

Supporting Information for

Efficient asymmetric hydrogenation of quinolines in neat water catalyzed by chiral cationic Ru-diamine complexes

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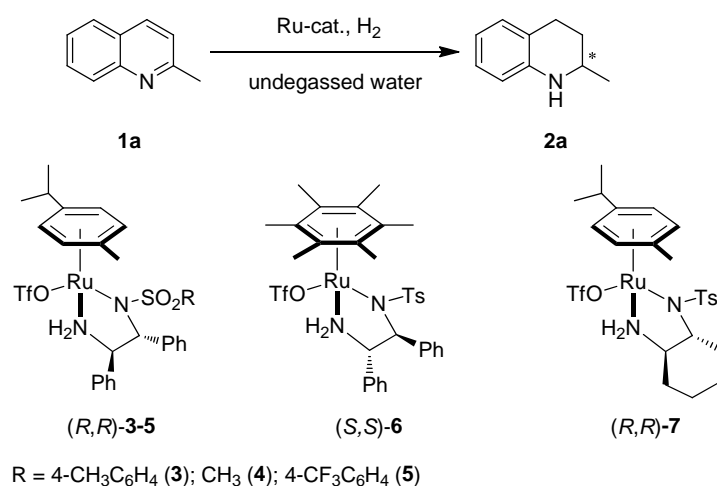
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1. General information

Unless otherwise noted, all experiments were carried out by using standard Schlenk techniques in undegassed water without the use of glovebox. ^1H NMR and ^{13}C NMR spectra were recorded on a Bruker Model Avance DMX 300 Spectrometer (^1H 300 MHz and ^{13}C 75 MHz, respectively). Chemical shifts (δ) were given in ppm and were referenced to residual solvent or TMS peaks. Optical rotations were measured with Rudolph Autopl VI polarimeter. HPLC analyses were performed on a VARIAN PROSTAR 210 liquid chromatograph. The water was purified by ULUPURE UPHW-III-90T ultra-pure grade water system. All other chemicals were used as received from Aldrich or Acros without further purification. The catalysts were prepared according to the published method.^[1] Quinolines **1a-1k** were prepared according to the literature procedure and their ^1H and ^{13}C NMR data are consistent with the literature data.^[1]

2. Optimization of conditions for asymmetric hydrogenation of **1a**

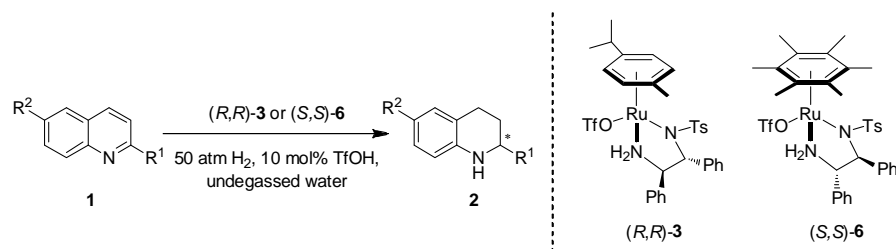
Table S1: Optimization of reaction conditions for the asymmetric hydrogenation of **1a** in water.^[a]



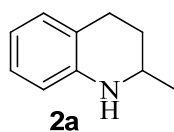
Entry	Catalyst	H ₂ (atm); T (°C)	TfOH (equiv)	Time (h)	Conv. (%) ^[b]	Ee (%) ^[c]
1 ^[d]	(<i>R,R</i>)- 3	50; 30	0	3	26	95
2 ^[d]	(<i>R,R</i>)- 3	50; 30	10	3	71	94
3	(<i>R,R</i>)- 3	50; 30	5	3	>99	95
4	(<i>R,R</i>)- 3	50; 30	10	3	>99	95
5	(<i>R,R</i>)- 3	50; 30	10	0.5	30	95
6	(<i>R,R</i>)- 3	50; 30	20	3	>99	94
7	(<i>R,R</i>)- 3	10; 30	10	3	64	95
8	(<i>R,R</i>)- 3	30; 30	10	3	>99	96
9	(<i>R,R</i>)- 3	80; 30	10	3	>99	96
10	(<i>R,R</i>)- 3	50; 20	10	3	>99	95
11	(<i>R,R</i>)- 3	50; 50	10	3	>99	94
12	(<i>R,R</i>)- 4	50; 30	10	3	>99	92
13	(<i>R,R</i>)- 5	50; 30	10	3	>99	94
14	(<i>S,S</i>)- 6	50; 30	10	3	>99	99
15	(<i>S,S</i>)- 6	50; 30	10	3	>99	99
16	(<i>R,R</i>)- 7	50; 30	10	3	>99	82
17 ^[f]	(<i>R,R</i>)- 3	50; 50	10	12	>99	94
18 ^[f]	(<i>S,S</i>)- 6	50; 50	10	12	87	98

[a] Reaction conditions: **1a** (0.2 mmol) in undegassed water (1 mL), Ru-cat. (1.0 mol %), reaction for 3 h. All manipulations were conducted in air, and the autoclave was purged with H₂ three times before reaction. [b] The conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. [c] The enantiomeric excesses were determined by HPLC with a chiral OJ-H column. [d] All manipulations were conducted under anaerobic conditions (with the use of degassed methanol and glovebox), and the autoclave was purged with H₂ three times before reaction. [e] All manipulations were conducted in air, without purging the autoclave with H₂ before reaction. [f] Substrate/catalyst = 500 (143 mg substrate in 5 mL H₂O).

3. Asymmetric hydrogenation of quinolines in undegassed water



Typical procedure for the asymmetric hydrogenation of quinolones in water: quinolines **1** (0.20 mmol), (*R,R*)-**3** or (*S,S*)-**6** (0.002 mmol, 1.0 mol%), undegassed water (1 mL) and undegassed TfOH (3.0 mg, 0.02 mmol, 10 mol%) were added to a glass tube in air. Then, the tube was placed into a 30 mL stainless steel autoclave and the autoclave was purged with hydrogen gas for 3 times before pressurized with H₂ to 50 atm. The hydrogenation was performed at 30 °C or 50 °C for the specified period of time. After careful releasing of the hydrogen, the reaction mixture was diluted with ethyl acetate (3 mL) and saturated sodium carbonate aqueous solution (2 mL), then stirred for 2 min. The aqueous layer was extracted with ethyl acetate (3 x 3 mL). The combined organic layer was dried over anhydrous sodium sulfate and concentrated to afford the crude product. The conversion of the reaction was based on ¹H NMR spectra. Purification was performed by a silica gel column eluted with petroleum ether/triethylamine (95:5, v/v) to give the pure product. The enantiomeric excesses were determined by chiral HPLC with a chiral column.

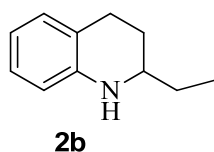


(*S*)-2-Methyl-1,2,3,4-tetrahydroquinoline (**2a**). (Known

compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He,

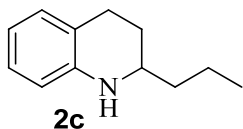
Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.*

2011, *133*, 9878-9891). Isolated yield 95%, 99% ee, $[\alpha]_D^{25} = -81.0$ (*c* 0.50, CHCl₃), [Lit.^[1] $[\alpha]_D^{RT} = +84.3$ (*c* 0.20, CHCl₃) 99% ee for *R* enantiomer]; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.04-7.00 (m, 2H), 6.60 (t, *J* = 7.35 Hz, 1H), 6.53-6.50 (m, 1H), 3.74 (b, 1H), 3.50-3.39 (m, 1H), 2.95-2.73 (m, 2H), 2.02-1.94 (m, 1H), 1.71-1.58 (m, 1H), 1.26 (d, *J* = 6.30 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 144.83, 129.35, 126.77, 121.19, 117.07, 114.11, 47.24, 30.21, 26.68, 22.69; The enantiomeric excess was determined by HPLC on Chiralcel OJ-H column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{R1} = 10.1$ min (major), $t_{R2} = 11.3$ min (minor).



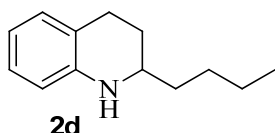
(R)-2-Ethyl-1,2,3,4-tetrahydroquinoline (2b). (Known compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He, Q.-H. Fan, J.

Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 95%, 94% ee, $[\alpha]_D^{25} = +73.0$ (*c* 0.50, CHCl₃), [Lit.^[1] $[\alpha]_D^{RT} = +80.3$ (*c* 0.19, CHCl₃) 99% ee for *R* enantiomer]; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 6.99 (t, *J* = 7.05, 2H), 6.63 (t, *J* = 7.20 Hz, 1H), 6.51 (d, *J* = 8.10 Hz, 1H), 3.78 (b, 1H), 3.24-3.16 (m, 1H), 2.89-2.73 (m, 2H), 2.05-1.96 (m, 1H), 1.69-1.51 (m, 3H), 1.02 (t, *J* = 7.50 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 144.84, 129.34, 126.82, 121.51, 116.99, 114.12, 53.15, 29.51, 27.69, 26.53, 10.17; The enantiomeric excess was determined by HPLC on Chiralcel OJ-H column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{R1} = 9.4$ min (minor), $t_{R2} = 10.3$ min (major).



(R)-2-Propyl-1,2,3,4-tetrahydroquinoline (2c). (Known

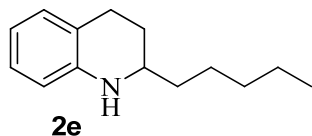
compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 93%, 94% ee, $[\alpha]_D^{25} = +78.6$ (*c* 0.50, CHCl₃), [Lit.^[1] $[\alpha]_D^{RT} = +89.0$ (*c* 0.16, CHCl₃) 99% ee for *R* enantiomer]; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 6.99 (t, *J* = 7.05 Hz, 2H), 6.62 (t, *J* = 7.20 Hz, 1H), 6.50 (d, *J* = 8.10 Hz, 1H), 3.77 (b, 1H), 3.28 (d, *J* = 6.00 Hz 1H), 2.90-2.71 (m, 2H), 2.01-1.94 (m, 1H), 1.69-1.43 (m, 5H), 1.00 (d, *J* = 7.20 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 144.84, 129.37, 126.81, 121.50, 116.99, 114.15, 51.42, 39.01, 28.24, 26.55, 19.02, 14.32; The enantiomeric excess was determined by HPLC on Chiralcel OJ-H column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{R1} = 8.7$ min (minor), $t_{R2} = 10.9$ min (major).



(R)-2-Butyl-1,2,3,4-tetrahydroquinoline (2d). (Known

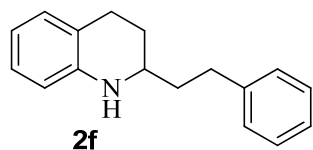
compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 96%, 93% ee, $[\alpha]_D^{25} = +73.0$ (*c* 0.50, CHCl₃), [Lit.^[1] $[\alpha]_D^{RT} = +90.4$ (*c* 0.19, CHCl₃) 99% ee for *R* enantiomer]; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.00 (t, *J* = 7.05 Hz, 2H), 6.64 (t, *J* = 7.35 Hz, 1H), 6.51 (d, *J* = 8.10 Hz, 1H), 3.79 (b, 1H), 3.29-3.25 (m, 1H), 2.86-2.78 (m, 2H), 2.03-1.96 (m, 1H), 1.67-1.40 (m, 7H), 0.98 (t, *J* = 6.90 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 144.84, 129.34, 126.79, 121.47, 116.96, 114.12, 51.68, 36.53, 28.23, 28.03, 26.55, 22.96, 14.21; The enantiomeric excess was determined by HPLC on Chiralcel OJ-H

column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{R1} = 7.6$ min (minor), $t_{R2} = 8.7$ min (major).



(R)-2-Pentyl-1,2,3,4-tetrahydroquinoline (2e). (Known compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding,

Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 91%, 95% ee, $[\alpha]_D^{25} = +106.0$ (*c* 0.50, CHCl₃), [Lit.^[1] $[\alpha]_D^{RT} = +87.3$ (*c* 0.20, CHCl₃) 99% ee for *R* enantiomer]; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 6.99 (t, *J* = 7.05 Hz, 2H), 6.63 (t, *J* = 7.20 Hz, 1H), 6.51 (d, *J* = 8.10 Hz, 1H), 3.77 (b, 1H), 3.31-3.23 (m, 1H), 2.89-2.71 (m, 2H), 2.04-1.95 (m, 1H), 1.70-1.37 (m, 9H), 0.95 (t, *J* = 6.75 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 144.83, 129.35, 126.80, 121.50, 116.99, 114.15, 51.71, 36.80, 32.08, 28.23, 26.56, 25.52, 22.77, 14.18; The enantiomeric excess was determined by HPLC on Chiralcel OJ-H column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{R1} = 6.8$ min (minor), $t_{R2} = 7.4$ min (major).

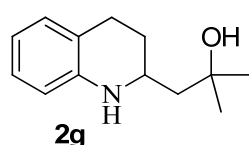


(R)-2-Phenethyl-1,2,3,4-tetrahydroquinoline (2f).

(Known compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen,

Z. Ding, Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 95%, 93% ee, $[\alpha]_D^{25} = +78.6$ (*c* 0.50, CHCl₃), [Lit.^[1] $[\alpha]_D^{RT} = +88.9$ (*c* 0.27, CHCl₃) 99% ee for *R* enantiomer]; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.31-7.19 (m, 5H), 6.95 (t, *J* = 7.05 Hz, 2H), 6.59 (t, *J* = 7.35 Hz, 1H), 6.43 (d, *J* = 7.80 Hz, 1H), 3.72 (b, 1H), 3.32-3.23 (m, 1H), 2.80-2.69 (m, 4H),

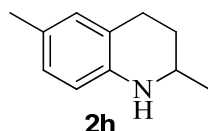
2.02-1.93 (m, 1H), 1.85-1.78 (m, 2H), 1.72-1.61 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 144.60, 141.95, 129.36, 128.59, 128.46, 126.84, 126.07, 121.39, 117.14, 114.25, 51.22, 38.35, 32.27, 28.07, 26.32; The enantiomeric excess was determined by HPLC on Chiralcel AS-H column (hexane : isopropanol = 95 : 5, flowing rate = 0.6 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{\text{R1}} = 9.4$ min (minor), $t_{\text{R2}} = 10.5$ min (major).



(S)-2-methyl-1-(1,2,3,4-tetrahydroquinolin-2-yl)propan-2-ol

(2g). (Known compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen,

Z. Ding, Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 92%, 93% ee, $[\alpha]_{\text{D}}^{25} = +52.6$ (c 0.50, CHCl_3), [Lit.^[1] $[\alpha]_{\text{D}}^{\text{RT}} = +80.3$ (c 0.19, CHCl_3) 99% ee for *S* enantiomer]; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 6.96 (t, $J = 7.20$ Hz, 2H), 6.60 (t, $J = 7.35$ Hz, 1H), 6.49 (d, $J = 7.80$ Hz, 1H), 3.62-3.53 (m, 2H), 2.89-2.83 (m, 1H), 2.78-2.71 (m, 1H), 1.88-1.82 (m, 1H), 1.77-1.57 (m, 2H), 1.33-1.28 (m, 6H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 144.70, 129.35, 126.81, 121.01, 116.80, 114.54, 72.08, 48.91, 48.50, 32.92, 29.88, 27.90, 26.69; The enantiomeric excess was determined by HPLC on Chiralcel OD-H column (hexane : isopropanol = 95 : 5, flowing rate = 0.8 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{\text{R1}} = 13.0$ min (major), $t_{\text{R2}} = 15.7$ min (minor).

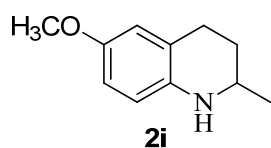


(S)-2,6-Dimethyl-1,2,3,4-tetrahydroquinoline (2h). (Known

compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He,

Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*,

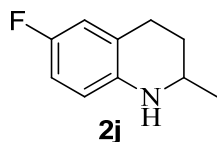
9878-9891). Isolated yield 94%, 99% ee, $[\alpha]_{\text{D}}^{25} = -81.4$ (c 0.50, CHCl_3), [Lit.^[1] $[\alpha]_{\text{D}}^{\text{RT}} = +80.3$ (c 0.19, CHCl_3) 98% ee for *R* enantiomer]; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 6.81 (d, $J = 6.60$ Hz, 2H), 6.44 (d, $J = 8.40$ Hz, 1H), 3.59 (b, 1H), 3.42-3.36 (m, 1H), 2.91-2.68 (m, 2H), 2.24 (s, 3H), 1.99-1.90 (m, 1H), 1.67-1.56 (m, 1H), 1.23 (d, $J = 6.30$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 142.55, 129.93, 127.32, 126.34, 121.32, 114.35, 47.42, 30.46, 26.70, 22.71, 20.52; The enantiomeric excess was determined by HPLC on Chiralcel OJ-H column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{\text{R}1} = 12.1$ min (major), $t_{\text{R}2} = 15.0$ min (minor).



(S)- 6-Methoxy-2-methyl-1,2,3,4-tetrahydroquinoline (2i).

