

Electronic Supplementary Information

From ultrafast events to equilibrium - uncovering the unusual dynamics of ESIPT reaction. The case of dually fluorescent diethyl-2,5-(dibenzoxazolyl)-hydroquinone.

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1. Experimental techniques

1a. Stationary spectra

Stationary absorption spectra were measured on JASCO-V550 spectrophotometer. Fluorescence spectra were recorded with the TCSPC system which can also work as a typical spectrofluorometer. In such configuration, instead of the time between excitation and the emission of first photon (TCSPC technique) the total numbers of photons per second vs. wavelength were measured. In this case the signal from the photomultiplier was sent directly to the photon counter. Additionally, the picosecond laser was used as a quasi-stationary source of the excitation light (4 MHz repetition rate).

1b. Chemicals

The diethyl-2,5(dibenzoxazolyl)-hydroquinone (DE-BBHQ)¹ was kindly given to us by professor J. M. Kaufmann from Philadelphia College of Pharmacy, USA. The sample was purified by chromatography and checked with NMR and UV spectra. The molar extinction coefficients determined at $S_0 \rightarrow S_1$ absorption maximum in CCl_4 and C_2Cl_4 are equal: $2.24 \times 10^4 \text{ M}^{-1}\text{cm}^{-1}$ at 416 nm and $2.12 \times 10^4 \text{ M}^{-1}\text{cm}^{-1}$ at 418 nm, respectively. Quantum yield (η_f) of the total fluorescence of DE-BBHQ was also measured using BBHQ in 3-MP as reference.² The $\eta_f = 0.021$ and 0.024 in CCl_4 and C_2Cl_4 , respectively ($\lambda_{ex} = 400$ nm).

Solvents: dichloromethane (DCM), Merck, HPLC grade; tetrachloromethane (CCl_4), Merck, for spectroscopy; 1,2-dichloroethane (DCE), Aldrich, HPLC grade; tetrachlorethylene: (C_2Cl_4) (UV/IR) grade, Roth and Sigma-Aldrich, anhydrous 99%; tetrahydrofuran (THF), Sigma-Aldrich, anhydrous 99.9%; heptane (HEP), Sigma-Aldrich, anhydrous 99%; acetonitrile (ACN), Sigma-Aldrich, HPLC grade.

1c. Femtosecond UV/VIS and IR spectrometers

The ultrafast infrared laser system consists of a short-pulse titanium-sapphire oscillator (Coherent, Vitesse, 50 fs) followed by a high-energy titanium-sapphire regenerative amplifier (Coherent, Positive Light, Legend HE USP).³ The 800 nm beam is split into two beams to generate second harmonic in BBO crystal (400 nm generation) and to pump OPA (Optical Parametric Amplifier, OPerA Coherent). The OPA with a DFG (Difference Frequency Generation) module generates IR pulses (tunable from 2 to 10 μm , 5 μJ at 4 μm). A Ge

beamsplitter splits the IR beam into reference and probe beams and both are focused into a 1 mm path length sample cell (Harrick Scientific), but only the probe beam overlaps with the pump beam in the sample. After passing through the sample the probe and reference beams were spectrally dispersed in a grating spectrometer (Horiba, iHR 320) and independently imaged on a liquid nitrogen cooled HgCdTe detector (2×32 elements) with 17 nm resolution. To avoid any interferences with residual absorption of water vapors, IR beams are permanently purged with dry air. Every other UV pump pulse is blocked by a synchronized chopper to eliminate long-term drift effects. The pump pulse energy was about 2 μ J at the sample position.

The broadband UV/VIS femtosecond transient absorption system has been described elsewhere.^{4,5} The sample concentration was adjusted for an optical density of 1 at the excitation wavelength with a 1 mm path length. A sample volume of 50 mL was continuously flowed through the sample cell to ensure that fresh sample was photoexcited. The entire set of pump-probe kinetics is recorded at least three times, to ensure data reproducibility. To avoid rotational diffusion effects, the angle between polarizations of the pump beam and the probe beam was set to the magic angle (54.7°). Kinetic traces are analyzed by fitting to a sum of exponential terms. Convolution with a Gaussian response function is included in the global fitting procedure. The instrument response was approximately 300 fs Full Width at Half Maximum (FWHM). All experiments were performed at room temperature.

