

**Supplementary Table 1.** Source, reference, company, endotoxin levels and purity of the Ch formulations used *in vitro* and *in vivo*.

Source	# Reference	Company	Endotoxin Levels	Purity (%)	Ref
Squid Pen	NR	France-Chitine	NR	*	(1)
NR	NR	Sigma-Aldrich	Negative	NR	(2)
NR	NR	Fluka Chemika	NR	NR	(3)
NR	NR	Sigma-Aldrich	NR	NR	(4)
NR	NR	Sigma-Aldrich	NR	NR	(5)
Shrimp shell	NR	Marinard Biotech	NR	*	(6)
NR	NR	Zhejiang Golden-shell Pharmaceutical	NR	NR	(7)
NR	NR	Xingcheng Biological Industrial Limited	NR	NR	(8)
NR	NR	Yuhuan County Marine Chemical Company	NR	NR	(9)
Squid Pen	NR	France-Chitine	Negative	*	(10)
NR	NR	Sigma-Aldrich	NR	NR	(11)
Shrimp shell	NR	Primex	NR	*	(12)
NR	NR	Sigma-Aldrich	NR	*	(13)
NR	NR	Ao'xing Biotechnology	NR	**	(14)
NR	NR	NR	NR	NR	(15)
Squid Pen	NR	France-Chitine	Negative	*	(16)
NR	NR	NR	Negative	NR	(17)
NR	NR	Yuhuan Ocean Biochemical	NR	*	(18)
NR	NR	Zhejiang Aoxing Biotechnology	NR	NR	(19)
NR	448869, 42344, 448877, 419419	Sigma-Aldrich	NR	NR	(20)
NR	NR	Sigma-Aldrich	NR	NR	(21)
NR	NR	Sigma-Aldrich	NR	NR	(22)
NR	NR	Sigma-Aldrich	NR	*	(23)
NR	NR	Sigma-Aldrich	NR	NR	(24)
NR	NR	Sigma-Aldrich	NR	NR	(25)
NR	NR	Aoxing Bio-Technology	NR	NR	(26)
NR	NR	Zhejiang Aoxing Biotechnology	NR	NR	(27)
NR	9012-76-4	Solarbio Company	NR	NR	(28)
NR	NR	NR	NR	NR	(29)
Crab Shell	9012-76-4	Coolaber	Negative	NR	(30)
NR	NR	Koyo Chemical	Negative	NR	(31)
Shrimp Shell	C-3646	Sigma-Aldrich	NR	NR	(32)
NR	NR	Sigma-Aldrich	NR	NR	(33)
NR	NR	Golden-shell Biochemical	NR	NR	(34)
Shrimp Shell	NR	Primex	NR	*	(35)
Shrimp Shell	NR	Primex	NR	*	(36)
NR	NR	Heppe Medical Chitosan	NR	NR	(37)
NR	NR	Golden-shell Biochemical	NR	*	(38)
NR	NR	NR	NR	NR	(39)
NR	Protasan UP CL110	Pronova Biopolymer	NR	NR	(40)
NR	NR	Immunophotonics	NR	NR	(41)

NR	Protosan UP G213	NovaMatrix	NR	NR	(42)
NR	NR	Immunophotonics	NR	NR	(43)
NR	NR	NR	NR	NR	(44)
NR	NR	NR	NR	NR	(45)
NR	NR	Intermedpharm	Negative	NR	(46)
NR	Protosan G213	NovaMatrix	NR	NR	(47)
NR	NR	Pronova Biopolymer	NR	NR	(48)
NR	Protosan G213	NovaMatrix	NR	NR	(49)
NR	Protosan G213	NovaMatrix	NR	NR	(50)
NR	NR	Koyo Chemical	NR	NR	(51)
NR	Protosan G 213	NovaMatrix	NR	NR	(52)
NR	NR	NR	NR	NR	(53)
NR	NR	NR	NR	NR	(54)
NR	NR	Pronova Biopolymer	NR	NR	(55)

**Notes:**

**NR** – Not Reported

\* The authors mention performing a purification protocol.

\*\* The authors mention not performing further purification.

