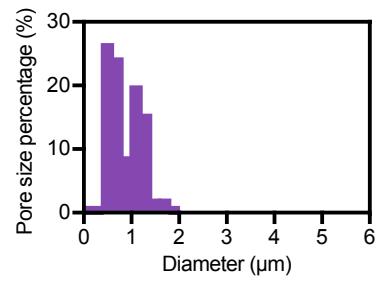


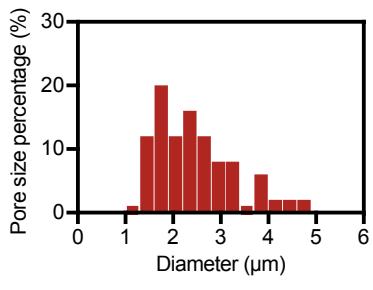
**Figure S1. Oscillating rheology of 2 mM CATCH(+-) hydrogels.** (a-d) Storage and loss modulus of 2 mM CATCH(4+/4-), CATCH(4+/6-), CATCH(6+/4-), and CATCH (6+/6-) at different angular frequencies. The filled circles correspond to the storage modulus and the open circles to the loss modulus. (e) Frequency at which the gel stops behaving as a viscoelastic solid and behaves as a liquid ("gel-sol transition point"). Data are presented as the mean  $\pm$  standard deviation ( $n = 3$ ).

### Average pore size

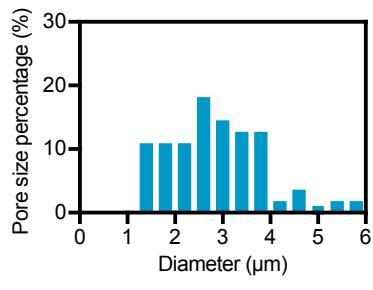
CATCH(4+/4-)



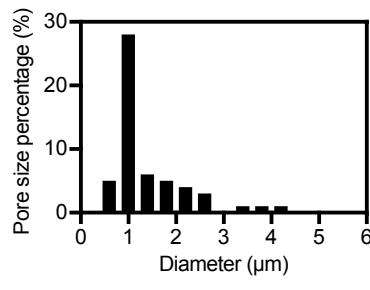
CATCH(4+/6-)



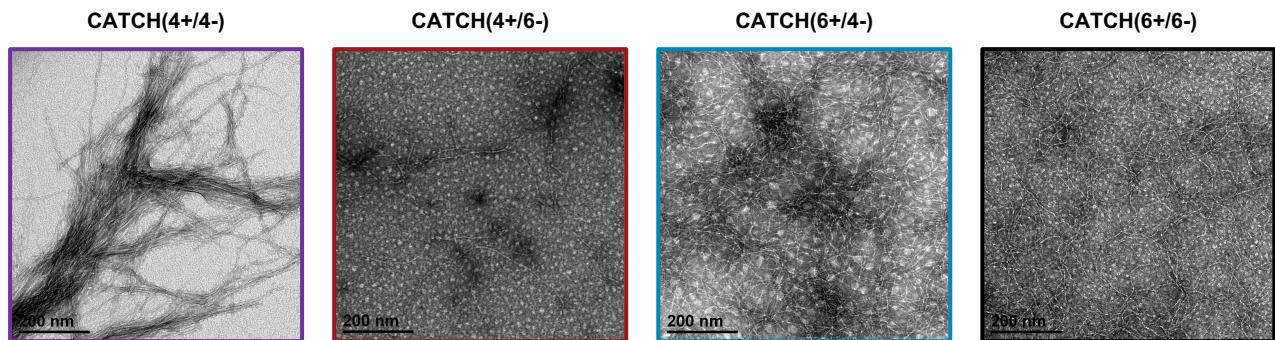
CATCH(6+/4-)



