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General methods and instrumentation

Commercial reagents were used without further purification. Solvents were freshly distilled from the appropriate drying agents or purified under nitrogen with the mBraun MBSPS-800 before use. The analytical TLCs were performed with silica gel 60 F254 plates. Column chromatography was performed by using silica gel 60 (200-300 mesh ASTM). The NMR spectra were recorded on a Bruker Avance II spectrometer, operating at 500 MHz for ¹H and 125 MHz for ¹³C or a Bruker Avance II spectrometer operating at 600 MHz for ¹H and 150 MHz for ¹³C. TMS was used as an internal reference for ¹H and ¹³C chemical shifts and CDCl₃ was used as solvent. Standard pulse programs from the Bruker library were used for 2D experiments. Mass spectrometry measurements were conducted by using the electrospray ionization technique on a Bruker Daltonics microTOF-Q or Finnigan LCQ Advantage MAX mass spectrometer. MALDI mass spectrometry was performed using Bruker ultrafleXtreme spectrometer with trans-2-[3-(4-tert-Butylphenyl)-2-methyl-2-propenylidene]malononitrile (DCTB) matrix and potassium trifluoroacetate. Absorption UV/Vis/NIR spectra were recorded by using a Varian Cary 50 Bio and Jasco V-770 spectrophotometers. The applied dichloromethane was freshly distilled over calcium hydride. Electrochemical measurements were performed by means of Autolab (Metrohm) potentiostat/galvanostat system for dichloromethane solutions with a glassy carbon, a platinum wire, and Ag/AgCl as the working, auxiliary, and reference electrodes, respectively. Tetrabutylammonium hexafluorophosphate was used as a supporting electrolyte. The potentials were referenced with the ferrocene/ferrocenium couple used as an internal standard. Crystals were obtained by slow diffusion of the dichloromethane solution into hexane. Crystal data for was collected at low temperature using Oxford Cryosystem device on Xcalibur Onyx four-circle diffractometer with graphite-monochromated Cu K α radiation ($\lambda = 1.54184 \text{ \AA}$). Data reduction and analysis were carried out with the CrysAlisPRO programs¹. The structures were solved by using the SHELXT² and refined by the full-matrix least-squares method on all F² data using the SHELXL.³ All hydrogen atoms, including those located in the difference density map, were placed in calculated positions and refined as the riding model. The symmetry-independent unit of **2e** contains a half of a molecule which is disordered by the inversion center. See Tables S1and S2 for detailed data. The full structures and diffraction data are available in Cambridge Crystallographic Data Center under the CCDC numbers 1957894 and 1977696.

Table S1 Crystal data for **2e**

| Crystal data | |
|---|---|
| Chemical formula | C ₅₄ H ₄₇ N ₅ O ₈ ·C ₆ H ₁₄ |
| M_r | 980.13 |
| Crystal system, space group | Triclinic, <i>P</i> -1 |
| Temperature (K) | 100 |
| a, b, c (Å) | 8.915 (1), 9.991(1), 14.328(2) |
| α, β, γ (°) | 98.93(1), 91.79(1), 92.21(1) |
| V (Å³) | 1259.0(3) |
| Z | 1 |
| Radiation type | Cu <i>K</i> α |
| μ (mm⁻¹) | 0.69 |
| Crystal size (mm) | 0.49 × 0.17 × 0.09 |
| Data collection | |
| Diffractometer | Xcalibur, Onyx |
| Absorption correction | — |
| No. of measured, independent and observed [<i>I</i> > 2σ(<i>I</i>)] reflections | 6859, 3952, 2398 |
| R_{int} | 0.095 |
| (sin θ/λ) _{max} (Å ⁻¹) | 0.575 |
| Refinement | |
| R[F² > 2σ(F²)], wR(F²), <i>S</i> | 0.099, 0.293, 1.00 |
| No. of reflections | 3952 |
| No. of parameters | 347 |
| Δρ_{max}, Δρ_{min} (e Å⁻³) | 0.39, -0.42 |

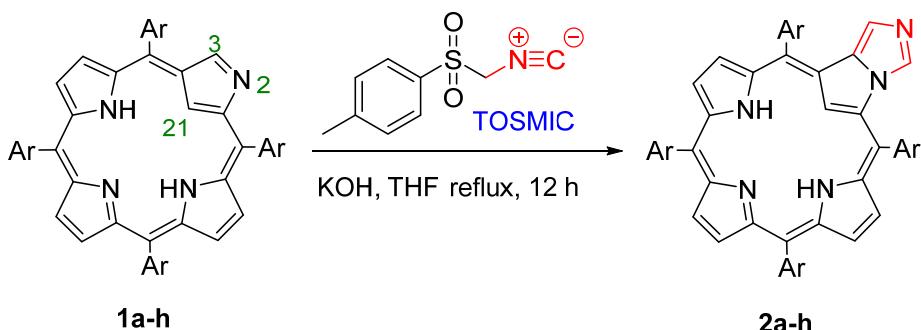
Table S2 Crystal data for **5a**

| Crystal data | |
|---|---|
| Chemical formula | C _{48.50} H ₃₃ AgAuCl ₄ N ₅ |
| M _r | 1132.43 |
| Crystal system, space group | Triclinic, P-1 |
| Temperature (K) | 100 |
| a, b, c (Å) | 9.9193 (5), 13.7034 (8), 15.7321 (8) |
| α, β, γ (°) | 75.013 (5), 84.724 (4), 78.550 (4) |
| V (Å ³) | 2022.72 (19) |
| Z | 2 |
| Radiation type | Cu Kα |
| μ (mm ⁻¹) | 13.38 |
| Crystal size (mm) | 0.34 × 0.03 × 0.03 |
| Data collection | |
| Diffractometer | Xcalibur, Ruby, Gemini ultra |
| Absorption correction | Analytical |
| Tmin, Tmax | 0.212, 0.736 |
| No. of measured, independent and observed [I > 2σ(I)] reflections | 13937, 6395, 5470 |
| Rint | 0.069 |
| (sin θ/λ) _{max} (Å ⁻¹) | 0.575 |
| Refinement | |
| R[F2 > 2σ(F2)], wR(F2), S | 0.045, 0.118, 1.02 |
| No. of reflections | 6395 |
| No. of parameters | 552 |
| H-atom treatment | H-atom parameters constrained |
| Δρ _{max} , Δρ _{min} (e Å ⁻³) | 1.74, -1.47 |

Synthesis and characterization

Synthesis of the precursor. Starting porphyrins **1a -1h** were obtained as described previously.⁴

Scheme S1. Synthesis of imidazole fused-21-carbachlorin derivatives **2a-h**



- 2a:** Ar = Ph, yield 69%; **2b:** Ar = 4-CH₃C₆H₄, yield 74%;
2c: Ar = 3,5-(CH₃)₂C₆H₃, yield 65%; **2d:** Ar = 4-CH₃OC₆H₄, yield 70%;
2e: Ar = 3,5-(CH₃O)₂C₆H₃, yield 71%; **2f:** Ar = 3,4,5-(CH₃O)₃C₆H₂, yield 76%;
2g: Ar = 4-FC₆H₄, yield 68%; **2h:** Ar = 4-CO₂CH₃C₆H₄, yield 69%;

General procedure for synthesis of Imidazole fused-21-carbachlorin:

A solution of 2-aza-21-carbaporphyrin **1** (0.1 mmol), toluenesulphonylmethyl isocyanide (0.3 mmol) and KOH (1 mmol) in THF (20 ml) was stirred at reflux temperature for 12 hr. The solvent was then evaporated under vacuum. The residue was chromatographed on a silica gel column with dichloromethane/ethyl acetate (V/V = 100:3) as eluent. The first red fraction was collected, followed by removal of solvent to afford imidazole fused-21-carbachlorin derivative **2**.

Selected data for **2a-h**

2a: yield 69%. ¹H NMR (500MHz, CDCl₃, 298K) δ = -5.58 (s, 1H, -HC=), -2.48 (br, 2H, -NH), 6.24 (s, 1H, -HC=), 6.84 (s, 1H, -HC=), 7.69-7.73 (m, 6H, ArH), 7.75-7.78 (m, 3H, ArH), 7.79-7.84 (m, 3H, ArH), 8.08-8.10 (m, 2H, ArH), 8.11-8.13 (m, 6H, ArH), 8.52-8.53 (m, 5H, pyrrH), 8.56 (d, J = 5.0 Hz, 1H, pyrrH); ¹³C NMR (125MHz, CDCl₃, 298 K) δ_C = 104.2, 104.3, 110.9, 118.2, 120.3, 120.5, 121.4, 125.3, 125.7, 126.1, 126.5, 126.9, 127.8, 127.9, 128.56, 128.66, 128.74, 129.4, 129.8, 131.0, 132.8, 132.9, 133.2, 133.63, 133.65, 134.3, 137.1, 137.3, 137.4, 138.6, 139.0, 139.2, 140.9, 141.8, 141.9, 154.7, 154.8. UV-vis (CH₂Cl₂) λ_{max}/nm (logε): 451 (5.22), 533 (4.38), 645 (3.67). ESI-HRMS calc. for [C₄₆H₃₂N₅]⁺ (M+H)⁺: 654.2652, Found: 654.2650.

2b: yield 74%. ¹H NMR (500MHz, CDCl₃, 298K) δ = -5.55 (s, 1H, -HC=), -2.35 (br, 2H, -NH), 2.67 (s, 12H, H₃C-), 6.28 (s, 1H, -HC=), 6.89 (s, 1H, -HC=), 7.52 (d, J = 8.0 Hz, 4H, ArH), 7.58 (d, J = 7.5 Hz, 2H, ArH), 7.64 (d, J = 7.5 Hz, 2H, ArH), 7.98-8.01 (m, 8H, ArH), 8.54-8.55 (m, 5H, pyrrH), 8.59 (d, J = 5.0 Hz, 1H, pyrrH); ¹³C NMR (125MHz, CDCl₃, 298 K) δ_C = 21.53, 21.58, 21.6, 104.2, 110.9, 118.1, 120.2, 120.3, 121.2, 125.2, 125.6, 126.29, 126.34, 127.6, 128.8, 129.3, 130.5, 130.9, 132.7, 132.9, 133.1, 133.49, 133.52, 134.3, 136.2, 137.2, 137.4, 137.5, 138.2, 138.3, 138.7, 138.9, 139.0, 139.2, 139.3, 154.7, 154.8. UV-vis (CH₂Cl₂) λ_{max}/nm (logε): 452 (5.25), 540 (4.39), 642 (3.62). ESI-HRMS calc. for [C₅₀H₄₀N₅]⁺ (M+H)⁺: 710.3278, Found: 710.3277.

2c: yield 65%. ¹H NMR (500MHz, CDCl₃, 298K) δ = -5.60 (s, 1H, -HC=), -2.37 (br, 2H, -NH), 2.58 (s, 24H, H₃C-), 6.33 (s, 1H, -HC=), 6.92 (s, 1H, -HC=), 7.37 (s, 2H, ArH), 7.40 (s, 1H, ArH), 7.44 (s, 1H, ArH), 7.73 (s, 4H, ArH), 7.76 (s, 4H, ArH), 8.57-8.59 (m, 5H, pyrrH), 8.65 (d, J = 5.0 Hz, 1H, pyrrH); ¹³C NMR (125MHz, CDCl₃, 298 K) δ_C = 21.5, 21.6, 104.2, 111.2, 118.4, 120.4, 120.6, 125.3, 125.7, 126.3, 129.4, 130.0, 130.5, 130.9, 131.0, 132.6, 132.7, 133.5, 133.6, 136.1, 137.2, 137.4, 138.0, 139.0, 139.2, 139.4,

141.0, 141.77, 141.81, 154.6, 154.8. UV-vis (CH_2Cl_2) λ_{\max}/nm ($\log\epsilon$): 451 (5.21), 535 (4.40), 641 (3.75). ESI-HRMS calc. for $[\text{C}_{54}\text{H}_{48}\text{N}_5]^+$ ($\text{M}+\text{H}$)⁺: 766.3904, Found: 766.3903.

2d: yield 70%. ^1H NMR (500MHz, CDCl_3 , 298K) δ = -5.58 (s, 1H, -HC=), -2.23 (br, 2H, -NH), 4.06 (s, 12H, - CH_3O), 6.34 (s, 1H, -HC=), 6.95 (s, 1H, -HC=), 7.26 (d, J = 8.0 Hz, 4H, ArH), 7.29 (d, J = 8.5 Hz, 2H, ArH), 7.35 (d, J = 8.5 Hz, 2H, ArH), 7.99-8.03 (m, 8H, ArH), 8.50-8.52 (m, 6H, pyrrH); ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 55.58, 55.62, 104.2, 110.5, 112.4, 113.1, 114.0, 115.2, 117.8, 119.79, 119.82, 120.9, 125.1, 125.4, 126.2, 126.3, 128.9, 130.7, 131.5, 133.0, 133.4, 133.5, 133.7, 134.1, 134.3, 134.4, 134.5, 135.5, 137.4, 137.5, 137.6, 138.8, 139.2, 154.95, 154.97, 159.5, 160.0, 160.5. UV-vis (CH_2Cl_2) λ_{\max}/nm ($\log\epsilon$): 454 (5.22), 539 (4.37), 644 (3.73). ESI-HRMS calc. for $[\text{C}_{50}\text{H}_{40}\text{N}_5\text{O}_4]^+$ ($\text{M}+\text{H}$)⁺: 774.3075, Found: 774.3074.

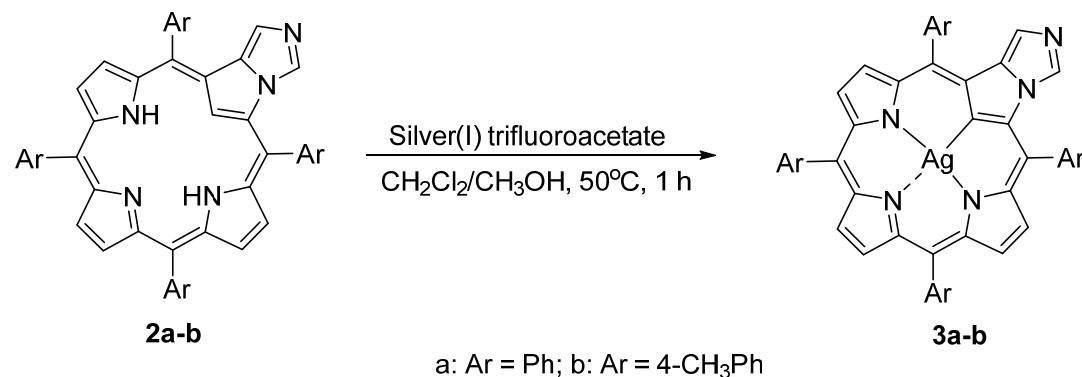
2e: yield 71%. ^1H NMR (500MHz, CDCl_3 , 298K) δ = -5.62 (s, 1H, -HC=), -2.41 (br, 2H, -NH), 3.95 (s, 18H, $\text{H}_3\text{CO}-$), 3.96 (s, 6H, $\text{H}_3\text{CO}-$), 6.53 (s, 1H, -HC=), 6.87 (t, J = 2.5 Hz, 2H, ArH), 6.89 (t, J = 2.5 Hz, 1H, ArH), 6.91 (t, J = 2.5 Hz, 1H, ArH), 7.12 (s, 1H, ArH), 7.24 (d, J = 2.0 Hz, 2H, ArH), 7.25 (d, J = 2.0 Hz, 2H, ArH), 7.34 (d, J = 2.0 Hz, 4H, ArH), 8.66-8.70 (m, 5H, pyrrH), 8.73 (d, J = 5.0 Hz, 1H, pyrrH); ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 55.6, 55.7, 55.8, 100.1, 100.8, 101.4, 110.76, 110.79, 111.5, 113.5, 117.9, 119.9, 120.1, 121.7, 125.5, 125.9, 126.40, 126.42, 128.5, 131.4, 132.6, 133.7, 136.9, 134.3, 137.2, 137.3, 138.5, 138.8, 140.7, 142.8, 143.68, 143.72, 154.4, 154.6, 159.1, 160.8, 161.8. UV-vis (CH_2Cl_2) λ_{\max}/nm ($\log\epsilon$): 449 (5.31), 535 (4.50), 638 (3.85). ESI-HRMS calc. for $[\text{C}_{54}\text{H}_{48}\text{N}_5\text{O}_8]^+$ ($\text{M}+\text{H}$)⁺: 894.3497, Found: 894.3497.

2f: yield 76%. ^1H NMR (500MHz, CDCl_3 , 298K) δ = -5.58 (s, 1H, -HC=), -1.95 (br, 2H, -NH), 3.98 (s, 18H, - CH_3O), 3.99 (s, 6H, - CH_3O), 4.17 (s, 12H, - CH_3O), 6.55 (s, 1H, -HC=), 7.12 (s, 1H, -HC=), 7.33 (s, 2H, ArH), 7.36 (s, 2H, ArH), 7.41 (s, 2H, ArH), 7.42 (s, 2H, ArH), 8.69-8.70 (m, 3H, pyrrH), 8.73 (d, J = 5.0 Hz, 2H, pyrrH), 8.76 (d, J = 5.0 Hz, 1H, pyrrH); ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 56.4, 56.55, 56.58, 61.3, 61.5, 61.6, 104.2, 109.8, 110.7, 110.9, 112.4, 118.1, 120.2, 120.4, 121.6, 125.5, 125.6, 126.1, 126.5, 128.8, 131.2, 132.9, 133.7, 134.4, 135.8, 136.5, 137.2, 137.3, 137.4, 137.5, 138.0, 138.7, 138.8, 138.9, 139.3, 151.7, 153.3, 154.4, 154.7, 154.8. UV-vis (CH_2Cl_2) λ_{\max}/nm ($\log\epsilon$): 452 (5.16), 534 (4.35), 642 (3.71). ESI-HRMS calc. for $[\text{C}_{58}\text{H}_{56}\text{N}_5\text{O}_{12}]^+$ ($\text{M}+\text{H}$)⁺: 1014.3920, Found: 1014.3919.

2g: yield 68%. ^1H NMR (500MHz, CDCl_3 , 298K) δ = -5.61 (s, 1H, -HC=), -2.53 (br, 2H, -NH), 6.31 (s, 1H, -HC=), 6.92 (s, 1H, -HC=), 7.43 (t, J = 8.5 Hz, 4H, ArH), 7.49 (t, J = 8.5 Hz, 2H, ArH), 7.56 (t, J = 8.5 Hz, 2H, ArH), 8.06-8.11 (m, 8H, ArH), 8.49-8.52 (m, 6H, pyrrH); ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 104.2, 109.8, 113.9 (d, 2J = 21 Hz), 115.8 (d, 2J = 21 Hz), 116.9 (d, 2J = 21 Hz), 117.0, 119.2, 119.4, 121.4, 125.2, 125.6, 126.4, 126.5, 128.9, 130.8, 132.9, 133.61, 133.63, 134.58, 134.65, 134.78, 134.8, 134.90, 134.93, 135.5, 135.6, 136.83, 136.86, 137.2, 137.4, 137.54, 137.57, 137.6, 138.7, 139.1, 154.8, 154.9, 162.9 (d, 1J = 250 Hz), 163.3 (d, 1J = 250 Hz), 163.7 (d, 1J = 250 Hz). UV-vis (CH_2Cl_2) λ_{\max}/nm ($\log\epsilon$): 450 (5.28), 535 (4.46), 644 (3.79). ESI-HRMS calc. for $[\text{C}_{46}\text{H}_{28}\text{F}_4\text{N}_5]^+$ ($\text{M}+\text{H}$)⁺: 726.2275, Found: 726.2273.

2h: yield 69%. ^1H NMR (500MHz, CDCl_3 , 298K) δ = -5.84 (s, 1H, -HC=), -1.96 (br, 2H, -NH), 4.11 (s, 6H, - $\text{H}_3\text{C}-$), 4.12 (s, 3H, - $\text{H}_3\text{C}-$), 4.13 (s, 3H, - $\text{H}_3\text{C}-$), 6.16 (s, 1H, -HC=), 6.78 (s, 1H, -HC=), 8.04 (d, J = 8.0 Hz, 2H, ArH), 8.10-8.15 (m, 6H, ArH), 8.28 (d, J = 4.5 Hz, 1H, pyrrH), 8.33 (d, J = 4.5 Hz, 1H, pyrrH), 8.35 (d, J = 4.5 Hz, 1H, pyrrH), 8.40-8.44 (m, 9H), 8.49 (d, J = 8.0 Hz, 2H, ArH); ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 52.5, 52.6, 104.2, 104.3, 110.0, 117.1, 119.6, 119.7, 121.5, 125.3, 125.5, 126.3, 126.5, 128.3, 129.87, 129.88, 129.9, 130.3, 130.9, 131.0, 131.1, 132.2, 133.0, 133.2, 133.7, 134.3, 136.6, 136.8, 137.0, 138.1, 138.7, 142.9, 144.8, 146.1. UV-vis (CH_2Cl_2) λ_{\max}/nm ($\log\epsilon$): 454 (5.29), 537 (4.47), 644 (3.80). ESI-HRMS calc. for $[\text{C}_{54}\text{H}_{40}\text{N}_5\text{O}_8]^+$ ($\text{M}+\text{H}$)⁺: 886.2871, Found: 886.2869.

Scheme S2. Synthesis of Ag^{III} complexes of imidazole fused-21-carbachlorin derivatives **3**



General procedure for synthesis of Ag^{III} complex of imidazole fused-21-carbachlorin 3:

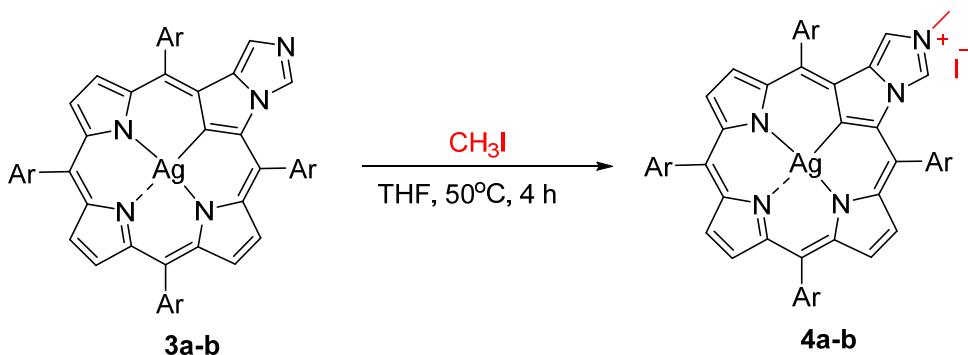
A 20 mg (0.028 mmol) sample of imidazole fused-21-carbachlorin **2** was dissolved in 12 mL of CH_2Cl_2 , and 24 mg (0.11 mmol, 4 equiv) of silver(I) trifluoroacetate dissolved in 0.5 mL of MeOH was added. The reaction mixture was protected from light and stirred for 1 h at 50°C. The solvent was removed under vacuum and the crude compound was loaded on a silica column and eluted with CH_2Cl_2 . The first fraction was collected, followed by removal of solvent to afford silver(III) complex of imidazole fused-21-carbachlorin derivative **3**.

Selected data for 3a-b

3a: yield 85%. ^1H NMR (500MHz, CDCl_3 , 298K) δ = 6.36 (s, 1H, -HC=), 6.89 (s, 1H, -HC=), 7.70-7.79 (m, 9H, ArH), 7.83-7.85 (m, 3H, ArH), 7.97-8.01 (m, 4H, ArH), 8.11-8.12 (m, 4H, ArH), 8.62-8.64 (m, 3H, pyrrH), 8.67-8.70 (m, 2H, pyrrH), 8.75-8.76 (m, 1H, pyrrH); ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 114.37, 118.21, 121.13, 121.15, 121.60, 122.11, 122.45, 124.72, 126.27, 126.30, 126.73, 126.75, 127.09, 128.05, 128.07, 128.51, 128.62, 128.79, 128.93, 128.95, 129.31, 129.39, 129.73, 130.04, 130.06, 131.31, 131.59, 132.11, 133.73, 133.77, 136.88, 137.06, 137.59, 137.61, 137.89, 137.98, 138.94, 138.99, 139.02, 139.15, 141.13, 141.61. UV-vis (CH_2Cl_2) $\lambda_{\text{max}}/\text{nm}$ ($\log \epsilon$): 389 (4.42), 449 (5.30), 523 (4.29), 555 (4.06), 575 (4.09), 629 (3.85). ESI-HRMS calc. for $[\text{C}_{46}\text{H}_{29}\text{AgN}_5]^+$ ($\text{M}+\text{H}$) $^+$: 758.1468, Found: 758.1473.

3b: yield 81%. ^1H NMR (500MHz, CDCl_3 , 298K) δ = 2.68 (s, 12H, $-\text{CH}_3$), 6.40 (s, 1H, $-\text{HC=}$), 6.94 (s, 1H, $-\text{HC=}$), 7.53 (d, J = 7.5 Hz, 4H, ArH), 7.58 (d, J = 8.0 Hz, 2H, ArH), 7.64 (d, J = 8.0 Hz, 2H, ArH), 7.84-7.88 (m, 4H, ArH), 7.99 (d, J = 8.0 Hz, 4H, ArH), 8.63-8.66 (m, 3H, pyrrH), 8.69-8.72 (m, 2H, pyrrH), 8.76-8.78 (m, 1H, pyrrH); ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 21.56, 21.63, 21.67, 114.35, 118.20, 121.15, 121.55, 122.06, 124.76, 126.13, 126.16, 126.59, 126.62, 127.77, 128.69, 128.80, 129.21, 129.29, 129.96, 130.42, 131.07, 131.41, 132.06, 133.63, 133.67, 136.14, 136.94, 137.15, 137.68, 137.71, 137.98, 138.07, 138.15. UV-vis (CH_2Cl_2) $\lambda_{\text{max}}/\text{nm}$ ($\log \epsilon$): 387 (4.31), 453 (5.17), 524 (4.21), 558 (3.94), 574 (3.94), 627 (3.71). ESI-HRMS calc. for $[\text{C}_{50}\text{H}_{37}\text{AgN}_5]^+$ ($\text{M}+\text{H}$) $^+$: 814.2094, Found: 814.2098.

Scheme 3. Synthesis of 3'-methyl- imidazole fused-21-carbachlorin silver(III) Derivatives **4**



3a: Ar = Ph; **3b:** Ar = 4-CH₃Ph

General procedure for synthesis of *N*-methylated Ag-imidazole fused-21-carbachlorin **4:**

A 20 mg (0.028 mmol) sample of Ag^{III} complex of imidazole fused-21-carbachlorin **3** was dissolved in 3 mL of THF, and 100 µL of CH₃I was added. The reaction mixture was stirred for 4 h at 50°C. The solvent was removed under vacuum and the crude compound was loaded on a silica column and eluted with CH₂Cl₂/CH₃OH (V:V = 95:5). The last fraction was collected, followed by removal of solvent to afford 3'-methylated Ag-imidazole fused-21-carbachlorin derivative **4**.

Selected data for [4a-b]I.

[4a]I: yield 68%. ¹H NMR (500MHz, CDCl₃, 298K) δ = 4.18 (s, 3H, -CH₃), 6.37 (s, 1H, -HC=), 7.57 (s, 1H, -HC=), 7.76-7.82 (m, 6H, ArH), 7.88-7.89 (m, 3H, ArH), 7.99-8.05 (m, 5H, ArH), 8.08-8.10 (m, 2H, ArH), 8.13-8.15 (m, 4H, ArH), 8.76-8.77 (m, 3H, pyrrH), 8.80 (d, J = 5.0 Hz, 1H, pyrrH), 8.82 (d, J = 5.0 Hz, 1H, pyrrH), 8.87 (d, J = 4.5 Hz, 1H, pyrrH); ¹³C NMR (125MHz, CDCl₃, 298 K) δ_C = 39.53, 113.33, 114.35, 116.44, 121.54, 122.77, 122.99, 123.39, 127.28, 127.85, 127.99, 128.49, 128.99, 129.53, 129.71, 130.34, 130.57, 130.62, 130.78, 131.22, 131.51, 133.82, 136.76, 137.25, 137.93, 138.59, 138.69, 138.77, 138.85, 139.62, 139.68; UV-vis (CH₂Cl₂) λ_{max}/nm (logε): 438 (5.54), 518 (4.38), 558 (4.30). ESI-HRMS calc. for [C₄₇H₃₁AgN₅]⁺ (M-I)⁺: 772.1625, Found: 772.1627.