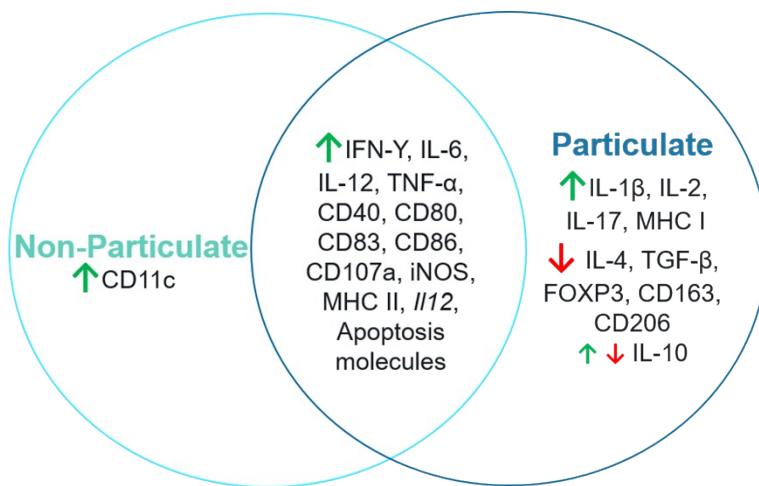
**Supplementary Table 2.** Ch formulations explored in vitro and in vivo, as well as the controls used to obtain the values presented in the results section.

Type of Formulation	Designation	Control	Ref
Films	Ch/γ-PGA PEMs incorporating IFN-γ	IFN-γ	(1)
	Ch solution for well coating	TCPS	(2)
Gel	Poloxamer 407-Ch hydrogel containing PLGA-OVA NPs, MPLA and QA	PLGA-OVA NPs	(3)
	Ch-DOX hydrogel containing Sig/E7/LAMP-1 vaccine	Sig/E7/LAMP-1 vaccine	(4)
	Ch hydrogel vaccine incorporating OVA and QA	DCs vaccine + OVA	(5)
	Ch thermogel	NT	(6)
Microneedles	Core-shell (Ch shell and PVA/PVP core) microneedle system incorporating anti-PD-L1 drug and 1-MT/Polymer solution	Anti-PD-L1/1-MT injection	(7)
Nanoparticles	IL-12 loaded TPP-Ch NPs	IL-12	(8)
	NKG2D-IL-21 loaded Ch NPs	<i>In vitro</i> : IL-21 <i>In vivo</i> : PBS	(9)
	Ch/γ-PGA NPs carrying IFN-γ	IFN-γ	(10)
	Ch NPs carrying CD40 and ICOSL mRNA	Lipofectamine as a transfection agent	(11)
	ChLa NPs carrying CD73 siRNA + TCL pulsed DCs	DCs vaccine	(12)
	PEG-ChLa NPs carrying A2AR siRNA	NT	(13)
	Mannose coated-Ch NPs carrying TCL	TCL	(14)
	CMCS-coated SF-CLN and IMD-CLN cationic lipid based NPs	SF-IMD solution	(15)
	Ch/γ-PGA NPs	NT	(16)
	Ch NPs carrying MG	MG or NT	(17)
	GC/GM-CSF-IL-21 NPs	GM-CSF-IL-21	(18)
	Biotinylated Ch NPs with plasmids encoding GM-CSF and IL-21	PlasmidS encoding GM-CSF and IL-21	(19)
	Ch/RA Polyplexes or Ch/RA Polyplexes + OVA	RA or RA + OVA	(20)
	Ch-conjugated green copper oxide NPs or Ch-conjugated green copper oxide NPs carrying BSA	Green Copper Oxide NPs or BSA	(21)
	Ch NPs carrying plasmid pBudCE4.1-azurin-MAM-A	Plasmid pBudCE4.1-azurin-MAM-A	(22)
	Ch coated HCuS NPs-CpG nanocomplexes + laser therapy	Laser therapy	(23)
	Ch NPs carrying OVA and poly I:C	<i>In vitro</i> : Soluble OVA <i>In vivo</i> : Soluble OVA + Poly I:C / Soluble OVA	(24)
	Ch-dextran sulfate NPs carrying PD-1 and PD-L1 siRNA	NT	(25)
	FA-Ch NPs carrying IP-10 gene + TRP2-specific CD8 <sup>+</sup> CD28 <sup>+</sup> T cells	TRP2 CD8 <sup>+</sup> CD28 <sup>+</sup> T cells	(26)
	MCMC/HA/PS/ODN NPs	HA/PS/ODN NPs	(27)
	FA-Ch NPs carrying IP-10 gene + DC/TCL fusion vaccine	DC/TCL Vaccine	(28)
	Ch NPs carrying EphrinA1-PE38/GM-CSF Ch NPs	Saline	(29)
	Ch NPs	NT	(30)
	Chitosan-carboxymethyl dextran NPs carrying cefmetazole, 5-FU or ICG	NT	(31)
	Alginic-acid coated Ch NPs carrying legumain DNA vaccine	PBS	(32)
	Nanogel of amphoteric methacrylamide N-carboxyethyl Ch and N-(2-hydroxy)propyl-3-trimethylammonium Ch Chloride coated with RBCM and	PTX + IL-2	(33)

