ELECTRONIC SUPPORTING INFORMATION

A Highly versatile convergent/divergent "onion peel" synthetic strategy toward potent multivalent glycodendrimers

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Contents

1.	Materials and methods	2
2.	Synthetic protocols and characterization	4
3.	Surface plasmon resonance studies	63
4.	Sensorgrams	64

1. Materials and methods:

All reactions in organic medium were performed in standard oven dried glassware under an inert atmosphere of nitrogen using freshly distilled solvents. CH_2Cl_2 and DMF were distilled from CaH_2 and ninhydrin respectively, and kept over molecular sieves. Solvents and reagents were deoxygenated when necessary by purging with nitrogen. Water used for lyophilization of final dendrimers was nanopure grade, purified through Barnstead NANOPure II Filter with Barnstead MegOhm-CM Sybron meter. All reagents were used as supplied without prior purification unless otherwise stated, and obtained from Sigma-Aldrich Chemical Co. Ltd. Reactions were monitored by analytical thin-layer chromatography (TLC) using silica gel 60 F254 precoated plates (E. Merck) and compounds were visualized by 254 nm light, a mixture of iodine/silica gel and/or mixture of ceric ammonium molybdate solution (100 ml H₂SO₄, 900 ml H₂O, 25g (NH₄)₆Mo₇O₂₄H₂O, 10g Ce(SO₄)₂) and subsequent development by gentle warming with a heat-gun. Purifications were performed by flash column chromatography using silica gel from Silicycle (60 Å, 40-63 μ m) with the indicated eluent.

¹H NMR and ¹³C{¹H} NMR spectra were recorded at 300 or 600 MHz and 75 or 150 MHz, respectively, on a Bruker spectrometer (300 MHz) and Varian spectrometer (600 MHz). All NMR spectra were measured at 25°C in indicated deuterated solvents. Proton and carbon chemical shifts (δ) are reported in ppm and coupling constants (J) are reported in Hertz (Hz). The resonance multiplicity in the ¹H NMR spectra are described as "s" (singlet), "d" (doublet), "t" (triplet), "quint" (quintuplet) and "m" (multiplet) and broad resonances are indicated by "br". Residual protic solvent of CDCl₃ (¹H, δ 7.27 ppm; ¹³C, δ 77.0 ppm (central resonance of the triplet)), D₂O (¹H, δ4.79 ppm and 30.9 ppm for CH₃ of Acetone for ¹³C spectra of de-Oacetylated compounds), MeOD (¹H, $\delta 3.31$ ppm and ¹³C, δ 49.0 ppm. 2D Homonuclear correlation ¹H-¹H COSY experiments were used to confirm NMR peak assignments. Gel Permeation Chromatography (GPC) was performed using THF as the eluent, at 40°C with a 1 mL/min flow rate on a Viscotek VE 2001 GPCmax (SEC System) with Wyatt DSP/Dawn EOS and refractive index RI/LS system as detectors. 2 PLGel mixed B LS (10 µm, 300×7.5 mm) and LS-MALLS detection with performances verified with polystyrene 100 kDa and 2000 kDa were used to determine the number-average molecular weight (M_n) and polydispersity index (M_w/M_n) . Calculations were performed with Zimm Plot (model). Fourier transform infrared (FTIR) spectra were obtained with Thermo-scientific, Nicolet model 6700 equipped with ATR. The absorptions are given in wavenumbers (cm^{-1}) .

Accurate mass measurements (HRMS) were performed on a LC-MSD-ToF instrument from Agilent Technologies in positive electrospray mode. Low-resolution mass spectra were performed on the same apparatus or on a LCQ Advantage ion trap instrument from Thermo Fisher Scientific in positive electrospray mode (Mass Spectrometry Laboratory (Université de Montréal), or Plateforme analytique pour molécules organiques (Université du Québec à Montréal), Québec, Canada). Either protonated molecular ions $[M+nH]^{n+}$ or adducts $[M+nX]^{n+}$ (X = Na, K, NH₄) were used for empirical formula confirmation. MALDI-TOF experiments were performed on an Autoflex III from Brucker Smarteam in linear positive mode (Mass Spectrometry Laboratory (McGill University)) to afford adducts $[M+nX]^{n+}$ (X = Na, K or Li). Samples were solubilized in H₂O for a final concentration of 6 mg/mL. Dihydroxybenzoic acid was used as the matrix. Cationization was eased by the use of the corresponding sodium salt (2 mg/mL).

2. Synthetic protocols and characterization:



Synthesis of compound 2: A flame dried two-neck round bottom flask (250mL) was charged with dipentaerythritol **1** (4.00g, 15.7mmol) and sodium hydride (3.77g, 157.4mmol). To this, DMF (60ml) was added slowly at 0°C under nitrogen atmosphere. The reaction mixture was stirred for 20 minutes at 0°C. It was followed by the addition of allyl bromide (16.3mL, 188.8mmol) and stirred for 5 hrs at room temp. The completion of reaction was monitored by TLC. The reaction mixture was then quenched with methanol at 0°C. Solvent was evaporated and the residue was dissolved in ethyl acetate (100mL) and washed with water. Organic layer was separated, dried with Na₂SO₄, filtered and concentrated under reduced pressure. The crude compound was purified by column chromatography using 7% ethyl acetate in hexanes as eluent to afford desired compound **2** (6.22g, 12.6mmol) in 80% yield as light yellow oil.

¹**H** NMR (300 MHz, CDCl₃) δ 5.88 (ddt, J = 17.1, 10.5, 5.3 Hz, 6H), 5.34–5.08 (m, 12H), 3.94 (dt, J = 5.2, 1.4 Hz, 12H), 3.46 (d, J = 7.9 Hz, 12H), 3.40 (s, 4H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 135.3, 116.0, 72.3, 70.1, 69.4, 45.6. IR (cm⁻¹) 3079, 2980, 2903, 2867, 1646, 1478, 1420, 1349, 1269, 1173, 989, 920. HRMS (ESI⁺) for C₂₈H₄₆O₇ *m/z* calc for C₂₈H₄₆O₇, 495.3316 [*M*+H]⁺; found: 495.3299.



Figure S1. ¹H NMR spectrum of compound 2 (CDCl₃, 300 MHz).



Figure S2. ¹³C{¹H} NMR spectrum of compound 2 (CDCl₃, 75 MHz).



Figure S3. COSY spectrum of compound 2.



Figure S4. HRMS (ESI⁺) spectrum of compound 2.



Figure S5. IR spectrum of compound 2.



Synthesis of compound 3: To a stirring solution of hexa-allyl derivative 2 (300mg, 0.606mmol), 2,2'-dimethoxy-2-phenylacetophonone (DMPAP) (155mg, 0.606mmol) in dry DMF (3ml) was added cysteamine hydrochloride (1.03g, 9.07mmol) under nitrogen. The vial was then purged with N_2 for 10 min and irradiated for 4-6 hrs with UV lamp (365nm) at room temperature. Upon completion of the reaction, the contents of the vial were washed three times with diethyl ether to remove excess of thiol, affording a clear viscous liquid. It was further purified using dialysis bag (cut-off 1000 Da, spectrum). Dialysis bath water was changed 4-5 times in the span of 6 hrs to remove all the impurities. The compound was lyophilized to yield white hygroscopic solid 3 (534mg, 0.455mmol) in 75% yield.

