

Supporting Information

Rhodium-Catalyzed Sommelet Hauser Type Rearrangement of α -Diazoimines: Synthesis of Functionalized Enamides

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1. General Comments:

All reactions were carried out under nitrogen atmosphere using oven dried reaction tubes. Dry toluene was prepared by distilling over sodium metal and stored over molecular sieves 4Å under nitrogen atmosphere. All the *N*-sulfonyl 1,2,3-triazoles were synthesized from corresponding acetylene and tosyl azide employing literature procedure.¹ All the α -thioesters were achieved from thiol and bromoesters.²

Column chromatography was performed using Rankem Silica gel (100-200 mesh) and the solvent system used unless otherwise specified, was ethyl acetate-hexanes with various percentage of polarity depending on the nature of the substrate.

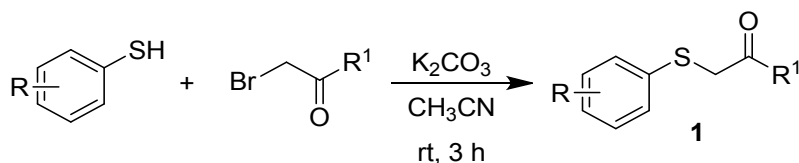
2. Analytical Methods:

NMR data were recorded on 400 and 500 MHz spectrometers. ¹³C and ¹H NMR spectra were referenced to signals of deuterio solvents and residual protiated solvents, respectively. Infrared spectra were recorded on a Thermo Nicolet iS10 FT spectrometer. HRMS were recorded by electron spray ionization (ESI) method on a Q-TOF Micro with lock spray source. Melting points are corrected.

¹ J. Raushel, V. V. Fokin, *Org. Lett.* **2010**, *12*, 4952-4955.

² R. VenkatRagavan, V. Vijaykumar, N. K. Suchetha, *Eur. J. Med. Chem.* **2009**, *44*, 3852-3857

3. General procedure for synthesis of α -thiocarbonyl derivatives 1



To a solution of arylthiols (2.72 mmol, 1 equiv) in CH₃CN (6 mL), was added K₂CO₃ (564 mg, 4.09 mmol, 1.5 equiv) followed by solution of bromocarbonyl compound (4.09 mmol, 1.5 equiv) in CH₃CN (4 mL) was added drop wise over 3 min. After approximately 3 h analyzed by TLC, ethyl acetate was added to the crude products, washed with water, brine and then dried over Na₂SO₄. The organic layer was filtered and evaporated under the reduced pressure. The obtained crude product was purified through column chromatography to afford the pure thiocarbonyl compounds in high yields.

4. Properties of isolated α -thiocarbonyl derivatives 1

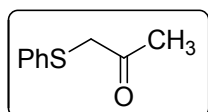
Thioester (2a): 395 mg; Light yellow liquid; Yield: 89%; R_f = 0.63 in 10:90 ethyl acetate/hexane; FTIR (Neat): 3213, 3149, 3142, 2922, 2337, 1635, 1544, 1507, 1421, 1401, 1336, 1260, 1195, 1098, 961 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, 24 °C): δ 7.44-7.38 (m, 2H), 7.33-7.19 (m, 3H), 4.16 (q, 2H, J = 7.1 Hz), 3.63 (s, 3H), 1.22 (t, 3H, J = 7.1 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C): δ 145.2, 136.2, 135.7, 134.5, 130.5, 129.9, 129.2, 128.6, 127.2, 122.0, 28.9; HRMS: calcd. for C₁₀H₁₂O₂S+H: 197.0631; found: 197.0632.

Thioester (2b): 347 mg; Yellow liquid; yield: 81%; R_f = 0.63 in 10:90 ethyl acetate/hexane; FTIR (Neat): 3272, 3109, 2928, 2856, 2356, 1712, 1642, 1550, 1396, 1270, 1225, 1079, 877 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, 24 °C): δ 7.45-7.37 (m, 2H), 7.35-7.27 (m, 2H), 7.27-7.19 (m, 1H), 3.71 (s, 3H), 3.65 (s, 2H); ¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C): δ 170.3, 135.0, 130.0, 129.2, 127.1, 52.6, 36.6; HRMS: calcd. for C₉H₁₀O₂S+H: 183.0474; found: 183.0479.

Thioester (2c): 330 mg; White solid; yield: 70%; R_f = 0.63 in 10:90 ethyl acetate/hexane; mp: 94-96°C; FTIR (KBr): 3064, 2928, 2859, 1737, 1652, 1581, 1471, 1440, 1272, 1138, 991 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, 24 °C): δ 7.43-7.39 (m, 2H), 7.32-7.27 (m, 2H), 7.25-7.20 (m, 1H), 5.85 (ddt, 1H, J = 17.1, 10.1, 5.6 Hz), 5.28 (dq, 1H, J = 17.1, 1.4 Hz), 5.22 (dt, 1H, J = 10.4, 1.2 Hz), 4.60 (dt, 1H, J = 5.7, 1.3 Hz).

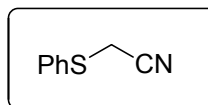
Hz), 3.66 (s, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 169.5, 135.0, 131.7, 130.2, 129.1, 127.1, 118.8, 66.2, 36.8; HRMS: calcd. for $\text{C}_{11}\text{H}_{12}\text{O}_2\text{S}+\text{Na}$: 231.0450; found: 231.0451.

Thioketone (2d): 297 mg; Colourless liquid; yield: 79%; R_f = 0.60 in 10:90 ethyl



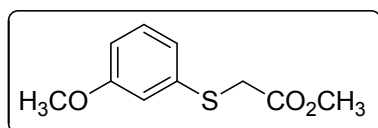
acetate/hexane; FTIR (Neat): 3061, 3004, 2925, 1714, 1638, 1583, 1477, 1430, 1404, 1359, 1278, 1230, 1151, 1088, 743 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3 , 24 °C): δ 7.38-7.24 (m, 4H), 7.24-7.18 (m, 1H), 3.66 (s, 2H), 2.27 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 203.7, 134.7, 129.6, 129.2, 127.0, 44.7, 28.1; HRMS: calcd. for $\text{C}_9\text{H}_{10}\text{OS}+\text{Na}$: 189.0345; found: 189.0345.

2-(phenylthio)acetonitrile (2g): 219 mg; Colorless liquid; yield: 65%; R_f = 0.70 in 10:90



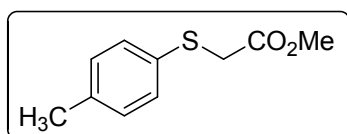
ethyl acetate/hexane; FTIR (Neat): 3405, 3390, 3376, 3362, 3344, 3298, 3289, 3362, 3298, 3171, 3057, 2927, 2856, 2307, 1739, 1586, 1472, 1431, 1266, 1195, 1034, 894 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3 , 24 °C): δ 7.65-7.49 (m, 2H), 7.46-7.31 (m, 3H), 3.56 (s, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 132.5, 132.2, 129.7, 129.1, 116.6, 21.5; HRMS: calcd. for $\text{C}_8\text{H}_7\text{NS}+\text{K}$: 187.9931; found: 188.0137.

Thioester (2h): 356 mg; Colorless liquid; yield: 74%; R_f = 0.57 in 10:90 ethyl



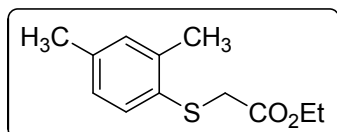
acetate/hexane; FTIR (Neat): 3057, 2987, 2848, 2685, 2522, 2412, 2306, 2136, 1738, 1598, 1427, 1266, 1159, 1039, 985 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3 , 24 °C): δ 7.21 (t, 1H, J = 7.9 Hz), 7.02-6.89 (m, 2H), 6.81-6.69 (m, 1H), 3.79 (s, 3H), 3.73 (s, 3H), 3.66 (s, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 170.2, 160.0, 136.3, 130.0, 121.7, 114.9, 112.8, 55.4, 52.7, 36.3; HRMS: calcd. for $\text{C}_{10}\text{H}_{12}\text{O}_3\text{S}+\text{Na}$: 235.0399; found: 235.0393.

Thioester (2i): 391 mg; Light green liquid; yield: 88%; R_f = 0.64 in 10:90 ethyl



acetate/hexane; FTIR (Neat): 3299, 3197, 3135, 3120, 3055, 2982, 2930, 2357, 1733, 1638, 1605, 1472, 1464, 1407, 1274, 1135, 1030 810 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3 , 24 °C): δ 7.36-7.28 (m, 2H), 7.14-7.07 (m, 2H), 3.69 (s, 3H), 3.59 (s, 2H), 2.31 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3 , 24 °C): δ 170.4, 137.5, 131.0, 129.9, 52.5, 37.3, 21.1; HRMS: calcd. for $\text{C}_{10}\text{H}_{12}\text{O}_2\text{S}+\text{H}$: 197.0631; found: 197.0634.

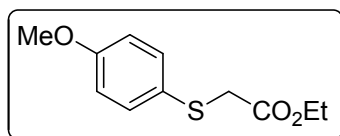
Thioester (2j): 361 mg; Colorless liquid; yield: 71%; R_f = 0.67 in 10:90 ethyl acetate/hexane;



FTIR (Neat): 3227, 3056, 2981, 2871, 2346, 2333, 1733, 1651, 1606, 1448, 1271, 1133, 1028, 741 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3 , 24 °C): δ 7.28 (d, 1H, J = 7.7 Hz), 7.01 (m, 1H), 6.96 (m, 1H), 4.13 (q, 2H, J = 7.1 Hz), 3.54 (s, 2H), 2.39 (s, 3H), 2.28 (s, 3H), 1.21 (t, 3H, J = 7.1 Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 169.9, 139.1, 137.4, 131.3, 131.2, 130.3,

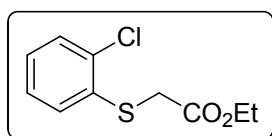
127.4, 61.5, 36.7, 21.0, 20.4, 14.1; HRMS: calcd. for $C_{12}H_{16}O_2S+Na$: 247.0763; found: 247.0764.

Thioester (2k): 356 mg; Colorless liquid; yield: 57%; $R_f = 0.40$ in 15:85 ethyl



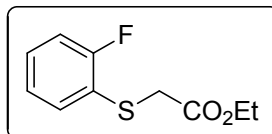
acetate/hexane; FTIR (Neat): 1739, 1494, 1285, 1247, 1030, 911, 762, 669 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 7.42 (d, 2H, $J = 8.9$ Hz), 7.84 (d, 2H, $J = 8.8$ Hz), 4.14 (q, 2H), 3.79 (s, 3H), 3.50 (s, 2H), 1.21 (t, 3H); $^{13}C\{^1H\}$ NMR (125 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 170.1, 159.8, 134.3, 125.1, 114.8, 61.5, 55.5, 38.8, 14.2; HRMS (ESI) m/z : $[M+Na]^+$ calcd for $C_{11}H_{14}O_3S+Na$ 249.2813; Found; 249.2818.

Thioester (2l): 402 mg; Colorless liquid; yield: 77%; $R_f = 0.63$ in 10:90 ethyl acetate/hexane;



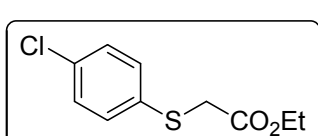
FTIR (Neat): 3057, 2986, 2306, 1734, 1609, 1579, 1444, 1271, 1134, 1030, 897 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 7.39-7.30 (m, 2H), 7.30-7.22 (m, 2H), 4.16 (q, 2H, $J = 7.1$ Hz), 3.60 (s, 2H), 1.22 (t, 3H); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 169.5, 133.5, 133.2, 131.5, 129.2, 61.7, 36.9, 14.1; HRMS: calcd. for $C_{10}H_{11}ClO_2S+H$: 231.0241; found: 231.0242.

Thioester (2m): 408 mg; Light green liquid; yield: 84%; $R_f = 0.62$ in 10:90 ethyl



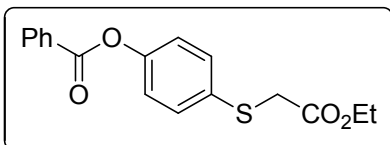
acetate/hexane; FTIR (Neat): 3058, 2987, 2306, 1733, 1580, 1465, 1432, 1272, 1143, 1028, 898 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 7.67-7.51 (m, 1H), 7.49-7.39 (m, 1H), 7.31-7.21 (m, 2H), 4.25 (q, 2H, $J = 7.0$ Hz), 4.12 (s, 2H), 1.29 (t, 3H, $J = 7.0$ Hz); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 168.0, 163.4, 152.1, 141.8, 124.3 (d, $J = 30.4$ Hz), 118.7, 110.1, 62.3, 34.3, 14.2; HRMS: calcd. for $C_{10}H_{12}FO_2S+Na$: 237.0356; found: 237.0356.

Thioester (2n): 417 mg; Light yellow liquid; yield: 80%; $R_f = 0.64$ in 10:90 ethyl



acetate/hexane; FTIR (KBr): 3127, 3055, 2981, 2934, 1733, 1625, 1588, 1457, 1283, 1155, 1019, 946 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 7.42-7.32 (m, 2H), 7.31-7.22 (m, 2H), 4.16 (q, 2H, $J = 7.1$ Hz), 3.60 (s, 2H), 1.22 (t, 3H, $J = 7.1$ Hz); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 169.5, 133.5, 133.2, 131.5, 129.2, 61.7, 36.9, 14.1; HRMS: calcd. for $C_{10}H_{11}ClO_2S+Na$: 253.0060; found: 253.0062.

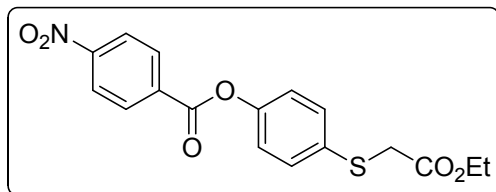
Thioester (2o): 564 mg; White semi solid; yield: 78%; $R_f = 0.56$ in 15:85 ethyl



acetate/hexane; FTIR (KBr): 2971, 2952, 2926, 2858, 1738, 1678, 1648, 1630, 1590, 1545, 1457, 1396, 1264, 1201, 1170, 1069, 1085, 935 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$, 24 $^{\circ}C$): δ 8.23-8.14 (m, 2H), 7.67-7.59 (m, 1H), 7.55-7.46 (m, 4H), 7.21-7.13 (m, 2H), 4.17 (q,

2H, $J = 7.0$ Hz), 3.63 (s, 2H), 1.24 (t, 3H, $J = 7.0$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 169.7, 165.0, 150.4, 133.8, 132.3, 132.0, 130.3, 129.4, 128.7, 122.5, 61.7, 37.4, 14.2; HRMS: calcd. for $\text{C}_{17}\text{H}_{16}\text{O}_4\text{S}+\text{H}$: 317.0842; found: 317.0842.

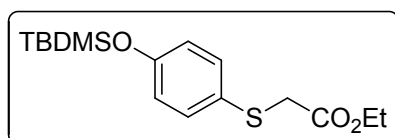
Thioester (2p): 420 mg; White semi solid; yield: 49%; $R_f = 0.56$ in 20:85 ethyl



acetate/hexane; FTIR (KBr): 3051, 2971, 2825, 1738, 1678, 1658, 1634, 1590, 1545, 1457, 1396, 1264, 1201, 1172, 1069, 1084 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3 , 24 °C): δ 8.42-8.19 (m, 4H), 7.61-7.51

(m, 2H), 7.25-7.19 (m, 2H), 4.17 (q, 2H, $J = 7.0$ Hz), 3.63 (s, 2H), 1.23 (t, 3H, $J = 7.0$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 169.6, 163.2, 151.0, 149.6, 134.7, 133.1, 131.7, 131.4, 123.8, 122.1, 61.7, 37.1, 14.1; HRMS: calcd. for $\text{C}_{17}\text{H}_{16}\text{O}_4\text{S}+\text{H}$: 361.3691; found: 361.3697.

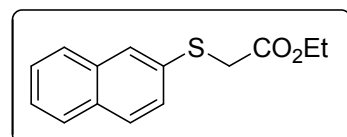
Thioester (2q): 518 mg; Colorless liquid; yield: 70%; $R_f = 0.45$ in 15:85 ethyl



acetate/hexane; FTIR (Neat): 3056, 2948, 2863, 2307, 1731, 1591, 1487, 1419, 1266, 1156, 1027, 910 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3 , 24 °C): δ 7.21-7.10 (m, 2H), 6.68-6.49

(m, 2H), 3.94 (q, 2H, $J = 7.0$ Hz), 3.33 (s, 2H), 1.02 (t, 3H, $J = 7.1$ Hz), 0.78 (s, 9H), 0.00 (s, 6H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 170.0, 155.9, 133.9, 125.9, 120.9, 61.4, 38.5, 25.7, 18.3, 14.2, -4.3; HRMS: calcd. for $\text{C}_{16}\text{H}_{26}\text{O}_3\text{SSi}+\text{Na}$: 349.1264; found: 349.1265.

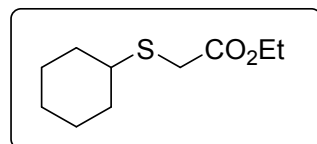
Thioester (2r): 429 mg; liquid; yield: 77%; $R_f = 0.64$ in 10:90 ethyl acetate/hexane; FTIR



(Neat): 3273, 3166, 3057, 2978, 2929, 1733, 1656, 1623, 1588, 1454, 1397, 1388, 1376, 1275, 1134, 1027 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3 , 24 °C): δ 7.84 (s, 1H), 7.82-7.70 (m, 3H), 7.51-

7.39 (m, 3H), 4.16 (q, 2H, $J = 7.1$ Hz), 3.73 (s, 2H), 1.20 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 169.8, 133.7, 132.5, 132.2, 128.7, 128.2, 127.8, 127.6, 127.3, 126.7, 126.1, 61.7, 36.7, 14.2; HRMS: calcd. for $\text{C}_{14}\text{H}_{14}\text{SO}_2+\text{H}$: 247.0780; found: 247.0787.

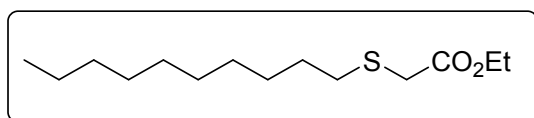
Thioester (2t): 1.68 g; liquid; yield: 89%; $R_f = 0.64$ in 5:95 ethyl acetate/hexane; FTIR



(Neat): 2981, 2927, 2854, 1731, 1450, 1365, 1268, 1133 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3 , 24 °C): δ 4.17-4.12 (s, 2H), 3.22 (s, 2H), 2.77 (s, 1H), 1.96 (m, 2H), 1.75 (m, 2H), 1.59 (d, 1H, $J = 10$ Hz),

1.30-1.23 (m, 7H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C): δ 171.0, 61.3, 44.1, 33.2, 32.2, 26.0, 25.8, 14.2; HRMS: calcd. for $\text{C}_{10}\text{H}_{18}\text{SO}_2+\text{Na}$: 225.0920; found: 225.0920.

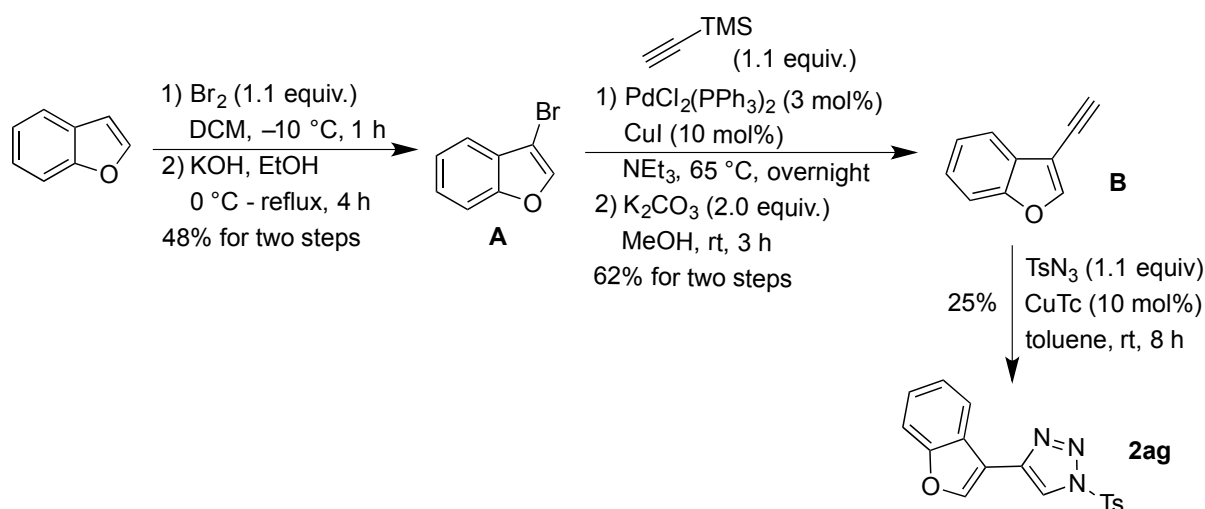
Thioester (2u): 670 mg; liquid; yield: 87%; R_f = 0.64 in 5:95 ethyl acetate/hexane; FTIR



(Neat): 2980, 2925, 28332, 1735, 1445, 1360, 1264, 1132 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3 , 24

$^\circ\text{C}$): δ 4.24-4.15 (m, 2H), 3.19 (s, 2H), 2.62 (t, 2H, J = 8.4 Hz), 1.59 (m, 2H), 1.38-1.25 (m, 17H), 0.90 (t, 3H, J = 6.9 Hz), 1.30-1.23 (m, 7H).; $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 $^\circ\text{C}$): δ 170.8, 61.4, 33.9, 32.8, 32.0, 29.7, 29.6, 29.4, 29.3, 29.1, 28.9, 22.8, 14.3.; HRMS: calcd. for $\text{C}_{14}\text{H}_{28}\text{SO}_2 + \text{Na}$: 283.1702; found: 283.1695.

5. Synthesis of 4-(Benzofuran-3-yl)-1-tosyl-1H-1,2,3-triazole (2ag):



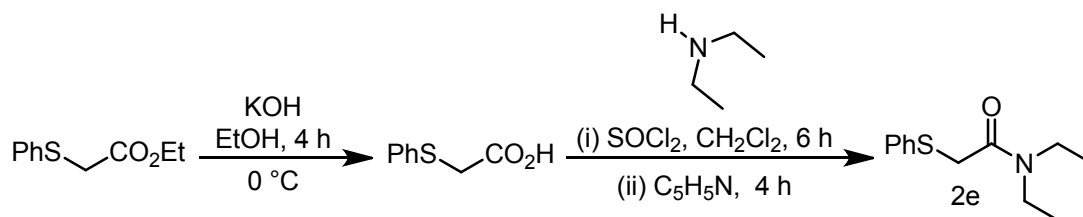
In a round bottom flask, benzofuran (1 g, 1.0 equiv.) was dissolved in 20 mL of DCM and cooled to $-10\text{ }^\circ\text{C}$. Bromine (0.6 mL, 1.1 equiv.) in DCM (3 mL) was slowly added at same temperature. The reaction mixture was stirred for 1 h at $-10\text{ }^\circ\text{C}$. Subsequently, aq. NaOH solution (1 mL, 1 M) was added and the reaction mixture was diluted with water (40 mL) followed by $\text{Na}_2\text{S}_2\text{O}_3$ solution (20 mL) was added. The resultant reaction mixture was extracted with DCM and the combined organic extract was dried over anhydrous Na_2SO_4 . The organic layer was filtered and concentrated under reduced pressure and the residue obtained was subjected for next step without any further purification. The dibromide obtained was dissolved in ethanol (20 mL) and saturated solution of KOH in ethanol (20 mL) was added dropwise at $0\text{ }^\circ\text{C}$. Next, the reaction mixture was kept for refluxing for 4 h. After refluxing for 4 h, water was added (10 mL) and the reaction mixture containing EtOH was removed under reduced pressure. The aqueous residue was extracted with ethyl acetate and the organic extract was washed successively with water and brine followed by dried over anhydrous Na_2SO_4 . Upon filtration, this organic extract was concentrated under reduced

pressure and the residue was purified by silica gel flash column chromatography (petroleum ether: EtOAc, 20:1) and the product **A** was obtained in 56% yield as a dark yellow oil.

The 3-bromobenzofuran **A** was taken in round bottom flask along with trimethylsilylacetylene (1.1 equiv), PdCl₂(PPh₃)₂ (3 mol%), CuI (10 mol%) and Et₃N (10 mL). The reaction flask was sealed and kept for reflux overnight. The reaction mixture was cooled to room temperature and filtered through Celite. Evaporation solvent followed by column chromatography using ethylacetate : hexane (1:9) as an eluent afforded the trimethylsilylalkyne in 72% yield. The isolated trimethylsilylalkyne was treated with K₂CO₃ (2.0 equiv) in MeOH (8 mL) at room temperature for 3 hours. The reaction was monitored by TLC, after complete consumption of alkyne, the reaction mixture was diluted with water (10 mL) and extracted with DCM. Evaporation of solvent followed by column chromatography furnished the compound **B** in 85% yield.

The compound **B** (1.0 equiv) was taken in round bottom flask, followed by TsN₃ (1.1 equiv), CuTc (10 mol%) and toluene (4 mL) were added at room temperature and stirred for 8 hours. Evaporation of solvent followed by column purification using ethyl acetate : hexane (20:80) gave the compound **2ag** in 25% yield. FTIR (KBr): 2950, 2514, 1605, 1415, 123, 586cm⁻¹; ¹H NMR (400 MHz, CDCl₃, 24 °C): δ 8.38 (s, 1H), 8.14 (s, 1H), 8.05 (d, 1H, *J* = 7.8 Hz); 7.90 (d, 1H, *J* = 7.1 Hz), 7.54 (d, 1H, *J* = 7.5 Hz), 7.40-7.28 (m, 6H), 2.44 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C): δ 155.6, 147.6, 143.3, 139.8, 133.1, 132.6, 128.9, 125.3, 123.6, 120.7, 118.9, 111.9, 111.4, 111.1, 104.8, 21.9.; HRMS: *m/z*: [M+H]⁺ Calcd. for C₁₇H₁₃N₃O₃S+Na: 362.0570; found: 362.0569.

6. Synthesis of *N,N*-diethyl-2-(phenylthio)acetamide:



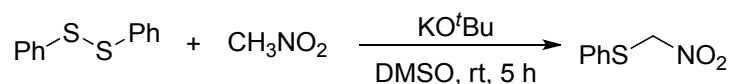
A solution of ethyl 2-(phenylthio)acetate (0.80 g, 4.0 mmol) in EtOH (10 mL) was added to an aqueous solution (10 mL) of potassium hydroxide (2.1 g, 38.0 mmol) at 0 °C. After being stirred for 4 h, by the TLC analysis, reaction mixture was evaporated and was acidified by 2 M HCl and extracted with EtOAc. The organic layer was washed with brine,

dried over anhydrous sodium sulfate, and concentrated in vacuum, purified through column chromatography to give 2-(phenylthio)acetic acid as a colorless liquid.

To a solution 2-(phenylthio)acetic acid (0.60 g, 3.57 mmol, 1 equiv) in dichloromethane, thionyl chloride (0.77 mL, 10.71 mmol, 3 equiv) was added and stirred at room temperature for 6 h. After consumption of acid (by TLC), excess thionyl chloride was then removed under reduced pressure to afford (phenylsulfanyl)acetyl chloride as a yellow liquid.

Pyridine (0.56 mL, 7.14 mmol, 2 equiv) was added to a stirred solution of diethylamine (0.74 mL, 7.14 mmol, 2 equiv) in dichloromethane at 0 °C and (phenylsulfonyl)acetyl chloride was also added drop wise to the reaction mixture at same temperature. By TLC analysis the reaction was quenched with water and extracted with dichloromethane, dried over Na₂SO₄ and concentrated in vacuum. Purified through column chromatography to afford corresponding amide as light yellow liquid; 210 mg; yield: 41%; FTIR (KBr): 3415, 2358, 2121, 1647, 1555, 1160, 670, 586 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, 24 °C): δ 7.49-7.41 (m, 2H), 7.34-7.25 (m, 2H), 7.25-7.18 (m, 1H), 3.73 (s, 2H), 3.43-3.28 (m, 4H), 1.19 (t, 3H, *J* = 7.0 Hz), 1.10 (t, 3H, *J* = 7.0 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C): δ 167.6, 135.4, 130.5, 129.0, 127.0, 42.6, 40.5, 37.1, 14.4, 12.9; HRMS: *m/z*: [M+H]⁺ Calcd. for C₁₂H₁₇NOS: 224.1104; found: 224.1109.

7. Synthesis of (nitromethyl)(phenyl)sulfide 2f



In a 50 mL three-neck round-bottomed flask equipped with nitrogen inlet, 5 mL of dry and degassed dimethylsulfoxide and potassium *t*-butoxide (123 mg, 1.10 mmol) were added. The mixture was stirred till the complete dissolution of the base, the nitromethane (44 µL, 0.82 mmol) was added and stirred for 5 min. To the reaction mixture, diphenyldisulfide (92 mg, 0.42 mmol) was added as a suspension. The reaction mixture was stirred for 5 h and quenched with an excess of ammonium nitrate and 30 mL of water was added. Subsequently, the reaction mixture was acidified with dilute HCl to pH-4 and extracted with dichloromethane (3 × 10 mL). The organic layer obtained was washed with water (2 × 10 mL) to remove the traces of dimethylsulfoxide. Subsequently, it was dried with MgSO₄, filtered, and concentrated to give a crude product, which was further purified by column chromatography with silica gel using ethylacetate/hexane as an eluent gave the product in 55

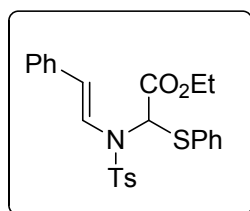
mg, 78% yield as yellow oil. $R_f = 0.40$ in 10:90 ethyl acetate/hexane; FTIR (KBr): 3061, 3002, 2951, 2843, 1739, 1524, 1471, 1357, 1281, 1150, 1039, 855 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3 , 24 $^\circ\text{C}$): δ 7.53-7.45 (m, 2H), 7.40-7.32 (m, 3H), 5.44 (s, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3 , 24 $^\circ\text{C}$): δ 132.2, 131.7, 129.7, 129.3, 79.5; HRMS: calcd. for $\text{C}_7\text{H}_7\text{NO}_2\text{S} + \text{Na}$: 192.0090; found: 192.0096.

8. Typical procedure for the rhodium catalyzed rearrangement reaction

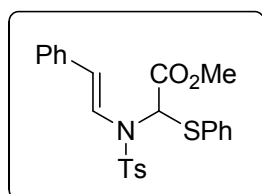
To a 10 mL reaction tube equipped with stir bar, *N*-sulfonyl-1,2,3-triazole **1** (0.20 mmol, 1 equiv), substituted arylsulfide **2** (0.40 mmol, 2 equiv) and $\text{Rh}_2(\text{Oct})_4$ (0.004 mmol, 2 mol%) were introduced. Next, 1.5 mL of dry toluene was added under an argon atmosphere, and the reaction mixture was transferred to preheated oil-bath at 120 $^\circ\text{C}$ and allowed to stir for 8 h. After completion of the reaction, monitored by TLC, the reaction mixture was cooled to room temperature and the solvent was evaporated under reduced pressure. The crude product that obtained was purified by column chromatography using hexane/ethyl acetate mixture as an eluent to afford enamide derivatives **3** in high yield and purity.

9. Properties of isolated enamides

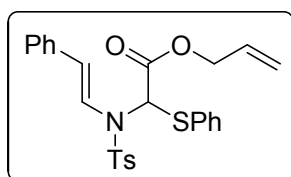
Enamide (4a): 74 mg; Colorless liquid; yield: 80%; $R_f = 0.55$ in 20:80 ethyl acetate/hexane; FTIR (neat): 3196, 3068, 2985, 2928, 2810, 2206, 1953, 1886, 1795, 1737, 1599, 1489, 1341, 1266, 1156, 1092, 905 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.63-7.57 (m, 2H), 7.56-7.50 (m, 2H), 7.40-7.33 (m, 3H), 7.30-7.25 (m, 4H), 7.23-7.16 (m, 3H), 6.94 (d, 1H, $J = 14.6$ Hz), 6.39-6.25 (m, 2H), 4.18-4.02 (m, 2H), 2.38 (s, 3H), 1.12 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.1, 144.2, 136.2, 136.0, 134.6, 133.3, 131.7, 129.6, 129.3, 129.1, 128.7, 127.8, 127.2, 126.0, 123.1, 119.4, 67.3, 62.7, 21.6, 14.0; HRMS: calcd. for $\text{C}_{25}\text{H}_{25}\text{NO}_4\text{S}_2 + \text{Na}$: 490.1117; found: 490.1129.



Enamide (4b): 71 mg; Light green gummy liquid; yield: 79%; $R_f = 0.55$ in 20:80 ethyl acetate/hexane; FTIR (neat): 3056, 2986, 2357, 1686, 1427, 1265, 1171, 897, 740 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.60-7.55 (m, 2H), 7.54-7.48 (m, 2H), 7.38-7.33 (m, 3H), 7.31-7.26 (m, 4H), 7.24-7.18 (m, 3H), 6.94 (d, 1H, $J = 14.7$ Hz), 6.35 (s, 1H), 6.29 (s, 1H, $J = 14.7$ Hz), 3.65 (s, 3H), 2.39 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.6, 144.3, 135.9, 134.5, 133.3, 131.6, 129.6, 129.4, 129.2, 128.7, 127.8, 127.2, 126.0, 123.0, 119.4, 67.1, 53.4, 21.7; HRMS: calcd. for $\text{C}_{24}\text{H}_{23}\text{NO}_4\text{S}_2 + \text{Na}$: 476.0961; found: 476.0966.

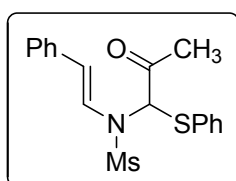


Enamide (4c): 62 mg; Colorless liquid; yield: 65%; R_f = 0.52 in 20:80 ethyl acetate/hexane;



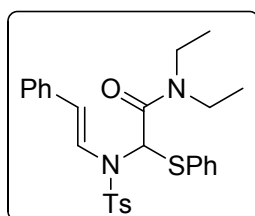
FTIR (neat): 3061, 2933, 2865, 2805, 1952, 1883, 1742, 1645, 1597, 1444, 1355, 1264, 1167, 1089, 1000, 937 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.63 (m, 2H), 7.56-7.50 (m, 2H) 7.40-7.33 (m, 3H), 7.31-7.25 (m, 4H), 7.23-7.18 (m, 3H), 6.94 (d, 1H, J = 14.6 Hz), 6.37 (s, 1H), 6.30 (d, 1H, J = 14.6 Hz), 5.72 (ddt, 1H, J = 17.2, 10.5, 5.9 Hz), 5.30-5.16 (m, 2H), 4.61-4.44 (m, 2H), 2.38 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 166.8, 144.3, 135.9(8), 135.9(5), 134.6, 131.6, 130.9, 129.6, 129.4, 129.2, 128.7, 127.8, 127.2, 126.0, 122.9, 119.5, 119.4, 67.2, 67.1, 21.7; HRMS: calcd. for $\text{C}_{26}\text{H}_{25}\text{NO}_4\text{S}_2 + \text{Na}$: 502.1117; found: 502.1126

Enamide (4d): 40 mg; Light green gummy liquid; yield: 56%; R_f = 0.49 in 20:80 ethyl acetate/hexane;



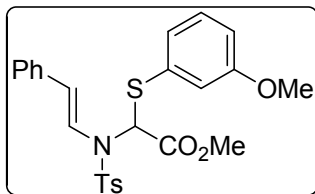
FTIR (neat): 3281, 3063, 2926, 2857, 2477, 2357, 1953, 1896, 1716, 1597, 1442, 1406, 1327, 1213, 1156, 976 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.61-7.55 (m, 2H), 7.38-7.34 (m, 3H), 7.33-7.31 (m, 4H), 7.25-7.19 (m, 1H), 6.97 (d, 1H, J = 14.7 Hz), 6.32 (d, 1H, J = 14.7 Hz), 6.28 (s, 1H), 2.70 (s, 3H), 2.39 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 200.9, 135.4, 134.0, 132.1, 129.7, 129.1, 128.8, 127.5, 126.0, 122.8, 118.0, 75.1, 39.8, 27.0; HRMS: calcd. for $\text{C}_{18}\text{H}_{19}\text{NO}_3\text{S}_2 + \text{H}$: 362.0879; found: 362.0882.

Enamide (4e): 70 mg; White solid; yield: 71%; R_f = 0.45 in 20:80 ethyl acetate/hexane;



FTIR (KBr): 3183, 3057, 2984, 2935, 2411, 2363, 2306, 2127, 2056, 1933, 1890, 1800, 1732, 1653, 1435, 1350, 1266, 1163, 1039, 949 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.56-7.43 (m, 4H), 7.39-7.30 (m, 5H), 7.31-7.23 (m, 2H), 7.22-7.13 (m, 4H), 6.75 (d, 1H, J = 14.3 Hz), 6.46 (s, 1H), 3.71-3.41 (m, 2H), 3.44-3.16 (m, 2H), 2.35 (s, 3H), 1.36 (t, 3H, J = 7.1 Hz), 1.07 (t, 3H, J = 7.1 Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 165.2, 144.0, 136.2, 135.9, 132.4, 129.5, 129.4, 128.6, 128.5, 127.7, 127.2, 126.3, 124.9, 124.3, 64.9, 42.7, 41.3, 21.6, 14.5, 12.8; HRMS: calcd. for $\text{C}_{27}\text{H}_{30}\text{N}_2\text{O}_3\text{S}_2 + \text{Na}$: 517.1590; found: 517.1595.

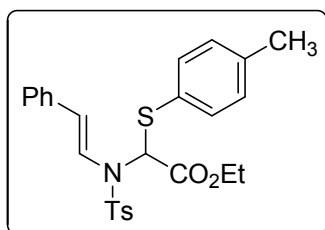
Enamide (4h): 78 mg; Colorless liquid; yield: 81%; R_f = 0.49 in 20:80 ethyl acetate/hexane;



FTIR (neat): 3061, 3018, 2938, 2852, 2357, 1746, 1645, 1589, 1470, 1449, 1351, 1247, 116, 1089, 1032, 940 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.63-7.54 (m, 2H), 7.31-7.26 (m, 4H), 7.24-7.15 (m, 4H), 7.09 (d, 1H, J = 7.6 Hz), 7.04 (s, 1H), 6.98-6.85 (m, 2H), 6.37 (s, 1H), 6.30 (d, 1H, J = 14.6 Hz), 3.78 (s, 3H), 3.67 (s, 3H), 2.39 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.6, 160.0, 144.3, 136.0, 135.9, 132.9,

130.1, 129.6, 128.7, 127.8, 127.3, 126.1, 126.0, 123.1, 119.7, 118.7, 115.5, 67.1, 55.4, 53.4, 21.7; HRMS: calcd. for $C_{25}H_{25}NO_5S_2 + NH_4$: 501.1512; found: 501.1522.

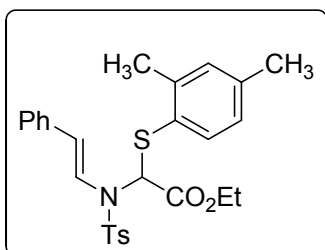
Enamide (4i): 75 mg; Colorless liquid; yield: 72%; R_f = 0.54 in 20:80 ethyl acetate/hexane;



FTIR (neat): 3057, 3028, 2974, 2928, 2865, 2357, 1740, 1645, 1599, 1483, 1450, 1354, 1260, 1166, 1091, 936 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$): δ 7.62-7.53 (m, 2H), 7.44-7.40 (m, 1H), 7.34-7.25 (m, 5H), 7.24-7.13 (m, 5H), 7.01-6.90 (d, 1H, J = 14.6 Hz), 6.39-6.22 (m, 2H), 4.16-4.03 (m, 2H), 2.38 (s, 3H), 2.35 (s,

3H), 1.11 (t, 3H, J = 7.1 Hz); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$): δ 167.1, 144.2, 139.4, 136.1, 136.0, 134.9, 130.1, 129.5, 128.7, 127.7, 127.1, 125.9, 123.1, 118.8, 67.4, 62.7, 21., 21.4, 13.9; HRMS: calcd. for $C_{26}H_{27}NO_4S_2 + Na$: 504.1274; found: 504.1283.

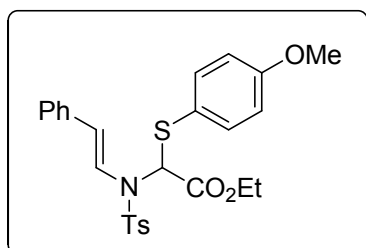
Enamide (4j): 80 mg; Colorless liquid; yield: 81%; R_f = 0.55 in 20:80 ethyl acetate/hexane;



FTIR (neat): 3288, 2974, 2929, 2356, 1740, 1645, 1600, 1450, 1358, 1260, 1166, 1091, 937 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$): δ 7.60-7.49 (m, 2H), 7.46- 7.37 (m, 1H), 7.34 -7.26 (m, 4H), 7.23-7.16 (m, 3H), 7.06 (s, 1H), 7.02-6.87 (m, 2H), 6.36 (d, 1H, J = 14.7 Hz), 6.25 (s, 1H), 4.17-4.02 (m, 2H), 2.44 (s, 3H), 2.38 (s,

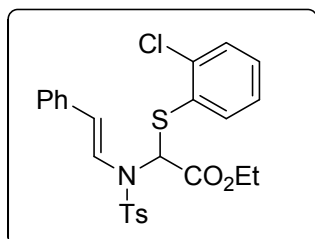
3H), 2.33 (s, 3H), 1.12 (t, 3H, J = 7.1 Hz); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$): δ 167.3, 144.1, 142.3, 139.7, 136.3, 136.2, 136.1, 131.6, 129.5, 128.7, 127.7 (6), 127.7 (2), 127.1, 127.0, 125.9, 123.2, 118.7, 66.5, 62.6, 21.6, 21.3, 20.9, 14.0; HRMS: calcd. for $C_{27}H_{29}NO_4S_2 + H$: 496.1611; found: 496.1615.

Enamide (4k): 56 mg; Colorless liquid; yield: 57%; R_f = 0.48 in 20:80 ethyl acetate/hexane;



FTIR (neat): 3142, 3044, 2928, 2852, 2366, 2250, 1934, 1885, 1739, 1645, 1529, 1490, 1350, 1251, 1167, 1021, 937 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$): δ 7.64-7.58 (m, 2H), 7.48-7.44 (m, 2H), 7.31-7.26 (m, 4H), 7.25-7.19 (m, 3H), 6.96 (d, 1H, J = 14.7 Hz), 6.88-6.84 (m, 2H), 6.31(d, 1H, J = 14.7 Hz), 6.22

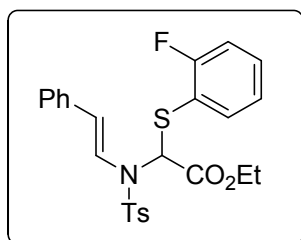
(s, 1H), 4.16-4.03 (m, 2H), 3.81 (s, 3H), 2.39 (s, 3H), 1.11(t, 3H, J = 7.1 Hz); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$): δ 167.1, 160.8, 144.2, 137.2, 136.2, 136.1, 129.6, 128.7, 127.8, 127.1, 125.9, 123.1, 121.7, 118.5, 114.8, 67.7, 62.7, 55.4, 21.7, 14.0; HRMS: calcd. for $C_{26}H_{27}NO_5S_2 + H$: 498.1403; found: 498.1417.



Enamide (4l): 74 mg; Colorless liquid; yield: 74%; R_f = 0.53 in 20:80 ethyl acetate/hexane; FTIR (neat): 3059, 2974, 2928, 2859,

2357, 1904, 1740, 1645, 1599, 1471, 1359, 1262, 1167, 1092, 936 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.66-7.55 (m, 3H), 7.51-7.42 (m, 1H), 7.36-7.17 (m, 9H), 6.91 (1H, d, $J = 14.7$ Hz), 6.54 (s, 1H), 6.31 (1H, d, $J = 14.7$ Hz), 4.18-4.04 (m, 2H), 2.38 (s, 3H), 1.10 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 166.8, 144.3, 139.0, 137.0, 136.0, 130.7, 130.2, 130.0, 129.6, 128.7, 127.8, 127.5, 127.2, 126.0, 122.9, 119.0, 65.4, 62.9, 21.6, 14.0; HRMS: calcd. for $\text{C}_{25}\text{H}_{24}\text{ClNO}_4\text{S}_2 + \text{H}$: 502.0908; found: 502.0919.

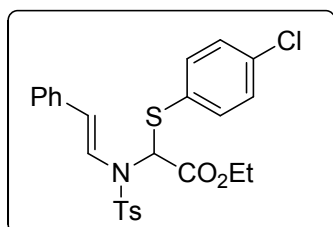
Enamide (4m): 82 mg; Colorless liquid; yield: 84%; $R_f = 0.53$ in 20:80 ethyl acetate/hexane;



FTIR (neat): 3063, 2996, 2858, 2357, 1922, 1740, 1646, 1597, 1468, 1360, 1262, 1167, 1017, 936 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.69-7.56 (m, 2H), 7.50 (t, 1H, $J = 7.4$ Hz), 7.45-7.34 (m, 1H), 7.34-7.08 (m, 10H), 6.89 (d, 1H, $J = 14.7$ Hz), 6.48 (s, 1H), 6.30 (d, 1H, $J = 14.7$ Hz), 4.17-4.03 (m, 2H), 2.39 (s, 3H), 1.09 (t,

3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 166.8, 144.3, 137.6, 136.1 ($J = 8.0$ Hz), 131.9 ($J = 8.2$ Hz), 129.6, 128.7, 127.8, 127.2, 126.0, 124.8 ($J = 3.7$ Hz), 122.7, 119.0, 116.3, 116.1, 65.6, 62.9, 21.6, 14.0; HRMS: calcd. for $\text{C}_{25}\text{H}_{24}\text{FNO}_4\text{S}_2 + \text{Na}$: 508.1023; found: 501.1033.

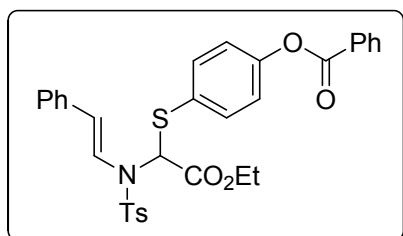
Enamide (4n): 80 mg; Colorless liquid; yield: 80%; $R_f = 0.52$ in 20:80 ethyl acetate/hexane;



FTIR (neat): 3061, 2982, 2860, 1740, 1646, 1599, 1448, 1408, 1348, 1264, 1164, 1021, 968 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.60-7.55 (m, 2H), 7.50-7.40 (m, 2H), 7.35-7.26 (m, 6H), 7.26-7.18 (m, 3H), 6.94 (d, 1H, $J = 14.7$ Hz), 6.31 (s, 1H), 6.28 (d, 1H, $J = 14.7$ Hz), 4.20-4.00 (m, 2H), 2.40 (s, 3H), 1.11 (t, 3H, J

$= 7.0$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 166.9, 144.4, 136.2, 136.0, 135.9, 135.7, 129.9, 129.7, 129.5, 128.8, 127.7, 127.2, 125.9, 122.8, 118.9, 67.2, 62.9, 21.7, 14.0; HRMS: calcd. for $\text{C}_{25}\text{H}_{24}\text{ClNO}_4\text{S}_2 + \text{K}$: 540.0467; found: 540.0457.

Enamide (4o): 78 mg; White semi solid; yield: 67%; $R_f = 0.46$ in 20:80 ethyl acetate/hexane;

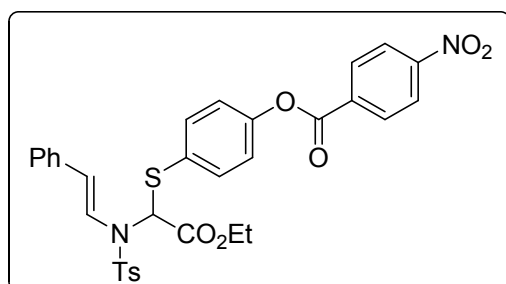


FTIR (neat): 3062, 2977, 2932, 2356, 1740, 1645, 1596, 1487, 1452, 1357, 1260, 1200, 1168, 1063, 937 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 8.23-8.15 (m, 2H), 7.66-7.57 (m, 5H), 7.54-7.49 (m, 2H), 7.32-7.27 (m, 4H), 7.27-7.22 (m, 4H), 7.22-7.20 (m, 1H), 6.97 (d, 1H, $J = 14.7$ Hz), 6.33

(s, 1H), 6.31 (d, 1H, $J = 14.7$ Hz), 4.19-4.06 (m, 2H), 2.39 (s, 3H), 1.13 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.0, 164.9, 151.9, 144.3, 136.2, 133.8, 130.3, 129.7,

129.4, 128.9, 128.8, 128.7, 127.8, 127.2, 126.0, 123.1, 122.7, 119.0, 67.4, 62.8, 21.7, 14.0; HRMS: calcd. for $C_{32}H_{29}NO_6S_2 + Na$: 610.1329; found: 610.1341.

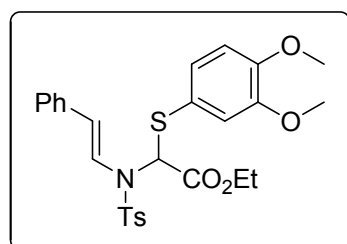
Enamide (4p): 78 mg; Colorless semi solid; yield: 62%; $R_f = 0.42$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 3295, 3166, 3056, 2982, 2926, 2856, 2358, 1797, 1644, 1599, 1437, 1347, 1267, 1201, 1167, 1060, 1201, 1167, 1060, 899 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$): δ 8.40-8.35 (m, 4H), 7.69-7.60 (m, 4H), 7.34-7.28 (m, 4H), 7.28-7.24 (m, 5H), 6.96 (d, 1H, $J = 14.7$ Hz), 6.30

(s, 1H), 6.28 (d, 1H, $J = 14.7$ Hz), 4.19-4.05 (m, 2H), 2.40 (s, 3H), 1.11 (t, 3H, $J = 7.0$ Hz); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$): δ 166.9, 163.0, 151.4, 151.0, 144.4, 136.5, 136.0, 135.9, 134.8, 131.4, 129.7, 129.4, 128.8, 128.2, 127.8, 127.2, 125.9, 123.9, 122.8, 122.4, 118.6, 67.3, 62.9, 21.7, 14.0; HRMS: calcd. for $C_{32}H_{28}N_2O_8S_2 + Na$: 655.1179; found: 655.1182.