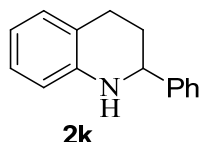
(Known compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z.

Ding, Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 94%, 99% ee, $[\alpha]_{\text{D}}^{25} = -81.0$ (c 0.50, CHCl_3), [Lit.^[1] $[\alpha]_{\text{D}}^{\text{RT}} = +80.3$ (c 0.19, CHCl_3) 99% ee for *R* enantiomer]; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 6.61 (t, $J = 8.55$ Hz, 2H), 6.45 (d, $J = 8.40$ Hz, 1H), 3.73 (s, 3H), 3.49-3.28 (m, 2H), 2.91-2.67 (m, 2H), 1.96-1.88 (m, 1H), 1.65-1.51 (m, 1H), 1.20 (d, $J = 6.30$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 151.98, 139.04, 122.64, 115.44, 114.76, 112.97, 55.93, 47.61, 30.45, 27.05, 22.69; The enantiomeric excess was determined by HPLC on Chiralcel OJ-H column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{\text{R}1} = 8.2$ min (minor), $t_{\text{R}2} = 9.0$ min (major).



(S)-6-Fluoro-2-methyl-1,2,3,4-tetrahydroquinoline (2j). (Known

compound, see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 94%, 99% ee, $[\alpha]_D^{25} = -81.2$ (*c* 0.50, CHCl₃), [Lit.^[1] $[\alpha]_D^{RT} = +80.3$ (*c* 0.19, CHCl₃) 98% ee for *R* enantiomer]; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 6.71-6.65 (m, 2H), 6.43-6.38 (m, 1H), 3.54 (b, 1H), 3.40-3.31 (m, 1H), 2.89-2.66 (m, 2H), 1.96-1.88 (m, 1H), 1.63-1.50 (m, 1H), 1.21 (d, *J* = 6.30 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 154.07, 141.09, 122.64, 122.55, 115.65, 115.37, 114.88, 114.78, 113.42, 113.13, 47.43, 30.01, 26.84, 22.62; The enantiomeric excess was determined by HPLC on Chiralcel OD-H column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{R1} = 5.2$ min (minor), $t_{R2} = 5.9$ min (major).

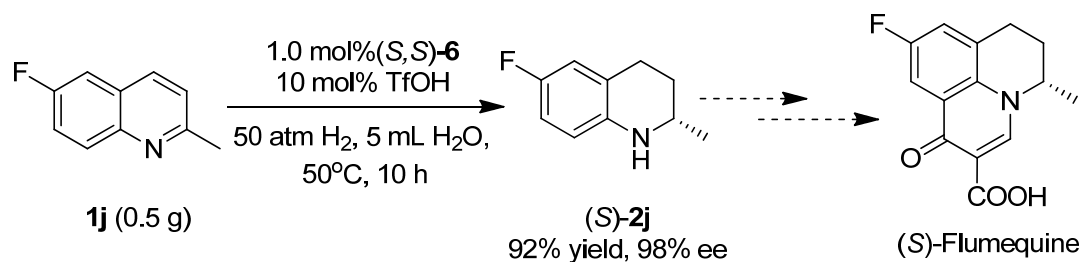


(S)-2-Phenyl-1,2,3,4-tetrahydroquinoline (2k). (Known compound,

see: T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891). Isolated yield 92%, 63% ee, $[\alpha]_D^{25} = -35.6$ (*c* 0.50, CHCl₃), [Lit.^[1] $[\alpha]_D^{RT} = +36.8$ (*c* 0.95, CHCl₃) 92% ee for *R* enantiomer]; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.44-7.32 (m, 5H), 7.05 (t, *J* = 6.90 Hz, 2H), 6.69 (t, *J* = 7.35 Hz, 1H), 6.57 (d, *J* = 8.10 Hz, 1H), 4.49-4.45 (m, 1H), 4.06 (b, 1H), 2.96-2.91 (m, 1H), 2.81-2.73 (m, 1H), 2.19-2.01 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 144.93, 144.84, 129.41, 128.69, 127.55, 127.02, 126.66, 120.98, 117.26, 114.09, 56.36, 31.10, 26.50; The enantiomeric excess was determined by HPLC on Chiralcel OD-H column (hexane : isopropanol = 90 : 10,

flowing rate = 0.6 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{R1} = 15.4$ min (major),
 $t_{R2} = 20.8$ min (minor).

4. Scaled-up synthesis of 6-fluoro-2-methyl-1,2,3,4-tetrahydroquinoline



Quinoline **1j** (0.5 g, 3.10 mmol), (*S,S*)-**6** (0.031 mmol, 1.0 mol%), undegassed water (5 mL) and undegassed CF₃SO₃H (46.5 mg, 0.31 mmol, 10 mol%) were added to a glass tube in air. Then, the tube was placed into a 30 mL stainless steel autoclave and the autoclave was purged with hydrogen gas for 3 times before pressurized with H₂ to 50 atm. The hydrogenation was performed at 50 °C for 10 h. After careful releasing of the hydrogen, the reaction mixture was diluted with ethyl acetate (3 mL) and saturated sodium carbonate aqueous solution (2 mL), then stirred for 2 min. The aqueous layer was extracted with ethyl acetate (3 x 3 mL). The combined organic layer was dried over anhydrous sodium sulfate and concentrated to afford the crude product. The full conversion of the reaction was based on ¹H NMR spectra. Purification was performed by a silica gel column eluted with petroleum ether/triethylamine (95:5, v/v) to give the pure product as a yellow solid (460 mg, 92% yield, 98% ee). The enantiomeric excess was determined by HPLC on Chiralcel OD-H column (hexane : isopropanol = 90 : 10, flowing rate = 1.0 mL/min, 25 °C, UV detection at $\lambda = 254$ nm) $t_{R1} = 5.2$ min (minor), $t_{R2} = 5.9$ min (major).

5. The determination of substrate solubility in water

5.1 The determination of solubilities for quinolines **1a and **1j**:**

2-Methylquinoline (**1a**) (27 μ L, 0.2 mmol) and D₂O (1 mL) were added to a glass tube and stirred for 30 min at 20 °C. Then, the reaction mixture was standed overnight. After that, 0.5 mL clear solution was carefully transferred into a NMR tube, then 1.12 mg isopropanol was added into the NMR tube as an internal standard. The solubility of **1a** (4.04 mg/mL) was obtained based on its ¹H NMR spectrum.

The solubility of 6-fluoro-2-methylquinoline (**1j**) (0.84 mg/mL) was obtained with the same method described above.

5.2 The determination of solubilities for TfOH protonated quinoline salts:

The TfOH protonated 2-methylquinoline salt was added into 0.2 mL deionized water in portions until the salt was undissolved and the solubility was obtained as 5.68 g/mL.

The solubility of TfOH protonated 6-fluoro-2-methylquinoline salt (2.12 g/mL) was obtained with the same method described above.

5.3 The difference between oil substrate 1a and solid substrate 1j:

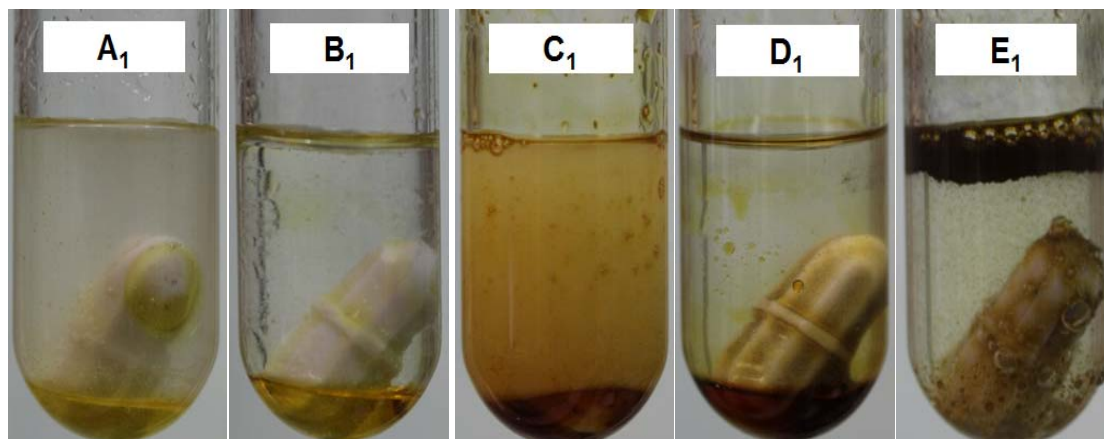


Figure 1. Unless noted otherwise, 500 mg 2-methylquinoline in 5 mL deionized water. A₁: stirred at room temperature; B₁: added 10 mol% CF₃SO₃H; C₁: added 10 mol% TfOH and 1 mol% catalyst; D₁: heated in an oil bath at 50 °C for 0.5 h; E₁: reacted with 50 atm hydrogen at 50 °C for 6 h.

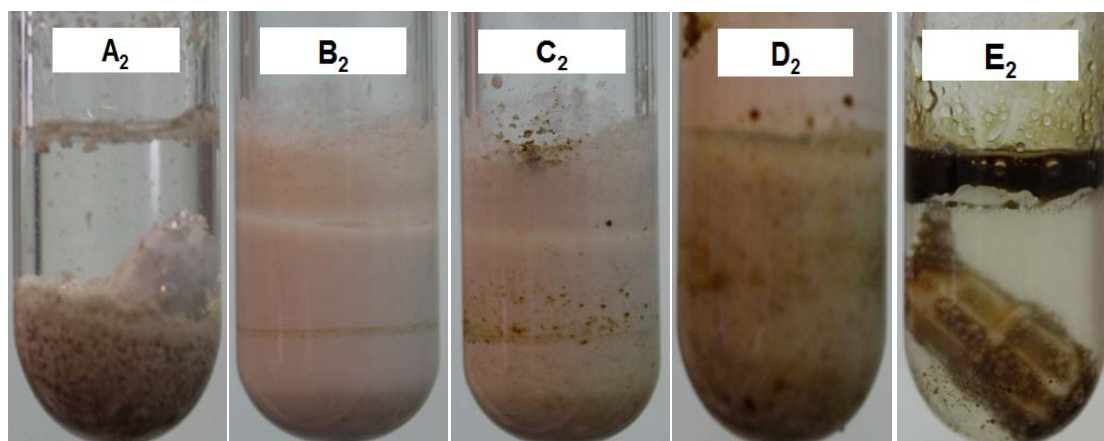
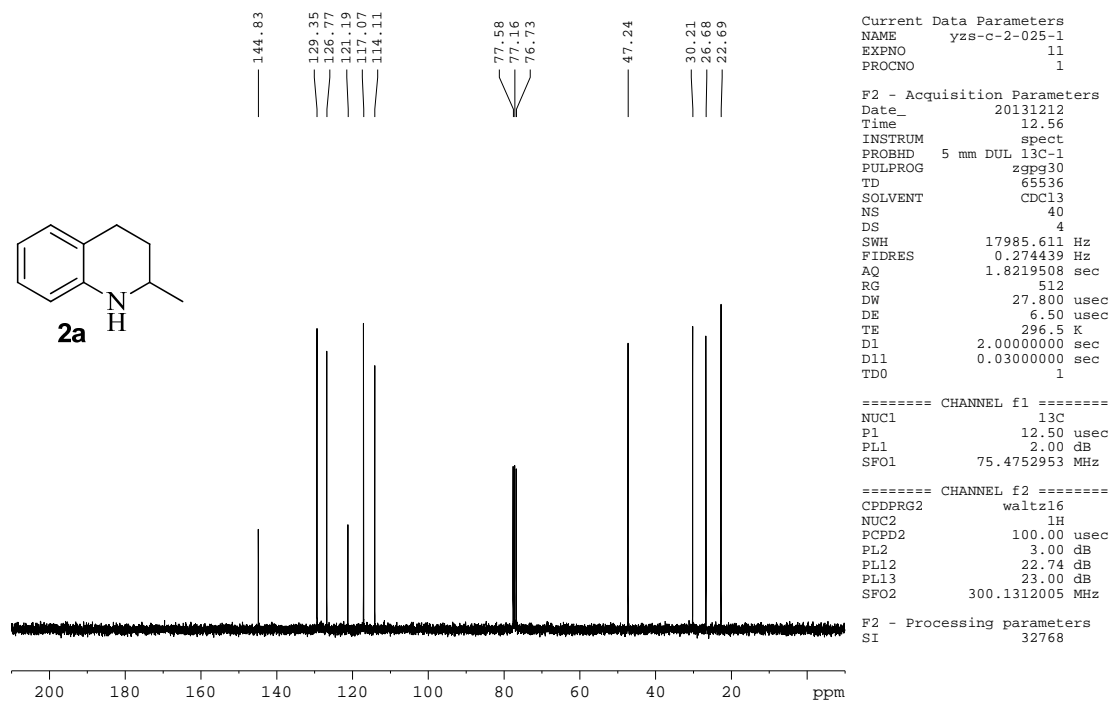
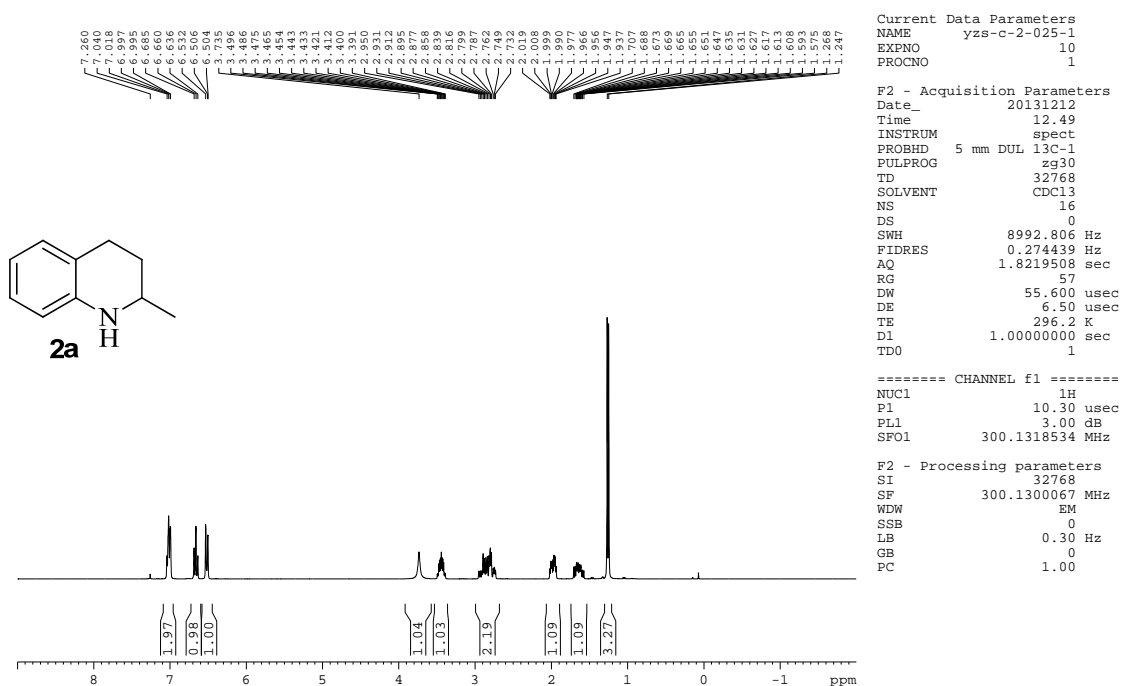


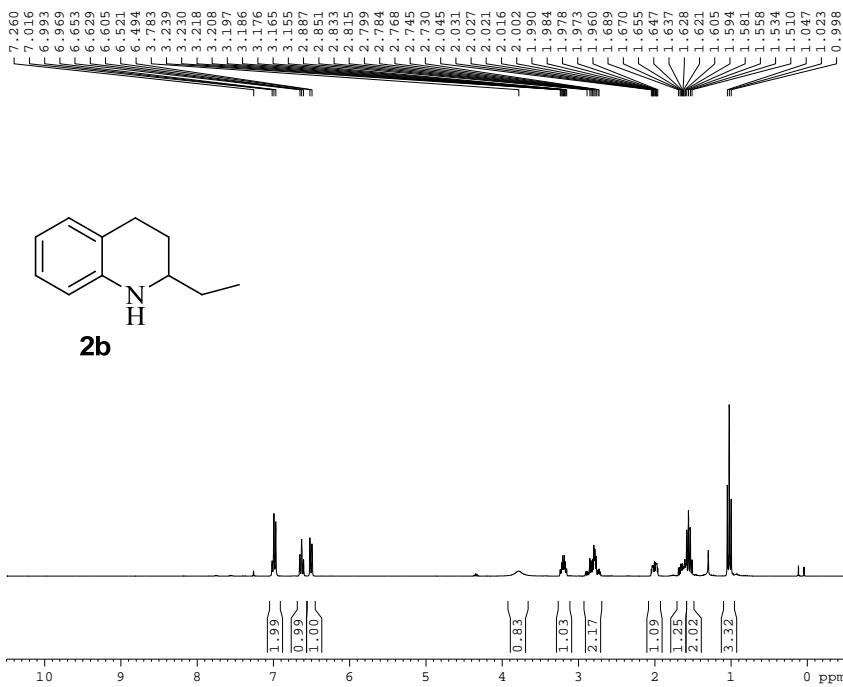
Figure 2. Unless noted otherwise, 500 mg 6-fluoro-2-methylquinoline in 5 mL deionized water. A₂: stirred at room temperature. B₂: added 10 mol% TfOH; C₂: added 1 mol% catalyst; D₂: heated in an oil bath at 50 °C for 0.5 h; E₂: reacted with 50 atm hydrogen at 50 °C for 6 h.

