

Supplementary Material

Efficient solvent boundary potential for hybrid potential simulations

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Force field determination for acetyl-Coenzyme A and oxaloacetate

As part of this study, we report a force field model for acetyl-Coenzyme A (COA) and oxaloacetate (OAA). The force field was developed so as to be compatible with the CHARMM27 force field for proteins and nucleic acids [1, 2], and with the TIP3P water model [3].

Force field parameters were developed for the keto, enol and enolate forms of COA, and for the dianionic state of oxaloacetate that predominates at neutral pH. Structures were taken from the PDB. Atomic charges were derived from a supermolecule *ab initio* approach and Lennard-Jones parameters were adopted from the CHARMM27 force field [1, 2, 4]. We calculated the *ab initio* energies and geometries of a water molecule interacting with each compound at a few, selected positions. This supermolecule approach is known to give a good balance between solute–water and water–water interactions in the force field [1, 2]. Good agreement was obtained between the *ab initio* and force field data with a rms deviation for the energies of 0.3 kcal/mol. The model also reproduces well the conformational *ab initio* energies and geometries of the individual species, and the interactions between acetyl-CoA, oxaloacetate and protein.

Optimization of the intermolecular force field parameters

We adopted a force field of the CHARMM27 form [1] and used it to simulate complexes between acetyl-CoA, oxaloacetate and citrate synthase in aqueous solution. The intermolecular energy consists of Lennard-Jones and Coulomb terms. As in the development of the CHARMM27 force field, we employ supermolecule, quantum chemical

calculations on complexes between a model compound and a single water molecule to parametrize the intermolecular force field terms. In these calculations, the internal geometries of the model compound and the water molecule are held fixed. The model compounds' geometries were determined by optimizing them at the HF/6-31G(d) level, whereas the water geometry was taken from the TIP3P model [3].

A few water positions were considered for each model compound. Each supermolecule structure was optimized at the HF/6-31G(d) level by varying the interaction distance and a single angle, to find the local minimum for the water position. From the resulting optimal structure, the interaction energy was calculated. No correction for basis set superposition error was made. The *ab initio* interaction energies were scaled by a factor of 1.16 and the *ab initio* interaction distances were reduced by 0.2 Å to compensate for overestimated interaction distances with the Hartree-Fock model (due to neglected electron correlation)[1].

The force field parameters to be optimized were adjusted to reproduce these “corrected” *ab initio* interaction energies and water positions. Initial partial charges were obtained from a Mulliken population analysis of the HF/6-31G(d) wavefunction and were compared with charges of similar molecules for which CHARMM parameters are known. Lennard-Jones parameters were adapted from the CHARMM27 force field for similar chemical groups.

For acetyl-CoA and OAA, we parametrized 3 model compounds: oxaloacetate, methylphosphate, and different tautomeric forms of ethyl thioacetate: keto, enolate, and enol. We optimized the interaction distance and the (corrected) *ab initio* data were then fitted by varying manually the model compound charges. This involved reoptimizing the compound-water distance after each parameter change. In addition to the water-compound interactions, the *ab initio* dipole moments were also used as target data in the partial charge optimization.

We first consider the supermolecule calculations for model compounds interacting with individual water molecules. Detailed results are given in Tables 3–7 for the five species that were considered. The first corresponds to oxaloacetate; the second, third, and fourth correspond to the thioacetate of acetyl-CoA and the last compound corresponds to the phosphate group of acetyl-CoA. Overall, very good agreement was obtained between the *ab initio* and force field data. The rms deviation for the energies was 0.3 kcal/mol, averaged over all fragments, all water positions, and all water orientations. To obtain this agreement, only small adjustments were required to the starting values that were chosen for the atomic charges.

The dipole moments of the compounds are presented in Table 8. It can be seen

that the force field values are systematically larger than the gas phase QC values, which is desirable due to the omission of explicit electronic polarizability from the force field [4]. Angles between the empirical and QC dipole moments are also given in Table 8, showing that they are almost collinear.

