

Supporting Information

DNA-cellulose: An economical, fully recyclable and highly effective chiral biomaterial for asymmetric catalysis

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Table of Contents

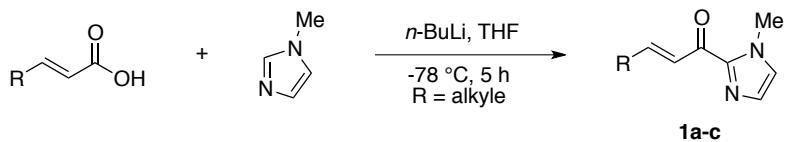
General Methods	3
Experimental and Spectral data	4
<i>General procedure A. Synthesis of α,β-unsaturated substrates 1a-c</i>	4
<i>General Procedure B. Synthesis of α,β-unsaturated substrates 1d-h</i>	7
<i>General Procedure C. Racemic Friedel-Crafts alkylations</i>	10
<i>General Procedure D. Enantioselective Friedel-Crafts alkylations</i>	17
<i>General Procedure E. Racemic Michael Additions</i>	22
<i>General Procedure F. Enantioselective Michael Additions</i>	25
<i>General Procedure G. Enantioselective Friedel Crafts alkylations – Recycling of CS-ct-DNA with MOPS buffer</i>	27
<i>General Procedure H. Enantioselective Friedel Crafts alkylations – Recycling of CS-ct-DNA with a MOPS buffer/MeOH (30:1) solution</i>	28
<i>General Procedure (I). Enantioselective Friedel Crafts alkylations in a continuous flow process</i>	29
¹ H NMR and ¹³ C NMR spectra	32
SFC chromatograms	57

General Methods

All reactions were run under argon atmosphere in oven-dried glassware unless otherwise specified. All commercially available compounds were purchased from Aldrich Chemical Co., Worthington Chemicals (CS-ct-DNA), GFS Chemicals, Strem Chemicals, Acros Organics or Alfa Aesar and used as received. Dichloromethane (DCM) was distilled from calcium hydride. Tetrahydrofuran (THF) and diethyl ether (Et₂O) were distilled from sodium/benzophenone. *N,N*-dimethylformamide (DMF) was distilled under vacuum over anhydrous MgSO₄. Analytical thin layer chromatography (TLC) was performed on silica gel plates (Merck 60F₂₅₄) visualized either with a UV lamp (254 nm) or by using solutions of *p*-anisaldehyde/sulfuric acid/acetic acid (AcOH) in ethanol (EtOH) or KMnO₄/K₂CO₃/AcOH in water followed by heating. Flash chromatographies were performed on silica gel (60-230 mesh). Organic extracts were dried over anhydrous Na₂SO₄ or MgSO₄. Infrared spectra (IR) were recorded on a Bruker TENSOR™ 27 (IRTF) and wavenumbers are indicated in cm⁻¹. ¹H NMR spectra were recorded on a Bruker AVANCE 400 at 400 MHz in CDCl₃ (unless otherwise specified) and the observed signals are reported as follows: chemical shift in parts per million from tetramethylsilane with the solvent as an internal indicator (CDCl₃ δ 7.26 ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, m = multiplet or overlap of nonequivalent resonances), integration. ¹³C NMR spectra were recorded at 100 MHz in CDCl₃ (unless otherwise specified) and the observed signals were reported as follows: chemical shift in parts per million from tetramethylsilane with the solvent as an internal indicator (CDCl₃ δ 77.0 ppm), multiplicity with respect to proton (deduced from DEPT experiments, s = quaternary C, d = CH, t = CH₂, q = CH₃). Coupling constants (*J*) are reported in hertz (Hz). All NMR spectra were obtained at room temperature unless otherwise specified. Enantiomeric excess (*e*_{es}) determinations were performed by supercritical fluid chromatography (SFC) analysis on chiral phase. The sign before the *e*_{es} values is arbitrary. Mass spectra (MS) were recorded using a Hewlett-Packard tandem 5890A/5971 GCMS (70 eV). High-resolution mass spectra were performed by "Groupe de Spectrométrie de masse de l'Université Pierre et Marie Curie (Paris)".

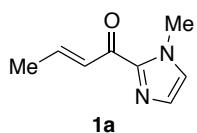
Experimental and Spectral data

General procedure A. Synthesis of α,β -unsaturated acyl imidazoles **1a-c**



The α,β -unsaturated substrates **1a-c** were synthesized *via* a modification of the procedure originally reported by Evans and co-workers.¹ An oven-dried, 250 mL round-bottomed flask under an argon atmosphere was charged with 1-methylimidazole (24 mmol, 2.4 equiv) and dry THF (50 mL). The solution was cooled to -78 °C in a dry ice/acetone bath for 15 min, then *n*-BuLi (2.5 M in *n*-hexane, 24 mmol, 2.4 equiv) was added drop-wise over 10 min. The mixture was warmed to rt and stirred for 30 min, then cooled back to -78 °C. The desired acid (10 mmol, 1 equiv) in dry THF (10 mL) was added drop-wise over a 10 min period. The resulting solution was stirred at -78 °C for 15 min, then warmed at rt and stirred for an additional 2 h. The reaction was eventually quenched with a saturated aqueous NaHCO₃ solution (50 mL) and the aqueous phase was extracted with EtOAc (3 x 40 mL). The combined organic layers were washed with brine (2 x 40 mL), dried over MgSO₄, gravity filtered, and concentrated under reduced pressure. The reaction residue was purified by silica gel flash chromatography, eluting with EtOAc/pentane (4:6) as the eluent.

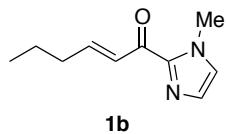
(E)-1-(1-Methyl-1*H*-imidazol-2-yl)but-2-en-1-one (**1a**)



Following general procedure A. 1-Methylimidazole (2 g, 2.2 mL, 27.6 mmol), *n*-BuLi (2.5 M in *n*-hexane, 11 mL, 27.6 mmol), and crotonic acid (0.95 g, 11.4 mmol) in dry THF (60 mL). The title compound (molecular formula: C₈H₁₀N₂O, MW = 150.18 g/mol, 0.91 g) was isolated as a colorless oil in 53% yield. **Rf:** 0.36 (EtOAc/pentane = 3:2). **IR** (neat): 3107, 2963, 1664, 1617, 1402, 1283, 1030, 909 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.36 (dq, *J* = 15.5, 1.8 Hz, 1H), 7.13-7.01 (m, 2H), 6.99 (s, 1H), 3.98 (s, 3H), 2.20-1.58 (m, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 180.5 (s), 143.7 (d), 143.5 (s), 129.0 (d), 127.7 (d), 126.9 (d), 36.1 (q), 18.3 (q). Spectroscopic data were consistent with the literature data for this compound.¹

¹ D. A. Evans, K. R. Fandrick, H.-J. Song *J. Am. Chem. Soc.* **2005**, 127, 8942.

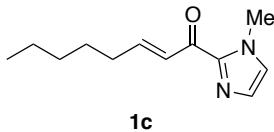
(E)-1-(1-Methyl-1*H*-imidazol-2-yl)hex-2-en-1-one (1b)



1b

Following general procedure A. 1-Methylimidazole (1 g, 1 mL, 12.6 mmol), *n*-BuLi (2.5 M in *n*-hexane, 5 mL, 13.7 mmol), and (*E*)-hexenoic acid (0.65 g, 0.7 mL, 5.7 mmol) in dry THF (50 mL). The title compound (molecular formula: C₁₀H₁₄N₂O, MW = 178.23 g/mol, 0.505 g) was isolated as a colorless oil in 51% yield. **Rf:** 0.37 (EtOAc/pentane 3:2). **IR** (neat): 2960, 2931, 2837, 2665, 1618, 1405, 1022, 978, 917, 728 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.40 (dt, *J* = 15.6, 1.5 Hz, 1H), 7.20-7.07 (m, 2H), 7.04 (d, *J* = 0.5 Hz, 1H), 4.12-4.03 (m, 3H), 2.41-2.20 (m, 2H), 1.62-1.39 (m, 2H), 0.96 (t, *J* = 7.4 Hz, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 180.4 (s), 148.3 (d), 143.5 (s), 128.9 (d), 126.9 (d), 126.2 (d), 36.0 (q), 34.4 (t), 21.2 (t), 13.6 (q). Spectroscopic data were consistent with the literature data for this compound.²

(E)-1-(1-Methyl-1*H*-imidazol-2-yl)oct-2-en-1-one (1c)



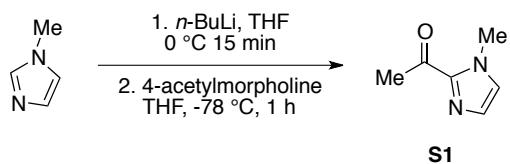
1c

Following general procedure A. 1-Methylimidazole (1 g, 1 mL, 12.2 mmol), *n*-BuLi (2.5 M in *n*-hexane, 5 mL, 13.3 mmol), and (*E*)-oct-2-enoic acid (0.79 g, 5.5 mmol) in dry THF (50 mL). The title compound (molecular formula: C₁₂H₁₈N₂O, MW = 206.28 g/mol, 0.56 g) was isolated as a colorless oil in 49% yield. **Rf:** 0.39 (EtOAc/pentane 6:4). **IR** (neat): 2957, 2928, 2858, 1666, 1618, 1406, 991, 916, 732 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.40 (dt, *J* = 15.6, 1.5 Hz, 1H), 7.19-7.06 (m, 2H), 7.08-6.99 (m, 1H), 4.12-3.97 (m, 3H), 2.40-2.21 (m, 2H), 1.58-1.44 (m, 2H), 1.41-1.23 (m, 4H), 1.12-0.76 (m, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 180.8 (s), 149.1 (d), 143.8 (s), 129.2 (d), 127.1 (d), 126.1 (d), 36.3 (q), 32.7 (t), 31.5 (t), 27.9 (t), 22.5 (t), 14.0 (q). Spectroscopic data were consistent with the literature data for this compound.³

² Wang, X.; Wang, D. Z. *Tetrahedron* **2011**, *67*, 3406.

³ Boersma, A. J.; Feringa, B. L.; Roelfes, G. *Angew. Chem, Int. Ed.* **2009**, *48*, 3346.

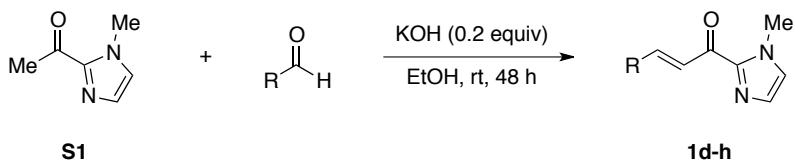
Synthesis of 1-(1-methyl-1*H*-imidazol-2-yl)ethanone (**S1**)



Compound **S1** was synthesized *via* a modification of the procedure originally reported by Scheidt and co-workers.³ An oven-dried 100 mL round-bottomed flask under an argon atmosphere was charged with 1-methylimidazole (4.5 g, 4.4 mL, 55.2 mmol, 1.1 equiv) and dry THF (60 mL). The solution was cooled to 0 °C in an ice bath for 15 min, then *n*-BuLi (2.5 M in *n*-hexane, 24 mL, 55.2 mmol, 1.1 equiv) was added dropwise over 10 min. The mixture was stirred at 0 °C for 15 min, then cannulated into a solution of 4-acetylmorpholine (6.5 g, 5.8 mL, 50.1 mmol, 1 equiv) in dry THF (40 mL) at -78 °C. The reaction mixture was then stirred at -78 °C for 1 h and quenched with a 1 N aqueous solution of HCl (5 mL), stirred for 5 min and diluted with a saturated aqueous solution of NaHCO₃ (15 mL) and brine (15 mL). The aqueous phase was extracted with EtOAc (3 x 40 mL) and the combined organic layers were dried over anhydrous MgSO₄, gravity filtered and concentrated under reduced pressure. The reaction residue was purified by silica gel flash chromatography, eluting with Et₂O/pentane (9:1) to provide the title compound (molecular formula: C₆H₈N₂O, MW = 124.14 g/mol, 6.2 g) as a colorless oil in 79% yield. **Rf:** 0.31 (Et₂O/pentane = 9:1). **IR** (neat) 3108, 2959, 1674, 1402, 915, 776 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.14 (d, J = 0.9 Hz, 1H), 7.02 (br, s, 1H), 3.99 (s, 3H), 2.66 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 190.2 (s), 142.9 (s), 128.7 (d), 126.7 (d), 35.9 (q), 26.8 (q). Spectroscopic data were consistent with the literature data for this compound.⁴

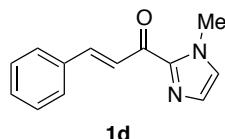
⁴ M. C. Myers, A. R. Bharadwaj, B. C. Milgram, K. A. Scheidt *J. Am. Chem. Soc.* **2005**, 127, 14675.

General procedure B. Synthesis of α,β -unsaturated acyl imidazoles **1d-h**



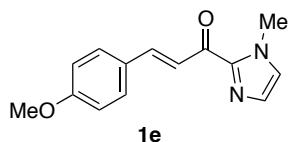
The α,β -unsaturated substrates **1d-h** were synthesized *via* a modification of the procedure originally reported by Scheidt and co-workers.³ An oven-dried 100 mL round-bottomed flask under an argon atmosphere was charged with 1-methylimidazole (10.0 mmol, 1.0 equiv) and EtOH (20 mL). The appropriate aromatic aldehyde (10.0 mmol, 1.0 equiv) and a catalytic amount of KOH (2 mmol, 0.2 equiv) were added and the solution was stirred at rt for 48 h. The formation of a precipitate was observed. The reaction was quenched with H₂O (50 mL) and the aqueous phase was extracted with EtOAc (3 x 40 mL). The combined organic layers were washed with brine (2 x 50 mL), dried over anhydrous MgSO₄, gravity filtered and concentrated under reduced pressure. The crude product was purified by flash chromatography over silica gel using EtOAc/pentane (1:1) as the eluent.

(E)-1-(1-Methyl-1*H*-imidazol-2-yl)-3-phenylprop-2-en-1-one (1d**)**



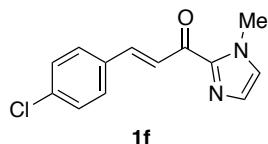
Following general procedure B. Compound **S1** (1.3 g, 10.5 mmol), benzaldehyde (1.1 g, 1.1 mL, 10.5 mmol), and KOH (0.11 g, 2 mmol), in EtOH (20 mL). The title compound (molecular formula: C₁₃H₁₂N₂O, MW = 212.25 g/mol, 1.3 g) was isolated as a white solid in 57% yield. **Rf:** 0.40 (Et₂O/pentane = 9:1). **IR** (neat): 3124, 3100, 3022, 1655, 1603, 1400, 1017, 769 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 8.07 (d, *J* = 16.0 Hz, 1H), 7.81 (d, *J* = 16.0 Hz, 1H), 7.73-7.64 (m, 2H), 7.44-7.32 (m, 3H), 7.20 (d, *J* = 0.9 Hz, 1H), 7.07 (s, 1H), 4.09 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 180.4 (s), 143.9 (s), 143.1 (d), 134.8 (s), 130.4 (d), 129.2 (d), 128.8 (2C, d), 128.7 (2C, d), 127.2 (d), 122.7 (d), 36.3 (q). Spectroscopic data were consistent with the literature data for this compound.⁴

(E)-3-(4-Methoxyphenyl)-1-(1-methyl-1*H*-imidazol-2-yl)prop-2-en-1-one (1e)



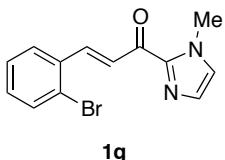
Following general procedure B. Compound **S1** (1.3 g, 10.5 mmol), 4-methoxybenzaldehyde (1.4 g, 1.3 mL, 10.5 mmol), and KOH (0.11 g, 2 mmol), in EtOH (20 mL). The title compound (molecular formula: C₁₄H₁₄N₂O₂, MW = 242.27 g/mol, 1.3 g) was isolated as a yellow solid in 51% yield. **Rf:** 0.36 (Et₂O/pentane = 9:1). **IR** (neat): 3129, 3105, 2938, 2844, 1649, 1586, 1567, 1510, 1398, 1250, 1155, 1021, 990, 730 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.93 (d, *J* = 16.0 Hz, 1H), 7.77 (d, *J* = 16.0 Hz, 1H), 7.62 (d, *J* = 8.8 Hz, 2H), 7.18 (s, 1H), 7.03 (s, 1H), 6.89 (d, *J* = 8.8 Hz, 2H), 4.05 (s, 3H), 3.81 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 180.4 (s), 161.5 (s), 144.0 (s), 143.1 (d), 130.4 (2C, d), 129.0 (d), 127.6 (s), 127.0 (d), 120.4 (d), 114.2 (2C, d), 55.3 (q), 36.3 (q). Spectroscopic data were consistent with the literature data for this compound.⁴

(E)-3-(4-Chlorophenyl)-1-(1-methyl-1*H*-imidazol-2-yl)prop-2-en-1-one (1f)



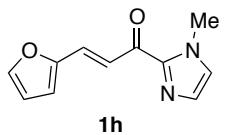
Following general procedure B. Compound **S1** (1.3 g, 10.5 mmol), 4-chlorobenzaldehyde (1.47 g, 10.5 mmol) and KOH (0.11 g, 2 mmol), in EtOH (20 mL). The title compound (molecular formula: C₁₃H₁₁ClN₂O, MW = 246.69 g/mol, 1.7 g) was isolated as a white solid in 67% yield. **Rf:** 0.40 (Et₂O/pentane = 9:1). **IR** (neat): 3129, 3109, 2957, 1654, 1594, 1397, 1286, 1014, 815, 783 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 8.02 (d, *J* = 16.0 Hz, 1H), 7.72 (d, *J* = 16.0 Hz, 1H), 7.59 (d, *J* = 8.4 Hz, 2H), 7.37 (d, *J* = 8.4 Hz, 2H), 7.19 (s, 1H), 7.06 (s, 1H), 4.06 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 180.1 (s), 143.8 (s), 141.7 (d), 136.2 (s), 133.3 (s), 129.8 (2C, d), 129.3 (d), 129.0 (2C, d), 127.3 (d), 123.2 (d), 36.3 (q). Spectroscopic data were consistent with the literature data for this compound.⁴

(E)-3-(2-Bromophenyl)-1-(1-methyl-1*H*-imidazol-2-yl)prop-2-en-1-one (1g)



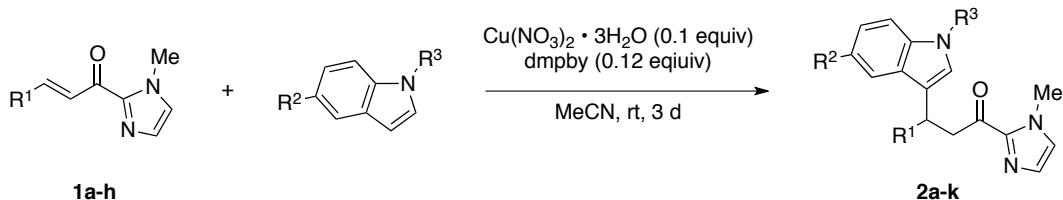
Following general procedure B. Compound **S1** (1.3 g, 10.5 mmol), 2-bromobenzaldehyde (1.9 g, 1.2 mL, 10.5 mmol) and KOH (0.11 g, 2 mmol), in EtOH (20 mL). The title compound (molecular formula: C₁₃H₁₁BrN₂O, MW = 291.14 g/mol, 2.0 g) was isolated as a white solid in 67% yield. **Rf:** 0.37 (Et₂O/pentane = 9:1). **IR** (neat): 3128, 3102, 2954, 1654, 1599, 1402, 1019, 735 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 8.21 (d, *J* = 15.9 Hz, 1H), 8.03 (d, *J* = 15.9 Hz, 1H), 7.86 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.62 (dd, *J* = 8.0, 1.1 Hz, 1H), 7.35-7.30 (m, 1H), 7.28-7.18 (m, 2H), 7.09 (s, 1H), 4.10 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 179.9 (s), 143.9 (s), 141.5 (d), 134.8 (s), 133.4 (d), 131.3 (d), 129.4 (d), 128.1 (d), 127.6 (d), 127.4 (d), 126.1 (s), 125.2 (d), 36.4 (q). Spectroscopic data were consistent with the literature data for this compound.⁴

(E)-3-(Furan-2-yl)-1-(1-methyl-1*H*-imidazol-2-yl)prop-2-en-1-one (1h)



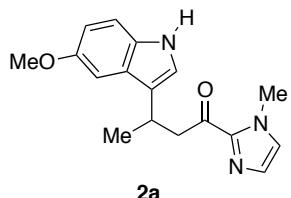
Following general procedure B. Compound **S1** (1.3 g, 10.5 mmol), furan-2-carbaldehyde (1 g, 0.87 mL, 10.5 mmol) and KOH (0.11 g, 2 mmol), in EtOH (20 mL). The title compound (molecular formula: C₁₁H₁₀N₂O₂, MW = 202.21 g/mol, 0.95 g) was isolated as a brown solid in 45% yield. **Rf:** 0.30 (EtOAc/pentane = 1:1). **IR** (neat): 3129, 3109, 2957, 1654, 1594, 1397, 1286, 1014, 815, 783 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.87 (d, *J* = 15.8 Hz, 1H), 7.56 (d, *J* = 15.8 Hz, 1H), 7.48 (d, *J* = 1.7 Hz, 1H), 7.17 (s, 1H), 7.04 (s, 1H), 6.70 (d, *J* = 3.4 Hz, 1H), 6.46 (dd, *J* = 3.4, 1.7 Hz, 1H), 4.04 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 180.2 (s), 151.8 (s), 144.9 (d), 143.9 (s), 129.4 (d), 129.2 (d), 127.1 (d), 120.7 (d), 115.6 (d), 112.4 (d), 36.2 (q). Spectroscopic data were consistent with the literature data for this compound.³

General Procedure C. Racemic Friedel-Crafts alkylations



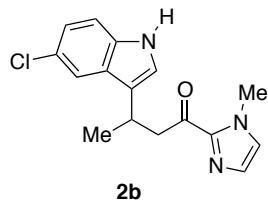
An oven-dried 25 mL round-bottomed flask was charged with $\text{Cu}(\text{NO}_3)_2 \cdot 3\text{H}_2\text{O}$ (0.035 mmol, 0.1 equiv), 4,4'-dimethyl-2,2'-bipyridyl (dmbpy, 0.042 mmol, 0.12 equiv), and MeCN (5 mL). The mixture was stirred at rt for 10 min then the α,β -unsaturated substrate (0.35 mmol, 1.0 equiv) and the desired indole (0.53 mmol, 1.5 equiv) were added. The solution was stirred at rt for 3 d. The reaction was eventually diluted with brine (10 mL) and extracted with Et_2O (3 x 5 mL). The combined organic layers dried over anhydrous MgSO_4 , gravity filtered and concentrated under reduced pressure. The crude product was purified by flash chromatography over silica gel using EtOAc/pentane (1:1) as the eluent.

3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2a)



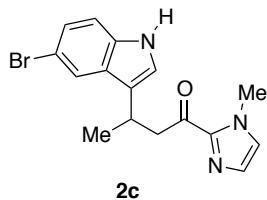
Following general procedure C. Compound **1a** (0.053 g, 0.35 mmol), 5-methoxyindole (0.078 g, 0.53 mmol), $\text{Cu}(\text{NO}_3)_2 \cdot 3\text{H}_2\text{O}$ (0.008 g, 0.035 mmol), dmbpy (0.008 g, 0.042 mmol), and MeCN (5 mL). The title compound (molecular formula: $\text{C}_{17}\text{H}_{19}\text{N}_3\text{O}_2$, MW = 297.35 g/mol, 0.094 g) was isolated as a brown solid in 91% yield. **Rf:** 0.29 (EtOAc/pentane = 1:1). **IR** (neat): 3047, 2958, 2926, 1671, 1484, 1405, 1215, 923, 795 cm^{-1} . **¹H NMR** (400 MHz, CDCl_3) δ 8.18 (br, s, 1H), 7.20 (d, J = 8.8 Hz, 1H), 7.17-7.15 (m, 2H), 7.02 (d, J = 2.5 Hz, 1H), 7.00 (s, 1H), 6.82 (dd, J = 8.8, 2.5 Hz, 1H), 3.94 (s, 3H), 3.86 (s, 3H), 3.83–3.72 (m, 1H), 3.58 (dd, J = 15.6, 6.5 Hz, 1H), 3.40 (dd, J = 15.6, 8.8 Hz, 1H), 1.40 (d, J = 6.5 Hz, 3H). **¹³C NMR** (100 MHz, CDCl_3) δ 192.3 (s), 153.7 (s), 143.3 (s), 131.5 (s), 128.8 (d), 126.9 (s), 126.8 (d), 121.1 (s), 120.9 (d), 112.1 (d), 111.7 (d), 101.0 (d), 55.9 (t), 46.8 (q), 36.2 (d), 27.1 (q), 21.4 (q). Spectroscopic data were consistent with the literature data for this compound.³

3-(5-Chloro-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2b)



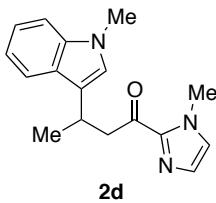
Following general procedure C. Compound **1a** (0.020 g, 0.133 mmol), 5-chloroindole (0.024 g, 0.160 mmol), Cu(NO₃)₂•3H₂O (0.0032 g, 0.013 mmol), dmbpy (0.0034 g, 0.019 mmol), and MeCN (0.5 mL). The title compound (molecular formula: C₁₆H₁₆ClN₃O, MW = 301.77 g/mol, 0.040 g) was isolated as a brown oil in quantitative yield. **Rf:** 0.32 (EtOAc/pentane 1:1). **IR** (neat): 3173, 2961, 2928, 2869, 1672, 1462, 1406, 1104, 986 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 8.34 (s, 1H), 7.56 (d, *J* = 2.0 Hz, 1H), 7.22 (dd, *J* = 8.6, 0.5Hz, 1H), 7.16 (d, *J* = 0.9 Hz, 1H), 7.12-7.06 (m, 1H), 7.06-6.99 (m, 2H), 3.93 (s, 3H), 3.80-3.68 (m, 1H), 3.46 (qd, *J* = 15.7, 7.3 Hz, 2H), 1.40 (d, *J* = 6.9 Hz, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 192.0 (s), 143.2 (s), 134.6 (s), 128.9 (d), 127.7 (s), 127.0 (d), 124.8 (s), 122.1 (d), 121.6 (d), 121.2 (s), 118.7 (d), 112.0 (d), 46.9 (t), 36.1 (d), 27.0 (q), 21.5 (q). **HRMS (ESI):** *m/z* calcd for C₁₆H₁₆ClN₃ONa [M + Na]⁺: 324.08741, found: 324.08711.

3-(5-Bromo-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2c)



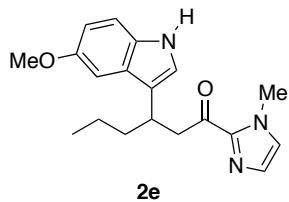
Following general procedure C. Compound **1a** (0.029 g, 0.19 mmol), 5-bromoindole (0.057 g, 0.29 mmol), Cu(NO₃)₂•3H₂O (0.004 g, 0.018 mmol), dmbpy (0.005 g, 0.026 mmol) and MeCN (1.5 mL). The title compound (molecular formula: C₁₆H₁₆BrN₃O, MW = 346.22 g/mol, 0.052 g) was isolated as a brown oil in 78% yield. **Rf:** 0.17 (EtOAc/pentane = 2:3). **¹H NMR** (400 MHz, CDCl₃) δ 8.68 (br, s, 1H), 7.62 (s, 1H), 7.19-7.06 (m, 3H), 7.95 (s, 1H), 6.89 (s, 1H), 3.85 (s, 3H), 3.69 (m, 1H), 3.37 (d, *J* = 7.8 Hz, 2H), 1.30 (d, *J* = 1.30 Hz, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 192.0 (s), 143.2 (s), 135.0 (s), 128.9 (d), 128.3 (s), 127.15 (d), 124.6 (s), 121.7 (d), 121.6 (d), 121.0 (s), 112.6 (d), 112.0 (d), 47.1 (t), 36.2 (d), 27.0 (q), 21.6 (q). Spectroscopic data were consistent with the literature data for this compound.³

1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)butan-1-one (2d)



Following general procedure C. Compound **1a** (0.074 g, 0.35 mmol), *N*-methylindole (0.069 g, 0.066 mL, 0.53 mmol), Cu(NO₃)₂•3H₂O (0.008 g, 0.035 mmol), dmbpy (0.008 g, 0.042 mmol), and MeCN (5 mL). The title compound (molecular formula: C₂₂H₂₁N₃O, MW = 343.42 g/mol, 0.096 g) was isolated as a brown solid in 80% yield. **Rf:** 0.31 (EtOAc/pentane = 1:1). **IR** (neat): 3026, 2916, 1673, 1472, 1406, 1154, 915, 741 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.48 (dt, *J* = 8.0, 1.0 Hz, 1H), 7.44-7.38 (m, 2H), 7.29-7.20 (m, 3H), 7.20-7.08 (m, 3H), 7.04-6.94 (m, 3H), 5.05 (t, *J* = 7.5 Hz, 1H), 4.01 (dd, *J* = 16.5, 7.5 Hz, 1H), 3.88 (s, 3H), 3.86 (dd, *J* = 16.5, 7.5 Hz, 1H), 3.71 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 191.0 (s), 144.6 (s), 143.1 (s), 137.1 (s), 128.8 (d), 128.3 (2C, d), 127.9 (2C, d), 127.1 (s), 126.8 (d), 126.2 (d), 126.0 (d), 121.5 (d), 119.6 (d), 118.7 (d), 117.8 (s), 109.0 (d), 45.5 (t), 38.0 (d), 36.1 (q), 32.7 (q). Spectroscopic data were consistent with the literature data for this compound.³

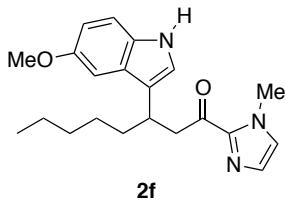
3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)hexan-1-one (2e)



Following general procedure C. Compound **1b** (0.020 g, 0.112 mmol), 5-methoxylindole (0.025 g, 0.168 mmol), Cu(NO₃)₂•3H₂O (0.0027 g, 0.011 mmol), dmbpy (0.0029 g, 0.016 mmol), and MeCN (0.5 mL). The title compound (molecular formula: C₁₉H₂₃N₃O₂, MW = 325.40 g/mol, 0.025 g) was isolated as a brown oil in 68% yield. **Rf:** 0.32 (EtOAc/pentane 1:1). **IR** (neat): 3405, 2955, 2927, 1672, 1483, 1458, 1405, 12122, 1029, 754 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 8.20 (s, 1H), 7.18 (d, *J* = 8.7 Hz, 1H), 7.13 (s, 1H), 7.08 (d, *J* = 2.4 Hz, 1H), 6.99 (d, *J* = 2.1 Hz, 1H), 6.96 (s, 1H), 6.80 (dd, *J* = 8.7, 2.4 Hz, 1H), 3.85 (s, 3H), 3.84 (s, 3H), 3.69 (p, *J* = 7.2 Hz, 1H), 3.51 (d, *J* = 7.2 Hz, 2H), 1.84-1.67 (m, 2H), 1.37-1.22 (m, 2H), 0.86 (t, *J* = 7.2 Hz, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 192.4 (s), 153.6 (s),

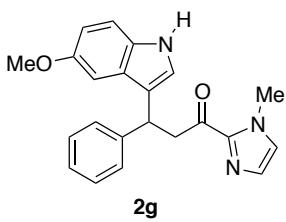
143.3 (s), 131.4 (s), 128.7 (d), 127.4 (s), 126.7 (d), 121.8 (d), 119.3 (s), 111.8 (d), 111.6 (d), 101.1 (d), 55.9 (q), 45.5 (t), 38.4 (t), 36.0 (q), 32.0 (d), 20.6 (t), 14.1 (q). **HRMS (ESI)**: m/z calcd for $C_{19}H_{23}N_3O_2Na$ [M + Na]⁺: 348.16825, found: 348.16772.

3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)octan-1-one (2f)



Following general procedure C. Compound **1c** (0.023 g, 0.112 mmol), 5-methoxyindole (0.025 g, 0.168 mmol), Cu(NO₃)₂•3H₂O (0.0027 g, 0.011 mmol), dmbpy (0.0029 g, 0.016 mmol), and MeCN (0.5 mL). The title compound (molecular formula: C₂₁H₂₇N₃O₂, MW = 353.46 g/mol, 0.0295 g) was isolated as a brown oil in 73% yield. **R_f**: 0.38 (EtOAc/pentane 1:1). **IR** (neat): 3049, 2954, 2926, 2855, 1672, 1484, 1459, 1407, 1214, 1032, 917, 754 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 8.07 (s, 1H), 7.19 (d, *J* = 8.7 Hz, 1H), 7.13 (d, *J* = 0.9 Hz, 1H), 7.07 (d, *J* = 2.4 Hz, 1H), 7.00 (d, *J* = 2.2 Hz, 1H), 6.96 (s, 1H), 6.80 (dd, *J* = 8.7, 2.4 Hz, 1H), 3.85 (s, 3H), 3.84 (s, 3H), 3.66 (d, *J* = 7.2 Hz, 1H), 3.50 (dd, *J* = 7.3, 4.3 Hz, 2H), 1.83-1.69 (m, 2H), 1.34-1.16 (m, 6H), 0.85-0.76 (m, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 192.4 (s), 153.6 (s), 143.4 (s), 131.4 (s), 128.8 (d), 127.4 (s), 126.7 (d), 121.8 (d), 119.4 (s), 111.9 (d), 111.6 (d), 101.1 (d), 55.9 (q), 45.5 (t), 36.1 (t), 36.0 (d), 32.3 (q), 31.9 (t), 27.1 (t), 22.6 (t), 14.1 (q). Spectroscopic data were consistent with the literature data for this compound.⁵

1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)-3-phenylpropan-1-one (2g)

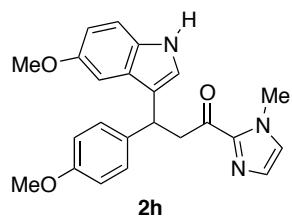


Following general procedure C. Compound **1d** (0.042 g, 0.20 mmol), 5-methoxyindole (0.044 g, 0.30 mmol), Cu(NO₃)₂•3H₂O (0.005 g, 0.020 mmol), dmbpy (0.005 g, 0.026 mmol),

⁵ Megens, R. P.; Roelfes, G. *Org. Biomol. Chem.* **2010**, 8, 1387.

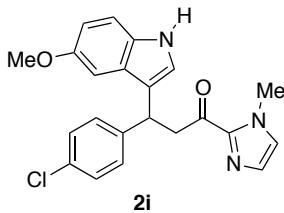
and MeCN (5 mL). The title compound (molecular formula: C₂₂H₂₁N₃O₂, MW = 359.42 g/mol, 0.056 g) was isolated as a brown solid in 79% yield. **Rf:** 0.31 (EtOAc/pentane = 1:1). **IR** (neat): 2919, 2830, 1672, 1511, 1401, 1247, 1030, 791, 735 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 8.31 (br, s, 1H), 7.27 (d, J = 7.2 Hz), 7.16-7.12 (m, 2H), 7.08-7.02 (m, 3H), 6.97 (d, J = 2.0 Hz, 1H), 6.89 (s, 1H), 6.83 (d, J = 2.4 Hz), 6.68 (dd, J = 88, 2.4 Hz, 1H), 4.92 (t, J = 7.6 Hz, 1H), 3.94-3.88 (m, 1H), 3.79 (s, 3H), 3.78-3.74 (m, 1H), 3.66 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 191.1 (s), 153.7 (s), 144 (s), 143.2 (s), 131.7 (s), 131.0 (d), 128.9 (d), 128.4 (2C, d), 128.0 (2C, d), 127.1 (s), 126.2 (d), 122.3 (d), 119.0 (s), 122.1 (d), 111.8 (d), 101.3 (d), 55.8 (q), 45.4 (t), 38.1 (d), 36.2 (q). Spectroscopic data were consistent with the literature data for this compound.³

3-(5-Methoxy-1*H*-indol-3-yl)-3-(4-methoxyphenyl)-1-(1-methyl-1*H*-imidazol-2-yl)propan-1-one (2h)



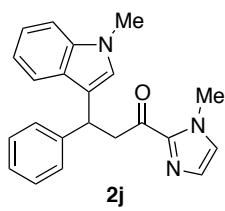
Following general procedure C. Compound **1g** (0.085 g, 0.35 mmol), 5-methoxyindole (0.078 g, 0.53 mmol), Cu(NO₃)₂·3H₂O (0.008 g, 0.035 mmol), dmbpy (0.008 g, 0.042 mmol) and MeCN (5 mL). The title compound (molecular formula: C₂₃H₂₃N₃O₃, MW = 389.4 g/mol, 0.135 g) was isolated as a brown solid in quantitative yield. **Rf:** 0.35 (EtOAc/pentane = 1:1). **IR** (neat): 2917, 2833, 1673, 1509, 1405, 1245, 1033, 789, 734 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.97 (br, s, 1H), 7.35-7.28 (m, 2H), 7.20 (d, J = 8.8 Hz, 1H), 7.17 (d, J = 0.9 Hz, 1H), 7.13 (d, J = 2.4 Hz, 1H), 7.04-6.98 (m, 1H), 6.93 (d, J = 2.4 Hz, 1H), 6.83-6.72 (m, 3H), 4.97 (t, J = 7.6 Hz, 1H), 4.00 (dd, J = 16.3, 7.6 Hz, 1H), 3.92 (s, 3H), 3.85 (dd, J = 16.3, 7.6 Hz, 1H), 3.78 (s, 3H), 3.76 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 191.2 (s), 157.8 (s), 153.7 (s), 143.2 (s), 136.4 (s), 131.6 (s), 128.8 (2C, d), 128.7 (d), 127.2 (d), 126.9 (s), 122.1 (d), 119.5 (s), 113.7 (2C, d), 112.1 (d), 111.6 (d), 101.5 (d), 55.8 (q), 55.2 (q), 45.5 (t), 37.4 (d), 36.2 (q). Spectroscopic data were consistent with the literature data for this compound.³

3-(4-Chlorophenyl)-3-(5-methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)propan-1-one (2i)



Following general procedure C. Compound **1f** (0.086 g, 0.35 mmol), 5-methoxyindole (0.078 g, 0.53 mmol) $\text{Cu}(\text{NO}_3)_2 \cdot 3\text{H}_2\text{O}$ (0.008 g, 0.035 mmol), dmbpy (0.008 g, 0.042 mmol) and MeCN (5 mL). The title compound (molecular formula: $\text{C}_{22}\text{H}_{20}\text{ClN}_3\text{O}_2$, MW = 393.87 g/mol, 0.137 g) was isolated as a brown solid in quantitative yield. **R_f**: 0.25 (EtOAc/pentane = 1:1). **IR** (neat): 2916, 1672, 1486, 1405, 1212, 1035, 916, 732 cm^{-1} . **¹H NMR** (400 MHz, CDCl_3) δ 8.41 (br, s, 1H), 7.32-7.21 (m, 2H), 7.22-7.10 (m, 4H), 7.05 (d, J = 2.4 Hz, 1H), 7.0 (s, 1H), 6.87 (d, J = 2.4 Hz, 1H), 6.79 (dd, J = 8.8, 2.4 Hz, 1H), 4.97 (t, J = 7.6 Hz, 1H), 3.94 (dd, J = 16.5, 8.3 Hz, 1H), 3.90 (s, 3H), 3.83 (dd, J = 16.5, 8.3 Hz, 1H), 3.76 (s, 3H). **¹³C NMR** (100 MHz, CDCl_3) δ 190.7 (s), 153.7 (s), 143.0 (s), 142.8 (s), 131.7 (s), 131.6 (s), 129.3 (2C, d), 128.9 (d), 128.4 (2C, d), 127.1 (d), 126.9 (s), 122.2 (d), 118.5 (s), 112.1 (d), 111.7 (d), 101.2 (d), 55.8 (q), 45.1 (t), 38.1 (s), 36.1 (q). Spectroscopic data were consistent with the literature data for this compound.³

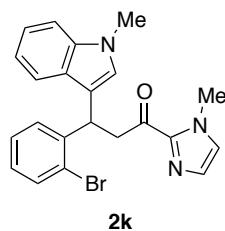
1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)-3-phenylpropan-1-one (2j)



Following general procedure C. Compound **1d** (0.074 g, 0.35 mmol), *N*-methylindole (0.069 g, 0.066 mL, 0.53 mmol), $\text{Cu}(\text{NO}_3)_2 \cdot 3\text{H}_2\text{O}$ (0.008 g, 0.035 mmol), dmbpy (0.008 g, 0.042 mmol), and MeCN (5 mL). The title compound (molecular formula: $\text{C}_{22}\text{H}_{21}\text{N}_3\text{O}$, MW = 343.42 g/mol, 0.096 g) was isolated as a brown solid in 80% yield. **R_f**: 0.31 (EtOAc/pentane = 1:1). **IR** (neat): 3026, 2916, 1673, 1472, 1406, 1154, 915, 741 cm^{-1} . **¹H NMR** (400 MHz, CDCl_3) δ 7.48 (dt, J = 8.0, 1.0 Hz, 1H), 7.44-7.38 (m, 2H), 7.29-7.20 (m, 3H), 7.20-7.08 (m, 3H), 7.04-6.94 (m, 3H), 5.05 (t, J = 7.5 Hz, 1H), 4.01 (dd, J = 16.5, 7.5 Hz, 1H), 3.88 (s, 3H), 3.86 (dd, J = 16.5, 7.5 Hz, 1H), 3.71 (s, 3H). **¹³C NMR**

(100 MHz, CDCl₃) δ 191.0 (s), 144.6 (s), 143.1 (s), 137.1 (s), 128.8 (d), 128.3 (2C, d), 127.9 (2C, d), 127.1 (s), 126.8 (d), 126.2 (d), 126.0 (d), 121.5 (d), 119.6 (d), 118.7 (d), 117.8 (s), 109.0 (d), 45.5 (t), 38.0 (d), 36.1 (q), 32.7 (q). Spectroscopic data were consistent with the literature data for this compound.³

3-(2-Bromophenyl)-1-(1-methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)propan-1-one (2k)



Following general procedure C. Compound **1g** (0.102 g, 0.35 mmol), *N*-methylindole (0.069 g, 0.066 mL, 0.53 mmol), Cu(NO₃)₂•3H₂O (0.008 g, 0.035 mmol), dmbpy (0.008 g, 0.042 mmol), and MeCN (5 mL). The title compound (molecular formula: C₂₂H₂₀BrN₃O, MW = 422.32 g/mol, 0.115 g) was isolated as a brown solid in 78% yield. **Rf:** 0.46 (EtOAc/pentane = 1:1). **IR** (neat): 2917, 1673, 1467, 1406, 1020, 740 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.55 (dd, *J* = 8.0, 1.3 Hz, 1H), 7.48 (dt, *J* = 8.0, 0.9 Hz, 1H), 7.32-7.20 (m, 3H), 7.17-7.11 (m, 3H), 7.06-6.94 (m, 3H), 5.51 (dd, *J* = 9.0, 6.0 Hz, 1H), 4.20 (dd, *J* = 16.6, 9.0 Hz, 1H), 3.92 (s, 3H), 3.70 (s, 3H), 3.53 (dd, *J* = 16.6, 6.0 Hz, 1H). **¹³C NMR** (100 MHz, CDCl₃) δ 190.4 (s), 143.4 (s), 143.0 (s), 137.2 (s), 132.9 (d), 129.5 (d), 128.8 (d), 127.7 (d), 127.5 (d), 127.2 (s), 127.0 (d), 126.7 (d), 124.1 (s), 121.7 (d), 119.8 (d), 118.8 (d), 116.5 (s), 109.0 (d), 44.5 (d), 37.2 (t), 36.2 (q), 32.8 (q). Spectroscopic data were consistent with the literature data for this compound.³

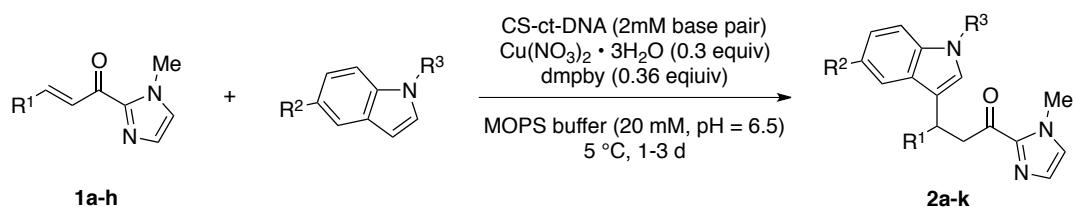
Preparation of a 20 mM MOPS buffer (pH 6.5)

209 mg of 3-(*N*-morpholino)propanesulfonic acid (MOPS, MW = 209.26 g/mol) were dissolved in 10 mL of H₂O RNase-free to obtain a 0.1 M solution. 1.9 mL of a 0.1 M solution of KOH (MW = 56.11 g/mol) in H₂O RNase-free was added followed by 38.1 mL of H₂O RNase-free.