	carrying IL-2 and PTX		
	PEG and mannose-trimethyl Ch and citraconic anhydride grafted PC carrying VEGF and PIGF siRNA	PBS	(34)
	Ch NPs carrying plasmid encoding E7 vaccine and IL-12	E7-DNA + IL-12	(35)
	Ch NPs carrying plasmid encoding E7 vaccine	E7-DNA	(36)
	ChNPs carrying siRNA for Gal-1	NT	(37)
	TMC NPs conjugated with DOX via CA linkage and carrying IL-2	DOX + IL-2	(38)
Solution	Ch solution	NT	(39)
	GC + HIFU	HIFU	(40)
	N-dihydrogalactochitosan solution	NT	(41)
	Ch glutamate/IL-12/OVA solution	IL-12/OVA solution	(42)
	N-dihydrogalactochitosan + PDT-generated vaccine	PDT-generated vaccine	(43)
	SWNT-GC solution + PTT or SWNT-GC + PTT + anti-CTLA-4	<i>In vitro</i> : PTT <i>In vivo</i> : anti-CTLA-4	(44)
	N-dihydrogalactochitosan solution + laser treatment	NT	(45)
	WSCO and IFN-Y solution	NT	(46)
	Ch glutamate/IL-12 solution	PBS	(47)
	Glycated Ch solution	NT	(48)
	Ch/IL-12 solution	NT	(49)
	Ch/IL-12 solution	IL-12 solution	(50)
	Ch solution	Acetic Acid	(51)
	Ch/IL-12 solution	IL-12 or PBS	(52)
	COS solution	NT	(53)
	COS solution + CTX	CTX	(54)
	Glycated Ch solution + Laser	Laser	(55)

### Abbreviations List:

1-MT: 1-methyl-D,L-tryptophan; 5-FU: 5-Fluorouracil; A2AR: Adenosine A2A Receptor; CA: *cis*-aconitic anhydride; CD: Cluster of Differentiation; Ch: Chitosan; ChLa: Chitosan Lactate; CLN: Cationic Lipid-based Nanoparticles; CMCS: O-Carboxymethyl-Chitosan; COS: Chitosan Oligosaccharide; CpG: Cytosine-Guanine; CTX: Cyclophosphamide; DCs: Dendritic Cells; DOX: Doxorubicin; Ephrin1A: Specific ligand for tyrosine kinase family receptor EphA2; FA: Folate; Gal-1: Galectin-1; GC: Glycated Chitosan; GM-CSF: Granulocyte-Monocyte Colony Stimulating Factor; HA: Hyaluronan; HCuS: Hollow Copper Sulfide; HIFU: High-Intensity Focused Ultrasound; ICG: Indocyanine Green; ICSL: Inducible Costimulator Ligand; IFN- $\gamma$ : Interferon  $\gamma$ ; IL: Interleukin; IMD: IMD-0354; IP-10: Interferon- $\gamma$ -inducible Protein 10; MCMC: Mannosylated Carboxymethyl Chitosan; MG: Methylglyoxal; MPLA: Monophosphoryl Lipid A; NPs: Nanoparticles; NT: No treatment; ODN: Oligodeoxynucleotides; OVA: Ovalbumin; PBS: Phosphate-buffered saline; PC: poly (allylamine hydrochloride); PDT: Photodynamic Therapy; PE38: *Pseudomonas aeruginosa* toxin PE38; PEG: Polyethylene Glycol; PIGF: Placenta Growth Factor; PLGA: Poly (lactic-co-glycolic acid); Poly I:C: Polyinosinic-Polycytidylic Acid Sodium Salt; PS: Protamine sulfate; PTT: Photothermal Therapy; PTX: Paclitaxel; PVA: Polyvinyl Alcohol; PVP: Poly (N-vinylpyrrolidone); QA: Quil A; RA: Toll-like receptor 3-recognizing RNA adjuvant; RBCM: Red Blood Cell Membrane; SF: Sorafenib; SigE7/LAMP-1: Vaccinia vector carrying E7 antigen and sorting signal of the lysosome-associated membrane protein 1; siRNA: Small Interfering RNA; SWNT: Single Walled Carbon Nanotube; TCL: Tumor Cell Lysate; TCPS: Tissue Culture Polystyrene; TMC: Trimethyl Chitosan; TRP2: Melanoma-specific antigen TRP2; TPP: Tripolyphosphate; VEGF: Vascular Endothelial Growth Factor; WSCO: Water Soluble Chitosan Oligomers;  $\gamma$ -PGA: Poly-( $\gamma$ -glutamic acid);