Optimization of the intramolecular force field parameters

In a force field treatment, the intramolecular geometry is mainly determined by the minimum energy values of the bond length and bond angle terms, and by the phase and multiplicity of the dihedrals. Initial values for the minimum energy bond lengths, bond angles and torsion angles were taken directly from the *ab initio* structures that were obtained by geometry optimizations at the HF/6-31G(d) level. Starting guesses for the bond and angle force constants, and for the dihedral force constants and phases were taken from the CHARMM27 force field [1] for related, small molecular fragments. The geometrical and dihedral parameters were then optimized by fitting to the *ab initio* structures. At each parameter optimization step, the structure was minimized with the force field model, using a Powell conjugate gradient algorithm and stopping when the rms energy gradient reached 10^{-6} kcal/mol/Å. The quality of the parameters was assessed by the rms coordinate deviation between the force field and *ab initio* structures, and the similarities of the *ab initio* and force field torsional energy profiles (see below). The bond and angle geometries, and the dihedral geometries were then updated manually and a new round of optimization was performed. This procedure was repeated until a satisfactory agreement was achieved, namely a coordinate rms deviation of less than 0.1 Å and energy differences of at most 0.5 kcal/mol.

To characterize molecular flexibility, we examined large, lower-frequency fluctuations, which involve the soft dihedral angles. Fig. 1–3 illustrate the softer dihedral fluctuations in oxaloacetate and acetyl-CoA. For OAA, energy profiles are shown for the dihedral angle that links the carboxyl group to the rest of the molecule, whereas for acetyl-CoA, energy profiles are shown for the dihedral angles that link the acetyl-pantetheine and thioacetic groups (Fig. 2 and 3, respectively). Again, agreement is very good between the force field and the *ab initio* profiles, not only for the energy wells, but also for the barriers between wells. These softer, dihedral degrees of freedom are very important for accurately describing the conformational free energy surfaces of the molecules.

Acetyl-CoA, OAA, and citrate synthase interactions: molecular dynamics simulations

We performed MD simulations of the acetyl-CoA:CS complex for 2 nanoseconds. The protein:acetyl-CoA complex was solvated by a 24 Å water sphere centered on the oxaloacetate binding site whereas the more distant regions were modeled as a dielectric continuum (see Methods).

We computed rms deviations from the crystal structure, averaging over the last 1 ns of the 2 ns simulations (Table 1). By superimposing the oxaloacetate on the crystal structure, we obtained intramolecular OAA deformations of less than 0.30 Å. The equivalent value for acetyl-CoA was 0.47 Å. Superimposing the protein backbone on the crystal structure, we obtained rms deviations of 0.49, 0.69 and 0.56 Å for OAA, COA and the protein backbone, respectively. Figure 4 shows the good structural agreement between the MD simulation of acetyl-CoA:CS and the experimental X-ray structure [5] (PDB entry code 4CSC). The interactions between OAA, acetyl-CoA and the protein are also well-reproduced (Tables 2).

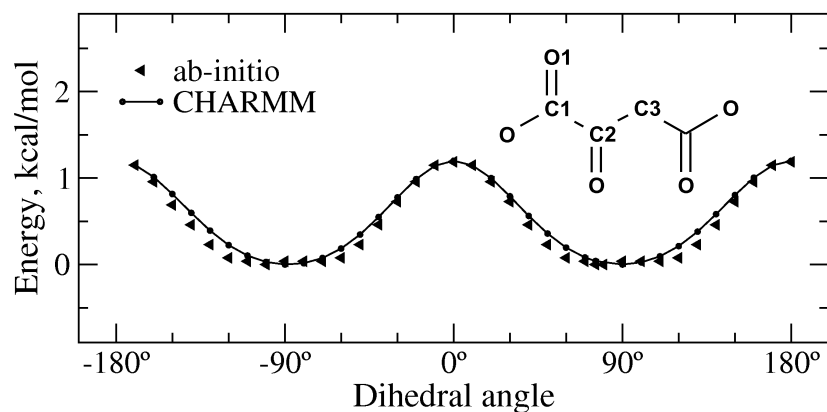


Figure 1: Comparing *ab initio* and force field energies for the dihedral angle of oxaloacetate. The dihedral angle is defined by the atoms labelled (C3-C2-C1-O1). The solid line corresponds to force field energies.