6. Reference

- [1] T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He, Q.-H. Fan, J. Xiang, Z.-X. Yu, A. S. C. Chan, *J. Am. Chem. Soc.* **2011**, *133*, 9878-9891.

7. ¹H and ¹³C NMR of products



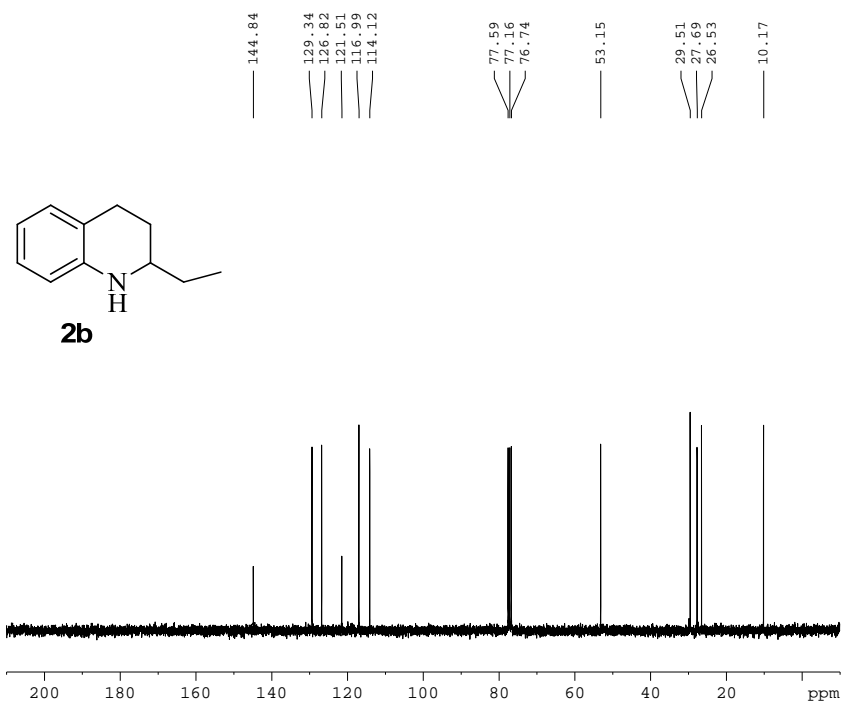


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 PULPROG zg30
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 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 71.8
 DW 55.600 usec
 DE 6.50 usec
 TE 297.4 K
 D1 1.0000000 sec
 TD0 1

==== CHANNEL f1 =====
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 P1 10.30 usec
 PL1 3.00 dB
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F2 - Processing parameters
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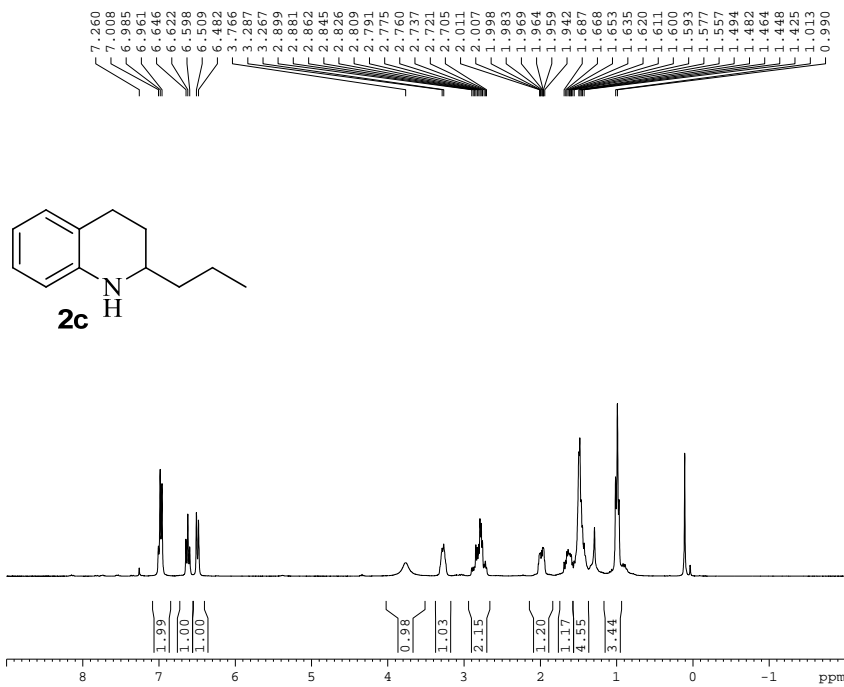
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 SOLVENT CDCl3
 NS 55
 DS 4
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 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 1625.5
 DW 27.800 usec
 DE 6.50 usec
 TE 297.4 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768

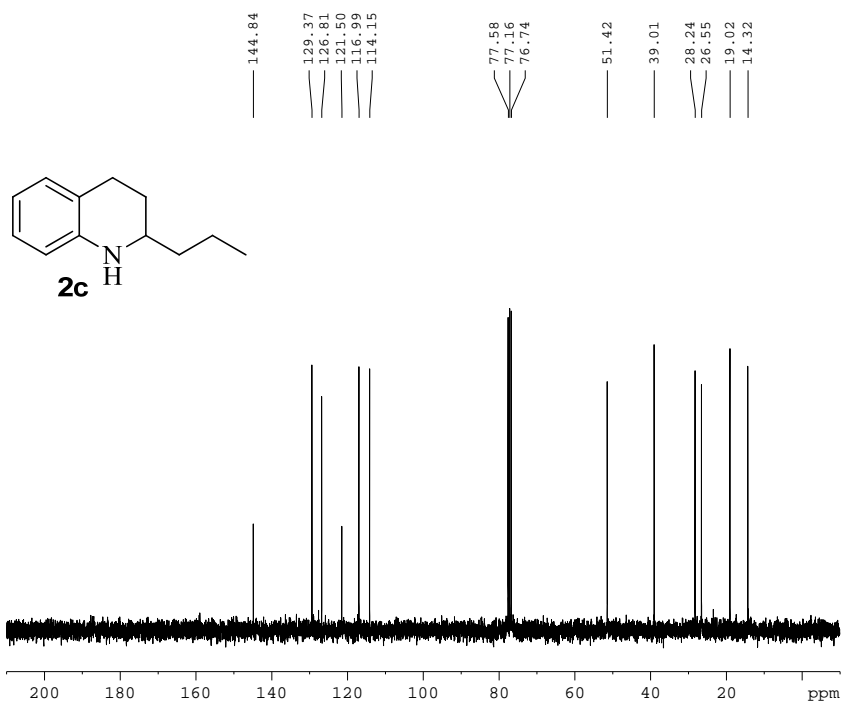


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 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 71.8
 DW 55.600 usec
 DE 6.50 usec
 TE 295.3 K
 D1 1.00000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
 SFO1 300.1318534 MHz

F2 - Processing parameters
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 GB 0
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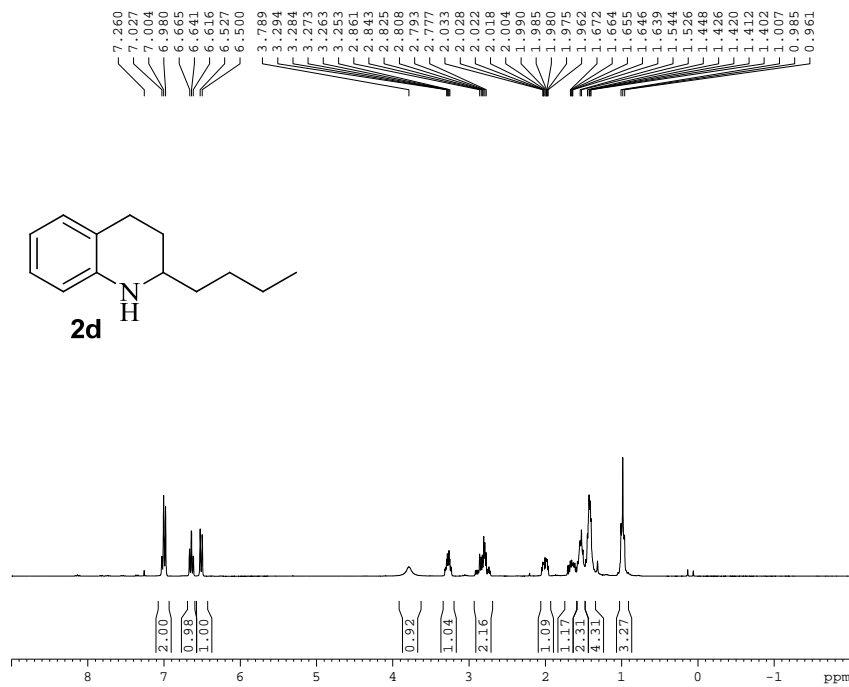
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 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 45
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 1625.5
 DW 27.800 usec
 DE 6.50 usec
 TE 297.7 K
 D1 2.00000000 sec
 D11 0.03000000 sec
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==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768

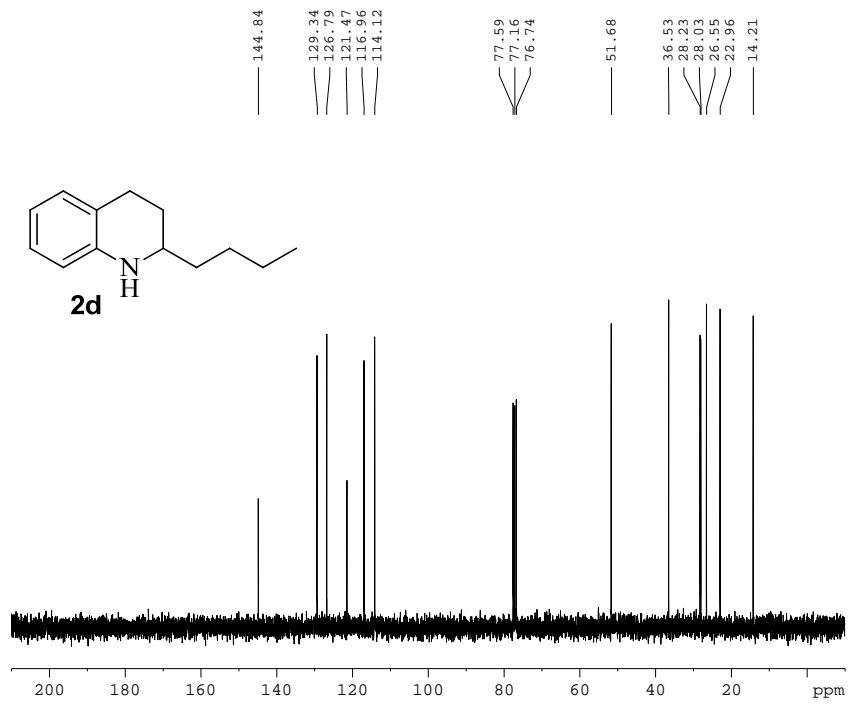


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 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 40.3
 DW 55.600 usec
 DE 6.50 usec
 TE 296.9 K
 D1 1.00000000 sec
 TDO 1

==== CHANNEL f1 =====
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 P1 10.30 usec
 PL1 3.00 dB
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F2 - Processing parameters
 SI 32768
 SF 300.1300067 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



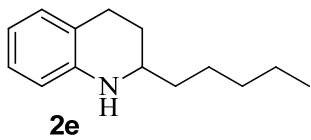
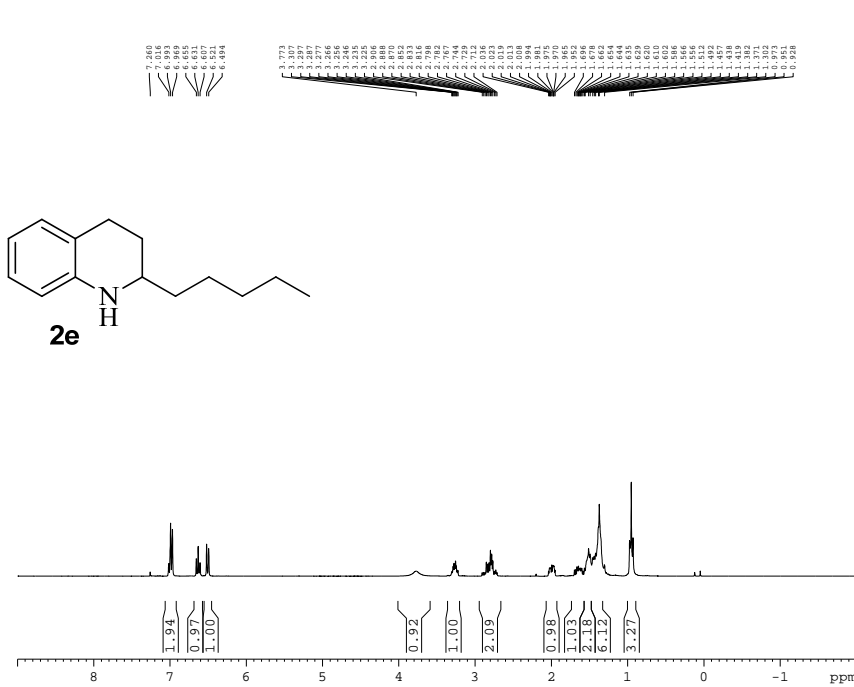
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 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 15
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 574.7
 DW 27.800 usec
 DE 6.50 usec
 TE 296.9 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768

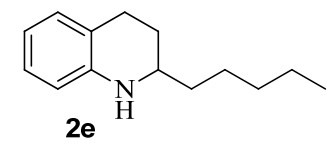
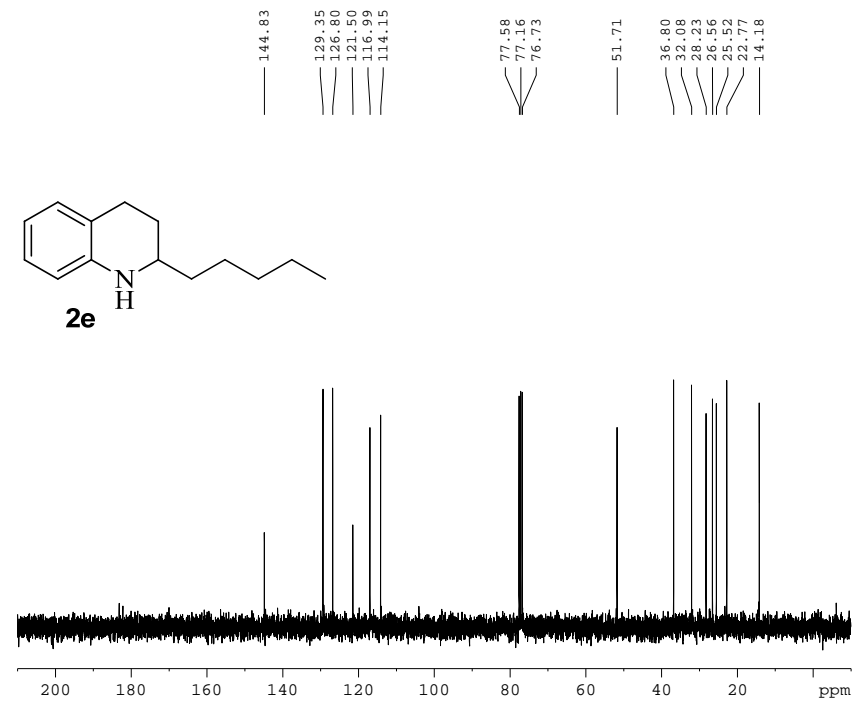


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 PROCNO 1

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 INSTRUM spect
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 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 57
 DW 55.600 usec
 DE 6.50 usec
 TE 296.8 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300067 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



