

LIMPID: A Versatile Method for Visualization of Brain Vascular Networks

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Supplementary Information

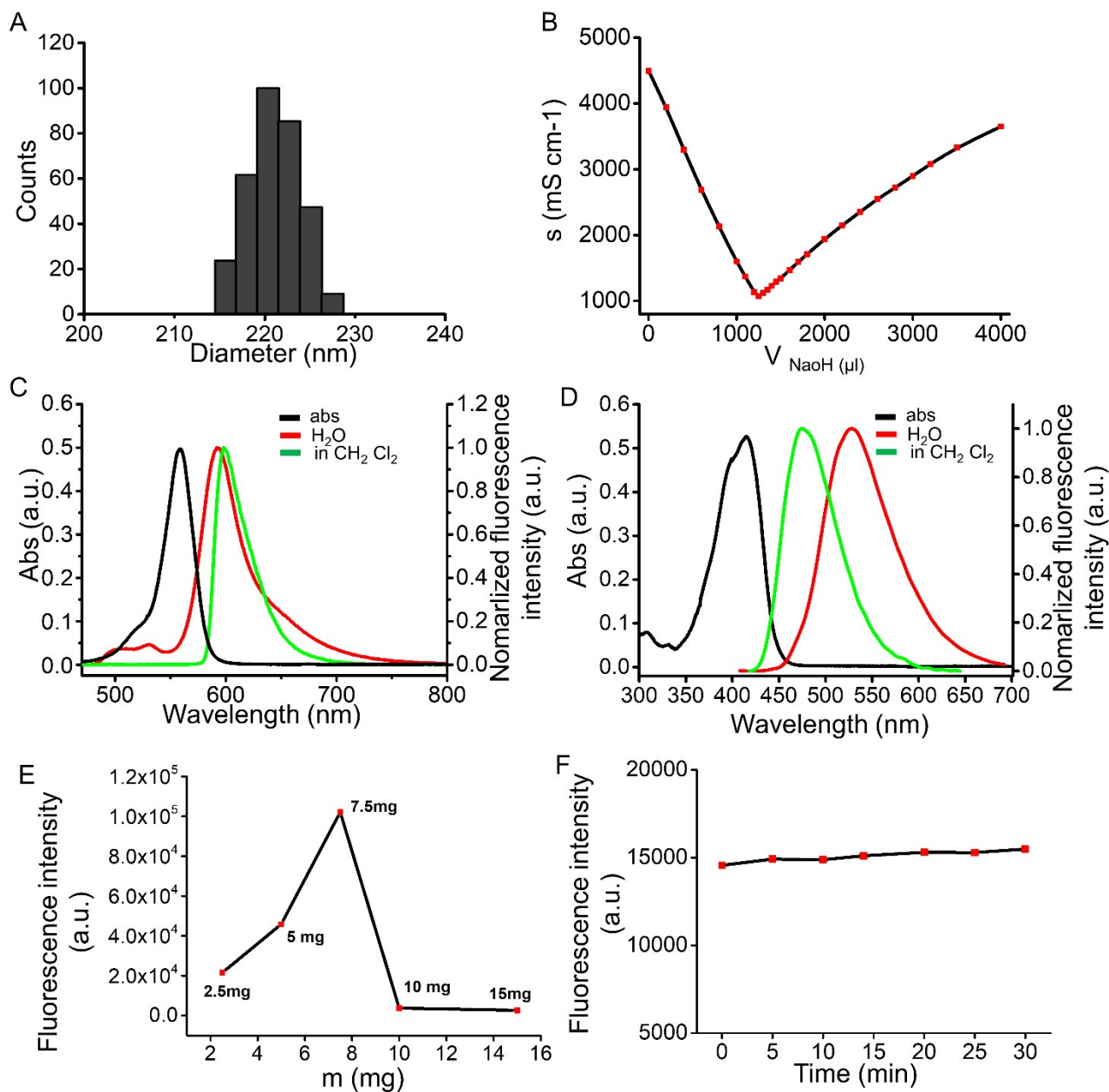


Fig. S1 Fluorescence spectroscopy measurement of fluorescent dyes and fluorescent nanoparticles. (A) Dynamic light scattering (DLS) measurements of SQ7-ANPs. (B) Conductometric titration detection of ANPs. (C) Absorption (black line) and fluorescence spectra (green line) of dye SQ7 in CH₂Cl₂ solution (1 \times 10⁻⁵ M, Ex: 450 nm), and fluorescence spectra (red line) of SQ7-ANPs (Ex: 450 nm). (D) Absorption (black line) and fluorescence spectra (green line) of BDT in CH₂Cl₂ solution (1 \times 10⁻⁵ M, Ex: 400 nm), and fluorescence spectra (red line) of