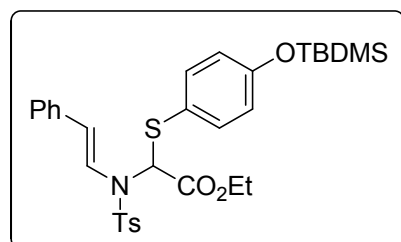
Enamide (4q): 71 mg; White semi solid; yield: 68%; $R_f = 0.45$ in 20:80 ethyl acetate/hexane;



FTIR (neat): 3061, 2948, 2848, 2578, 2328, 1974, 1740, 1644, 1587, 1502, 1453, 1353, 1336, 1252, 1167, 1022, 937 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$): δ 7.63-7.58 (m, 2H), 7.33-7.27 (m, 4H), 7.24-7.19 (m, 3H), 7.13 (dd, 1H, $J = 8.3, 2.0$ Hz), 7.05 (d, 1H, $J = 2.0$ Hz), 6.97 (d, 1H, $J = 14.7$ Hz), 6.82 (d, 1H, $J = 8.3$

Hz), 6.31 (d, 1H, $J = 14.7$ Hz), 6.29 (s, 1H), 4.16-4.03 (m, 2H), 3.89 (s, 3H), 3.84 (s, 3H), 2.39 (s, 3H), 1.12 (t, 3H, $J = 7.1$ Hz); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$): δ 167.1, 150.3, 149.1, 144.2, 136.2, 136.1, 129.6, 128.8, 128.4, 127.7, 127.1, 125.9, 123.2, 122.0, 118.6, 118.0, 111.4, 67.7, 62.7, 56.0 (4), 56.0 (1), 21.7, 14.0; HRMS: calcd. for $C_{27}H_{29}NO_6S_2 + H$: 528.1509; found: 528.1522.

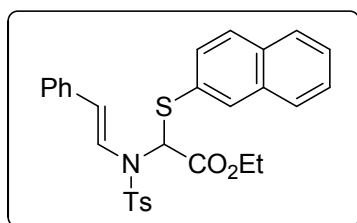
Enamide (4r): 88 mg; Colorless liquid; yield: 74%; $R_f = 0.48$ in 20:80 ethyl acetate/hexane;



FTIR (neat): 3172, 3071, 2950, 2796, 2305, 1893, 1856, 1776, 1592, 1489, 1361, 1265, 1168, 1091, 910 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$): δ 7.42-7.38 (m, 2H), 7.22-7.17 (m, 2H), 7.10-7.06 (m, 4H), 7.03-7.00 (m, 2H), 6.75 (d, 1H, $J = 14.7$ Hz), 6.61-6.56 (m, 2H), 6.11 (d, 1H, $J = 14.7$ Hz),

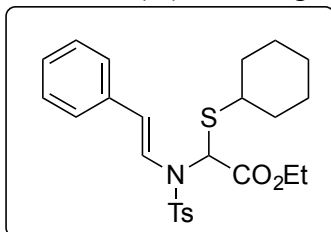
6.01 (s, 1H), 3.94-3.82 (m, 2H), 2.18 (s, 3H), 0.90 (t, 3H, $J = 7.0$ Hz), 0.77 (s, 9H), 0.01 (s, 6H); $^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$): δ 167.2, 157.2, 144.2, 137.0, 136.3, 136.1, 129.6, 128.7, 127.8, 127.1, 125.9, 123.2, 122.7, 121.1, 118.7, 67.6, 62.6, 25.7, 21.6, 18.3, 14.0, -4.3; HRMS: calcd. for $C_{31}H_{39}NO_5S_2Si + Na$: 620.1931; found: 620.1946.

Enamide (4s): 65 mg; White gummy liquid; yield: 63%; $R_f = 0.53$ in 20:80 ethyl



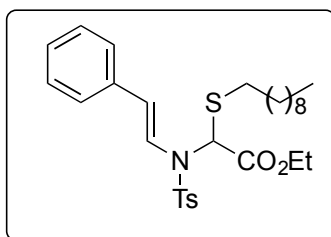
acetate/hexane; FTIR (neat): 3295, 3056, 2969, 2927, 2859, 2356, 1920, 1740, 1644, 1596, 1492, 1450, 1355, 1262, 1167, 1090, 938 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.97-7.91 (m, 1H), 7.85-7.80 (m, 2H), 7.75-7.72 (m, 1H), 7.60 (dd, 1H, $J = 1.7, 8.5$ Hz), 7.53-7.49 (m, 2H), 7.48-7.44 (m, 2H), 7.32-7.27 (m, 4H), 7.24-7.19 (m, 1H), 7.05-6.94 (m, 3H), 6.46(s, 1H), 6.35(d, 1H, $J = 14.7$ Hz), 4.19-4.09 (m, 2H), 2.30 (s, 3H), 1.16 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.2, 144.1, 136.0, 135.9, 134.3, 133.6, 133.2, 130.8, 129.5, 129.1, 128.9, 128.7, 127.9, 127.8, 127.6, 127.2, 127.0, 126.7, 126.0, 123.1, 119.3, 67.4, 62.9, 21.6, 14.0; HRMS: calcd. for $\text{C}_{29}\text{H}_{27}\text{NO}_4\text{S}_2 + \text{NH}_4$: 535.1720; found: 535.1728.

Enamide (4t): 122 mg; Light green gummy liquid; yield: 76%; $R_f = 0.50$ in 20:80 ethyl



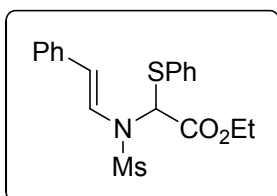
acetate/hexane; FTIR (neat): 2923, 2854, 2358, 1739, 1643, 1450, 1357, 1257, 1222, 1160, 938 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): δ 7.76 (d, 2H, $J = 7.8$ Hz), 7.29-7.25 (m, 7H), 6.35 (d, 1H, $J = 14.9$ Hz), 6.14 (s, 1H), 4.07-3.96 (m, 2H), 2.90 (t, 1H, $J = 9.5$ Hz), 2.41 (s, 3H), 2.16 (d, $J = 11.6$ Hz, 1H), 1.92 (d, 1H, $J = 8.3$ Hz), 1.81-1.72 (m, 3H), 1.49-1.20 (m, 8H), 1.08 (t, 3H, $J = 7.2$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.7, 144.3, 136.2, 129.7, 128.7, 127.7, 127.1, 126.0, 123.1, 119.7, 62.4, 62.1, 44.2, 33.8, 32.9, 26.1, 25.8, 25.8, 21.7, 13.9; HRMS: calcd. for $\text{C}_{25}\text{H}_{31}\text{NO}_4\text{S}_2 + \text{Na}$: 496.1587; found: 496.1583.

Enamide (4u): 122 mg; Light green gummy liquid; yield: 81%; $R_f = 0.50$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 2920, 2851, 2357, 1732, 1634, 1452, 1358, 1241, 1214, 938 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): δ 7.76 (d, 2H, $J = 7.8$ Hz), 7.29-7.25 (m, 7H), 6.35 (d, 1H, $J = 14.9$ Hz), 6.14 (s, 1H), 4.07-3.96 (m, 2H), 2.90 (t, 1H, $J = 9.5$ Hz), 2.41 (s, 3H), 2.16 (d, $J = 11.6$ Hz, 1H), 1.92 (d, 1H, $J = 8.3$ Hz), 1.81-1.72 (m, 3H), 1.49-1.20 (m, 8H), 1.08 (t, 3H, $J = 7.2$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.7, 144.3, 136.2, 129.7, 128.7, 127.7, 127.1, 126.0, 123.1, 119.7, 62.4, 62.1, 44.2, 33.8, 32.9, 26.1, 25.8, 25.8, 21.7, 13.9; HRMS: calcd. for $\text{C}_{25}\text{H}_{31}\text{NO}_4\text{S}_2 + \text{Na}$: 496.1587; found: 496.1583.

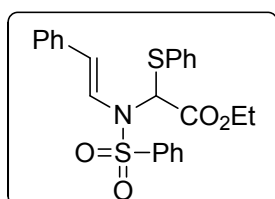
Enamide (4v): 57 mg; Colorless liquid; yield: 73%; $R_f = 0.51$ in 20:80 ethyl acetate/hexane;



FTIR (neat): 3193, 3061, 2982, 2933, 2860, 1740, 1646, 1599, 1448, 1408, 1348, 1264, 1164, 1021, 968 cm^{-1} ; ^1H NMR (400 MHz,

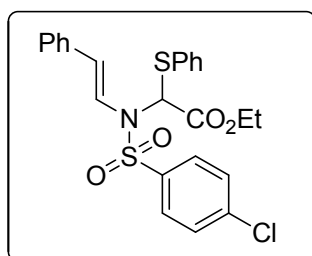
CDCl₃): δ 7.67-7.48 (m, 2H), 7.42-7.28 (m, 7H), 7.27-7.18 (m, 1H), 6.95 (d, 1H, J = 14.7 Hz), 6.38 (d, 1H, J = 14.8 Hz), 6.31 (s, 1H), 4.45-4.16 (m, 2H), 2.86 (s, 3H), 1.31 (t, 3H, J = 7.0 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 167.3, 135.8, 134.8, 131.2, 129.4, 129.3, 128.8, 127.3, 125.9, 122.8, 118.2, 67.3, 63.2, 40.1, 14.2; HRMS: calcd. for C₁₉H₂₁NO₄S₂⁺ Na: 414.0804; found: 414.0819.

Enamide (4w): 67 mg; Colorless liquid; yield: 75%; R_f = 0.53 in 20:80 ethyl acetate/hexane;



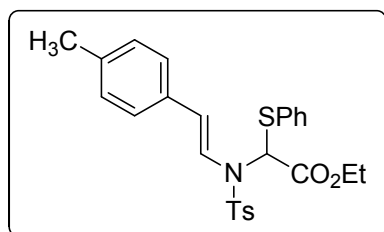
FTIR (neat): 3065, 2979, 2930, 2357, 1741, 1645, 1585, 1473, 1364, 1260, 1170, 1091, 1014, 937 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.75-7.67 (m, 2H), 7.57-7.51 (m, 3H), 7.45-7.41 (m, 2H), 7.38-7.33 (m, 3H), 7.32-7.26 (m, 4H), 7.23-7.19 (m, 1H), 6.95 (d, 1H, J = 14.7 Hz), 6.40-6.23 (m, 2H), 4.15-4.02 (m, 2H), 1.10 (t, 3H, J = 7.1 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 167.0, 139.1, 135.9, 134.7, 133.3, 131.6, 129.4, 129.2, 129.0, 128.8, 127.7, 127.3, 126.0, 122.9, 119.8, 67.4, 62.8, 14.0; HRMS: calcd. for C₂₄H₂₃NO₄S₂ + Na: 476.0961; found: 476.0966.

Enamide (4x): 68 mg; Colorless liquid; yield: 70%; R_f = 0.51 in 20:80 ethyl acetate/hexane;



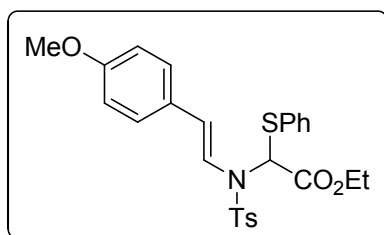
FTIR (neat): 3059, 2983, 2357, 1740, 1646, 1585, 1474, 1443, 1363, 1265, 1171, 1091, 937 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.75-7.67 (m, 2H), 7.57-7.51 (m, 3H), 7.45-7.41 (m, 2H), 7.38-7.33 (m, 3H), 7.32-7.26 (m, 4H), 7.23-7.19 (m, 1H), 6.95 (d, 1H, J = 14.7 Hz), 6.40-6.23 (m, 2H), 4.15-4.02 (m, 2H), 1.10 (t, 3H, J = 7.1 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 167.0, 139.1, 135.9, 134.7, 133.3, 131.6, 129.4, 129.2, 129.0, 128.8, 127.7, 127.3, 126.0, 122.9, 119.8, 67.4, 62.8, 14.0; HRMS: calcd. for C₂₄H₂₂ClNO₄S₂ + Na: 510.0571; found: 510.0575.

Enamide (4y): 71 mg; Colorless liquid; yield: 74%; R_f = 0.51 in 20:80 ethyl acetate/hexane;



FTIR (neat): 3057, 3028, 2974, 2928, 2865, 2357, 1740, 1645, 1599, 1483, 1354, 1260, 1166, 1091, 936 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.62-7.56 (m, 2H), 7.55-7.51 (m, 2H), 7.37-7.33 (m, 3H), 7.22-7.16 (m, 4H), 7.13-7.07 (m, 2H), 6.87 (d, 1H, J = 14.6 Hz), 6.33 (s, 1H), 6.28 (d, 1H, J = 14.6 Hz), 4.18-4.04 (m, 2H), 2.38 (s, 3H), 2.32 (s, 3H), 1.11 (t, 3H, J = 7.1 Hz); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 167.2, 144.2, 137.1, 136.1, 134.6, 133.0, 131.7, 129.6, 129.4, 129.3, 129.1, 127.8, 125.9, 122.1, 119.9, 67.3, 62.7, 21.6, 21.2, 14.0; HRMS: calcd. for C₂₆H₂₇NO₄S₂ + Na: 504.1274; found: 504.1284.

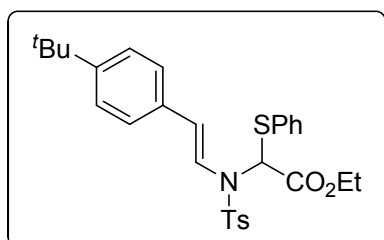
Enamide (4z): 78 mg; Colorless gummy solid; yield: 79%; $R_f = 0.48$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 3061, 2948, 2848, 2573, 2328, 1974, 1740, 1644, 1587, 1502, 1453, 1353, 1336, 1252, 1167, 1092, 937 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.61-7.56 (m, 2H), 7.56-7.48 (m, 2H), 7.39-7.33 (m, 3H), 7.24-7.18 (m, 4H), 6.88-6.80 (m, 2H), 6.79-6.72 (m, 1H), 6.45-

6.20 (m, 2H), 4.16-4.04 (m, 2H), 3.80 (s, 3H), 2.39 (s, 3H), 1.13 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.2, 159.1, 144.1, 136.1, 134.5, 131.7, 129.6, 129.3, 129.0, 128.8, 128.4, 127.8, 127.6, 127.3, 121.0 (9), 121.0(5), 114.2, 67.4, 62.7, 55.4, 21.7, 14.0; HRMS: calcd. for $\text{C}_{26}\text{H}_{27}\text{NO}_5\text{S}_2 + \text{Na}$: 520.1223; found: 520.1229.

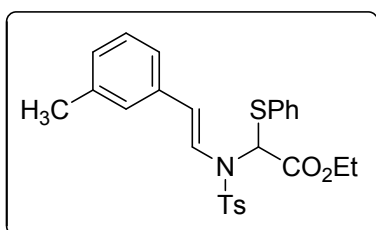
Enamide (4aa): 94 mg; Colorless gummy liquid; yield: 90%; $R_f = 0.51$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 3058, 2959, 2869, 2357, 1741, 1645, 1599, 1463, 1359, 1262, 1167, 1093, 935 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.64-7.56 (m, 2H), 7.56 (m, 2H), 7.37-7.30 (m, 5H), 7.25-7.17 (m, 4H), 6.90 (d, 1H, $J = 14.6$ Hz), 6.34 (s, 1H), 6.29 (d, 1H, $J = 14.6$ Hz), 4.17-4.03 (m,

2H), 2.38 (s, 3H), 1.31 (s, 9H), 1.11 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.1, 150.3, 144.2, 136.1, 134.6, 133.1, 131.6, 129.6, 129.3, 129.1, 127.8, 125.7, 125.6, 122.3, 119.4, 67.3, 62.7, 34.6, 31.3, 21.6, 14.0; HRMS: calcd. for $\text{C}_{29}\text{H}_{33}\text{NO}_4\text{S}_2 + \text{Na}$: 546.1743; found: 546.1758.

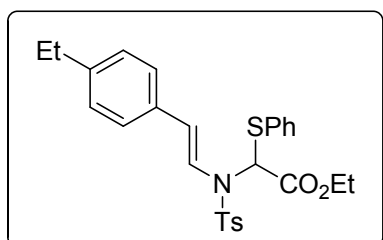
Enamide (4ab): 45 mg; Colorless gummy liquid; yield: 80%; $R_f = 0.52$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 3262, 3053, 2968, 2927, 2863, 2357, 1740, 1645, 1598, 1451, 1357, 1259, 1168, 1091, 934 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.63-7.56 (m, 2H), 7.56-7.51 (m, 2H), 7.39-7.32 (m, 3H), 7.24-7.15 (m, 3H), 7.13-7.05 (m, 2H), 7.05-7.00 (m, 1H), 6.94 (d, 1H, $J = 14.6$ Hz),

6.33 (s, 1H), 6.26 (d, 1H, $J = 14.6$ Hz), 4.18-4.03 (m, 2H), 2.38 (s, 3H), 2.33 (s, 3H), 1.11 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.1, 144.2, 138.3, 136.1, 135.8, 134.7, 131.6, 129.6, 129.3, 129.1, 128.6, 128.0, 127.7, 126.7, 123.1, 122.8, 119.2, 67.2, 62.8, 21.6, 21.5, 14.0; HRMS: calcd. for $\text{C}_{26}\text{H}_{27}\text{NO}_4\text{S}_2 + \text{Na}$: 504.1274; found: 504.1274.

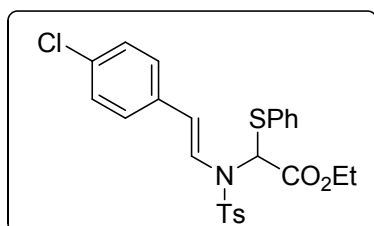
Enamide (4ac): 87 mg; Colorless gummy liquid; yield: 88%; $R_f = 0.53$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 3057, 2971, 2933, 2872, 2356, 1740, 1646, 1600, 1442, 1355, 1265, 1168, 1091, 934 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.61-7.56 (m, 2H), 7.56-7.50

(m, 2H), 7.38-7.32 (m, 3H), 7.23-7.18 (m, 4H), 7.15-7.11 (m, 2H), 6.88 (d, 1H, $J = 14.6$ Hz), 6.34 (s, 1H), 6.29 (d, 1H, $J = 14.6$ Hz), 4.16-4.03 (m, 2H), 7.66 (q, 2H, $J = 7.6$ Hz), 2.38 (s, 3H), 1.22 (t, 3H, $J = 7.6$ Hz), 1.11 (t, 3H, $J = 7.2$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.1, 144.1, 143.5, 136.1, 134.6, 133.3, 131.7, 129.6, 129.3, 129.1, 128.2, 127.8, 126.0, 122.2, 119.8, 67.3, 62.7, 28.6, 21.6, 15.7, 14.0; HRMS: calcd. for $\text{C}_{27}\text{H}_{29}\text{NO}_4\text{S}_2 + \text{Na}$: 518.1430; found: 518.1437.

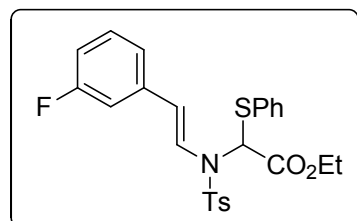
Enamide (4ad): 72 mg; Light green semi solid; yield: 71%; $R_f = 0.51$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 2925, 2857, 2357, 1892, 1740, 1645, 1609, 1447, 1351, 1257, 1166, 1092, 1020, 935 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): 7.64-7.59 (m, 2H), 7.58 (dd, 1H, $J = 7.5, 1.7$ Hz), 7.48 (dd, 1H, $J = 7.9, 1.3$ Hz), 7.34 (dd, 1H, $J = 7.6, 1.6$ Hz), 7.32-7.25 (m, 5H), 7.24-7.19 (m, 3H),

6.91 (d, 1H, $J = 14.7$ Hz), 6.54 (s, 1H), 6.30 (d, 1H, $J = 14.7$ Hz), 4.20-4.03 (m, 2H), 2.38 (s, 3H), 1.10 (s, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 166.8, 144.3, 139.0, 137.1, 136.0, 130.7, 130.2, 129.9, 129.6, 128.7, 127.8, 127.5, 127.2, 125.9, 122.8, 118.9, 65.4, 62.9, 21.6, 14.0; HRMS: calcd. for $\text{C}_{25}\text{H}_{24}\text{ClNO}_4\text{S}_2 + \text{NH}_4$: 519.1179; found: 519.1190.

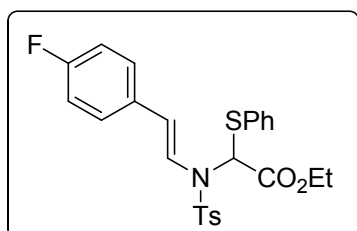
Enamide (4ae): 76 mg; Colorless gummy liquid; yield: 79%; $R_f = 0.51$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 3581, 3466, 3304, 3064, 2929, 2860, 2357, 1741, 1645, 1597, 1477, 1445, 1358, 1259, 1166, 1090, 931 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.65-7.56 (m, 2H), 7.56-7.50 (m, 2H), 7.40-7.33 (m, 3H), 7.26-7.20 (m, 3H), 7.06-7.02 (m, 1H), 7.01-6.94 (m, 2H), 6.92-6.85 (m, 1H),

6.33 (s, 1H), 6.23 (s, 1H, $J = 14.6$ Hz), 4.18-4.05 (m, 2H), 2.39 (s, 3H), 1.12 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.0, 163.2 (d, $J = 246.4$ Hz), 144.4, 138.4 (d, $J = 8.0$ Hz), 135.8, 134.7, 131.4, 130.2 (d, $J = 8.4$ Hz), 129.7, 129.4, 129.3, 127.7, 124.2, 121.8, 116.7, 113.8 (d, $J = 21.2$ Hz), 112.3 (d, $J = 22.0$ Hz), 67.1, 62.8, 21.7, 14.0; HRMS: calcd. for $\text{C}_{25}\text{H}_{24}\text{FNO}_4\text{S}_2 + \text{NH}_4$: 503.1469; found: 503.1482.

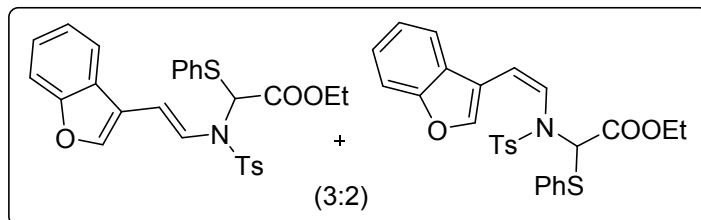
Enamide (4af): 68 mg; Light green gummy liquid; yield: 71%; $R_f = 0.50$ in 20:80 ethyl



acetate/hexane; FTIR (neat): 3057, 2971, 2933, 2872, 2356, 1740, 1646, 1600, 1442, 1355, 1265, 1168, 1091, 934 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 7.52-7.39 (m, 4H), 7.37-7.24 (m, 3H), 7.19-7.11 (m, 4H), 6.95-6.87 (m, 2H), 6.78 (d, 1H, $J = 14.7$ Hz), 6.25 (m, 1H), 6.20 (d, 1H, $J = 14.7$ Hz), 4.09-3.97

(m, 2H), 2.31 (s, 3H), 1.05 (t, 3H, $J = 7.1$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.1, 162.1 (d, $J = 246.3$ Hz), 144.3, 135.9, 134.6, 132.0 (d, $J = 3.5$ Hz), 131.5, 129.6, 129.3, 129.2, 127.7, 127.4 (d, $J = 7.8$ Hz), 122.8, 118.4, 115.6 (d, $J = 21.6$ Hz), 67.2, 62.8, 21.7, 14.0; HRMS: calcd. for $\text{C}_{25}\text{H}_{24}\text{FNO}_4\text{S}_2 + \text{Na}$: 508.1023; found: 508.1017.

Enamide(4g): 16 mg; Light green gummy liquid; yield: 27% (E/Z = 3/2); $R_f = 0.50$ in 20:80

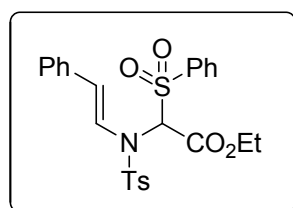


ethyl acetate/hexane; FTIR (neat): 3053, 2974, 2930, 2874, 2354, 1740, 1600, 1440, 1350, 1267, 934 cm^{-1} ; ^1H NMR (400

MHz, CDCl_3): δ 7.74 (d, 2H, $J = 7.8$ Hz), 7.57 (d, 2H, $J = 6.9$ Hz), 7.52-7.46 (m, 3H), 7.42-7.30 (m, 8H), 7.27-7.23 (m, 5H), 7.07 (d, 1H, $J = 14.1$ Hz), 6.83 (d, 1H, $J = 6.6$ Hz), 6.47 (d, 1H, $J = 14.4$ Hz), 6.37 (s, 1H), 6.04 (s, 1H), 5.99 (d, 1H, $J = 7.2$ Hz), 4.25-4.08 (m, 4H), 2.41 (s, 2.6H), 2.37 (s, 3H), 1.27 (t, 3H, $J = 7.0$ Hz), 1.16 (t, 3H, $J = 7.0$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 167.2, 167.1, 144.7, 144.3, 142.9, 136.1, 135.7, 134.5, 131.6, 130.1, 130.1, 129.7, 129.4, 129.2, 127.8, 127.2, 125.7, 125.0, 124.8, 124.1, 123.5, 123.5, 123.2, 122.6, 120.7, 111.9, 111.9, 111.8, 111.3, 100.7, 67.2, 62.7, 62.6, 56.7, 21.7, 14.2, 14.1; HRMS: calcd. for $\text{C}_{27}\text{H}_{25}\text{NO}_5\text{S}_2 + \text{Na}$: 530.1066; found: 530.1071.

10. Synthesis of sulfone 5

To a 25 mL round bottom flask equipped with stir bar, enamide **3a** (0.21 mmol, 1 equiv) was

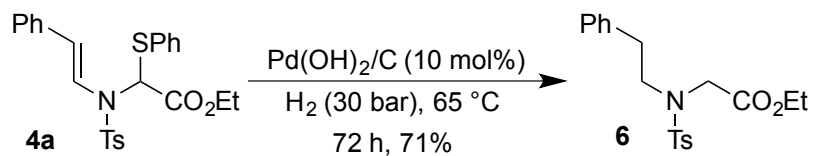


dissolved with dry CH_2Cl_2 (10 mL), transferred to a magnetic stir with ice-bath. Next, at 0 $^\circ\text{C}$ temperature m-CBPA (0.64 mmol, 3 equiv) was added *via* portion-wise and allowed to stir for 2h at room temperature. After completion of the reaction, by TLC, the reaction

was neutralized with NaHCO_3 solution and workup with CH_2Cl_2 , the organic layer was evaporated under reduced pressure. The crude product that obtained was purified by column chromatography using hexane/ethyl acetate mixture as eluent to afford sulfone derivatives **5** in high yield and purity. Green semi solid; 81 mg; yield: 78%; $R_f = 0.41$ in 20:80 ethyl acetate/hexane; FTIR (neat): 3288, 3174, 3064, 2934, 2747, 2661, 2594, 2550, 2380, 1956, 1912, 1742, 1650, 1594, 1448, 1334, 1274, 1164, 1086, 1022, 939, 809 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ 8.03-7.99 (m, 2H), 7.74-7.66 (m, 3H), 7.64-7.58 (m, 2H), 7.31-7.22 (m, 7H), 6.74 (d, 1H, $J = 14.5$ Hz), 6.62 (d, 1H, $J = 14.5$ Hz), 6.30 (s, 1H), 4.23 (q, 2H, $J = 7.2$ Hz), 2.39 (s, 3H), 1.21 (t, 3H, $J = 7.2$ Hz); $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 162.4, 145.0,

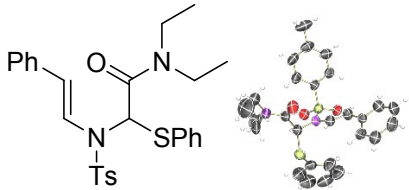
137.4, 135.4, 134.9, 134.8, 133.8, 129.8, 129.7, 129.2, 128.7, 128.2, 127.7, 126.3, 124.0, 122.6, 63.4, 21.7, 14.0; HRMS: calcd. for $C_{25}H_{25}NO_6S_2 + Na$: 522.1016; found: 522.1026.

11. Hydrogenation of enamide **4a**:



In a 250 mL autoclave Enamine **4a** was taken in 50 mg scale (1.0 equiv.) and 10% Pd-hydroxide on carbon (10 mol%) in MeOH was taken, followed by autoclave was closed tightly. Slowly hydrogen was purged three times. Finally 30 bar pressure was set inside the autoclave. The reaction was monitored for 72 h at 65 °C. After 72 h, the reaction mixture was filtered with celite three times using DCM solvent. The filtered solution was concentrated in high-vacuum pump, which was further purified by column chromatography with silica gel using ethylacetate/hexane as (5:95 ratio) an eluent gave the product **6** with 71% yield. FTIR (neat): 2992, 2931, 2887, 2350, 1741, 1633, 1432, 1351, 1260, 930 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3 , 24 °C): δ 7.73 (d, J = 8.0 Hz, 1H), 7.30-7.28 (m, 4H), 7.24 (d, J = 7.3 Hz, 1H), 7.17 (d, J = 7.3 Hz, 1H), 4.11-4.07 (m, 2H), 4.00 (s, 2H), 4.32 (s, 2H), 3.49 (d, J = 9.1 Hz, 2H) 2.90 (t, J = 9.1 Hz, 2H), 2.44 (s, 3H), 1.20 (t, J = 7.7 Hz, 3H).; $^{13}\text{C}\{^1\text{H}\}$ NMR (125 MHz, CDCl_3 , 24 °C): δ 169.1, 143.5, 138.4, 136.9, 129.7, 128.9, 128.7, 127.5, 126.7, 61.4, 50.2, 48.8, 35.3, 21.6, 14.1. HRMS: calcd. for $C_{19}H_{23}NO_4 + Na$: 384.1240; found: 384.1242.

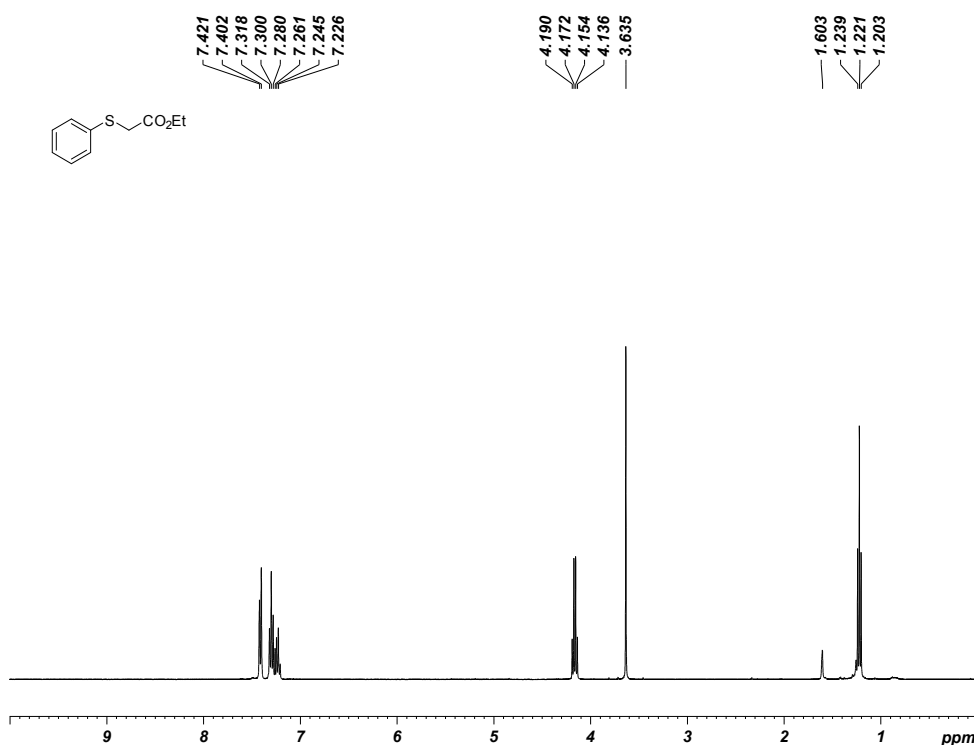
12. Crystallographic data and structure refinements summary for compound 4e

DATA	4e
Molecular Structure (ORTEP Structure)	
Formula	C ₂₇ H ₃₀ N ₂ O ₃ S ₂
Formula weight	494.65
Color	white
Temperature/K	293(2)
Radiation	Mo K α
Wavelength/Å	0.71073
Crystal system	Monoclinic
Space group	P2 ₁ /c
<i>a</i> (Å)	13.5570(9)
<i>b</i> (Å)	15.9431(9)
<i>c</i> (Å)	12.6454(8)
α (°)	90
β (°)	101.401(2)
γ (°)	90
Volume (Å ³)	2679.3(3)
<i>Z</i>	4
Density (g/ml)	1.226
μ (1/mm)	0.0077
<i>F</i> (000)	1048
θ (min, max)	2.217, 24.999
No. of unique reflns	4735
No. of parameters	500
<i>R</i> _{obs} , <i>wR</i> _{2_obs}	0.0605, 0.1537
$\Delta\rho_{\min}$, $\Delta\rho_{\max}$ (eÅ ⁻³)	-0.220, 0.403
GooF	1.066

13. NMR spectra of isolated compounds

Thioester 4a

^1H NMR (400 MHz, CDCl_3 , 24 °C)



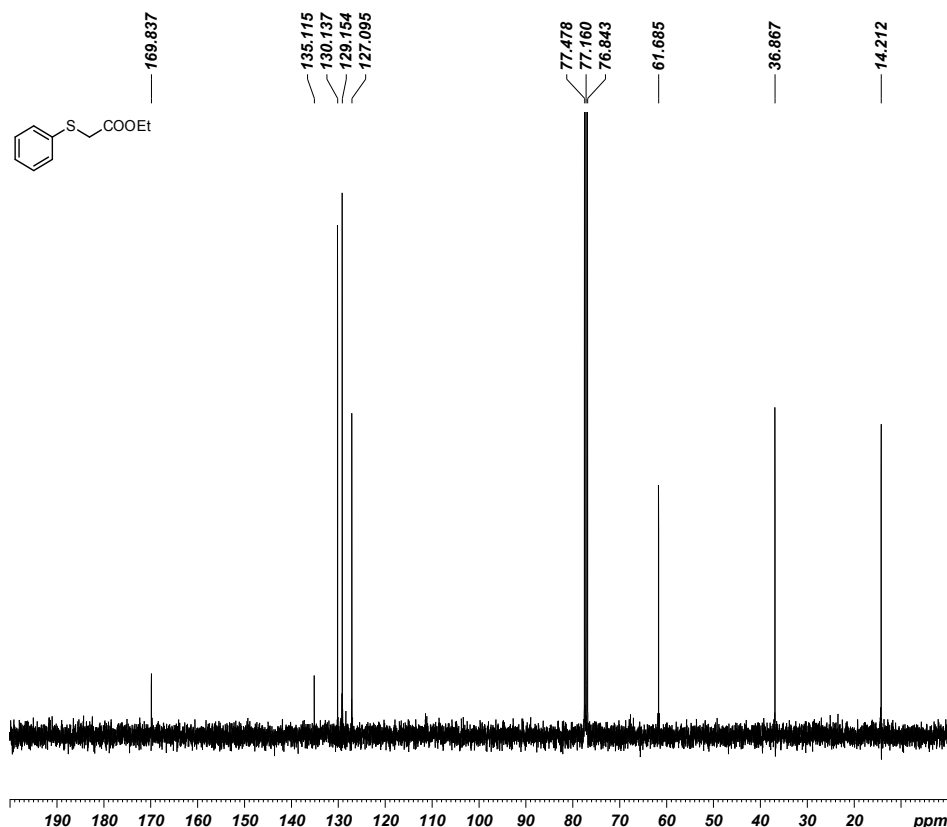
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NAME acr
EXPNO 128
PROCNO 1

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PULPROG zg30
TD 65536
SOLVENT CDCl_3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 153.13
DW 62.400 usec
DE 6.50 usec
TE 295.9 K
D1 0.50000000 sec
TD0 1

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NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
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WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Current Data Parameters
NAME spa41017
EXPNO 100
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171006
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PULPROG zgpg30
TD 16540
SOLVENT CDCl_3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 295.1 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

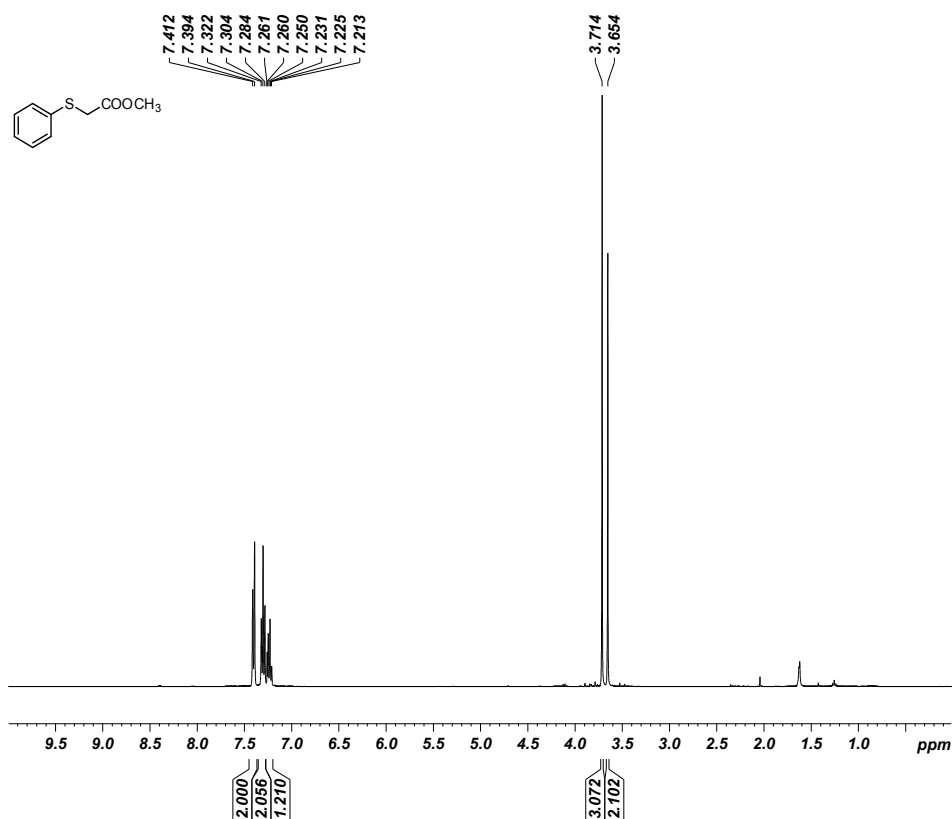
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SFO1 100.6228289 MHz
NUC1 ^{13}C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 ^1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127690 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

Thioester 1b

^1H NMR (400 MHz, CDCl_3 , 24 °C)



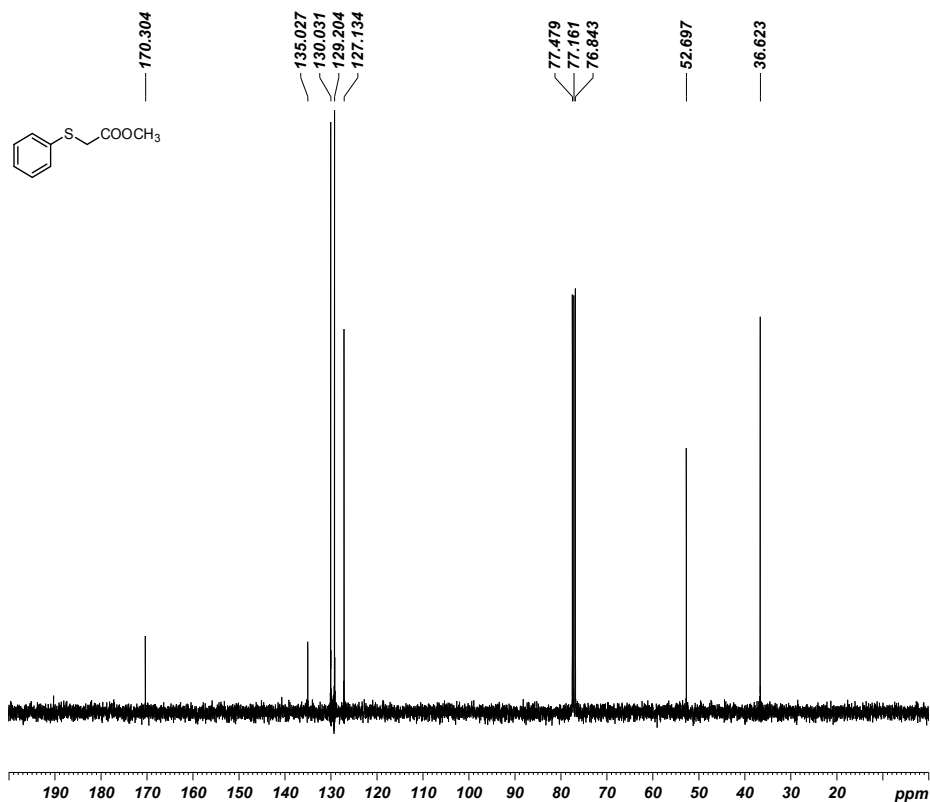
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NAME spa40717
EXPNO 29
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170701
Time 5.26
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PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl_3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 153.13
DW 62.400 usec
DE 6.50 usec
TE 294.5 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
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NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300093 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Current Data Parameters
NAME spa40717
EXPNO 30
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170701
Time 5.33
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 16540
SOLVENT CDCl_3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 294.9 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

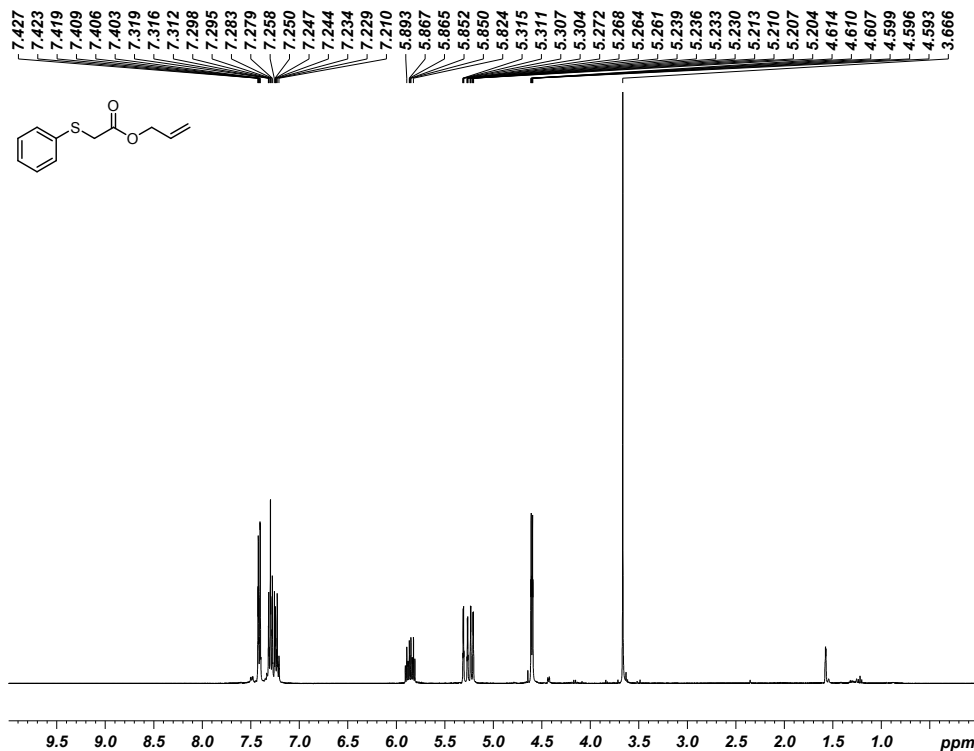
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NUC1 ^{13}C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
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NUC2 ^1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127690 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0

Thioester 1c

^1H NMR (400 MHz, CDCl_3 , 24 °C)



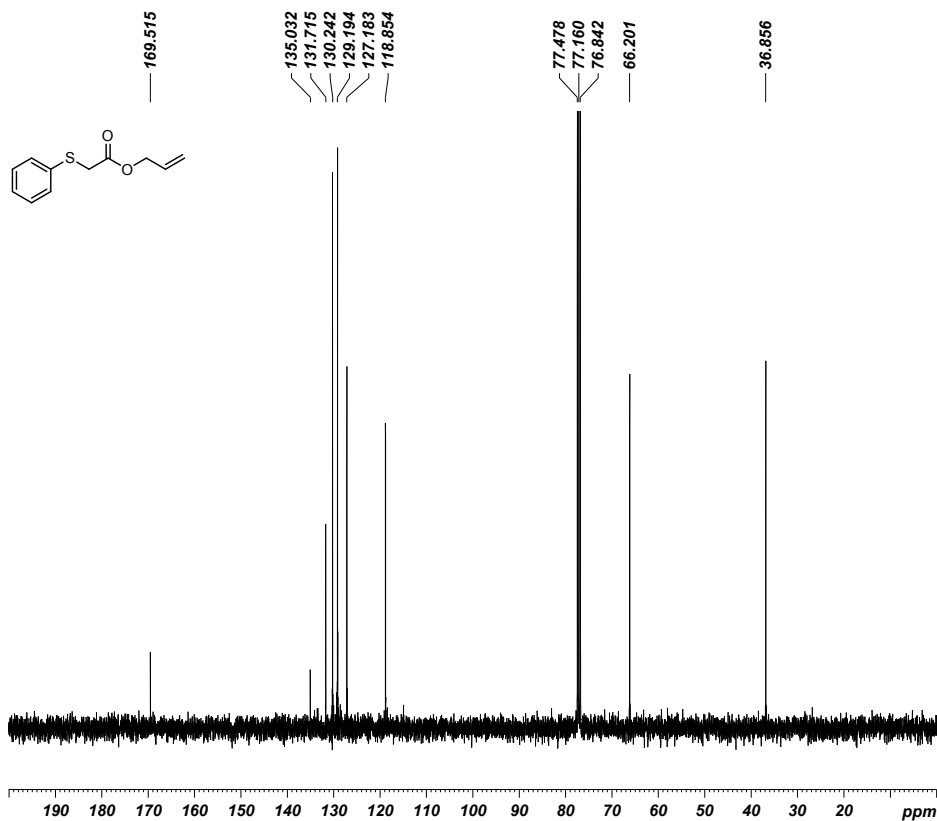
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NAME Sm spectra file
EXPNO 506
PROCNO 1

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PULPROG zg30
TD 65536
SOLVENT CDCl_3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 169.77
DW 62.400 usec
DE 6.50 usec
TE 298.6 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300106 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Current Data Parameters
NAME Sm spectra file
EXPNO 507
PROCNO 1

F2 - Acquisition Parameters
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Time 0.01
INSTRUM spect
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PULPROG zgpg30
TD 16540
SOLVENT CDCl_3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 299.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

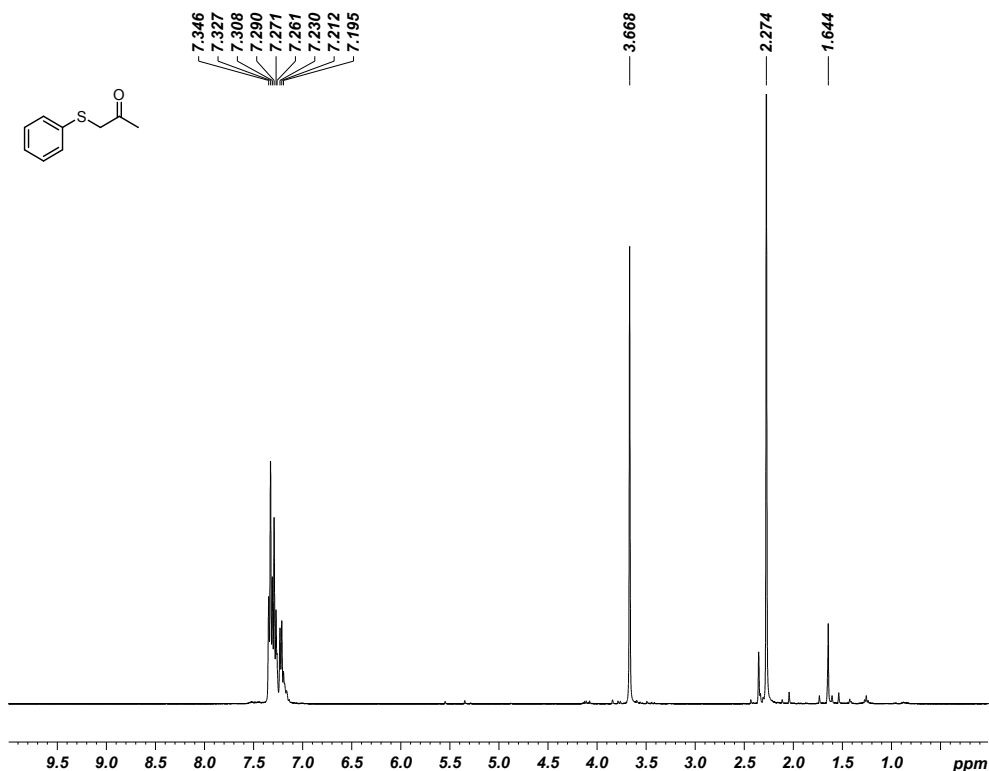
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P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
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NUC2 ^1H
PCPD2 waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127560 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Thioketone 1d

^1H NMR (400 MHz, CDCl_3 , 24 °C)



