

A STING-based Fluorescent Polarization Assay for Monitoring Activities of Cyclic Dinucleotide Metabolizing Enzymes

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Supplementary information

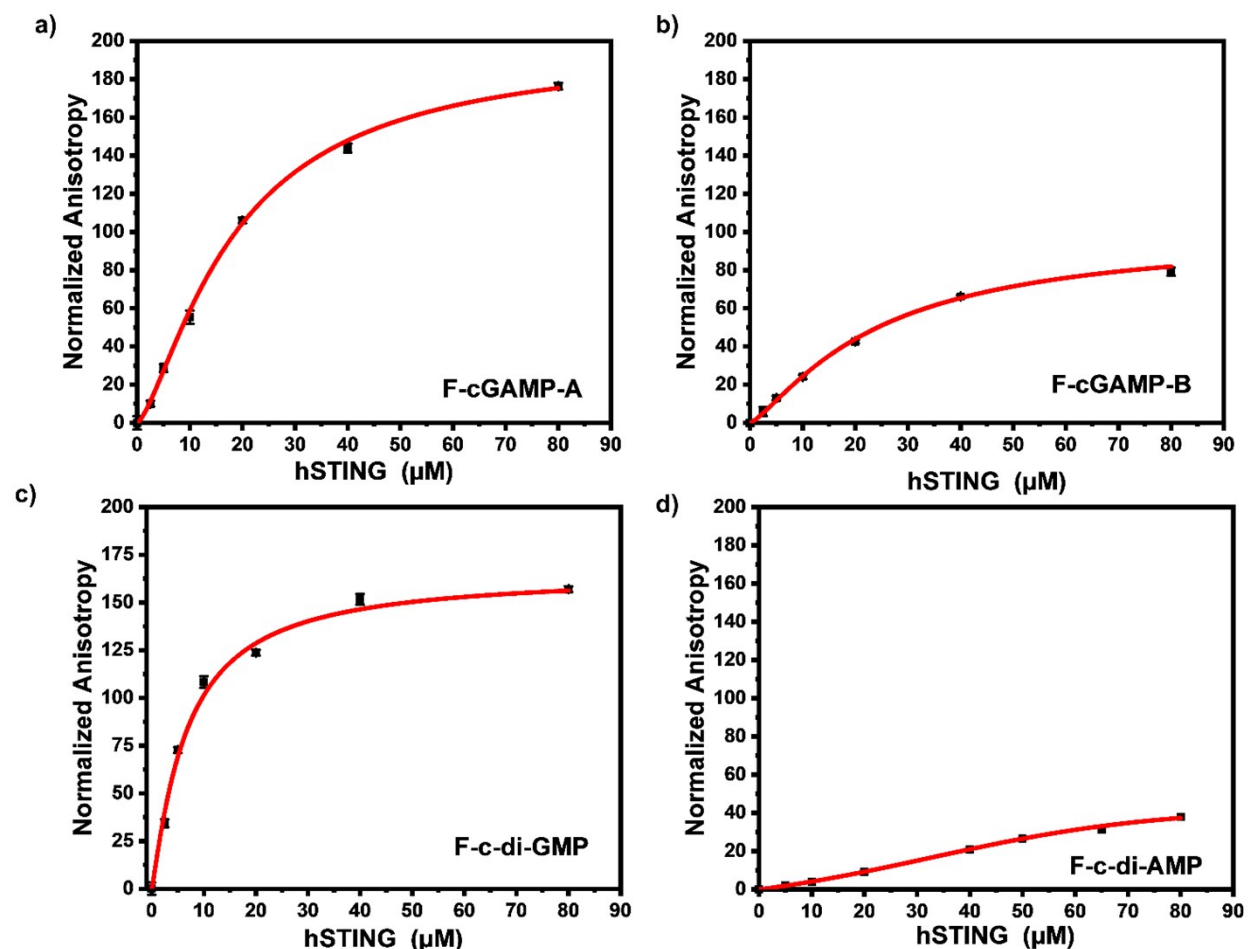


Fig. S1: hSTING titration curves a) Anisotropy of F-cGAMP-A in the presence of different concentrations of hSTING. b) Anisotropy of F-cGAMP-B in the presence of different concentrations of hSTING. c) Anisotropy of F-c-di-GMP in the presence of different concentrations of hSTING. d) Anisotropy of F-c-di-AMP in the presence of different concentrations of hSTING. Curves generated with origin in-built dose response function.

