

Electronic Supplementary Information File For

**A Novel Liquid Biopsy-Based Approach for Highly Specific Cancer
Diagnostics: Mitigating False Responses in Assaying Patient Plasma-Derived
Circulating microRNAs through Combined SERS and Plasmon-Enhanced
Fluorescence Analyses**

Adrianna N. Masterson,^a Thakshila Liyanage,^a Claire Berman,^a Hristos Kaimakliotis,^b Merrell
Johnson,^c and Rajesh Sardar^{*,a}

^aDepartment of Chemistry & Chemical Biology, Indiana University-Purdue University
Indianapolis, 402 N. Blackford Street, Indianapolis, Indiana 46202, United States

^bDepartment of Urology, Indiana University School of Medicine, 535 N. Barnhill Dr.
Indianapolis, Indiana 46202, United States

^cDepartment of Physics, Purdue University Fort Wayne, 2101 E. Coliseum Blvd.
Fort Wayne, Indiana 46805, United States

Table S1: -ssDNA (oligomer) sequences used for this study.

Name	Sequence	Modification
-ssDNA-10b	5' CAC AAA TTC GGT TCT ACA GGG TA 3'	3' Thiol-(CH ₂) ₃
-ssDNA-96	5' AGC AAA AAT GTG CTA GTG CCA AA 3'	3' Thiol-(CH ₂) ₃

Table S2: microRNA sequences used for this study.

Name	Sequence	Modification
microRNA-10b	5' UACCCUGUAGAACCGAAUUUGUG 3'	N/A
microRNA-96	5' UUUGGCACUAGCACAUUUUUGUG 3'	N/A
microRNA-145	5' GUCCAGUUUCCCCAGGAAUCCCU 3'	N/A
microRNA-143	5'GGUGCAGUGCUGCAUCUCUGGU 3'	N/A
microRNA-490-5p	5' CCAUGGAUCUCCAGGUGGGU 3'	N/A
microRNA-10b-FAM	5' UACCCUGUAGAACCGAAUUUGUG 3'	5' 6-FAM
microRNA-96-FAM	5' UUUGGCACUAGCACAUUUUUGUG 3'	3' 6-FAM

Table S3: microRNA-10b calibration curve raw data.

microRNA-10b Concentration (nM)	1255 cm ⁻¹ Peak Intensity (cm ⁻¹)	Standard Deviation (cm ⁻¹)
100	440.5	32.0
10	409.3	35.0
1	344.7	40.0
0.1	313.9	33.5
0.01	284.6	23.5
0.001	277.2	30.0
0.0001	206.1	27.5

Table S4: microRNA-96 calibration curve raw data.

microRNA-96 Concentration (nM)	1255 cm^{-1} Peak Intensity (cm^{-1})	Standard Deviation (cm^{-1})
100	497.7	50.0
10	428.3	35.0
1	397.1	22.9
0.1	362.7	30.0
0.01	346.1	27.5
0.001	329.4	17.3
0.0001	257.4	40.0