1d. Time Correlated Single Photon Counting (TCSPC) spectrometer

The time correlated single photon counting (TCSPC) system has been described elsewhere.⁶ Briefly, spectrometer consists of the picosecond titanium-sapphire laser oscillator (Tsunami, Spectra Physics), pumped by Millennia Prime. Repetition rate of the picosecond oscillator (1.5 ps pulse duration) was reduced to 4 MHz using an electrooptical pulse picker. The second harmonic (400 nm) was generated by focusing fundamental beam onto surface of a BBO nonlinear crystal. The excitation beam was then focused in the sample by a quartz lens. The excitation beam intensity was adjusted by a rotating neutral filter to eliminate the photon pile-up effect. The emission polarizer was set at the magic angle in respect to vertical polarization of the excitation beam. The monochromator of the focal length 300 mm and aperture $f/4.2$ was equipped with a diffraction grating of 1800 grooves/mm. Optical signal was registered with proximity type 6 μ m microchannel plate MCP-PMT R3809U-05 (Hamamatsu,

thermoelectrically cooled to -20°C), equipped with a high speed preamplifier C5594 (Hamamatsu). Electronic signal pathway was composed of an upgraded constant fraction discriminator CFD TC 454 (Tennelec), time to amplitude converter TAC-TC 864 (Tennelec), and a multichannel analyzer Norland 5000 MCA. The time per channel determined by the optical method was 0.61 ± 0.01 ps. The full width at half maximum of IRF was about 30 ps. All experiments were performed at room temperature in non degassed solutions.

1e. Femtosecond up-conversion spectrometer

For fluorescence measurements with femtosecond time resolution we used up-conversion spectrometer, similar to one described in Ref. 7. It consists of femtosecond Ti:Sapphire oscillator (Coherent Mira, 110 fs, 80 MHz), followed by home build regenerative amplifier (5 kHz repetition rate, 1 mJ). Amplified pulses were split into two parts by a beam splitter, one part was used to generate second harmonic beam at 400 nm (utilizing BBO nonlinear crystal), while the rest of the pulse was sent to Noncollinear Optical Parametric Amplifier (NOPA). Central wavelength of the pulses generated by NOPA was set at 1020 nm with compressed pulse energy of 10 μJ . Excitation beam at 400 nm was focused with a quartz lens on cuvette with 200 μm sample thickness. Fluorescence induced by excitation pulse was collected with all reflective Schwarzschild objective and imaged onto surface of BBO nonlinear crystal. Infrared pulses from NOPA which served as a ultrafast gate, were sent through a delay stage and were mixed with fluorescence signal inside nonlinear BBO crystal. Up-converted signal was then routed to a imaging spectrometer (Andor, SR-303i), equipped with EM-CCD camera (Hamamatsu C9100-13). Spectrum for each delay between pulses was recorded many times to increase signal to noise ratio. Scattered excitation beam, which would saturate camera, was blocked by a color filter before an up-conversion process. A number of dispersive elements (such as: color filter, cuvette cover glass, solvent and BBO gating nonlinear crystal) in a fluorescence beam path results in a wavelength dependent group delay. A precise value of dispersion introduced by mentioned elements was determined independently with a white light interferometric spectrometer. Measured fluorescence evolution map was then corrected for measured group delay, with achieved temporal correction accuracy better than 5 fs in range of 400-700 nm. Setup was used with two different up-converting BBO crystals i.e. 0.2 mm – for high temporal resolution measurements (IRF=160 fs FWHM) and 2 mm thick crystal – for detection of weak signal with

reduced temporal resolution (IRF=450 fs FWHM). In the first case, temporal resolution was limited by excitation pulse duration, while in the second case it was determined by group velocity mismatch in a thick up-converting crystal. Accuracy of retrieved decay time constants in our case, are limited by a noise of recorded data, in practice accuracy was not worse than $\pm 10\%$. For all experiments, polarization between excitation and gate beam was set at a magic angle configuration – to avoid rotational and diffusional effects of molecular motions.

2. Computational methods

All computations were done within the Gaussian 09 suite of programs.⁸ The geometry optimizations for the ground (S_0) and the first electronically excited singlet state (S_1) were performed using the Density Functional Theory (DFT) and its time dependent (TD-DFT) formalism, respectively. The B3LYP hybrid functional and the 6-31+G(d,p) basis set were applied.