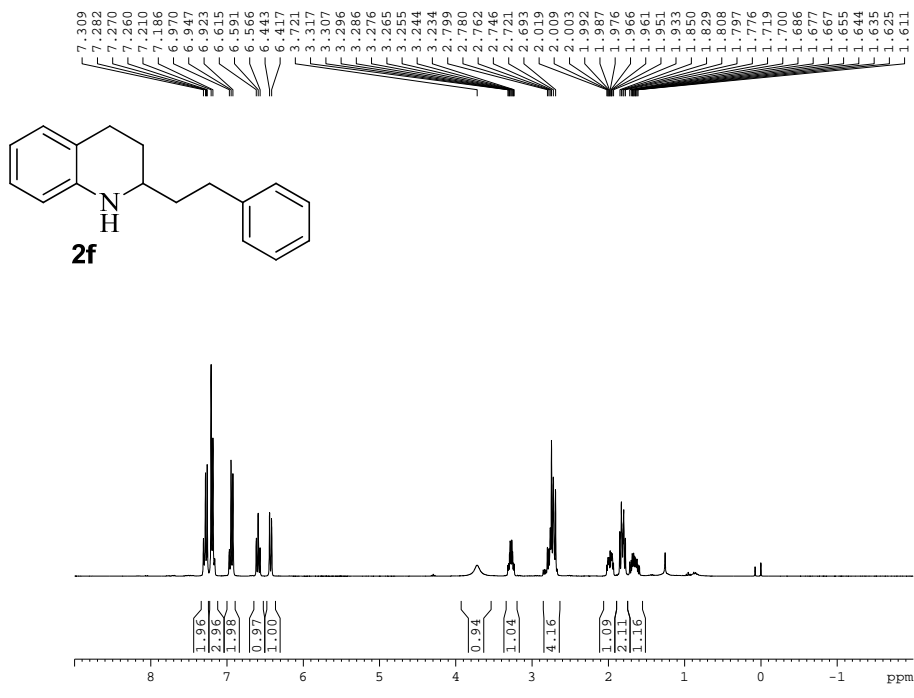
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 SOLVENT CDCl3
 NS 19
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 1149.4
 DW 27.800 usec
 DE 6.50 usec
 TE 296.9 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768

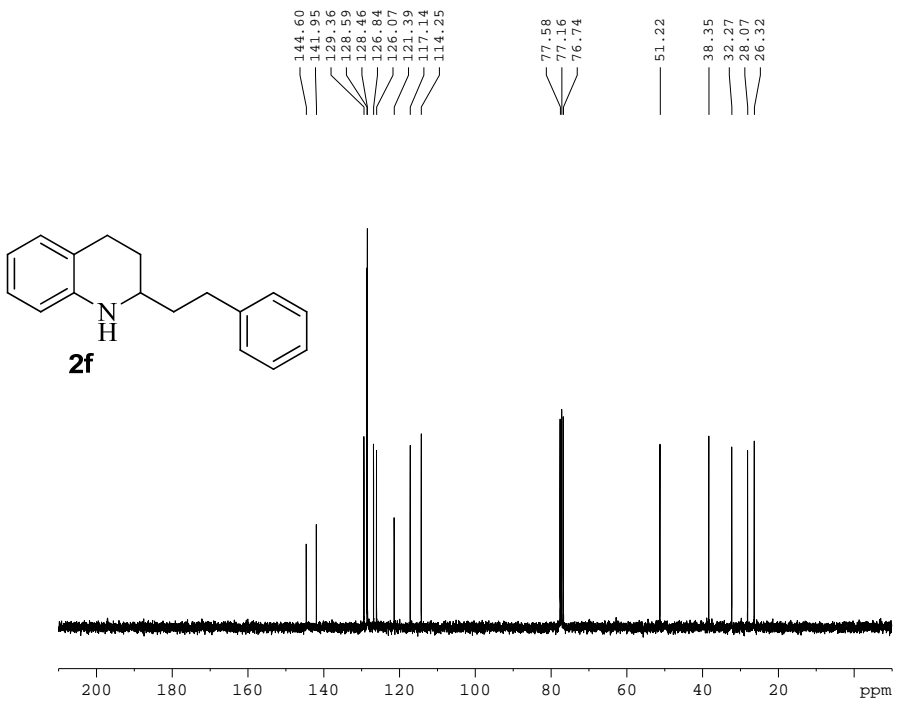


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 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 64
 DW 55.600 usec
 DE 6.50 usec
 TE 296.9 K
 D1 1.00000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
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F2 - Processing parameters
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 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



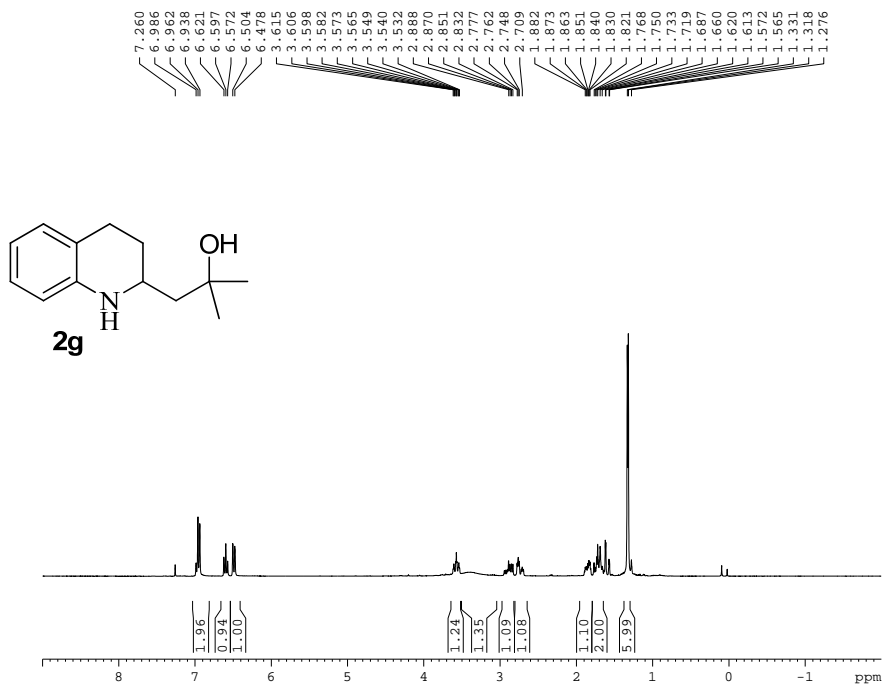
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 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 45
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 512
 DW 27.800 usec
 DE 6.50 usec
 TE 297.2 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768

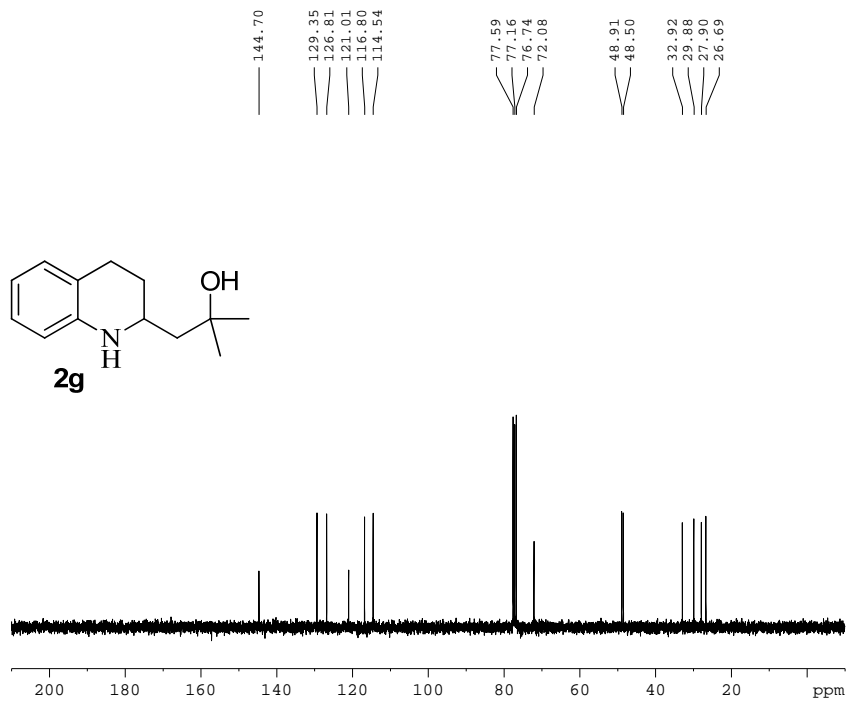


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 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 71.8
 DW 55.600 usec
 DE 6.50 usec
 TE 295.2 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
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F2 - Processing parameters
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 SSB 0
 LB 0.30 Hz
 GB 0
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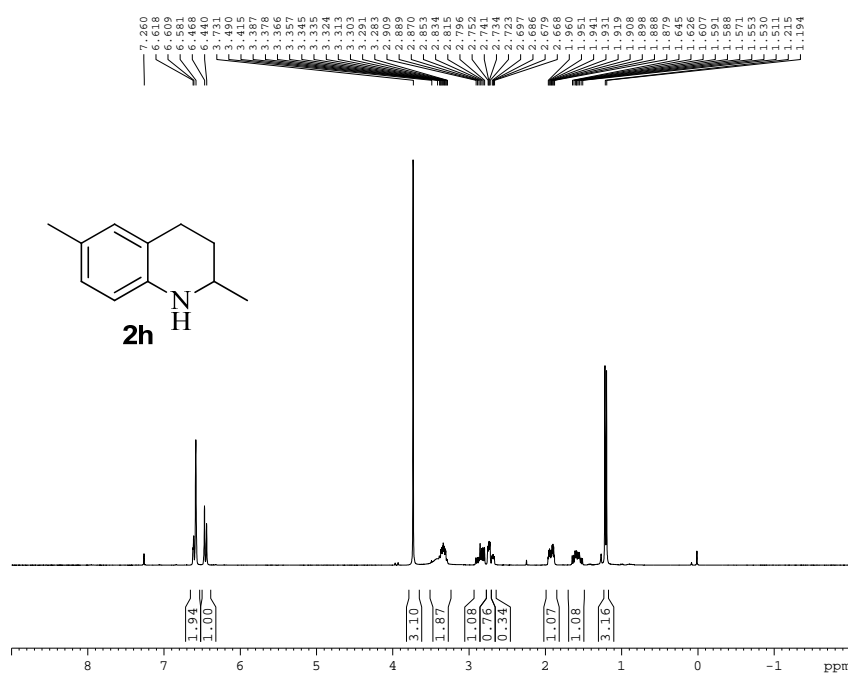
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 SOLVENT CDCl3
 NS 40
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 362
 DW 27.800 usec
 DE 6.50 usec
 TE 295.6 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
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 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768

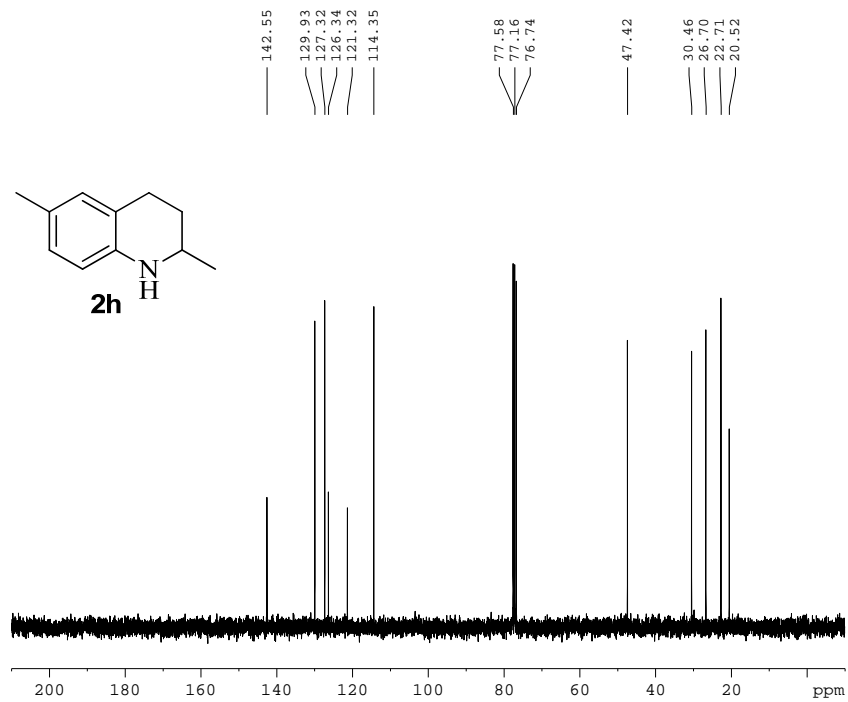


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 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 181
 DW 55.600 usec
 DE 6.50 usec
 TE 296.8 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
 SF01 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300067 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



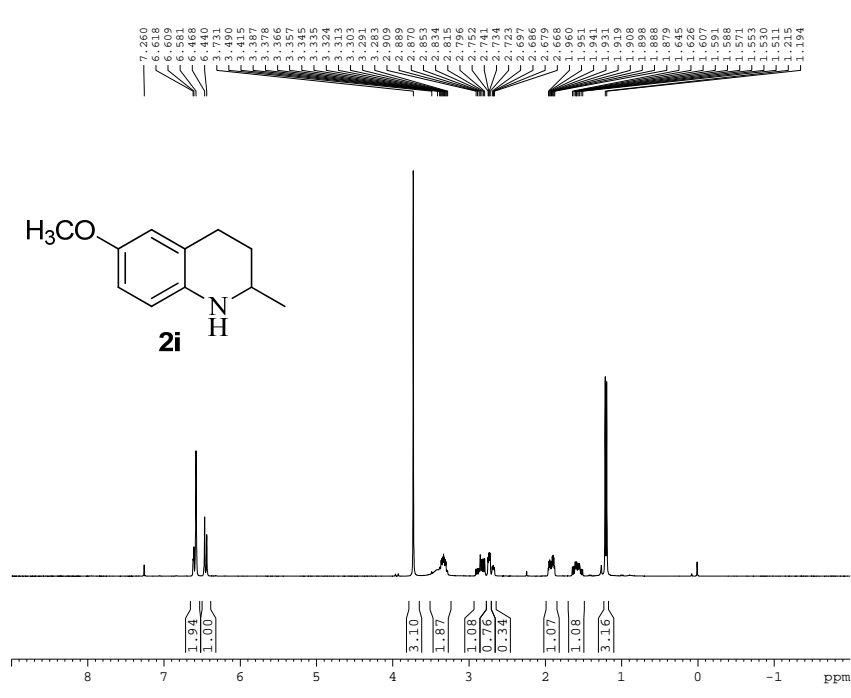
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 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 512
 DW 27.800 usec
 DE 6.50 usec
 TE 296.8 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SF01 75.4752953 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
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 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SF02 300.1312005 MHz

F2 - Processing parameters
 SI 32768

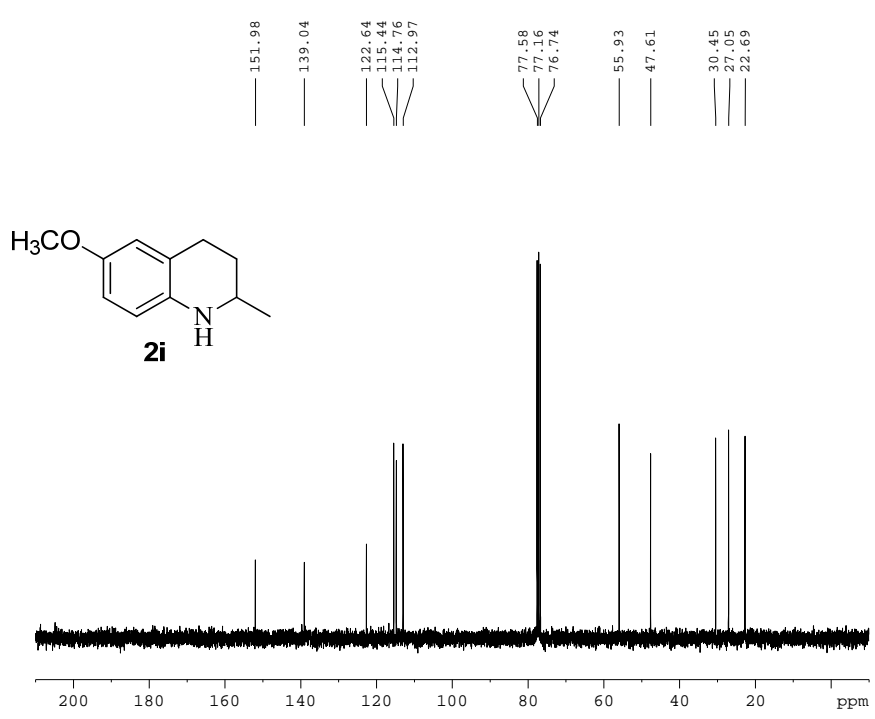


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 SOLVENT CDCl3
 NS 16
 DS 0
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 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 181
 DW 55.600 usec
 DE 6.50 usec
 TE 296.8 K
 D1 1.0000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300067 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



