

Supplementary information for:

Label-free analysis of gingival crevicular fluid (GCF) by surface enhanced Raman scattering (SERS)

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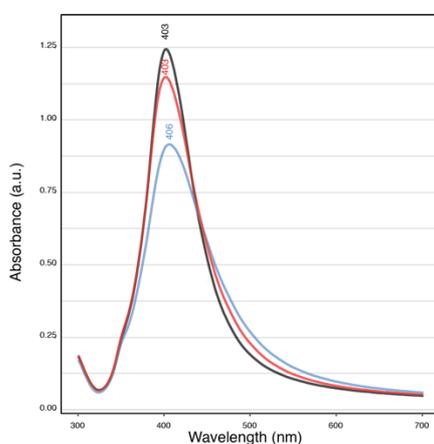


Figure S1 Representative UV-Vis absorption spectra of three different batches of Ag nanoparticles respectively synthesized by a modified Lee-Meisel (Lee and Meisel, *J. Phys. Chem.* 1982, 86 (17), 3391-3395) method. Spectra were chosen to represent the spectral variation between batches, which in some cases was indicative of the presence of some nanoparticle aggregates. The extinction maxima were 403-406 nm, which, according to literature (see for instance Steinigeweg and Schluecker, *Chem.Comm.* 2012, 48, 8682-8684; Wan et al. *J. Colloid Interf. Sci.* 2013, 394, 263-268, but there are many others which characterized the Lee-Meisel colloids), corresponds to nanoparticles having a variety of shapes and a broad size distribution (40-100 nm).

