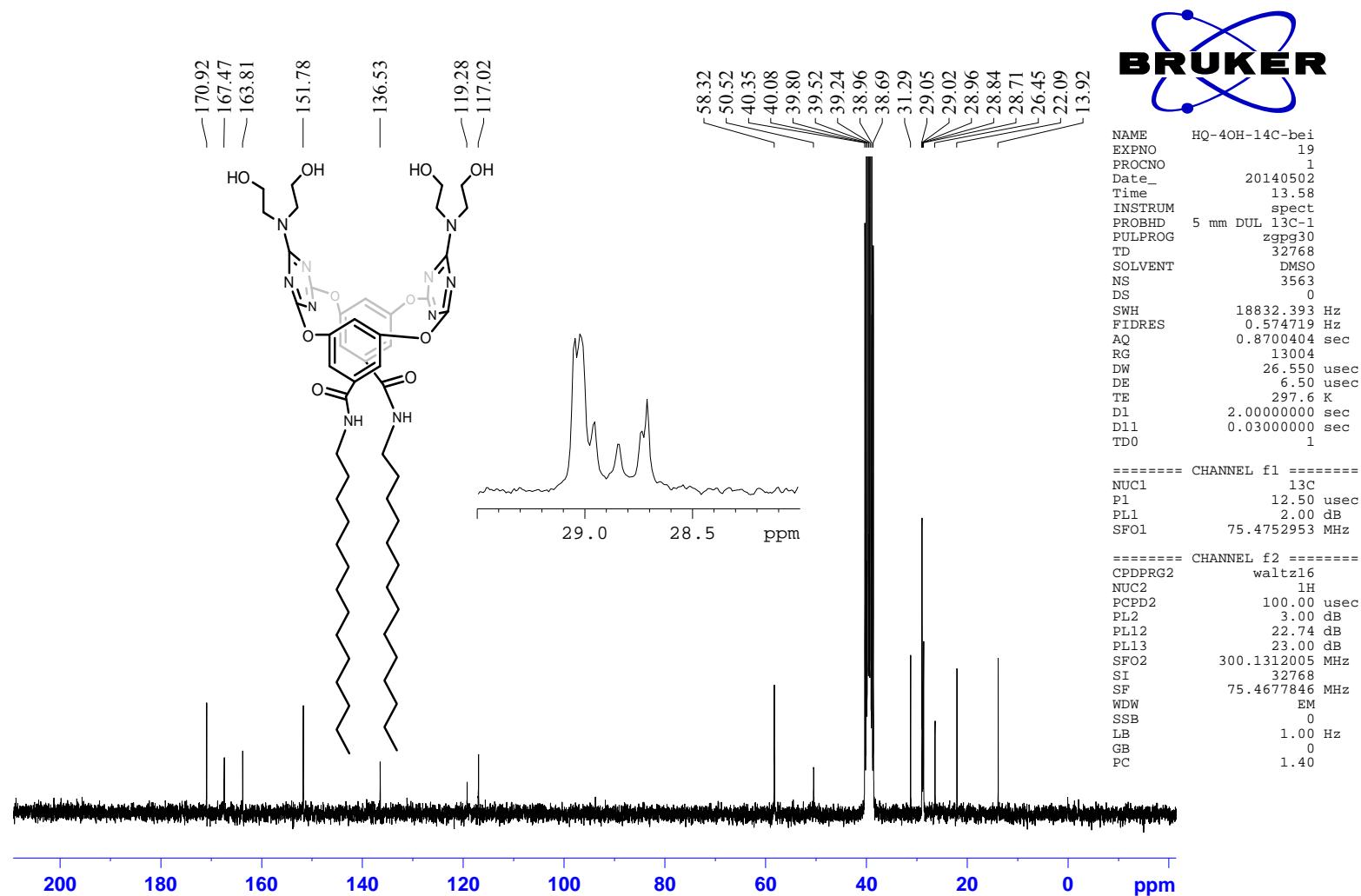


Figure S27  $^{13}\text{C}$  NMR spectrum of 3a.

