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Iridium-Catalyzed double C-H Amidation of s-Tetrazines

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I. General Information.

All commercially available organic compounds were purchased from Sigma-Aldrich, adamas-beta and Shanghai GeorGene Biotech Co., Ltd. in China. Unless otherwise noted, all commercial reagents and solvents were used without additional purification. The substrates **1** were prepared according to literature methods.^[1] Water was purified with a Millipore Milli-Q system. NMR spectra were recorded on Bruker AM-500 instruments. Chemical shifts are reported in δ (ppm) referenced to TMS as an internal standard for ¹H NMR and CDCl₃ (δ 77.0) or DMSO-*d*₆ (δ 39.5) for ¹³C NMR. High-resolution mass spectra were obtained on High-resolution mass spectra (HRMS-ESI) were obtained on an Agilent Technologies 6230 Accurate Mass TOF LC/MS instrument or an AB Sciex 4600 QTOF MS instrument.

All reagents and DNA headpiece HP-NH2 (5'- / 5phos / GAGTCA / iSp9 / iUniAmM / iSp9 / TGACTCCC-3') were obtained from commercial sources unless otherwise noted and used as received. All on-DNA reactions were performed in 1.5 mL or 5 mL Eppendorf tubes. On-DNA reactions in the studies of reaction condition optimization and substrate scope extension were analyzed by UPLC-MS. Typically, samples were dissolved in an appropriate amount of distilled and deionized water (ddH₂O) and injected into a reverse-phase chromatography column (Xbridge Oligonucleotide BEH C18 column, 1.7 μ m, 2.1×50 mm). The elution was carried out as followings: 5–95% solvent B over 4.5 min, 0.4 mL/min, λ = 260 nm; solvent A: 0.75% v/v hexafluoroisopropanol/ 0.038% v/v triethylamine in methanol/water = 5/95; solvent B: 0.75% v/v hexafluoroisopropanol/ 0.038% v/v triethylamine in methanol/water = 90/10. The effluents were analyzed by a Xevo G2-XS Q-TOF with electrospray ionization source was used for detection.

II. Experimental Procedures and Characterizations:

General procedure for the Synthesis and Characterization of 1.^[1]

A solution of 4-methylbenzonitrile (586 mg, 5 mmol), acetonitrile (1.7 mL, 40 mmol) and 3mercaptopropionic acid (0.44 mL, 5 mmol) in ethanol (1 mL) was cooled to 0 °C. To this mixture was added dropwise hydrazine hydrate (3.9 mL, 80 mmol). The reaction mixture was stirred at room temperature for 18 h. An ice water solution of NaNO₂ (5.175, 75 mmol) was slowly added into the reaction mixture, followed by slow addition of 1M HCl during which the solution was stirred intensely and turned bright red, and gas evolved. Addition of 1M HCl continued until gas evolution ceased and the pH value was 3-4. Then, the reaction mixture was extracted with dichloromethane before washing with saturated salt water. The extract was combined, dried over sodium sulfate and concentrated in vacuo. The crude residue was purified on silica column chromatography (PE : DCM = 2 : 1) to afford the desired product **1b** as a pink solid in 41% yield (382 mg).



¹H NMR (500 MHz, CDCl₃) δ 8.47 (d, *J* = 7.9 Hz, 1H), 7.38 (d, *J* = 8.0 Hz, 1H), 3.07 (s, 2H), 2.47 (s, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 167.0, 164.1, 143.2, 130.0, 129.0, 127.8, 21.6, 21.1. HRMS (ESI): Calcd for C₁₀H₁₁N₄: [M+H]⁺ 187.0984, found: m/z 187.0982.



Pink solid, 35% yield. ¹H NMR (500 MHz, CDCl₃) δ 8.52 (d, *J* = 8.6 Hz, 2H), 7.06 (d, *J* = 8.6 Hz, 2H), 3.90 (s, 3H), 3.04 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.5, 163.7, 163.2, 129.7, 124.1, 114.6, 55.4, 21.0. HRMS (ESI): Calcd for C₁₀H₁₁N₄O: [M+H]⁺ 203.0933, found: m/z 203.0937.