Preparation of a 0.3 mM stock solution of [Cu(dmbpy)(NO₃)₂]

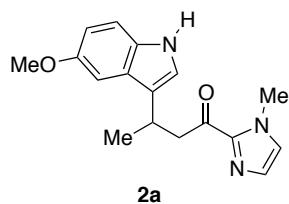
4.46 mg of Cu(NO₃)₂•3H₂O (MW = 241.60 g/mol), and 4.24 mg of 4,4'-dimethyl-2,2'-bipyridyl (dmbpy, MW = 184.24 g/mol) were dissolved in 3.3 mL of the previously prepared 20 mM MOPS buffer (pH 6.5). The mixture was stirred at rt for 5 h in a sealed tube under air. 1.8 mL of this solution were then transferred to another vial, and diluted with 8.2 mL of the 20 mM MOPS buffer (pH 6.5).

General Procedure D. Enantioselective Friedel-Crafts alkylations



To 163 mg of cellulose-supported-ct-DNA (CS-ct-DNA, 2 mM base pair) in a 20 mM MOPS buffer (400 μL) was added the 0.3 mM stock solution of [Cu(dmbpy)(NO₃)₂] (200 μL). To the mixture was then added a 0.5 M solution of enone in MeCN (1.2 μL), followed by a 2.5 M solution of substituted indole in MeCN (1.2 μL). The reaction was mixed by inversion at 5 °C in a cold room. After 1-3 d, the mixture was warmed to rt. The solution was filtered, and the CS-ct-DNA was washed with Et₂O (5 mL) and H₂O MilliQ (10 mL). The layers were separated and the aqueous phase was extracted with Et₂O (3 x 2 mL). The combined organic layers were washed with brine (2 x 5 mL), dried over Mg₂SO₄, gravity filtered and concentrated under reduced pressure to give the crude product which was subjected to SFC analysis without further purification.

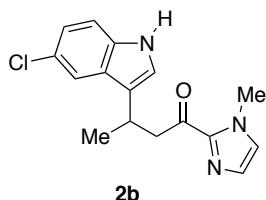
3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (**2a**)



Following general procedure D. CS-ct-DNA (2 mM base pair), **1a** (1 mM), 5-methoxyindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL), 1 d. SFC analysis of the crude residue indicated a ratio between **1a** and **2a** of 0:100 and an

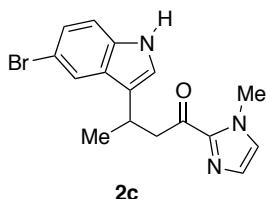
enantiomeric excess of (+) 82% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; λ = 220 nm; major enantiomer t_R = 8.56 min; minor enantiomer t_R = 5.78 min].

3-(5-Chloro-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2b)



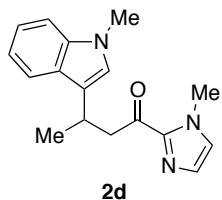
Following general procedure D. CS-ct-DNA (2 mM base pair), **1a** (1 mM), 5-chloroindole (5 mM), $[\text{Cu}(\text{dmbpy})(\text{NO}_3)_2]$ (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL), 1 d. SFC analysis of the crude residue indicated a ratio between **1a** and **2b** of 0:100 and an enantiomeric excess of (+) 66% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; λ = 220 nm; major enantiomer t_R = 7.00 min; minor enantiomer t_R = 5.80 min].

3-(5-Bromo-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2c)



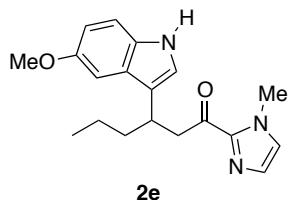
Following general procedure D. CS-ct-DNA (2 mM base pair), **1a** (1 mM), 5-bromoindole (5 mM), $[\text{Cu}(\text{dmbpy})(\text{NO}_3)_2]$ (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL), 1 d. SFC analysis of the crude residue indicated a ratio between **1a** and **2c** of 8:92 and an enantiomeric excess of (+) 66% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; λ = 220 nm; major enantiomer t_R = 7.40 min; minor enantiomer t_R = 6.14 min].

1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)-3-phenylpropan-1-one (2d)



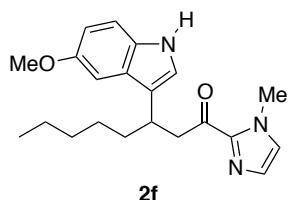
Following general procedure D. CS-ct-DNA (2 mM base pair), **1a** (1 mM), *N*-methylindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL); 1 d. SFC analysis of the crude residue indicated a ratio between **1a** and **2d** of 23:77 and an enantiomeric excess of (+) 76% [DAICEL AD-H column; 100 bar; flow: 5.0 mL/min; 15% *i*-PrOH; λ = 220 nm; major enantiomer t_R = 3.33 min; minor enantiomer t_R = 4.19 min].

3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)hexan-1-one (2e)



Following general procedure D. CS-ct-DNA (2 mM base pair), **1b** (1 mM), 5-methoxyindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL); 1 d. SFC analysis of the crude residue indicated a ratio between **1b** and **2e** of 1:99 and an enantiomeric excess of (+) 83% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 12% MeOH; λ = 220 nm; major enantiomer t_R = 6.76 min; minor enantiomer t_R = 5.47 min].

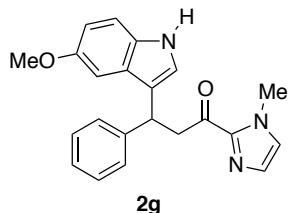
3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)octan-1-one (2f)



Following general procedure D. CS-ct-DNA (2 mM base pair), **1c** (1 mM), 5-methoxyindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL); 1 d. SFC analysis of the crude residue indicated a ratio between **1c** and **2f** of 1:99 and an enantiomeric excess of (+) 72 % [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 12%

MeOH; $\lambda = 220$ nm; major enantiomer $t_R = 6.50$ min; minor enantiomer $t_R = 5.35$ min].

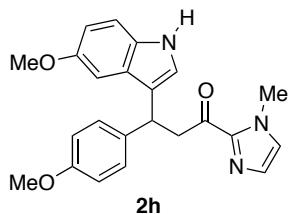
1-(1-Methyl-1*H*-imidazol-2-yl)-3-(5-methoxy-1*H*-indol-3-yl)-3-phenylpropan-1-one (2g)



2g

Following general procedure D. CS-ct-DNA (2 mM base pair), **1d** (1 mM), 5-methoxyindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μ L); 3 d. SFC analysis of the crude residue indicated a ratio between **1d** and **2g** of 1:99 and an enantiomeric excess of (+) 62% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 20% MeOH; $\lambda = 220$ nm; major enantiomer $t_R = 8.67$ min; minor enantiomer $t_R = 4.92$ min].

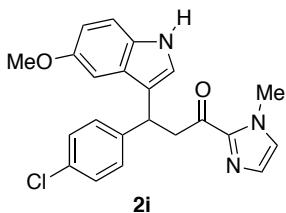
3-(5-Methoxy-1*H*-indol-3-yl)-3-(4-methoxyphenyl)-1-(1-methyl-1*H*-imidazol-2-yl)propan-1-one (2h)



2h

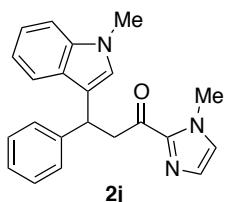
Following general procedure D. CS-ct-DNA (2 mM base pair), **1e** (1 mM), 5-methoxyindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μ L); 3 d. SFC analysis of the crude residue indicated a ratio between **1e** and **2h** of 10:90 and an enantiomeric excess of (+) 54% [DAICEL AD-H column; 100 bar; flow: 6.0 mL/min; 30% *i*-PrOH; $\lambda = 220$ nm; major enantiomer $t_R = 6.56$ min; minor enantiomer $t_R = 4.07$ min].

3-(4-Chlorophenyl)-3-(5-methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)propan-1-one (2i)



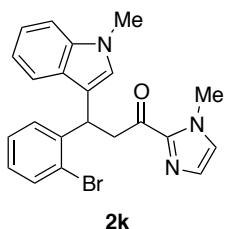
Following general procedure D. CS-ct-DNA (2 mM base pair), **1f** (1 mM), 5-methoxyindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL); 3 d. SFC analysis of the crude residue indicated a ratio between **1f** and **2i** of 1:99 and an enantiomeric excess of (+) 48% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 20% MeOH; λ = 220 nm; major enantiomer t_R = 11.77 min; minor enantiomer t_R = 6.69 min].

1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)-3-phenylpropan-1-one (2j)



Following general procedure D. CS-ct-DNA (2 mM base pair), **1d** (1 mM), *N*-methylindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL); 3 d. SFC analysis of the crude residue indicated a ratio between **1d** and **2j** of 1:99 and an enantiomeric excess of (+) 76% [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 10% MeOH; λ = 220 nm; major enantiomer t_R = 14.80 min; minor enantiomer t_R = 12.33 min].

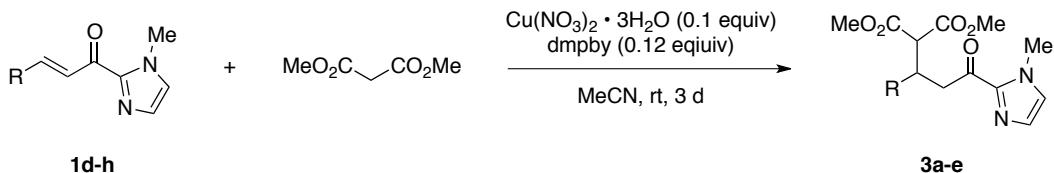
3-(2-Bromophenyl)-1-(1-methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)propan-1-one (2k)



Following general procedure D. CS-ct-DNA (2 mM base pair), **1g** (1 mM), *N*-methylindole (5 mM), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL); 3 d.

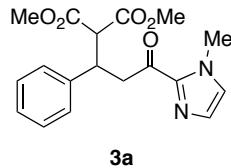
SFC analysis of the crude residue indicated a ratio between **1g** and **2k** of 39:61 and an enantiomeric excess of (+) 65% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 10% MeOH; $\lambda = 220$ nm; major enantiomer $t_R = 11.36$ min; minor enantiomer $t_R = 9.74$].

General Procedure E. Racemic Michael Additions



An oven-dried 10 mL round-bottomed flask was charged with $\text{Cu}(\text{NO}_3)_2 \cdot 3\text{H}_2\text{O}$ (0.066 mmol, 0.2 equiv), 4,4'-dimethyl-2,2'-bipyridyl (dmbpy, 0.073 mmol, 0.22 equiv), and MeCN (1 mL). The mixture was stirred at rt for 10 min then the α,β -unsaturated substrate (0.33 mmol, 1.0 equiv), and dimethylmalonate (6.65 mmol, 20 equiv) were added. The resulting solution was stirred at rt for an additional 3 d. The reaction was then diluted with brine (10 mL) and extracted with Et_2O (3 x 5 mL). The combined organic layers were dried over anhydrous MgSO_4 , gravity filtered and concentrated under reduced pressure. The crude product was purified by flash chromatography over silica gel using EtOAc/pentane (1:1) as the eluent.

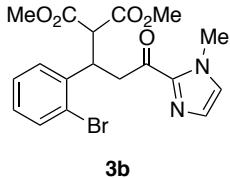
Dimethyl 2-(3-(1-methyl-1*H*-imidazol-2-yl)-3-oxo-1-phenylpropyl)malonate (3a)



Following general procedure E. Compound **1d** (0.070 g, 0.33 mmol), dimethylmalonate (0.878 g, 0.760 mL, 6.65 mmol), $\text{Cu}(\text{NO}_3)_2 \cdot 3\text{H}_2\text{O}$ (0.016 g, 0.066 mmol), dmbpy (0.013 g, 0.073 mmol) and MeCN (1 mL). The title compound (molecular formula: $\text{C}_{18}\text{H}_{20}\text{N}_2\text{O}_5$, MW = 344.36 g/mol, 0.046 g) was isolated as a colorless oil in 40% yield. **Rf:** 0.47 (EtOAc/pentane = 1:1). **IR** (neat): 3030, 2954, 1733, 1676, 1408, 1255, 1153, 736, 701 cm^{-1} . **¹H NMR** (400 MHz, CDCl_3) δ 7.32-7.20 (m, 4H), 7.19-7.13 (m, 1H), 7.07 (d, $J = 0.9$ Hz, 1H), 6.94 (s, 1H), 4.16 (td, $J = 10.0, 4.4$ Hz, 1H), 3.85 (s, 3H), 3.84-3.74 (m, 2H), 3.71 (s, 3H), 3.49-3.40 (m, 4H). **¹³C NMR** (100 MHz, CDCl_3) δ 189.6 (s), 168.5 (s), 168.0 (s), 142.8 (s), 140.4 (s), 128.9 (d), 128.3 (2C, d), 128.2 (2C, d), 127.0 (d), 126.8 (d), 57.6 (d), 52.6 (q), 52.3 (q), 42.7 (t), 40.3 (d), 36.0 (q). Spectroscopic data were consistent with the

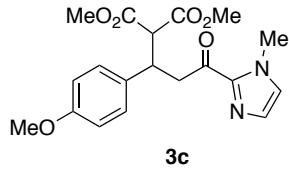
literature data for this compound.⁶

Dimethyl 2-(1-(2-bromophenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate (3b)



Following general procedure E. Compound **1g** (0.096 g, 0.33 mmol), dimethylmalonate (0.878 g, 0.760 mL, 6.65 mmol), Cu(NO₃)₂•3H₂O (0.016 g, 0.066 mmol), dmbpy (0.013 g, 0.073 mmol), and MeCN (1 mL). The title compound (molecular formula: C₁₈H₁₉BrN₂O₅, MW = 423.26 g/mol, 0.061 g) was isolated as a colorless oil in 44% yield. **Rf:** 0.45 (EtOAc/pentane = 1:1). **IR** (neat): 2953, 1735, 1677, 1409, 1254, 1155, 1022, 761 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.51 (dd, *J* = 8.0, 1.0 Hz, 1H), 7.30 (dd, *J* = 7.8, 1.4 Hz, 1H), 7.23-7.11 (m, 1H), 7.07 (s, 1H), 7.01 (td, *J* = 7.8, 1.4 Hz, 1H), 6.95 (s, 1H), 4.62 (td, *J* = 8.9, 4.6 Hz, 1H), 4.05-3.88 (m, 2H), 3.86 (s, 3H), 3.66 (s, 3H), 3.62 (dd, *J* = 17.8, 4.6 Hz, 1H), 3.55 (s, 3H). **¹³C NMR** (100 MHz, CDCl₃) δ 189.5 (s), 168.3 (s), 168.0 (s), 142.7 (s), 139.6 (s), 133.3 (d), 128.9 (d), 128.5 (2C, d), 127.5 (d), 126.9 (d), 125.0 (s), 55.6 (d), 52.5 (q), 52.4 (q), 41.3 (t), 38.9 (d), 36.1 (q). Spectroscopic data were consistent with the literature data for this compound.⁶

Dimethyl 2-(1-(4-methoxyphenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl) malonate (3c)

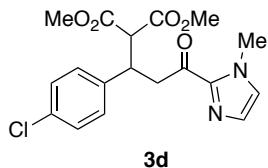


Following general procedure E. Compound **1e** (0.080 g, 0.33 mmol), dimethylmalonate (0.878 g, 0.760 mL, 6.65 mmol), Cu(NO₃)₂•3H₂O (0.016 g, 0.066 mmol), dmbpy (0.013 g, 0.073 mmol) and MeCN (1 mL). The title compound (molecular formula: C₁₉H₂₂N₂O₆, MW = 374.39 g/mol, 0.053 g) was isolated as a colorless oil in 43% yield. **Rf:** 0.49 (EtOAc/pentane = 1:1). **IR** (neat): 3002, 2954, 2838, 1735, 1675, 1514, 1410, 1250, 1030, 776 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.25-7.17 (m, 2H), 7.09 (d, *J* = 0.9 Hz, 1H), 6.95 (d,

⁶ Coquiere, D.; Feringa, B. L.; Roelfes, G. *Angew. Chem., Int. Ed.* **2007**, *46*, 9308-9311.

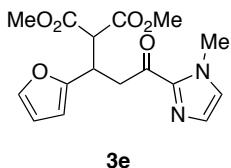
J = 0.9 Hz, 1H), 6.81-6.74 (m, 2H), 4.11 (td, *J* = 10.1, 4.4 Hz, 1H), 3.87 (s, 3H), 3.79-3.74 (m, 2H), 3.74 (s, 3H), 3.72 (s, 3H), 3.47 (s, 3H), 3.41 (dd, *J* = 17.4, 4.4 Hz, 1H). **¹³C NMR** (100 MHz, CDCl₃) δ 189.7 (s), 168.6 (s), 168.2 (s), 158.4 (s), 142.8 (s), 132.4 (s), 129.3 (2C, d), 128.8 (d), 126.8, (d) 113.8 (2C, d), 57.8 (q), 55.1 (d), 52.6 (q), 52.3 (q), 42.9 (t), 39.6 (q), 36.0 (d). Spectroscopic data were consistent with the literature data for this compound.⁶

Dimethyl 2-(1-(4-chlorophenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate (3d)



Following general procedure E. Compound **1f** (0.081 g, 0.33 mmol), dimethylmalonate (0.878 g, 0.760 mL, 6.65 mmol), Cu(NO₃)₂•3H₂O (0.016 g, 0.066 mmol), dmbpy (0.013 g, 0.073 mmol), and MeCN (1 mL). The title compound (molecular formula: C₁₈H₁₉ClN₂O₅, MW = 378.81 g/mol, 0.051 g) was isolated as a colorless oil in 41% yield. **Rf:** 0.42 (EtOAc/pentane = 1:1). **IR** (neat): 2953, 1733, 1674, 1408, 1255, 1154, 1014, 914, 730 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.26-7.18 (m, 4H), 7.08 (d, *J* = 0.9 Hz, 1H), 6.98-6.94 (m, 1H), 4.13 (td, *J* = 10.1, 4.3 Hz, 1H), 3.86 (s, 3H), 3.86-3.74 (m, 2H), 3.72 (s, 3H), 3.48 (s, 3H), 3.41 (dd, *J* = 17.5, 4.3 Hz, 1H). **¹³C NMR** (100 MHz, CDCl₃) δ 189.3 (s), 168.3 (s), 167.9 (s), 142.7 (s), 139.0 (s), 132.8 (s), 129.7 (2C, d), 129.0 (d), 128.5 (2C, d), 127.0 (d), 57.3 (d), 52.7 (q), 52.4 (q), 42.6 (t), 39.7 (d), 36.0 (q). Spectroscopic data were consistent with the literature data for this compound.⁶

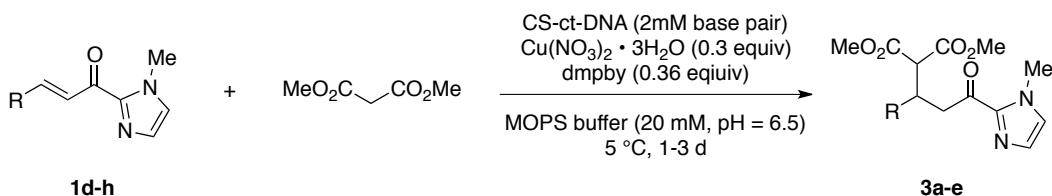
Dimethyl 2-(1-(furan-2-yl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate (3f)



Following general procedure E. Compound **1h** (0.067 g, 0.33 mmol), dimethylmalonate (0.878 g, 0.760 mL, 6.65 mmol), Cu(NO₃)₂•3H₂O (0.016 g, 0.066 mmol), dmbpy (0.013 g, 0.073 mmol), and MeCN (1 mL). The title compound (molecular formula: C₁₆H₁₈N₂O₆, MW = 334.32 g/mol, 0.039 g) was isolated as a colorless oil in 35% yield. **Rf:** 0.44

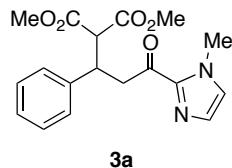
(EtOAc/pentane = 1:1). **IR** (neat): 3115, 2955, 1736, 1677, 1411, 1257, 1157, 1014, 915, 741 cm⁻¹. **¹H NMR** (400 MHz, CDCl₃) δ 7.29-7.24 (m, 1H), 7.12 (d, *J* = 0.9 Hz, 1H), 7.00 (s, 1H), 6.22 (dd, *J* = 3.2, 1.8 Hz, 1H), 6.14 (dt, *J* = 3.2, 0.7 Hz, 1H), 4.29 (td, *J* = 9.2, 4.4 Hz, 1H), 3.94 (s, 3H), 3.91-3.80 (m, 2H), 3.72 (s, 3H), 3.62 (s, 3H), 3.45 (dd, *J* = 17.7, 4.4 Hz, 1H). **¹³C NMR** (100 MHz, CDCl₃) δ 189.4 (s), 168.2 (s), 168.1 (s), 153.5 (s), 142.7 (s), 141.7 (d), 129.1 (d), 126.9 (d), 110.1 (d), 106.9 (d), 55.2 (d), 52.7 (q), 52.6 (q), 40.2 (t), 36.1 (q), 33.9 (d). Spectroscopic data were consistent with the literature data for this compound.⁶

General Procedure F. Enantioselective Michael Additions



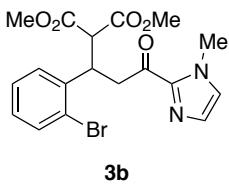
To 163 mg of CS-ct-DNA in a 20 mM MOPS buffer (400 μL) was added the 0.3 mM stock solution of [Cu(dmbpy)(NO₃)₂] (200 μL). To the resulting mixture was added a 0.5 M solution of enone in MeCN (1.2 μL), followed by dimethyl malonate (6.9 μL). The reaction was mixed by inversion at 5 °C in a cold room. After 1-3 d, the solution was filtered, and the CS-ct-DNA was washed with Et₂O (5 mL), and H₂O MilliQ (10 mL). The layers were separated and the aqueous phase was extracted with Et₂O (3 x 2 mL). The combined organic layers were washed with brine (2 x 5 mL), dried over Mg₂SO₄, gravity filtered and concentrated under reduced pressure to give the crude product which was subjected to SFC analysis without further purification.

Dimethyl 2-(3-(1-methyl-1*H*-imidazol-2-yl)-3-oxo-1-phenylpropyl)malonate (**3a**)



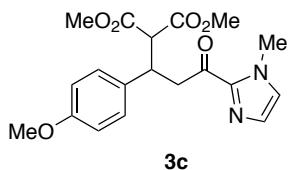
Following general procedure F. CS-ct-DNA (2 mM base pair), **1d** (1 mM), dimethyl malonate (6.9 μL), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μL); 3 d. SFC analysis of the crude residue indicated a ratio between **1d** and **3a** of 2:98 and an enantiomeric excess of (+) 97% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 10% MeOH; λ = 220 nm; major enantiomer t_R = 3.55 min; minor enantiomer t_R = 2.85 min].

Dimethyl 2-(1-(2-bromophenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl) malonate (3b)



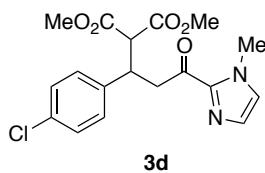
Following general procedure F. CS-ct-DNA (2 mM base pair), **1g** (1 mM), dimethyl malonate (6.9 μ L), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μ L); 3 d. SFC analysis of the crude residue indicated a ratio between **1g** and **3b** of 2:98 and an enantiomeric excess of (+) 96% [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 7% MeOH; λ = 220 nm; major enantiomer t_R = 7.40 min; minor enantiomer t_R = 8.70 min].

Dimethyl 2-(1-(4-methoxyphenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl) malonate (3c)



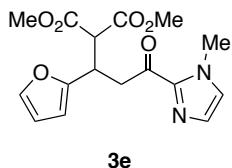
Following general procedure F. CS-ct-DNA (2 mM base pair), **1e** (1 mM), dimethyl malonate (6.9 μ L), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μ L); 3 d. SFC analysis of the crude residue indicated a ratio between **1e** and **3c** of 1:99 and an enantiomeric excess of (+) 81% [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 10% MeOH; λ = 220 nm; major enantiomer t_R = 7.44 min; minor enantiomer t_R = 6.83 min].

Dimethyl 2-(1-(4-chlorophenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl) malonate (3d)



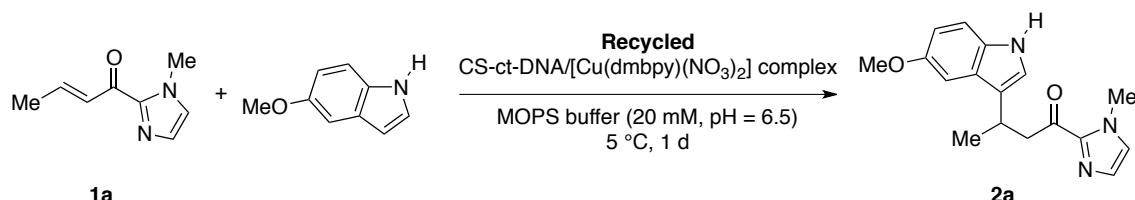
Following general procedure F. CS-ct-DNA (2 mM base pair), **1f** (1 mM), dimethyl malonate (32 μ L), [Cu(dmbpy)(NO₃)₂] (0.3 mM), and MOPS buffer (20 mM, pH 6.5, 600 μ L); 1 d. SFC analysis of the crude residue indicated a ratio between **1f** and **3d** of 11:89 and an enantiomeric excess of (+) 93% [DAICEL AD-H column; 100 bar; flow: 6.0 mL/min; 30% *i*-PrOH; λ = 220 nm; major enantiomer t_R = 3.73 min; minor enantiomer t_R = 3.03 min].

Dimethyl 2-(1-(furan-2-yl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate (3e**)**



Following general procedure F. CS-ct-DNA (2 mM base pair), **1h** (1 mM), dimethyl malonate (6.9 μ L), $[\text{Cu}(\text{dmbpy})(\text{NO}_3)_2]$ (0.3 mM) and MOPS buffer (20 mM, pH 6.5, 600 μ L); 3 d. SFC analysis of the crude residue indicated a ratio between **1h** and **3e** of 72:28 and an enantiomeric excess of (+) 89% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; major enantiomer $t_R = 1.99$ min; minor enantiomer $t_R = 1.73$ min].

General Procedure G. Enantioselective Friedel Crafts alkylations - Recycling of CS-ct-DNA with MOPS buffer

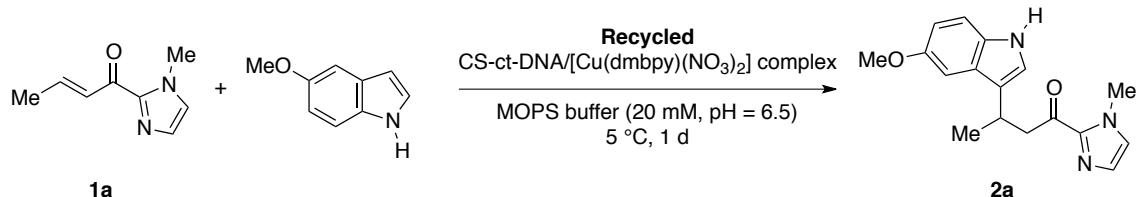


To 163 mg of CS-ct-DNA in a 20 mM MOPS buffer (400 μ L) was added the 0.3 mM stock solution of $[\text{Cu}(\text{dmbpy})(\text{NO}_3)_2]$ (200 μ L). To the resulting mixture was added a 0.5 M solution of **1a** in MeCN (1.2 μ L), followed by a 2.5 M solution of 5-methoxyindole (1.2 μ L). The reaction was mixed by inversion at 5 °C in a cold room. After 1 d, the solution was warmed to rt. The CS-ct-DNA was filtered off, and washed with a 20 mM MOPS buffer (3 x 5 mL). The aqueous solution was then extracted with Et₂O (2 x 5 mL), the combined organic layers were washed with brine (2 x 5 mL), dried over Mg₂SO₄, gravity filtered, and concentrated under reduced pressure. The crude product was subjected to SFC analysis without further purification. The CS-ct-DNA was recovered and used without any further treatment in the next run simply by adding a 20 mM MOPS buffer (600 μ L), compound **1a** (0.5 M solution in MeCN, 1.2 μ L), and 5-methoxyindole (2.5 M solution in MeCN, 1.2 μ L). Every recycling run was performed with no additional $[\text{Cu}(\text{dmbpy})(\text{NO}_3)_2]$. The reaction was achieved ten times, and the SFC analyses of the crude residues indicated a complete conversion of the starting material into the product. The observed enantiomeric excesses are reported in the following table.

Number of runs	<i>e</i> es (%)
1 st run	80
2 nd run	81
3 rd run	80
4 th run	79
5 th run	79
6 th run	79
7 th run	78
8 th run	77
9 th run	76
10 th run	76

SFC conditions: [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm].

General Procedure H. Enantioselective Friedel Crafts alkylations – Recycling of CS-ct-DNA with a MOPS buffer/MeOH 30:1 solution



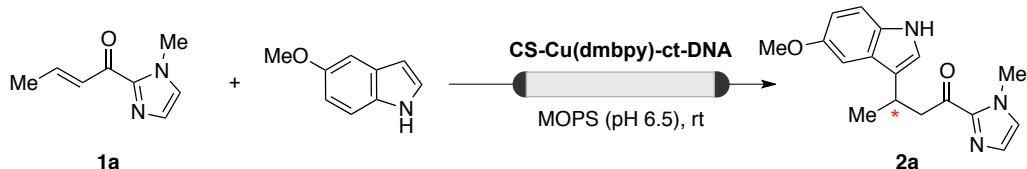
To 163 mg of CS-ct-DNA in a 20 mM MOPS buffer/MeOH 20:1 solution (400 μ L) was added the 0.3 mM stock solution of [Cu(dmbpy)(NO₃)₂] (200 μ L). To the resulting mixture was added a 0.5 M solution of **1a** in MeCN (1.2 μ L), followed by a 2.5 M solution of 5-methoxyindole (1.2 μ L). The reaction was mixed by inversion at 5 °C in a cold room. After 1 d, the solution was warmed to rt. The CS-ct-DNA was filtered off, and washed with a 20 mM MOPS buffer/MeOH 30:1 solution (3 x 5 mL). The aqueous solution was then extracted with Et₂O (2 x 5 mL), the combined organic layers were washed with brine (2 x 5 mL), dried over Mg₂SO₄, gravity filtered, and concentrated under reduced pressure. The crude product was subjected to SFC analysis without further purification. The CS-ct-DNA was recovered and used without any further treatment in the next run simply by adding a 20 mM MOPS buffer/MeOH 30:1 solution (600 μ L), compound **1a** (0.5 M solution in MeCN, 1.2 μ L), and 5-methoxyindole (2.5 M solution in MeCN, 1.2 μ L). Every recycling run was performed with no additional [Cu(dmbpy)(NO₃)₂]. The reaction was achieved ten times, and the SFC analyses of the crude

residues indicated a complete conversion of the starting material into the product. The observed enantiomeric excesses are reported in the following table.

Number of runs	<i>ees</i> (%)
1 st run	82
2 nd run	82
3 rd run	80
4 th run	80
5 th run	76
6 th run	74
7 th run	76
8 th run	75
9 th run	75
10 th run	75

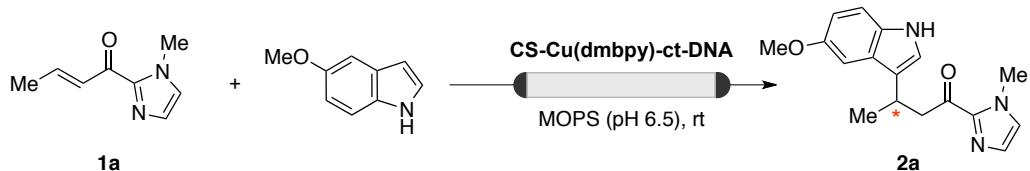
SFC conditions: [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm].

General Procedure I. Enantioselective Friedel Crafts alkylations in a continuous flow process.



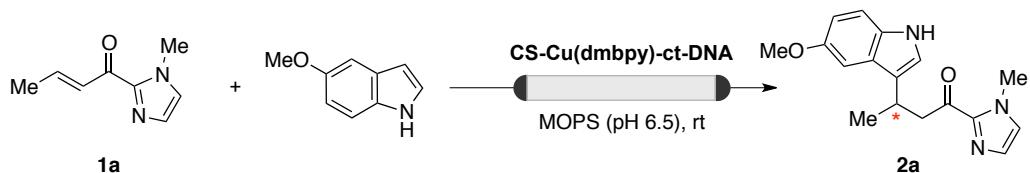
In a 4 g MPLC plastic cartridge ($L = 65$ mm, $\varnothing = 13$ mm) was packed 1.1 g of the preformed CS-ct-DNA-Cu(dmbpy) biohybrid catalyst [prepared by mixing CS-ct-DNA with a 0.3 mM solution of $[\text{Cu}(\text{dmbpy})(\text{NO}_3)_2]$ in a 20 mM MOPS buffer (1.35 mL)]. The cartridge was connected to a syringe-pump and washed with a 20 mM MOPS buffer (50 mL) at a 1 mL/min flow-rate. The syringe-pump was then loaded with a 5:1 mixture of 5-methoxyindole (0.15 mmol) and **1a** (0.03 mmol) in a 20 mM MOPS buffer/MeOH 30:1 solution (31 mL). The reagents were then passed through the column at a 0.25 mL/min flow-rate followed by a 20 mM MOPS buffer/MeOH 30:1 solution (25 mL). The resulting solution was then extracted with Et_2O (3 x 15 mL), the combined organic layers were washed with brine (2 x 10 mL), dried over anhydrous MgSO_4 , gravity filtered, and concentrated under reduced pressure to afford a crude residue that was subjected to SFC analysis without further purification.

(2a): 0.25 mL/min flow rate, 1.1 g of CS-ct-DNA



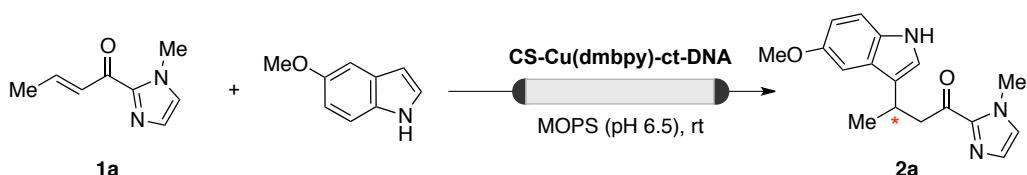
Following general procedure I. CS-ct-DNA (2 mM base pair) (1.1 g), **1a** (0.03 mmol), 5-methoxyindole (0.15 mmol) in a MOPS/MeOH 30:1 solution (31 mL); [Cu(dmbpy)(NO₃)₂] (0.3 mM) in a MOPS buffer (20 mM, pH 6.5, 1.35 mL). Flow rate: 0.25 mL/min. SFC analysis of the crude residue indicated a ratio between **1a** and **2a** of 50:50 and an enantiomeric excess of (+) 81% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; major enantiomer $t_R = 8.58$ min; minor enantiomer $t_R = 5.74$ min].

(2a): 0.125 mL/min flow rate, 2.2 g of CS-ct-DNA



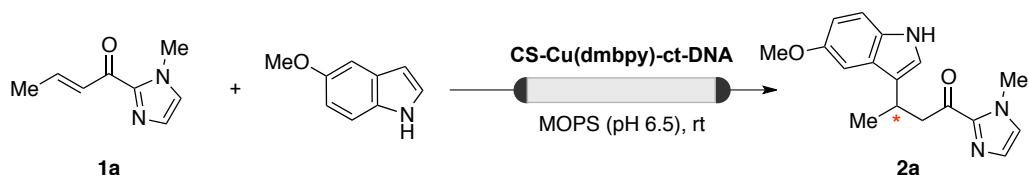
Following general procedure I. CS-ct-DNA (2 mM base pair) (2.2 g), **1a** (0.03 mmol), 5-methoxyindole (0.15 mmol) in a MOPS/MeOH 30:1 solution (31 mL); [Cu(dmbpy)(NO₃)₂] (0.3 mM) in a MOPS buffer (20 mM, pH 6.5, 2.70 mL). Flow rate: 0.125 mL/min. SFC analysis of the crude residue indicated a ratio between **1a** and **2a** of 10:90 and an enantiomeric excess of (+) 82% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; major enantiomer $t_R = 8.47$ min; minor enantiomer $t_R = 5.68$ min].

(2a): 0.125 mL/min flow rate, 4.4 g of CS-ct-DNA



Following general procedure I. CS-ct-DNA (2 mM base pair) (4.4 g), **1a** (0.03 mmol), 5-methoxyindole (0.15 mmol) in a MOPS/MeOH 30:1 solution (31 mL); [Cu(dmbpy)(NO₃)₂] (0.3 mM) in a MOPS buffer (20 mM, pH 6.5, 5.40 mL). Flow rate: 0.125 mL/min. SFC analysis of the crude residue indicated a ratio between **1a** and **2a** of 1.5:98.5 and an enantiomeric excess of (+) 79% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; major enantiomer $t_R = 9.41$ min; minor enantiomer $t_R = 6.28$ min]. The crude was purified using petroleum ether/EtOAc (3:2). The title compound was obtained as a brownish oil (9.1 mg, 92 %).