¹**H NMR** (300 MHz, D₂O) δ 3.62 (t, J = 6.2 Hz,12H), 3.48 (s, 12H), 3.39 (s, 4H), 3.26 (t, J = 6.7 Hz, 12H), 2.90 (t, J = 6.8 Hz, 12H), 2.70 (t, J = 7.2 Hz, 12H), 1.98–1.83 (m, 12H). ¹³C{¹H} **NMR** (75 MHz, D₂O) δ 70.5, 69.9, 45.8, 38.9, 28.8, 28.0. **IR** (cm⁻¹) 3637, 2980, 2971, 2883, 1382, 1150, 1070, 954. **HRMS** (ESI+) m/z calc. for C₄₀H₈₈N₆O₇S₆, 957.5112 [M+H]⁺; found, 957.5134.







¹¹⁵ ¹¹⁰ ¹⁰⁵ ¹⁰⁰ ⁹⁵ ⁹⁰ ⁸⁵ ⁸⁰ ⁷⁵ ⁷⁰ ⁶⁵ ⁶⁰ ⁵⁵ ⁵⁰ ⁴⁵ ⁴⁰ ³⁵ ³⁰ ²⁵ ²⁰ ¹⁵ ¹⁰ ⁵ **Figure S7**. ¹³C{¹H} NMR spectrum of compound **3** (75 MHz, D_2O).



Figure S8. COSY spectrum of compound 3.





Figure S10. IR spectrum of compound 3.



Synthesis of compound 5: A solution of hexa-amine hydrochloride **3** (150mg, 0.127mmol) and *N*,*N*-diisopropylethylamine (0.26ml, 1.48mmol) in DMF (2ml) was stirred for 30 minutes. In another two neck flask, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC.HCl) (243mg, 1.27mmol), and 4-(dimethylamino)pyridine (DMAP) (155mg, 1.27mmol) were added in DMF (4mL) and stirred for 10 minutes followed by the addition of tripropargylated gallic acid **4** (434mg, 1.53mmol). The free amine solution from first flask was then transferred to the reaction mixture of the second flask with the help of canula syringe and was heated at 50°C for overnight. The completion of reaction was monitored by TLC. Upon completion, the reaction mixture was diluted with water (40mL) and extracted with ethyl acetate (3×40mL). The combined organic extracts were washed with 0.1 N HCl (3×15mL), followed by saturated NaHCO₃ solution and brine. The organic layer was dried over anhydrous sodium sulfate, filtered and evaporated under reduced pressure. The crude mixture was then purified with the help of flash column chromatography using 80% EtOAc in hexanes as eluent. The desired compound **5** (232.5mg, 0.0910mmol) was achieved in 72% yield as light yellow oil.

¹**H** NMR (300 MHz, CDCl₃) δ 7.27–7.23 (m, 12H), 7.07 (br s, 6H), 4.78 (dd, J = 2.3, 2.3 Hz, 36H), 3.60 (d, J = 6.0 Hz, 12H), 3.43 (t, J = 5.9 Hz, 12H), 3.31 (d, J = 8.1 Hz, 16H), 2.74 (t, J = 6.6 Hz, 12H), 2.59 (t, J = 7.2 Hz, 12H), 2.55 (t, J = 2.3 Hz, 12H), 2.48 (t, J = 2.4 Hz, 6H), 1.92–1.70 (m, 12H).

¹³C{¹H} NMR (75 MHz, CDCl₃) δ 166.7, 151.5, 140.0, 130.2, 107.9, 78.7, 78.1, 76.4, 75.7, 69.6, 60.3, 57.3, 45.6, 39.2, 31.6, 29.8, 28.4.

IR (cm⁻¹) 3290, 3005, 2922, 2867, 2122, 1637, 1581, 1541, 1492, 1428, 1365, 1323, 1275, 1261, 1207, 1106, 1032, 992, 764, 750, 671.

HRMS (**ESI**⁺) m/z for C₁₃₆H₁₄₈N₆O₃₁S₆, 1277.4329 [*M*+2H]²⁺; found, 1277.4359, 2576.8406 [*M*+Na]⁺; found 2575.8346.



Figure S11. ¹H NMR spectrum of compound 5 (300 MHz, CDCl₃).



Figure S12. ¹³C{¹H} NMR spectrum of compound **5** (75 MHz, CDCl₃).



Figure S13. COSY spectrum of compound 5.



Figure S14. IR spectrum of compound 5.



Figure S15. HRMS (ESI^+) spectrum of compound **5**.



Synthesis of compound 7: Divergent approach

To a solution of compound **5** (50.0mg, 0.0195mmol) in THF (3mL) was added azido derivative **6** (261mg, 0.702mmol) dissolved in THF (2mL), followed by the addition of sodium ascorbate (70.0mg, 0.351mmol). An aqueous solution of $CuSO_4 \cdot 5H_2O$ (88.0 mg, 0.351mmol) was then added to the reaction mixture. The final ratio of H_2O to THF was kept 1:1. The reaction mixture was stirred at 40°C for 12 hrs. The progress of the reaction was monitored with the help of TLC. Upon completion, reaction mixture was diluted with EtOAc (25mL) and washed with a saturated solution of EDTA (2×15mL). Organic layer was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. Purification of the crude compound was achieved *via* flash column chromatography on silica gel using 2-4% MeOH in DCM as eluent gradient to afford desired compound **7** (146.5mg, 0.0158mmol) in 81% yield as a white solid.

¹**H NMR** (300 MHz, CDCl₃) δ 8.16 (s, 6H), 8.09 (s, 12H), 7.19 (s, 12H), 5.94 (d, J = 9.2 Hz, 18H), 5.68 (t, J = 9.8 Hz, 6H), 5.56 (dd, J = 10.9, 6.9 Hz, 32H), 5.38–5.12 (m, 55H), 4.32 (t, J = 6.4 Hz, 18H), 4.27–4.06 (m, 38H), 3.58 (d, J = 5.5 Hz, 12H), 3.45 (s, 12H), 3.34 (s, 16H), 2.74 (t, J = 6.5 Hz, 12H), 2.63 (t, J = 6.9 Hz, 12H), 2.20 (s, 58H), 2.01 (d, J = 8.4 Hz, 118H), 1.77 (d, J = 18.0 Hz, 54H).

¹³C{¹H} NMR (75 MHz, CDCl₃) δ 170.3, 170.3, 170.1, 170.1, 169.9, 169.8, 169.6, 169.0, 168.7, 166.7, 151.9, 144.6, 144.0, 140.1, 130.3, 123.3, 122.3, 107.2, 86.1, 85.7, 73.7, 73.5, 73.2, 71.0, 70.7, 69.7, 67.9, 67.7–67.5, 66.8, 66.1, 62.8, 61.0, 31.3, 29.8, 28.5, 20.5, 20.1.

IR (cm⁻¹): 3628, 2994, 2947, 1751, 1651, 1583, 1491, 1428, 1370, 1218, 1093, 1064, 923, 732, 667.

MALDI-TOF: m/z calc. for C₃₈₈H₄₉₀N₆₀O₁₉₃S₆, 9274.7; found, 9296.4. $[M+Na]^+$; m/z calc. for C₃₈₈H₄₉₀N₆₀O₁₉₃S₆, 9274.7; found, 9312.05. $[M+K]^+$



Figure S16. ¹H NMR spectrum of compound 7 (300 MHz, CDCl₃).





