Ditopic CMPO-pillar[5]arenes as unique receptors for efficient separation of americium(III) and europium(III)

Yuyu Fang,^a Xiangyang Yuan,^a Lei Wu,^a Zhiyong Peng,^a Wen Feng,^{*a}

Ning Liu,^a Dingguo Xu,^a Shoujian Li,^a Arijit Sengupta,^b Prasanta K.

Mohapatra^b and Lihua Yuan*a

 ^a Institute of Nuclear Science and Technology, Key Laboratory for Radiation Physics and Technology of Ministry of Education, College of Chemistry, Sichuan University, Chengdu 610064, China. Fax: +86-28-85418755; Tel: +86-28-85412890; E-mail: wfeng9510@scu.edu.cn; Ihyuan@scu.edu.cn
 ^b Radiochemistry Division, Bhabha Atomic Research Centre, Trombay, Mumbai-400085, India

Supporting Information

Contents

1. General information	S3
2. Synthesis and characterization	S4
3. NMR and ESI-HRMS spectra	S8
4. ESI-HRMS for complexes	S18
5. NMR titration of 1 with La ³⁺	S21
6. Fluorescencetitration of Eu ³⁺ with 1	S24
7. Job's plot of 1 and Eu ³⁺	S26
8. UV-vis spectrophotometric titration of 1 with Eu ³⁺	S27
9. FT-IR spectra of free and Eu ³⁺ -loaded 1	S29
10. Extraction properties of Am(III) and Eu(III)	S31
11. Fluorescence studies	S33
12. References	S36

1. General information

The ¹H NMR and ¹³C NMR spectra were recorded on Bruker AVANCE AV II-400 MHz (¹H: 400 MHz; ¹³C: 100 MHz; ³¹P: 163 MHz). Chemical shifts are reported in δ values in ppm using tetramethlysilane (TMS) and coupling constants (J) are denoted in Hz. Multiplicities are denoted as follows: s = singlet, d = doublet, t = triplet, dd = double doublet and m = multiplet. High resolution mass (HRMS) data were obtained by WATERS Q-TOF Premier. Solvents for extraction and chromatography were reagent grade. CH₂Cl₂ was distilled from CaH₂. CDCl₃ and DMSO-*d*₆ were from Cambridge Isotope Laboratories (CIL)

2. Synthesis and characterization



Synthetic route

Scheme S1 Synthesis of CMPO-functionalized pillar[5]arenes **1a-c**. Reagents: (1) (CH₂O)_n, BF₃•OEt₂, CH₂Cl₂, r.t.; (2) NaN₃, DMF, 90 °C; (3) H₂, Pd/C, CH₃OH, 48 h, 50 °C; (4) **6**, CHCl₃, NEt₃, 48 h, reflux.

Synthesis of compound 3



The general procedure for compounds **3b-c** was exemplified by the synthesis of **3a**: To a solution of **2a** (3.03 g, 9.34 mmol) in dry dichloromethane (80 mL) was added paraformaldehyde (291 mg, 9.34 mmol) under nitrogen atmosphere. Then boron trifluoride diethyl etherate (1.32 g, 11.5 mmol) was added to the solution and the mixture was stirred at room temperature for 2 h. Water (100 mL) was added to quench the reaction. The organic layer was washed twice with H₂O (2 × 100 mL) and dried over anhydrous Na₂SO₄. The solvent was removed under reduced pressure. The obtained solid was purified by column chromatography on silica gel with petroleum ether/dichloromethane (1:1, v/v) as the eluent to afford a white powder.

3a^[S1]: Yield 41%. ¹H NMR (400 MHz, CDCl₃) δ 6.92 (s, 10 H), 4.23 (t, *J* = 5.5 Hz, 20 H), 3.84 (s, 10 H), 3.64 (t, *J* = 5.5 Hz, 20 H).

3b^[S2]: Yield 40%.¹H NMR (400 MHz, CDCl₃) δ 6.74 (s, 10 H), 3.99 (t, *J* = 5.7 Hz, 20 H), 3.76 (s, 10 H), 3.52 (t, *J* = 6.4 Hz, 20 H), 2.23 (m, 20 H).

