

**Supporting Information for:**

**A Fluorescent Probe for Monitoring PTP-PEST Enzymatic Activity**

Garrett R. Casey<sup>1,2</sup> and Cliff I. Stains<sup>1,3,4,5,6\*</sup>

<sup>1</sup>Department of Chemistry, University of Nebraska – Lincoln, Lincoln, NE 68588 (USA)

<sup>2</sup>Department of Chemistry and Physics, Southeast Missouri State University, Cape Girardeau, MO 63701 (USA)

<sup>3</sup>Department of Chemistry, University of Virginia, Charlottesville, VA 22904 (USA)

<sup>4</sup>Nebraska Center for Integrated Biomolecular Communication, University of Nebraska – Lincoln, Lincoln, NE 68588 (USA)

<sup>5</sup>Cancer Genes and Molecular Regulation Program, Fred & Pamela Buffet Cancer Center, University of Nebraska Medical Center, Omaha NE 68198 (USA)

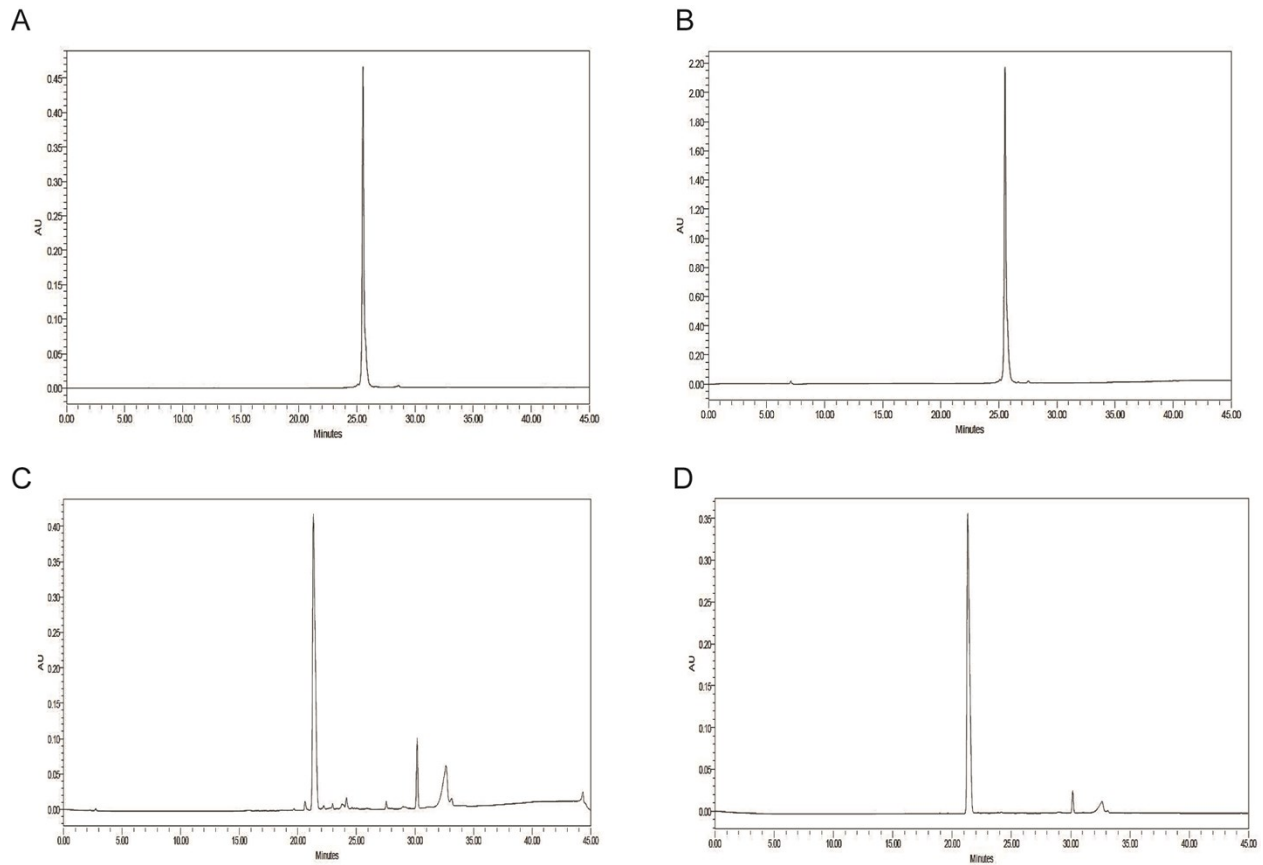
<sup>6</sup>University of Virginia Cancer Center, University of Virginia, Charlottesville, VA, 22904 (USA)