**Supplementary Figure 1.** Summary of the immune-related alterations observed in *in vitro* studies, using particulate and non-particulate Ch.

	Non-particulate	Common	Particulate
TME	↑ ↓ Macrophages	↑ CTL, DCs, Th1, NK cells ↓ Tregs	↑ M1-macrophages ↓ M2-macrophages, MDSCs
Systemic	↑ ↓ Macrophages	↑ B cells, CTL, DCs, Th, NK cells, IgGs ↓ MDSCs, Tregs	↑ ↓ IL-4, IL-10, IL-17, TGF- $\beta$

**Supplementary Figure 2.** Summary of the immune-related alterations observed in *in vivo* studies, using particulate and non-particulate Ch.

## **References**

1. Cardoso AP, Goncalves RM, Antunes JC, Pinto ML, Pinto AT, Castro F, et al. An interferon-gamma-delivery system based on chitosan/poly(gamma-glutamic acid) polyelectrolyte complexes modulates macrophage-derived stimulation of cancer cell invasion in vitro. *Acta biomaterialia*. 2015;23:157-71.
2. Lin YC, Lou PJ, Young TH. Chitosan as an adjuvant-like substrate for dendritic cell culture to enhance antitumor effects. *Biomaterials*. 2014;35(31):8867-75.
3. Bobbala S, Gibson B, Gamble AB, McDowell A, Hook S. Poloxamer 407-chitosan grafted thermoresponsive hydrogels achieve synchronous and sustained release of antigen and adjuvant from single-shot vaccines. *Immunology and cell biology*. 2018;96(6):656-65.
4. Han HD, Song CK, Park YS, Noh KH, Kim JH, Hwang T, et al. A chitosan hydrogel-based cancer drug delivery system exhibits synergistic antitumor effects by combining with a vaccinia viral vaccine. *International journal of pharmaceutics*. 2008;350(1-2):27-34.
5. Highton AJ, Girardin A, Bell GM, Hook SM, Kemp RA. Chitosan gel vaccine protects against tumour growth in an intracaeal mouse model of cancer by modulating systemic immune responses. *BMC immunology*. 2016;17(1):39.
6. Monette A, Ceccaldi C, Assaad E, Lerouge S, Lapointe R. Chitosan thermogels for local expansion and delivery of tumor-specific T lymphocytes towards enhanced cancer immunotherapies. *Biomaterials*. 2016;75:237-49.
7. Yang P, Lu C, Qin W, Chen M, Quan G, Liu H, et al. Construction of a Core-shell Microneedle System to Achieve Targeted Co-delivery of Checkpoint Inhibitors for Melanoma Immunotherapy. *Acta biomaterialia*. 2020.
8. Xu Q, Guo L, Gu X, Zhang B, Hu X, Zhang J, et al. Prevention of colorectal cancer liver metastasis by exploiting liver immunity via chitosan-TPP/nanoparticles formulated with IL-12. *Biomaterials*. 2012;33(15):3909-18.
9. Tan L, Han S, Ding S, Xiao W, Ding Y, Qian L, et al. Chitosan nanoparticle-based delivery of fused NKG2D-IL-21 gene suppresses colon cancer growth in mice. *International journal of nanomedicine*. 2017;12:3095-107.
10. Castro F, Pinto ML, Almeida R, Pereira F, Silva AM, Pereira CL, et al. Chitosan/poly(gamma-glutamic acid) nanoparticles incorporating IFN-gamma for immune response modulation in the context of colorectal cancer. *Biomaterials science*. 2019;7(8):3386-403.
11. Daneshmandi S, Pourfathollah AA, Forouzandeh-Moghaddam M. Enhanced CD40 and ICOSL expression on dendritic cells surface improve anti-tumor immune responses; effectiveness of mRNA/chitosan nanoparticles. *Immunopharmacology and immunotoxicology*. 2018;40(5):375-86.
12. Jadidi-Niaragh F, Atyabi F, Rastegari A, Kheshtchin N, Arab S, Hassannia H, et al. CD73 specific siRNA loaded chitosan lactate nanoparticles potentiate the antitumor effect of a dendritic cell vaccine in 4T1 breast cancer bearing mice. *Journal of controlled release : official journal of the Controlled Release Society*. 2017;246:46-59.
13. Masjedi A, Hassannia H, Atyabi F, Rastegari A, Hojjat-Farsangi M, Namdar A, et al. Downregulation of A2AR by siRNA loaded PEG-chitosan-lactate nanoparticles restores the T cell mediated anti-tumor responses through blockage of PKA/CREB signaling pathway. *International journal of biological macromolecules*. 2019;133:436-45.
14. Shi GN, Zhang CN, Xu R, Niu JF, Song HJ, Zhang XY, et al. Enhanced antitumor immunity by targeting dendritic cells with tumor cell lysate-loaded chitosan nanoparticles vaccine. *Biomaterials*. 2017;113:191-202.