[4b]I: yield 65%. ¹H NMR (500MHz, CDCl₃, 298K) δ = 2.70 (s, 6H, -CH₃), 2.70 (s, 3H, -CH₃), 2.77 (s, 3H, -CH₃), 4.24 (s, 3H, -CH₃), 6.47 (s, 1H, -HC=), 7.57 (d, J = 7.5 Hz, 4H, ArH), 7.63 (s, 1H, -HC=), 7.67 (d, J = 7.5 Hz, 2H, ArH), 7.80 (d, J = 7.5 Hz, 2H, ArH), 7.89 (d, J = 7.5 Hz, 2H, ArH), 7.93 (d, J = 7.5 Hz, 2H, ArH), 8.01 (d, J = 7.5 Hz, 4H, ArH), 8.74-8.87 (m, 6H, pyrrH); ¹³C NMR (125MHz, CDCl₃, 298 K) δ_C = 21.57, 21.81, 22.03, 39.88, 113.33, 114.30, 116.49, 121.60, 122.79, 123.03, 123.50, 127.82, 127.98, 129.60, 130.24, 130.69, 130.95, 131.27, 131.38, 133.75, 136.70, 137.44, 137.94, 137.97, 138.04, 138.23, 138.25, 138.71, 138.85, 139.44, 139.72, 140.77. UV-vis (CH₂Cl₂) λ_{max}/nm (logε): 442 (5.52), 518 (4.42), 558 (4.31). ESI-HRMS calc. for [C₅₁H₃₉AgN₅]⁺ (M-I)⁺: 828.2251, Found: 828.2251.

Iodide counteranion was exchanged into BF₄⁻ by addition of 10 mg of silver(I) tetrafluoroborate in 1 mL of ethanol into the solution of **[4a]I** (15 mg) in dichloromethane (5 mL) and stirring for 10 min at room temperature. The solvent was then removed by evaporation, and the compound was extracted with CH₂Cl₂, solution was filtered, and passed down a short silica gel column with dichloromethane/ethanol solution (99/1). Substitution of I⁻ with chloride was accomplished by stirring of a sample of **[4a]I** (10 mg) in chloroform with brine for 1 h. The organic layer was then separated and washed with three portion of water, evaporated and passed down a short silica gel column.

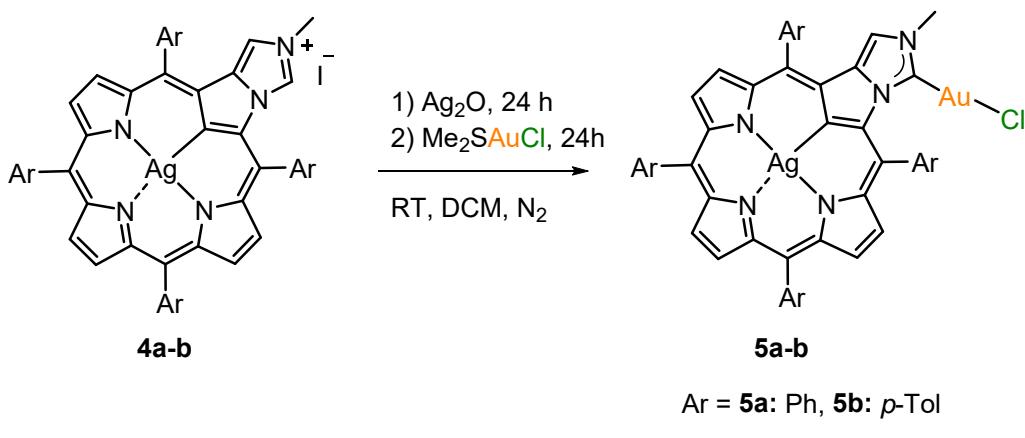
Iodide counteranion was exchanged into Cl⁻ by stirring a sample of **[4a]I** or **[4b]I** (10 mg) in dichloromethane (5 mL) with brine (2 mL) for 20 h at room temperature. The organic layer was then

separated and washed with four portion of water, dried in vacuo and crystallized from chloroform/hexane.

[4a]BF₄: yield 85%. ¹H NMR (500MHz, CDCl₃, 300K) δ = 4.03 (m, 3H, 3'-CH₃), 6.31, (dq, *J*_d= 1.2 Hz, *J*_q = 0.4 Hz, 1H, 4'), 7.51 (dq, *J*_d= 1.2 Hz, *J*_q = 0.6 Hz, 1H, 2'), 7.77 (m, 6H, meta+para Ph), 7.87 (m, 3H, meta Ph), 8.00 (m, 5H, meta+ortho Ph), 8.01 (m, 2, ortho Ph), 8.12 (m, 4H, ortho Ph), 8.74 (dd, ³*J*_{HH} = 4.8 Hz, ⁴*J*_{AgH} = 0.4 Hz, 1H, pyrrH), 8.75 (dd, ³*J*_{HH} = 4.9 Hz, ⁴*J*_{AgH} = 0.3 Hz, 1H, pyrrH), 8.75 (dd, ³*J*_{HH} = 4.8 Hz, ⁴*J*_{AgH} = 0.3 Hz, 1H, pyrrH), 8.79 (dd, ³*J*_{HH} = 4.9 Hz, ⁴*J*_{AgH} = 1.1 Hz, 1H, pyrrH), 8.81 (dd, ³*J*_{HH} = 4.9 Hz, ⁴*J*_{AgH} = 1.2 Hz, 1H, pyrrH), 8.86 (dd, ³*J*_{HH} = 4.9 Hz, ⁴*J*_{AgH} = 1.6 Hz, 1H, pyrrH).

[4a]Cl: yield 90%. ¹H NMR (500MHz, CDCl₃, 300K) δ = 4.25 (s, 3H, 3'-CH₃), 6.36, (d, ⁴*J* = 0.9 Hz, 1H, 4'), 7.77 (m, 6H, meta+para Ph), 7.87 (m, 3H, meta Ph), 7.92 (b, 1H, 2'), 7.99 (m, 5H, meta+ortho Ph), 8.02 (m, 2H, ortho Ph), 8.11 (m, 4H, ortho Ph), 8.72 (dd, ³*J*_{HH} = 5.0 Hz, ⁴*J*_{AgH} = 1.4 Hz, 1H, pyrrH), 8.73 (d, ³*J*_{HH} = 4.6 Hz, 1H, pyrrH), 8.75 (d, ³*J*_{HH} = 4.6 Hz, 1H, pyrrH), 8.78 (dd, ³*J*_{HH} = 5.0 Hz, ⁴*J*_{AgH} = 1.0 Hz, 1H, pyrrH), 8.81 (dd, ³*J*_{HH} = 4.8 Hz, ⁴*J*_{AgH} = 1.1 Hz, 1H, pyrrH), 8.85 (dd, ³*J*_{HH} = 4.8 Hz, ⁴*J*_{AgH} = 1.5 Hz, 1H, pyrrH).

[4b]Cl: yield 93%. ¹H NMR (500MHz, CDCl₃, 300K) δ = 2.69 (s, 6H, Tol(CH₃)x2), 2.77 (s, 3H, Tol(CH₃)), 2.73 (s, 3H, Tol(CH₃)), 4.30 (s, 3H, 3'-CH₃), 6.46 (d, ⁴*J* = 1.1 Hz, 1H, 4'), 7.55 (m, 4H, meta Tol), 7.65 (m, 2H, meta Tol), 7.77 (m, 2H, meta Tol), 7.83 (m, 1H, 2'), 7.87 (m, 2H, ortho Tol), 7.89 (m, 2H, ortho Tol), 7.99 (m, 4H, ortho Tol), 8.72 (dd, ³*J*_{HH} = 5.0 Hz, ⁴*J*_{AgH} = 1.5 Hz, 1H, pyrrH), 8.74 (d, ³*J*_{HH} = 4.9 Hz, 1H, pyrrH), 8.76 (d, ³*J*_{HH} = 4.9 Hz, 1H, pyrrH), 8.79 (dd, ³*J*_{HH} = 5.0 Hz, ⁴*J*_{AgH} = 1.0 Hz, 1H, pyrrH), 8.81 (dd, ³*J*_{HH} = 5.0 Hz, ⁴*J*_{AgH} = 1.1 Hz, 1H, pyrrH), 8.85 (dd, ³*J*_{HH} = 5.0 Hz, ⁴*J*_{AgH} = 1.4 Hz, 1H, pyrrH),



General procedure for synthesis of chlorogold(I) carbene complex of methylated Ag-imidazole fused-21-carbachlorin 5:

A sample of [**4a**]I or [**4b**]I (20 mg, 0.025 mmol) was dissolved in 10 mL of dichloromethane, and Ag_2O (0.05 mmol, 12 mg) was added. The slurry was deaerated for 20 min by a stream of N_2 . The reaction mixture was then stirred for 24 h at room temperature under inert atmosphere. After that time 0.05 mmol of AuClSiMe_2 (15 mg) was added and the reaction mixture was stirred for further 24 h. The mixture was then filtered, the solvent was evaporated and the product was redissolved in a small amount of dichloromethane and charged on the silica gel column. The fastest moving brown fraction migrating with dichloromethane was collected. The product **5a** or **5b** was recrystallized with dichloromethane/hexane and dried on air.

Selected data for **5a-b.**

5a: yield 72%. ^1H NMR (500MHz, CDCl_3 , 300K) δ = 8.74 (dd, $^3J_{\text{HH}} = 4.9$ Hz, $^4J_{\text{AgH}} = 1.5$ Hz, 1H, pyrrH), 8.69 (dd, $^3J_{\text{HH}} = 4.9$ Hz, $^4J_{\text{AgH}} = 1.1$ Hz, 1H, pyrrH), 8.65 (d, $^3J_{\text{HH}} = 4.8$ Hz, 1H, pyrrH), 8.6 (dd, $^3J_{\text{HH}} = 5.0$ Hz, $^4J_{\text{AgH}} = 1.2$ Hz, 1H, pyrrH), 8.63 (d, $^3J_{\text{HH}} = 4.8$ Hz, 1H, pyrrH), 8.61 (dd, $^3J_{\text{HH}} = 4.9$ Hz, $^4J_{\text{AgH}} = 1.5$ Hz, 1H, pyrrH), 8.13 (dd, $^3J_{\text{HH}} = 7.6$ Hz, $^4J_{\text{HH}} = 1.5$ Hz, 2H, ortho ArH), 8.10 (m, 4H, ortho ArH), 8.00 (m, 2H, ortho ArH), 7.83 (m, 6H, meta+para ArH), 7.73 (m, 6H, meta+para ArH), 6.00 (s, 1H, 4'-H), 3.92 (s, 3H, 3'- CH_3). ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 170.1, 142.0, 141.3, 141., 140.9, 139.6, 139.5, 139.3, 138.9, 138.6, 137.7, 137.1, 133.9, 133.8, 133.7, 131.6, 130.2, 129.9, 129.6, 129.5, 129.3, 129.0, 128.9, 128.8, 128.3, 128.2, 127.2, 127.1, 126.2, 122., 121.7, 120.2, 117.4, 113.0, 40.2. UV-vis (CH_2Cl_2) λ_{max} /nm (log ϵ): 266 (4.57), 303 sh, 363 (4.06), 384 (4.08), 427 sh, 452 (5.23), 491 sh, 530 (4.19), 572 (4.06), 620 (3.36). MALDI-TOF-HRMS calc. for $[\text{C}_{47}\text{H}_{30}\text{AgAuClN}_5]^+$ (M) $^+$: 1003.0906, Found: 1003.093; calc. for $[\text{C}_{47}\text{H}_{30}\text{AgAuClKN}_5]^+$ (M+K) $^+$: 1042.0538, Found: 1042.056.

5b: yield 60%. ^1H NMR (500MHz, CDCl_3 , 300K) δ = 8.71 (dd, $^3J_{\text{HH}} = 4.9$ Hz, $^4J_{\text{AgH}} = 1.4$ Hz, 1H, pyrrH), 8.69 (dd, $^3J_{\text{HH}} = 4.9$ Hz, $^4J_{\text{AgH}} = 1.1$ Hz, 1H, pyrrH), 8.65 (d, $^3J_{\text{HH}} = 4.8$ Hz, 1H, pyrrH), 8.64 (dd, $^3J_{\text{HH}} = 4.9$ Hz, $^4J_{\text{AgH}} = 1.5$ Hz, 1H, pyrrH), 8.637 (d, $^3J_{\text{HH}} = 4.8$ Hz, 1H, pyrrH), 8.59 (dd, $^3J_{\text{HH}} = 4.9$ Hz, $^4J_{\text{AgH}} = 1.4$ Hz, 1H, pyrrH), 7.99 (m, 3H, ortho ArH), 7.98 (m, 3H, ortho ArH), 7.86 (m, 2H, ortho ArH), 7.63 (m, 2H, meta ArH), 7.61 (m, 2H, meta ArH), 7.53 (m, 4H, meta ArH), 6.07 (s, 1H, 4'-H), 3.95 (s, 3H, 3'- CH_3), 2.71 (s, 3H, *p*- CH_3), 2.70 (s, 3H, *p*- CH_3), 2.67 (s, 6H, *p*- CH_3). ^{13}C NMR (125MHz, CDCl_3 , 298 K) δ_{C} = 169.8, 142.3, 140.0, 139.8, 139.6, 139.1, 138.6, 138.4. 138.4, 138.0, 137.9, 137.8, 137.3, 136.5, 133.9, 133.8, 133.3, 131.5, 130.4, 130.2, 129.8, 129.5, 129.1, 129.0, 128.8, 127.9, 127.0, 126.4, 122.7, 121.6, 120.2, 117.5, 115.6, 112.9, 40.3, 22.0, 21.7, 21.5. UV-vis (CH_2Cl_2) λ_{max} /nm (log ϵ): 266 (4.56), 303 sh, 361 (4.07), 381

(4.11), 427 sh, 453 (5.23), 491 sh, 531 (4.22), 572 (4.06), 618 (3.31). MALDI-TOF-HRMS calc. for $[C_{51}H_{38}AgAuClN_5]^+$ (M)⁺: 1059.1532, Found: 1059.190; calc. for $[C_{51}H_{38}AgAuClKN_5]^+$ ($M+K$)⁺: 1098.1164, Found: 1098.157.

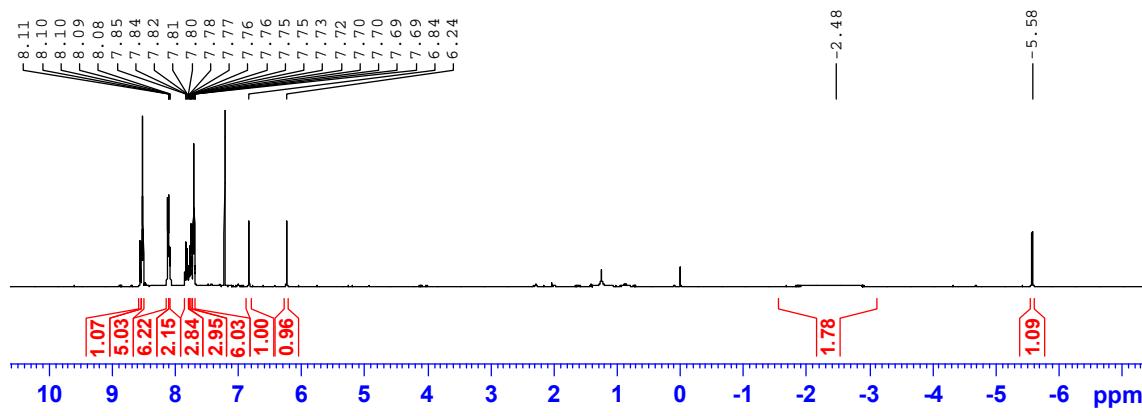


Figure S1. ^1H NMR spectrum of **2a**, 298 K, CDCl_3 .

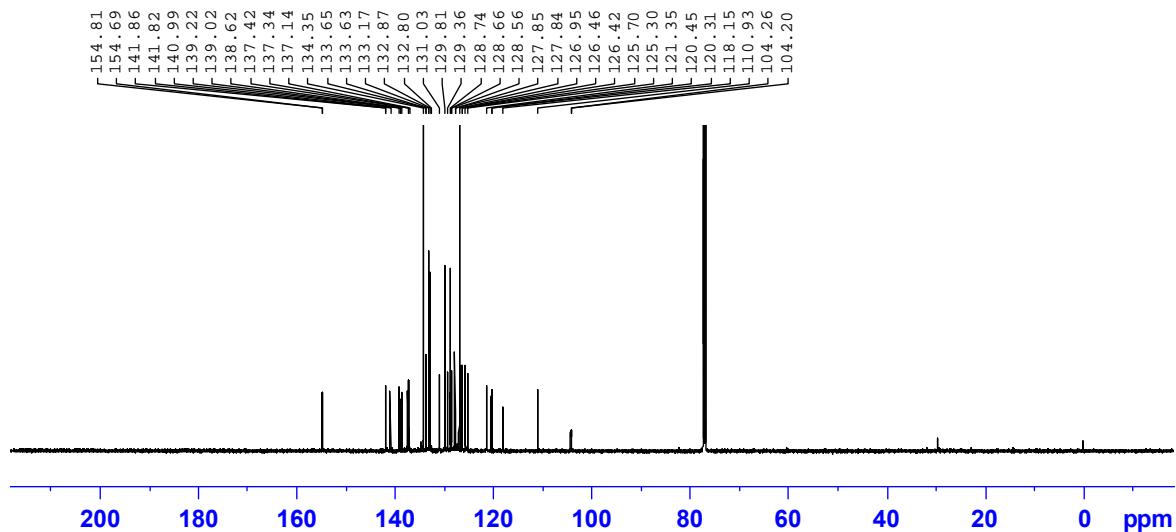


Figure S2. ^{13}C NMR spectrum of **2a**, 298 K, CDCl_3 .

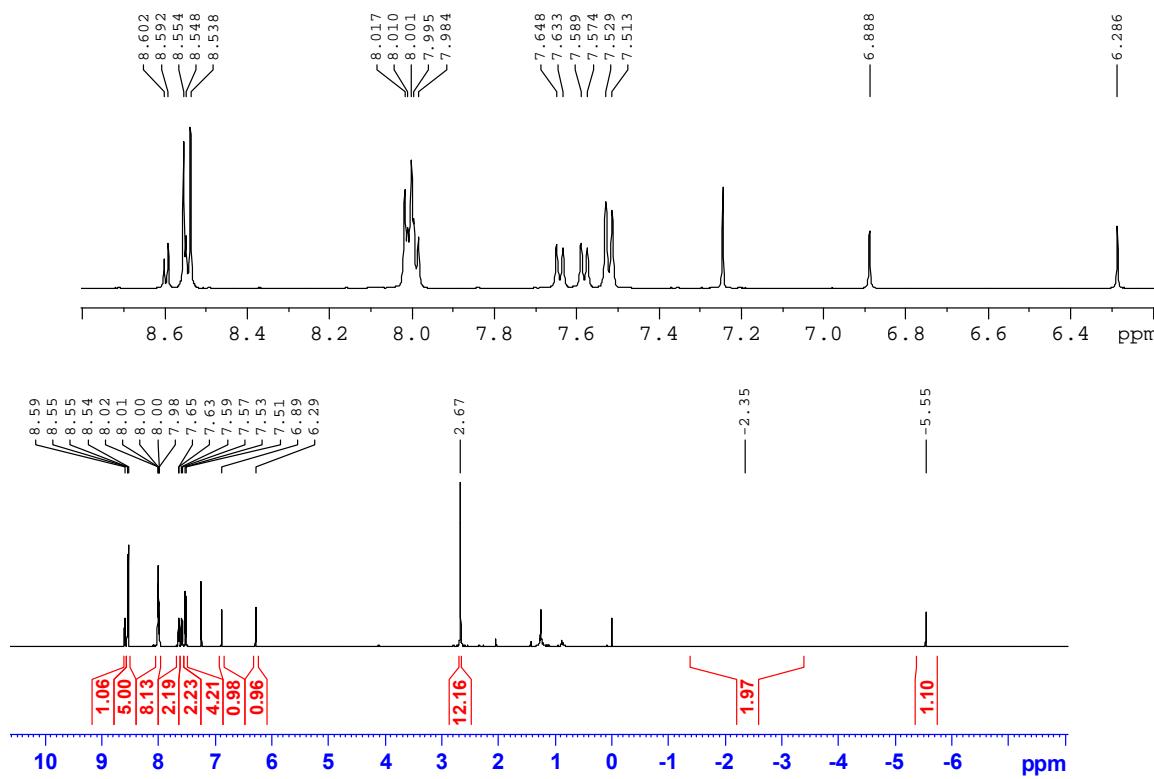


Figure S3. ^1H NMR spectrum of **2b**, 298 K, CDCl_3

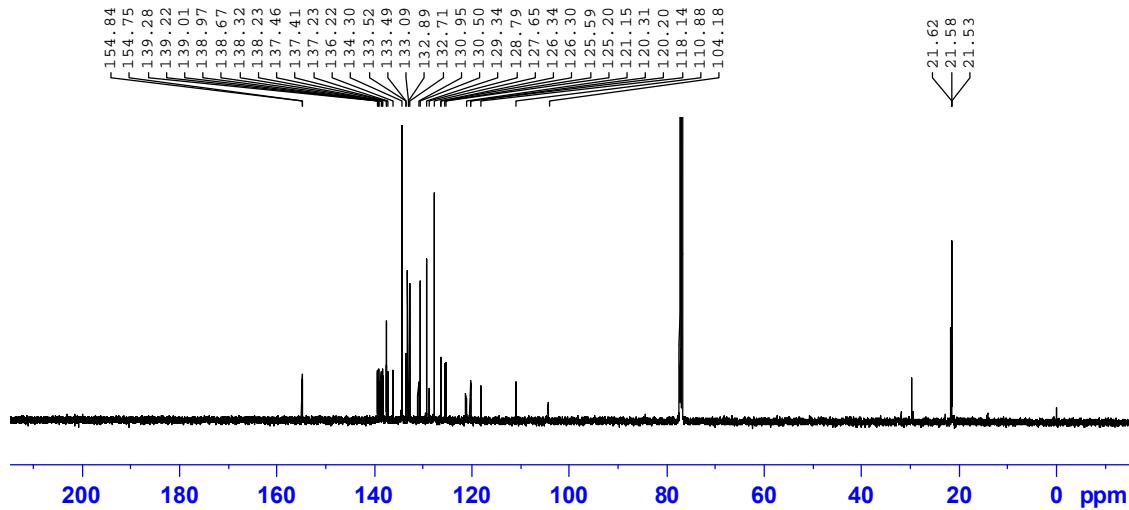


Figure S4. ^{13}C NMR spectrum of **2b**, 298 K, CDCl_3 .

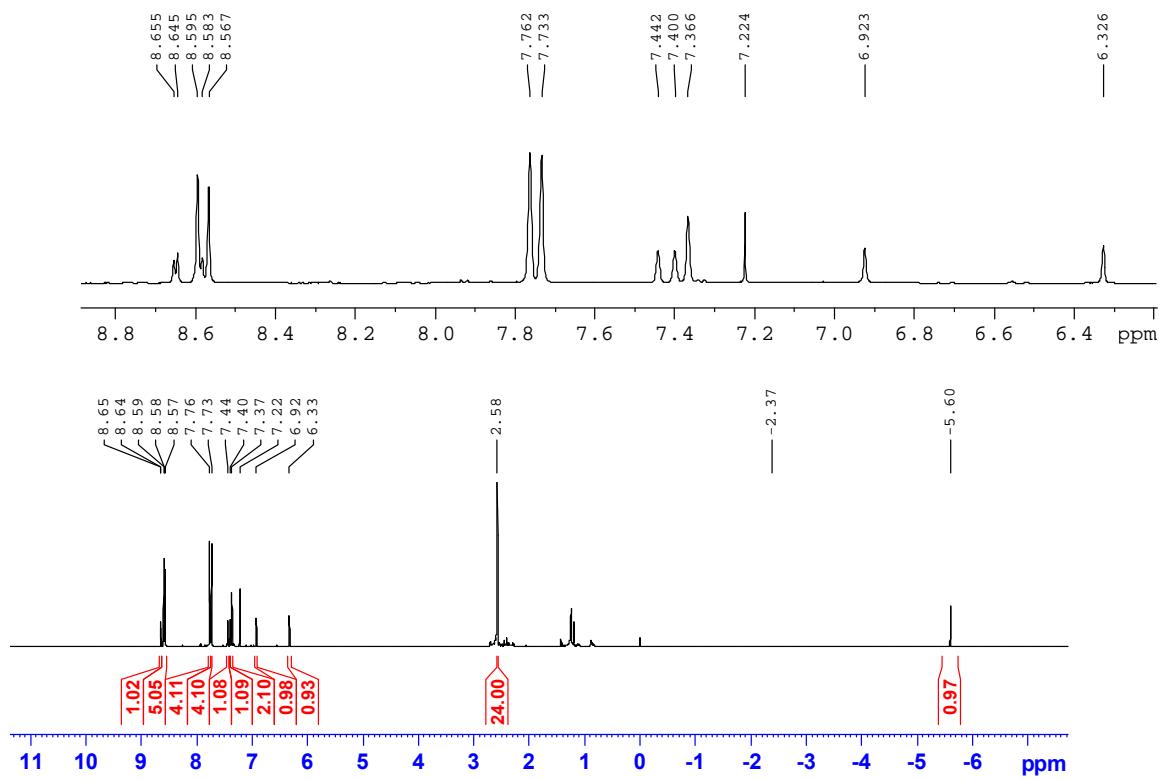


Figure S5. ¹H NMR spectrum of **2c**, 298 K, CDCl₃.

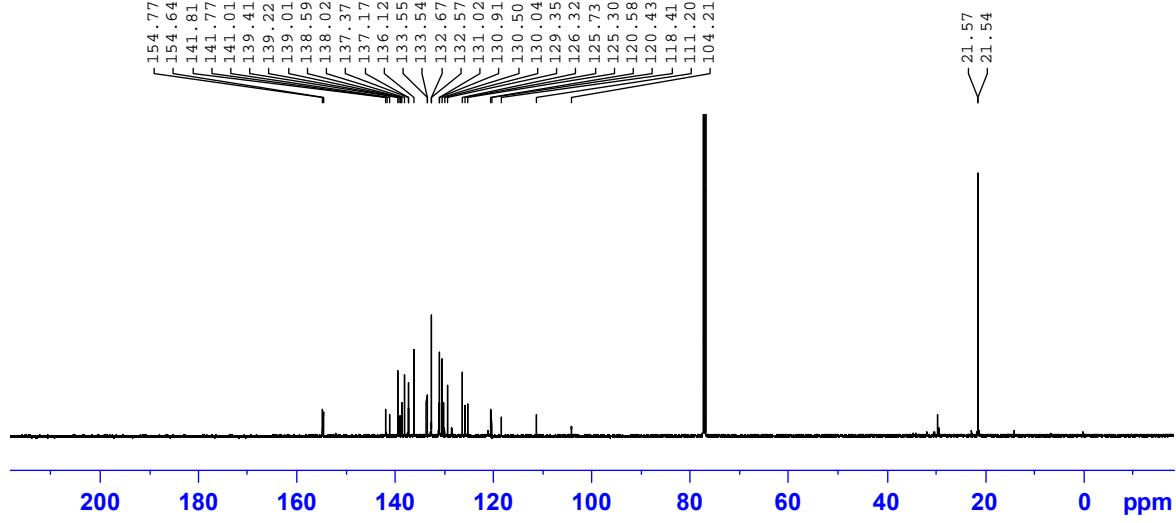


Figure S6. ¹³C NMR spectrum of **2c**, 298 K, CDCl₃.