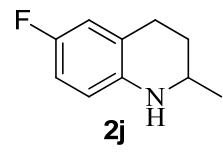
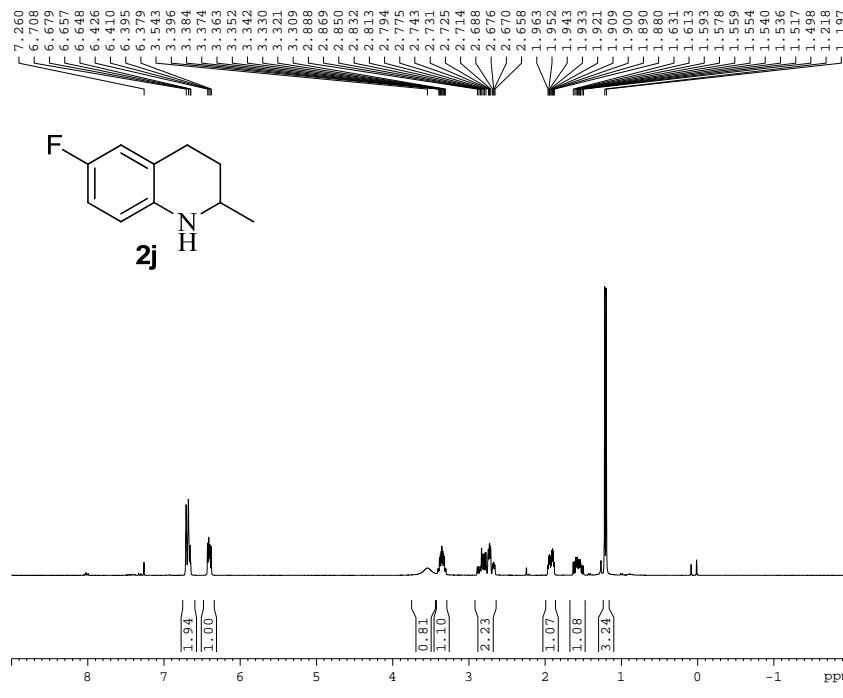
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 AQ 1.8219508 sec
 RG 512
 DW 27.800 usec
 DE 6.50 usec
 TE 296.9 K
 D1 2.0000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768

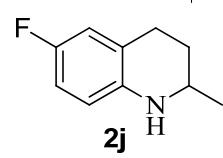
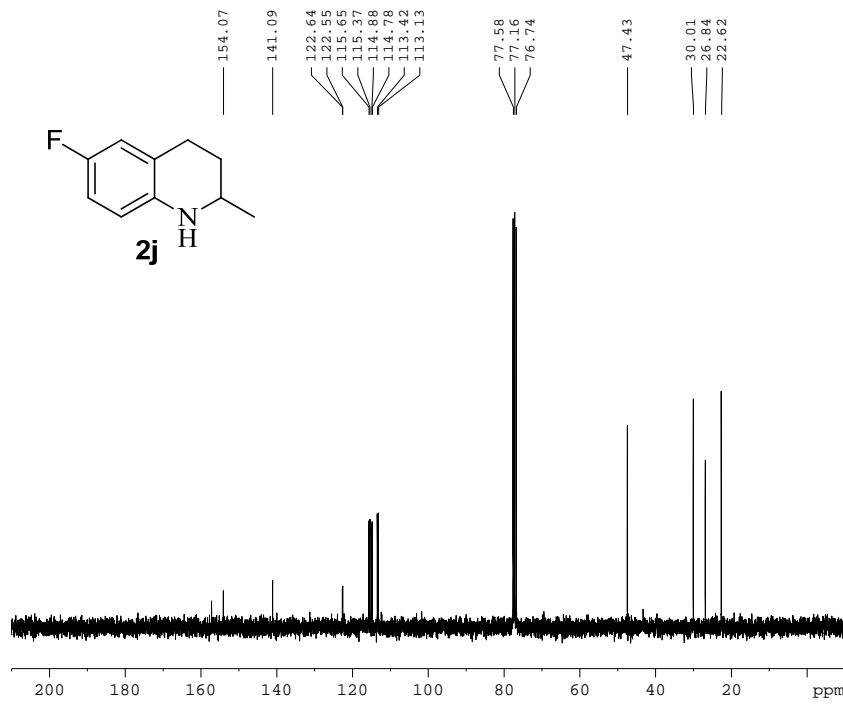


Current Data Parameters
 NAME YZS-C-2-026-2
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20131211
 Time 21.41
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 181
 DW 55.600 usec
 DE 6.50 usec
 TE 296.8 K
 D1 1.00000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300067 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



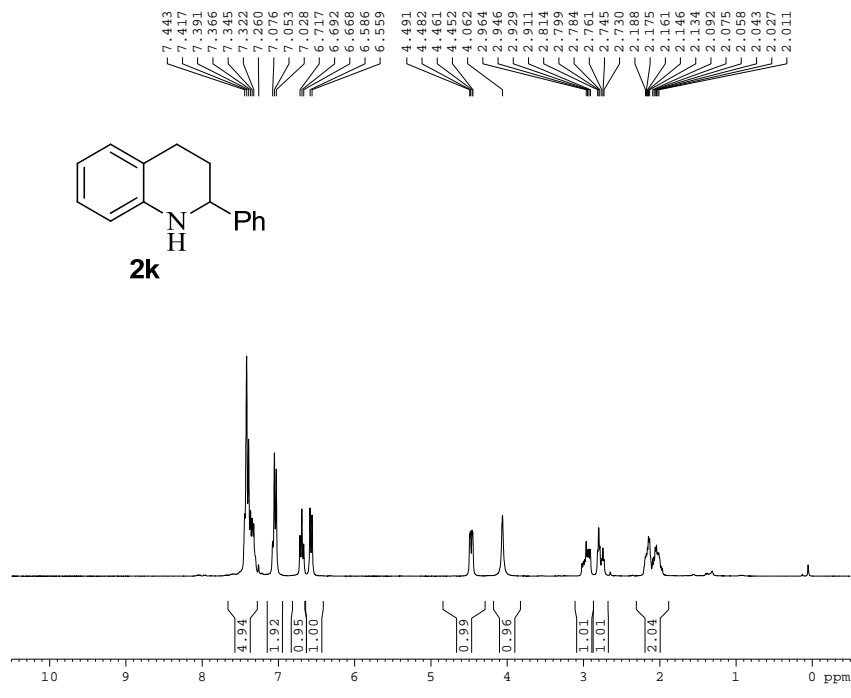
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 NAME YZS-C-2-026-2
 EXPNO 21
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20131211
 Time 21.55
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 92
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 1149.4
 DW 27.800 usec
 DE 6.50 usec
 TE 297.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768

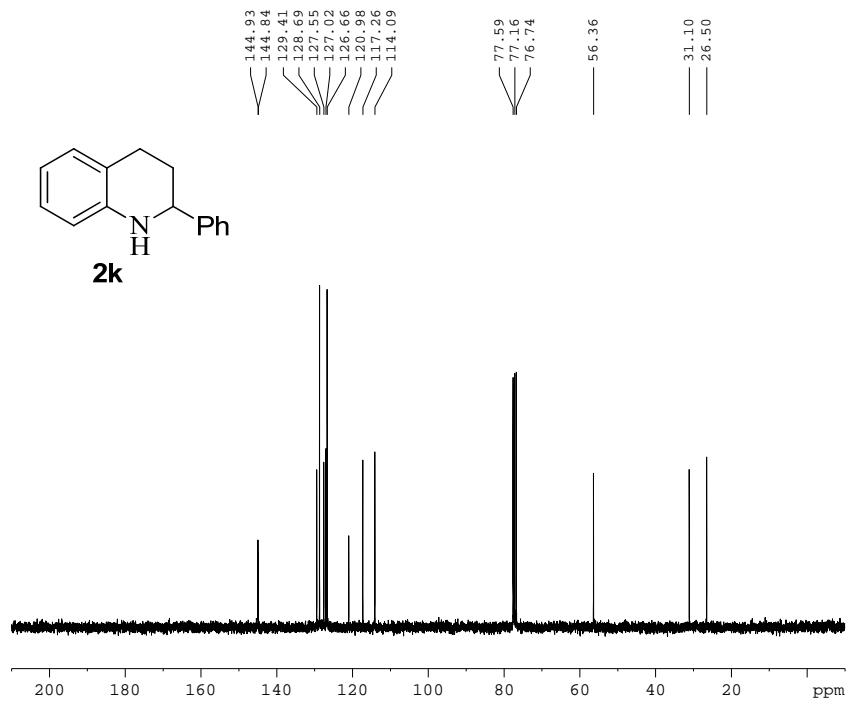


Current Data Parameters
 NAME yzs-c-2-ph-r
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20140121
 Time 14.35
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 128
 DW 55.600 usec
 DE 6.50 usec
 TE 295.9 K
 D1 1.00000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300070 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME yzs-c-2-ph-r
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20140121
 Time 14.45
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 103
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 2580.3
 DW 27.800 usec
 DE 6.50 usec
 TE 296.5 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

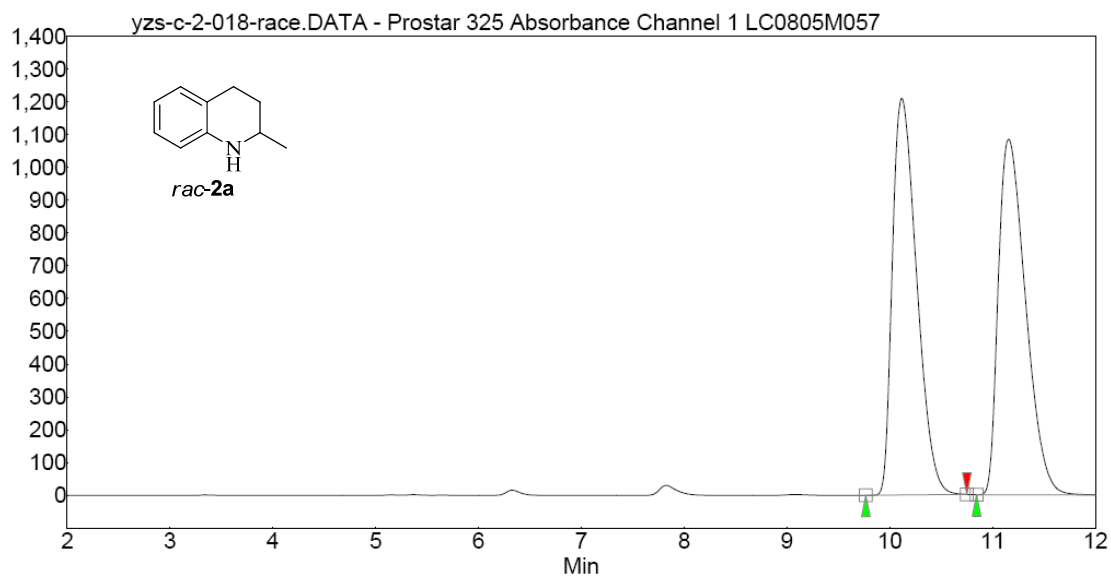
==== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

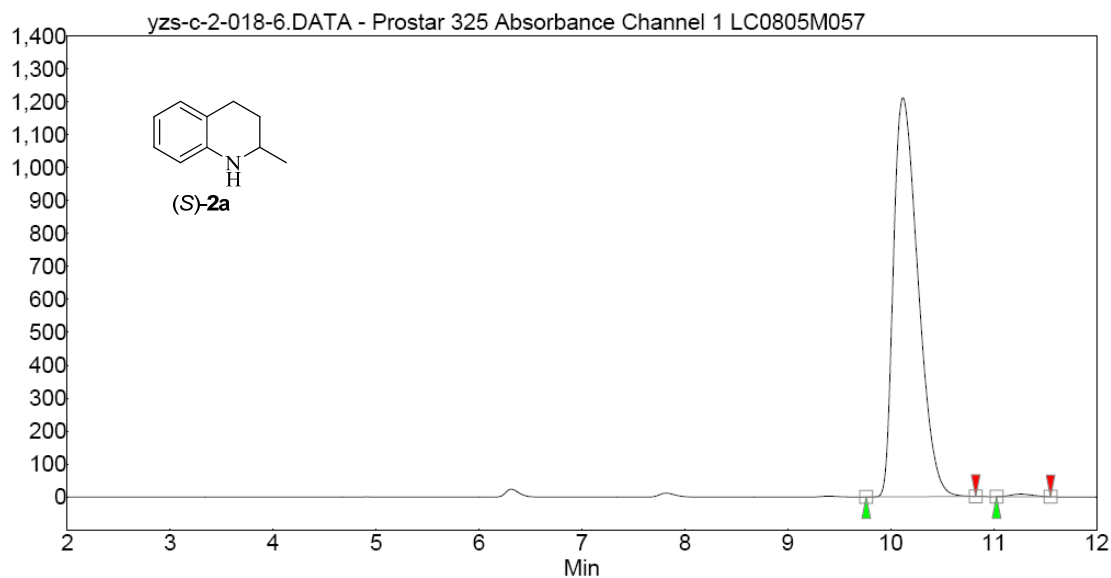
F2 - Processing parameters
 SI 32768

8. HPLC spectra of products catalyzed by (S,S)-6

7.1 HPLC spectra of 2a catalyzed by (S,S)-6

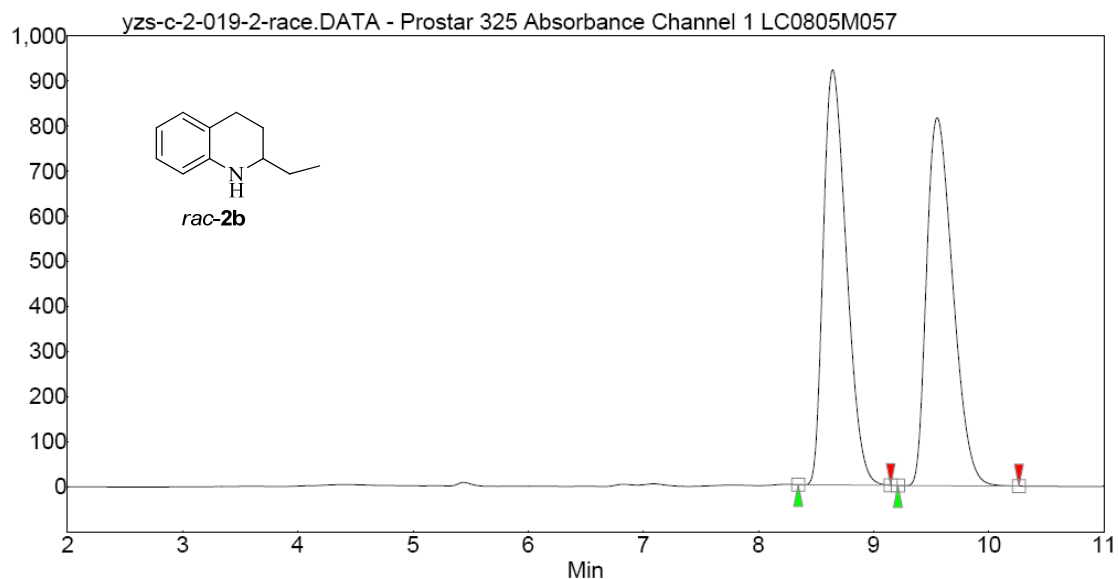


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	10.11	49.84	1208.8	331.2	49.836
2	未知	11.15	50.16	1083.9	333.4	50.164
Total			100.00	2292.7	664.6	100.000

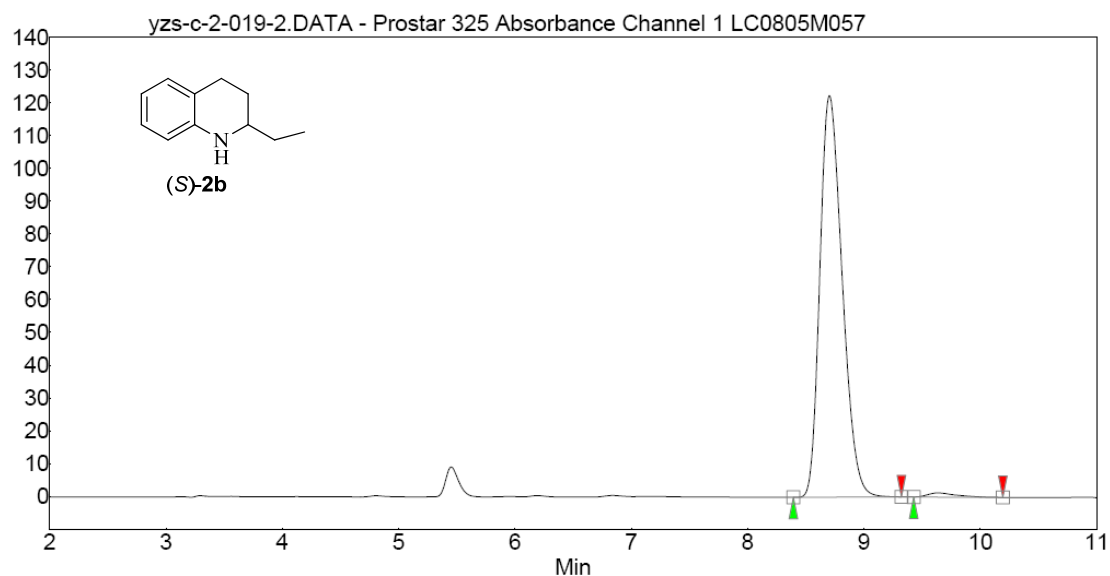


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	10.12	99.42	1211.7	334.5	99.421
2	未知	11.26	0.58	7.7	1.9	0.579
Total			100.00	1219.4	336.5	100.000