\*E-mail: [cstains@virginia.edu](mailto:cstains@virginia.edu)

**Contents**

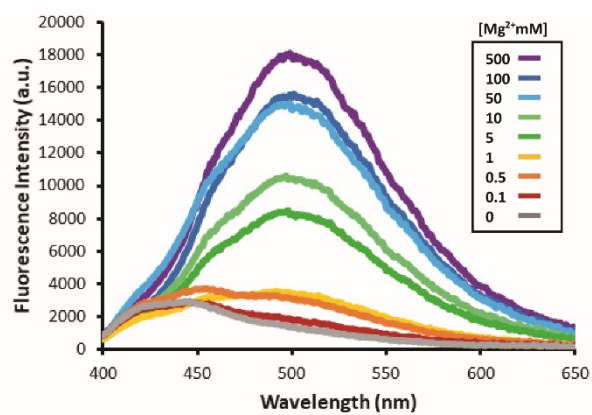
Figure S1 .....	2
Figure S2 .....	3
Figure S3 .....	4
Figure S4 .....	5
Figure S5 .....	6
Figure S6 .....	7
Table S1 .....	8
Table S2 .....	9

**Figure S1**



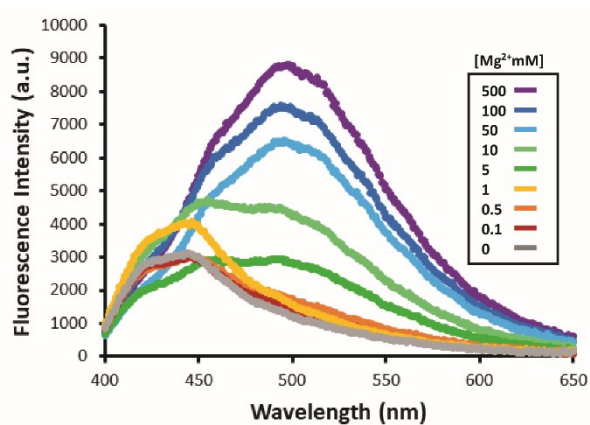
HPLC chromatograms for PTP-PEST peptides. pPEST1tide (A,B) and PEST1tide (C,D). Samples were monitored at 228 nm (A,C) and 316 nm (B,D). Buffer A is nanopure H<sub>2</sub>O with 0.1% (v/v) TFA and buffer B is acetonitrile with 0.1% (v/v) TFA. Peptides were separated using a linear gradient of buffer B from 5 to 95% (v/v) between 5 and 35 minutes with a flow rate of 1 mL/min on a C<sub>18</sub> analytical column (YMC, AA12S05-2546WT).

**Figure S2**



Emission spectra for pPEST1tide (1 μM) in the presence of increasing concentrations of Mg<sup>2+</sup>.

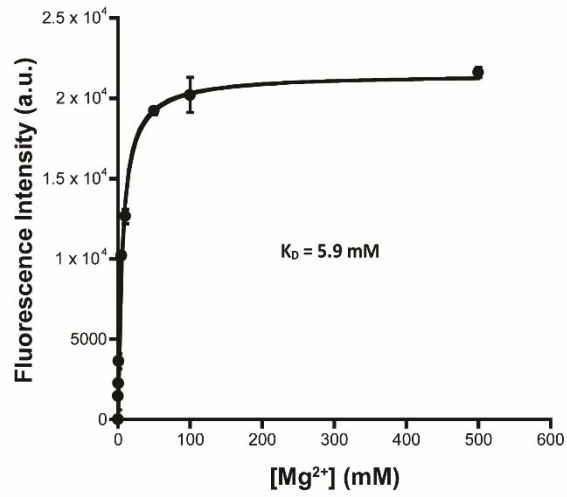
**Figure S3**



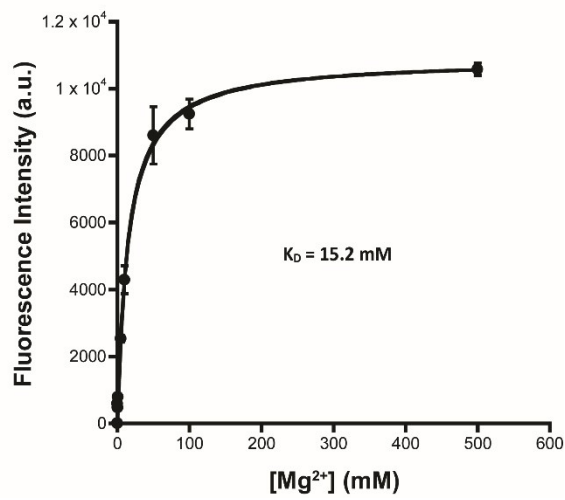
Emission spectra for PEST1tide (1 μM) in the presence of increasing concentrations of Mg<sup>2+</sup>.

