

## Supporting Information

### Amphiphilic Dendrimers Control Protein Binding and Corona Formation on Liposome Nanocarriers

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# 1 Synthesis of dendrimers with different surface patterns

## 1.1 Materials and Instruments

**Chemicals and Solvents.** All chemicals were obtained from commercial suppliers such as Sigma Aldrich, TCI, Acros Organics, Fisher Scientific, etc., and were utilized without further purification. The solvents (HPLC grade) were purchased from Fisher Scientific or Acros Organics. Chloroform (Rotisolv<sup>®</sup>, HPLC grade) for the *recycling*-gel permeation chromatography (GPC) system was purchased from Carl Roth. The deuterated solvents were purchased from Sigma Aldrich. Argon (Westfalen AG) was used for reactions under inert gas.

**Silica Gel Chromatography.** For thin-layer chromatography (TLC) Alugram Sil G/UV254 plates from Macherey-Nagel were used and the detection of substances was performed under UV light at 254 nm or 366 nm. Macherey-Nagel silica gel with particle size of 0.04–0.063 mm or 0.063–0.2 mm was used for preparative silica gel chromatography.

**Gel Permeation Chromatography (GPC).** Dendrimers with peripheral neopentyl-protected sulfonic acid groups were purified on a *recycling*-GPC purification system from Shimadzu. Dendrimers with deprotected peripheral sulfonic acid groups were purified by size exclusion chromatography using Sephadex<sup>®</sup> LH-20 in DMF.

**Nuclear Magnetic Resonance Spectroscopy (NMR).** All <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on a Bruker Avance III 300 MHz, Avance III 500 MHz, Avance III 700 MHz or Bruker Avance III 850 MHz spectrometer in deuterated solvents such as CD<sub>2</sub>Cl<sub>2</sub>, THF-*d*<sub>8</sub> and DMSO-*d*<sub>6</sub> at 298 K. <sup>13</sup>C NMR were recorded in *j*-modulated spin-echo (JMOD) mode. Spectra were analyzed by applying the software MestReNova.

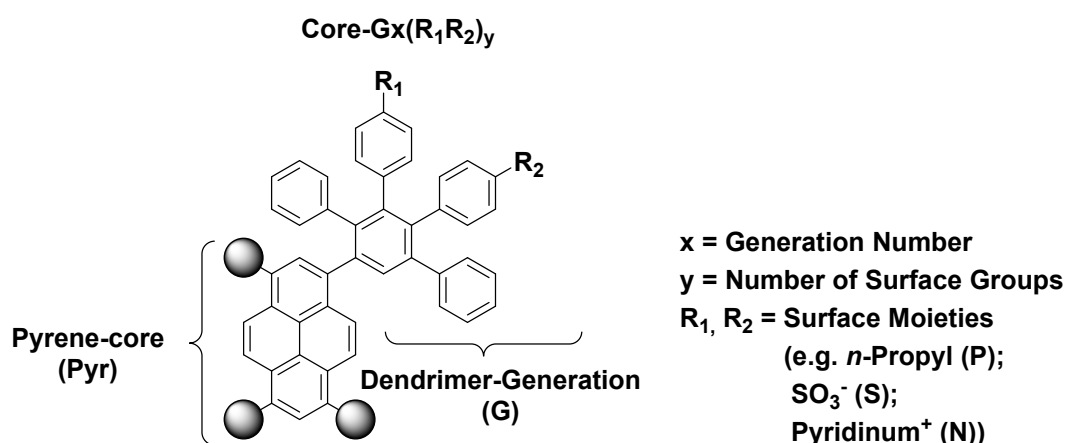
**Matrix-assisted Laser Desorption/Ionization–Time of Flight (MALDI-TOF) Mass Spectrometry.** MALDI-TOF mass spectra were measured on a Waters MALDI Synapt G2-SI or Bruker rapifleX MALDI-TOF/TOF. Precursors and uncharged dendrimers were dissolved in THF and measured by applying *trans*-2-[3-(4-*tert*-butylphenyl)-2-methyl-2-propenylidene]-malononitrile (DCTB) or dithranol as matrix. Deprotected sulfonic acid based dendrimers were solvated in DMF and diluted in a saturated  $\alpha$ -cyano-4-hydroxycinnamic acid (CHCA) solution in water:acetonitrile (1:1) + 0.1% TFA. All dendrimers with peripheral pyridine groups were measured in tetrahydrofuran (THF) with tetracyanoquinodimethane (TCNQ) as matrix. Processing of data was performed in mMass.

**Electrospray Ionization (ESI) Spectrometry.** ESI spectra were measured on a Waters Synapt G2-SI. The samples were dissolved in methanol.

**Atmospheric Pressure Chemical Ionization Mass Spectrometry (APCI-MS).** APCI mass spectra of building blocks and intermediates were measured on an Advion expression-L Compact Mass Spectrometer (CMS) (Advion Inc. 61 Brown Rd, Suite 100, Ithaca, NY 14850, USA) by either applying the sample (solid) as an atmospheric solid analysis probe (ASAP) or measuring directly from TLC plates by an automated TLC plate reader (Plate express).

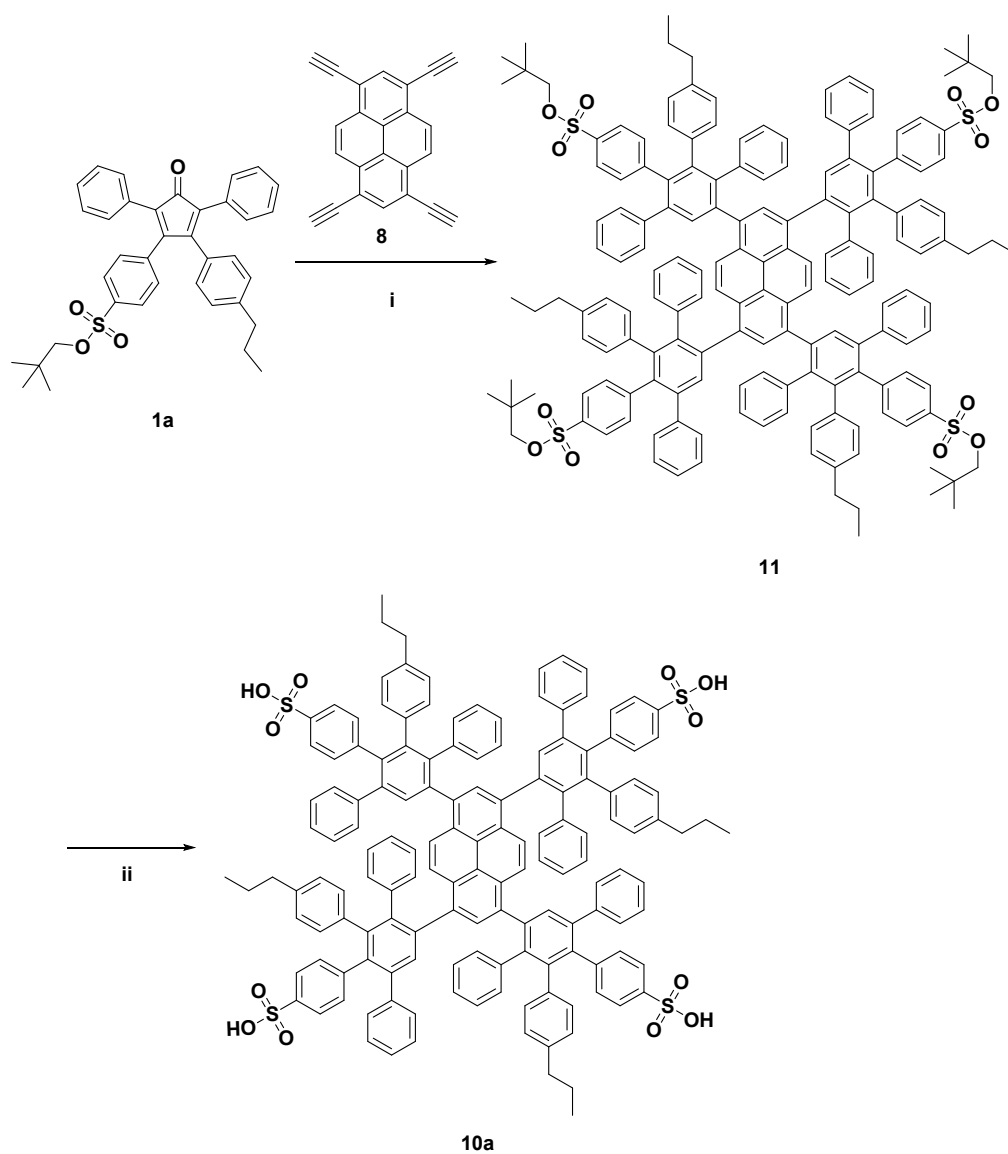
**Field Desorption Mass Spectrometry (FD-MS).** FD mass spectra of intermediates were measured on a VG Instruments ZAB 2-SE-FPD using an 8 kV accelerating voltage.

### Nomenclature for Polyphenylene Dendrimers.



**Figure S1.** Systematic designation of polyphenylene dendrimers (PPDs)

## 1.2 Synthesis of a first-generation negatively charged amphiphilic polyphenylene dendrimer – (PS)<sub>4</sub>



**Figure S2.** Synthesis of first-generation amphiphilic dendrimer **10a** with alternating sulfonic acid and *n*-propyl groups according to Stangenberg *et al.*<sup>1</sup> i) *o*-xylene, 145 °C, 48 h, 45% yield; ii) DMF, 180 °C, 48 h, 85% yield.

*Neopentyl 4-(3-oxo-2,4-diphenyl-5-(4-propylphenyl)cyclopenta-1,4-dien-1-yl)benzenesulfonate (1a)*

Surface building block **1a** was synthesized according to the literature in a four-step synthesis.<sup>2</sup>

<sup>3</sup> All spectral data were in agreement with the literature.<sup>2, 3</sup>

<sup>1</sup>H NMR (300 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ(ppm) = 7.70 (d, *J* = 8.5 Hz, 2H), 7.31–7.22 (m, 8H), 7.20–7.12 (m, 4H), 7.01 (d, *J* = 8.3 Hz, 2H), 6.83 (d, *J* = 8.2 Hz, 2H), 3.64 (s, 2H), 2.54 (t, *J* = 7.8, 7.2 Hz, 2H), 1.61 (dt, *J* = 13.7, 7.4 Hz, 2H), 0.92 (t, *J* = 7.3 Hz, 3H), 0.87 (s, 9H).

<sup>13</sup>C NMR (75 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ(ppm) = 200.22, 155.08, 152.76, 144.50, 139.78, 135.83, 131.36, 130.72, 130.66, 130.32, 129.56, 128.84, 128.74, 128.56, 128.09, 127.61, 125.62, 80.56, 38.26, 32.05, 26.27, 24.77, 14.02.

FD-MS: *m/z* calcd. for C<sub>37</sub>H<sub>36</sub>O<sub>4</sub>S: 576.7, found 576.8 [M]<sup>+</sup>.