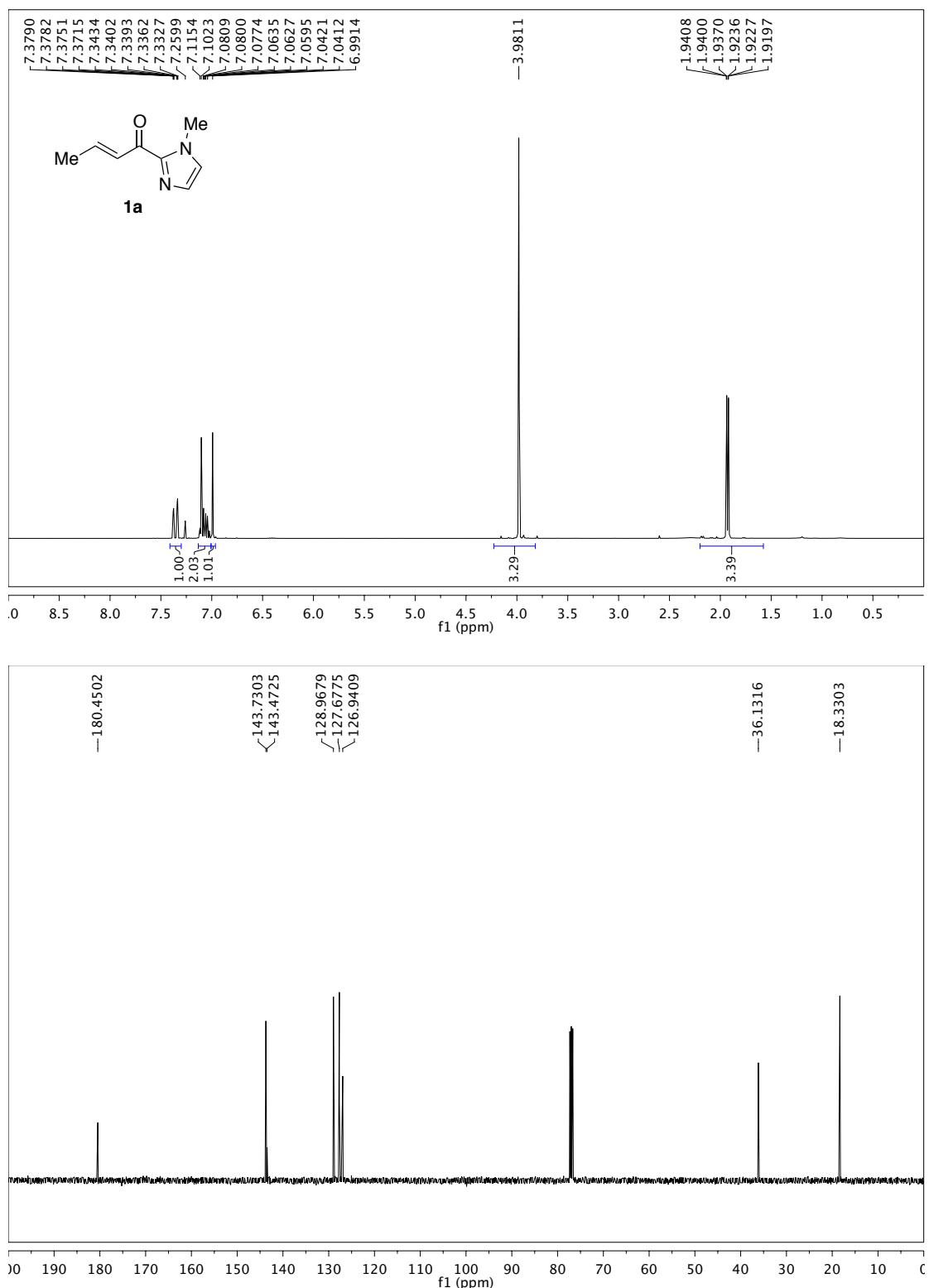
(2a): 0.125 mL/min flow rate, 4.4 g of CS-ct-DNA



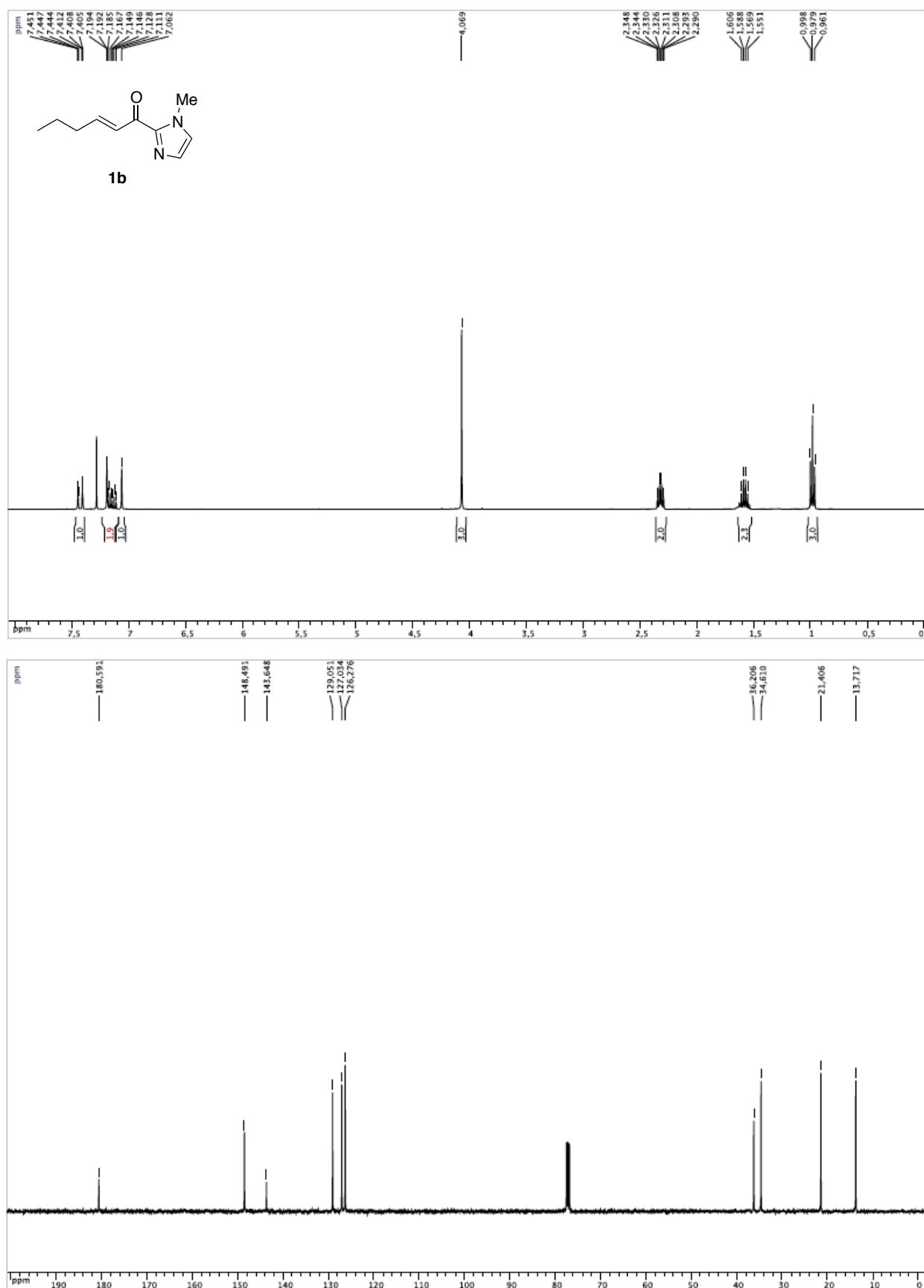
Following general procedure I. CS-ct-DNA (2 mM base pair) (4.4 g), **1a** (0.3 mmol), 5-methoxyindole (1.5 mmol) in a MOPS/MeOH 30:1 solution (310 mL); [Cu(dmbpy)(NO₃)₂] (0.3 mM) in a MOPS buffer (20 mM, pH 6.5, 5.40 mL). Flow rate: 0.125 mL/min. SFC analysis of the crude residue indicated a ratio between **1a** and **2a** of 2:98 and an enantiomeric excess of (+) 78% [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; major enantiomer $t_R = 8.51$ min; minor enantiomer $t_R = 5.76$ min]. The crude was purified using petroleum ether/EtOAc (3:2). The title compound was obtained as a brownish oil (79.2 mg, 89 %).

Copies of ^1H and ^{13}C NMR Spectra

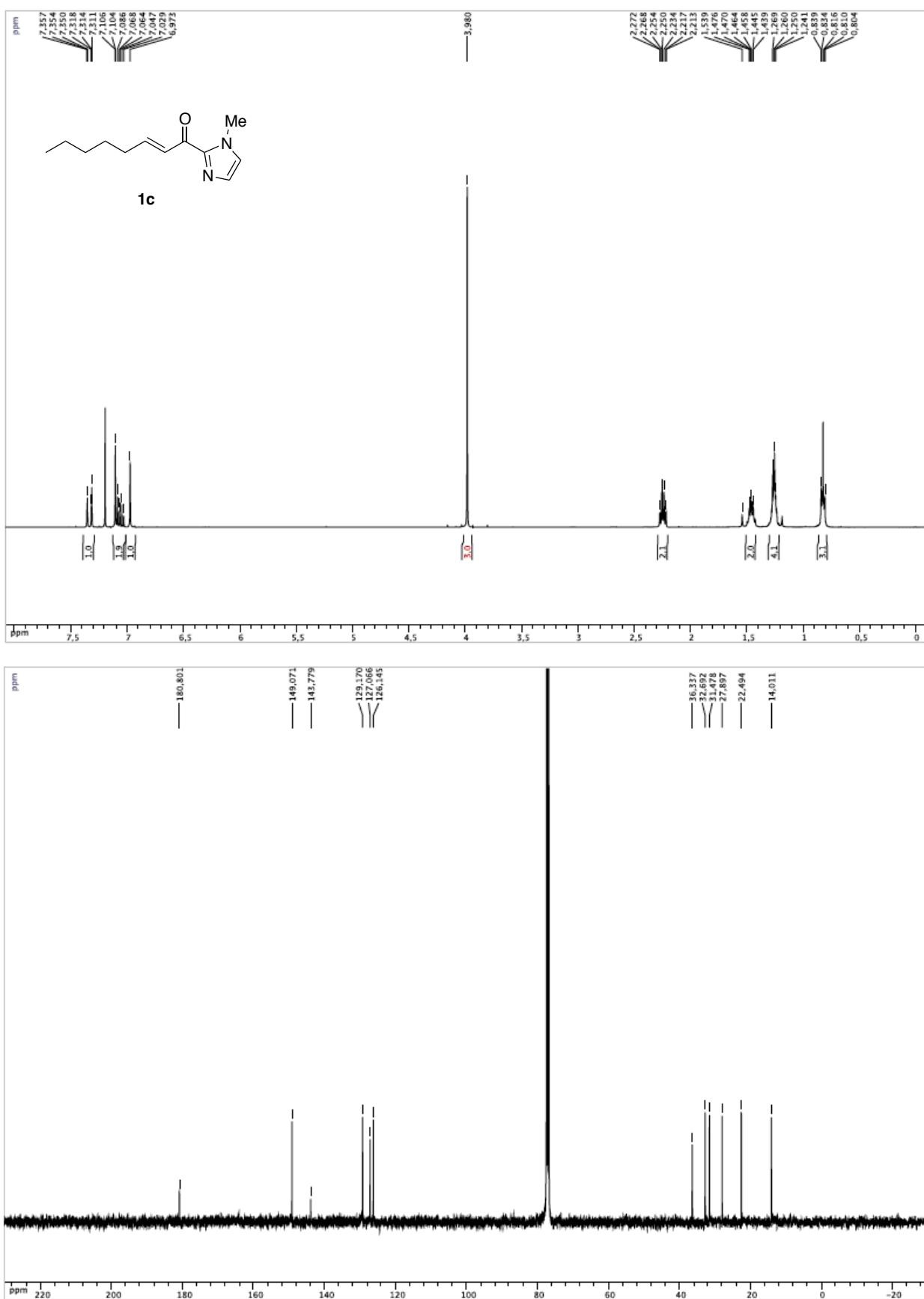
(E)-1-(1-Methyl-1*H*-imidazol-2-yl)but-2-en-1-one (**1a**)



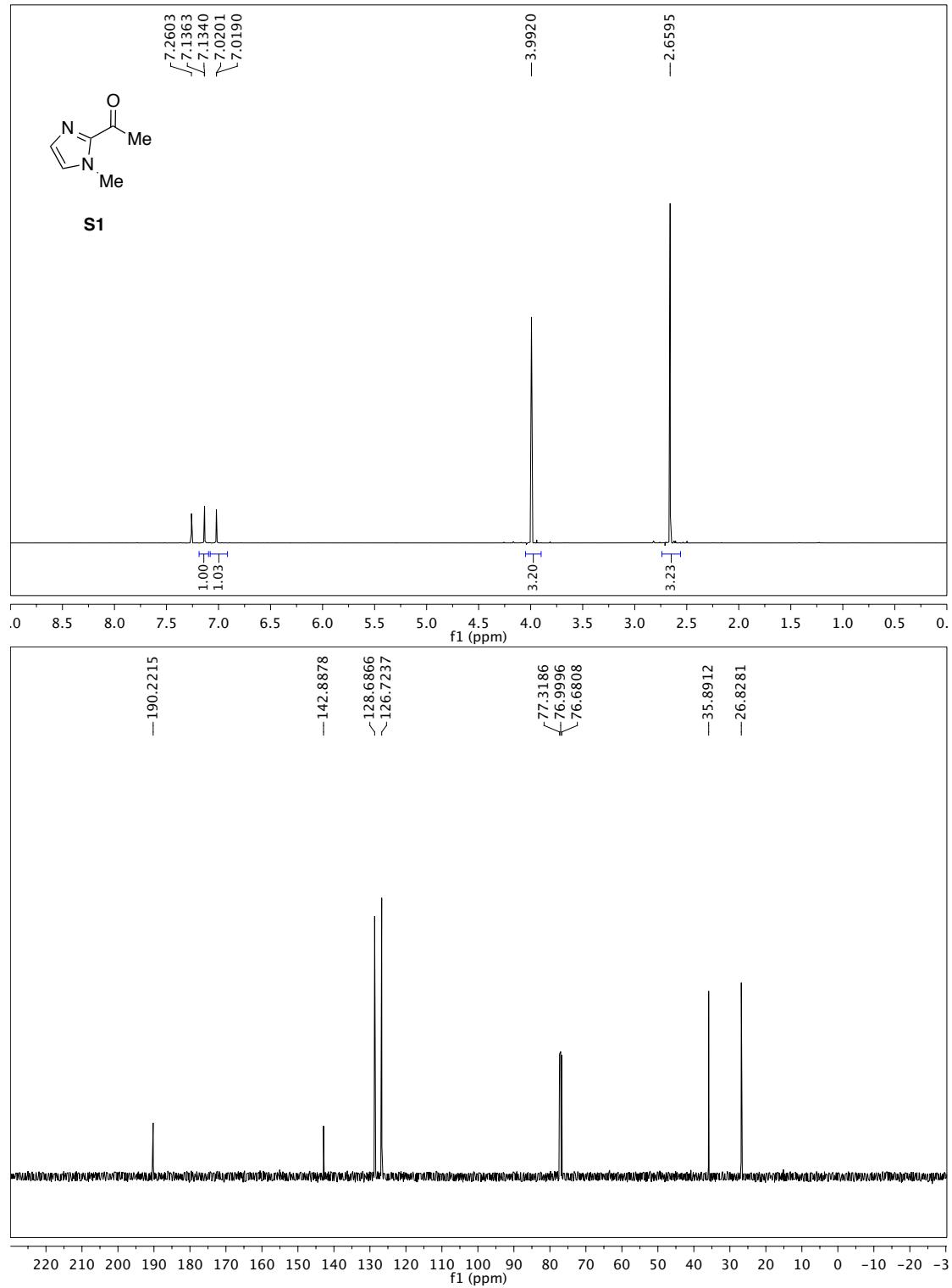
(E)-1-(1-Methyl-1*H*-imidazol-2-yl)hex-2-en-1-one (1b)



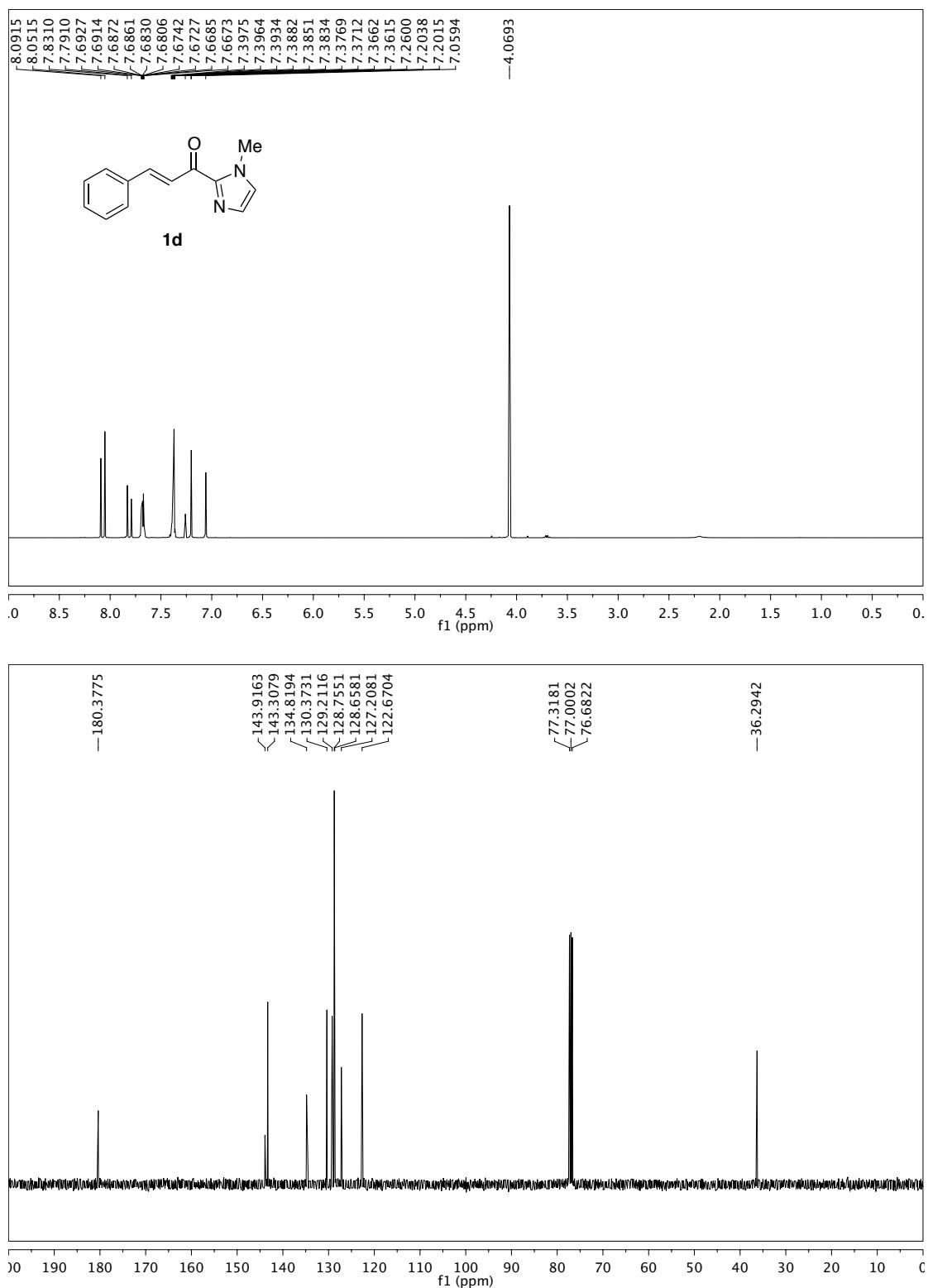
(E)-1-(1-Methyl-1*H*-imidazol-2-yl)oct-2-en-1-one (1c)



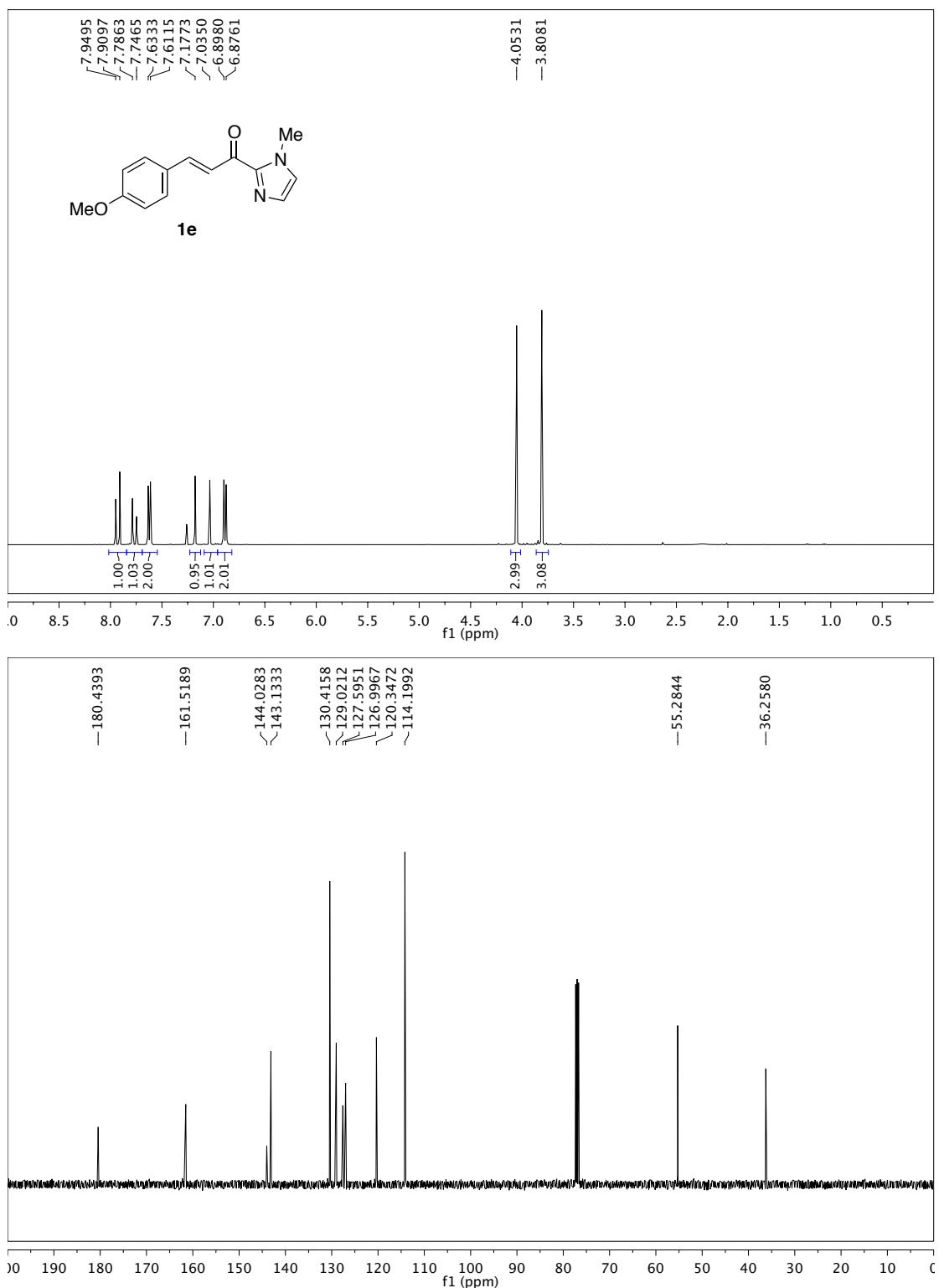
1-(1-methyl-1*H*-imidazol-2-yl)ethanone (S1**)**



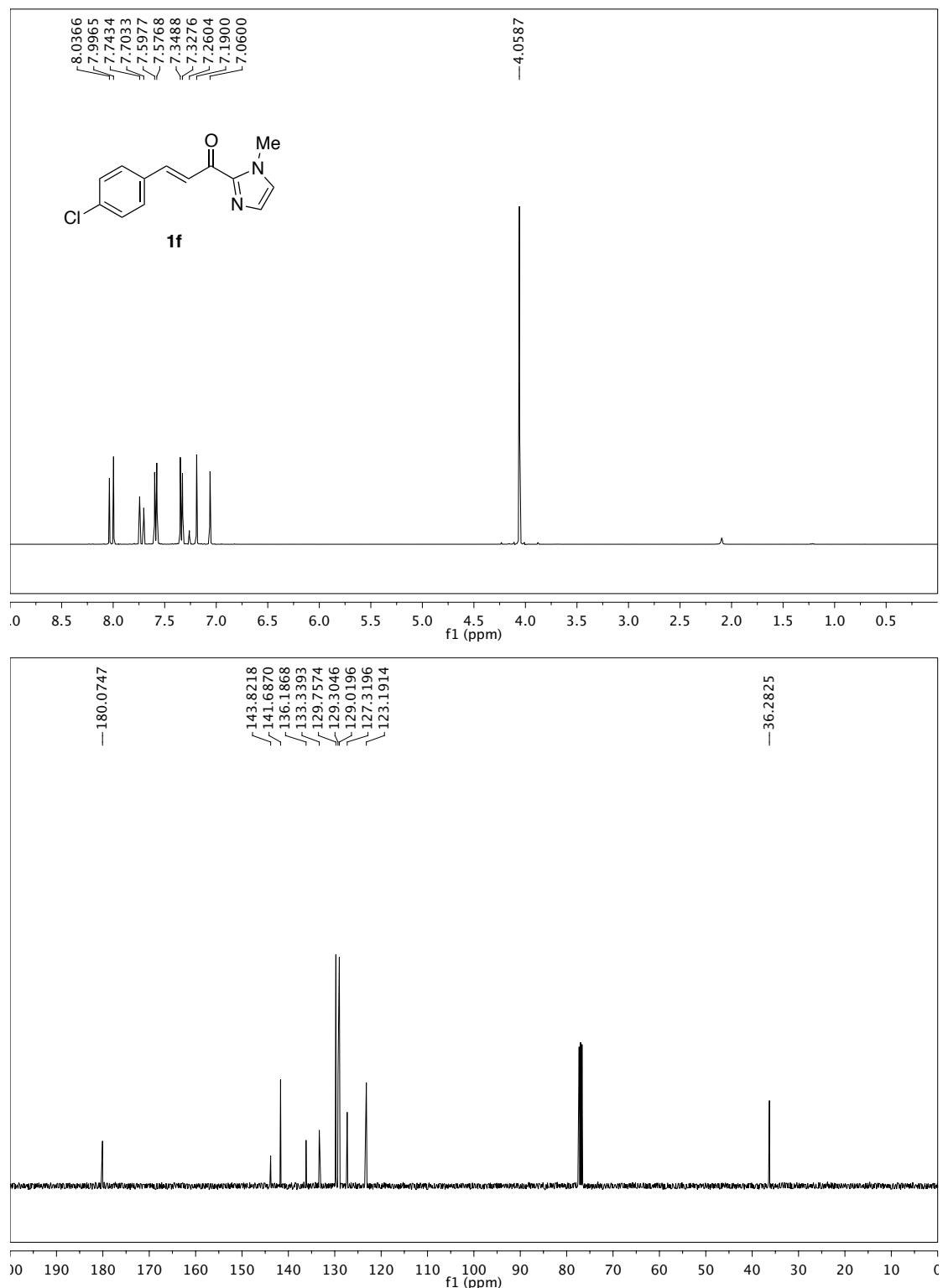
(E)-1-(1-Methyl-1*H*-imidazol-2-yl)-3-phenylprop-2-en-1-one (1d**)**



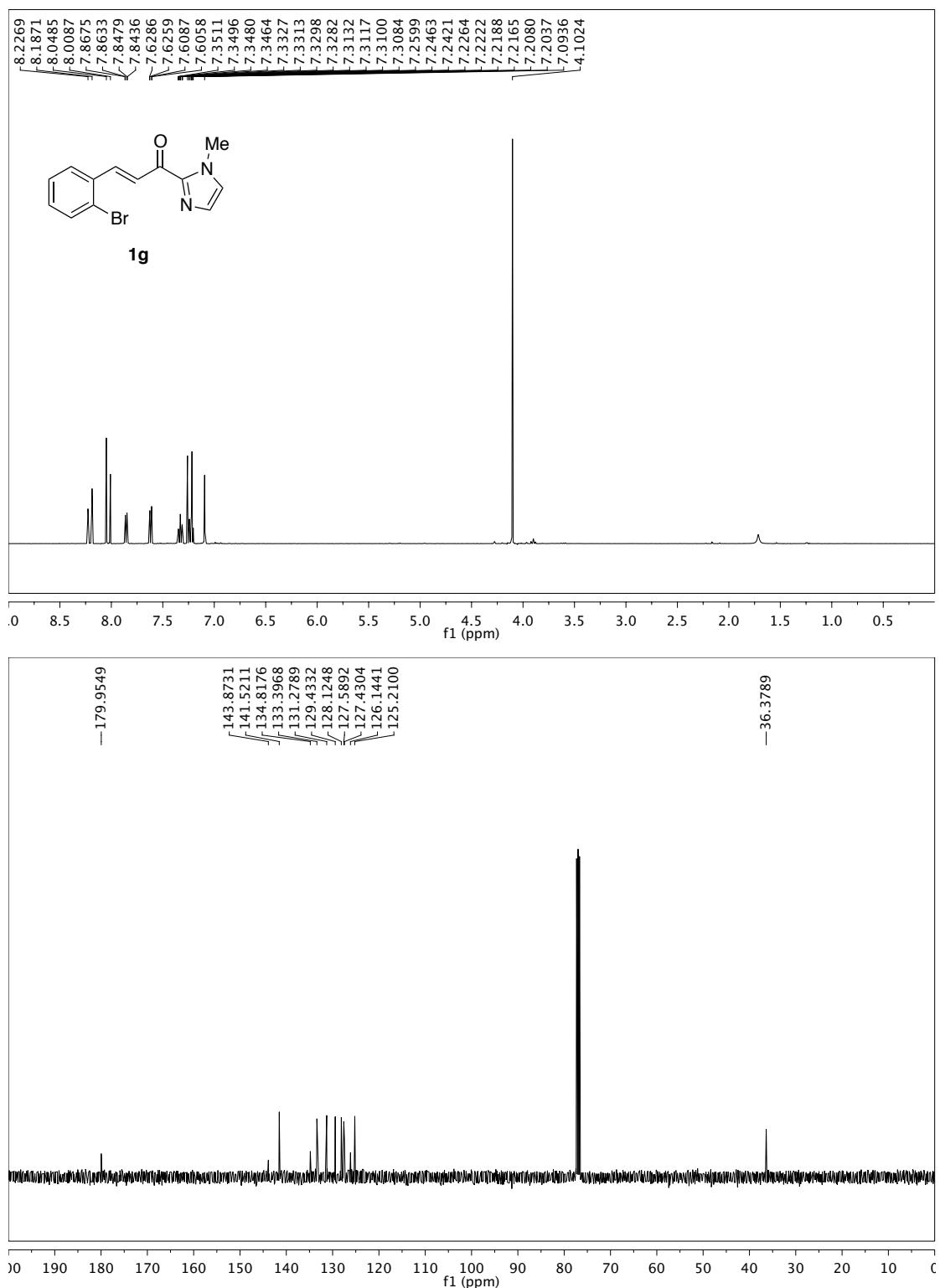
(E)-3-(4-Methoxyphenyl)-1-(1-methyl-1*H*-imidazol-2-yl)prop-2-en-1-one (1e)



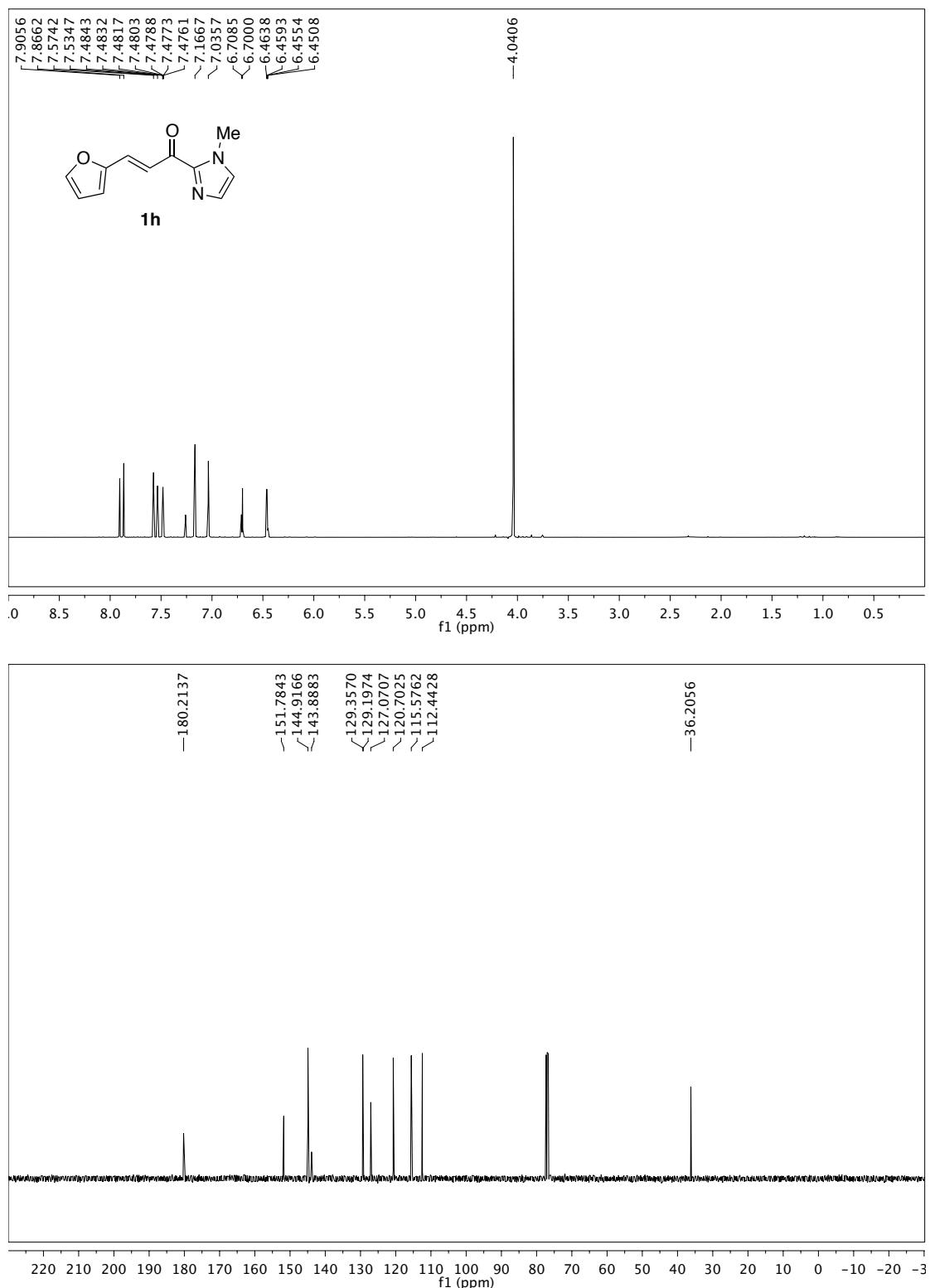
(E)-3-(4-Chlorophenyl)-1-(1-methyl-1*H*-imidazol-2-yl)prop-2-en-1-one (1f**)**



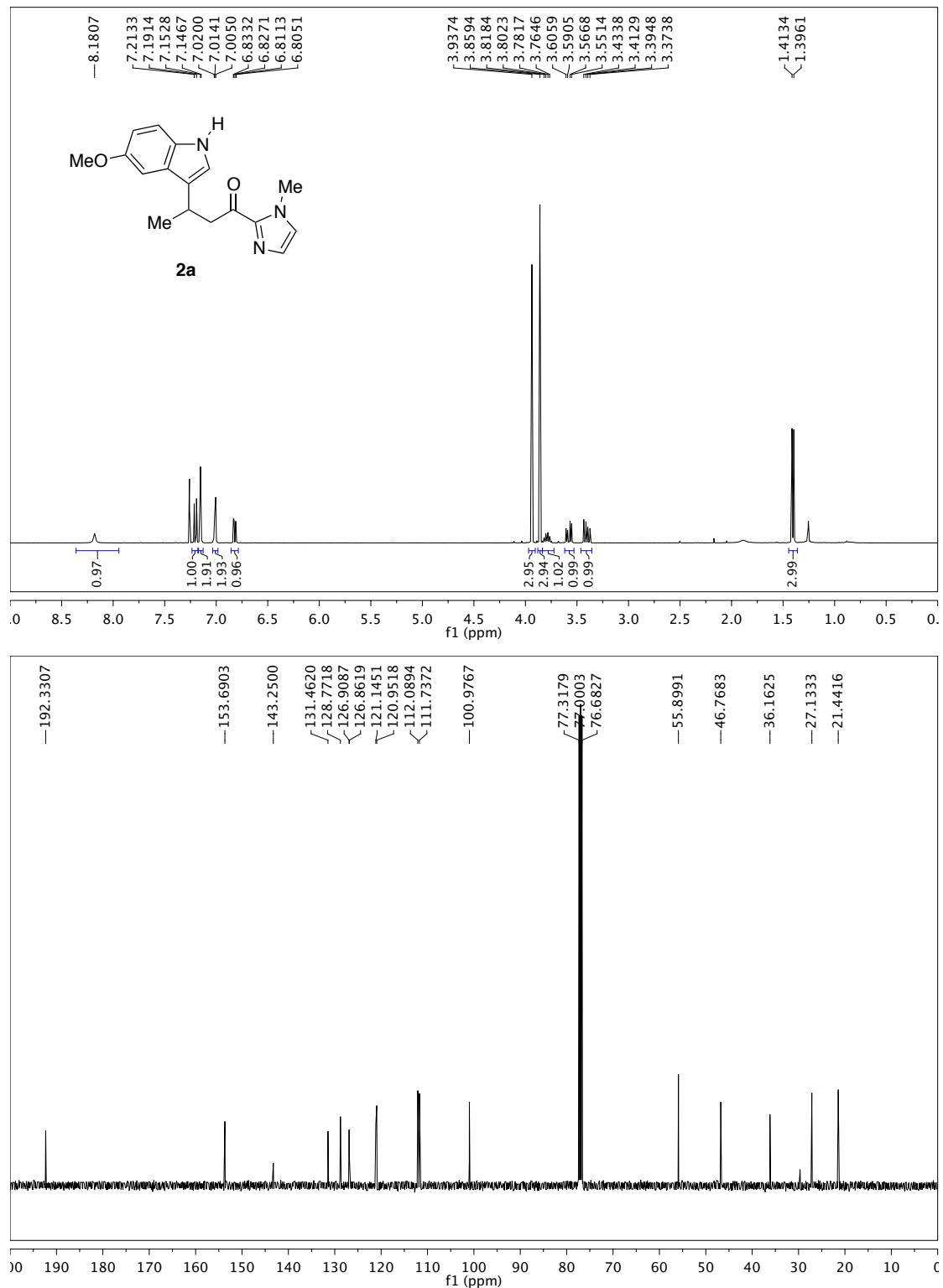
(E)-3-(2-Bromophenyl)-1-(1-methyl-1*H*-imidazol-2-yl)prop-2-en-1-one (1g)



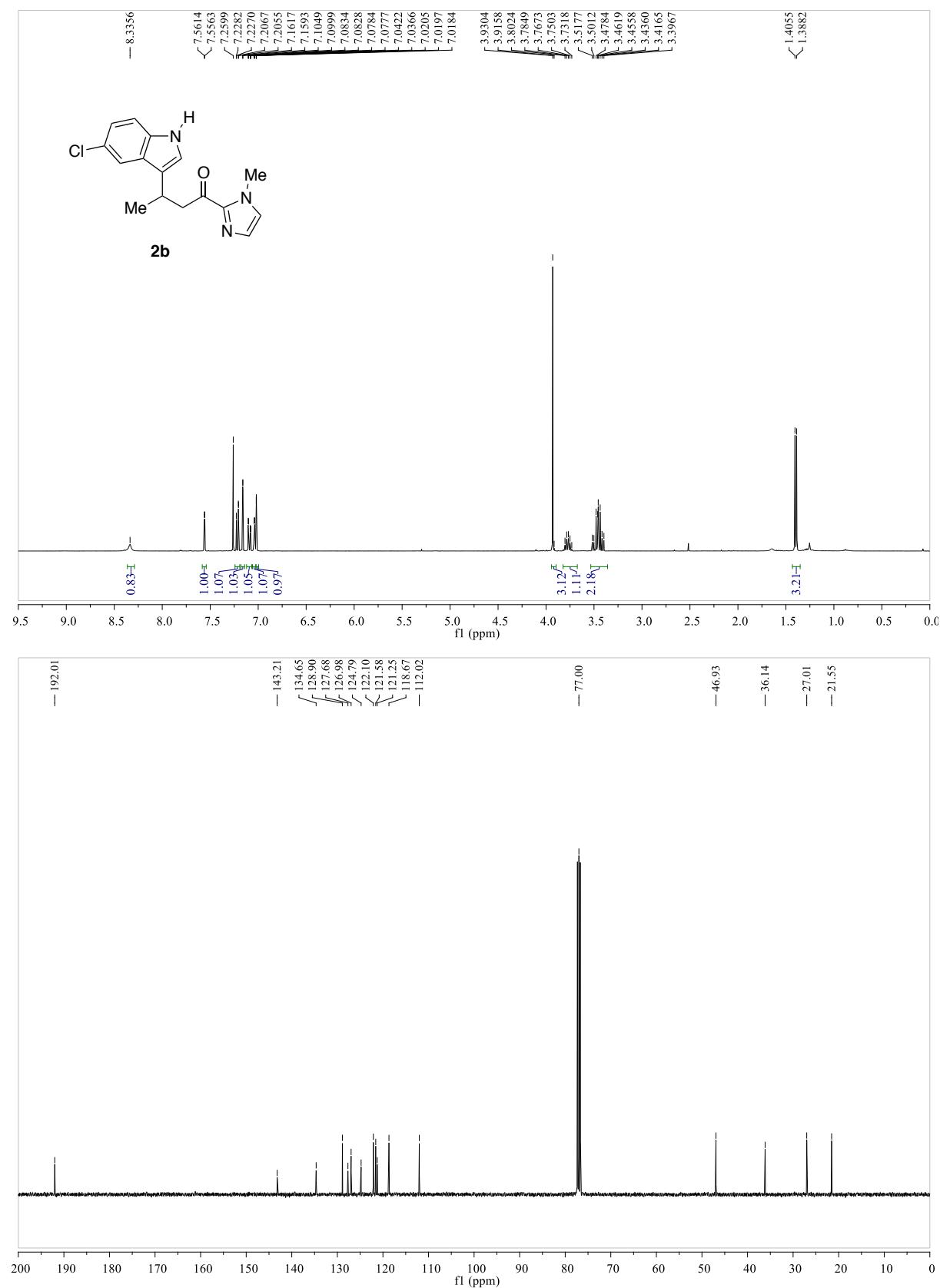
(E)-3-(Furan-2-yl)-1-(1-methyl-1*H*-imidazol-2-yl)prop-2-en-1-one (1h)



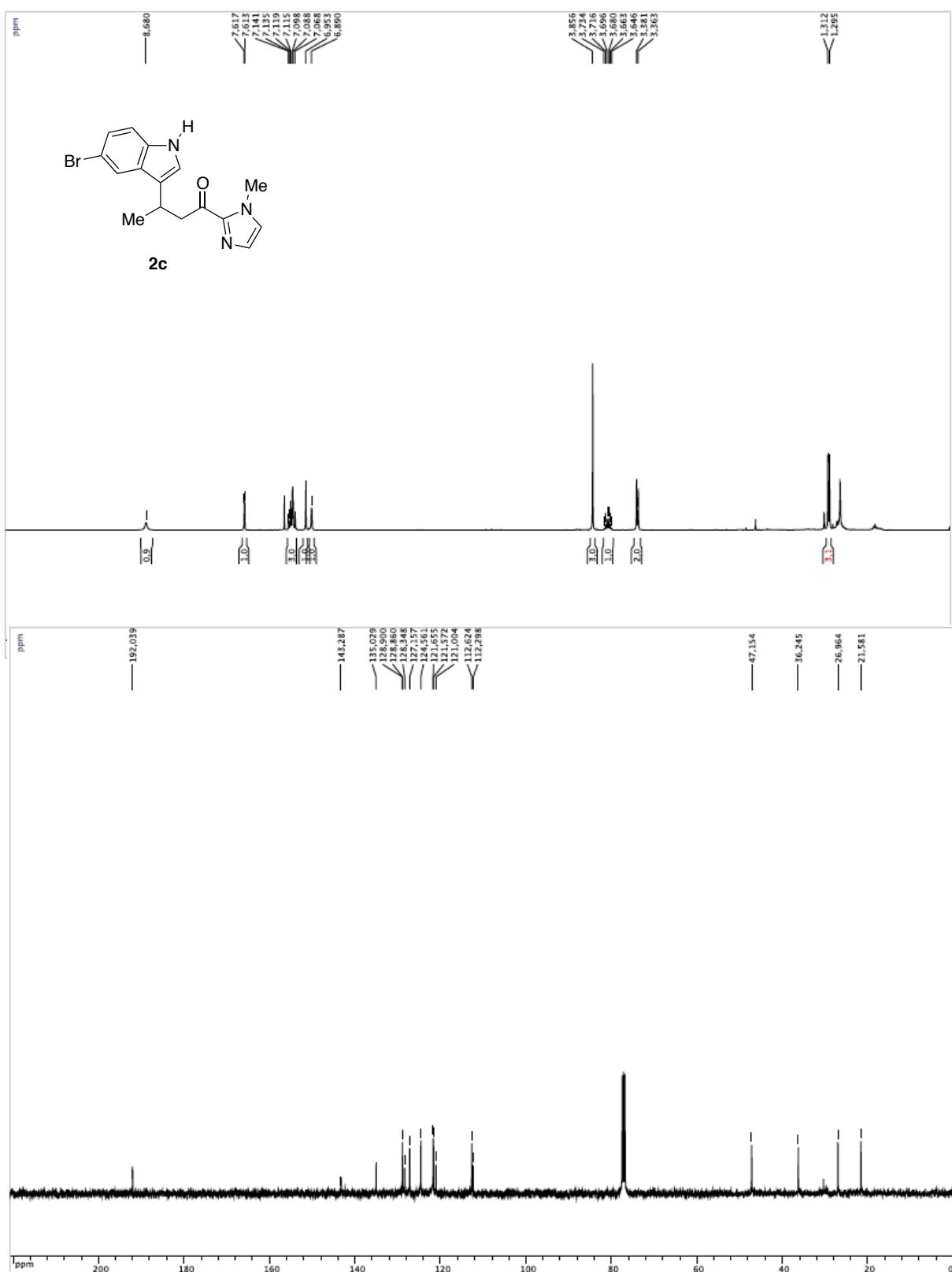
3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2a)