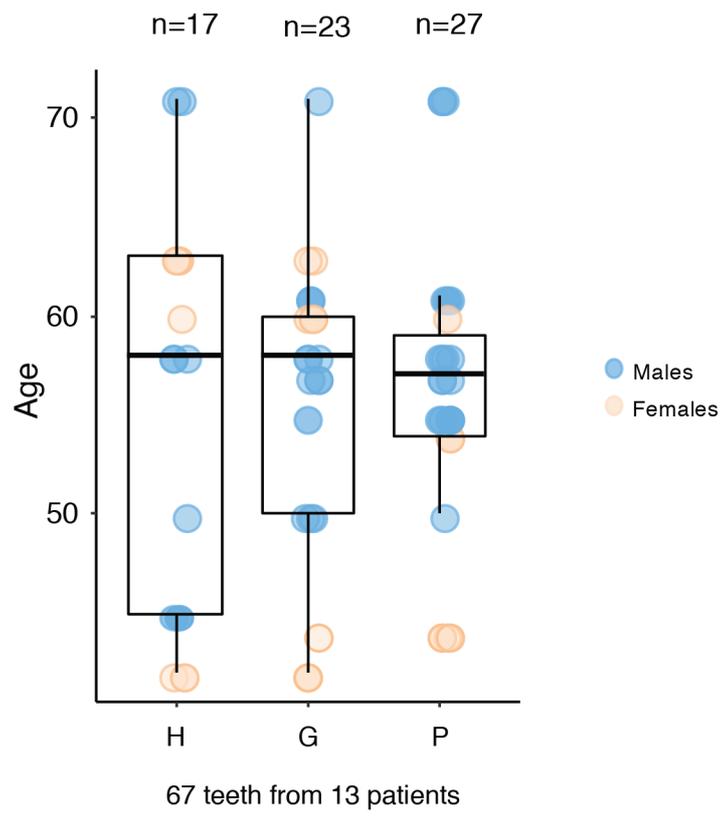


Figure S2 Age distribution

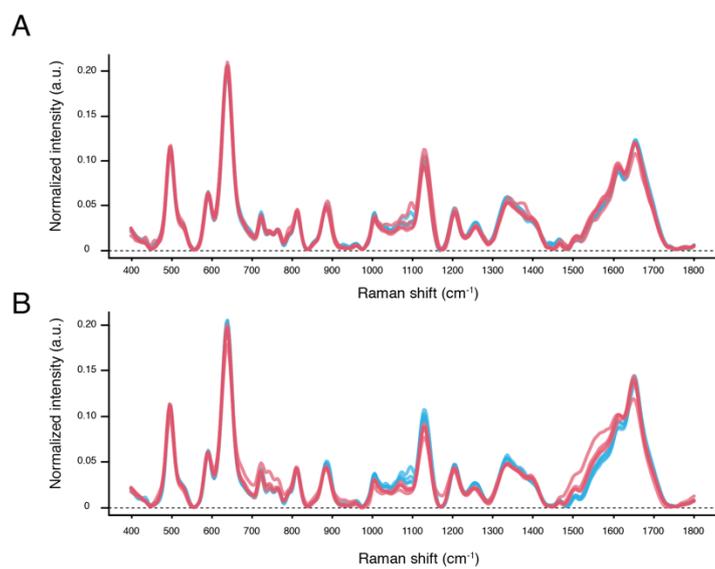


Figure S3 Stability of the SERS signal of human serum (ERM[®] certified Reference Material) on our substrate after one hour (A), and 48 hours (B). Four technical replicates from two different paper strips are shown overlaid. All spectra were collected with a 785 nm excitation.

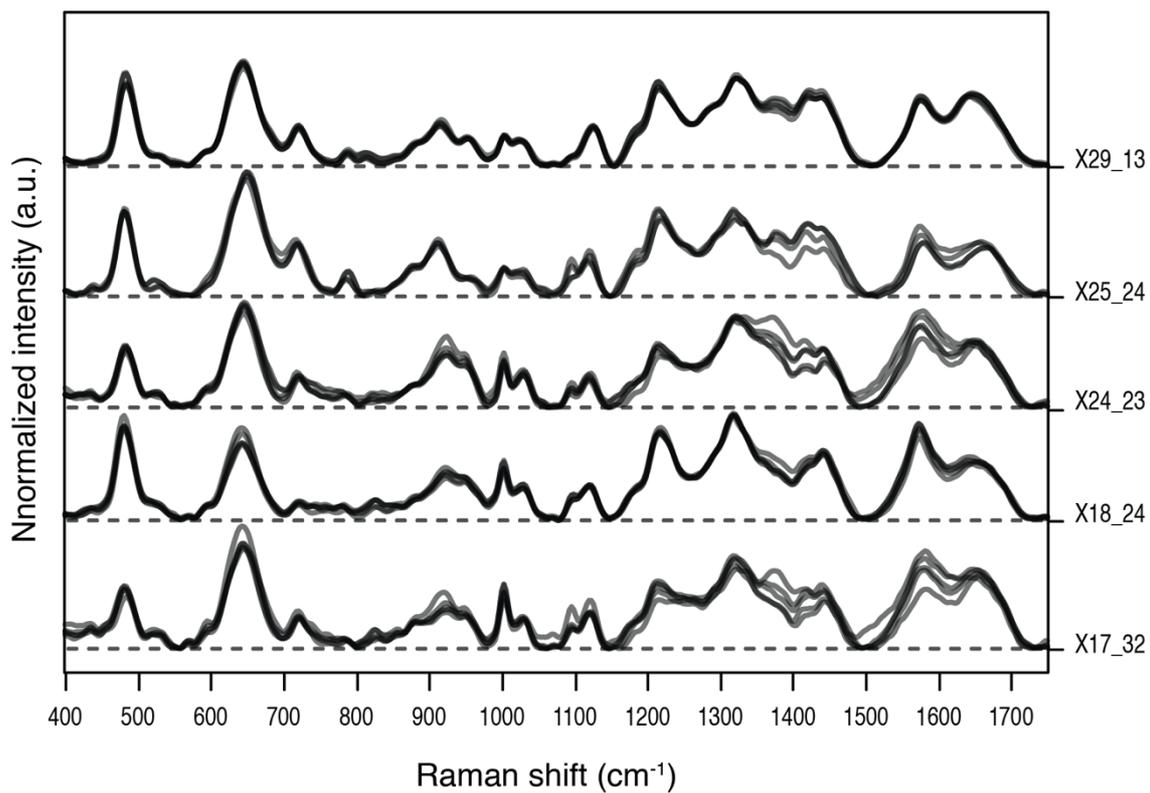


Figure S4 Intra-substrate variability. Five spectra (taken from different positions randomly selected on the substrates surface) from five different samples are shown overlaid. All spectra were collected with a 785 nm excitation. Each sample is labelled with a "patient_element" code.