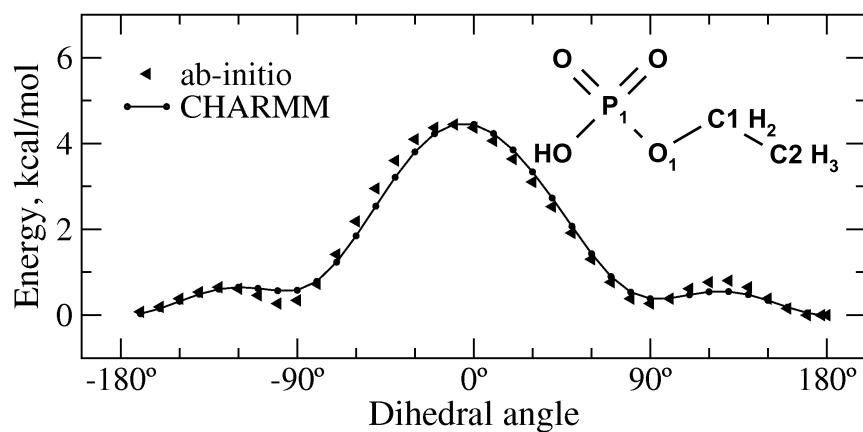


Figure 2: Comparing *ab initio* and force field energies for the dihedral angle of ethyl-phosphate. The dihedral angle is defined by the atoms labelled (C2-C1-O1-P1). The solid line corresponds to force field energies.

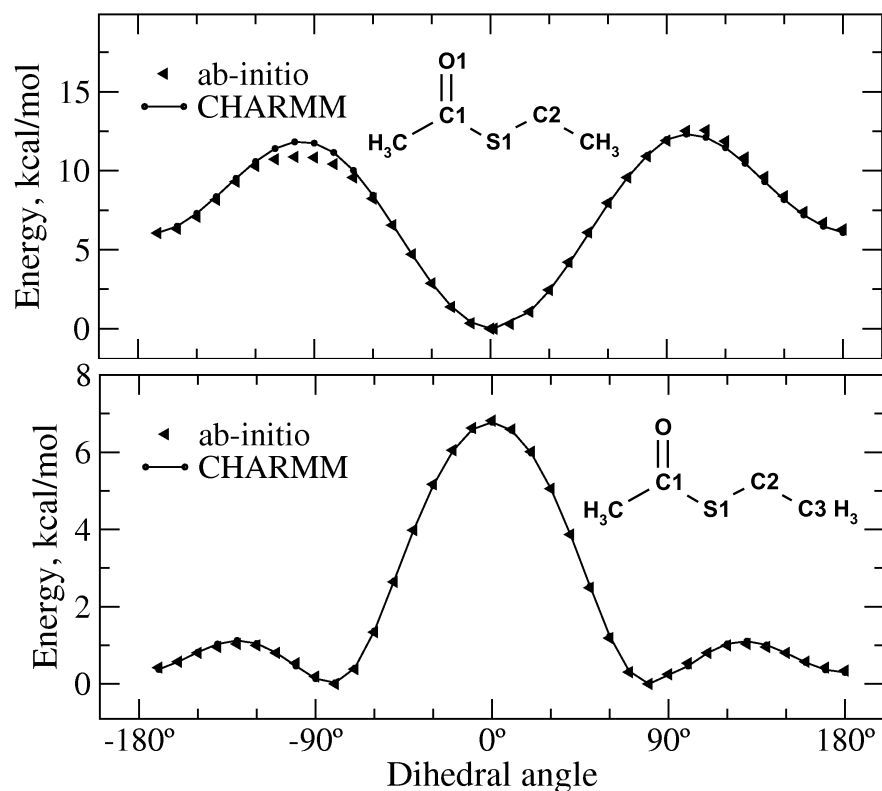


Figure 3: Comparing *ab initio* and force field energies for two dihedral angles that link the acetyl group. The dihedral angles are defined by the atoms labelled in each inset (C2-S1-C1-O1 and C3-C2-S1-C1). In each panel, the solid line corresponds to force field energies.