3c^[S3]: Yield 36%. ¹H NMR (400 MHz, CDCl₃) δ 6.81 (s, 10 H), 3.93 (t, *J* = 5.8 Hz, 20 H), 3.75 (s, 10 H), 3.43 (t, *J* = 6.5 Hz, 20 H), 2.05 (m, 20 H), 1.92 (m, 20 H).

Synthesis of compound 4



The general procedure for compounds **4b-c** was exemplified by the synthesis of **4a**: To a solution of **3a** (3.03 g, 1.80 mmol) in dry DMF (80 mL) was added sodium azide (1.41 g, 21.6 mmol) under nitrogen atmosphere. The mixture was stirred at 90 °C for 12 h and cooled to room temperature. After slow addition of ice water to the solution, the precipitate was collected by filtration, and washed with water (2 × 100 mL) and methanol (2 × 100 mL) to afford a white solid.

4a^[S4]: Yield 97%. ¹H NMR (400 MHz, CDCl₃) δ 6.83 (s, 10 H), 4.01 (t, J = 4.8 Hz, 20 H), 3.84 (s, 10 H), 3.55 (t, J = 4.7 Hz, 20 H).

4b: Yield 98%. ¹H NMR (400 MHz, CDCl₃) δ 6.72 (s, 10 H), 3.91 (t, *J* = 5.9 Hz, 20 H), 3.75 (s, 10 H), 3.43 (t, *J* = 6.7 Hz, 20 H), 1.97 (m, 20 H).¹³C NMR (100 MHz, CDCl₃) δ 149.8, 128.6, 115.3, 65.5, 48.4, 29.8, 29.1. ESI-HRMS (m/z) calcd. for C₆₅H₈₀N₃₀O₁₀ [M+Na]⁺ 1463.6571; found [M+Na]⁺ 1463.6572.

4c: Yield 98%. ¹H NMR (400 MHz, CDCl₃) δ 6.81 (s, 10 H), 3.90 (t, J = 4.7 Hz, 20 H), 3.75 (s, 10 H), 3.34 (t, J = 6.3 Hz, 20 H), 1.82 (m, 40 H).¹³C NMR (100 MHz, CDCl₃) δ 149.8, 128.4, 115.1, 67.9, 51.3, 29.6, 27.1, 26.0. ESI-HRMS (m/z) calcd. for C₇₅H₁₀₀N₃₀O₁₀ [M+Na]⁺ 1063.8136; found [M+Na]⁺ 1063.8134.

Synthesis of compound 1



The general procedure for compounds **1b-c** was exemplified by the synthesis of **1a**: A mixture of **4a** (2.00 g, 1.54 mmol) and Pd/C (400 mg, 20% w.t.) in methanol was stirred at 50 °C under hydrogen atmosphere (0.4 MPa) for 48 h. After removal of the catalyst by filtration, the solvent was concentrated under reduced pressure to give the amine **5a** as a white solid which was used without further purification. The resulting solid **5a** was dissolved together with *p*-nitrophenyl(diphenylphosphoryl) acetate **6**^[S5] (8.81 g, 23.1 mmol) and triethylamine (2.34 g, 23.1 mmol) in ethanol-free dry chloroform (120 mL) at 60 °C for 48 h. Water (100 mL) was added to quench the reaction. The organic layer was washed repeatedly with 5% Na₂CO₃ (3 × 100 mL), water (3 × 100 mL) and dried over anhydrous Na₂SO₄. The solvent was removed under reduced pressure. The obtained pale yellow solid was washed with cold ethyl acetate (3 × 20 mL) and purified by column chromatography on silica gel with dichloromethane/methanol (15:1, v/v) as the eluent to afford a white powder.

1a: Yield 48%. ¹H NMR (400 MHz, CDCl₃) δ 8.08 (s, 10 H), 7.75 (m, 40 H), 7.38 (m, 60 H), 6.50 (s, 10 H), 3.59 (s, 10 H), 3.37-3.50 (m, 40 H), 3.28 (m, 20 H). ¹³C NMR (100 MHz, CDCl₃) δ 165.3, 149.9, 132.2, 131.0, 128.8, 128.7, 128.6, 115.9, 67.2, 39.5, 38.7, 29.3. ESI-HRMS (m/z) calcd. for C₁₉₅H₁₉₀O₃₀P₁₀N₁₀ [M+2Na]²⁺ 1754.5444; found [M+2Na]²⁺ 1754.5402.

