

1 Quantitative ¹H-NMR Spectroscopy as an Efficient Method for 2 Identification and Quantification of PVC, ABS and PA Microparticles

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6 **Supporting Information:**

- 7 Table SI 1 (2) Possible solvents for PVC, ABS and PA based on the literature. Given are the
8 solvents already described for NMR spectroscopic investigations of the
9 respective polymers, the corresponding ¹H-Signals of the polymers and the
10 residual proton signals in ppm as well as the solvents tested in this survey.
- 11 Figure SI 1 (2) ¹H-NMR spectra of matrix effects of natural aquatic biofilm. **a)** ~11 mg/mL
12 biofilm solved in CDCl₃/FA 3:2. **b)** ~16 mg/mL biofilm solved in CDCl₃. **c)**
13 ~14 mg/mL biofilm solved in DMSO-d₆.
- 14 Figure SI 2 (2) ¹H-NMR spectrum of PVC after line fitting for the PF-method in the
15 range of 3.0-1.5 ppm. Blue represent the individual fitted peaks and
16 pink represent the sum of the fitted peaks.
- 17 Figure SI 3 (3) ¹H-NMR spectra of ABS in CDCl₃ (left) and in DMSO-d₆ (right) after line
18 fitting for the PF-method in the range of 8.0-6.0 ppm and 3.0-2.0 ppm. Blue
19 represent the fitted peaks and pink represent the sum of the fitted peaks. ¹H
20 NMR spectrum of ABS after line fitting for the PF-method in the
21 range of 8.0-6.0 ppm. Blue represent the individual fitted peaks and
22 pink represent the sum of the fitted peaks.
- 23 Figure SI 4 (3) Calibration curves for the signal which can also be used for quantification for
24 the MP particles of PVC, ABS and PA with the appropriate confidence interval
25 (CI). Plotted is the MP concentration in mg/mL against the normalized
26 intensity.
- 27 Table SI 2 (4) Linearity data of MP particles (PVC, ABS and PA). Data were calculated from
28 five measurement points.
- 29 Table SI 3 (4) Quantitative results for model samples using the signal which can also be used
30 for quantification for the MP particles of PVC, ABS and PA.
- 31 Table SI 4 (5) Overview of the ¹H-Signals of the MP polymer types, residual solvent signals
32 and matrix effects of biofilm in the corresponding solvent conditions.

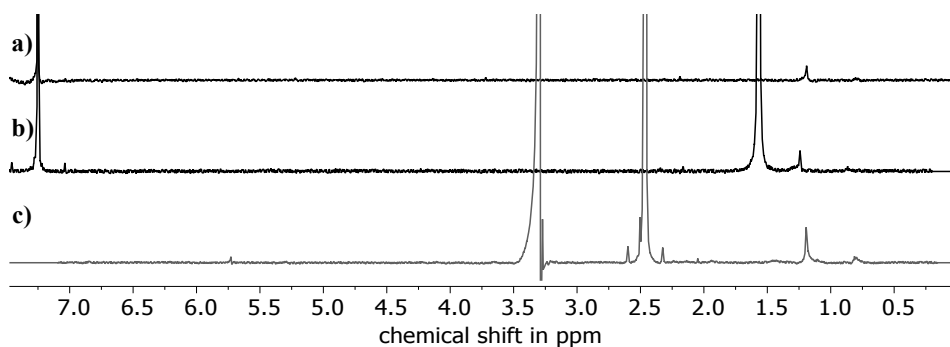
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34 **Table SI 1.** Possible solvents for PVC, ABS and PA based on the literature. Given are the solvents already
 35 described for NMR spectroscopic investigations of the respective polymers, the corresponding ¹H-Signals of the
 36 polymers and the residual proton signals in ppm as well as the solvents tested in this survey.

MP	Solvents used in the literature	MP ¹ H signals in ppm	Solvent ¹ H signals in ppm	Solvents tested in this survey
PVC	DMSO ²⁸	4.7-4.3; 2.4-2.1 ²⁸	2.5	DMSO
	CDCl ₃ ²⁷	4.45; 2.1 ²⁷	7.2	
ABS	CDCl ₃ ³⁰	7.2-6.4; 5.5; 2.5-1.2 ³⁰	7.2	CDCl ₃
	DMSO ²⁹	7.2-6.8; 5.3; 2.5-1.4 ²⁹	2.5	DMSO
PA 6.6	TFA ³¹	3.3; 2.5; 1.7; 1.3 ³¹	~ 12	CDCl ₃ /FA 3:2
	Cresol/ODCB ²⁶	not specified ²⁶	7.2; 6.6; 5.0; 2.3	

