

Supporting Information for

**Tryptamine-functionalized magnetic nanoparticles for highly
sensitive detection of *Salmonella* Typhimurium**

Table S1. Specific sequence and primers we used to identify DNA of *S. Typhimurium*.

Target	Sequences (5'-3')	T _m (°C)	Product size (bp)
<i>S. Typhimurium-Forward</i>	5'-TATCGCCACGTTCGGGCAA-3'	59.7	275
<i>S. Typhimurium-Reverse</i>	5'-TCGCACCGTCAAAGGAACC-3'	58.1	

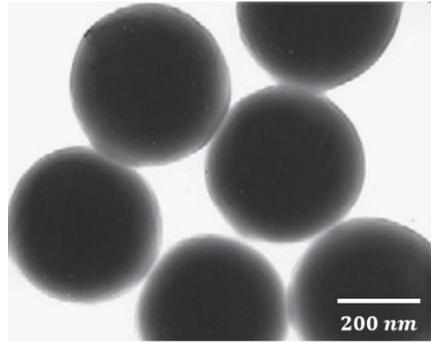


Figure S1. A TEM image of Indole@MNPs

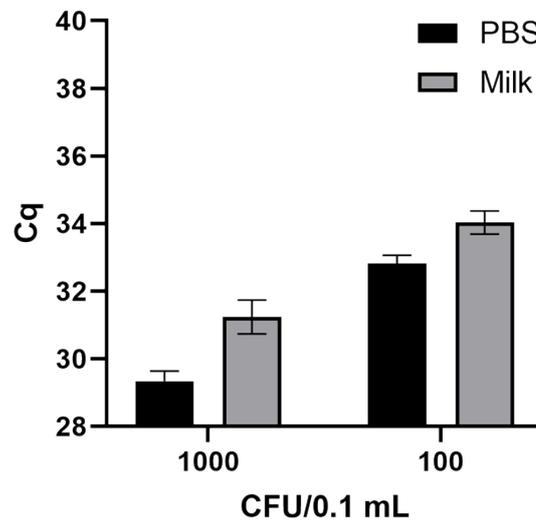


Figure S2. Evaluation of interference of milk during sample preparation using commercial kit. No Cq values were obtained after amplification of no-template controls (NTCs) in any experiments. Error bars indicate standard deviation from mean based on three independent experiments.

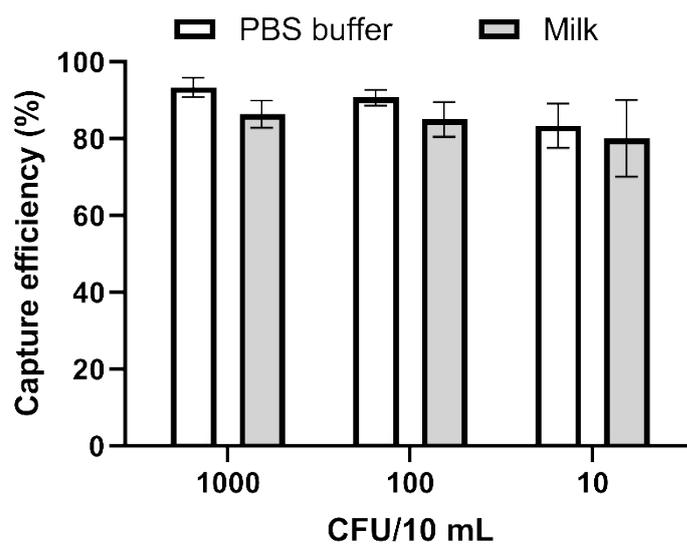


Figure S3. Capture efficiencies of Indole@MNPs with concentrations of *S. Typhimurium* (1000, 100, 10 CFU in 10 mL PBS buffer and milk). The original samples without binding with Indole@MNPs were counted as 100 % and served as controls. Error bars indicate standard deviation from mean based on three independent experiments.

