Identifying the sarco(endo)plasmic reticulum Ca²⁺ ATPase (SERCA) as a potential target for hypericin – A theoretical study

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References

Fig. S1: Sequence alignment of human SERCA1A and rabbit SERCA1A with ClustalW2.

"*": identical residues; ":": conserved constitution; and ".": semiconserved residues.

SERCA1A_human	MEAAHAKTTEECLAYFGVSETTGLTPDQVKRNLEKYGLNELPAEEGKTLWELVIEQFEDL	60
SERCA1A_rabbit	${\tt MEAAHSKSTEECLAYFGVSETTGLTPDQVKRHLEKYGHNELPAEEGKSLWELVIEQFEDL}$	60
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SERCA1A human	LVRILLLAACISFVLAWFEEGEETITAFVEPFVILLILIANAIVGVWOERNAENAIEALK	120
SERCA1A rabbit	LVRTLLLAACTSEVLAWFEEGEETTTAEVEPEVILLILLANAIVGVWOERNAENAIEALK	120
	***************************************	100
SERCA1A_human	EYEPEMGKVYRADRKSVQRIKARDIVPGDIVEVAVGDKVPADIRILAIKSTTLRVDQSIL	180
SERCA1A_rabbit	EYEPEMGKVYRADRKSVQRIKARDIVPGDIVEVAVGDKVPADIRILSIKSTTLRVDQSIL ************************************	180
SERCA1A human	TGESVSVIKHTEDVEDERAVNODKKNMI.ESGTNIAAGKAI.GIVATTGVGTEIGKIRDOMA	240
SERCAIA rabbit	TGESVSVIKHTEPVPDPRAVNODKKNMLFSGTNIAAGKALGIVATTGVSTEIGKIRDOMA	240
	***************************************	210
SERCA1A_human	ATEQDKTPLQQKLDEFGEQLSKVISLICVAVWLINIGHFNDPVHGGSWFRGAIYYFKIAV	300
SERCA1A_rabbit	$\verb ATEQDKTPLQQKLDEFGEQLSKVISLICVAVWLINIGHFNDPVHGGSWIRGAIYYFKIAV $	300

SERCA1A_human	ALAVAAIPEGLPAVITTCLALGTRRMAKKNAIVRSLPSVETLGCTSVICSDKTGTLTTNQ	360
SERCA1A_rabbit	ALAVAAIPEGLPAVITTCLALGTRRMAKKNAIVRSLPSVETLGCTSVICSDKTGTLTTNQ	360

SERCA1A_human	MSVCKMFIIDKVDGDICLLNEFSITGSTYAPEGEVLKNDKPVRPGQYDGLVELATICALC	420
SERCA1A_rabbit	${\tt MSVCKMFIIDKVDGDFCSLNEFSITGSTYAPEGEVLKNDKPIRSGQFDGLVELATICALC}$	420

SERCA1A human	NDSSLDFNEAKGVYEKVGEATETALTTLVEKMNVFNTDVRSLSKVERANACNSVIRQLMK	480
SERCA1A_rabbit	NDSSLDFNETKGVYEKVGEATETALTTLVEKMNVFNTEVRNLSKVERANACNSVIRQLMK	480

SERCA1A human	KEFTLEFSRDRKSMSVYCSPAKSSRAAVGNKMFVKGAPEGVIDRCNYVRVGTTRVPLTGP	540
SERCA1A_rabbit	$\tt KEFTLEFSRDRKSMSVYCSPAKSSRAAVGNKMFVKGAPEGVIDRCNYVRVGTTRVPMTGP$	540