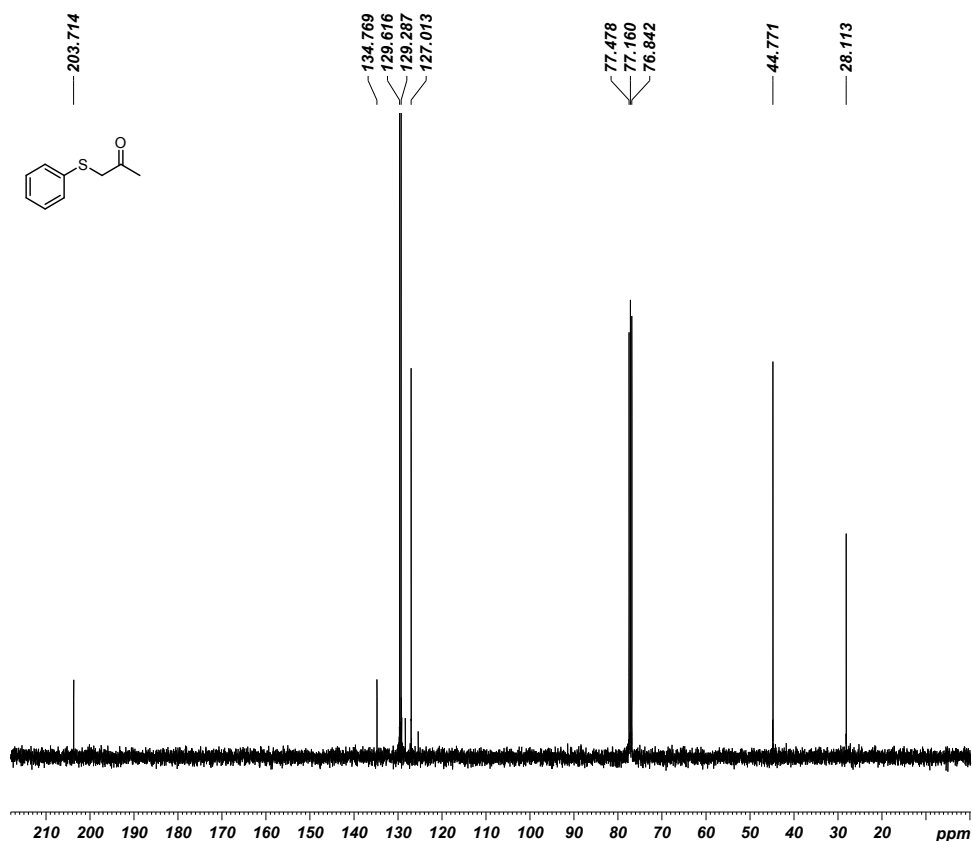
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NAME spa40717
EXPNO 23
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170701
Time 3.50
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl_3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 124.58
DW 62.400 usec
DE 6.50 usec
TE 294.3 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Current Data Parameters
NAME spa40717
EXPNO 24
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170701
Time 3.57
INSTRUM spect
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PULPROG zgpg30
TD 16540
SOLVENT CDCl_3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 294.9 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

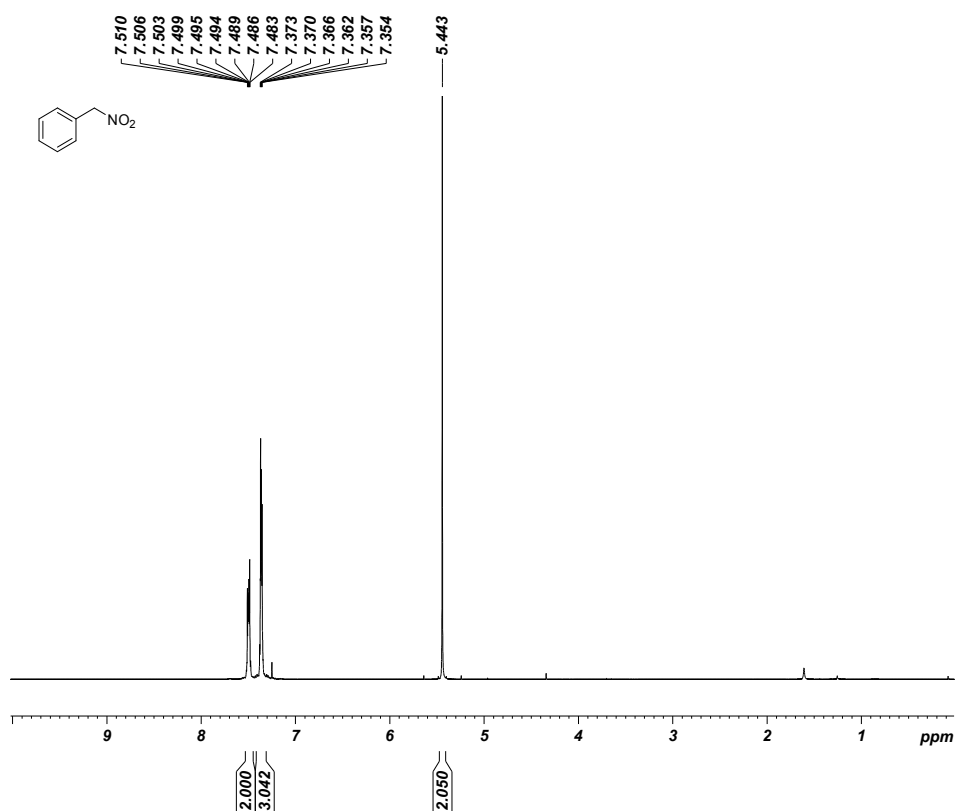
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SFO1 100.6228289 MHz
NUC1 ^{13}C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 ^1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127690 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

(Nitromethyl)benzene 1f

^1H NMR (400 MHz, CDCl_3 , 24 °C)



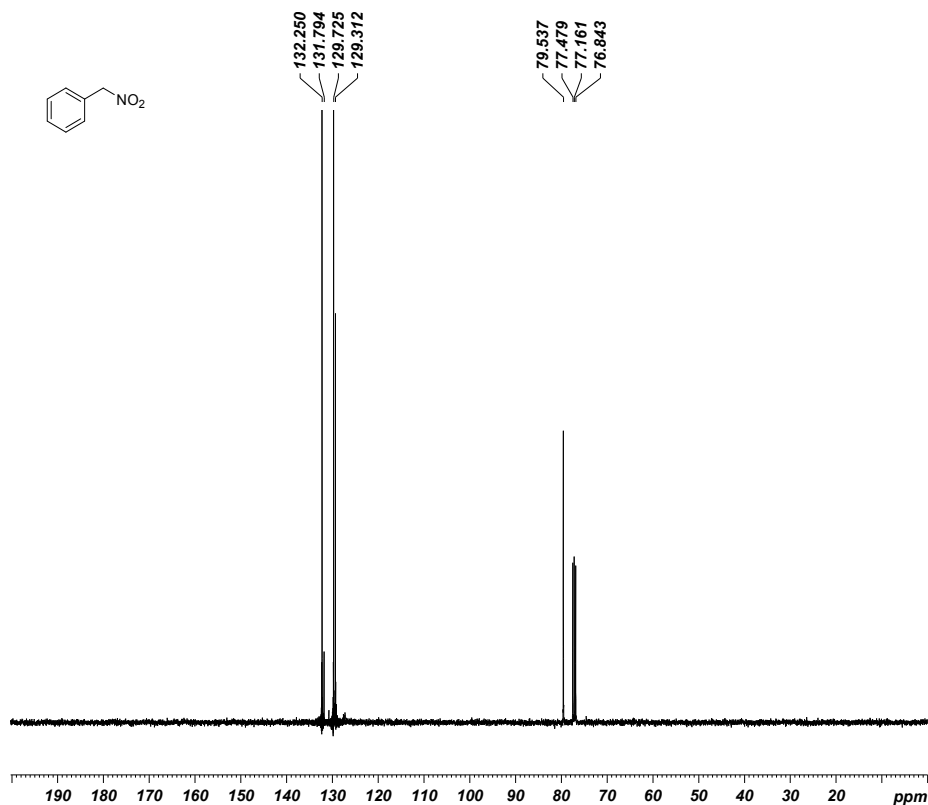
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EXPNO 17
PROCNO 1

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PULPROG zg30
TD 65536
SOLVENT CDCl_3
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DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 88.51
DW 62.400 usec
DE 6.50 usec
TE 294.4 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300136 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Current Data Parameters
NAME Rar\$DRa0.918
EXPNO 18
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170701
Time 3.27
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 16540
SOLVENT CDCl_3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 294.9 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

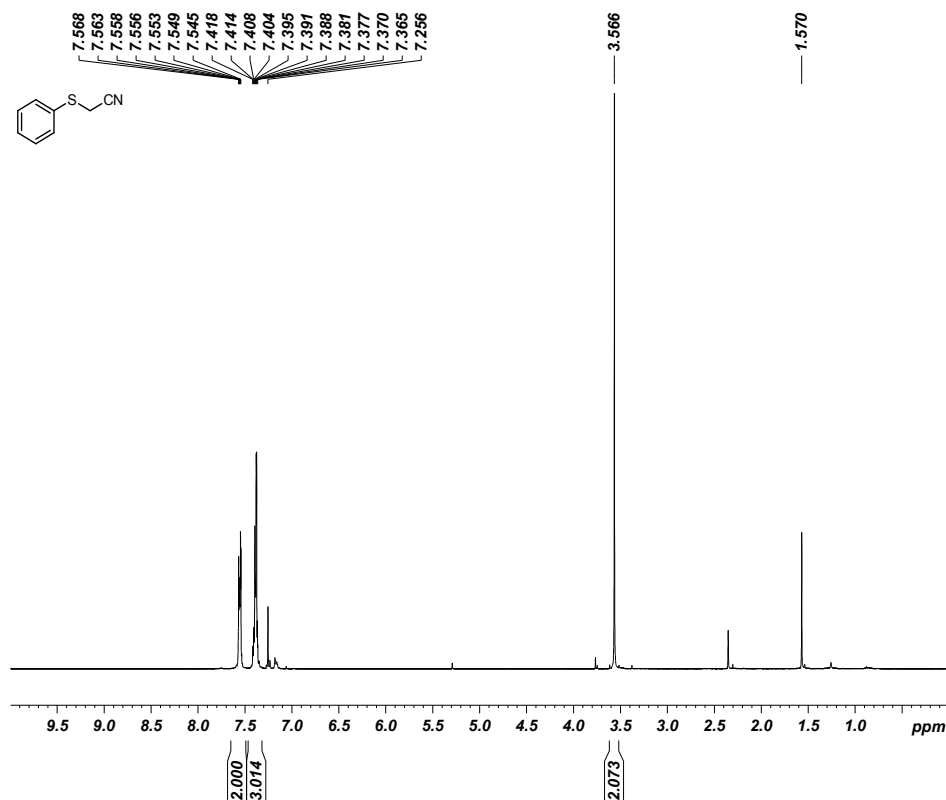
===== CHANNEL f1 =====
SFO1 100.6228289 MHz
NUC1 ^{13}C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 ^1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127640 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0

2-(Phenylthio)acetonitrile **1g**

^1H NMR (400 MHz, CDCl_3 , 24 °C)



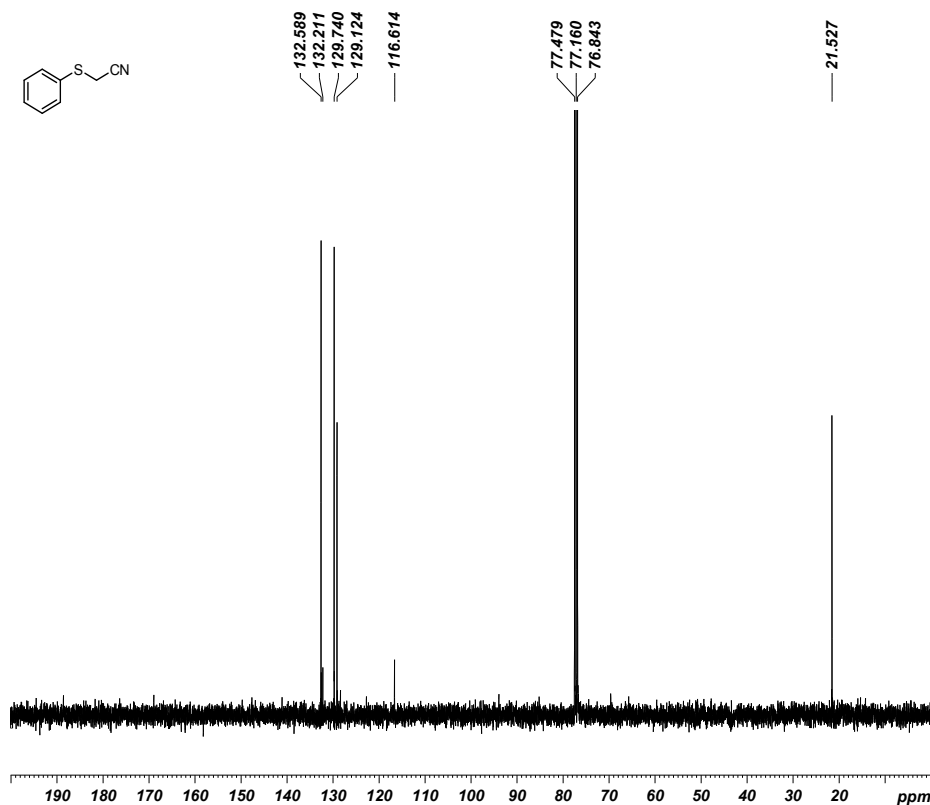
Current Data Parameters
NAME spa41217
EXPNO 71
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171202
Time 22.51
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl_3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 200.34
DW 62.400 usec
DE 6.50 usec
TE 298.5 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300111 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Current Data Parameters
NAME spa41217
EXPNO 72
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171202
Time 22.57
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 16540
SOLVENT CDCl_3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 298.9 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

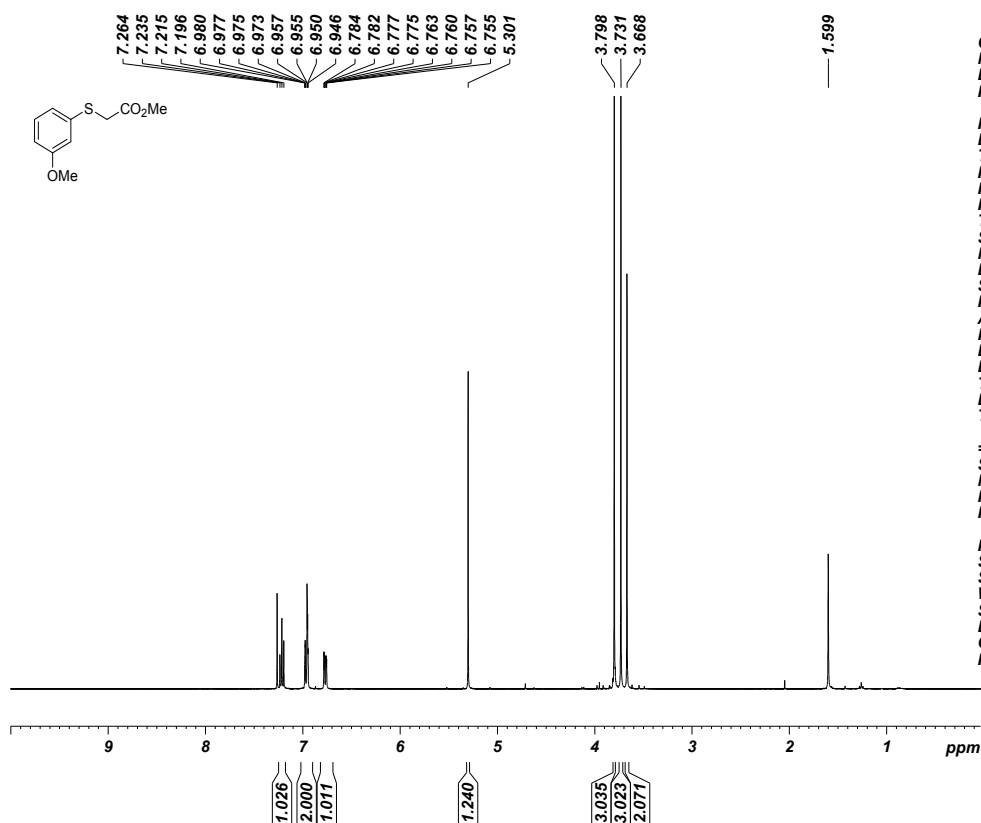
===== CHANNEL f1 =====
SFO1 100.6228289 MHz
NUC1 ^{13}C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 ^1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127690 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0

Thioester 1h

^1H NMR (400 MHz, CDCl_3 , 24 °C)



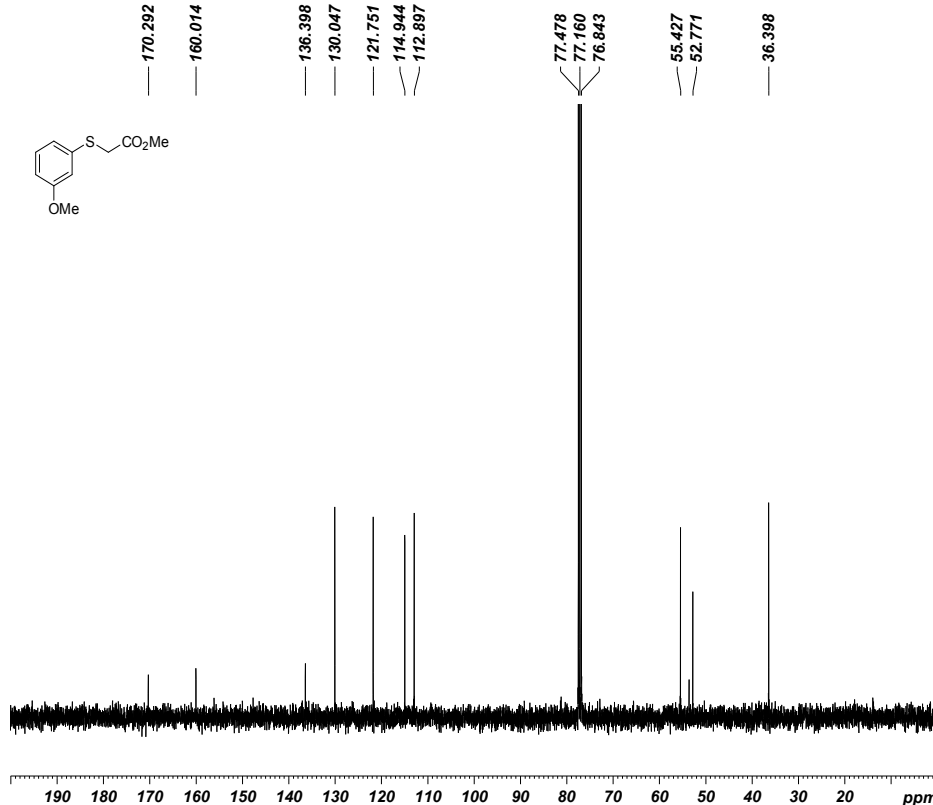
Current Data Parameters
NAME Rar\$DRa0.695
EXPNO 32
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170701
Time 5.38
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl_3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 200.34
DW 62.400 usec
DE 6.50 usec
TE 294.5 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300082 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Current Data Parameters
NAME Rar\$DRa0.116
EXPNO 33
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170701
Time 5.45
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 16540
SOLVENT CDCl_3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 294.9 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

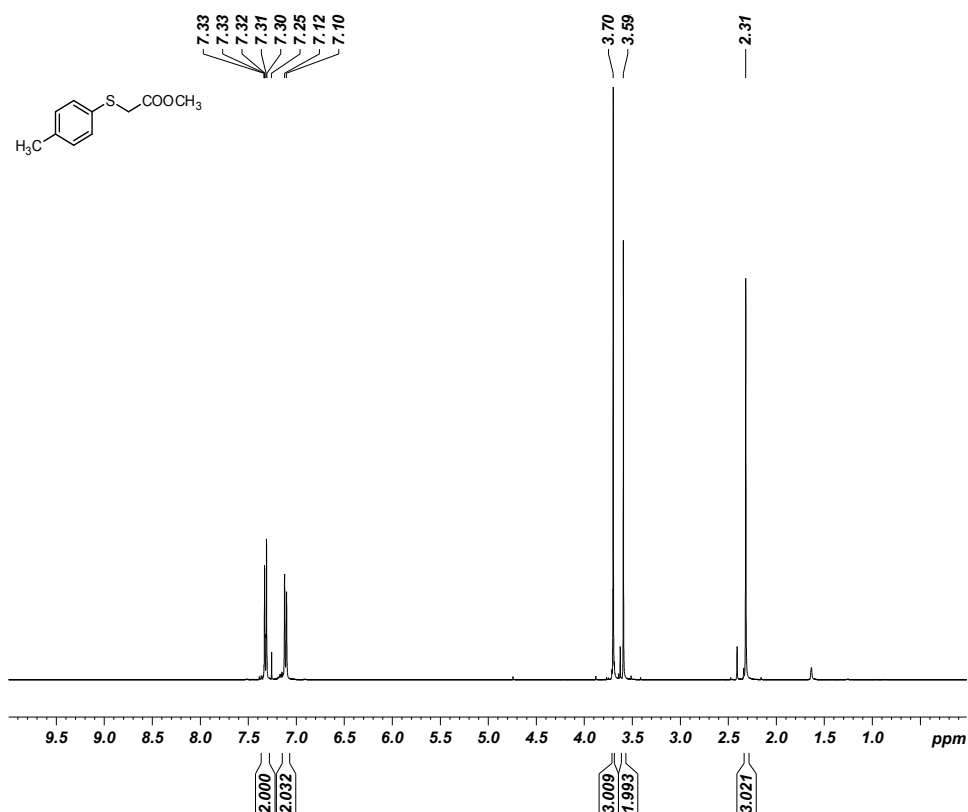
===== CHANNEL f1 =====
SFO1 100.6228289 MHz
NUC1 ^{13}C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 ^1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127563 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB n

Thioester 1i

¹H NMR (400 MHz, CDCl₃, 24 °C)



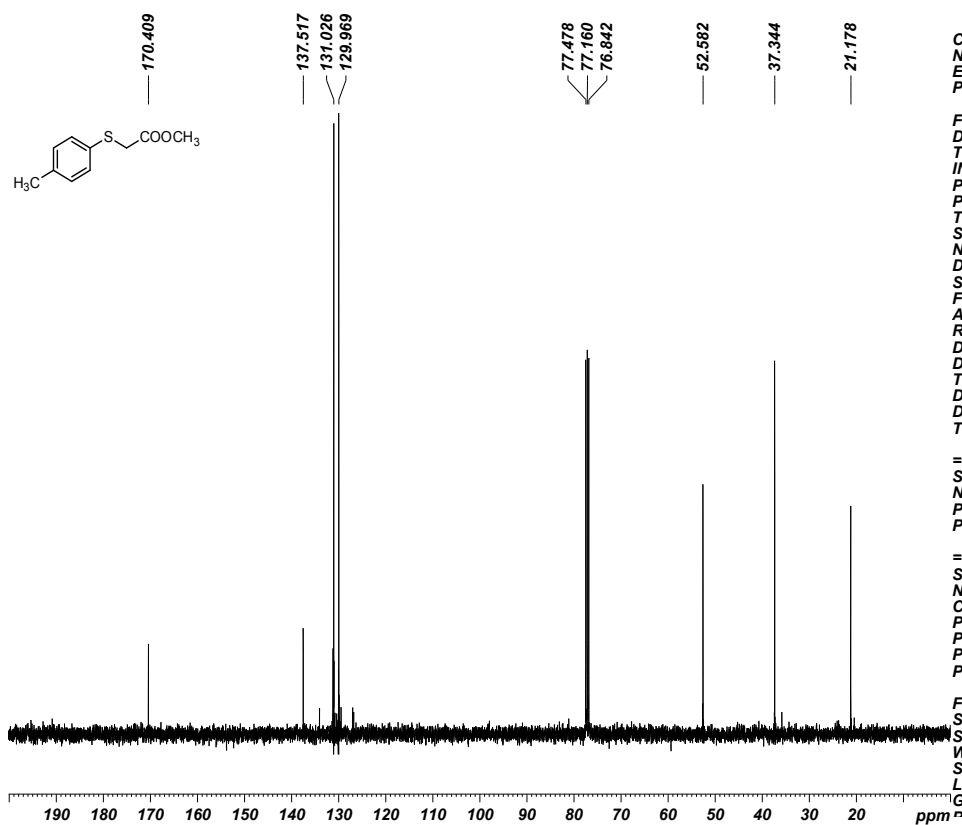
Current Data Parameters
NAME spa40617
EXPNO 398
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170613
Time 1.17
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 108.26
DW 62.400 usec
DE 6.50 usec
TE 296.2 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300105 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Current Data Parameters
NAME spa40617
EXPNO 399
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170613
Time 1.21
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 16540
SOLVENT CDCl3
NS 162
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 296.6 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

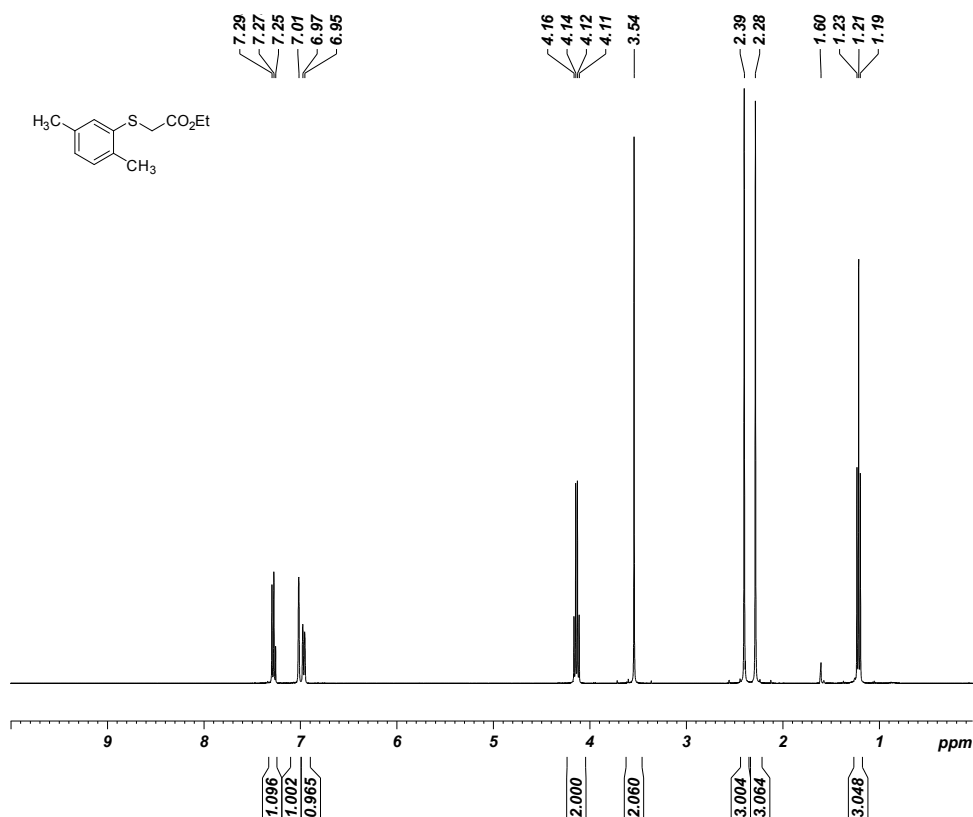
===== CHANNEL f1 =====
SFO1 100.6228289 MHz
NUC1 13C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127690 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0

Thioester 1j

¹H NMR (400 MHz, CDCl₃, 24 °C)



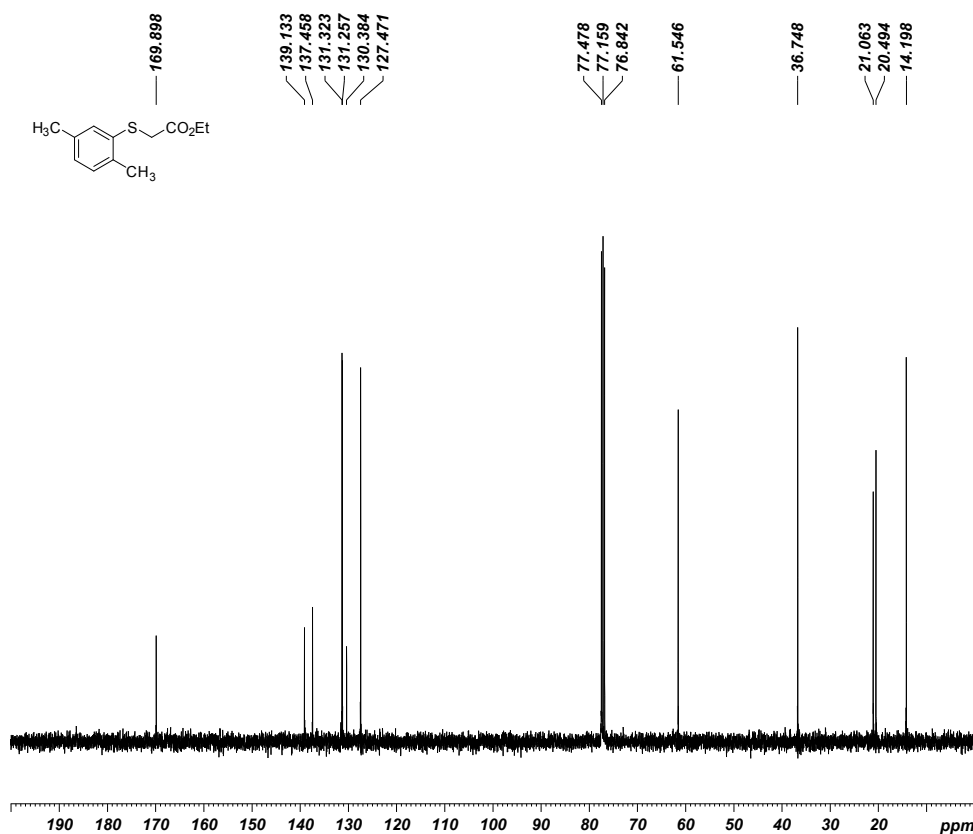
Current Data Parameters
NAME acr
EXPNO 100
PROCNO 1

F2 - Acquisition Parameters
Date_ 20191005
Time 4.03
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 108.26
DW 62.400 usec
DE 6.50 usec
TE 296.1 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300115 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Current Data Parameters
NAME acr
EXPNO 101
PROCNO 1

F2 - Acquisition Parameters
Date_ 20191005
Time 4.10
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 16540
SOLVENT CDCl3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 296.4 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

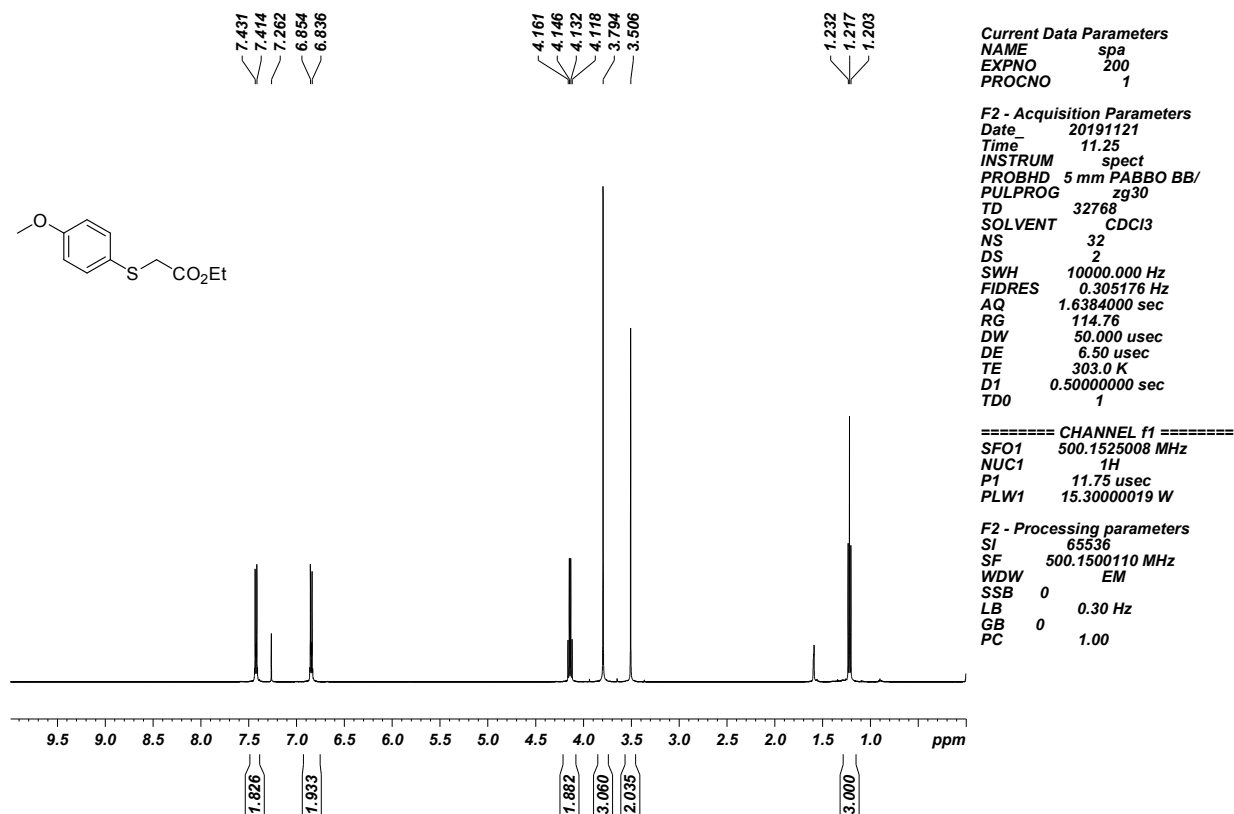
===== CHANNEL f1 =====
SFO1 100.6228289 MHz
NUC1 13C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

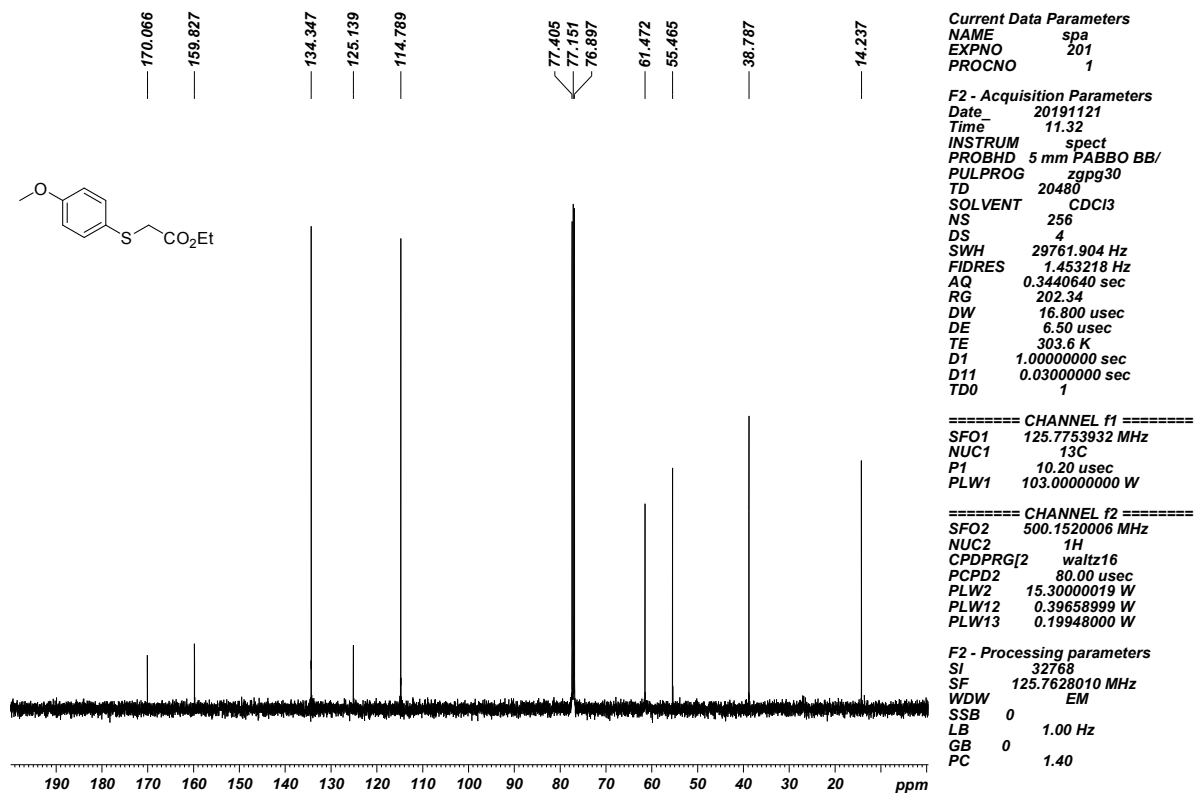
F2 - Processing parameters
SI 32768
SF 100.6127582 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Thioester 1k

¹H NMR (500 MHz, CDCl₃, 24 °C)

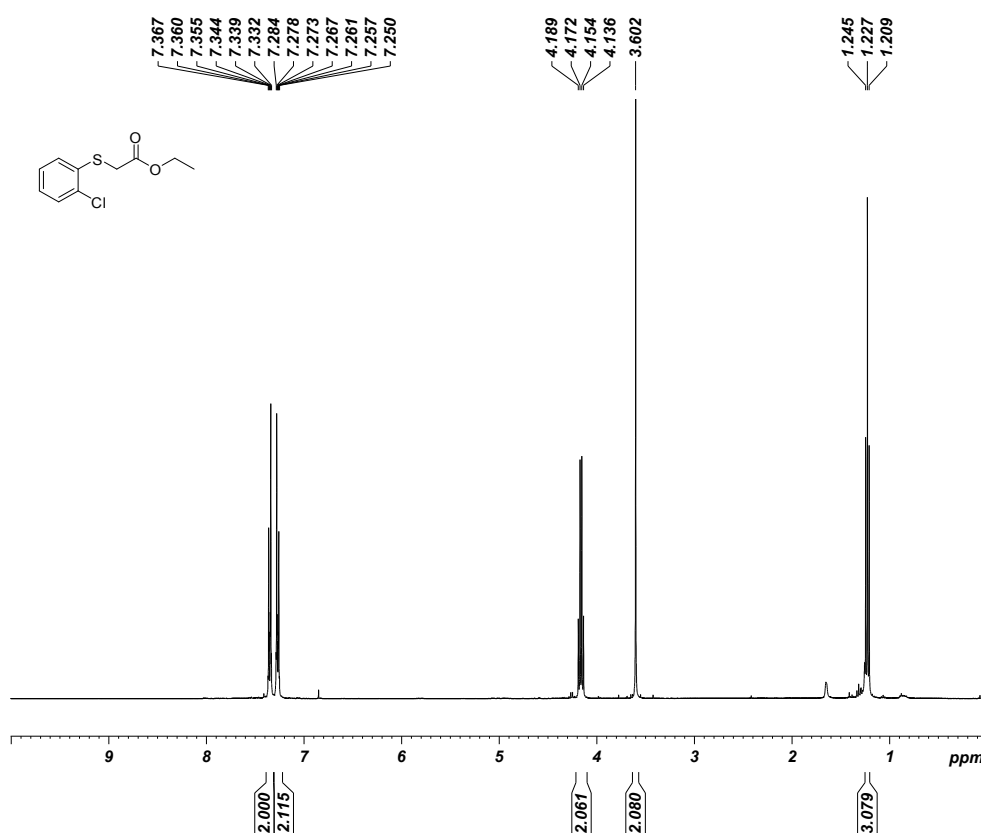


¹³C{¹H} NMR (125 MHz, CDCl₃, 24 °C)



Thioester 11

^1H NMR (400 MHz, CDCl_3 , 24 °C)



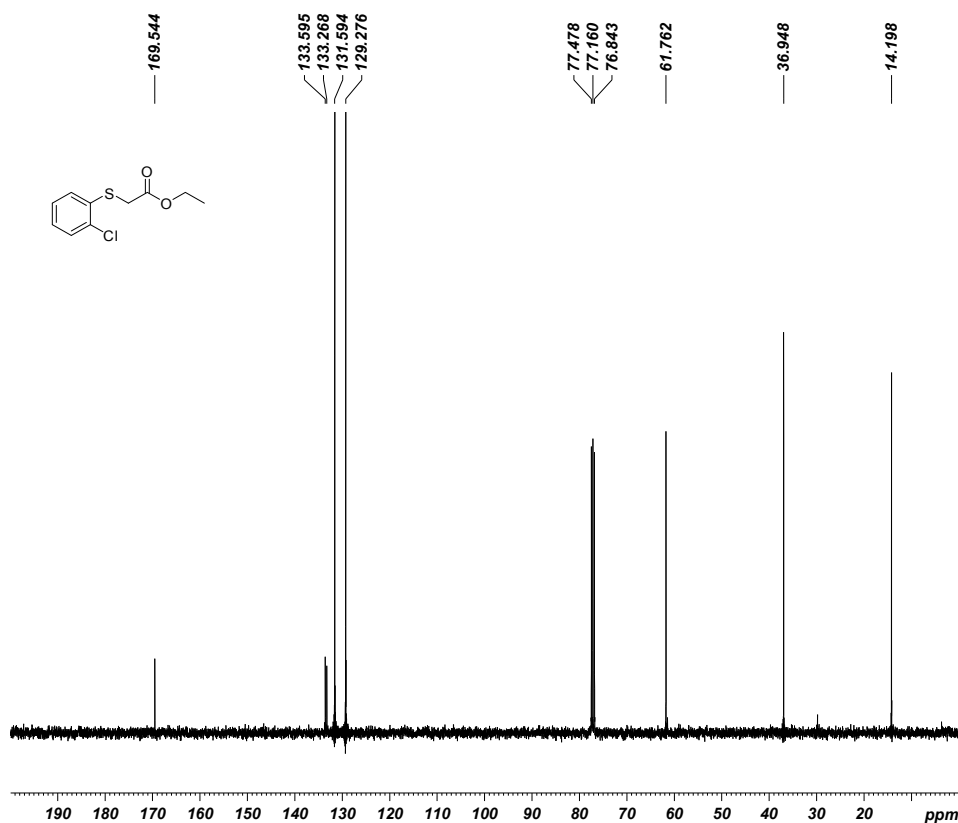
Current Data Parameters
NAME acr
EXPNO 459
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170817
Time 13.18
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl_3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 108.26
DW 62.400 usec
DE 6.50 usec
TE 298.0 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300068 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Current Data Parameters
NAME acr
EXPNO 460
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170817
Time 13.24
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 16540
SOLVENT CDCl_3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 298.4 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

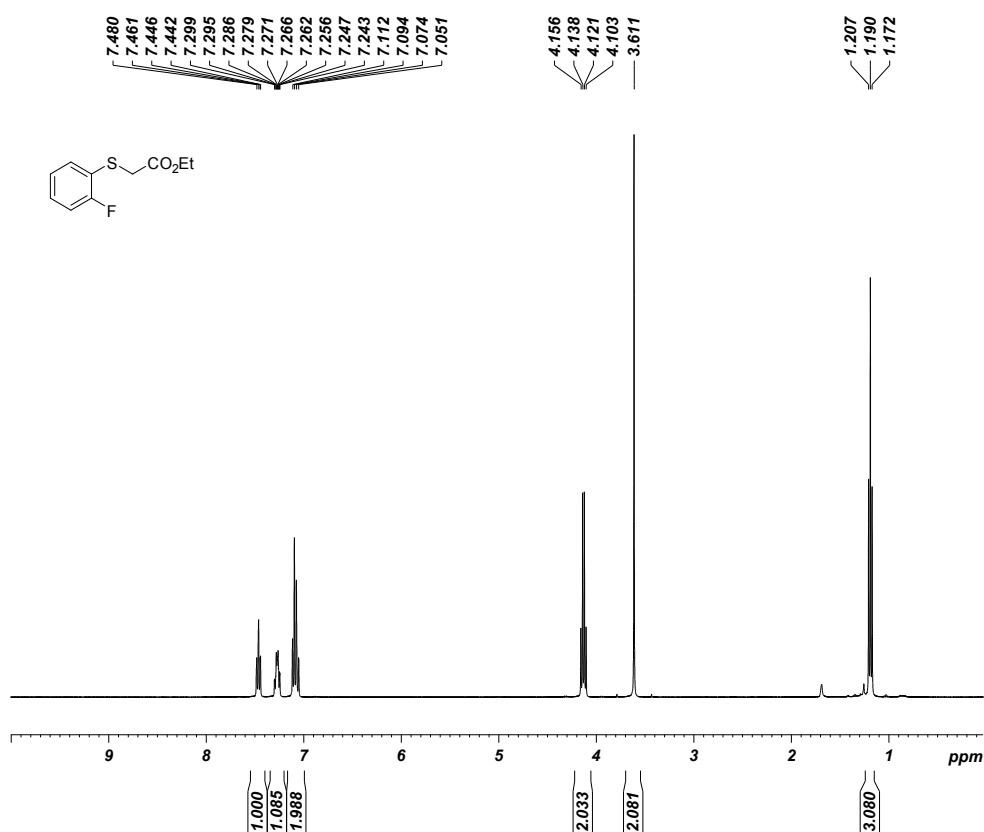
===== CHANNEL f1 =====
SFO1 100.6228289 MHz
NUC1 ^{13}C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 ^1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

F2 - Processing parameters
SI 32768
SF 100.6127581 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Thioester 1m

^1H NMR (400 MHz, CDCl_3 , 24 °C)



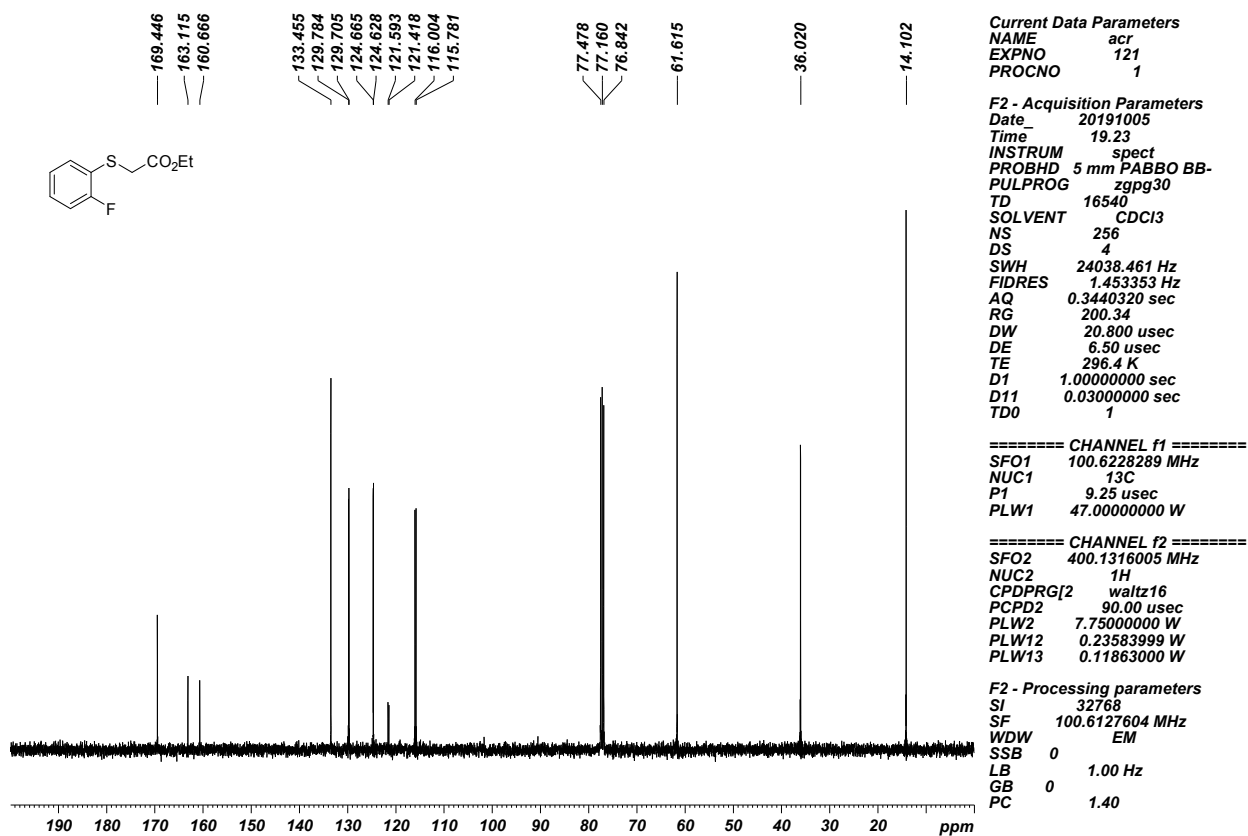
Current Data Parameters
NAME acr
EXPNO 120
PROCNO 1

F2 - Acquisition Parameters
Date 20191005
Time 19.17
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl_3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 88.51
DW 62.400 usec
DE 6.50 usec
TE 296.0 K
D1 0.5000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 ^1H
P1 15.70 usec
PLW1 7.75000000 W

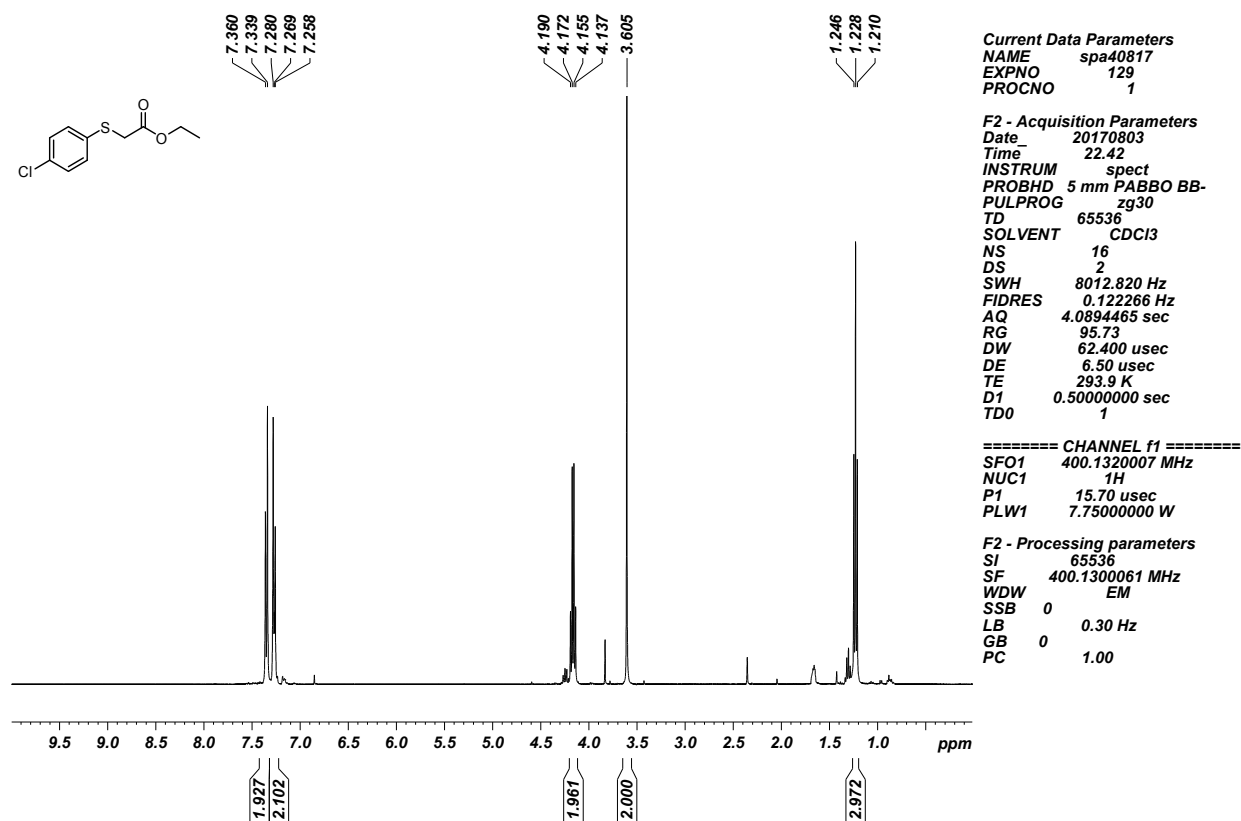
F2 - Processing parameters
SI 65536
SF 400.1300056 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