*1,3,6,8-Tetraethynylpyrene (8)*

Pyrene core **8** was synthesized in three steps according to the literature.<sup>4</sup>

*Pyr-G1-(PSpen)<sub>4</sub> 11*

The synthesis of neopentyl-protected dendrimer **11** was modified from the previously reported method.<sup>1</sup> 1,3,6,8-Tetraethynylpyrene (**8**) (10 mg, 33.5 μmol) and building block **1a** (116 mg, 201 μmol, 6 equiv) were dissolved in 3 mL *o*-xylene. The mixture was degassed with argon and stirred at 145 °C for 48 h in a sealed microwave tube. The product was precipitated in methanol, filtered off and purified by silica gel chromatography using a mixture of cyclohexane and THF (3:1). The product was further purified by *recycling*-GPC in chloroform to obtain dendrimer **11** as a yellow solid (38 mg, 45%).

<sup>1</sup>H NMR (700 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ (ppm) = 7.98–7.73 (m, 5H), 7.60–7.31 (m, 11H), 7.26–6.23 (m, 66H), 3.54–3.30 (m, 8H), 2.50–2.27 (m, 8H), 1.52–1.38 (m, 8H), 0.88–0.66 (m, 48H).

<sup>13</sup>C NMR (176 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ(ppm) = 147.44, 147.16, 142.17, 141.36, 141.06, 140.65, 140.53, 140.10, 139.77, 138.10, 137.26, 137.01, 136.65, 136.32, 132.99, 132.81, 131.67, 130.37, 130.28, 128.20, 128.15, 127.92, 127.79, 127.45, 126.99, 126.72, 80.04, 37.68, 26.10, 24.67, 13.49.

MALDI-TOF: *m/z* calcd. for C<sub>168</sub>H<sub>154</sub>O<sub>12</sub>S<sub>4</sub> 2491.03, found 2492.98 [M+H]<sup>+</sup>.

### *Pyr-G1-(PS)<sub>4</sub> 10a*

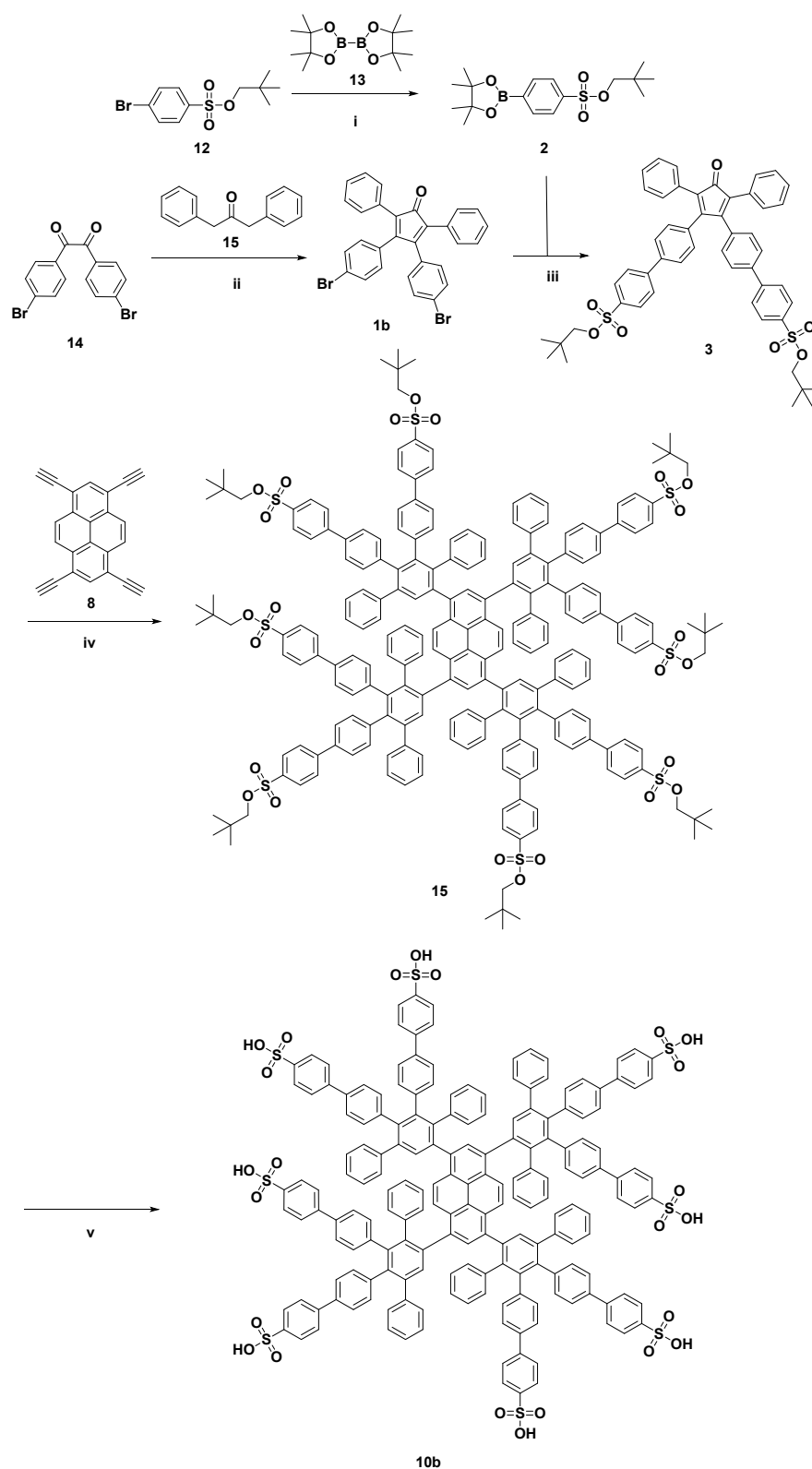
Negatively charged amphiphilic dendrimer **10a** was synthesized according to the literature<sup>1</sup> with a modified purification protocol. Briefly, neopentyl-protected dendrimer **11** (12.0 mg, 4.81  $\mu$ mol) was dissolved in 2 mL dry DMF, the resulting solution was degassed with argon and stirred at 180 °C for 48 h in a sealed microwave tube. Subsequently, DMF was removed *in vacuo*. The residue was dissolved in methanol and precipitated in diethyl ether. The resulting product was further purified by gel permeation chromatography (GPC) applying Sephadex LH-20 in DMF to obtain *Pyr-G1-(PS)<sub>4</sub> 10a* as a light yellow solid (9 mg, 85%).

<sup>1</sup>H NMR (700 MHz, DMSO):  $\delta$  (ppm) = 8.02–6.20 (m, 82H), 2.44–2.28 (m, 8H), 1.52–1.31 (m, 8H), 0.79–0.58 (m, 12H).

<sup>13</sup>C NMR (176 MHz, DMSO):  $\delta$  (ppm) = 145.51, 145.41, 141.11, 141.02, 139.64, 139.27, 137.01, 130.93, 130.36, 129.61, 129.54, 127.74, 127.70, 127.46, 127.43, 127.12, 126.72, 125.09, 124.18, 123.89, 36.54, 23.50, 13.12.

MALDI-TOF: *m/z* calcd. for C<sub>148</sub>H<sub>114</sub>O<sub>12</sub>S<sub>4</sub> 2210.72, found 2210.73 [M]<sup>•+</sup>.

### 1.3 Synthesis of a first-generation negatively charged polyphenylene dendrimer – S<sub>8</sub>



**Figure S3.** Synthesis of first-generation negatively charged dendrimer **10b**. i) KOAc, Pd(dppf)Cl<sub>2</sub>, 1,4-dioxane, 90 °C, 6 h, 99% yield; ii) TBAH, ethanol, 85 °C, 0.5 h, 85% yield; iii) Pd(PPh<sub>3</sub>)<sub>4</sub>, K<sub>2</sub>CO<sub>3</sub> (aq), 1,4-dioxane, 90 °C, 15 h, 34% yield; iv) *o*-xylene, 145 °C, 36 h, 27% yield; v) DMF, 180 °C, 48 h, 88% yield.



### Neopentyl 4-bromobenzenesulfonate (**12**)

Compound **17** was synthesized according to the literature.<sup>2, 3</sup> All spectral data were in agreement with the literature.<sup>2</sup>

<sup>1</sup>H NMR (300 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ(ppm) = 7.79–7.70 (m, 4H), 3.67 (s, 2H), 0.89 (s, 9H).

<sup>13</sup>C NMR (75 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ(ppm) = 135.64, 133.12, 129.95, 129.27, 80.63, 32.08, 26.22.

FD-MS: *m/z* calcd. for C<sub>11</sub>H<sub>15</sub>BrO<sub>3</sub>S: 307.2, found 308.7 [M+H]<sup>+</sup>.

### Neopentyl 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzenesulfonate (**2**)

In a dry Schlenk flask neopentyl 4-bromobenzenesulfonate (**12**) (1.00 g, 3.26 mmol), bis(pinacolato)diboron (**13**) (1.24 g, 4.88 mmol, 1.5 equiv) and potassium acetate (703 mg, 7.16 mmol, 2.2 equiv) were dissolved in 15 mL dry 1,4-dioxane. After degassing with argon [1,1'-bis(diphenylphosphino)ferrocene]dichloropalladium(II) (Pd(dppf)Cl<sub>2</sub>) (53.2 mg, 65.1 μmol, 0.02 equiv) was added under argon atmosphere. The reaction mixture was stirred at 90 °C for 6 h under argon atmosphere. Subsequently, ethyl acetate was added and the organic layer was washed with water. The organic layer was dried over magnesium sulphate. After removing the solvent *in vacuo* the crude product was purified by silica gel column chromatography using a mixture of cyclohexane and ethyl acetate (10:1) to obtain compound **2** as a colourless solid (1.14 g, 99%).

<sup>1</sup>H NMR (300 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ (ppm) = 7.95 (d, *J* = 8.1 Hz, 2H), 7.86 (d, *J* = 8.1 Hz, 2H), 3.65 (s, 2H), 1.35 (s, 12H), 0.88 (s, 9H).

<sup>13</sup>C NMR (75 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ (ppm) = 138.47, 135.62, 127.23, 122.32, 84.95, 83.63, 79.89, 31.89, 26.50, 25.26, 25.06.

APCI-MS: *m/z* calcd. for C<sub>17</sub>H<sub>27</sub>BO<sub>5</sub>S: 354.2, found 371.7 [M+NH<sub>4</sub>]<sup>+</sup>.

### 3,4-Bis(4-bromophenyl)-2,5-diphenylcyclopenta-2,4-dien-1-one (**1b**)

1,2-Bis(4-bromophenyl)ethane-1,2-dione (**14**) (1.5 g, 4.08 mmol) and 1,3-diphenylacetone (**15**) (857 mg, 4.08 mmol) were dissolved in 150 mL ethanol and heated up to 85 °C. Subsequently, 1 M methanolic tetrabutylammonium hydroxide (TBAH) solution (1.06g, 4.08 mL, 4.08 mmol) was added. After stirring at 85 °C for 0.5 h, the reaction mixture was diluted in ethyl acetate. The organic layer was washed with water, dried over sodium sulphate and concentrated *in vacuo*. The product was precipitated in an ethanol water mixture to afford building block **1b** as a dark red solid (1.89 g, 85%).

<sup>1</sup>H NMR (300 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ (ppm) = 7.41–7.32 (m, 4H), 7.32–7.25 (m, 6H), 7.24–7.15 (m, 4H), 6.90–6.77 (m, 4H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  (ppm) = 153.28, 132.24, 131.81, 131.38, 130.85, 130.49, 128.54, 128.18, 126.38, 123.35.