Figure S18. COSY spectrum of compound 7.





Figure S19. MALDI ToF spectrum of compound 7.



Figure S20. IR spectrum of compound 7.



Figure S21. GPC profile of compound 7.



Synthesis of compound 8: To a stirring solution of compound **7** (100mg, 0.0107mmol) in MeOH (3mL) was slowly added 1M solution of MeONa in MeOH to adjust the pH 9–10. Reaction mixture was left for overnight stirring at room temperature. The reaction pH was then adjusted with H^+ resin to pH 6. Solvent was evaporated and the residue was dissolved in 3mL of water and washed with diethyl ether (3×15mL) to remove impurities. Aqueous layer was finally lyophilized to yield **8** (58.8mg, 9.42µmol) as a white solid with a 88% yield.

¹**H** NMR (600 MHz, D_2O) δ 8.29 (s, 12H), 7.98 (s, 6H), 7.15 (s, 12H), 5.59 (d, J = 65.2 Hz, 18H), 5.04–4.79 (m, 112H), 4.29–3.22 (m, 150H), 2.68 (t, J = 56.2 Hz, 24H), 1.97–1.70 (m, 12H).

¹³C{¹H} NMR (151 MHz, D₂O) δ 168.6, 152.2, 144.0, 143.7, 139.3, 130.6, 125.2, 124.5, 107.3, 88.8, 88.6, 78.8, 73.7, 70.3, 69.8, 69.2, 65.8, 62.7, 61.4, 40.5, 30.1, 28.9.

IR (cm⁻¹) 3350, 2879, 1637, 1583, 1494, 1428, 1327, 1233, 1095, 1057, 891, 825, 760, 703. **HRMS** (**ESI**⁺) m/z calc. for C₂₄₄H₃₄₆N₆₀O₁₂₁S₆, 1584.0165 ([*M*+4Na]⁴⁺); found, 1584.0122.



Figure S22. ¹H NMR spectrum of compound 8 (600 MHz, D₂O).



Figure S23. $^{13}C{^{1}H}$ NMR spectrum of compound 8 (151 MHz, D₂O).



Figure S24. COSY spectrum of compound 8 (300 MHz, D₂O).



Figure S25. HRMS (ESI⁺) spectrum of compound 8.



Figure S26. IR spectrum of compound 8.



Synthesis of compound 10: To a stirring solution of tripropargyl gallic acid **4** (562mg, 1.98mmol) in DMF (3mL) were added EDC·HCl (529mg, 2.77mmol) and DMAP (290mg, 2.37mmol). The reaction mixture was stirred under nitrogen atmosphere for 15 minutes. Amine terminated compound **9** (600mg, 1.98mmol) was then added and the reaction mixture was stirred at 50°C for overnight. The completion of reaction was monitored by TLC. Upon completion, the reaction mixture was diluted with water (50mL) and extracted with ethyl acetate (3×30mL). The combined organic extracts were washed with 0.1N HCl (3×10mL), followed by saturated NaHCO₃ solution and brine. The organic layer was dried over anhydrous Na₂SO, filtered and evaporated under reduced pressure. The crude mixture was then purified by flash column chromatography using 2% MeOH in DCM as eluent to furnish **10** (905mg, 1.54mmol) in a 78% yield as an off-white solid.

¹**H** NMR (300 MHz, CDCl₃) δ 7.40–7.27 (m, 5H), 7.24–7.04 (m, 12H), 6.20 (s, 1H), 4.72 (dd, *J* = 2.4, 2.4 Hz, 6H), 3.20 (m, 2H), 2.46 (t, *J* = 6.2 Hz, 2H), 2.38 (t, *J* = 2.4 Hz, 1H), 2.32 (t, *J* = 2.3 Hz, 2H).

¹³C{¹H} NMR (75 MHz, CDCl₃) δ 166.1, 151.1, 144.3, 139.4, 130.0, 129.2, 127.7, 126.5, 107.5, 78.52, 77.8, 76.2, 75.5, 66.5, 60.0, 56.9, 38.4, 31.7.

IR (cm⁻¹): 3284, 3057, 2937, 2121, 1750, 1647, 1582, 1537, 1489, 1444, 1427, 1368, 1323, 1214, 1106, 1063, 952, 923, 734, 700.

HRMS (**ESI**⁺) m/z calc for C₃₇H₃₁NO₄S, 608.1866 [M+Na]⁺; found, 608.1863.



Figure S27. ¹H NMR spectrum of compound 10 (300 MHz, CDCl₃).



Figure S28. $^{13}C{^{1}H}$ NMR spectrum of compound 10 (75 MHz, CDCl₃).



Figure S29. COSY spectrum of compound 10.



Figure S30. HRMS (ESI⁺) spectrum of compound 10.



Figure S31. IR spectrum of compound 10.



Synthesis of compound 11: To a solution of compound **10** (200mg, 0.341mmol) in THF (2mL) was added galactosyl azide **6** (442mg, 1.196mmol) dissolved in THF (3mL), followed by the addition of sodium ascorbate (67mg, 0.34mmol). An aqueous solution of $CuSO_4 \cdot 5H_2O$ (85mg, 0.34mmol) was added and the final ratio of H₂O and THF was kept 1:1. The reaction mixture was stirred at 40°C for 12 hrs. The progress of the reaction was monitored with the help of TLC. Upon completion, reaction mixture was diluted with EtOAc (25mL) and washed with saturated solution of EDTA (2×15mL). Organic layer was finally washed with brine solution, dried with anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. Purification of the residue was achieved *via* flash column chromatography on silica gel using 2% MeOH in DCM as eluent to afford the desired compound **11** (488mg, 0.286mmol) in a 84% yield as an off-white solid.

¹**H** NMR (300 MHz, CDCl₃) δ 8.16 (s, 1H), 8.04 (s, 2H), 7.49–7.38 (m, 6H), 7.30–7.17(m, 15H), 7.09 (s, 2H), 6.47 (s, 1H), 5.90 (m, 3H), 5.69 (t, *J* = 9.8 Hz, 1H), 5.63–5.45 (m, 5H), 5.39–5.20 (m, 9H), 4.37–4.10 (m, 9H), 3.34–3.13 (m, 2H), 2.51 (t, *J* = 6.5 Hz, 2H), 2.22 (s, 9H), 2.08–1.98 (m, 18H), 1.82 (s, 6H), 1.77 (s, 3H).

¹³C{¹H} NMR (75 MHz, CDCl₃) δ 170.1, 170.0, 169.9, 169.8, 169.4, 168.8, 168.5, 166.2, 151.6, 144.4, 143.8, 140.0, 130.0, 129.2, 127.7, 126.5, 122.9, 121.9, 107.1, 85.9, 85.5, 73.5, 70.8, 70.4, 67.8, 67.6, 66.6, 65.9, 62.7, 60.9, 38.7, 31.7, 20.3, 19.9.

IR (cm⁻¹) 2980, 1750, 1654, 1584, 1535, 1490, 1428, 1369, 1324, 1216, 1100, 1047, 953, 923, 733, 702.

HRMS (**ESI**⁺) m/z calc for C₇₉H₈₈N₁₀O₃₁S, 1727.5230 [M+Na]⁺; found, 1727.5256, 1705.5410 [M+H]⁺; found 1705.5452.



Figure S32. ¹H NMR spectrum of compound 11 (300 MHz, CDCl₃).