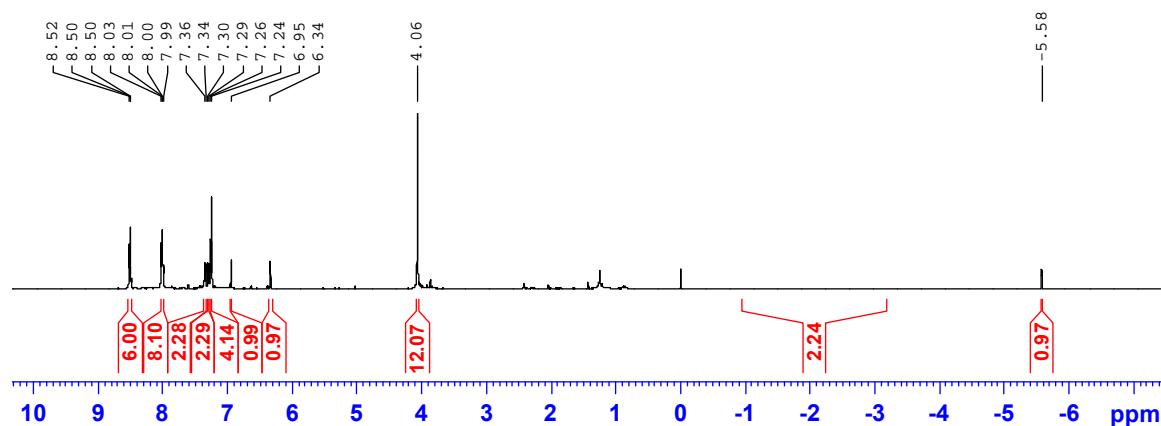


Figure S7. ¹H NMR spectrum of **2d**, 298 K, CDCl₃.

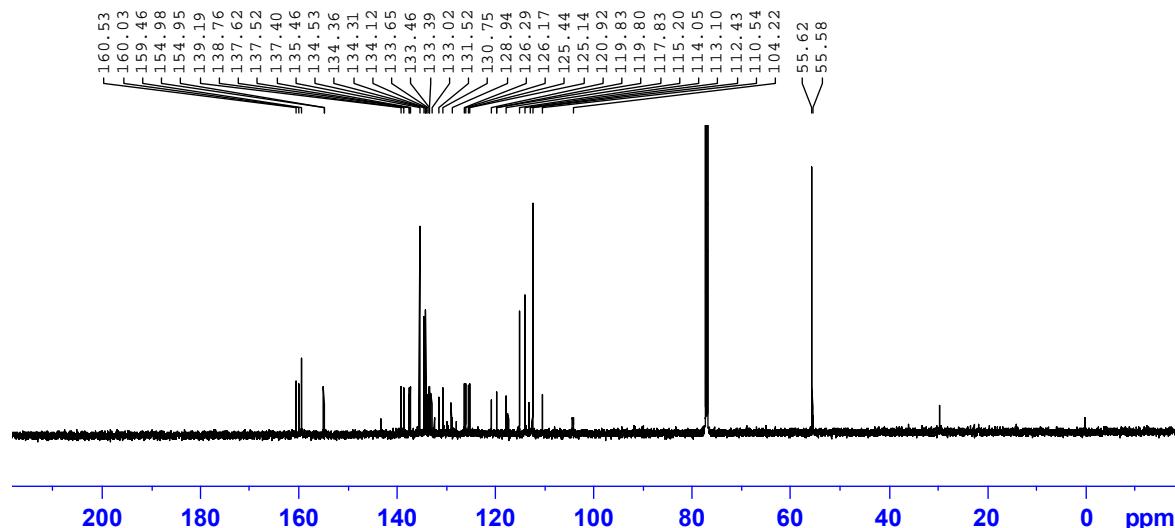


Figure S8. ¹³C NMR spectrum of **2d**, 298 K, CDCl₃.

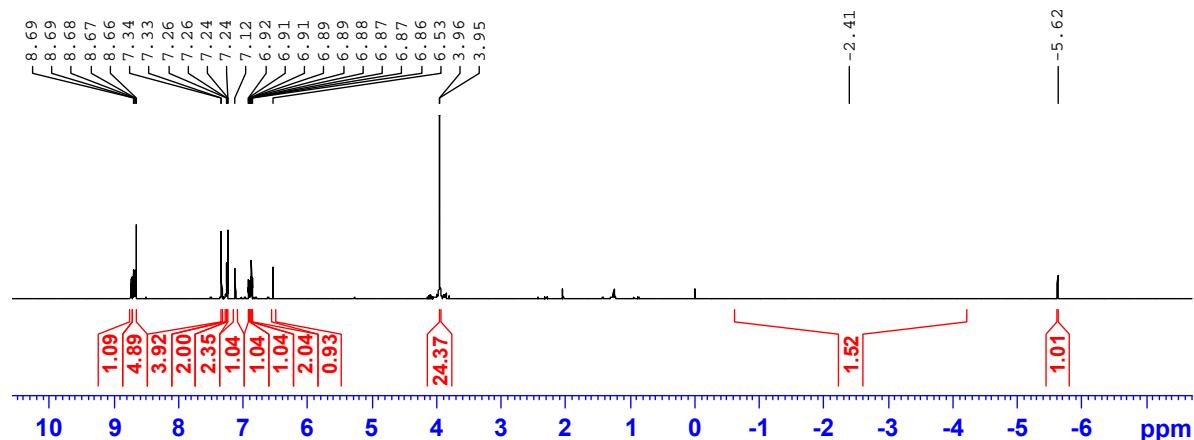


Figure S9. ¹H NMR spectrum of **2e**, 298 K, CDCl₃.

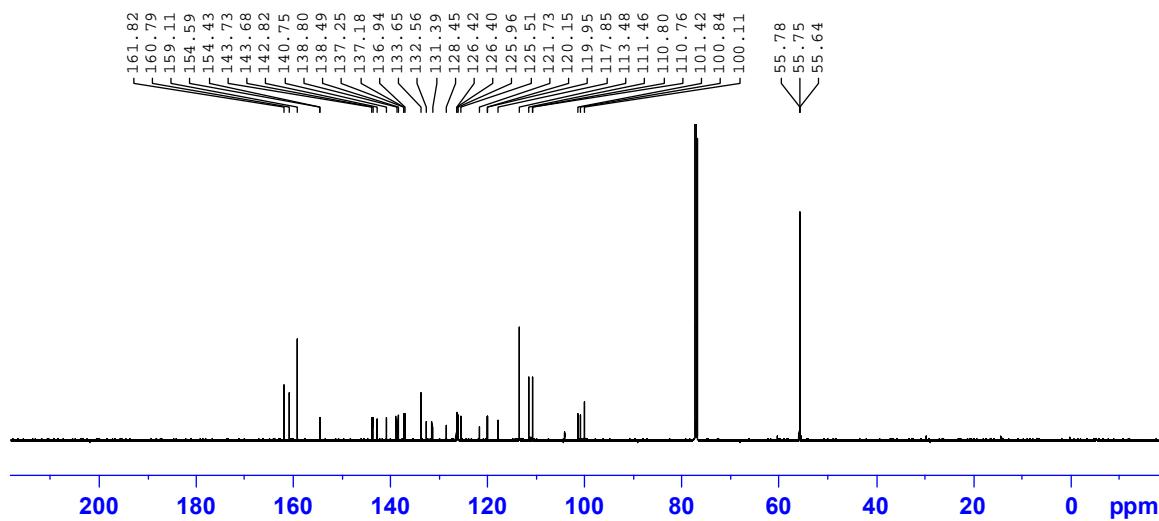


Figure S10. ¹³C NMR spectrum of **2e**, 298 K, CDCl₃.

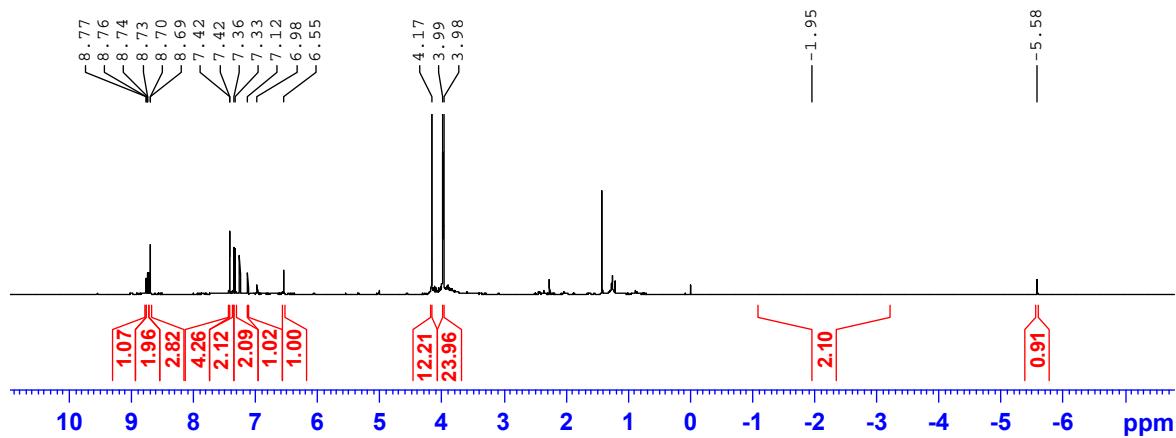


Figure S11. ¹H NMR spectrum of **2f**, 298 K, CDCl₃.

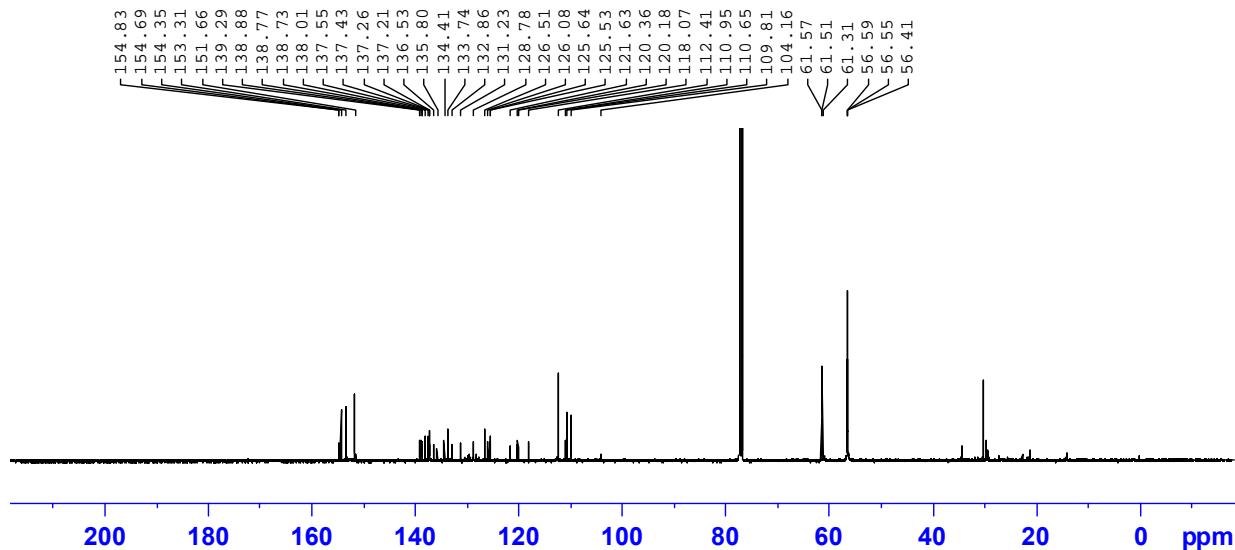


Figure S12. ¹³C NMR spectrum of **2f**, 298 K, CDCl₃.

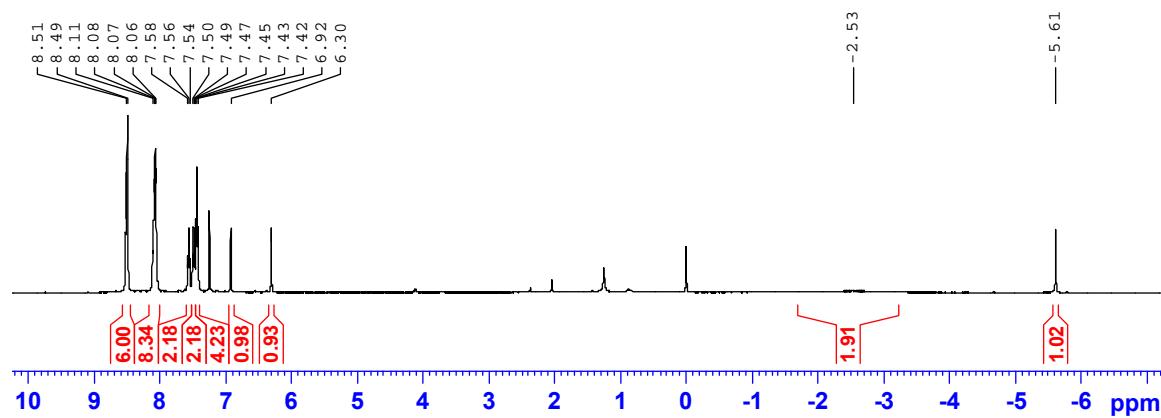


Figure S13. ¹H NMR spectrum of **2g**, 298 K, CDCl₃.

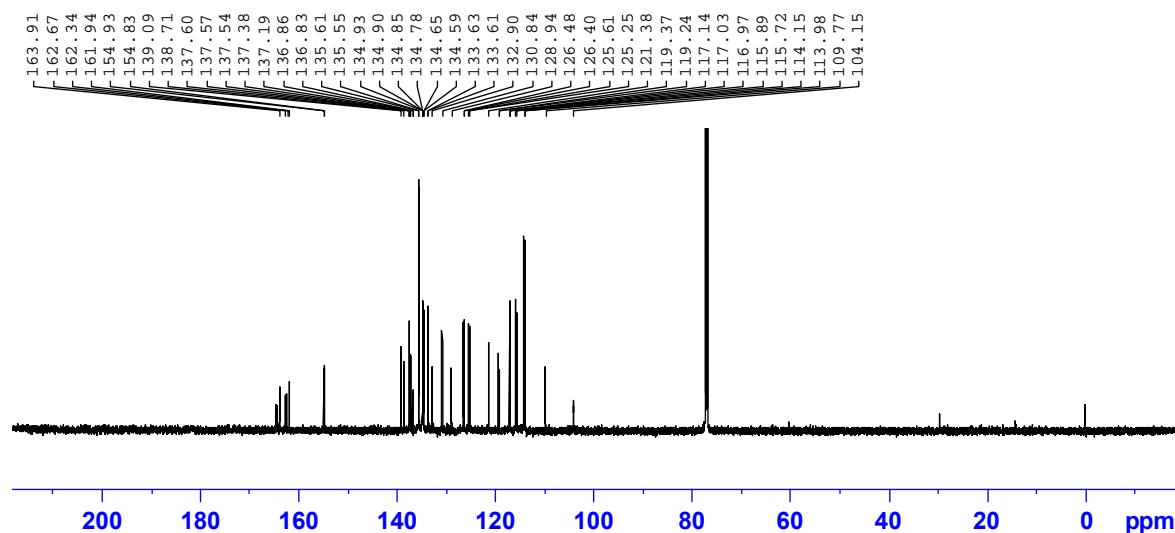


Figure S14. ¹³C NMR spectrum of **2g**, 298 K, CDCl₃.

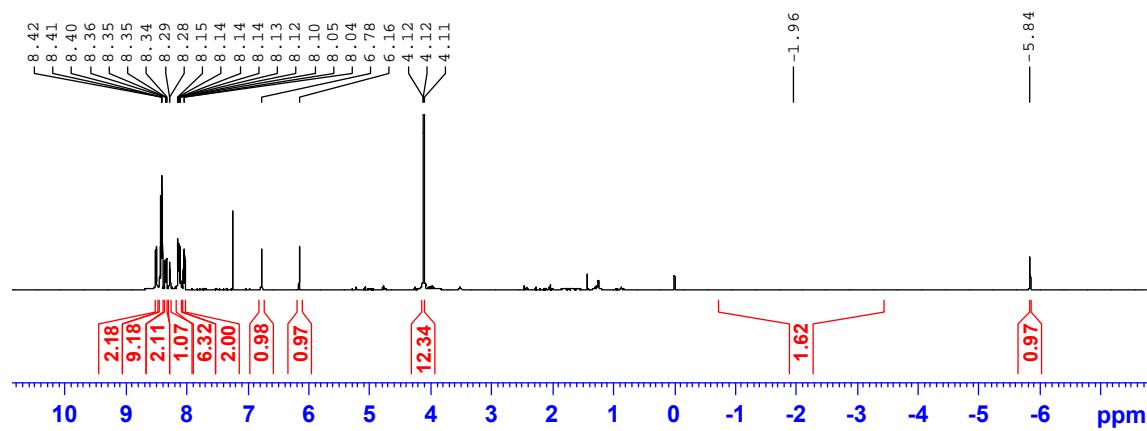


Figure S15. ¹H NMR spectrum of **2h**, 298 K, CDCl₃.

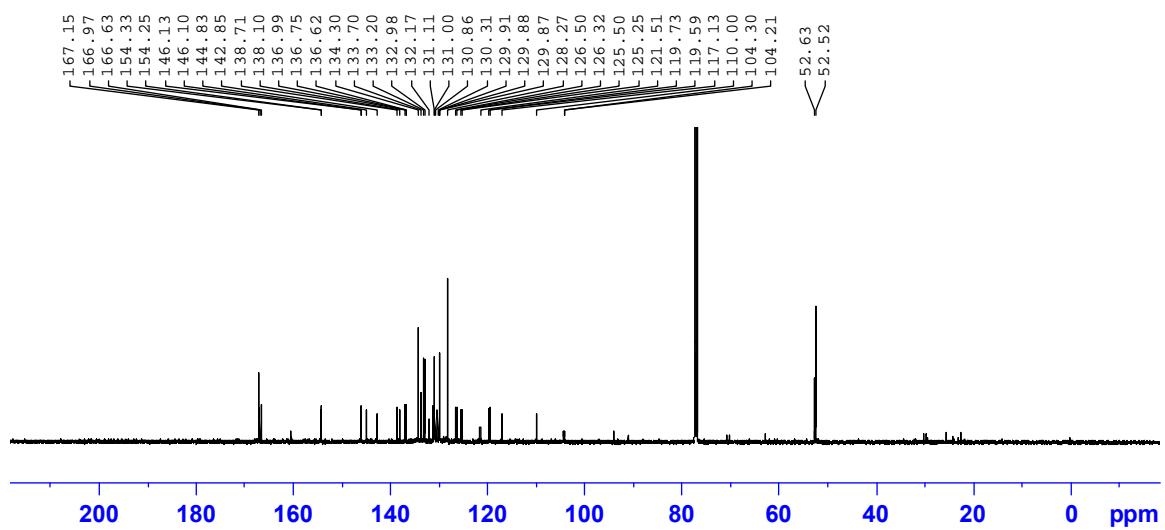
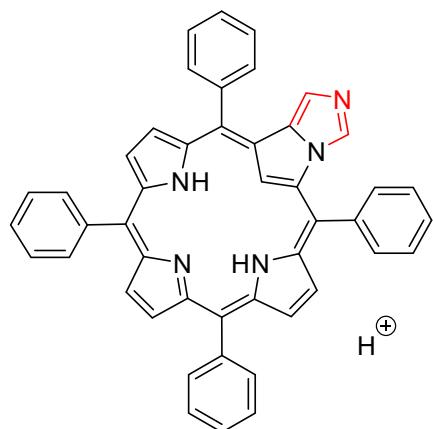


Figure S16. ¹³C NMR spectrum of **2h**, 298 K, CDCl_3 .



Chemical Formula: C₄₆H₃₂N₅⁺
Exact Mass: 654.2652

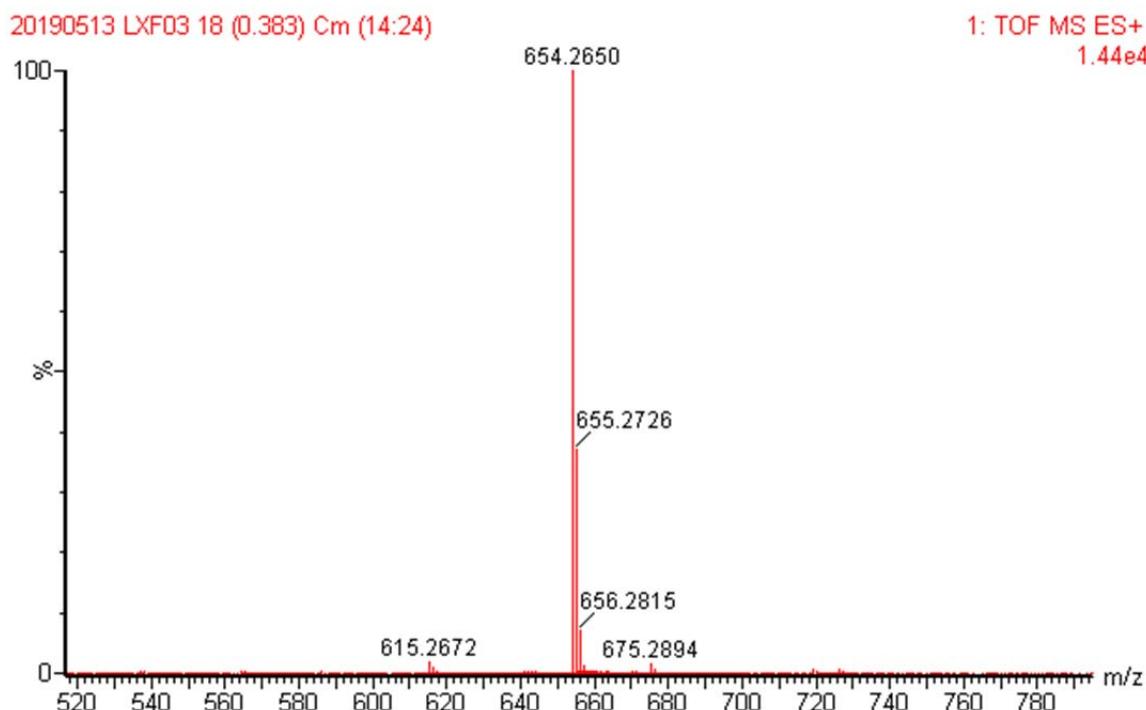
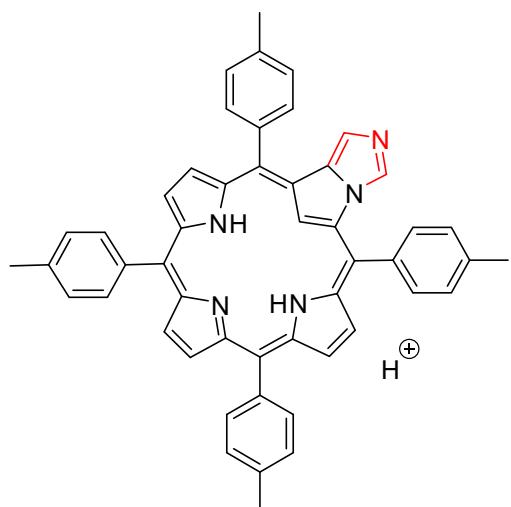


Figure S17. HRMS spectrum of 2a.



Chemical Formula: $C_{50}H_{40}N_5^+$
Exact Mass: 710.3278

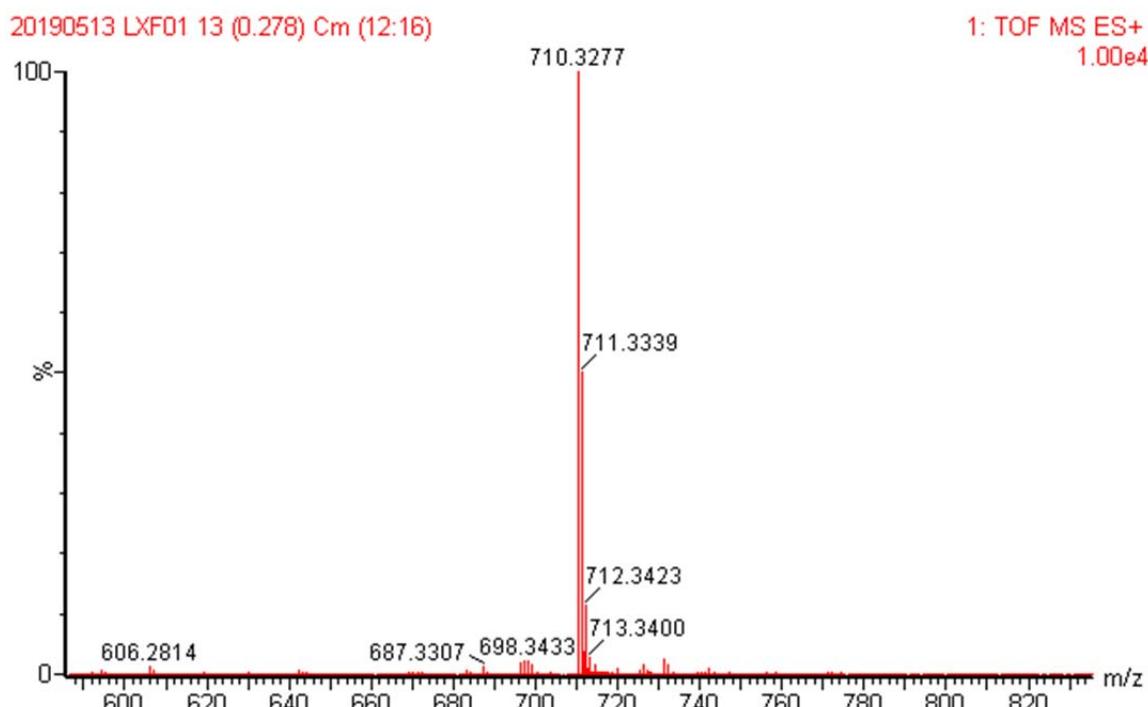
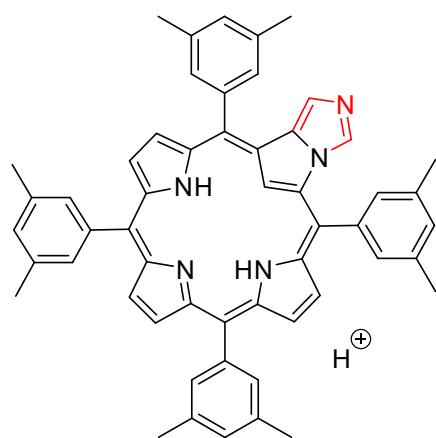


Figure S18. HRMS spectrum of **2b**.