HPLC spectra of 2b catalyzed by (S,S)-6

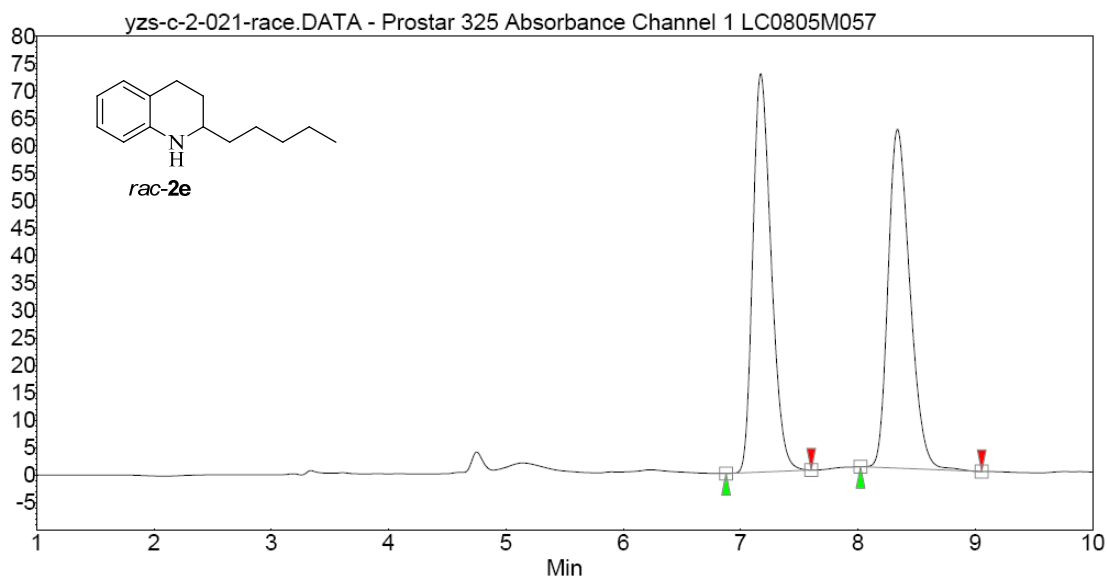


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	8.65	49.84	920.1	214.2	49.836
2	未知	9.55	50.16	816.0	215.7	50.164
Total			100.00	1736.0	429.9	100.000

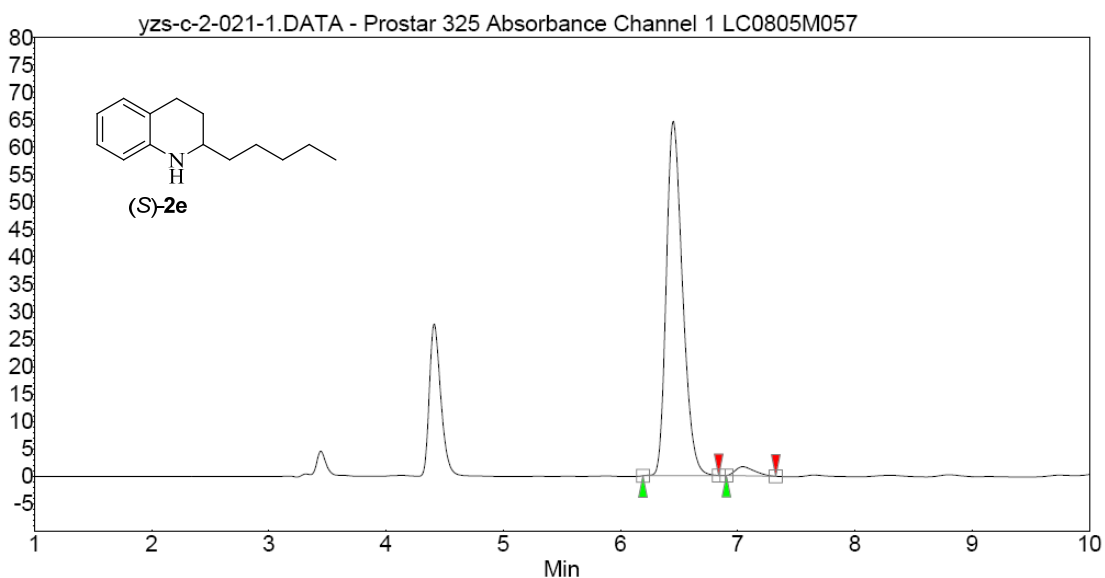


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	8.70	98.68	122.3	27.2	98.685
2	未知	9.64	1.32	1.3	0.4	1.315
Total			100.00	123.6	27.6	100.000

HPLC spectra of 2e catalyzed by (S,S)-6

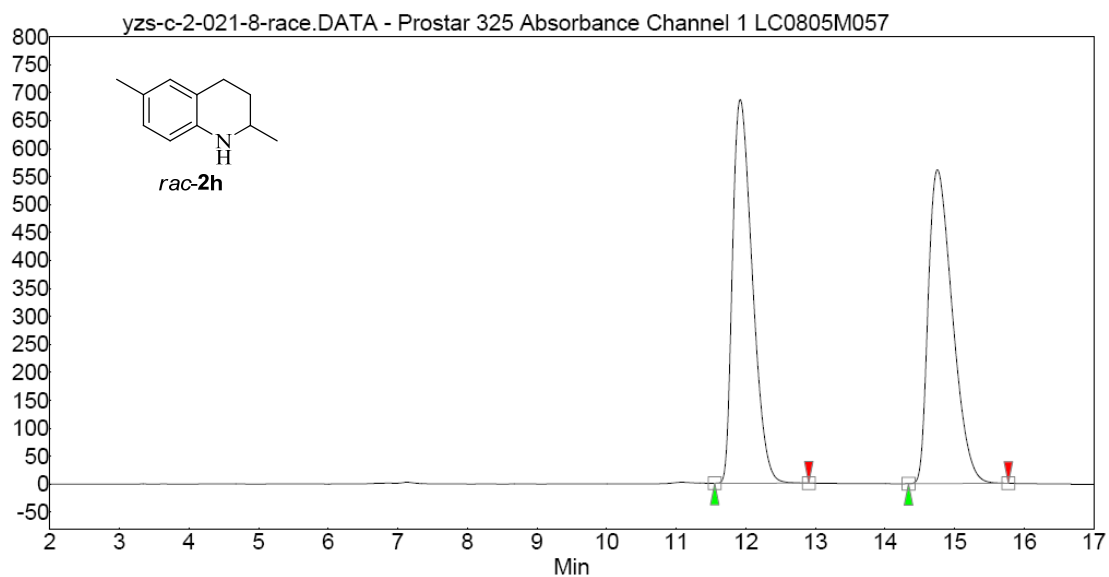


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	7.17	49.66	72.6	13.2	49.662
2	未知	8.33	50.34	61.7	13.3	50.338
Total			100.00	134.3	26.5	100.000

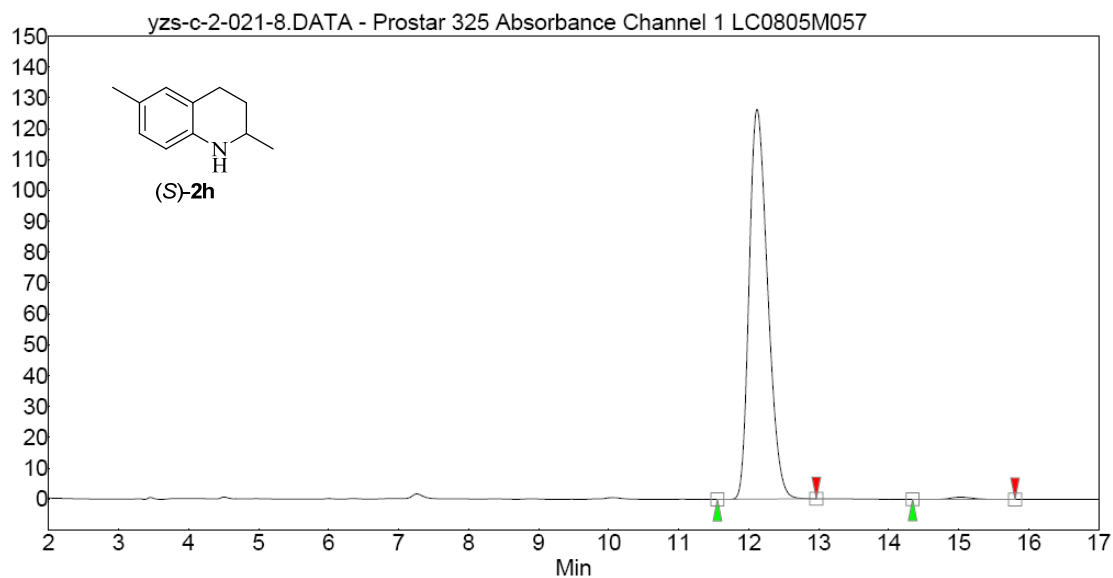


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	6.45	97.11	64.6	10.4	97.109
2	未知	7.05	2.89	1.7	0.3	2.891
Total			100.00	66.2	10.7	100.000

HPLC spectra of 2h catalyzed by (S,S)-6

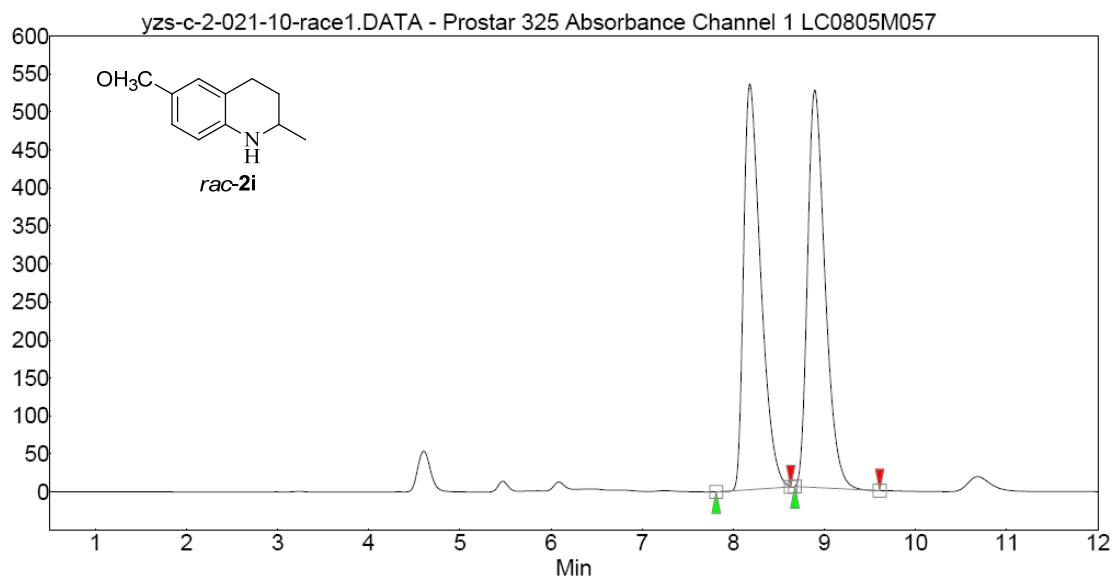


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	12.12	99.32	126.3	38.7	99.325
2	未知	15.02	0.68	0.7	0.3	0.675
Total			100.00	127.1	38.9	100.000

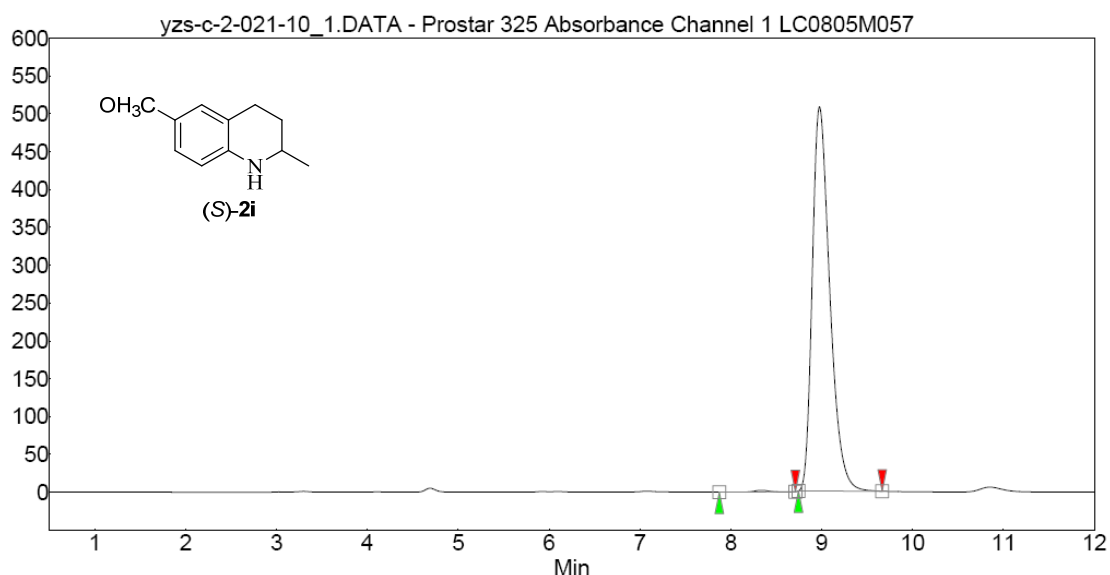


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	12.12	99.32	126.3	38.7	99.325
2	未知	15.02	0.68	0.7	0.3	0.675
Total			100.00	127.1	38.9	100.000

HPLC spectra of 2i catalyzed by (S,S)-6

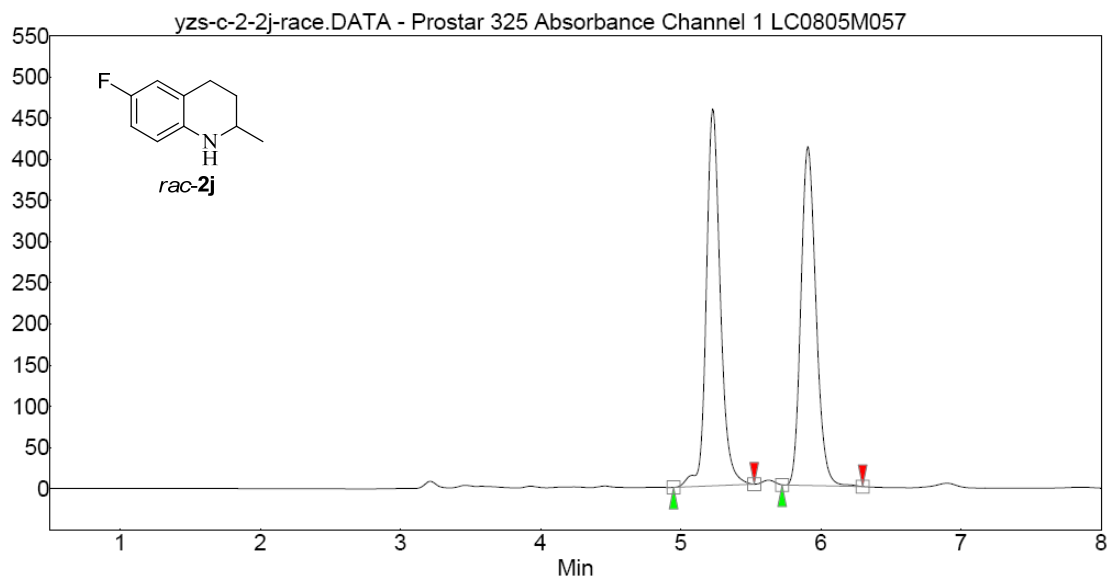


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	8.18	49.97	533.8	116.5	49.965
2	未知	8.89	50.03	523.1	116.6	50.035
Total			100.00	1056.9	233.1	100.000

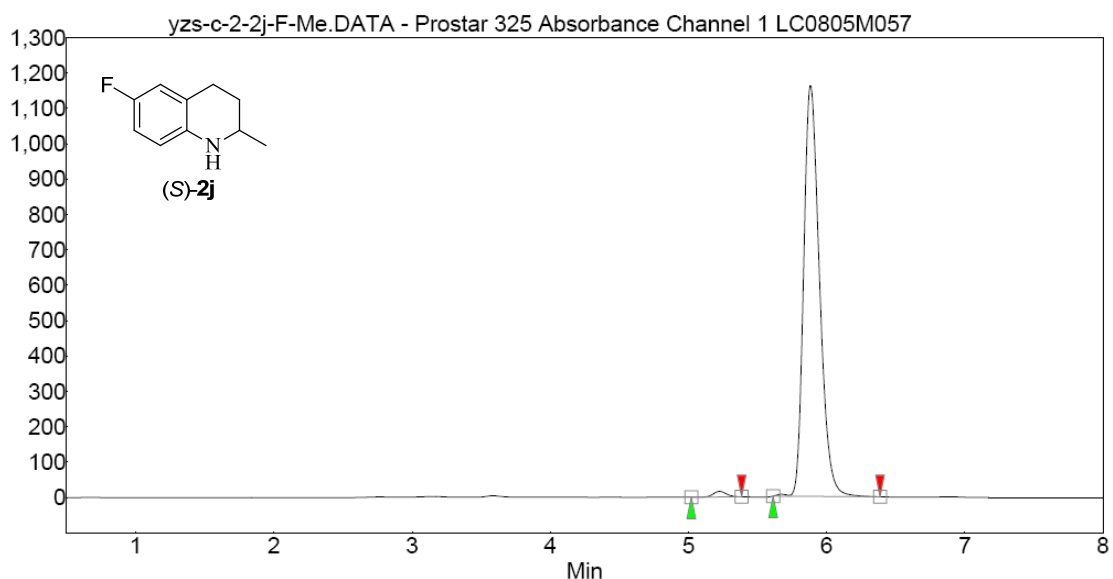


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	8.33	0.31	2.3	0.4	0.311
2	未知	8.97	99.69	507.8	113.5	99.689
Total			100.00	510.1	113.8	100.000