Figure S33. ¹³C{¹H} NMR spectrum of compound **11** (75 MHz, CDCl₃).



Figure S34. COSY spectrum of compound 11.



Figure S35. HRMS (ESI⁺) spectrum of compound 11.



Figure S36. IR spectrum of compound 11.



Synthesis of compound 12: To a stirring solution of 3% trifluoroacetic acid in DCM (1.5mL), was added a solution of 11 (300mg, 0.176mmol) in dichloromethane (1.5mL). Et₃SiH (28 μ L, 0.18mmol) was added to the orange solution. After 3 hrs, toluene (4mL) was added and the solvents were evaporated under vacuum. The addition of toluene and evaporation was repeated twice. The product 12 was isolated by quick flash chromatography using 3% MeOH in DCM in a 86% yield (221mg, 0.151mmol).

¹**H** NMR (300 MHz, CDCl₃) δ 8.15 (s, 1H), 8.04 (s, 2H), 7.16 (s, 2H), 6.91 (s, 1H), 5.91 (m, 3H), 5.67 (t, *J* = 9.8 Hz, 1H), 5.64–5.48 (m, 5H), 5.39–5.17 (m, 9H), 4.39–4.06 (m, 9H), 3.73–3.46 (m, 2H), 2.76 (dd, *J* = 14.9, 6.5 Hz, 2H), 2.22 (d, *J* = 2.3 Hz, 9H), 2.14–1.96 (m, 18H), 1.81 (d, *J* = 18.4 Hz, 9H), 1.48 (t, S*H*, *J* = 8.5 Hz, 1H).

¹³C{¹H} NMR (75 MHz, CDCl₃) δ 170.3, 169.9, 169.2, 168.8, 151.9, 144.2, 107.4, 86.3, 74.0, 70.7, 68.2, 66.8, 62.9, 61.1, 43.0, 24.5, 20.6, 20.2.

IR (cm⁻¹) 3649, 2980, 2888, 1754, 1382, 1249, 1153, 1079, 955.

HRMS (ESI⁺) m/z calc for C₆₀H₇₄N₁₀O₃₁S 1463.4315 [*M*+H]⁺; found, 1463.4317.



Figure S37. ¹H NMR spectrum of compound 12 (300 MHz, CDCl₃).







·8.0 ·8.5

1.5

2.0

Figure S40. HRMS (ESI⁺) spectrum of compound 12.



Figure S41. I.R spectrum of compound 12 (75 MHz, CDCl₃).

Synthesis of galactodendrimer 7 according to a convergent strategy

Compound 2 (3mg, 6µmol) and compound 12 (263mg, 0.180mmol, 5eq/alkene) were suspended in dioxane (1mL) in a 5 mL glass vial equipped with a small magnetic stirring bar. To this was added AIBN (1mg, 0.5µmol, 0.15eq/alkene), and the vial was tightly sealed with an aluminum/Teflon® crimp top. The mixture was then heated at 75°C for 5 hrs. After completion of the reaction, the vial was cooled to 25°C before it was opened. Dioxane was removed under vacuum. Purification of the crude compound was achieved *via* flash column chromatography on silica gel using 2-4% MeOH in DCM as eluent gradient to afford desired compound 7 (29.8mg, 3.22µmol) in 53% yield as a white solid. *Spectroscopic data for compound* 7 *obtained via convergent strategy are in full agreement with those of one originated from divergent approach.*



Synthesis of compound 15: β -D-Galactopyranose pentaacetate 13 (300mg, 0.769mmol) and monotosylated tetra(ethylene)glycol 14 (669mg, 1.92mmol) were mixed in dry DCM (5mL) and stirred for 1 hr with 4Å molecular sieves. The reaction mixture was then cooled to 0°C, followed by the addition of BF₃·Et₂O (660µL, 5.38mmol). The reaction mixture was stirred for 4 hrs at room temperature. Upon completion of reaction, the mixture was diluted with DCM (30mL), washed with water, saturated NaHCO₃ solution followed by brine. Drying over Na₂SO₄ and concentration under vacuum afforded crude compound that was purified by column chromatography (60% EtOAc in hexanes as eluent) to give 15 (287mg, 0.423mmol) colourless oil in 55% yield.

¹**H NMR** (300 MHz, CDCl₃) δ 7.81 (d, *J* = 8.2 Hz, 2H), 7.35 (d, *J* = 8.2 Hz, 2H), 5.39 (d, *J* = 3.3 Hz, 1H), 5.21 (dd, *J* = 10.4, 8.0 Hz, 1H), 5.02 (dd, *J* = 10.5, 3.4 Hz, 1H), 4.57 (d, *J* = 7.9 Hz, 1H), 4.15 (dt, *J* = 15.5, 7.8 Hz, 4H), 4.05 – 3.87 (m, 2H), 3.82–3.53 (m, 13H), 2.46 (s, 3H), 2.15 (s, 3H), 2.06 (d, *J* = 1.9 Hz, 6H), 1.99 (s, 3H).

¹³C{¹H} NMR (75 MHz, CDCl₃) δ 170.2, 170.1, 169.9, 169.3, 144.6, 132.8, 129.6, 127.7, 101.1, 77.2, 70.5, 70.4, 70.4, 70.3, 68.8, 66.9, 61.1, 21.4, 20.5, 20.5, 20.4.

IR (cm⁻¹) 2923, 1748, 1367, 1221, 1176, 1075.

HRMS (ESI⁺) m/z calc for C₂₉H₄₂O₁₆S, 701.2086 [*M*+Na]⁺; found, 701.2073.



Figure S42. ¹H NMR spectrum of compound 15 (300 MHz, CDCl₃).





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8.0 7.5 7.0 6.5 6.0 5.5

5.0 4.5 f2 (ppm)

•••

7.5

• • •

4.0 3.5 3.0 2.5 2.0

5.0

· 5.5 · 6.0 · 6.5 · 7.0

7.5

1.5



Figure S45. HRMS (ESI⁺) spectrum of compound 15.



Figure S46. IR spectrum of compound 15.



Synthesis of compound 16: Compound 15 (2.000g, 2.949mmol) and sodium azide (958mg, 15.0mmol) in dry DMF (20mL) were stirred at 80°C for 6 hrs. The solvent was evaporated under reduced pressure and crude was diluted with ethyl acetate (50mL), washed with water (2×30mL) and brine, dried over Na_2SO_4 and concenterated in *vacuo*. Column chromatography of the residual crude mixture was performed using 60% EtOAc in hexanes as eluent to give 16 (1.329g, 2.410mmol) as a yellowish oil in 82% yield.

¹**H NMR** (600 MHz, CDCl₃) δ 5.42–5.35 (m, 1H), 5.24–5.15 (m, 1H), 5.00 (ddd, J = 10.5, 3.4, 1.7 Hz, 1H), 4.56 (dd, J = 8.0, 1.6 Hz, 1H), 4.14 (dddd, J = 25.6, 11.3, 6.7, 1.5 Hz, 2H), 4.01–3.87 (m, 2H), 3.80–3.70 (m, 1H), 3.72–3.58 (m, 12H), 3.38 (t, J = 4.3 Hz, 2H), 2.13 (d, J = 1.7 Hz, 3H), 2.11–2.00 (m, 6H), 1.97 (d, J = 1.7 Hz, 3H).