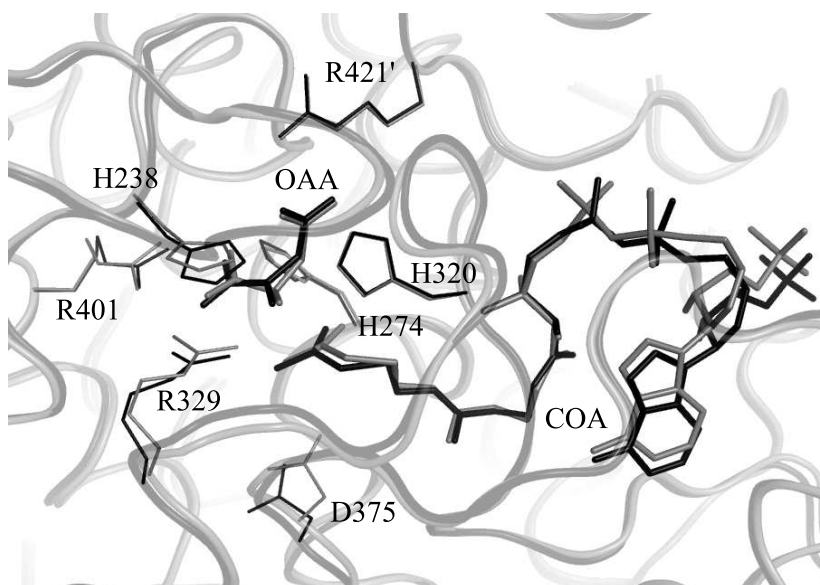


Figure 4: View of the acetyl-coenzyme A binding site from the final nanosecond of the MD trajectory and the experimental X-ray structure (PDB entry 4CSC). COA and OAA are in stick representation, important residues are shown as lines, the experimental structure is gray, and the MD structure is black.

Table 1: Rms deviations (\AA) for the acetyl-CoA:CS complex

atoms	RMSD
OAA ^a	0.29
OAA ^b /backbone	0.49
COA ^a	0.47
COA ^b /backbone	0.69
backbone	0.56

Rms deviations between the MD and crystal structures. ^aAfter superimposing the MD ligand on the ligand in the crystal structure. ^bAfter superimposing the protein backbone on the crystal structure. Non-hydrogen atoms only in each case.

Table 2: Selected distances (\AA) between atoms of acetyl-CoA, OAA, and citrate-synthase

atom pair	MD simulation	^a Xray structure
NH2 _{R401} O1A _{OAA}	2.8	2.8
NH1 _{R401} O1B _{OAA}	2.7	2.8
N ϵ _{R421} O4A _{OAA}	2.8	2.7
NH2 _{R421} O4B _{OAA}	2.7	2.8
N δ 1 _{H238} O4B _{OAA}	2.7	2.8
N δ 1 _{H274} O _{COA}	3.0	2.9
O _{L273} N4P _{COA}	3.1	2.9
N _{G317} O5P _{COA}	2.9	2.8
O _{Y318} N8P _{COA}	3.1	3.0

All atoms (left) are labelled by the amino acid to which they belong.

^aPDB entry 4CSC.

Table 3: Interactions between a probe water and selected oxaloacetate sites

probe site	<i>ab initio</i> /force field results		
	energy (kcal/mol)	distance (Å)	angle (°)
O2C2	-14.04/-14.27	1.80/1.72	0.0
O2C2	-13.16/-13.17	1.80/1.73	60.0
O2C2	-12.09/-11.60	1.80/1.74	120.0
O2C2	-11.78/-10.99	1.80/1.74	180.0
O2C2	-11.90/-11.39	1.80/1.74	240.0
O2C2	-13.09/-13.09	1.80/1.73	300.0

Table 4: Interactions between a probe water and selected sites of the keto form of ethyl thioacetate

probe site	<i>ab initio</i> /force field results		
	energy (kcal/mol)	distance (Å)	angle (°)
OC	-4.21/-4.43	1.81/1.83	0.0
OC	-4.29/-4.41	1.81/1.83	60.0
OC	-4.60/-4.46	1.81/1.83	120.0
OC	-4.74/-4.49	1.81/1.83	180.0
OC	-4.69/-4.50	1.81/1.83	240.0
OC	-4.47/-4.47	1.81/1.83	300.0

Table 5: Interactions between a probe water and selected sites of the enolate form of ethyl thioacetate

probe site	<i>ab initio</i> /force field results		
	energy (kcal/mol)	distance (Å)	angle (°)
OC	-12.43/-12.54	1.90/1.77	0.0
OC	-12.36/-12.44	1.90/1.77	60.0
OC	-12.47/-12.41	1.90/1.77	120.0
OC	-12.51/-12.51	1.90/1.77	180.0
OC	-12.51/-12.51	1.90/1.77	240.0
OC	-12.47/-12.47	1.90/1.77	300.0
CH3C	-8.64/-7.76	2.26/2.07	0.0
CH3C	-11.59/-12.38	2.22/1.99	180.0