HPLC spectra of 2j catalyzed by (S,S)-6



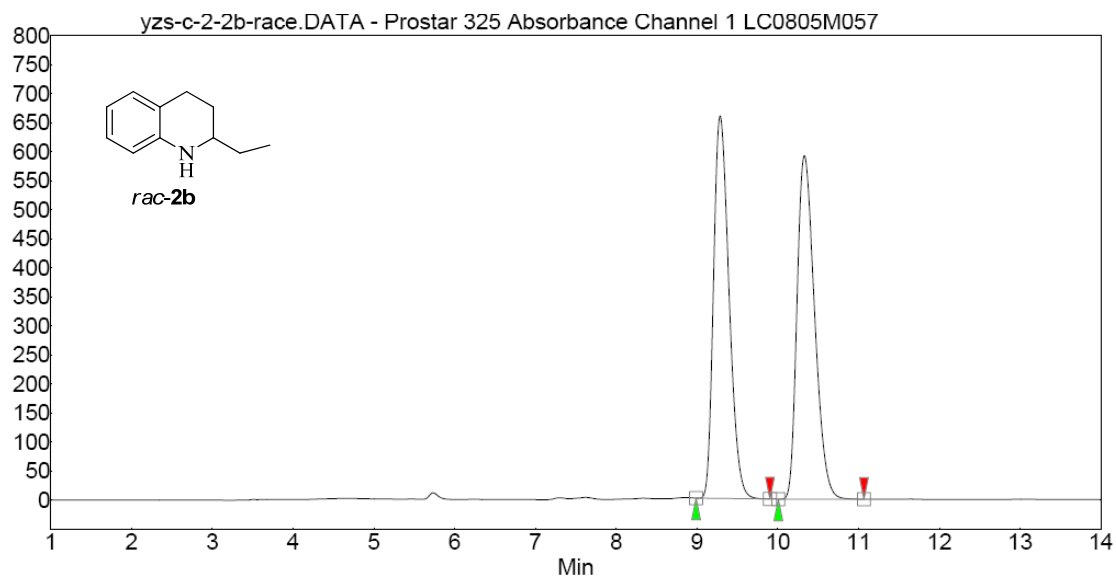
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	5.23	50.85	457.6	53.1	50.853
2	未知	5.91	49.15	411.7	51.3	49.147
Total			100.00	869.2	104.4	100.000



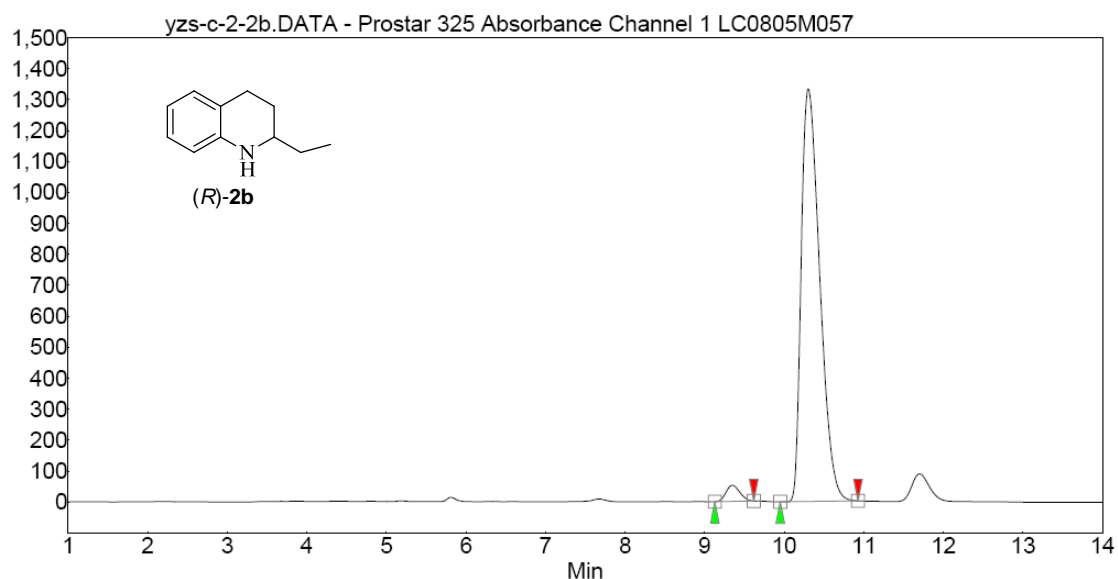
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	5.23	1.13	16.0	1.8	1.125
2	未知	5.88	98.87	1161.0	154.8	98.875
Total			100.00	1177.0	156.6	100.000

8.2 HPLC spectra of products catalyzed by (R,R)-3

HPLC spectra of 2b catalyzed by (R,R)-3

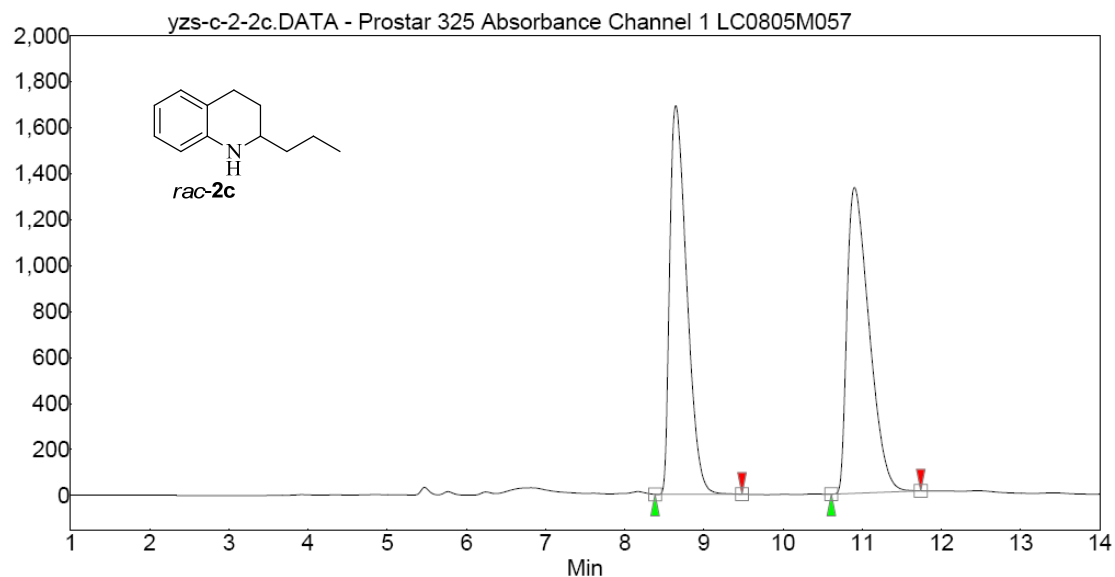


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	9.29	49.50	658.5	146.1	49.501
2	未知	10.33	50.50	591.5	149.0	50.499
Total			100.00	1249.9	295.1	100.000

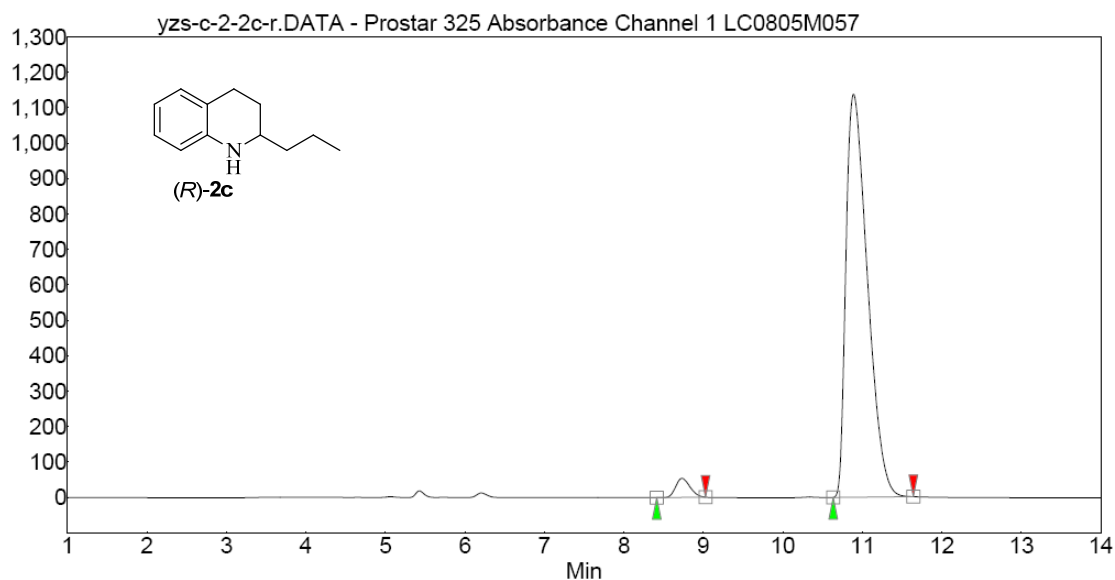


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	9.35	2.84	52.2	10.5	2.839
2	未知	10.30	97.16	1332.9	359.1	97.161
Total			100.00	1385.1	369.6	100.000

HPLC spectra of 2c catalyzed by (R,R)-3

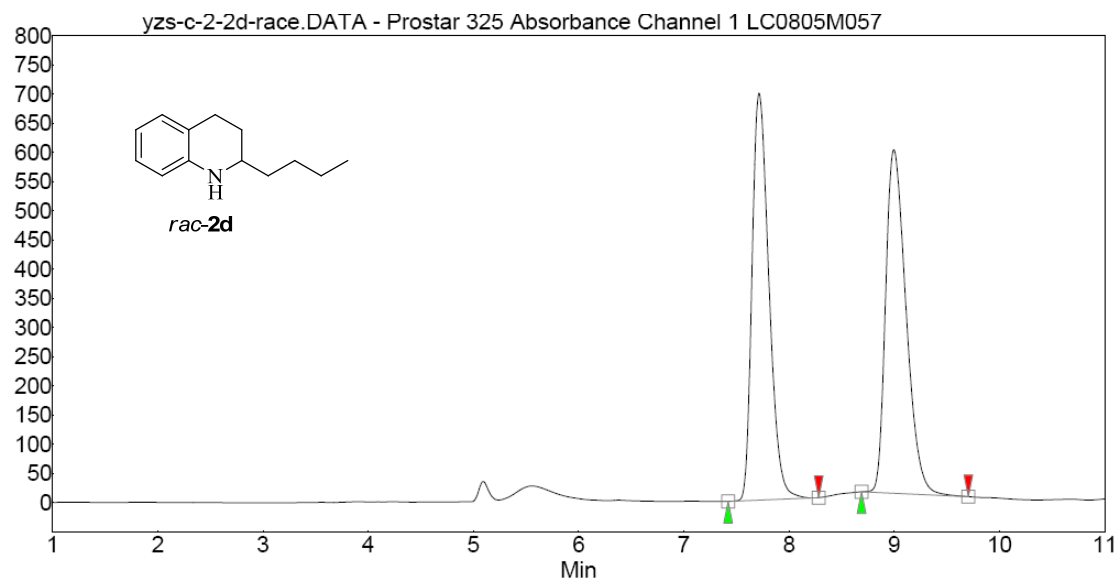


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	8.65	49.42	1690.7	421.4	49.420
2	未知	10.90	50.58	1330.2	431.3	50.580
Total			100.00	3020.8	852.8	100.000

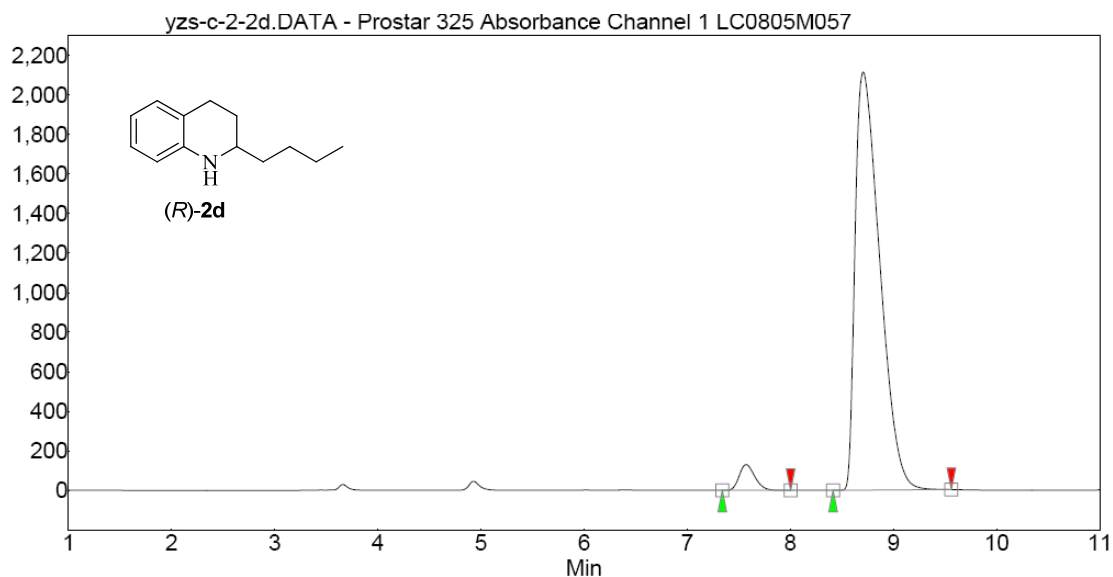


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	8.73	3.05	53.8	11.2	3.050
2	未知	10.89	96.95	1137.9	356.0	96.950
Total			100.00	1191.7	367.2	100.000

HPLC spectra of 2d catalyzed by (R,R)-3

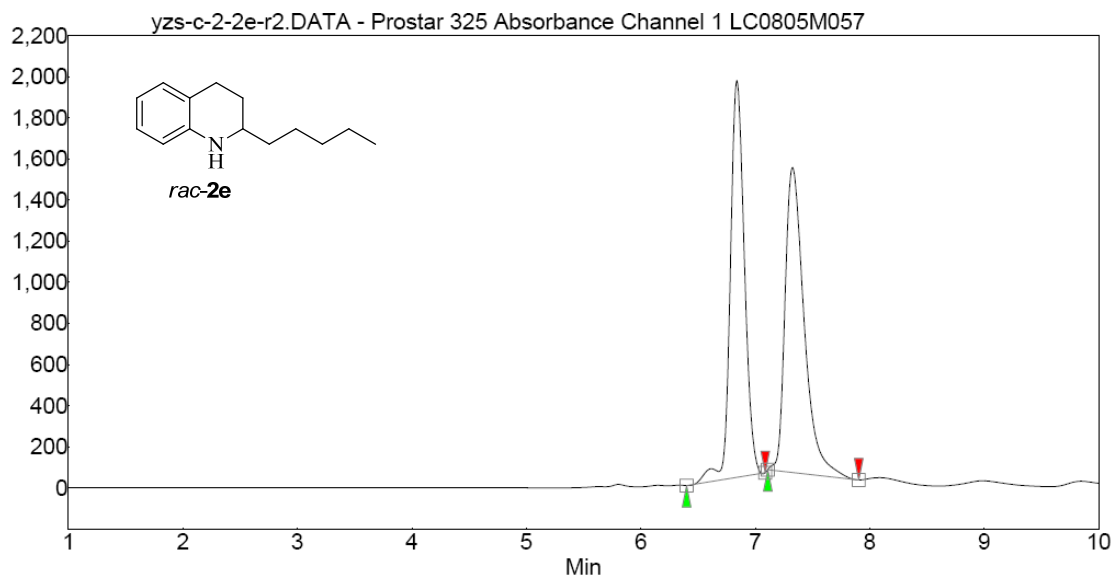


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	7.71	49.36	697.3	129.4	49.365
2	未知	8.99	50.64	588.2	132.7	50.635
Total			100.00	1285.5	262.2	100.000