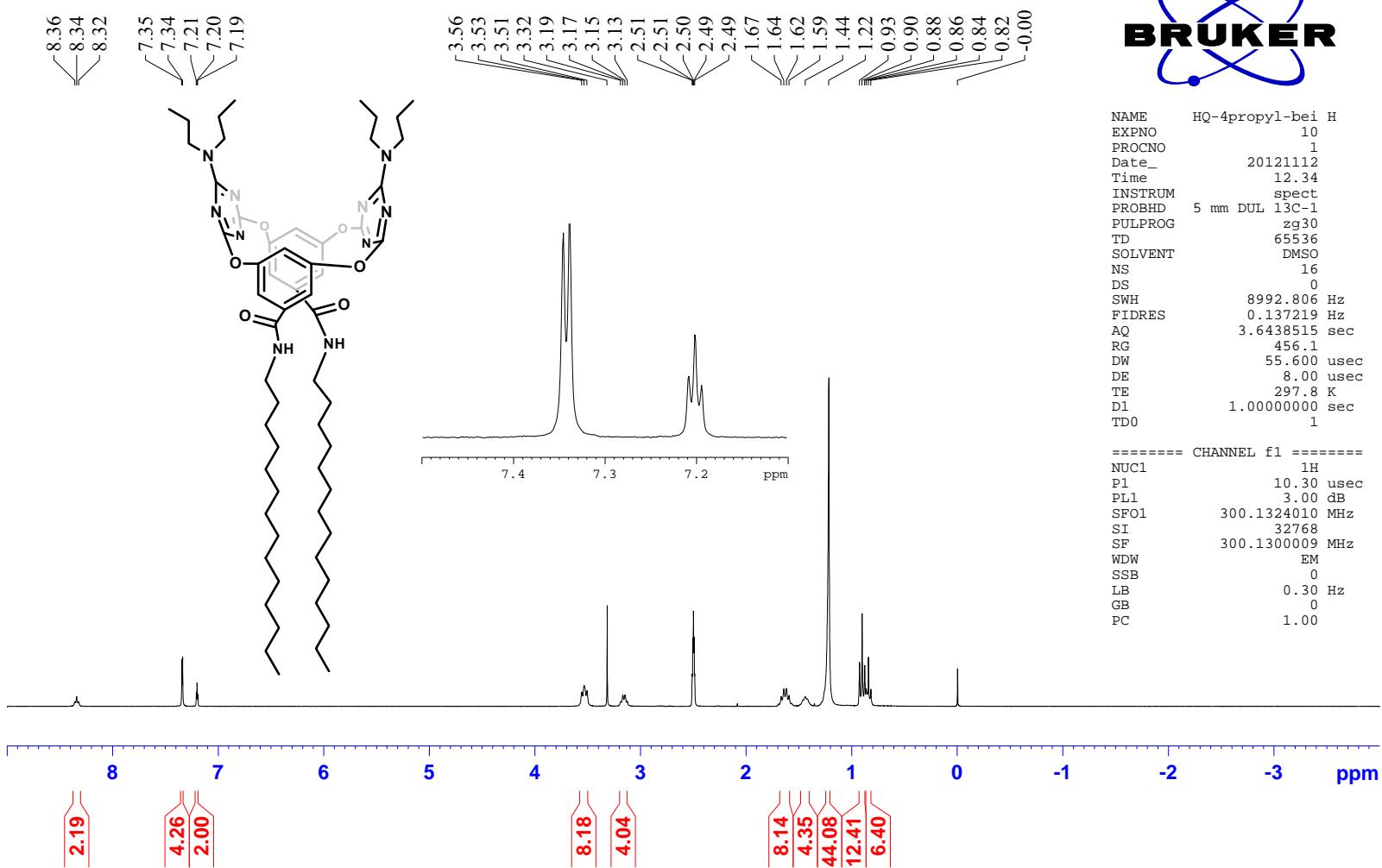
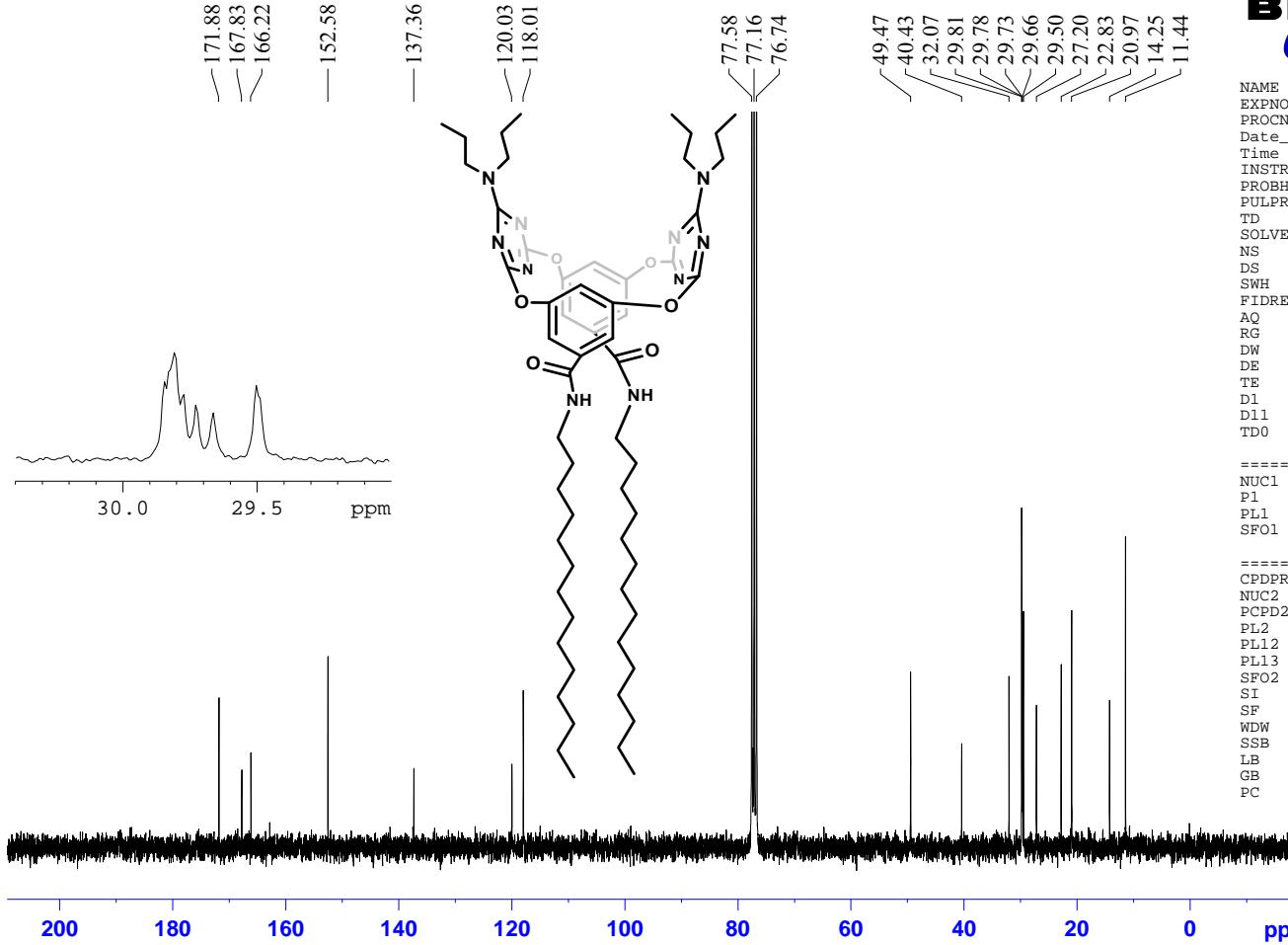


Figure S28  $^1\text{H}$  NMR spectrum of 3b.



