

Electronic Supplementary Information (ESI)

Extracellular pH-Manipulated In Situ Reconfiguration of Aptamer Functionalized DNA Monomer Enable Specifically Improved Affinity, Detection and Drug Delivery

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Table S1 All of the oligonucleotides used in this work.¹

Probe	Sequence (5'-3')
Y _{1i}	CGA CCG ATG AAT AGC <i>T</i> ATC CGT ACC TAC TCG TTTTT CCC CCC T CCC CCC
Y _{2i}	CGA GTC GTT CGC ACG <i>T</i> GCT ATT CAT CGG TCG TTTTT CCC CCC T CCC CCC
Y ₃	CGA GTA GGT ACG GAT <i>T</i> CGT GCG AAC GAC TCG GCT GTG AAC CAA GTC
Cy5-apt	Cy5- <i>T</i> GAC TTG GTT CAC AGC <i>TTT TTTT TTT</i> <u>ACG CGC GCG CGC ATA GCG CGC TGA GCT GAA GAT CGT ACC GTG AGC GCG T</u>
Cy5-con ²	Cy5- <i>T</i> GAC TTG GTT CAC AGC <i>TTT TTTT TTT</i> ACT CAT AGT GTG TTT CAC ACT ATT TTA TCT TTG TTC TTA TCT TAT GAG T
Y _{1c} ³	CGA CCG ATG AAT AGC <i>T</i> ATC CGT ACC TAC TCG TTTTT TTT TTT T TTT TTT
Y _{2c} ³	CGA GTC GTT CGC ACG <i>T</i> GCT ATT CAT CGG TCG TTTTT TTT TTT T TTT TTT
Y _{1cp} ⁴	CGA CCG ATG AAT AGC TAT CCG TAC CTA CTC GTA GTC GAA TGT CTC GTT A
Y _{2cp} ⁴	CGA GTC GTT CGC ACG TGC TAT TCA TCG GTC GTA GTC GAA TGT CTC GTT A
Y _{1cp'} ⁴	CGA CCG ATG AAT AGC TAT CCG TAC CTA CTC GTT AAC GAG ACA TTC GAC T
Y _{2cp'} ⁴	CGA GTC GTT CGC ACG TGC TAT TCA TCG GTC GTT AAC GAG ACA TTC GAC T

¹ In all sequences, split i-motif DNA sequences are in bold. The complementary ssDNA sequences used to replace split i-motif sequences are shown in blue. ZY11 sequence against SMMC-7721 cell is showed underlined, poly T in italic is used as linker for minimizing steric resistance.

² ConDM is assembled from Cy5-con, Y_{1i}, Y_{2i}, Y₃, which shows little affinity to target SMMC-7721 cells. ³ AptDM_{Con} is assembled from Cy5-apt, Y_{1c}, Y_{2c}, and Y₃, which is incapable of crosslinking each other into bulk nanostructure in whatever pH. While, ConDM_{Con} is assembled from Cy5-con, Y_{1c}, Y_{2c}, Y₃, which is neither affinitive to target cells nor sensitive to pH.

⁴ AptDM-C₁ is assembled from Cy5-apt, Y_{1cp}, Y_{2cp} and Y₃, while AptDM-C₂ is assembled from Cy5-apt, Y_{1cp'}, Y_{2cp'}, and Y₃. AptDM-C₁ and AptDM-C₂ can spontaneously crosslink each other into bulk nanostructure (MDA_{cp}) via sticky-end hybridization in whatever pH.