Chemical Formula: C₅₄H₄₈N₅⁺

Exact Mass: 766.3904

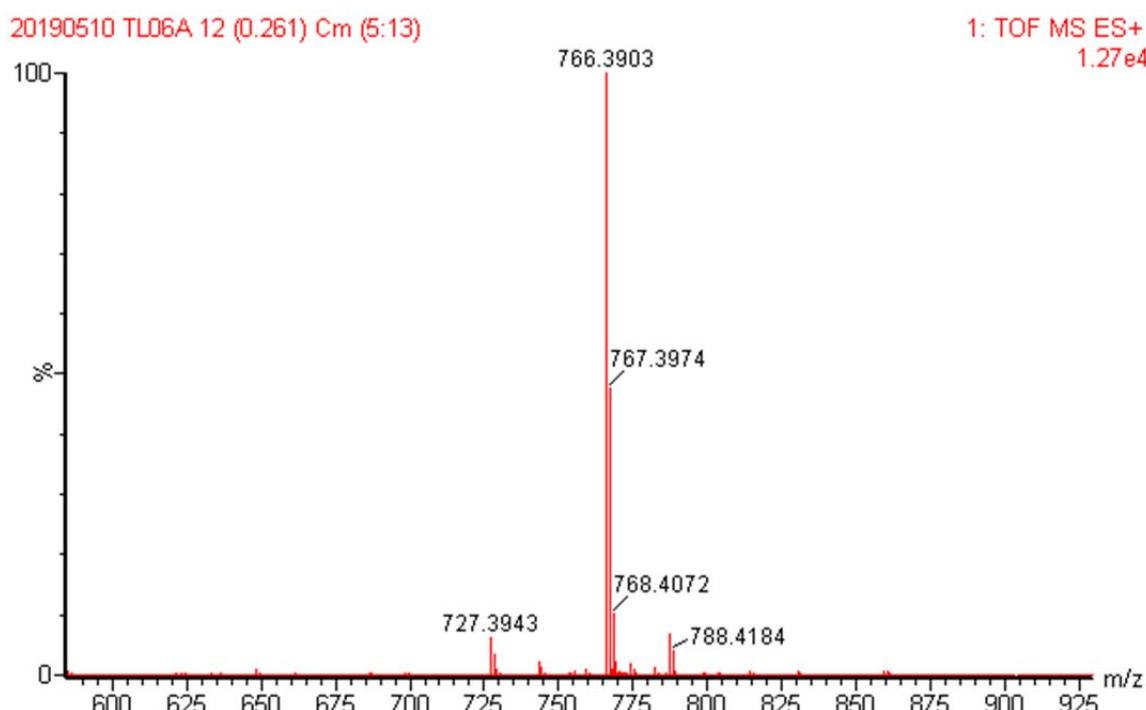
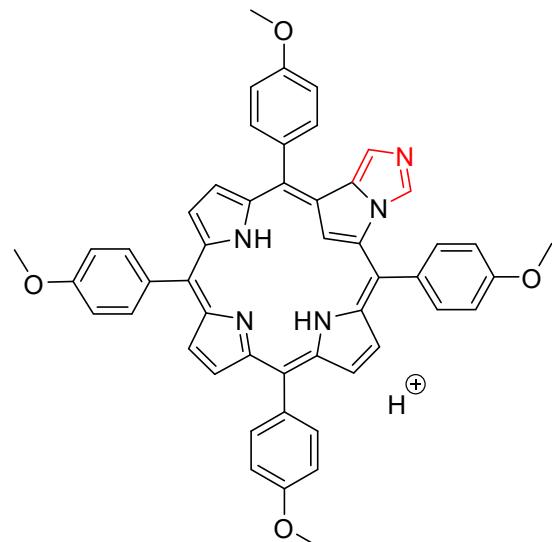


Figure S19. HRMS spectrum of **2c**.



Chemical Formula: $C_{50}H_{40}N_5O_4^+$
Exact Mass: 774.3075

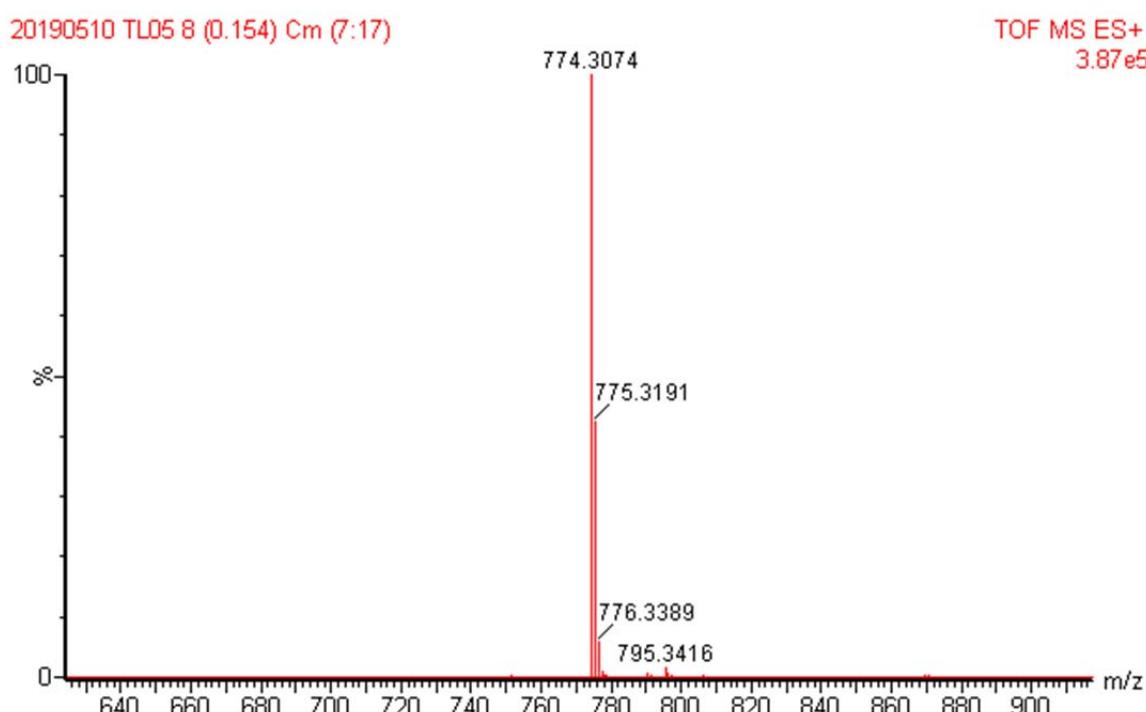
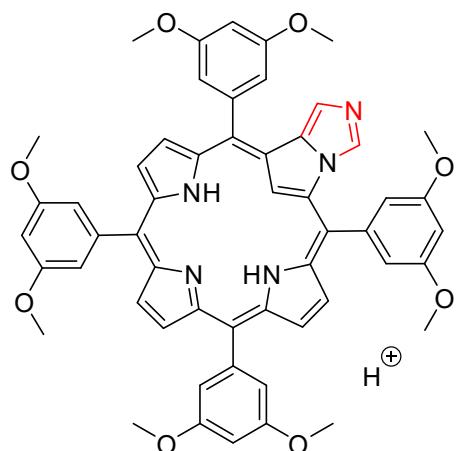


Figure S20. HRMS spectrum of **2d**.



Chemical Formula: $\text{C}_{54}\text{H}_{48}\text{N}_5\text{O}_8^+$
Exact Mass: 894.3497

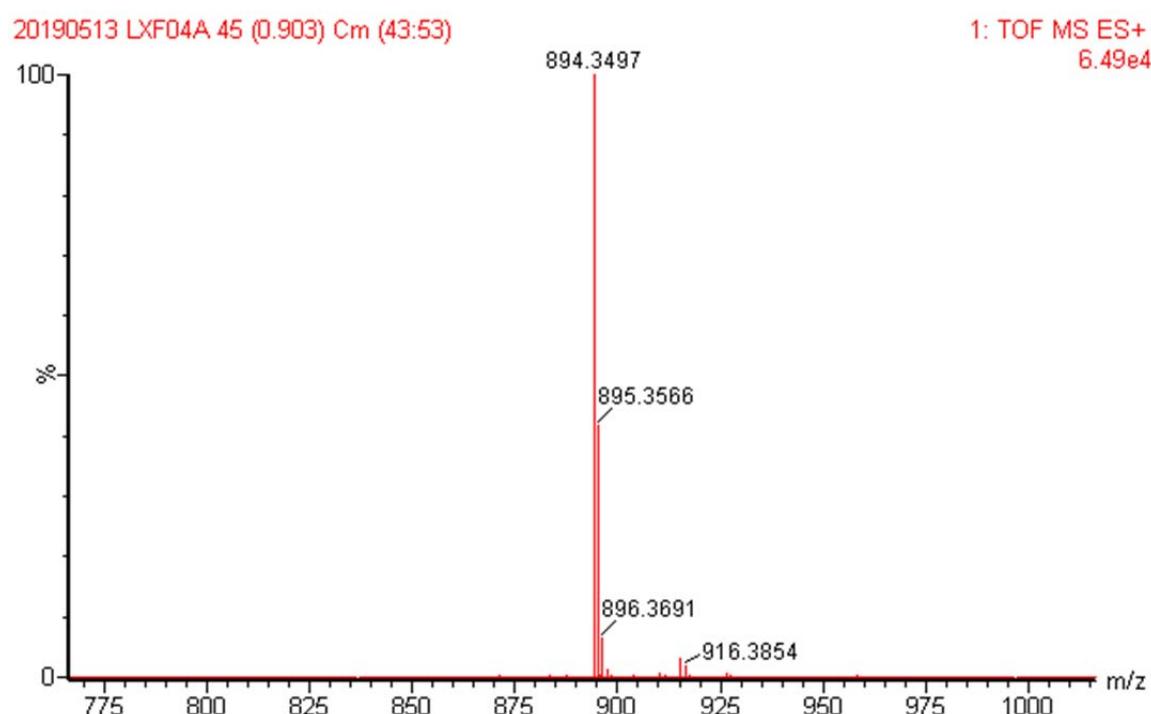
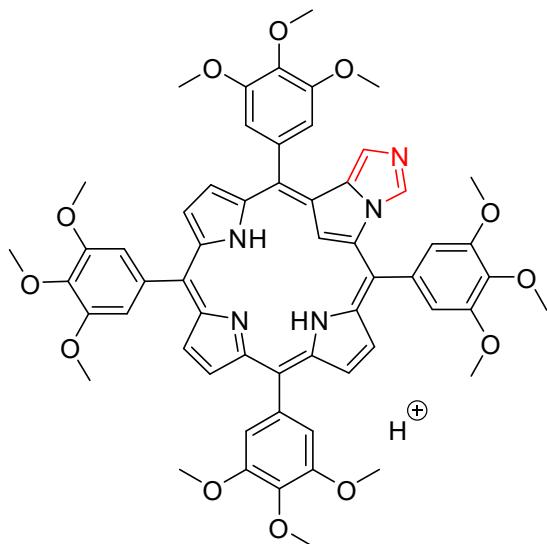


Figure S21. HRMS spectrum of **2e**.



Chemical Formula: C₅₈H₅₆N₅O₁₂⁺

Exact Mass: 1014.3920

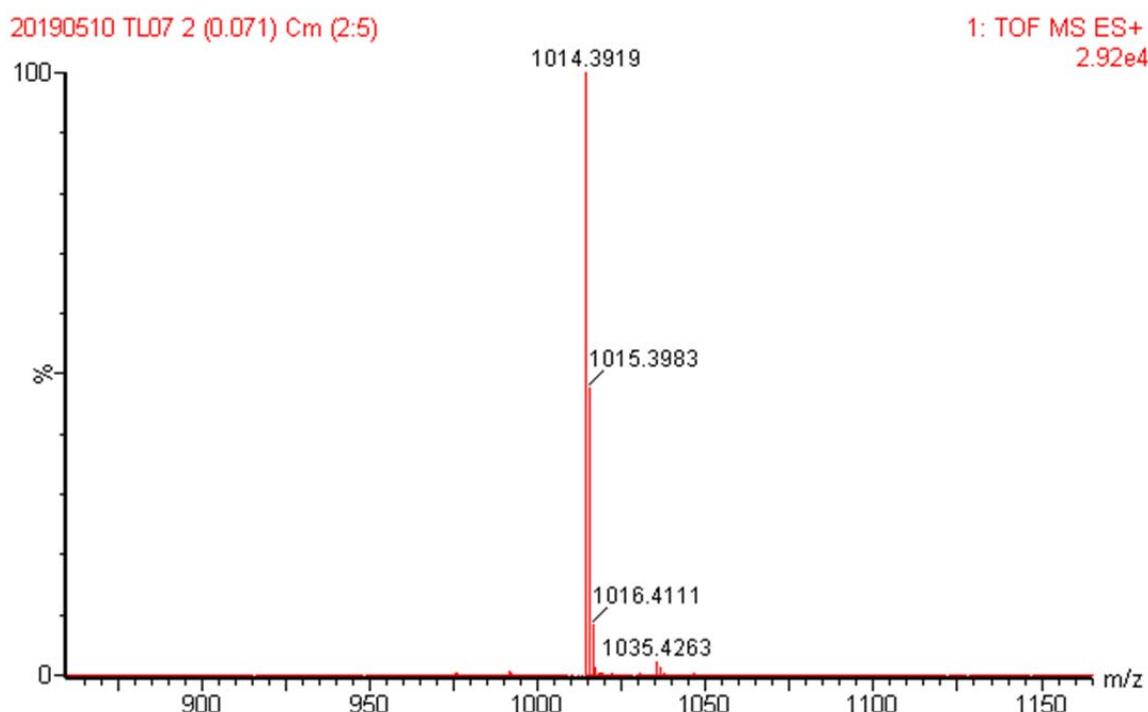


Figure S22. HRMS spectrum of **2f**.

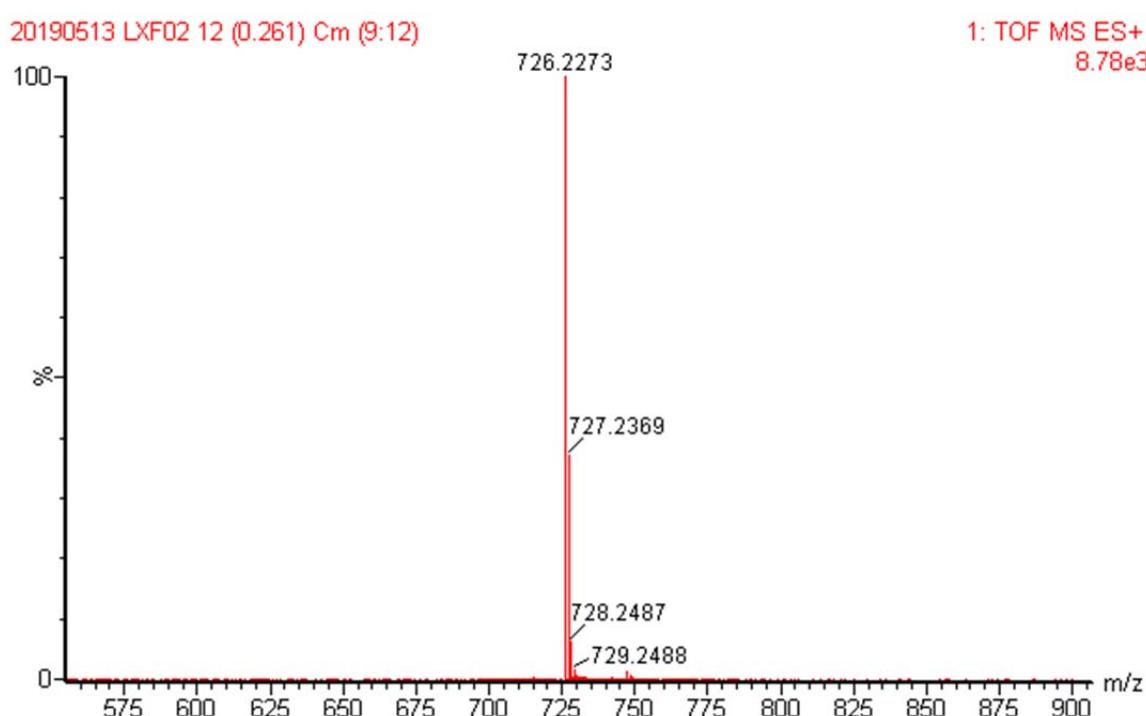
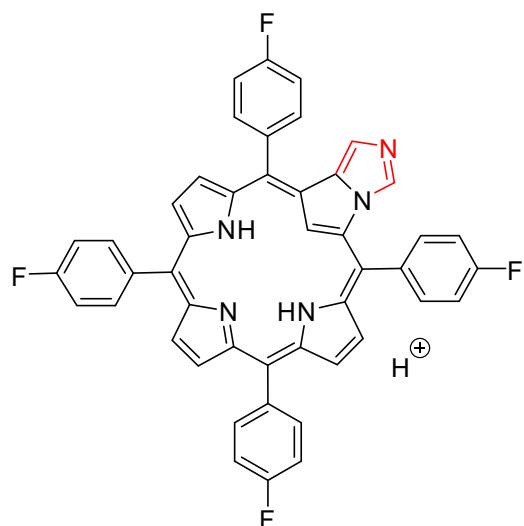


Figure S23. HRMS spectrum of **2g**.

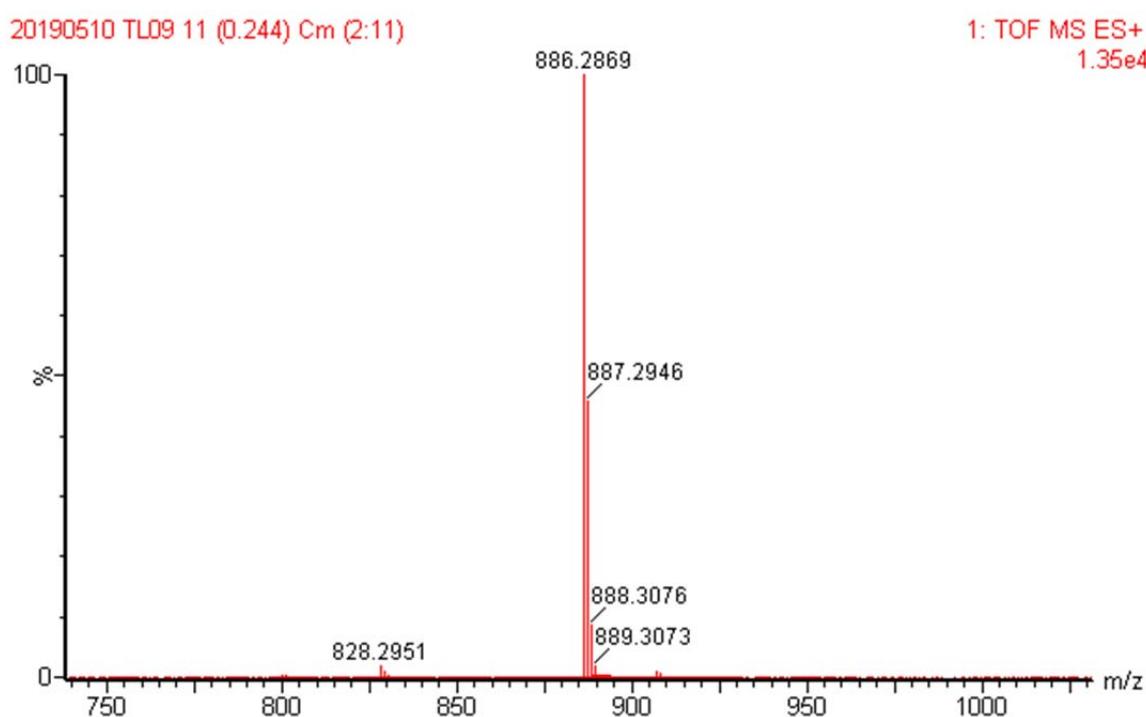
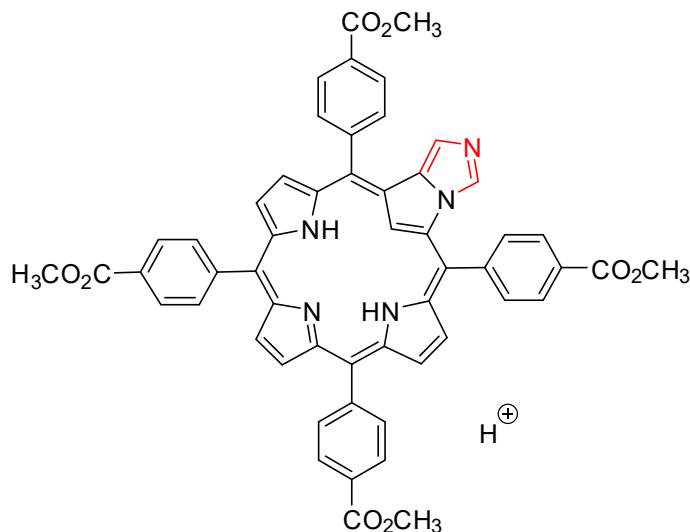


Figure S24. HRMS spectrum of **2h**.

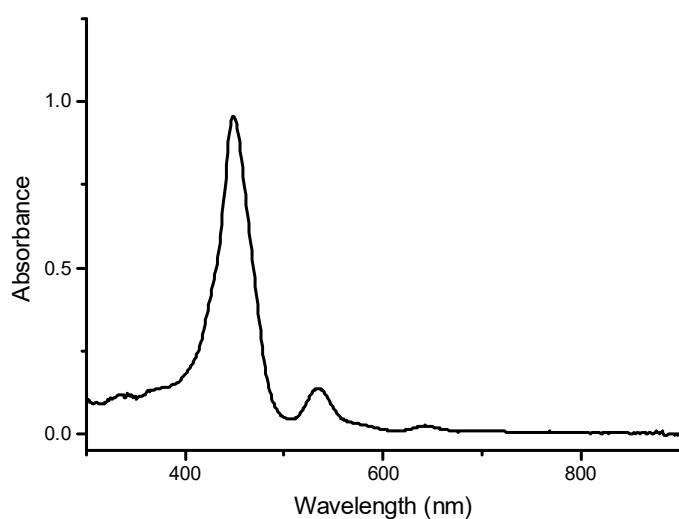


Figure S25. UV-vis spectra of **2a** (CH_2Cl_2 , 5.67×10^{-6}).

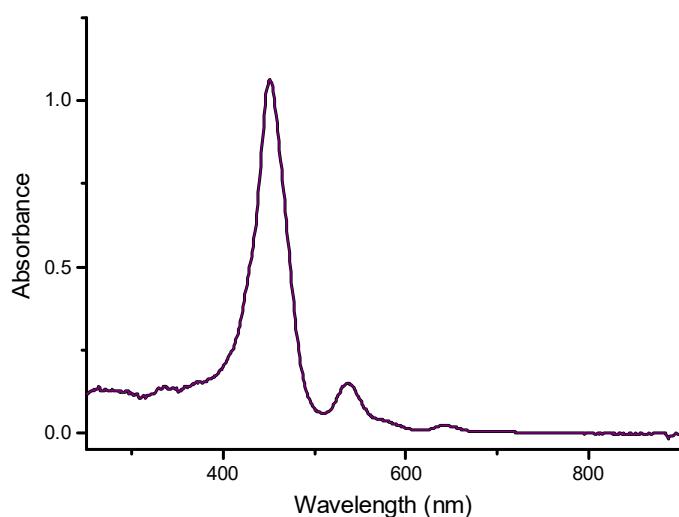


Figure S26. UV-vis spectra of **2b** (CH_2Cl_2 , 6.01×10^{-6}).

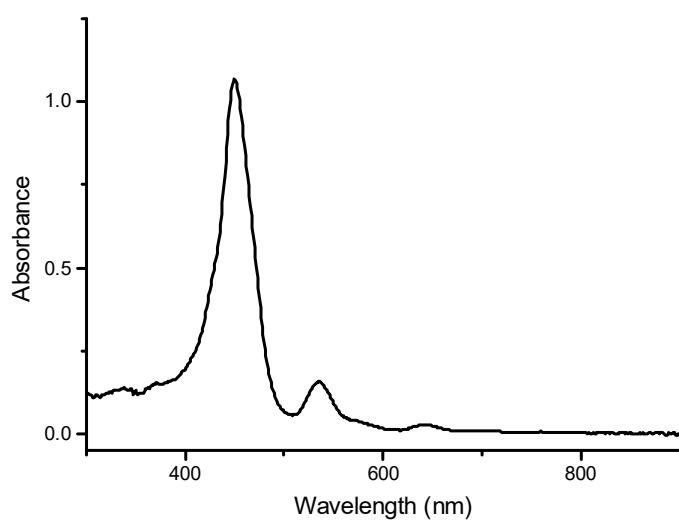


Figure S27. UV-vis spectra of **2c** (CH_2Cl_2 , 6.45×10^{-6})

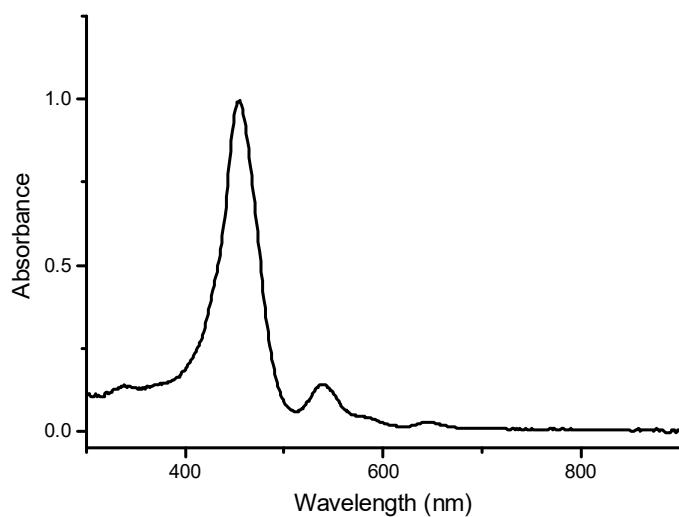


Figure S28. UV-vis spectra of **2d** (CH_2Cl_2 , 5.99×10^{-6}).

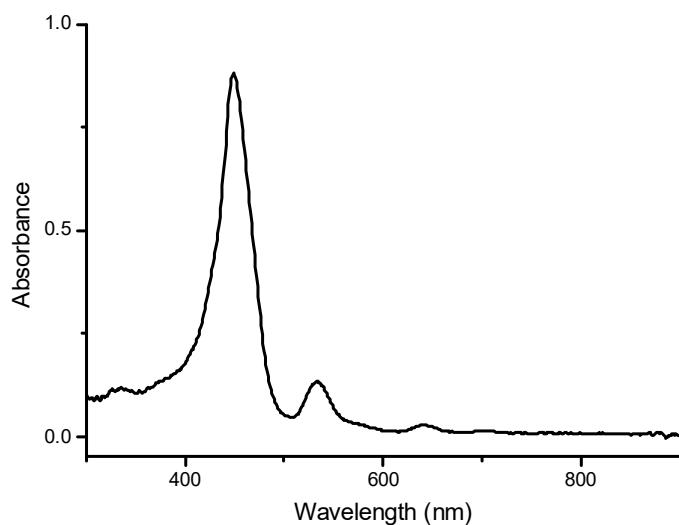


Figure S29. UV-vis spectra of **2e** (CH_2Cl_2 , 4.29×10^{-6}).

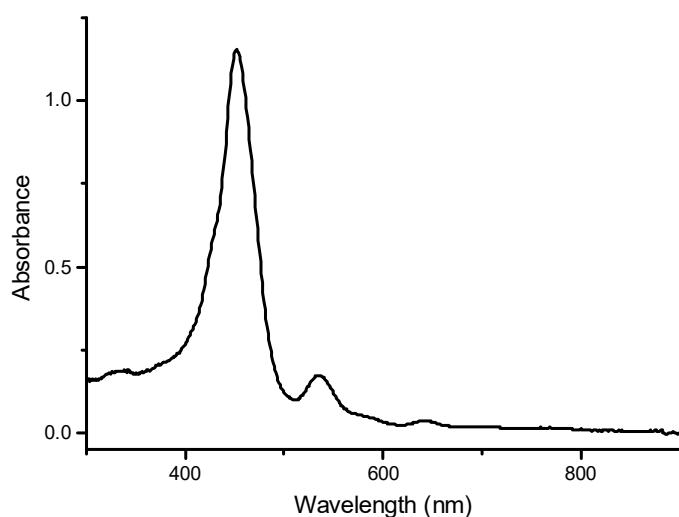


Figure S30. UV-vis spectra of **2f** (CH_2Cl_2 , 7.96×10^{-6}).

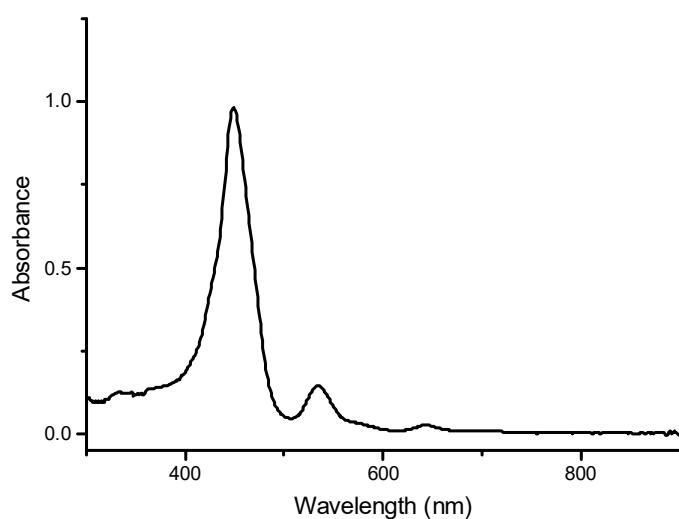


Figure S31. UV-vis spectra of **2g** (CH_2Cl_2 , 5.15×10^{-6}).