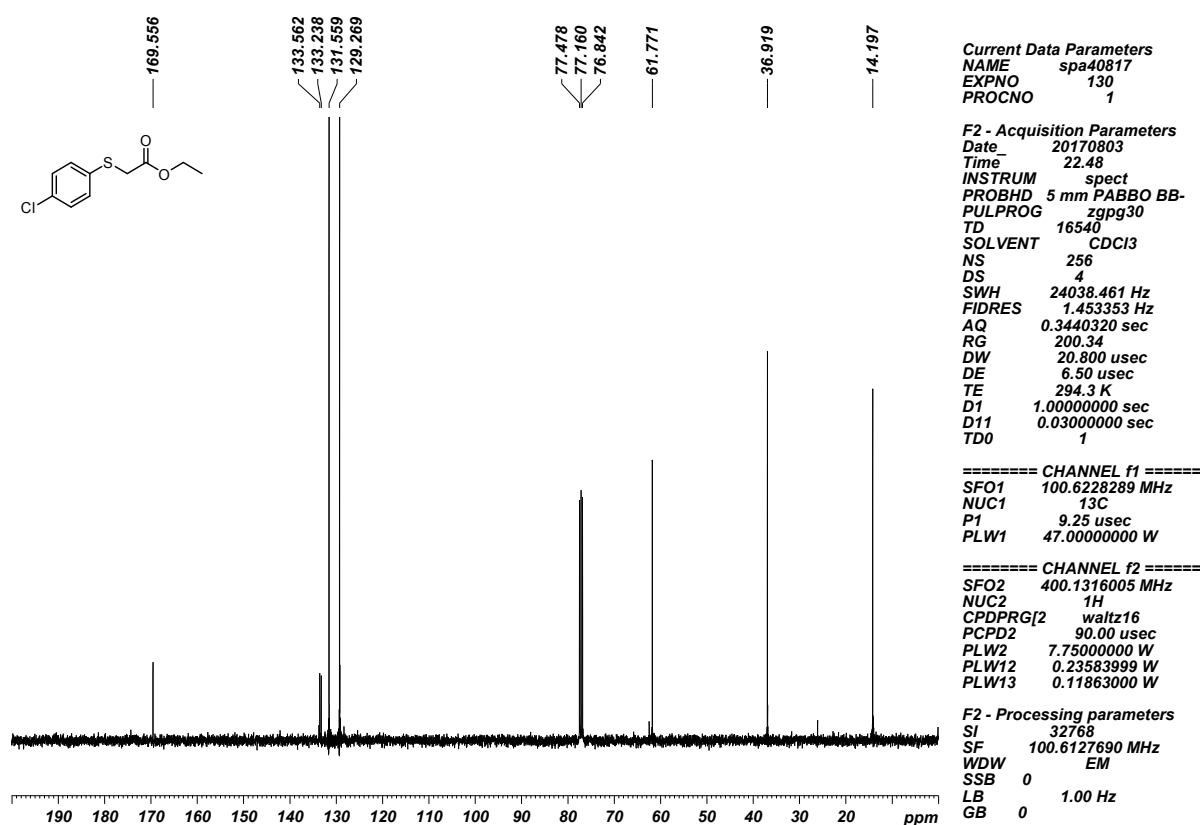


Thioester 1n

^1H NMR (400 MHz, CDCl_3 , 24 $^\circ\text{C}$)

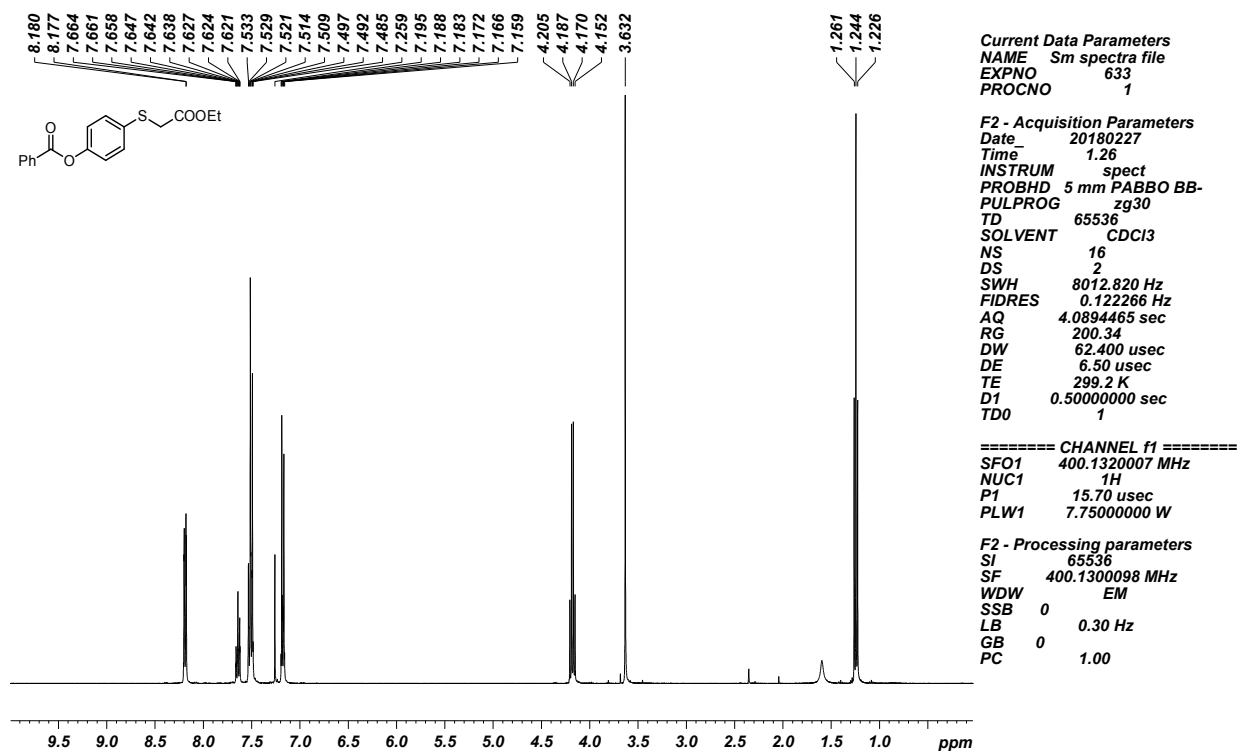


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

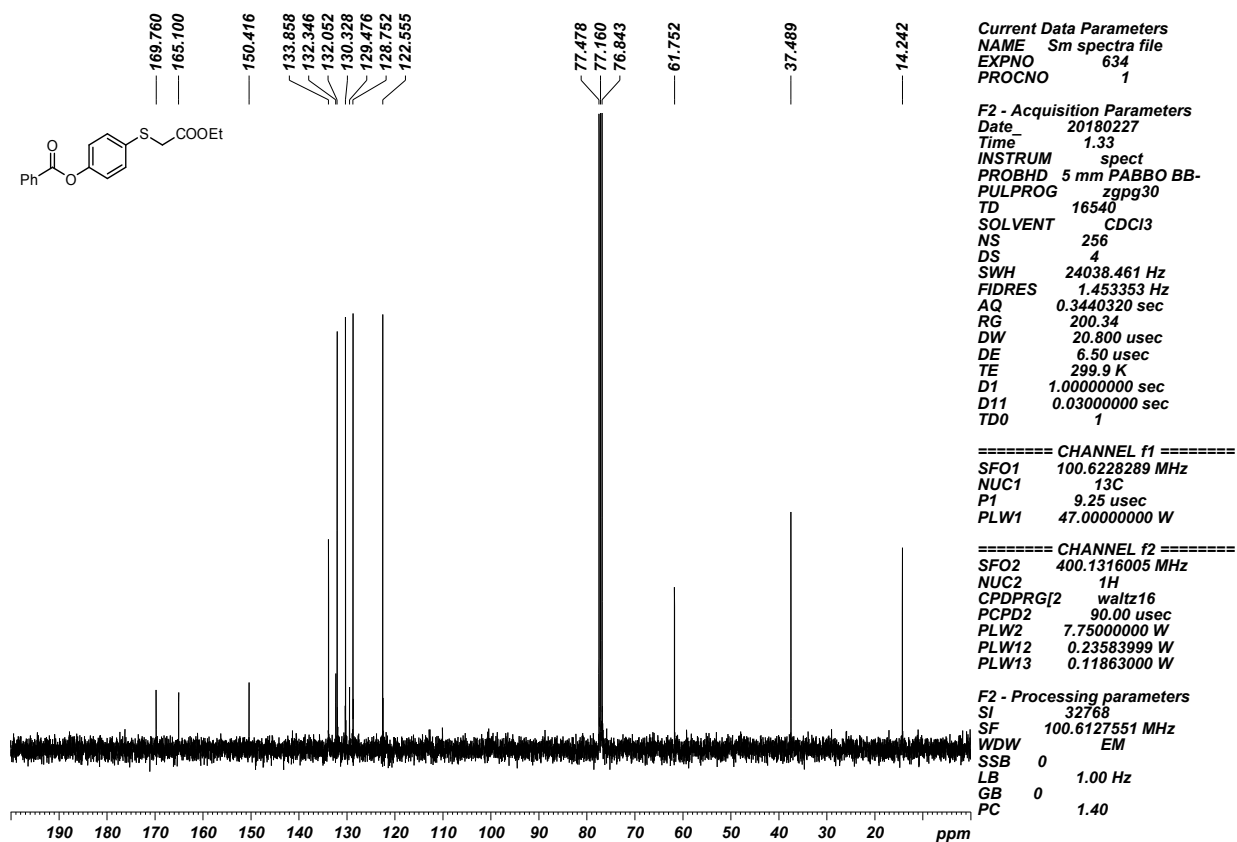


Thioester 1o

^1H NMR (400 MHz, CDCl_3 , 24 °C)

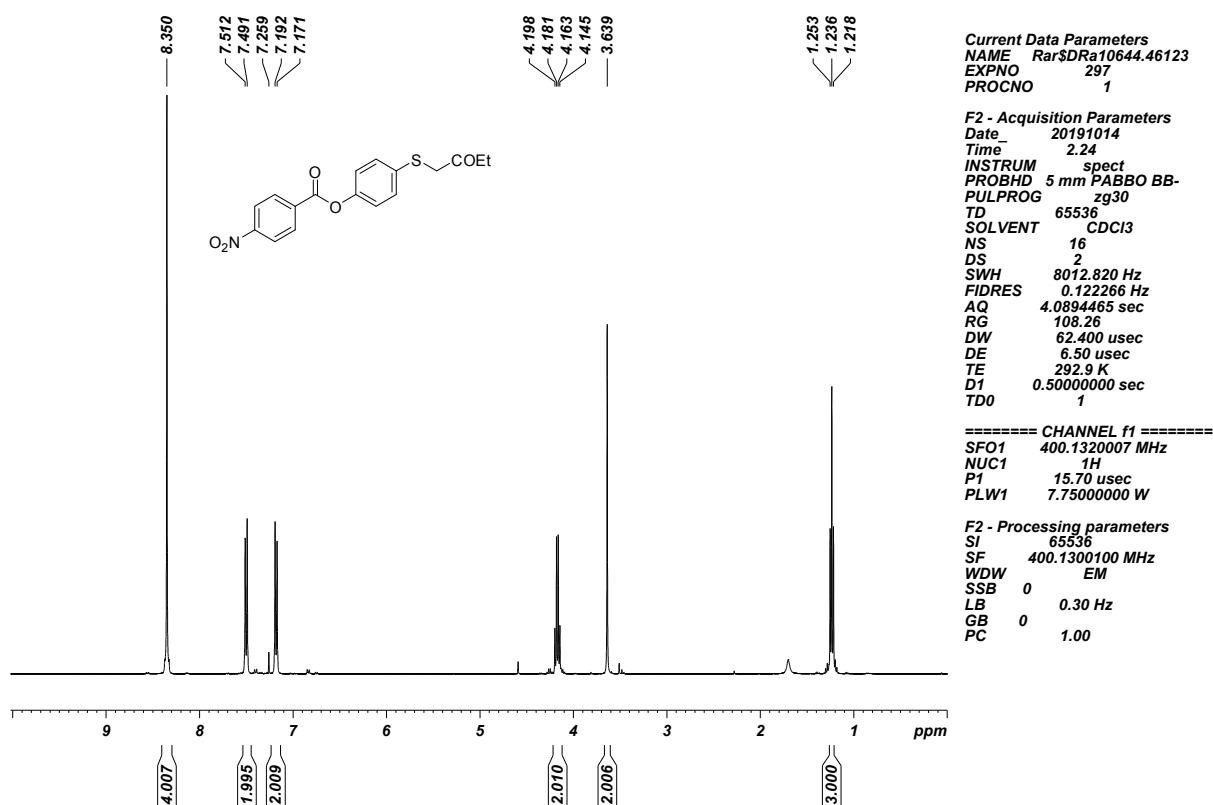


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

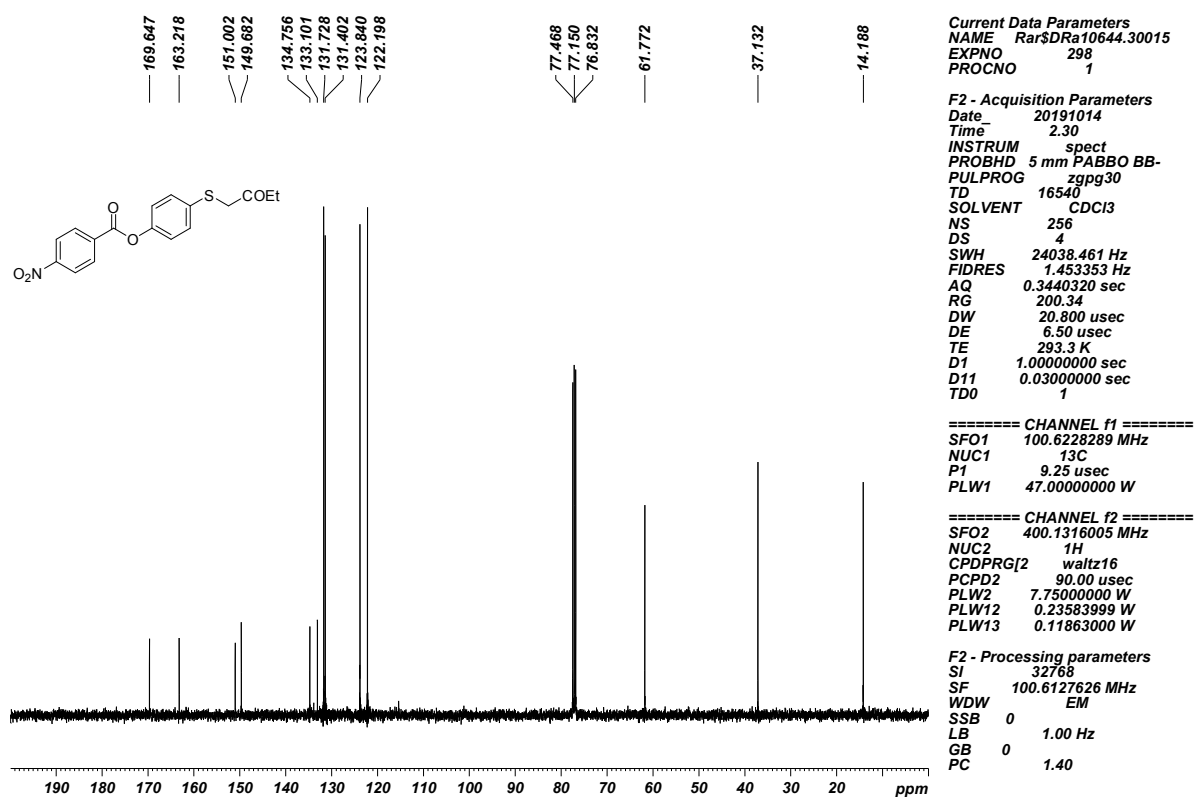


Thioester 1p

^1H NMR (400 MHz, CDCl_3 , 24 °C)

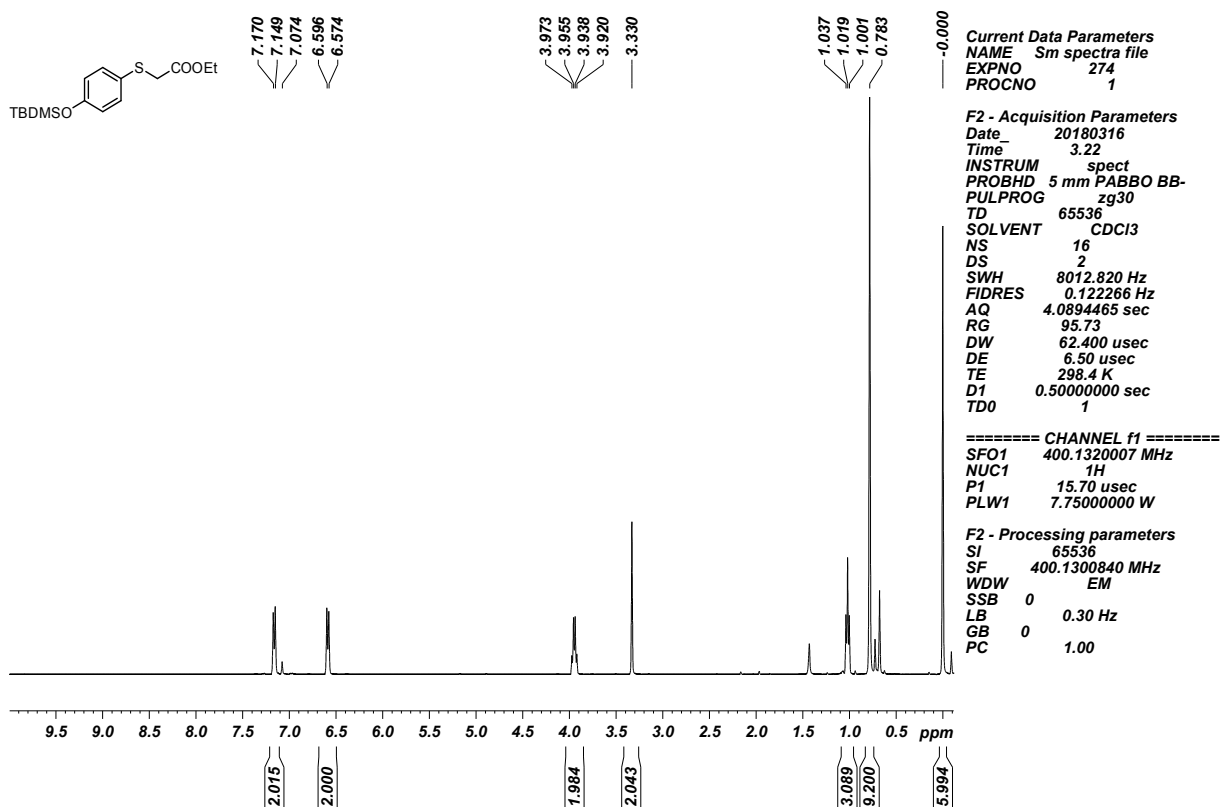


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

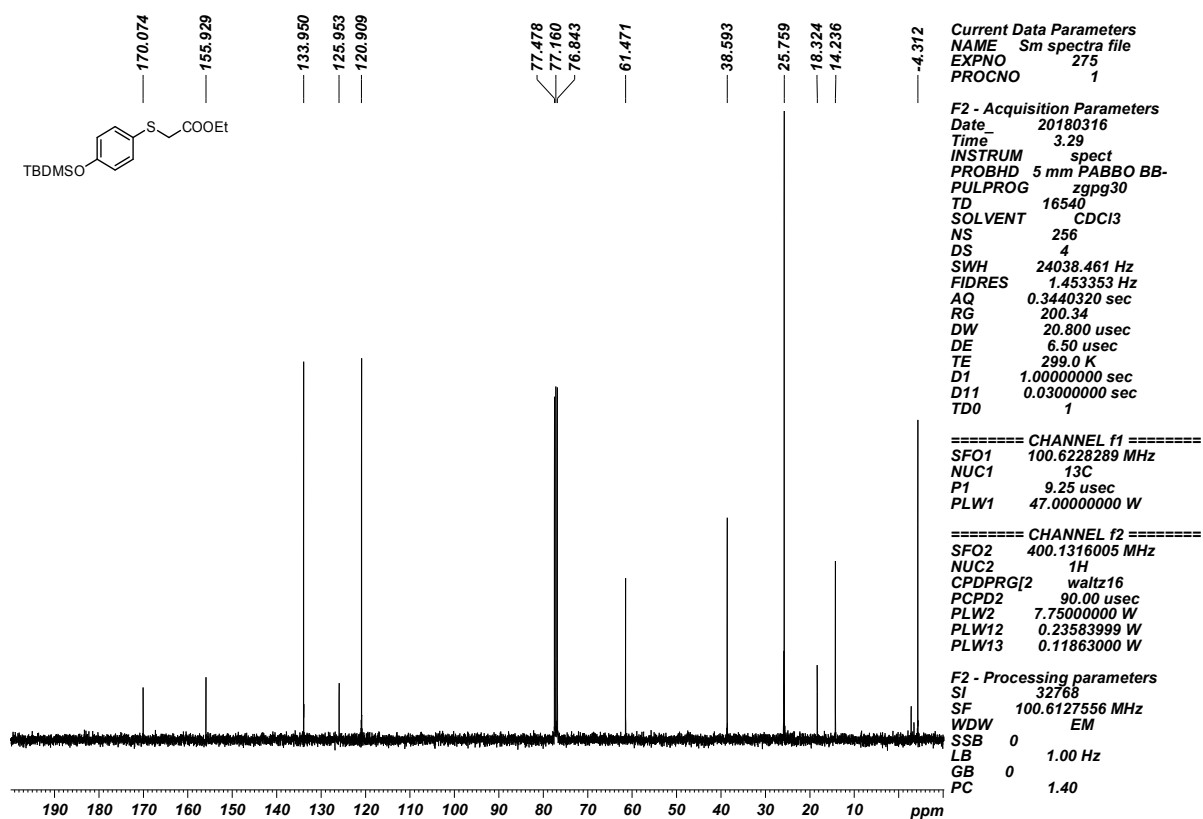


Thioester 1r

^1H NMR (400 MHz, CDCl_3 , 24 °C)

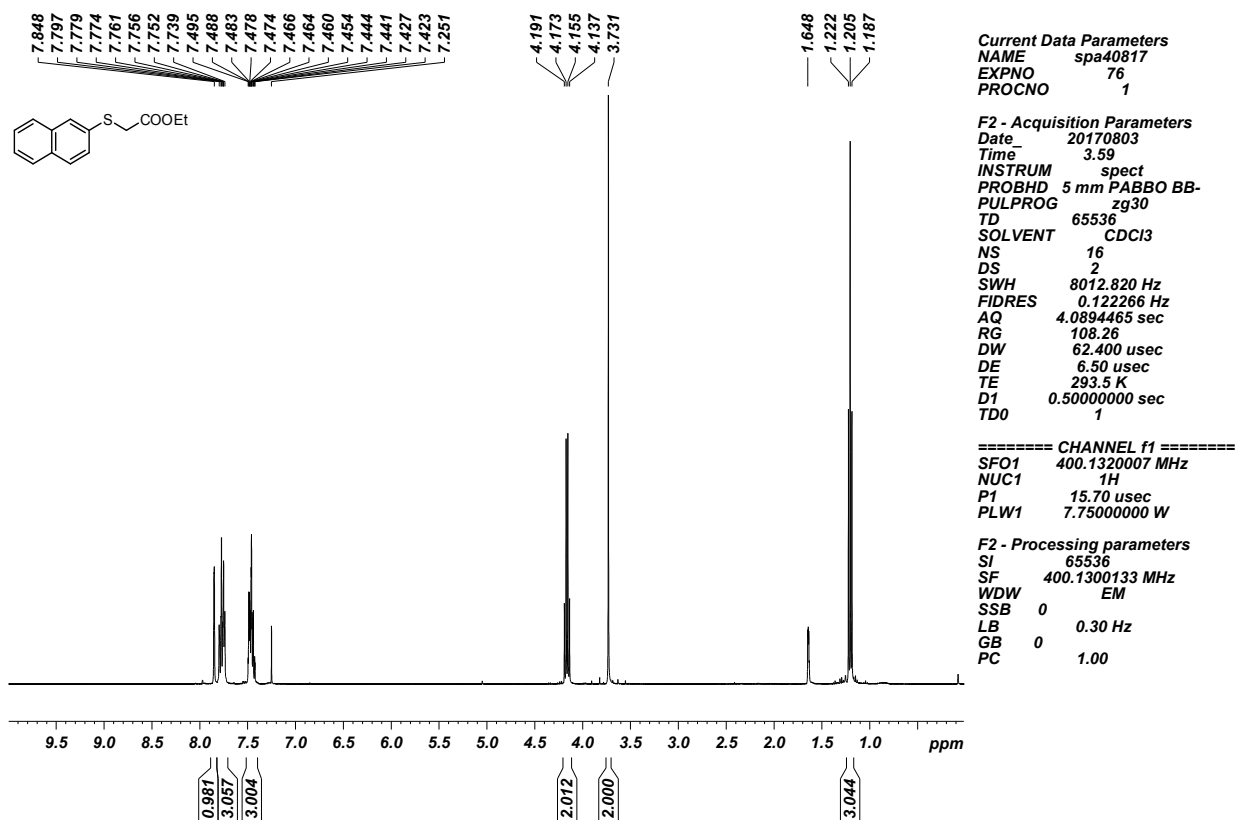


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

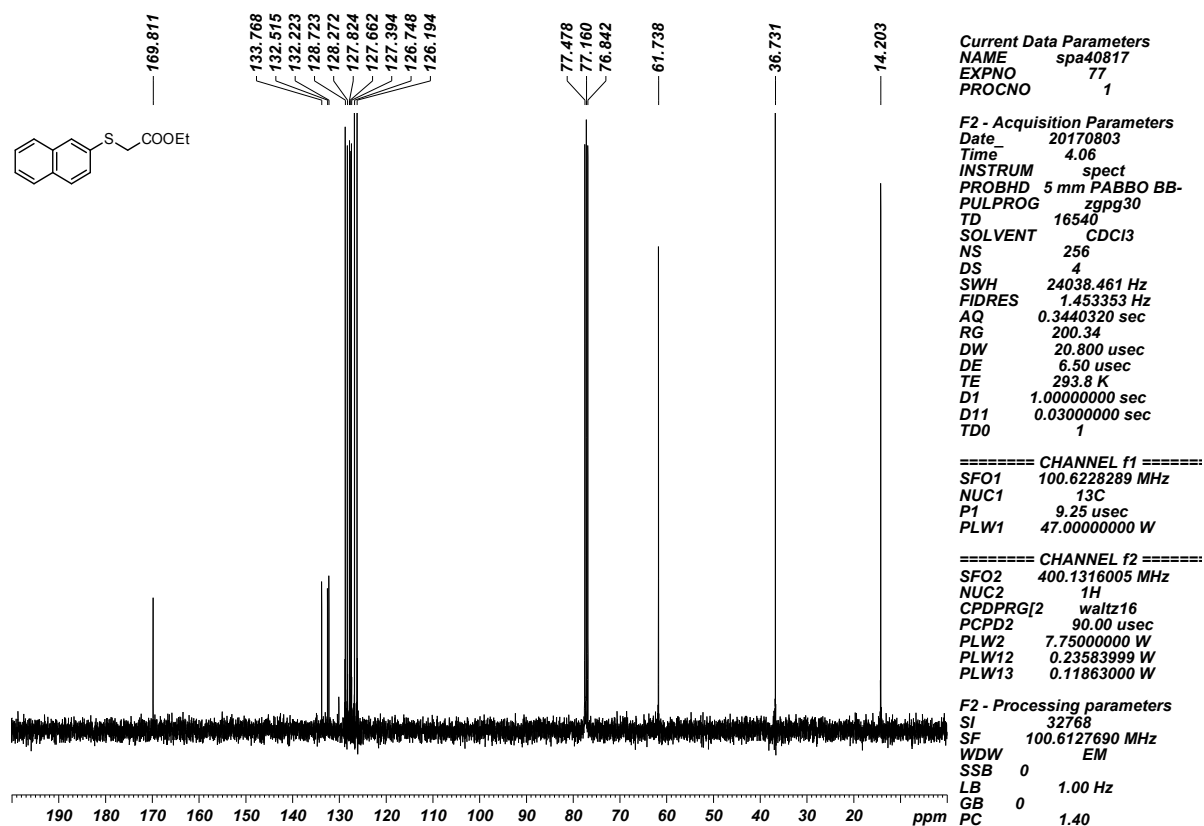


Thioester 1s

^1H NMR (400 MHz, CDCl_3 , 24 °C)

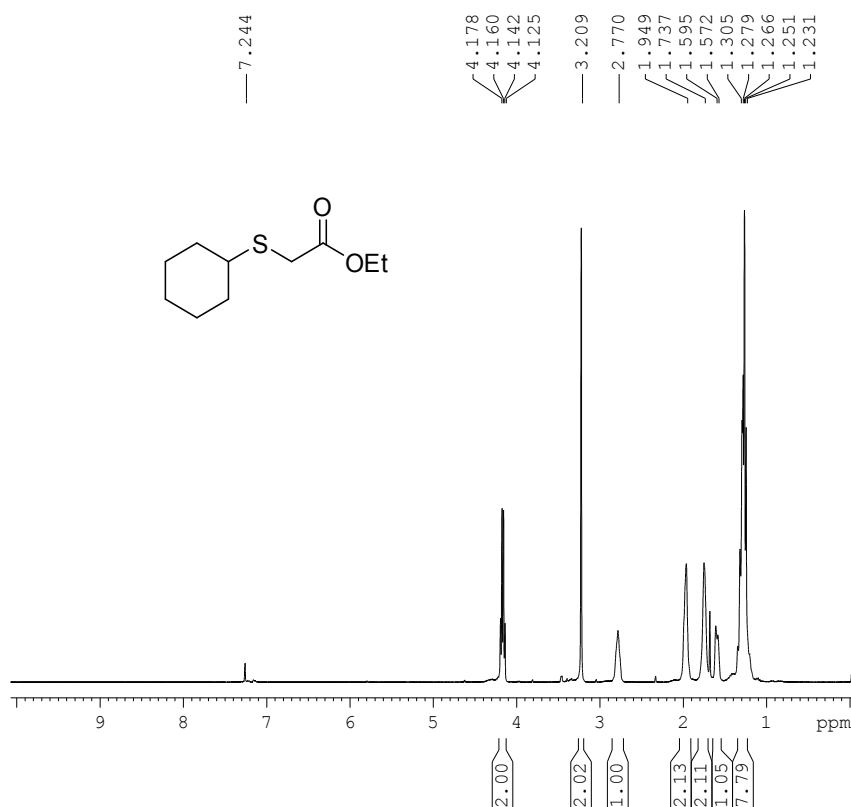


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Thioester 2t

¹H NMR (400 MHz, CDCl₃, 24 °C)



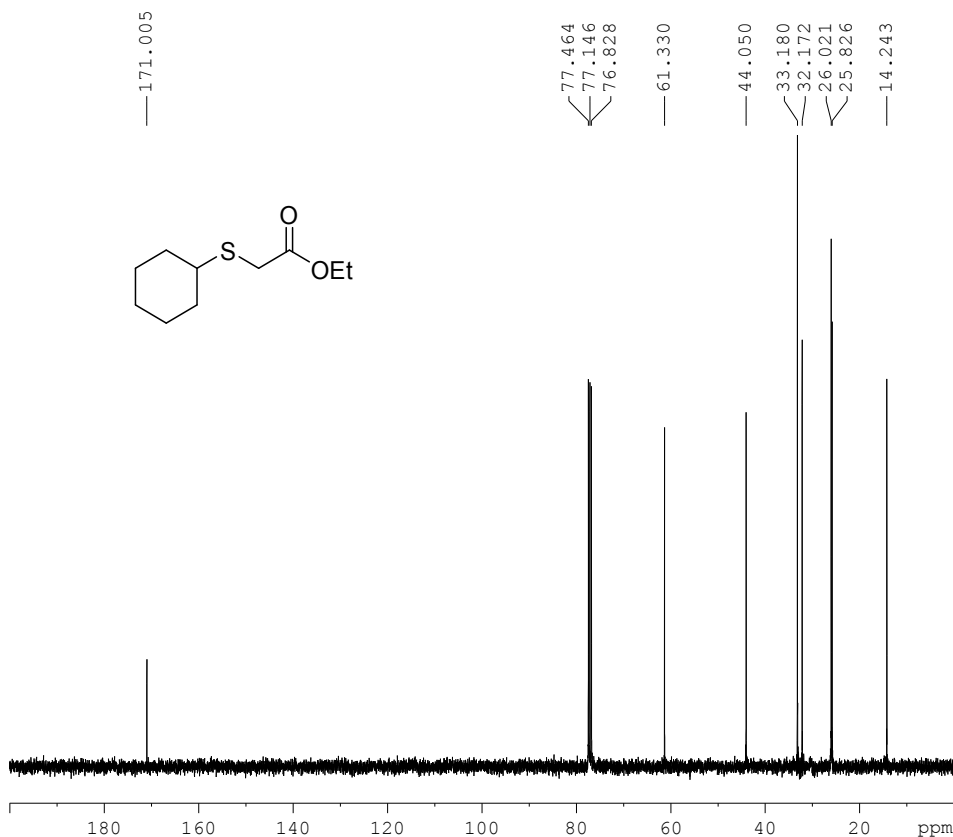
Current Data Parameters
 NAME spa40220
 EXPNO 413
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200229
 Time_ 16.16
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 108.26
 DW 62.400 usec
 DE 6.50 usec
 TE 297.4 K
 D1 0.50000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1320007 MHz
 NUC1 1H
 P1 15.70 usec
 PLW1 7.75000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1300005 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Current Data Parameters
 NAME 411 thio cy
 EXPNO 4
 PROCNO

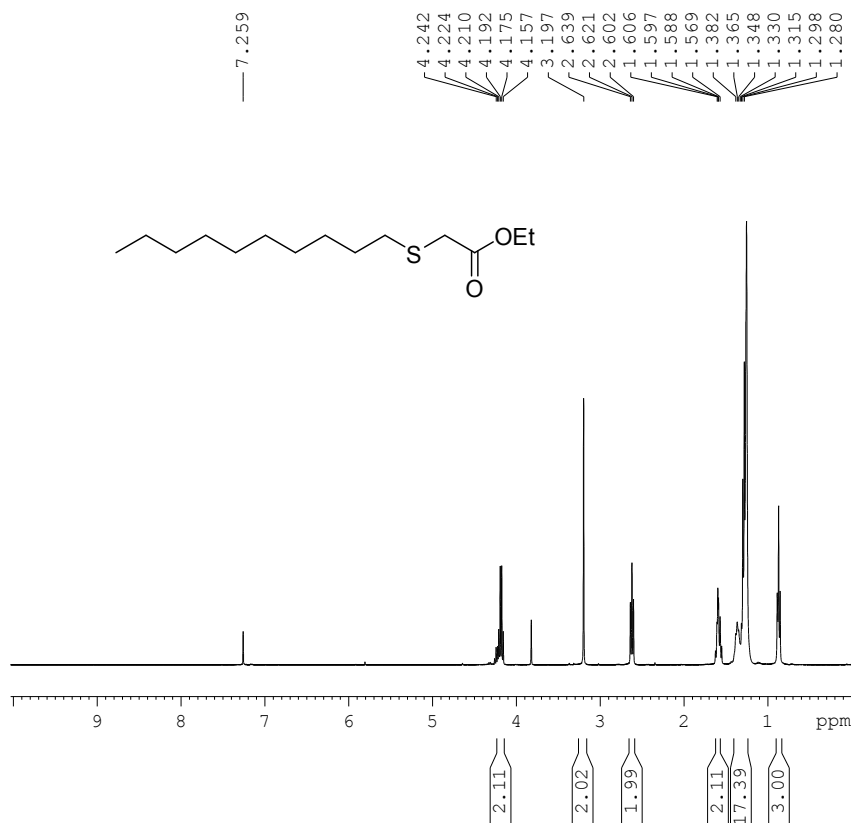
F2 - Acquisition Parameters
 Date_ 202002
 Time_ 16.
 INSTRUM spe
 PROBHD 5 mm PABBO B
 PULPROG zgpg
 TD 165
 SOLVENT CDC
 NS 2
 DS
 SWH 24038.4
 FIDRES 1.4533
 AQ 0.34403
 RG 200.
 DW 20.8
 DE 6.
 TE 297
 D1 1.000000
 D11 0.030000
 TD0

===== CHANNEL f1 =
 SFO1 100.62282
 NUC1 1
 P1 9.
 PLW1 47.000000

===== CHANNEL f2 =
 SFO2 400.13160
 NUC2
 CPDPRG[2] waltz
 PCPD2 90.
 PLW2 7.7500000

Thioester 2u

¹H NMR (400 MHz, CDCl₃, 24 °C)



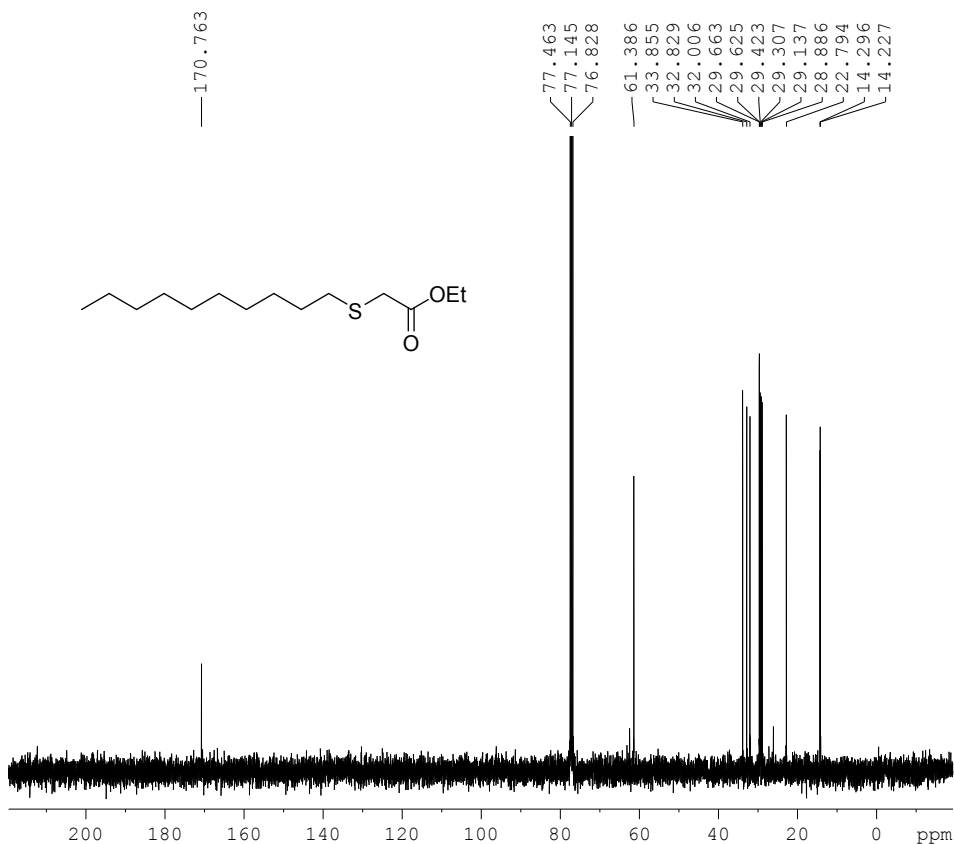
Current Data Parameters
NAME spa40320
EXPNO 201
PROCNO 1

F2 - Acquisition Parameters
Date_ 20200315
Time_ 11.52
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 153.13
DW 62.400 usec
DE 6.50 usec
TE 297.0 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300064 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Current Data Parameters
NAME spa403
EXPNO 2
PROCNO

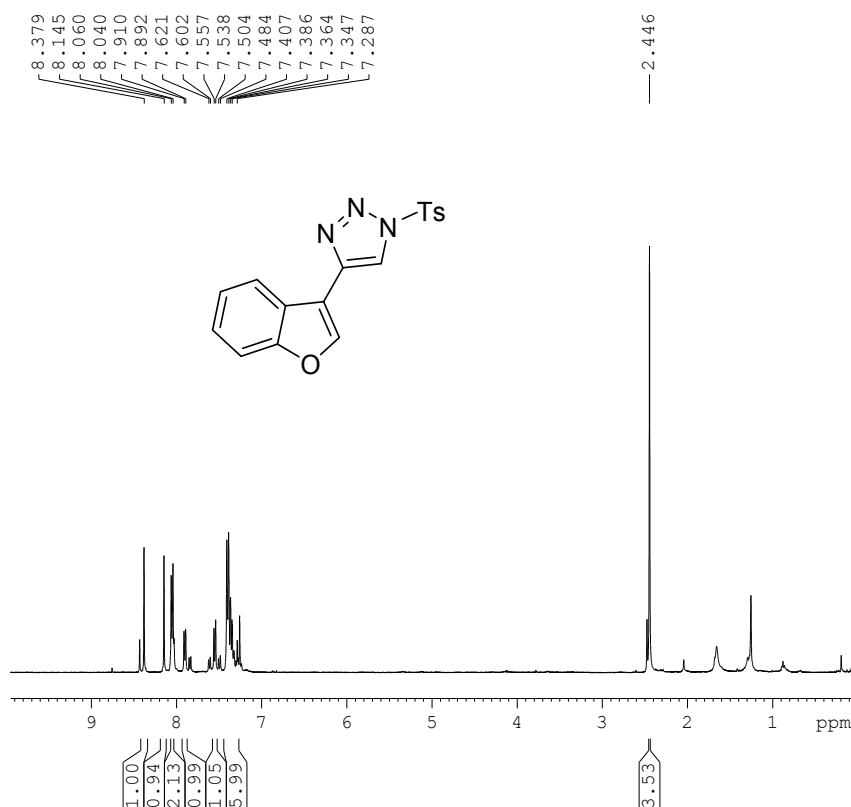
F2 - Acquisition Parameters
Date_ 202003
Time_ 11.
INSTRUM spe
PROBHD 5 mm PABBO B
PULPROG zgpg
TD 165
SOLVENT CDC
NS 2
DS
SWH 24038.4
FIDRES 1.4533
AQ 0.34403
RG 200.
DW 20.8
DE 6.
TE 297
D1 1.000000
D11 0.030000
TD0

===== CHANNEL f1 =
SFO1 100.62282
NUC1 1
P1 9.
PLW1 47.000000

===== CHANNEL f2 =
SFO2 400.13160
NUC2
CPDPRG[2] waltz
PCPD2 90.
PLW2 7.750000

Thioester 2ag

¹H NMR (400 MHz, CDCl₃, 24 °C)



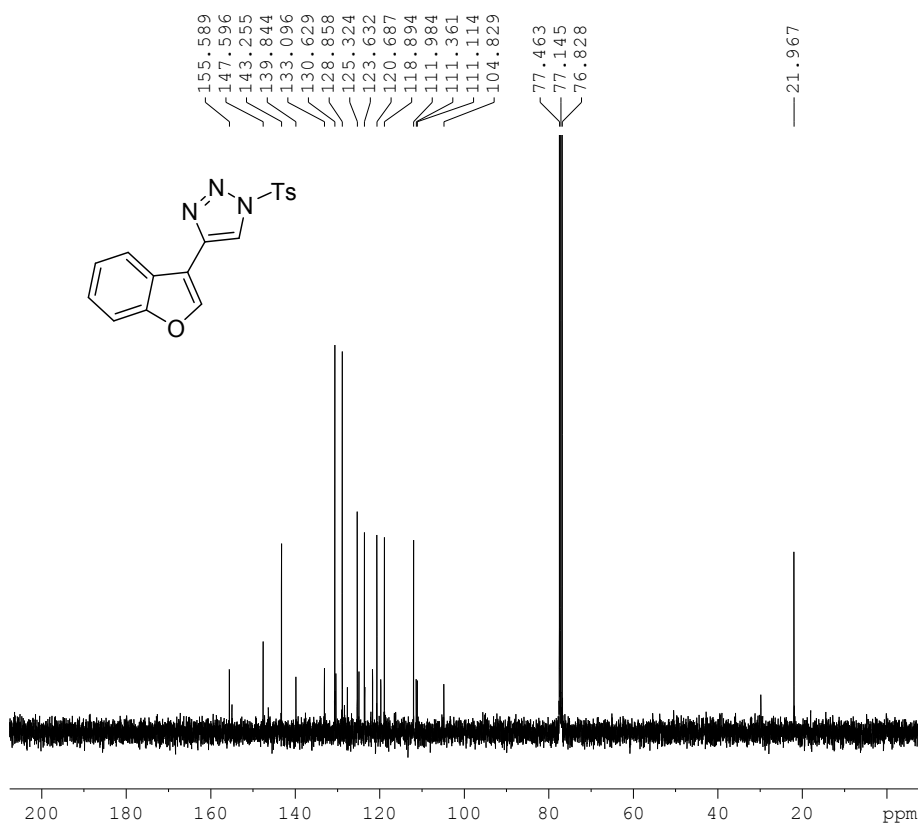
Current Data Parameters
NAME spa40320
EXPNO 209
PROCNO 1

F2 - Acquisition Parameters
Date_ 20200315
Time 22.17
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 200.34
DW 62.400 usec
DE 6.50 usec
TE 297.0 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Current Data Parameters
NAME spa403
EXPNO 2
PROCNO

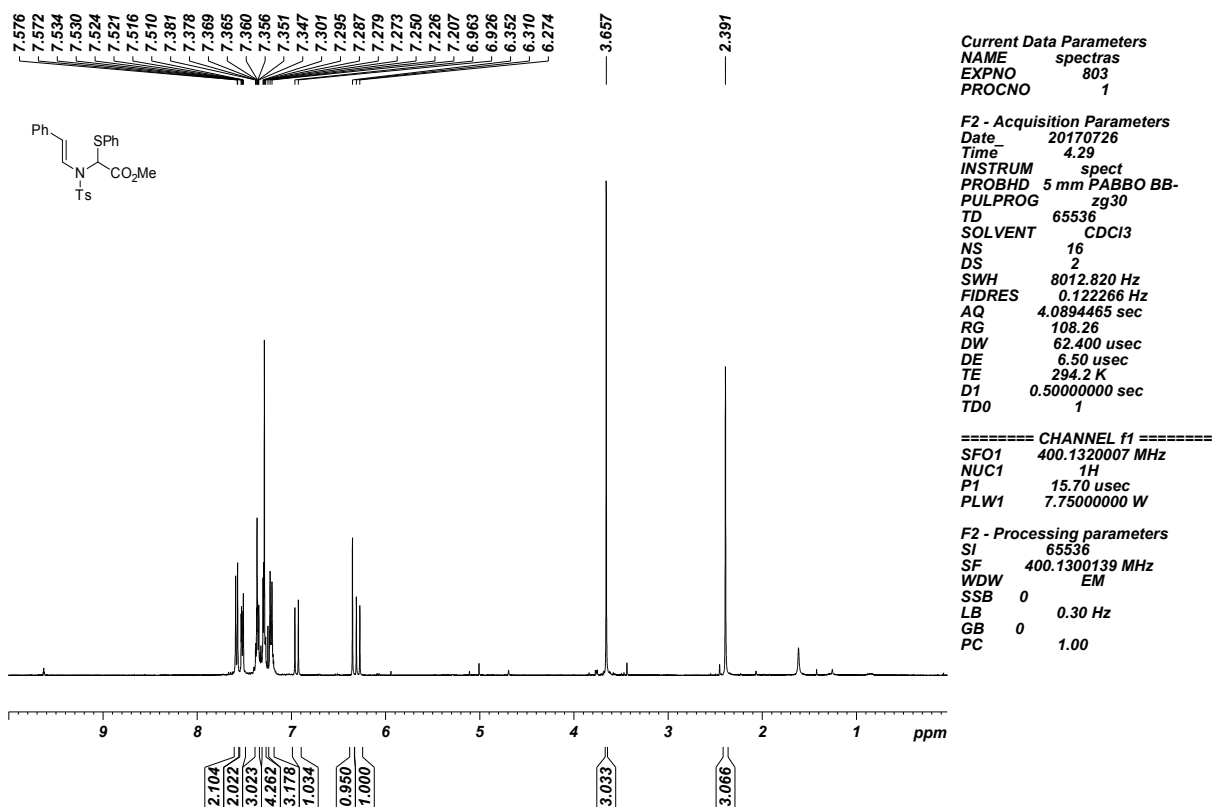
F2 - Acquisition Parameters
Date_ 202003
Time 22.
INSTRUM spe
PROBHD 5 mm PABBO B
PULPROG zgpg
TD 165
SOLVENT CDC
NS 2
DS
SWH 24038.4
FIDRES 1.4533
AQ 0.34403
RG 200.
DW 20.8
DE 6.
TE 297
D1 1.000000
D11 0.030000
TD0

===== CHANNEL f1 =
SFO1 100.62282
NUC1 1
P1 9.
PLW1 47.000000

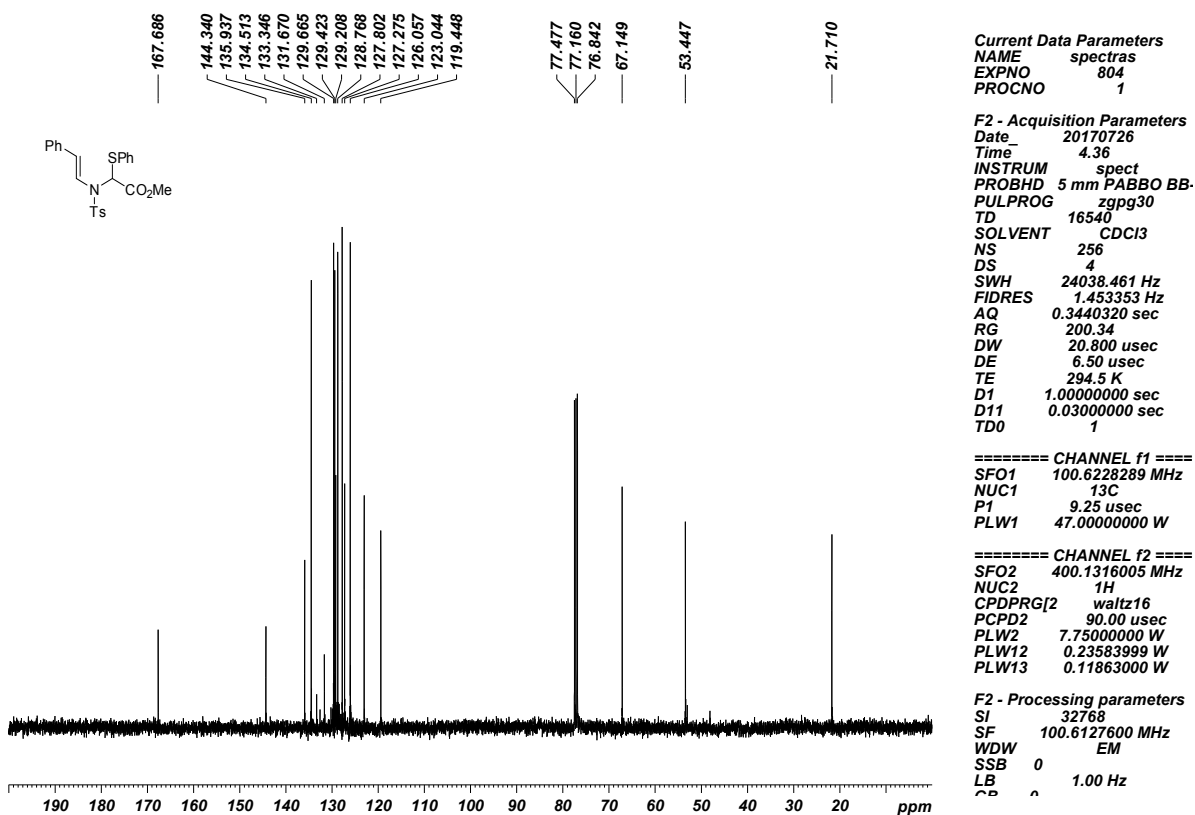
===== CHANNEL f2 =
SFO2 400.13160
NUC2
CPDPRG[2] waltz
PCPD2 90.
P1 7.750000