SERCA1A_human	VKEKIMAVIKEWGTGRDTLRCLALATRDTPPKREEMVLDDSARFLEYETDLTFVGVVGML	600
SERCA1A_rabbit	$\tt VKEKILSVIKEWGTGRDTLRCLALATRDTPPKREEMVLDDSSRFMEYETDLTFVGVVGML$	600
	*****: <u>*</u> ******************************	
SERCA1A human	DPPRKEVTGSIQLCRDAGIRVIMITGDNKGTAIAICRRIGIFGENEEVADRAYTGREFDD	660
SERCA1A_rabbit	${\tt DPPRKEVMGSIQLCRDAGIRVIMITGDNKGTAIAICRRIGIFGENEEVADRAYTGREFDD}$	660
	****** ********************************	
SERCA1A human	LPLAEQREACRRACCFARVEPSHKSKIVEYLQSYDEITAMTGDGVNDAPALKKAEIGIAM	720
SERCA1A_rabbit	LPLAEQREACRRACCFARVEPSHKSKIVEYLQSYDEITAMTGDGVNDAPALKKAEIGIAM	720
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SERCA1A human	GSGTAVAKTASEMVLADDNFSTIVAAVEEGRAIYNNMKOFIRYLISSNVGEVVCIFLTAA	780
SERCA1A rabbit	GSGTAVAKTASEMVLADDNFSTIVAAVEEGRAIYNNMKQFIRYLISSNVGEVVCIFLTAA	780
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SERCA1A_human	LGLPEALIPVQLLWVNLVTDGLPATALGFNPPDLDIMDRPPRSPKEPLISGWLFFRYMAI	840
SERCA1A_rabbit	$\verb"LGLPEALIPVQLLWVNLVTDGLPATALGFNPPDLDIMDRPPRSPKEPLISGWLFFRYMAI"$	840

SERCA1A human	GGYVGAATVGAAAWWFLYAEDGPHVNYSQLTHFMQCTEDNTHFEGIDCEVFEAPEPMTMA	900
SERCA1A_rabbit	GGYVGAATVGAAAWWFMYAEDGPGVTYHQLTHFMQCTEDHPHFEGLDCEIFEAPEPMTMA	900

SERCA1A_human	LSVLVTIEMCNALNSLSENQSLLRMPPWVNIWLLGSICLSMSLHFLILYVDPLPMIFKLR	960
SERCA1A_rabbit	$\verb"LSVLVTIEMCNALNSLSENQSLMRMPPWVNIWLLGSICLSMSLHFLILYVDPLPMIFKLK"$	960

SERCA1A_human ALDLTQWLMVLKISLPVIGLDEILKFVARNYLEG 994 SERCA1A_rabbit ALDLTQWLMVLKISLPVIGLDEILKFIARNYLEG 994

Section S2: Membrane embedding in YASARA

First, a cell is created around the protein. The cell was extended by 15 Å on each side of the protein in the x and y directions (the membrane plane), giving a box that is 30 Å larger than the protein. The cell was then extended by 10 Å on each side of the protein in the z direction (normal to the membrane plane), giving a water extension of 20 Å in total in this direction.

The protein was initially searched for exposed hydrophobic side-chains of amino acids in transmembrane helices and strands. After the transmembrane region in the protein was identified, a previously equilibrated POPE membrane fragment was loaded and replicated to reach the size of the simulation cell. The membrane of required size was constructed in the absence of the protein. The protein was then loaded and lipids deleted in order to fit the protein within the membrane. To ensure that not too many lipids were deleted, which would be the case if a single overlap was enough to delete a lipid, the protein was first compressed and lipids that overlapped with the compressed protein were deleted. The protein was then grown back to its original size during an energy minimization of the lipids to fill the membrane pore and generate a system where lipids smoothly cover the protein. The entire system was then energy minimized.

Next, the cell was filled with water of density 0.998 g/ml and a NaCl concentration of 0.9 % and pH 7.4. Water molecules inside the membrane region were deleted. The system was then minimized and equilibrated in a 250 ps MD simulation, during which waters were prevented from entering the membrane. A 38 ns MD simulation followed in which a time step of 2×1.25 fs was used. During the simulation the temperature was set to 298 K. The AMBER03 force field was applied throughout with a force cutoff of 7.86 Å. Periodic boundary conditions were applied and the PME algorithm was used to treat long-range electrostatics.

Section. S3: Energy minimization in YASARA.

