

How protein structure affects redox reactivity: example of Human centrin 2.

Abdeslam Et Taouil, Emilie Brun, Patricia Duchambon, Yves Blouquit, Manon Gilles, Emmanuel Maisonhaute*, Cécile Sicard-Roselli*

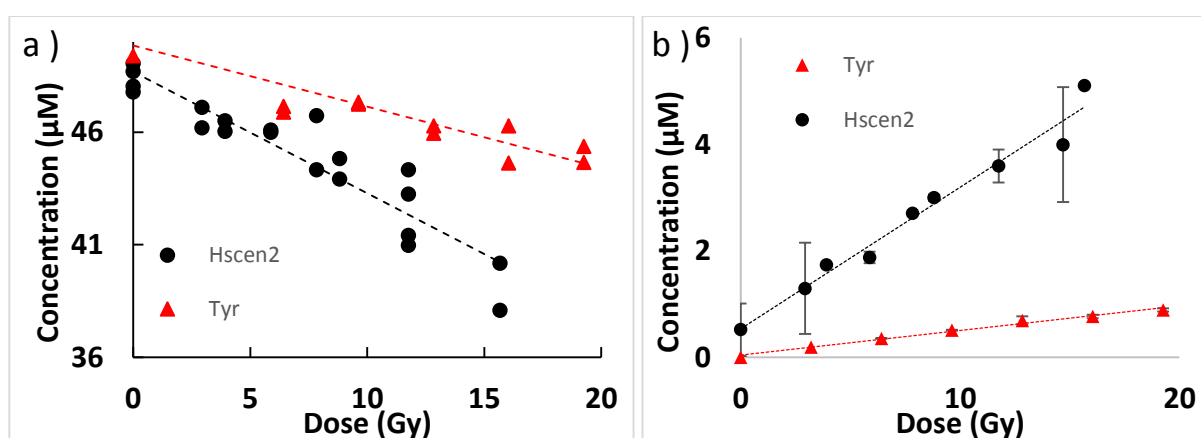
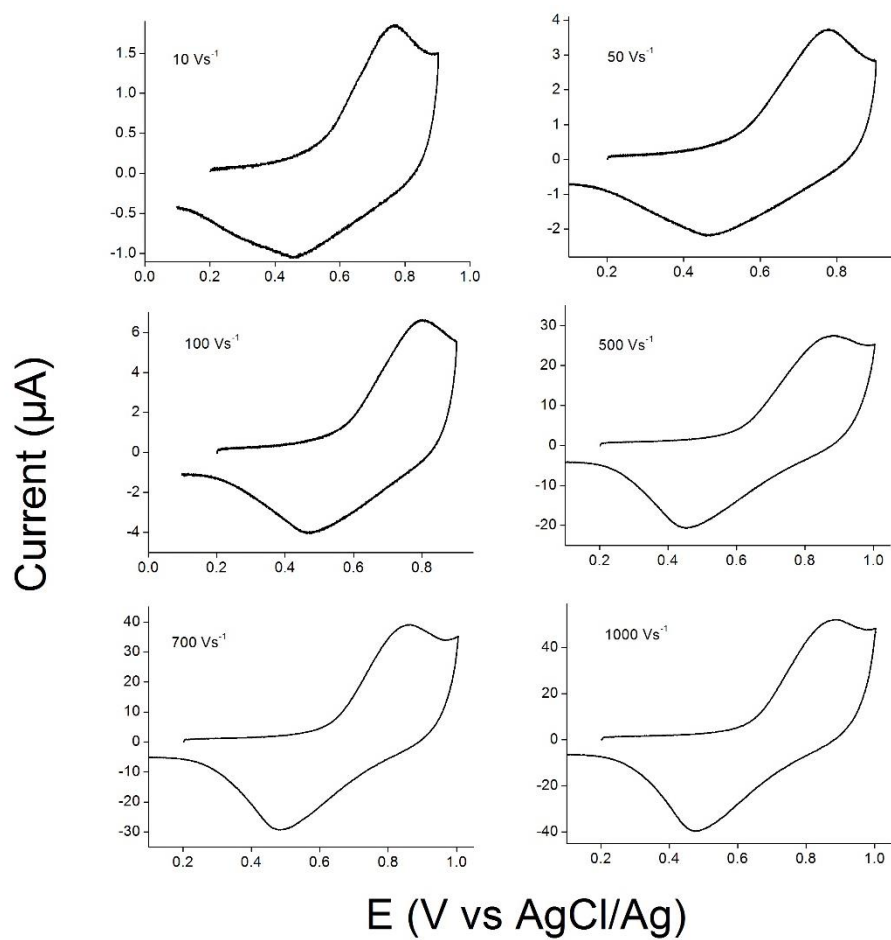