Enamide 4a

¹H NMR (400 MHz, CDCl₃, 24 °C)

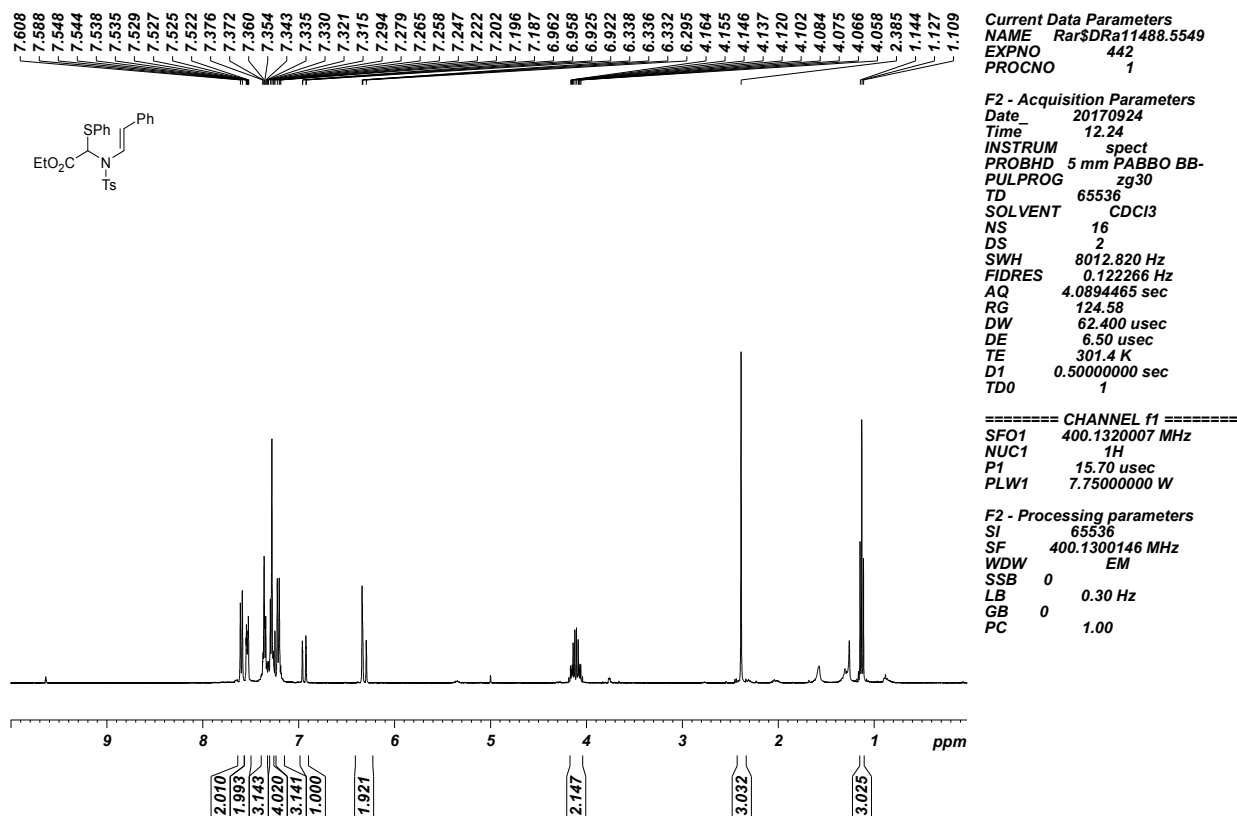


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

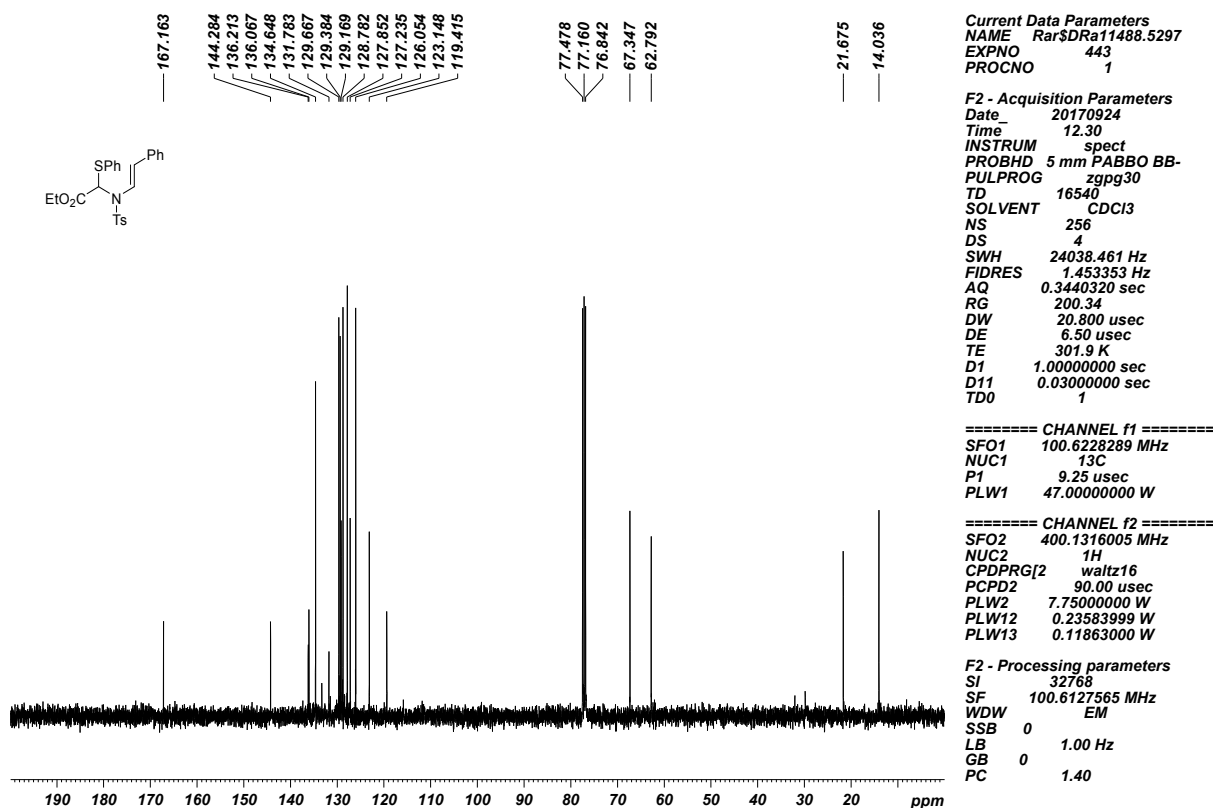


Enamide 4b

^1H NMR (400 MHz, CDCl_3 , 24 °C)

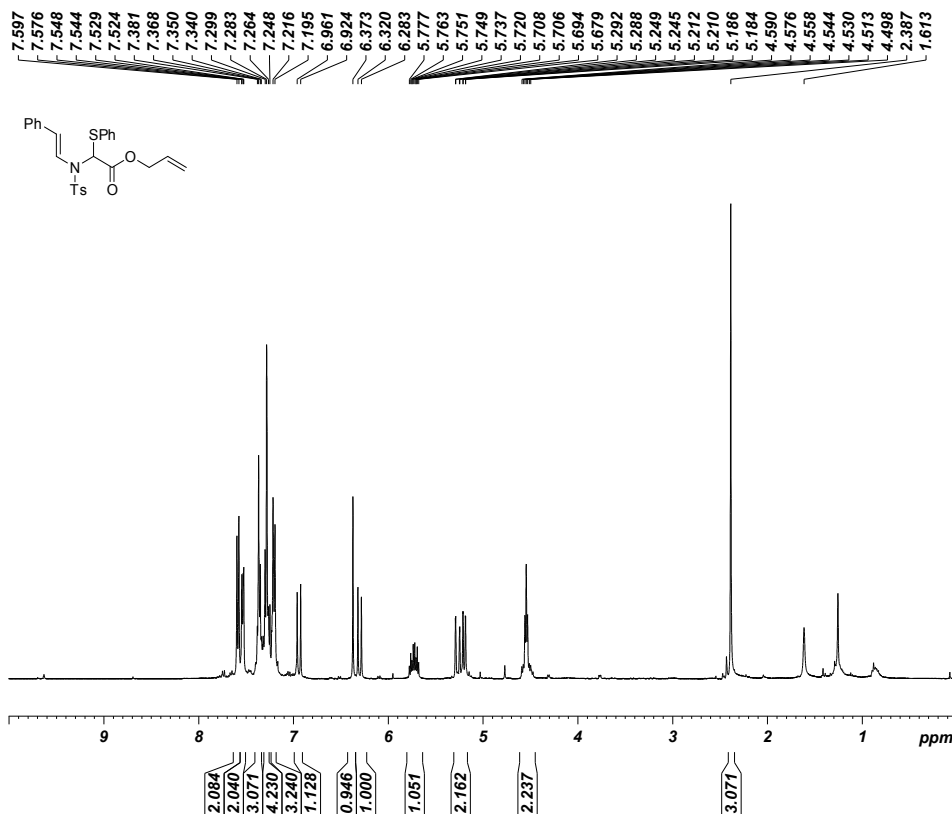


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Enamide 4c

^1H NMR (400 MHz, CDCl_3 , 24 °C)



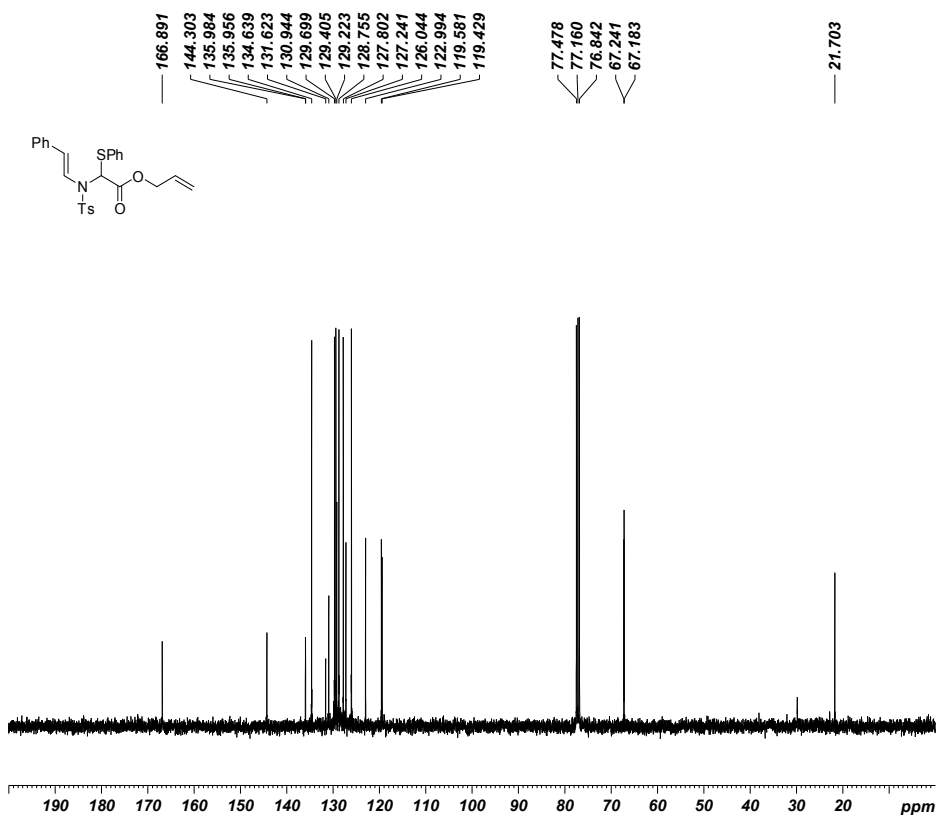
Current Data Parameters
NAME spectras
EXPNO 402
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171218
Time 15.54
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 108.26
DW 62.400 usec
DE 6.50 usec
TE 293.4 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300142 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Current Data Parameters
NAME spectras
EXPNO 403
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171218
Time 16.01
INSTRUM spect
PROBHD 5 mm PABBO BB
PULPROG zgpg30
TD 16540
SOLVENT CDCl3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 293.5 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

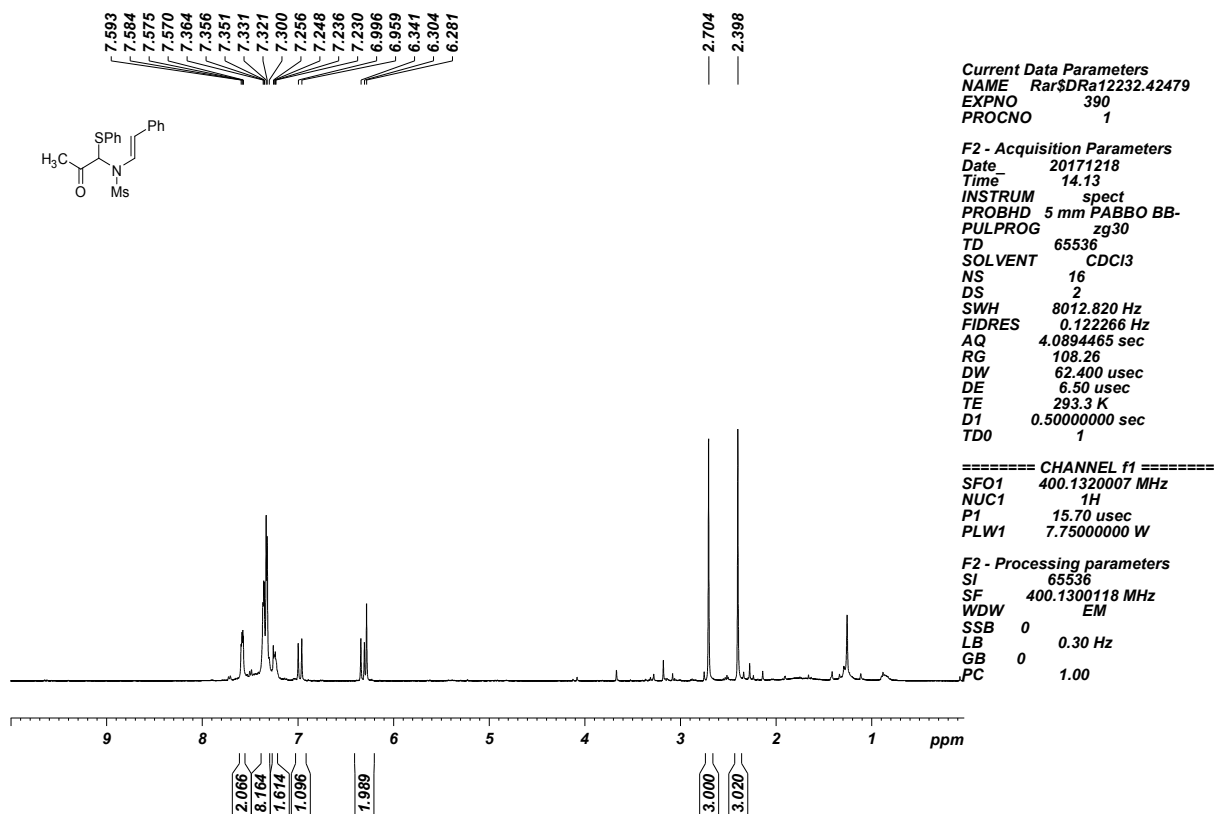
===== CHANNEL f1 =====
SFO1 100.6228289 MHz
NUC1 13C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

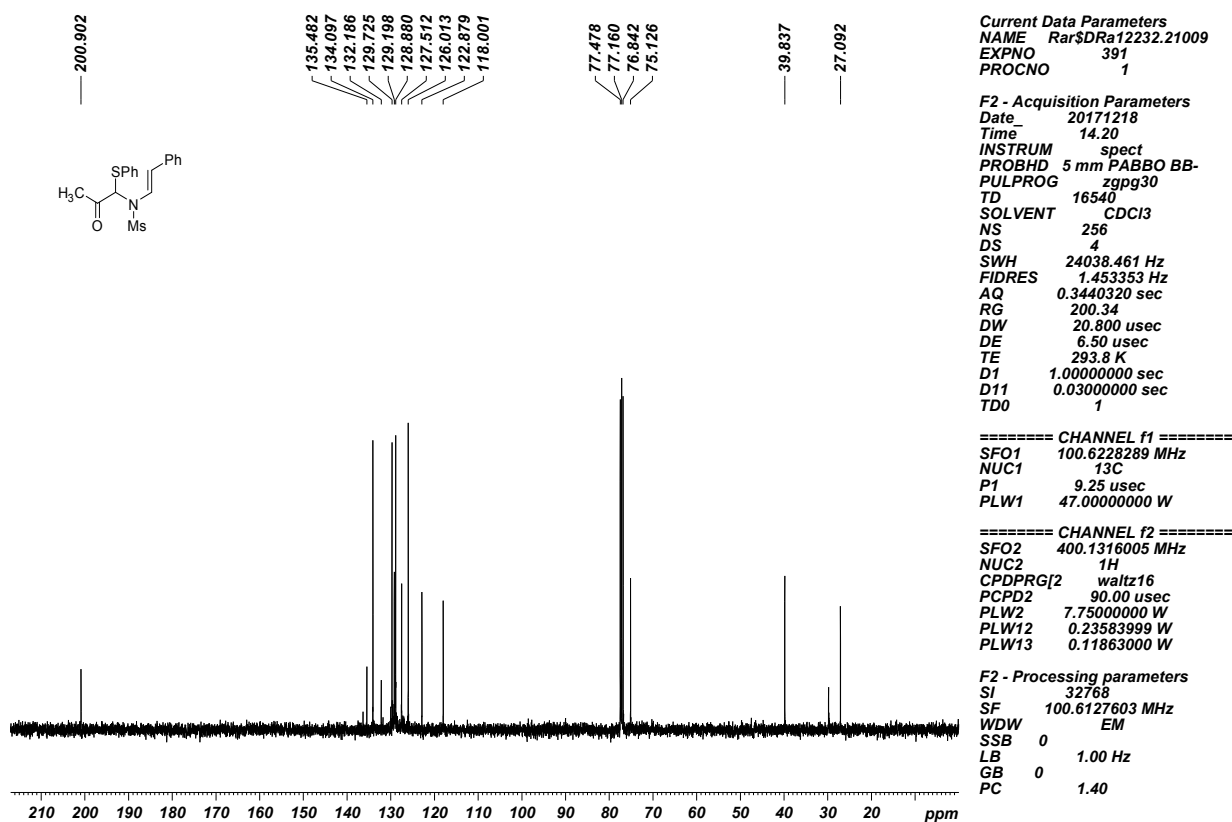
F2 - Processing parameters
SI 32768
SF 100.6127607 MHz
WDW EM
SSB 0
LB 1.00 Hz

Enamide 4d

¹H NMR (400 MHz, CDCl₃, 24 °C)

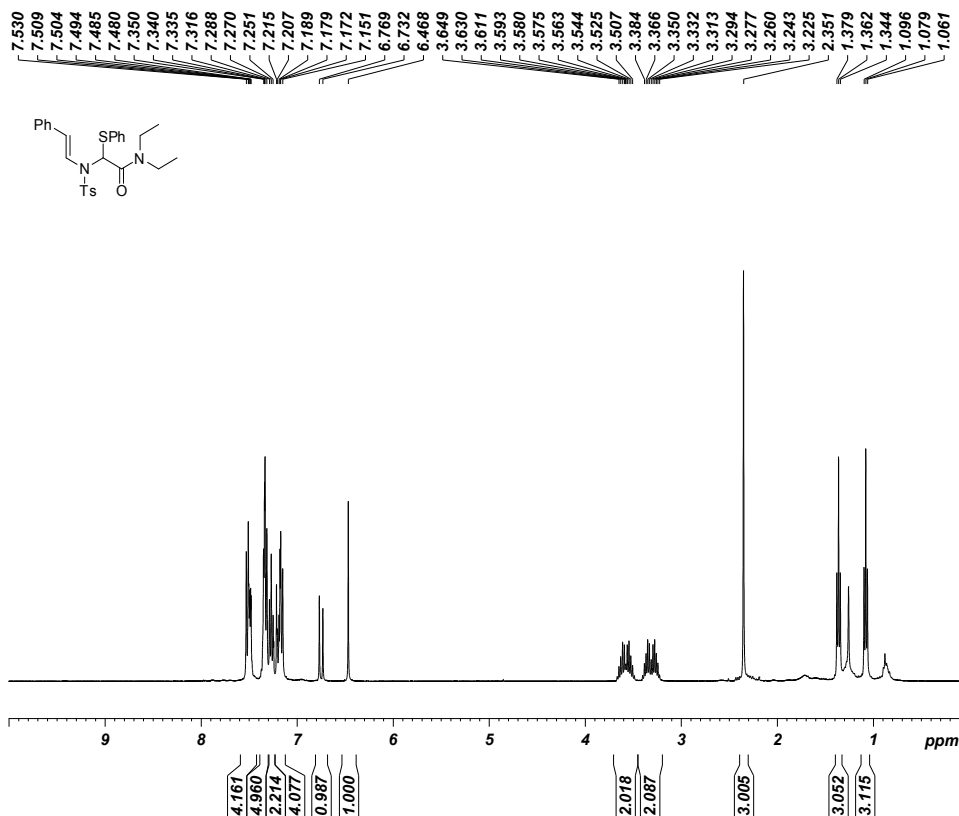


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Enamide 4e

¹H NMR (400 MHz, CDCl₃, 24 °C)



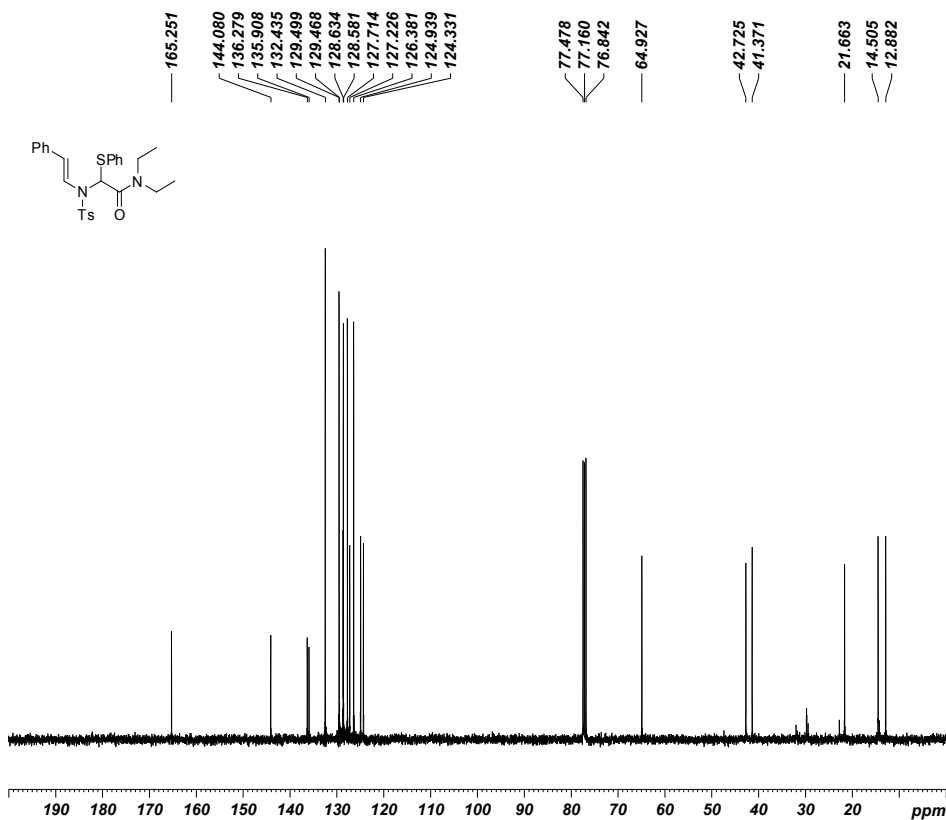
Current Data Parameters
NAME spectras
EXPNO 405
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171218
Time 16.12
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 67.99
DW 62.400 usec
DE 6.50 usec
TE 293.4 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1320007 MHz
NUC1 1H
P1 15.70 usec
PLW1 7.75000000 W

F2 - Processing parameters
SI 65536
SF 400.1300160 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



Current Data Parameters
NAME spectras
EXPNO 406
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171218
Time 16.18
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 16540
SOLVENT CDCl3
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 1.453353 Hz
AQ 0.3440320 sec
RG 200.34
DW 20.800 usec
DE 6.50 usec
TE 293.9 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

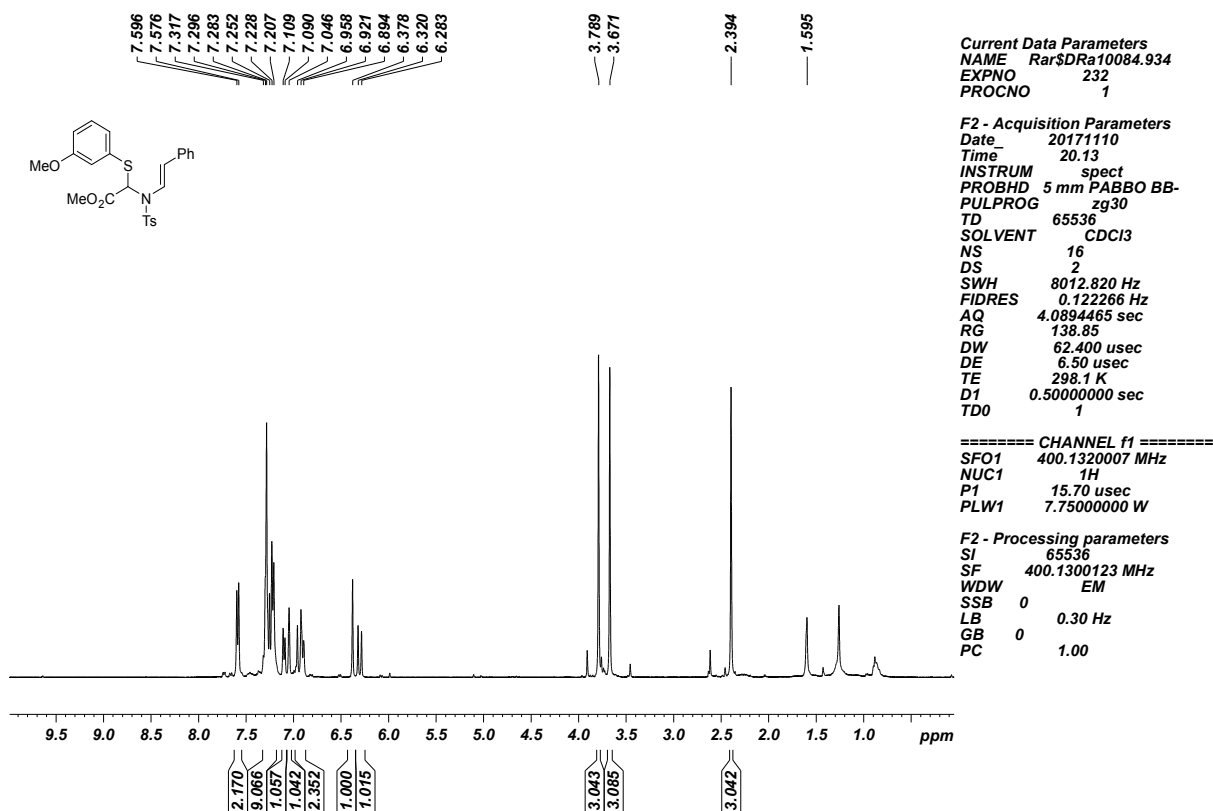
===== CHANNEL f1 =====
SFO1 100.6228289 MHz
NUC1 13C
P1 9.25 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1316005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 7.75000000 W
PLW12 0.23583999 W
PLW13 0.11863000 W

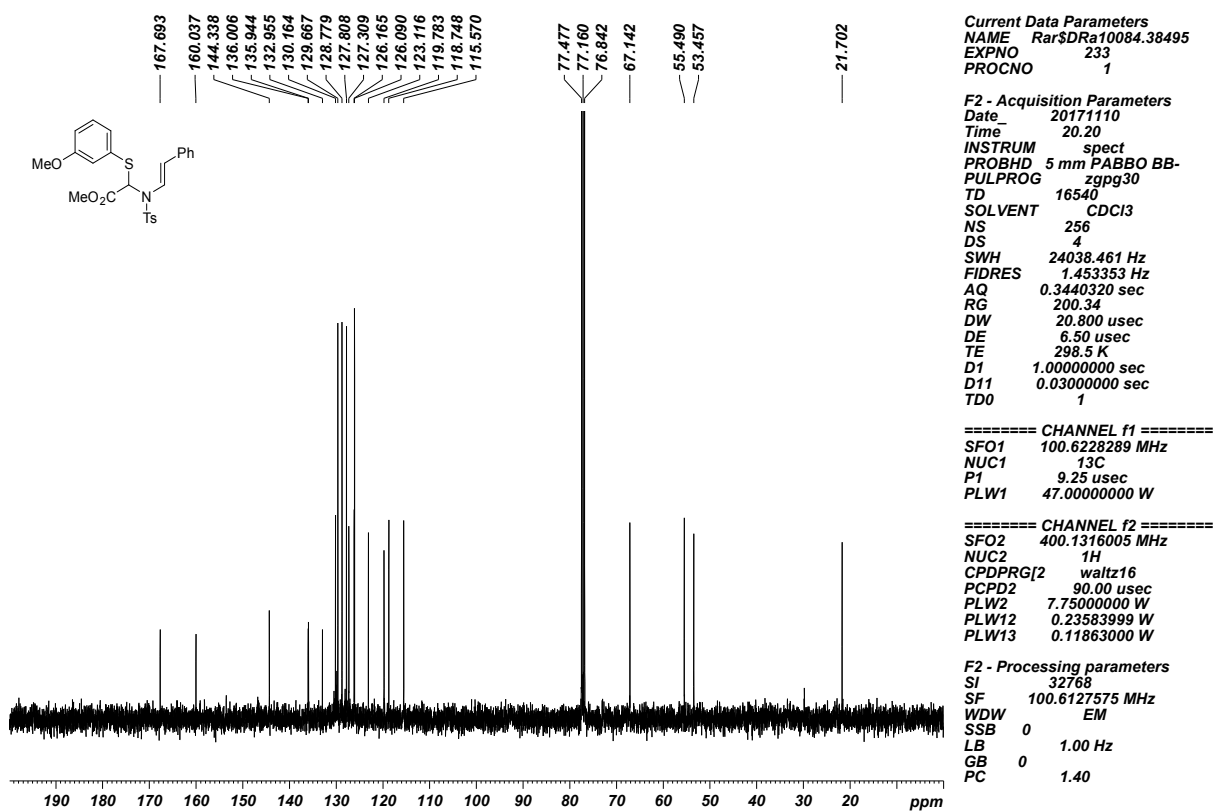
F2 - Processing parameters
SI 32768
SF 100.6127642 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

Enamide 4h

¹H NMR (400 MHz, CDCl₃, 24 °C)

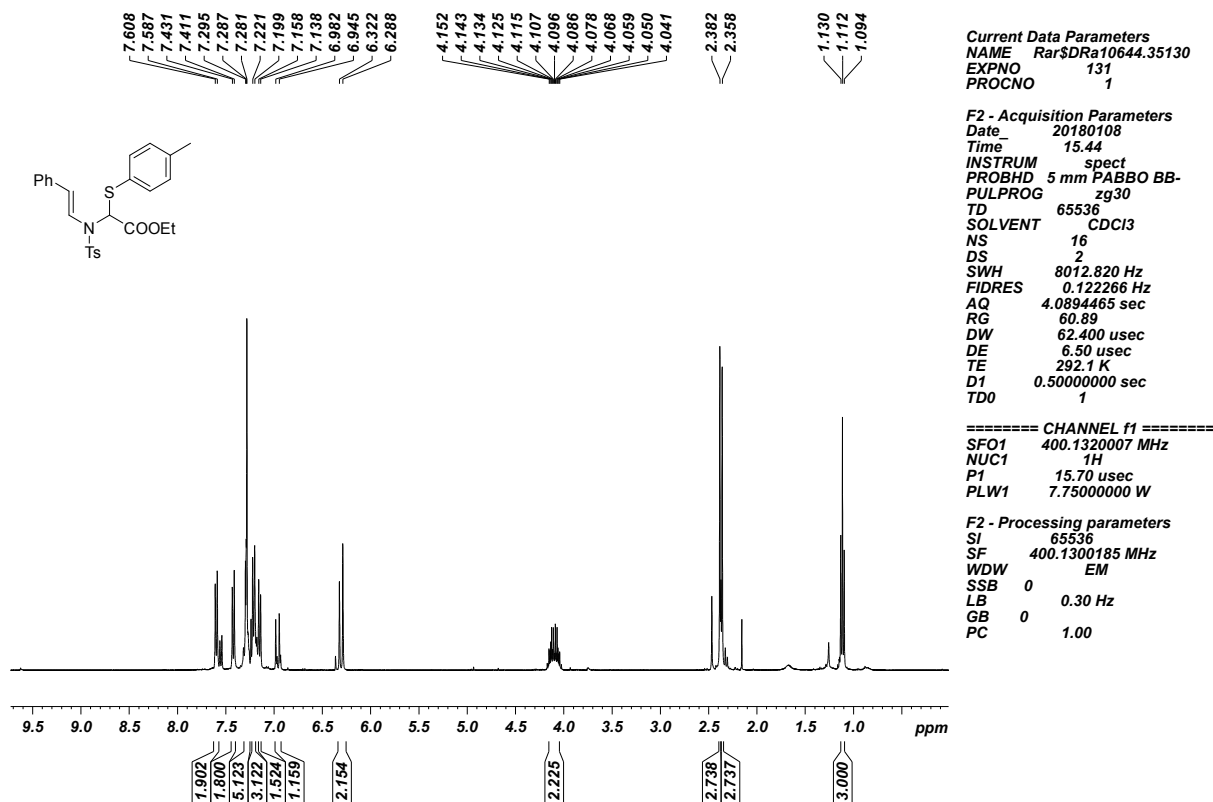


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

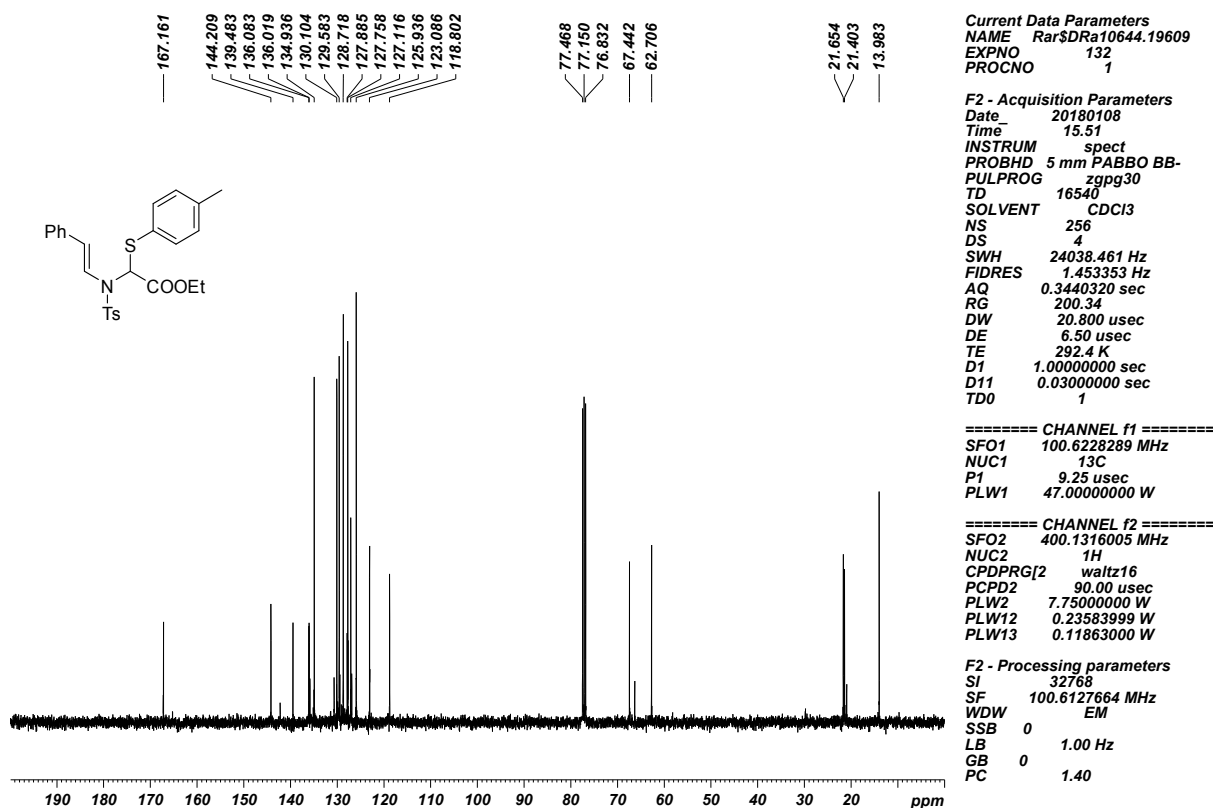


Enamide 4i

¹H NMR (400 MHz, CDCl₃, 24 °C)

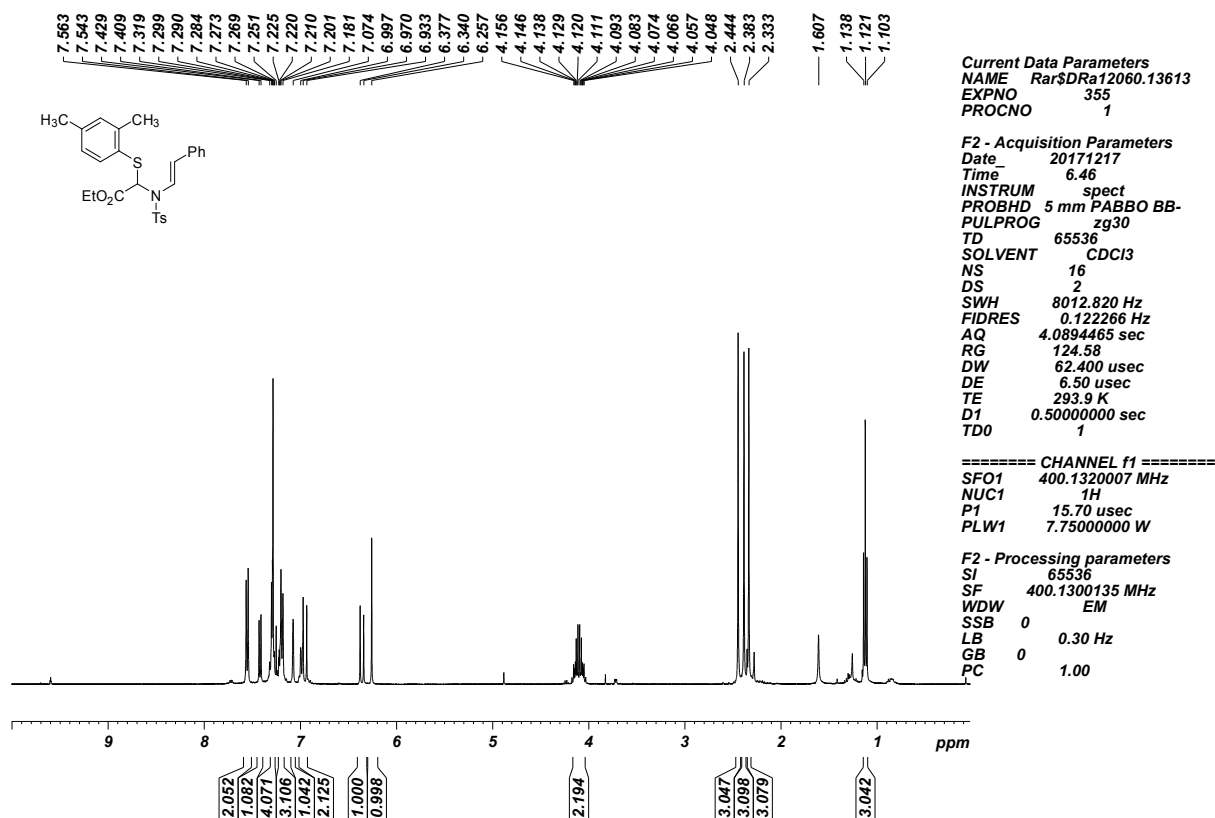


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

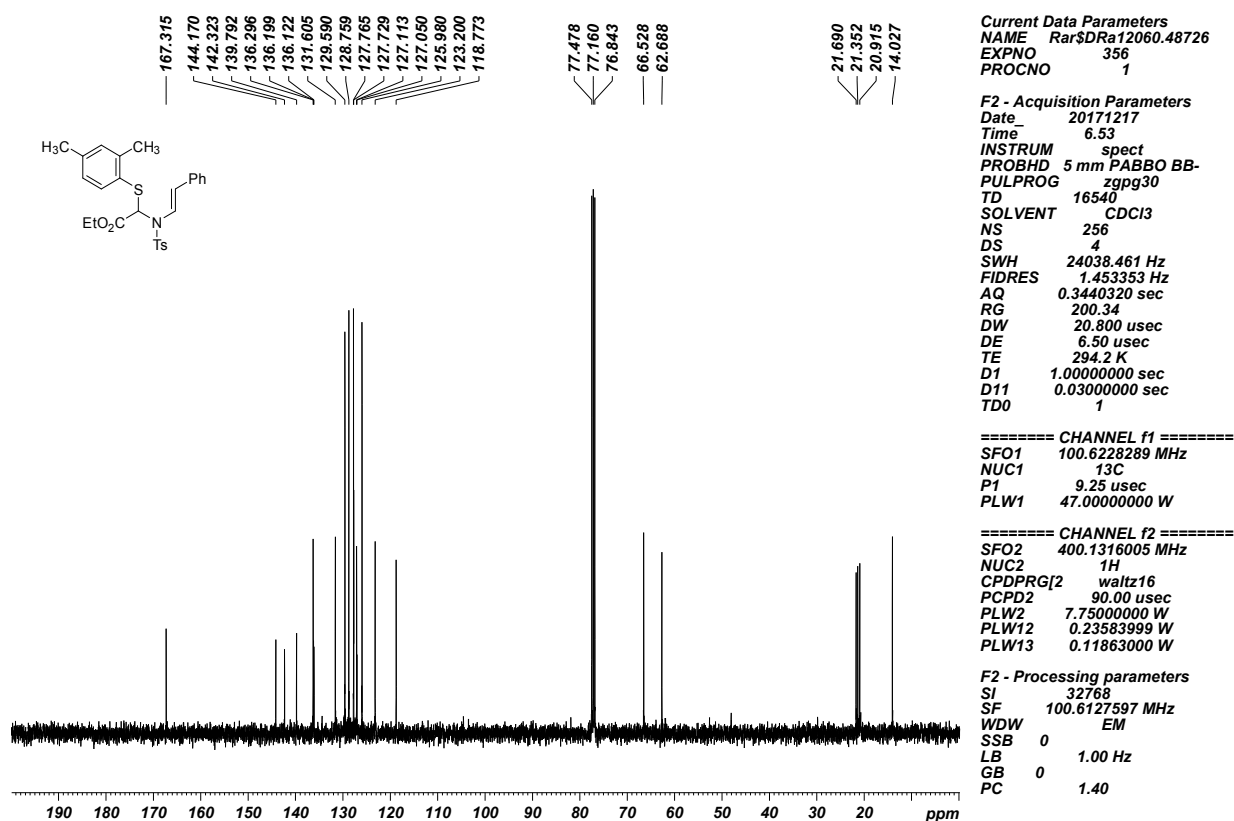


Enamide 4j

¹H NMR (400 MHz, CDCl₃, 24 °C)

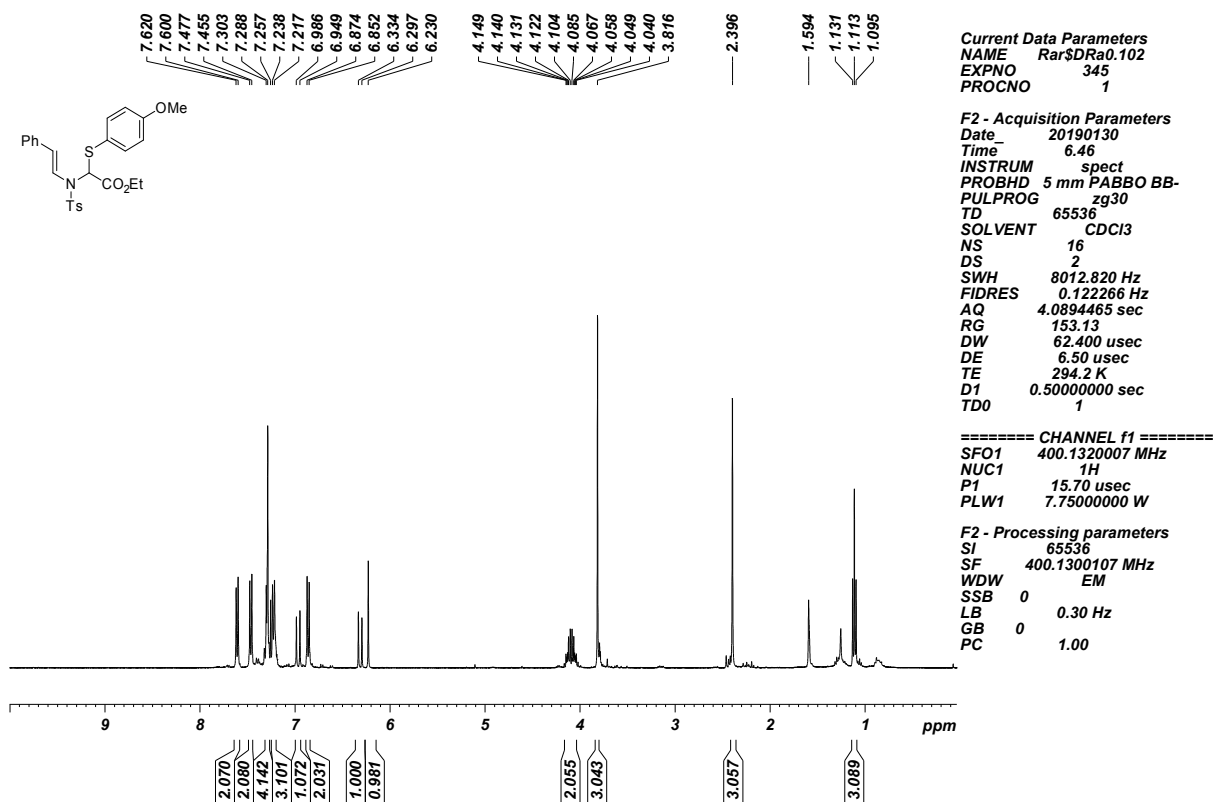


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

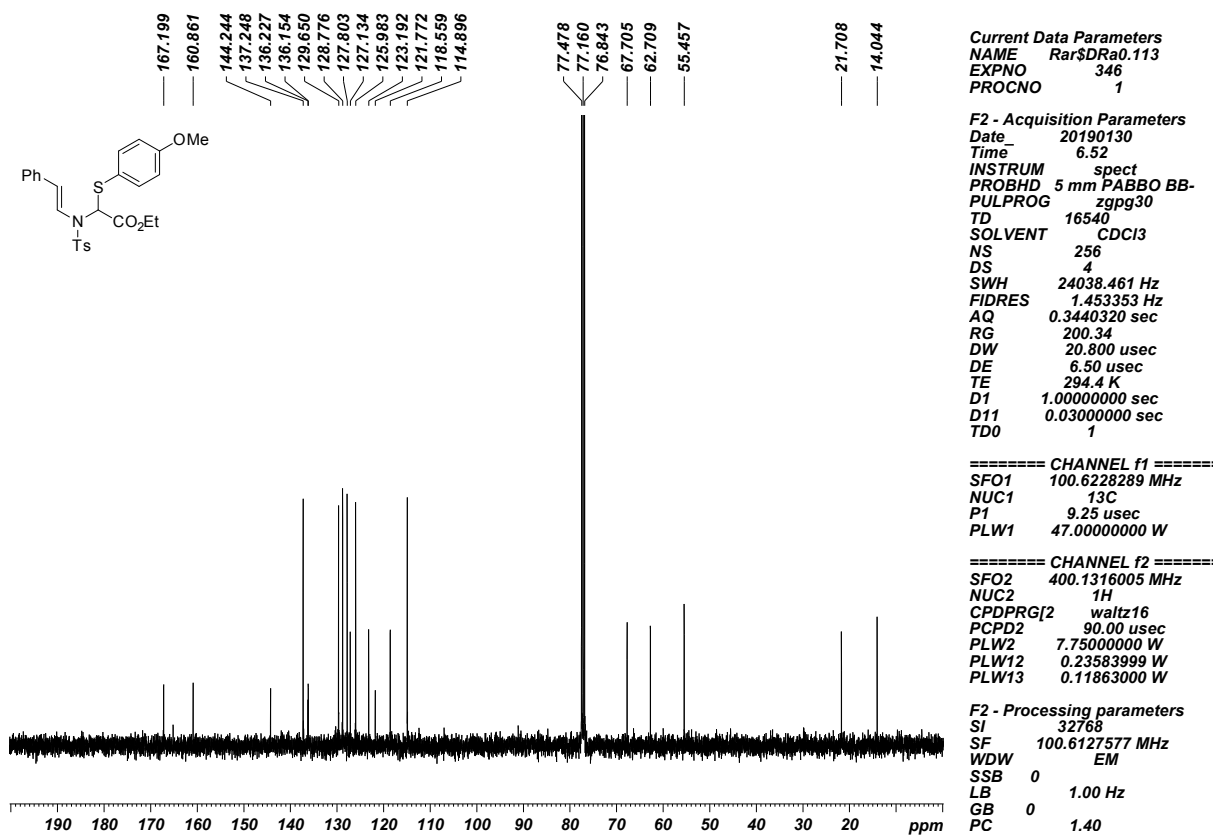


Enamide 4k

^1H NMR (400 MHz, CDCl_3 , 24 °C)