A steepest descent minimization is first performed to remove bumps and correct the covalent geometry, followed by a simulated annealing minimization (time step 2 fs, atom velocities scaled down by 0.9 every 10th step) until convergence is reached, i.e. the energy improved by less than the convergence parameter per atom during 200 steps.

In a system containing solvent, the solvent alone is first subject to minimization. The convergence parameter for this minimization is set to 0.1 kJ/mol. The total system is then minimized, using a convergence parameter set to 0.05 kJ/mol.

Fig. S4: Sequence alignment of SERCA2A (human), SERCA2B (human), and SERCA1A (rabbit) with ClustalW2.

"*": identical residues; ":": conserved constitution; and ".": semiconserved residues.

SERCA2A SERCA2B SERCA1A	MENAHTKTVEEVLGHFGVNESTGLSLEQVKKLKERWGSNELPAEEGKTLLELVIEQFEDL MENAHTKTVEEVLGHFGVNESTGLSLEQVKKLKERWGSNELPAEEGKTLLELVIEQFEDL MEAAHSKSTEECLAYFGVSETTGLTPDQVKRHLEKYGHNELPAEEGKSLWELVIEQFEDL ** **:*:.** *.:***: :***: *::*	60 60 60
SERCA2A SERCA2B SERCA1A	LVRILLLAACISFVLAWFEEGEETITAFVEPFVILLILVANAIVGVWQERNAENAIEALK LVRILLLAACISFVLAWFEEGEETITAFVEPFVILLILVANAIVGVWQERNAENAIEALK LVRILLLAACISFVLAWFEEGEETITAFVEPFVILLILIANAIVGVWQERNAENAIEALK ************************************	120 120 120
SERCA2A SERCA2B SERCA1A	EYEPEMGKVYRQDRKSVQRIKAKDIVPGDIVEIAVGDKVPADIRLTSIKSTTLRVDQSIL EYEPEMGKVYRQDRKSVQRIKAKDIVPGDIVEIAVGDKVPADIRLTSIKSTTLRVDQSIL EYEPEMGKVYRADRKSVQRIKARDIVPGDIVEVAVGDKVPADIRILSIKSTTLRVDQSIL ********** **********	180 180 180
SERCA2A SERCA2B SERCA1A	TGESVSVIKHTDPVPDPRAVNQDKKNMLFSGTNIAAGKAMGVVVATGVNTEIGKIRDEMV TGESVSVIKHTDPVPDPRAVNQDKKNMLFSGTNIAAGKAMGVVVATGVNTEIGKIRDEMV TGESVSVIKHTEPVPDPRAVNQDKKNMLFSGTNIAAGKALGIVATTGVSTEIGKIRDQMA **********	240 240 240
SERCA2A SERCA2B SERCA1A	ATEQERTPLQQKLDEFGEQLSKVISLICIAVWIINIGHFNDPVHGGSWIRGAIYYFKIAV ATEQERTPLQQKLDEFGEQLSKVISLICIAVWIINIGHFNDPVHGGSWIRGAIYYFKIAV ATEQDKTPLQQKLDEFGEQLSKVISLICVAVWLINIGHFNDPVHGGSWIRGAIYYFKIAV ****::*******************************	300 300 300
SERCA2A SERCA2B SERCA1A	ALAVAAIPEGLPAVITTCLALGTRRMAKKNAIVRSLPSVETLGCTSVICSDKTGTLTTNQ ALAVAAIPEGLPAVITTCLALGTRRMAKKNAIVRSLPSVETLGCTSVICSDKTGTLTTNQ ALAVAAIPEGLPAVITTCLALGTRRMAKKNAIVRSLPSVETLGCTSVICSDKTGTLTTNQ *******	360 360 360
SERCA2A SERCA2B SERCA1A	MSVCRMFILDRVEGDTCSLNEFTITGSTYAPIGEVHKDDKPVNCHQYDGLVELATICALC MSVCRMFILDRVEGDTCSLNEFTITGSTYAPIGEVHKDDKPVNCHQYDGLVELATICALC MSVCKMFIIDKVDGDFCSLNEFSITGSTYAPEGEVLKNDKPIRSGQFDGLVELATICALC ****:**::::::::::::::::::::::::::::::	420 420 420
SERCA2A SERCA2B SERCA1A	NDSALDYNEAKGVYEKVGEATETALTCLVEKMNVFDTELKGLSKIERANACNSVIKQLMK NDSALDYNEAKGVYEKVGEATETALTCLVEKMNVFDTELKGLSKIERANACNSVIKQLMK NDSSLDFNETKGVYEKVGEATETALTTLVEKMNVFNTEVRNLSKVERANACNSVIRQLMK ***:**:**:***	480 480 480
SERCA2A SERCA2B SERCA1A	KEFTLEFSRDRKSMSVYCTPNKPSRTSMS-KMFVKGAPEGVIDRCTHIRVGSTKVPMTSG KEFTLEFSRDRKSMSVYCTPNKPSRTSMS-KMFVKGAPEGVIDRCTHIRVGSTKVPMTSG KEFTLEFSRDRKSMSVYCSPAKSSRAAVGNKMFVKGAPEGVIDRCNYVRVGTTRVPMTGP ************************************	539 539 540
SERCA2A SERCA2B SERCA1A	VKQKIMSVIREWGSGSDTLRCLALATHDNPLRREEMHLEDSANFIKYETNLTFVGCVGML VKQKIMSVIREWGSGSDTLRCLALATHDNPLRREEMHLEDSANFIKYETNLTFVGCVGML VKEKILSVIKEWGTGRDTLRCLALATRDTPPKREEMVLDDSSRFMEYETDLTFVGVVGML **:**:**:**:**:* *********************	599 599 600
SERCA2A SERCA2B SERCA1A	DPPRIEVASSVKLCRQAGIRVIMITGDNKGTAVAICRRIGIFGQDEDVTSKAFTGREFDE DPPRIEVASSVKLCRQAGIRVIMITGDNKGTAVAICRRIGIFGQDEDVTSKAFTGREFDE DPPRKEVMGSIQLCRDAGIRVIMITGDNKGTAIAICRRIGIFGENEEVADRAYTGREFDD **** ** .*::***:***********************	659 659 660
SERCA2A SERCA2B SERCA1A	LNPSAQRDACLNARCFARVEPSHKSKIVEFLQSFDEITAMTGDGVNDAPALKKAEIGIAM LNPSAQRDACLNARCFARVEPSHKSKIVEFLQSFDEITAMTGDGVNDAPALKKAEIGIAM LPLAEQREACRRACCFARVEPSHKSKIVEYLQSYDEITAMTGDGVNDAPALKKAEIGIAM * : **:** .* ******	719 719 720