1b: Yield 55%. ¹H NMR (400 MHz, CDCl₃) δ 8.03 (s, 10 H), 7.75 (dd, 40 H), 7.34 (m, 60 H), 6.60 (s, 10 H), 3.62 (t, *J* = 6.0 Hz, 20 H), 3.52 (m, 10 H), 3.41 (m, 30 H),

3.29 (m, 10 H), 1.77 (m, 20 H). ¹³C NMR (100 MHz, CDCl₃) δ 165.0, 149.7, 132.3, 130.9, 130.8, 128.8, 128.7, 115.0, 66.2, 39.5, 38.9, 37.3, 29.3. ESI-HRMS (m/z) calcd. for C₂₀₅H₂₁₀O₃₀P₁₀N₁₀ [M+2Na]²⁺ 1824.6226; found [M+2Na]²⁺ 1824.6232.

1c: Yield 67%. ¹H NMR (400 MHz, CDCl₃) δ 7.92 (s, 10 H), 7.76 (m, 40 H), 7.39 (m, 60 H), 6.69 (s, 10 H), 3.72 (m, 10 H), 3.67 (s, 10 H), 3.60 (m, 10 H), 3.18 (m, 20 H), 1.67 (m, 40 H). ¹³C NMR (100 MHz, CDCl₃) δ 164.9, 149.8, 132.2, 131.3, 130.9, 128.8, 128.2, 115.0, 67.9, 39.6, 38.8, 29.3, 27.1, 26.1. ESI-HRMS (m/z) calcd. for C₂₁₅H₂₃₀O₃₀P₁₀N₁₀ [M+2Na]²⁺ 1894.1992; found [M+2Na]²⁺ 1894.1949.

3. NMR and ESI-HRMS spectra







Figure S3. ¹H NMR spectrum (400 MHz, CDCl₃) of 3c at 298 K.



Figure S5. ¹H NMR spectrum (400 MHz, CDCl₃) of 4b at 298 K.



Figure S6. ¹³C NMR spectrum (100 MHz, CDCl₃) of 4b at 298 K.



Figure S7. ESI-HRMS spectrum of 4b.



Figure S9. ¹³C NMR spectrum (100 MHz, CDCl₃) of 4c at 298 K.



Figure S11. ¹H NMR spectrum (400 MHz, CDCl₃) of 1a at 298 K.



Figure S12. ¹³C NMR spectrum (100 MHz, CDCl₃) of 1a at 298 K.



Figure S13. ESI-HRMS spectrum of 1a.



S15







Figure S17. ¹H NMR spectrum (400 MHz, CDCl₃) of 1c at 298 K.



Figure S18. ¹³C NMR spectrum (100 MHz, CDCl₃) of 1c at 298 K.



Figure S19. ESI-HRMS spectrum of 1c.

4. ESI-HRMS for complexes



Figure S20. ESI-HRMS spectrum of the 1:1 (M:L) complex formed between **1a** and Eu³⁺ on addition of 1 equiv. of Eu(NO₃)₃ (inset: experimental isotope distribution (blue) and computer simulation (red)).



Figure S21. ESI-HRMS spectrum of the 2:1 (M:L) complex formed between **1a** and Eu³⁺ on addition of 3 equiv. of Eu(NO₃)₃ (inset: experimental isotope distribution (blue) and computer simulation (red)).



Figure S22. ESI-HRMS spectrum of the 1:1 (M:L) complex formed between **1b** and Eu³⁺ on addition of 1 equiv. of Eu(NO₃)₃ (inset: experimental isotope distribution (blue) and computer simulation (red)).



Figure S23. ESI-HRMS spectrum of the 2:1 (M:L) complex formed between 1b and Eu^{3+} on addition of 3 equiv. of $Eu(NO_3)_3$ (inset: experimental isotope distribution (blue) and computer simulation (red)).



(blue) and computer simulation (red)).



5. NMR titration of 1 with La³⁺





Figure S26. Titration of a **1a** solution (4.0 mM) by La^{3+} in CDCl₃/DMSO- d_6 (4:1, v/v) as monitored by ¹H NMR spectroscopy at 298 K.

Figure S27. Titration of a 1a solution (4.0 mM) by La³⁺ in CDCl₃/DMSO- d_6 (4:1, v/v)

as monitored by ³¹P NMR spectroscopy at 298 K.