Pink solid, 31% yield. ¹H NMR (500 MHz, CDCl₃) δ 8.72 (d, J = 8.2 Hz, 2H), 7.85 (d, J = 8.2 Hz, 2H), 3.13 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 167.9, 163.2, 135.1, 134.1 (q, J = 32.9 Hz), 128.2, 126.2 (q, J = 3.8 Hz), 123.7 (q, J = 272.4 Hz), 21.2. HRMS (ESI): Calcd for C₁₀H₈F₃N₄: [M+H]⁺ 241.0701, found: m/z 241.0701.



Pink solid, 45% yield.¹H NMR (500 MHz, CDCl₃) δ 8.25 (d, J = 3.7 Hz, 1H), 7.66 (d, J = 4.9 Hz, 1H), 7.24 (t, J = 4.3 Hz, 1H), 3.03 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.7, 162.0, 135.7, 132.3, 131.0, 128.8, 21.1. HRMS (ESI): Calcd for C₇H₇N₄S: [M+H]⁺ 179.0391, found: m/z 179.0391.



Pink solid, 48% yield. ¹H NMR (500 MHz, CDCl₃) δ 8.76 (s, 1H), 8.53 (d, *J* = 7.8 Hz, 1H), 7.75 (d, *J* = 7.9 Hz, 1H), 7.47 (t, *J* = 7.9 Hz, 1H), 3.12 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 167.7, 163.1, 135.5, 133.7, 130.8, 130.8, 126.4, 123.4, 21.2. HRMS (ESI): Calcd for C₉H₈N₄Br: [M+H]⁺ 250.9932, found: m/z 250.9958.



To a 25 mL test tube was combined (4-(6-methyl-1,2,4,5-tetrazin-3-yl)phenyl)methanamine

(commercial available, 1.006 g, 5 mmol, 1 equiv), TEA (1.1 mL, 7.5 mmol, 1.5 equiv) and 20 mL CH_2Cl_2 , then Ac_2O (0.57 mL, 6 mmol, 1.2 equiv) was added at 0 °C and the mixture was stirred at room temperature for 2 h. Then water (20 mL) was added and extracted with CH_2Cl_2 (15 mL x 2), dried over Na_2SO_4 , and concentrated under reduced pressure. The residue was purified on silica gel column chromatography ($CH_2Cl_2/MeOH$, 100 : 1) to afford **1h** (1.156 g, 95%).

¹H NMR (600 MHz, CDCl₃) δ 8.53 (d, *J* = 8.0 Hz, 2H), 7.48 (d, *J* = 8.0 Hz, 2H), 6.02 (s, 1H), 4.54 (d, *J* = 5.9 Hz, 2H), 3.09 (s, 3H), 2.07 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 170.1, 167.2, 163.8, 143.2, 130.9, 128.4, 128.2, 43.3, 23.2, 21.1. HRMS (ESI): Calcd for C₁₂H₁₄N₅O: [M+H]⁺ 244.1198, found: m/z 244.1195.

Procedure for the Synthesis and Characterization of 2.

Methyl 4-(chlorosulfonyl)benzoate (2.347 g, 10.0 mmol) was dissolved in acetone (30 mL) and water (30 mL). The solution was cooled to 0 °C and NaN₃ (715 mg, 11.0 mmol) was added. The reaction was stirred for 2 h. The acetone was removed under vacuum and the remaining water layer was extracted with EtOAc (50 mL), the EtOAc layer was washed with brine, dried on Na₂SO₄ and concentrated. The residue was purified on silica gel column chromatography (PE/DCM, 10 : 1) to afford **2f** (2.29 g, 95%).



¹H NMR (500 MHz, CDCl₃) δ 8.26 (d, *J* = 7.9 Hz, 2H), 8.03 (d, *J* = 8.0 Hz, 2H), 3.98 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 165.0, 142.0, 135.7, 130.8, 127.5, 52.9.



97% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.98 (d, J = 7.8 Hz, 1H), 7.66 (d, J = 7.8 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 139.0, 138.0, 128.6, 103.0.



93% yield. ¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, J = 2.0 Hz, 1H), 7.72 (dd, J = 8.8, 2.0 Hz, 1H), 7.00 (d, J = 8.9 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 156.0, 138.8, 132.6, 129.0, 114.1, 112.5, 56.7.

General Procedure for the Synthesis and Characterization of 4 and/or 3.