Figure S1. a) Hscen 2 and Tyrosine concentration evolution as a function of the irradiation dose determined by liquid chromatography. Yield determined: $G_{(-\text{Hscen2})} = 0.57 \pm 0.05 \mu\text{mol}\cdot\text{J}^{-1}$, $G_{(-\text{Tyr})} = 0.27 \pm 0.04 \mu\text{mol}\cdot\text{J}^{-1}$. b) Dimer concentration evolution as a function of the irradiation dose determined by liquid chromatography. Yield determined: $G_{(\text{Hscen2Dimer})} = 0.26 \pm 0.02 \mu\text{mol}\cdot\text{J}^{-1}$, $G_{(\text{TyrDimer})} = 0.046 \pm 0.002 \mu\text{mol}\cdot\text{J}^{-1}$.



Figures S2. Cyclic voltammograms of Hscen2 (0.2 mM) in 0.2 M NaCl aqueous solution at different scan rates onto a 125 μm diameter gold electrode.

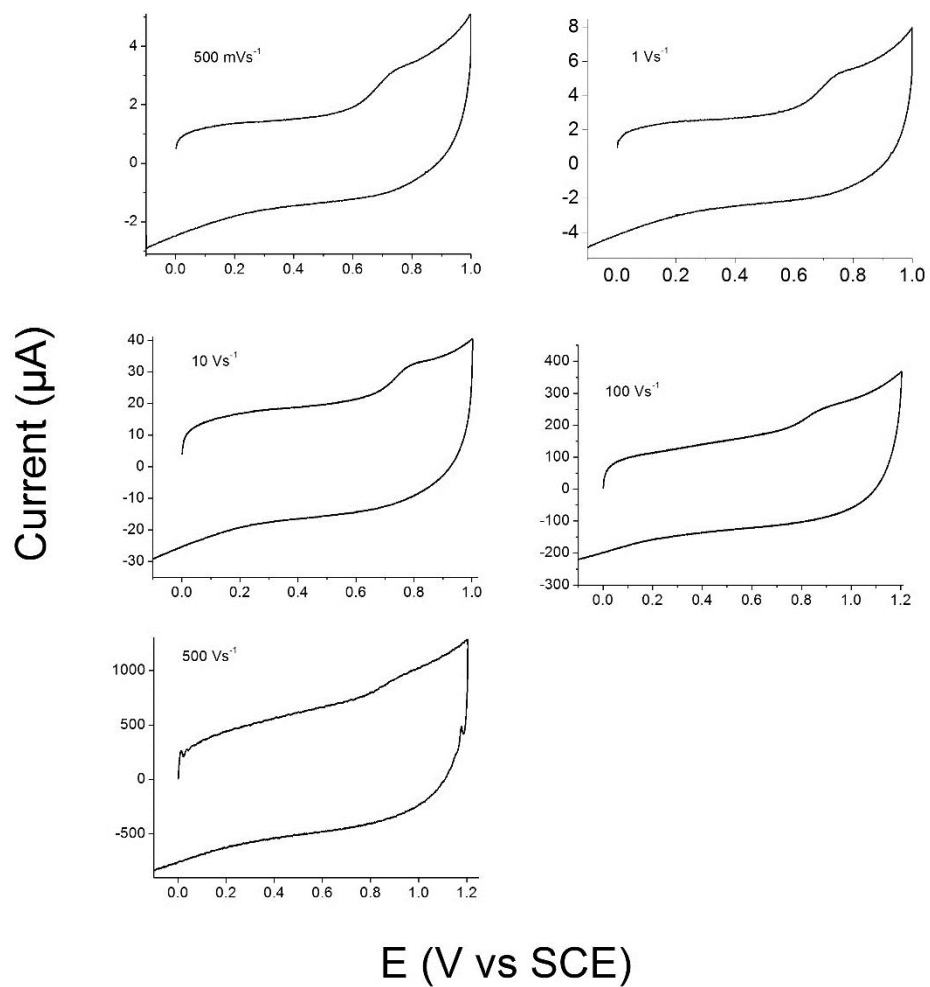


Figure S3. Cyclic voltammograms of 0.2 mM tyrosine in 0.5 M LiNO₃ aqueous solution on a 1 mm diameter glassy carbon electrode at different scan rates.

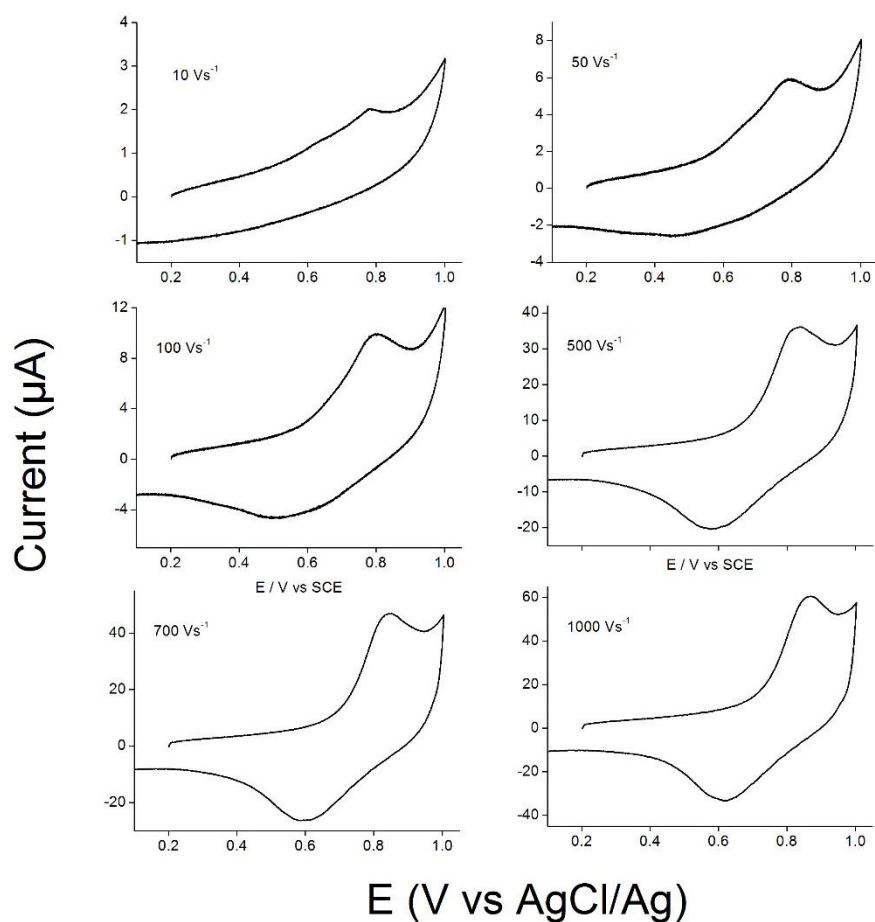


Figure S4. Cyclic voltammograms of $\Delta 25$ Human centrin 2 (0.2 mM) in 0.2 M NaCl aqueous solution at different scan rates onto a 125 μm diameter gold electrode.