APCI-MS:  $m/z$  calcd. for  $\text{C}_{29}\text{H}_{18}\text{Br}_2\text{O}$  539.97, found 540.7  $[\text{M}+\text{H}]^+$ .

*Dineopentyl 4',4'''-(4-oxo-3,5-diphenylcyclopenta-2,5-diene-1,2-diyl)bis-([1,1'-biphenyl]-4-sulfonate) (3)*

Building block **1b** (700mg, 1.29 mmol) and neopentyl 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzenesulfonate (**2**) were dissolved in 40 mL 1,4-dioxane. Then, an aqueous 2 M potassium carbonate solution (2.68 g, 9.68 mL, 19.4 mmol) was added and the mixture was degassed with argon. Tetrakis(triphenylphosphine)palladium(0) ( $\text{Pd}(\text{PPh}_3)_4$ ) was added and the reaction mixture was stirred at 90 °C overnight under argon atmosphere. The crude mixture was diluted with dichloromethane and the organic layer was washed with water and dried over sodium sulfate. After removing the solvent *in vacuo* the crude product was purified by silica gel column chromatography using a mixture of cyclohexane and THF (5:1) to obtain building block **3** as a red solid (340 mg, 34%).

$^1\text{H}$  NMR (300 MHz,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  (ppm) = 7.85 (dd,  $J$  = 48.4, 8.3 Hz, 8H), 7.52 (d,  $J$  = 8.1 Hz, 4H), 7.33–7.22 (m, 10H), 7.12 (d,  $J$  = 8.2 Hz, 4H), 0.89 (s, 18H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  = 200.21, 153.98, 145.80, 139.36, 135.30, 133.95, 131.15, 130.58, 128.84, 128.52, 128.11, 127.98, 127.30, 126.54, 80.23, 31.92, 26.10.

MALDI-TOF:  $m/z$  calcd. for  $\text{C}_{51}\text{H}_{48}\text{O}_7\text{S}_2$  3531.24, found 836.39  $[\text{M}]^{++}$ .

### *Pyr-G1-(Spen)<sub>8</sub> (15)*

1,3,6,8-Tetraethynylpyrene (**8**) (6 mg, 20.1  $\mu$ mol) and building block **3** (101 mg, 121  $\mu$ mol, 6 equiv) were dissolved in 3 mL *o*-xylene, degassed with argon and stirred at 145 °C for 36 h in a sealed microwave tube. The solvent was removed *in vacuo* and the crude product was purified by silica gel chromatography using a mixture of cyclohexane and THF (2:1). The product was further purified by *recycling*-GPC in chloroform to obtain dendrimer **15** as a yellow solid (19 mg, 27%).

<sup>1</sup>H NMR (700 MHz, CD<sub>2</sub>Cl<sub>2</sub>):  $\delta$  (ppm) = 8.03 – 7.80 (m, 21H), 7.71 – 7.48 (m, 19H), 7.38 – 6.38 (m, 75H), 3.71 – 3.57 (m, 16H), 0.93 – 0.81 (m, 72H).

<sup>13</sup>C NMR (176 MHz, CD<sub>2</sub>Cl<sub>2</sub>):  $\delta$ (ppm) = 146.12, 141.80, 141.54, 141.24, 140.17, 138.98, 136.90, 136.54, 136.19, 134.78, 132.82, 130.42, 130.33, 128.72, 128.68, 128.12, 128.07, 127.70, 127.61, 126.88, 126.28, 125.94, 80.12, 31.89, 26.09.

MALDI-TOF: *m/z* calcd. for C<sub>224</sub>H<sub>202</sub>O<sub>24</sub>S<sub>8</sub> 3531.24, found 3532.72 [M+H]<sup>+</sup>.

### *Pyr-G1-S<sub>8</sub> (10b)*

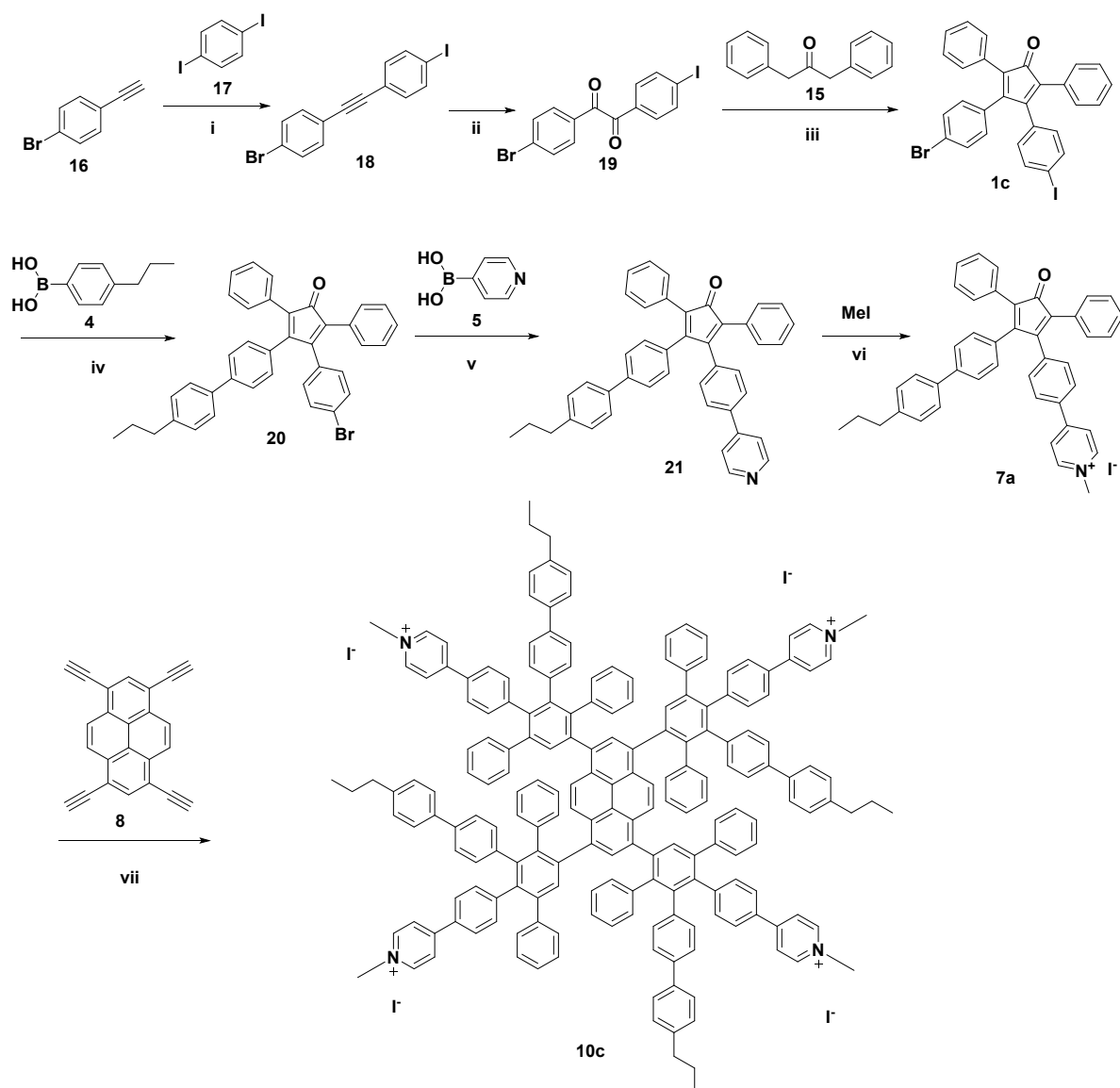
Pyr-G1-(Spen)<sub>8</sub> **15** (7.00 mg, 1.98  $\mu$ mol) was dissolved in 2 mL dry DMF, the resulting solution was degassed with argon and stirred at 180 °C in a sealed microwave tube for 48 h. The solvent was evaporated *in vacuo* and the product was precipitated in diethyl ether. The product was further purified by gel permeation chromatography (Sephadex LH-20) in DMF. Dendrimer **10b** was obtained as a slight yellow solid (5.2 mg, 88%).

<sup>1</sup>H NMR (700 MHz, DMSO):  $\delta$  (ppm) = 7.98–7.86 (m, 2H), 7.85–7.75 (m, 2H), 7.65–6.31 (m, 110H).

<sup>13</sup>C NMR (176 MHz, DMSO):  $\delta$  (ppm) = 147.06, 146.43, 141.53, 140.92, 139.74, 139.06, 138.45, 136.59, 136.14, 131.79, 129.62, 127.63, 126.61, 125.99, 125.94, 125.43, 125.35, 125.01, 124.96, 124.51.

MALDI-TOF: *m/z* calcd. for C<sub>184</sub>H<sub>122</sub>O<sub>24</sub>S<sub>8</sub> 2970.61, found 2971.59 [M+H]<sup>+</sup>, 2994.56 [M+Na]<sup>+</sup>, 3009.56 [M+K]<sup>+</sup>.

## 1.4 Synthesis of a first-generation positively charged amphiphilic polyphenylene dendrimer –(PN)<sub>4</sub>



**Figure S4.** Synthesis of positively charged first-generation dendrimer **10c** with alternating pyridinium and *n*-propyl groups. i) CuI, PPh<sub>3</sub>, PdCl<sub>2</sub>(Ph<sub>3</sub>P)<sub>2</sub>, 1,4-dioxane, 40 °C, 15 h, 60% yield; ii) I<sub>2</sub>, [Ru(*p*-cymene)Cl<sub>2</sub>]<sub>2</sub>, *tert*-butyl hydroperoxide (70% in H<sub>2</sub>O), 1,4-dioxane, 50 °C, 15 h, 86% yield; iii) TBAH, ethanol, 85 °C, 20 min, 60% yield; iv) Pd(PPh<sub>3</sub>)<sub>4</sub>, K<sub>2</sub>CO<sub>3</sub> (aq), 1,4-dioxane, 40 °C, 15 h; v) Pd(PPh<sub>3</sub>)<sub>4</sub>, K<sub>2</sub>CO<sub>3</sub> (aq), 1,4-dioxane, 80 °C, 15 h, 60 % yield; vi) methanol, RT, 48 h, 70% yield; vii) DMSO, 140 °C, 3 d, 47% yield.

### 1-Bromo-4-((4-iodophenyl)ethynyl)benzene (**18**)

4-Bromophenylacetylene (2.0 g, 11.05 mmol) (**16**), 1,4-diiodobenzene (**17**) (7.29 g, 22.10 mmol, 2 equiv), copper(I) iodide (210 mg, 1.1 mmol, 0.1 equiv) and triphenylphosphine (579 mg, 2.21 mmol, 0.2 equiv) were dissolved in 200 mL 1,4-dioxane in a 500 mL flask. To this mixture 50 mL triethylamine was added and the solution was degassed with argon. Bis(triphenylphosphine)palladium(II) dichloride ( $\text{PdCl}_2(\text{Ph}_3\text{P})_2$ ) (338 mg, 0.552 mmol) was added and the solution was stirred at 40 °C under argon atmosphere overnight. Then, the solution was filtered and the solvent was removed *in vacuo* followed by an extraction with dichloromethane. The organic layer was dried over magnesium sulphate and purified by silica gel flash chromatography using hexane as eluent to obtain compound **18** as a bright yellow solid (2.54 g, 60 %).