Figure S4

A



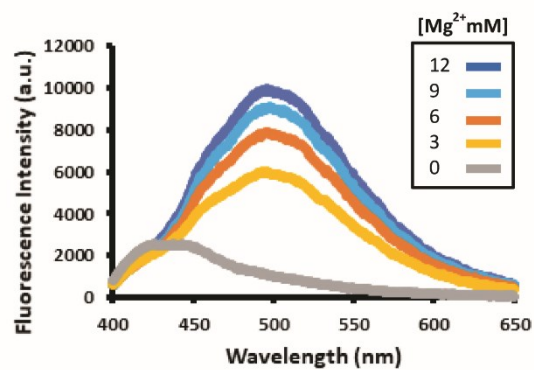
B



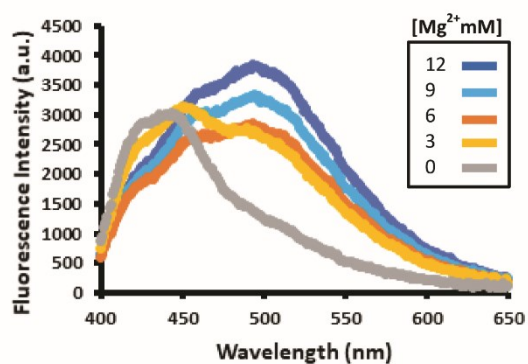
$Mg^{2+}$   $K_D$  determination for pPEST1tide (1  $\mu$ M, A) and PEST1tide (1  $\mu$ M, B). Error bars represent the standard deviation of triplicate experiments.

Figure S5

A



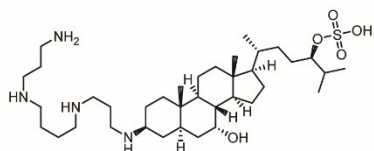
B



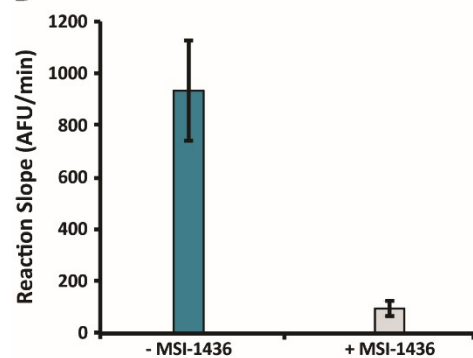
Emission spectra with increasing concentrations of Mg<sup>2+</sup> for pPEST1tide (A) and PEST1tide (B). Peptides (10  $\mu$ M) were incubated with the indicated concentrations of Mg<sup>2+</sup>.

**Figure S6**

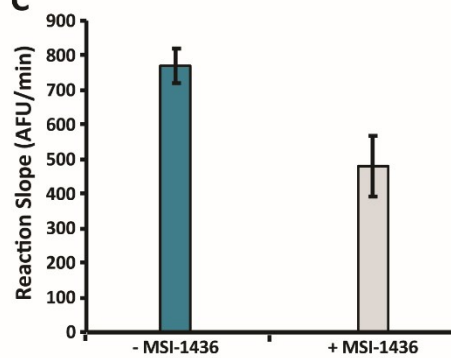
**A**



**B**



**C**



MSI-1436 selectively inhibits PTP1B activity. (A) The structure of MSI-1436. Recombinant PTP1B (2.5 nM, B) or PTP-PEST (2.5 nM, C) in the absence or presence of MSI-1436 (10  $\mu$ M) assayed with 100  $\mu$ M DiFMUP. Error bars represent the standard deviation of triplicate experiments.

**Table S1**

ESI-MS data for PTP-PEST peptides.

Peptide	Expected Mass (M+H) <sup>+</sup>	Observed Mass (M+H) <sup>+</sup>
pPEST1tide	1731.7	1731.2
PEST1tide	1651.7	1652.4



**Table S2**

Fold fluorescence increases of pPEST1tide at varying  $Mg^{2+}$  concentrations.

$[Mg^{2+}]$ (mM) <sup>a</sup>	Fold Fluorescence Increase <sup>b</sup>
12	3.6
9	4.0
6	4.4
3	3.6
0	-

<sup>a</sup>Concentrations of  $Mg^{2+}$  represent 0.5, 1.0, 1.5, and 2.0 times the  $Mg^{2+}$   $K_D$  for the substrate peptide. <sup>b</sup>The fold fluorescence increase is defined as the average fluorescence of the substrate peptide divided by the average fluorescence of the non-phosphorylated peptide assayed at the indicated concentration of  $Mg^{2+}$ .