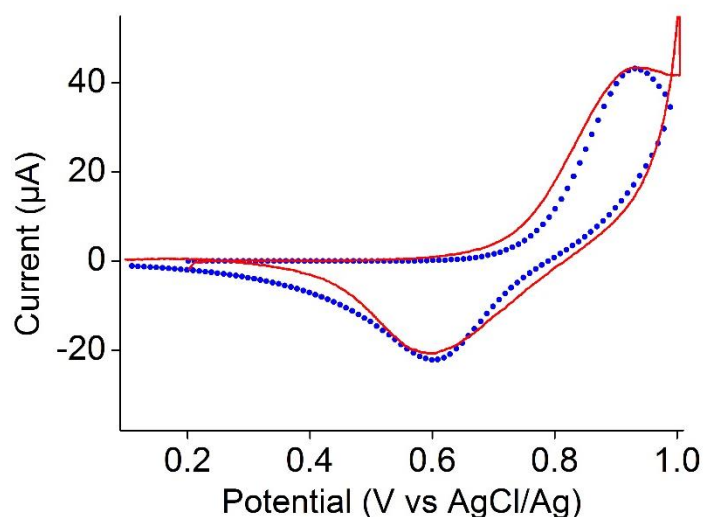


Figure S5. Experimental (solid line) and simulated cyclic voltammograms (dots) of $\Delta 25$ Human Centrin 2 (0.2 mM) in 0.2 M NaCl aqueous solution at 1000 Vs^{-1} onto a 125 μm diameter gold electrode. The background current is subtracted. Simulation parameters: $E^0 = 0.765$ V vs AgCl/Ag diffusion coefficient $D_{\Delta 25} = 8.2 \times 10^{-7}$ $\text{cm}^2 \text{s}^{-1}$, $k_{\text{dim}} = 1.45 \times 10^4$ $\text{L mol}^{-1} \text{s}^{-1}$, layer thickness = 180 nm, $[\Delta 25] = 59$ mM, pH = 7.5.