Table 6: Interactions between a probe water and selected sites of the enol form of methyl thioacetate

probe site	<i>ab initio</i> /force field results		
	energy (kcal/mol)	distance (Å)	angle (°)
HO	-7.27/-6.86	1.73/1.83	0.0
HO	-6.87/-6.87	1.74/1.83	60.0
HO	-6.29/-6.77	1.77/1.83	120.0
H21CH3	-1.63/-1.72	3.02/2.76	0.0
H21CH3	-2.16/-1.83	2.89/2.74	60.0
H21CH3	-2.12/-1.87	2.93/2.73	120.0
H11CH3	-0.78/-0.78	2.66/2.58	0.0
H11CH3	-0.74/-0.77	2.67/2.59	60.0
H11CH3	-0.65/-0.71	2.69/2.59	120.0
OC	-4.07/-4.06	1.93/1.91	0.0
OC	-4.43/-4.51	1.91/1.90	60.0
OC	-4.43/-4.95	1.91/1.88	120.0
OC	-4.34/-4.70	1.92/1.89	180.0
OC	-4.07/-4.22	1.93/1.90	240.0
OC	-4.16/-4.08	1.91/1.91	300.0

Table 7: Interactions between a probe water and selected monoanionic methylphosphate sites

<i>ab initio</i> /force field results			
probe site	energy (kcal/mol)	distance (Å)	angle (°)
O2C2	-7.68/-8.34	1.80/1.77	0.0
O2C2	-7.30/-7.56	1.80/1.78	60.0
O2C2	-6.88/-6.87	1.80/1.79	120.0
O2C2	-6.96/-6.96	1.80/1.79	180.0
O2C2	-7.46/-7.49	1.80/1.78	240.0
O2C2	-7.76/-8.27	1.80/1.77	300.0

Table 8: *Ab initio* and force field dipole moments

compound	dipole moment (debye)		angle (°)
	<i>abinitio</i>	force field	
oxaloacetate	1.67	2.04	3.1
keto ethyl thioacetate	1.16	1.70	16.3
enolate ethyl thioacetate	4.74	6.35	11.7
enol methyl thioacetate	2.25	1.69	8.7
monoanionic methylphosphate	6.58	8.96	1.1

Force field for acetyl-CoA and oxaloacetate

The force field is presented below in the format appropriate for the CHARMM simulation program [6]. Atom types, atom charges, and chemical bonding information are contained in a “topology file”. Bond lengths and force constants, bond angle values and associated force constants, dihedral parameters and van der Waals parameters are contained in a “parameter file”.

References

- [1] Mackerell, A. et al. (1998) An all-atom empirical potential for molecular modelling and dynamics study of proteins. *J. Phys. Chem. B* 102, 3586–3616.
- [2] Mackerell, A., Wiorkiewicz-Kuczera, J., and Karplus, M. (1995) An all-atom empirical energy force-field for the study of nucleic acids. *J. Am. Chem. Soc.* 117, 11946–11975.
- [3] Jorgensen, W., Chandrasekar, J., Madura, J., Impey, R., and Klein, M. (1983) Comparison of simple potential functions for simulating liquid water. *J. Chem. Phys.* 79, 926–935.
- [4] Foloppe, N., and MacKerell, A. (2000) All-atom empirical force field for nucleic acids: I. Parameter optimization based on small molecule and condensed phase macromolecular target data. *J. Comp. Chem.* 21, 86–104.
- [5] Karpusas, M., Holland, D., and Remington, S. (1991) 1.9-Å structures of ternary complexes of citrate synthase with D- and L-malate: mechanistic implications. *Biochemistry* 30, 6024–31.
- [6] Brooks, B. R. et al. (2009) CHARMM: the biomolecular simulation program. *J. Comp. Chem.* 30, 1545–1614.