NAME HQ-4propyl-bei  
 EXPNO 11  
 PROCNO 1  
 Date\_ 20121029  
 Time 12.45  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 876  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 RG 5160.6  
 DW 27.800 usec  
 DE 6.50 usec  
 TE 300.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 12.50 usec  
 PL1 2.00 dB  
 SFO1 75.4752953 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 100.00 usec  
 PL2 3.00 dB  
 PL12 22.74 dB  
 PL13 23.00 dB  
 SFO2 300.1312005 MHz  
 SI 32768  
 SF 75.4677366 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

**Figure S29**  $^{13}\text{C}$  NMR spectrum of **3b**.

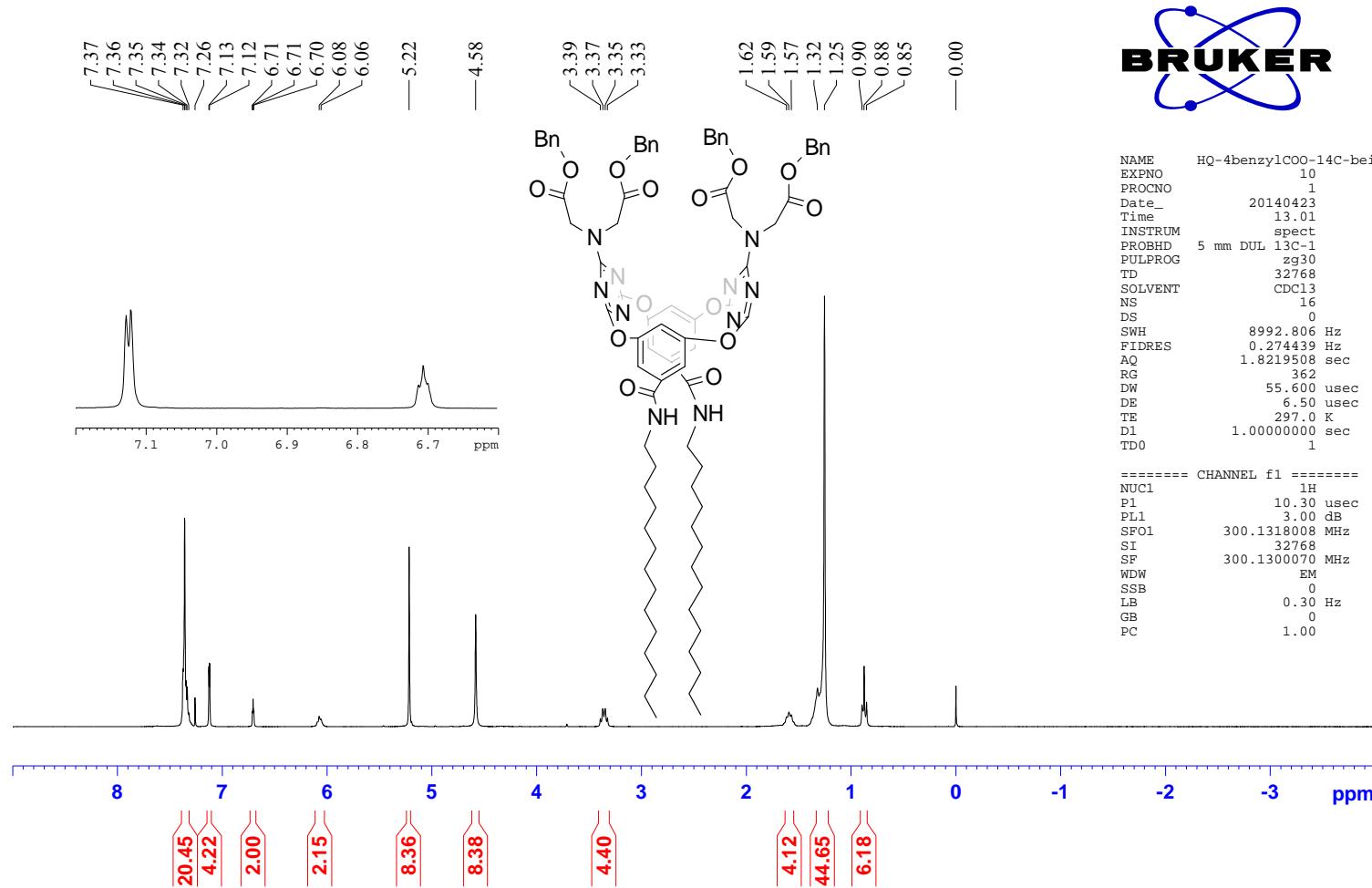


Figure S30 <sup>1</sup>H NMR spectrum of 3c`.