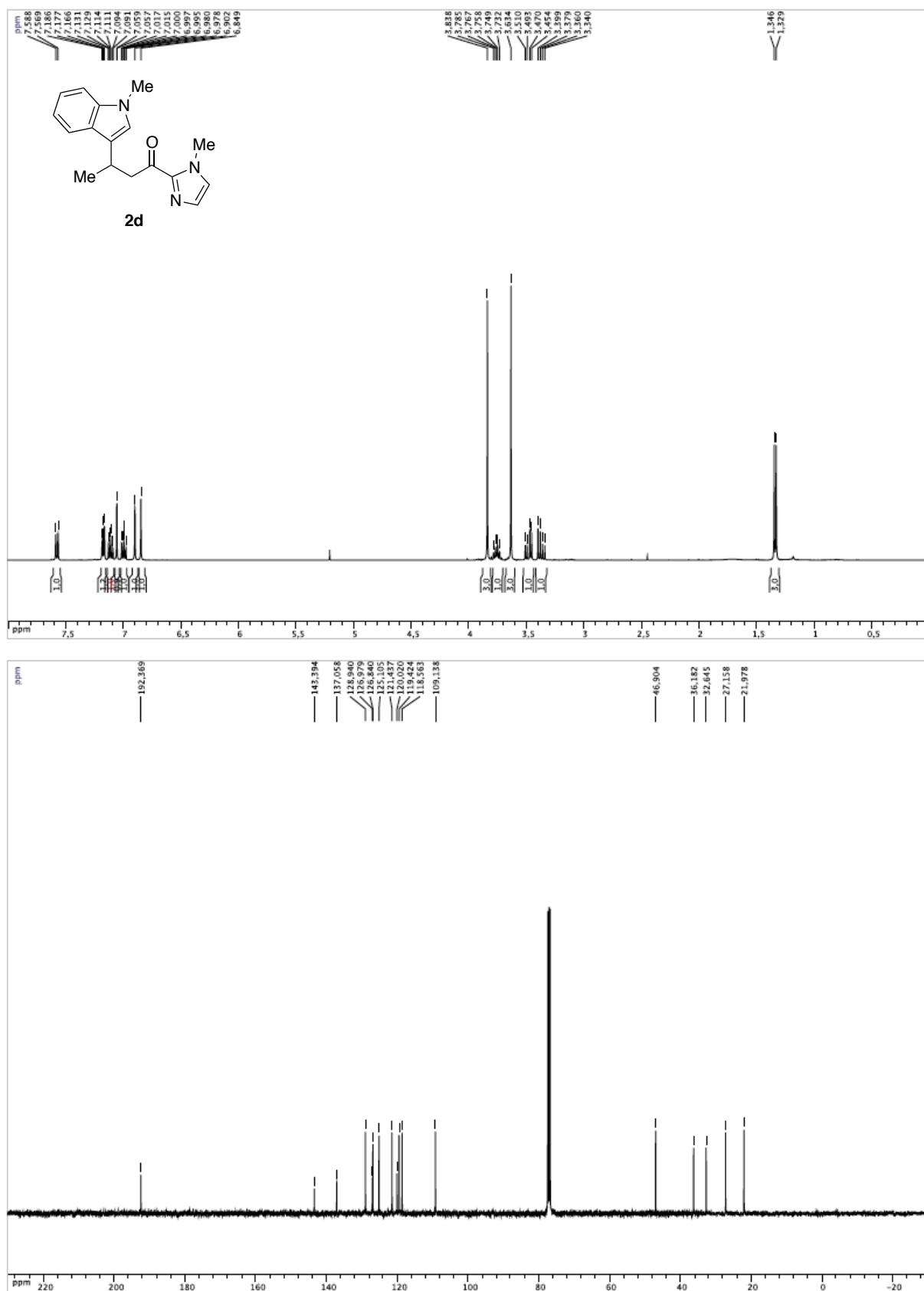
3-(5-Chloro-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2b)



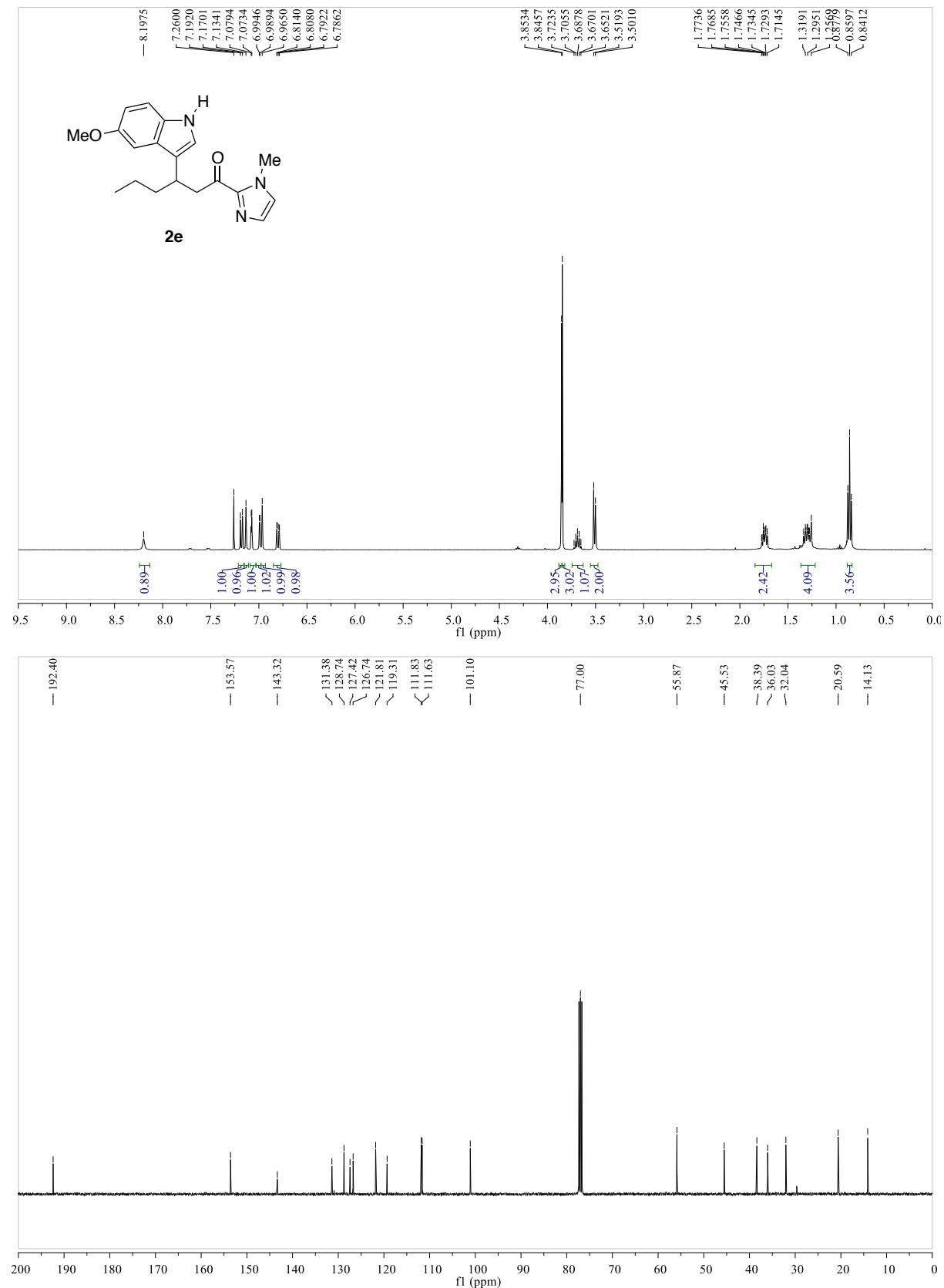
3-(5-Bromo-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2c)



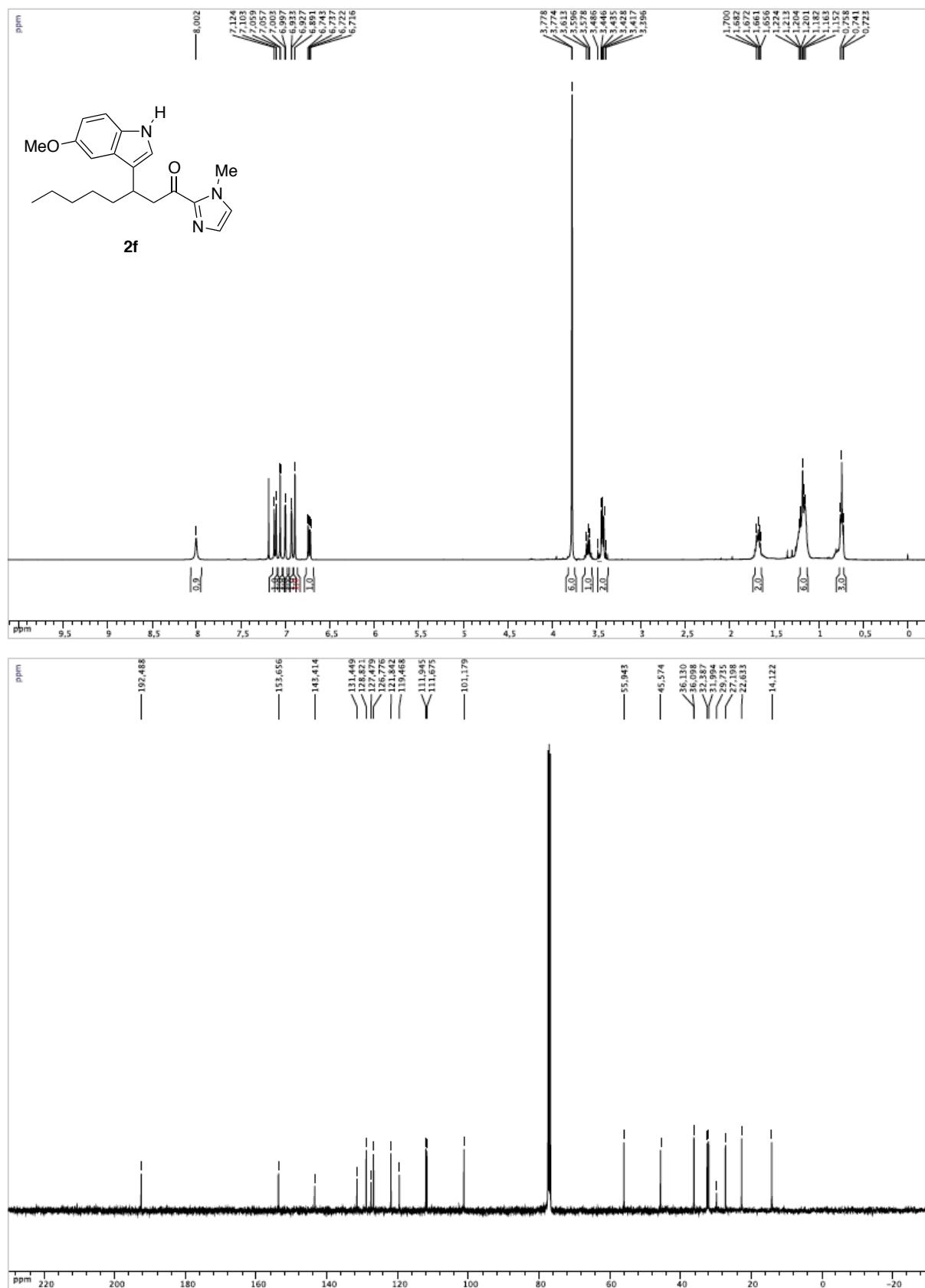
1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)butan-1-one (2d)



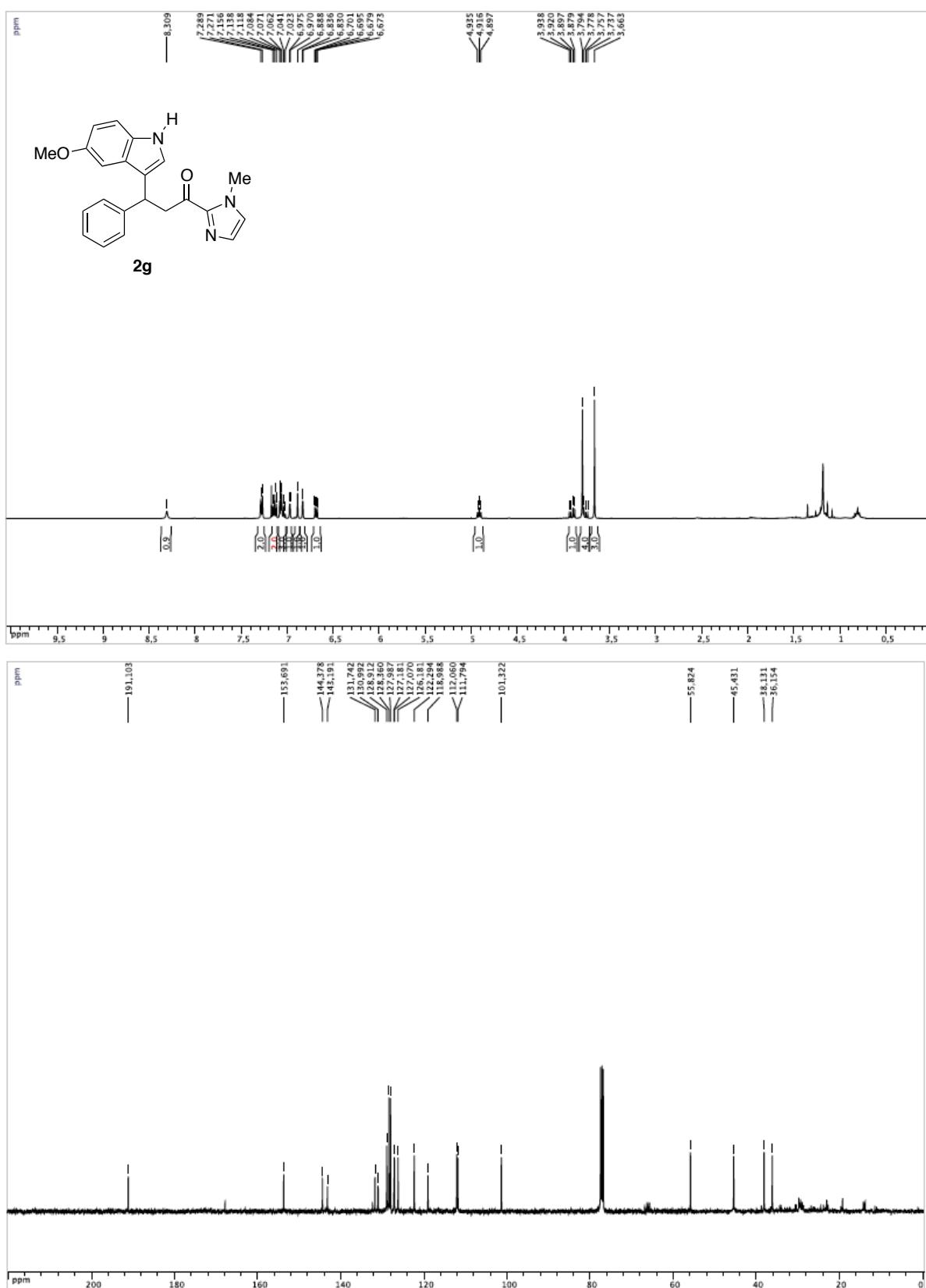
3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)hexan-1-one (2e)



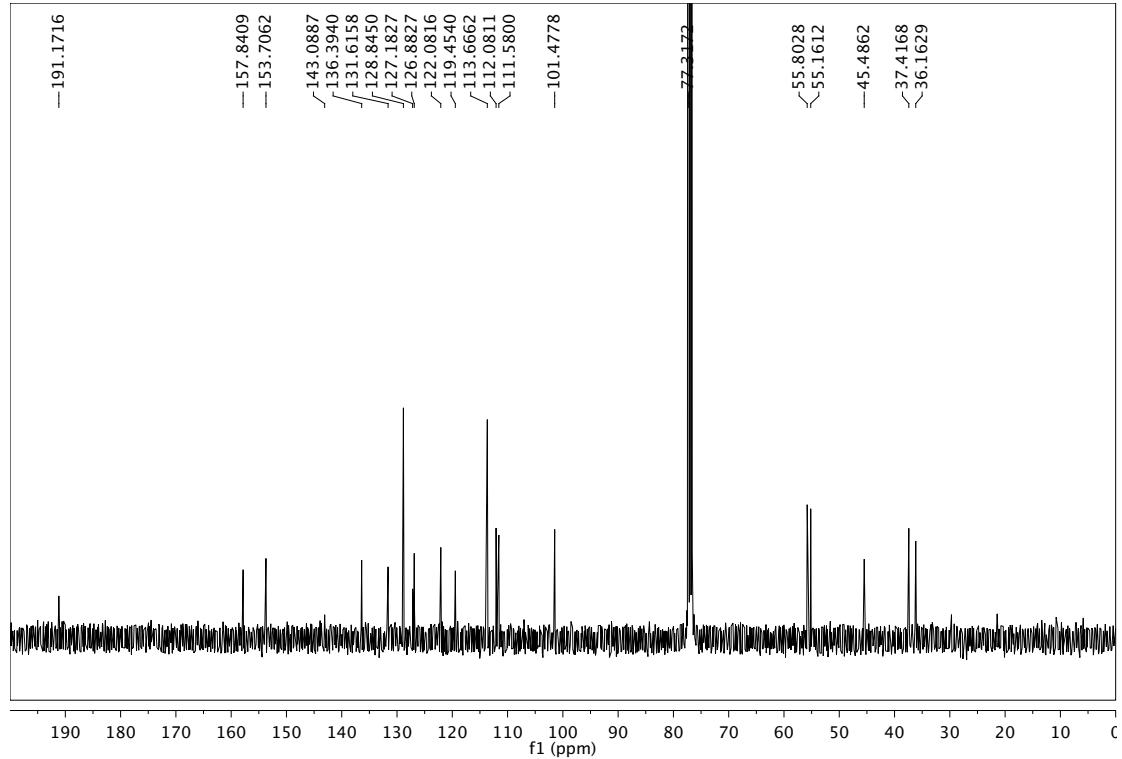
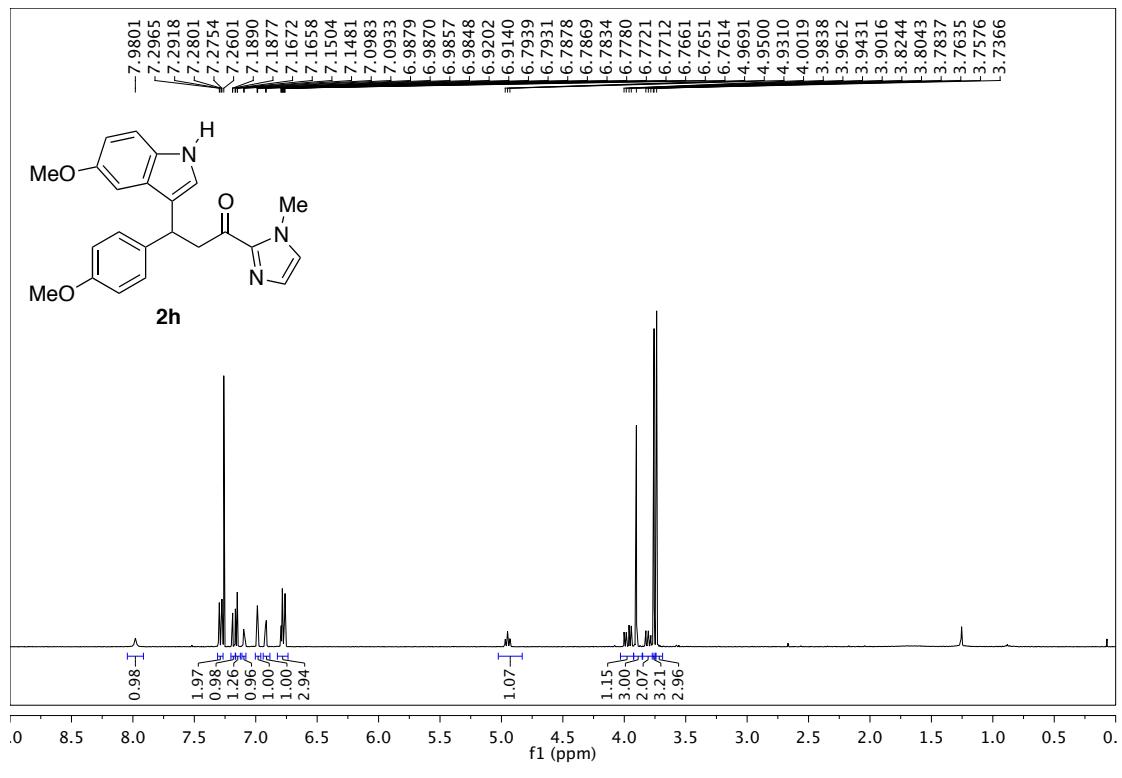
3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)octan-1-one (2f**)**



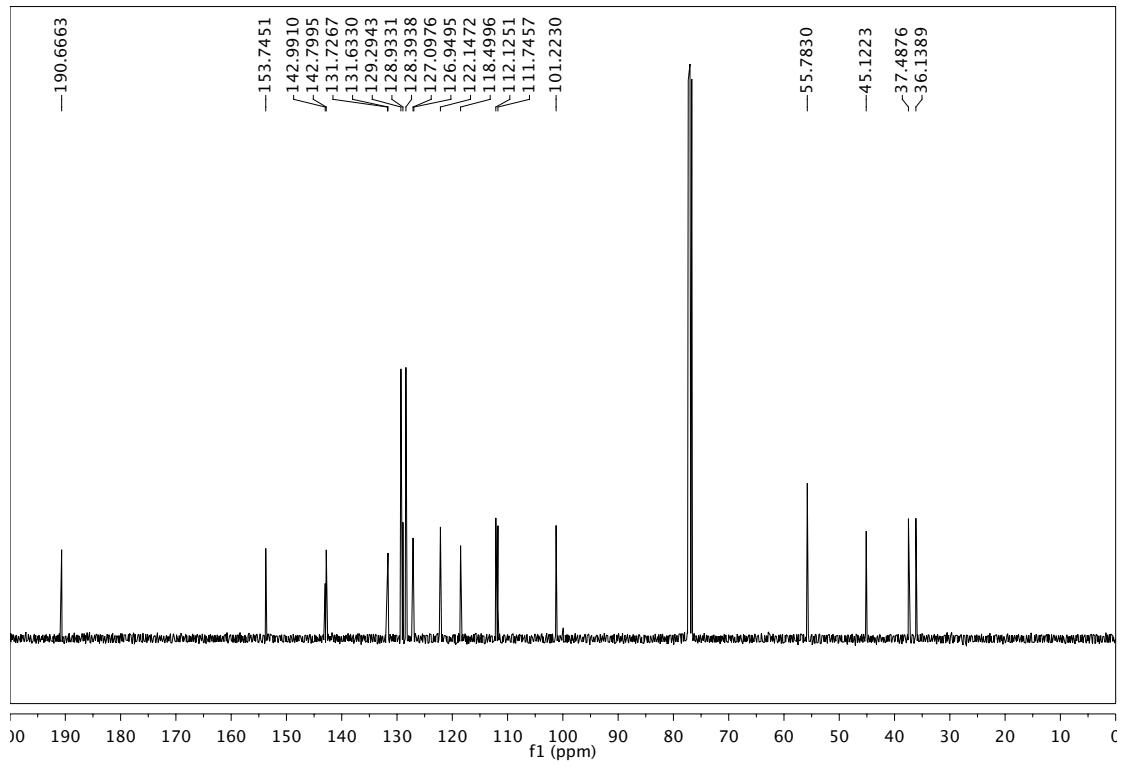
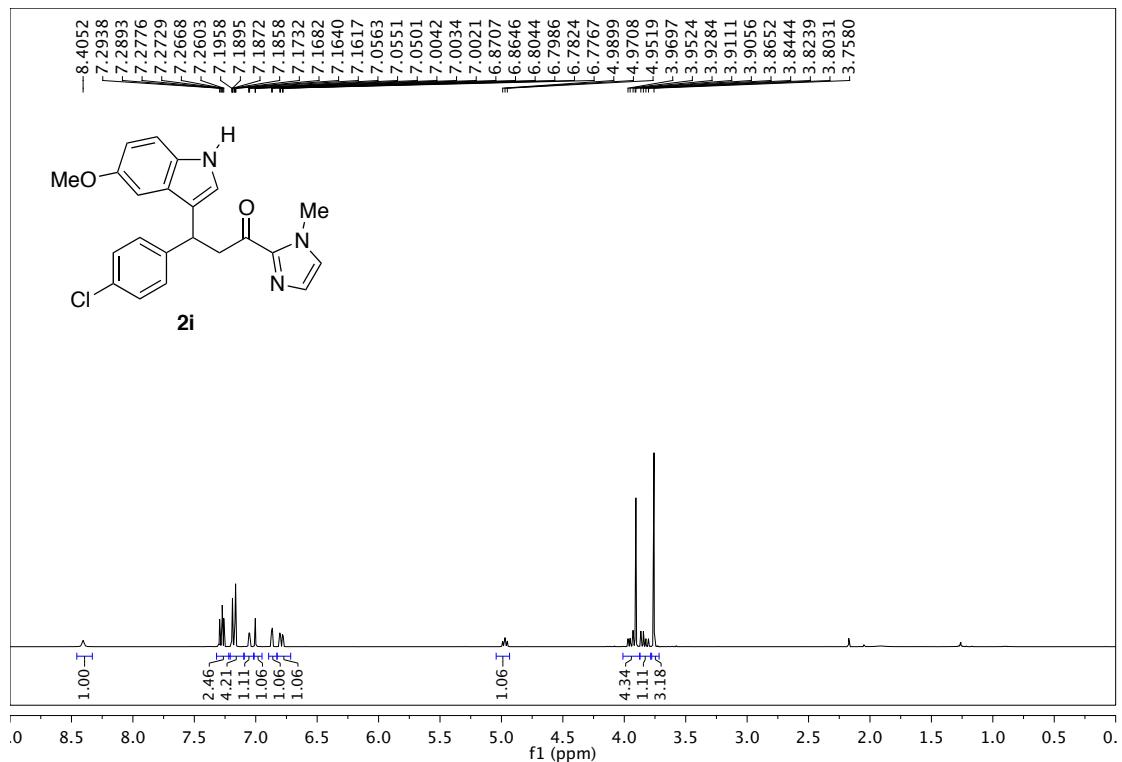
1-(1-Methyl-1*H*-imidazol-2-yl)-3-(5-methoxy-1*H*-indol-3-yl)-3-phenylpropan-1-one (2g)



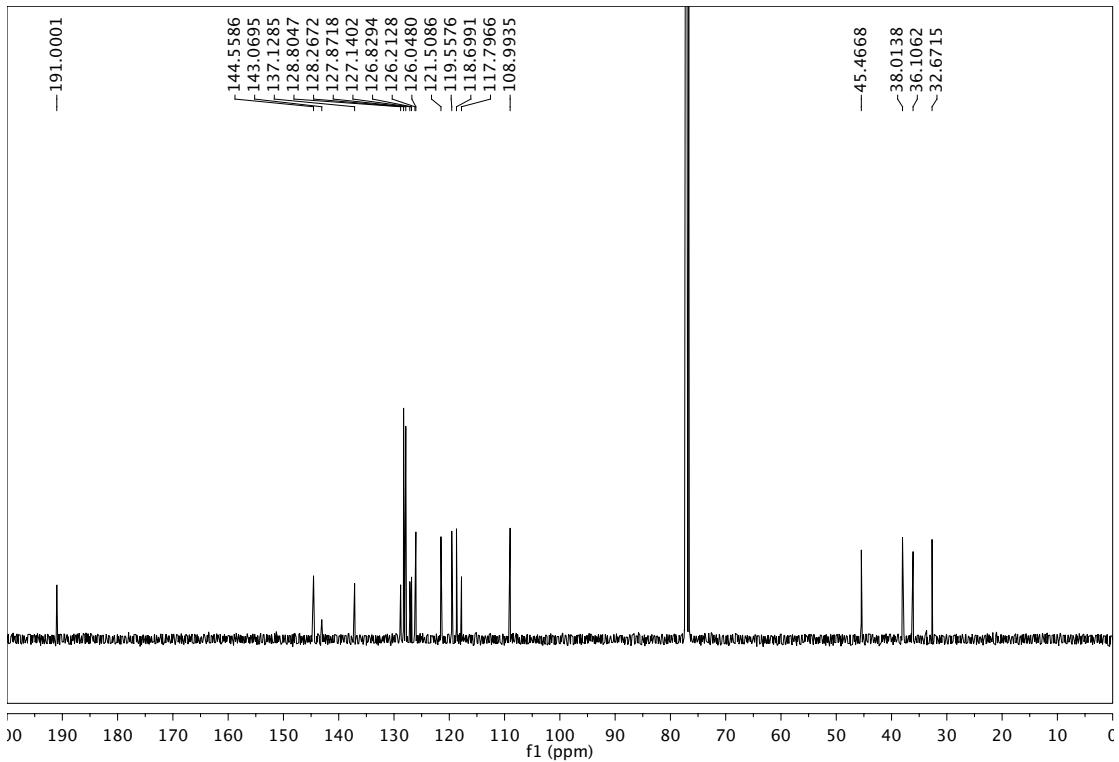
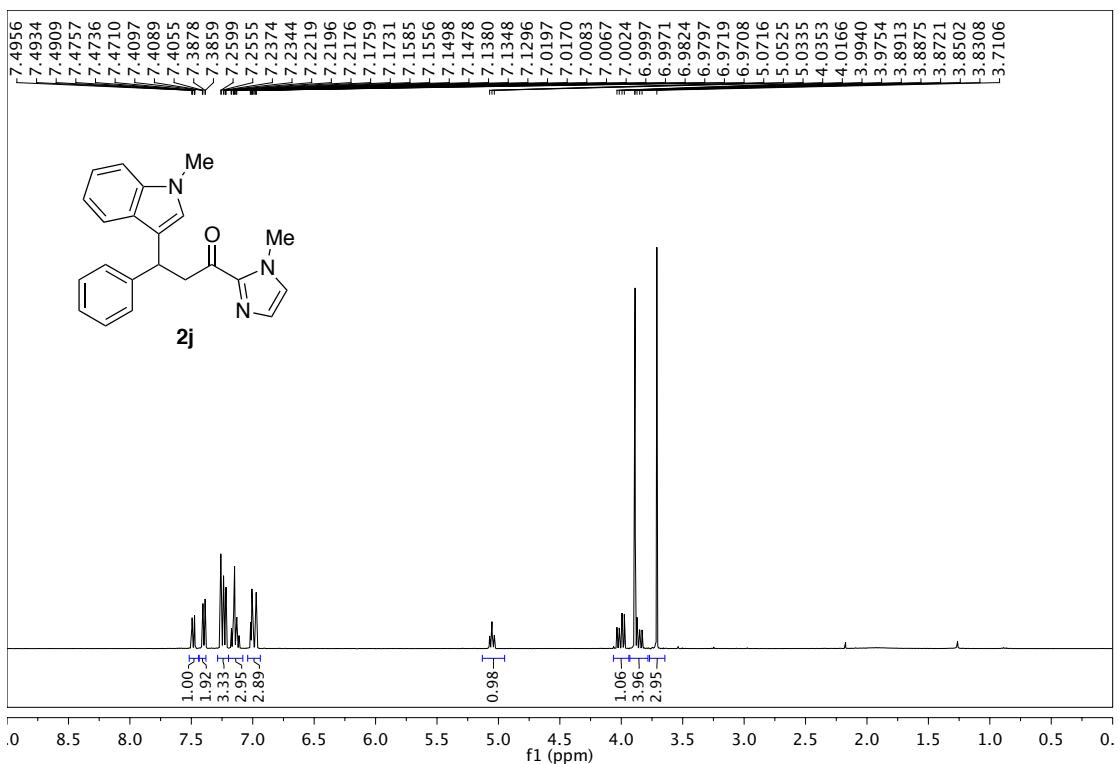
3-(5-Methoxy-1*H*-indol-3-yl)-3-(4-methoxyphenyl)-1-(1-methyl-1*H*-imidazol-2-yl)propan-1-one (2h)



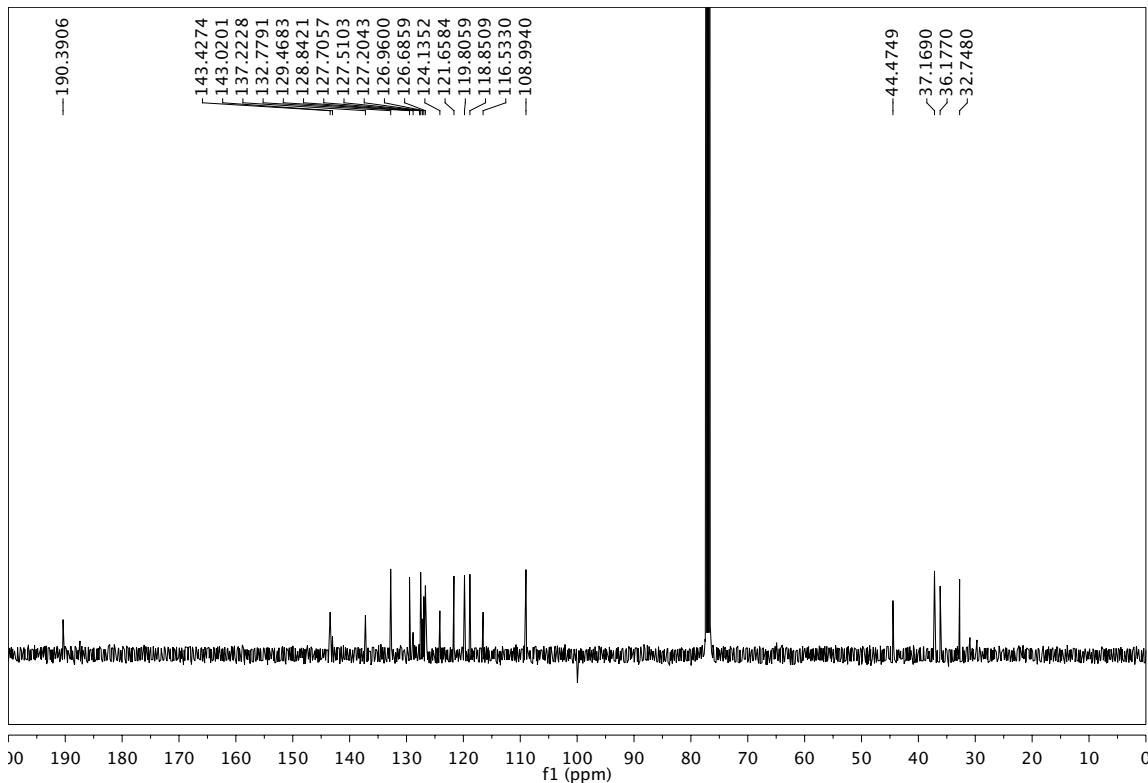
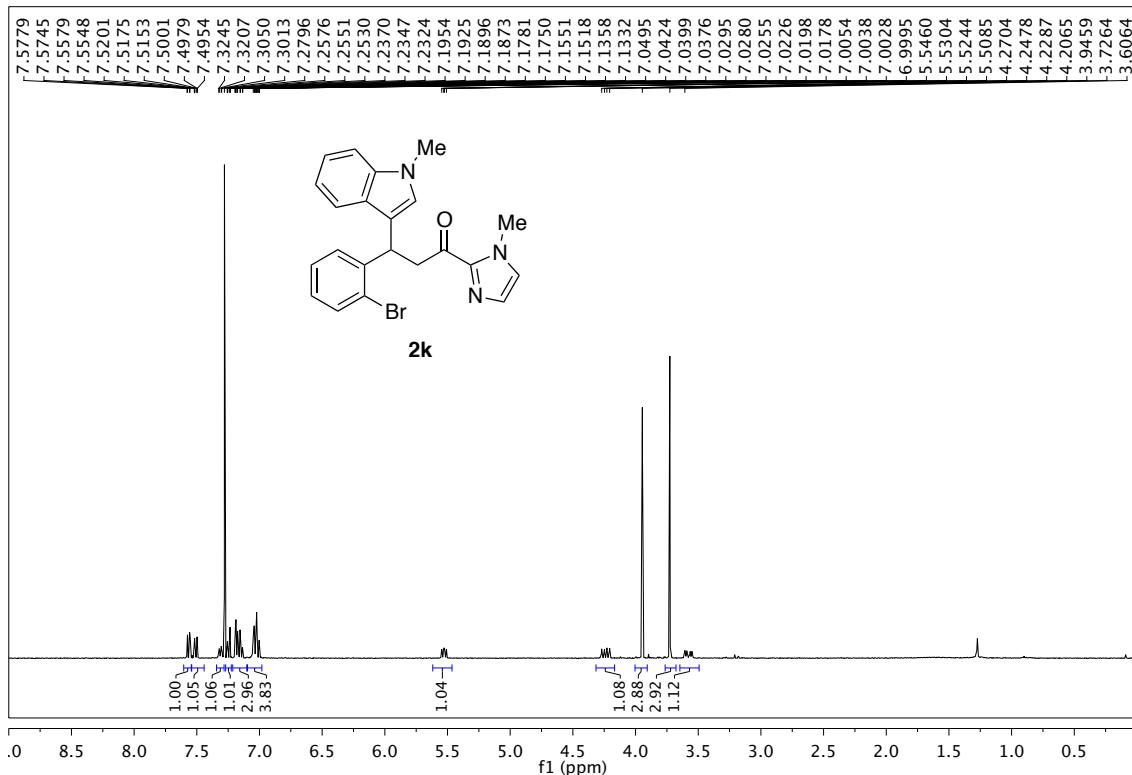
3-(4-Chlorophenyl)-3-(5-methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)propan-1-one (2i)



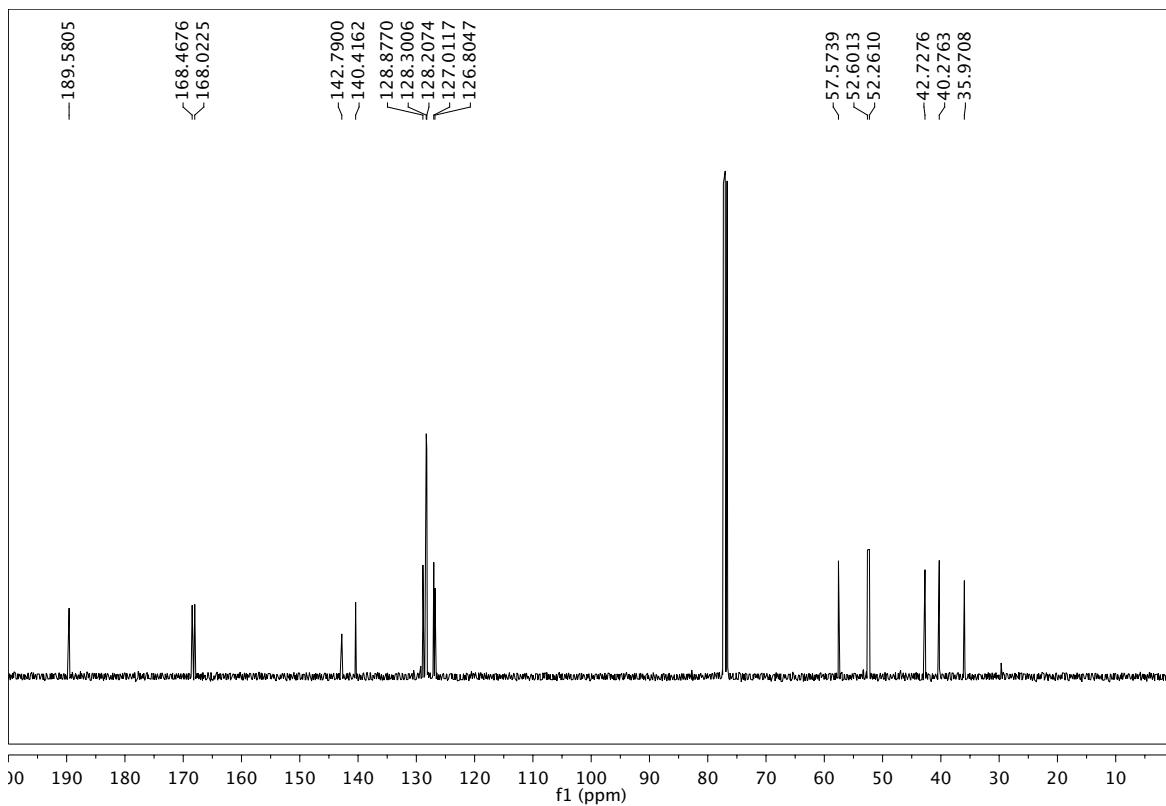
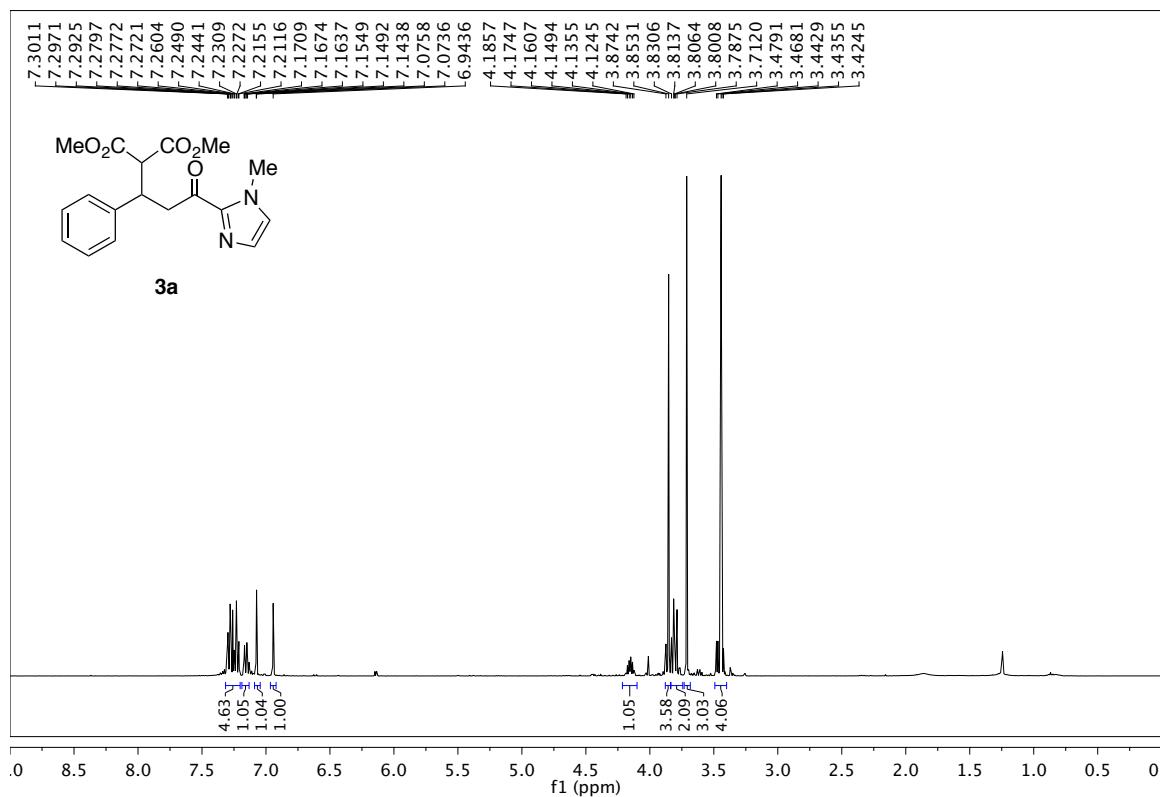
1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)-3-phenylpropan-1-one (2j)