To a 10 mL test tube was combined **1** (0.2 mmol, 1 equiv), **2** (0.5 mmol, 2.5 equiv), $[IrCp*Cl_2]_2$ (6.4 mg, 4 mol %), AgNTf₂ (12 mg, 16 mol %), AgOCOCF₃ (9 mg, 20 mol %) and 2 mL DCE, then the reaction was stirred at 100 °C for 3 h. The solvent was removed and the residue was purified by silica gel chromatography using PE/EA to afford the title compounds.



Pink solid, 55% yield (60 °C). ¹H NMR (500 MHz, CDCl₃) δ 10.56 (s, 1H), 8.49 (d, J = 8.0 Hz, 1H), 7.81 (d, J = 8.3 Hz, 1H), 7.56 (t, J = 7.8 Hz, 1H), 7.52 (d, J = 7.8 Hz, 2H), 7.30 (t, J = 7.7 Hz, 1H), 7.06 (d, J = 7.9 Hz, 2H), 3.10 (s, 3H), 2.30 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.3, 164.5, 143.9, 138.1, 136.1, 133.7, 130.3, 129.4, 127.0, 124.9, 122.9, 120.4, 21.4, 21.1. HRMS (ESI): Calcd for C₁₆H₁₆N₅O₂S: [M+H]⁺ 342.1025, found: m/z 342.1016.



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Pink solid, 93% yield.¹H NMR (500 MHz, CDCl₃) δ 8.99 (s, 2H), 7.66 – 7.56 (m, 3H), 7.08 (d, *J* = 7.8 Hz, 4H), 6.92 (d, *J* = 7.8 Hz, 4H), 3.06 (s, 3H), 2.30 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 165.6, 163.4, 144.1, 136.5, 135.0, 133.2, 129.3, 126.3, 125.8, 118.9, 21.4, 21.2. HRMS (ESI): Calcd for C₂₃H₂₃N₆O₄S₂: [M+H]⁺ 511.1222, found: m/z 511.1214.



Pink solid, 88% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.02 (s, 2H), 7.44 (s, 2H), 7.06 (d, J = 8.0 Hz, 4H), 6.89 (d, J = 8.0 Hz, 4H), 3.01 (s, 3H), 2.46 (s, 3H), 2.27 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 165.3, 163.4, 144.6, 144.0, 136.3, 134.9, 129.1, 126.4, 126.2, 115.9, 21.8, 21.3, 21.1.



Pink solid, 92% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.71 (s, 2H), 7.23 (d, *J* = 7.9 Hz, 4H), 7.11 (s, 2H), 6.97 (d, *J* = 7.9 Hz, 4H), 3.89 (s, 3H), 3.03 (s, 3H), 2.30 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 165.0, 163.8, 162.8, 144.1, 139.0, 135.2, 129.3, 126.5, 109.1, 108.3, 55.8, 21.4, 21.0. HRMS (ESI): Calcd for C₂₄H₂₅N₆O₅S₂: [M+H]⁺ 541.1328, found: m/z 541.1321.



Pink solid, 85% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.79 (s, 2H), 7.33 – 7.27 (m, 6H), 7.03 (d, *J* = 7.8 Hz, 4H), 3.09 (s, 3H), 2.33 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 165.78, 164.4 (d, *J* = 254.7 Hz), 163.6, 144.4, 139.7 (d, *J* = 13.4 Hz), 135.1, 111.1 (d, *J* = 3.7 Hz), 109.8 (d, *J* = 25.6 Hz), 21.5, 21.2. HRMS (ESI): Calcd for C₂₃H₂₂FN₆O₄S₂: [M+H]⁺ 529.1128, found: m/z 529.1103.



Pink solid, 73% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.29 (s, 2H), 7.75 (s, 2H), 7.18 (d, J = 7.9 Hz, 2H), 6.98 (d, J = 7.9 Hz, 2H), 3.06 (s, 3H), 2.31 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 165.8, 163.4, 144.4, 137.8, 134.9, 129.4, 127.3, 127.0, 126.5, 115.8, 21.4, 21.2. HRMS (ESI): Calcd for C₂₃H₂₂BrN₆O₄S₂: [M+H]⁺ 589.0327, found: m/z 589.0317



Pink solid, 78% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.31 (s, 2H), 7.77 (s, 2H), 7.19 (d, *J* = 7.8 Hz, 4H), 6.98 (d, *J* = 7.9 Hz, 4H), 3.07 (s, 3H), 2.33 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 165.8, 163.4, 144.4, 137.8, 134.9, 129.4, 127.4, 127.2, 126.5, 115.8, 21.5, 21.2. HRMS (ESI): Calcd for C₂₃H₂₂IN₆O₄S₂: [M+H]⁺ 637.0189, found: m/z 637.0185.