37 TFA, trifluoroacetic acid; FA, formic acid; ODCB, 1,2-dichlorobenzene.

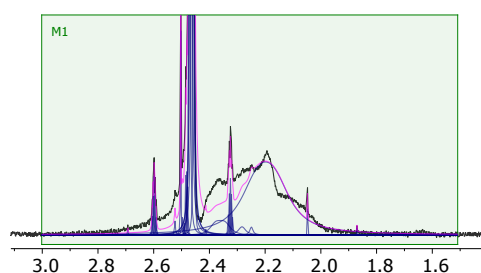
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40 **Figure SI 1** ¹H-NMR spectra of matrix effects of natural aquatic biofilm in the respective deuterated solvent. **a)**
 41 ~11 mg/mL biofilm solved in CDCl₃/FA 3:2. **b)** ~16 mg/mL biofilm solved in CDCl₃. **c)** ~14 mg/mL biofilm solved
 42 in DMSO-d₆. **Aquatic biofilm, i.e. a complex and dynamic community of algae, cyanobacteria, heterotrophic**
 43 **microorganisms, and detritus attached to surfaces in most aquatic exosystems. Biofilm samples were collected at**
 44 **the river Moselle (position 50° 21' 49.8" N 7° 33' 54.6" E) by brushing light-exposed grown surfaces of submerges**
 45 **stones and and dried at 60° C.)**

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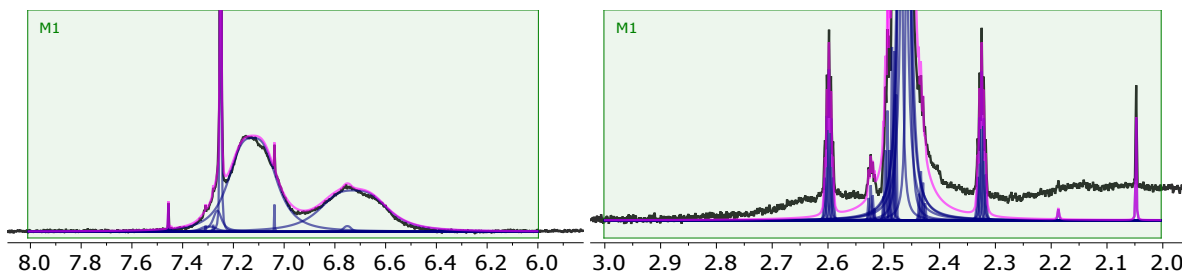
48 **Figure SI 2** ¹H-NMR spectrum of PVC after line fitting for the PF-method in the range of 3.0-1.5 ppm. Blue
 49 represent the fitted peaks and pink represent the sum of the fitted peaks.

Comment [NP]: To Referee 2 Suggestion 2

We here refer to aquatic biofilm (i.e. stream periphyton) which is a complex and dynamic community of algae, cyanobacteria, heterotrophic microorganisms, and detritus. Under natural conditions, bacteria generally exist in the biofilm state, i.e. cells attached to each other and also to a surface, embedded in a protective matrix. In aquatic ecosystems aquatic biofilm represents an important food source for grazing macroinvertebrates or fish habitats and can adsorb contaminants from the free-flowing water.

Analyzed biofilm samples were taken from natural biofilm of the river Moselle (at position 50° 21' 49.8" N 7° 33' 54.6" E) by brushing grown surfaces (i.e. stones), as it not commercially available.