¹³C{¹H} NMR (151 MHz, CDCl₃) δ 170.4, 170.2, 170.1, 169.5, 101.3, 70.9, 70.7, 70.6, 70.3, 70.0, 69.0, 68.8, 67.0, 61.3, 50.6, 20.7, 20.6, 20.6, 20.6.

IR (cm⁻¹) 2980, 2881, 2098, 1749, 1369, 1223, 1073.

HRMS (**ESI**⁺) m/z calc for C₂₂H₃₅N₃O₁₃, 567.2508 [*M*+NH₄]⁺; found, 567.2480, 588.1802 [*M*+K]⁺; 588.1789.



Figure S47. ¹H NMR spectrum of compound 16 (600 MHz, CDCl₃).



Figure S48. ¹³C{¹H} NMR spectrum of compound 16 (151 MHz, CDCl₃).



Figure S49. COSY NMR spectrum of compound 16.



Figure S50. HRMS (ESI⁺) spectrum of compound 16.



Figure S51. IR spectrum of compound 16.



Synthesis of compound 17: To a solution of per-O-acetylated lactose (β anomer,¹ 5.00g, 7.40mmol) and tetra(ethylene)glycol monotosylate (synthesized as previously described,² 8.10g, 22.1mmol) in dry DCM (60mL) under a nitrogen atmosphere and at 0°C was added dropwise BF₃·Et₂O (2.7mL, 22.1mmol) over a 15 minutes period. After stirring overnight (12 hrs) at r.t., the solvent was removed and EtOAc was added, then the solution was washed successively with NaHCO₃ (40mL), water (40mL) and brine (40mL). The organic phase was then dried over $MgSO_4$ and concentrated under reduced pressure. The crude residue was directly re-dissolved in DMF (70mL) under a nitrogen atmosphere and sodium azide (962mg, 14.8mmol) together with sodium iodide (11.1mg, 0.11mmol) were added. After stirring overnight (16 h) at 70°C, the solvent was removed and EtOAc (100mL) was added, then the solution was washed successively with water (4×40mL) and brine (3×50mL). The organic phase was then dried over MgSO₄ and concentrated under reduced pressure. After a short flash column chromatography on silica (EtOAc/Hexanes 6:4 to 8:2), the crude was subjected to de-O-acetylation protocol and dissolved in MeOH (40mL). To this solution was slowly added 1M MeONa/MeOH to adjust the pH 9-10. Reaction mixture was left for stirring overnight. The reaction pH was then adjusted with H⁺ resin to adjust pH to 6. Solvent was evaporated and the residue was benzoylated with benzoyl chloride (20.8g, 17.2mL, 148mmol) in pyridine (50mL) for overnight stirring at room temperature. Upon completion solvents were removed and reaction mixture was dissolved in DCM (100mL) and washed with 0.1N HCl (3×50 mL) followed by saturated solution of NaHCO₃ (3×75mL) and finally with brine. Organic layer was then dried with anhydrous sodium sulphate filtered and evaporated. Crude compound was then purified with the help of column chromatography using Hexane/Ethyl Acetate (1:1) as eluent. Desired compound 17 (5.93g, 4.66mmol) was isolated in a 63% overall yield as a yellow oil.

¹**H** NMR (**300** MHz, CDCl₃) δ 8.12–7.94 (m, 10H), 7.94–7.87 (m, 2H), 7.77–7.69 (m, 2H), 7.69–7.29 (m, 17H), 7.18 (m, 4H), 5.87–5.68 (m, 3H), 5.53–5.41 (m, 1H), 5.37 (dd, *J* = 10.3, 3.4 Hz, 1H), 4.86 (dd, *J* = 10.8, 7.9 Hz, 2H), 4.61 (d, *J* = 11.1 Hz, 1H), 4.49 (dd, *J* = 12.1, 4.0 Hz, 1H), 4.26 (t, *J* = 9.5 Hz, 1H), 3.87 (dd, *J* = 13.3, 8.1 Hz, 3H), 3.69 (ddd, *J* = 14.0, 10.4, 5.0 Hz, 5H), 3.62–3.48 (m, 6H), 3.47–3.33 (m, 6H).

¹³C NMR (75 MHz, CDCl₃) δ 165.8, 165.5, 165.4, 165.2, 165.1, 164.8, 133.5, 133.4, 133.2, 133.7, 130.0, 129.8, 129.7, 129.6, 129.6, 129.5, 129.4, 128.8, 128.7, 128.6, 128.5, 128.5, 128.31, 128.20, 101.2, 101.0, 76.0, 72.9, 72.9, 71.8, 70.6, 70.5, 70.5, 70.3, 69.9, 69.9, 69.2, 67.5, 62.4, 61.0, 50.6.

IR (cm⁻¹) 2980, 2883, 2104, 1728, 1601, 1451, 1314, 1267, 1176, 1094, 1069, 1027, 709. **HRMS** (**ESI**⁺) m/z calc for C₆₉H₆₅N₃O₂₁, 1294.4003 [M+Na]⁺; found, 1294.4031

¹ Wolfrom, M. L., Thompson, A. Methods. Carbohydr. Chem. 1963, 211-215.

² Zhang, L.; Sun, L.; Cui, Z.; Gottlieb, R. L.; Zhang, B. *Bioconjugate Chem.* 2001, 12, 939-948.



Figure S52. ¹H NMR spectrum of compound 17 (300 MHz, CDCl₃).



Figure S53. ¹³C{¹H} NMR spectrum of compound 17 (300 MHz, CDCl₃).



Figure S54. COSY spectrum of compound 17 (300 MHz, CDCl₃).



Figure S55. HRMS (ESI $^+$) spectrum of compound 17.



Figure S56. IR spectrum of compound 17.



Synthesis of compound 19a: To a solution of azide terminated compound 16 (100mg, 0.181mmol) in THF (4mL), was added propargyl alcohol (0.021ml, 0.363mmol), followed by sodium ascorbate (36mg, 0.181mmol). An aqueous solution of $CuSO_4 \cdot 5H_2O$ (45mg, 0.181mmol) was added and the final ratio of H₂O to THF was kept 1:1. The reaction mixture was stirred at 40°C for 12 hrs. Progress of the reaction was monitored with the help of TLC. Upon completion, reaction mixture was diluted with EtOAc (25mL) and washed with a saturated solution of EDTA (2×15mL). Organic layer was washed with brine solution, dried with anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. Purification of the crude was achieved *via* flash coloumn chromatography on silica gel using 2% MeOH in DCM as eluent to afford acetylated compound 19a (93.2mg, 0.154mmol) in a 85% yield as a yellowish oil.

¹**H** NMR (300 MHz, CDCl₃) δ 7.79 (s, 1H), 5.38 (d, *J* = 3.2 Hz, 1H), 5.19 (dd, *J* = 10.4, 7.9 Hz, 1H), 5.02 (dd, *J* = 10.5, 3.4 Hz, 1H), 4.79 (d, *J* = 5.0 Hz, 2H), 4.55 (dd, *J* = 6.3, 4.9 Hz, 3H), 4.25–4.06 (m, 2H), 4.04–3.82 (m, 4H), 3.78–3.51 (m, 11H), 2.14 (s, 3H), 2.04 (s, 6H), 1.98 (s, 3H).

¹³C{¹H} NMR (75 MHz, CDCl₃) δ 170.4, 170.2, 170.2, 169.3, 70.8, 70.7, 70.6, 70.5, 70.5, 70.4, 70.2, 69.4, 69.1, 68.8, 67.0, 61.2, 56.6, 50.2, 20.7, 20.7, 20.6, 20.6.