Pink solid, 83% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.27 (s, 2H), 7.81 (s, 2H), 7.21 (d, J = 7.9 Hz, 4H), 7.00 (d, J = 7.8 Hz, 4H), 3.11 (s, 3H), 2.33 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 166.3, 163.1, 144.6, 137.9, 134.9, 134.8 (q, J = 34.3 Hz), 134.6, 126.6, 122.5 (q, J = 273.3 Hz), 120.5 (q, J = 3.6 Hz), 119.7, 21.5, 21.3. HRMS (ESI): Calcd for C₂₄H₂₂F₃N₆O₄S₂: [M+H]⁺ 579.1096, found: m/z 579.1090.



Pink solid, 88% yield. ¹H NMR (500 MHz, DMSO- d_6) δ 9.60 (s, 2H), 8.37 (t, J = 6.0 Hz, 1H), 7.44 (d, J = 8.0 Hz, 4H), 7.29 (d, J = 8.0 Hz, 4H), 7.02 (s, 2H), 4.11 (d, J = 6.1 Hz, 2H), 3.06 (s, 3H), 2.35 (s, 6H), 1.84 (s, 3H). ¹³C NMR (125 MHz, DMSO- d_6) δ 169.1, 165.7, 163.7, 143.8, 143.3, 136.5, 129.5, 126.7, 120.6, 119.1, 41.5, 22.4, 21.0. HRMS (ESI): Calcd for C₂₆H₂₈N₇O₅S₂: [M+H]⁺ 582.1593, found: m/z 582.1573.



Pink solid, 98% yield. ¹H NMR (500 MHz, CDCl₃) δ 10.12 (s, 1H), 7.75 (d, *J* = 7.7 Hz, 2H), 7.56 – 7.50 (m, 2H), 7.21 (d, *J* = 7.9 Hz, 2H), 3.04 (s, 3H), 2.35 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 165.79, 162.55, 144.25, 141.00, 136.44, 132.54, 129.79, 127.04, 121.99, 114.18, 77.00, 21.53, 21.21. HRMS (ESI): Calcd for C₁₄H₁₄N₅O₂S₂: [M+H]⁺ 348.0589, found: m/z 348.0595.



Pink solid, 87% yield. ¹H NMR (500 MHz, CDCl₃) δ 10.56 (s, 1H), 8.63 (s, 1H), 7.69 (d, J = 8.8 Hz, 1H), 7.63 (d, J = 8.8 Hz, 1H), 7.52 (d, J = 7.8 Hz, 2H), 7.09 (d, J = 7.8 Hz, 2H), 3.12 (s, 3H), 2.31 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.7, 163.6, 144.2, 137.2, 136.4, 135.8, 132.6, 129.5, 126.9, 124.1, 121.6, 117.9, 21.4, 21.2. HRMS (ESI): Calcd for C₁₆H₁₅BrN₅O₂S: [M+H]⁺ 420.0130, found: m/z 420.0126.



Pink solid, 91% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.12 (s, 2H), 7.68 – 7.59 (m, 3H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.30 – 7.26 (m, 4H), 7.19 (t, *J* = 7.5 Hz, 4H), 3.08 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 165.6, 163.5, 138.0, 136.5, 133.3, 133.0, 128.8, 126.3, 125.5, 118.4, 21.2. HRMS (ESI): Calcd for C₂₁H₁₉N₆O₄S₂: [M+H]⁺ 483.0909, found: m/z 483.0894.