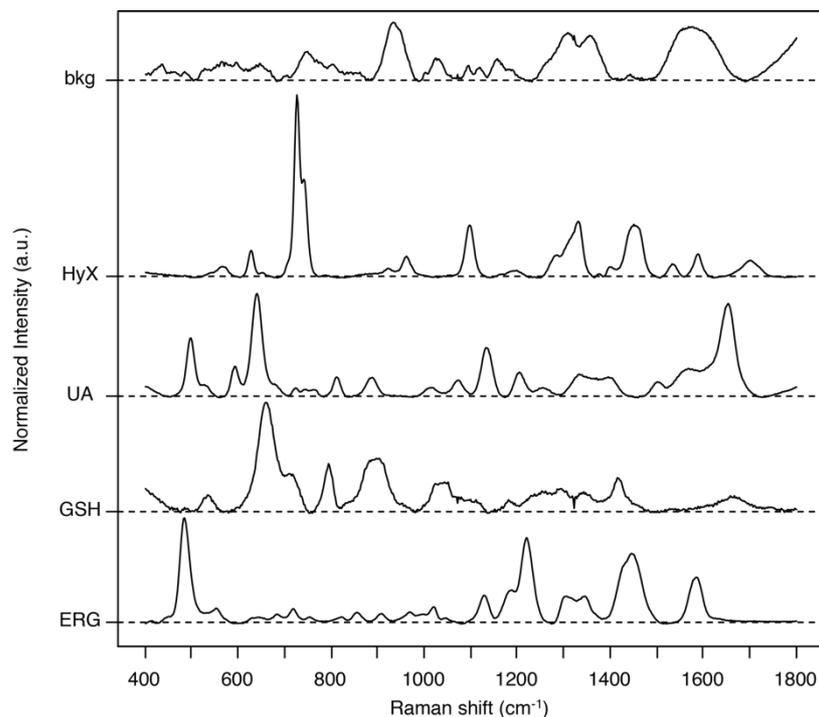


Figure S5 Normalized Surface-enhanced Raman scattering (SERS) spectra of reference compounds. ERG, ergothioneine; GSH, reduced glutathione; UA, uric acid; HyX, hypoxanthine; bkg, naked Periopaper strip covered with Ag NPs.

Supplementary methods

To evaluate the performance of the spectral fitting, the lack of fit (LoF) explained variance (R^2) and relative fitting error (RFE) were calculated as follows:

$$LOF = \sqrt{\frac{\sum_{i=1}^I \sum_{j=1}^J (g_{ij} - \hat{g}_{ij})^2}{\sum_{i=1}^I \sum_{j=1}^J g_{ij}^2}} \times 100$$

$$R^2 = \left(1 - \frac{\sum_{i=1}^I \sum_{j=1}^J (g_{ij} - \hat{g}_{ij})^2}{\sum_{i=1}^I \sum_{j=1}^J g_{ij}^2} \right) \times 100$$

Where g_{ij} is the original elements of the matrix, with I rows and J columns, and \hat{g}_{ij} is the elements of the matrix predicted by the fitting method. Although LOF and R^2 have the same interpretation, LOF is more sensitive to fitting differences.

Another common approach to quantify the reliability of a spectral fitting routine is to compare the norm of the fit to the norm of the input signal, a metric that has been termed the relative fitting error, RFE:

$$RFE = \frac{\|S - R\|}{\|S\|}$$

where S is the measured spectral data and R are the residuals (minimized by least-squares algorithm). $\| \cdot \|$ indicates the norm of the vector. An RFE of 100% indicates a perfect fit and approaches zero as the fit degrades (i.e. the optimal fit lacks the ability to represent the input signal as a linearly weighted sum of the references supplied).