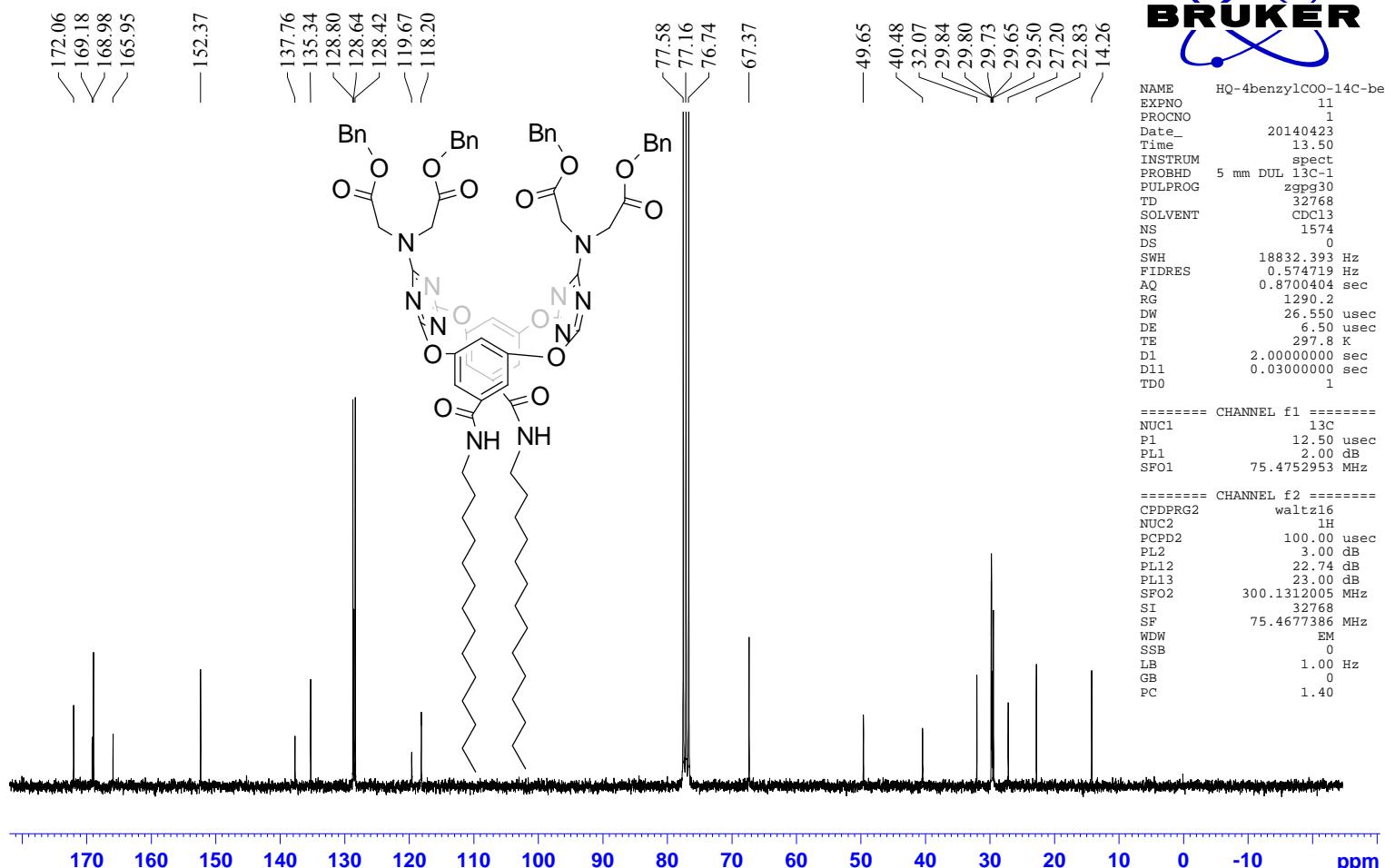


Figure S31  $^{13}\text{C}$  NMR spectrum of  $3\text{c}'$ .