The potential energy profiles for the proton transfer reactions were obtained as a potential energy scan (PES) along the so-called coordinate-driven reaction path (CDRP, all coordinates are optimized except one, appropriately chosen, which is fixed and varied systematically). The fixed coordinate was the longer of the O-H and N-H distances in a hydrogen bond with a translocating proton. For the double proton transfer C_{2h} symmetry of the system was preserved along the path (concerted symmetric reaction). The real transition states were also localized, with the use of the TS Gaussian 09 option. Vibrational frequency analysis performed for all of the localized stationary points, confirmed their minimum/saddle point nature.

The calculations, which included the presence of a solvent, used a Polarizable Continuum Model (PCM) in its state-specific implementation. Full geometry optimization in solution was performed for the ground state, whereas for the excited S_1 state vacuum geometry was used. Non-equilibrium solvation energies of the vertically excited states have been calculated to obtain vertical excitation energies in solution.

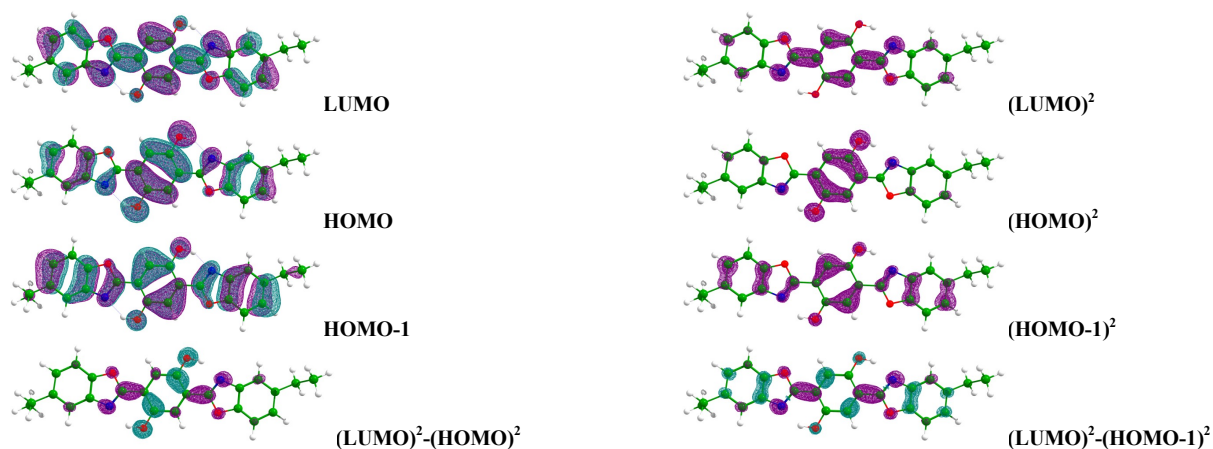


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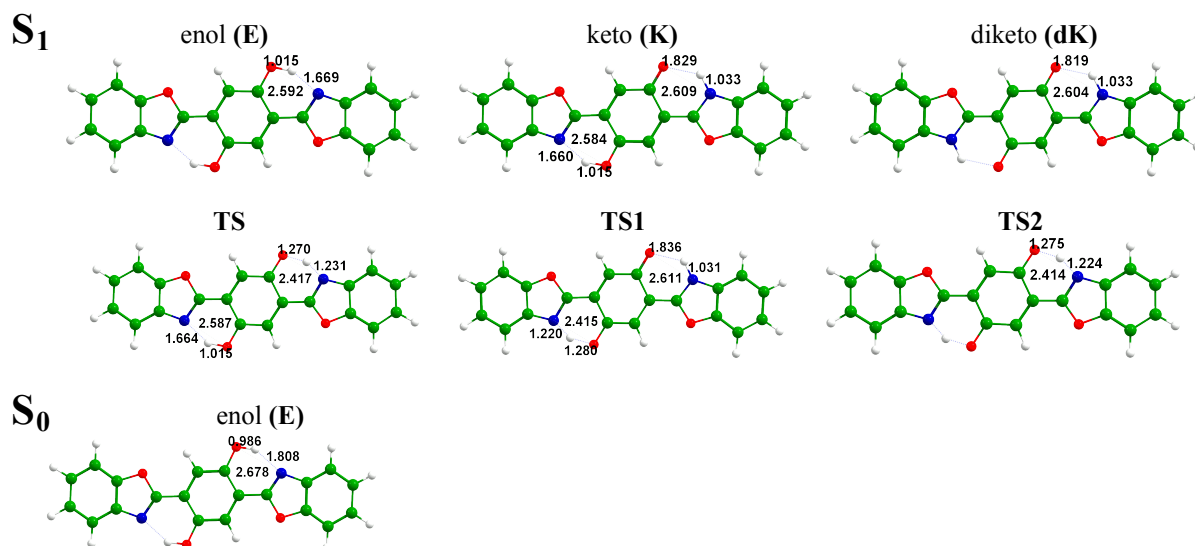