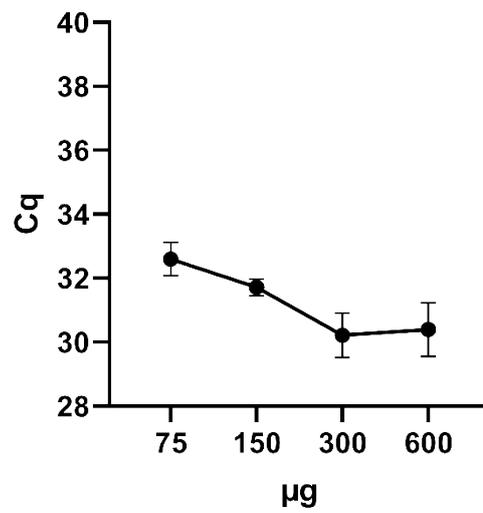


Figure S4. Evaluation of different Indole@MNP amounts in 10 mL PBS sample using 10^3 CFU of *S. Typhimurium*. Error bars indicate standard deviation from mean based on three independent experiments.

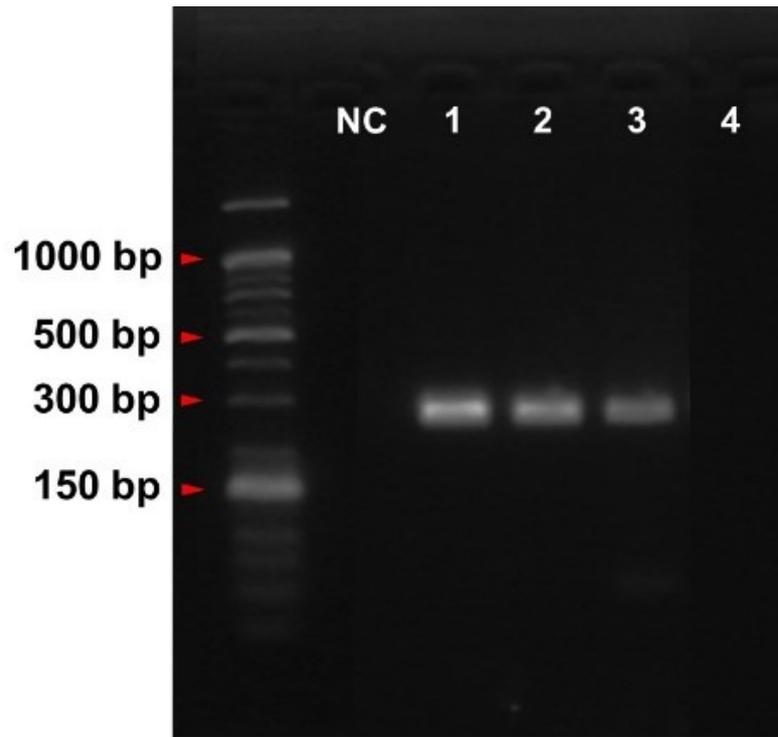


Figure S5. Gel electrophoresis image of PCR product. 25/100 bp mixed DNA ladder (Bioneer Inc.); NC: negative control with no DNA template; Lane 1: 1000 CFU/10 mL sample; Lane 2: 100 CFU/10 mL sample; Lane 3: 10 CFU/10 mL sample; Lane 4: 5 CFU/10 mL sample.