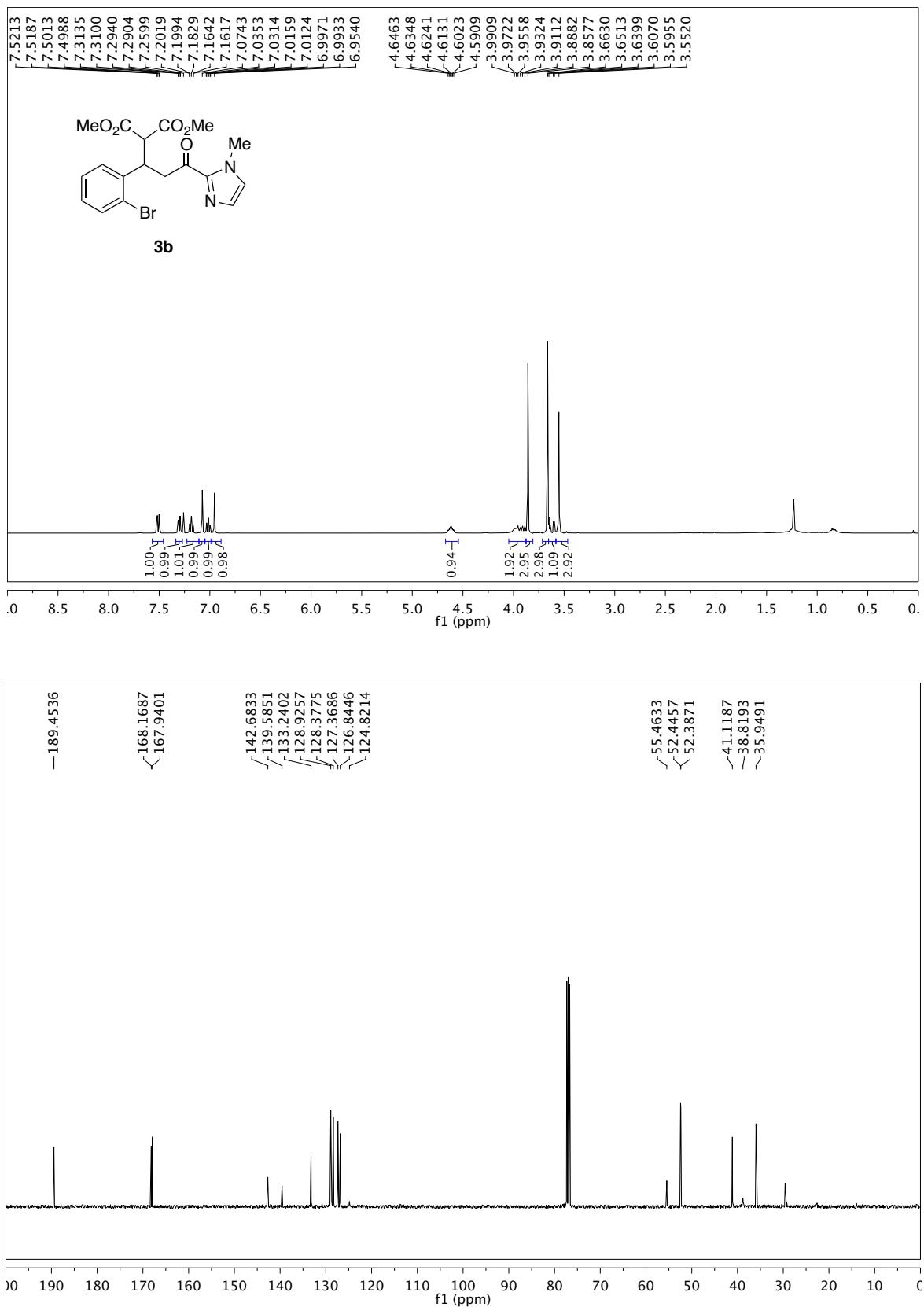
3-(2-Bromophenyl)-1-(1-methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)propan-1-one (2k)



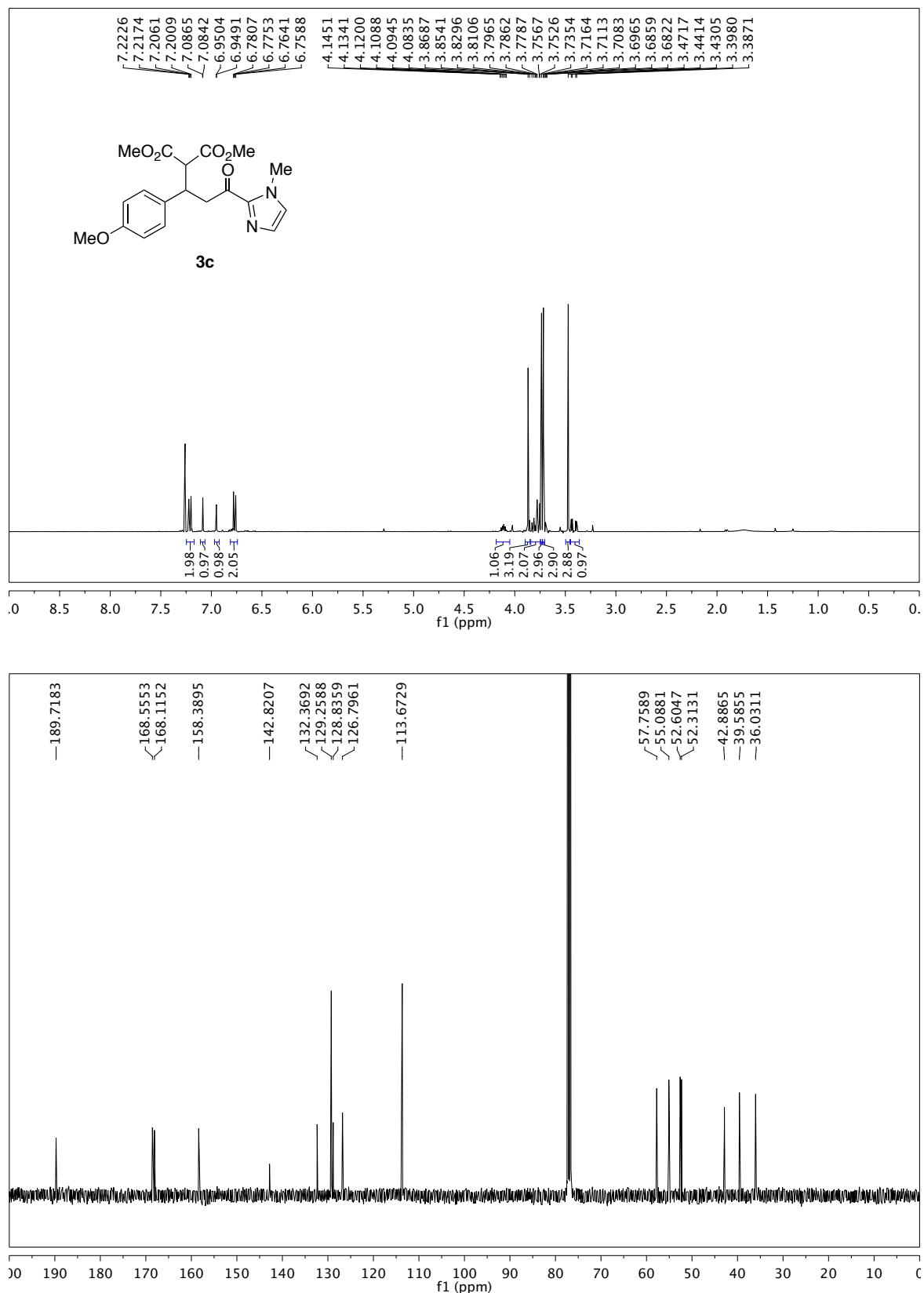
Dimethyl 2-(3-(1-methyl-1*H*-imidazol-2-yl)-3-oxo-1-phenylpropyl)malonate (3a)



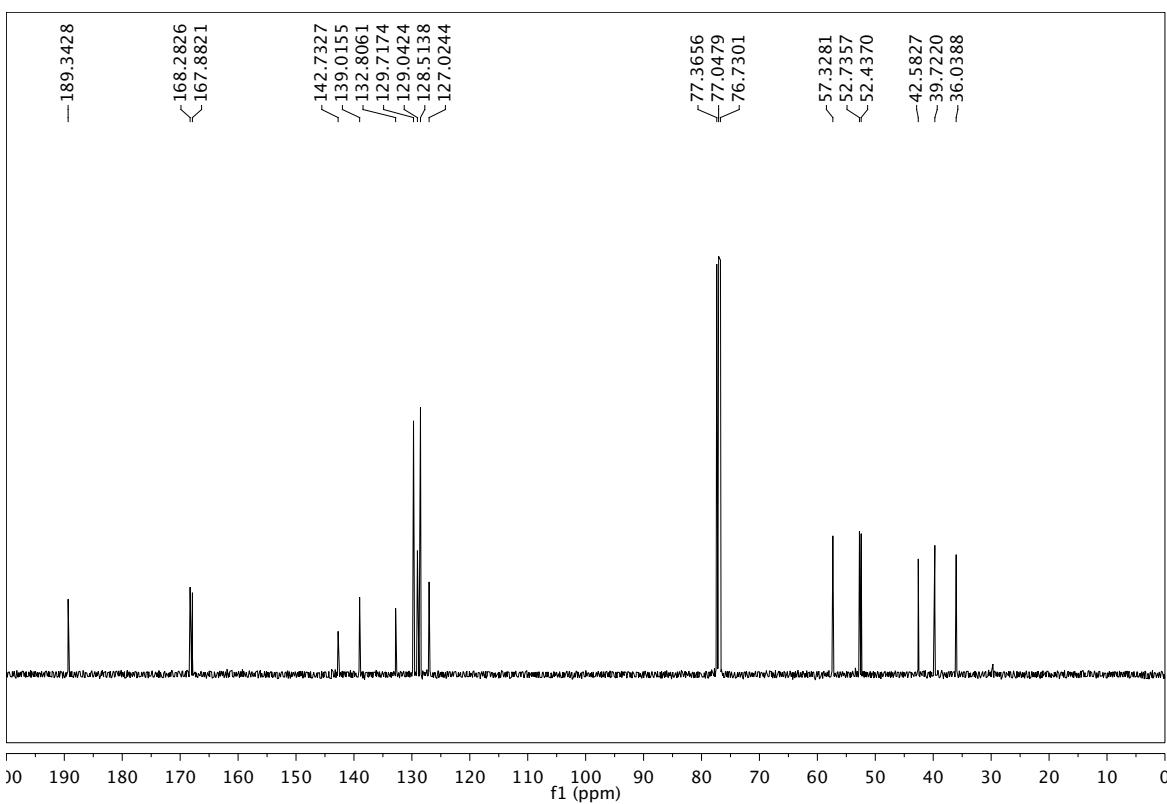
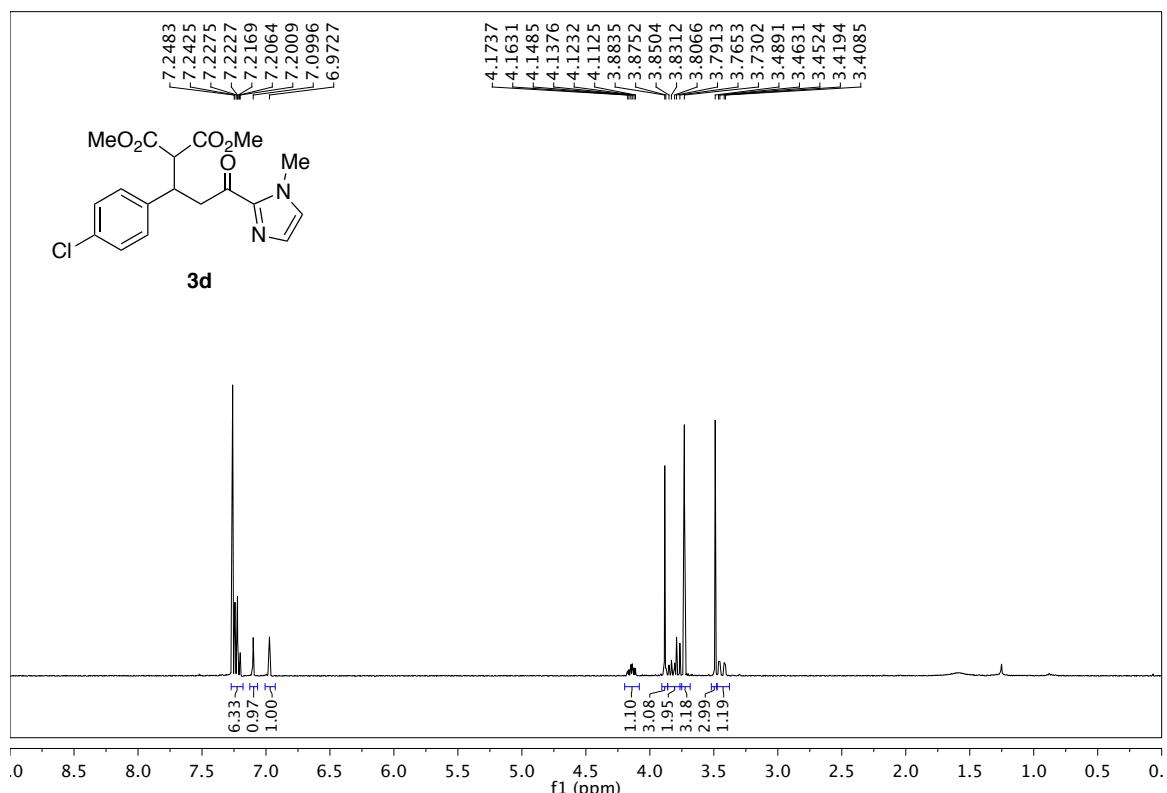
**Dimethyl 2-(1-(2-bromophenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate
(3b)**



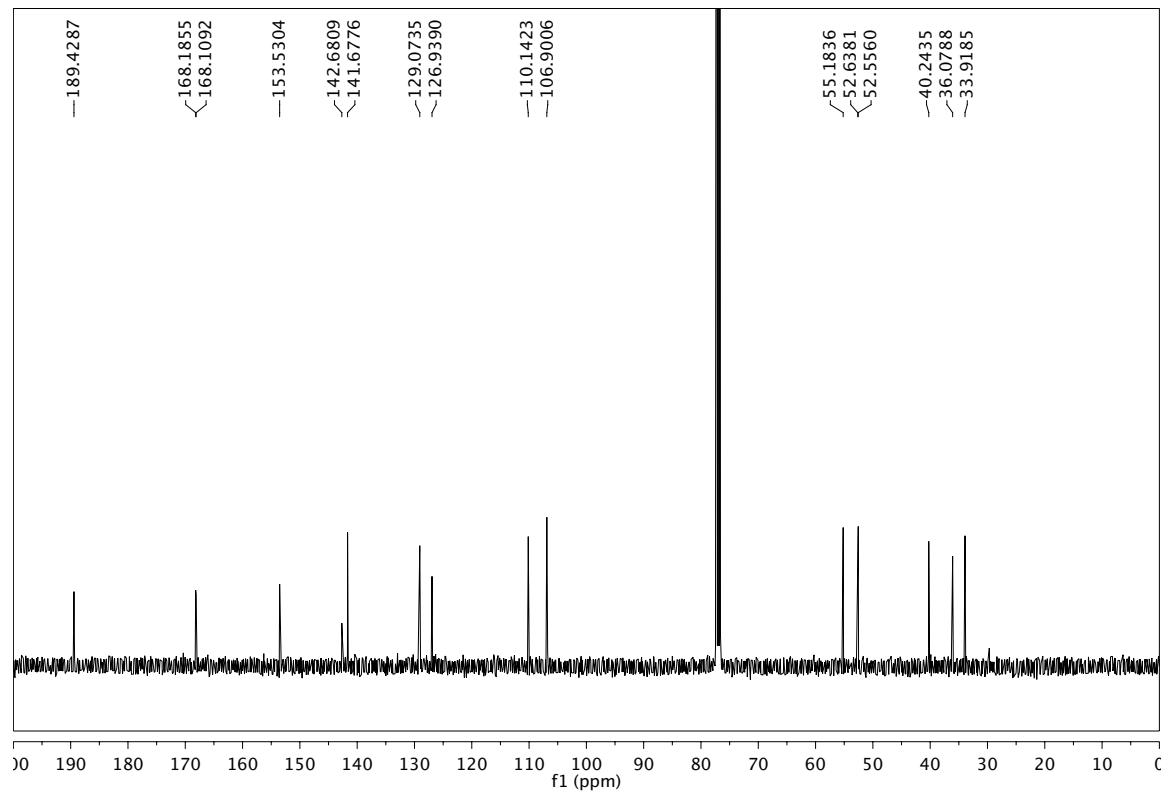
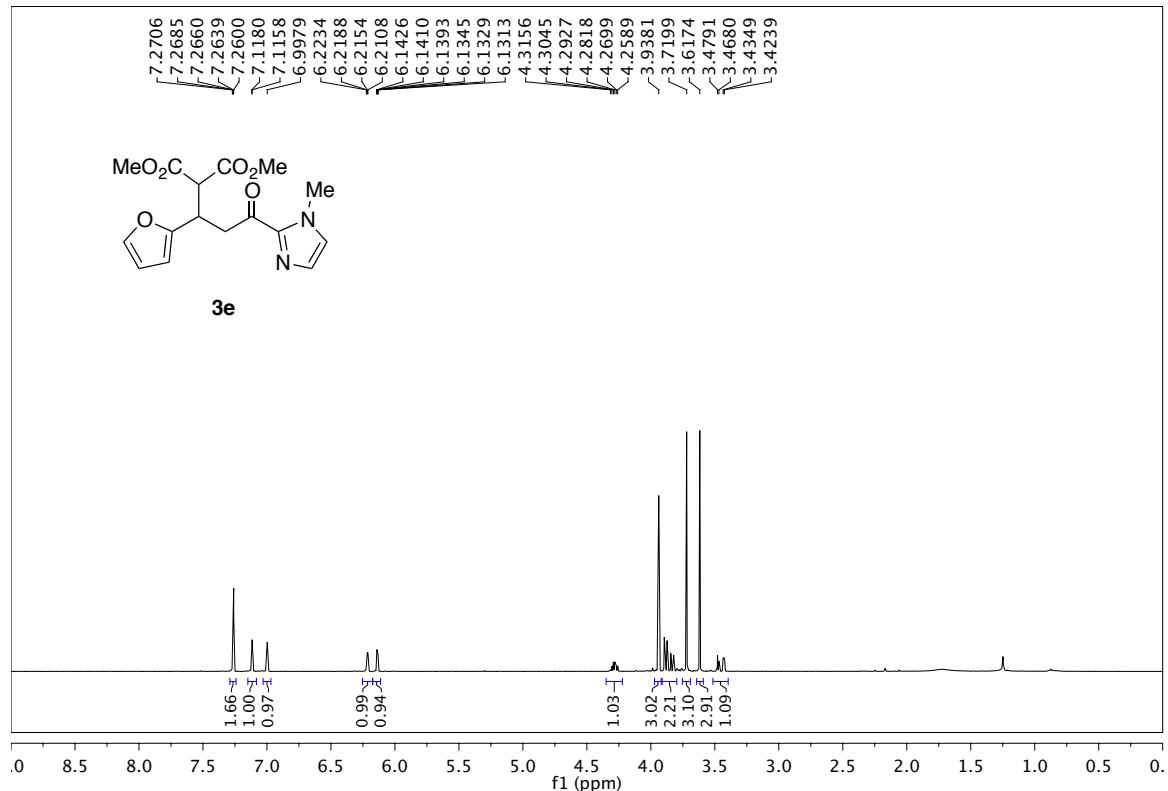
**Dimethyl 2-(1-(4-methoxyphenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate
(3c)**



Dimethyl 2-(1-(4-chlorophenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate (3d)



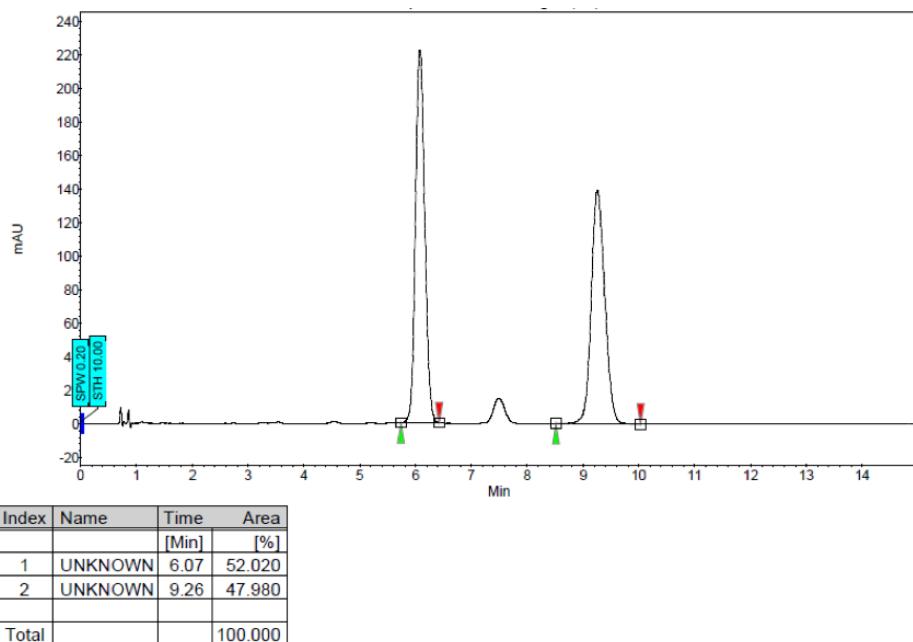
Dimethyl 2-(1-(furan-2-yl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate (3e)



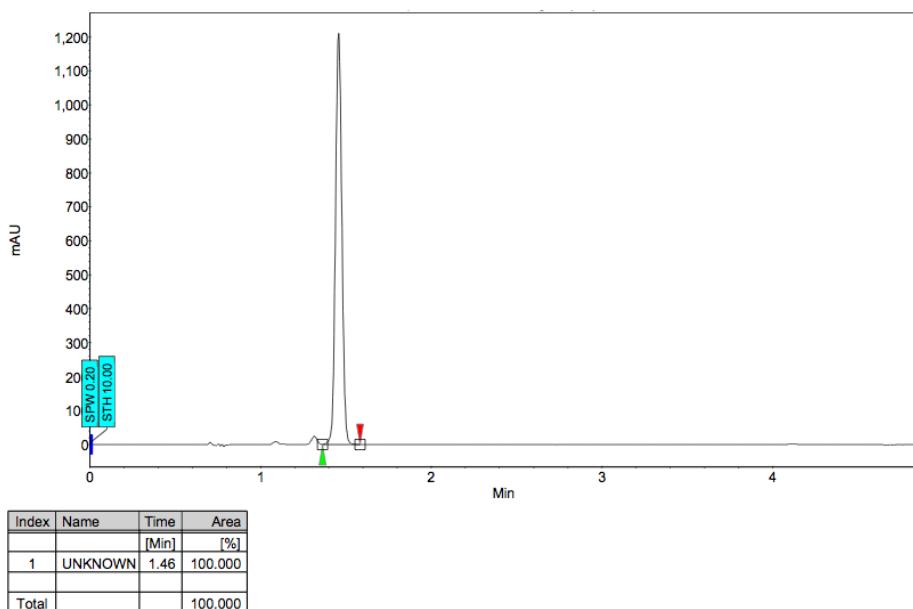
SFC Chromatograms

3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2a)

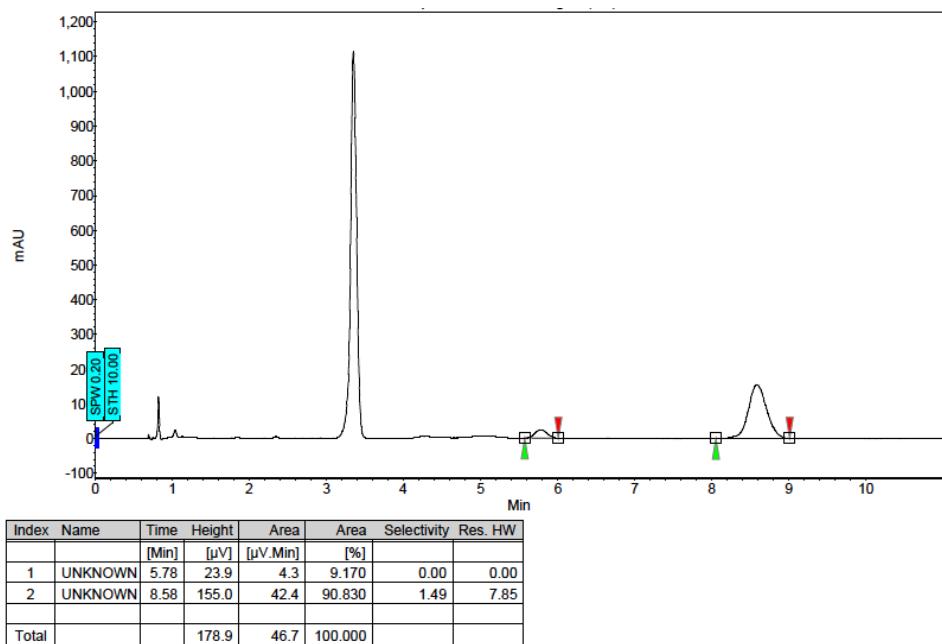
Racemic (2a) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 6.07$; second enantiomer $t_R = 9.26$ min].



Starting material (1a) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; $t_R = 1.46$ min].

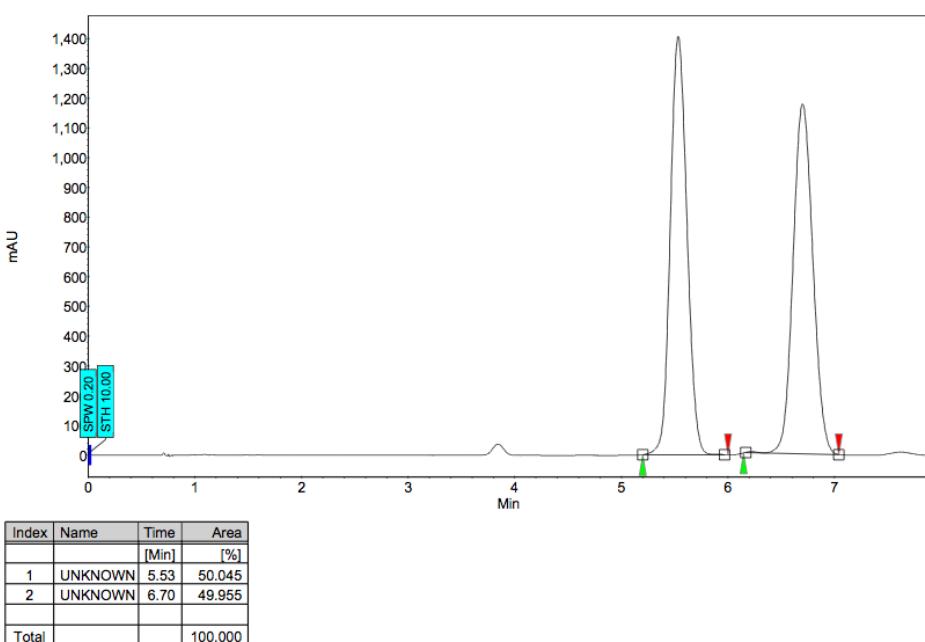


Following general procedure **D** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.56$ min; minor enantiomer $t_R = 5.78$ min].

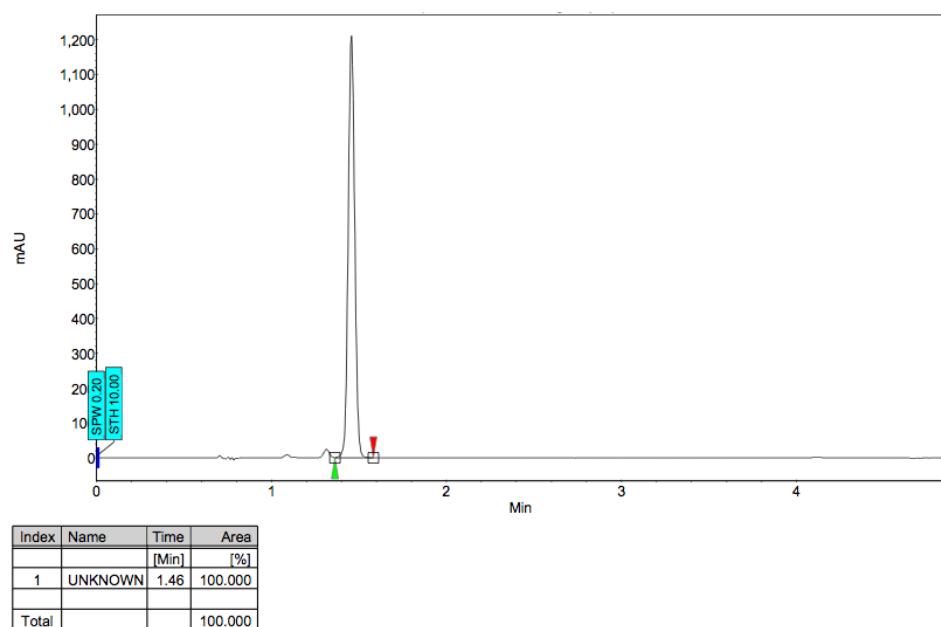


3-(5-Chloro-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2b)

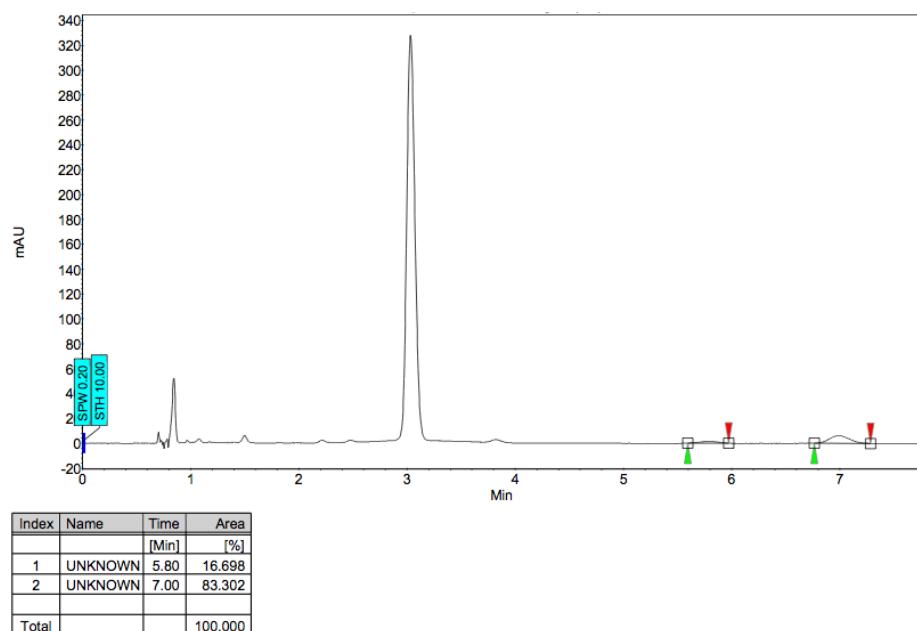
Racemic 2b: [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 5.53$ min; second enantiomer $t_R = 6.70$ min].



Starting material 1a: [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; $t_R = 1.46$ min].

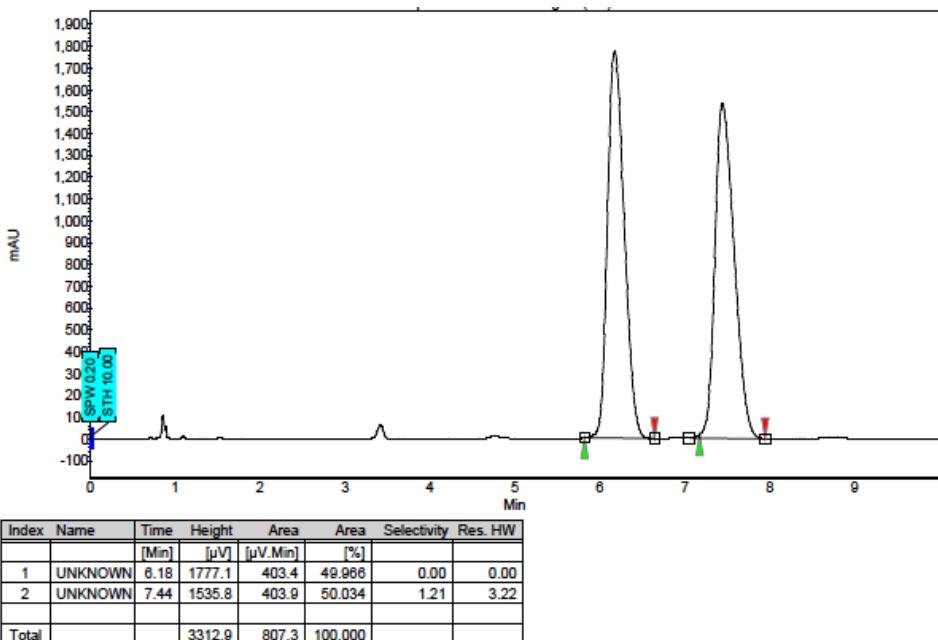


Following general procedure **D** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 7.00$ min; minor enantiomer $t_R = 5.80$ min].

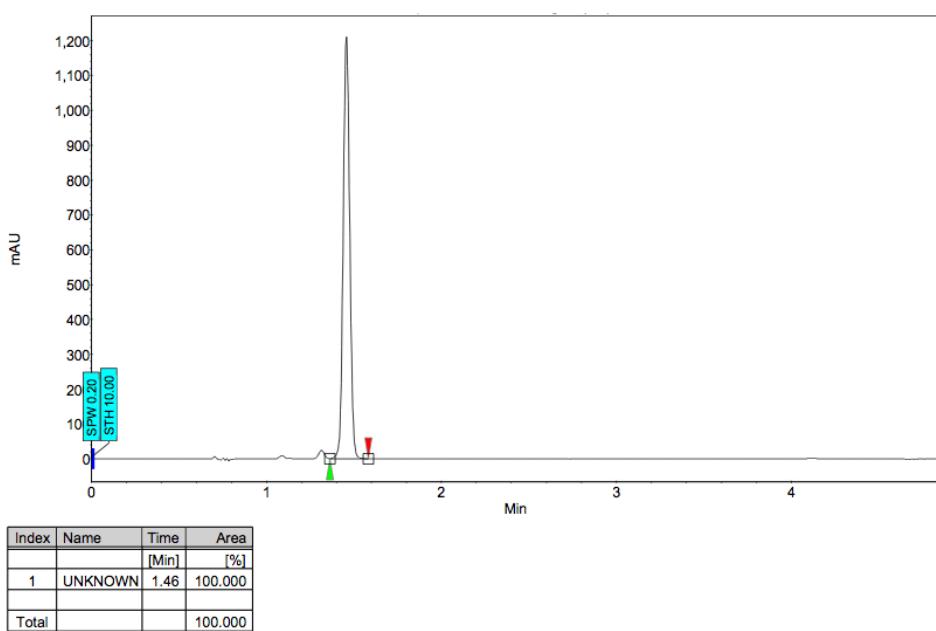


3-(5-Bromo-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)butan-1-one (2c)

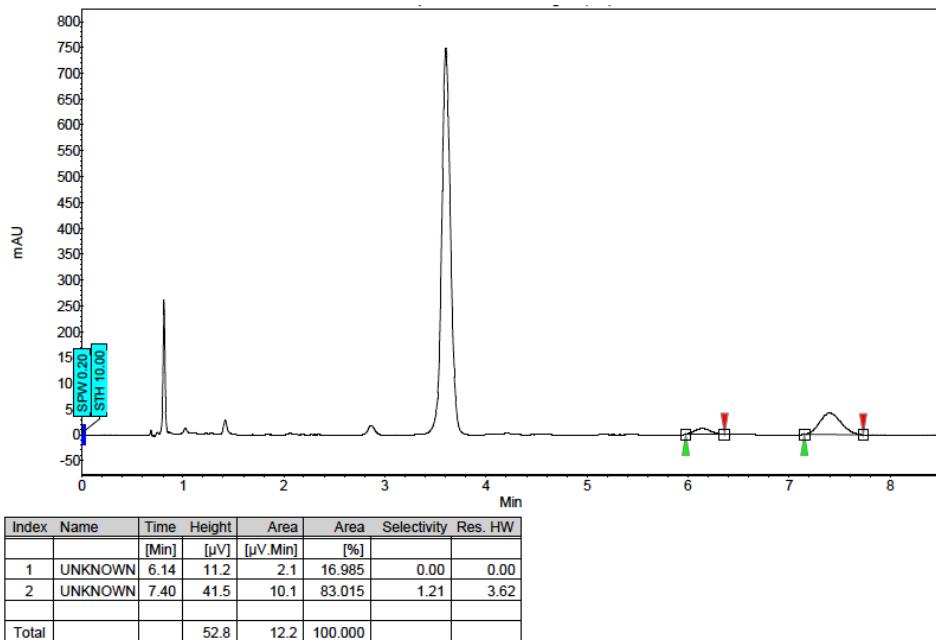
Racemic (2c) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 6.18$ min; second enantiomer $t_R = 7.44$ min].



Starting material (1a) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; $t_R = 1.46$ min].

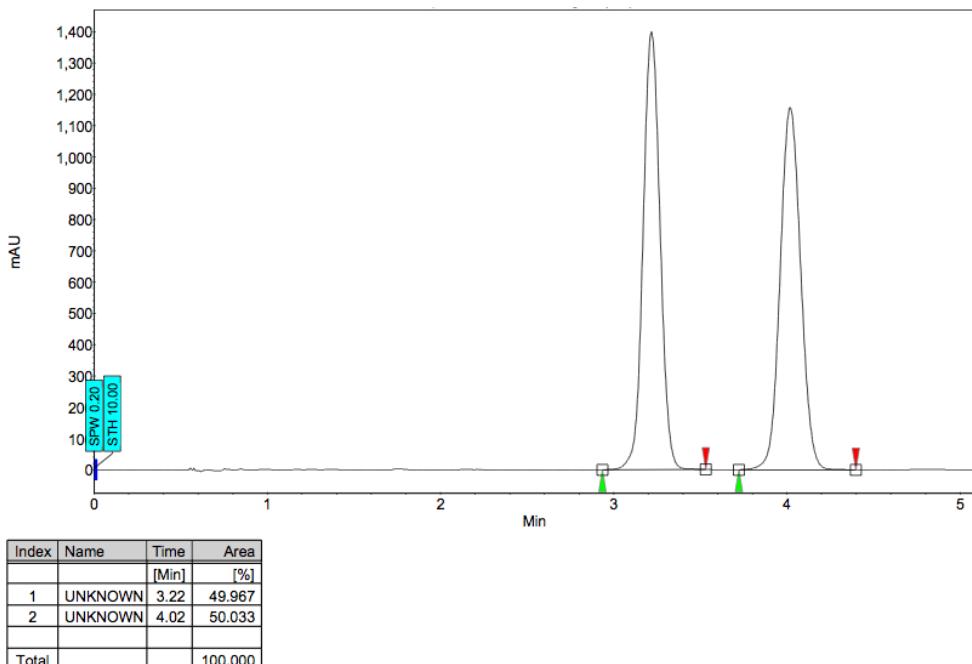


Following general procedure **D** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 7.40$ min; minor enantiomer $t_R = 6.14$ min].