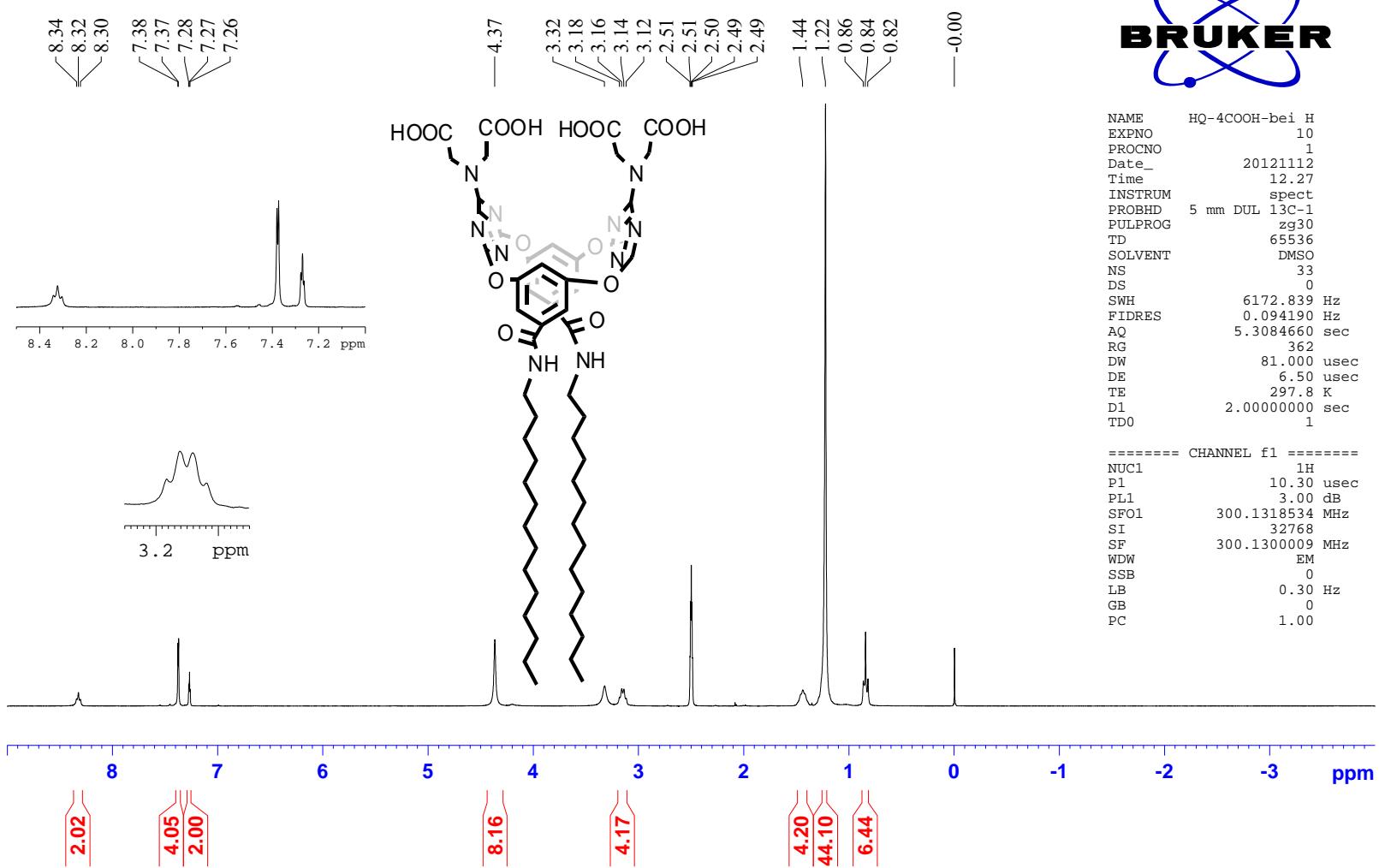


Figure S32  $^1\text{H}$  NMR spectrum of **3c**.

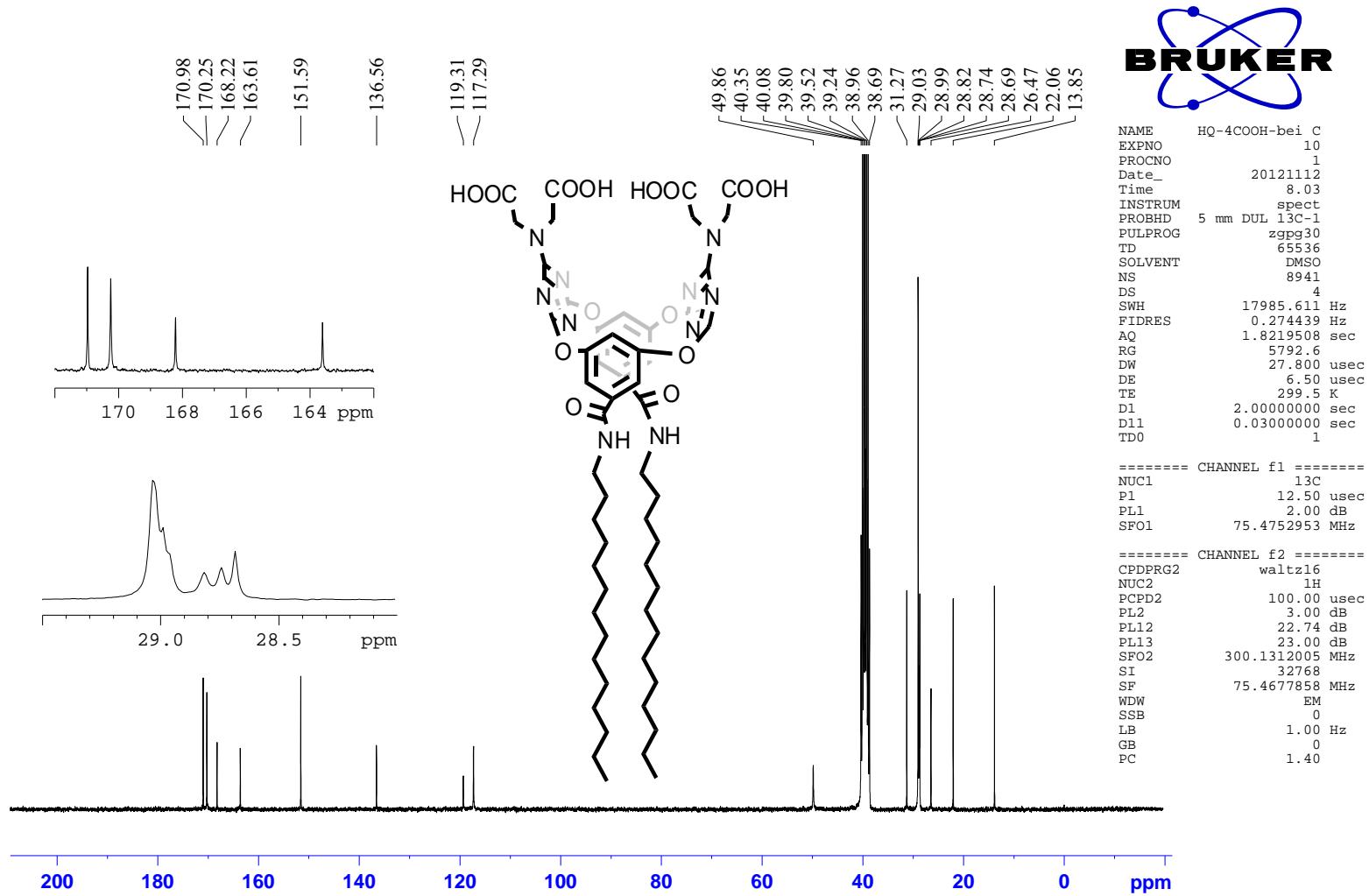


Figure S33  $^{13}\text{C}$  NMR spectrum of 3c.