Figure S28. Titration of a **1b** solution (4.0 mM) by La^{3+} in CDCl₃/DMSO- d_6 (4:1, v/v) as monitored by ¹H NMR spectroscopy at 298 K.

Figure S29. Titration of a **1b** solution (4.0 mM) by La³⁺ in CDCl₃/DMSO- d_6 (4:1, v/v) as monitored by ³¹P NMR spectroscopy at 298 K.



S23



Figure S30. Titration of a **1c** solution (4.0 mM) by La^{3+} in CDCl₃/DMSO- d_6 (4:1, v/v) as monitored by ¹H NMR spectroscopy at 298 K.

Figure S31. Titration of a **1c** solution (4.0 mM) by La^{3+} in CDCl₃/DMSO- d_6 (4:1, v/v) as monitored by ³¹P NMR spectroscopy at 298 K.

6. Fluorescencetitration of Eu³⁺ with 1



Figure S32. (a) Fluorescence titration of $Eu(NO_3)_3$ (2 × 10⁻⁴ M) with ligand 1a in

CH₃OH; (b) Corresponding asymmetry factors (AF = I_{617} / I_{592}) as a function of [1a]/[M] mole ratio ($\lambda_{ex} = 394$ nm).



Figure S33. (a) Fluorescence titration of Eu(NO₃)₃ (2 × 10⁻⁴ M) with ligand 1b in CH₃OH; (b) Corresponding asymmetry factors (AF = I_{617} / I_{592}) as a function of [1b]/[M] mole ratio ($\lambda_{ex} = 394$ nm).



Figure S34. (a) Fluorescence titration of Eu(NO₃)₃ (2 × 10⁻⁴ M) with ligand 1c in CH₃OH; (b) Corresponding asymmetry factors (AF = I_{617} / I_{592}) as a function of [1c]/[M] mole ratio ($\lambda_{ex} = 394$ nm).

7. Job's plot of 1 and Eu³⁺



Figure S35. Job's plot of 1a and Eu³⁺ in CH₃OH. The emission intensity at 617 nm is plotted against the molar fraction of [1a] to ([1a] + [Eu³⁺]) at a fixed total concentration 2.0×10^{-4} M ($\lambda_{ex} = 394$ nm).



Figure S36. Job's plot of 1b and Eu³⁺ in CH₃OH. The emission intensity at 617 nm is plotted against the molar fraction of [1b] to ([1b] + [Eu³⁺]) at a fixed total concentration 2.0×10^{-4} M ($\lambda_{ex} = 394$ nm).



Figure S37. Job's plot of 1c and Eu³⁺ in CH₃OH. The emission intensity at 617 nm is plotted against the molar fraction of [1c] to ([1c] + [Eu³⁺]) at a fixed total concentration 2.0×10^{-4} M ($\lambda_{ex} = 394$ nm).

8. UV-vis spectrophotometric titration of 1 with Eu³⁺

The stability constants β defined as the concentration ratio ($\beta = [M_x L_y^{n+}]^x/[M^{n+}]^x[L]^y)$ were determined in methanol by UV-vis spectrophotometric titration at 25 °C. The concentration of ligand **1** in methanol was 4.0×10^{-5} M and 0.01 M tetraethylammonium nitrate (Et₄NNO₃) was added to control the ionic strength. The experiments were carried out in a 1.0 cm quartz cell. Factor analysis and mathematical treatment of the spectrophotometric data were performed and fitted with the ReactLab EQUILIBRIA software.^[S6]



Figure S38. UV-vis spectrophotometric titration of 1a $(4.0 \times 10^{-5} \text{ M})$ with Eu(NO₃)₃



in methanol ($0 \le R = C_M/C_L \le 5$).

Figure S39. UV-vis spectrophotometric titration of 1b (4.0×10^{-5} M) with Eu(NO₃)₃



Figure S40. UV-vis spectrophotometric titration of 1c (4.0×10^{-5} M) with Eu(NO₃)₃

in methanol ($0 \le R = C_M/C_L \le 4$).



9. FT-IR spectra of free and Eu³⁺-loaded 1

Figure S41. FT-IR spectra of 1a in the absent and present of different equivalent of $Eu(NO_3)_3$.