15. Wang T, Zhang J, Hou T, Yin X, Zhang N. Selective targeting of tumor cells and tumor associated macrophages separately by twin-like core-shell nanoparticles for enhanced tumor-localized chemoimmunotherapy. *Nanoscale*. 2019;11(29):13934-46.
16. Castro F, Pinto ML, Silva AM, Pereira CL, Teixeira GQ, Gomez-Lazaro M, et al. Pro-inflammatory chitosan/poly(gamma-glutamic acid) nanoparticles modulate human antigen-presenting cells phenotype and revert their pro-invasive capacity. *Acta biomaterialia*. 2017;63:96-109.
17. Chakrabarti A, Talukdar D, Pal A, Ray M. Immunomodulation of macrophages by methylglyoxal conjugated with chitosan nanoparticles against Sarcoma-180 tumor in mice. *Cellular immunology*. 2014;287(1):27-35.
18. Cheng M, Li Q, Wan T, Hong X, Chen H, He B, et al. Synthesis and efficient hepatocyte targeting of galactosylated chitosan as a gene carrier in vitro and in vivo. *Journal of biomedical materials research Part B, Applied biomaterials*. 2011;99(1):70-80.
19. Cheng M, Zhu W, Li Q, Dai D, Hou Y. Anti-cancer efficacy of biotinylated chitosan nanoparticles in liver cancer. *Oncotarget*. 2017;8(35):59068-85.
20. Choi JJ, Le QV, Kim D, Kim YB, Shim G, Oh YK. High Molecular Weight Chitosan-Complexed RNA Nanoadjuvant for Effective Cancer Immunotherapy. *Pharmaceutics*. 2019;11(12).
21. Dey A, Manna S, Kumar S, Chattopadhyay S, Saha B, Roy S. Immunostimulatory effect of chitosan conjugated green copper oxide nanoparticles in tumor immunotherapy. *Cytokine*. 2020;127:154958.
22. Ghasemi-Dehkordi P, Doosti A, Jami MS. The concurrent effects of azurin and Mammaglobin-A genes in inhibition of breast cancer progression and immune system stimulation in cancerous BALB/c mice. *3 Biotech*. 2019;9(7):271.
23. Guo L, Yan DD, Yang D, Li Y, Wang X, Zalewski O, et al. Combinatorial photothermal and immuno cancer therapy using chitosan-coated hollow copper sulfide nanoparticles. *ACS nano*. 2014;8(6):5670-81.
24. Han HD, Byeon Y, Jang JH, Jeon HN, Kim GH, Kim MG, et al. In vivo stepwise immunomodulation using chitosan nanoparticles as a platform nanotechnology for cancer immunotherapy. *Scientific reports*. 2016;6:38348.
25. Hassannia H, Ghasemi Chaleshtari M, Atyabi F, Nosouhian M, Masjedi A, Hojjat-Farsangi M, et al. Blockage of immune checkpoint molecules increases T-cell priming potential of dendritic cell vaccine. *Immunology*. 2020;159(1):75-87.
26. He J, Duan S, Yu X, Qian Z, Zhou S, Zhang Z, et al. Folate-modified Chitosan Nanoparticles Containing the IP-10 Gene Enhance Melanoma-specific Cytotoxic CD8(+)CD28(+) T Lymphocyte Responses. *Theranostics*. 2016;6(5):752-61.
27. He XY, Liu BY, Wu JL, Ai SL, Zhuo RX, Cheng SX. A Dual Macrophage Targeting Nanovector for Delivery of Oligodeoxynucleotides To Overcome Cancer-Associated Immunosuppression. *ACS applied materials & interfaces*. 2017;9(49):42566-76.
28. Hu Z, Chen J, Zhou S, Yang N, Duan S, Zhang Z, et al. Mouse IP-10 Gene Delivered by Folate-modified Chitosan Nanoparticles and Dendritic/tumor Cells Fusion Vaccine Effectively Inhibit the Growth of Hepatocellular Carcinoma in Mice. *Theranostics*. 2017;7(7):1942-52.
29. Li M, Wang B, Wu Z, Shi X, Zhang J, Han S. Treatment of Dutch rat models of glioma using EphrinA1-PE38/GM-CSF chitosan nanoparticles by in situ activation of dendritic cells. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2015;36(10):7961-6.
30. Lin L, He J, Li J, Xu Y, Li J, Wu Y. Chitosan Nanoparticles Strengthen Vgamma9Vdelta2 T-Cell Cytotoxicity Through Upregulation Of Killing Molecules And Cytoskeleton Polarization. *International journal of nanomedicine*. 2019;14:9325-36.

