

## Electronic Supplementary Material for $\beta$ -FeOOH/Fe-TiO<sub>2</sub> heterojunctions on Ti for bacterial inactivation under light irradiation and biosealing

Kai Li, Yang Xue, Lan Zhang\*, Yong Han\*

State-key Laboratory for Mechanical Behavior of Materials, Xi'an Jiaotong University, Xi'an  
710049, China

### Preparation of Fe-TiO<sub>2</sub> and MFC coatings on Ti.

Ti plates ( $\phi$  14 mm) were MAOed at a voltage of 480 V and pulse frequency of 500 Hz for 2 min in an aqueous electrolyte containing various amounts of EDTA-FeNa and 0.02 M  $\beta$ -glycerophosphate disodium. The Fe-TiO<sub>2</sub> coatings were designated as Fe1, Fe2, Fe3 and Fe4 according to the EDTA-FeNa concentrations of 0.001M, 0.003 M, 0.006 M and 0.012 M, respectively. With the increased concentration of EDTA-FeNa in the electrolyte, the average content of Fe increased. They are 1.2 wt% on Fe1, 3.1 wt% on Fe2, 6.0 wt% on Fe3 and 11.8 wt% on Fe4, respectively.

In order to confirm the necessity of alkali-heat pretreatment in forming the HFx samples. A part of MC samples were directly HTed in 20 ml of aqueous solution with 0.1 M FeCl<sub>3</sub>·6H<sub>2</sub>O at 100°C for 2 h, and denoted as MFC.

### Preparation of $\beta$ -FeOOH nanoparticles.

Pure  $\beta$ -FeOOH nanoparticles were prepared by HT for 20 ml of aqueous solution with 0.1 M FeCl<sub>3</sub>·6H<sub>2</sub>O at 100°C for 2 h. The obtained precipitates were rinsed with deionized water and dried in a vacuum drying chamber overnight at room temperature.

Table S1 Primers used for qRT-PCR and the corresponding annealing temperatures

Gene	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')	Annealing temperature(°C)	Product size(bp)
IL-1 $\beta$	GCAACTGTCCTGAACTCAACT	ATCTTTGGGGTCCGTCAACT	60	89
IL-6	ACAAAGCCAGAGTCCTTCAGAGAG	TTGGATGGTCTTGGTCCTTAGCCA	62	164
TNF- $\alpha$	CCCTCACACTCAGATCATCTTCT	GCTACGACGTGGGCTACAG	62	61
CTGF	CCTACCGCGTCCCGATCAT	GAGAGCGAGGAGCACCAAG	60	71
Col-I	ACGCCATCAAGGTCTACTGC	CGTACTCGAACGGGAATCCA	59	162
$\alpha$ -SMA	CCTGAAGAGCATCCGACACT	AGAGTCCAGCACAAATACCAGT	62	174
GAPDH	CCACCCTGTGCTGTAGCC	CCCCTCCTCCACCTTGA	60	105

\*Corresponding author, e-mail: lan.zhang@mail.xjtu.edu.cn (Lan Zhang), yonghan@mail.xjtu.edu.cn (Yong Han)  
Tel.: +86 02982665580;

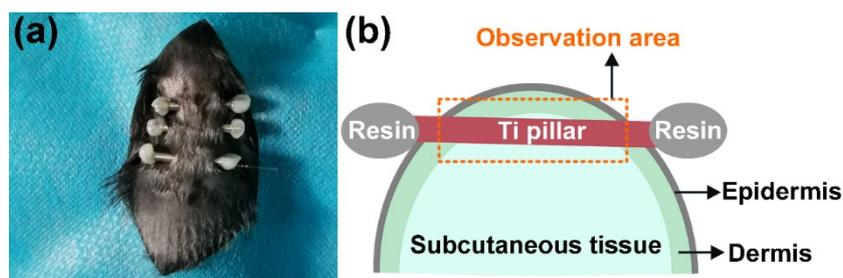


Figure S1 Schematic diagram showing the rectangle-marked region for histological analysis of the coated Ti pillar percutaneously implanted in mice.

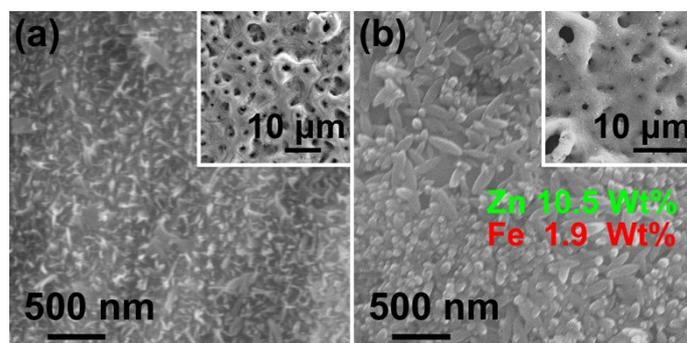


Figure S2(a) SEM morphology of MC after alkali-heat pretreatment and (b) SEM morphology of MFC coatings.