Figure S42. FT-IR spectra of 1b in the absent and present of different equivalent of $Eu(NO_3)_3$.



Figure S43. FT-IR spectra of 1c in the absent and present of different equivalent of

Eu(NO₃)₃.

10. Extraction properties of Am(III) and Eu(III)

The ligands $(1.0 \times 10^{-3} \text{ M})$ were dissolved in a mixed solvent system consisting of *m*-nitro(trifluoromethyl)benzene and *n*-octanol (*m*-NTFB/*n*-octanol, 95:5, v:v). The organic phase was mixed with an aqueous phase containing europium(III) nitrate (C_{Eu} = 10^{-5} M) with ²⁴¹Am(III) or ¹⁵²Eu(III) (radiochemical purity, *ca*. 1.0×10^{-7} mol·L⁻¹) as well as HNO₃ at different concentrations (0.1, 1.0, 2.0, 3.0 and 4.0 M). Liquid-liquid extraction experiments were performed by shaking 1 mL of organic phase and 1 mL of aqueous phase in a stoppered tube immersed in a thermostated bath at 25 ± 0.5 °C for 1 hour which is long enough to reach the extraction equilibrium. After separation of the two phases, 0.5 mL of each phase was collected and the concentration was determined by gamma counting (NaI(TI) well detector). The distribution ratios D_M were calculated as the ratio of the cation gamma activity in the aqueous phase. The selectivity for Am(III) over Eu(III) is expressed as SF_{Am/Eu} = D_{Am}/D_{Eu}. The reported D_M values are the averages of at least two experiments. The errors in the extraction percentages of the duplicate experiments are less than 5%.



Figure S44. Chemical structures of CMPO-functionalized calixarenes I-III.

		°C.				
Compounds	HNO ₃ / mol·L ⁻¹			•L ⁻¹		
Compounds		0.1	1.0	2.0	3.0	4.0
1 a	D_{Am}	4.30	8.69	2.66	0.69	0.13
	\mathbf{D}_{Eu}	2.10	4.54	1.30	0.44	0.12
	$SF_{Am/Eu}$	2.05	1.91	2.05	1.57	1.08
1b	D_{Am}	4.46	10.7	3.30	0.78	0.43
	\mathbf{D}_{Eu}	2.12	6.17	1.37	0.47	0.19
	$SF_{Am/Eu}$	2.10	1.73	2.41	1.66	2.26
1c	D_{Am}	57.7	171	5.10	1.56	0.50
	\mathbf{D}_{Eu}	7.54	13.1	2.90	1.16	0.34
	$SF_{Am/Eu}$	7.65	13.0	1.76	1.34	1.47
\mathbf{I} - \mathbf{a}^a	D_{Am}	48	51	61	63	nr
	\mathbf{D}_{Eu}	28	33	44	48	nr
	$SF_{Am/Eu}$	1.7	1.5	1.4	1.3	
I-b ^{<i>a</i>}	D_{Am}	11.1	> 100	> 100	> 100	> 100
	\mathbf{D}_{Eu}	5.1	26.4	27.7	35.5	26.6
	$SF_{Am/Eu}$	2.2	> 3	> 3	> 3	> 3
I-c ^{<i>a</i>}	D_{Am}	18.3	32.8	nd	nd	nd
	\mathbf{D}_{Eu}	9.2	21.9	nd	nd	nd
	$SF_{Am/Eu}$	2.0	1.5	nd	nd	nd
\mathbf{II}^{a}	D_{Am}	19	195	275	150	100
	\mathbf{D}_{Eu}	2.3	30	52	37	19
	$SF_{Am/Eu}$	8.3	6.5	5.3	4.0	5.26
$\mathbf{III}^{\ b}$	D_{Am}	397	nd	> 10 ³	nd	nd
	\mathbf{D}_{Eu}	93	nd	> 10 ³	nd	nd
	$SF_{Am/Eu}$	4.27				
2	D_{Am}	< 0.01	< 0.01	0.04	0.02	< 0.01
	\mathbf{D}_{Eu}	< 0.01	< 0.01	0.03	< 0.01	< 0.01

Table S1 Extraction data of Am(III) and Eu(III) from different HNO₃ solution by the ligands **1a-c** (1 mM) and CMPO **2** (10 mM) into *m*-NTFB/*n*-octanol (95:5, v:v) at 25