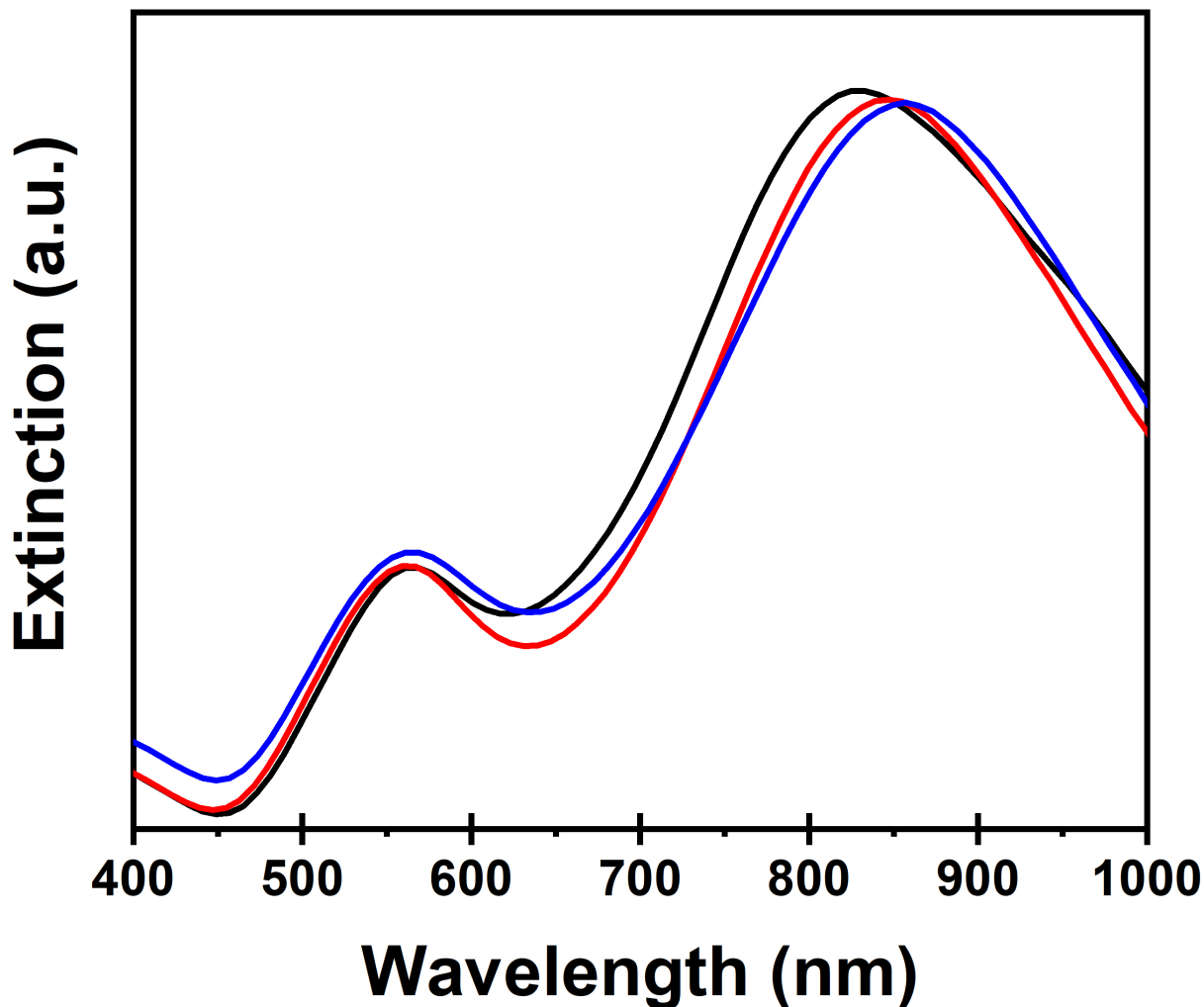


Fig. S1. UV-Vis extinction spectra of Au TNPs during the different stages of surface functionalization.

Black curve: Extinction spectrum of as synthesized Au TNPs attached onto a silanized glass substrate.

The LSPR dipole peak position (λ_{LSPR}) of Au TNPs is ~ 830 nm. **Red curve:** Functionalization of TNPs

with $-S(CH_2)_3$ -ssDNA-10b/96-FAM, $\lambda_{LSPR} = \sim 845$ nm. Red shifting of the LSPR dipole peak is due to the change in the local refractive index around the nanoprisms upon attachment of $-S(CH_2)_3$ -ssDNA-10b/96-

FAM. **Blue curve:** Further functionalization of TNPs with 3-MP produces additional red shifts, $\lambda_{LSPR} =$

~ 860 nm. All spectra were collected in RNAs free water.

