

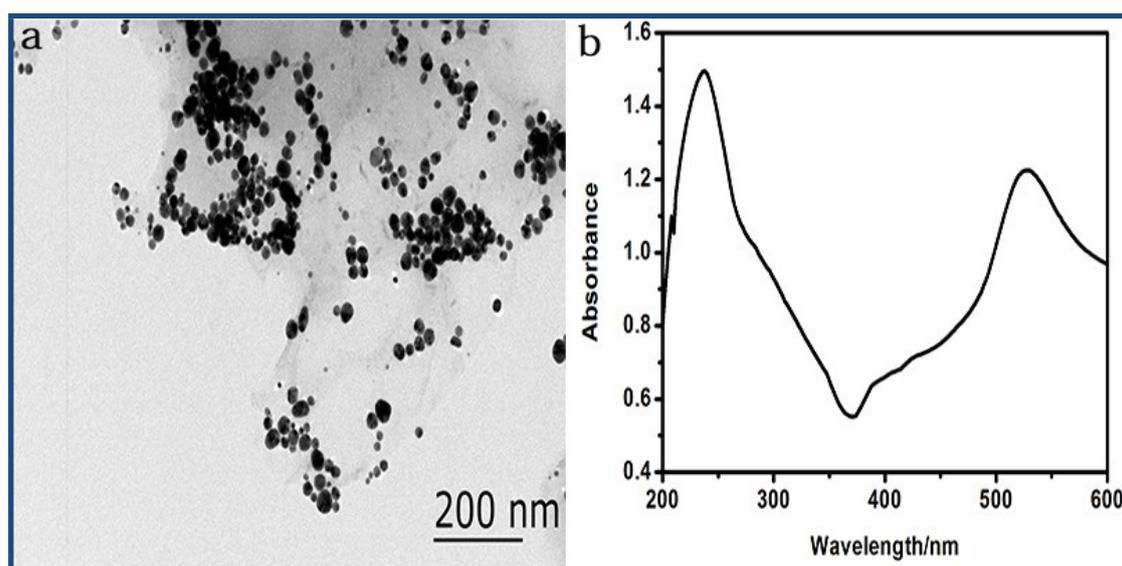
## Supporting Materials

Electrogenerated chemiluminescence resonance energy transfer  
between ZnGa<sub>2</sub>O<sub>4</sub>/g-C<sub>3</sub>N<sub>4</sub> and gold nanoparticles/graphene and  
its application in the detection of thrombin

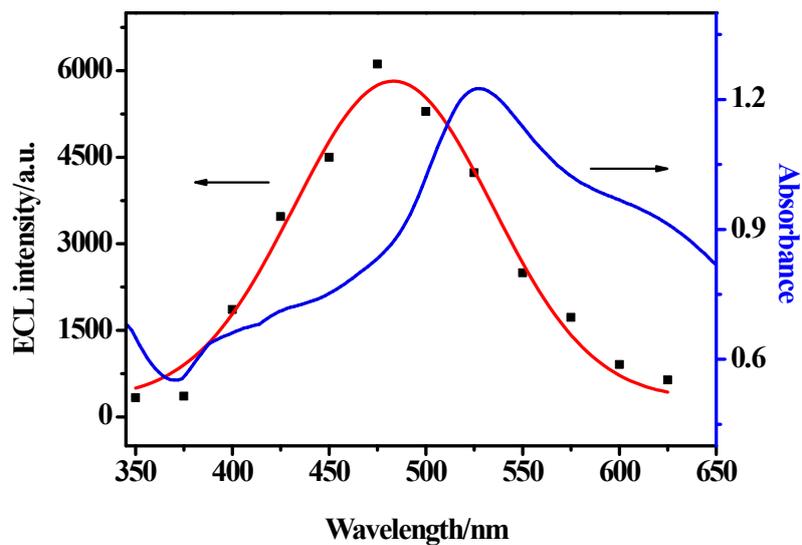
Hui Liu, Hao Yin, TingTing Yang, HouCheng Ding, YongPing Dong\*

*School of Chemistry and Chemical Engineering, Anhui Province Key Laboratory  
of Coal Clean Conversion and High Valued Utilization, Hexian Development Institute  
of Chemical Industry, Anhui University of Technology, Maanshan243002, China*