Table T1 Quality metrics to evaluate the performance of the spectral fitting

Quality metrics	value
LoF (%)	4.67 (1.75 – 8.73)
R ² (%)	99.07 (98.52 – 99.60)
RFE (%)	97.10 (96.94 – 99.15)
LoF, Lack of Fit; R ² , explained variance; RFE, relative fitting error	

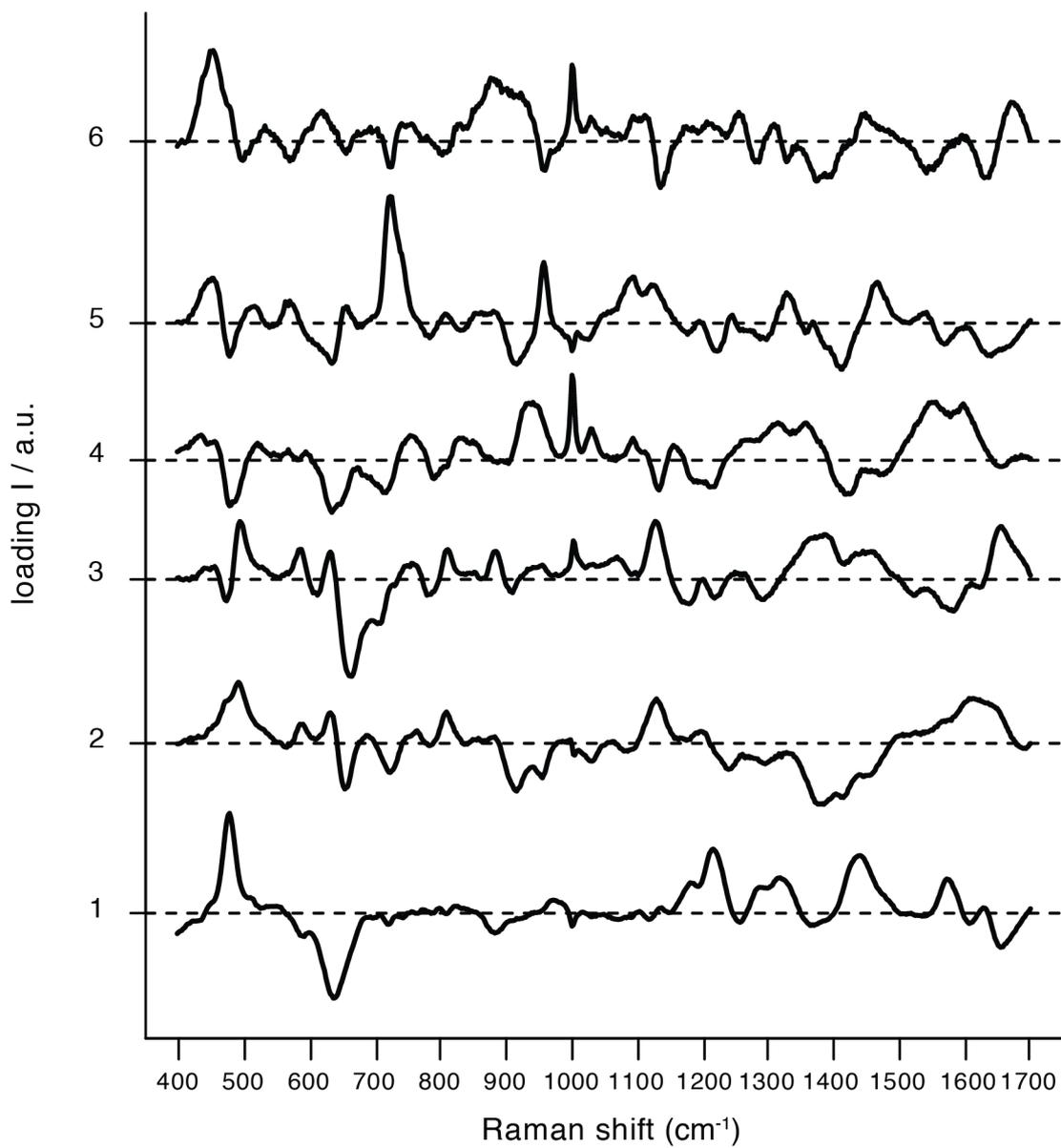


Figure S7 Loadings for first six principal components.