Topology file: top_coa.inp

* CHARMM topology for acetyl-CoA and oxaloacetate
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27 1

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RESI COA          -3.00 ! acetyl-CoA, keto form
GROUP
ATOM CH3  CT3     -0.27
ATOM H11  HA       0.09
ATOM H21  HA       0.09 !      0          H12
ATOM H31  HA       0.09 !      \\         |
ATOM C    C        0.44 !      C--S1P--C2P--
ATOM O    O       -0.39 !      /          |
ATOM S1P  S       -0.09 ! H11-CH3        H22
ATOM C2P  CT2     -0.14 !      | \
ATOM H12  HA       0.09 !      H21  H31
ATOM H22  HA       0.09

GROUP
ATOM C3P  CT2     -0.02
ATOM H13  HA       0.09 !      H13          O5P  H16
ATOM H23  HA       0.09 !      |          ||   |
ATOM N4P  NH1     -0.47 ! --C3P--N4P--C5P--C6P--
ATOM H4P  H        0.31 !      |      |          |
GROUP                    !      H23  H4P          H26
ATOM C5P  C        0.51
ATOM O5P  O       -0.51
GROUP
ATOM C6P  CT2     -0.18
ATOM H16  HA       0.09
ATOM H26  HA       0.09

GROUP
ATOM C7P  CT2     -0.02
ATOM H17  HA       0.09 !      H17          O9P
ATOM H27  HA       0.09 !      |          ||
ATOM N8P  NH1     -0.47 ! --C7P--N8P--C9P--
ATOM H8P  H        0.31 !      |      |
GROUP                    !      H27  H8P
ATOM C9P  C        0.51

```

ATOM O9P O -0.51

GROUP

ATOM CAP CT1 0.14

ATOM OAP OH1 -0.66 ! HAP CDP(H3)

ATOM HO2 H 0.43 ! | |

ATOM HAP HA 0.09 ! --CAP--CBP--

GROUP ! | |

ATOM CBP CT1 0.00 ! OAP CEP(H3)

GROUP ! |

ATOM CEP CT3 -0.27 ! HO2

ATOM H1EP HA 0.09

ATOM H2EP HA 0.09

ATOM H3EP HA 0.09

GROUP

ATOM CDP CT3 -0.27

ATOM H1DP HA 0.09

ATOM H2DP HA 0.09

ATOM H3DP HA 0.09

GROUP

ATOM CCP CN9 -0.08

ATOM H1CP HN9 0.09 ! H1CP 01P2

ATOM H2CP HN9 0.09 ! | |

ATOM O6A ON2 -0.62 ! --CCP--O6A--P2--O12--

ATOM P2 P 1.46 ! | |

ATOM O1P2 ON3 -0.83 ! H2CP 02P2

ATOM O2P2 ON3 -0.83

ATOM O12 ON2 -0.63

GROUP

ATOM P P 1.50 ! ADE

ATOM O1P ON3 -0.82 ! H61 H62!

ATOM O2P ON3 -0.82 ! \ /

ATOM O5' ON2 -0.61 ! N6

ATOM C5' CN8B -0.08 ! |

ATOM H5'	HN8	0.09	!	C6
ATOM H5''	HN8	0.09	!	// \
GROUP			!	N1 C5--N7\\
ATOM C4'	CN7	0.16	!	C8-H8
ATOM H4'	HN7	0.09	!	C2 C4--N9/
ATOM O4'	ON6B	-0.50	!	/ \\ / \
ATOM C1'	CN7B	0.16	!	H2 N3 \