Pink solid, 95% yield. ¹H NMR (500 MHz, DMSO- d_6) δ 9.58 (s, 2H), 7.48 (d, J = 8.7 Hz, 4H), 7.35 (t, J = 8.2 Hz, 1H), 7.06 (d, J = 8.2 Hz, 2H), 7.00 (d, J = 8.7 Hz, 4H), 3.81 (s, 6H), 3.08 (s, 3H). ¹³C NMR (125 MHz, DMSO- d_6) δ 165.7, 163.8, 162.5, 136.7, 131.5, 130.8, 128.9, 122.6, 120.8, 114.2, 55.6, 21.0. HRMS (ESI): Calcd for C₂₃H₂₃N₆O₆S₂: [M+H]⁺ 543.1120, found: m/z 543.1093.



Pink solid, 56% yield. ¹H NMR (500 MHz, DMSO) δ 10.34 (s, 2H), 9.63 (s, 2H), 7.67 (d, J = 8.6 Hz, 4H), 7.47 (d, J = 8.5 Hz, 4H), 7.35 (t, J = 8.2 Hz, 1H), 7.04 (d, J = 8.2 Hz, 2H), 3.06 (s, 3H), 2.08 (s, 6H). ¹³C NMR (125 MHz, DMSO- d_6) δ 169.0, 165.7, 163.8, 143.2, 136.6, 132.7, 131.4, 127.8, 123.2, 121.3, 118.3, 24.1, 21.0. HRMS (ESI): Calcd for C₂₅H₂₅N₈O₆S₂: [M+H]⁺ 597.1338, found: m/z 597.1327.



Pink solid, 63% yield. ¹H NMR (500 MHz, DMSO-*d*₆) δ 10.19 (s, 2H), 8.32 (d, *J* = 8.4 Hz, 4H), 7.73 (d, *J* = 8.4 Hz, 4H), 7.50 (t, *J* = 8.1 Hz, 1H), 7.24 (d, *J* = 8.1 Hz, 2H), 3.02 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 165.9, 163.4, 149.7, 144.8, 135.8, 132.0, 128.1, 124.9, 124.4, 123.7, 20.9. HRMS (ESI): Calcd for C₂₁H₁₇N₈O₈S₂: [M+H]⁺ 573.0611, found: m/z 573.0600.



Pink solid, 65% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.15 (s, 2H), 7.83 (d, *J* = 8.0 Hz, 4H), 7.60 (s, 3H), 7.36 (d, *J* = 8.1 Hz, 4H), 3.95 (s, 6H), 3.08 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.1, 165.0, 163.5, 141.9, 136.3, 134.2, 133.5, 129.9, 126.5, 125.3, 118.2, 52.8, 21.2. HRMS (ESI): Calcd for C₂₅H₂₃N₆O₈S₂: [M+H]⁺ 599.1019, found: m/z 599.1017.



Pink solid, 85% yield. ¹H NMR (500 MHz, DMSO) δ 9.79 (s, 2H), 7.58 (dd, J = 8.1, 5.3 Hz, 4H), 7.43 (t, J = 8.2 Hz, 1H), 7.35 (t, J = 8.6 Hz, 4H), 7.14 (d, J = 8.2 Hz, 2H), 3.06 (s, 3H). ¹³C NMR (125 MHz, DMSO- d_6) δ 165.8, 164.4 (d, J = 251.5 Hz), 163.6, 136.3, 135.5, 131.6, 129.7 (d, J = 9.7 Hz), 123.8, 123.8, 122.1, 116.3 (d, J = 22.9 Hz), 21.0. HRMS (ESI): Calcd for C₂₁H₁₇F₂N₆O₄S₂: [M+H]⁺ 519.0721, found: m/z 519.0715.



Pink solid, 72% yield. ¹H NMR (500 MHz, DMSO-*d*₆) δ 9.87 (s, 2H), 7.89 (d, *J* = 7.9 Hz, 4H), 7.44 (t, *J* = 8.2 Hz, 1H), 7.26 (d, *J* = 7.9 Hz, 4H), 7.14 (d, *J* = 8.2 Hz, 2H), 3.07 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 165.8, 163.6, 138.9, 138.0, 136.2, 131.8, 128.2, 124.0, 122.4, 101.3, 21.0. HRMS (ESI): Calcd for C₂₁H₁₇I₂N₆O₄S₂: [M+H]⁺ 734.8842, found: m/z 734.8847.