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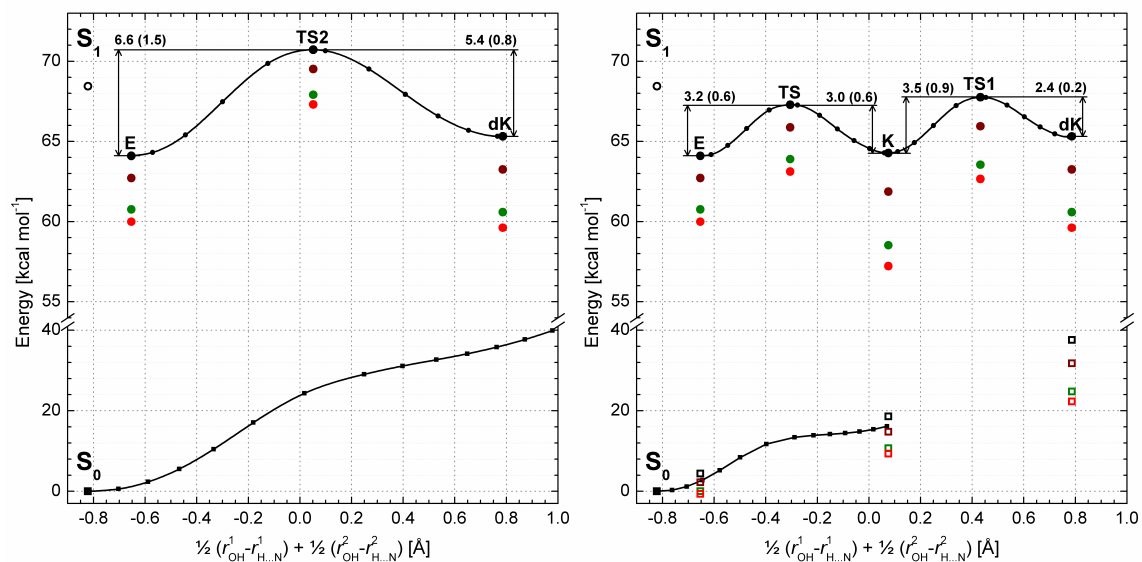


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Table S1. Relative energies of different tautomeric forms and transition states between them in the S_1 optimized state ($E_{S1}/\Delta E_{S1}/\Delta E_{S1\ ZPE}$) and vertically excited the S_0 state (E_{S0}), solvation energies (E_{solv}), dipole moments (μ), vertical excitation energies (Ex), oscillator strengths (f) and relative fluorescence intensities under equilibrium conditions (I_i/I_{total}) of BBHQ/DE-BBHQ in vacuum and in solutions calculated at the DFT level.