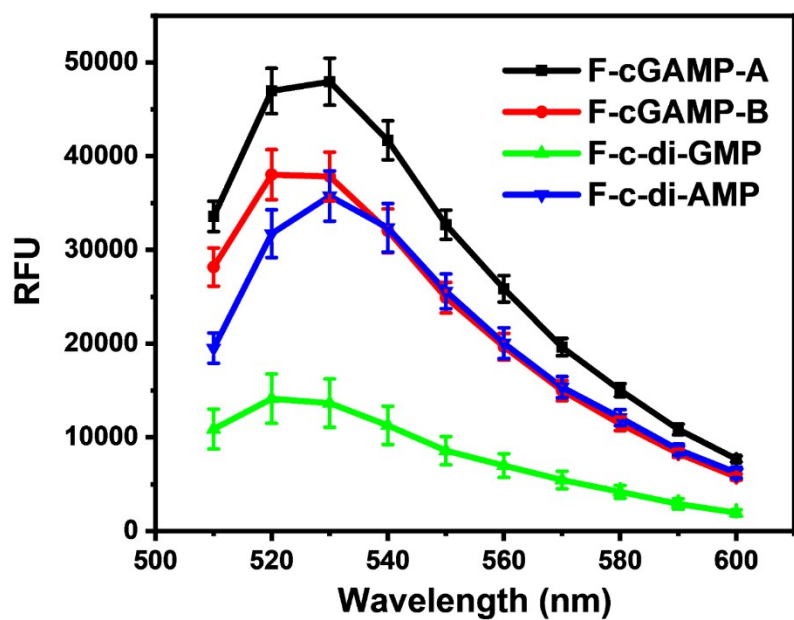


Fig. S2: F-cGAMP-A, F-cGAMP-B, F-c-di-GMP and F-c-di-AMP emission spectra. Emission spectra generated using 12.5 nM probe in 1x PBS. Excitation: 480/20 nm. Error bars represent standard deviation of n=3.