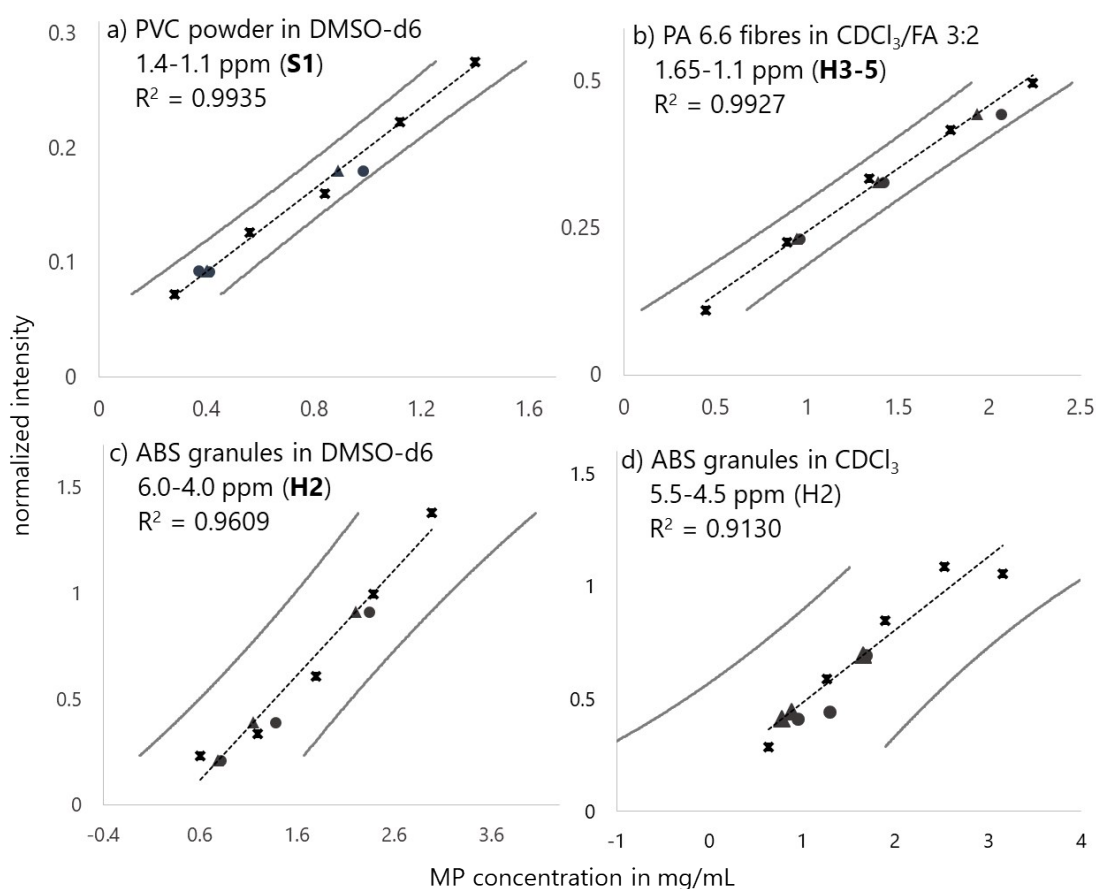
Relevant information on the general composition of biofilm and the collection site was added to the caption of figure SI 2.



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51 **Figure SI 3** $^1\text{H-NMR}$ spectra of ABS in CDCl_3 (left) and in DMSO-d_6 (right) after line fitting for the PF-method
 52 in the range of 8.0-6.0 ppm and 3.0-2.0 ppm. Blue represent the fitted peaks and pink represent the sum of the
 53 fitted peaks.

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56 **Figure SI 4.** Calibration curves for the signal which can also be used for quantification for the MP particles of
 57 PVC, ABS and PA with the appropriate confidence interval (CI). Plotted is the MP concentration in mg/mL against
 58 the normalized intensity. In addition, the true and calculated values of the model samples are given. a) signal
 59 range of 1.4-1.1 ppm (S1) for PVC particles, b) signal range of 1.65-1.1 ppm (H3-5) for PA fibres, c) signal range
 60 of 6.0-4.0 ppm (H2) for ABS particles in DMSO-d_6 and d) signal range of 5.5-4.5 ppm (H2) for ABS particles in
 61 CDCl_3 are compared. For linearity data see supporting information SI 6.

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65 **Table SI 2.** Linearity data of MP particles (PVC, ABS and PA). Data were calculated from five measurement
66 points. PVC: Integration area 4.7-4.05 ppm (H1) and 1.4-1.1 ppm (S1). For integration the peak-fitting method
67 was used. 0.28-1.40 mg/mL concentration range. ABS in CDCl₃: Integration area 7.1-6.0 ppm (H1) and 5.5-4.5
68 ppm (H2) are given. For integration the peak-fitting method was used. 0.63-3.15 mg/mL concentration range. ABS
69 in DMSO-d₆: Integration area 7.5-6.0 ppm (H1) and 6.0-4.0 ppm (H2) are given. For integration the peak-fitting
70 method was used. 0.63-3.15 mg/mL concentration range. PA: Integration area 3.2-3.05 ppm (H1) and 1.65-1.1
71 ppm (H3-5) are given. For integration the peak-fitting method was used. 0.45-2.24 mg/mL concentration range.

	MP particles	Slope ± SD	Intercept ± SD	R ²
Signal H1	PVC in DMSO-d ₆	1.3487 ± 0.0330	- 0.0455 ± 0.0306	0.9982
Signal H1	ABS in CDCl ₃	3.8863 ± 0.2247	+ 0.6164 ± 0.4699	0.9901
Signal H1	ABS in DMSO-d ₆	3.8158 ± 0.0312	+ 0.0832 ± 0.0617	0.9998
Signal H1	PA in CDCl ₃ /FA	0.0714 ± 0.0029	+ 0.0106 ± 0.0043	0.9950
Signal S1	PVC in DMSO-d ₆	0.1797 ± 0.0084	+ 0.0204 ± 0.0078	0.9935
Signal H2	ABS in CDCl ₃	0.3235 ± 0.0577	+ 0.1609 ± 0.1206	0.9130
Signal H2	ABS in DMSO-d ₆	0.4960 ± 0.0578	- 0.1774 ± 0.1143	0.9609
Signal H3-5	PA in CDCl ₃ /FA	0.2156 ± 0.0107	+ 0.0281 ± 0.0158	0.9927