$^1\text{H}$  NMR (300 MHz,  $\text{THF-}d_8$ ):  $\delta$  (ppm) = 7.75 (d,  $J$  = 7.9 Hz, 2 H), 7.55 (d,  $J$  = 8.0 Hz, 2 H), 7.42 (d,  $J$  = 8.1 Hz, 2 H), 7.27 (d,  $J$  = 8.0 Hz, 2 H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{THF-}d_8$ ):  $\delta$  (ppm) = 138.44, 134.62, 133.71, 133.62, 132.64, 132.43, 123.31, 123.17, 122.81, 95.06, 90.08.

APCI-MS:  $m/z$  calcd. for  $\text{C}_{14}\text{H}_8\text{BrI}$ : 381.9, found 383.9  $[\text{M}+2\text{H}]^+$ .

### 1-(4-Bromophenyl)-2-(4-iodophenyl)ethane-1,2-dione (**19**)

Tolane **18** (1.0 g, 2.61 mmol), iodine (132 mg, 0.522 mmol, 0.2 equiv) and dichloro(*p*-cymene)ruthenium(II) dimer (16 mg, 0.026 mmol, 0.01 equiv) were added to a 50 mL flask and dissolved in as little 1,4-dioxane as possible. 1.5 mL (10.44 mmol, 4 equiv) of a 70 % *tert*-butyl hydroperoxide solution was slowly added to the solution. The mixture was stirred at 50 °C overnight. After evaporation of the solvent and purification via silica gel flash column chromatography using hexane as eluent followed by THF benzil **19** was obtained as a yellow solid (931 mg, 86 %).

$^1\text{H}$  NMR (300 MHz,  $\text{THF-}d_8$ ):  $\delta$  (ppm) = 7.97 (d,  $J$  = 8.5 Hz, 2H), 7.88 (d,  $J$  = 8.6 Hz, 2H), 7.75 (d,  $J$  = 8.5 Hz, 2H), 7.70 (d,  $J$  = 8.5 Hz, 2H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{THF-}d_8$ ):  $\delta$  (ppm) = 193.57, 193.21, 139.43, 138.19, 133.33, 132.99, 132.27, 131.86, 130.91, 128.89, 125.95, 121.07, 104.30.

### *3-(4-Bromophenyl)-4-(4-iodophenyl)-2,5-diphenylcyclopenta-2,4-dien-1-one (1c)*

Benzil **19** (500 mg, 1.2 mmol) and 1,3-diphenylacetone **3** (279 mg, 1.33 mmol, 1.1 equiv) were dissolved in 15 mL ethanol in a 50 mL flask. The solution was heated under reflux and 0.24 mL tetrabutylammonium hydroxide (1M in THF) were added. After 20 min the reaction was stopped by cooling in an ice bad. After cooling at  $-20\text{ }^{\circ}\text{C}$  for a few hours the product was filtered off. Building block **1c** was received as a purple solid (424 mg, 60 %).

$^1\text{H}$  NMR (300 MHz,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  (ppm) = 7.56 (d,  $J$  = 8.3 Hz, 2 H), 7.35 (d,  $J$  = 8.5 Hz, 2 H), 7.31–7.24 (m, 6 H), 7.24–7.17 (m, 4 H), 6.82 (d,  $J$  = 8.5 Hz, 2 H), 6.68 (d,  $J$  = 8.5 Hz, 2 H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  (ppm) = 200.17, 153.53, 153.42, 137.91, 132.92, 132.38, 131.96, 131.59, 131.53, 131.02, 131.01, 130.64, 128.70, 128.33, 126.57, 126.50, 123.50, 95.41.

APCI-MS:  $m/z$  calcd. for  $\text{C}_{29}\text{H}_{18}\text{BrIO}$ : 588.0, found 589.1  $[\text{M}+\text{H}]^+$ .

### *3-(4-Bromophenyl)-2,5-diphenyl-4-(4'-propyl-[1,1'-biphenyl]-4-yl)cyclopenta-2,4-dien-1-one (20)*

Building block **1c** (230 mg, 0.390 mmol) and 4-propylbenzeneboronic acid (**4**) (83 mg, 0.507 mmol, 1.3 equiv) were added to a 50 mL flask and were dissolved in 25 mL 1,4-dioxane. To this solution 0.97 mL of a 2 M potassium carbonate solution were added and degassed with argon. Then,  $\text{Pd}(\text{PPh}_3)_4$  (27 mg, 0.023 mmol, 0.06 equiv) was added and the reaction mixture was stirred at  $40\text{ }^{\circ}\text{C}$  overnight. The reaction mixture was filtered and extracted with DCM followed by a filtration over a silica gel flash column using a mixture of THF and hexane (1:19). Product **20** was used without further purification.

APCI-MS:  $m/z$  calcd. for  $\text{C}_{38}\text{H}_{29}\text{BrO}$ : 580.1, found 581.5  $[\text{M}+\text{H}]^+$ .

### *2,5-Diphenyl-3-(4'-propyl-[1,1'-biphenyl]-4-yl)-4-(4-(pyridin-4-yl)phenyl)cyclopenta-2,4-dien-1-one (21)*

Building block **20** (380 mg, 0.653 mmol) and 4-pyridineboronic acid (**5**) (321 mg, 2.61 mmol, 4 equiv) were dissolved in 25 mL 1,4-dioxane in a 50 mL flask. 3.27 mL of a 2M potassium carbonate solution were added and the mixture was degassed. After adding  $\text{Pd}(\text{PPh}_3)_4$  (45 mg, 0.038 mmol, 0.06 equiv) the mixture was stirred at  $80\text{ }^{\circ}\text{C}$  for two days under argon atmosphere. The solvent was removed followed by an extraction with DCM. The purification was performed by silica gel flash chromatography using a mixture of THF and hexane (1:2). Building block **21** was received as a violet solid (236 mg, 60%).

$^1\text{H}$  NMR (300 MHz,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  (ppm) = 8.64–8.57 (m, 2 H), 7.58–7.43 (m, 8 H), 7.32–7.20 (m, 12 H), 7.12 (d,  $J$  = 8.4 Hz, 2 H), 7.03 (d,  $J$  = 8.4 Hz, 2 H), 2.66–2.55 (m, 2 H), 1.70–1.56 (m, 2 H), 0.94 (t,  $J$  = 7.3 Hz, 3 H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  (ppm) = 200.53, 154.81, 154.22, 152.06, 150.89, 150.83, 148.64, 147.60, 143.18, 141.65, 138.71, 138.51, 137.80, 136.47, 134.64, 132.17, 131.58, 131.44, 130.83, 130.78, 130.50, 129.66, 129.60, 129.58, 129.17, 128.87, 128.68, 128.65, 128.21, 128.12, 127.53, 127.15, 127.03, 126.80, 126.70, 126.13, 125.99, 122.09, 121.78, 38.16, 25.15, 14.17.

APCI-MS:  $m/z$  calcd. for  $\text{C}_{43}\text{H}_{33}\text{NO}$ : 579.3, found 580.4  $[\text{M}+\text{H}]^+$ .

*1-Methyl-4-(4-(3-oxo-2,4-diphenyl-5-(4'-propyl-[1,1'-biphenyl]-4-yl)cyclopenta-1,4-dien-1-yl)phenyl)pyridin-1-ium (7a)*

Building block **21** (220 mg, 0.38 mmol) was dissolved in 1 mL methyl iodide in a 10 mL Schlenk tube followed by the addition of 1 mL methanol. After two days, the product was precipitated by the addition of diethyl ether. After filtration, compound **7a** was obtained as a red-brown solid (192 mg, 70%).

$^1\text{H}$  NMR (300 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  (ppm) = 8.96 (d,  $J$  = 6.6 Hz, 2 H), 8.46 (d,  $J$  = 7.1 Hz, 2 H), 8.00 (d,  $J$  = 8.5 Hz, 2 H), 7.59 (d,  $J$  = 2.7 Hz, 2 H), 7.56 (d,  $J$  = 2.5 Hz, 2 H), 7.35–7.16 (m, 14 H), 7.08 (d,  $J$  = 8.4 Hz, 2 H), 4.29 (s, 3 H), 2.56 (t,  $J$  = 7.5 Hz, 2 H), 1.58 (h,  $J$  = 7.4 Hz, 2 H), 0.87 (t,  $J$  = 7.3 Hz, 3 H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  (ppm) = 199.29, 154.27, 153.12, 153.03, 145.53, 141.99, 139.93, 136.65, 136.13, 133.09, 131.26, 130.43, 130.31, 130.20, 129.86, 129.59, 128.16, 128.11, 127.79, 127.60, 126.29, 126.11, 125.82, 124.93, 123.88, 47.04, 36.74, 23.91, 13.52.

ESI-MS:  $m/z$  calcd. for  $\text{C}_{44}\text{H}_{36}\text{NO}^+$ : 594.28, found 592.22  $[\text{M}+\text{H}]^+$ .

### *Pyr-G1(PN)<sub>4</sub> (10c)*

Building block **7a** (100 mg, 0.17 mmol) and 1,3,6,8-tetraethynylpyrene (**8**) (6.9 mg, 0.023 mmol, 0.14 equiv) were dissolved in 5 mL DMSO in a 10 mL Schlenk tube. After degassing with argon, the mixture was heated up to 140 °C for three days. After the addition of diethyl ether product **10c** precipitated and was filtered off. Purification was performed applying dialyses (1000 MWCO) in methanol. Dendrimer **10c** was received as yellow crystals (33.3 mg, 47 %).