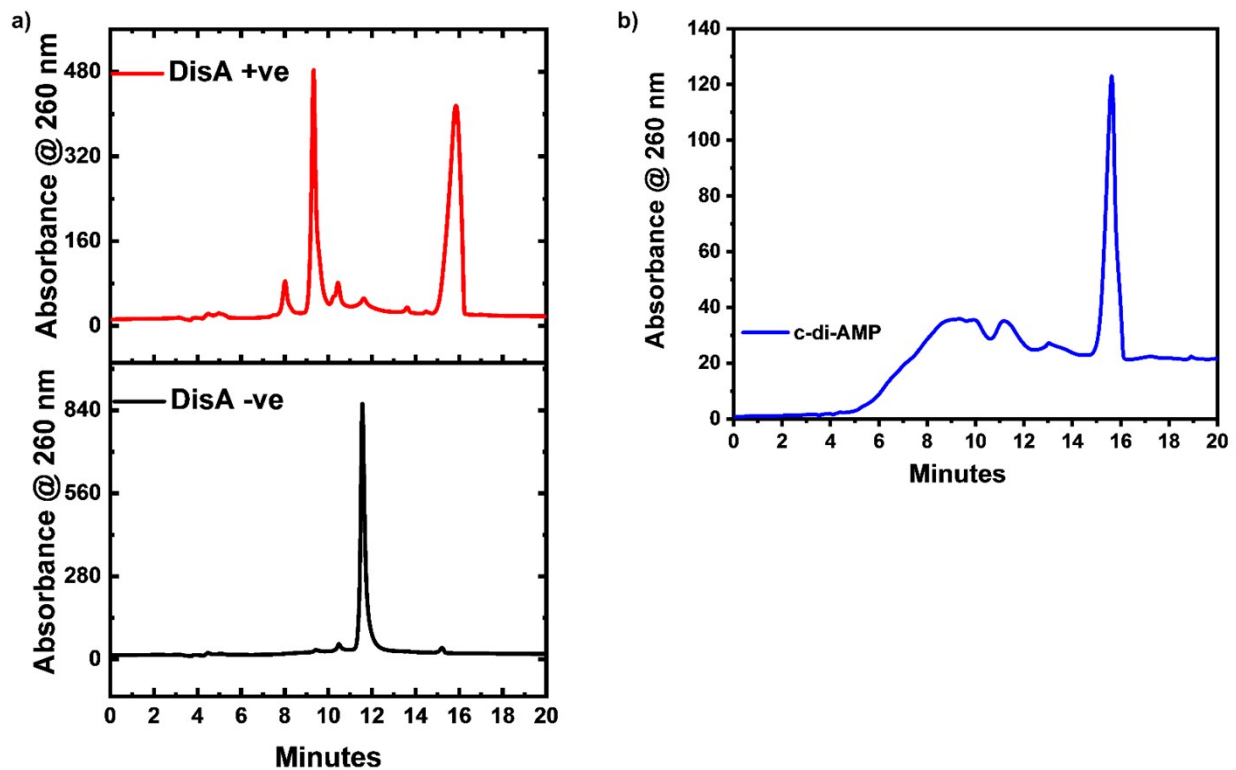


Fig. S3: HPLC traces of DisA reactions. a) HPLC traces of DisA +ve and DisA -ve reactions. DisA reactions: 1 μM enzyme and 100 μM ATP in 1x reaction buffer (40 mM Tris-HCl pH 7.5, 100 mM NaCl, and 10mM MgCl_2) and incubate at 37 $^\circ\text{C}$ for 16 h. DisA -ve = Reactions containing no DisA. DisA +ve = Reactions containing DisA. b) HPLC trace of c-di-AMP standard.

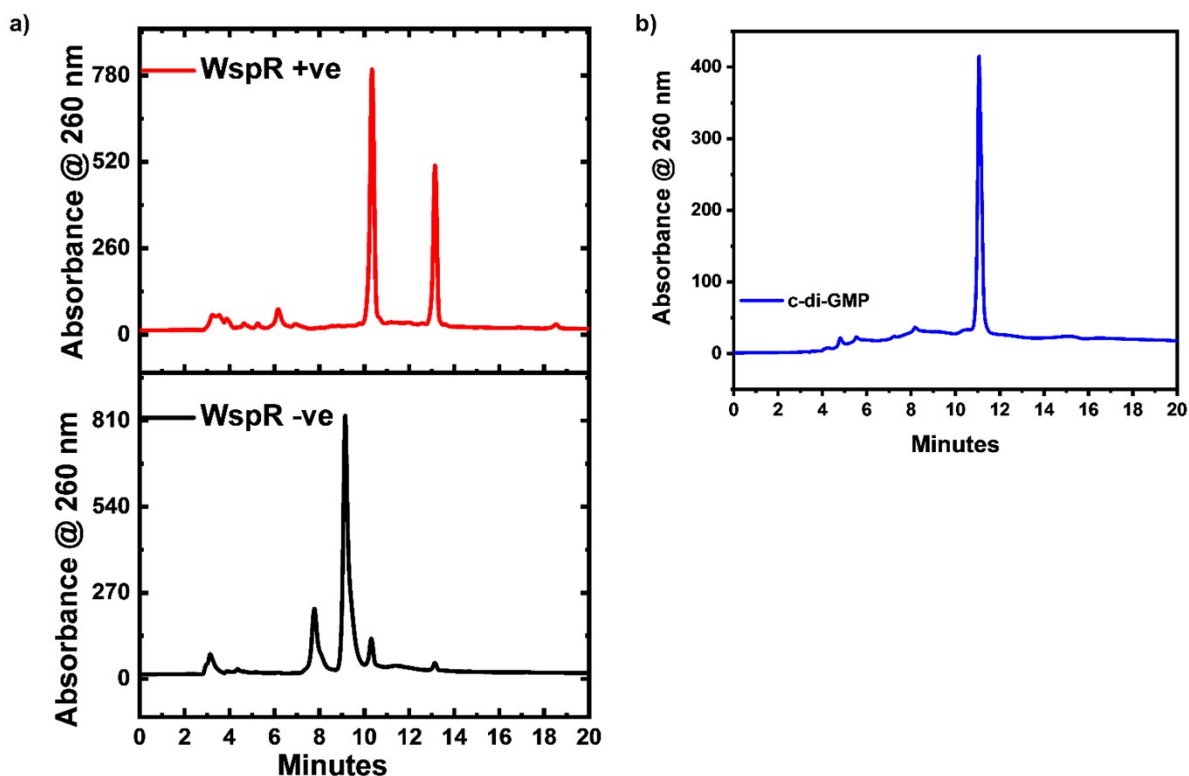


Fig. S4: HPLC traces of WspR reactions. a) HPLC traces of WspR +ve and WspR -ve reactions. WspR reactions: 11 μM enzyme and 100 μM GTP in with 1x reaction buffer (10 mM Tris-HCl pH 7.5, 100 mM NaCl, and 5mM MgCl_2) and incubate at 37 $^\circ\text{C}$ for 16 h. WspR -ve = Reactions containing no WspR. WspR +ve = Reactions containing WspR. b) HPLC trace of c-di-GMP standard.

