Electronic Supporting Information

FRET-based sensor for visualizing pH variation with colorimetric/ratiometric and application for bioimaging in living cells, bacteria and zebrafish

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Scheme S1 Synthetic routes of Rh-TPE sensor, and the structure of Rh-TPE in neutral and acidic condition.



Figure S1 ¹H NMR spectra of Rh-TPE sensor in DMSO-*d*₆.



Figure S2 ¹³C NMR spectra of Rh-TPE sensor in DMSO- d_6 .



Figure S3 ¹H NMR spectra of Rh-TPE sensor after addition of H⁺ in DMSO- d_6 .



Figure S4 Mass spectrum of Rh-TPE with and without addition of H⁺ in EtOH.



Figure S5 The overlap spectra of the absorption of Rhodamine and emission of TPE.



Figure S6 The excitation spectrum of Rh-TPE in buffer (1% DMSO) at pH 8.4.



Figure S7 (A) Absorption spectra (10 μ M) and (B) fluorescence spectra (5 μ M) of **Rh-TPE** in BR buffer (include 1% DMSO) at pH 8.4 (black line) and pH 2.0 (red line), respectively.



Figure S8 Time-depend fluorescence responses (F_{593}/F_{455}) of 5 μ M Rh-TPE at pH 2.0 buffer solution.



Figure S9 Detection the effect of viscosity on the sensor in water-glycerol system. (A) Absorption spectra of **Rh-TPE** (10 μ M) in the water (containing 1% DMSO) and glycerol (containing 1% DMSO) at pH 8.0 and pH 2.0. (B) Fluorescence spectra of **Rh-TPE** (5 μ M) in the water (containing 1% DMSO) and glycerol (containing 1% DMSO) at pH 8.0 and pH 2.0.



Figure S10 Cell viability test using the MTT assay in hPDLCs cells at 24 h of culture. PBS treatment as a control group. The concentration of **Rh-TPE** sensor varied from 5 to 80 μ M. Results are presented as the mean of the three measurements \pm standard deviation. (n = 3)