Pink solid, 81% yield. ¹H NMR (500 MHz, DMSO-*d*₆) δ 9.71 (s, 2H), 7.75 (dd, *J* = 8.8, 1.5 Hz, 2H), 7.48 (d, *J* = 2.1 Hz, 2H), 7.46 (t, *J* = 8.0 Hz, 1H), 7.17 (d, *J* = 8.2 Hz, 2H), 7.10 (d, *J* = 8.9 Hz, 2H), 3.63 (s, 6H), 3.01 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 165.6, 163.7, 155.6, 149.0, 137.3, 136.4, 131.4, 130.8, 128.6, 123.5, 115.3, 110.7, 56.2, 21.0. HRMS (ESI): Calcd for C₂₃H₂₁Br₂N₆O₆S₂: [M+H]⁺ 698.9331, found: m/z 698.9328.



Pink solid, 61% yield. ¹H NMR (500 MHz, CDCl₃) δ 10.96 (s, 1H), 8.60 (d, J = 8.0 Hz, 1H), 7.85 (d, J = 8.1 Hz, 2H), 7.79 (d, J = 8.3 Hz, 1H), 7.63 (d, J = 8.1 Hz, 2H), 7.57 (t, J = 7.8 Hz, 1H), 7.33 (t, J = 7.7 Hz, 1H), 3.13 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.6, 164.4, 143.4, 137.5, 134.0, 132.7, 130.6, 127.6, 125.2, 121.7, 119.6, 116.9, 116.7, 21.2. HRMS (ESI): Calcd for C₁₆H₁₃N₆O₂S: [M+H]⁺ 353.0821, found: m/z 353.0816.



Pink solid, 93% yield. ¹H NMR (500 MHz, DMSO-*d*₆) δ 10.10 (s, 2H), 8.26 (d, *J* = 8.6 Hz, 2H), 8.15 (d, *J* = 8.2 Hz, 2H), 8.02 (d, *J* = 8.2 Hz, 2H), 7.77 (d, *J* = 7.3 Hz, 2H), 7.62 (t, *J* = 7.5 Hz, 2H), 7.52 (t, *J* = 7.7 Hz, 2H), 7.47 (t, *J* = 7.8 Hz, 2H), 7.29 (t, *J* = 8.2 Hz, 1H), 7.02 (d, *J* = 8.2 Hz, 2H), 2.71 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 165.1, 163.2, 135.8, 134.3, 133.5, 131.3, 128.9, 127.8, 126.8, 126.8, 124.9, 124.3, 124.1, 21.0. HRMS (ESI): Calcd for C₂₉H₂₃N₆O₄S₂: [M+H]⁺ 583.1222, found: m/z 583.1437.



Pink solid, 75% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.28 (s, 2H), 7.71 (d, *J* = 8.0 Hz, 2H), 7.67 – 7.62 (m, 1H), 7.35 (d, *J* = 4.9 Hz, 2H), 7.04 (d, *J* = 3.6 Hz, 2H), 6.80 (t, *J* = 4.3 Hz, 2H), 3.11 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 165.9, 163.6, 138.7, 136.4, 133.4, 132.6, 132.3, 127.3, 125.6, 21.2. HRMS (ESI): Calcd for C₁₇H₁₅N₆O₄S₄: [M+H]⁺ 495.0038, found: m/z 495.0029.



Pink solid, 92% yield. ¹H NMR (500 MHz, CDCl₃) δ 9.58 (s, 2H), 7.60 – 7.52 (m, 3H), 3.16 (s, 3H), 3.14 – 3.06 (m, 4H), 1.67 (p, *J* = 7.5 Hz, 4H), 1.41 – 1.31 (m, 4H), 0.85 (t, *J* = 7.3 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 166.4, 164.9, 138.7, 133.7, 117.5, 112.1, 52.6, 25.3, 21.3, 13.4. HRMS (ESI): Calcd for C₁₇H₂₇N₆O₄S₂: [M+H]⁺ 443.1535, found: m/z 443.1523.

Procedure for the deprotection of 5.

3j (126 mg, 0.3 mmol) was added cold H_2SO_4 (1.0 mL) slowly at 0 °C and stirred at room temperature for 2 h. The reaction mixture was cooled to 0 °C and quenched by cold water (30 mL) and then saturated aqueous NaHCO₃ (20 mL) slowly, extracted with CH₂Cl₂ (20 mL x 3), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified on silica gel column chromatography (CH₂Cl₂/MeOH, 20 : 1) to afford **5** (69 mg, 87%) as a pink solid.