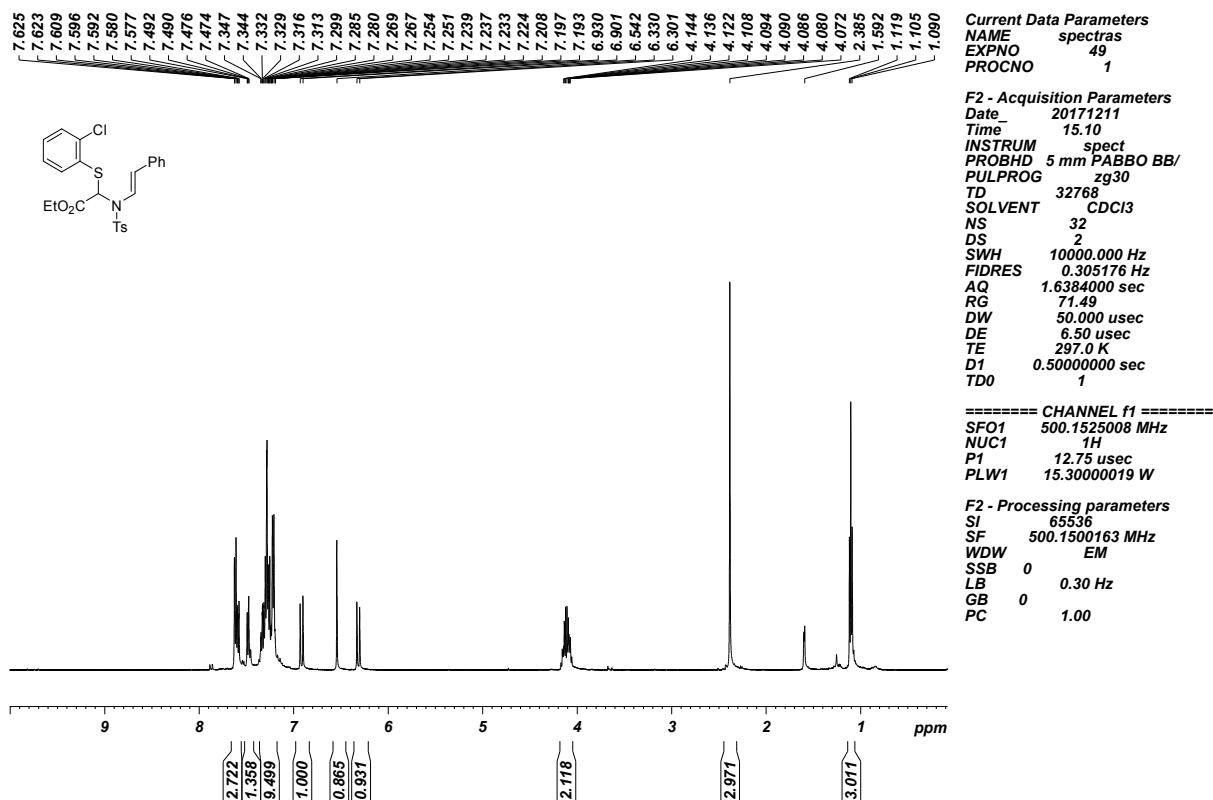


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

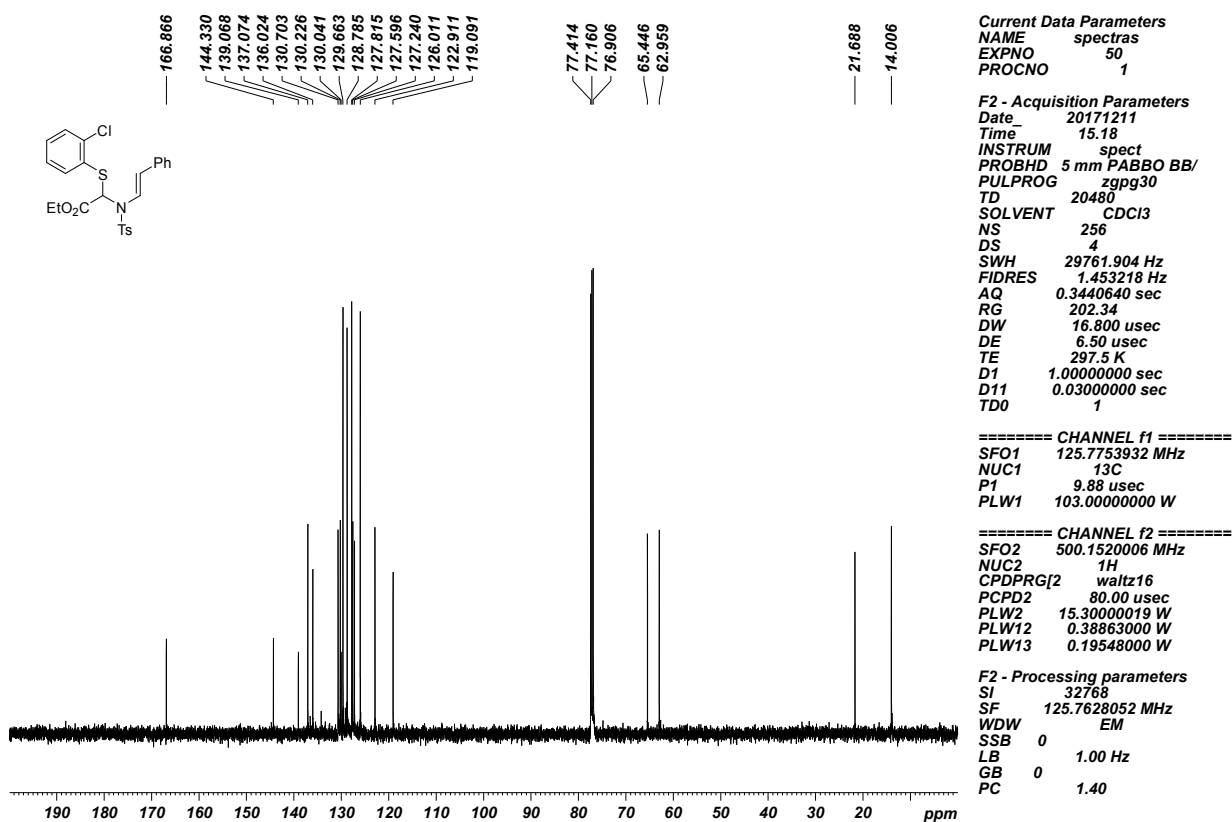


Enamide 4l

¹H NMR (400 MHz, CDCl₃, 24 °C)

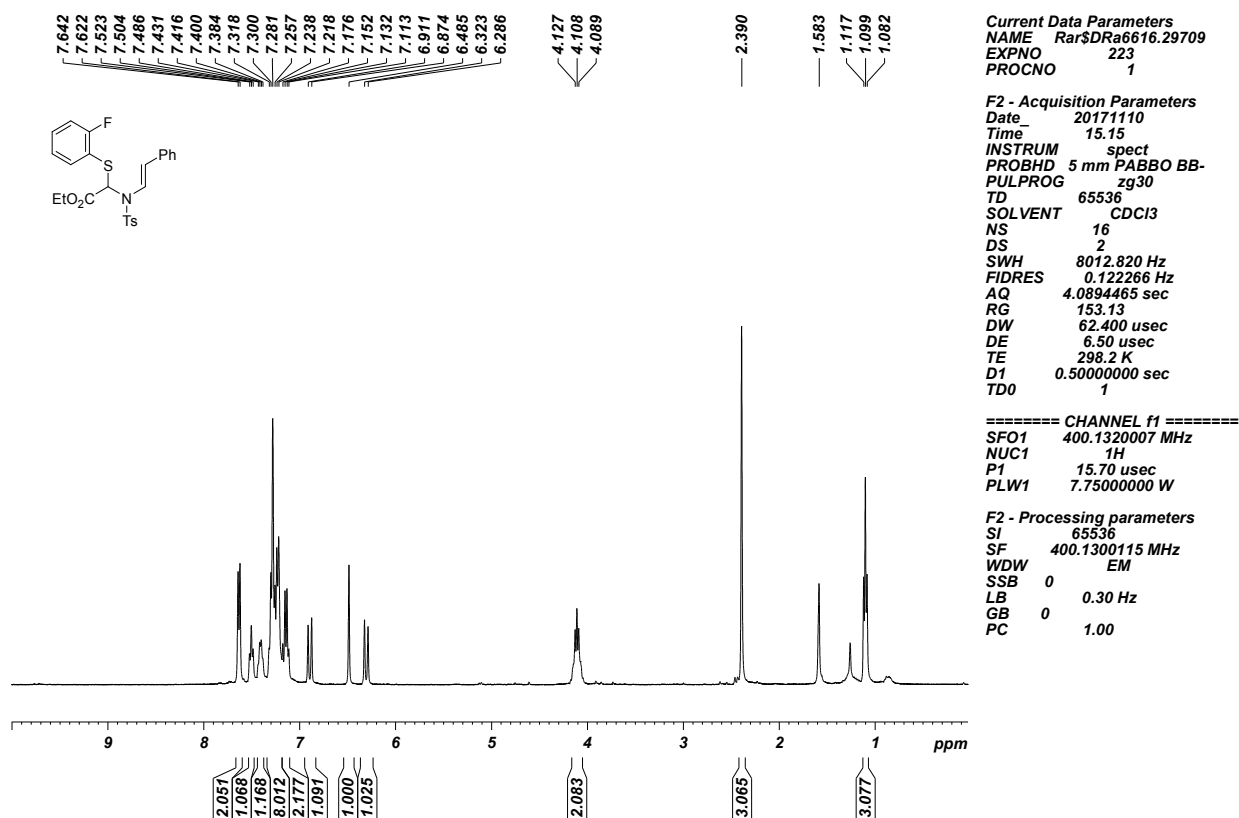


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

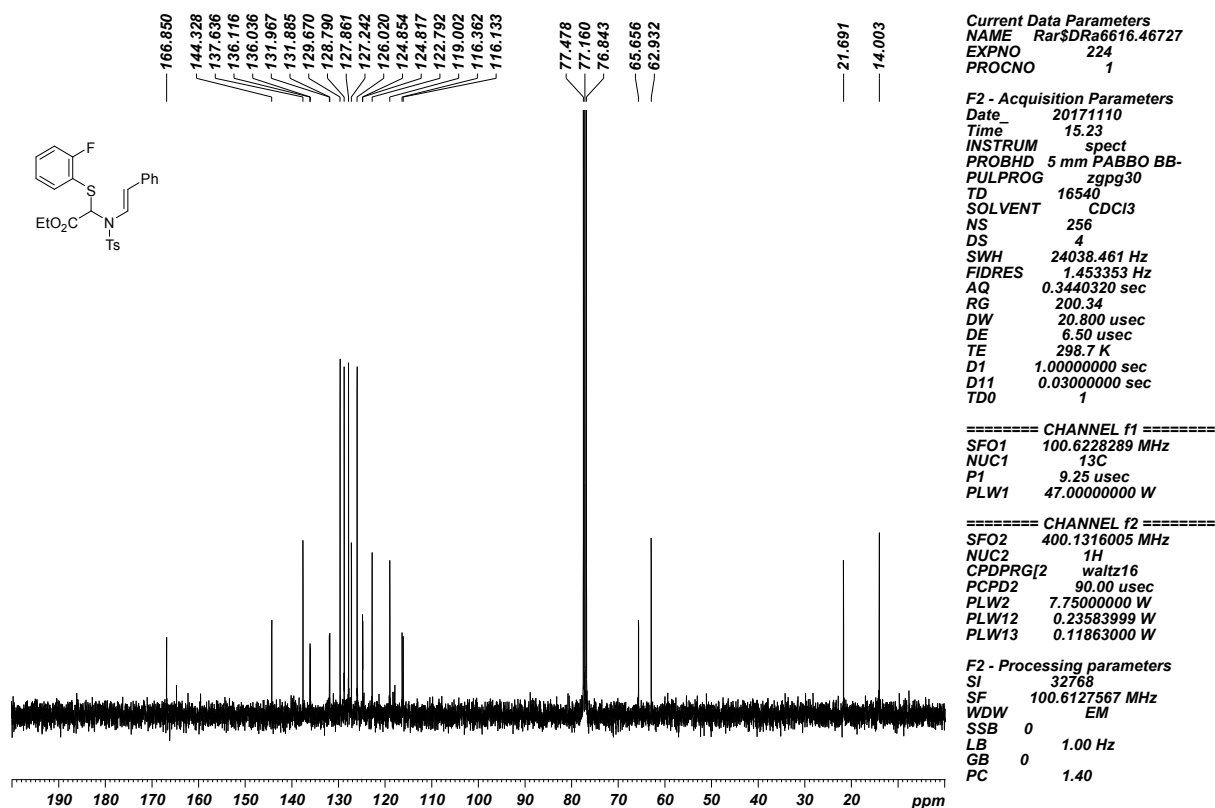


Enamide 4m

¹H NMR (400 MHz, CDCl₃, 24 °C)

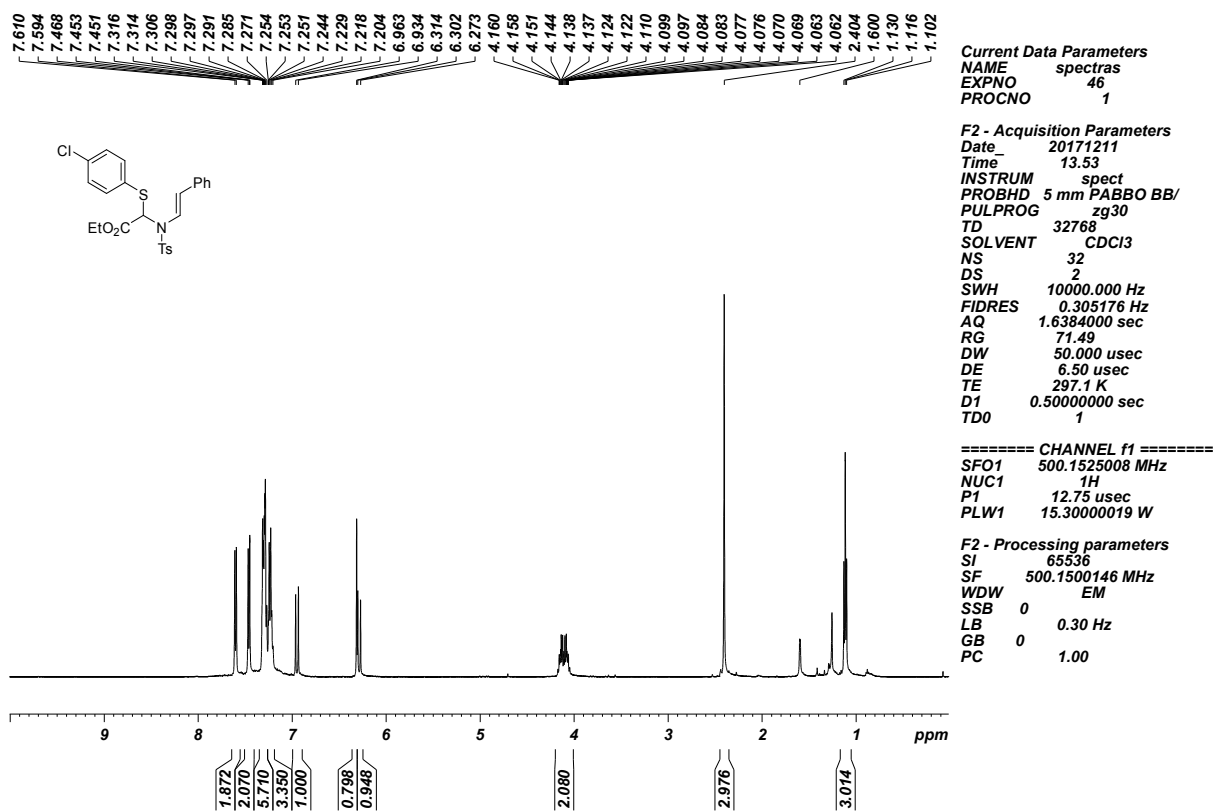


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

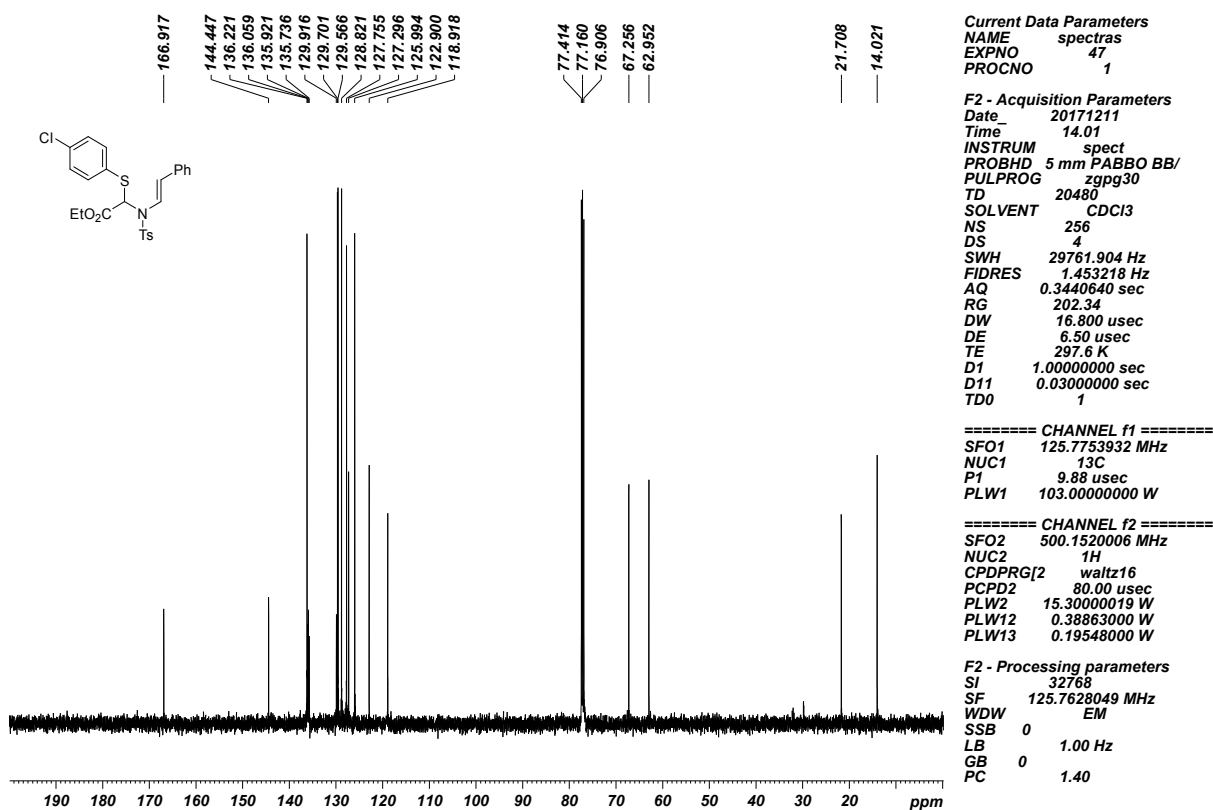


Enamide 4n

^1H NMR (400 MHz, CDCl_3 , 24 °C)

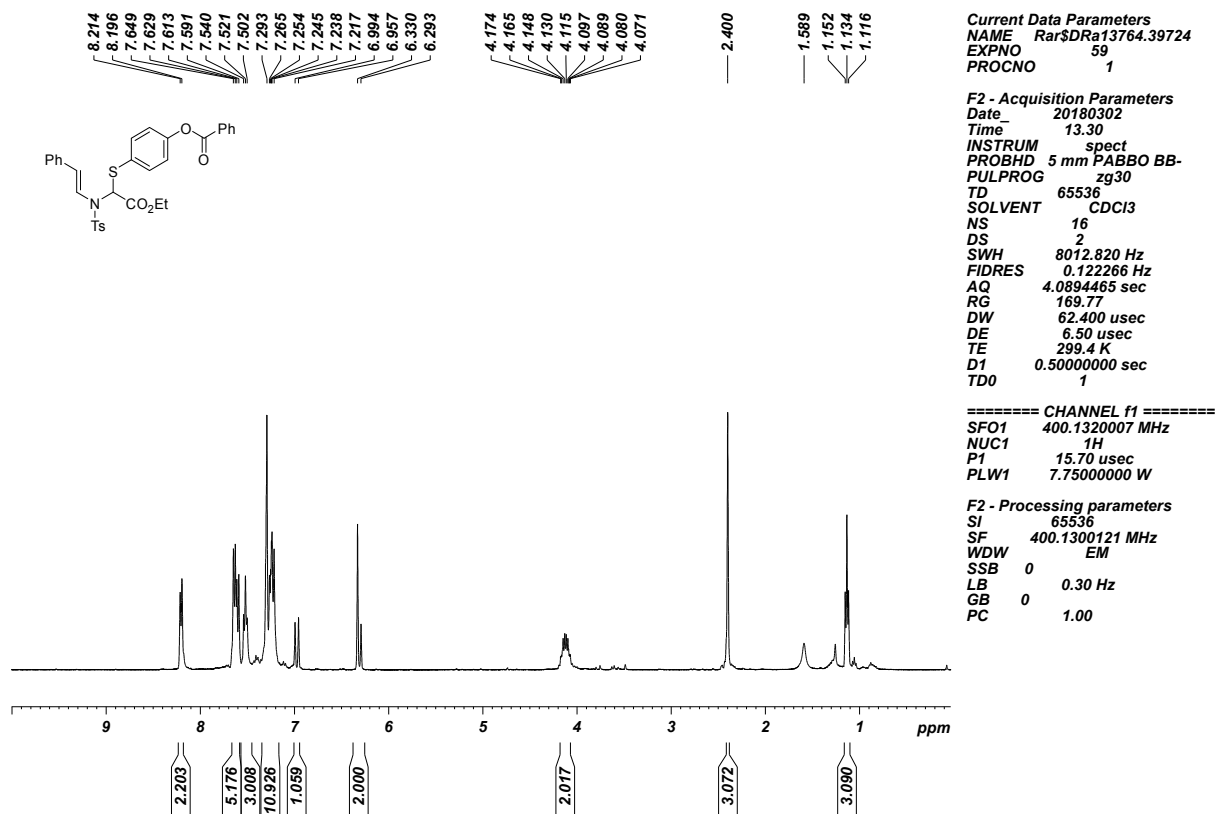


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

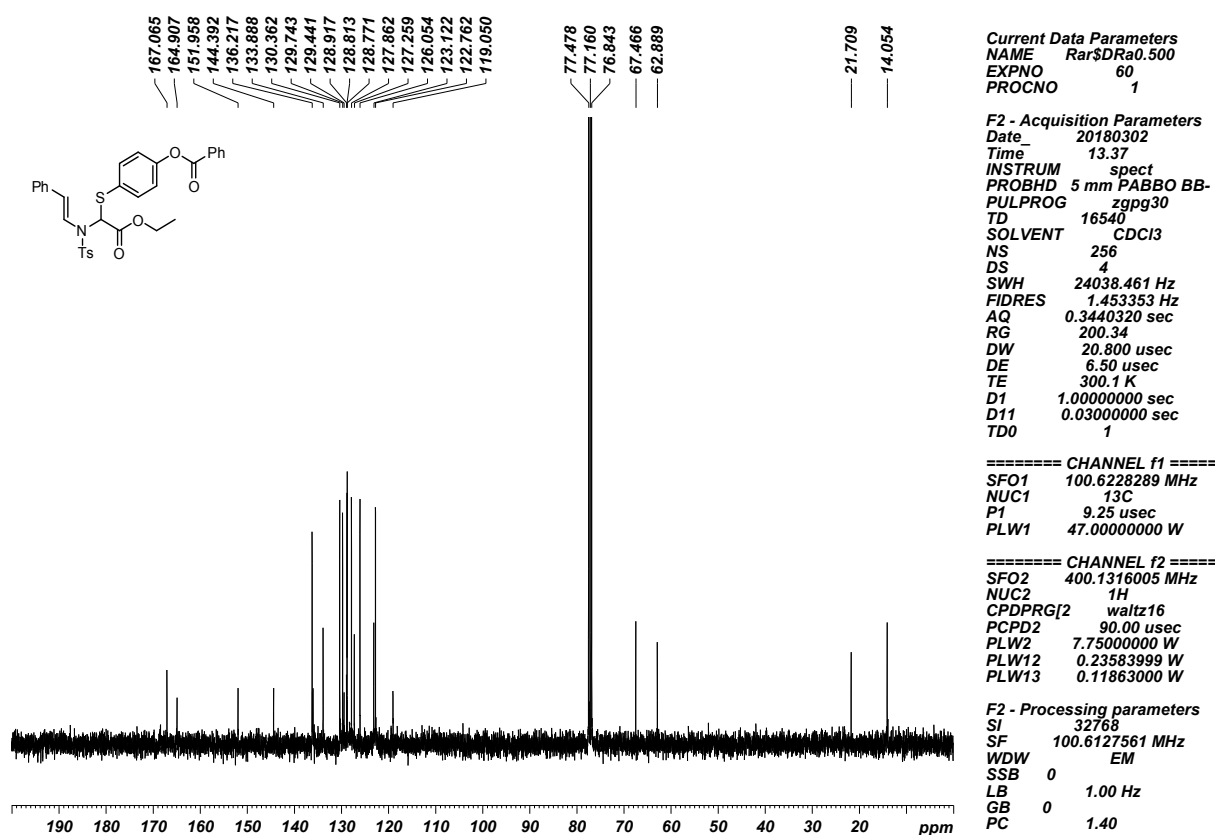


Enamide 4o

^1H NMR (400 MHz, CDCl_3 , 24 °C)

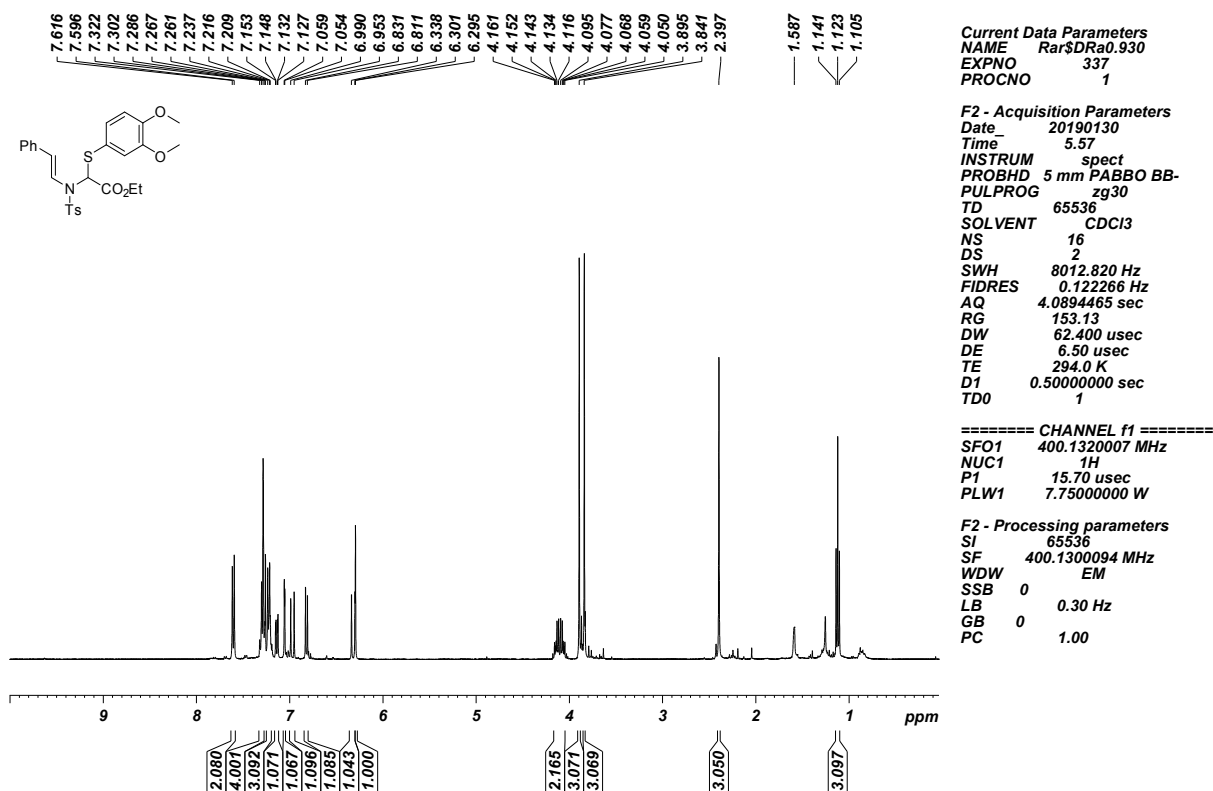


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

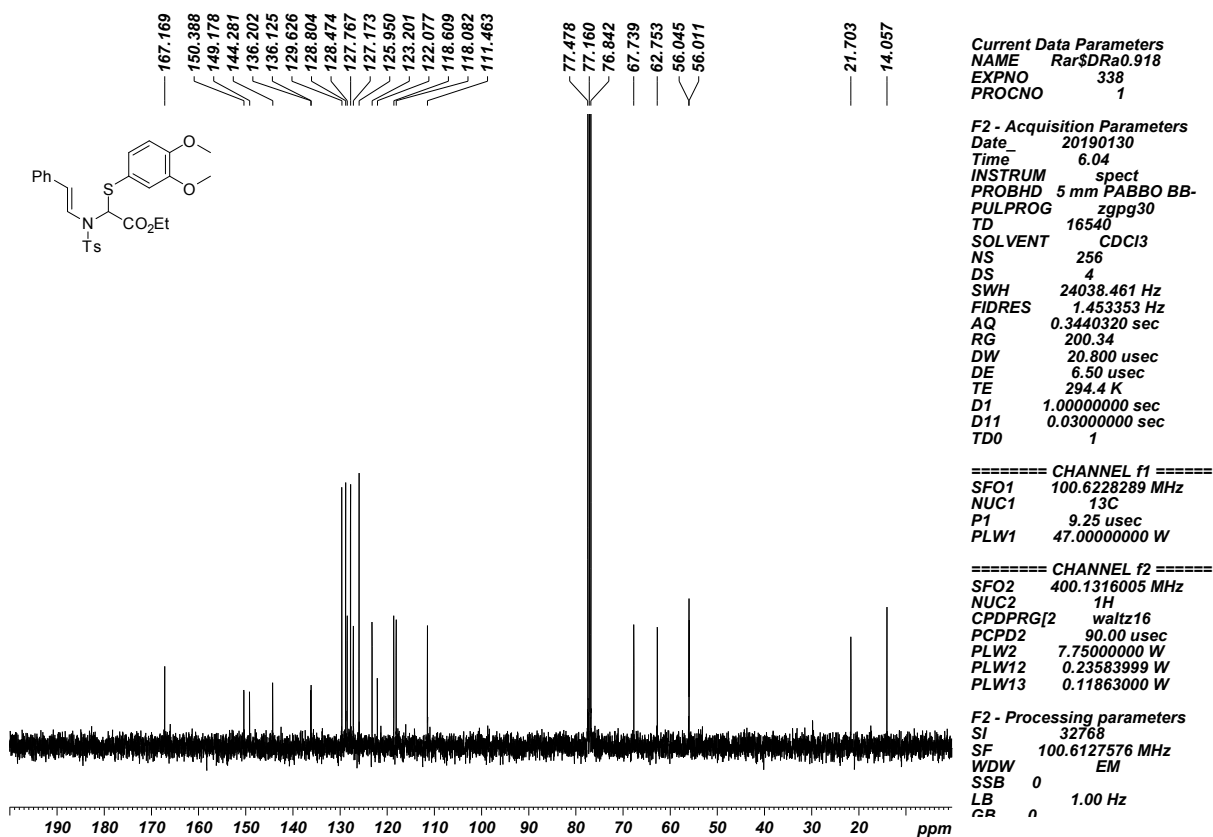


Enamide 4p

¹H NMR (400 MHz, CDCl₃, 24 °C)

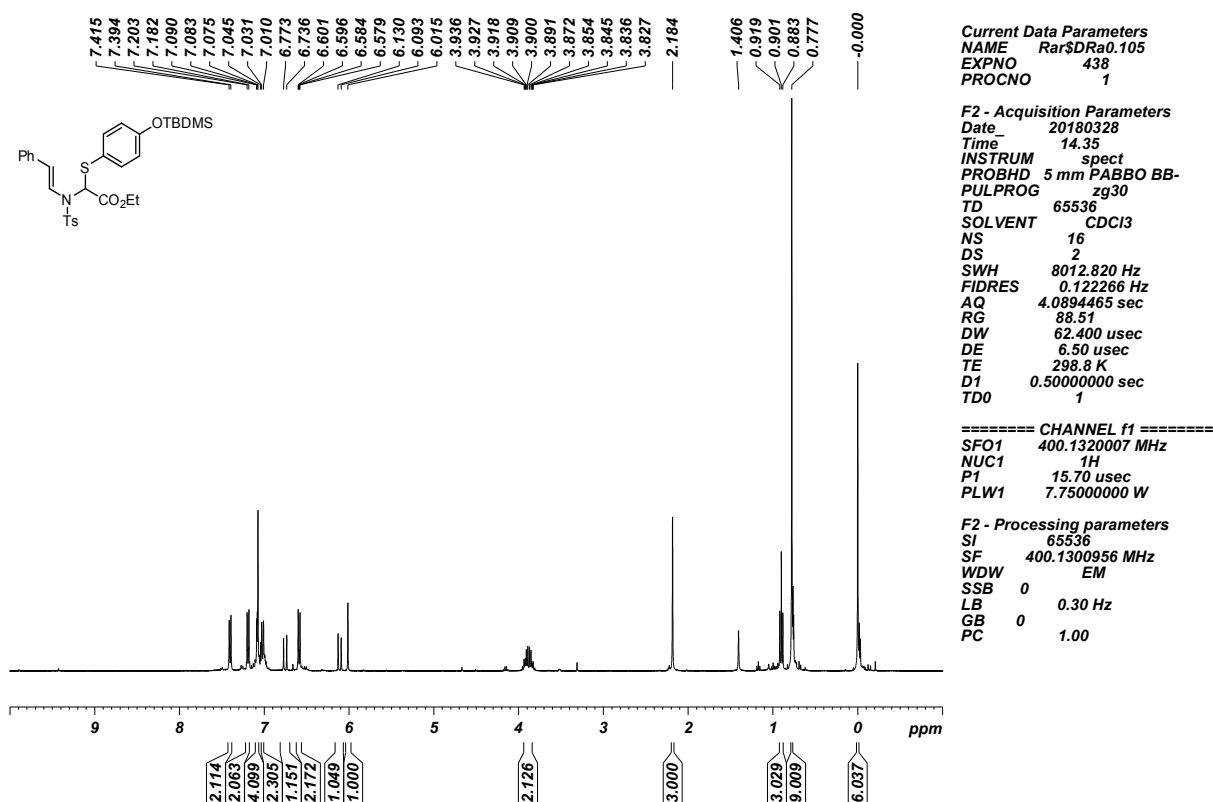


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

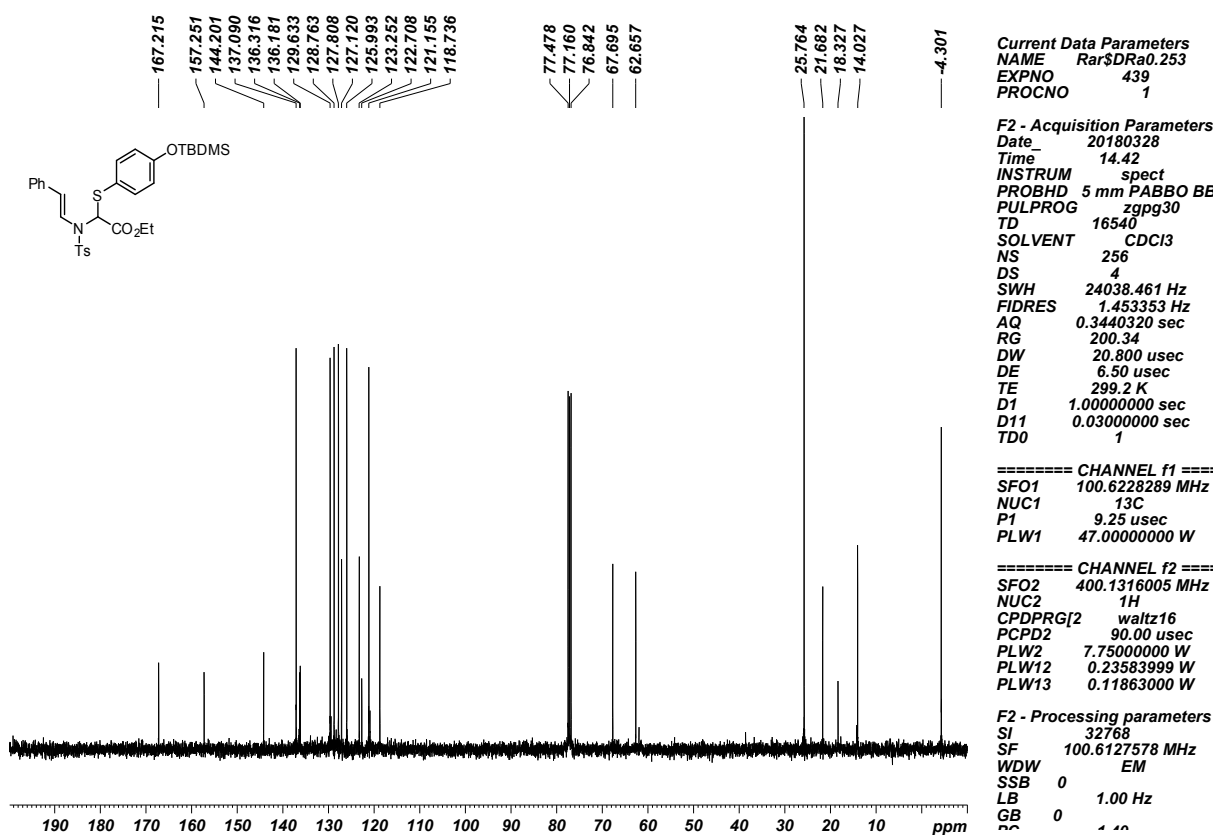


Enamide 4q

^1H NMR (400 MHz, CDCl_3 , 24 °C)

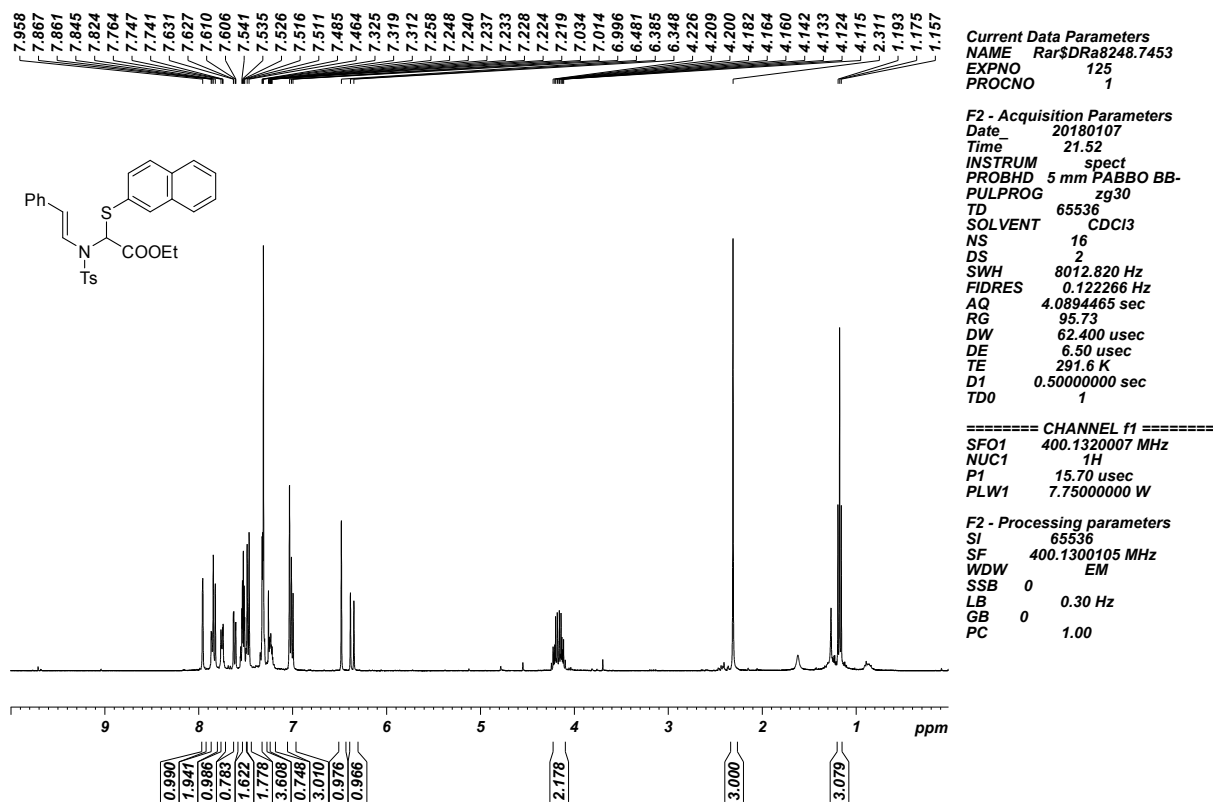


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

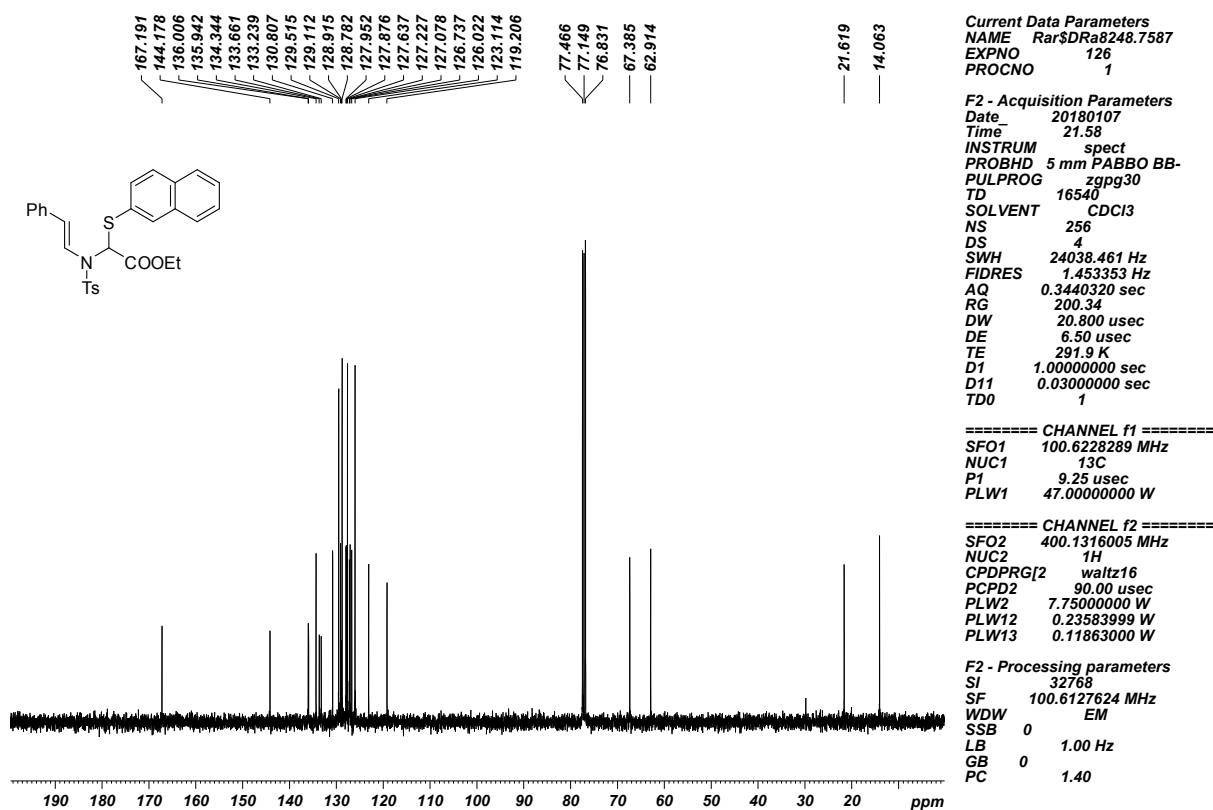


Enamide 4r

¹H NMR (400 MHz, CDCl₃, 24 °C)

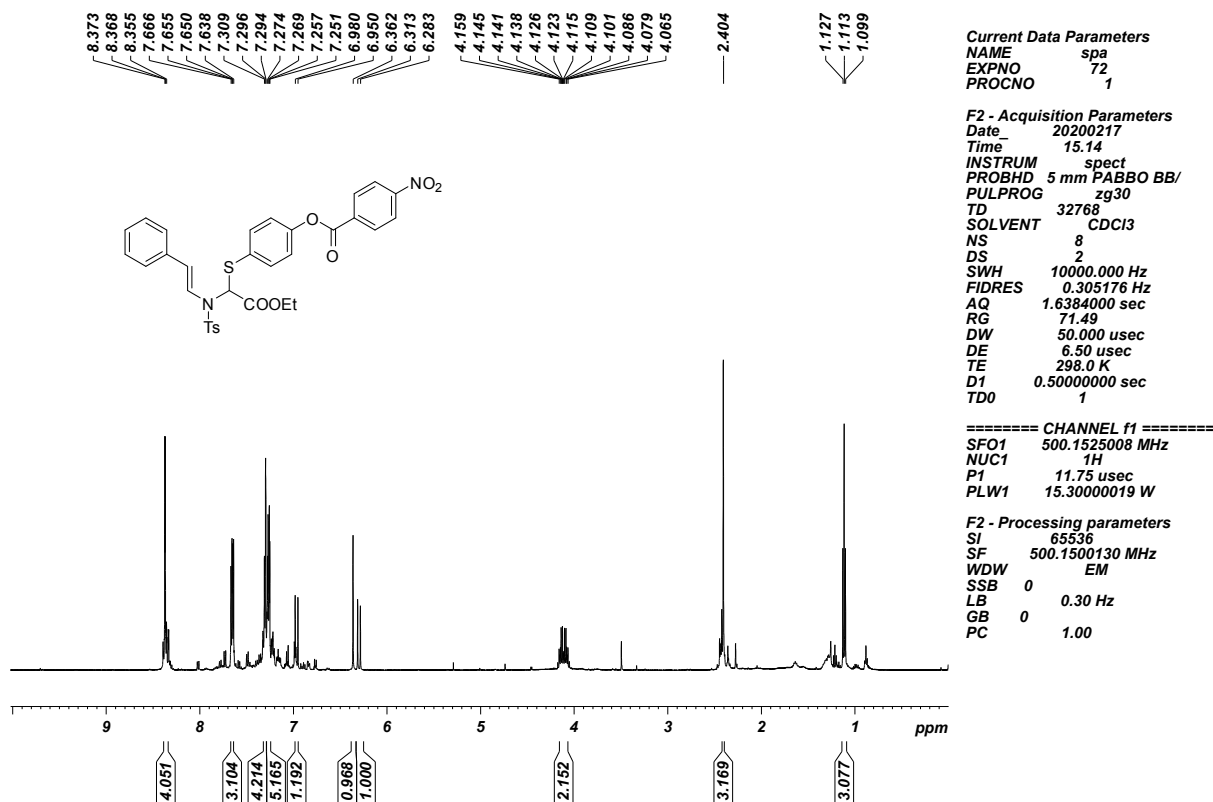


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

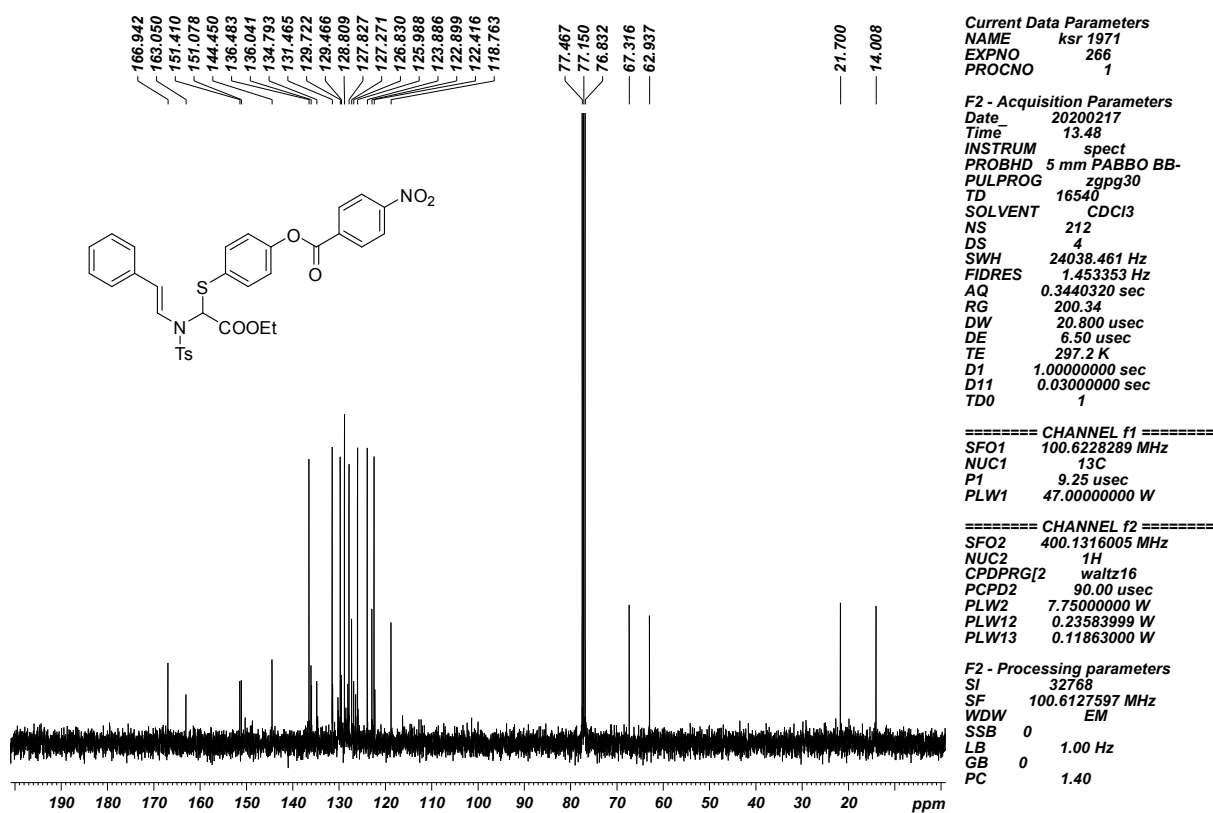


Enamide 4s

¹H NMR (400 MHz, CDCl₃, 24 °C)