Purity analysis of fluorescent cyclic dinucleotides via HPLC. Fluorescent cyclic - dinucleotides were dissolved in water to yield a 100 μM solution, and 100 μL of the solution was subjected to HPLC analysis using a AGILENT Eclipse Plus C18- Packed column (particle size 3.5 μm ; 4.6 x 100mm) (mobile phase = water and acetonitrile). Gradient is as follows: 0 – 16 min: 99 % - 87 % water, 1% -13% acetonitrile, 16 – 23 min: 87 % - 10 % water, 13 % - 90 % acetonitrile, 23 – 25 min: 10 % - 99 % water, 90 % -1 % acetonitrile. Nucleotides were detected by measuring absorbance at 260 nm.

Mass analysis of fluorescent cyclic dinucleotides via MALDI-TOF MS. Fluorescent cyclic dinucleotides were dissolved to yield a 10 μM solution and was desalted prior to analysis using ZipTip pipette tips. A 10 mg/mL solution of α -cyano-4-hydroxycinnamic acid in a 1:1 mixture of acetonitrile and 0.1% trifluoroacetic acid in water was used as the matrix. Aliquots of the desalted sample were combined with an equal volume of the matrix and transferred in 1 μL quantities onto a 400-well MALDI plate. MALDI-TOF/TOF spectra was obtained using a 4800 Proteomics Analyzer (Applied Biosystems). Spectra were acquired in the positive ion mode.