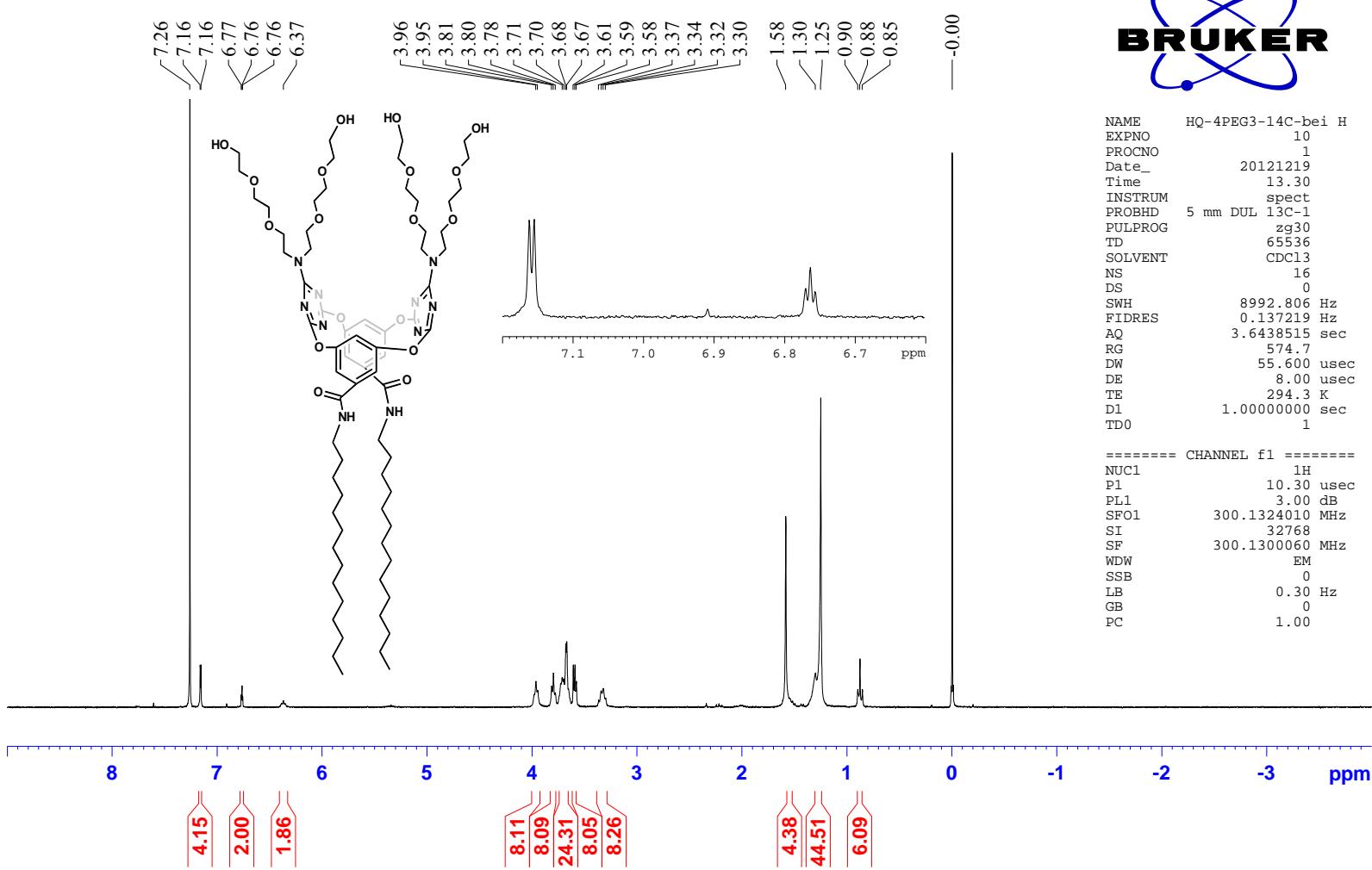


Figure S34  $^1\text{H}$  NMR spectrum of 3d.