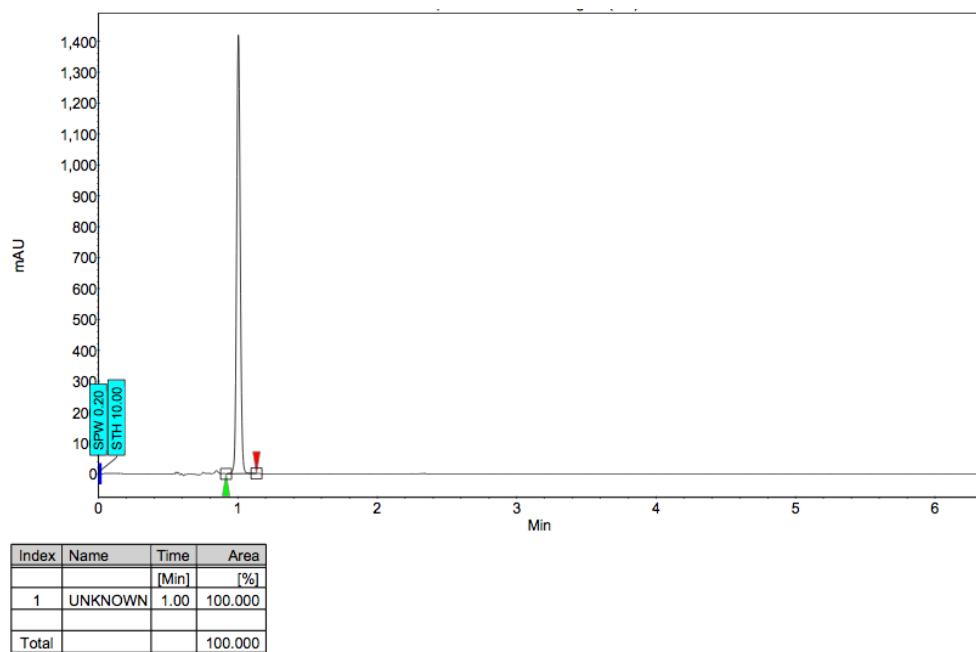


1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)butan-1-one (2d)

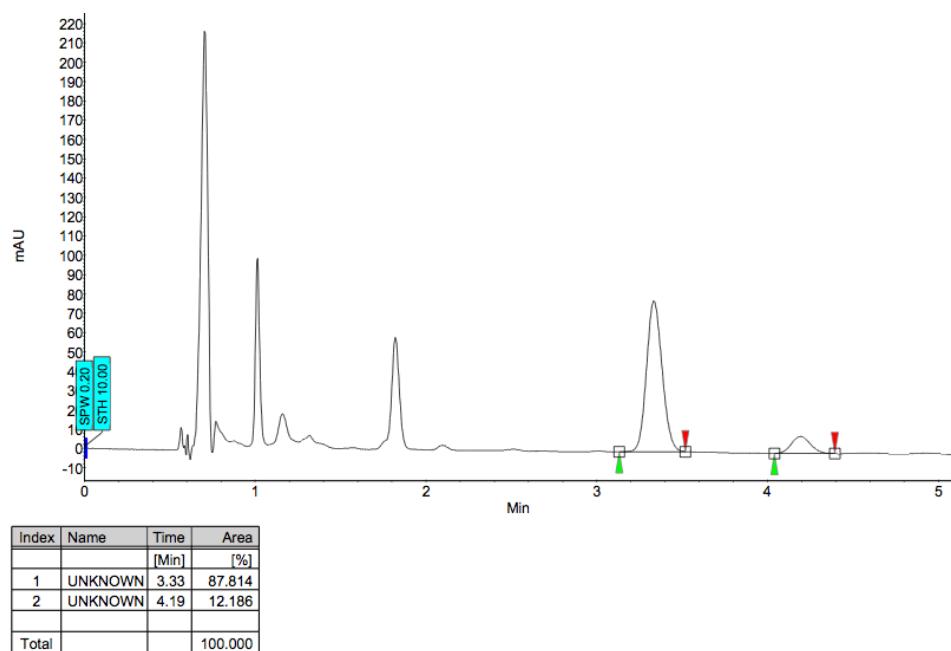
Racemic (2d) [DAICEL AD-H column; 100 bar; flow: 5.0 mL/min; 15% *i*-PrOH; $\lambda = 220$ nm; first enantiomer $t_R = 3.22$ min; second enantiomer $t_R = 4.02$ min].



Starting material (1a) [DAICEL AD-H column; 100 bar; flow: 15.0 mL/min; 15% *i*-PrOH; $\lambda = 220$ nm; $t_R = 1.00$ min].

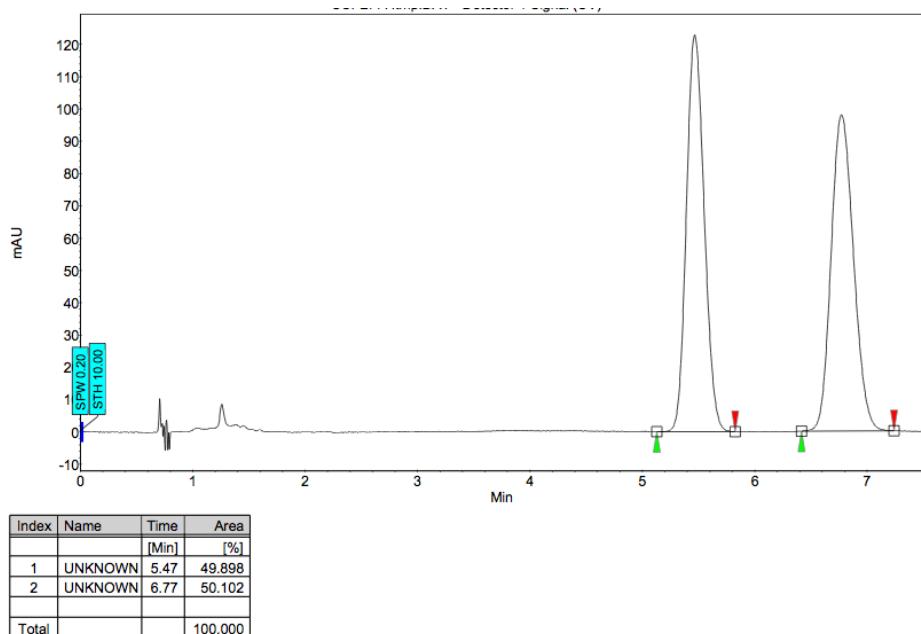


Following the general procedure **D** [DAICEL AD-H column; 100 bar; flow: 15.0 mL/min; 15% *i*-PrOH; $\lambda = 220$ nm, major enantiomer $t_R = 3.33$ min; minor enantiomer $t_R = 4.19$ min]

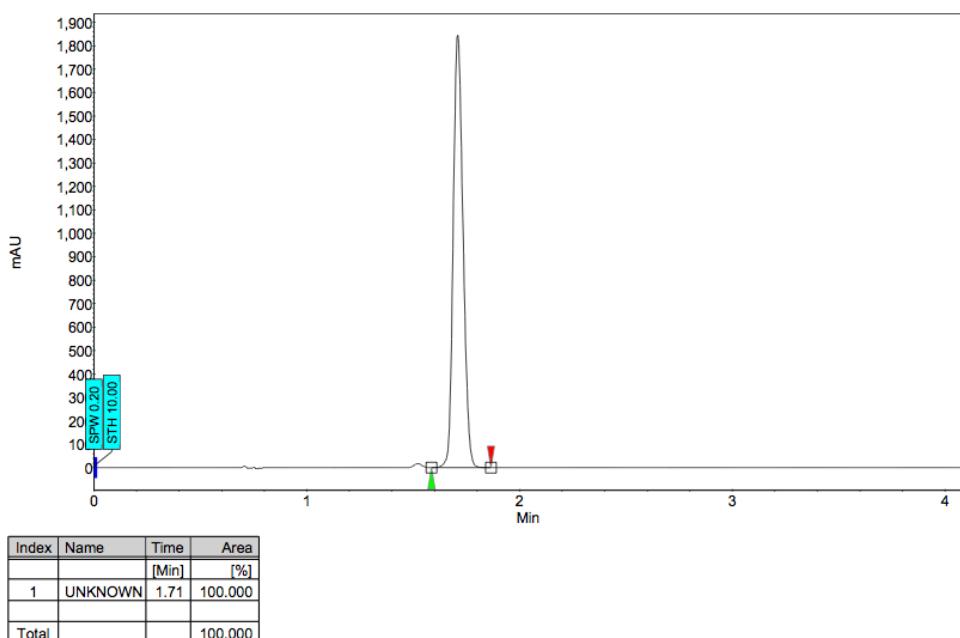


3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)hexan-1-one (2e)

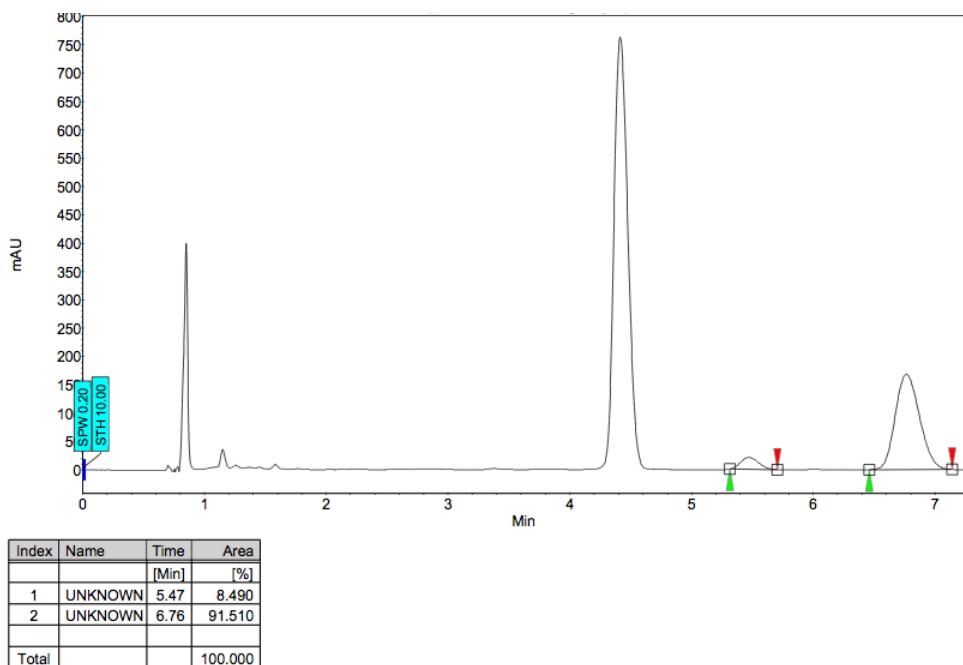
Racemic 2e: [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 12% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 5.47$ min; second enantiomer $t_R = 6.77$ min].



Starting material 1b: [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 12% MeOH; $\lambda = 220$ nm; $t_R = 1.71$ min].

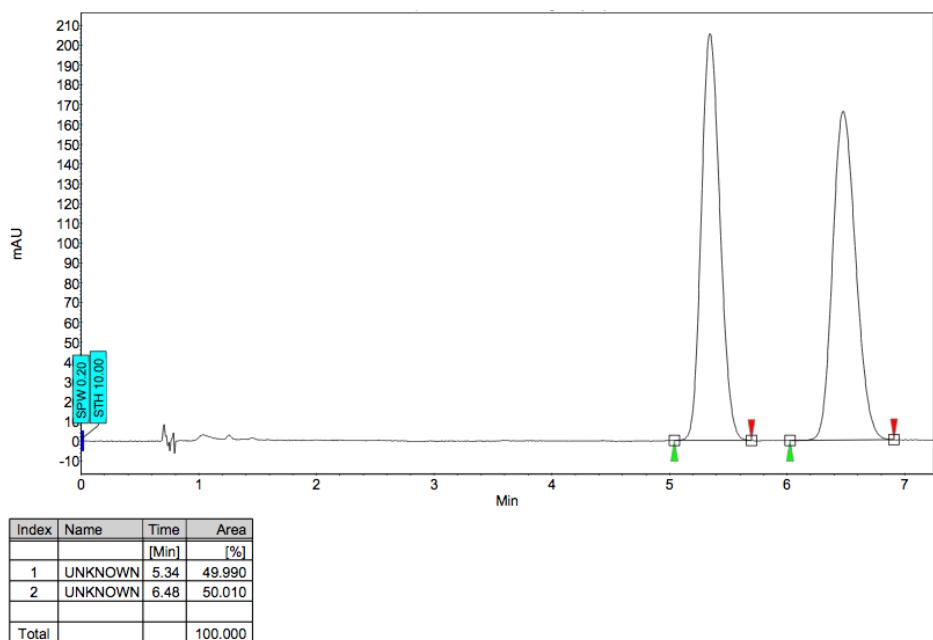


Following the general procedure **D** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 12% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 6.76$ min; minor enantiomer $t_R = 5.47$ min]

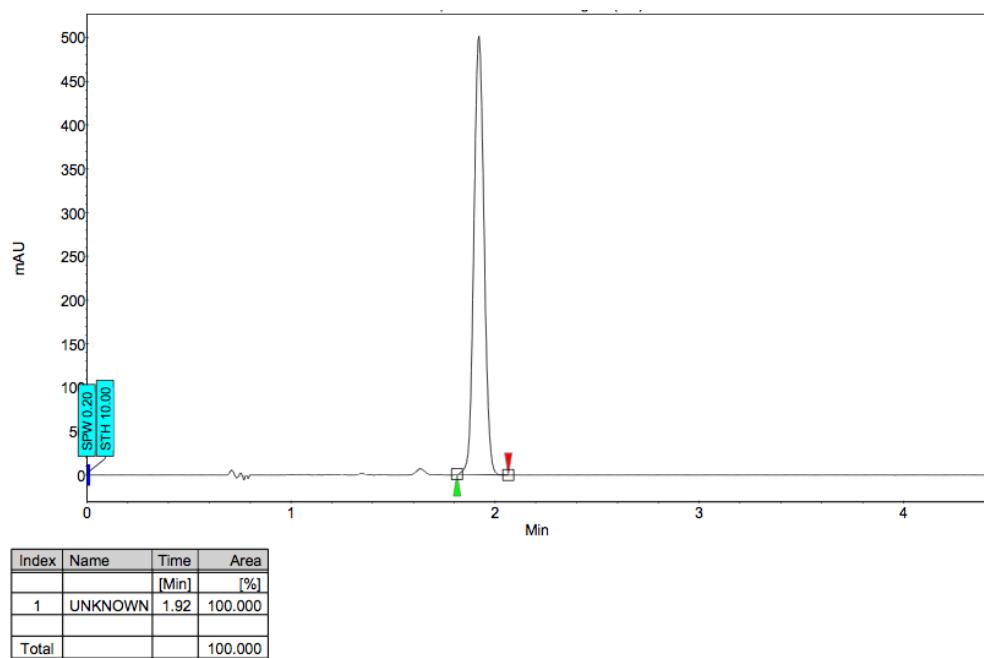


3-(5-Methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)octan-1-one (**2f**)

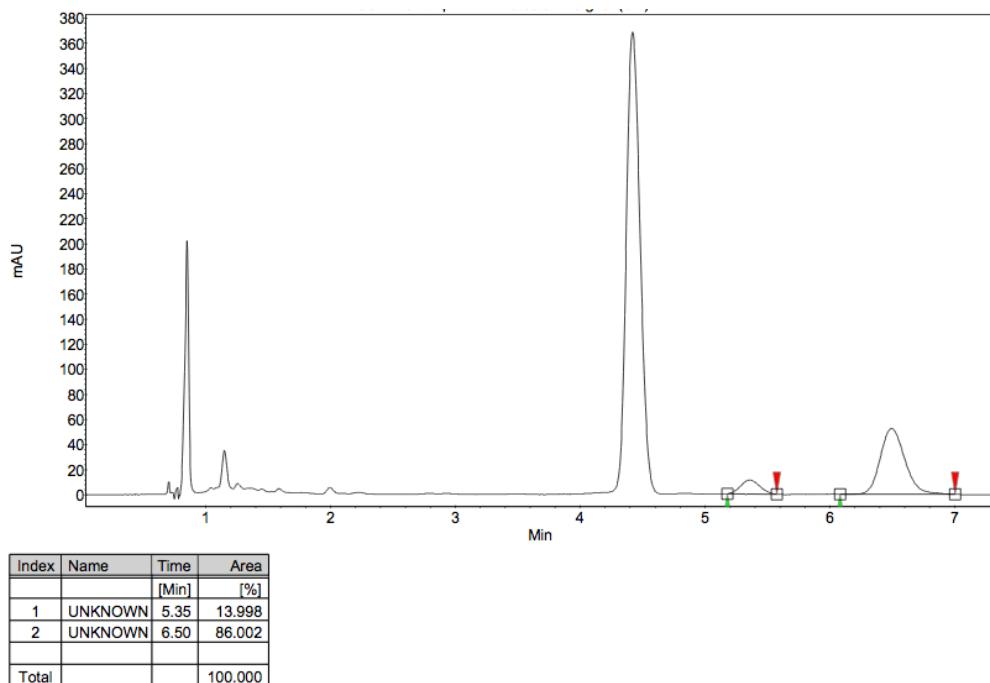
Racemic **2f**: [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 12% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 5.34$ min; second enantiomer $t_R = 6.48$ min].



Starting material (1c) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 12% MeOH; $\lambda = 220$ nm; $t_R = 1.92$ min]

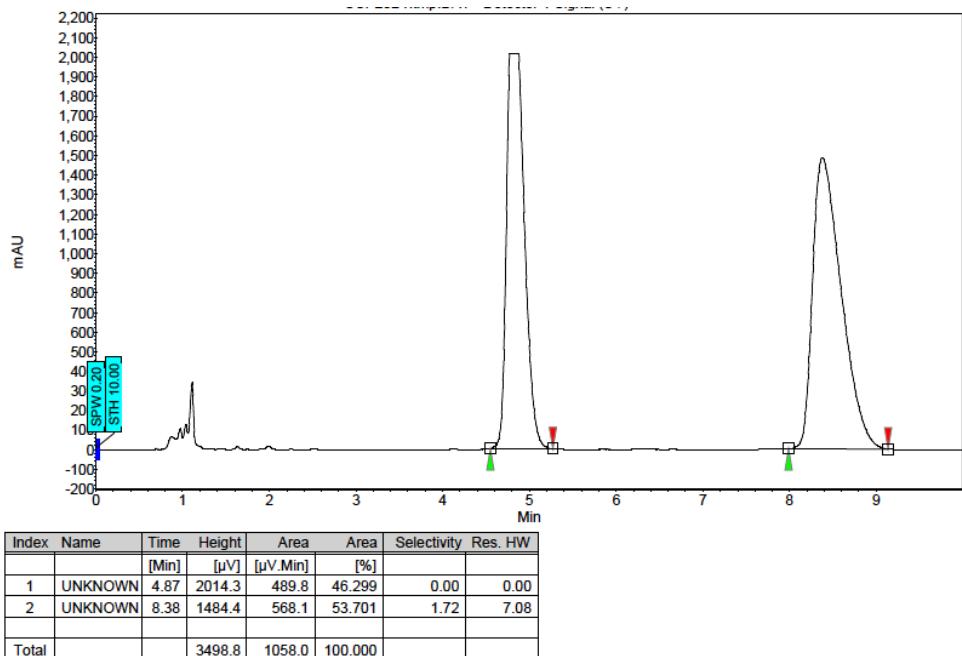


Following the general procedure **D** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 12% MeOH; $\lambda = 220$ nm; major enantiomer; $t_R = 6.50$ min; minor enantiomer; $t_R = 5.35$ min]

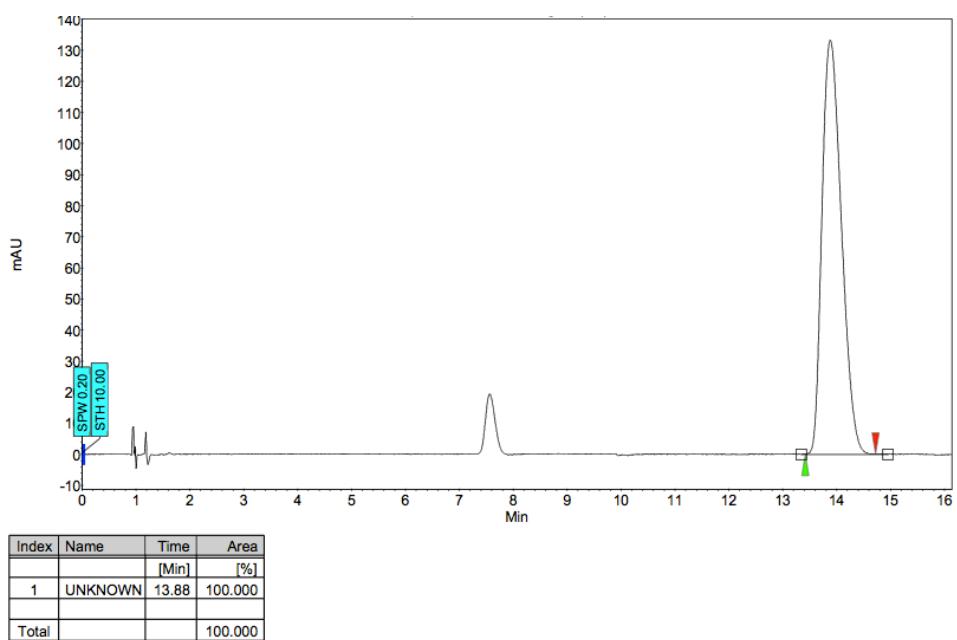


1-(1-Methyl-1*H*-imidazol-2-yl)-3-(5-methoxy-1*H*-indol-3-yl)-3-phenylpropan-1-one (2g)

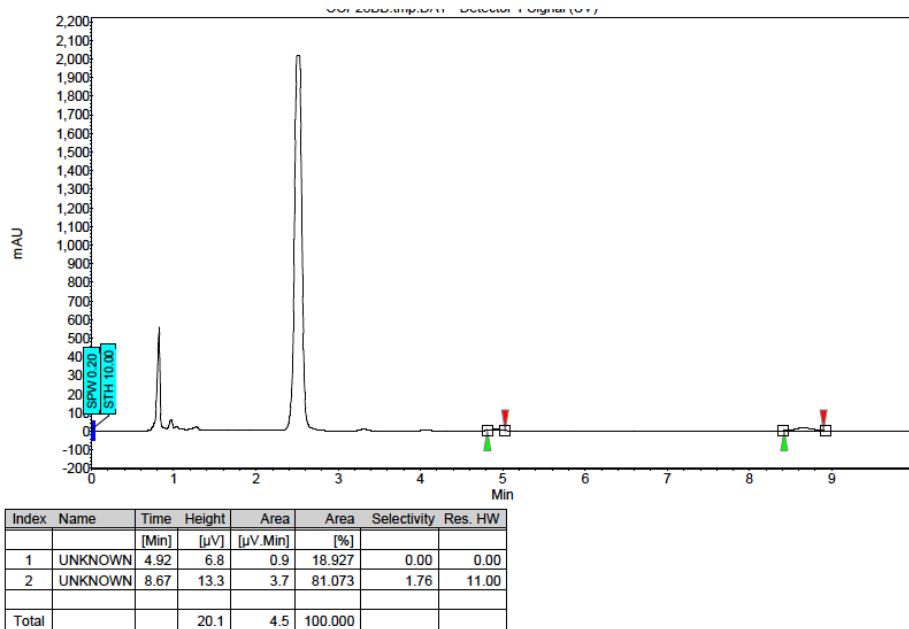
Racemic (2g) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 20% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 4.87$ min; second enantiomer $t_R = 8.38$ min]



Starting material (1d) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 20% MeOH; $\lambda = 220$ nm; $t_R = 13.88$ min]

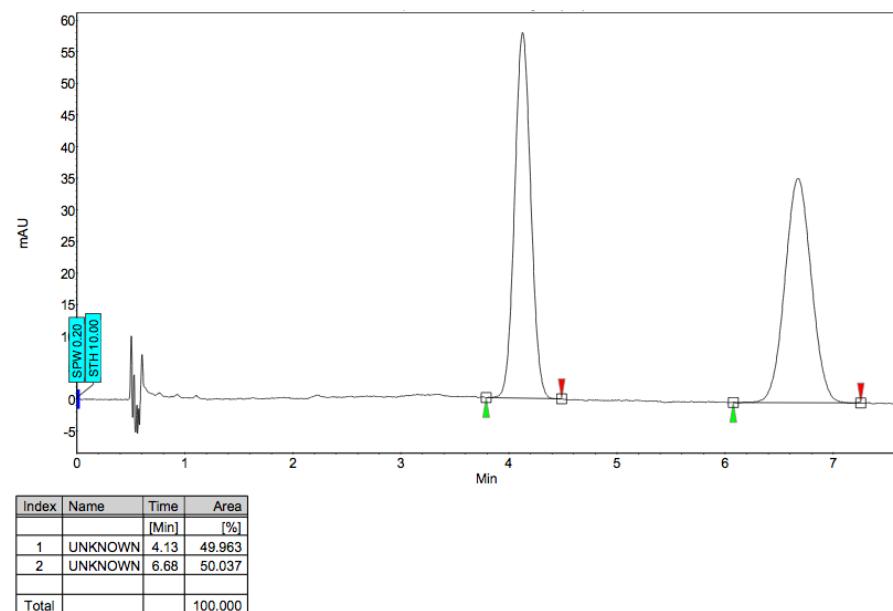


Following the general procedure **D** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 20% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.67$ min; minor enantiomer $t_R = 4.92$ min].

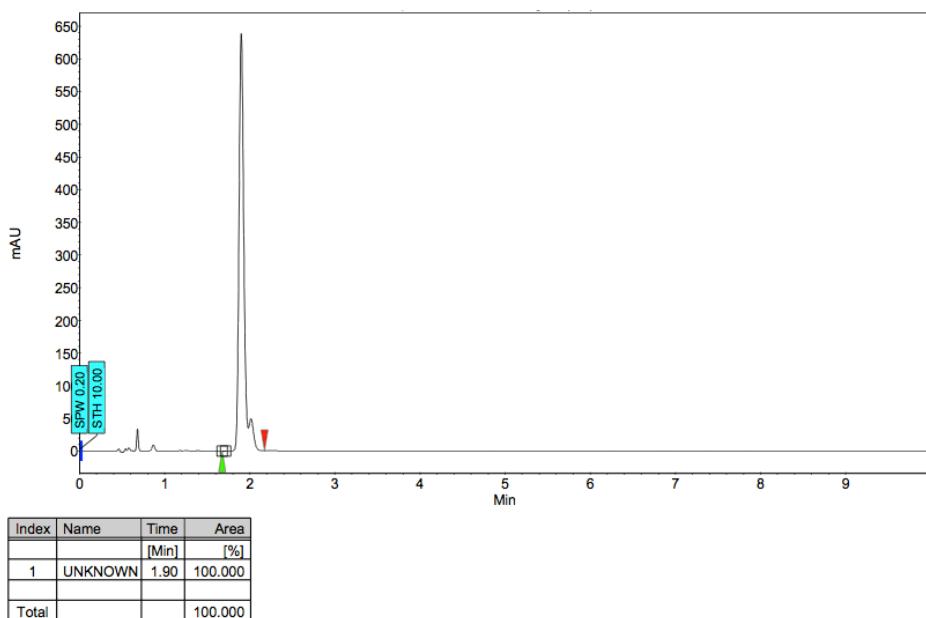


3-(5-Methoxy-1H-indol-3-yl)-3-(4-methoxyphenyl)-1-(1-methyl-1H-imidazol-2-yl)propan-1-one (2h**)**

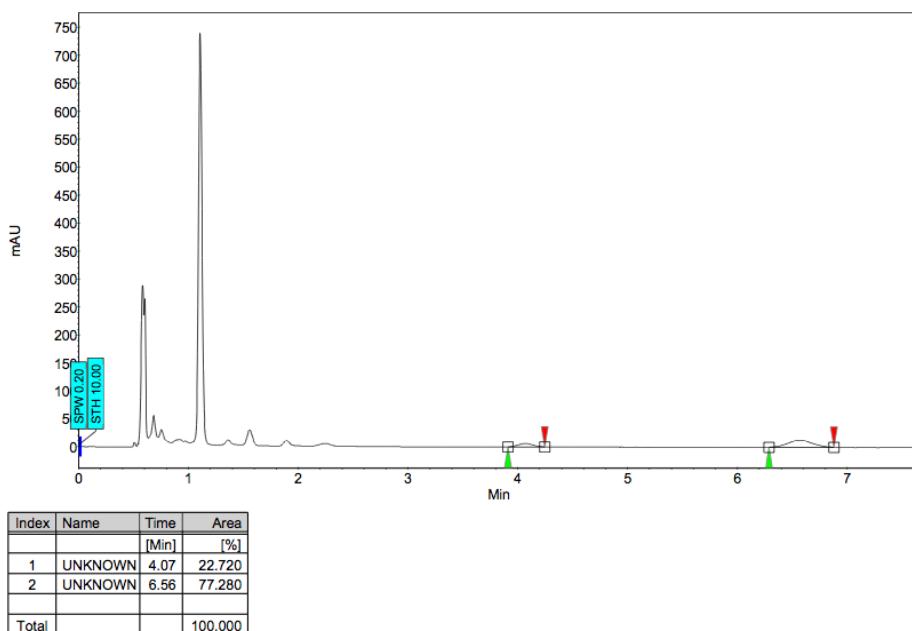
Racemic (2h**)** [DAICEL AD-H column; 100 bar; flow: 6.0 mL/min; 30% *i*-PrOH; $\lambda = 220$ nm; first enantiomer $t_R = 4.13$ min; second enantiomer $t_R = 6.68$ min]



Starting material (1e) [DAICEL AD-H column; 100 bar; flow: 6.0 mL/min; 30% *i*-PrOH; $\lambda = 220$ nm; $t_R = 1.90$ min]

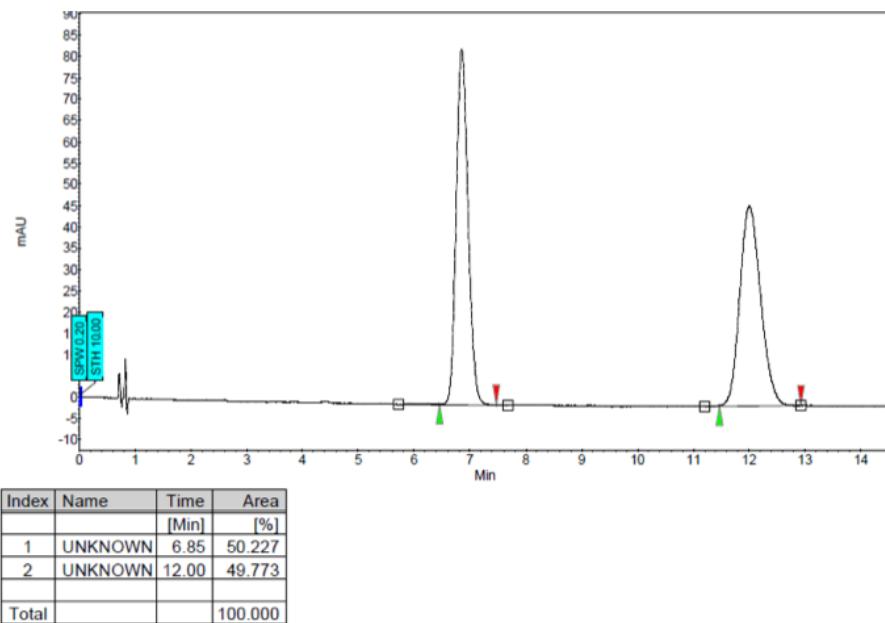


Following the general procedure **D** [DAICEL AD-H column; 100 bar; flow: 6.0 mL/min; 30% *i*-PrOH; $\lambda = 220$ nm, major enantiomer $t_R = 6.56$ min; minor enantiomer $t_R = 4.07$ min].

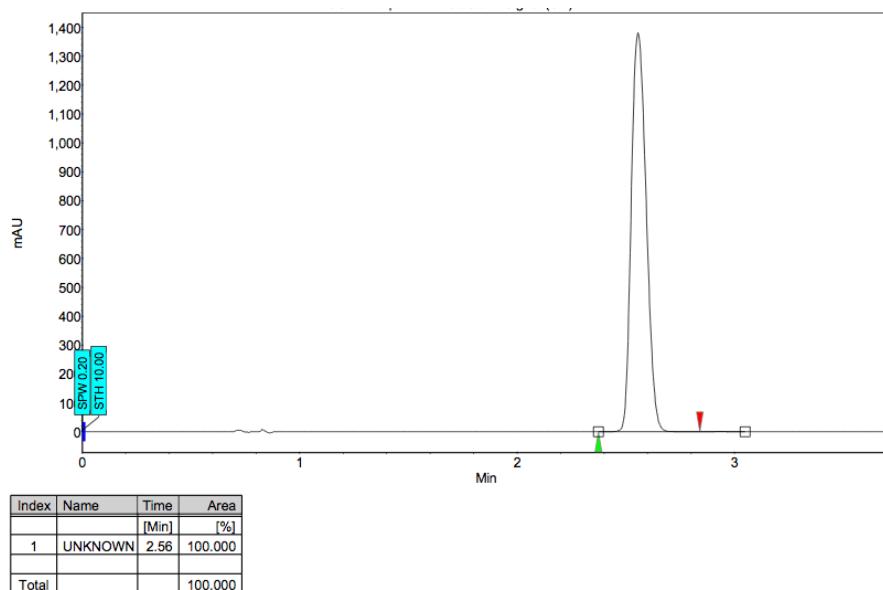


3-(4-Chlorophenyl)-3-(5-methoxy-1*H*-indol-3-yl)-1-(1-methyl-1*H*-imidazol-2-yl)propan-1-one(2i)

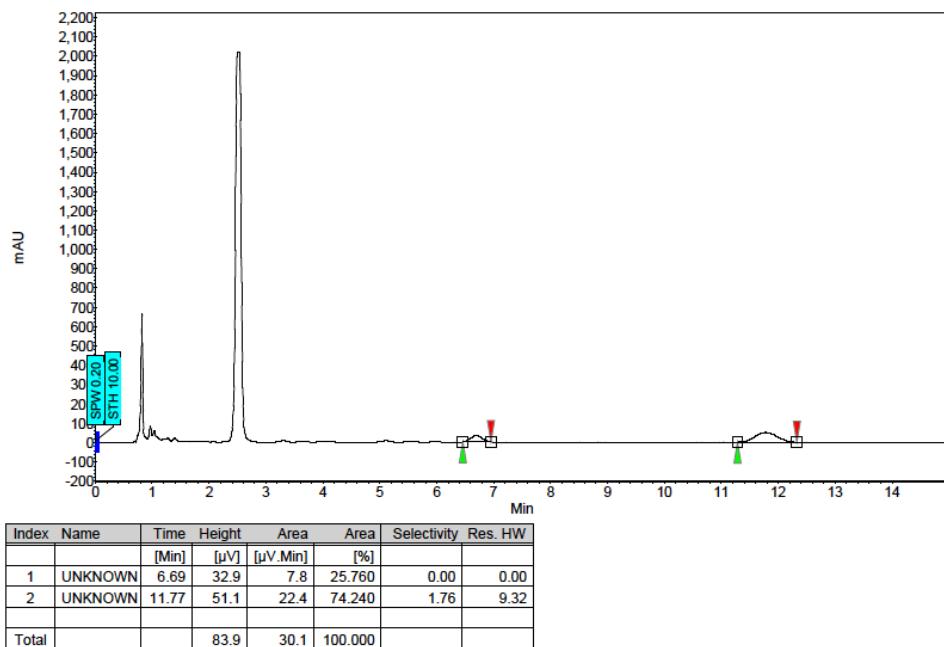
Racemic (2i) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 20% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 6.85$ min; second enantiomer $t_R = 12.00$ min]



Starting material **1f**: [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 20% MeOH; $\lambda = 220$ nm; $t_R = 6.11$ min].

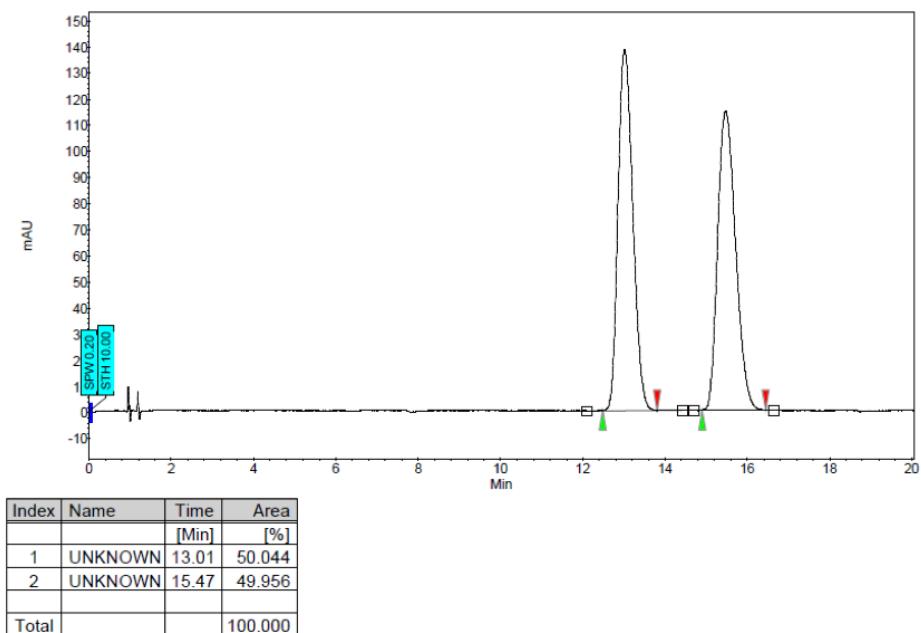


Following the general procedure **D** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 20% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 11.77$ min; minor enantiomer $t_R = 6.69$ min].