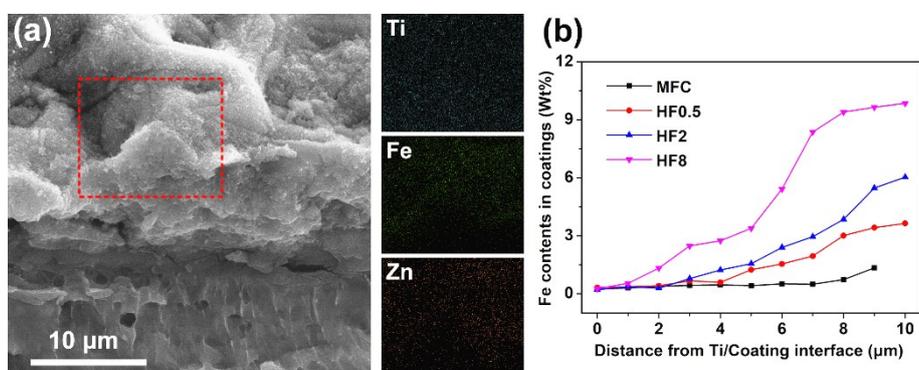


Figure S3(a) Cross-sectional SEM morphologies and the distributions of detected elements (Ti, Zn and Fe) of HF8 coating, (b) Fe profiles in the cross-sections of coatings for different treatments.

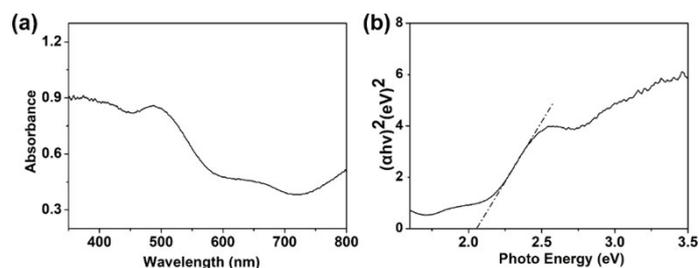


Figure S4(a) UV-vis diffuse reflectance spectra and (b)  $(\alpha h\nu)^2$  as a function of  $h\nu$  calculated based on the band gap energy of  $\beta$ -FeOOH nanoparticles.

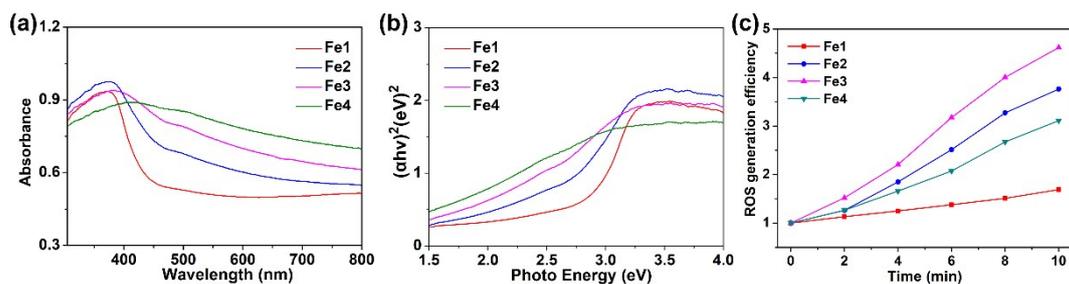


Figure S5(a) UV-vis diffuse reflectance spectra and (b)  $(ahv)^2$  as a function of  $h\nu$  calculated based on the bandgap energies of Fe-TiO<sub>2</sub> coatings, (c) ROS production curves of Fe-TiO<sub>2</sub> samples detected by DCFH-DA under 635 nm light irradiation for 10 min.

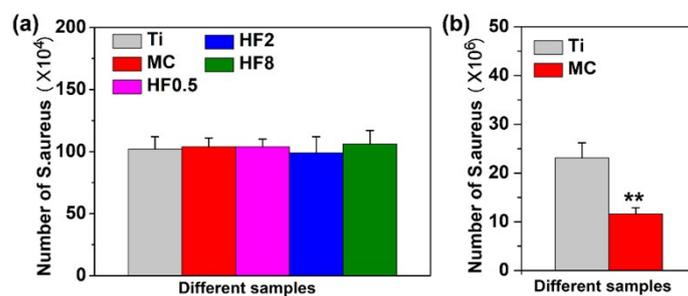


Figure S6 Numbers of *S. aureus* on different samples for 10 min (a) and 24 h (b) in dark. \*\* $p < 0.01$  compared with the Ti control.

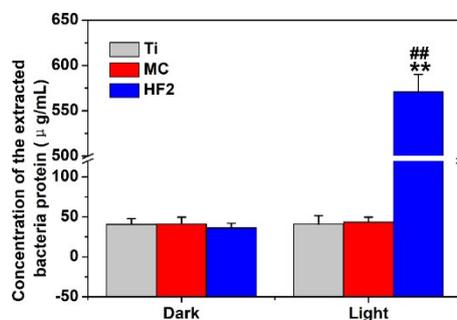


Figure S7 Concentration of the extracted bacteria protein on different surfaces with and without light irradiation for 10 min. (\*\*) $p < 0.01$  compared with the Ti control, and (##) $p < 0.01$  compared with the MC.