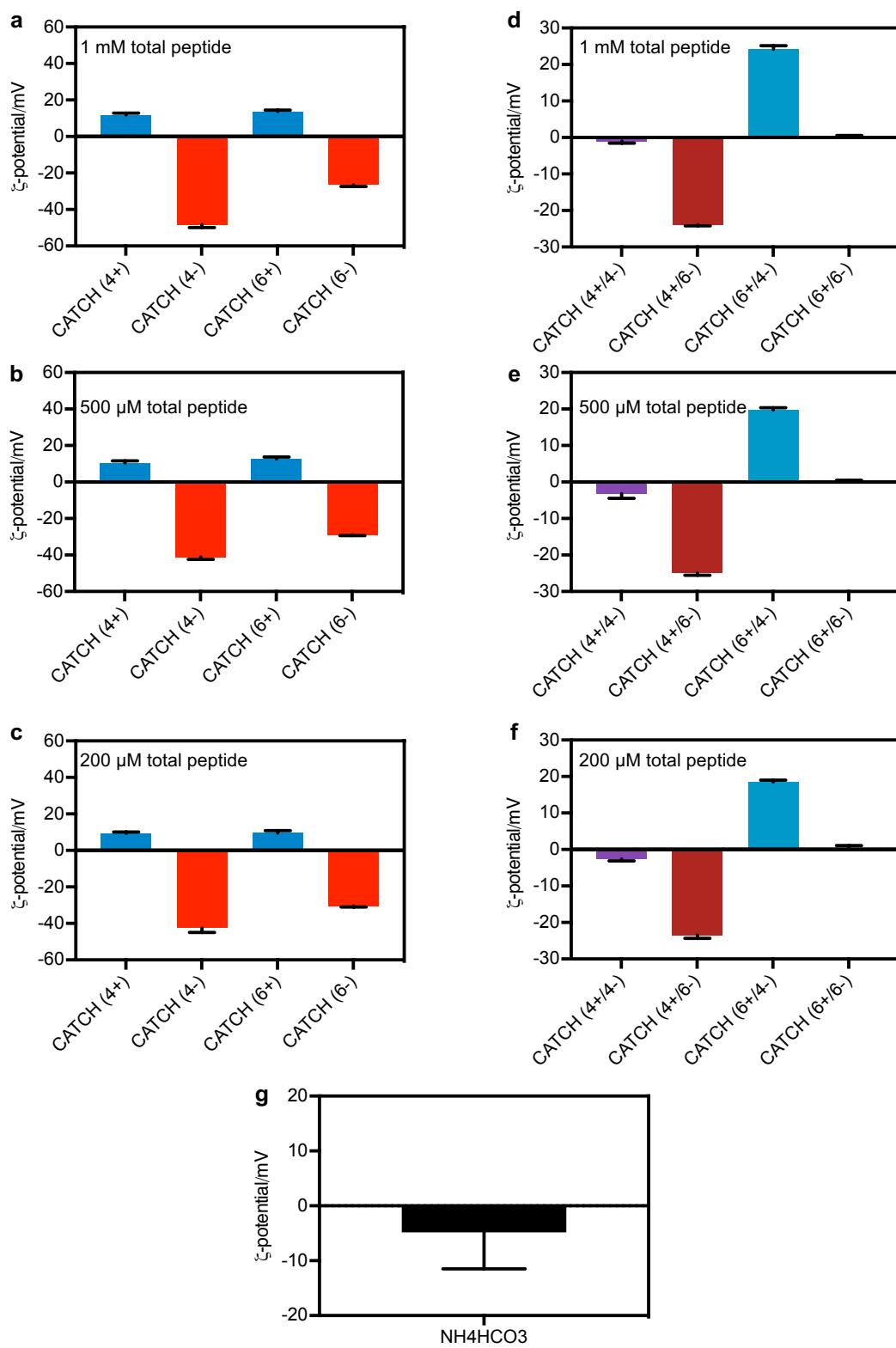
CATCH(6+/6-)