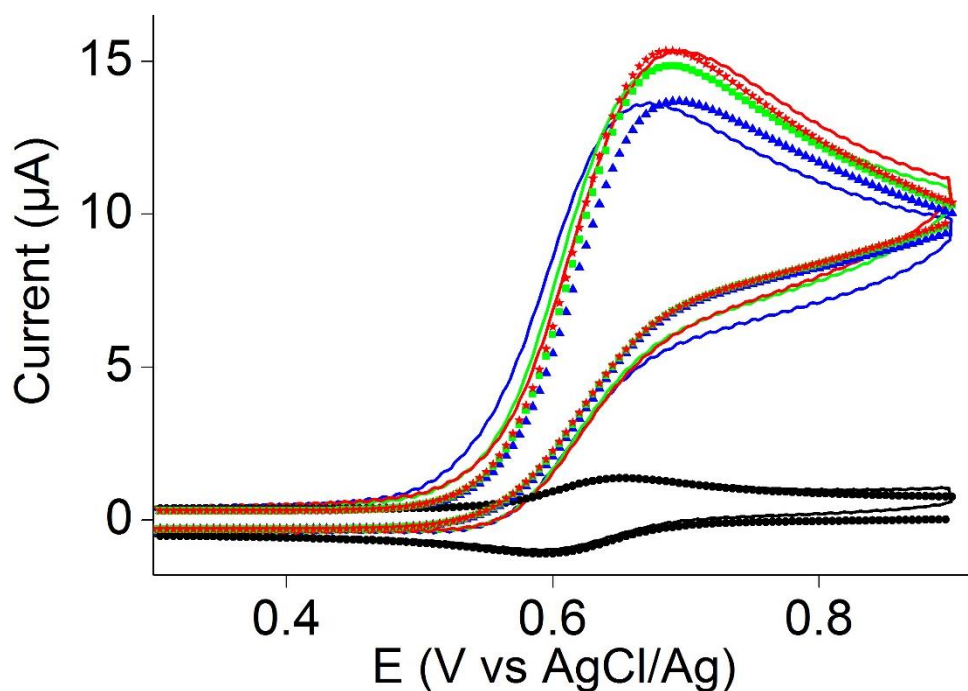


Figure S6. Redox catalysis of 0.1 mM tyrosine in the presence of 11 μM $\text{Os}(\text{bpy})^{2+}$ ($E^{\circ}_{\text{Os}(\text{bpy})^{2+/3+}} = 623$ mV/AgCl/Ag; $E^{\circ}_{\text{Tyrosine}} = 730$ mV/AgCl/Ag; $D_{\text{Os}(\text{bpy})^{2+/3+}} = 6.10^{-6}$ $\text{cm}^2\cdot\text{s}^{-1}$; $D_{\text{Tyrosine}} = 4.10^{-5}$ $\text{cm}^2\cdot\text{s}^{-1}$; $k_{\text{dimerization}} = 10^7$ $\text{M}^{-1}\cdot\text{s}^{-1}$; $K_{\text{eq}} = 1.5.10^5$). Phosphate buffer concentrations of 10 (blue), 50 (green) and 100 (red) mM, k_{app} were found to be 3.5×10^6 , 4.5×10^6 and 5.1×10^6 $\text{M}^{-1}\cdot\text{s}^{-1}$, respectively. The fits are performed taking a diffusion coefficient D_{Tyrosine} of 4×10^{-5} $\text{cm}^2\cdot\text{s}^{-1}$ in line with Fecenko *et al.*¹ Such high value nevertheless seems overestimated in comparison with the one published recently by Wiegang *et al.*² Nevertheless, introducing a larger k_{app} and a smaller D_{tyrosine} did not lead to a correct fit since then the system shifted to the "total catalysis" zone as described by Saveant.³ Revisiting the model of Fecenko *et al.* was however beyond the scope of this study.

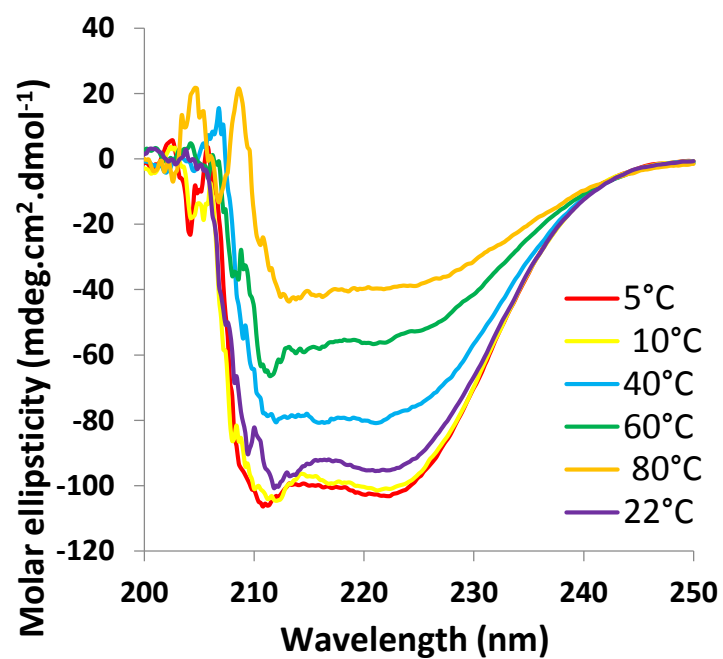


Figure S7. Hscen2 circular dichroism signal evolution as a function of the temperature.

References.

1. C. J. Fecenko, T. J. Meyer and H. H. Thorp, *J. Am. Chem. Soc.*, 2006, 128, 11020-11021.
2. Z. L. Wang, H. Kriegs and S. Wiegand, *J. Phys. Chem. B*, 2012, 116, 7463-7469.
3. J.-M. Saveant, *Elements Molecular and Biomolecular Electrochemistry*, John Wiley and Son, 2002.