

## Supporting Information

# Highly Sensitive and Selective Detection of miRNA: DNase I-Assisted Target Recycling using DNA Probes Protected by Polydopamine Nanospheres

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## Experimental Section

### Materials and Reagents

Dopamine hydrochloride was purchased from Sangon Biotechnology Co., Ltd (Shanghai, China). DNase I, HPLC-purified RNA, RNase inhibitor, and DEPC-treated water were purchased from Takara Biotechnology Co. Ltd. (Dalian, China). The DNA probes were synthesized on a PolyGen Column 12 DNA synthesizer and the reagents were purchased from Glen Research (Sterling, VA, USA). Cell lysate from breast cancer cell MDA-MB-231 was obtained by the reported procedure adding 5mM MgCl<sub>2</sub> and 5mM CaCl<sub>2</sub>.<sup>1</sup> All DNA/RNA sequences are listed in Tables S1. Polydopamine nanospheres (PDANSs) were synthesized using the reported procedure.<sup>2</sup>

### Fluorescence measurements

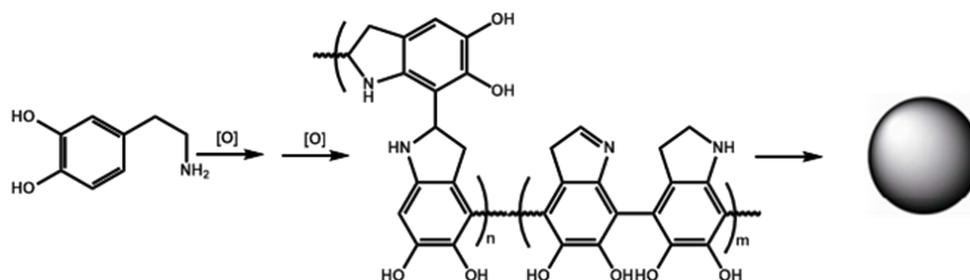
Fluorescence measurements were carried out on a FluoroMax-4P Fluorescence Spectrophotometer (Horiba, Japan). The excitation and emission wavelengths were set at 490 and 520 nm, respectively, with a 3 nm bandwidth and 0.3s integration time. The emission spectra were obtained by exciting the samples at 490 nm and scanning the emission from 500 to 650 nm in steps of 1 nm/s. The reaction buffer contained 20 mM Tris-HCl (pH 8.0), 6% PEG, 5 mM MgCl<sub>2</sub>, 5mM CaCl<sub>2</sub> and 50 mM NaCl. The amplified detection of miRNA was performed in 200 μL solution with 50 nM probe incubated with 0.09 mg/mL PDANSs for 30 min at 37°C. After the addition of 20 units of DNase I and different concentrations of miRNA, the solution was incubated at 37°C for 90 min and the fluorescence intensities were detected afterwards.

### Gel electrophoresis

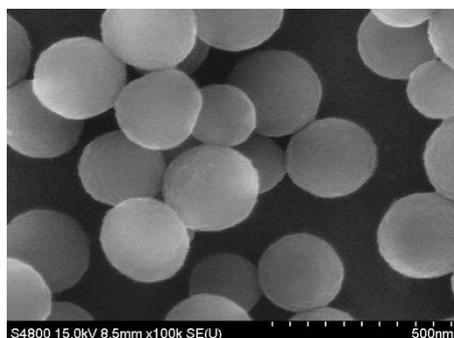
A 20% denaturing PAGE analysis of the products from the cyclic enzymatic amplification reaction was carried out in 1×TBE (pH 8.3) at 1W power for about 1.5hr. After Stains-All staining, gels were scanned.