BDT-ANPs. (E) Fluorescence intensity of SQ7-ANPs containing different weight of SQ7 in SDS solution (Ex: 557 nm). (F) Fluorescence spectra of SQ7-ANPs in SDS solution (1×10^{-5} M, Ex: 557 nm) exposing to light for different period of time.

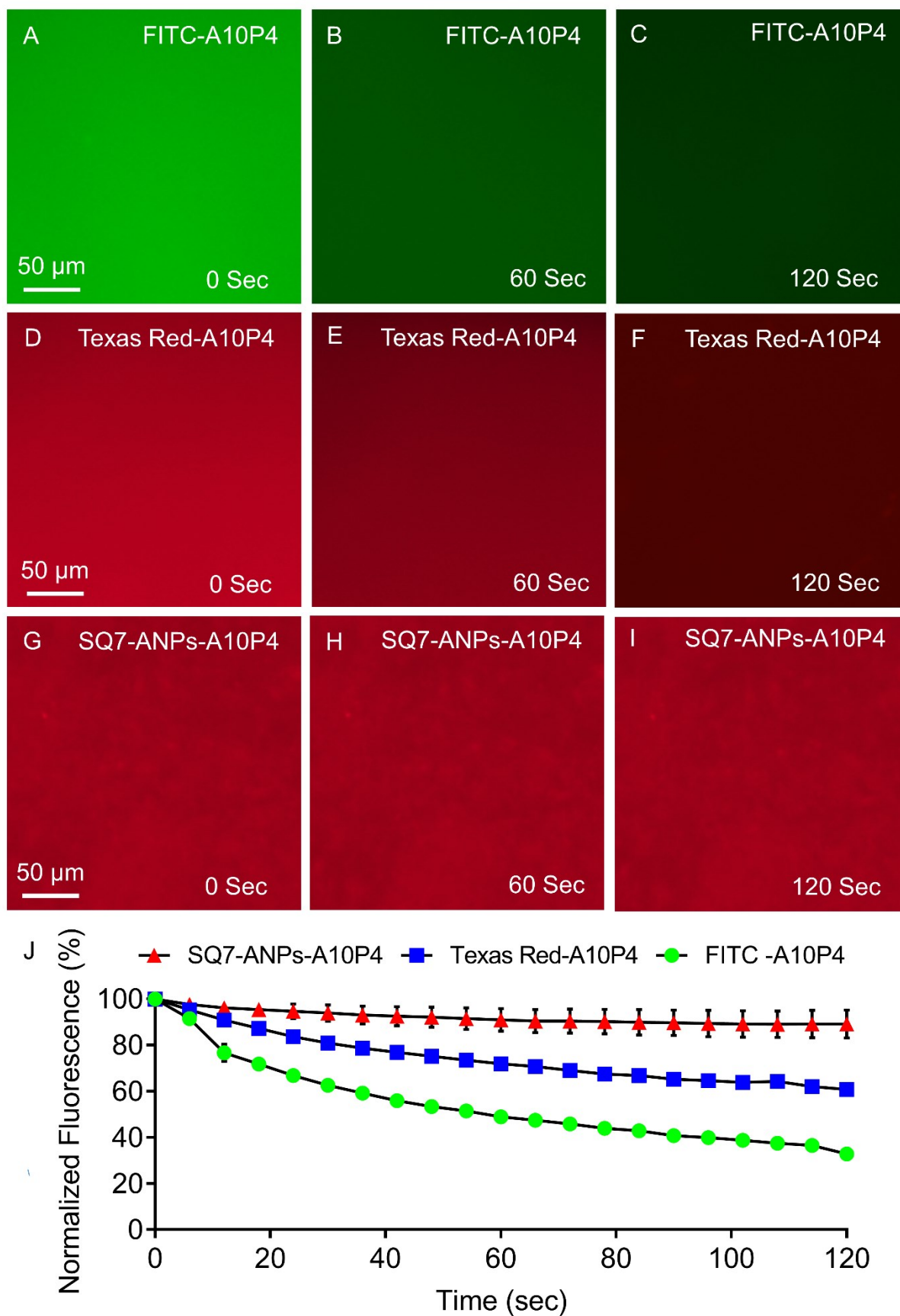


Fig. S2 Measurement of fluorescence loss in photobleaching of fluorescent hydrogels. (A-C) The representative