		E_{S1} [kcal/mol]	$E_{solv\ S1}$ [kcal/mol]	ΔE_{S1}^a [kcal/mol]	$\Delta E_{S1\ ZPE}^{a,b}$ [kcal/mol]	μ_{S1} [D]	E_{S0} [kcal/mol]	$E_{solv\ S0}$ [kcal/mol]	μ_{S0} [D]	$Ex_{S1\rightarrow S0}$ [cm ⁻¹]	$f_{S1\rightarrow S0}$	I_i/I_{total}^c	
BBHQ	vacuum	E	64.10	0.00	0.00	0.00	4.43		0.00	20870	0.431	0.619	
		TS	67.29		3.18	0.64	2.24	14.95		1.48			
		K	64.28		0.18	0.05	5.24	18.60		2.51	15980	0.238	0.369
		TS1	67.76		3.66	0.98	2.74	31.38		0.68			
		dK	65.32		1.21	0.78	0.00	37.61		0.00	9690	0.145	0.012
		TS2	70.72		6.62	1.53	0.00	26.62		0.00			
	hexane	E	62.72	-1.38	0.00	0.00	0.00	2.25	-2.19	0.00	21150	0.432	0.218
		TS	65.88	-1.41	3.16	0.61	2.74	12.07	-2.88	1.74			
		K	61.87	-2.41	-0.85	-0.97	6.38	14.81	-3.79	3.02	16460	0.242	0.764
		TS1	65.95	-1.81	3.23	0.55	3.33	26.71	-4.67	0.91			
		dK	63.25	-2.07	0.53	0.09	0.00	31.78	-5.83	0.00	11010	0.156	0.018
		TS2	69.52	-1.21	6.80	1.71	0.00	22.94	-3.67	0.00			
	THF	E	60.76	-3.34	0.00	0.00	0.00	0.05	-4.38	0.00	21230	0.429	0.025
		TS	63.90	-3.39	3.14	0.59	3.50	9.07	-5.88	2.22			
		K	58.53	-5.75	-2.24	-2.36	8.00	10.70	-7.90	4.17	16730	0.247	0.965
		TS1	63.55	-4.21	2.79	0.11	4.12	21.41	-9.97	1.57			
		dK	60.59	-4.73	-0.17	-0.61	0.00	24.79	-12.82	0.00	12520	0.170	0.010
		TS2	67.91	-2.81	7.15	2.06	0.00	18.93	-7.68	0.00			
acetonitrile	E	59.99	-4.11	0.00	0.00	0.00	-0.65	-5.08	0.00	21210	0.426	0.010	
	TS	63.12	-4.17	3.13	0.58	3.82	8.09	-6.85	2.44				
	K	57.22	-7.06	-2.78	-2.90	8.64	9.35	-9.25	4.67	16740	0.248	0.984	
	TS1	62.66	-5.10	2.67	-0.01	4.42	19.60	-11.78	1.86				
	dK	59.62	-5.70	-0.38	-0.81	0.00	22.32	-15.29	0.00	13050	0.176	0.006	
	TS2	67.31	-3.41	7.32	2.22	0.00	17.59	-9.03	0.00				
DE-BBHQ	vacuum	E	64.08		0.00	0.00	0.00	4.41		0.00	20870	0.501	0.589
		K	64.18		0.10	-0.04	5.49	18.44		2.66	16000	0.264	0.392
		dK	64.99		0.90	0.47	0.00	37.08		0.00	9760	0.160	0.019
	hexane	E	62.79	-1.29	0.00	0.00	0.00	2.20	-2.21	0.00	21190	0.506	0.198
		K	61.87	-2.31	-0.92	-1.06	6.57	14.62	-3.81	3.15	16520	0.270	0.774
		dK	62.98	-2.01	0.18	-0.26	0.00	31.24	-5.84	0.00	11100	0.172	0.028
	acetonitrile	E	60.23	-3.85	0.00	0.00	0.00	-0.84	-5.25	0.00	21360	0.509	0.009
		K	57.39	-6.80	-2.85	-2.99	8.66	9.04	-9.40	4.69	16910	0.280	0.979
		dK	59.36	-5.63	-0.87	-1.31	0.00	21.70	-15.38	0.00	13170	0.195	0.012

^a relative to the enol form in a given environment

^b zero point vibrational energy corrected values (vibrational energy scaled by 0.964)

^c calculated according to $I_i/I_{total} = g_i \times \exp(-E_i/(k \times T)) \times Ex_i^2 \times f_i / \sum_i (g_i \times \exp(-E_i/(k \times T)) \times Ex_i^2 \times f_i)$, where: g – degeneracy (equal 1 for **E** and **dK** and 2 for **K**), k – Boltzmann constant, T – temperature (298 K), $E = \Delta E_{S1\ ZPE}$ and summation covers **E**, **K** and **dK** forms