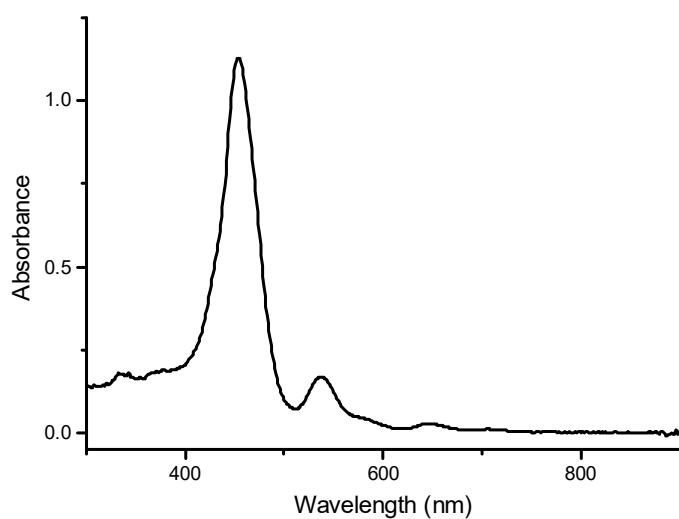


Figure S32. UV-vis spectra of **2h** (CH_2Cl_2 , 5.76×10^{-6}).

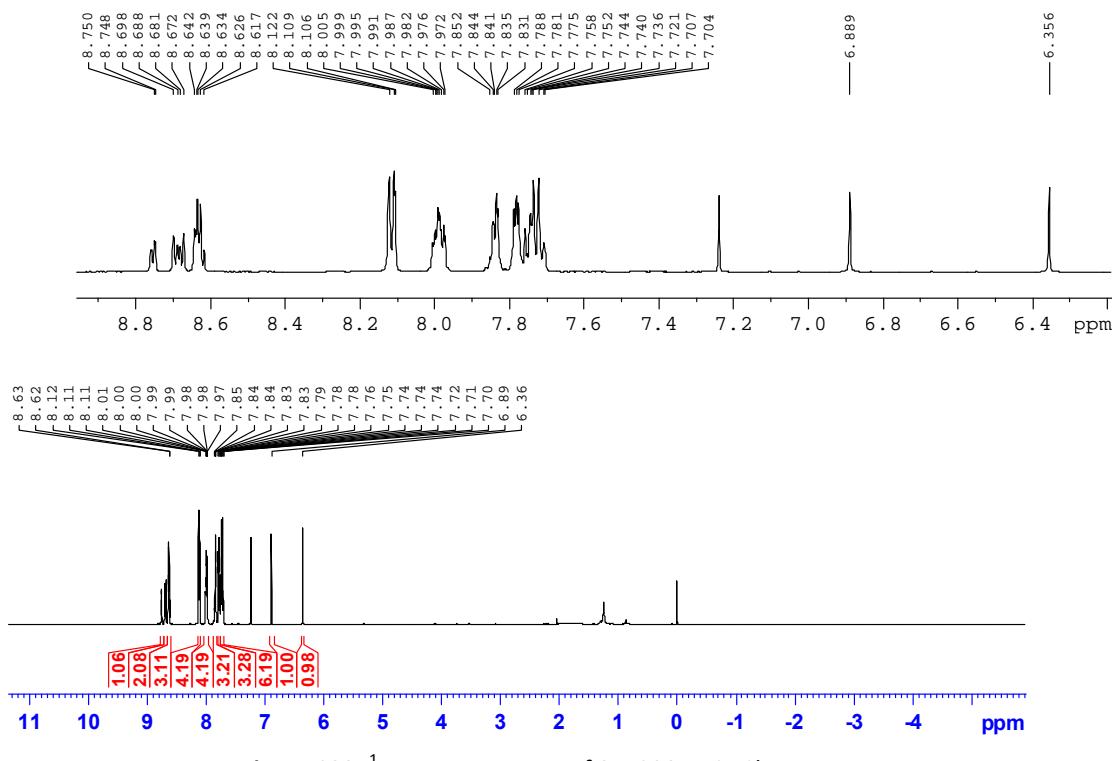


Figure S33. ¹H NMR spectrum of 3a, 298 K, CDCl₃.

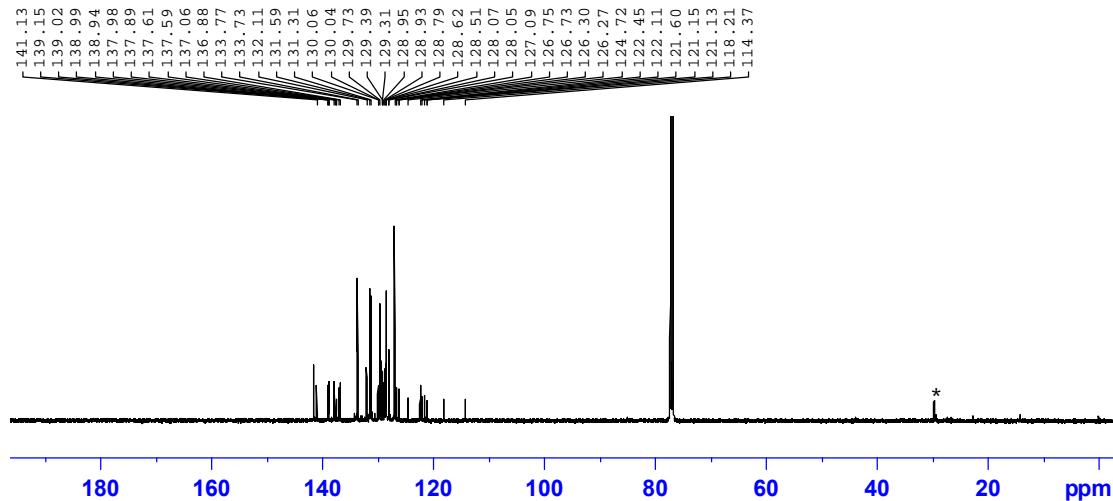


Figure S34. ¹³C NMR spectrum of 3a, 298 K, CDCl₃.

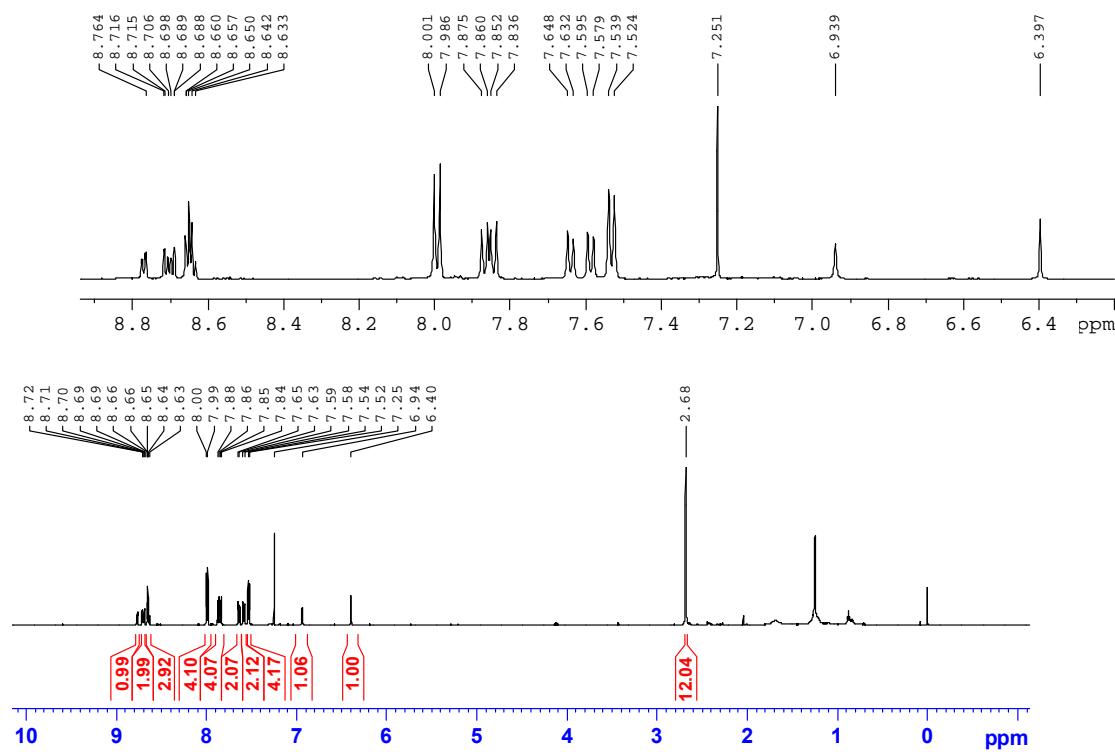


Figure S35. ¹H NMR spectrum of **3b**, 298 K, CDCl₃

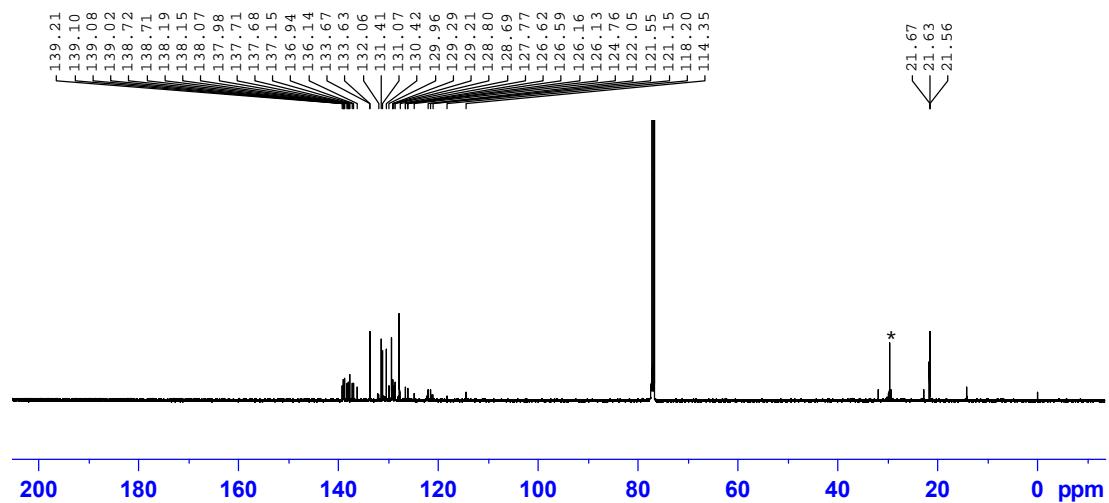
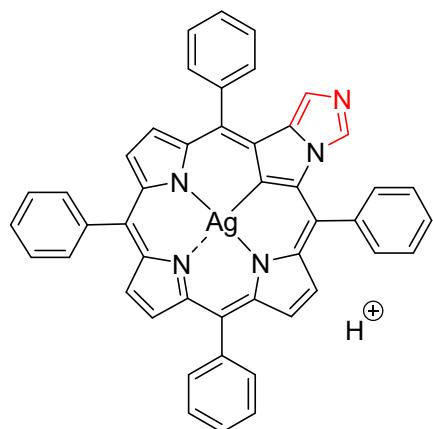


Figure S36. ¹³C NMR spectrum of **3b**, 298 K, CDCl₃.



Chemical Formula: $C_{46}H_{29}AgN_5^+$
Exact Mass: 758.1468

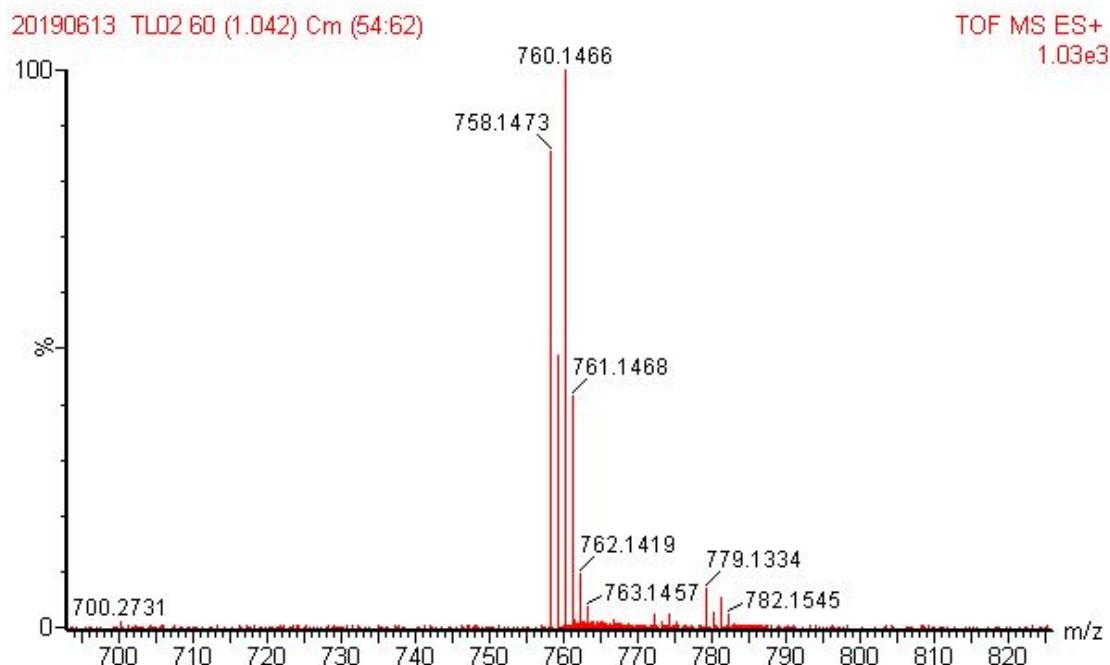
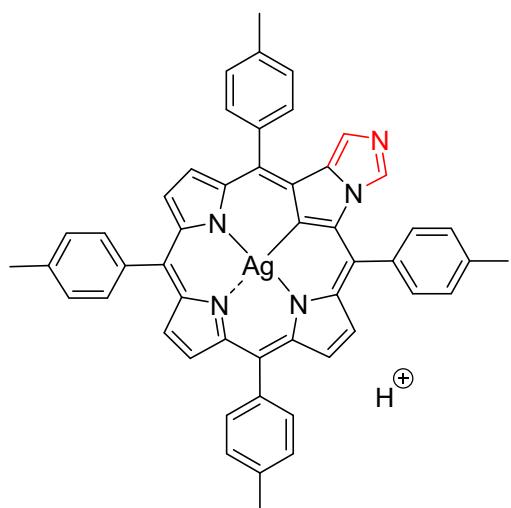


Figure S37. HRMS spectrum of 3a.



Chemical Formula: $C_{50}H_{37}AgN_5^+$
Exact Mass: 814.2094

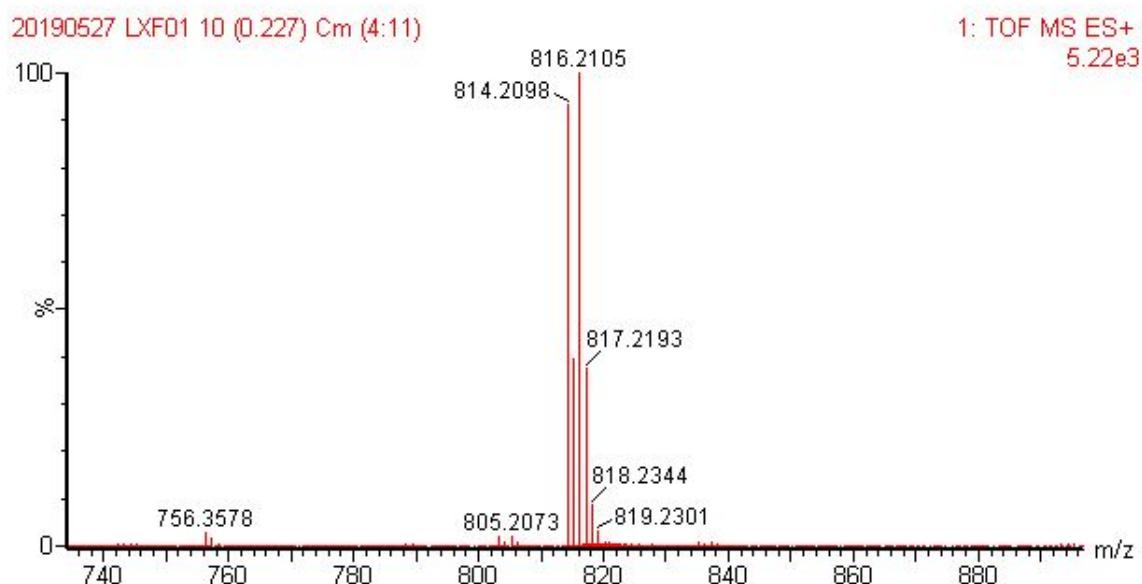


Figure S38. HRMS spectrum of **3b**.

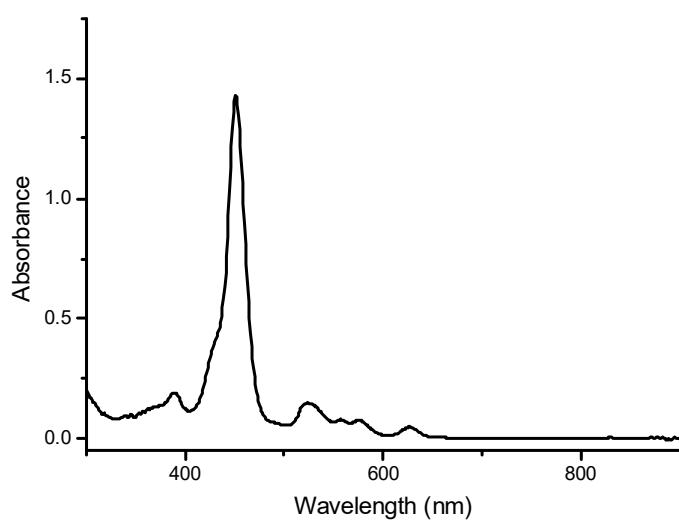


Figure S39. UV-vis spectra of **3a** (CH_2Cl_2 , 7.13×10^{-6})

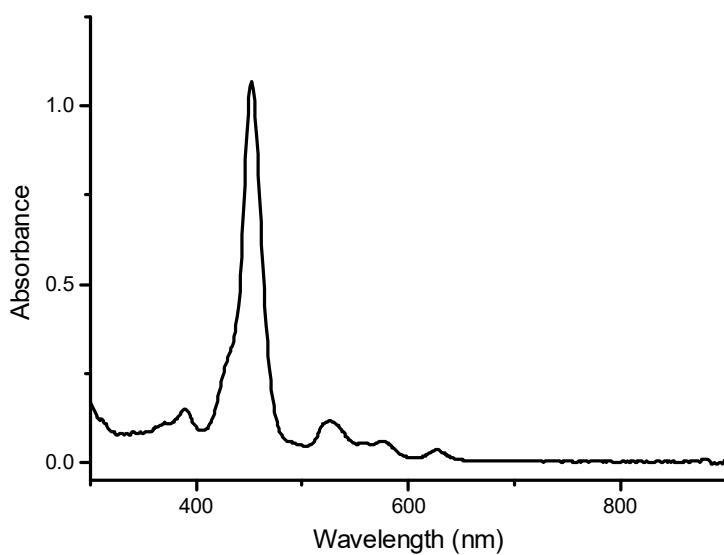


Figure S40. UV-vis spectra of **3b** (CH_2Cl_2 , 7.22×10^{-6}).

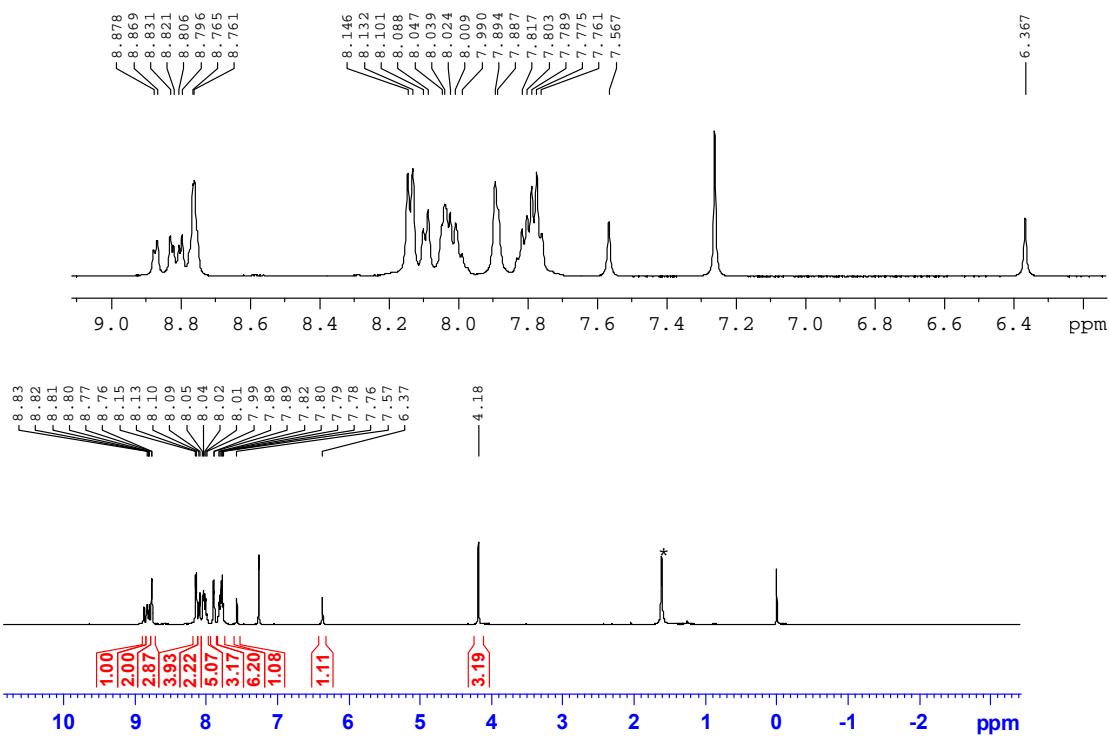


Figure S41. ¹H NMR spectrum of [4a]I, 298 K, CDCl₃.

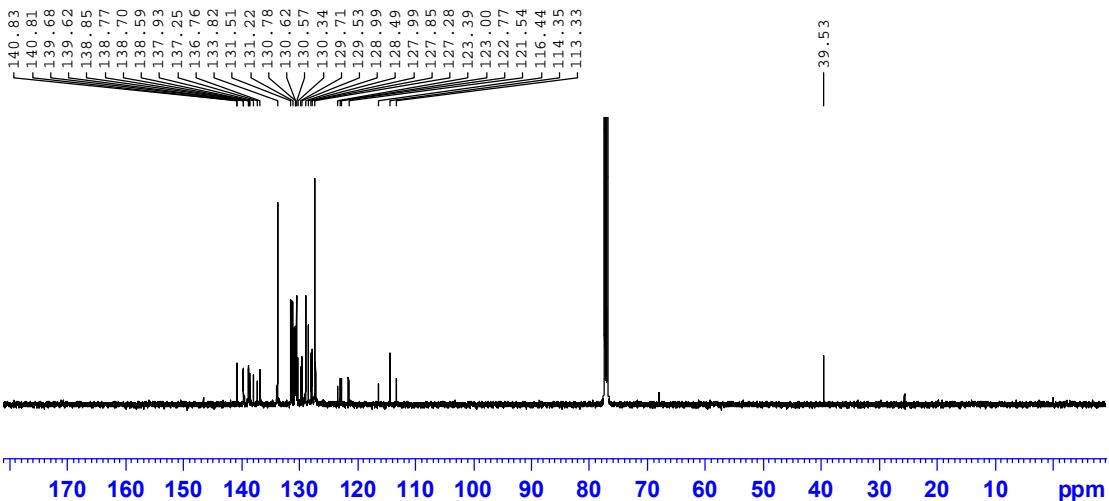


Figure S42. ¹³C NMR spectrum of [4a]I, 298 K, CDCl₃.

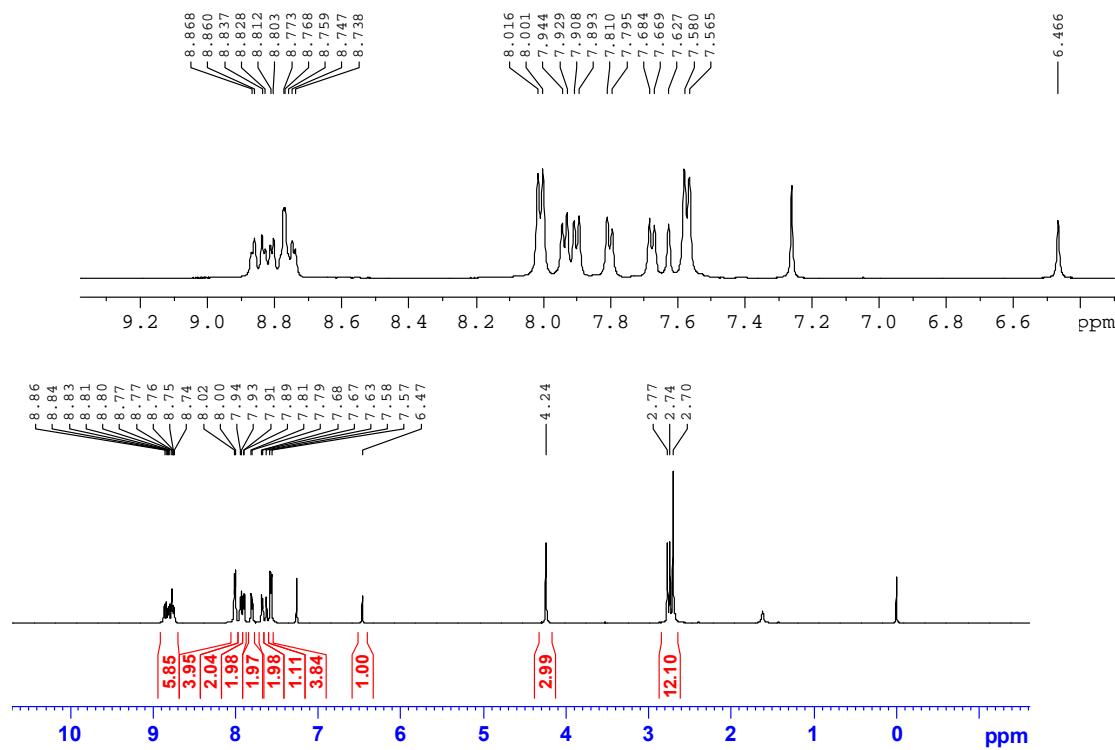


Figure S43. ¹H NMR spectrum of [4b]I, 298 K, CDCl₃

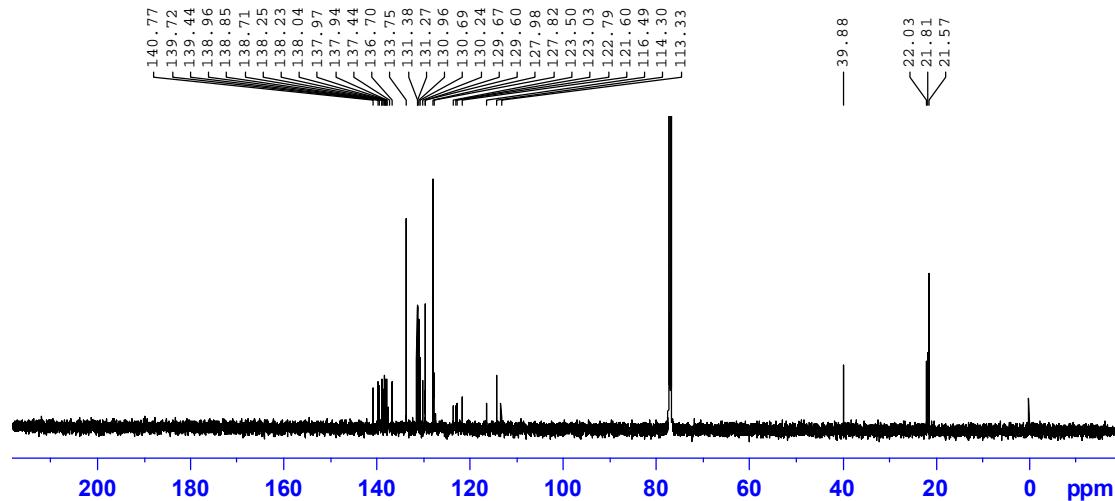
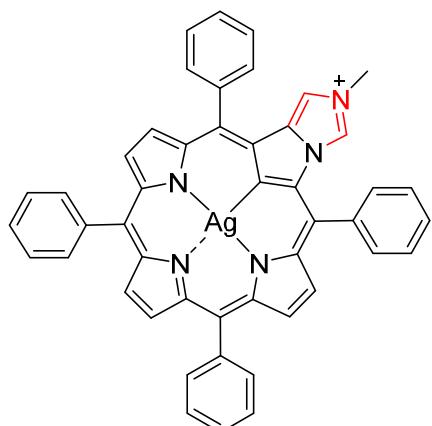


Figure S44. ¹³C NMR spectrum of [4b]I, 298 K, CDCl₃.