**Table S1. Sequences of oligonucleotides used in this study**

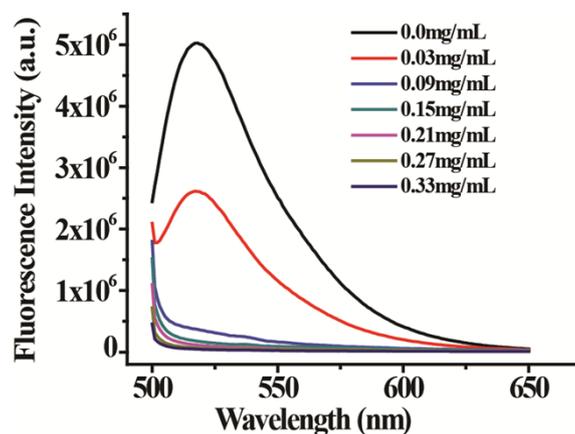
Name	Sequence
Probe	5'- AAC TAT ACA ACC TAC TAC CTC A -FAM-3'
Let-7a	5' - UGA GGU AGU AGG UUG UAU AGU U -3'
Let-7f	5'-UGA GGU AGU AGA UUG UAU AGU U-3'
Let-7i	5'-UGA GGU AGU AGU UUG UGC UGU U-3'
Mir122	5'- AAC GCC AUU AUC ACA CUA AAU A -3'



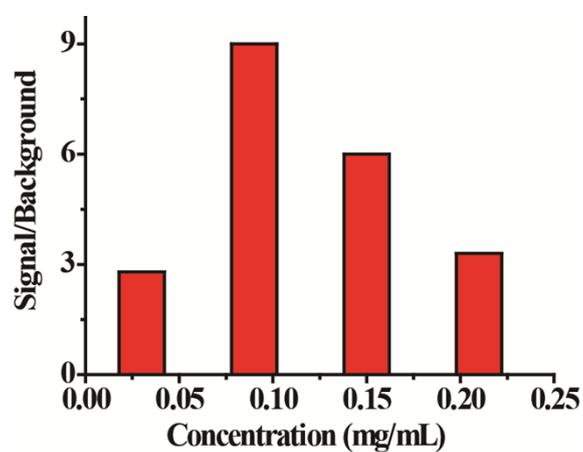
**Figure S1.** Simplified schematic of the oxidative polymerization of dopamine to prepare polydopamine nanospheres.



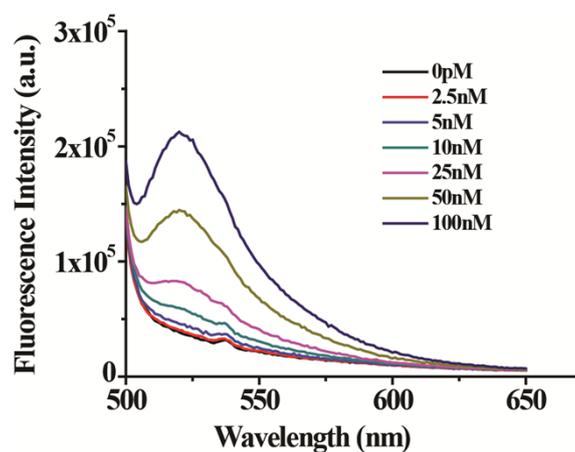
**Figure S2.** SEM image of PDANSs, the diameter is approximately 250 nm.



**Figure S3.** Fluorescence quenching of the DNA probe by different concentrations of PDANS.



**Figure S4.** Signal-to-background ratio upon addition of different concentrations of PDANS.



**Figure S5.** Response of the probe to different concentration of miRNA target in the absence of DNaseI.

**Table S2. Comparison of different analysis assays for miRNA**

	Sensitivity	Selectivity	Time	Conjugation	Biocompatibility	Ref
<b>Gold Nanoparticle</b>	5-8 pM	N.D.	5 hr	Au-Thiol	good	[3]
<b>Graphene oxide</b>	9 pM	Single-base mismatch	20 min	Absorption	Moderate <sup>4,5</sup>	[6]
<b>PDANS</b>	2.3 pM	Single-base mismatch	1.5 hr	Absorption	Excellent	This work

**References:**

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3. F. Degliangeli, P. Kshirsagar, V. Brunetti, P. P. Pompa and R. Fiammengo, *J. Am. Chem. Soc.*, 2014, **136**, 2264-2267.
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