figures of fluorescence intensity change of FITC-A10P4 hydrogel (BP: 470-490 nm, n=3) at different time points

during photobleaching. (D-F) The representative figures of fluorescence intensity change of Texas Red-A10P4 hydrogel (BP: 510-550 nm, n=3) at different time points during photobleaching. (G-I) The representative figures of fluorescence intensity change of SQ7-ANPs-A10P4 hydrogel (BP: 510-550 nm, n=6) at different time points during photobleaching. (J) Statistical diagram showing fluorescence loss of different fluorescent hydrogels at different time points during photobleaching ($n \geq 3$). The fluorescence intensity is normalized to pre-bleach fluorescent intensity.

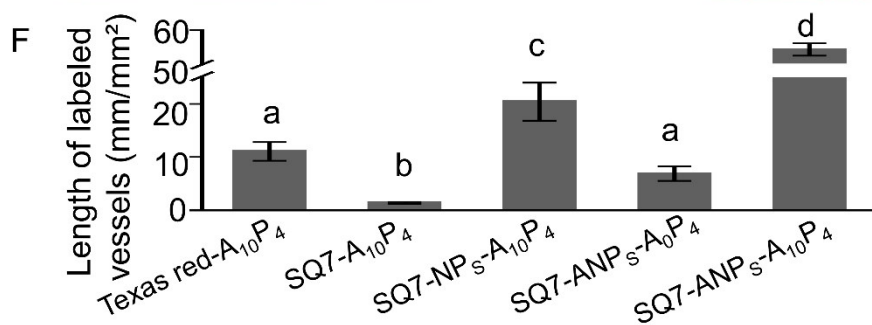
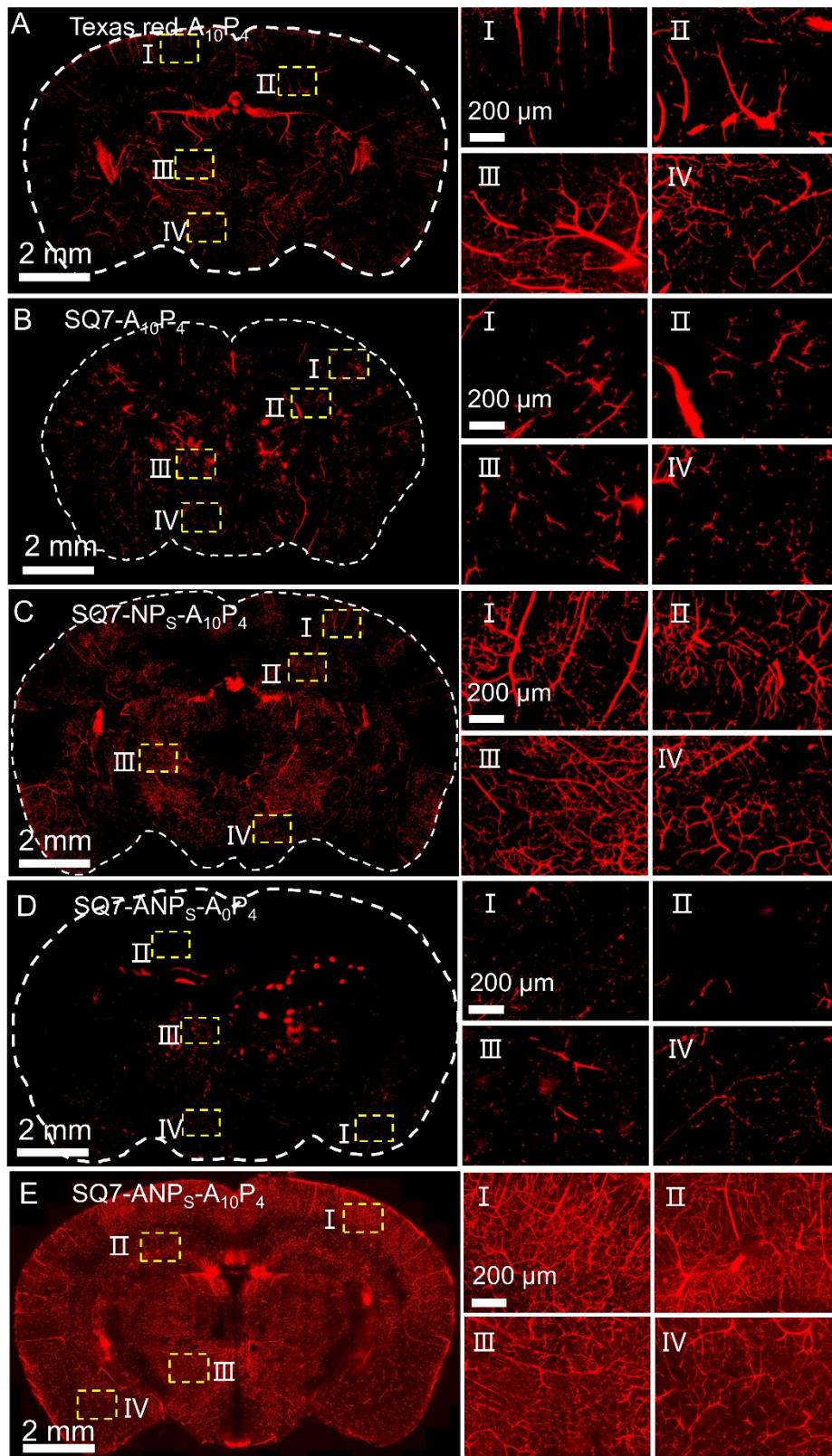