SERCA2A SERCA2B SERCA1A	GSGTAVAKTASEMVLADDNFSTIVAAVEEGRAIYNNMKQFIRYLISSNVGEVVCIFLTAA GSGTAVAKTASEMVLADDNFSTIVAAVEEGRAIYNNMKQFIRYLISSNVGEVVCIFLTAA GSGTAVAKTASEMVLADDNFSTIVAAVEEGRAIYNNMKQFIRYLISSNVGEVVCIFLTAA ***********************************	779 779 780
SERCA2A SERCA2B SERCA1A	LGFPEALIPVQLLWVNLVTDGLPATALGFNPPDLDIMNKPPRNPKEPLISGWLFFRYLAI LGFPEALIPVQLLWVNLVTDGLPATALGFNPPDLDIMNKPPRNPKEPLISGWLFFRYLAI LGLPEALIPVQLLWVNLVTDGLPATALGFNPPDLDIMDRPPRSPKEPLISGWLFFRYMAI **:**********************************	839 839 840
SERCA2A SERCA2B SERCA1A	GCYVGAATVGAAAWWFIAADGGPRVSFYQLSHFLQCKEDNPDFEGVDCAIFESPYPMTMA GCYVGAATVGAAAWWFIAADGGPRVSFYQLSHFLQCKEDNPDFEGVDCAIFESPYPMTMA GGYVGAATVGAAAWWFMYAEDGPGVTYHQLTHFMQCTEDHPHFEGLDCEIFEAPEPMTMA * ***********************************	899 899 900
SERCA2A SERCA2B SERCA1A	LSVLVTIEMCNALNSLSENQSLLRMPPWENIWLVGSICLSMSLHFLILYVEPLPLIFQIT LSVLVTIEMCNALNSLSENQSLLRMPPWENIWLVGSICLSMSLHFLILYVEPLPLIFQIT LSVLVTIEMCNALNSLSENQSLMRMPPWVNIWLLGSICLSMSLHFLILYVDPLPMIFKLK ***********************************	959 959 960
SERCA2A SERCA2B SERCA1A	PLNVTQWLMVLKISLPVILMDETLKFVARNYLEPPLNVTQWLMVLKISLPVILMDETLKFVARNYLEPGKECVQPATKSCSFSACTDGISWPFV ALDLTQWLMVLKISLPVIGLDEILKFIARNYLEG	993 1019 994
SERCA2A SERCA2B SERCA1A	-AILE 997 LLIMPLVIWVYSTDTNFSDMFWS 1042	