ATOM H1'	HN7	0.09	!	\
GROUP			!	\
ATOM N9	NN2	-0.05	!	\
ATOM C5	CN5	0.28	!	01P H5' H4' O4' \
ATOM N7	NN4	-0.71	!	\ / \ \
ATOM C8	CN4	0.34	!	-P-O5'-C5'---C4' C1'
ATOM H8	HN3	0.12	!	\ / \
ATOM N1	NN3A	-0.74	!	02P H5'' C3'--C2' H1'
ATOM C2	CN4	0.50	!	/ \ / \
ATOM H2	HN3	0.13	!	03' H3' 02' H2''
ATOM N3	NN3A	-0.75	!	
ATOM C4	CN5	0.43	!	H2'
ATOM C6	CN2	0.46		
ATOM N6	NN1	-0.77		
ATOM H61	HN1	0.38		
ATOM H62	HN1	0.38		
GROUP				
ATOM C2'	CN7B	0.14		
ATOM H2''	HN7	0.09		
ATOM O2'	ON5	-0.66		
ATOM H2'	HN5	0.43		
GROUP			!	3'terminal phosphate
ATOM C3'	CN7	0.01	!	/
ATOM H3'	HN7	0.09	!	03'
ATOM P3	P	1.50	!	
ATOM 01P3	ON3	-0.82	!	01P3=P3=02P3
ATOM 02P3	ON3	-0.82	!	
ATOM 03'	ON2	-0.62	!	03T-H3T

ATOM O3T ON4 -0.68 !
ATOM H3T HN4 0.34

BOND CH3 H11 CH3 H21 CH3 H31
BOND CH3 C C O S1P C
BOND C2P S1P C2P H12 C2P H22
BOND C2P C3P
BOND C3P H13 C3P H23 C3P N4P
BOND N4P H4P N4P C5P
BOND C5P O5P C5P C6P
BOND C6P H16 C6P H26
BOND C6P C7P
BOND C7P H17 C7P H27 C7P N8P
BOND N8P H8P N8P C9P
BOND C9P O9P
BOND C9P CAP
BOND CAP OAP CAP HAP CAP CBP OAP HO2
BOND CBP CDP CBP CEP
BOND CDP H1DP CDP H2DP CDP H3DP
BOND CEP H1EP CEP H2EP CEP H3EP
BOND CBP CCP
BOND P O12 O12 P2
BOND P2 O1P2 P2 O2P2 P2 O6A
BOND O6A CCP CCP H1CP CCP H2CP

! ADE

BOND P O1P P O2P P O5'
BOND O5' C5' C5' C4' C4' O4' C4' C3' O4' C1'
BOND C1' N9 C1' C2' N9 C4 N9 C8 C4 N3
BOND C2 N1 C6 N6
BOND N6 H61 N6 H62 C6 C5 C5 N7
BOND C2' C3' C2' O2' O2' H2' C3' O3'
BOND C1' H1' C2' H2'' C3' H3' C4' H4' C5' H5'
BOND C5' H5'' C8 H8 C2 H2
DOUBLE N1 C6 C2 N3 C4 C5 N7 C8

! 3PH0

BOND O3' P3 P3 O1P3 P3 O2P3 P3 O3T O3T H3T

IMPR C S1P CH3 O
IMPR N4P C5P C3P H4P
IMPR C5P C6P N4P O5P
IMPR N8P C9P C7P H8P
IMPR C9P CAP N8P O9P
IMPR N6 C6 H61 H62
IMPR C6 N1 C5 N6

PRES ELT -1.00 ! convert keto form of acetyl-CoA into enolate

GROUP

ATOM CH3 CE2 -0.83
ATOM H11 HE2 0.09
ATOM H21 HE2 0.09
ATOM C C 0.30
ATOM O O -0.60
ATOM S1P S -0.09
ATOM C2P CT2 -0.14
ATOM H12 HA 0.09
ATOM H22 HA 0.09
DELE ATOM H31

PRES ETE 0.00 ! converts keto form of acetyl-CoA into enol

GROUP

ATOM CH3 CE2 -0.55
ATOM H11 HE2 0.22
ATOM H21 HE2 0.22 ! H--O H12 H13
ATOM C C 0.28 ! \\
ATOM O OH1 -0.53 ! C--S1P--C2P--C3P-H23
ATOM H H 0.41 ! /
ATOM S1P S -0.09 ! H11-CH3 H22 H33
ATOM C2P CT2 -0.14 ! |