Chemical Formula: $C_{47}H_{31}AgN_5^+$

Exact Mass: 772.1625

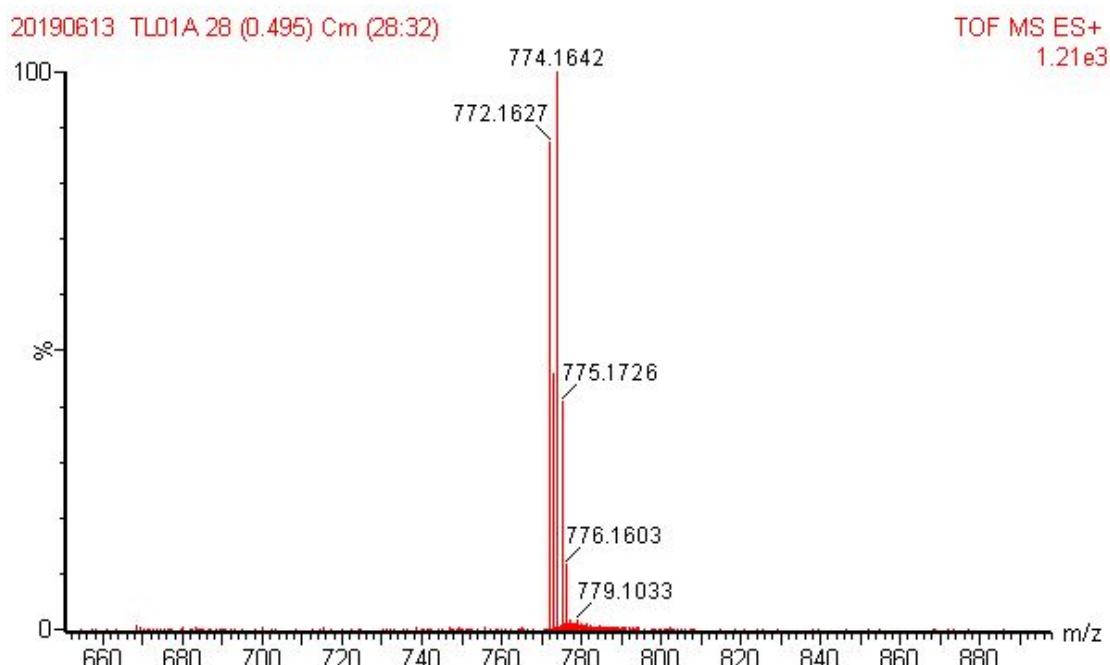
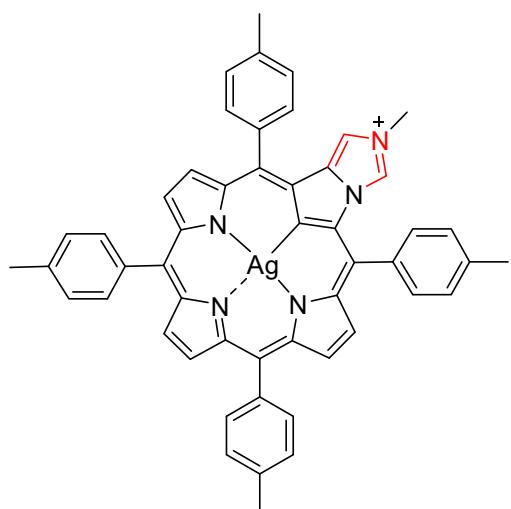


Figure S45. HRMS spectrum of **[4a]I**.



Chemical Formula: $C_{51}H_{39}AgN_5^+$
Exact Mass: 828.2251

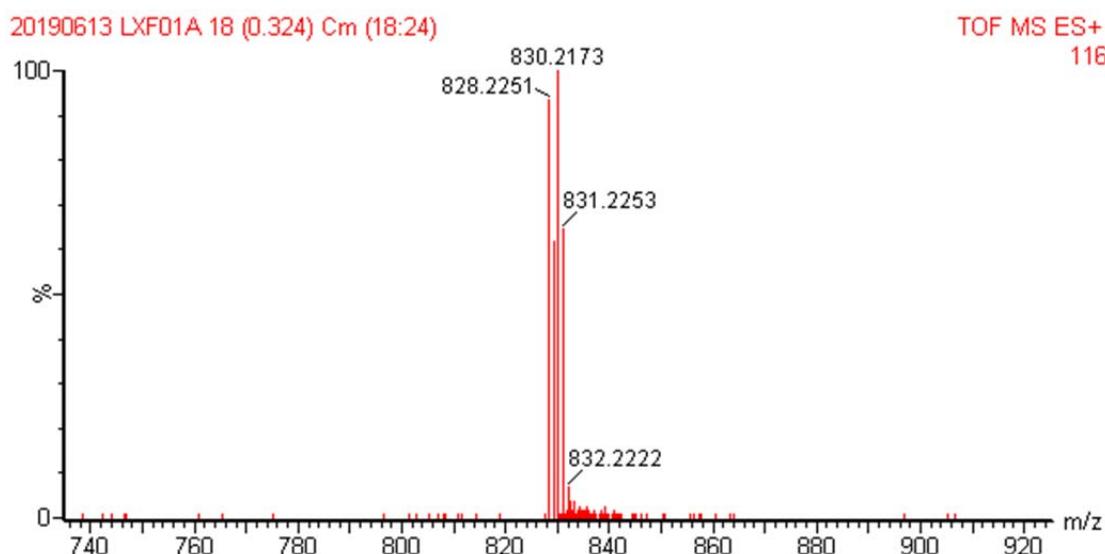


Figure S46. HRMS spectrum of **[4b]I**.

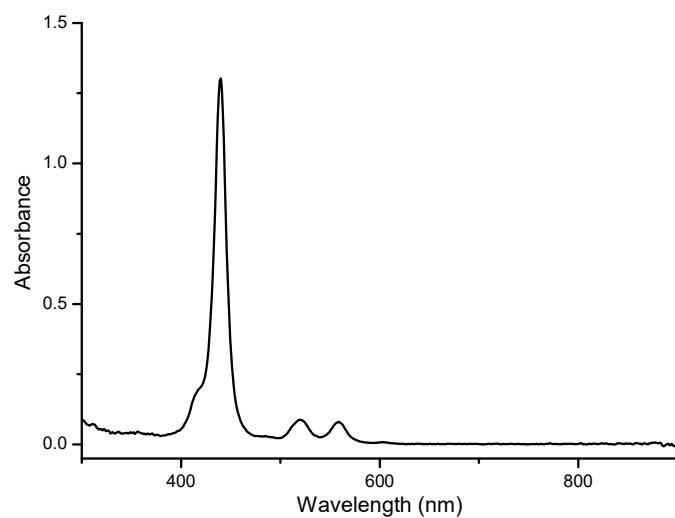


Figure S47. UV-vis spectra of **[4a]I** (CH_2Cl_2 , 3.74×10^{-6}).

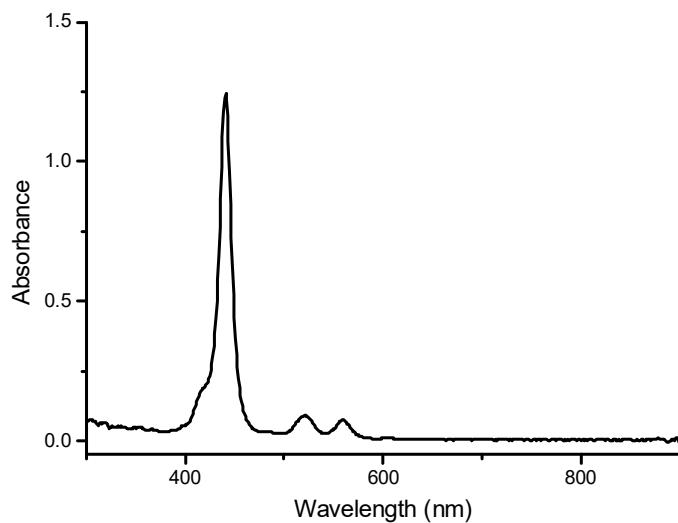


Figure S48. UV-vis spectra of **[4b]I** (CH_2Cl_2 , 3.66×10^{-6}).

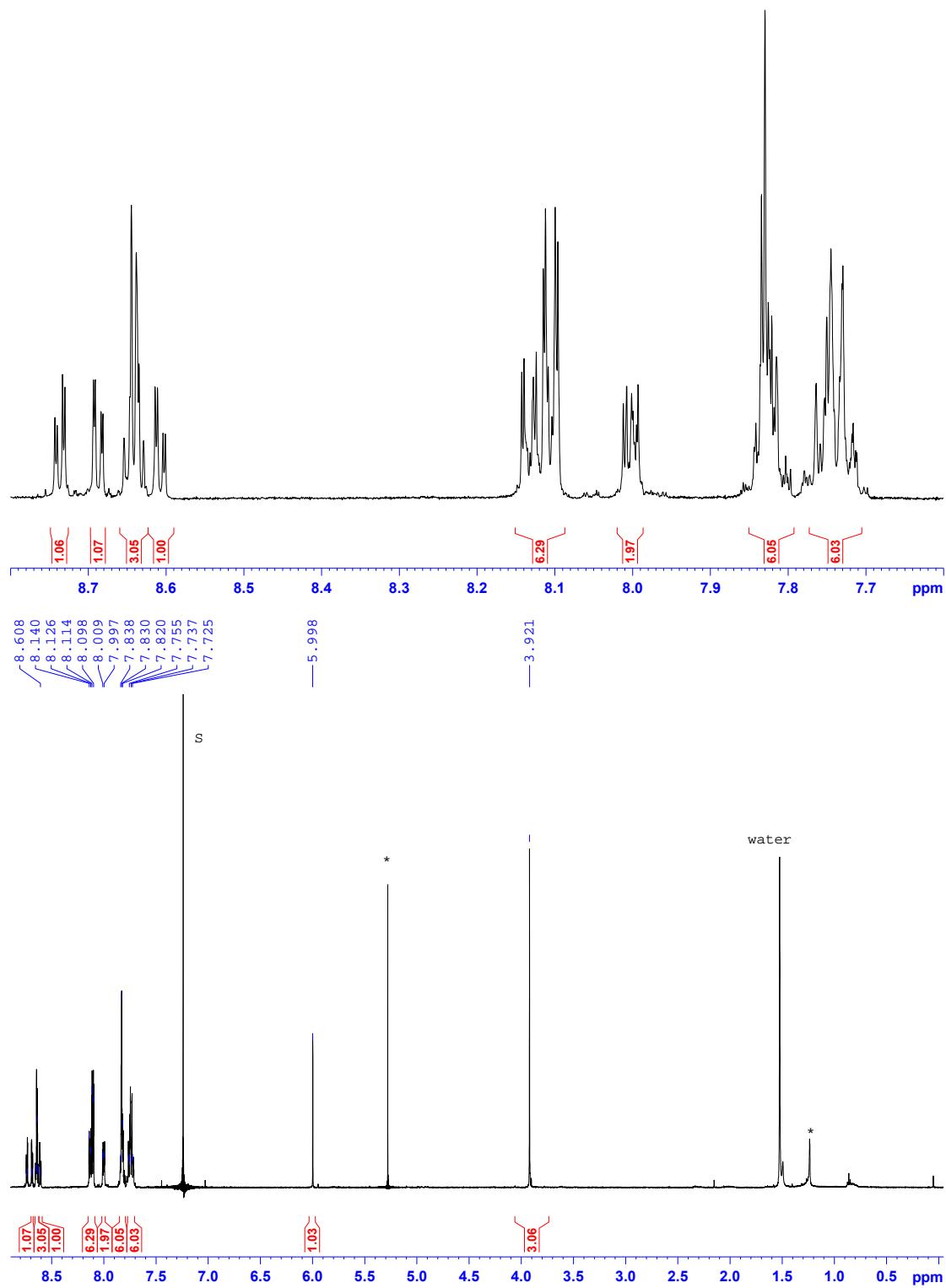


Figure S49. ^1H NMR spectrum of **5a**, 300 K, CDCl_3

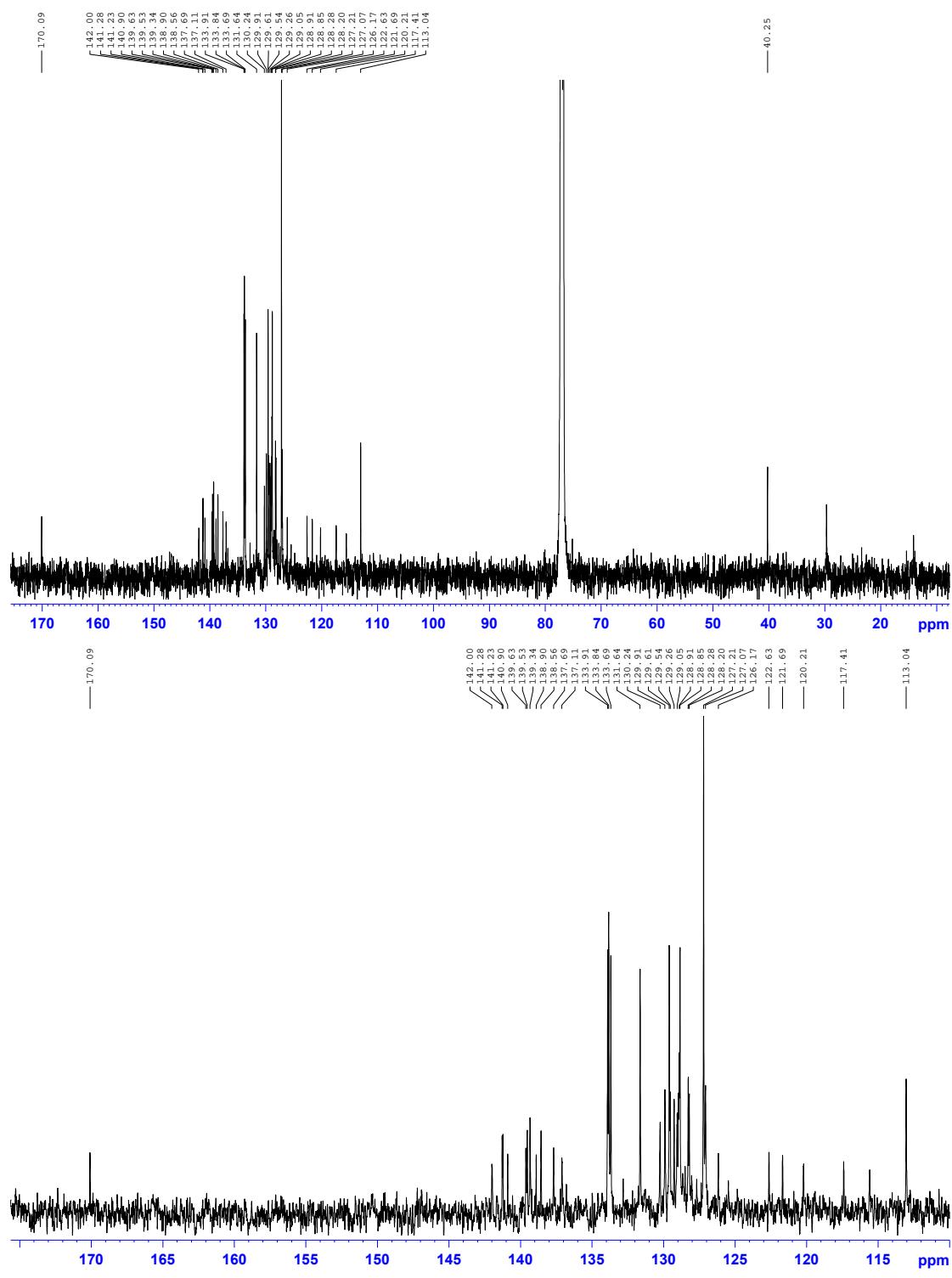
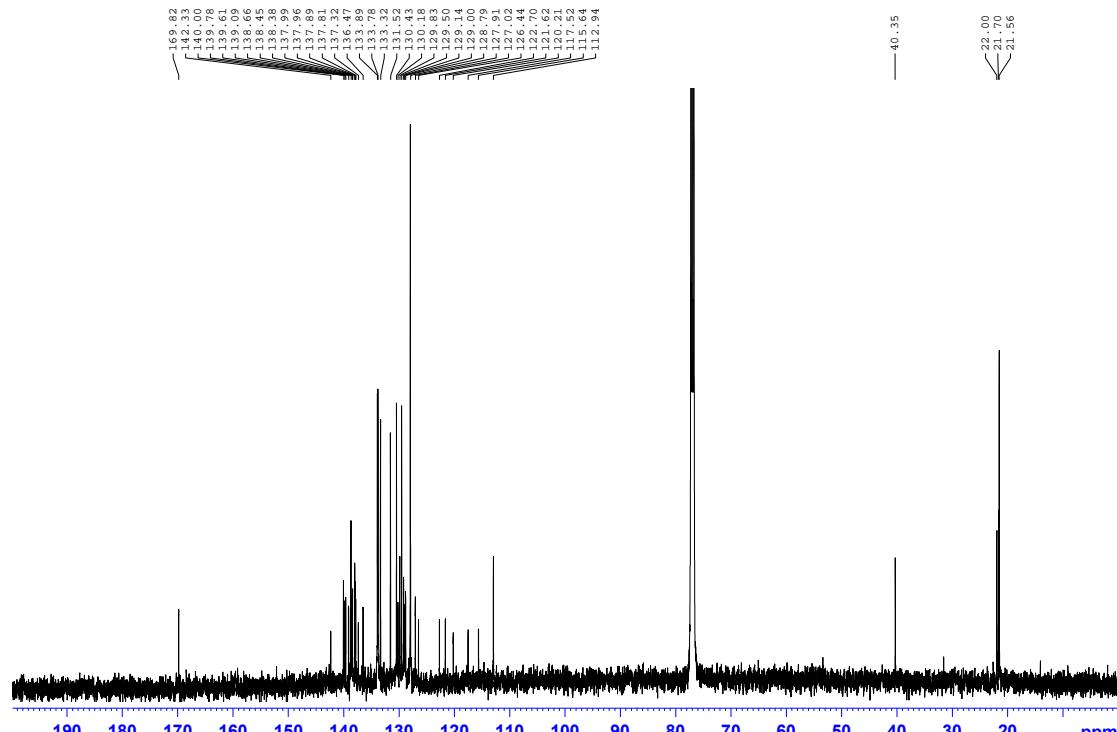
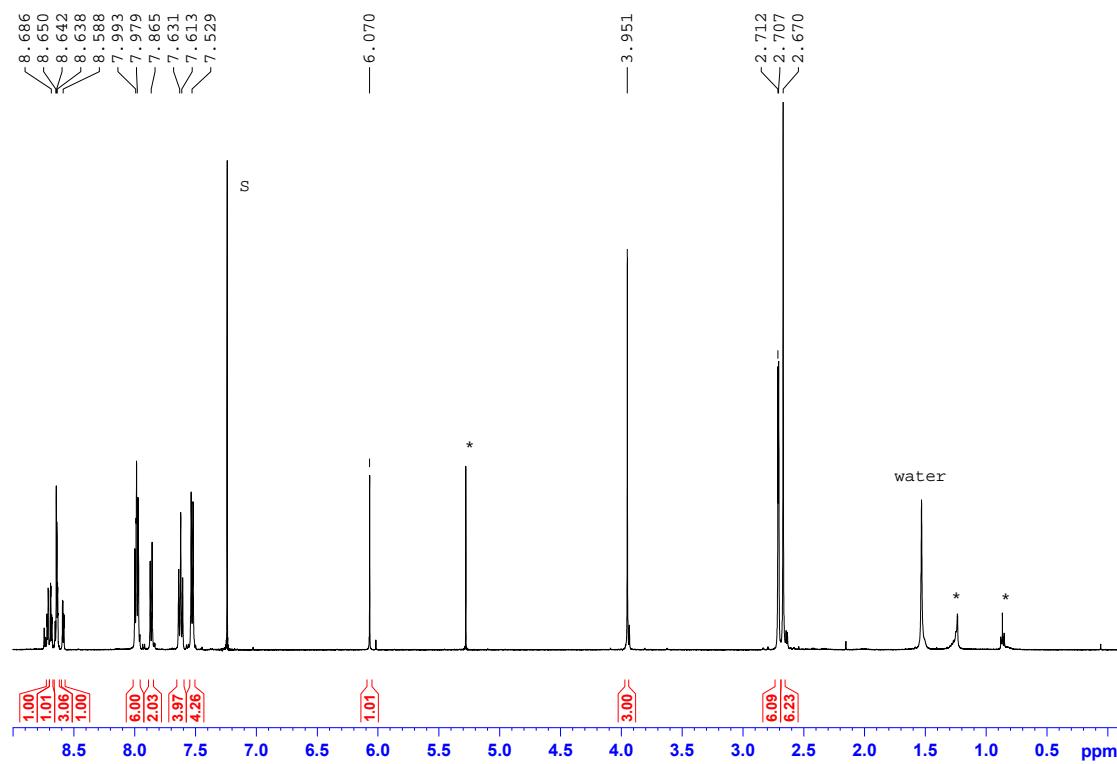


Figure S50. ^{13}C NMR spectrum of **5a**, 300 K, CDCl_3 .



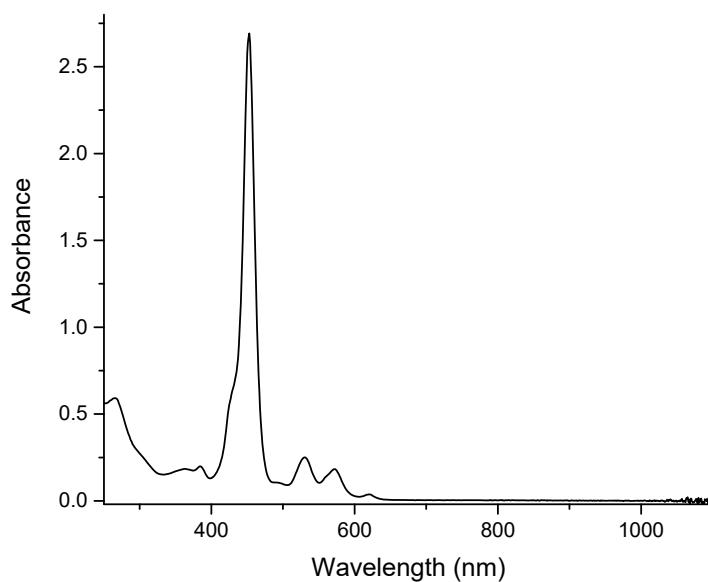


Figure S53. UV-vis spectrum of **5a** (CH_2Cl_2 , 1.51×10^{-5} M, 1 cm).

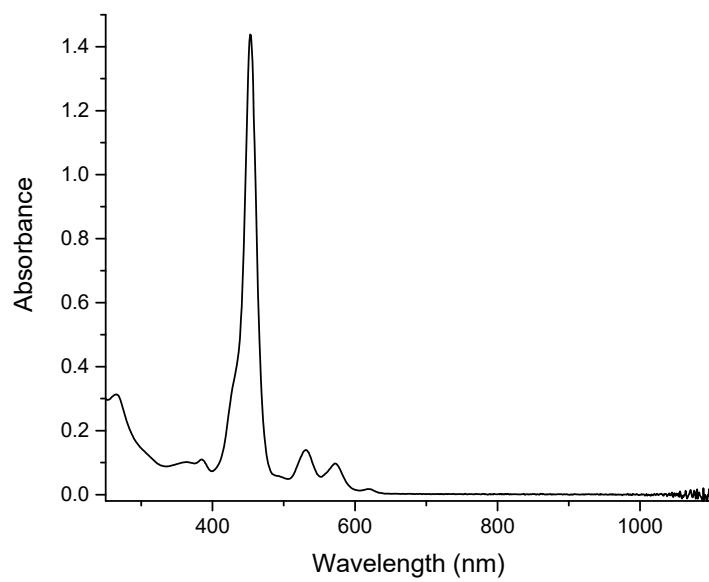
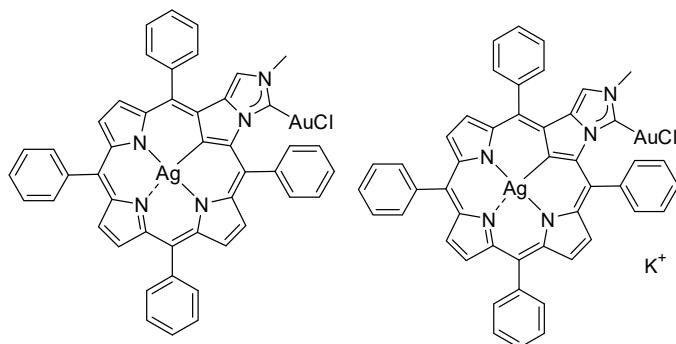


Figure S54. UV-vis spectrum of **5b** (CH_2Cl_2 , 8.47×10^{-6} M, 1 cm).



Chemical Formula: C₄₇H₃₀AgAuClN₅ Exact Mass: 1003.0906 Chemical Formula: C₄₇H₃₀AgAuClKN₅⁺ Exact Mass: 1042.0538

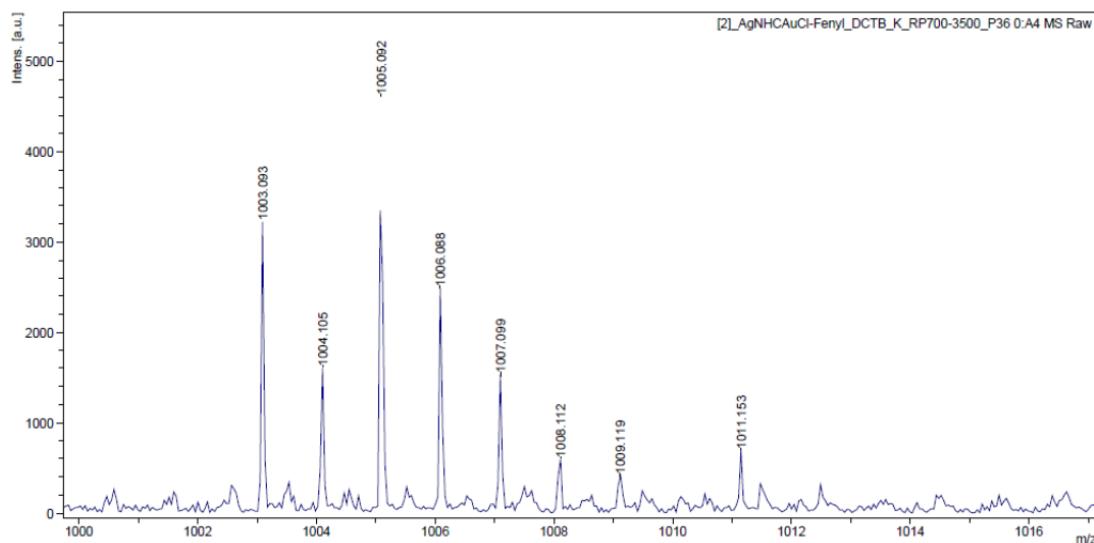


Figure S55. MALDI HRMS spectrum of **5a** (molecular ion).