Fig. S5: The transmembrane region of SERCA1A highlighted in cyan. Amino acids included in the transmembrane region:

SERCA1A: 48-79, 83-106, 253-277, 291-314, 763-805, 832-862, 892-915, 933-987. SERCA2A: 48-79, 83-106, 256-279, 290-314, 762-805, 833-861, 891-914, 932-984.



Fig. S6: Time evolution of the Cα RMSD of the transmembrane region of SERCA1A and SERCA2A during the MD simulations (800 snapshots) in the membranes, after alignment of these atoms (cf. S5).



Fig. S7: Superposed backbones of (A) SERCA1A and (B) SERCA2A before (red) and after (blue) membrane embedding, MD simulation and energy minimization. Magnifications of (C and D) the TG binding pocket and (E and F) the BHQ binding pocket.



Fig. S8: Ligand interaction diagrams for ligands complexed with SERCA1A and SERCA2A.



S8.1. (A) TG in the TG binding pocket in SERCA1A (A) with and (B) without lipid in pocket during docking. Hypericin in the TG binding pocket in SERCA1A (C) with and (D) without lipid in pocket during docking.





S8.2. (A) BHQ and (B) hypericin in the BHQ binding pocket in SERCA1A.

S8.3: Hypericin in site 2 in (A) SERCA1A and (B) SERCA2A. Hypericin in the cytosolic region of (C) SERCA1A and (D) SERCA2A.

B







S8.4: TG in the TG binding pocket in SERCA2A (A) with and (B) without lipid in pocket during docking. Hypericin in the TG binding pocket in SERCA2A (C) with and (D) without lipid in pocket during docking.





Fig. S9: Time evolution of the ligand heavy atom RMSD during the MD simulations (800 snapshots per simulation) after superposition of the protein transmembrane Cα's of SERCA1A.

- (A) TG in the TG pocket with a lipid present
- (B) TG in the TG pocket without a lipid present
- (C) Hypericin in the TG pocket with a lipid present
- (D) Hypericin in the TG pocket without a lipid present
- (E) BHQ in the BHQ pocket
- (F) Hypericin in the BHQ pocket
- (G) Hypericin in Site 2



Fig. S10: Time evolution of the ligand heavy atom RMSD during the MD simulations (800 snapshots per simulation) after superposition of the protein transmembrane Cα's of SERCA2A.

(A) TG in the TG pocket with a lipid present

- (B) TG in the TG pocket without a lipid present
- (C) Hypericin in the TG pocket with a lipid present
- (D) Hypericin in the TG pocket without a lipid present
- (E) Hypericin in Site 2

(F) Hypericin in the cytosolic pocket



Section S11: TG and BHQ in SERCA1A and SERCA2A

TG in SERCA1A

In SERCA1A, TG is docked deeper into the binding pocket when a lipid occupies a major part thereof, compared to the crystal structure. The three rings of the TG skeleton are positioned approximately the same as in the crystal structure, i.e. close to the interface between the lipids and the cytosole, however, the long tail of TG is positioned in the direction of the interior of the pocket instead of 'down' due to the presence of the lipid tail (Fig. S11A). During the MD simulation there is no significant change in the location of neither TG nor the lipid, and it is clear that the TG molecule is not able to push the lipid out of the active site.