Fig. S3 Effect of different fluorescent dyes and fluorescent nanoparticles on labeling vascular networks. (A) Images of labeled blood vessels in the brain perfused with Texas red-A10P4. Higher-magnification images of boxed regions in (A) are shown in (I -IV) (n=4); (B) Blood vessel image in the brain perfused with SQ7-A10P4. Higher-magnification images of boxed regions in (B) are shown in (I -IV) (n=6); (C) Blood vessel image in the brain perfused with SQ7-NPs-A10P4. Higher-magnification images of boxed regions in (C) are shown in (I -IV) (n=6); (D) Blood vessel image in the brain perfused with SQ7-ANPs-A0P4. Higher-magnification images of boxed regions in (D) are shown in (I -IV) (n=5). (E) Blood vessel image in the brain perfused with SQ7-ANPs-A10P4. Higher-magnification images of boxed regions in (E) are shown in (I -IV) (n=6). (F) Statistical analysis of total length of labeled vessels between different labeling dyes or nanoparticles. a-d in (F) indicate significant differences between the two groups [one-way ANOVA $F(4,22)=101.4$, $p<0.0001$].

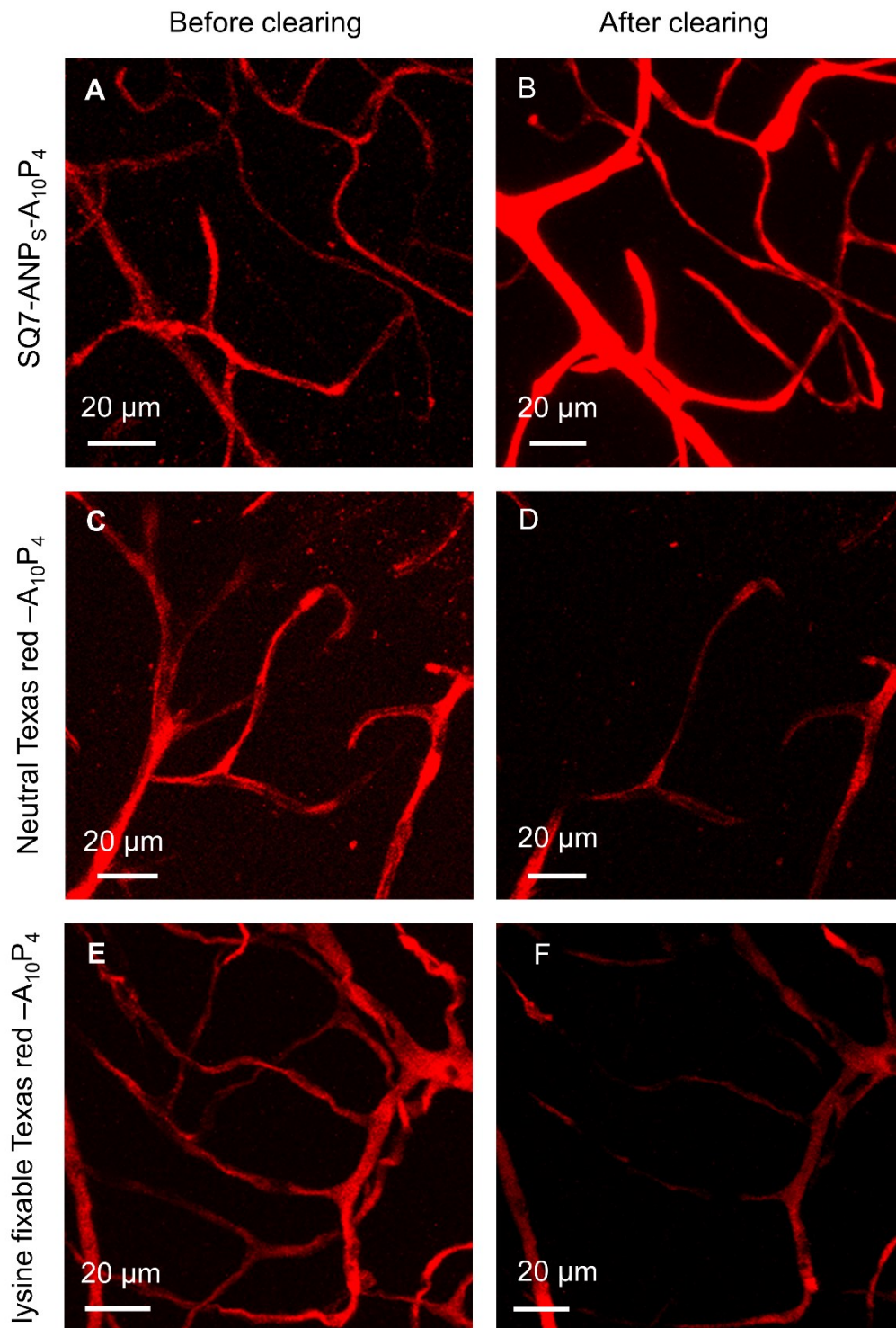


Figure S4. Comparison of the effect of blood vessel labeling using different fluorescent dyes before and after clearing. (A-B) Representative images of SQ7-ANPS-A10P4 labeled blood vessels before and after clearing. SQ7-ANPS-A10P4 can stably labelling blood vessels. (C-D) Representative images of neutral Texas red -A10P4 labeled blood vessels before and after clearing (E-F) Representative images of labeled blood vessels with lysine fixable

Texas red -A10P4 before and after clearing. Scale bar = 20 μ m.