IR (cm⁻¹): 3478, 2881, 1744, 1433, 1369, 1219, 1175, 1047, 954.

HRMS (**ESI**⁺) m/z calc for C₂₅H₃₉N₃O₁₄ 606.2505 [*M*+H]⁺; found, 606.2501.



Figure S57. ¹H NMR spectrum of compound **19a** (300 MHz, CDCl₃).



Figure S58. $^{13}C{^{1}H}$ NMR spectrum of compound 19a (75 MHz, CDCl₃).



Figure S59. COSY spectrum of compound 19a.





Figure S61. IR spectrum of compound 19a.



Synthesis of compound 19: To a stirring solution of compound 19a (100mg, 0.165mmol) in MeOH (3mL) was slowly added 1M MeONa/MeOH solution to adjust the pH to 9-10. Reaction mixture was left to stir overnight. The reaction pH was then adjusted to 6 with H^+ resin. The solvent was evaporated and the residue was dissolved in 3mL of water, and washed with diethyl ether (3×15mL) to remove impurities. Aqueous layer was lyophilized to provide 19 (65mg, 0.149mmol) as a white solid in a 90% yield.

¹**H** NMR (300 MHz, CD₃OD) δ 8.11 (br s, 1H), 4.74 (s, 2H), 4.63 (d, *J* = 4.7 Hz, 2H), 4.27 (d, *J* = 7.0 Hz, 1H), 4.09–3.97 (m, 1H), 3.93 (t, *J* = 5.0 Hz, 2H), 3.84 (d, *J* = 2.2 Hz, 1H), 3.79–3.57 (m, 13H), 3.57–3.44 (m, 3H). ¹³C{¹H} NMR (75 MHz, CD₃OD) δ 105.0, 76.7, 74.8, 72.5, 71.4, 70.3, 70.2, 69.6, 62.5, 51.8. IR (cm⁻¹) 3358, 2924, 2502, 1643, 1455, 1073.

HRMS (ESI⁺) m/z calc for C₁₇H₃₁N₃O₁₀, 438.2082 [*M*+H]⁺; found, 438.2107.



Figure S62. ¹H NMR spectrum of compound 19 (300 MHz, CD₃OD).



Figure S64. COSY spectrum of compound 19.



Figure S65. HRMS (ESI⁺) of compound 19.



Figure S66. IR spectrum of compound 19.



Synthesis of compound 20a: To a solution of per-*O*-acetylated galactose (100mg, 119µmol) in a 1:1 mixture of H_2O/THF_{anh} (5 mL), were added propargyl alcohol (29.1µL, 501µmol), CuSO₄·5H₂O (14.9mg, 59.7µmol) and sodium ascorbate (11.8mg, 59.7µmol). While stirring, the mixture was first heated at 50°C for 3 hrs and at room temperature for additional 18 hours. Ethyl acetate (15mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10mL), washed with saturated aqueous NH₄Cl (2×10mL), water (10mL) and brine (5mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 92:8) afforded desired multivalent compound **20a** (86.0mg, 96.6µmol) as a white foam in a 91% yield.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 7.75 (s, 1H), 5.29 (d_{app}, 1H), 5.14 (dd, ³*J*_{4,3} = 9.4 Hz, ³*J*_{3,2} = 9.1 Hz, 1H), 5.05 (dd, ³*J*_{2,1} = 10.5 Hz, ³*J*_{3,2} = 8.0 Hz, 1H), 4.93 (dd, ³*J*_{2,3} = 10.5 Hz, ³*J*_{3,4} = 3.4 Hz, 1H), 4.83 (dd, ³*J*_{2,1} = 9.4 Hz, ³*J*_{3,2} = 8.0 Hz, 1H), 4.73 (br s, 2H), 4.53 (d, ³*J*_{1,2} = 9.4 Hz, 1H), 4.50 (t_{app}, 2H), 4.48 (dd, ²*J*_{6a.6b} = 12.0 Hz, ³*J*_{5.6a} = 2.1 Hz, 1H), 4.47 (d, ³*J*_{1,2} = 7.9 Hz, 1H), 4.12–4.00 (m, 3H), 3.90-3.52 (m, 17H), 3.30 (br s, 1H), 2.15 (s, 3H), 2.12 (s, 3H), 2.06 (s, 3H), 2.04 (3s, 9H), 1.96 (s, 3H).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.2, 170.0, 169.9, 169.7, 169.6, 169.0, 147., 122.9, 100.9, 100.4, 76.6, 72.6, 72.5, 71.5, 70.8, 70.5, 70.3, 70.3, 70.2, 70.2, 69.3, 69.0, 68.9, 66.5, 61.8, 60.7, 56.3, 50.0, 20.8, 20.8, 20.7, 20.6, 20.6, 20.6, 20.5.

HRMS (**ESI**⁺) m/z for C₃₇H₅₅N₃O₂₂, 894.3350 [M+H]⁺; found 894.3361, 916.3169 [M+Na]⁺; found 916.3181.



Figure S67. ¹H NMR spectrum of compound 20a (600 MHz, CDCl₃).



Figure S68. COSY spectrum of compound 20a.



43



Figure S70. HRMS (ESI⁺) of compound 20a.



Synthesis of compound 20: Acetylated compound **20a** (86.0mg, 96.6µmol) was dissolved in dry MeOH (4mL) and a solution of sodium methoxide (1M in MeOH, 150 µL) was added until pH 9-10. The reaction mixture was stirred at room temperature for 24 hrs. The pH was adjusted to 6-7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected reference **20** as a white solid (52.5mg, 87.6µmol) in a 91% yield.

¹**H NMR** (300 MHz, D₂O, δ ppm): 8.03 (s, 1H), 4.73 (s, 2H), 4.64 (t, J = 5.0 Hz, 2H), 4.52 (d, J = 7.9 Hz, 1H), 4.46 (d, J = 7.7 Hz, 1H), 4.08-3.53 (m, 25H), 3.36 (m, 6H).

¹³**C NMR** (75 MHz, D₂O, δ ppm): 147.5, 125.1, 103.6, 102.7, 79.0, 76.0, 75.4, 75.0, 73.5, 73.2 (, 71.6, 70.3, 70.2, 70.1, 70.1, 69.4, 69.2, 61.7, 60.7, 55.3, 50.7.

HRMS (ESI⁺) m/z for C₂₃H₄₁N₃O₁₅,600.2610 [M+H]⁺; found 600.2618, 622.2430 [M+Na]⁺; found 622.2438.



Figure S71. ¹H NMR spectrum of compound **20** (300 MHz, D_2O).



Figure S72. COSY spectrum of compound 20.



Figure S73. ¹³C{¹H} NMR spectrum of compound 20 (75 MHz, CDCl₃).





MS Spectrum Peak List

Ion	Formula	Abund	Observed m/z	Calc m/z	Diff (ppm)
(M+H)+	C23H42N3O15	393220.81	600.26181	600.26104	1.28
(M+Na)+	C23H41N3NaO15	1074960.28	622.24379	622.24299	1.28

Figure S74. HRMS (ESI⁺) of compound 20.