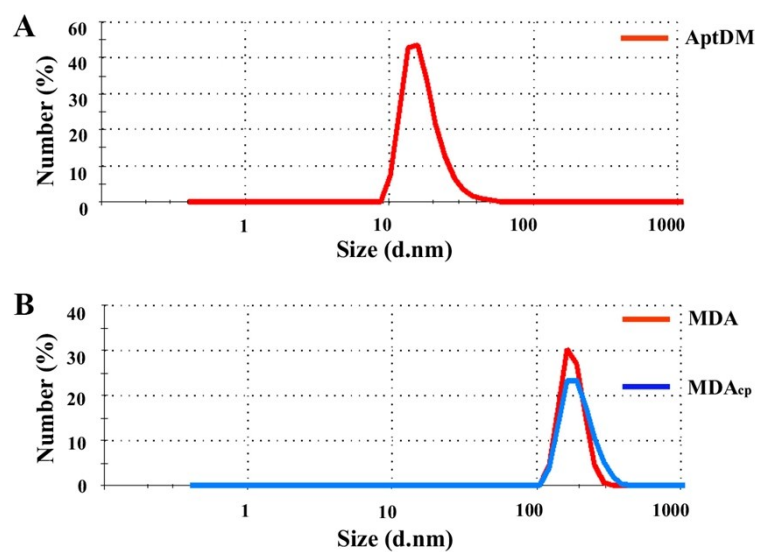


Fig. S1 Dynamic light scattering analysis of the size of AptDM, MDA and MDA_{cp}, showing the mean diameters at 17.1, 178.5 and 195.2 nm, respectively.

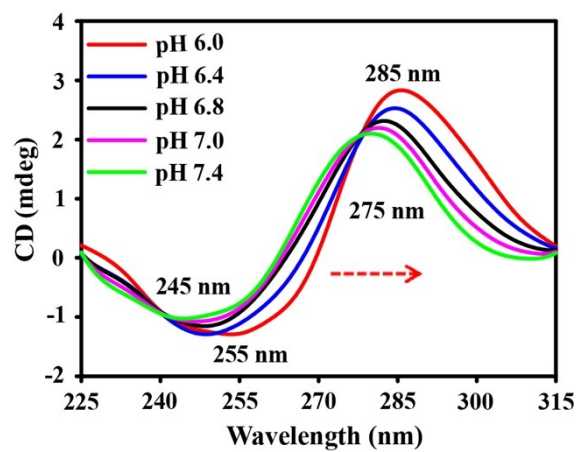


Fig. S2 CD spectra of AptDM in PBS with different pH values. (Probe concentration: 1 μ M.)

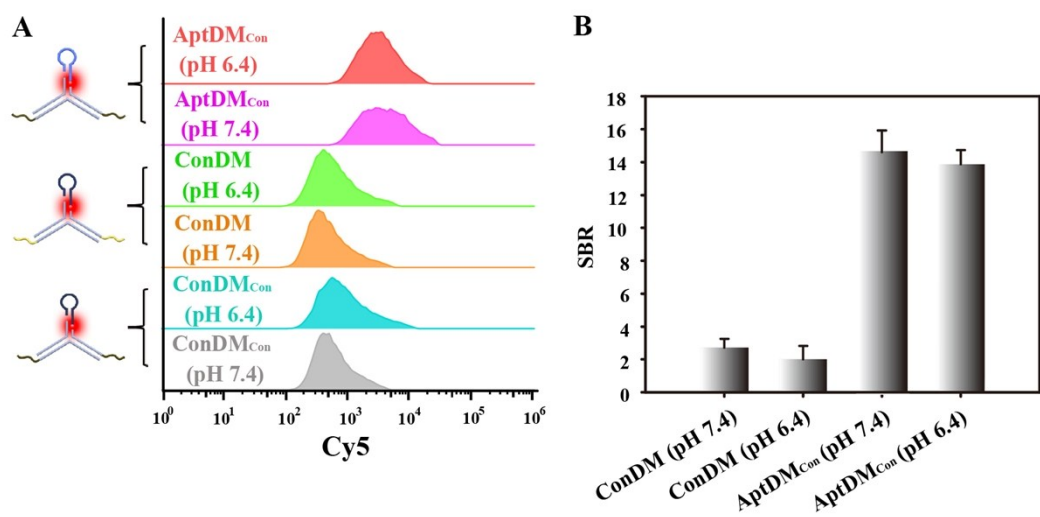


Fig. S3 (A) Flow cytometric assays of SMMC-7721 cells after incubation with different probes in binding buffer with different pH values, respectively. (B) The corresponding signal-to-background ratio (SBR) of probes in (A) for detecting SMMC-7721 cells at different pH values. (Probe concentration: 50 nM.)