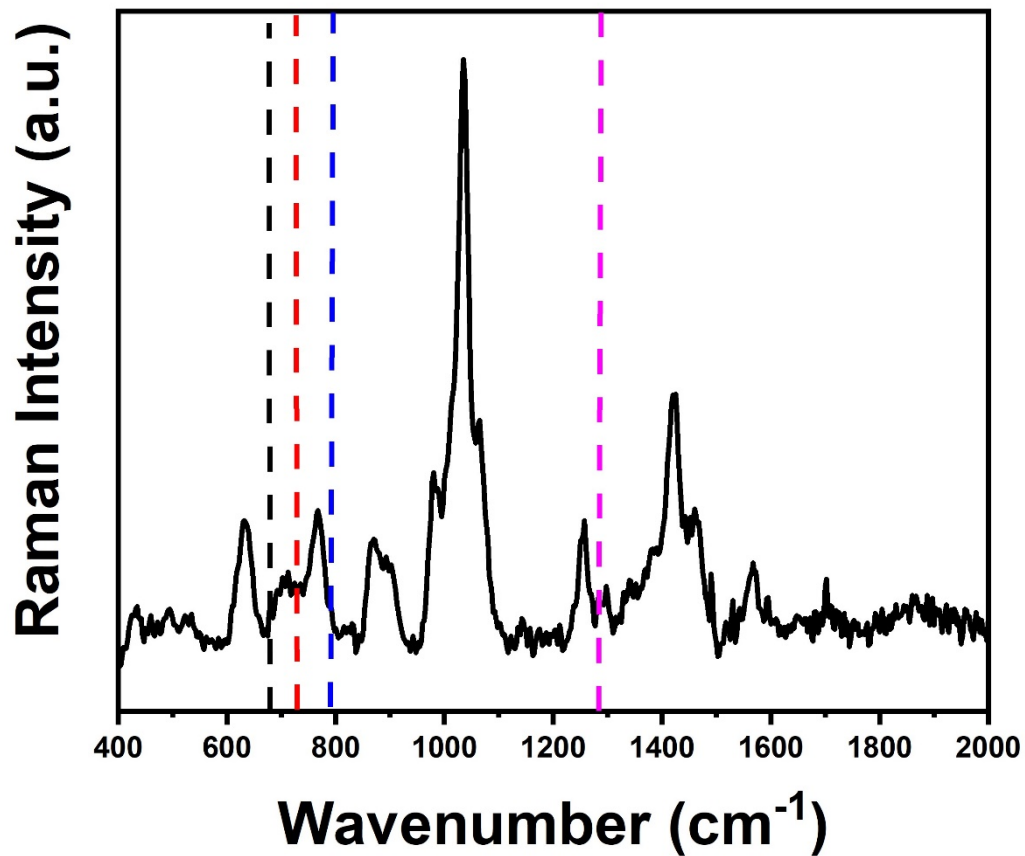


Fig. S2. SERS spectra of -ssDNA attached onto Au TNPs through the Au-S bond. Guanine ring breathing is represented at 688 cm^{-1} (black dashed line). Adenine ring breathing is represented at 733 cm^{-1} (red dashed line). Cytosine ring breathing is represented at 790 cm^{-1} (blue dashed line). Phosphodiester group of the nucleic acids is represented at 1289 cm^{-1} (pink dashed line).