Synthesis of compound 22: To a solution of propargylated scaffold **5** (55mg, 0.022mmol) in THF (5mL) was added galactoside **16** (354mg, 0.645mmol), followed by sodium ascorbate (68mg, 0.39mmol). An aqueous solution of $CuSO_4 \cdot 5H_2O$ (96mg, 0.39mmol) was added and the final ratio of H₂O to THF was kept 1:1. The reaction mixture was stirred at 40°C for 24 hrs. The progress of the reaction was monitored with the help of TLC. Upon completion, the reaction mixture was diluted with EtOAc (25mL) and washed with saturated solution of EDTA (2×15mL). Organic layer was washed with brine solution, dried with anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. Purification of the crude compound was achieved *via* flash column chromatography on silica gel using 5% MeOH in DCM as eluent to afford the desired compound **22** (208mg, 0.0167mmol)in 76% yield.

¹**H** NMR (600 MHz, CDCl₃) δ 7.92 (s,12H), 7.84 (s, 6H), 7.25-7.230 (m, 12H), 5.40–5.31 (m, 18H), 5.30–5.27 (m, 7H), 5.19–5.07 (m, 53H), 4.99 (dd, *J* = 10.5, 3.4 Hz, 19H), 4.52 (dd, *J* = 22.2, 19.3 Hz, 57H), 4.18–4.04 (m, 36H), 3.97–3.77 (m, 75H), 3.70 (ddd, *J* = 10.8, 6.9, 3.9 Hz,19H), 3.63–3.50 (m, 176H), 3.45 (br s, 18H), 3.34 (s, 18H), 2.75 (br s, 1 2H), 2.63 (br s, 12H), 2.12 (d, *J* = 1.4 Hz, 52H), 2.04–1.93 (m, 164H), 1.83 (s, 12H).

¹³C{¹H} NMR (151 MHz, CDCl₃) δ 170.2, 170.0, 170.0, 170.0, 169.3, 166.7, 152.0, 144.0, 143.2, 140.1, 130.1, 124.7, 124.4, 107.3, 101.2, 70.8, 70.6, 70.5, 70.5, 70.4, 70.4, 70.1, 69.7, 69.3, 69.2, 69.0, 68.8, 67.1, 66.3, 63.0, 61.2, 50.2, 39.6, 31.4, 29.7, 28.6.

IR (cm⁻¹) 2872, 1747, 1491, 1427, 1368, 1325, 1221, 1175, 1104, 1050, 732.

MALDI-TOF (DHB matrix) *m*/*z* calc. for C₅₃₂H₇₇₈N₆₀O₂₆₅S₆, 12446.5; found, 12446.0. **GPC** *Mn*= 12500 g/mol. *Mw/Mn*= 1.06.





Figure S76. ¹³C{¹H} NMR spectrum of compound 22 (151 MHz, CDCl₃).



Figure S77. COSY spectrum of compound 22.



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Figure S78. MALDI TOF spectrum of compound 22.



Figure S79. GPC traces of compound 22.



Figure S80. IR spectrum of compound 22.



Synthesis of compound 23: To a stirring solution of compound 22 (100mg, 0.080mmol) in MeOH (3mL), was slowly added a 1M solution of MeONa/MeOH to adjust the pH to 9-10. The reaction mixture was left stirring overnight. The reaction pH was then adjusted to 6 with H^+ resin. Solvent was evaporated and the residue was dissolved in 3 mL of water and washed with diethyl ether (3x15ml) to remove impurities. Aqueous layer was finally lyophilized to yield 23 (68mg, 0.072mmol) as a white solid in 90% yield.

¹**H** NMR (600 MHz, D_2O) δ 8.40–7.80 (m, 18H), 7.28 (br s, 12H), 5.30-5.00 (m, 30H), 4.62–4.49 (m, 36H), 4.38 (d, J = 8.0 Hz, 15H), 4.02 (d, J = 11.4 Hz, 18H), 3.93 (d, J = 13.8 Hz, 39H), 3.88–3.82 (m, 12H), 3.80-3.71 (m, 54H), 3.69–3.61 (m, 75H), 3.62–3.49 (m, 165 H), 3.45–3.38 (m, 16H), 3.37–3.35 (m, 100H), 3.28–3.22 (m, 9H), 2.86–2.70 (m, 12H), 2.68–2.49 (m, 12H), 1.85–1.65 (m, 12H).

¹³C{¹H} NMR (151 MHz, D_2O) δ 168.8, 152.7, 140.1, 130.7, 107.8, 103.9, 76.1, 73.7, 73.0, 71.7, 70.7, 70.6, 70.5, 70.5, 69.7, 69.6, 69.5, 63.5, 61.9, 61.4, 51.2, 40.5, 31.7, 29.9, 29.1 (*C* and *CH* of triazole rings not visible).

IR (cm⁻¹) 3377, 2917, 1653, 1586, 1495, 1239, 1104.

MS (**ESI**⁺) m/z calc for C₃₈₈H₆₃₄N₆₀O₁₉₃S₆, 9420.9 [M+H]⁺; found (deconvoluted), 9414.0.





Figure S82. $^{13}C{^{1}H}$ NMR spectrum of compound 23 (151 MHz, D₂O).



Figure S83. COSY spectrum of compound 23.







Figure S84. HRMS (ESI^+) spectrum of compound **23**.



Figure S85. IR spectrum of compound 23.



Synthesis of compound 24: To a solution of propargylated scaffold 5 (20mg, 7.8µmol) in THF (5mL) was added PEGylated lactoside 17 (302mg, 0.234mmol), followed by sodium ascorbate (28mg, 0.14mmol). An aqueous solution of $CuSO_4 \cdot 5H_2O$ (35mg, 0.14mmol) was added and the final ratio of H₂O to THF was kept 1:1. The reaction mixture was stirred at 40°C for 24 hrs. The progress of the reaction was monitored by TLC. Upon completion, the reaction mixture was diluted with EtOAc (25mL) and washed with sat. solution of EDTA (2×15mL). Organic layer was washed with brine solution, dried with anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. Purification of the residue was achieved *via* flash coloumn chromatography on silica gel using 5% MeOH in DCM as eluent to afford desired compound 24 (153mg, 6.0µmol) as an off-white viscous solid in 77% yield.

¹**H** NMR (300 MHz, CDCl₃) δ 8.06–7.14 (m, 666H), 5.83–5.67 (m, 54H), 5.49–5.34 (m, 36H), 5.11 (s, 32H), 4.88 (d, *J* = 7.9 Hz, 18H), 4.80 (d, *J* = 7.8 Hz, 18H), 4.65–4.55 (m, 18H), 4.53–4.38 (m, 54H), 4.26 (t, *J* = 9.4 Hz, 18H), 3.95–3.20 (m, 368H), 2.85–2.50 (m, 24H), 1.82 (br s, 12H).

¹³C{¹H} NMR (151 MHz, CDCl₃) δ 166.7, 165.8, 165.5, 165.4, 165.2, 165.1, 164.8, 152.0, 144.6, 140.0, 133.5, 130.2, 133.4, 133.3, 129.7, 129.6, 129.5, 129.3, 128.6, 128.6, 128.5, 128.4, 128.2, 107.9, 101.2, 101.0, 76.0, 75.7, 70.5, 69.9, 69.6, 67.5, 62.4, 61.0, 60.3, 57.3, 45.6, 39.2, 31.6, 29.8, 28.4 (Signals corresponding to CONH, C_{ar} H, CH₂ and C_{q} (*in italic*) located in inner positions are not visible in this case, even after 30000 scans).