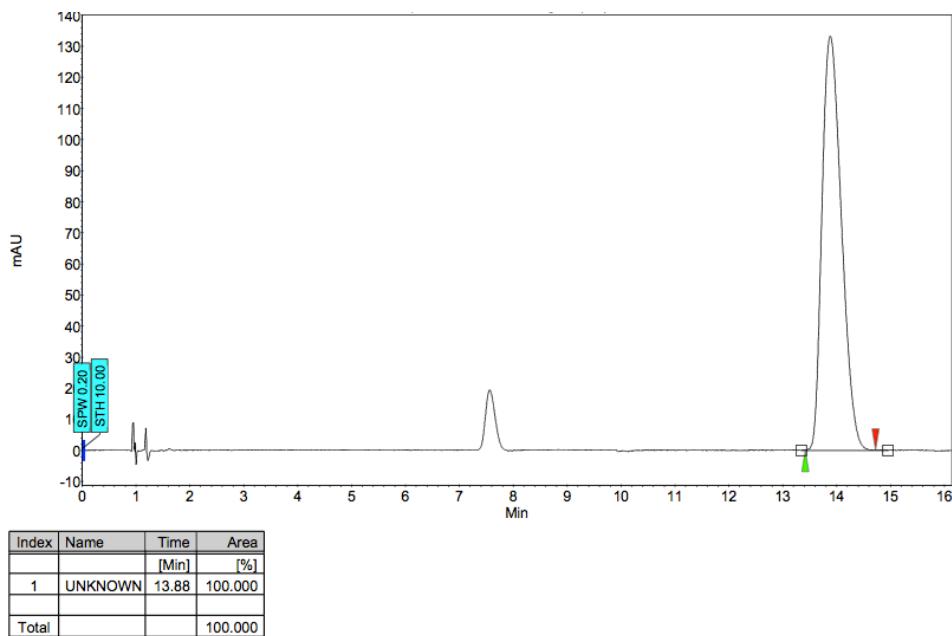


1-(1-Methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)-3-phenylpropan-1-one (2j)

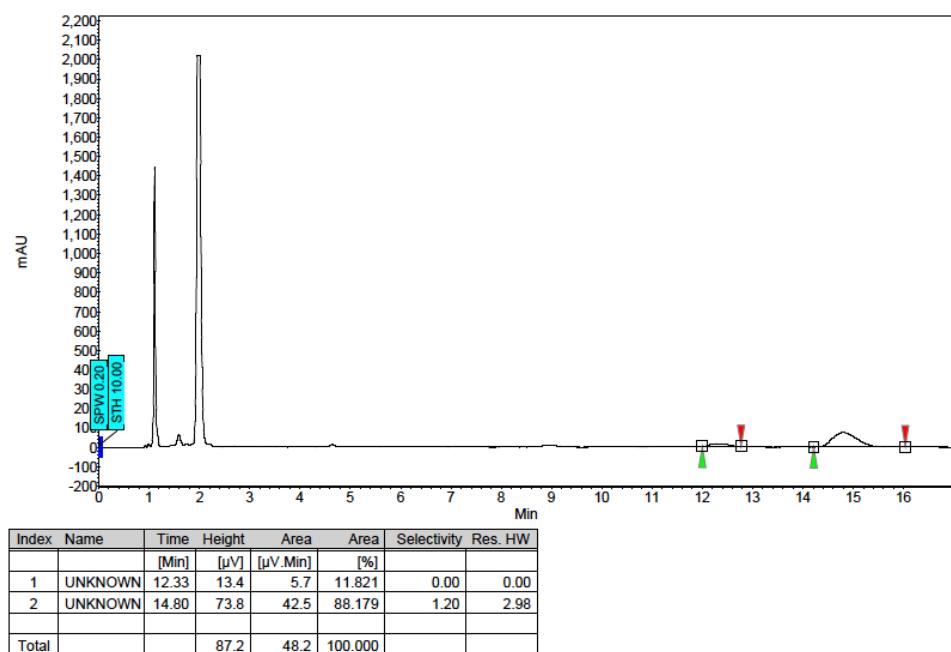
Racemic (2j) [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 10% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 13.01$ min; second enantiomer $t_R = 15.47$ min]



Starting material (1d) [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 10% MeOH; $\lambda = 220$ nm; $t_R = 13.88$ min]

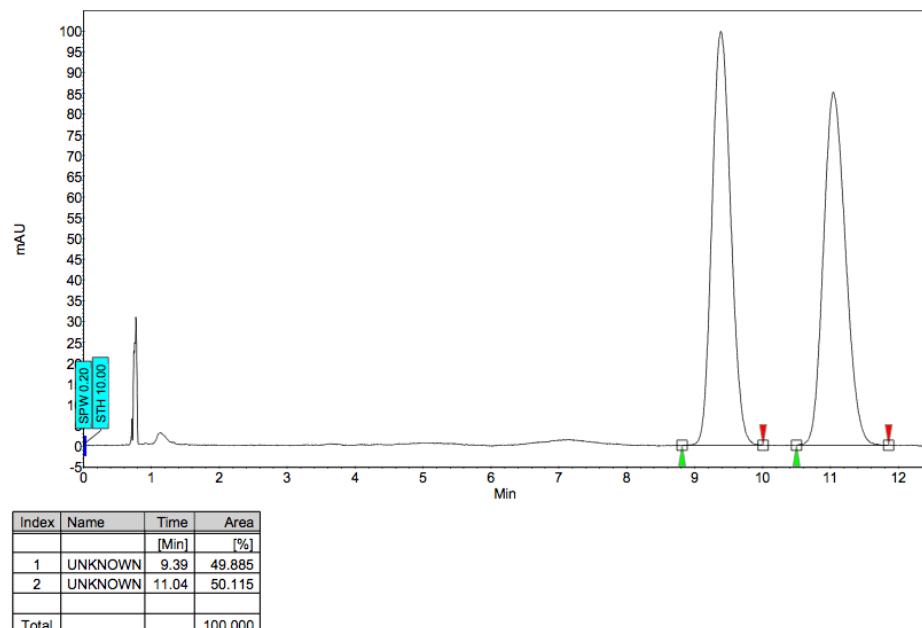


Following the general procedure **D** [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 10% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 14.80$ min; minor enantiomer $t_R = 12.33$ min]

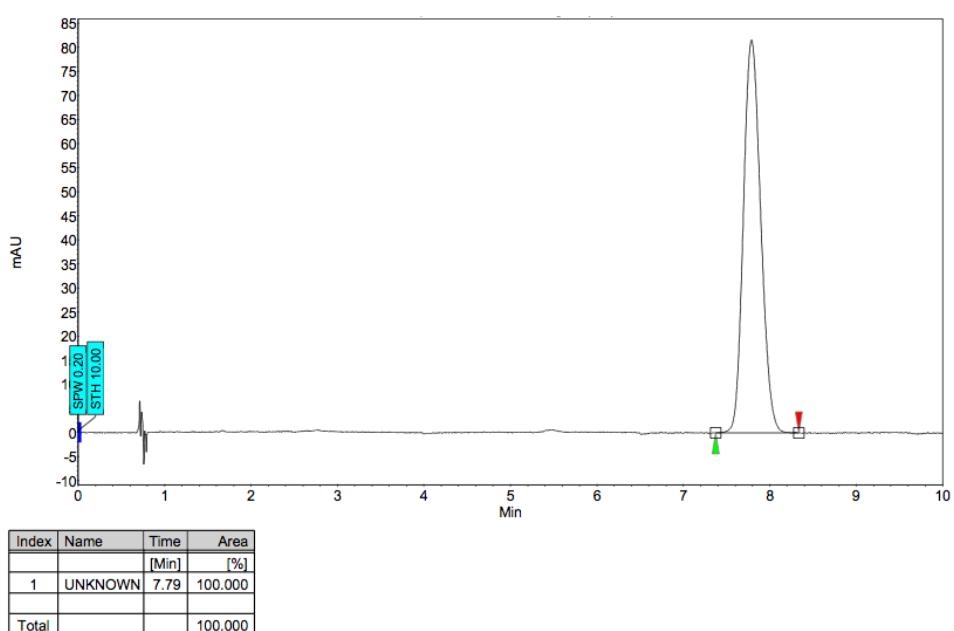


3-(2-Bromophenyl)-1-(1-methyl-1*H*-imidazol-2-yl)-3-(1-methyl-1*H*-indol-3-yl)propan-1-one (2k)

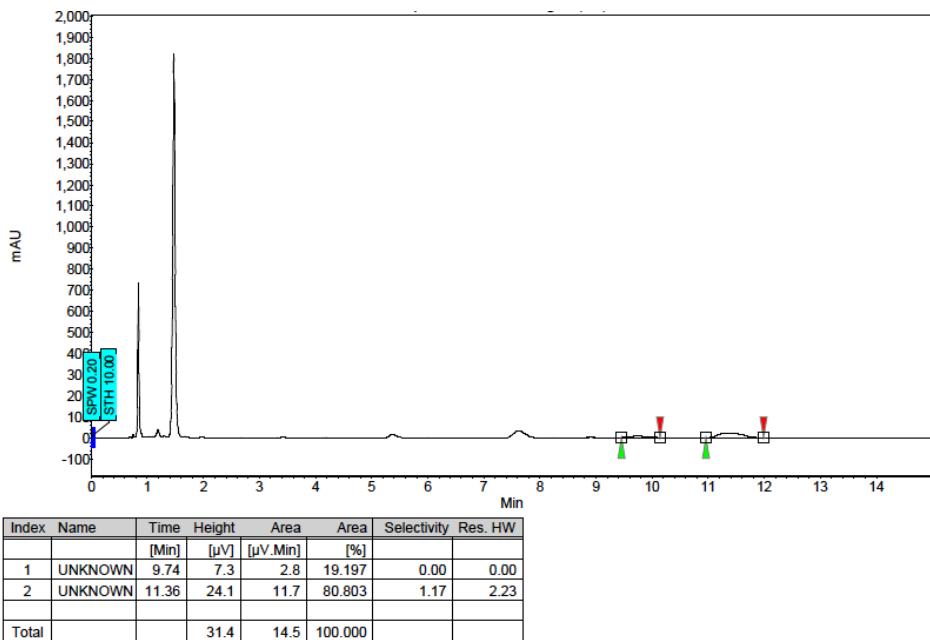
Racemic (2k) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 10% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 9.39$ min; second enantiomer $t_R = 11.04$ min]



Starting material (1g) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 10% MeOH; $\lambda = 220$ nm; $t_R = 7.79$ min]

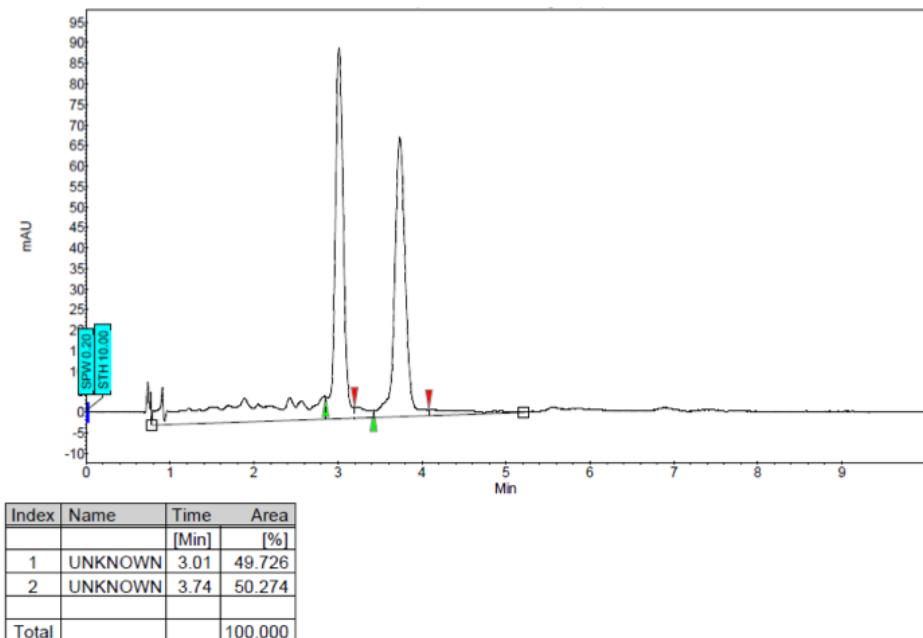


Following the general procedure **D** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 10% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 11.36$ min; minor enantiomer $t_R = 9.74$ min]

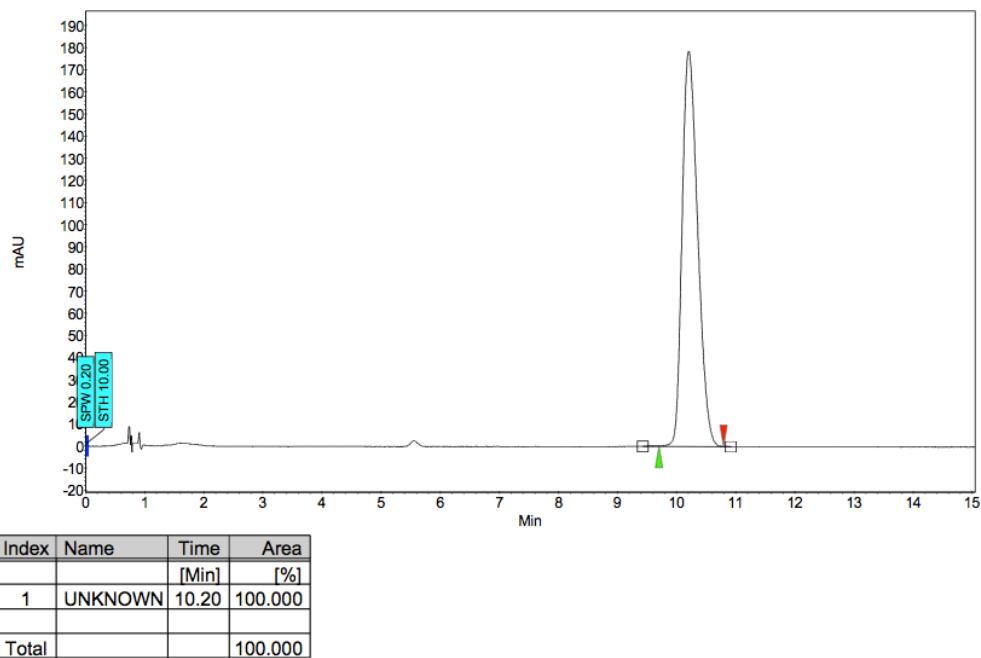


Dimethyl 2-(3-(1-methyl-1*H*-imidazol-2-yl)-3-oxo-1-phenylpropyl)malonate (**3a**)

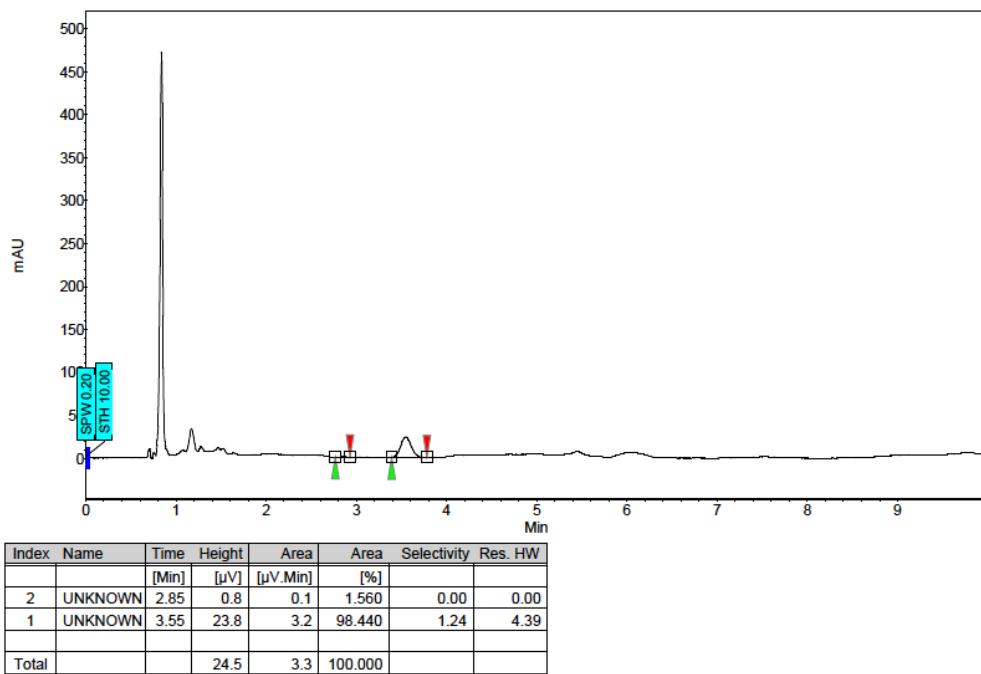
Racemic (3a**)** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 10% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 3.01$ min; second enantiomer $t_R = 3.74$ min]



Starting material (1d) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 10% MeOH; $\lambda = 220$ nm; $t_R = 10.20$ min]

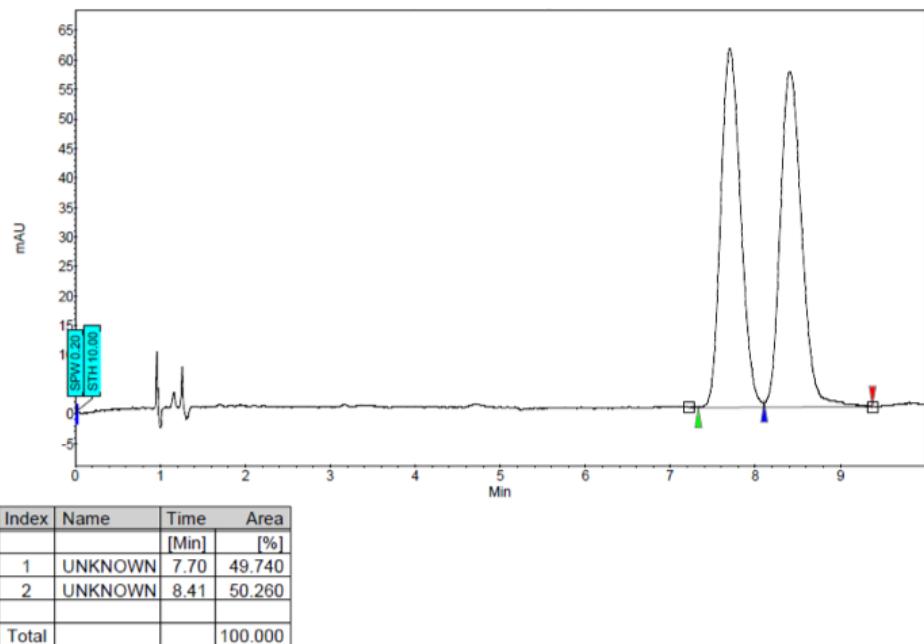


Following the general procedure F [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 10% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 3.55$ min; minor enantiomer $t_R = 2.85$ min]

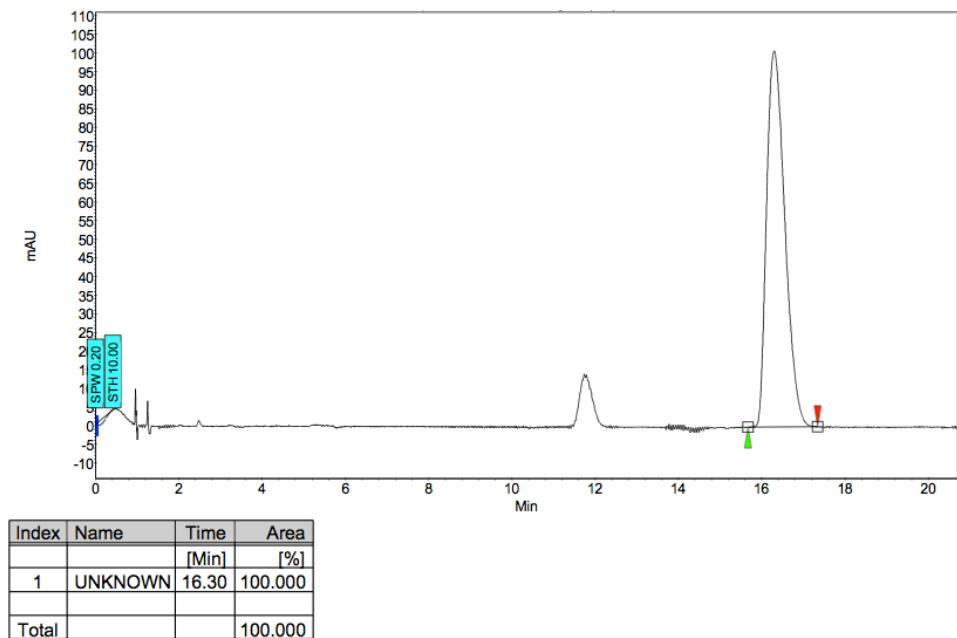


**Dimethyl 2-(1-(2-bromophenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate
(3b)**

Racemic (3b) [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 7% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 8.41$ min; second enantiomer $t_R = 7.70$ min]

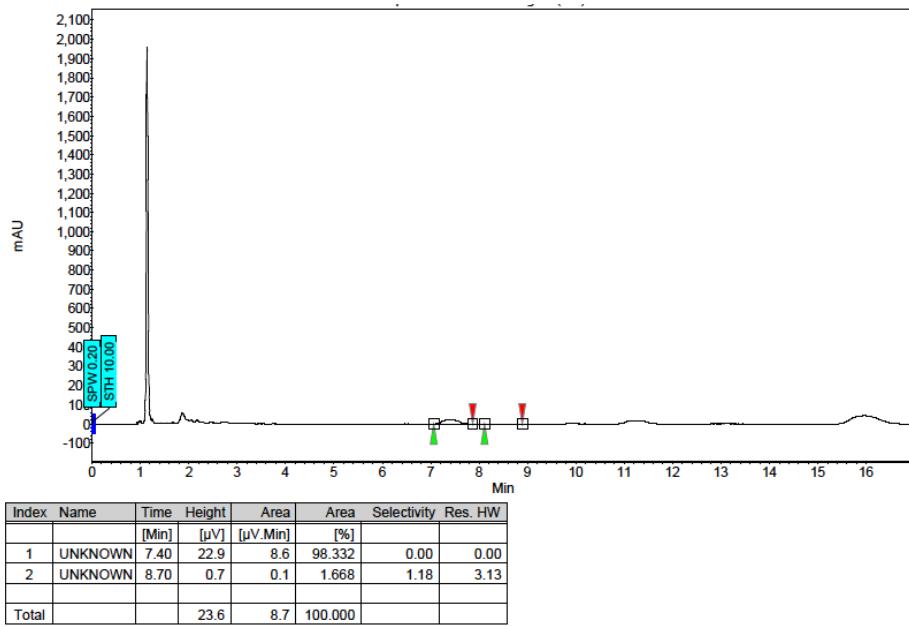


Starting material (1g) [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 7% MeOH; $\lambda = 220$ nm; $t_R = 16.30$ min]



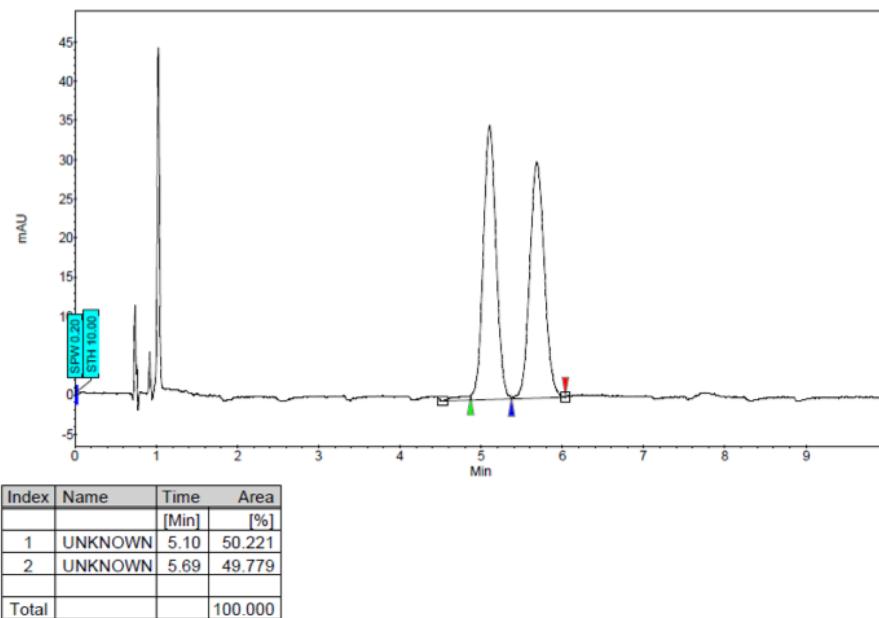
v

Following the general procedure F [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 7% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 7.40$ min; minor enantiomer $t_R = 8.70$ min]

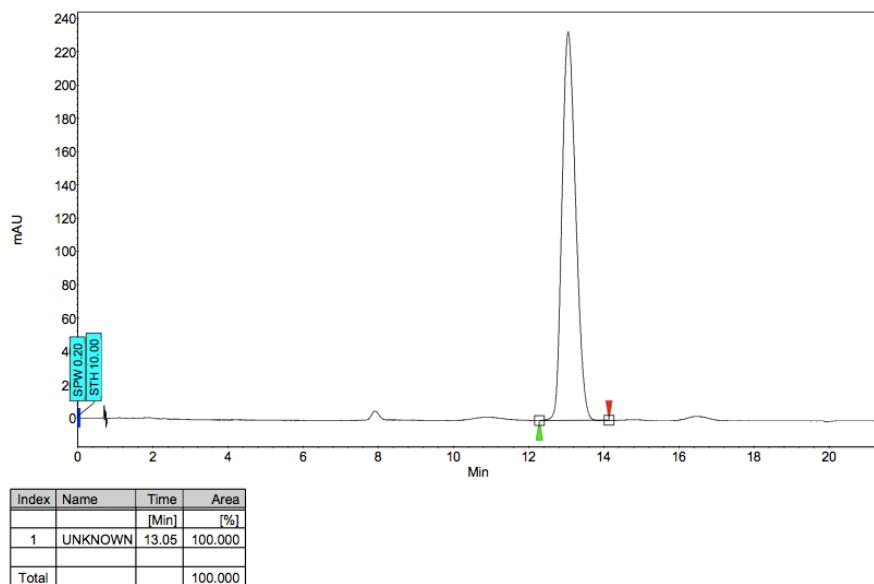


Dimethyl 2-(1-(4-methoxyphenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl) malonate (3c)

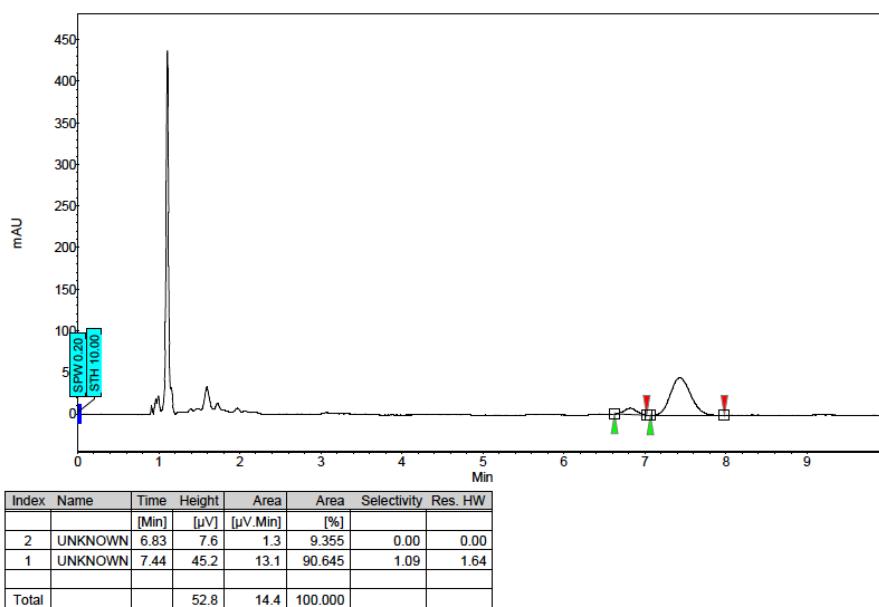
Racemic (3b) [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 10% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 5.10$ min; second enantiomer $t_R = 5.69$ min]



Starting material (1e) [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 10% MeOH; $\lambda = 220$ nm; $t_R = \text{min}$]

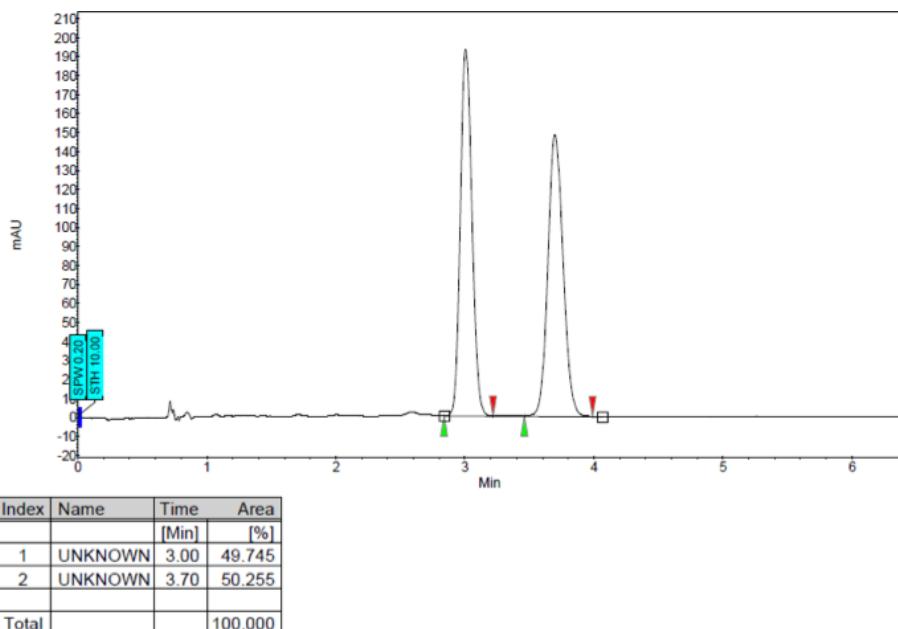


Following the general procedure F [DAICEL AD-H column; 100 bar; flow: 3.0 mL/min; 10% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 7.44$ min; minor enantiomer $t_R = 6.83$ min].

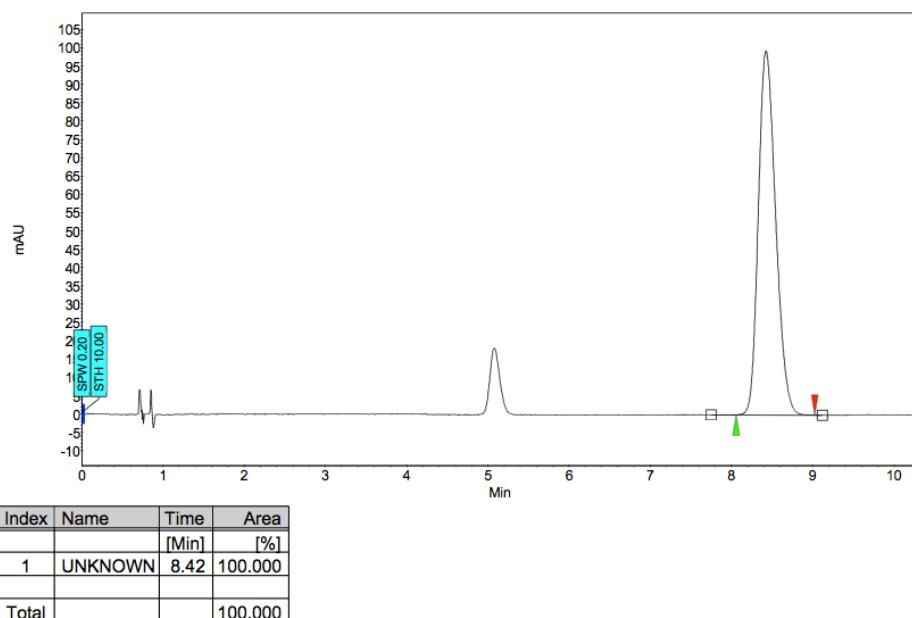


Dimethyl 2-(1-(4-chlorophenyl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl) malonate (3d)

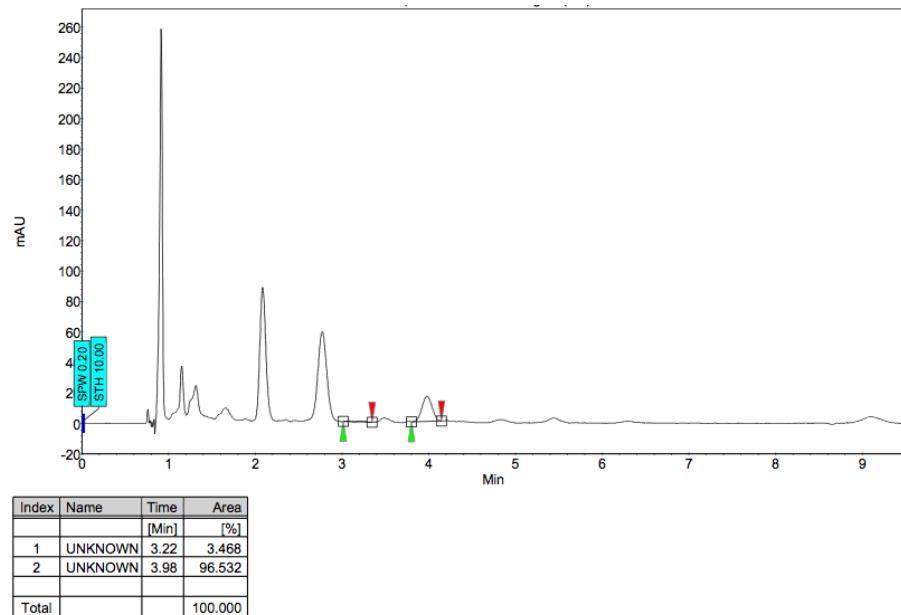
Racemic (3d) [DAICEL AD-H column; 100 bar; flow: 6.0 mL/min; 30% *i*-PrOH; $\lambda = 220$ nm; first enantiomer $t_R = 3.00$ min; second enantiomer $t_R = 3.70$ min].



Starting material (1f) [DAICEL AD-H column; 100 bar; flow: 6.0 mL/min; 30% *i*-PrOH; $\lambda = 220$ nm; $t_R = 8.42$ min].

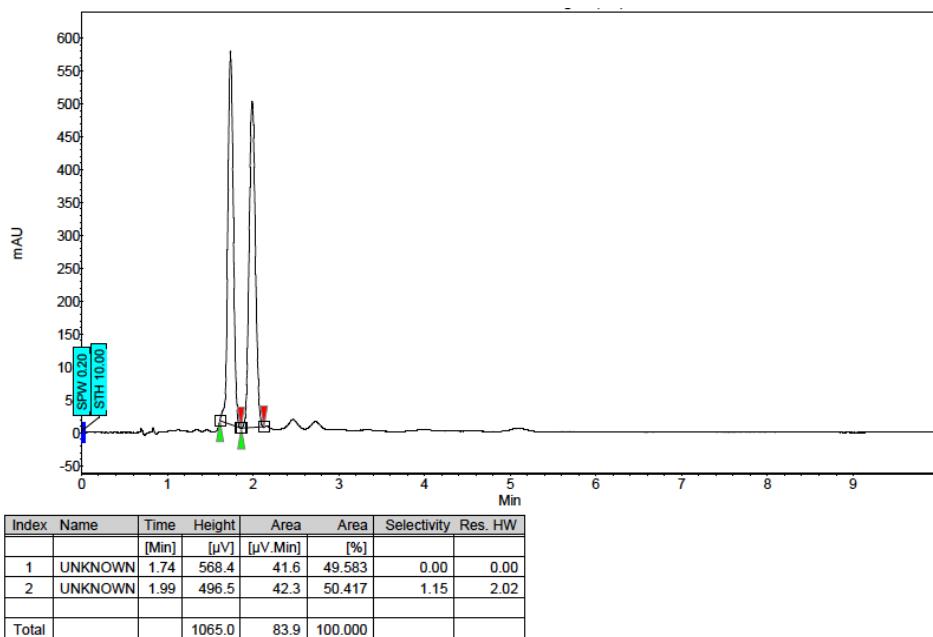


Following the general procedure F [DAICEL AD-H column; 100 bar flow: 6.0 mL/min; 30% *i*-PrOH; $\lambda = 220$ nm, major enantiomer $t_R = 3.98$ min; minor enantiomer $t_R = 3.22$ min].

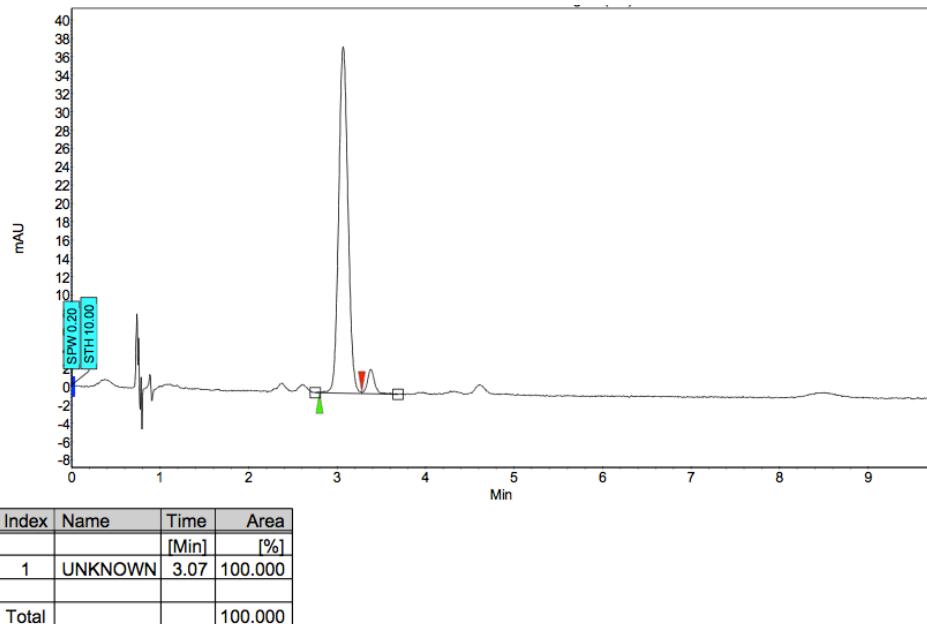


Dimethyl 2-(1-(furan-2-yl)-3-(1-methyl-1*H*-imidazol-2-yl)-3-oxopropyl)malonate (3e)

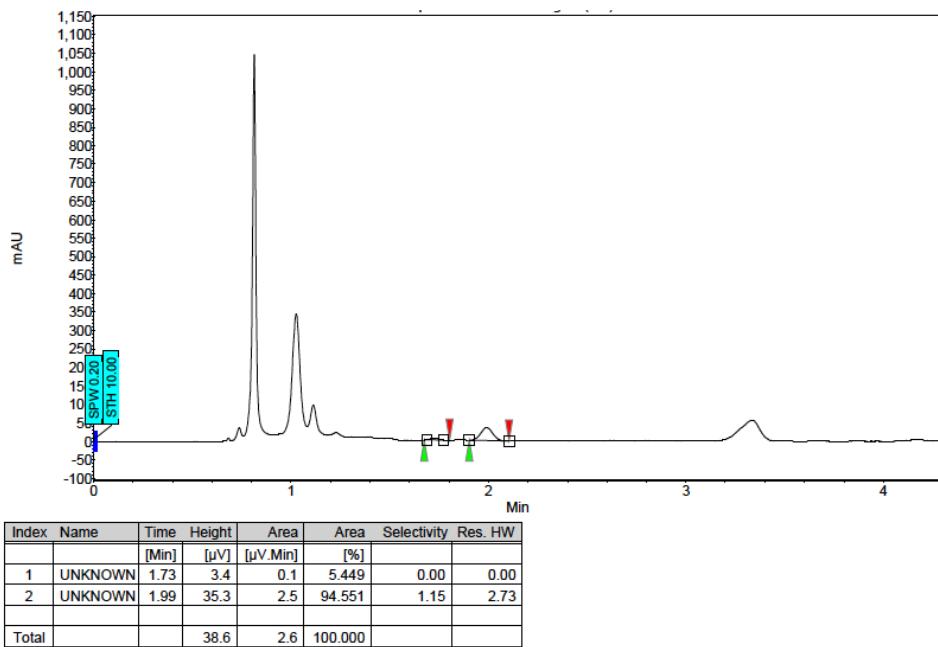
Racemic (3e) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; first enantiomer $t_R = 1.74$ min; second enantiomer $t_R = 1.99$ min].



Starting material (1h) [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm; $t_R = 3.07$ min].

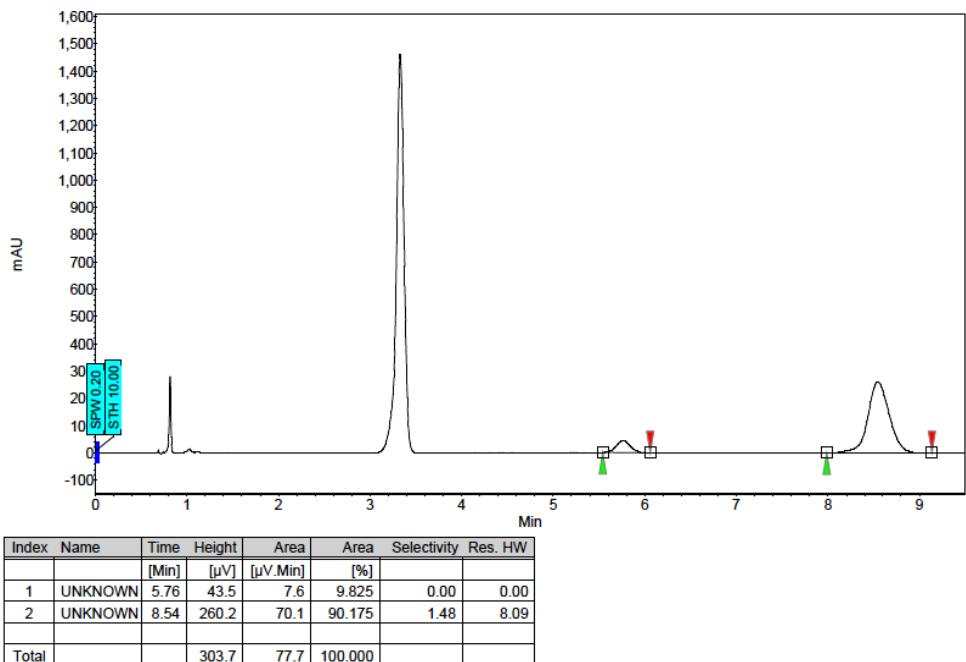


Following the general procedure F [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 1.99$ min; minor enantiomer $t_R = 1.73$ min].

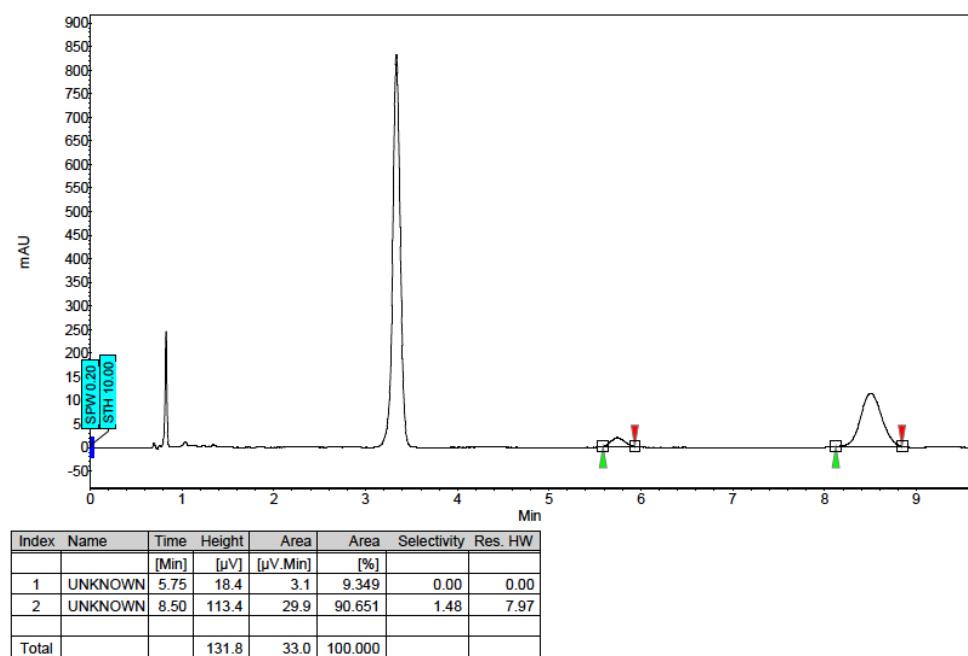


Enantioselective Recycling of Friedel Crafts alkylations

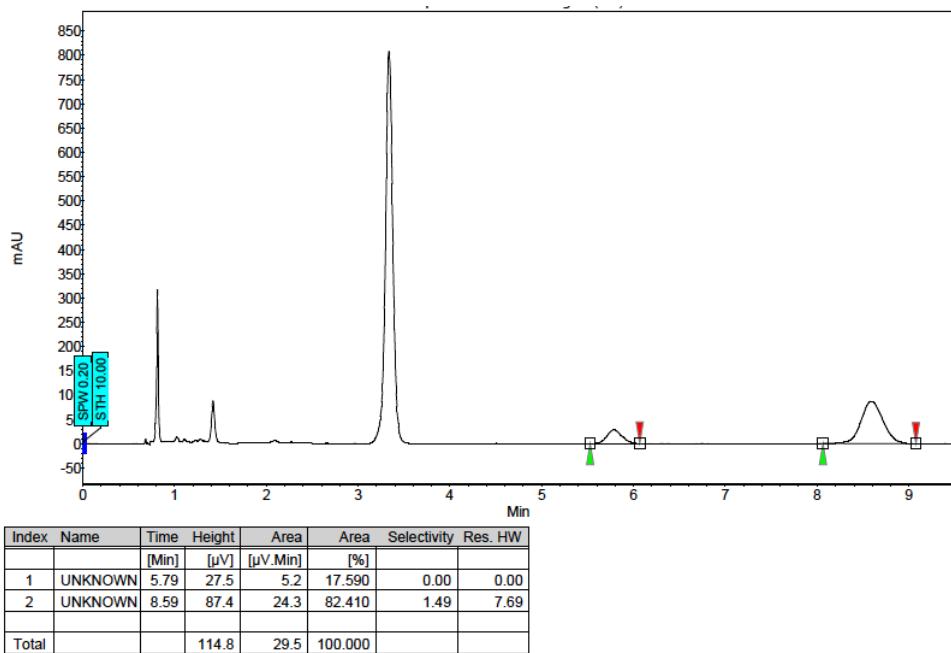
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.54$ min; minor enantiomer $t_R = 5.76$ min].



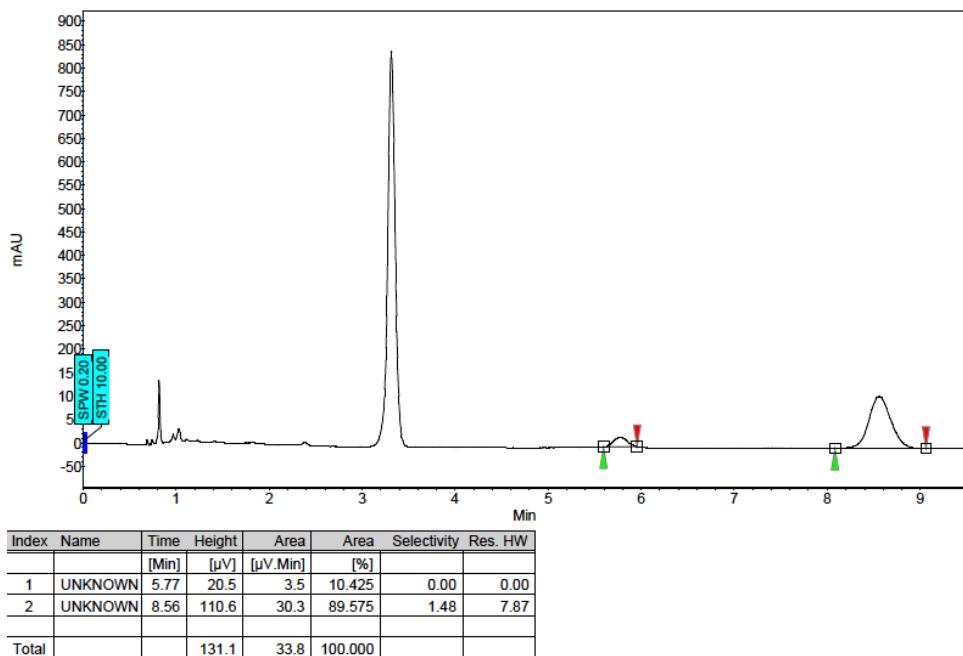
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.50$ min; minor enantiomer $t_R = 5.75$ min].