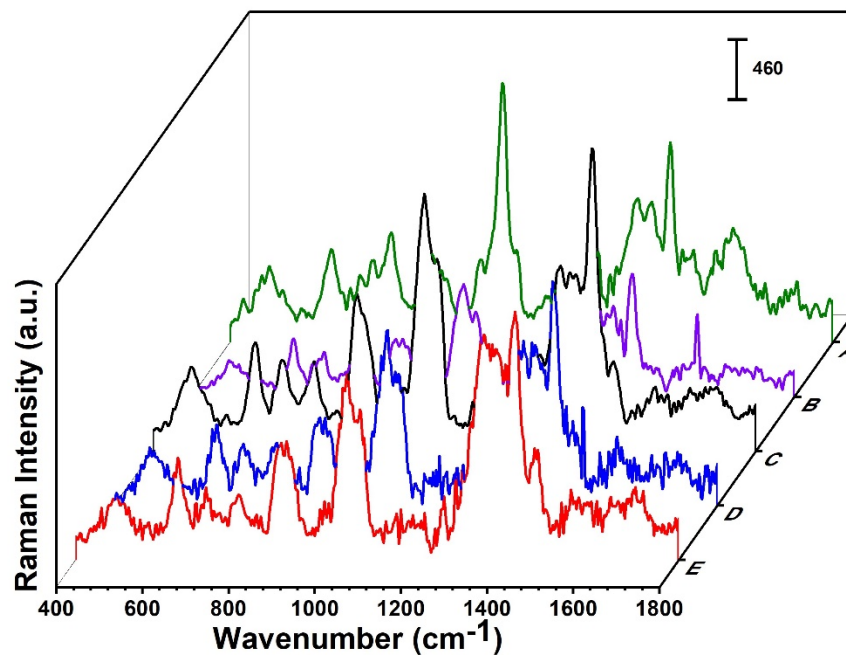


Fig. S3. SERS spectra of reversibility test. (A) SERS spectra after attachment of -ssDNA-10b, (B) SERS spectra after attachment of 3-MP, (C) SERS spectra after attachment of microRNA-10b-FAM, (D) SERS spectra after treating with RNase H enzyme solution, and (E) SERS spectra after re-attachment of microRNA-10b-FAM.

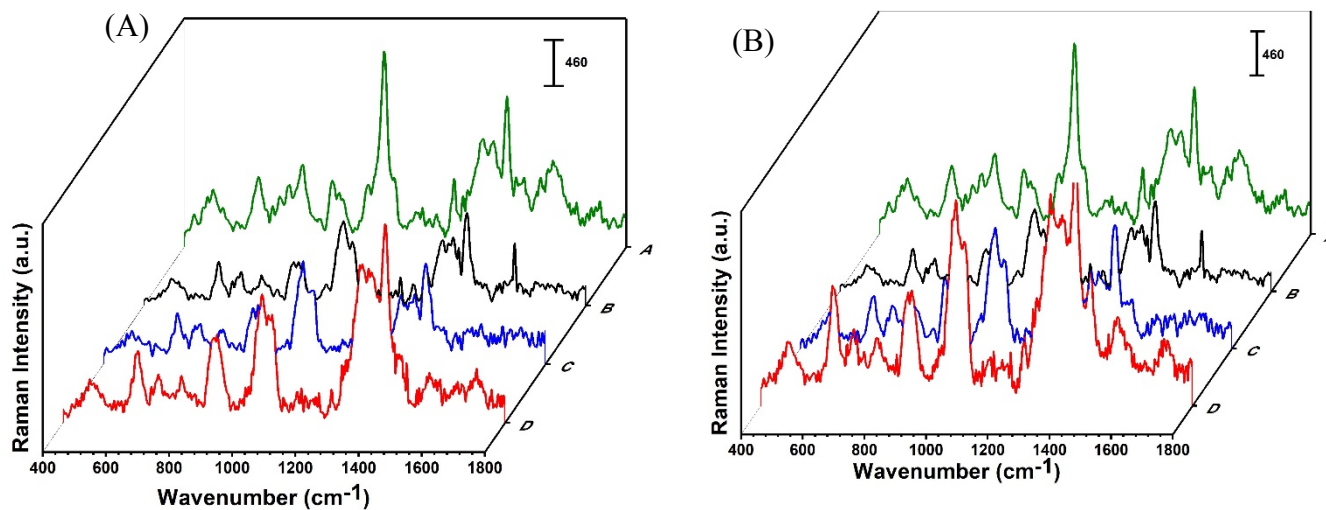


Fig. S4. SERS spectra of -ssDNA specificity tests. (A) Representation of SERS spectra for -ssDNA- 10b specificity test. [A = -ssDNA-10b, B = 3-MP, C = mixture of microRNA-96, 145, 143, and 490-5p. D = microRNA-10b.] (B) Representation of SERS spectra for -ssDNA-96 specificity test. [A = -ssDNA-96, B = 3-MP, C = mixture of microRNA-10b, 145, 143, and 490-5p. D = microRNA-96.]