¹H NMR (500 MHz, DMSO- d_6) δ 8.40 (s, 1H), 7.42 (d, J = 8.8 Hz, 1H), 7.11 (s, 2H), 6.90 (d, J = 8.8 Hz, 1H), 2.97 (s, 3H). ¹³C NMR (125 MHz, DMSO- d_6) δ 164.9, 164.0, 148.7, 135.4, 130.7, 119.4, 113.0, 106.2, 20.8. HRMS (ESI): Calcd for C₉H₉BrN₅: [M+H]⁺ 266.0041, found: m/z 266.0055.

Procedure for the synthesis of 6.

To a stirring solution of **5** (53 mg, 0.2 mmol) in DMF (1.0 mL) was added ESF (34 μ L, 0.4 mmol) at room temperature. After t stirring at 60 °C for 3h, the mixture was cooled to room temperature and concentrated under reduced pressure. The residue was purified on silica gel column chromatography (CH₂Cl₂/MeOH, 50 : 1) to afford **6** (60 mg, 80%) as a pink solid.



¹H NMR (500 MHz, CDCl₃) δ 8.86 (s, 1H), 8.69 (s, 1H), 7.56 (d, *J* = 8.8 Hz, 1H), 6.72 (d, *J* = 8.9 Hz, 1H), 4.03 – 3.95 (m, 2H), 3.75 – 3.68 (m, 2H), 3.10 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 165.5, 164.4, 146.4, 136.8, 133.0, 115.1, 112.8, 109.4, 49.6 (d, *J* = 14.8 Hz), 37.3, 21.1. HRMS (ESI): Calcd for C₁₁H₁₂BrFN₅O₂S: [M+H]⁺ 375.9879, found: m/z 375.9892.

Procedure for the synthesie of 7.

4e (177 mg, 0.3 mmol) was added cold H_2SO_4 (1.0 mL) slowly at 0 °C and stirred at room temperature for 2 h. The reaction mixture was cooled to 0 °C and quenched by cold water (30 mL) and then saturated aqueous NaHCO₃ (20 mL) slowly, extracted with CH₂Cl₂ (20 mL x 3), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified on silica gel column chromatography (CH₂Cl₂/MeOH, 20 : 1) to afford 5 (72 mg, 85%) as a pink solid.



¹H NMR (500 MHz, CDCl₃) δ 6.29 (s, 2H), 6.22 and 3.03 (a pair of s, 4H), 3.05 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 167.0, 163.2, 151.1, 128.0, 108.2, 97.4, 21.0. HRMS (ESI): Calcd for C₉H₁₀BrN₅: [M+H]⁺ 281.0150, found: m/z 281.0176.

Procedure for the synthesis of 8.

To a 25 mL test tube was combined 7 (28mg, 0.1 mmol), bromoacetyl chloride (18 μ L, 0.22 mmol) and 3 mL CH₂Cl₂, then TEA (42 μ L, 0.3 mmol) was added at 0 °C and the mixture was stirred at room temperature for 2 h. Then water (20 mL) was added and extracted with CH₂Cl₂ (10 mL x 2), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified on silica gel column chromatography (CH₂Cl₂/acetone, 2 : 1) to afford **8** (31 mg, 60%) as a pink solid.



¹H NMR (500 MHz, CDCl₃) δ 11.12 (s, 2H), 8.30 (s, 2H), 3.98 (s, 4H), 3.19 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.5, 164.5 (two peaks, 164.54 and 164.49), 137.9, 127.4, 123.6, 111.9, 29.3, 21.4. HRMS (ESI): Calcd for C₁₃H₁₂Br₃N₆O₂: [M+H]⁺ 520.8572, found: m/z 520.8583.

Procedure for the synthesis of HP-A-1 and HP-A-2.