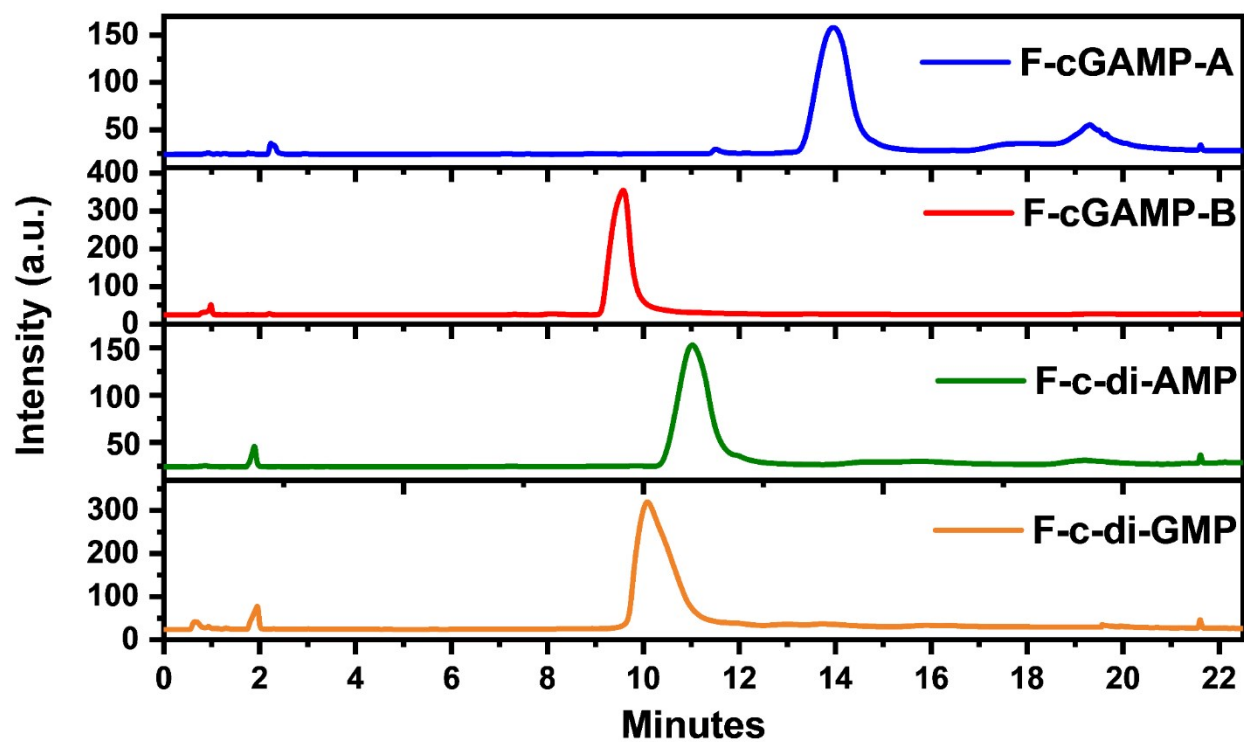
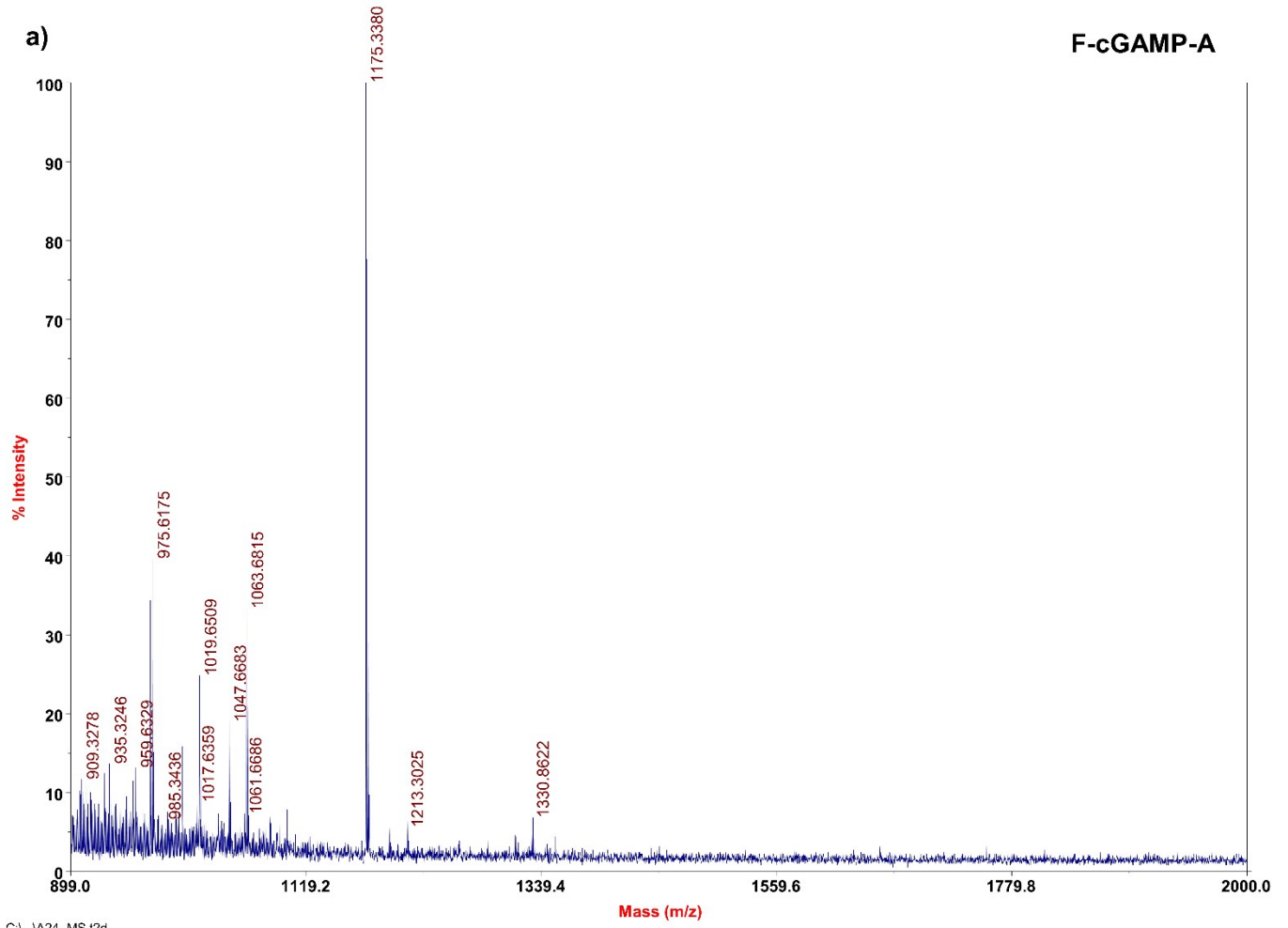