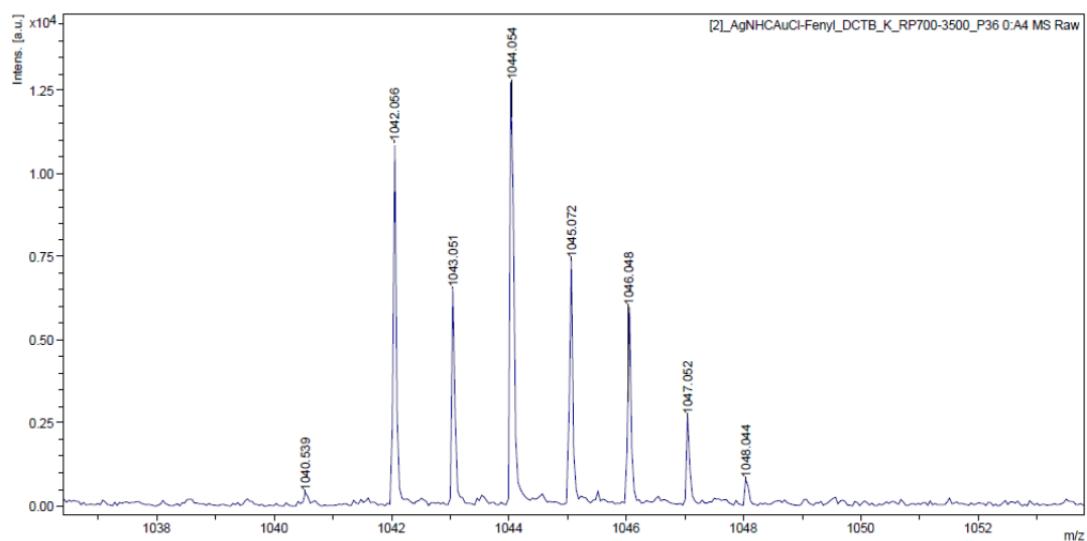


Figure S56. MALDI HRMS spectrum of **5a** ([5a+K]⁺).

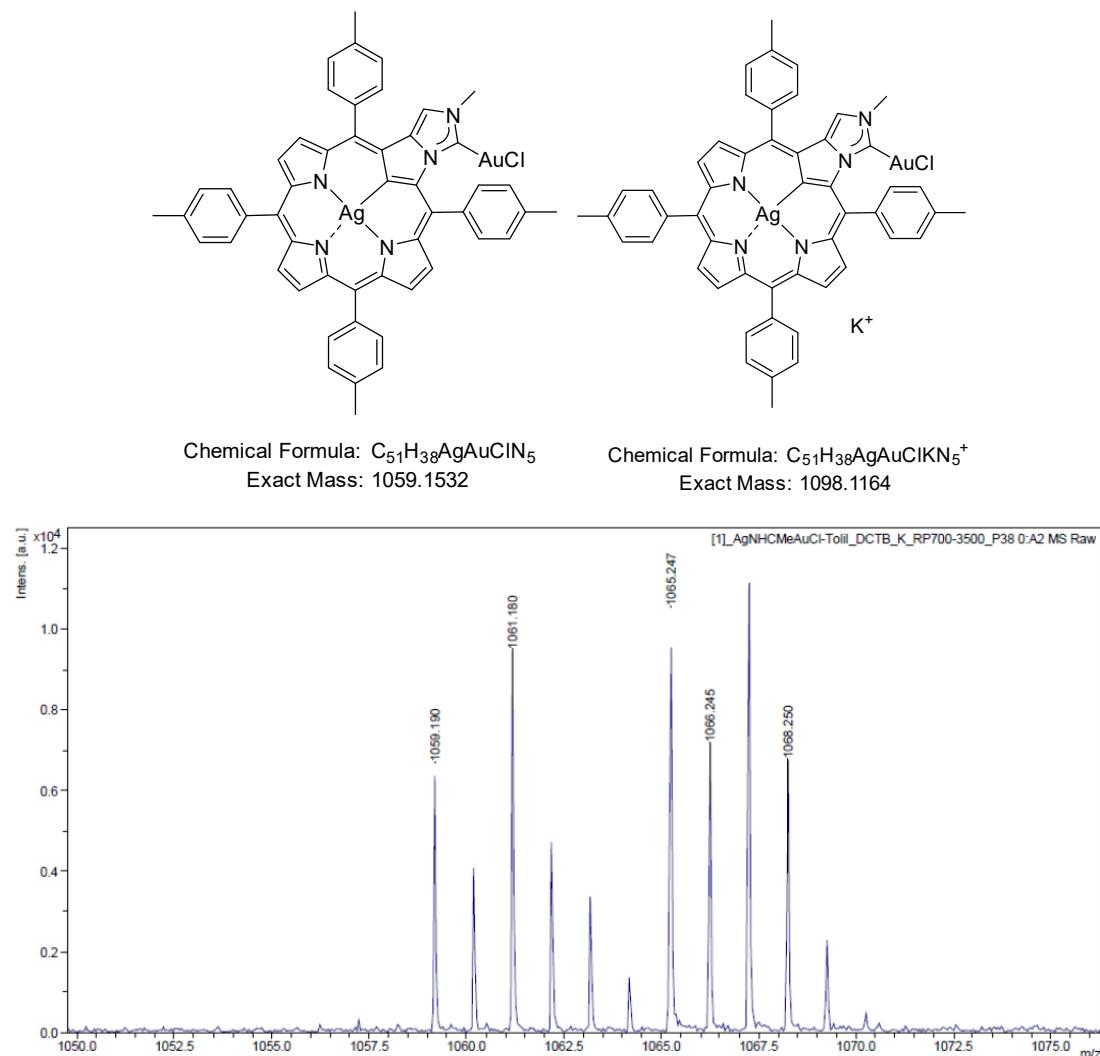


Figure S57. MALDI HRMS spectrum of **5b** (molecular ion).

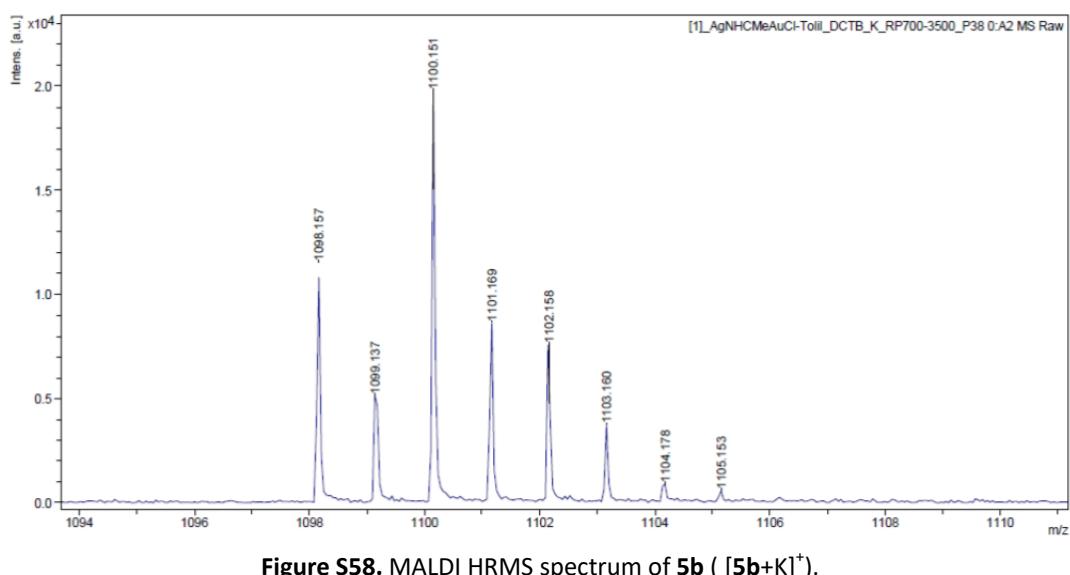


Figure S58. MALDI HRMS spectrum of **5b** ($[5b+K]^+$).

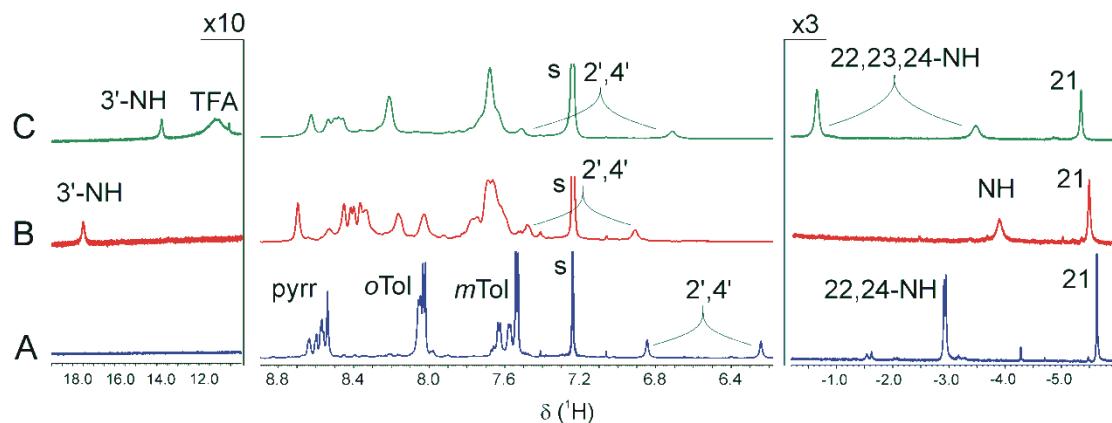


Figure S59. Protonation scheme of **2b** and selected regions of ^1H NMR spectra (600 MHz, CDCl_3 , 213 K) of **2b** (A) and the same solution after addition of 1.1 equiv. (B) and 4 equiv. (C) of trifluoroacetic acid (TFA).

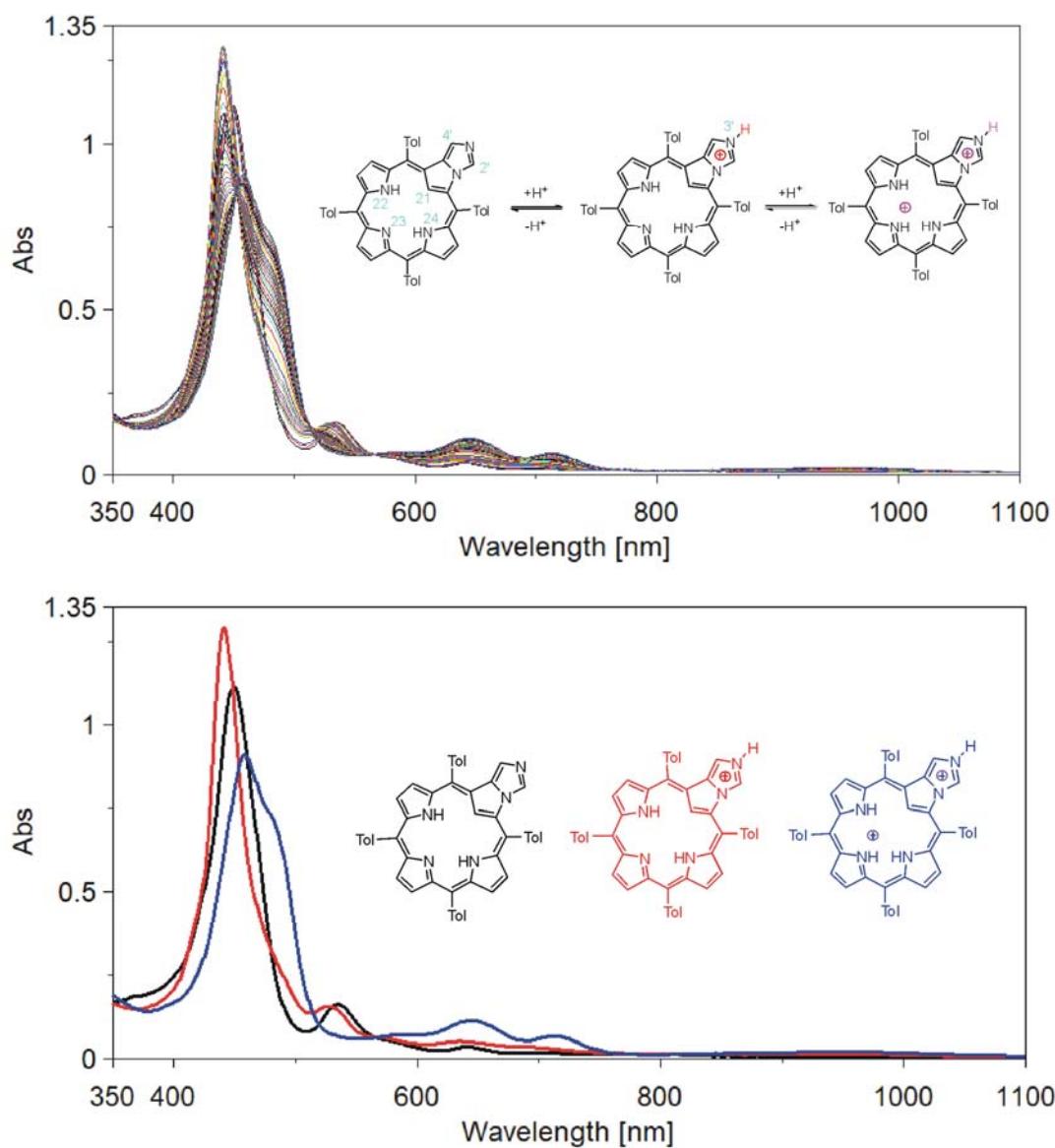


Figure S60. Top, spectrophotometric titration of **2b** with TFA (CHCl_3 , 298 K); bottom, selected spectra recorded after addition of 0 equiv. (black trace), 1.5 equiv. (red trace), and 6 equiv. (blue) of the acid.

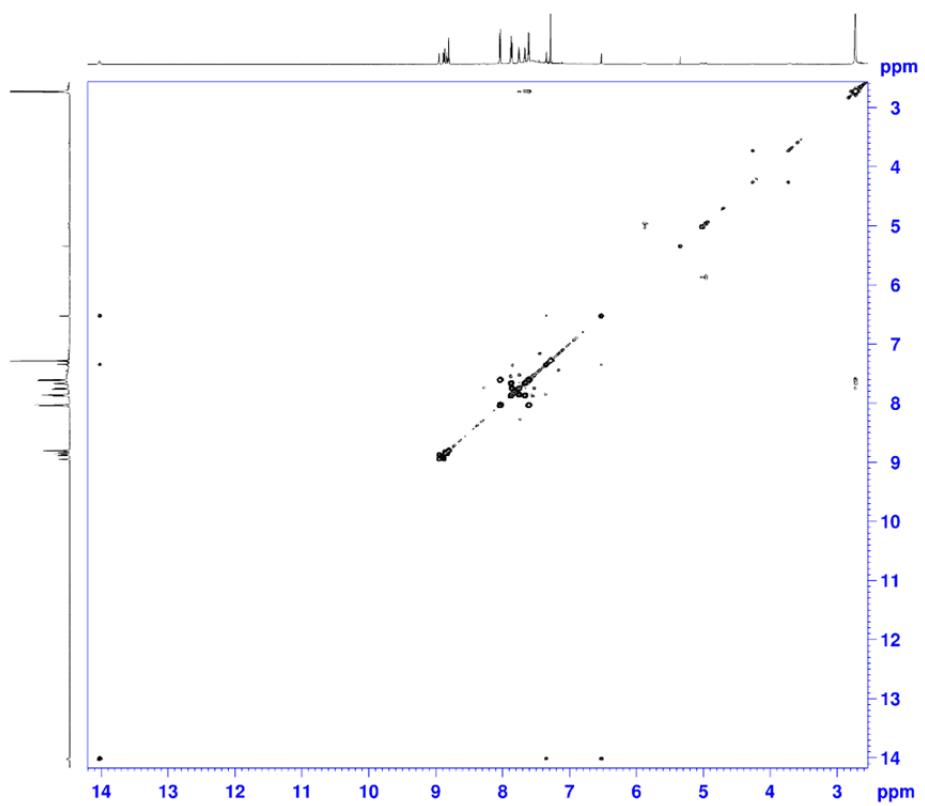


Figure S61. ¹H-¹H COSY spectrum of $[3\mathbf{b}\mathbf{H}]^+$ (600 MHz, 233 K, CDCl_3).

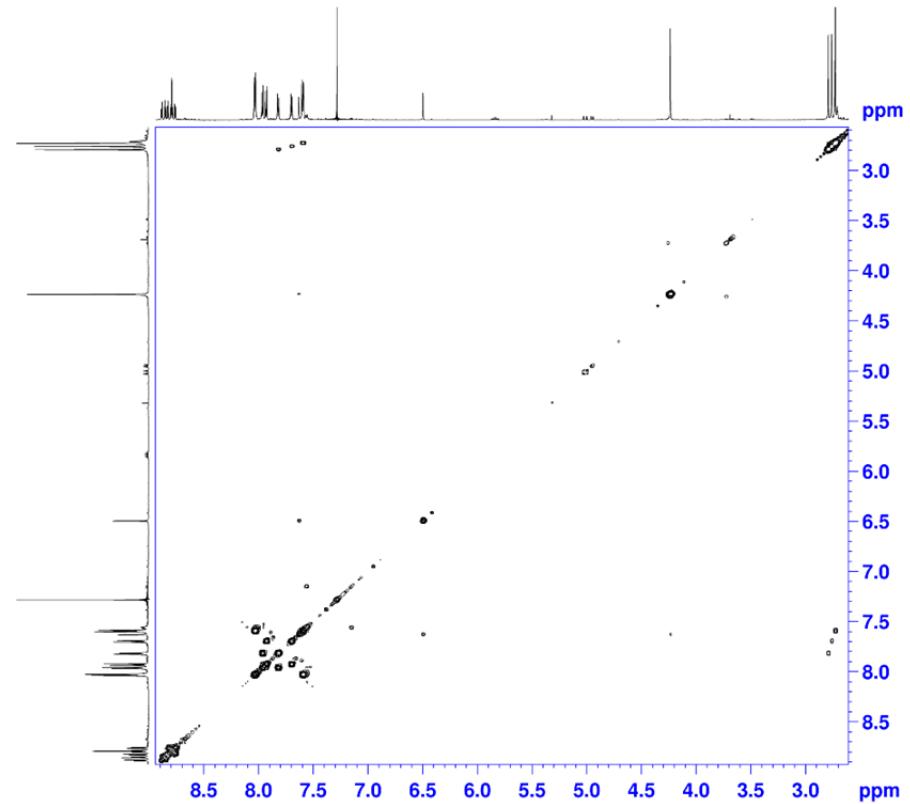


Figure S62. ¹H-¹H COSY spectrum of $[4\mathbf{b}]\mathbf{I}$ (600 MHz, 300 K, CDCl_3).

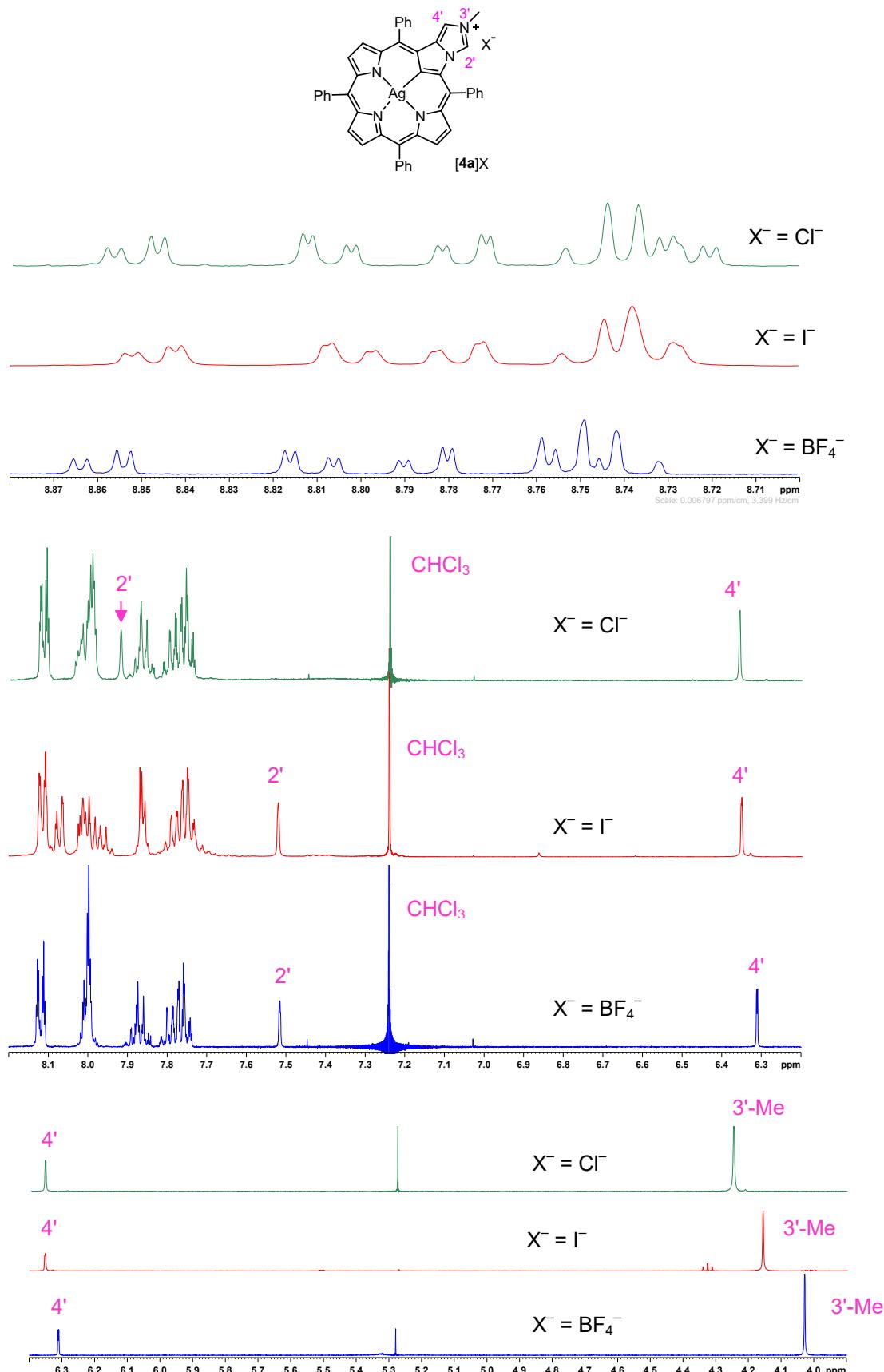


Figure S63. Selected regions of ^1H NMR spectra (500 MHz, 300 K, CDCl_3) of $[4\mathbf{a}]\text{X}$ (600 MHz, 300 K, CDCl_3) with specified counterions.

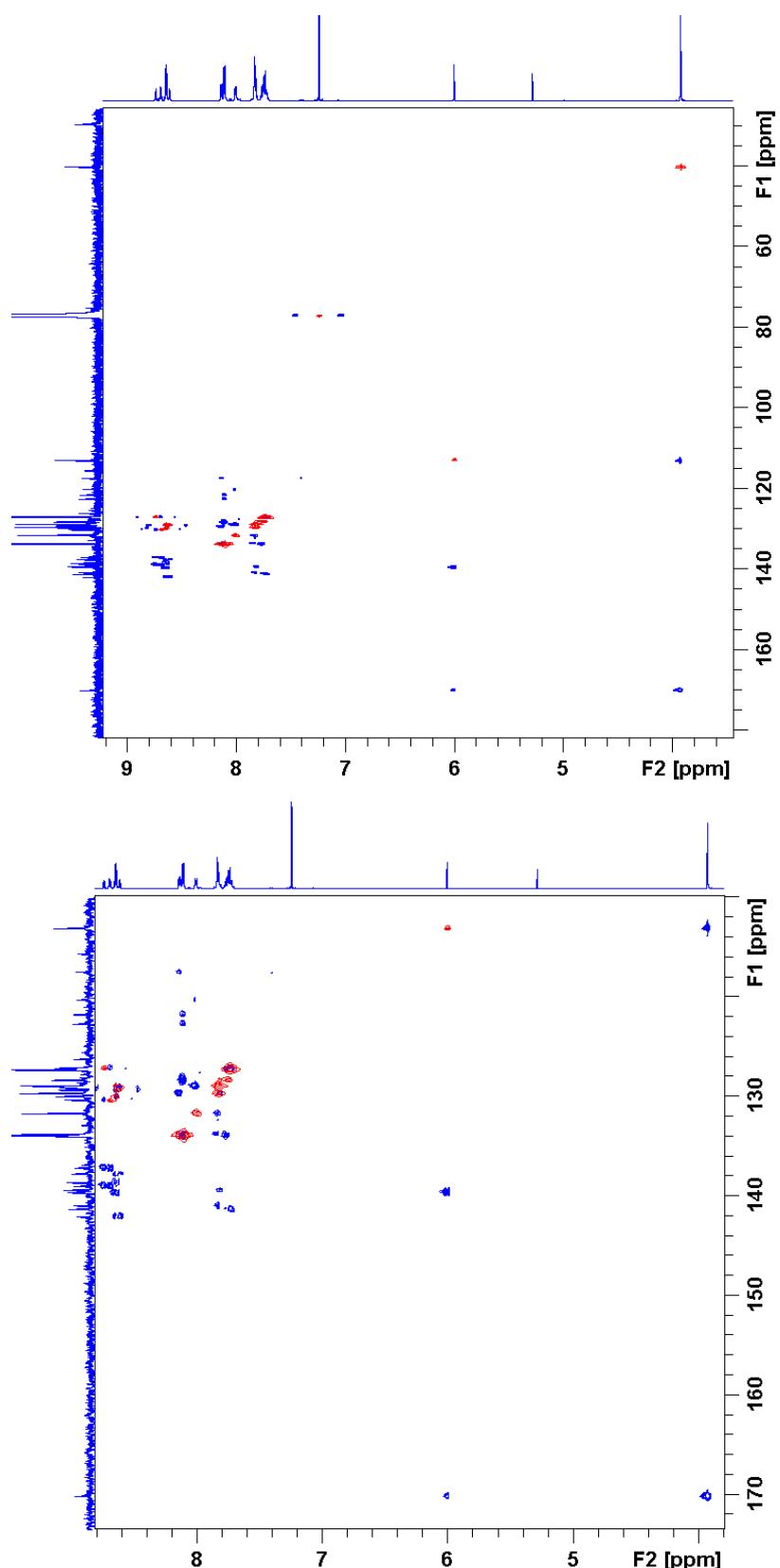


Figure S64. Superimposed ¹H, ¹³C HMQC (red spots) and HMBC (blue spots) spectra of **5a** (CDCl_3 , 300 K) and their down-field region expansions.