```

ATOM H12  HA      0.09 !      H21
ATOM H22  HA      0.09 !
DELE  ATOM  H31
BOND  H   O

RESI OAA      -2.00 ! oxaloacetate
GROUP
ATOM  C4  CC      0.62 !  01A      H3A      04A(-)
ATOM  04A OC     -0.76 !    \ \      |      /
ATOM  04B OC     -0.76 !      C1--C2--C3--C4
ATOM  C3  CT2    -0.28 !    /    ||  |    \ \
ATOM  H3A HA      0.09 ! 01B(-)  02 H3B      04B
ATOM  H3B HA      0.09 !
GROUP
ATOM  C2  CC      0.45
ATOM  O2  O      -0.55
ATOM  C1  CC      0.62
ATOM  01A OC     -0.76
ATOM  01B OC     -0.76
BOND   C4  04A    C4  04B    C4   C3
BOND   C3  H3A    C3  H3B    C3   C2
BOND   C2   O2    C2   C1
BOND   C1  01A    C1  01B
IMPR  C1 C2 01A 01B
IMPR  C4 C3 04A 04B
IMPR  C2 C3 C1  O2

END

```

Parameter file: par.coa.prm

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* CHARMM parameter file for acetyl-CoA and oxaloacetate
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BONDS

S	C	198.00	1.8180
CT1	CN9	222.50	1.5380
C	CE2	250.00	1.3550
C	OH1	230.00	1.4000
CC	CC	493.60	1.5070
CC	CD	493.60	1.5070

ANGLES

CT2	S	C	34.00	95.00		
S	C	O	80.00	121.00		
S	C	CT3	50.00	114.00		
C	CT1	OH1	50.00	110.50		
CN9	CT1	CT1	53.35	111.00	8.00	2.5610
CT3	CT1	CN9	53.35	114.00	8.00	2.5610
CT1	CN9	HN9	33.43	110.10	22.53	2.1790
CT1	CN9	ON2	45.00	111.50		
C	CE2	HE2	55.50	120.50		
O	C	CE2	80.00	133.50		
S	C	CE2	50.00	114.00		
S	C	OH1	58.00	116.20		
C	OH1	H	65.00	108.00		
OH1	C	CE2	45.20	120.00		
CC	CT2	CC	69.10	120.60		
CT2	CC	CC	57.80	113.40		
CC	CC	OC	47.40	116.60		
O	CC	CC	84.30	117.00		
CC	CD	OH1	47.40	116.60		
CC	CD	OB	47.40	116.60		
O	CC	CD	84.30	117.00		
CT2	CC	CD	57.80	113.40		

DIHEDRALS

C	S	CT2	HA	0.0	3	0.0
HA	CT3	C	S	0.04	3	0.0

C	S	CT2	CT2	0.24	1	180.0
C	S	CT2	CT2	0.37	3	0.0
C	S	CT2	CT3	0.24	1	180.0
C	S	CT2	CT3	0.37	3	0.0
O	C	S	CT2	3.73	1	180.0
O	C	S	CT2	2.07	2	180.0
CT3	C	S	CT2	2.63	1	180.0
CT3	C	S	CT2	2.56	2	180.0
NH1	C	CT2	CT2	2.98	1	0.0
CT2	CT2	NH1	C	1.44	1	180.0
CT2	CT2	NH1	C	0.30	3	180.0
NH1	C	CT1	OH1	0.00	3	0.0
O	C	CT1	OH1	0.00	3	0.0
NH1	C	CT1	HA	0.00	3	0.0
O	C	CT1	HA	0.00	3	0.0
CT1	CT1	CN9	HN9	0.04	3	0.0
CT3	CT1	CN9	HN9	0.04	3	0.0
CT3	CT1	CN9	ON2	0.19	3	0.0
CT1	CT1	CN9	ON2	0.16	1	180.0
CT1	CT1	CN9	ON2	0.39	2	0.0
P	ON2	CN9	CT1	1.55	1	0.0
P	ON2	CN9	CT1	0.55	2	0.0
P	ON2	CN9	CT1	0.20	3	0.0
HE2	CE2	C	S	5.20	2	180.0
HE2	CE2	C	O	5.20	2	180.0
HE2	CE2	C	OH1	5.20	2	180.0
CE2	C	S	CT2	2.46	1	180.0
CE2	C	S	CT2	1.43	2	180.0
CE2	C	S	CT2	0.42	3	180.0
OH1	C	S	CT2	1.77	1	180.0
OH1	C	S	CT2	0.73	3	0.0
H	OH1	C	S	0.02	1	0.0
H	OH1	C	S	0.31	2	180.0
H	OH1	C	CE2	0.57	1	180.0
H	OH1	C	CE2	0.85	2	180.0
OC	CC	CC	CT2	0.20	2	180.0

OC	CC	CC	O	0.00	2	180.0
OB	CD	CC	CT2	0.20	2	180.0
OH1	CD	CC	CT2	0.20	2	180.0
OH1	CD	CC	O	0.00	2	180.0
OB	CD	CC	O	0.00	2	180.0

IMPROPER

C	S	CE2	OH1	120.00	0	0.0
CC	CT2	CC	O	50.00	0	0.0
CD	CC	OB	OH1	50.00	0	0.0
CC	CT2	CD	O	50.00	0	0.0