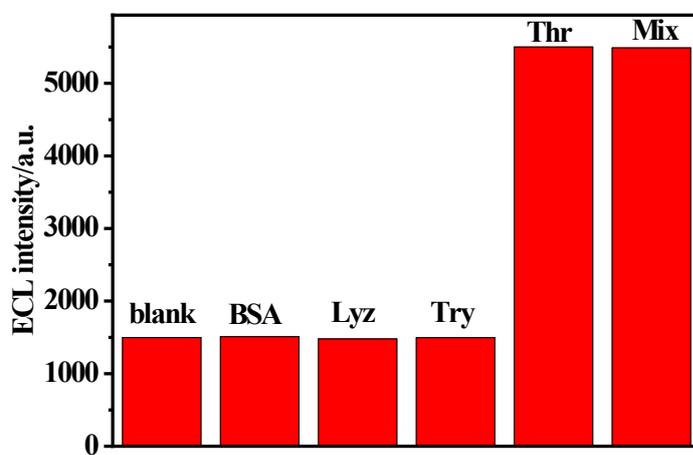
**Electrochemical and ECL measurements.** The electrochemical measurements were recorded with CHI 660D electrochemical workstation (CH Instruments Co., China). A conventional three-electrode system was adopted, including a modified GCE as the working electrode, a platinum wire as the counter electrode and a saturated calomel electrode (SCE) as the reference electrode, respectively. Electrochemical impedance spectroscopy (EIS) was carried out at open circuit potential in 0.1 mol L<sup>-1</sup> KCl solution containing K<sub>3</sub>[Fe(CN)<sub>6</sub>]/K<sub>4</sub>[Fe(CN)<sub>6</sub>] (5 mM, 1:1). The frequency range was selected as 0.01 Hz-100 kHz, and potential amplitude was 5 mV. The ECL measurement was carried out on a model MPI-M electrochemiluminescence analyzer (Xi'An Remax Electronic Science & Technology Co. Ltd., China) at room temperature, and the voltage of the photomultiplier tube (PMT) was set at -800V during the detection. A commercial 5 mL cylindroid glass cell was used as ECL cell and was placed directly in front of the PMT.



**Fig.S1** TEM image (a) and UV-vis absorption spectrum (b) of gold nanoparticles/graphene composite



**Fig.S2** ECL spectrum of ZnGa<sub>2</sub>O<sub>4</sub>/g-C<sub>3</sub>N<sub>4</sub> nanocomposites and UV-vis absorption spectrum of gold nanoparticles/graphene nanocomposites.



**Fig.S3** Selectivity of the fabricated sensor.

**Table S1** Comparison of different ECL biosensors for the detection of thrombin.

ECL system	Linear range (pM)	Detection limit (pM)	Ref.
CdTe QDs-S <sub>2</sub> O <sub>8</sub> <sup>2-</sup>	0.5 -800	0.35	1
Ru(phen) <sub>3</sub> <sup>2+</sup> -TPA	0.05 -50	0.02	2
Ru(bpy) <sub>3</sub> <sup>2+</sup> -TPA	0.05 -100	0.008	3
Ru(bpy) <sub>3</sub> <sup>2+</sup> -TPA	0.90 -226	0.40	4
Luminol-H <sub>2</sub> O <sub>2</sub>	5-50000	1.7	5
Luminol-QDs	0.01-100	0.0014	6
Pd NCs-TPA	0.01-100	0.00676	7
RuAg/SiO <sub>2</sub> NPs	0.002-2	0.001	8
ZnGa <sub>2</sub> O <sub>4</sub> /g-C <sub>3</sub> N <sub>4</sub>	0.00137-27.4	0.00055	This work

[1] Y. Chen, B. Y. Jiang, Y. Xiang, Y. Q. Chai and R. Yuan, *Chem. Commun.* 2011, **47**, 7758-7760.

[2] X. B. Yin, Y. Y. Xin and Y. Zhao, *Anal. Chem.* 2009, **81**, 9299-9305.

[3] J. Zhang, P. P. Chen, X. Y. Wu, J. H. Chen, L. J. Xu, G. N. Chen and F. F. Fu, *Biosens. Bioelectron.* 2011, **26**, 2645-2650.

[4] X. Y. Wang, A. Gao, C. C. Lu, X. W. He and X. B. Yin, *Biosens. Bioelectron.* 2013, **48**, 120-125.

[5] F. Li and H. Cui, *Biosens. Bioelectron.* 2013, **39**, 261-267.

[6] Y. P. Dong, T. T. Gao, Y. Zhou and J. J. Zhu, *Anal. Chem.* 2014, **86**, 11373-11379.

[7] H. M. Wang, Y. Fang, P. X. Yuan, A. J. Wang, X. L. Luo and J. J. Feng, *Electrochim. Acta*, 2019, **310**, 195-202.

[8] Y. Q. Sun, X. Jin, M. Gong, L. R. Lv, L. Y. Li, M. Jiang, X. Y. Wang and J. Xu, *Electroanalysis*, 2019, **31**, 1-11.