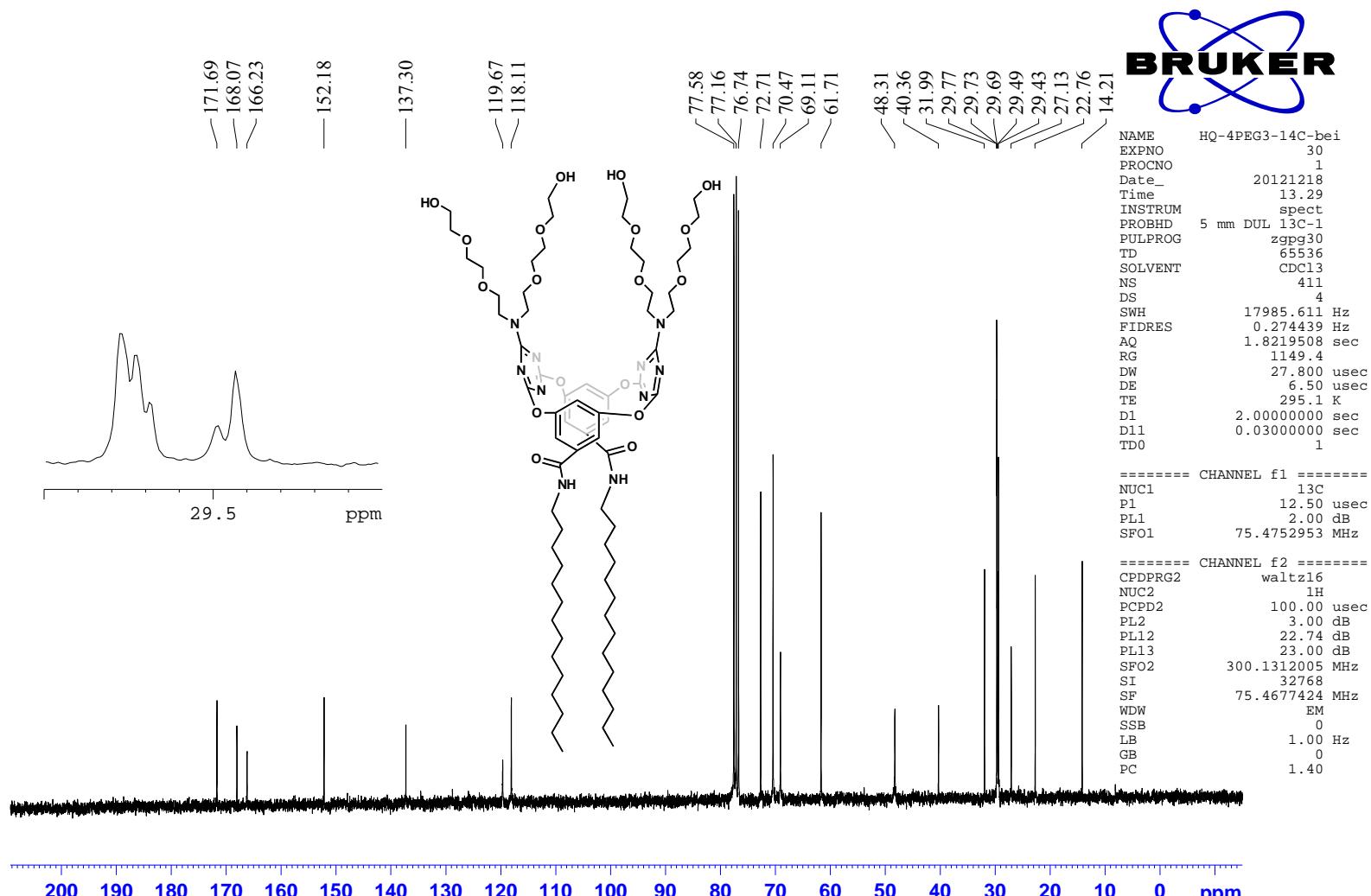


Figure S35  $^{13}\text{C}$  NMR spectrum of 3d.