To a solution of DNA headpiece (600 μ L, 300 nmol) in borate buffer (250 mM, pH = 9.4), was added a mixture of DMA solution of HATU (60 μ L, 200 mM), DIPEA (60 μ L, 200 mM) and acids (60 μ L, 200 mM). The resultant mixture was vortexed and stood at 25 °C for 8 hours. 5M NaCl (78 μ L) and cold ethanol (2.34 mL) were sequentially added, and the resultant mixture was stored at -80 °C for 30 min. The mixture was centrifuged at 4 °C for 30 min at 12000 rpm to remove the supernatant. The resulting pellet was re-dissolved in ddH₂O (300 μ L), which was used in following reaction without further purification.



The Structure of DNA Headpiece.

Procedure for the synthesis of HP-1-1 and HP-1-2.

The solution of oxidant system was prepared according the literature (*Org. Lett. 2018, 20, 7186–7191*) procedure: briefly,^[2] to a solution of Cu(ClO₄)₂ (100 µL, 100 mM in water) was added a solution of 2,2'-bipyridine (100 µL, 100 mM in DMSO). The resultant mixture was vortexed and stood at 20 °C for 5 min. After that, to the reaction mixture was added a solution of TEMPO (100 µL, 100 mM in DMSO) and the resultant mixture was vortexed to give the oxidant solution (300 µL, 33 mM). To a solution of DNA-conjugated alkenes 10 (10 nmol) in water (10 µL) was added a solution of tetrazine **5** (10 µL, 200 mM in DMSO, 2000 nmol). The mixture was vortexed and stood at 20 °C. When UPLC-MS showed that t DNA-conjugated alkenes was consumed completely, a solution of oxidant system (6 µL, 33 mM, 200 nmol) was added to the reaction mixture. The mixture was vortexed and stood at 20 °C for 4 h. Aqueous sodium diethyldithiocarbamic acid (10 µL, 100 mM, 1000 nmol) was added and the reaction mixture was vortexed and centrifuged. To the supernatant were sequentially added aqueous NaCl (3.6 µL, 5.0 M) and cold ethanol (108 µL). The resultant mixture was stored at -80 °C for 30 min and centrifuged at 4 °C for 30 min at 10000 rpm before the resultant supernatant was removed. The pellet was dissolved in deionized water and analyzed by UPLC-MS to determine the conversion.



III. Mechanistic Studies

Procedure for the synthesis of complex A.

AgOCOCF₃ (88.4 mg, 0.4 mmol) was added to a solution of $[Cp*IrCl_2]_2$ (80.0 mg, 0.1 mmol) in CH₂Cl₂ (20 mL) in the dark and stirred at room temperature for 30 min. **1a** (17.2 mg, 0.2 mmol) and NaOAc (24.6 mg, 0.3 mmol) were then added. The mixture was stirred at room temperature for 12 h and the solution was filtered. The filtrate was evaporated under vacuum and 10 mL (hexane and methyl *tert*-butyl ether) solvent was added to the residue, the title compound complex A was precipitated and collected by filtration as black solid in 92% yield (112.5 mg).



Complex A

¹H NMR (500 MHz, CDCl₃) δ 8.28 (d, J = 7.7 Hz, 1H), 7.98 (d, J = 7.6 Hz, 1H), 7.38 (t, J = 7.5 Hz,

1H), 7.22 (t, J = 7.5 Hz, 1H), 3.14 (s, 3H), 1.74 (s, 15H). ¹³C NMR (125 MHz, CDCl₃) δ 175.8, 165.8, 163.1 (q, J = 36.3 Hz), 160.9, 135.7, 135.6, 133.7, 127.3, 124.1, 113.9 (q, J = 291.6 Hz), 92.0, 20.8, 8.8. ¹⁹F NMR (471 MHz, CDCl₃) δ -74.7 (s). HRMS (ESI): Calcd for C₁₉H₂₂N₆Ir: [M-CF₃CO₂]⁺ 499.1474, found: m/z 499.1456.

IV. Reference

- [1] Mao, W.; Shi, W.; Li, J.; Su, D.; Wang, X.; Zhang, L.; Pan, L.; Wu, X.; Wu, H. Angew. Chem. Int. Ed. 2019, 58, 1106.
- [2] Li, H.; Sun, Z.; Wu, W.; Wang, X.; Zhang, M.; Lu, X.; Zhong, W.; Dai, D. Org. Lett. 2018, 20 (22), 7186–7191.

V. Copies of ¹H, ¹³C NMR, HPLC and MS Spectra.





























200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 fl (ppm)





































