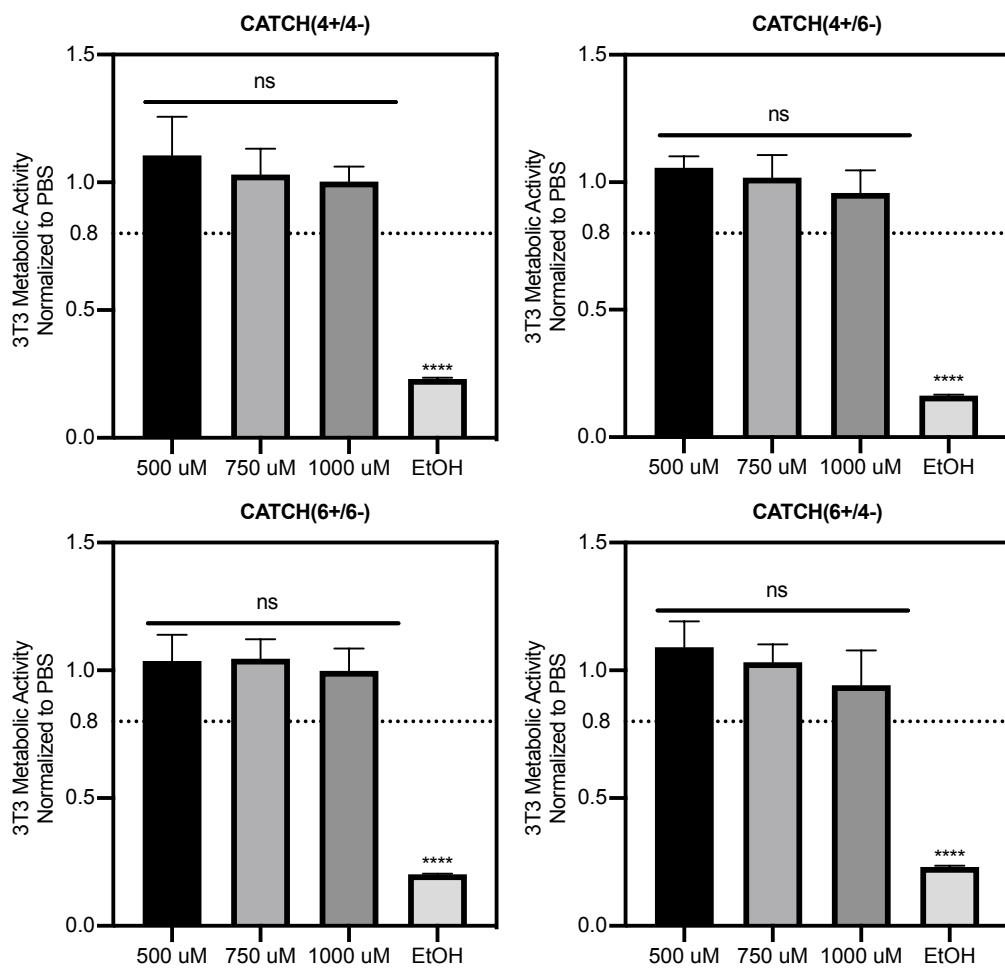
**Figure S2. Average pore size analysis of CATCH(+/-) hydrogels.** Histograms of the pore diameter measured from micrographs in (Figure 4) ( $n = 30$  measurements).



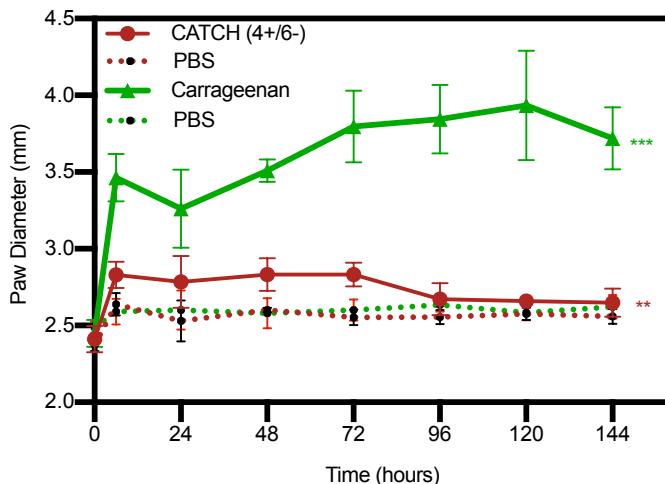
**Figure S3. Electron microscopy analysis of CATCH(+/-) nanofibers morphology.** TEM micrographs of 1 mM CATCH(4+/4-), CATCH(4+/6-), CATCH(6+/4-), and CATCH(6+/6-) nanofibers.