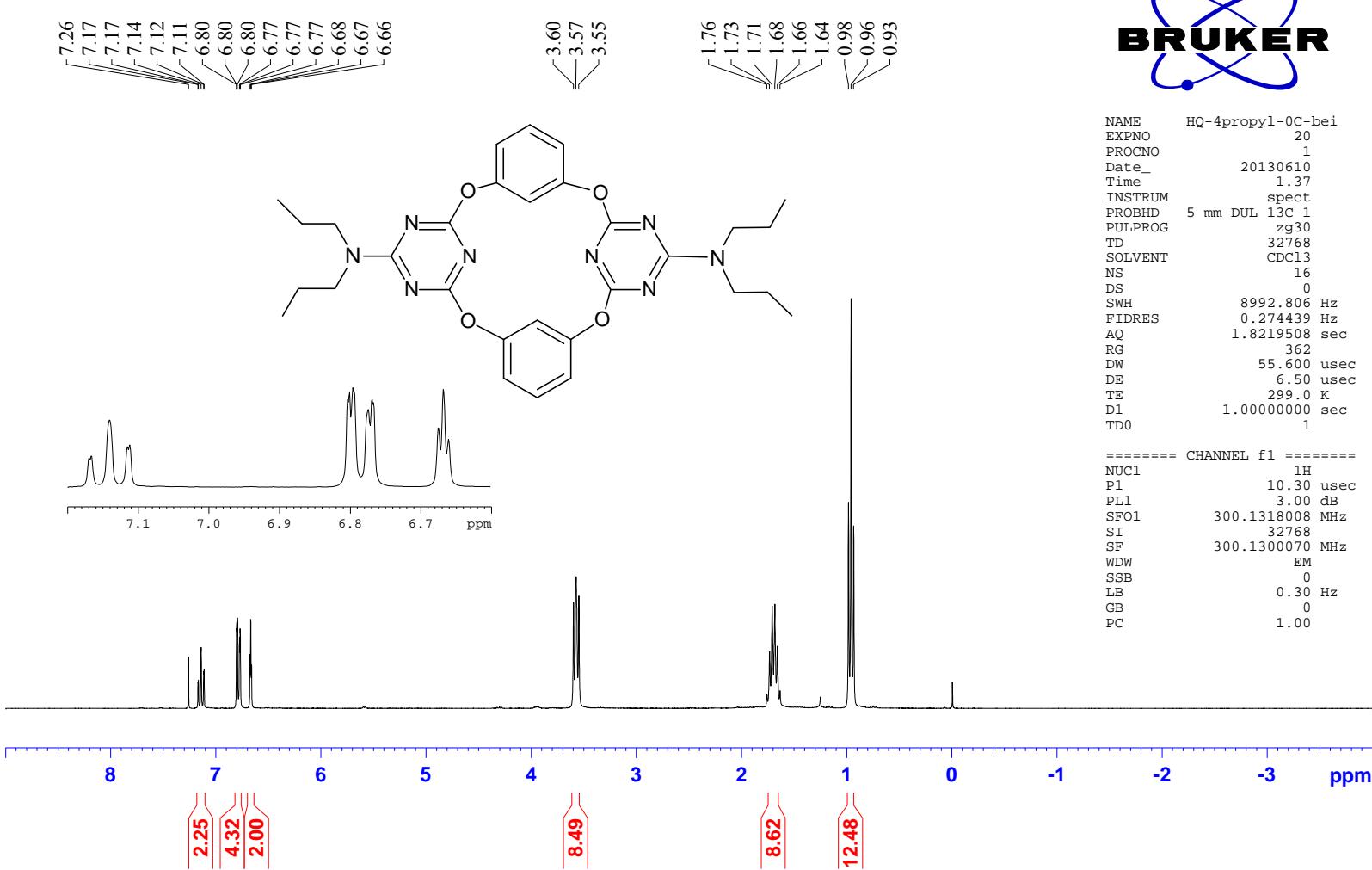
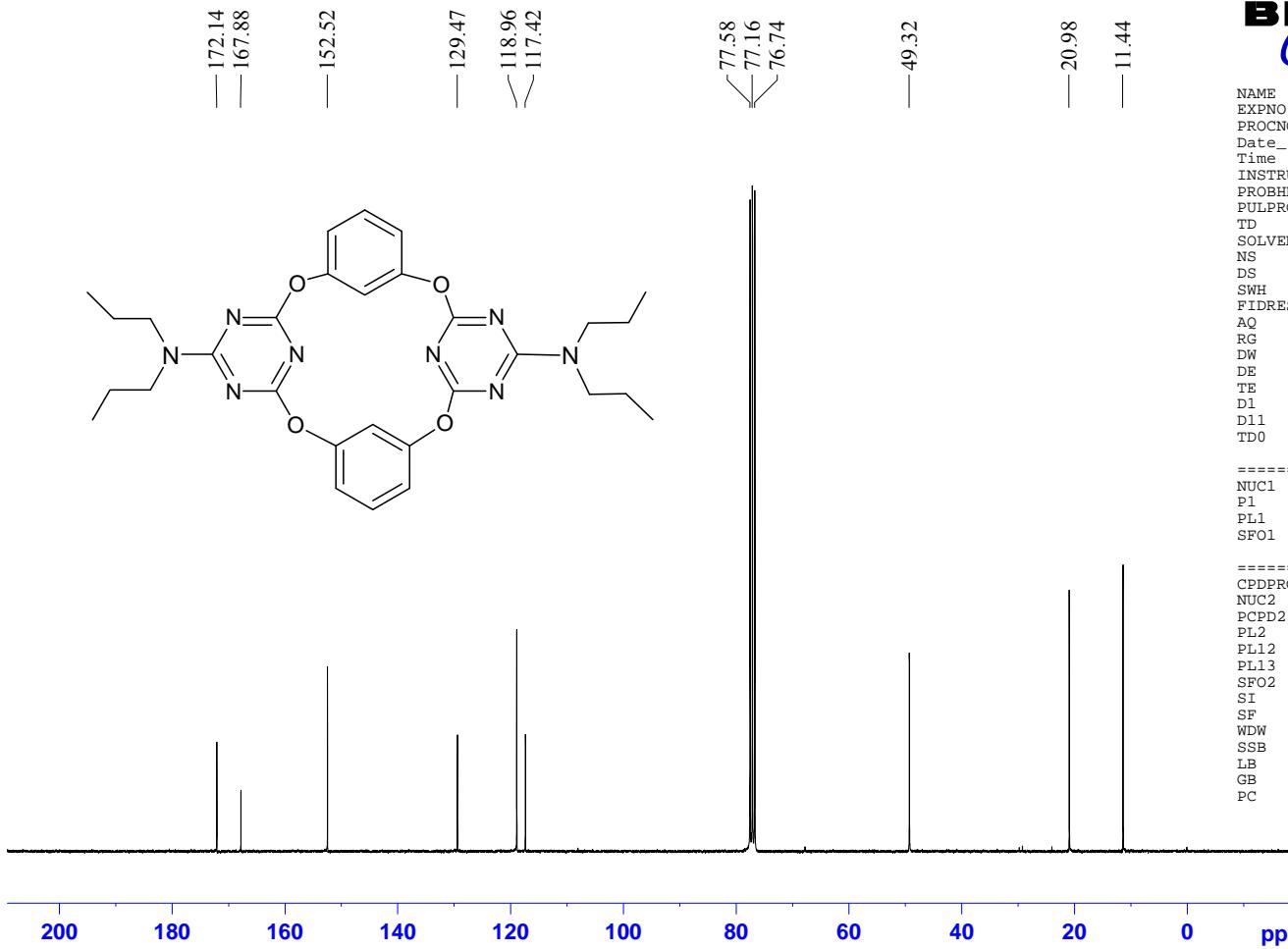


Figure S36 <sup>1</sup>H NMR spectrum of 5.



```

NAME      HQ-4propyl-0C-bei
EXPNO        30
PROCNO       1
Date_   20130610
Time    9.59
INSTRUM spect
PROBHD  5 mm DUL 13C-1
PULPROG zpgpg30
TD      32768
SOLVENT   CDCl3
NS       10240
DS        0
SWH     18832.393 Hz
FIDRES   0.574719 Hz
AQ      0.8700404 sec
RG      322.5
DW      26.550 usec
DE      6.50 usec
TE      300.0 K
D1      2.0000000 sec
D11     0.03000000 sec
TD0          1

===== CHANNEL f1 =====
NUC1      13C
P1        12.50 usec
PL1        2.00 dB
SFO1    75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     100.00 usec
PL2        3.00 dB
PL12      22.74 dB
PL13      23.00 dB
SFO2    300.1312005 MHz
SI        32768
SF      75.4677373 MHz
WDW        EM
SSB        0
LB      1.00 Hz
GB        0
PC      1.40

```

Figure S37  $^{13}\text{C}$  NMR spectrum of **5**.