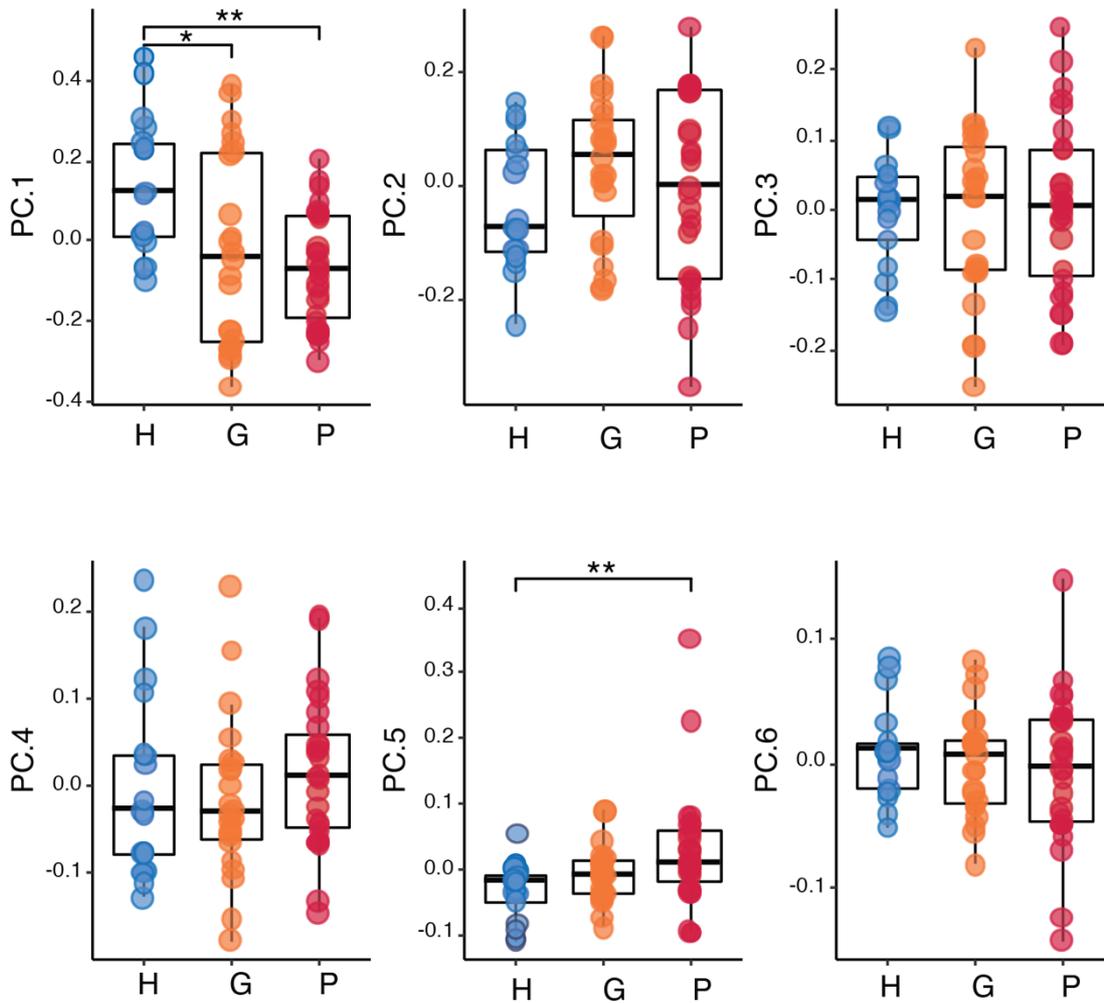


Figure S8. Box plot for the scores the first six principal components. The black line represents the median; top and bottom edges of the box are the upper and lower quartiles, whiskers extend to upper and lower quartiles plus and minus 1.5× the IQR. Kruskal-Wallis test with Dunn–Bonferroni correction for multiple comparisons test (confidence level 95%, * = $p < 0.05$, ** = $p < 0.01$) reveal a significant difference between the H and the other two groups.