DLVO model

The potential interaction energy profile between *S. Typhimurium* and Indole@MNP was calculated based on the DLVO theory. [1-4] Following equations were used to make the computation:

$$V_{Tot} = V_{LW} + V_{EL}$$

$$V_{LW} = -\frac{A(a_1 a_2)}{6d(a_1 + a_2)}$$

$$V_{EL} = \frac{\pi \epsilon a_1 a_2 (\zeta_1^2 + \zeta_2^2)}{(a_1 + a_2)} \left[\frac{2\zeta_1 \zeta_2}{\zeta_1^2 + \zeta_2^2} \ln \frac{1 + \exp(-kd)}{1 - \exp(-kd)} + \ln \{1 - \exp(-2kd)\} \right]$$

V_{LW} = Electrical double layer repulsive energy

V_{EL} = Van-der-Waals attractive energy

V_{Tot} = DLVO energy barrier

A = Hamaker constant where $A_{132} = (\sqrt{A_1} - \sqrt{A_3})(\sqrt{A_2} - \sqrt{A_3}) = 1.40 \times 10^{-21} J$ for the interaction between bacterial pathogen ($A_1 = 5.2 \times 10^{-20} J$) and iron oxide nanoparticle ($A_2 = 10 \times 10^{-21} J$) separated by a water medium ($A_3 = 3.7 \times 10^{-20} J$).

a_1 = Radii of microbial cells; 770 nm for *Salmonella Typhimurium*

a_2 = Radii of Indole@MNP (iron oxide nanoparticle) = 425 nm

d = Separation distance: 0–3000 nm (dependent variable)

$\epsilon = \epsilon_0 \times \epsilon_r = 7.083 \times 10^{-10} F/m$ where the relative permittivity of the medium, $\epsilon_r = 80$ (for water at 20°C) and ther permittivity of a vacuum, $\epsilon_0 = 8.854 \times 10^{-12} F/m$

ζ_1 = Zeta potential of bacteria where -11.7 mV for *Salmonella Typhimurium*

ζ_2 = Zeta potential of Indole@MNPs (pH 7) = -32 mV

k = Inverse Debye-Huckel length = 729 nm for pH 6.5

References:

- (1) G. Hwang, I. S. Ahn, B. J. Mhin, J. Y. Kim, *Colloids Surf., B* **2012** 97, 138–144.
- (2) H. Ohshima, Interaction of colloidal particles. In *Colloid and interface science in pharmaceutical research and development*; Ohshima, H., Makino, K., Eds.; Elsevier, New York, 2002; pp 1–28.
- (3) J. Israelachvili, *Intermolecular and surface forces*. Elsevier, New York 2011.
- (4) B. Faure, G. Salazar-Alvarez, L. Bergström, *Langmuir* **2011** 27, 8659-8664.

Table S2. An overview on recently reported MNP-based methods for the detection of *Salmonella* species.

Materials used	Detection method	Detection limit	References
Antibody functionalized MNPs	Multiplex PCR	10 CFU/ml (Pure culture) 100 CFU/ml (Meat)	[1]
Antibody functionalized MNPs	Fluorescence (DNA-QD-AuNP)	13.6 CFU/ml (PBS)	[2]
Antibody functionalized MNP chains	Electrochemistry	10 CFU/ml (Milk)	[3]
Aptamer functionalized MNPs	Chemiluminescence (Rolling circle amplification)	10 CFU/mL (PBS)	[4]
Aptamer functionalized MNPs	SERS	15 CFU/mL (PBS)	[5]
Oligonucleotide functionalized MNPs	RT-PCR	10 CFU/mL (Milk)	[6]
Indole functionalized MNPs	qPCR	10 CFU/10 mL (Milk)	This study

- [1] F. Li, F. Li, B. Chen, B. Zhou, P. Yu, S. Yu, W. Lai, H. Xu, *Food Control*, 2017, **73**, 587-594.
- [2] Q. Wang, X. Cheng, H. Li, F. Yu, Q. Wang, M. Yu, D. Liu, J. Xia, *Materials Today Communications*, 2020, **25**, 101428.
- [3] Y. Hou, W. Tang, W. Qi, X. Guo, J. Lin, *Biosensors and Bioelectronics*, 2020, **157**, 112160.
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- [6] Y. Bai, Y. Cui, Y. Suo, C. Shi, D. Wang, X. Shi, *Frontiers in microbiology*, 2019, **10**, 770.