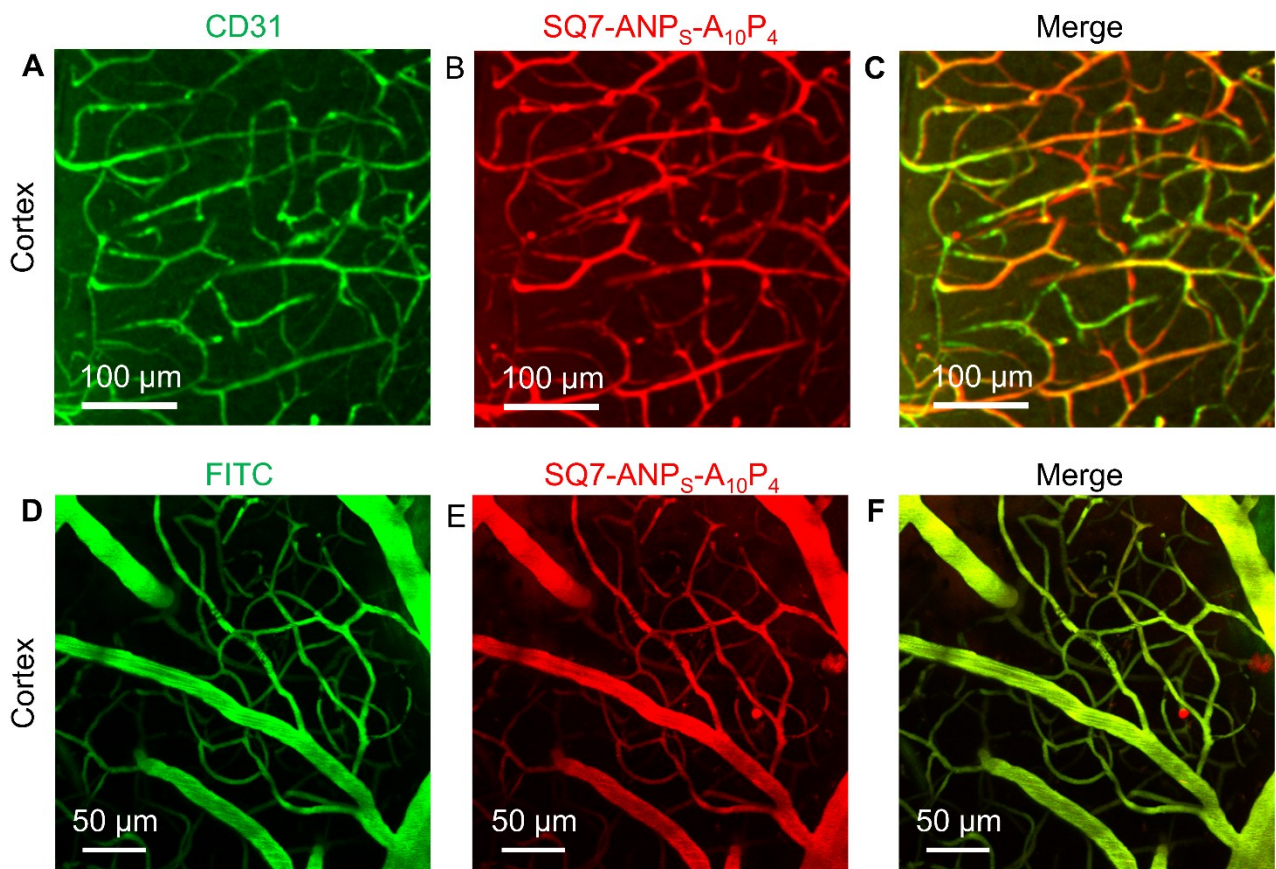


Figure S5. Visualization of cerebral vasculature using LIMPID. (A-B) Representative images showing the overlap of anti-CD31 antibody stained blood vessels (green) and SQ7-ANPS-A10P4 labelled blood vessels (red) in a mouse cortex. (C) Merged image showing the overlaps between anti CD31 antibody stained vessels and SQ7-ANPS-A10P4 labelled vessels. Scale bar = 100 μ m. (D) FITC dextran-labelled blood vessels imaged in a live mouse. (E) SQ7-ANPS-A10P4 labelled blood vessels in the same fields as showed in (D). (F) Merged image showing the overlaps between FITC dextran-labelled blood vessels and SQ7-ANPs-A10P4 labelled vessels. Scale bar = 50 μ m.