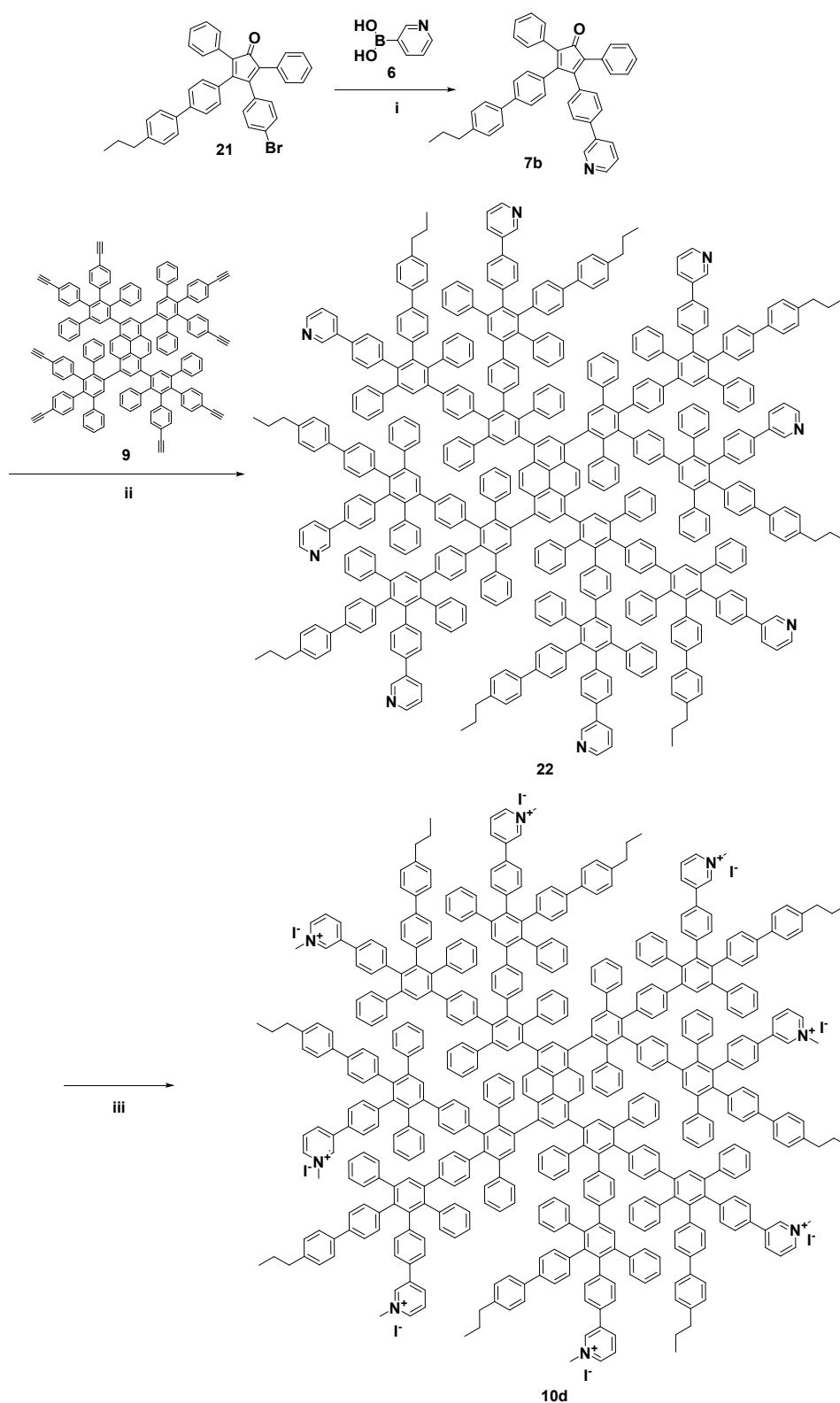
<sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>): δ (ppm) = 8.90 (d, J = 6.7 Hz, 8 H), 8.47–6.04 (m, 106 H), 4.25 (s, 12 H), 1.65–1.42 (m, 8 H), 0.86 (t, J = 6.5 Hz, 12 H).

<sup>13</sup>C-NMR (126 MHz, DMSO-*d*<sub>6</sub>): δ (ppm) = 153.09, 145.46, 141.13, 140.68, 136.32, 132.60, 131.58, 130.31, 129.66, 128.88, 127.88, 126.60, 125.91, 124.83, 123.33, 46.88, 36.71, 24.64, 13.53.

ESI-MS: *m/z* calcd. for (C<sub>196</sub>H<sub>154</sub>N<sub>4</sub>)<sup>4+</sup>: 641.31, found 641.14; *m/z* calcd. for (C<sub>196</sub>H<sub>154</sub>N<sub>4</sub>)<sup>3+</sup>: 897.38, found 897.31; *m/z* calcd. for (C<sub>196</sub>H<sub>154</sub>N<sub>4</sub>)<sup>2+</sup>: 1409.52, found 1409.44.



## 1.5 Synthesis of a second-generation positively charged amphiphilic polyphenylene dendrimer – (PN)<sub>8</sub>



**Figure S5.** Synthesis of second-generation dendrimer **15d** with alternating pyridinum and *n*-propyl groups. i) Pd(PPh<sub>3</sub>)<sub>4</sub>, K<sub>2</sub>CO<sub>3</sub> (aq), 1,4-dioxane, 80 °C, 15 h, 50 % yield; ii) *o*-xylene, 160 °C, 3 d, 64% yield; iii) methanol, RT, 15 h, 92% yield.

*2,5-Diphenyl-3-(4'-propyl-[1,1'-biphenyl]-4-yl)-4-(4-(pyridin-3-yl)phenyl)cyclopenta-2,4-dien-1-one (7b)*

Compound **21** (380 mg, 0.653 mmol) and 4-pyridineboronic acid (**6**) (321 mg, 2.61 mmol, 4 equiv) were dissolved in 25 mL 1,4-dioxane in a 50 mL flask. 3.27 mL of a 2M potassium carbonate solution were added and the mixture was degassed with argon. After adding Pd(PPh<sub>3</sub>)<sub>4</sub> (45 mg, 0.038 mmol, 0.06 equiv) the mixture was stirred at 80 °C for two days under argon atmosphere. The solvent was removed *in vacuo* followed by an extraction with dichloromethane (DCM). The purification was performed by silica gel flash chromatography using a mixture of THF and hexane (1:2). Building block **7b** was received as a violet solid (112 mg, 50%).

<sup>1</sup>H NMR (300 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ (ppm) = 8.82 (d, J = 2.4 Hz, 1 H), 8.55 (dd, J = 4.9, 1.6 Hz, 1 H), 7.88 (dt, J = 8.1, 2.1 Hz, 1 H), 7.55–7.43 (m, 5 H), 7.27 (d, J = 9.3 Hz, 14 H), 7.11 (d, J = 8.2 Hz, 2 H), 7.04 (d, J = 8.3 Hz, 2 H), 2.61 (t, J = 7.6 Hz, 2 H), 1.74–1.54 (m, 2 H), 0.94 (t, J = 7.3 Hz, 3 H).

<sup>13</sup>C NMR (75 MHz, CD<sub>2</sub>Cl<sub>2</sub>): δ (ppm) = 200.61, 154.85, 154.44, 149.31, 148.66, 143.16, 141.61, 138.41, 137.82, 136.12, 134.58, 133.56, 132.22, 131.62, 131.52, 130.79, 130.76, 130.50, 129.55, 128.63, 128.61, 128.14, 128.07, 127.14, 127.06, 126.76, 126.53, 126.13, 125.97, 124.08, 38.14, 25.12, 14.12.

APCI-MS: *m/z* calcd. for C<sub>43</sub>H<sub>33</sub>NO: 579.3, found 580.4 [M+H]<sup>+</sup>.

*Pyr-G1-(ethynyl)<sub>8</sub> 9*

First generation dendrimer **9** was synthesized according to the literature.<sup>1</sup>

<sup>1</sup>H NMR (300 MHz, THF-*d*<sub>8</sub>): δ (ppm) = 8.05–7.31 (m, 8 H), 7.29–7.02 (m, 28 H), 7.02–6.71 (m, 34 H), 6.71–6.43 (m, 12 H), 3.47–3.32 (m, 8 H).

<sup>13</sup>C NMR (126 MHz, THF-*d*<sub>8</sub>): δ (ppm) = 142.00, 141.69, 132.76, 131.84, 131.49, 131.42, 130.98, 130.85, 128.69, 128.61, 121.22, 120.89, 78.90.

MALDI-TOF: *m/z* calcd. for C<sub>152</sub>H<sub>90</sub>: = 1914.70, found 1914.62 [M]<sup>+</sup>.

### *Pyr-G2-(PN)<sub>8</sub> uncharged 22*

First generation dendrimer core **9** (50 mg, 0.0261 mmol) and building block **7b** (242 mg, 0.418 mmol, 16 equiv) were dissolved in 1 mL *o*-xylene in a 10 mL Schlenk tube. After degassing, the reaction mixture was heated up to 160 °C for three days. Subsequently, dendrimer **22** was precipitated in methanol and purified by GPC in THF. Dendrimer **22** was obtained as a yellow solid (106 mg, 64%).

<sup>1</sup>H NMR (500 MHz, THF-*d*<sub>8</sub>): δ (ppm) = 8.67 (d, *J* = 12.4 Hz, 8H), 8.40 (s, 8H), 7.80–7.72 (m, 8H), 7.56–6.30 (m, 274H), 1.60 (h, *J* = 6.1, 5.7 Hz, 16H), 0.90 (t, *J* = 7.3 Hz, 24H).

<sup>13</sup>C NMR (126 MHz, THF-*d*<sub>8</sub>): δ (ppm) = 149.21, 148.92, 142.99, 142.59, 142.41, 142.35, 142.33, 142.17, 142.10, 141.75, 141.66, 141.39, 141.25, 141.17, 140.37, 140.27, 139.97, 139.81, 139.74, 139.42, 139.05, 138.89, 138.85, 138.77, 136.66, 136.63, 135.99, 135.71, 134.19, 134.16, 133.43, 133.09, 130.88, 129.61, 129.59, 128.58, 127.95, 127.26, 127.24, 126.68, 126.27, 125.99, 125.72, 124.10, 68.07, 67.92, 67.75, 67.57, 38.53, 25.94, 25.79, 25.64, 25.48, 14.15.

MALDI-TOF: *m/z* calcd. for C<sub>488</sub>H<sub>354</sub>N<sub>8</sub> 6324.79, found 6325.89 [M+H]<sup>+</sup>.

### *Pyr-G2-(PN)<sub>8</sub> 10d*

Dendrimer **22** (106 mg, 0.017 mmol) was dissolved in 1 mL methyl iodide in a 10 mL Schlenk tube followed by the addition of 1 mL methanol. After 24 h diethyl ether was added leading to precipitation of the product which was filtered off to obtain dendrimer **10d** as a yellow-brown solid (116 mg, 92 %).

<sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>): δ (ppm) = 9.22 (s, 8 H), 8.81 (s, 8 H), 8.68 (s, 8 H), 8.03 (s, 8 H), 7.63–6.38 (m, 266 H), 4.27 (s, 24 H), 1.60–1.44 (m, 16 H), 0.83 (t, *J* = 7.0 Hz, 24 H).

<sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>): δ (ppm) = 153.21, 143.25, 140.89, 140.25, 137.99, 136.30, 132.30, 131.63, 130.83, 129.95, 129.48, 128.87, 127.78, 127.57, 125.88, 125.53, 124.62, 124.23, 47.95, 40.02, 39.86, 39.69, 39.52, 39.35, 39.19, 39.02, 36.69, 23.99, 13.52.

ESI-MS: *m/z* calcd. for (C<sub>496</sub>H<sub>378</sub>N<sub>8</sub>)<sup>8+</sup>: 806.37, found 806.24; *m/z* calcd. for (C<sub>496</sub>H<sub>378</sub>N<sub>8</sub>I)<sup>7+</sup>: 939.56, found 939.55; *m/z* calcd. for (C<sub>496</sub>H<sub>378</sub>N<sub>8</sub>I<sub>2</sub>)<sup>6+</sup>: 1117.30, found 1117.29; *m/z* calcd. for (C<sub>496</sub>H<sub>378</sub>N<sub>8</sub>I<sub>3</sub>)<sup>5+</sup>: 1366.14, found 1366.15; *m/z* calcd. for (C<sub>496</sub>H<sub>378</sub>N<sub>8</sub>I<sub>4</sub>)<sup>4+</sup>: 1739.40, found 1739.39.

