Supplementary Information

A Fluorescent Probe Based on Reversible Michael Addition– elimination Reaction for the Cycle Between Cysteine and H₂O₂

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Materials and instrumentation

All chemical reagents and materials used were purchased from commercial companies and used without further purification. Doubly distilled water was used throughout the experiments. Silica gel (200–300 mesh, Qingdao Haiyang Chemical Co.) was used for column chromatography. NMR spectra were measured on Bruker Avance III at 400 MHz for ¹H NMR and at 100 MHz for ¹³C NMR with chemical shifts reported as ppm (in DMSO-d6, TMS as internal standard). Mass spectra (MS) were recorded with Bruker Apex IV FTMS using electrospray ionization (ESI). Absorption spectra were recorded by a Purkinje TU-1901 spectrophotometer. Fluorescence data were taken through a Hitachi F-7000 fluorescence spectrometer with a 1 cm standard quartz cell. pH measurements were carried out with a pH acidometer (Mettler Toledo FE-30). Cell imaging was performed using a confocal laser scanning microscope (OLYPUS) with a 60× objective lens.



Scheme S1. Synthesis of Cys 1.



Figure S1. (A)Fluorescent spectra changes of the probe (10 μ M Cys1) and (b)fluorescence intensity of the probe (10 μ M Cys1) in 508 nm, recorded over 40 min. Each data was recorded in PBS buffer (10 mM, pH = 7.4, with 10% DMSO), $\lambda_{ex} = 460$ nm, slit width: $d_{ex} = d_{em} = 5$ nm.



Figure S2. MS Spectra of Cys 1 (upper) and the reaction addictive product (lower).



Figure S3. Fluorescence kinetics ratio intensity (F_{508}) responses of probe (10 µM) upon addition of 100 µM Cysteine in 10 mM, pH 7.4 PBS with 10% DMSO at room temperature. $\lambda_{ex} = 460$ nm, slit width: $d_{ex} = d_{em} = 5$ nm.



Figure S4. Fluorescence responses of probe (10 μ M) in the absence and presence of Cysteine (100 μ M) under different pHs with 10% DMSO at room temperature. $\lambda_{ex} = 460$ nm, slit width: $d_{ex} = d_{em} = 5$ nm.



Figure S5. (a) Fluorescent spectra responses of **Cys 1** (10 μ M) in the absence various analytes (100 μ M);(b) Fluorescent spectra responses of **Cys 1** (10 μ M) in the absence various analytes (100 μ M) and Cysteine (100 μ M);(c)Red column: Fluorescent intensity at 508nm of **Cys 1** (10 μ M) in the absence various analytes (100 μ M); Black column: Fluorescent intensity at 508nm of **Cys 1** (10 μ M) in the absence various analytes (100 μ M); Black column: Fluorescent intensity at 508nm of **Cys 1** (10 μ M) in the absence various analytes (100 μ M); Black column: Fluorescent intensity at 508nm of **Cys 1** (10 μ M) in the absence various analytes (100 μ M); Black column: Fluorescent intensity at 508nm of **Cys 1** (10 μ M) in the absence various analytes (100 μ M) and Cysteine (100 μ M). 0. Probe itself, 1. Ala, 2. Asn, 3. Glu, 4. Lys, 5. Gly, 6. Thr, 7. Tyr, 8. Fe²⁺, 9. Fe³⁺, 10. HSO3⁻, 11. SO3⁻, 12. SO4²⁻, 13. NO2⁻, 14. NO3⁻, 15. HClO, 16. ONOO⁻, 17. OH, 18. GSH, 19. Hcy, 20. Cys.



Figure S6. LC-MS analysis of the H_2O_2 reaction product. The LC spectrum(A) and mass spectrum(B) of **Cys 1**, the LC spectrum(C) and mass spectrum(D) of H_2O_2 reaction product.



Figure S7. Cell viability with Hela cells, which were cultured in the presence of 0-20 μ M Cys 1 for 24 h.

NMR and MS Spectra







Figure S9. ¹H NMR Spectra of Compound 1



Figure S10. ¹H NMR Spectra of Cys 1



Figure S11. ¹³C NMR Spectra of Cys 1