IR (cm⁻¹): 2879, 1730, 1601, 1584, 1451, 1369, 1315, 1269, 1250, 1175, 1069, 1026, and 708. **MALDI-TOF** m/z calc. for C₁₃₇₈H₁₃₁₈N₆₀O₄₀₉S₆, 25478.7 [M+Na]⁺; found, 25602.3.



Figure S86. ¹H NMR spectrum of compound 24 (300 MHz, CDCl₃).



Figure S87. ¹³C{¹H} NMR spectrum of compound 24 (151 MHz, CDCl₃).



Bruker Daltonics flexAnalysis Figure S88. MALDI TOF spectrum of compound 24.



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Figure S89. IR spectrum of compound 24.



Synthesis of compound 25: To a stirring solution of compound 24 (100mg, 3.93μ mol) in MeOH (3mL), was slowly added a 1M solution of MeONa/MeOH to adjust the pH to 9-10. The reaction mixture was left stirring overnight. The reaction pH was adjusted to 6 with H⁺ resin and the solvent was evaporated. The residue was dissolved in 3mL of water and washed with diethyl ether (3×15ml) to remove the impurities. Aqueous layer was lyophilized to afford 25 (41mg, 3.3µmol) as a white solid in a 85% yield.

¹**H** NMR (600 MHz, D_2O) δ 8.16–7.70 (m, 18H), 7.25–71 (m, 12H), 5.15–4.83 (m, 18H), 4.56–4.20 (m, 36H), 4.37–4.20 (m, 67H), 3.86–3.06 (m, 627H), 2.67–2.47 (br s, 12H), 2.45–2.30 (br s, 12H), 1.75–1.45 (br s, 12H).

¹³C{¹H} NMR (151 MHz, D₂O) δ: 168.5, 152.3, 150.4, 139.7, 130.3, 126.4, 107.4, 103.6, 102.8, 79.1, 75.9, 75.3, 74.9, 73.4, 73.1, 72.6, 71.5, 70.2, 70.1, 70.1, 69.2, 69.1, 63.0, 61.6, 60.7, 51.0, 45.9, 40.0, 31.3, 29.4, 28.6.

IR (cm⁻¹): 3695, 3384, 2937, 2843, 1646, 1429, 1347, 1055, and 1032.

MALDI-TOF m/z calc for C₄₉₆H₈₁₄N₆₀O₂₈₃S₆, 12361.4 [M+Na]⁺; found, 12368.4, 12401.9 (Cu adduct); found 12403.5.



Figure S90. ¹H NMR spectrum of compound 25 (600 MHz, D_2O).





Figure S92. COSY spectrum of compound 25.





Figure S93. Top: MALDI TOF spectrum of compound 25, bottom: MALDI-TOF spectrum of compound 25 (Positive mode with Cu)



Figure S94. IR spectrum of compound 25.

4. Surface plasmon resonance (SPR) studies:

The studies were conducted using a Biacore T200 SPR instrument with a CM5 sensor chip. A continuous flow of HEPES buffer (10 mm HEPES and 150 mm NaCl, 2 mM CaCl₂, pH 7.4) was maintained over the sensor surface at a flow rate of 10 µl/min. The CM5 sensor chip was activated with an injection of a solution containing N-ethyl-N'-(3-diethylaminopropyl) carbodiimide (EDC) (0.2M) and N-hydroxysuccinimide (NHS) (0.05M) for 7 minutes. Lactoside **21** (200 μ g/mL) and Et₃N (1 mM) in NaOAc buffer (pH 4.5) was injected over the activated flow cell at flow rate of 10 µl/min for 2 minute to achieve a ~230 RU immobilization. The immobilization procedure was completed by an injection of ethanolamine hydrochloride (1M) (70 μ L), followed by a flow of the buffer (100 μ L/min.), in order to eliminate physically adsorbed compounds. Ethanol amine alone was used in one of the flow-cell as a reference. The solutions of pre-incubated (1 hr) mixtures of glycodendrimers or monomers (with the various concentrations) and a PA-IL lectin (1.5 μ M) in running HEPES buffer are passed over flow cells of the galactoside and ethanol amine (Association: 3 min and dissociation: 3 min). The sensor chip was regenerated with the serial injections of D-lactose (0.25 M, 3 min), buffer (3 min), Dlactose (0.25 M, 3 min) and buffer (3 min). For each inhibition assay, PA-IL lectin (1.5 μ M) without inhibitor was injected to observe the full adhesion of the lectin onto the sugar-coated surface (0% inhibition). Response units from the surface of lactoside were subtracted from the surface of ethanol amine to eliminate non-specific interactions, as well as, bulk change in RU due to variation in refractive index of the medium. The primary subtracted sensorgrams were analyzed by 1:1 Langmuir model fitting, using the BIAevaluation software. For IC₅₀ evaluation, the response units at the equilibrium was considered as the amount of lectin bound to the sugar surface in the presence of a defined concentration of inhibitor. Inhibition curves were obtained by plotting the percentage of inhibition against the inhibitor concentration (on a logarithmic scale) by using Origin 7.0 software (OriginLab Corp.) and IC₅₀ values were extracted from a sigmoidal fit of the inhibition curve.

5. SPR Sensorgram:



Figure S95. (a) Sensorgrams obtained by injection of PA-IL (1.5 μ M) incubated with different concentrations of **18** varying from 10 μ M (top curve) to 320 μ M (bottom curve) on the surface of immobilized lactoside **21**. (b) The inhibitory curve for the **18**. IC₅₀ value was extracted from the sigmoidal fit of the inhibition curve.



Figure S96. (a) Sensorgrams obtained by injection of PA-IL (1.5 μ M) incubated with different concentrations of dendrimer **8** varying from 0.062 μ M (top curve) to 1 μ M (bottom curve) on the surface of immobilized lactoside **21**. (b) The inhibitory curve for the dendrimer **8**. IC₅₀ value was extracted from the sigmoidal fit of the inhibition curve.



Figure S97. (a) Sensorgrams obtained by injection of PA-IL (1.5 μ M) incubated with different concentrations of **19** varying from 1.87 μ M (top curve) to 120 μ M (bottom curve) on the surface of immobilized lactoside **21**. (b) The inhibitory curve for the **19**. IC50 value was extracted from the sigmoidal fit of the inhibition curve.



Figure S98. (a) Sensorgrams obtained by injection of PA-IL (1.5 μ M) incubated with different concentrations of glycodendrimer 23 varying from 0.005 μ M (top curve) to 0.16 μ M (bottom curve) on the surface of immobilized lactoside 21. (b) The inhibitory curve for the glycodendrimer 23. IC₅₀ value was extracted from the sigmoidal fit of the inhibition curve.



Figure S99. (a) Sensorgrams obtained by injection of PA-IL (1.5 μ M) incubated with different concentrations of **20** varying from 18 μ M (top curve) to 4.60 mM (bottom curve) on the surface of immobilized lactoside **21**. (b) The inhibitory curve for 20. IC50 value was extracted from the sigmoidal fit of the inhibition curve.



Figure S100. (a) Sensorgrams obtained by injection of PA-IL (1.5 μ M) incubated with different concentrations of dendrimer **25** varying from 0.5 μ M (top curve) to 16 μ M (bottom curve) on the surface of immobilized lactoside **21**. (b) The inhibitory curve for the dendrimer **25**. IC50 value was extracted from the sigmoidal fit of the inhibition curve.