Fig. S5: HPLC traces of Fluorescent cyclic dinucleotides used for the fluorescence polarization assay. Fluorescent cyclic dinucleotides used have a purity of >90% as quantified by HPLC analysis.

a)

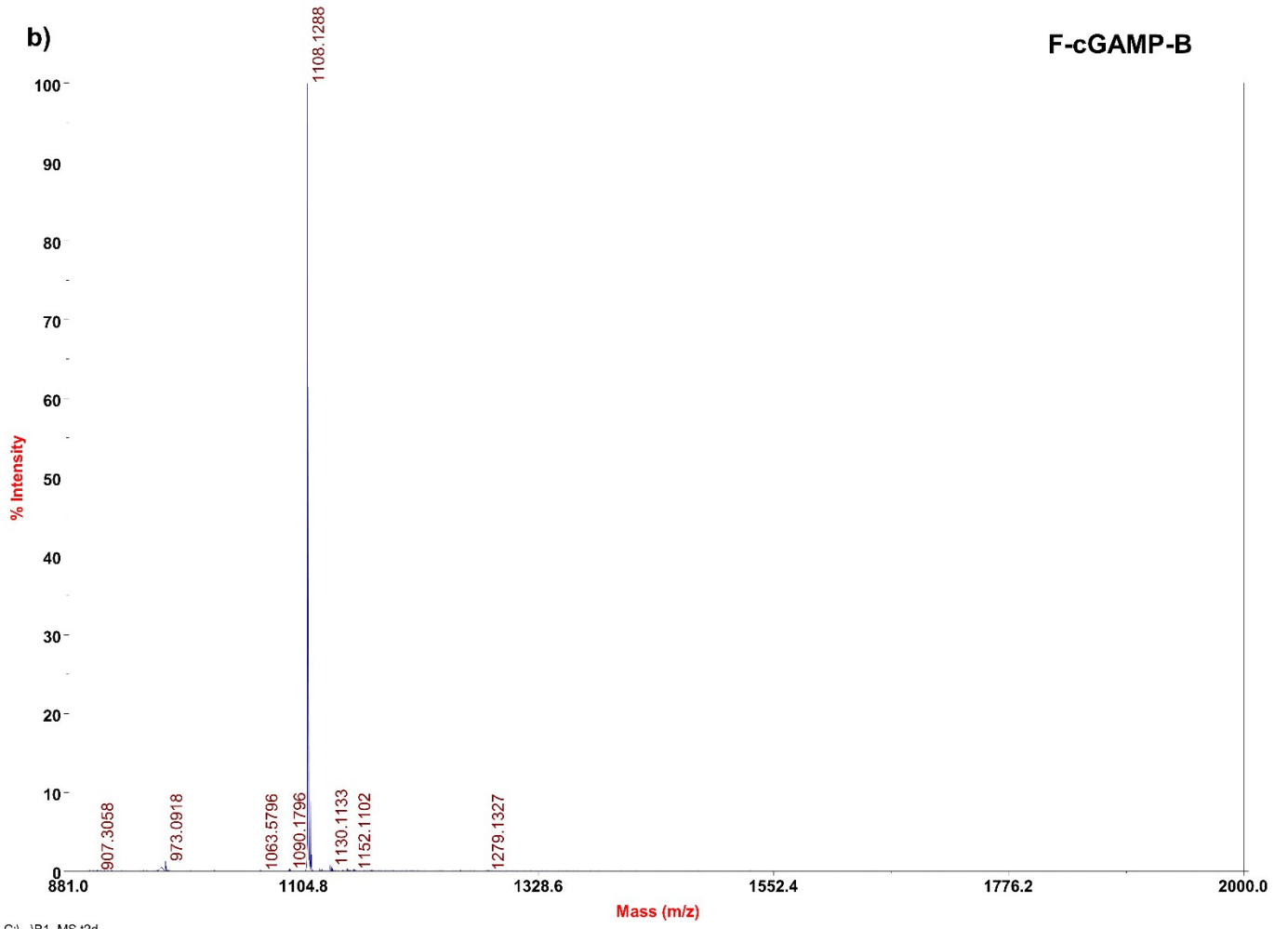
F-cGAMP-A



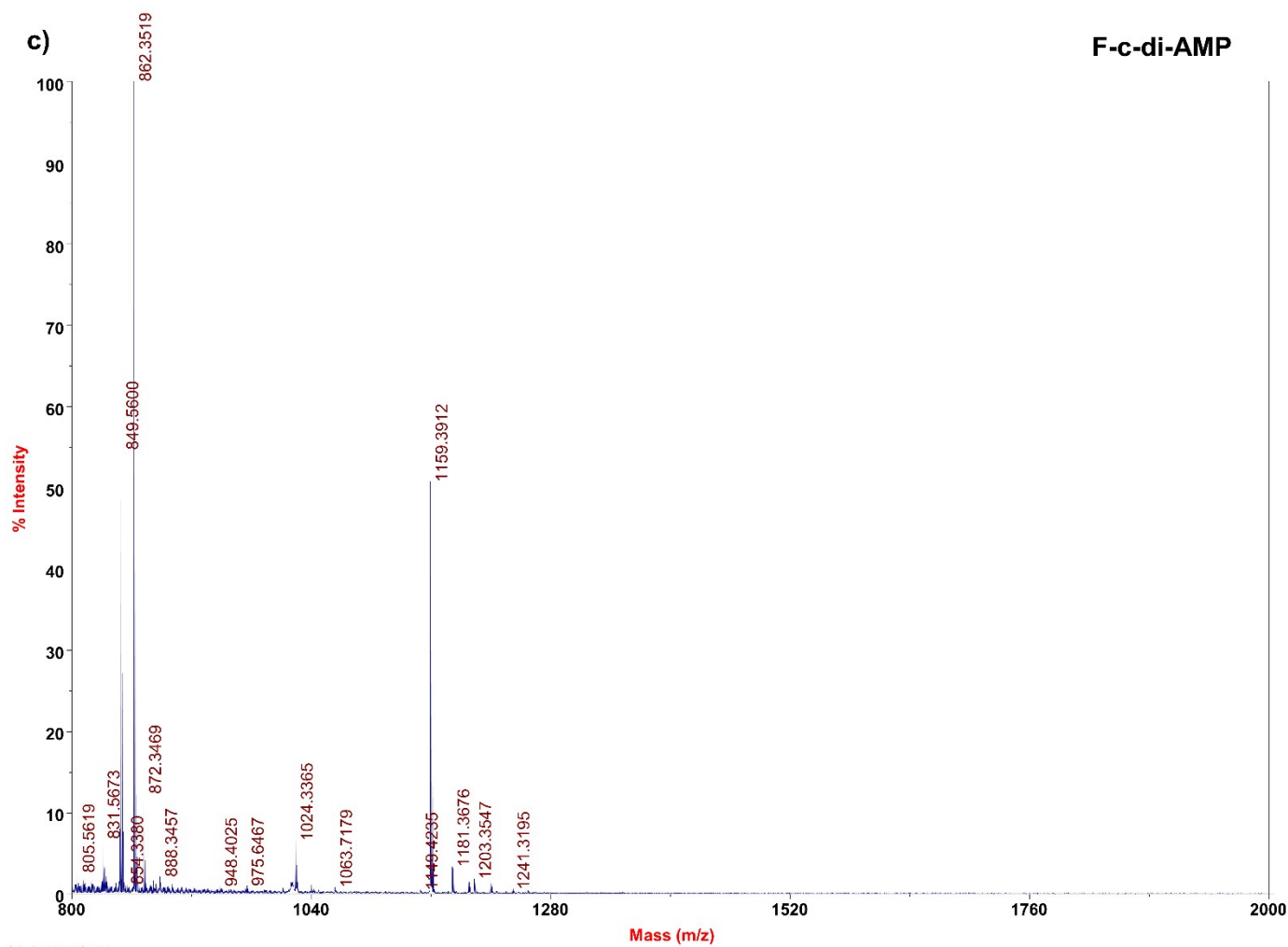
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b)