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73 **Table SI 3.** Quantitative results for model samples using the signal which can also be used for quantification for
74 the MP particles of PVC, ABS and PA. True Value ± weighting error and calculated values ± confidence interval
75 (CI) of the respective MP samples in mg/mL as well as the accuracy (bias in %) and precision (RSP in %) are
76 given. PVC samples: integration area 1.4-1.1 ppm (S1). For integration the PF-method was used. ABS samples in
77 DMSO-d₆: integration area 6.0-4.0 ppm (H2). For integration the PF-method was used. ABS samples in CDCl₃:
78 integration area 5.5-4.55 ppm (H2). For integration the PF-method was used. PA samples: integration area 1.65-
79 1.1 ppm (H3-5). For integration the INT-method was used.

	MP signal	True Value ± Δx (mg/mL)	Calculated ± CI (mg/mL)	Accuracy (bias in %)	Precision (RSD in %)
PVC 1 (in DMSO-d ₆)	S1	0.98 ± 0.01	0.94 ± 0.13	95.5	99.9
PVC 2 (in DMSO-d ₆)	S1	0.78 ± 0.01	0.68 ± 0.13	87.2	99.9
PVC 3 (in DMSO-d ₆)	S1	0.41 ± 0.01	0.44 ± 0.14	107	99.9
ABS 1 (in DMSO-d ₆)	H2	2.34 ± 0.01	2.18 ± 0.78	93.0	99.2
ABS 2 (in DMSO-d ₆)	H2	1.37 ± 0.01	1.15 ± 0.80	84.3	99.2
ABS 3 (in DMSO-d ₆)	H2	0.81 ± 0.01	0.78 ± 0.85	95.9	99.8
ABS 1 (in CDCl ₃)	H2	1.68 ± 0.01	1.70 ± 1.25	101	99.9
ABS 2 (in CDCl ₃)	H2	1.29 ± 0.01	0.88 ± 1.37	68.1	99.9
ABS 3 (in CDCl ₃)	H2	0.95 ± 0.01	0.70 ± 1.39	73.4	92.2
PA 1 (in CDCl ₃ /FA 3:2)	H3-5	2.06 ± 0.01	1.94 ± 0.26	94.1	99.2
PA 2 (in CDCl ₃ /FA 3:2)	H3-5	1.42 ± 0.01	1.39 ± 0.24	97.9	99.9
PA 3 (in CDCl ₃ /FA 3:2)	H3-5	0.96 ± 0.01	0.95 ± 0.25	98.6	99.9

81 **Table SI 4.** Overview of the ^1H -Signals of the MP polymer types, residual solvent signals and matrix effects of biofilm in the corresponding solvent conditions. Indicated
 82 are the ^1H -signals or the range of the chemical shift in ppm, whether the MP signals is suitable for quantitative analysis by qNMR and the reasons why the corresponding

		Signals [ppm]	Suitable qNMR	Reasons why not suitable for quantification
DMSO-d6	solvent	2.47-2.4 (DMSO)		
		3.36-3.24 (H ₂ O)		
	matrix	2.3-1.9; 1.5-0.6		
	PVC	4.6-4.2 (H1)	yes	-
		2.5-2.1 (H2)	no	Signal overlap with solvent signal, interference with matrix effects possible
		1.4-1.1 (S1)	(yes)	interference with matrix effects possible
		0.9-0.7 (S2)	no	interference with matrix effects possible, low signal intensity
	ABS	7.7-6.5 (H1)	yes	-
		5.9-4.5 (H2)	(yes)	Low signal intensity
		2.7-1.2 (H3-H7)	no	Signal overlap with solvent signal, Interference with matrix effects possible
0.9-0.7 (H8)		no	Interference with matrix effects possible, low signal intensity	
CDCl₃	solvent	7.25 (CDCl ₃)		
	matrix	1.6-0.6		
	ABS	7.4-6.3 (H1)	(yes)	Signal overlap with solvent signal
		5.8-4.7 (H2)	(yes)	low signal intensity
		3.0-1.0 (H3-H7)	(yes)	Interference with matrix effects possible
		0.9-0.7 (H8)	no	Interference with matrix effects possible, low signal intensity
CDCl₃/FA	solvent	7.25 (CDCl ₃)		
	Matrix	1.4-0.6		
	PA 6.6	3.2-3.0 (H1)	yes	-
		2.3-2.15 (H2)	(yes)	interference with matrix effects possible
		1.65-1.1 (H3-H5)	(yes)	interference with matrix effects possible
		7.31 (amines)	(yes)	low signal intensity

83 *signal is not suitable.*