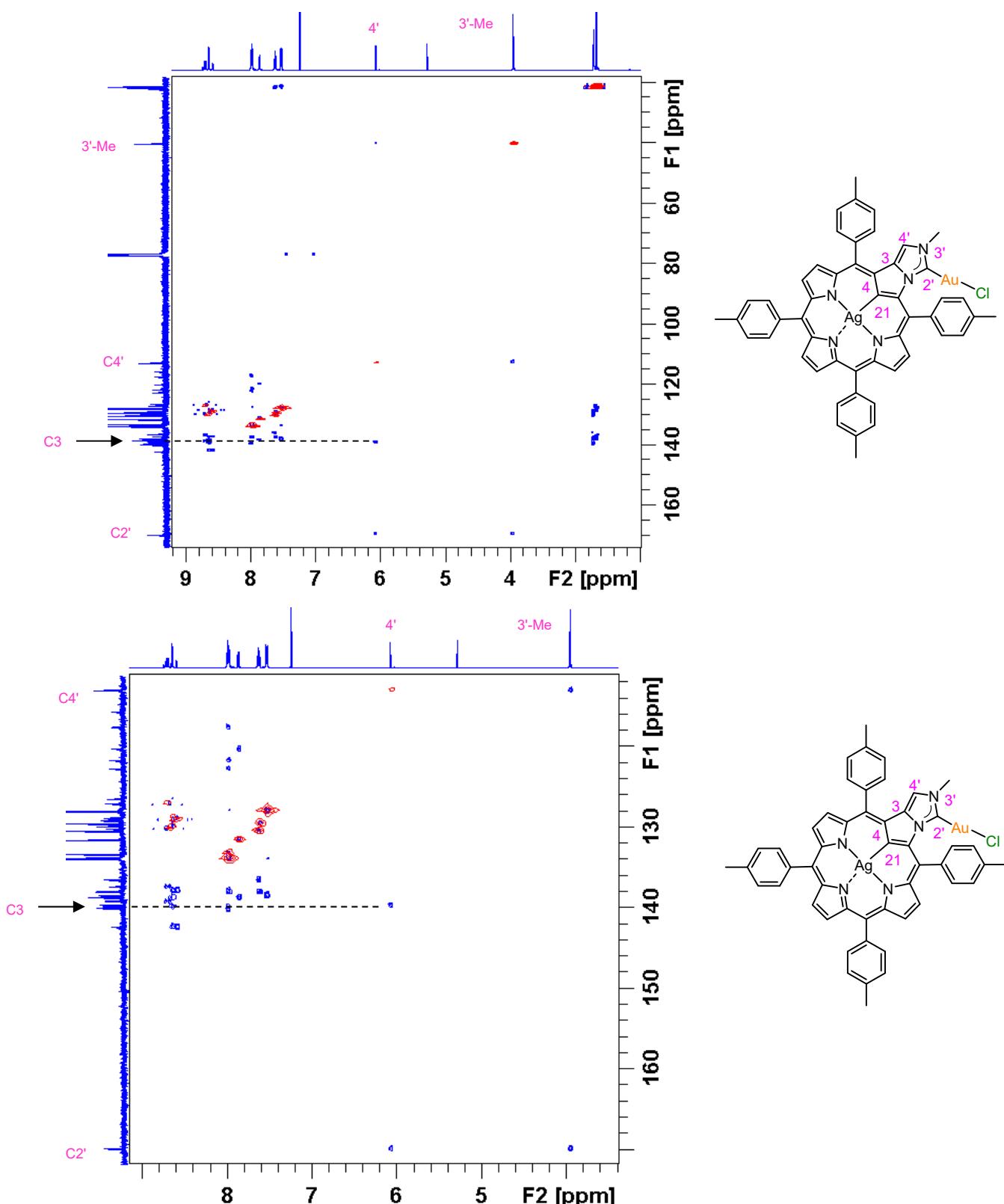


Figure S65. Superimposed ^1H , ^{13}C HMQC (red spots) and HMBC (blue spots) spectra of **5b** (CDCl_3 , 300 K) and their down-field region expansions.

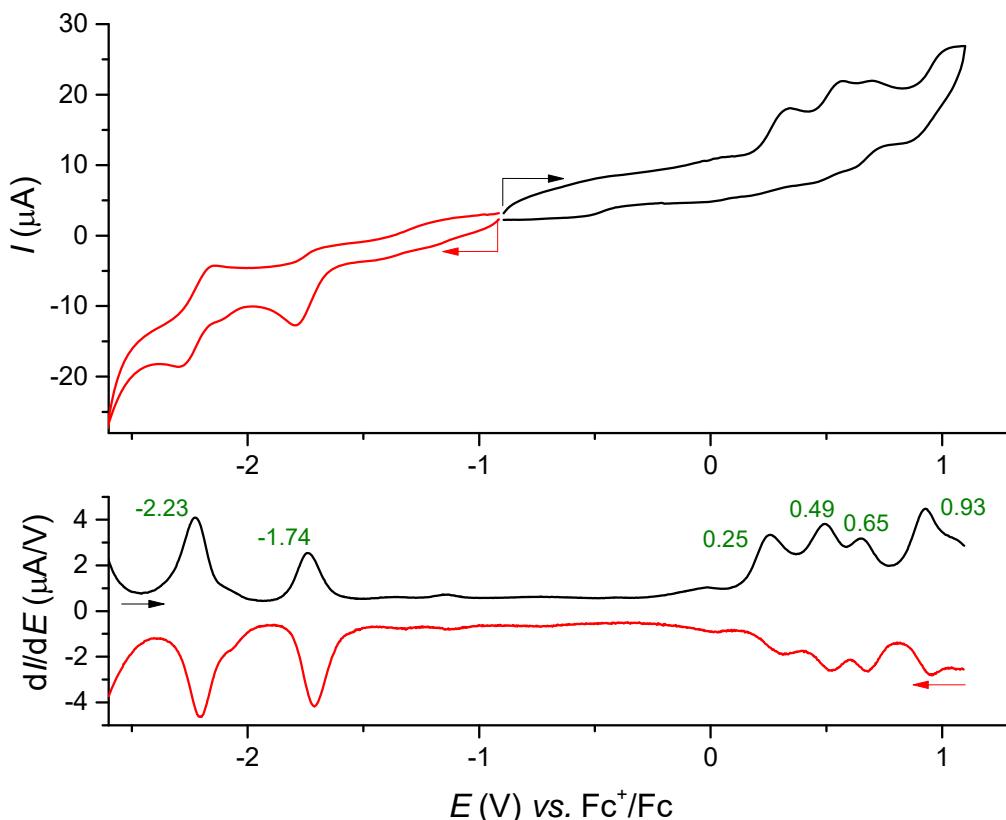


Figure S66. Cyclic voltammograms (top) and differential pulse voltammograms (bottom) for **2a** in DCM. The arrows indicate directions of the electrode potential advancements. Conditions: solvent , DCM; supporting electrolyte, tetrabutylammonium hexafluorophosphate; working electrode, glassy carbon; reference electrode, Ag/AgCl; auxiliary electrode, Pt rod.

Table S3. Electrochemical data for **2a-h***

| System | E_{Red1} | E_{Red2} | E_{Ox1} | E_{Ox2} | E_{Ox3} | E_{Ox4} | HLG ^a | $\Sigma \sigma^b$ |
|-----------|-------------------|-------------------|------------------|------------------|------------------|------------------|------------------|-------------------|
| 2a | -1.74 | -2.23 | 0.25 | 0.49 | 0.65 | 0.93 | 1.99 | 0 |
| 2b | -1.75 | -2.25 | 0.23 | 0.43 | 0.55 | 0.91 | 1.98 | -0.68 |
| 2c | -1.76 | -2.26 | 0.24 | 0.44 | 0.58 | 0.97 | 2.00 | -0.56 |
| 2d | -1.79 | -2.30 | 0.20 | 0.40 | 0.70 | n.d. | 1.99 | -1.08 |
| 2e | -1.75 | -2.28 | 0.31 | 0.52 | 0.66 | 1.01 | 2.06 | 0.96 |
| 2f | -1.76 | -2.30 | 0.30 | 0.45 | 0.68 | n.d. | 2.06 | -0.12 |
| 2g | -1.67 | -2.19 | 0.32 | 0.51 | 0.68 | 1.00 | 1.99 | 0.24 |
| 2h | -1.62 | -2.13 | 0.38 | 0.59 | 0.77 | 1.03 | 2.00 | 1.8 |

*Peak potentials in differential pulse voltammograms in volts. ^aElectrochemical HOMO-LUMO gap, HLG = $E_{\text{Ox1}} - E_{\text{Red1}}$. ^b Sum of the Hammett constants (Hansch, C.; Taft, R. W. A Survey of Hammett Substituent Constants and Resonance and Field Parameters *Chem. Rev.* **1991**, *91*, 165-195.) of the substituents present at the meso-aryls.

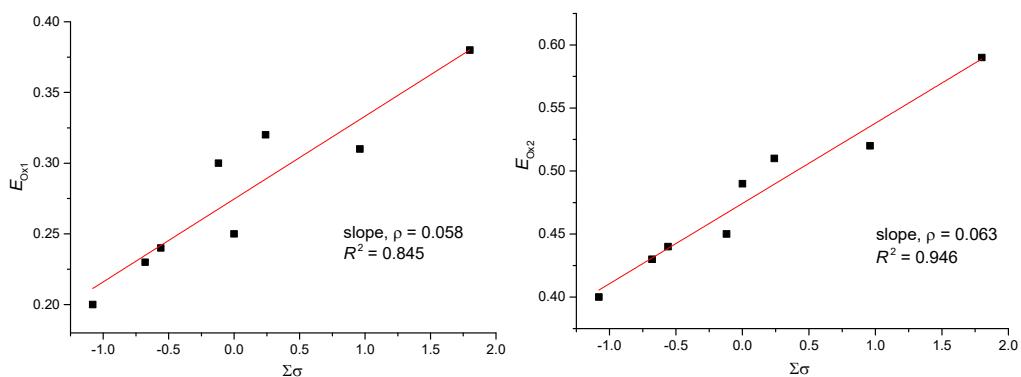


Figure S67. Dependencies of E_{Ox1} (top, left), E_{Ox2} (top, right), and E_{Red1} (bottom) for **2a-h** on the sum of the Hammett constants of the substituents present at the *meso*-aryls.

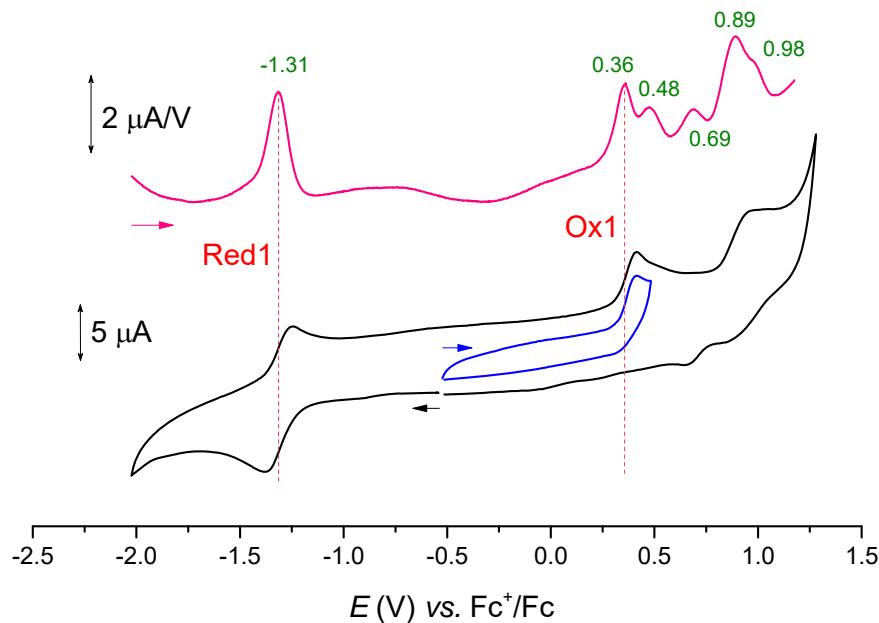


Figure S68. Cyclic voltammograms (bottom) and differential pulse voltammograms (top) for **3a** in DCM. The arrows indicate directions of the electrode potential advancements. The green numbers associated with peaks of the differential pulse voltammogram are electrode potentials in volts. Conditions: solvent, DCM; supporting electrolyte, tetrabutylammonium hexafluorophosphate; working electrode, glassy carbon; pseudoreference electrode, Ag/AgCl; auxiliary electrode, Pt rod.

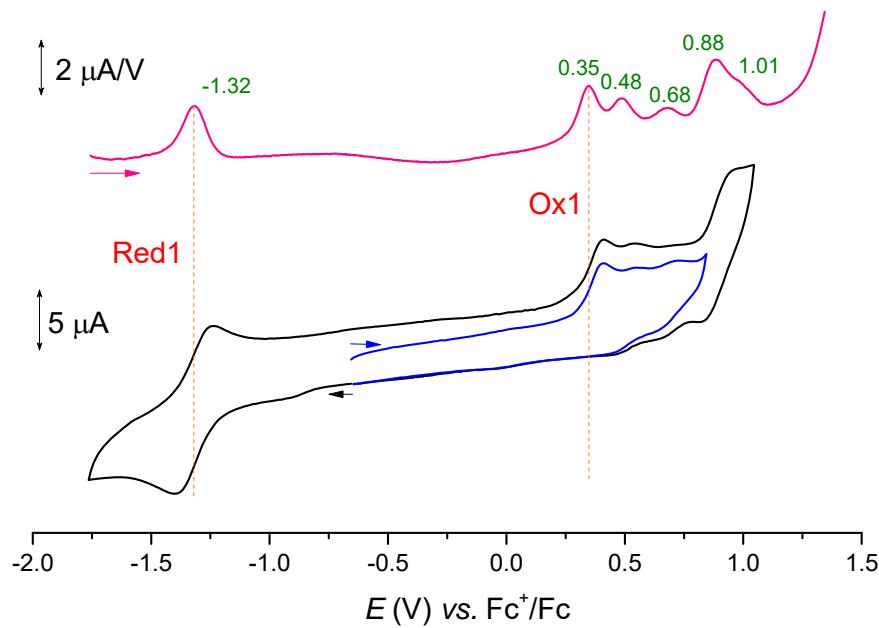


Figure S69. Cyclic voltammograms (bottom) and differential pulse voltammogram (top) for **3b** in DCM. The green numbers associated with peaks of the differential pulse voltammogram are electrode potentials in volts. The arrows indicate directions of the electrode potential advancements. Conditions: solvent, DCM; supporting electrolyte, tetrabutylammonium hexafluorophosphate; working electrode, glassy carbon; pseudoreference electrode, Ag/AgCl; auxiliary electrode, Pt rod.

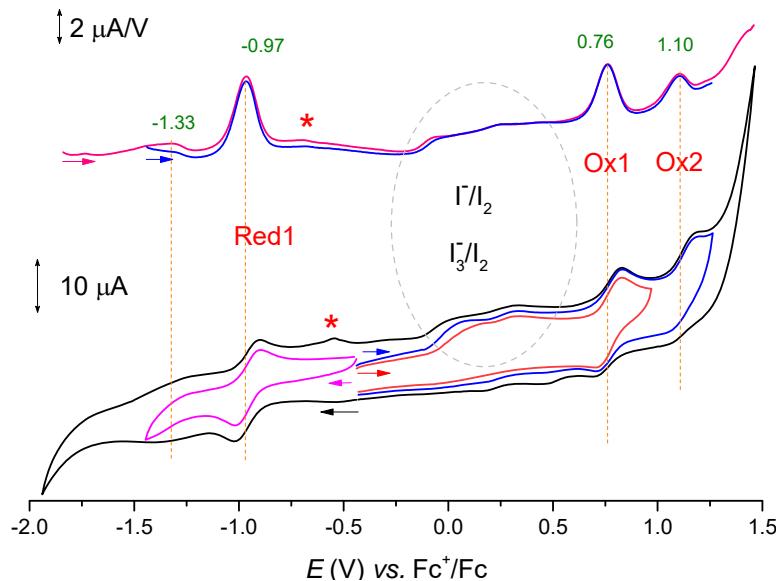


Figure S70. Cyclic voltammograms (bottom) and differential pulse voltammograms (top) for **[4a]I** in DCM. The arrows indicate directions of the electrode potential advancements. The green numbers associated with peaks of the differential pulse voltammogram are electrode potentials in volts. The red asterisks denote re-oxidation of the products of the irreversible second reduction at -1.33 V. The irreversible oxidation of iodide counteranion is marked with dashed ellipsis. Conditions: solvent, DCM; supporting electrolyte, tetrabutylammonium hexafluorophosphate; working electrode, glassy carbon; pseudoreference electrode, Ag/AgCl; auxiliary electrode, Pt rod.

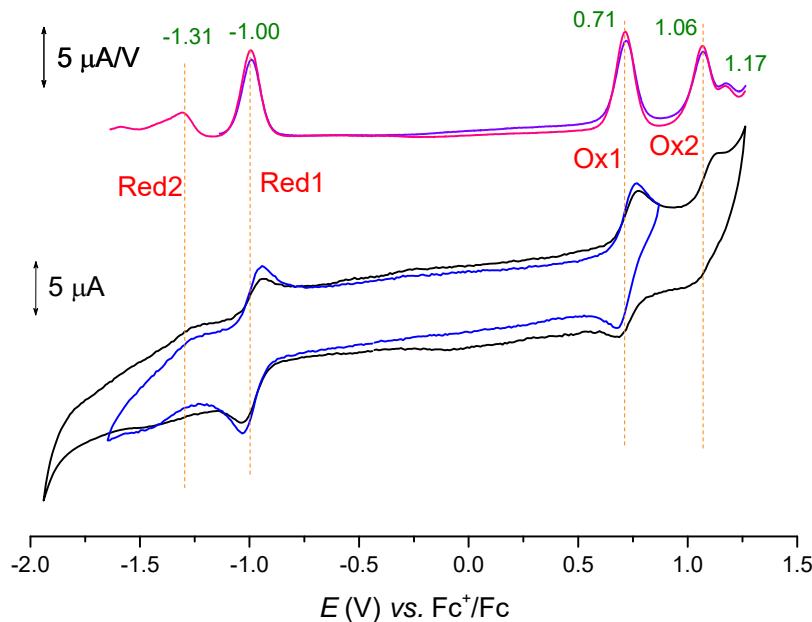


Figure S71. Cyclic voltammograms (bottom) and differential pulse voltammograms (top) for **[4a]BF₄** in DCM. The green numbers associated with peaks of the differential pulse voltammogram are electrode potentials in volts. Conditions: solvent, DCM; supporting electrolyte, tetrabutylammonium hexafluorophosphate; working electrode, glassy carbon; pseudoreference electrode, Ag/AgCl; auxiliary electrode, Pt rod.

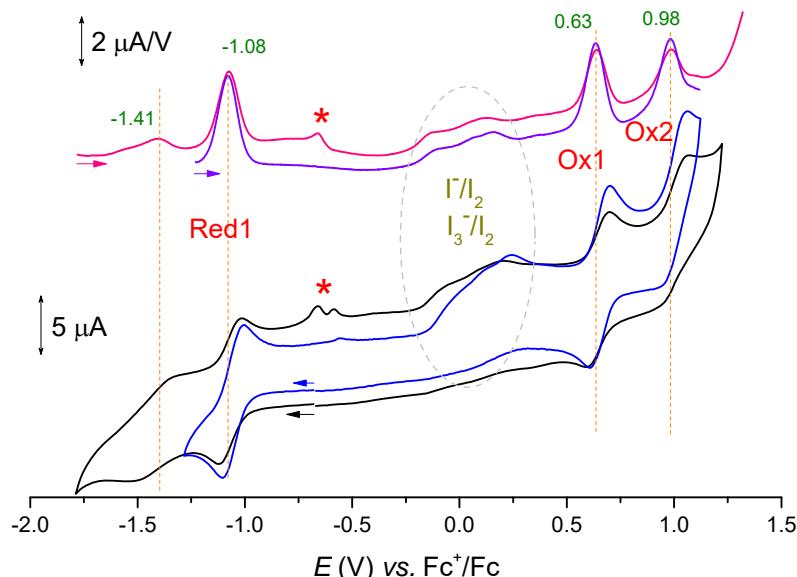


Figure S72. Cyclic voltammograms (bottom) and differential pulse voltammograms (top) for **[4b]I** in DCM. The arrows indicate directions of the electrode potential advancements. The green numbers associated with peaks of the differential pulse voltammogram are electrode potentials in volts. The red asterisks denote re-oxidation of the products of the irreversible second reduction at -1.41 V. The irreversible oxidation of iodide counteranion is marked with dashed ellipsis. Conditions: solvent, DCM; supporting electrolyte, tetrabutylammonium hexafluorophosphate; working electrode, glassy carbon; pseudoreference electrode, Ag/AgCl; auxiliary electrode, Pt rod.

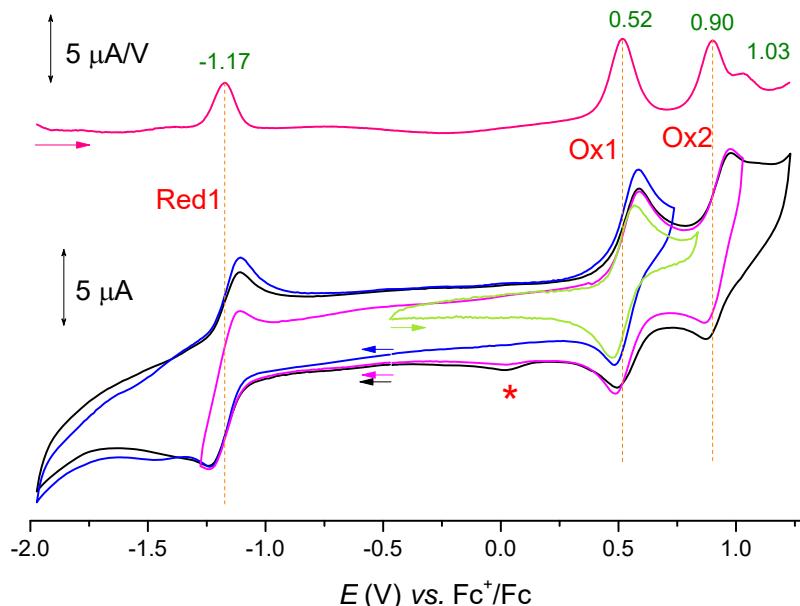


Figure S73. Cyclic voltammograms (bottom) and differential pulse voltammogram (top) for **5a** in DCM. The arrows indicate directions of the electrode potential advancements. The green numbers associated with peaks of the differential pulse voltammogram are electrode potentials in volts. The red asterisks denote re-reduction of the products of the irreversible third oxidation at 1.03 V. Conditions: solvent, DCM; supporting electrolyte, tetrabutylammonium hexafluorophosphate; working electrode, glassy carbon; pseudoreference electrode, Ag/AgCl; auxiliary electrode, Pt rod.

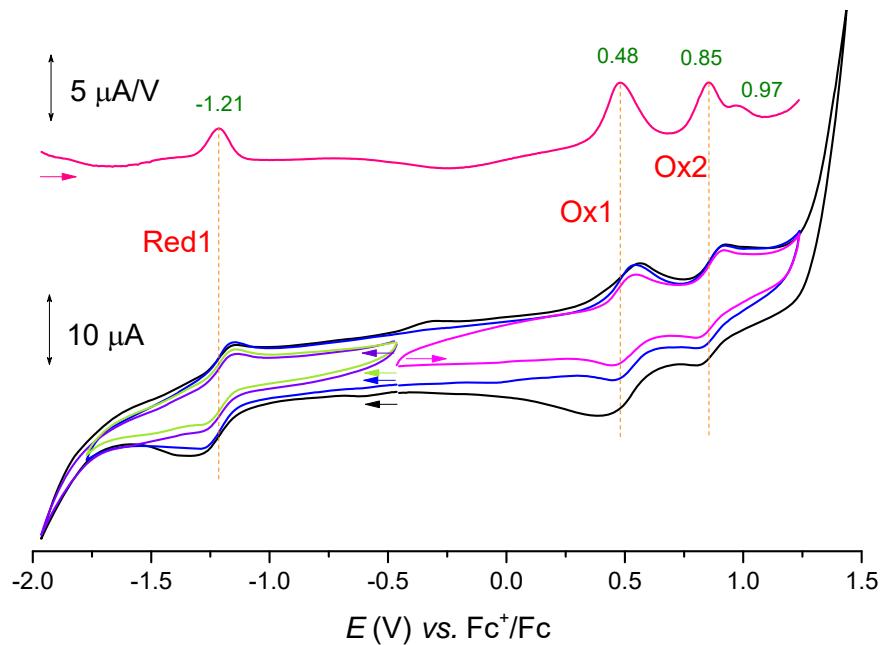


Figure S74. Cyclic voltammograms (bottom) and differential pulse voltammogram (top) for **5b** in DCM. The arrows indicate directions of the electrode potential advancements. The green numbers associated with peaks of the differential pulse voltammogram are electrode potentials in volts. Conditions: solvent, DCM; supporting electrolyte, tetrabutylammonium hexafluorophosphate; working electrode, glassy carbon; pseudoreference electrode, Ag/AgCl; auxiliary electrode, Pt rod.

Table S4. Electrochemical data^{*} for silver(III) complexes **3**, **4**, and **5**.

| System | $E_{\text{Red}1}$ | $E_{\text{Red}2}$ | $E_{\text{Ox}1}$ | $E_{\text{Ox}2}$ | $E_{\text{Ox}3}$ | $E_{\text{Ox}4}$ | HLG ^a |
|------------------------------|-------------------|--------------------|-------------------|-------------------|-------------------|---------------------------------------|------------------|
| 3a | -1.31 | – | 0.31 ^b | 0.48 ^b | 0.69 ^b | 0.89 ^b , 0.95 ^b | 1.67 |
| 3b | -1.32 | – | 0.35 ^b | 0.48 ^b | 0.68 ^b | 0.88 ^b , 1.01 ^b | 1.67 |
| [4a]I ^c | -0.97 | -1.33 ^b | 0.76 | 1.10 | – | – | 1.73 |
| [4a]BF ₄ | -1.00 | -1.31 ^b | 0.71 | 1.06 | 1.17 ^b | – | 1.71 |
| [4b]I ^c | -1.08 | -1.41 ^b | 0.63 | 0.98 | – | – | 1.71 |
| 5a | -1.17 | – | 0.52 | 0.90 | 1.03 ^b | – | 1.69 |
| 5b | -1.21 | – | 0.48 | 0.85 | 0.97 ^b | – | 1.69 |

*Peak potentials in differential pulse voltammograms in volts. ^a Electrochemical HOMO-LUMO gap, HLG = $E_{\text{Ox}1} - E_{\text{Red}1}$. ^b Irreversible. ^c Irreversible processes observed at about -0.15 and 0.15 V are related with I⁻ counteranion oxidation.

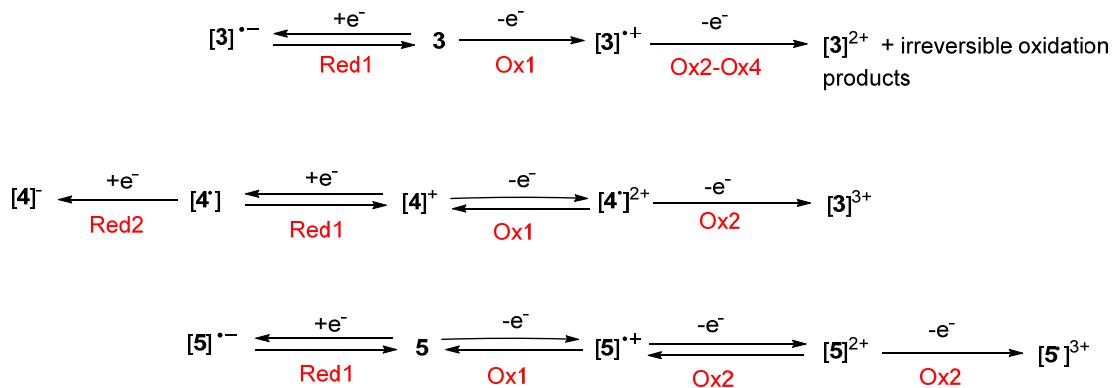


Figure S75 Schematic representations of the electrode reactions for the silver(III) complexes of imidazole-fused carbachlorins **3**, $[4]^+$, and **5**.

References

1. Rigaku Oxford Diffraction (2015). CrysAlis PRO. Rigaku Oxford Diffraction, Wrocław, Poland.
2. G. M. Sheldrick, *Acta Cryst.* 2015, **A71**, 3.
3. G. M. Sheldrick, *Acta Cryst.* 2015, **C71**, 3.
4. G. R. Geier, III, D. M. Haynes, and J. S. Lindsey, *Org. Lett.*, 1999, **1**, 1455.