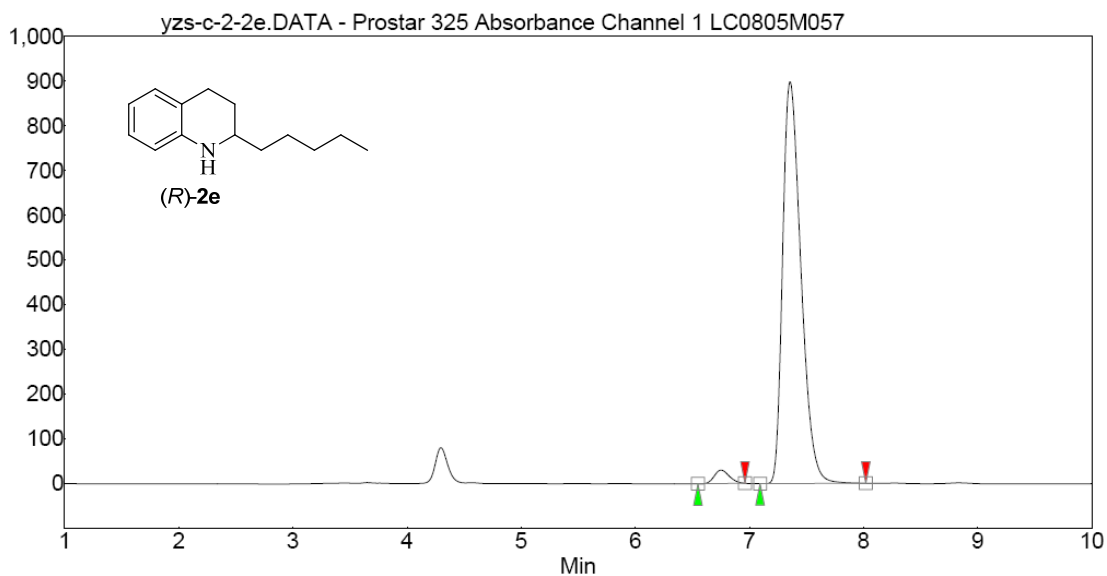


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	7.57	3.77	130.1	22.9	3.766
2	未知	8.71	96.23	2113.7	584.4	96.234
Total			100.00	2243.8	607.2	100.000

HPLC spectra of 2e catalyzed by (R,R)-3

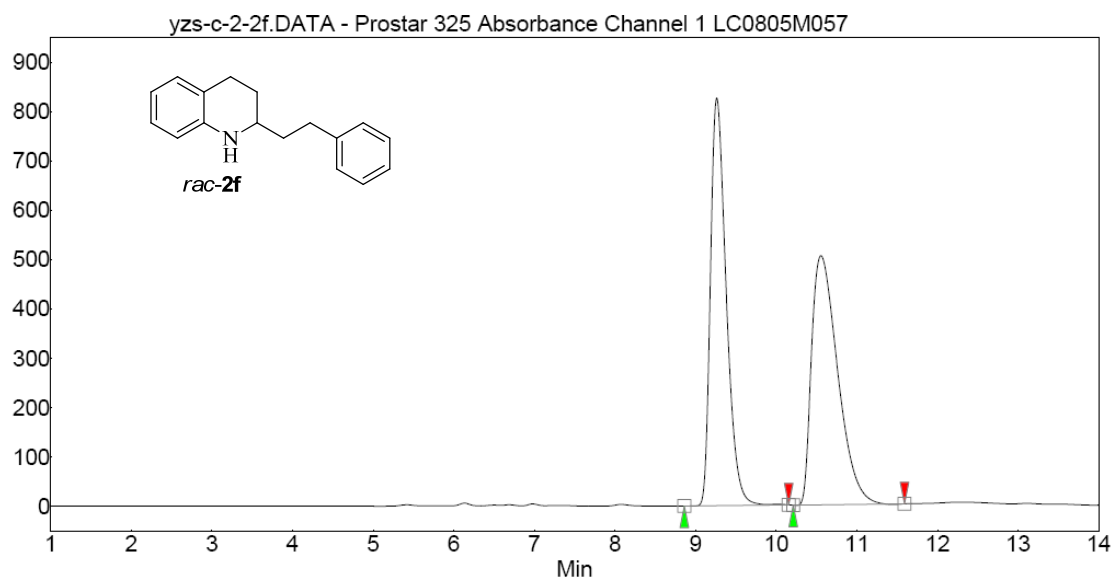


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	6.84	49.08	1929.8	277.1	49.077
2	未知	7.33	50.92	1483.1	287.5	50.923
Total			100.00	3413.0	564.6	100.000

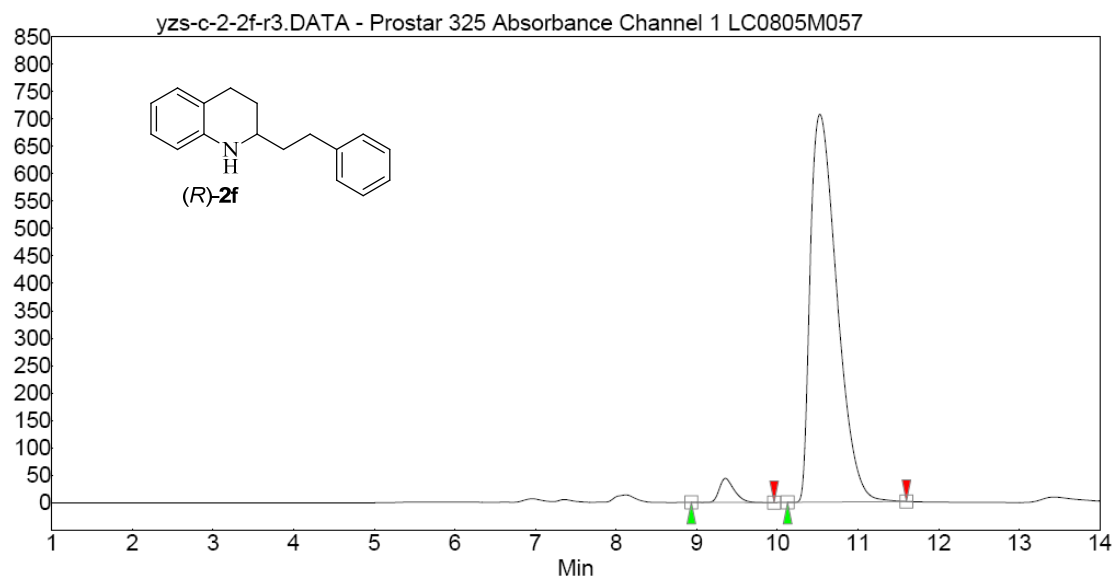


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	6.75	2.68	30.1	4.6	2.676
2	未知	7.35	97.32	898.2	166.8	97.324
Total			100.00	928.3	171.4	100.000

HPLC spectra of 2f catalyzed by (R,R)-3

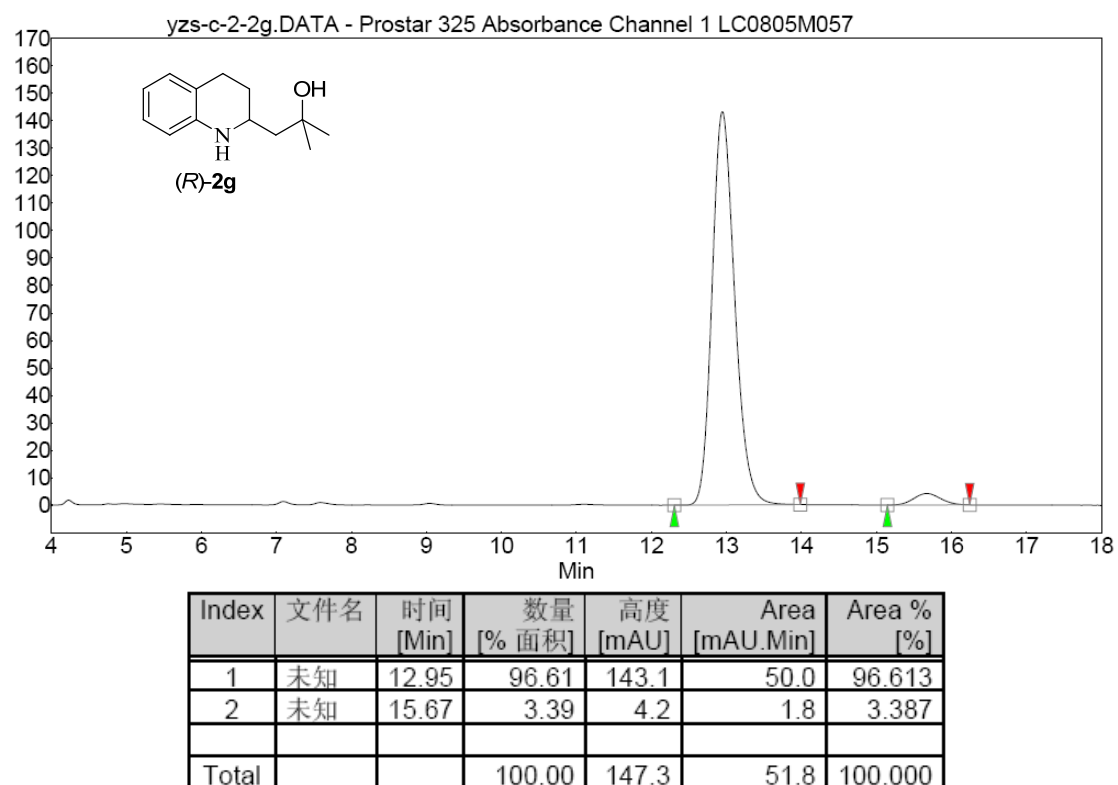
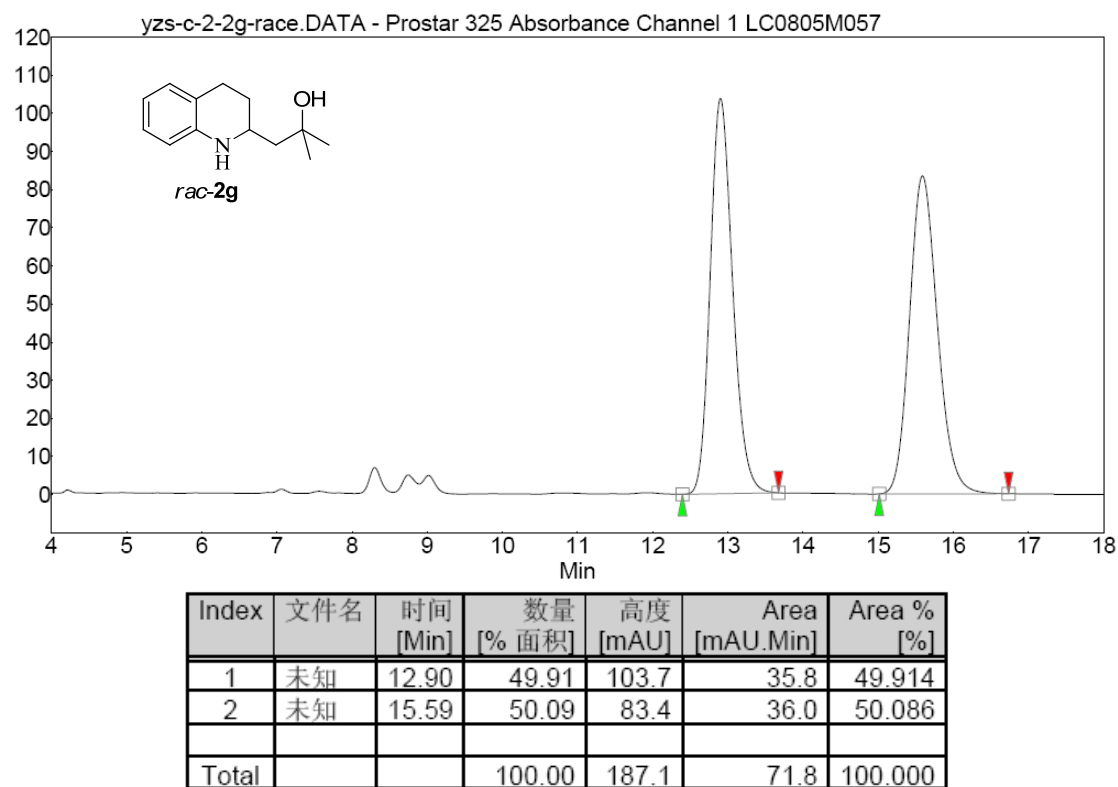


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	9.26	49.66	826.3	187.0	49.664
2	未知	10.55	50.34	504.9	189.5	50.336
Total			100.00	1331.2	376.5	100.000

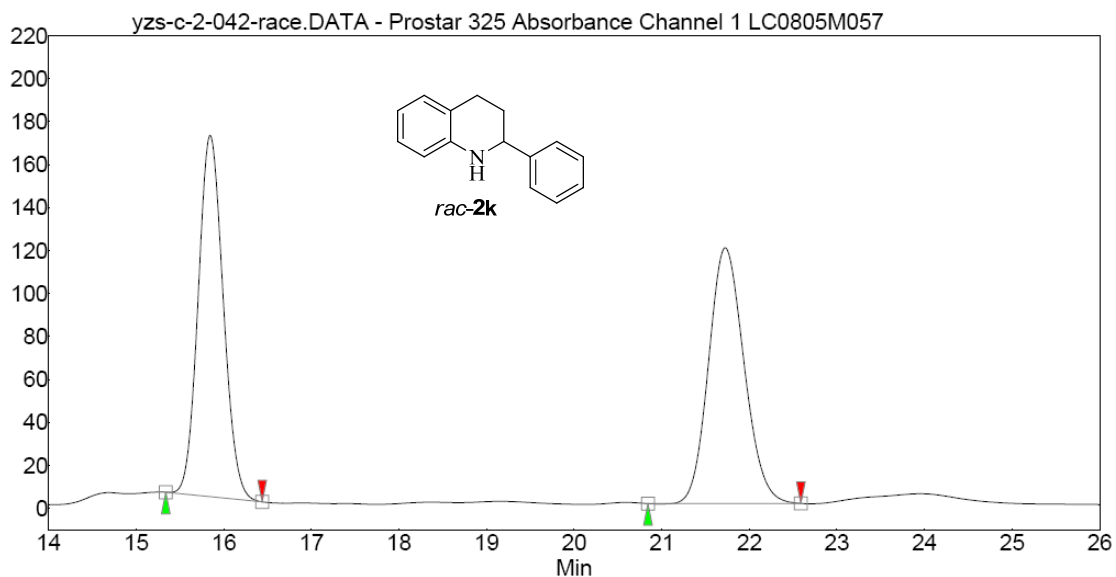


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	9.35	3.35	44.0	9.4	3.349
2	未知	10.53	96.65	707.2	270.7	96.651
Total			100.00	751.2	280.1	100.000

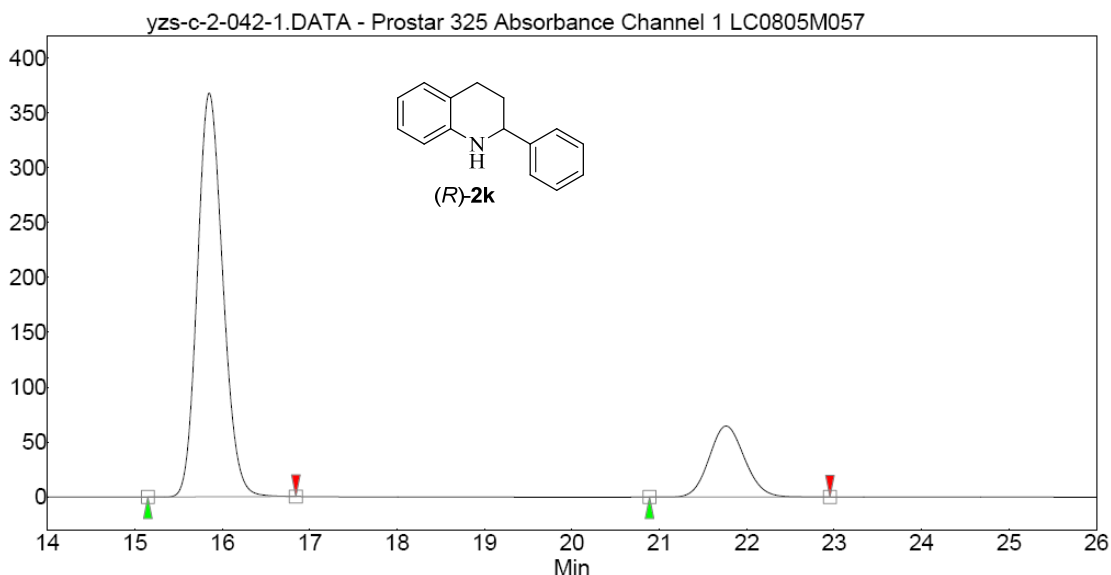
HPLC spectra of 2g catalyzed by (R,R)-3



HPLC spectra of 2k catalyzed by (R,R)-3



Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	15.35	49.64	462.4	150.3	49.645
2	未知	20.74	50.36	333.8	152.5	50.355
Total			100.00	796.2	302.8	100.000



Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	15.39	81.63	1211.4	409.8	81.629
2	未知	20.83	18.37	205.7	92.2	18.371
Total			100.00	1417.2	502.0	100.000