**Figure S4. Zeta potential of CATCH(+/-) peptides alone and in combination.** (a-c) Zeta potential measurements of CATCH(4+), CATCH(4-), CATCH(6+), and CATCH(6-) at 1 mM, 500  $\mu$ M, and 200  $\mu$ M total peptide. (d-f) Zeta potential measurements of equimolar mixtures of CATCH(4+/4-), CATCH(4+/6-), CATCH(6+/4-), and CATCH(6+/6-) at 1 mM, 500  $\mu$ M, and 200  $\mu$ M total peptide. (g) Zeta potential measurement of NH<sub>4</sub>HCO<sub>3</sub> buffer. Data are presented as the mean  $\pm$  standard deviation ( $n = 3$ ).

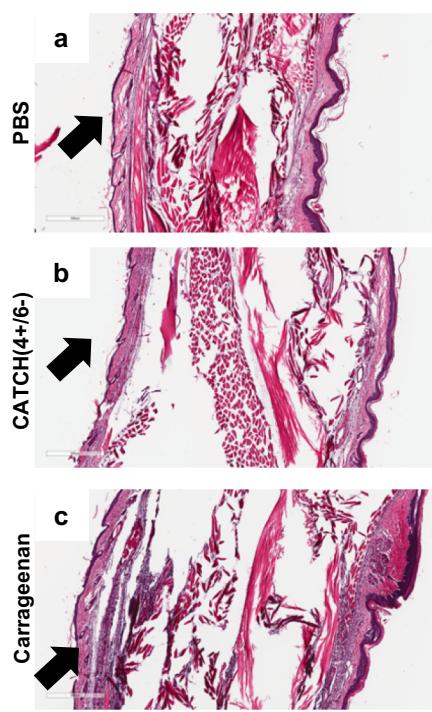


**Figure S5.** Metabolic activity of NIH 3T3 fibroblast cells after treatment with CATCH(+/-) nanofibers. Data are presented as mean  $\pm$  standard deviation ( $n = 5$ ). \*\*\*\* represents  $p < 0.0001$ , ANOVA with Tukey's post-hoc.