## 2 Protein corona on dendrimer-coated liposomes and polystyrene nanoparticles

### 2.1 Material and methods

**Liposome synthesis.** Based on previous reports,<sup>5</sup> amine functionalized liposomes were synthesized using 1,2-dioleoyl-*sn*-glycero-3-phosphoethanolamine (DOPE), L- $\alpha$ -phosphatidylcholine (egg PC) and cholesterol (Chol) with a molar ratio of egg PC:DOPE:Chol = 1:1:1 by film hydration followed by extrusion. Cholesterol and the lipids were dissolved in chloroform at a concentration of 10 mg mL<sup>-1</sup>. All components (Egg PC = 835  $\mu$ L, DOPE = 767  $\mu$ L and Cholesterol = 398  $\mu$ L) were mixed with 2 mL of chloroform containing 1 vol% EtOH in a 50 mL round-bottomed flask. The mixture was dried using a rotary evaporator at room temperature (30 min, 450 mbar) and further at 42°C (30 min, 3 mbar). To remove organic solvent residuals the flask was placed into a vacuum oven for 1 hour. (Diameter  $\varnothing$ : 242  $\pm$  6 nm,  $\zeta$  – Potential: - 52  $\pm$  6 mV)

**Polystyrene nanoparticle synthesis.** According to previous reports,<sup>6</sup> amine functionalised polystyrene nanoparticles were prepared via direct miniemulsion. For colloidal stability, the ionic surfactant cetyl trimethyl ammonium chloride was used. To generate amino groups, 2-aminoethyl methacrylate hydrochloride (2 wt% to styrene) was copolymerized with styrene. Excessive surfactant was removed via dialysis. A detailed protocol is given in previous reports.<sup>7</sup> (Diameter  $\varnothing$ : 98  $\pm$  10 nm,  $\zeta$  – Potential: + 34  $\pm$  12 mV)

**Coating of nanoparticles and liposomes with dendrimers.** All dendrimers were dissolved in DMSO at a concentration of 10 mg mL<sup>-1</sup>. PS-NH<sub>2</sub> nanoparticles (10 mg mL<sup>-1</sup>, 100  $\mu$ L) and liposomes-NH<sub>2</sub> (3 mg mL<sup>-1</sup>, 333  $\mu$ L) were mixed with the dendrimers (10 mg mL<sup>-1</sup>, 100  $\mu$ L) and incubated at room temperature for 1 h. To remove unbound dendrimers, the liposome dispersion was diluted with 1 mL of PBS and the PS-nanoparticle dispersion with 1 mL of water. Afterwards, the mixture was centrifuged (20 000 g, 15 min, 4°C). Finally, polystyrene nanoparticles were resuspended in 100  $\mu$ L of water and liposomes were resuspended in 100  $\mu$ L of PBS.

**Zeta Potential.** Measurements were performed using a Nano Z Zetasizer (Malvern Instruments GmbH, Herrenberg, Germany). Uncoated and coated liposomes/PS-

nanoparticles (20  $\mu\text{L}$ ) were diluted with 1 mL of a 1 mM KCl solution and measured instantly at 25 °C. Each measurement was repeated in technical triplicate.

**Human serum/plasma.** Human blood serum and plasma was obtained from healthy donors from the Transfusion Center of the University Clinic of Mainz, Germany. For plasma preparation, citrate was used as an anticoagulant. Six serum batches and ten plasma batches were pooled together and stored at  $-20\text{ }^{\circ}\text{C}$ . All experiments were performed in compliance with the relevant laws and institutional guidelines. The institutional ethics committee had approved the study (Landesärztekammer Rheinland-Pfalz, 837.439.12 (8540-F)). Written informed consent was obtained for any experimentation with human subjects.

**Protein corona analysis.** Uncoated and coated liposomes or polystyrene nanoparticles (10 mg  $\text{mL}^{-1}$ , 100  $\mu\text{L}$ ) were incubated with 1 mL of human serum or human plasma at 37 °C for 1 h. The protein corona coated liposomes and nanoparticles were centrifuged (20 000 g, 15 min, 4 °C) and washed with 1 mL PBS solution to remove unbound and loosely bound proteins. Overall, this step was repeated three times. The adsorbed corona proteins were detached from the surface by resuspending the liposome/nanoparticle pellet in 100  $\mu\text{L}$  of an aqueous solution containing 2% SDS supplemented with 62.5 mM Tris-HCL (Sigma Aldrich, Germany). Afterwards, the dispersion was incubated for 5 min at 95 °C and centrifuged as described above. The supernatant contained the detached corona proteins.

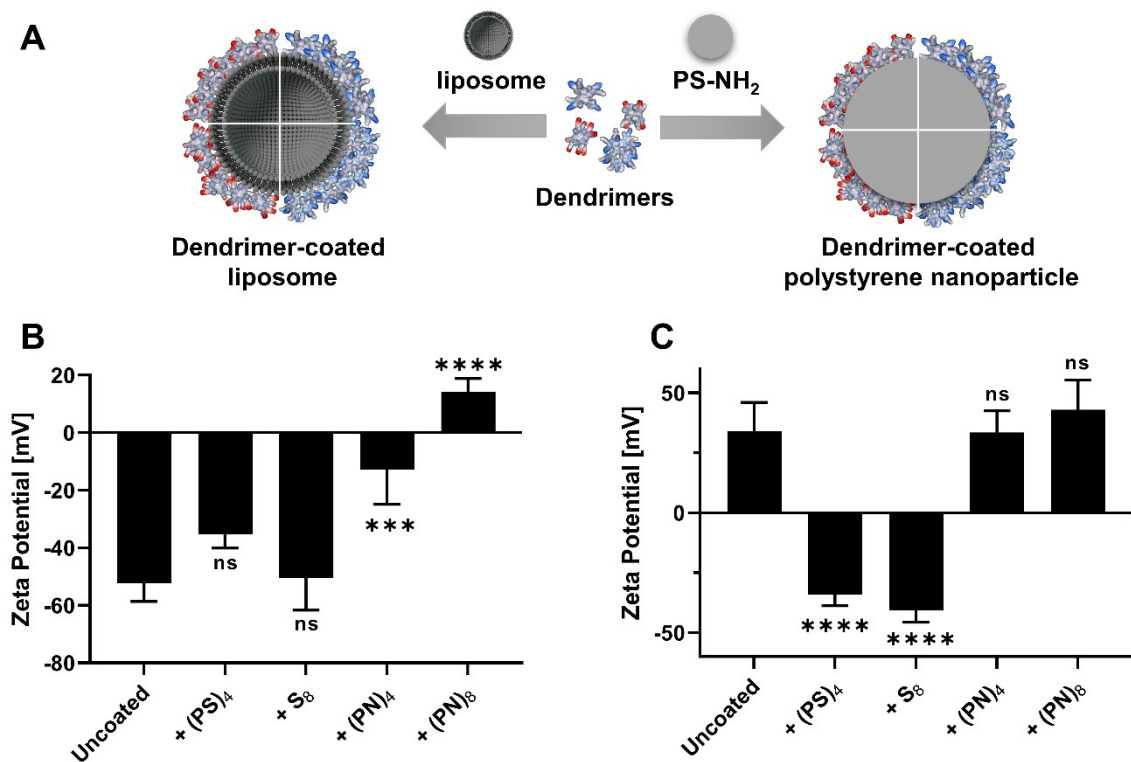
**Protein Assay.** The protein concentration was determined by Pierce 660 nm Protein Assay (Thermo Fisher, Germany) according to manufacturer's instructions.

**In solution digestion.** Prior to LC-MS analysis, proteins were digested according to former protocols.<sup>7,8</sup> Briefly, SDS was removed via Pierce detergent removal columns (Thermo Fisher, Germany) according to the manufacture's instruction. Afterwards, proteins were precipitated using ProteoExtract protein precipitation kit (Merck Millipore, Germany) overnight and the protein pellet was isolated via centrifugation (14 000 g, 10 min). Precipitated proteins were resuspended in RapiGest SF (Waters, USA) dissolved in 50 mM ammonium bicarbonate (Sigma Aldrich, Germany) and incubated at 80 °C for 15 min. Further, proteins were reduced with dithithreitol at 56 °C for 45 min (5 mM, Sigma Aldrich, Germany) and alkylated with idoacetoamide at room temperature in the dark for 1 h (15 mM, Sigma Aldrich, Germany). Digestion was carried out using a protein:trypsin ratio of 50:1 for 14–18 h at 37 °C and stopped by adding 2  $\mu\text{L}$  hydrochloric acid (Sigma Aldrich, Germany). To remove degradation products of RapiGest SF, the peptide sample was centrifuged (14 000 g, 15 min, 4 °C).

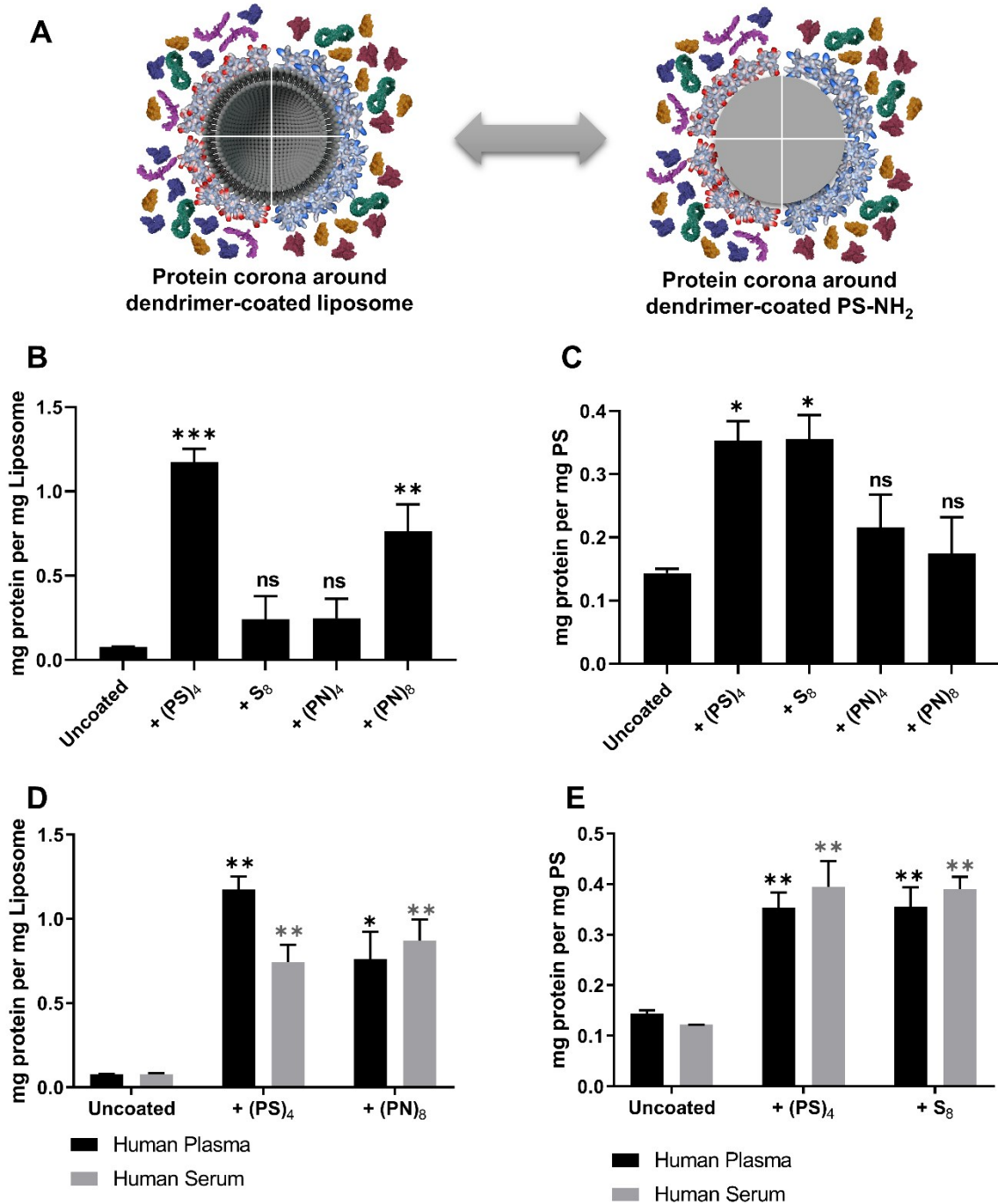
**Liquid chromatography-high resolution mass spectrometry (LC-MS).** The peptide samples were spiked with 50 fmol of Hi3 EColi Standard (Waters, USA) for absolute protein quantification. Peptides were separated using a nanoACQUITY UPLC system equipped with a C18 nanoACQUITY Trap Column (5  $\mu\text{m}$ , 180  $\mu\text{m}$  x 20 mm, Waters, USA) and C18 analytic reversed phase column (1.7  $\mu\text{m}$ , 75  $\mu\text{m}$  x 150 mm, Waters, USA). The UPLC system was coupled to a Synapt G2-Si mass spectrometer (Waters, USA) and electrospray ionization (ESI) was carried in positive ion with a nanoLockSpray source. The high-resolution mass spectrometer was operated in resolution mode performing data-independent experiments ( $\text{MS}^E$ ). Data was processed with MassLynx 4.1 and all proteins were identified with Progenesis Q1. A reviewed human data base downloaded from Uniprot and the peptide sequence of Hi3 Ecoli standard (Chaperone protein CLpB, Water, USA) was added for absolute protein quantification. Based on the TOP3/Hi3 approach<sup>9</sup> all proteins were quantified and the absolute amount in fmol is calculated.