F-cGAMP-B



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C:\...V19_MS.t2d
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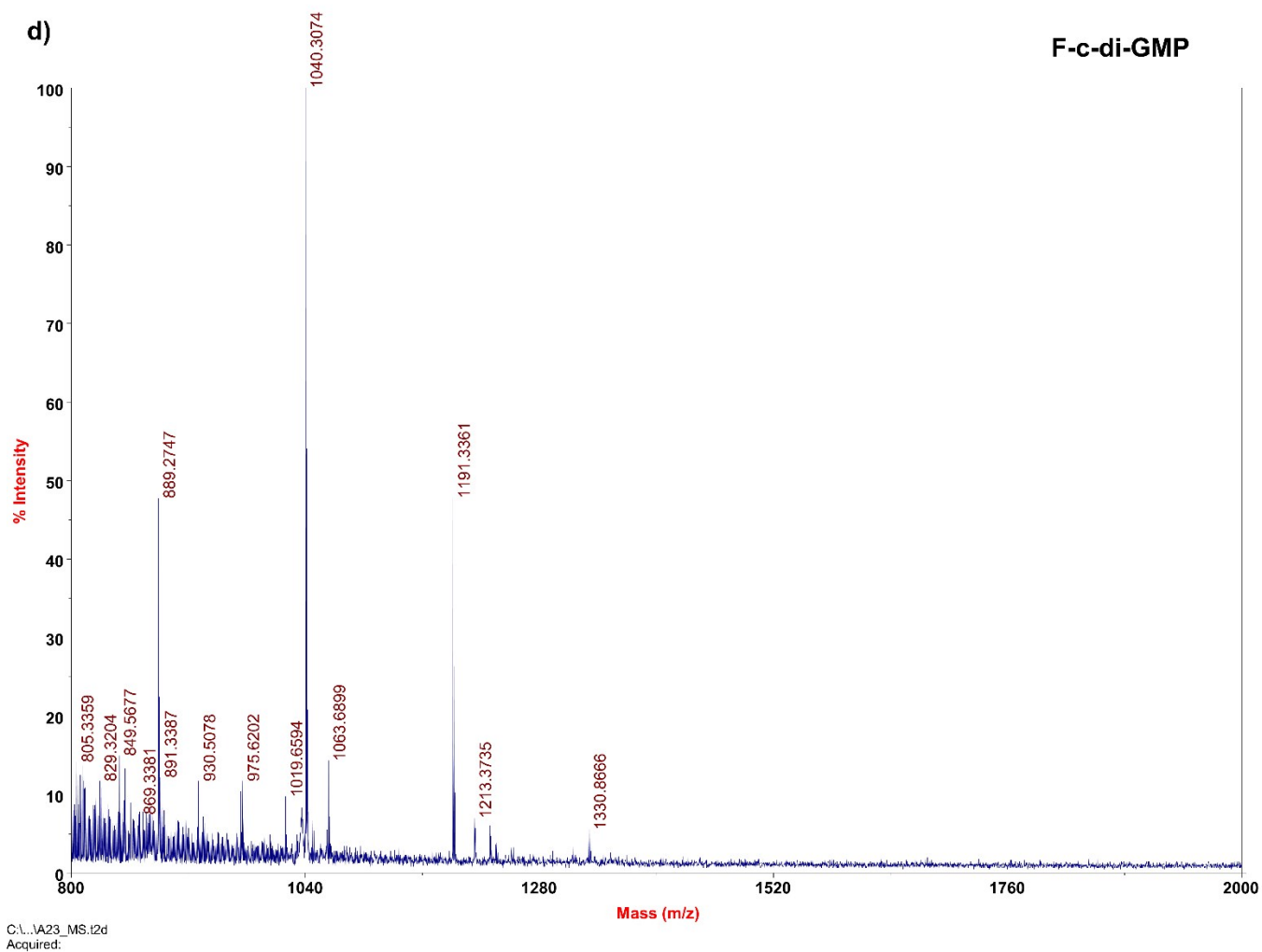


Fig. S6. MALDI-TOF Spectra of Fluorescent cyclic dinucleotides used for the fluorescence polarization assay. a) MALDI-TOF spectra of F-cGAMP-A. Manufacturer reported mass: 1174.9. Found: 1175.3380 ($M+H^+$); b) MALDI-TOF spectra of F-cGAMP-B. Manufacturer reported mass: 1107.9. Found: 1108.1288 ($M+H^+$); c) MALDI-TOF spectra of F-c-di-AMP. Manufacturer reported mass: 1158.9. Found: 1159.3912 ($M+H^+$); d) MALDI-TOF spectra of F-c-di-GMP. Manufacturer reported mass: 1190.9. Found: 1191.3361 ($M+H^+$).