31. Lin YS, Radzi R, Morimoto M, Saimoto H, Okamoto Y, Minami S. Characterization of chitosan-carboxymethyl dextran nanoparticles as a drug carrier and as a stimulator of mouse splenocytes. *Journal of biomaterials science Polymer edition*. 2012;23(11):1401-20.
32. Liu Z, Lv D, Liu S, Gong J, Wang D, Xiong M, et al. Alginic acid-coated chitosan nanoparticles loaded with legumain DNA vaccine: effect against breast cancer in mice. *PloS one*. 2013;8(4):e60190.
33. Song Q, Yin Y, Shang L, Wu T, Zhang D, Kong M, et al. Tumor Microenvironment Responsive Nanogel for the Combinatorial Antitumor Effect of Chemotherapy and Immunotherapy. *Nano letters*. 2017;17(10):6366-75.
34. Song Y, Tang C, Yin C. Combination antitumor immunotherapy with VEGF and PIGF siRNA via systemic delivery of multi-functionalized nanoparticles to tumor-associated macrophages and breast cancer cells. *Biomaterials*. 2018;185:117-32.
35. Tahamtan A, Barati M, Tabarraei A, Mohebbi SR, Shirian S, Gorji A, et al. Antitumor Immunity Induced by Genetic Immunization with Chitosan Nanoparticle Formulated Adjuvanted for HPV-16 E7 DNA Vaccine. *Iranian journal of immunology : IJI*. 2018;15(4):269-80.
36. Tahamtan A, Ghaemi A, Gorji A, Kalhor HR, Sajadian A, Tabarraei A, et al. Antitumor effect of therapeutic HPV DNA vaccines with chitosan-based nanodelivery systems. *Journal of biomedical science*. 2014;21:69.
37. Van Woensel M, Mathivet T, Wauthoz N, Rosiere R, Garg AD, Agostinis P, et al. Sensitization of glioblastoma tumor micro-environment to chemo- and immunotherapy by Galectin-1 intranasal knock-down strategy. *Scientific reports*. 2017;7(1):1217.
38. Wu J, Tang C, Yin C. Co-delivery of doxorubicin and interleukin-2 via chitosan based nanoparticles for enhanced antitumor efficacy. *Acta biomaterialia*. 2017;47:81-90.
39. Li X, Dong W, Nalin AP, Wang Y, Pan P, Xu B, et al. The natural product chitosan enhances the anti-tumor activity of natural killer cells by activating dendritic cells. *Oncoimmunology*. 2018;7(6):e1431085.
40. Chen YL, Wang CY, Yang FY, Wang BS, Chen JY, Lin LT, et al. Synergistic effects of glycated chitosan with high-intensity focused ultrasound on suppression of metastases in a syngeneic breast tumor model. *Cell death & disease*. 2014;5:e1178.
41. El-Hussein A, Lam SSK, Raker J, Chen WR, Hamblin MR. N-dihydrogalactochitosan as a potent immune activator for dendritic cells. *Journal of biomedical materials research Part A*. 2017;105(4):963-72.
42. Heffernan MJ, Zaharoff DA, Fallon JK, Schlom J, Greiner JW. In vivo efficacy of a chitosan/IL-12 adjuvant system for protein-based vaccines. *Biomaterials*. 2011;32(3):926-32.
43. Korbelik M, Banath J, Zhang W, Gallagher P, Hode T, Lam SSK, et al. N-dihydrogalactochitosan as immune and direct antitumor agent amplifying the effects of photodynamic therapy and photodynamic therapy-generated vaccines. *International immunopharmacology*. 2019;75:105764.
44. Li Y, Li X, Doughty A, West C, Wang L, Zhou F, et al. Phototherapy using immunologically modified carbon nanotubes to potentiate checkpoint blockade for metastatic breast cancer. *Nanomedicine : nanotechnology, biology, and medicine*. 2019;18:44-53.
45. Qi X, Lam SS, Liu D, Kim DY, Ma L, Alleruzzo L, et al. Development of inCVAX, In situ Cancer Vaccine, and Its Immune Response in Mice with Hepatocellular Cancer. *Journal of clinical & cellular immunology*. 2016;7(4).
46. Seo WG, Pae HO, Kim NY, Oh GS, Park IS, Kim YH, et al. Synergistic cooperation between water-soluble chitosan oligomers and interferon-gamma for induction of nitric oxide synthesis and tumoricidal activity in murine peritoneal macrophages. *Cancer letters*. 2000;159(2):189-95.