**Statistical Analysis.** GraphPad Prism 8.4.2 Software was used for statistical analysis applying a one-way ANOVA followed by Tukey's post-hoc multiple comparisons test for all protein corona results in the main text and Figure S11 in order to compare the difference between both uncoated liposome/ PS nanoparticle and lipo-dendrimers/PS-dendrimers as well as between the different dendrimer-coatings. For zeta-potential and Pierce Assay and concentration dependency measurements a one-way ANOVA followed by Dunnett's post-hoc multiple comparisons test was used to compare the coatings with the uncoated liposomes/PS nanoparticles. Significance was considered at  $p$ -values such as not significant (ns) for  $p > 0.05$ , and significant \* for  $p \leq 0.05$ , \*\* for  $p \leq 0.01$ , \*\*\* for  $p \leq 0.001$ , \*\*\*\* for  $p \leq 0.0001$ ).

## 2.2 Supplementary figures: Coating of liposomes and polystyrene nanoparticles and their respective protein corona

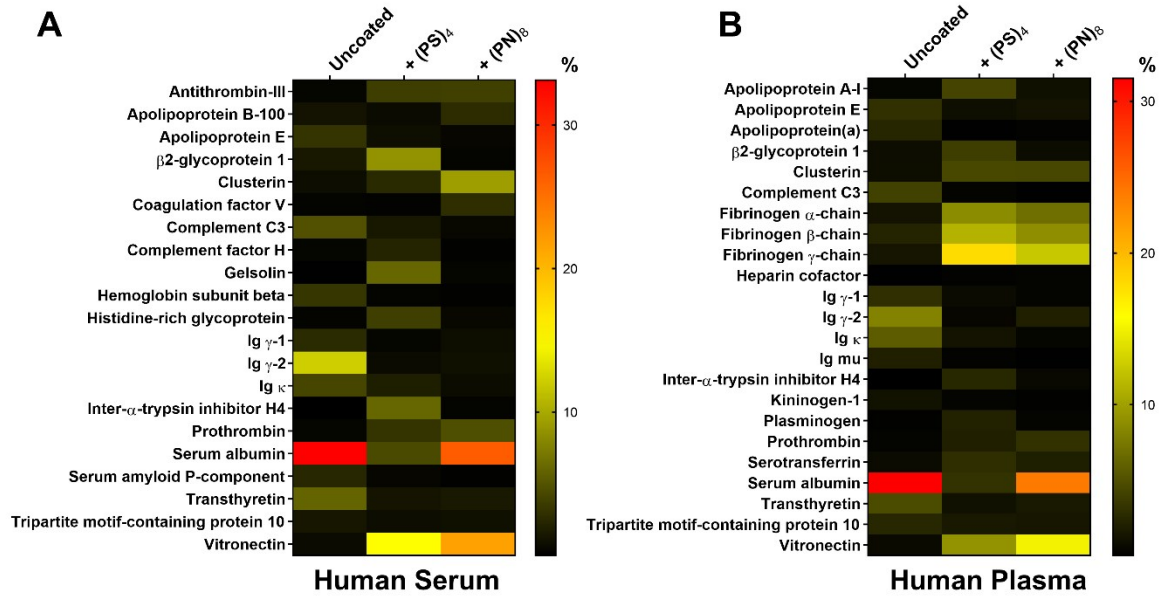


**Figure S6.** Verification of dendrimer-coating on liposomes and polystyrene nanoparticles (PS-NH<sub>2</sub>). **(A)** Dendrimers were mixed with liposomes and  $\zeta$ -potentials of **(B)** lipo-dendrimers and **(C)** PS-dendrimers were measured. (PN)<sub>4</sub> and S<sub>8</sub> did not show a clear change in  $\zeta$ -potential values when mixed with liposomes indicating an insufficient coating. The same was observed for (PN)<sub>4</sub> and (PN)<sub>8</sub> when mixed with PS nanoparticles ( $n = \text{minimum } 2$ ; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; ns for  $p > 0.05$ , \*\*\* for  $p \leq 0.001$ , \*\*\*\* for  $p \leq 0.0001$ ).

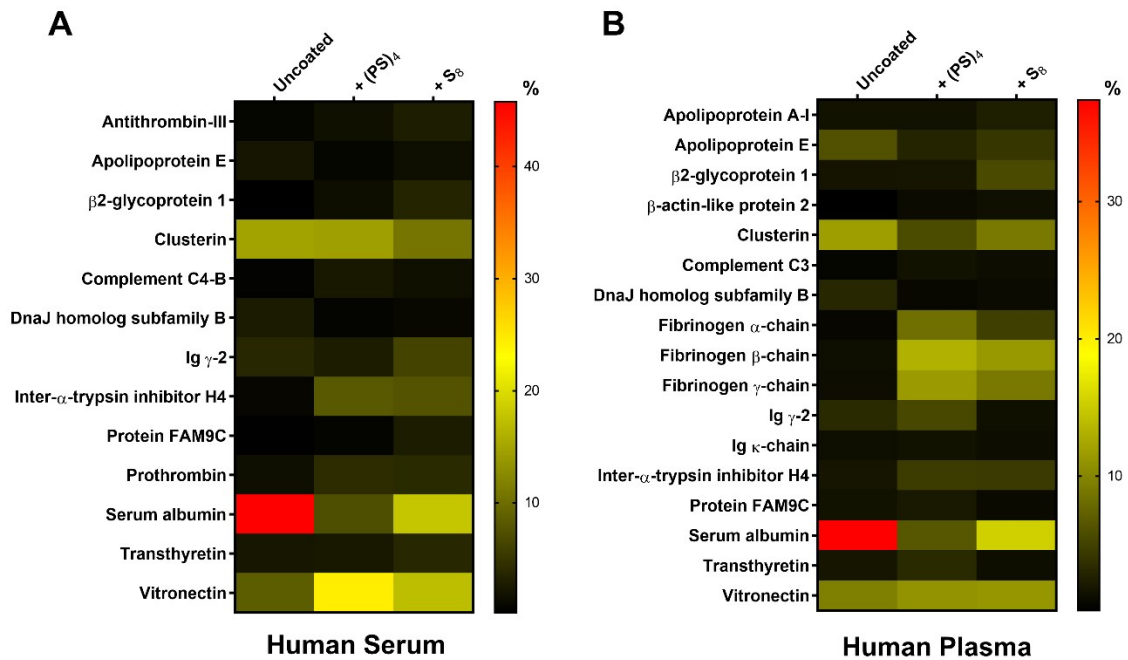


**Figure S7.** Verification of protein adsorption on lipo-dendrimers and PS-dendrimers: **(A)** Comparison of protein adsorption on lipo-dendrimers and PS-dendrimers. Quantification of protein adsorption by Pierce Assay for **(B)** lipo-dendrimers and **(C)** PS-dendrimers in blood plasma. Liposomes and PS nanoparticles with insufficient dendrimer binding did not show significant differences in protein quantities. Protein quantification by Pierce Assay for efficient dendrimer-coatings in human plasma and serum for **(D)** lipo-(PS)<sub>4</sub> and lipo-(PN)<sub>8</sub> as well as **(E)** PS-(PS)<sub>4</sub> and PS-S<sub>8</sub> (n = minimum 2; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; ns for  $p > 0.05$ , \* for  $p \leq 0.05$ , \*\* for  $p \leq 0.01$ , \*\*\* for  $p \leq 0.001$ ).