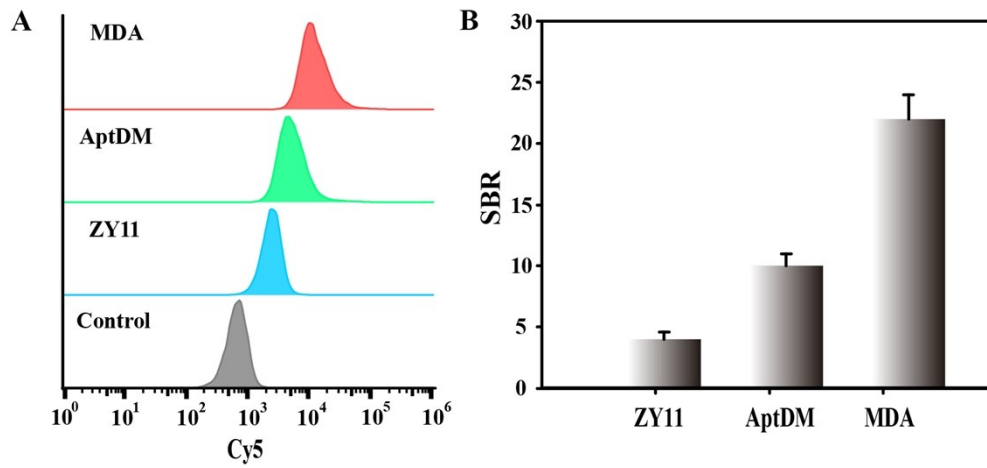


Fig. S4 (A) Flow cytometric assay of binding ability of different probes to SMMC-7721 cells at 37 °C in binding buffer containing 0 mM Mg^{2+} . (B) The corresponding SBR of ZY11, AptDM and MDA probes in (A).

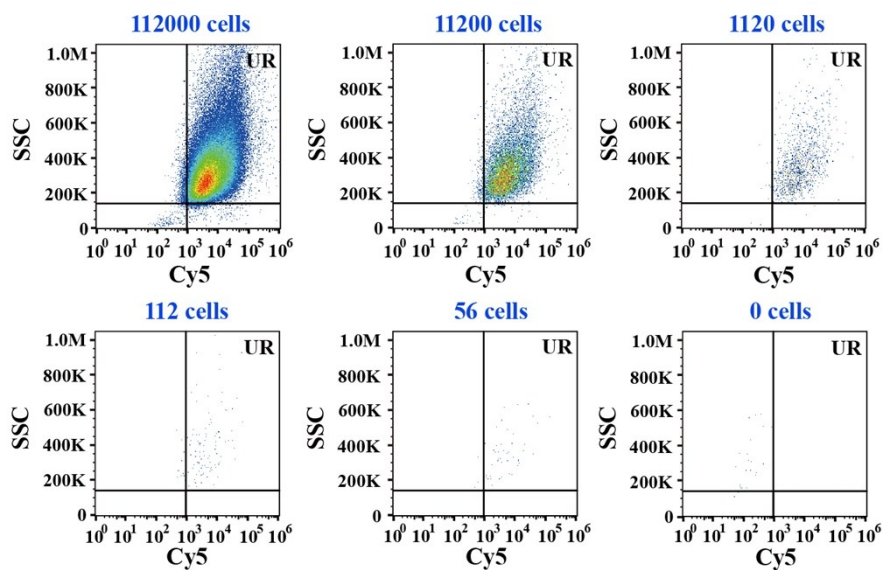


Fig. S5 Flow cytometric assays of SMMC-7721 cells with decreasing cell number from 1.12×10^5 to 0 in 150 μ L binding buffer after incubation with MDA.

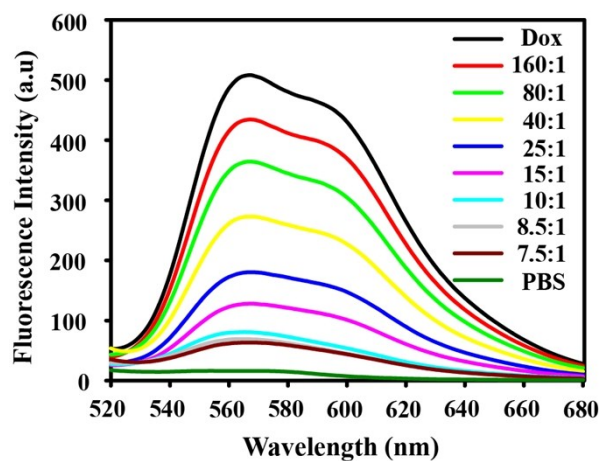


Fig. S6 Fluorescence spectra of Dox solutions (1 μ M) added with AptDM at various ratios of Dox to AptDM.