When TG is docked without the lipid occupying the binding pocket, the binding mode reproduces that of the crystal structure very well, and the similarity remains fairly well during the MD simulation (Fig. S11B). The flexibility of the TG tail contributes to slightly larger fluctuations in ligand RMSD compared to the other ligands (Fig. S9). A hydrogen bond is formed between one of the hydroxyl groups and the backbone of Lys-252 on the M3 helix, and close contacts are found with Phe-256, which constitutes a key residue in TG binding,¹⁻⁴ and Phe-834. During the MD simulation the neighboring lipids are closing in and tightly surround the TG molecule. Despite the highly different binding modes of TG when a lipid is present or absent, the calculated binding energies are the same (ensemble average and MM/GBVI). In both TG binding modes the molecule is found to reside relatively close to Ile-829 in the upper part of the binding pocket, a residue that is believed to form a hydrogen bond with TG.⁵

BHQ in SERCA1A

Docking of BHQ in this site generated the molecule in a position very similar to the one in the crystal structure and the binding was maintained during the MD simulation (Fig. S11E). Asp-59 and Pro-308, amino acids that are known to be important for BHQ binding,⁴ are positioned to enable interaction. Due to the presence of BHQ that occupies the space on this side of the M4 helix, the side-chain of Glu-309 is positioned on the opposite side of the M4 helix such that it blocks the entrance of Ca^{2+} , as observed in the crystal structure,⁴ cf. Fig. S11E. The binding energies for BHQ are lower than that for TG, in agreement with experimental inhibition constants (cf. Table 1).⁶ The binding energies are also lower than those for hypericin in all binding sites.

TG in SERCA2A

When TG is docked into SERCA2A with a lipid present in the binding pocket, the TG molecule is positioned at an almost 90 degree angle compared to the crystal structure of SERCA1A, with the long tail pointing in the direction of the interior of the protein. Initially during the MD simulation the ring system of the molecule becomes even more rotated, almost 180 degrees, with respect to the TG pose in the SERCA1A crystal structure, but with the long tail still pointing in the direction of the interior of the protein (Fig. S11C). The lipid in the pocket did not move during the MD simulation. The location of the molecule is even higher up in the pocket compared to in SERCA1A and approximately half of the molecule is located in the cytosolic region of the protein. Despite the fact that this binding mode differs significantly from TG in the crystal structure of SERCA1A, the binding energies are higher than the corresponding binding in SERCA1A and it is the overall highest binding energy found for any ligand in the present study. For TG docked into the protein without the lipid occupying the pocket, the binding mode is more similar to the binding mode of TG in the crystal structure of SERCA1A. However, during the

MD simulation the molecule moves out from the pocket and is after the simulation positioned on the surface of the pocket, a location that offers very few interactions with amino acids (Fig. S11D). The binding energies are considerably lower compared to the previous system with TG, and lower than that of both binding modes of TG in SERCA1A.



Pro-308

Glu-309

Fig. S12: Hypericin in the TG binding pocket in (A) SERCA1A and (B) SERCA2A with a lipid in the pocket during docking.



Fig. S13: The two possible conformers of the hypericin molecule; (A) propeller and (B) double-butterfly. Geometries obtained from B3LYP/6-31G(d,p) optimizations.



Fig. S14: Dihedral angles of hypericin as function of time during the MD simulations (800 snapshots) in SERCA1A and SERCA2A.

(A) Hypericin in the TG pocket with a lipid present

- (B) Hypericin in the TG pocket without a lipid present
- (C) Hypericin in the BHQ pocket
- (D) Hypericin in Site2
- (E) Hypericin in the cytosolic pocket

Propeller conformer: both dihedral angles positive. Double-butterfly conformer: one dihedral angle positive, one negative.



References

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