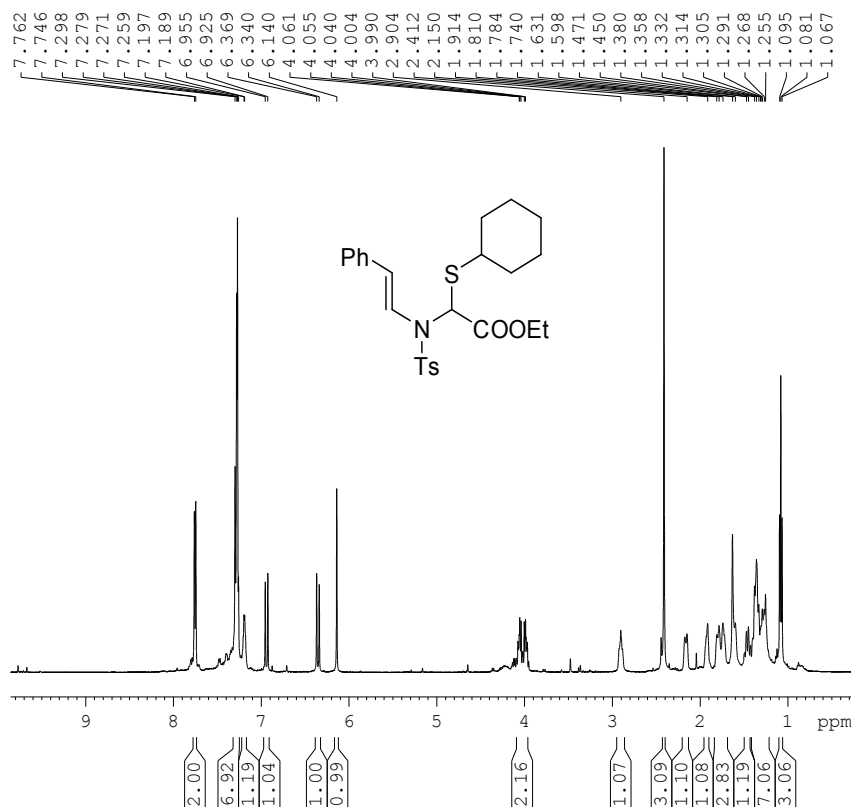


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Enamide 4t

^1H NMR (500 MHz, CDCl_3 , 24 °C)

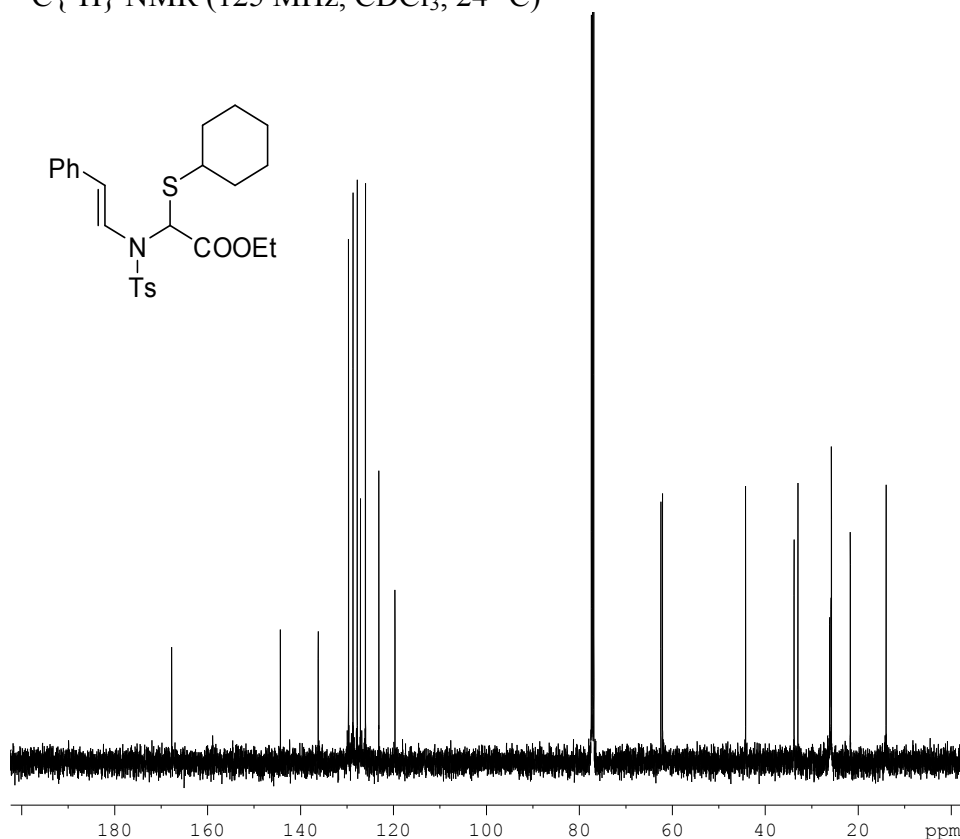


Current Data Parameters
 NAME spa50320
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200302
 Time 9.37
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 32768
 SOLVENT CDCl_3
 NS 15
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.305176 Hz
 AQ 1.6384000 sec
 RG 64
 DW 50.000 usec
 DE 6.50 usec
 TE 298.0 K
 D1 0.50000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 500.1525008 MHz
 NUC1 ^1H
 P1 11.75 usec
 PLW1 15.30000019 W

F2 - Processing parameters
 SI 65536
 SF 500.1500131 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME spa503
 EXPNO
 PROCNO

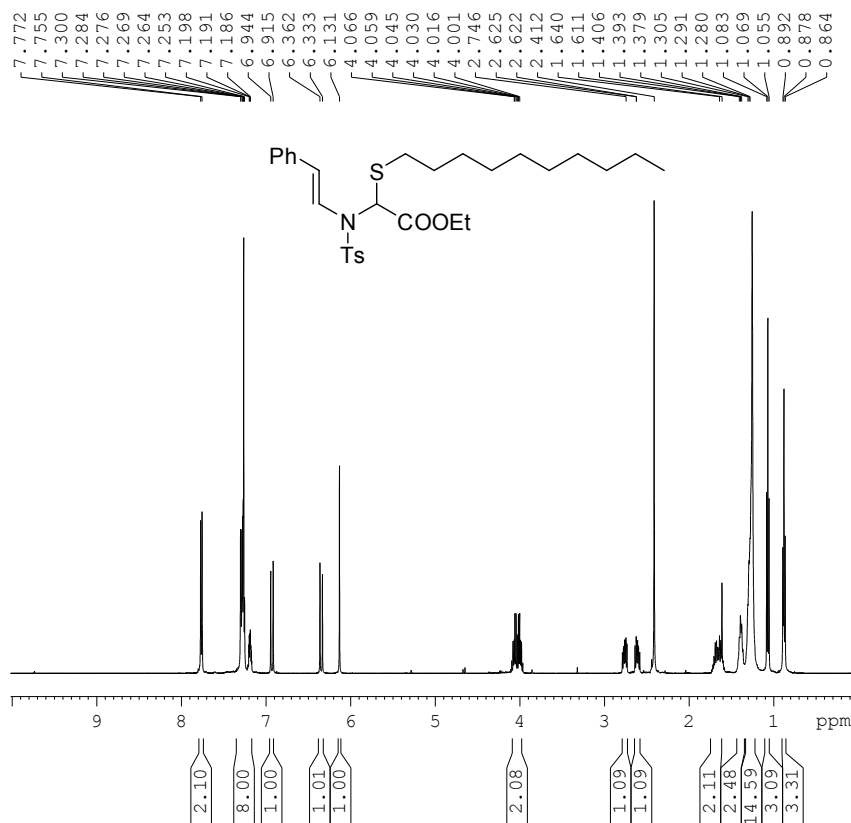
F2 - Acquisition Parameters
 Date_ 202003
 Time 9.
 INSTRUM spe
 PROBHD 5 mm PABBO B
 PULPROG zgpg
 TD 204
 SOLVENT CDCl_3
 NS 1
 DS
 SWH 29761.9
 FIDRES 1.4532
 AQ 0.34406
 RG 202.
 DW 16.8
 DE 6.
 TE 300
 D1 1.000000
 D11 0.030000
 TD0

===== CHANNEL f1 =
 SFO1 125.77539
 NUC1 ^{13}C
 P1 10.
 PLW1 103.000000

===== CHANNEL f2 =
 SFO2 500.15200
 NUC2 ^1H
 CPDPRG[2] waltz
 PCPD2 80.
 D11 15.300000

Enamide 4u

^1H NMR (500 MHz, CDCl_3 , 24 °C)



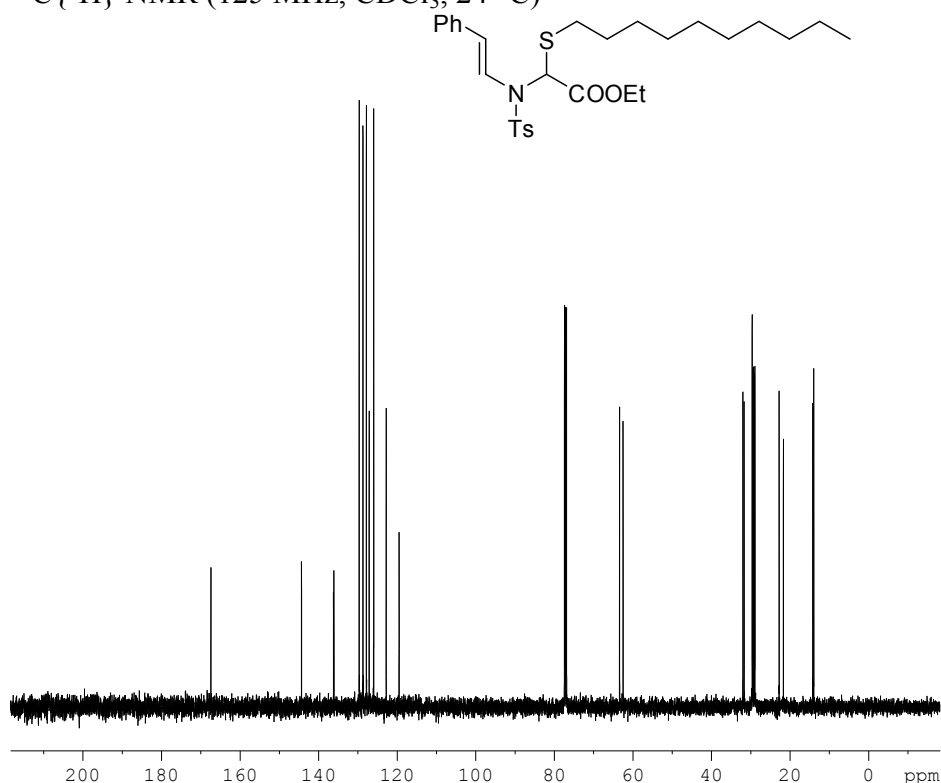
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 NAME spa50320
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200303
 Time_ 15.03
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.305176 Hz
 AQ 1.6384000 sec
 RG 31.24
 DW 50.000 usec
 DE 6.50 usec
 TE 298.0 K
 D1 0.50000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 500.1525008 MHz
 NUC1 1H
 P1 11.75 usec
 PLW1 15.30000019 W

F2 - Processing parameters
 SI 65536
 SF 500.1500146 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C{¹H} NMR (125 MHz, CDCl₃, 24 °C)



Current Data Parameters
 NAME spa503
 EXPNO
 PROCNO

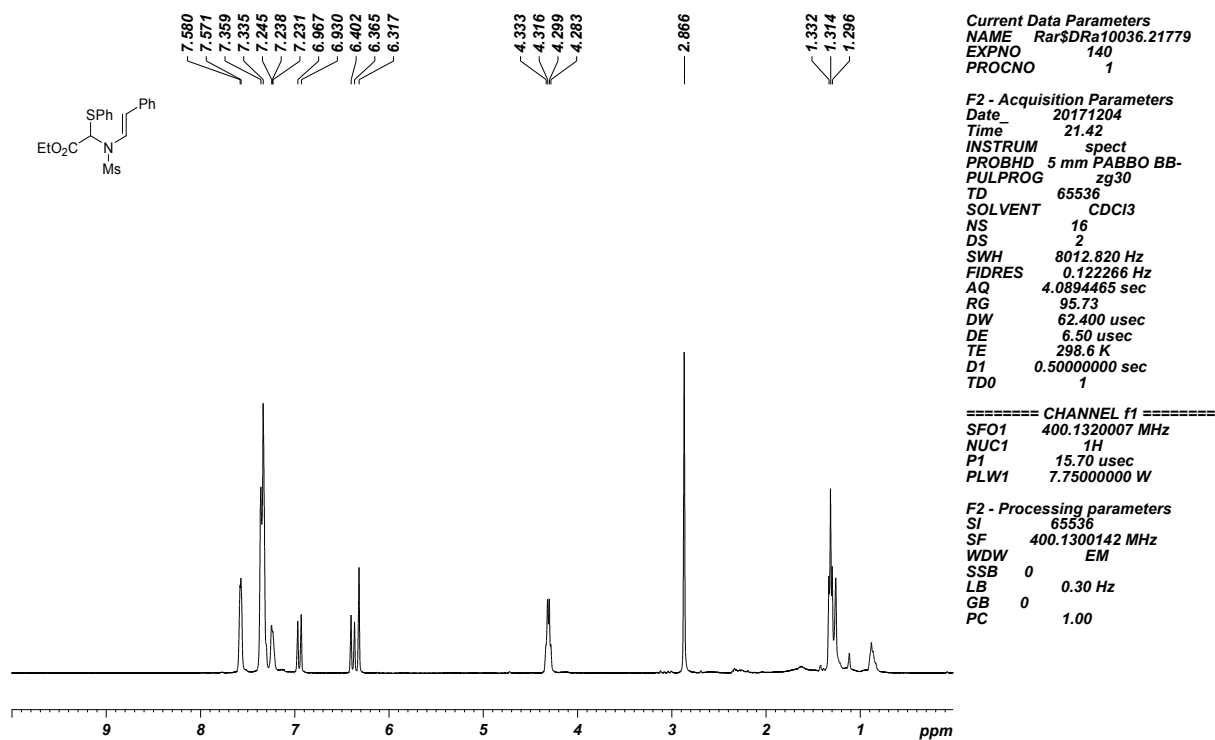
F2 - Acquisition Parameters
 Date_ 202003
 Time_ 15.
 INSTRUM spe
 PROBHD 5 mm PABBO B
 PULPROG zgpg
 TD 204
 SOLVENT CDC
 NS 2
 DS
 SWH 29761.9
 FIDRES 1.4532
 AQ 0.34406
 RG 202.
 DW 16.8
 DE 6.
 TE 300
 D1 1.000000
 D11 0.030000
 TD0

===== CHANNEL f1 =
 SFO1 125.77539
 NUC1 1
 P1 10.
 PLW1 103.000000

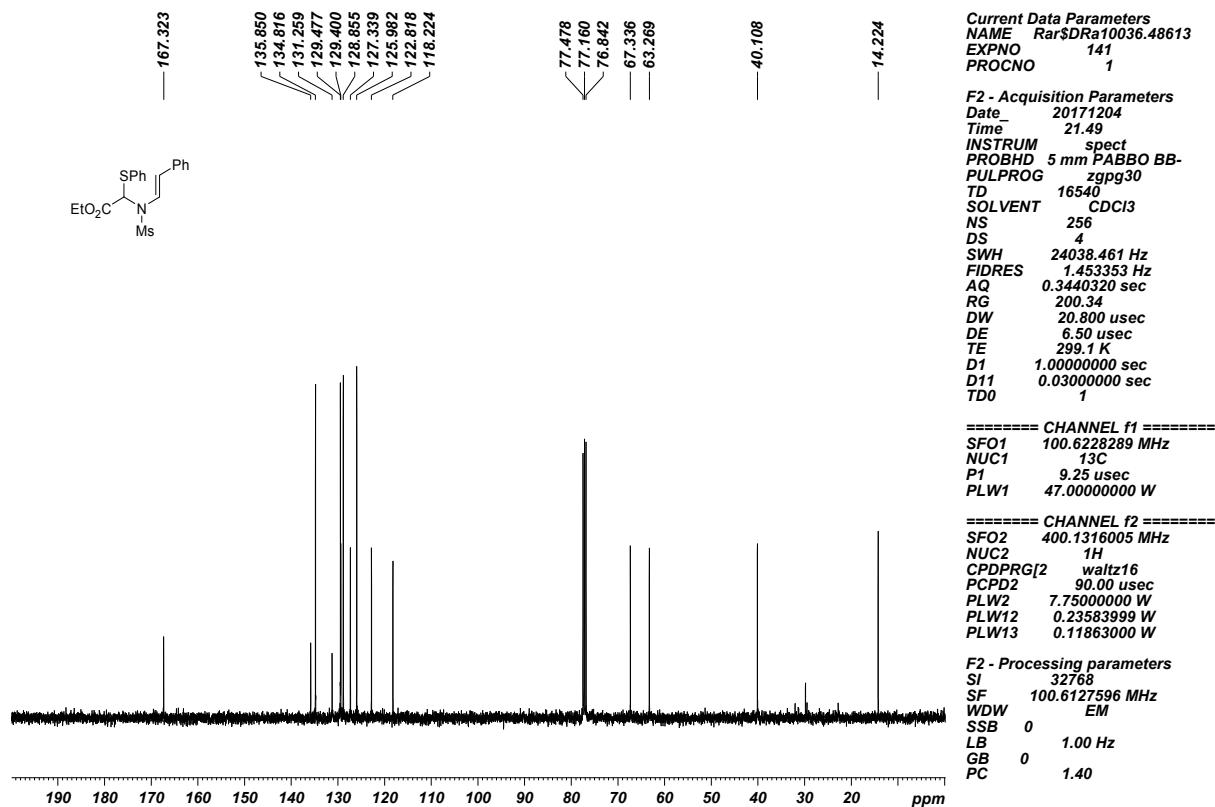
===== CHANNEL f2 =
 SFO2 500.15200
 NUC2
 CPDPRG[2] waltz
 PCPD2 80.
 RTM2 15.300000

Enamide 4v

¹H NMR (400 MHz, CDCl₃, 24 °C)

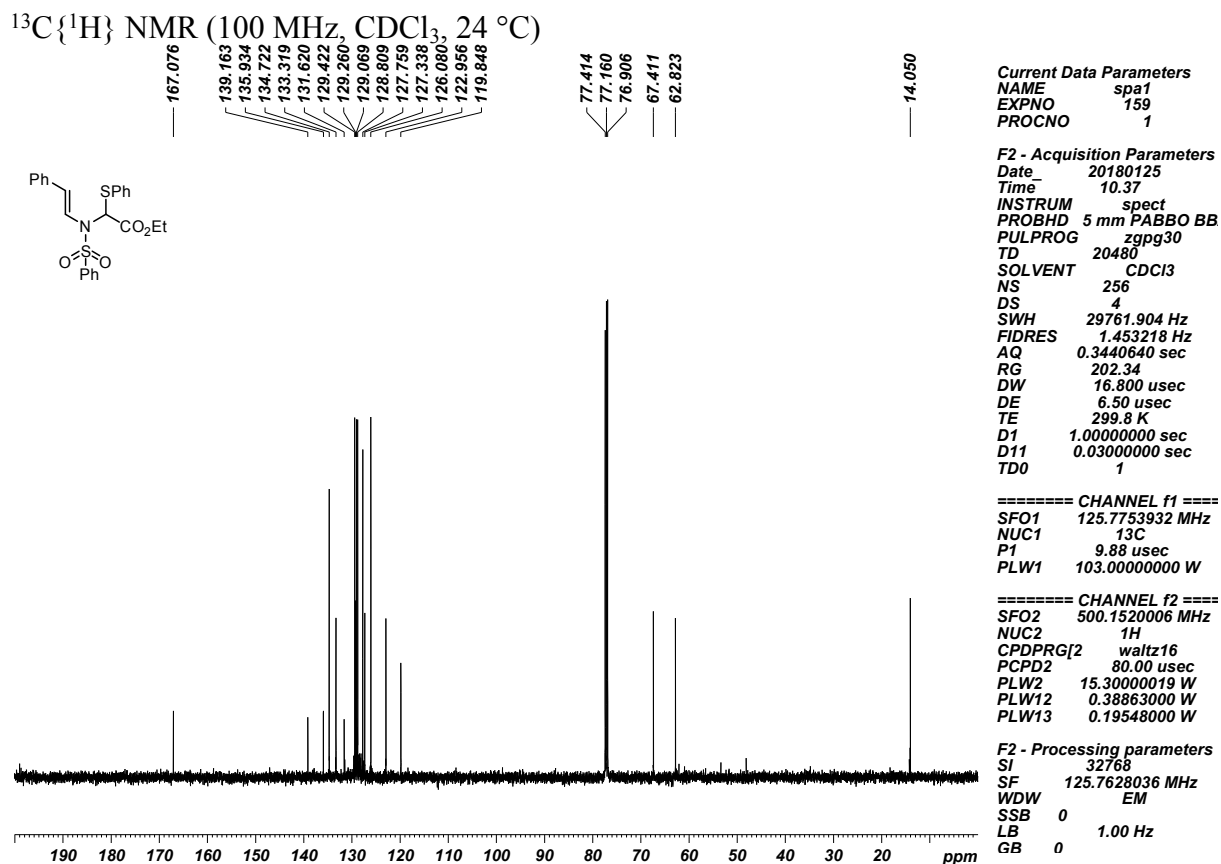
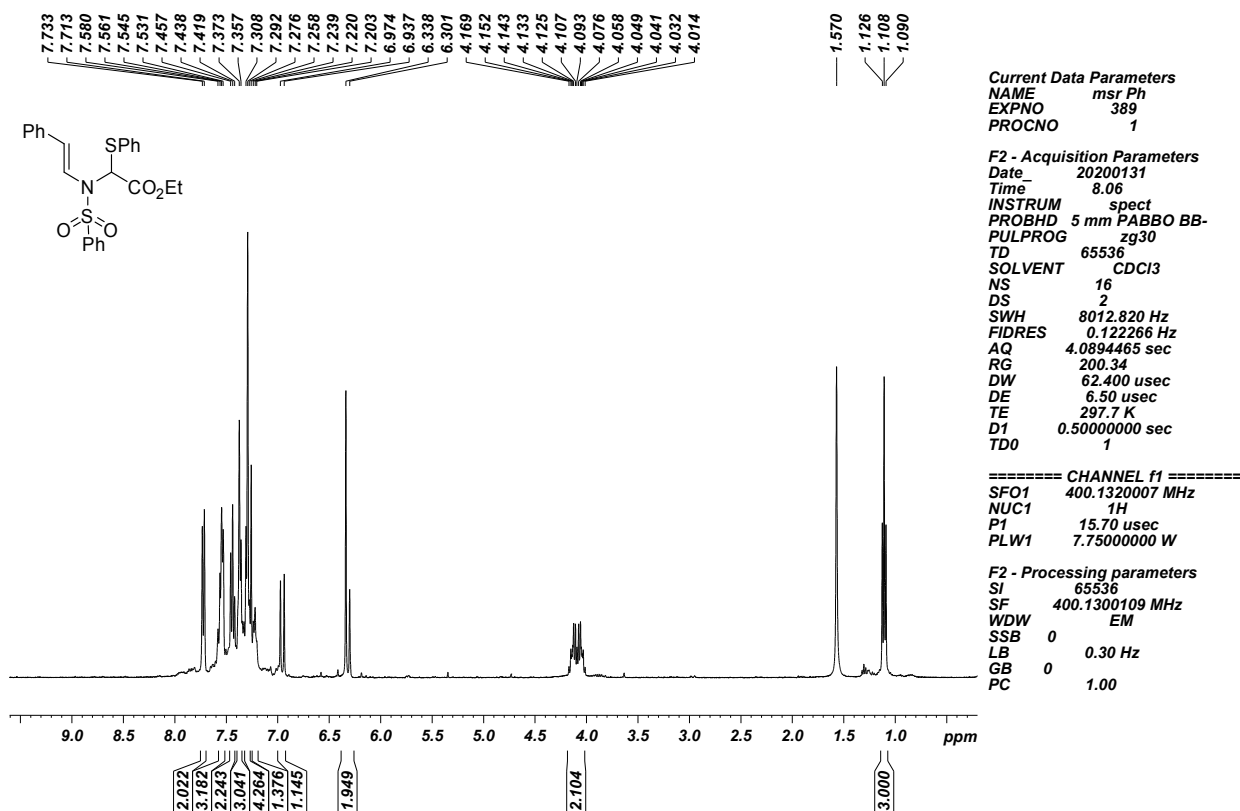


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)



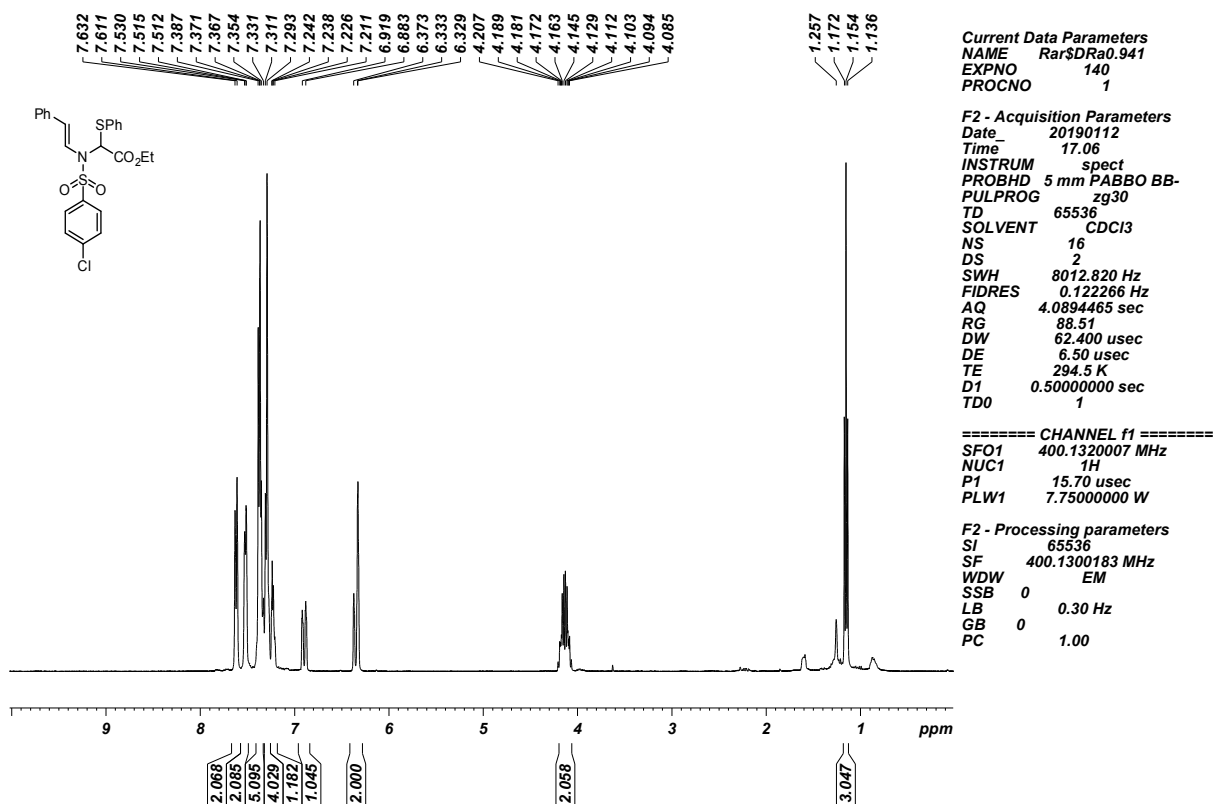
Enamide 4w

¹H NMR (400 MHz, CDCl₃, 24 °C)

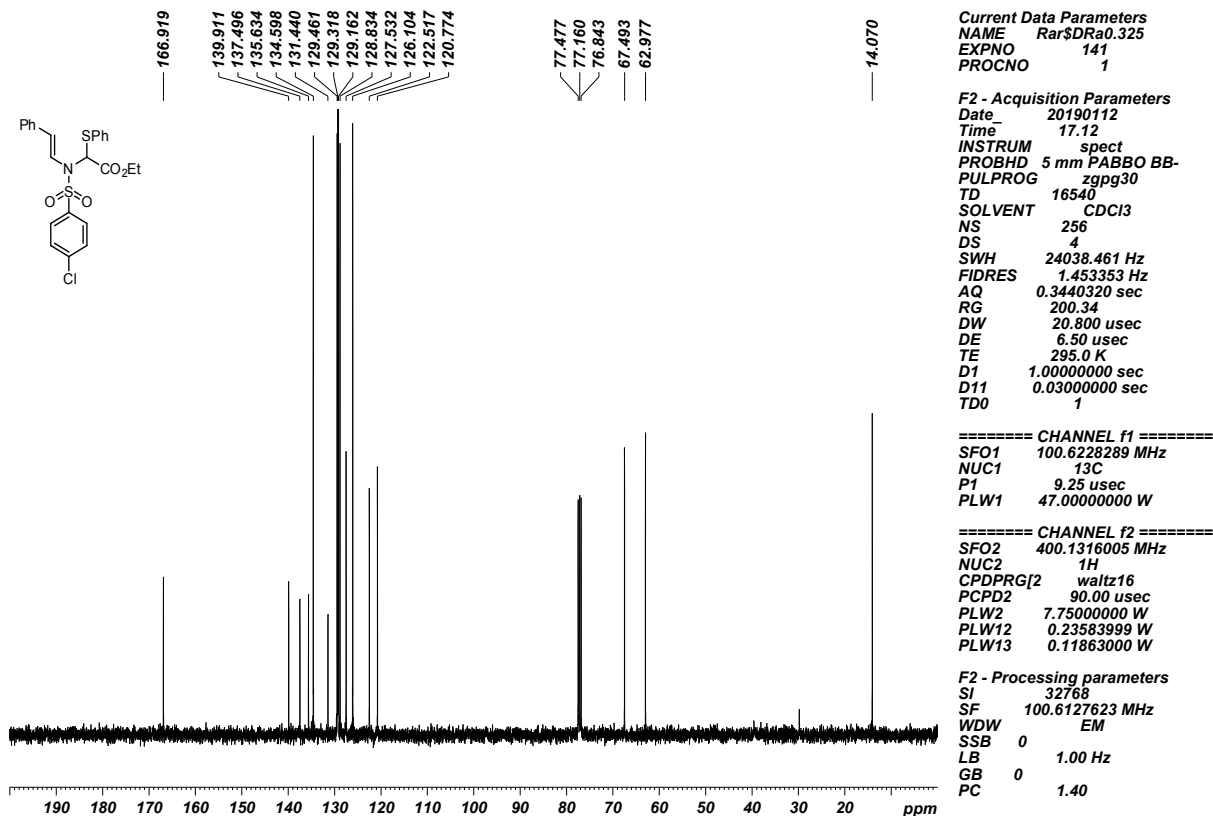


Enamide 4x

¹H NMR (400 MHz, CDCl₃, 24 °C)

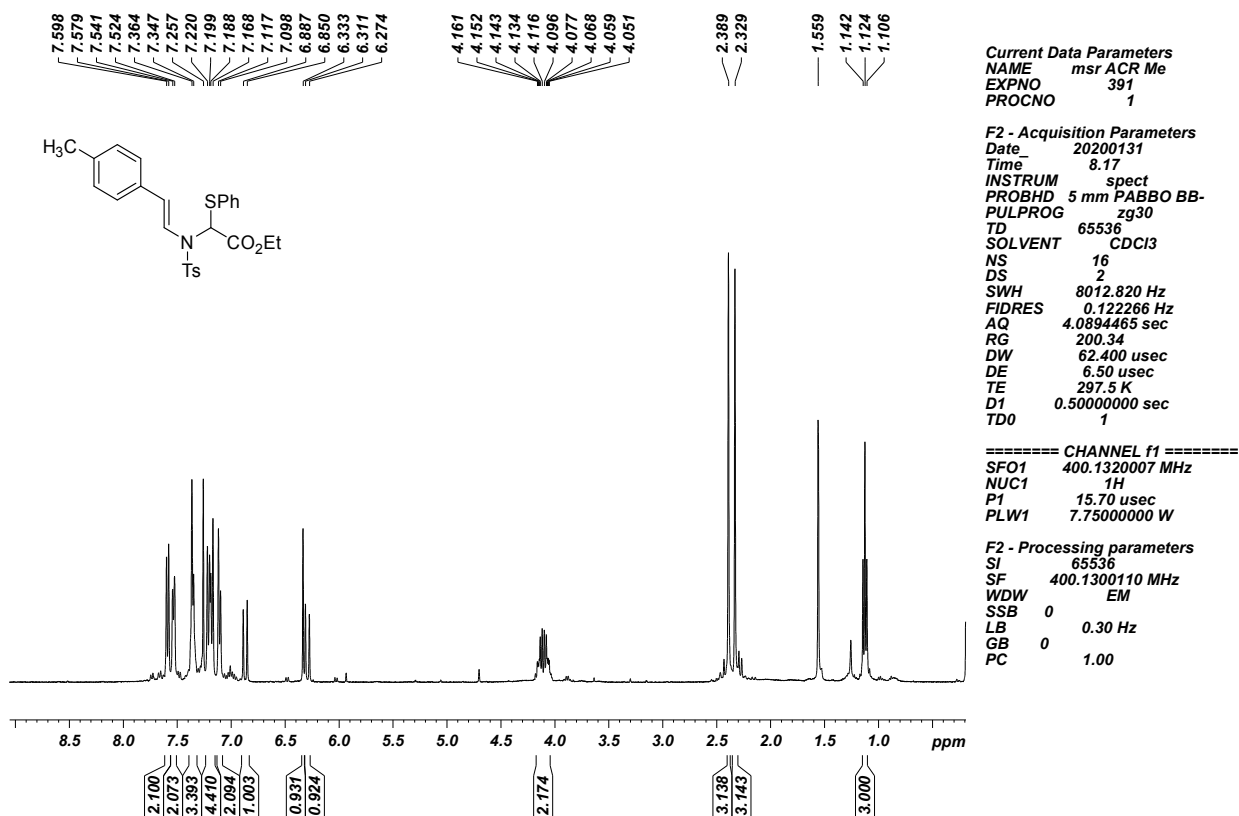


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

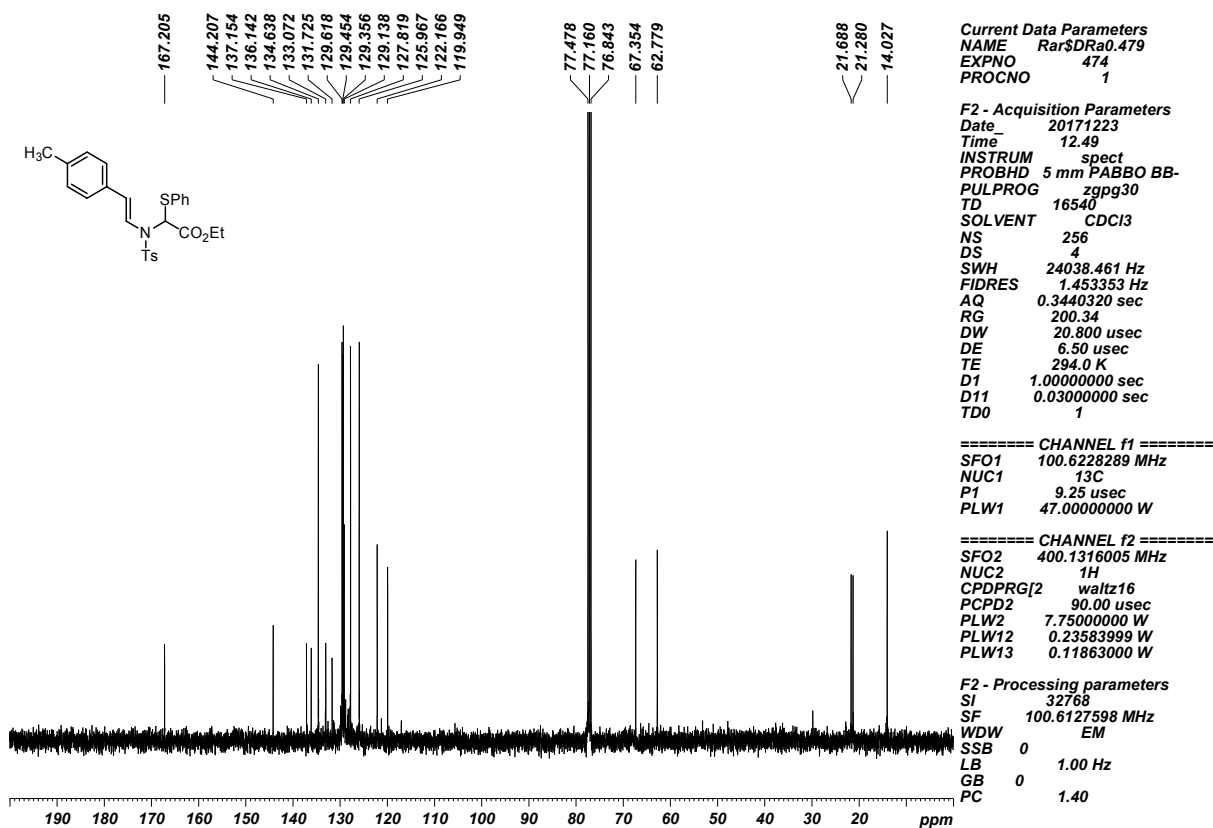


Enamide 4y

¹H NMR (400 MHz, CDCl₃, 24 °C)

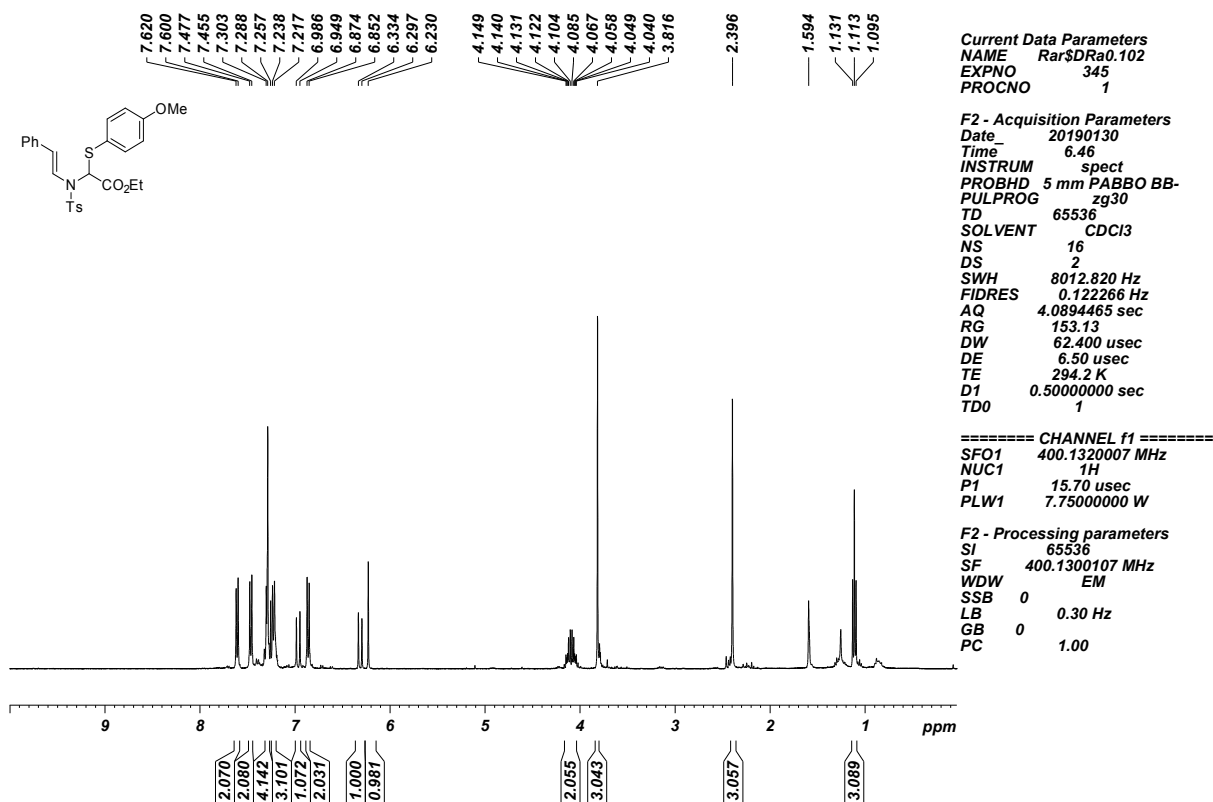


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

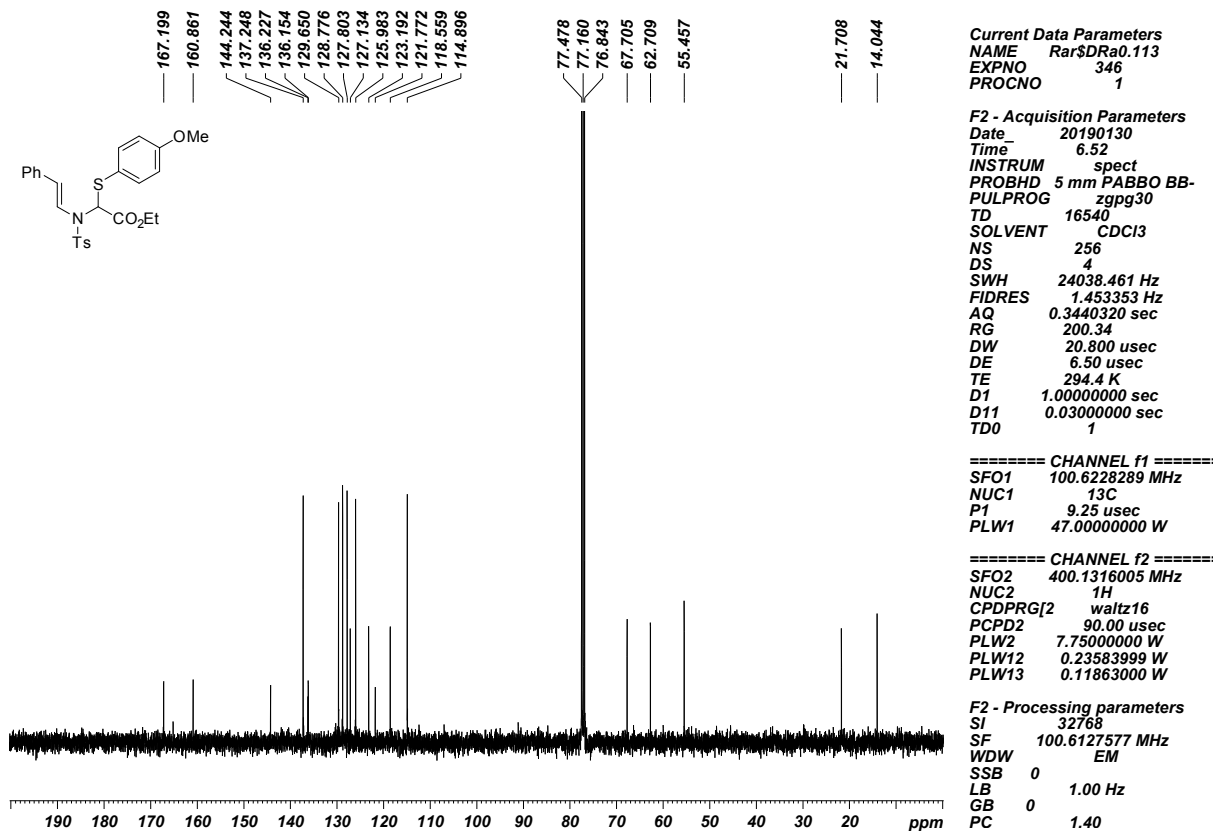


Enamide 4z

¹H NMR (400 MHz, CDCl₃, 24 °C)

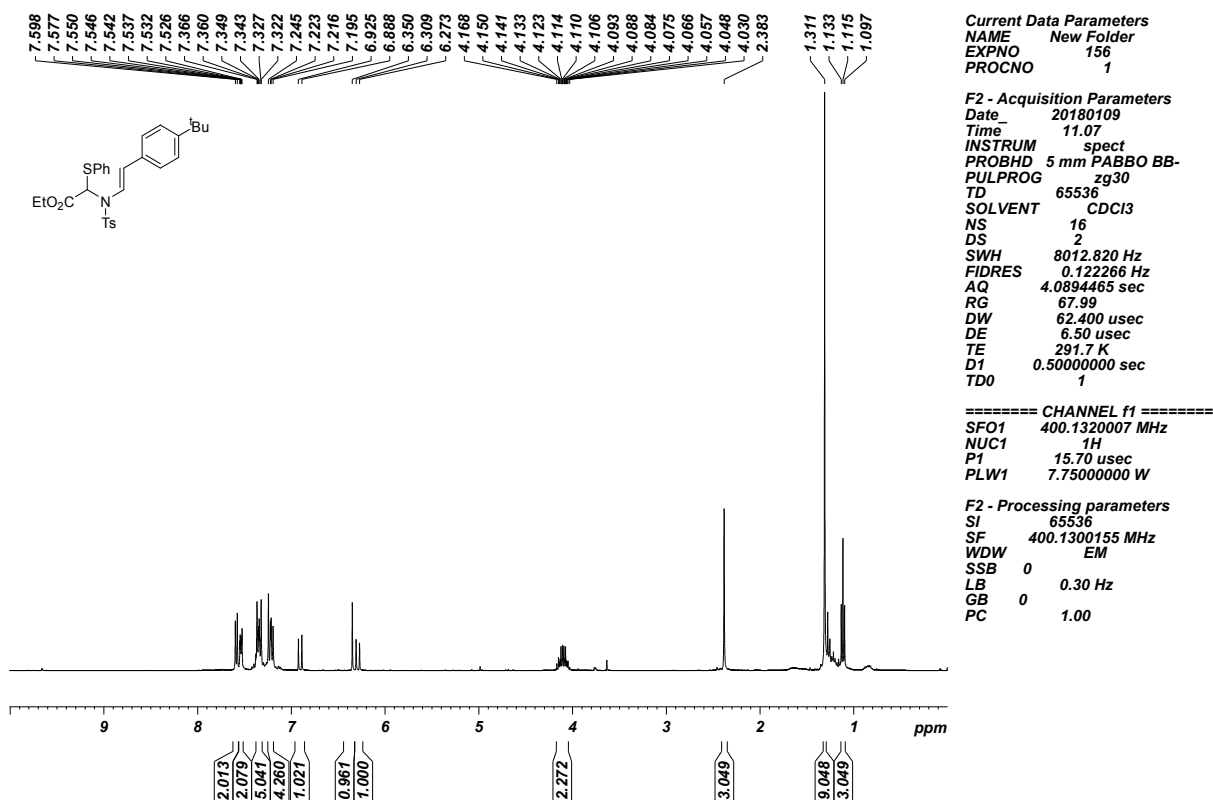


¹³C{¹H} NMR (100 MHz, CDCl₃, 24 °C)

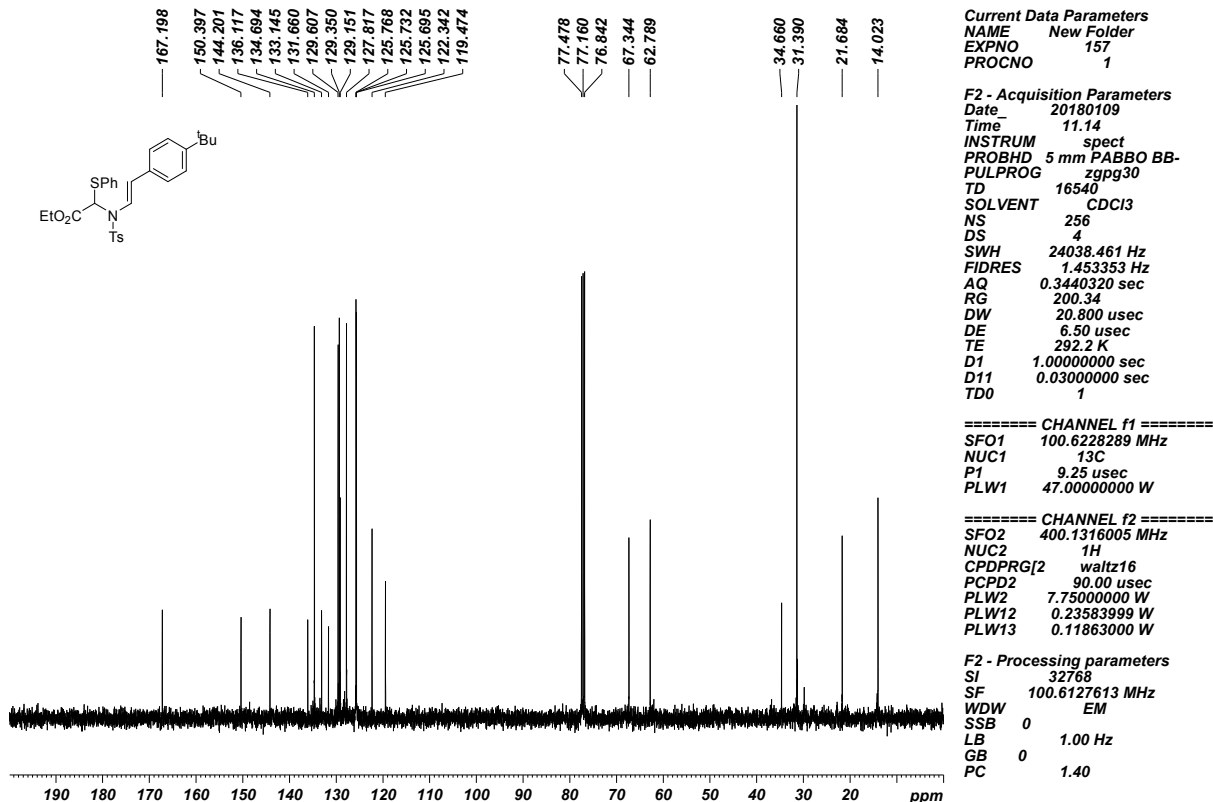


Enamide 4aa

¹H NMR (400 MHz, CDCl₃, 24 °C)