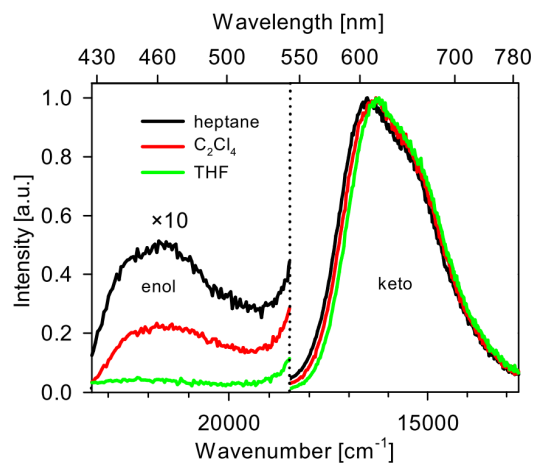


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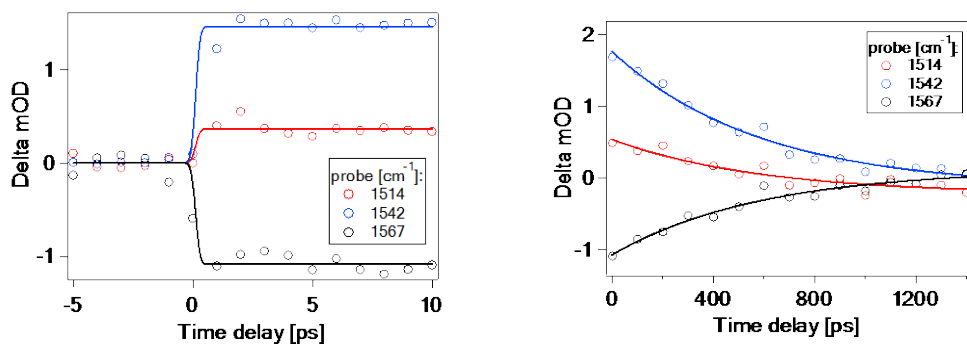


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Table S2. Kinetic parameters obtained with UV/VIS transient absorption technique.

Wavelength [nm]	A_1	τ_1	A_2	τ_2 [ps]	A_3	τ_3 [ps]	offset
430	-0.048	0.25	—	—	-0.033	271	-0.003
490	-0.049	0.18	—	—	0.031	383	0.002
650	0.041	0.33	0.003	7.7	-0.018	365	0.000

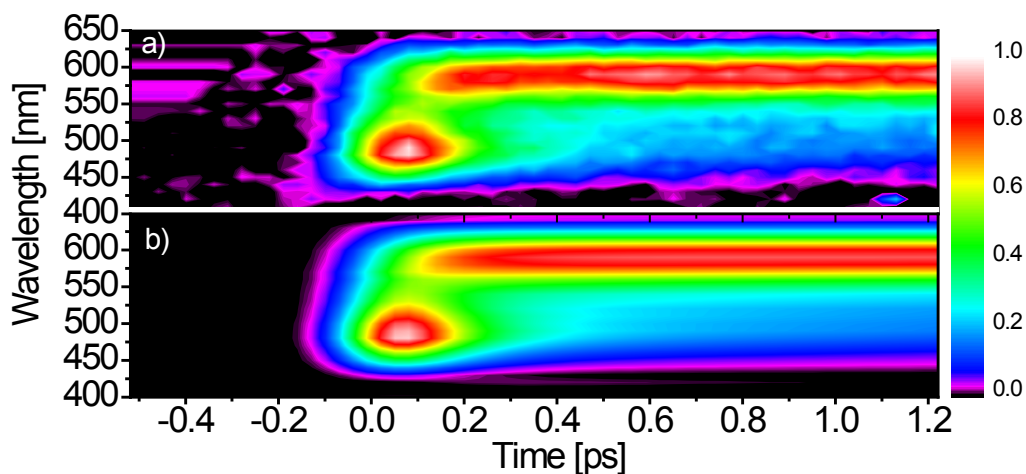


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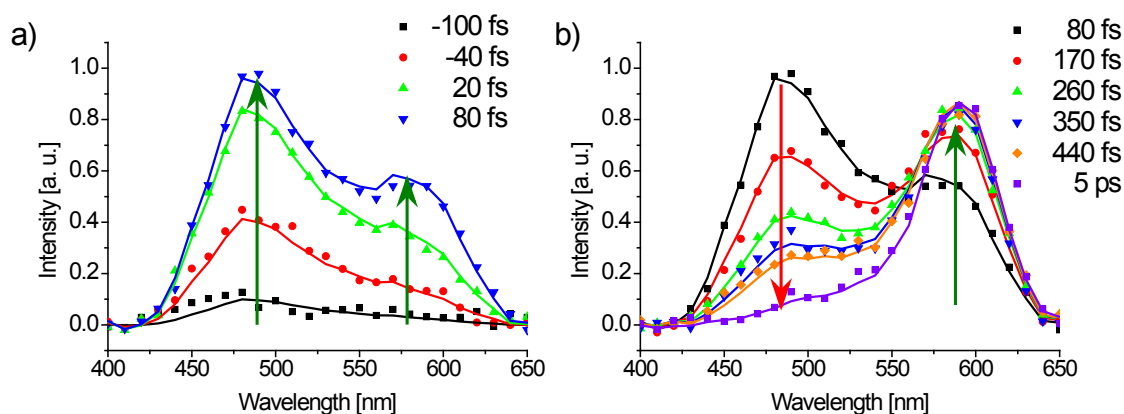


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