^{*a*} The data were obtained from the literature [S7]; ^{*b*} The data were obtained from the literature [S8] ([ligands] = 1 mM; the diluent: *o*-nitrophenylhexyl ether, NPHE). nd: not determined.

e	× /	× ×	· · ·	
Compounds	Extraction No.	D_{Am}	\mathbf{D}_{Eu}	$SF_{Am/Eu}$
	1	1.56	1.16	1.34
1c	2	12.0	2.97	4.04
	3	103	6.63	15.5

Table S2 Multiple extractions of Am(III) and Eu(III) from 3 M HNO₃ solution by the ligands **1c** (1 mM) into *m*-NTFB/*n*-octanol (95:5, v:v) at 25 °C.

11. Fluorescence studies

General

Eu(NO₃)₃·6H₂O (spectroscopic grade, >99.99%) was procured from Alpha Biochem and the solutions were made using Suprapur (Merck) nitric acid and HPLC grade methanol. Sample solutions for time resolved laser induced fluorescence spectroscopy (TRLFS) measurements were prepared by taking suitable aliquots of Eu(III) stock in the present (or absent) of **1a-c** in the mixed solvent system of MeOH/H₂O (5:1, v/v, pH = 3.0).

TRLFS studies of the Eu³⁺-solvent and Eu³⁺-1 complexes, in dilute nitric acid (pH = 3.0) as well as in the solvent, were carried out using a spectrometer (Edinburgh Analytical Instruments, UK) controlled by CD 920 controller and equipped with OPO laser. While the samples were excited at 394 nm, the emission spectra were recorded in the range of 575-750 nm. The emission decay curves were fitted into the exponential function to obtain the lifetimes/decay rates of the excited states using inbuilt software GEM/3 (Edinburgh) with reproducibility of lifetimes of the excited states within \pm 3 µs. The life time of the ⁵D₀ emitting level of Eu³⁺ depends on the number of OH oscillators linked to the cations.^[S9] An empirical relation has been established between primary hydration number of Eu³⁺ ion ($N_{H_{20}}$) and the lifetime of its ⁵D₀ emitting level:

$$N_{\rm H2O} = 1.06/\tau - 0.19$$

The number of water molecules in the inner coordination sphere calculated from

the lifetime has an uncertainty of ± 0.5 in these measurements.

System	Life time (ms)	No. of inner-sphere water molecules
Eu ³⁺ -solvent	$\tau = 0.144$	7
Eu ³⁺ -1a	$\tau_1 = 0.225 (61\%)$	4.5
	$\tau_2 = 0.619 (39\%)$	1.5
Eu ³⁺ -1b	$\tau_1 = 0.246 \ (9.0\%)$	4.0
	$\tau_2 = 0.411 (91\%)$	2.0
Eu ³⁺ -1c	$\tau_1 = 0.335 (22\%)$	3.0
	$\tau_2 = 0.673 (78\%)$	1.5

Table S3 Life time and number of inner-sphere water molecules (N_{H2O}) of Eu³⁺-1complexes when mixing 1.0 equiv. of 1 (λ_{ex} = 394 nm; λ_{em} =617 nm).

Table S4 Life time and number of inner-sphere water molecules ($N_{\rm H2O}$) of Eu³⁺-1

System	Life time (ms)	No. of inner-sphere water molecules
Eu ³⁺ -1a	$\tau = 0.334$	3
Eu ³⁺ -1b	$\tau = 0.401$	2.5
Eu ³⁺ -1c	$\tau = 0.578$	1.6

complexes when mixing 0.5 equiv. of 1 (λ_{ex} = 394 nm; λ_{em} =617 nm).



Figure S44. Fluorescence decay profiles of Eu³⁺-1 complexes on addition of 0.5 equiv. of 1 in methanol/water (5:1, v/v, pH=3.0); (a) Eu³⁺-1a complex; (b) Eu³⁺-1b complex; (c) Eu³⁺-1c complex.



Figure S45. Fluorescence decay profiles of Eu³⁺-1 complexes on addition of 1.0

equiv. of **1** in methanol/water (5:1, v/v, pH=3.0); (a) Eu³⁺-**1a** complex; (b) Eu³⁺-**1b** complex; (c) Eu³⁺-**1c** complex.

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