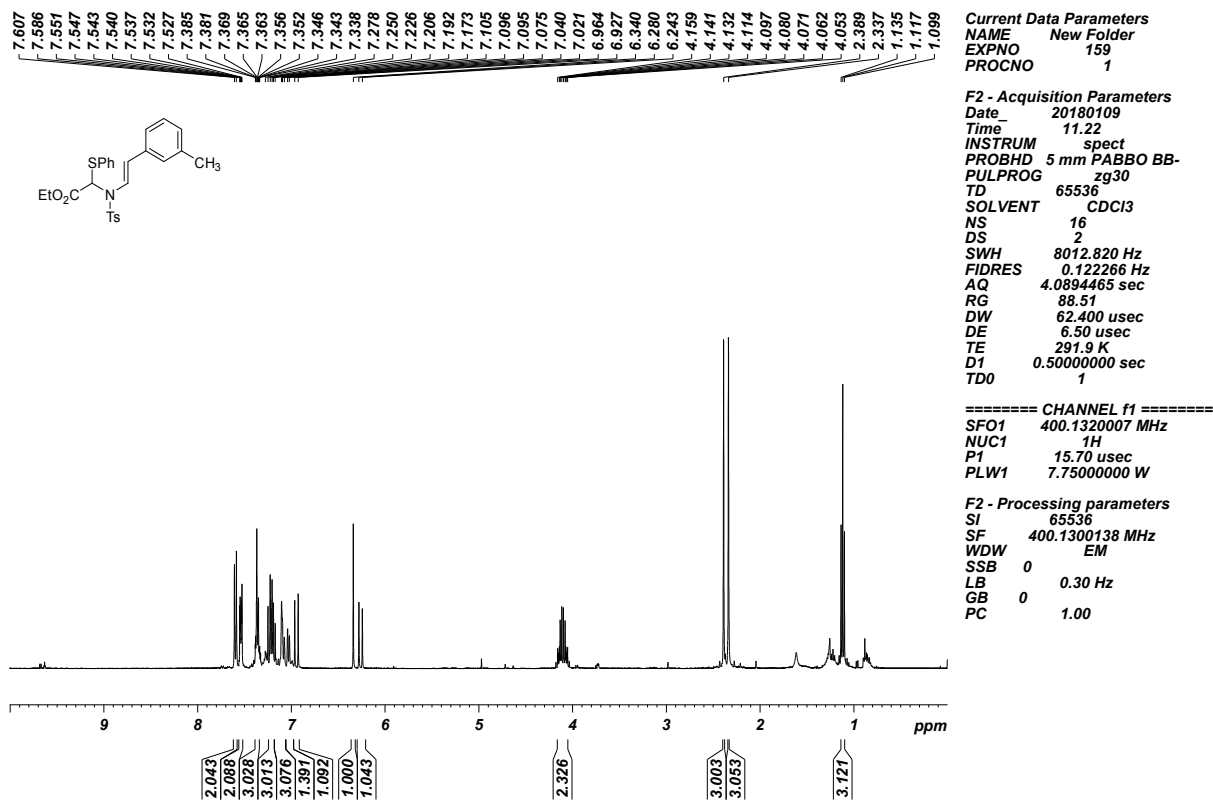


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

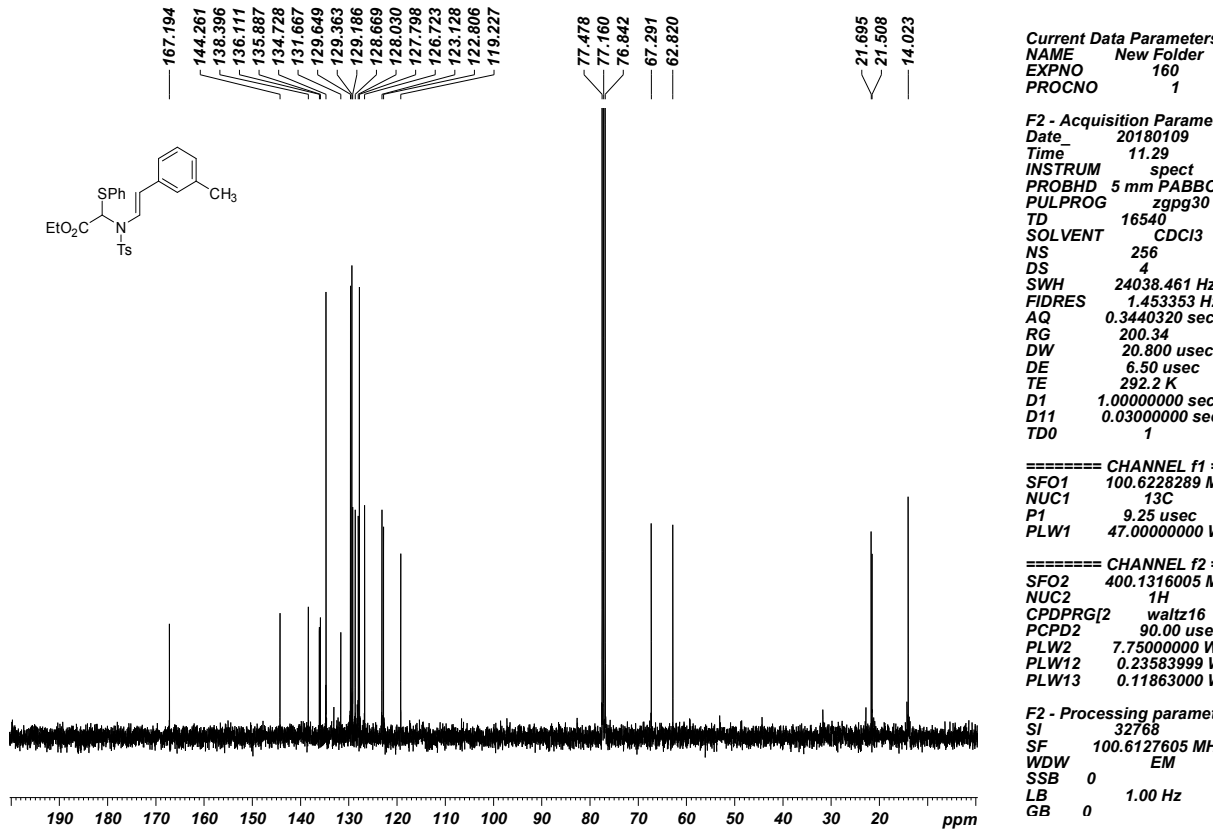


Enamide 4ab

^1H NMR (400 MHz, CDCl_3 , 24 °C)

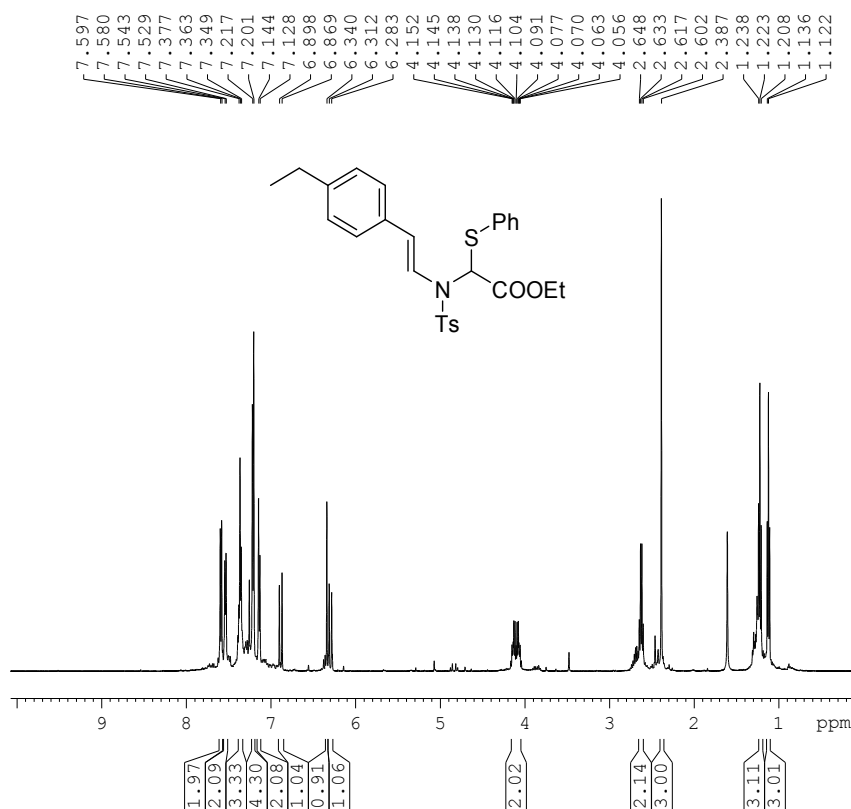


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Enamide 4ac

^1H NMR (400 MHz, CDCl_3 , 24 °C)



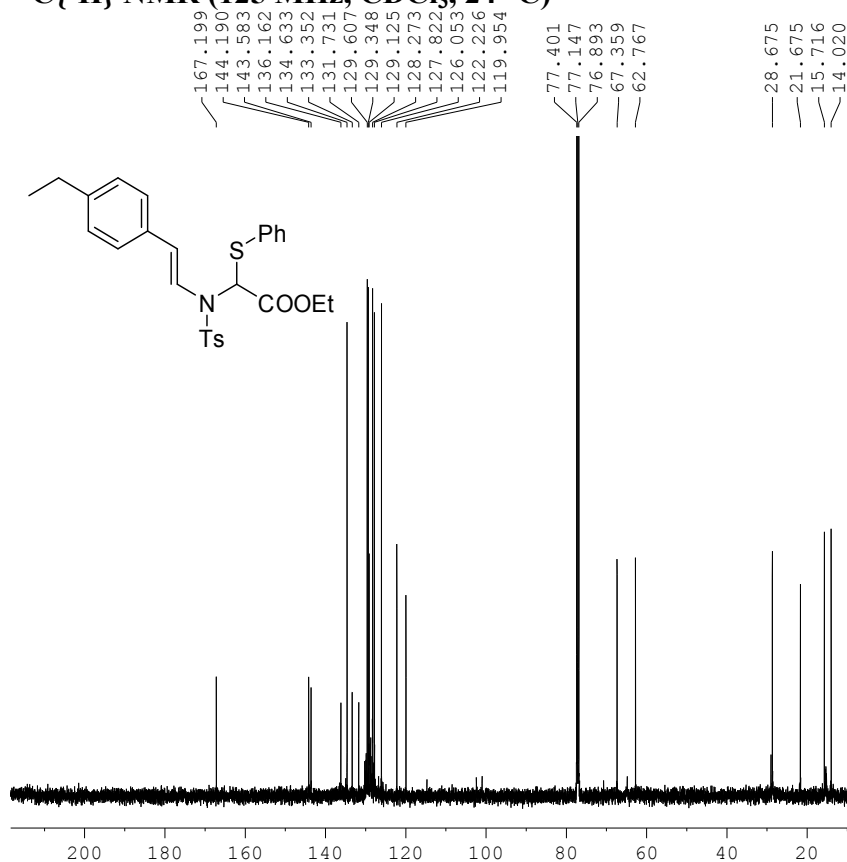
Current Data Parameters
 NAME spa50220
 EXPNO 148
 PROCNO 1

F2 - Acquisition Parameters
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 Time_ 6.06
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 32
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.305176 Hz
 AQ 1.6384000 sec
 RG 79.04
 DW 50.000 usec
 DE 6.50 usec
 TE 298.0 K
 D1 0.50000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 500.1525008 MHz
 NUC1 1H
 P1 11.75 usec
 PLW1 15.30000019 W

F2 - Processing parameters
 SI 65536
 SF 500.1500156 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C{¹H} NMR (125 MHz, CDCl₃, 24 °C)



Current Data Parameters
 NAME spa502
 EXPNO 1
 PROCNO

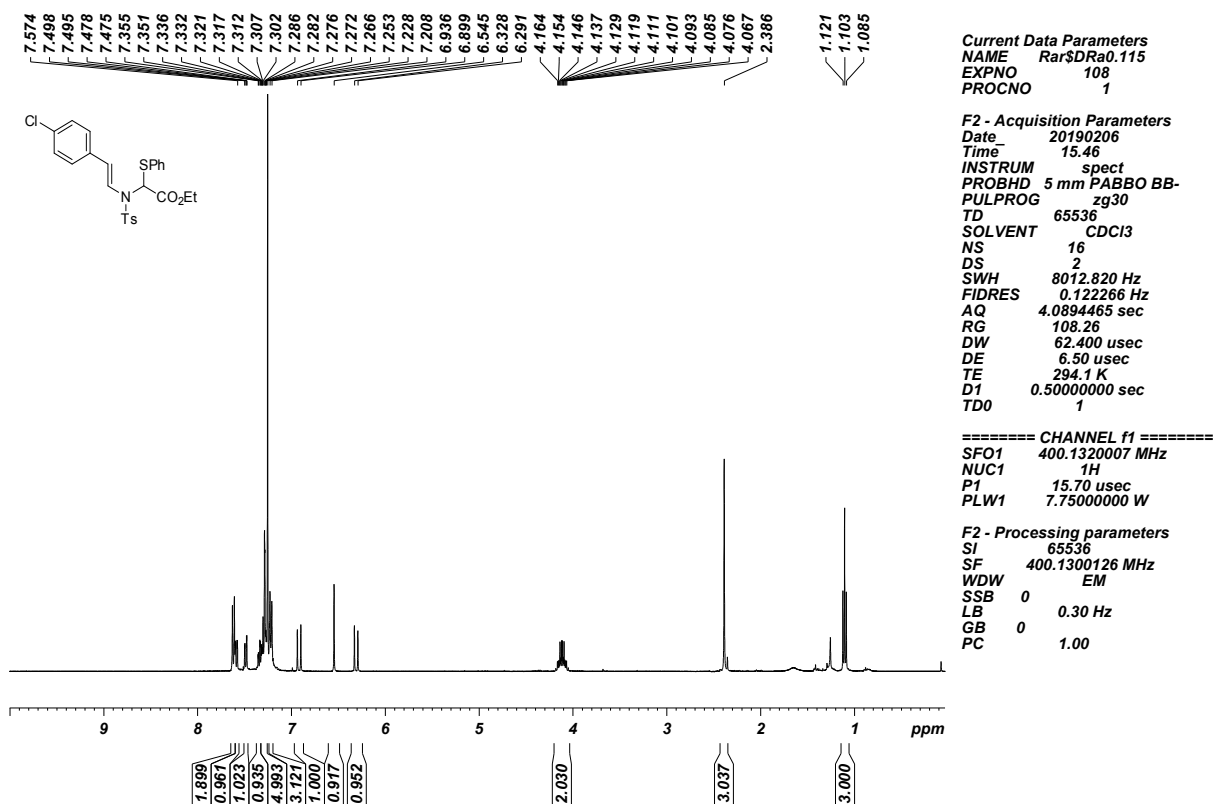
F2 - Acquisition Parameters
 Date_ 202002
 Time_ 6.
 INSTRUM spe
 PROBHD 5 mm PABBO B
 PULPROG zgpg
 TD 204
 SOLVENT CDC
 NS 5
 DS
 SWH 29761.9
 FIDRES 1.4532
 AQ 0.34406
 RG 202.
 DW 16.8
 DE 6.
 TE 300
 D1 1.000000
 D11 0.030000
 TD0

===== CHANNEL f1 =
 SFO1 125.77539
 NUC1 1
 P1 10.
 PLW1 103.000000

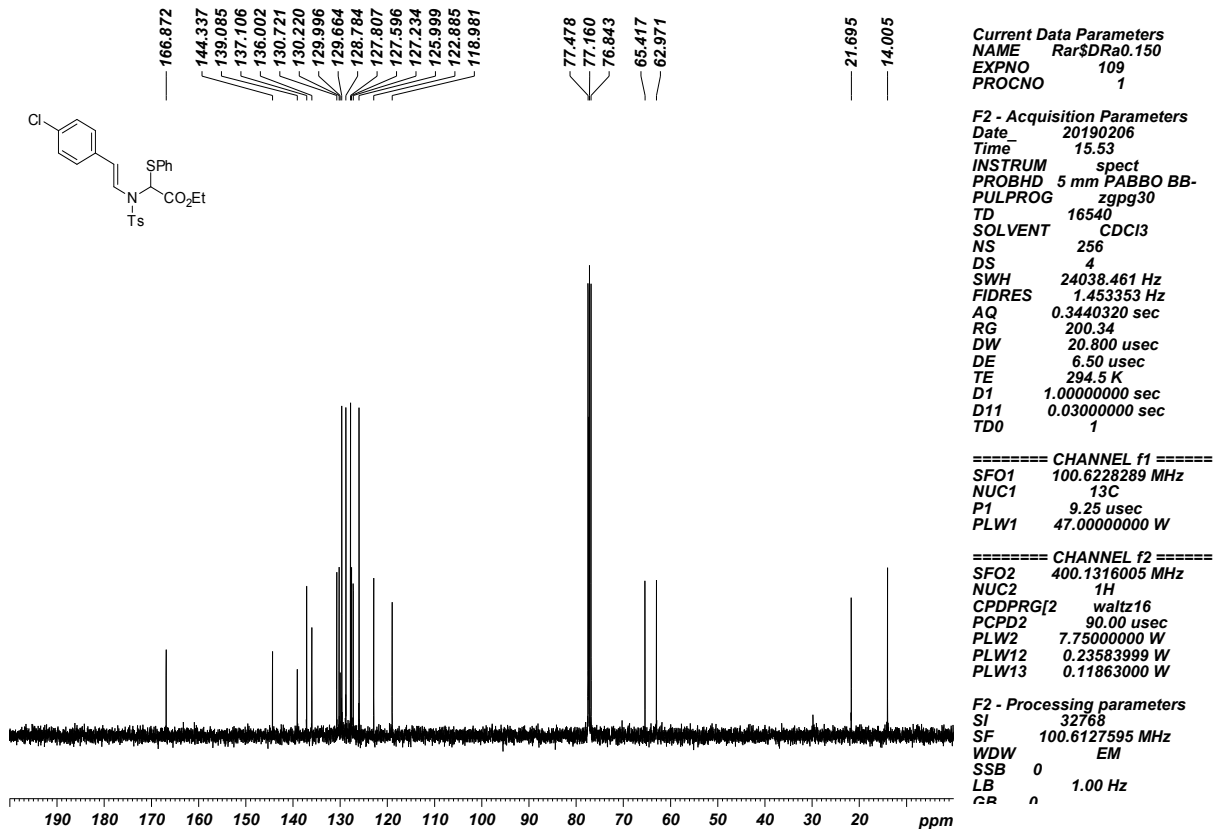
===== CHANNEL f2 =
 SFO2 500.15200
 NUC2
 CPDPRG[2 waltz
 PCPD2 80.
 PLW2 15.300000

Enamide 4ad

¹H NMR (400 MHz, CDCl₃, 24 °C)

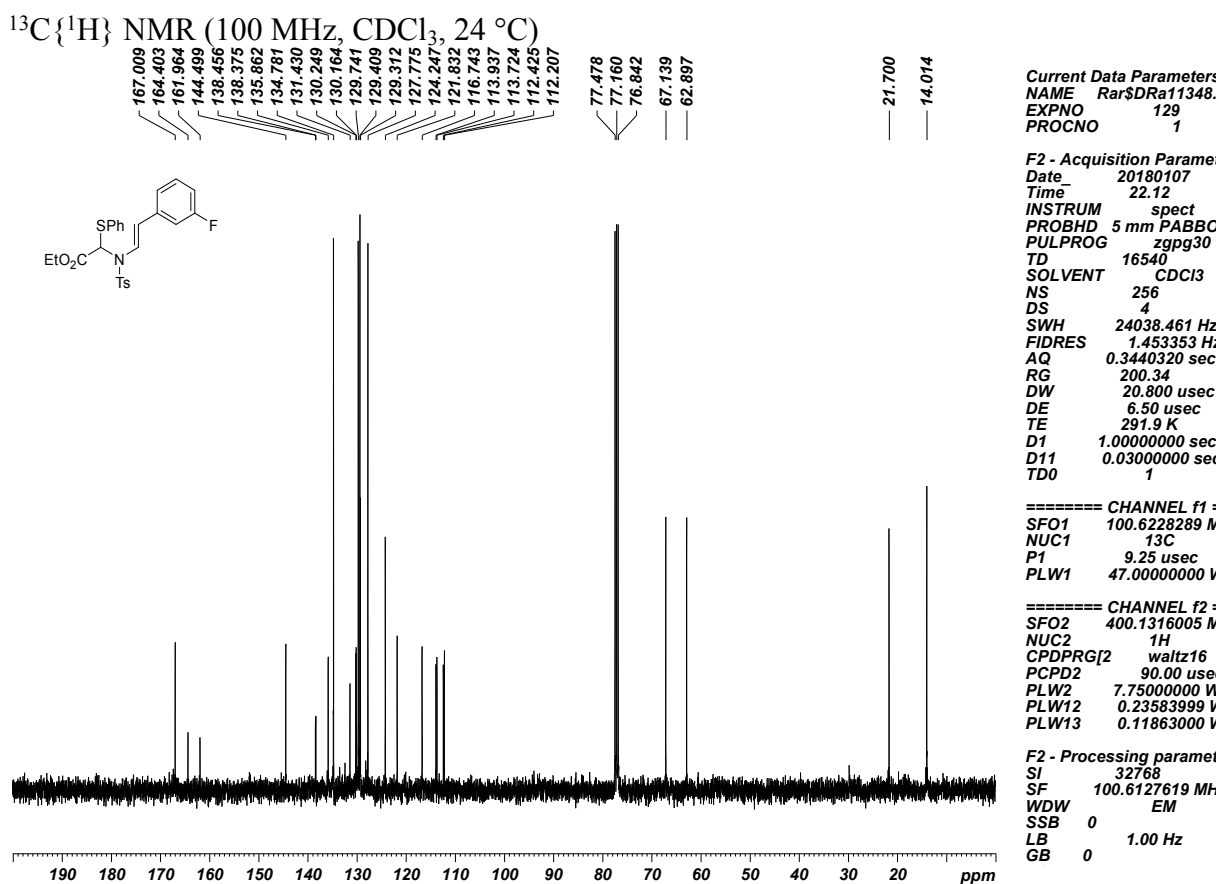
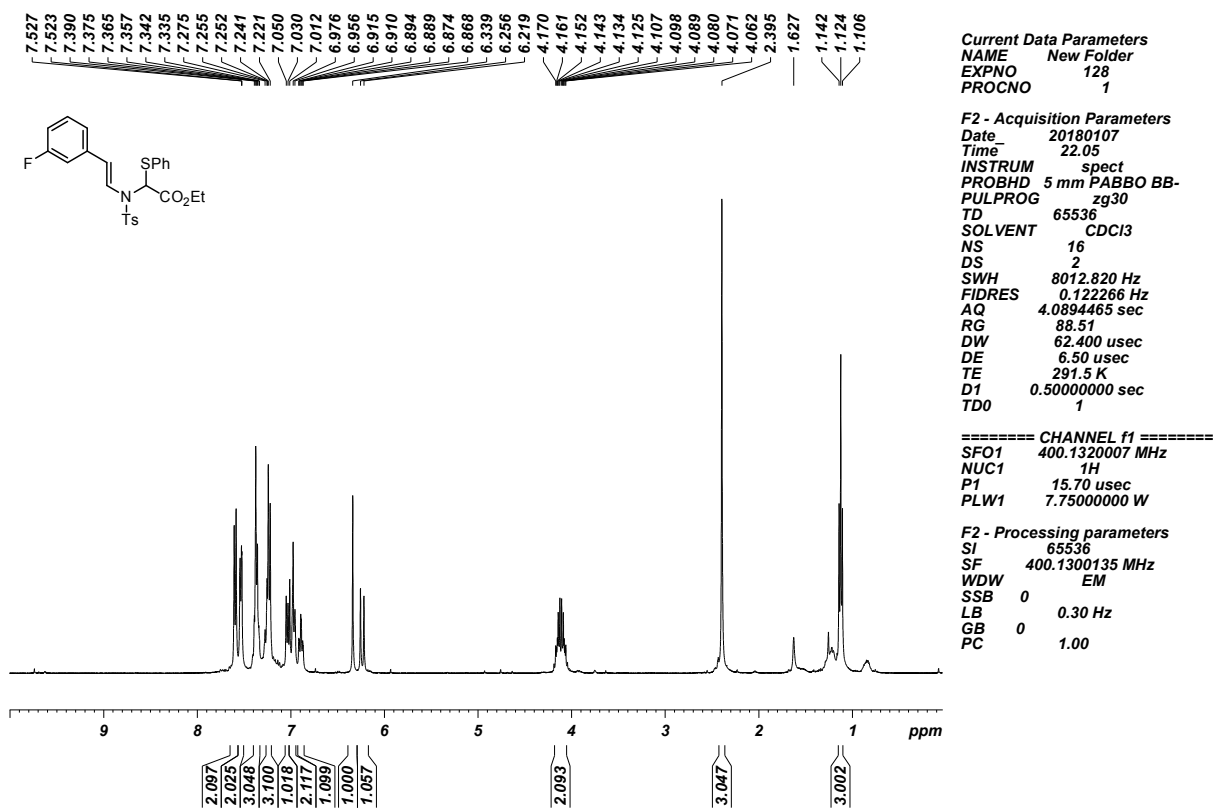


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



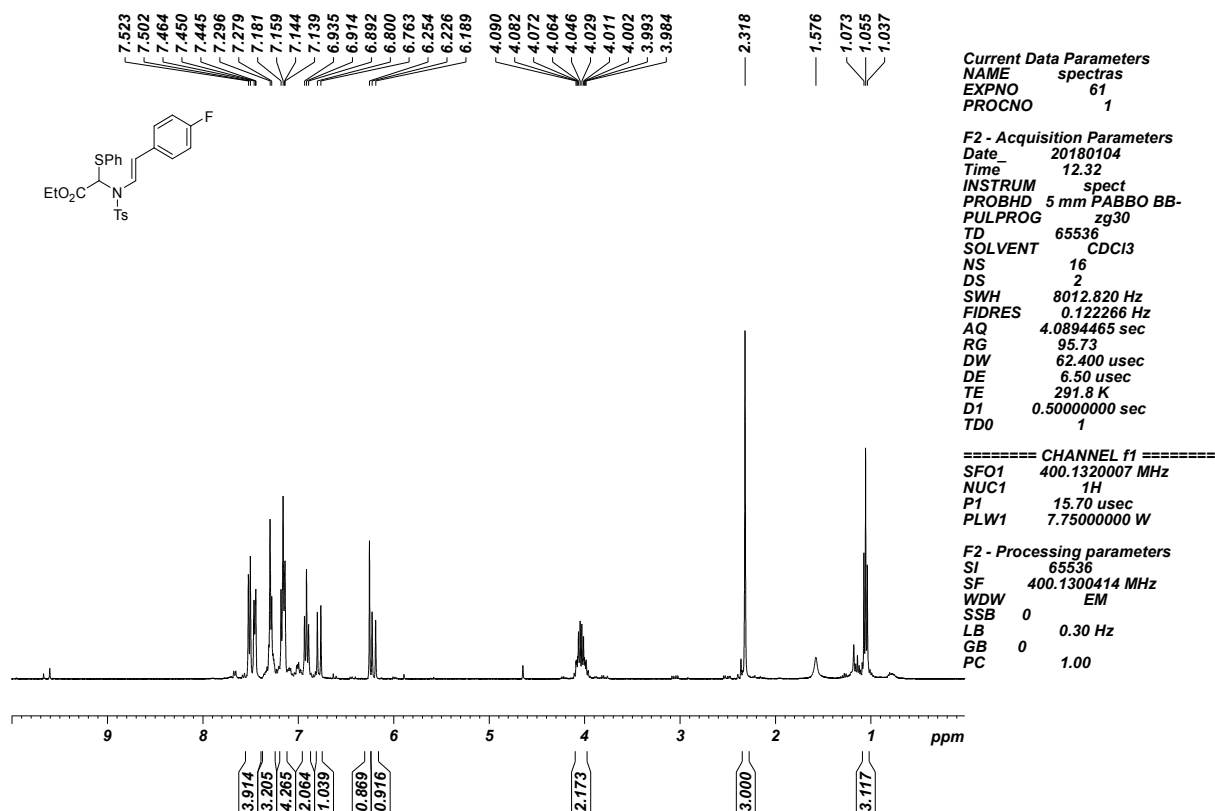
Enamide 4ae

^1H NMR (400 MHz, CDCl_3 , 24 °C)

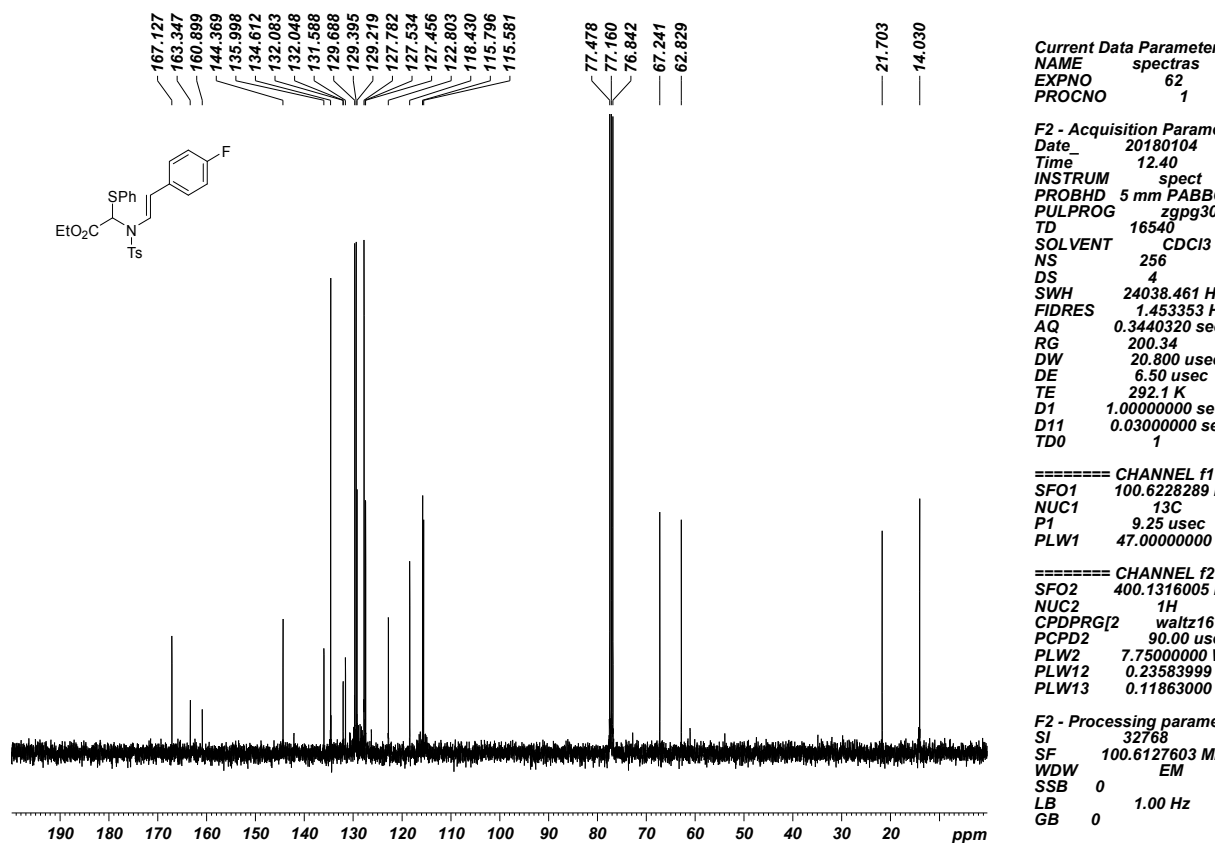


Enamide 4af

¹H NMR (400 MHz, CDCl₃, 24 °C)

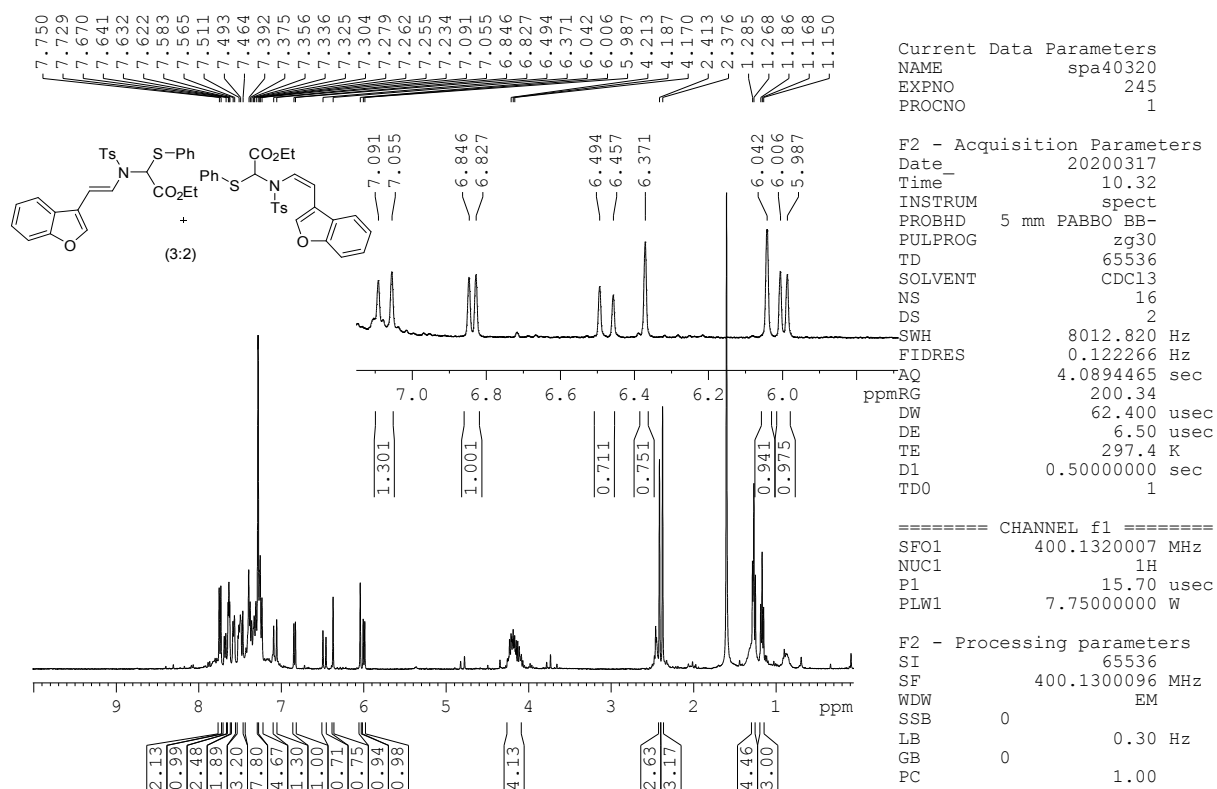


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)

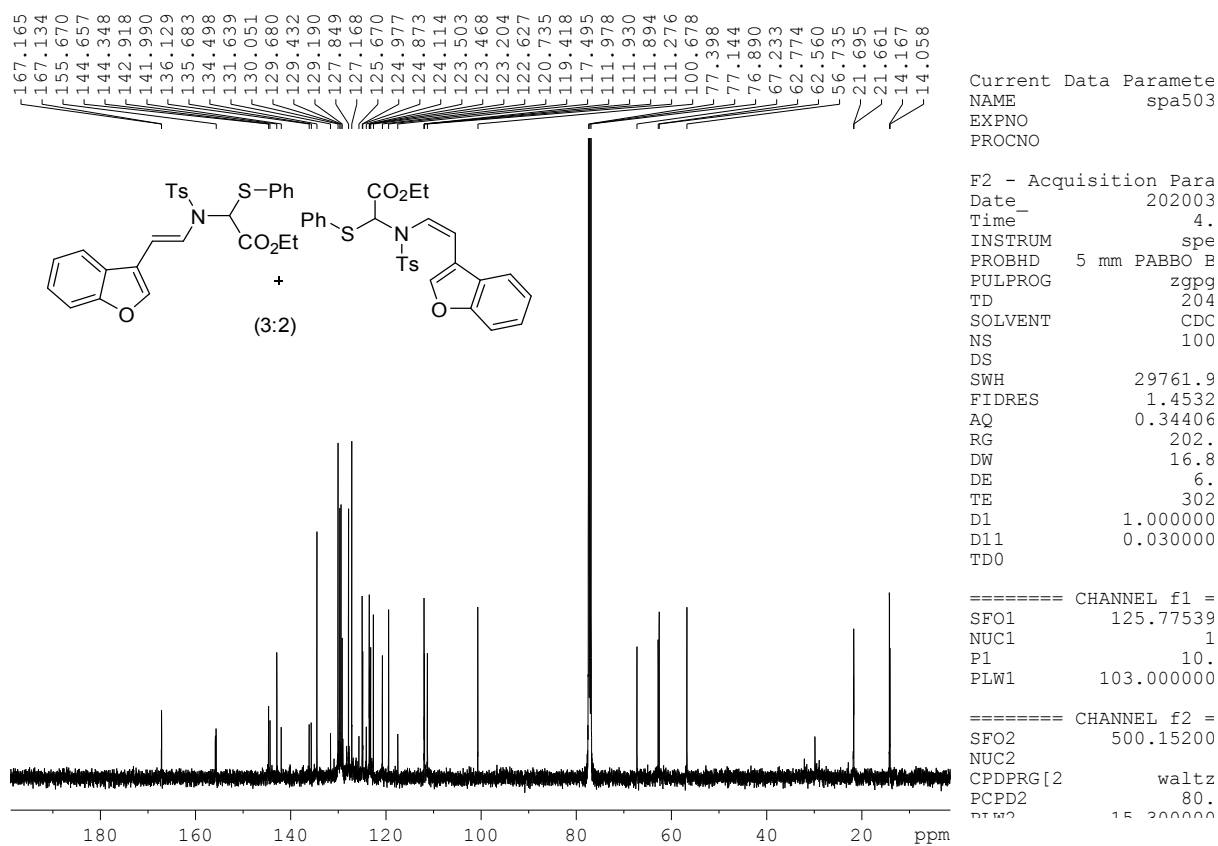


Enamide 4ag

^1H NMR (400 MHz, CDCl_3 , 24 °C)

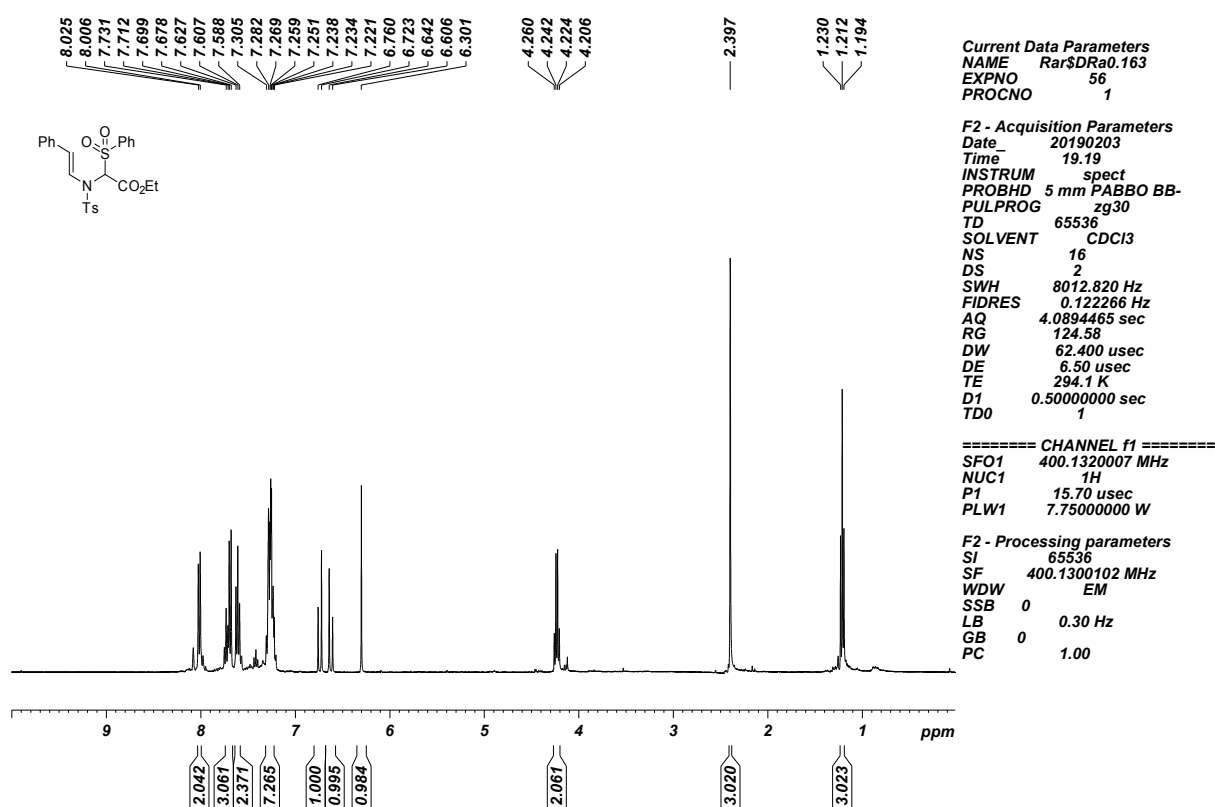


¹³C{¹H} NMR (125 MHz, CDCl₃, 24 °C)

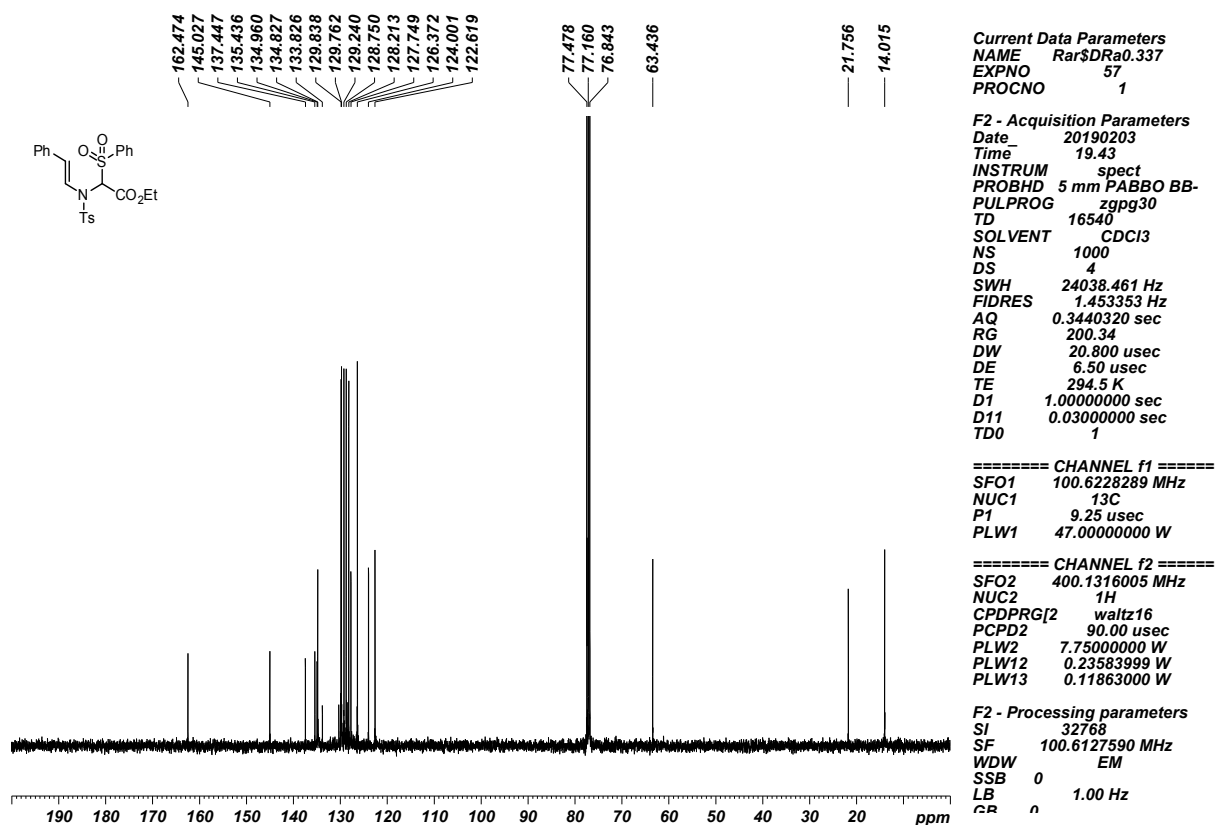


Sulfone 5

^1H NMR (400 MHz, CDCl_3 , 24 °C)

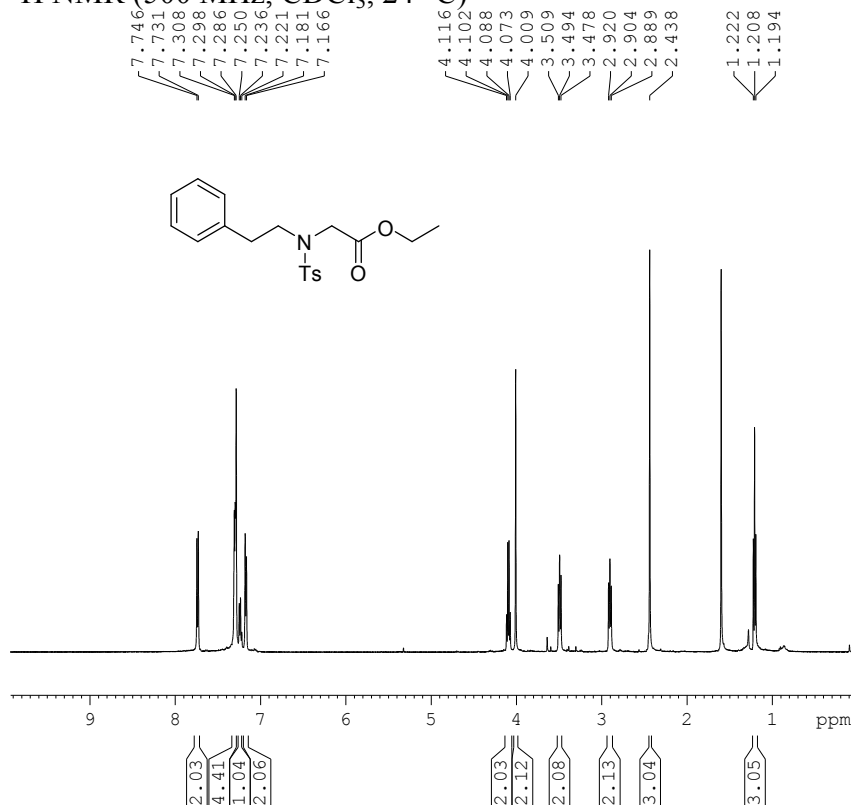


$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3 , 24 °C)



Compound 6

¹H NMR (500 MHz, CDCl₃, 24 °C)



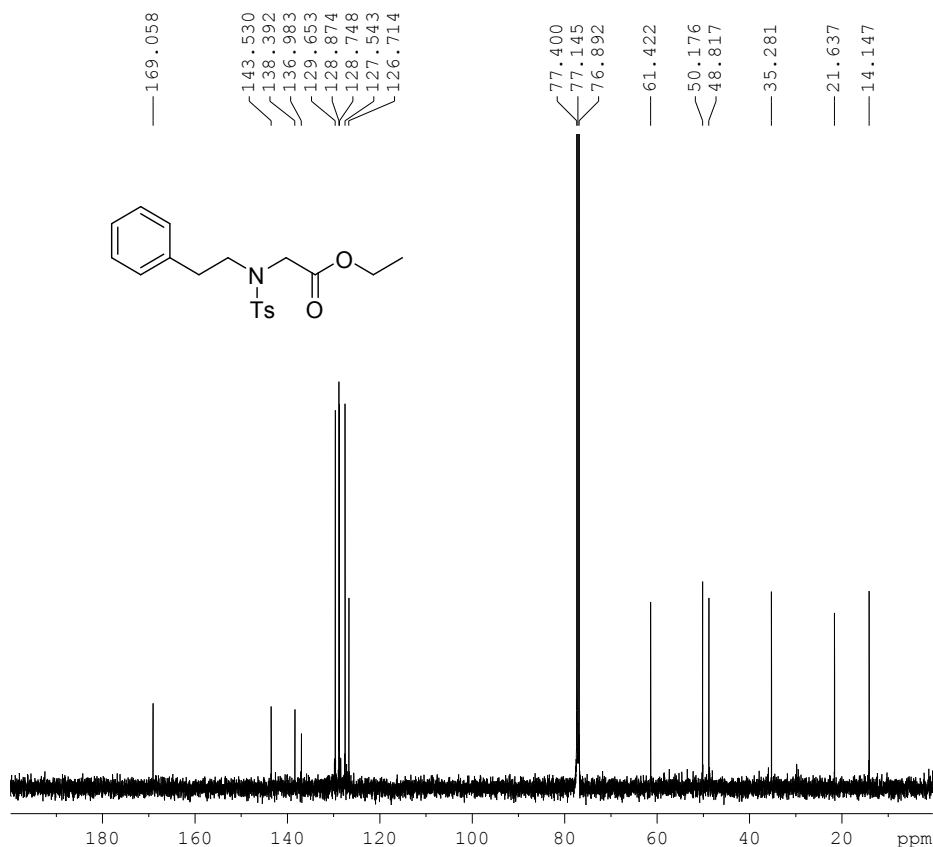
Current Data Parameters
NAME spa50320
EXPNO 23
PROCNO 1

F2 - Acquisition Parameters
Date_ 20200306
Time_ 10.22
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 10000.000 Hz
FIDRES 0.305176 Hz
AQ 1.6384000 sec
RG 138.53
DW 50.000 usec
DE 6.50 usec
TE 298.0 K
D1 0.50000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 500.1525008 MHz
NUC1 1H
P1 11.75 usec
PLW1 15.30000019 W

F2 - Processing parameters
SI 65536
SF 500.1500120 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C{¹H} NMR (125 MHz, CDCl₃, 24 °C)



Current Data Parameters
NAME spa503
EXPNO
PROCNO

F2 - Acquisition Parameters
Date_ 202003
Time_ 9.
INSTRUM spe
PROBHD 5 mm PABBO B
PULPROG zgpg
TD 204
SOLVENT CDC
NS 5
DS
LB
SWH 29761.9
FIDRES 1.4532
AQ 0.34406
RG 202.
DW 16.8
DE 6.
TE 300
D1 1.000000
D11 0.030000
TD0

===== CHANNEL f1 =
SFO1 125.77539
NUC1 1
P1 10.
PLW1 103.000000

===== CHANNEL f2 =
SFO2 500.15200
NUC2
PCPD2 waltz
PCPD2 80.
PCPD2 15.300000