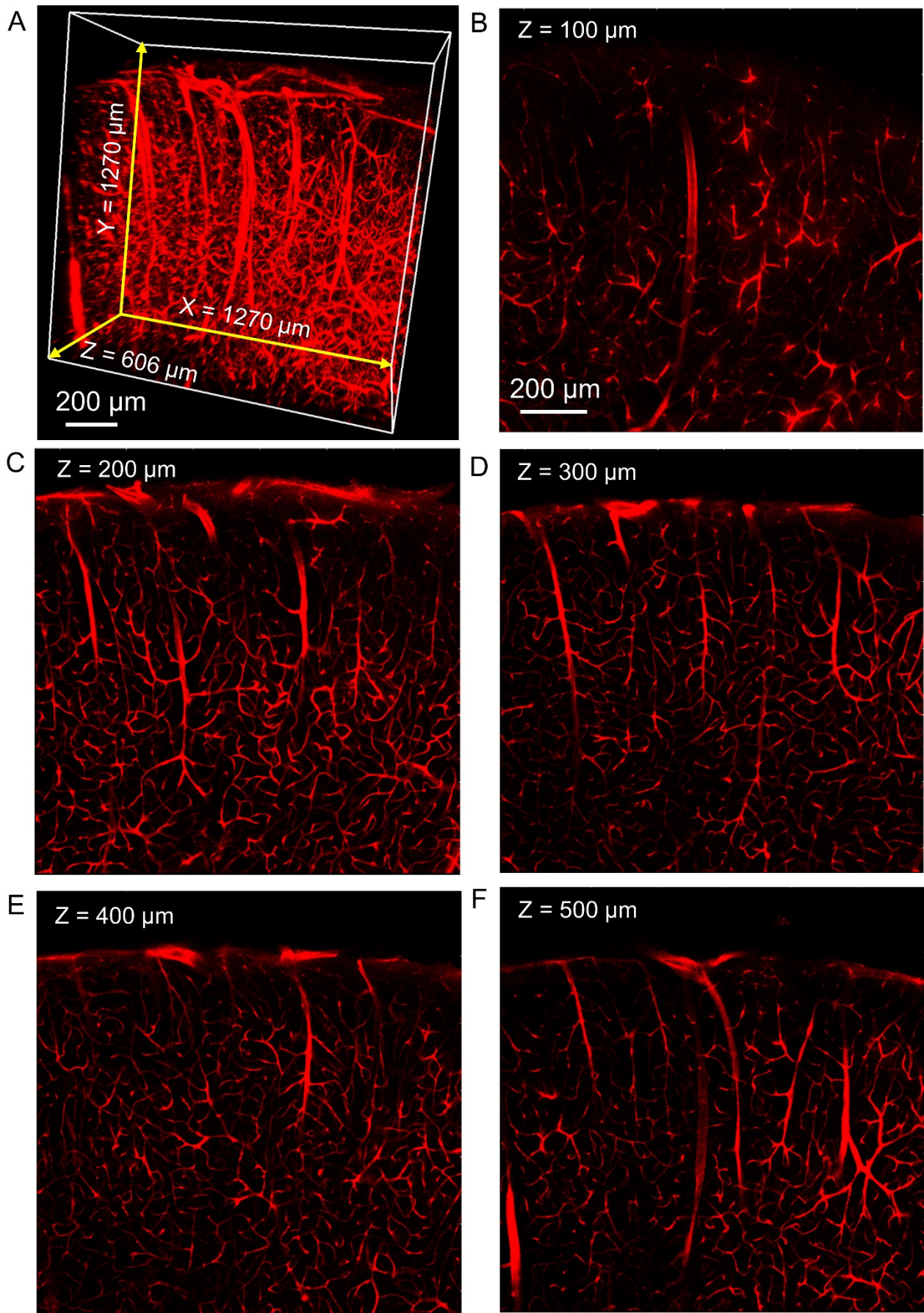


Figure S6. Images of vasculature in a mouse brain slice after LIMPID. (A) Representative 3D raw data view of the

vasculature in a cortical slice. Images were obtained using confocal microscopy and the image volume was $1270\ \mu\text{m} \times 1270\ \mu\text{m} \times 606\ \mu\text{m}$ with a voxel size of $1.242\ \mu\text{m} \times 1.242\ \mu\text{m} \times 1.25\ \mu\text{m}$. (B-F). Representative images at different depths of brain slice (100 μm , 200 μm , 300 μm , 400 μm , and 500 μm relative to the surface of the slice) showing that fluorescent nanoparticles penetrated into both surfaces and the deep tissues. Scale bar = 200 μm .