47. Smith SG, Baltz JL, Koppolu BP, Ravindranathan S, Nguyen K, Zaharoff DA. Immunological mechanisms of intravesical chitosan/interleukin-12 immunotherapy against murine bladder cancer. *Oncoimmunology*. 2017;6(1):e1259050.
48. Song S, Zhou F, Nordquist RE, Carubelli R, Liu H, Chen WR. Glycated chitosan as a new non-toxic immunological stimulant. *Immunopharmacology and immunotoxicology*. 2009;31(2):202-8.
49. Vo JL, Yang L, Kurtz SL, Smith SG, Koppolu BP, Ravindranathan S, et al. Neoadjuvant immunotherapy with chitosan and interleukin-12 to control breast cancer metastasis. *Oncoimmunology*. 2014;3(12):e968001.
50. Yang L, Zaharoff DA. Role of chitosan co-formulation in enhancing interleukin-12 delivery and antitumor activity. *Biomaterials*. 2013;34(15):3828-36.
51. Yeh MY, Shih YL, Chung HY, Chou J, Lu HF, Liu CH, et al. Chitosan promotes immune responses, ameliorating total mature white blood cell numbers, but increases glutamic oxaloacetic transaminase and glutamic pyruvic transaminase, and ameliorates lactate dehydrogenase levels in leukemia mice *in vivo*. *Molecular medicine reports*. 2017;16(3):2483-90.
52. Zaharoff DA, Hoffman BS, Hooper HB, Benjamin CJ, Jr., Khurana KK, Hance KW, et al. Intravesical immunotherapy of superficial bladder cancer with chitosan/interleukin-12. *Cancer research*. 2009;69(15):6192-9.
53. Zhai X, Yang X, Zou P, Shao Y, Yuan S, Abd El-Aty AM, et al. Protective Effect of Chitosan Oligosaccharides Against Cyclophosphamide-Induced Immunosuppression and Irradiation Injury in Mice. *Journal of food science*. 2018;83(2):535-42.
54. Zhai X, Yuan S, Yang X, Zou P, Shao Y, Abd El-Aty AM, et al. Growth-inhibition of S180 residual-tumor by combination of cyclophosphamide and chitosan oligosaccharides *in vivo*. *Life sciences*. 2018;202:21-7.
55. Zhou F, Song S, Chen WR, Xing D. Immunostimulatory properties of glycated chitosan. *Journal of X-ray science and technology*. 2011;19(2):285-92.