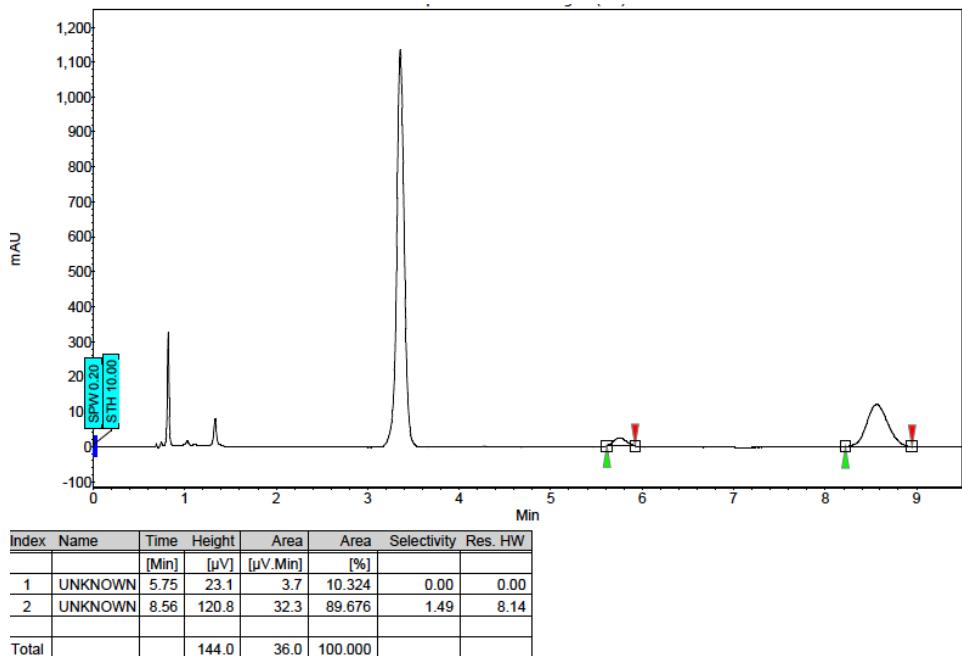
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.59$ min; minor enantiomer $t_R = 5.79$ min].



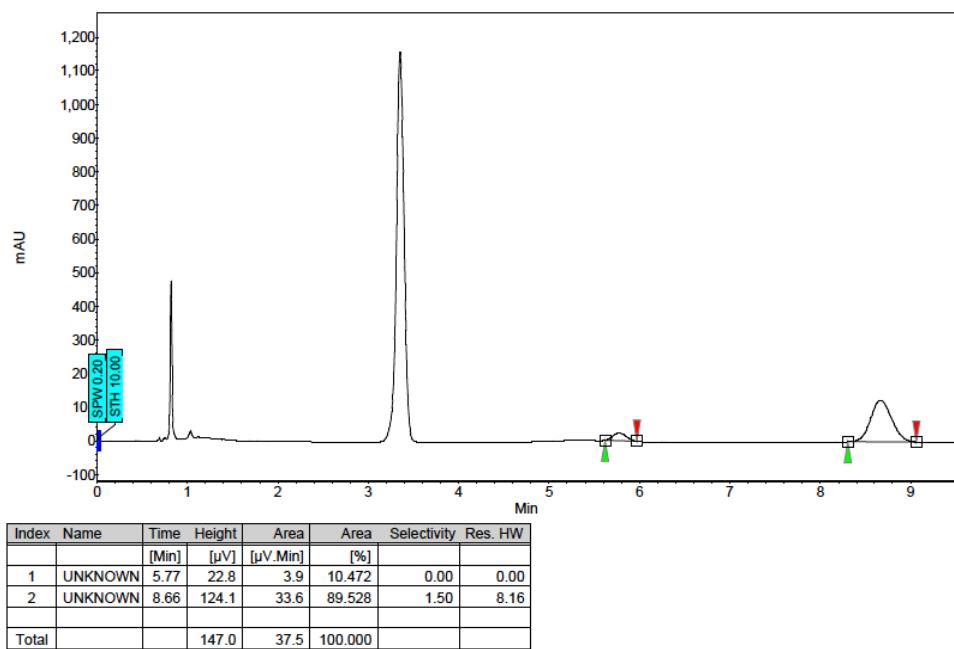
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.56$ min; minor enantiomer $t_R = 5.77$ min].



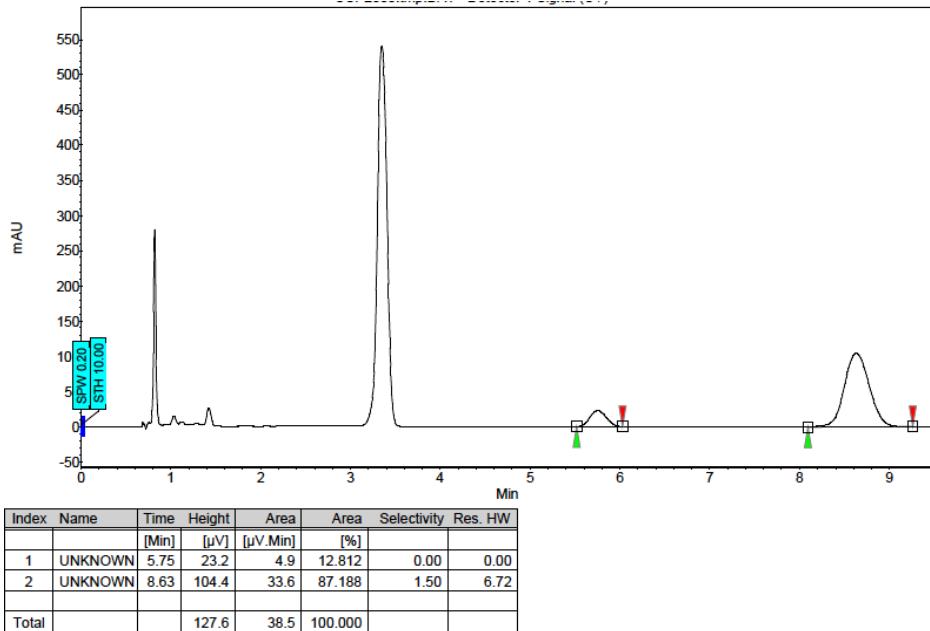
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.56$ min; minor enantiomer $t_R = 5.75$ min].



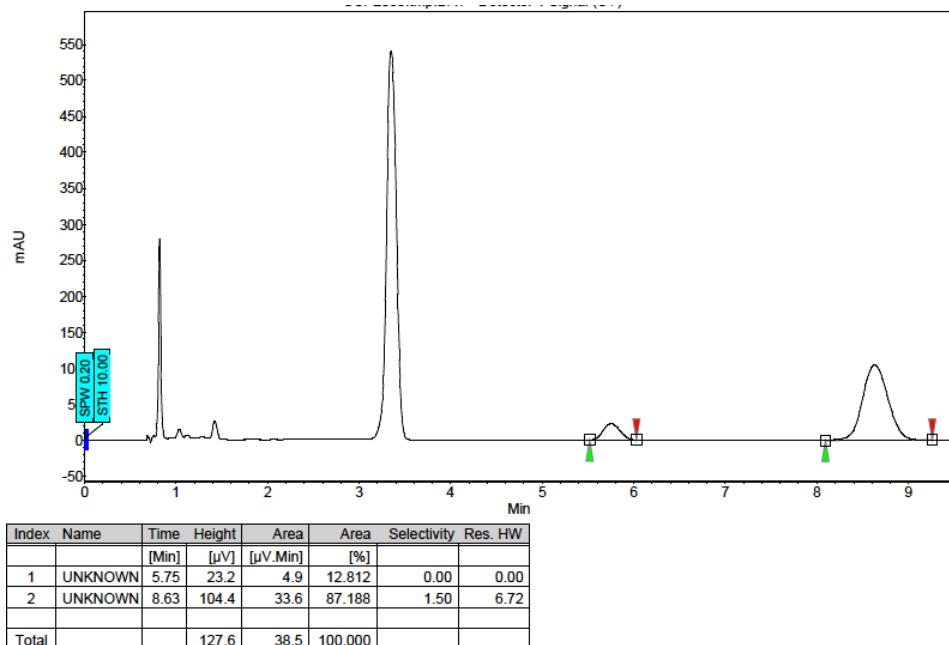
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.66$ min; minor enantiomer $t_R = 5.77$ min].



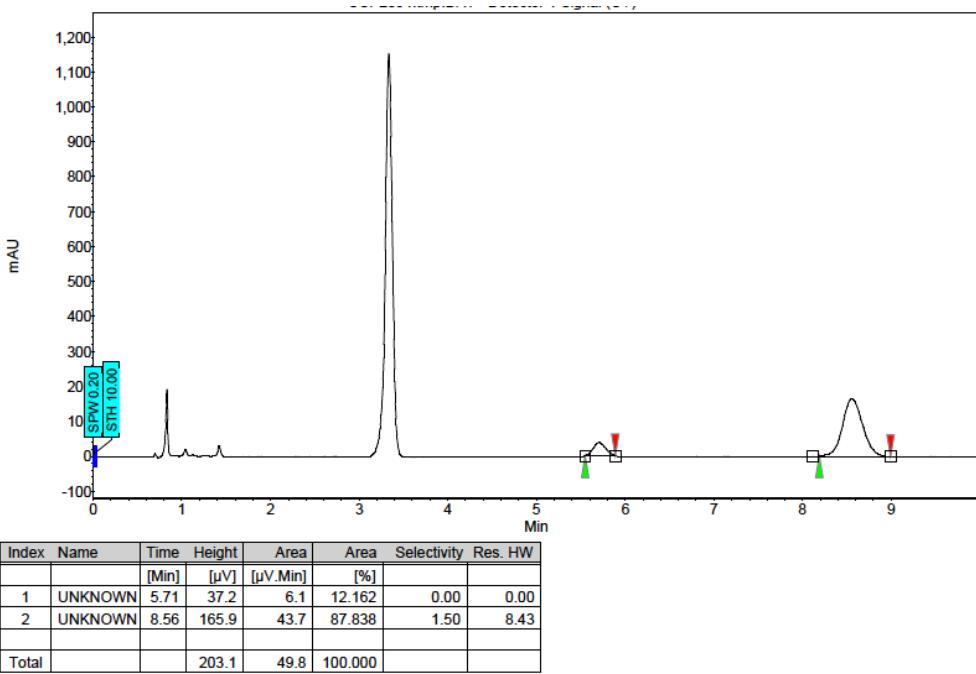
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.63$ min; minor enantiomer $t_R = 5.75$ min].



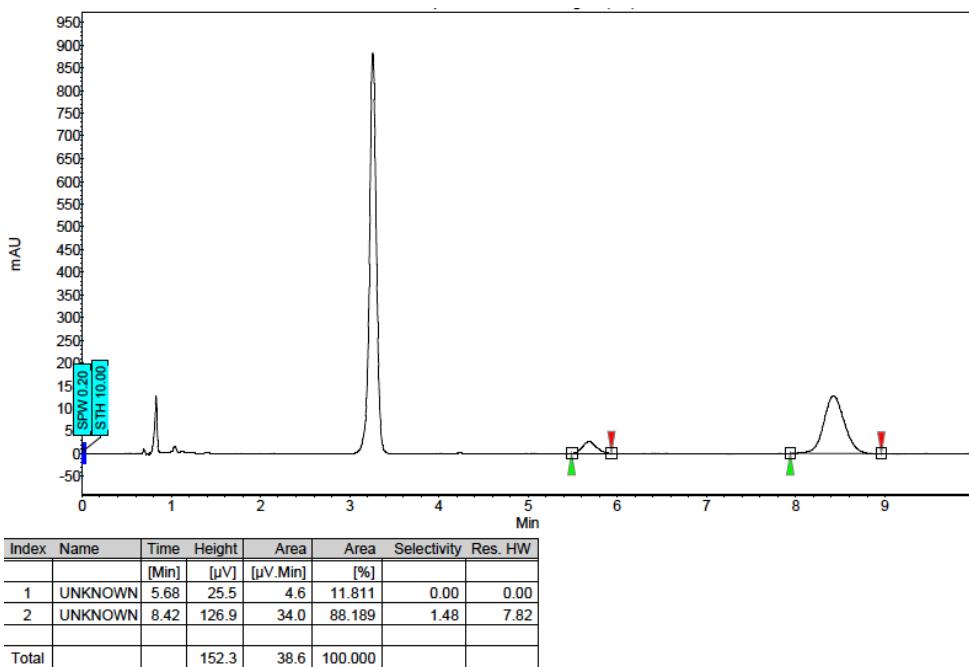
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.63$ min; minor enantiomer $t_R = 5.75$ min].



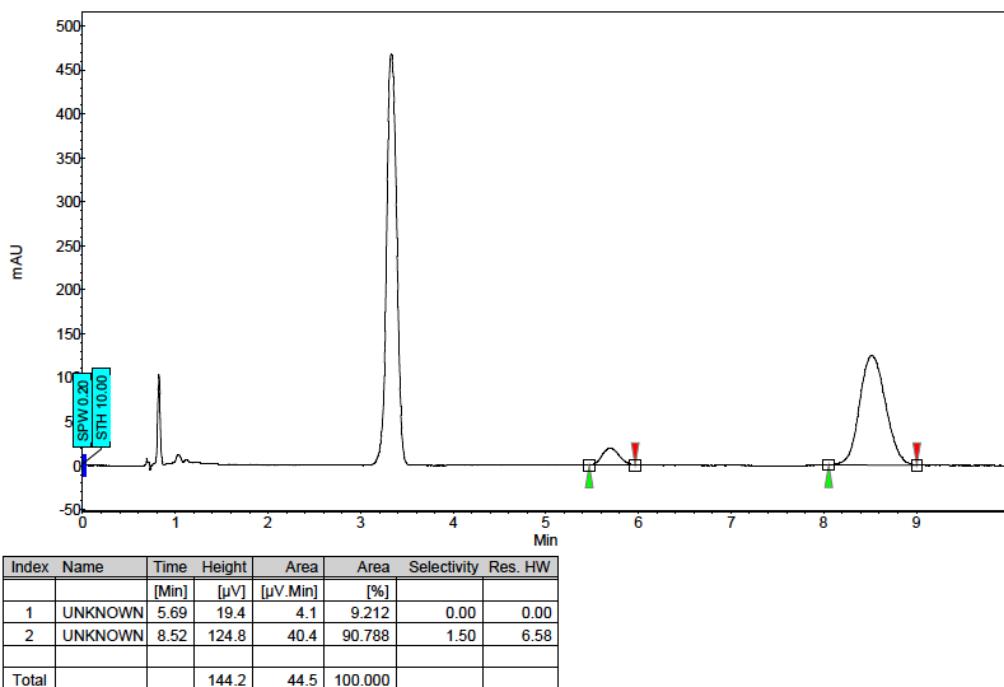
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.56$ min; minor enantiomer $t_R = 5.71$ min].



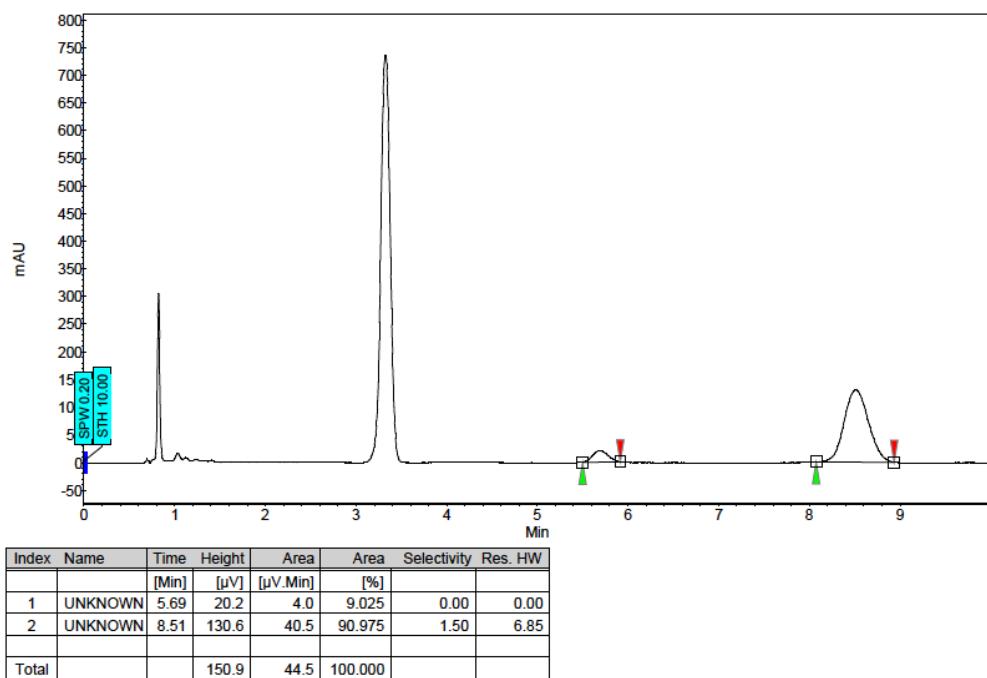
Following General procedure G [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.42$ min; minor enantiomer $t_R = 5.68$ min].



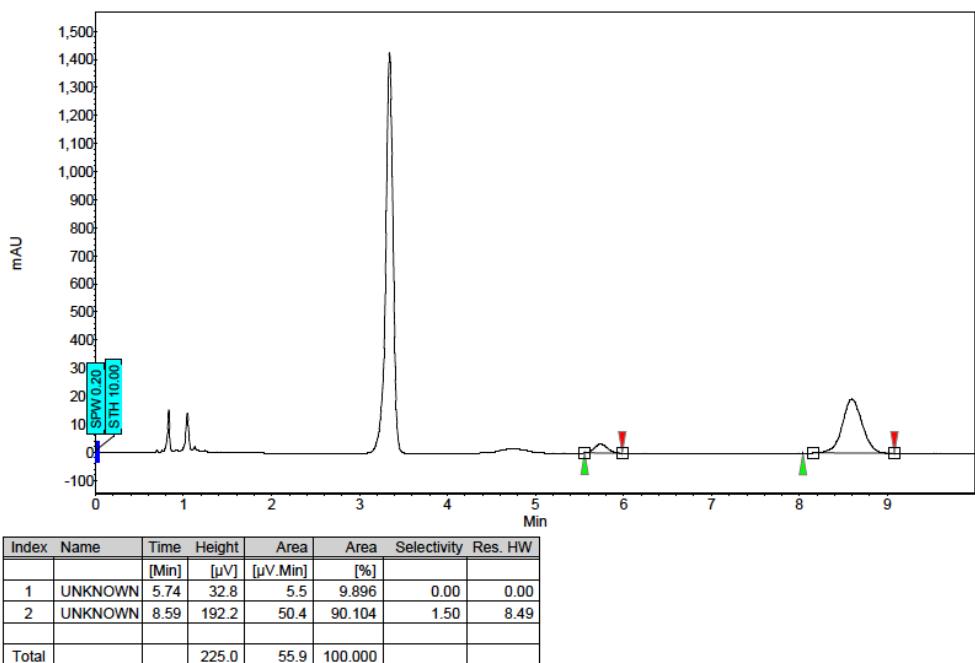
Following General procedure **H** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.52$ min; minor enantiomer $t_R = 5.69$ min].



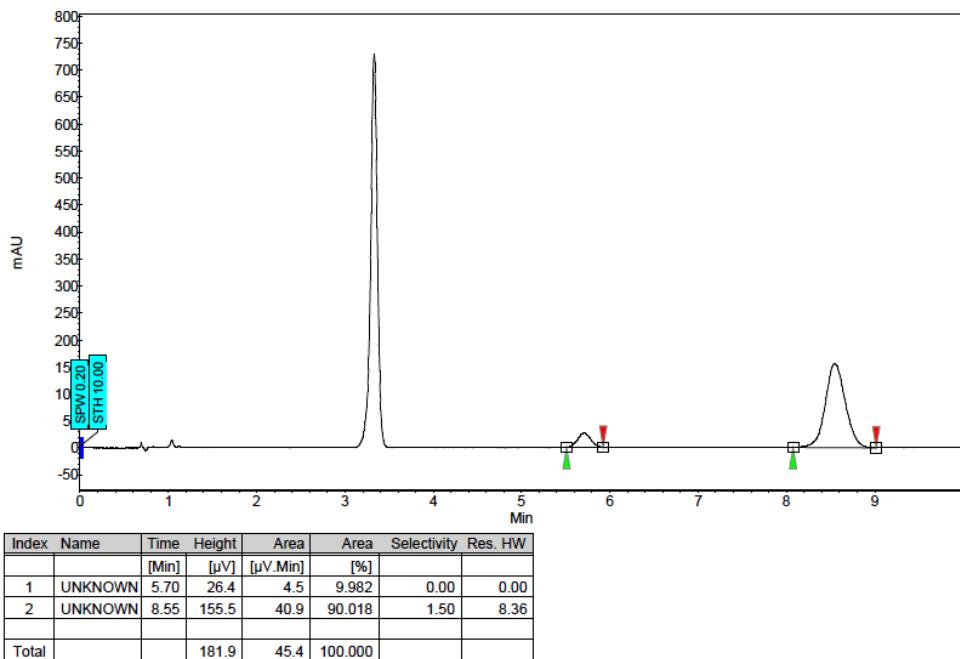
Following General procedure **H** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.51$ min; minor enantiomer $t_R = 5.69$ min].



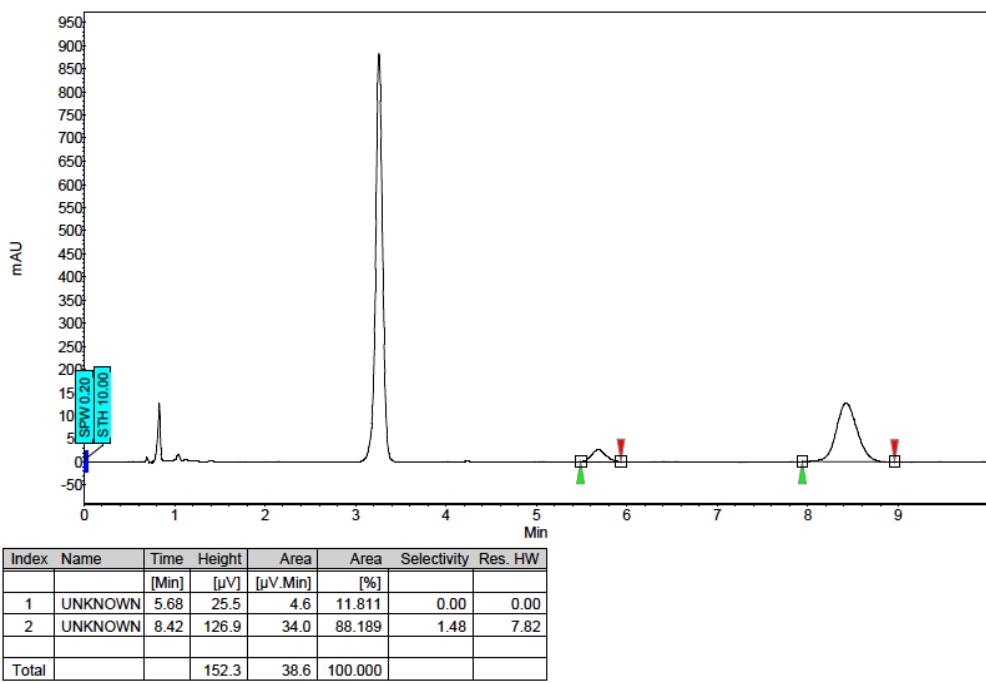
Following General procedure **H** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.49$ min; minor enantiomer $t_R = 5.66$ min].



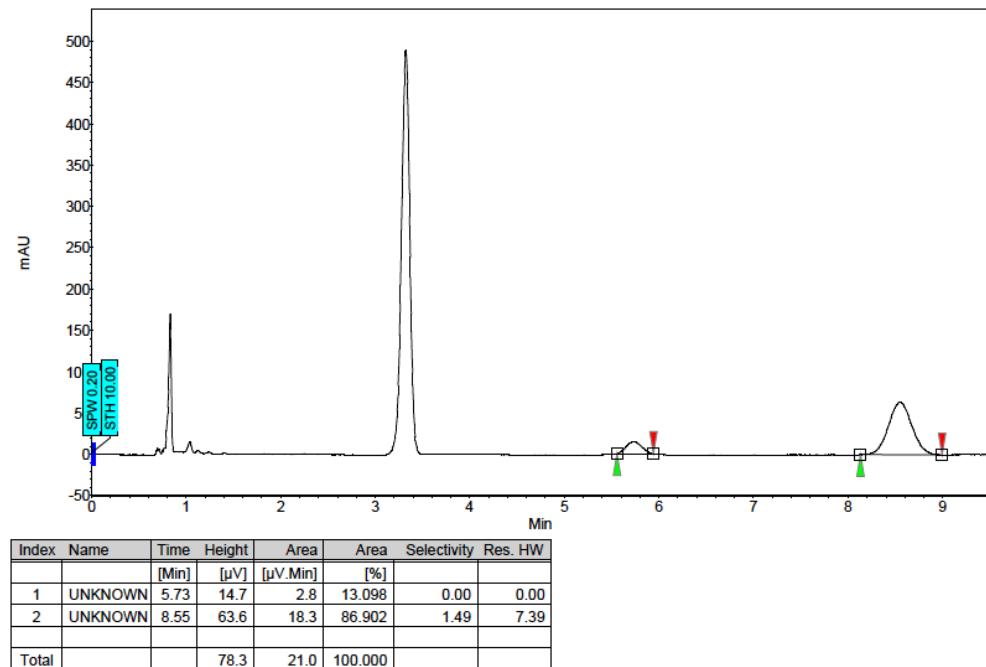
Following General procedure **H** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.55$ min; minor enantiomer $t_R = 5.70$ min].



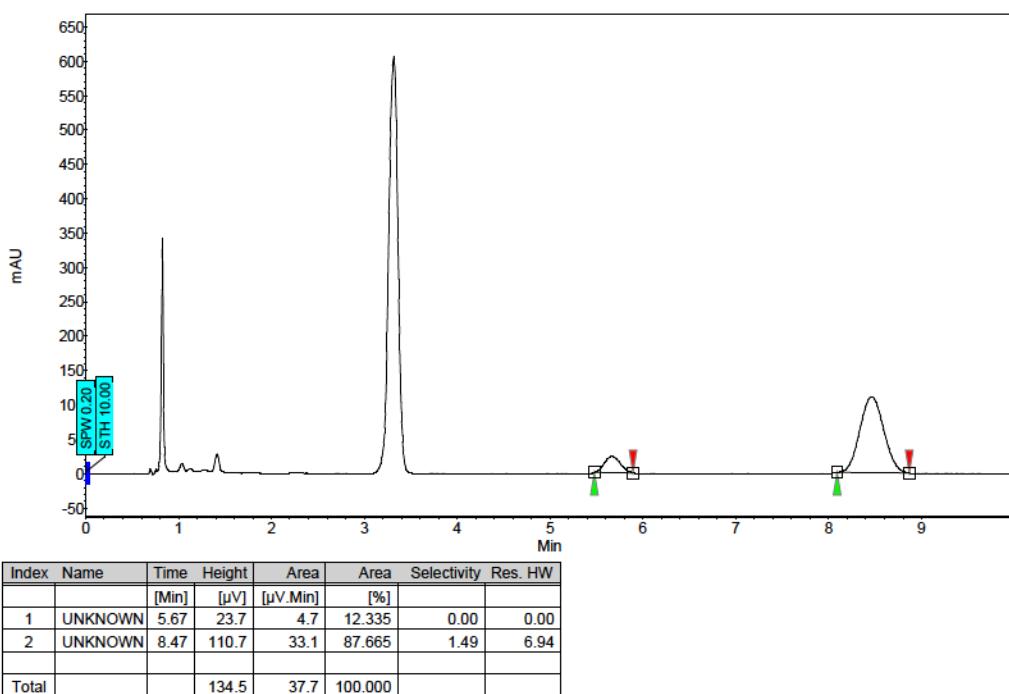
Following General procedure H [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.42$ min; minor enantiomer $t_R = 5.68$ min].



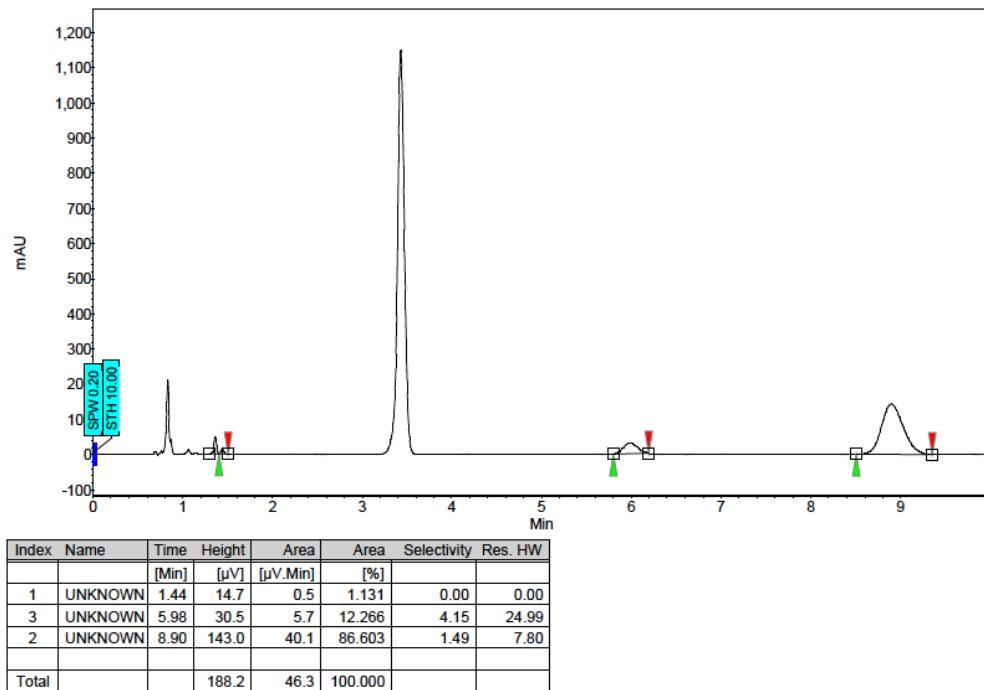
Following General procedure H [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.55$ min; minor enantiomer $t_R = 5.73$ min].



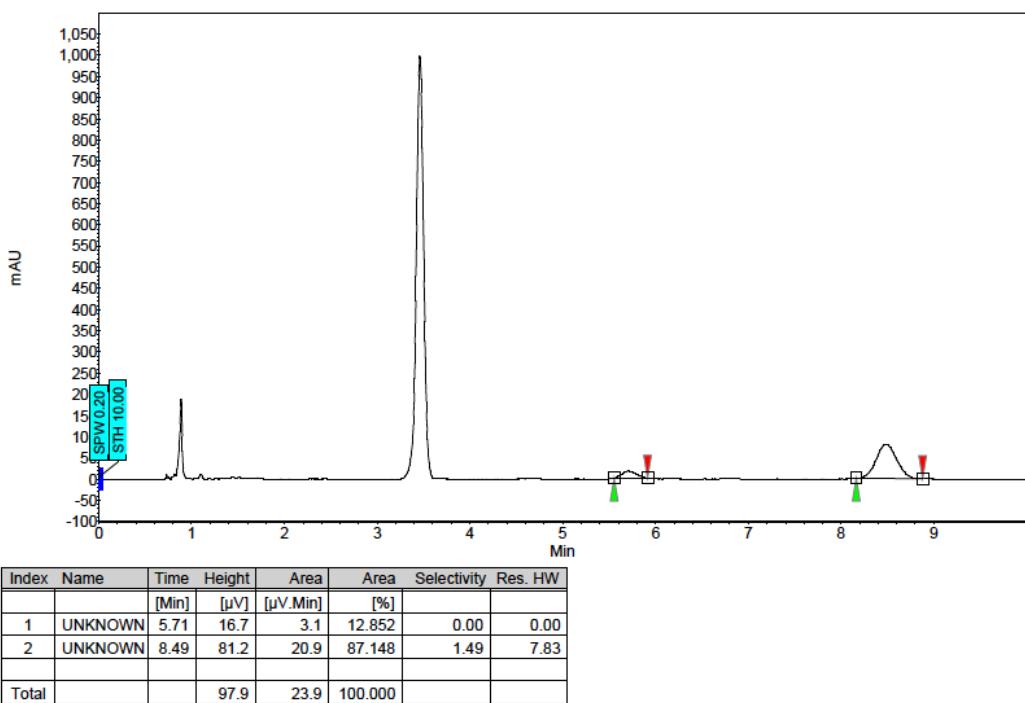
Following General procedure **H** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.47$ min; minor enantiomer $t_R = 5.67$ min].



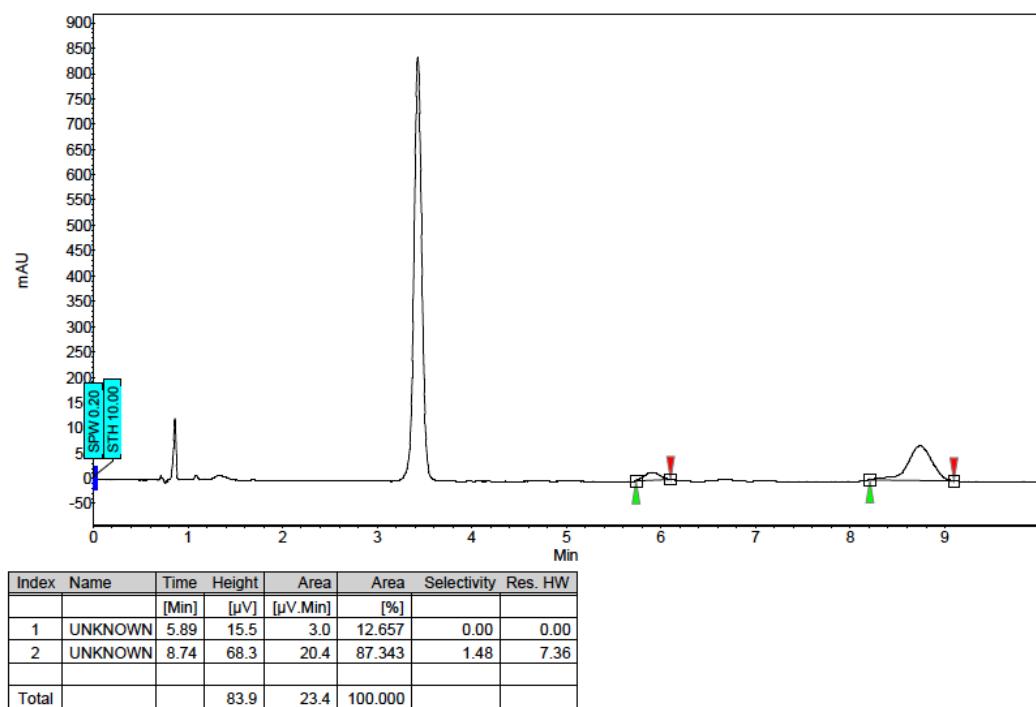
Following General procedure **H** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.90$ min; minor enantiomer $t_R = 5.98$ min].



Following General procedure **H** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.49$ min; minor enantiomer $t_R = 5.71$ min].

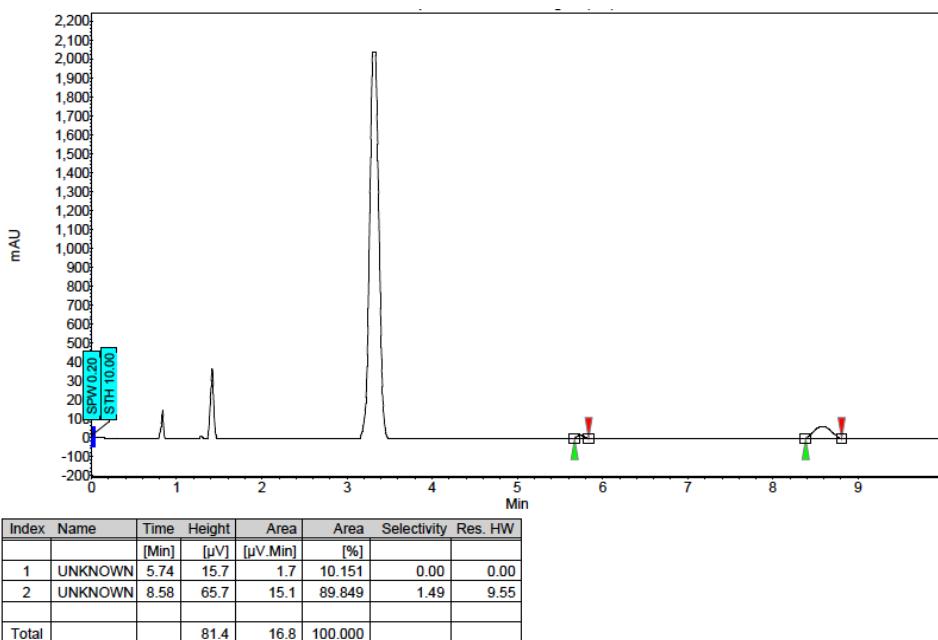


Following General procedure **H** [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.74$ min; minor enantiomer $t_R = 5.89$ min].

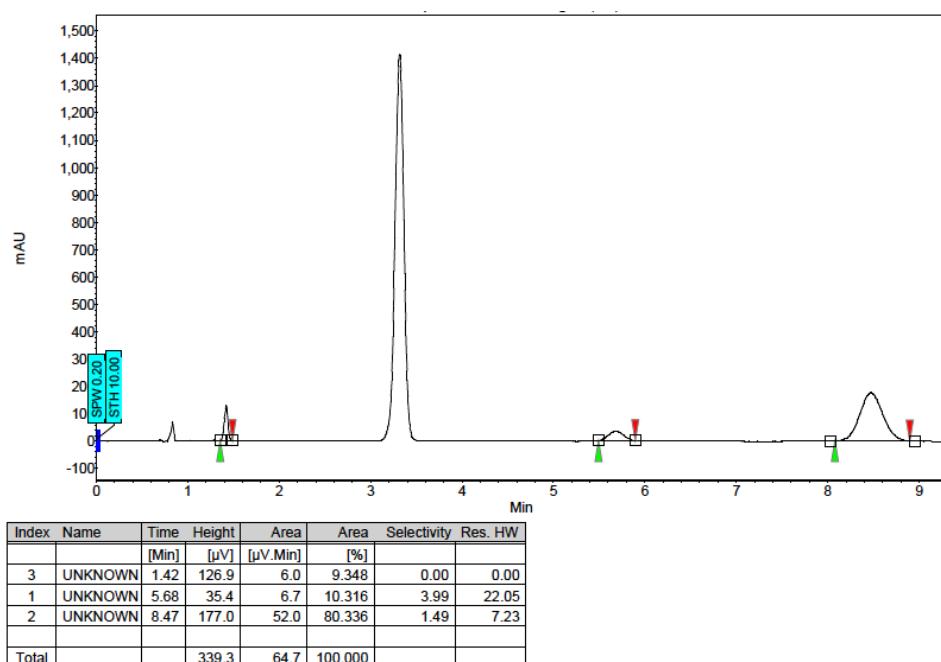


Enantioselective Friedel Crafts alkylations in a continuous flow process

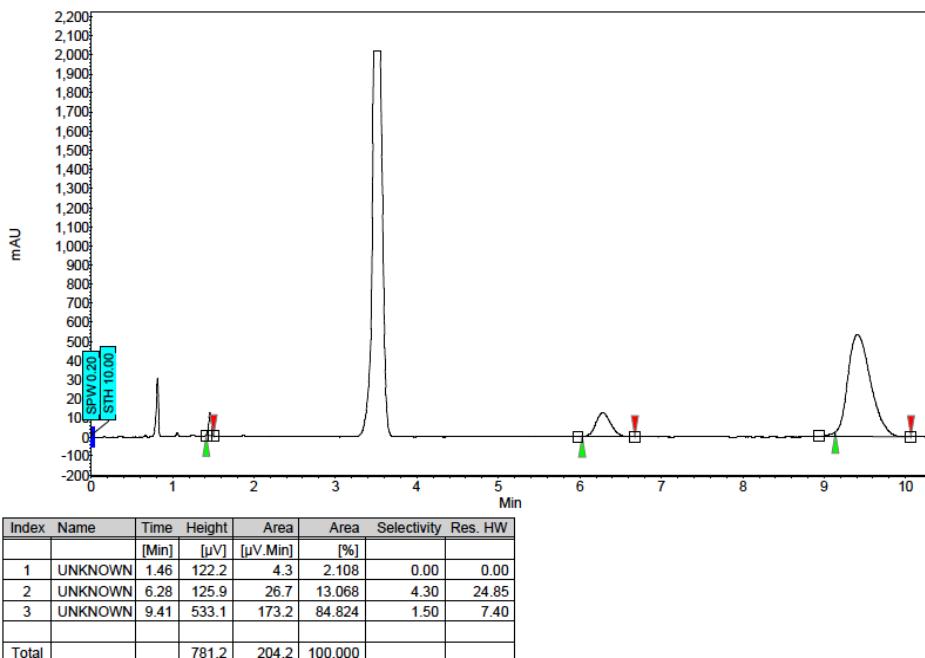
(2a): 0.25 mL/min flow rate, 1.1 g of CS-ct-DNA: Following General procedure I [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.58$ min; minor enantiomer $t_R = 5.74$ min].



(2a): 0.125 mL/min flow rate, 2.2 g of CS-ct-DNA: Following General procedure I [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.47$ min; minor enantiomer $t_R = 5.68$ min].



(2a): 0.125 mL/min flow rate, 4.4 g of CS-ct-DNA: Following General procedure I [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 9.41$ min; minor enantiomer $t_R = 6.28$ min].



(2a): 0.125 mL/min flow rate, 4.4 g of CS-ct-DNA (Large Scale): Following General procedure I [DAICEL AD-H column; 100 bar; flow: 4.0 mL/min; 15% MeOH; $\lambda = 220$ nm, major enantiomer $t_R = 8.51$ min; minor enantiomer $t_R = 5.76$ min].