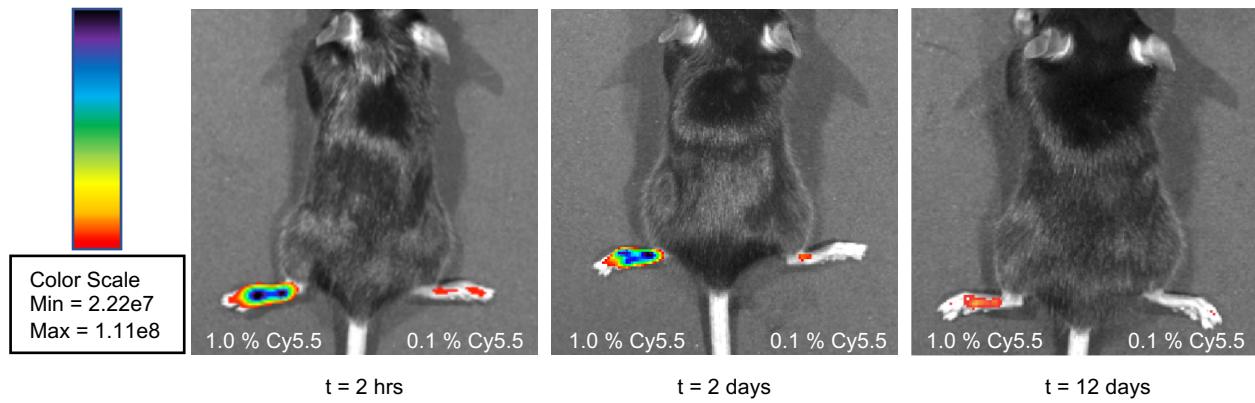


**Figure S6.** Caliper measurements of paw thickness after injection of CATCH(4+/6-) hydrogel,  $\lambda$ -Carrageenan, or PBS vehicle control into male C57BL/6 mice. Data presented as mean  $\pm$  standard deviation ( $n = 5$ ). \*\* represents  $p < 0.01$ , \*\*\* represents  $p < 0.001$ , repeated measures ANOVA with Dunnett's multiple comparison.

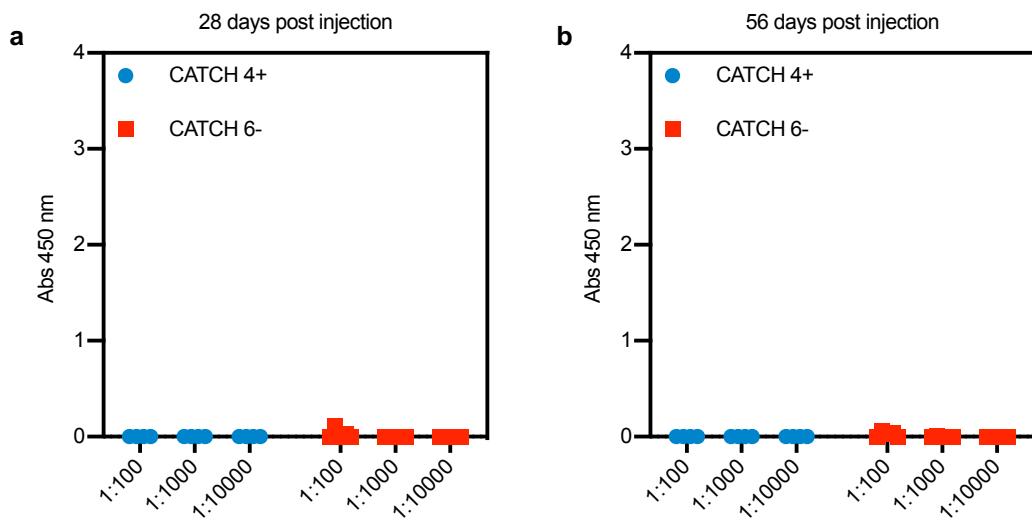
Histology at 24 hours



**Figure S7.** Representative histology sections of tissues collected 24 h after injection of CATCH(4+/6-),  $\lambda$ -Carrageenan, or PBS into the subcutaneous space of female C57BL/6 mice. Arrows show the site of injection.



**Figure S8.** In vivo fluorescence imaging of CATCH(4+/6-) hydrogels. Hydrogels with Cy5.5-labeled CATCH(4+) peptide (1.0 or 0.1% of total peptide), unlabeled CATCH(4+) peptide (49 or 49.9% of total peptide) and CATCH(6-) (50% of total peptide) were injected subcutaneously and imaged using an IVIS system. Color scale represents radiant efficiency ( $[p/sec/cm^2/sr] / [\mu W/cm^2]$ ).



**Figure S9.** Total IgG ELISA absorbance readings of dilutions of sera collected on day 28 and day 56 from female C57BL/6 mice that received a subcutaneous injection of 12 mM CATCH(4+/6-) hydrogel on day 0, followed by CATCH(4+) or CATCH(6-) in PBS on days 14 and 42. Data presented as a single point for each animal ( $n = 5$ ).

Peptide	Sequence	Adjusted Rank
CATCH(4+)	QQKFKFKFKQQ	322.81
CATCH(6-)	EQEFEFEFEQE	417.08
Q11	QQKFQFQFEQQ	331.38
OVA	ISQAVHAAHAEINEAGR	4.4

**Figure S10.** Immune Epitope Database (IEDB) predictions for binding of CATCH(4+), CATCH(6-), Q11, and OVA to C57BL/6 MHC II (H2-IAb background).