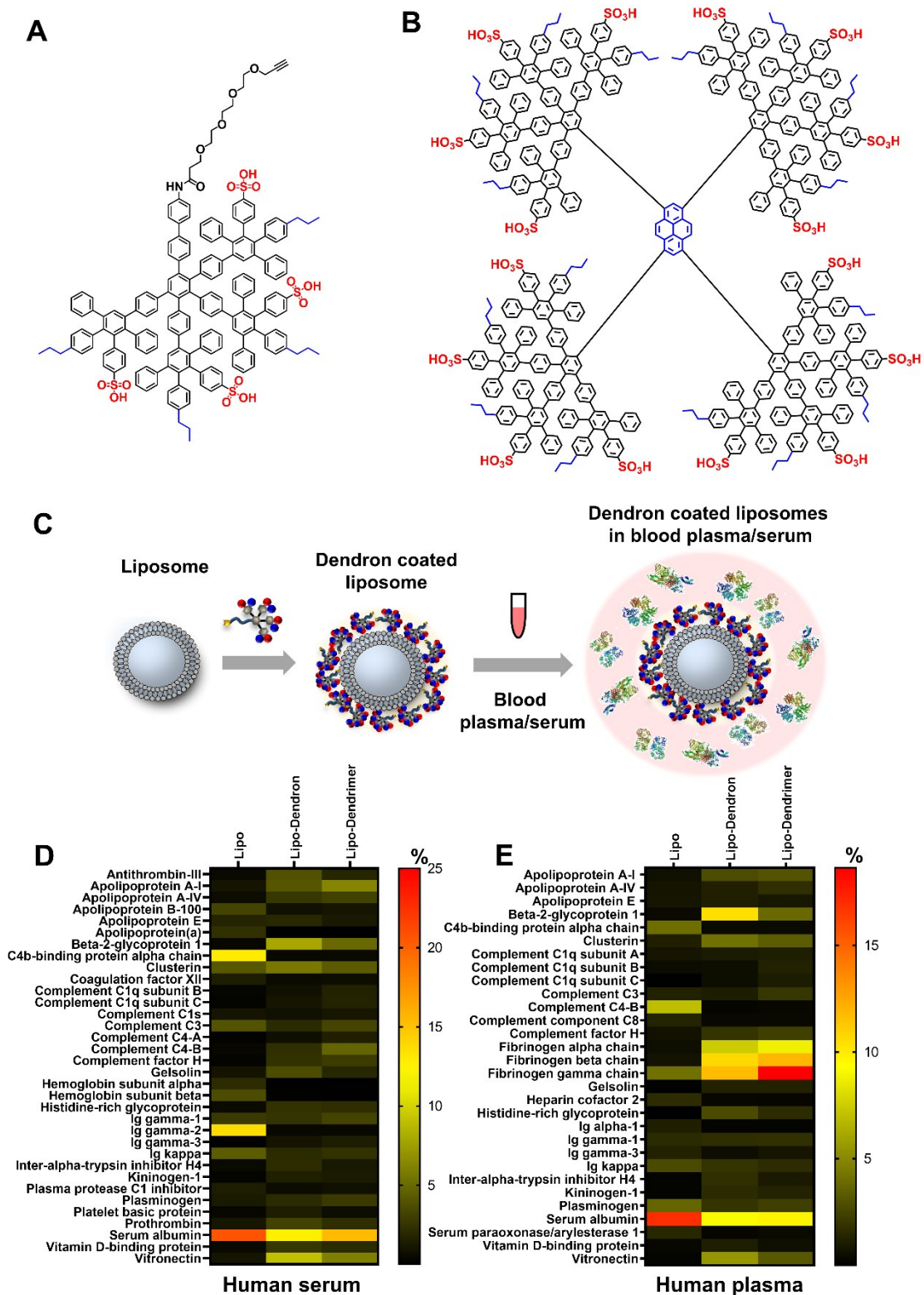




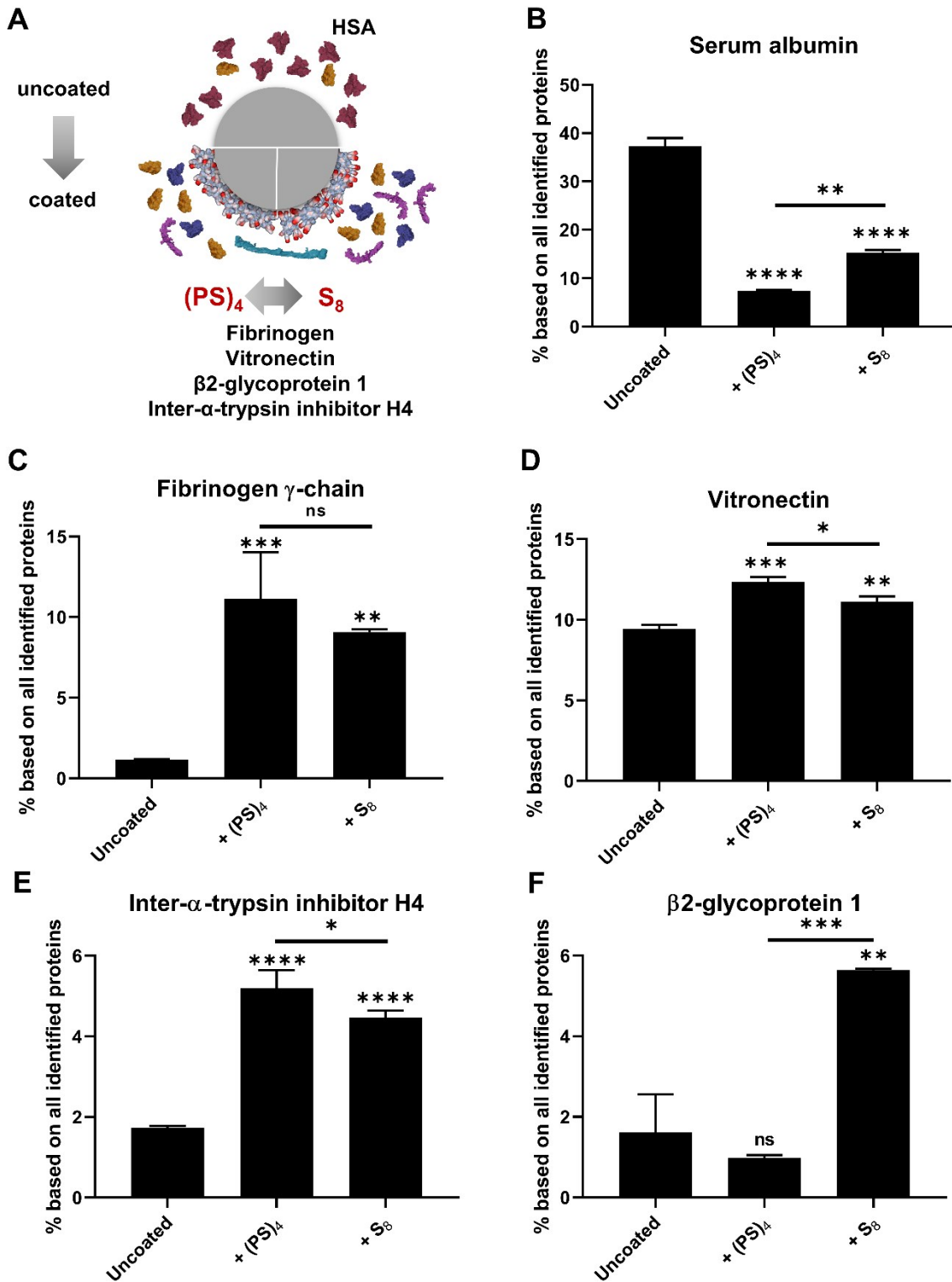
**Figure S8.** Heatmaps of adsorbed proteins to dendrimer-coated liposomes lipo-(PS)<sub>4</sub> and lipo-(PN)<sub>8</sub> in (A) blood serum and (B) blood plasma. A list of all identified proteins is provided in Table S1.



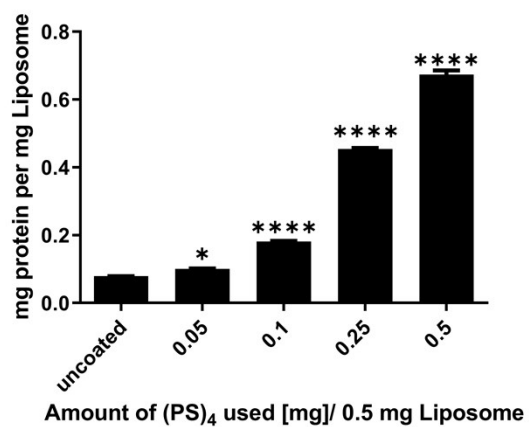
**Figure S9.** Heatmaps of adsorbed proteins to dendrimer-coated polystyrene nanoparticles PS-(PS)<sub>4</sub> and PS-S<sub>8</sub> in (A) blood serum and (B) blood plasma. A list of all identified proteins is provided in Table S2.



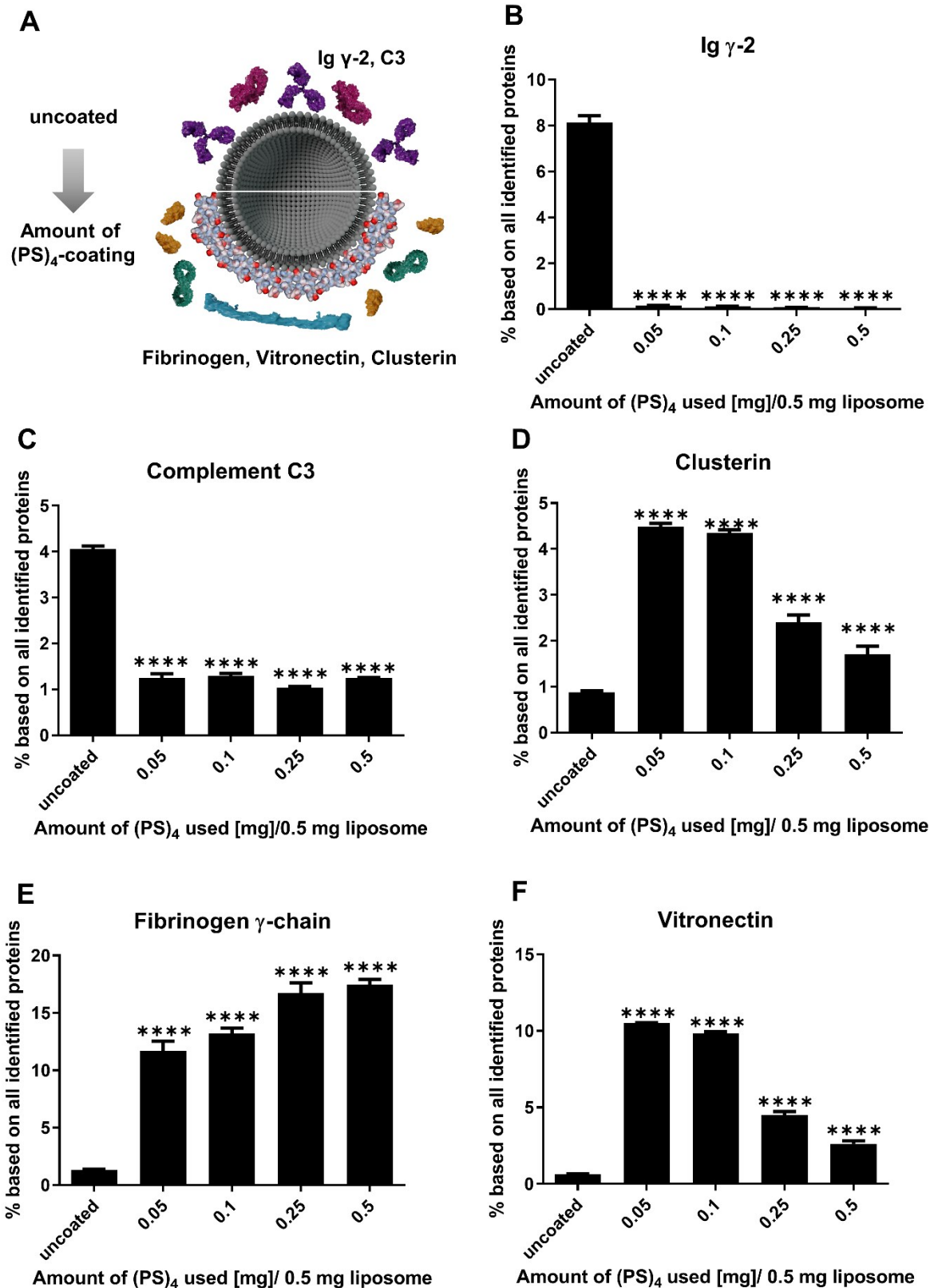
**Figure S10.** Protein corona on dendron- and dendrimer-coated liposomes. **(A)** Amphiphilic polyphenylene dendron<sup>3</sup> and **(B)** amphiphilic polyphenylene dendrimer<sup>1</sup> with high density of surface pattern. **(C)** Coating of liposomes and addition of human serum and plasma. Heat map of adsorbed proteins on lipo-dendron and lipo-dendrimer in **(D)** human serum and **(E)** human plasma. **(C-E)**. Reproduced from Wagner *et al.*<sup>3</sup> (<https://doi.org/10.1002/anie.201913708>). Licensed under a Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).



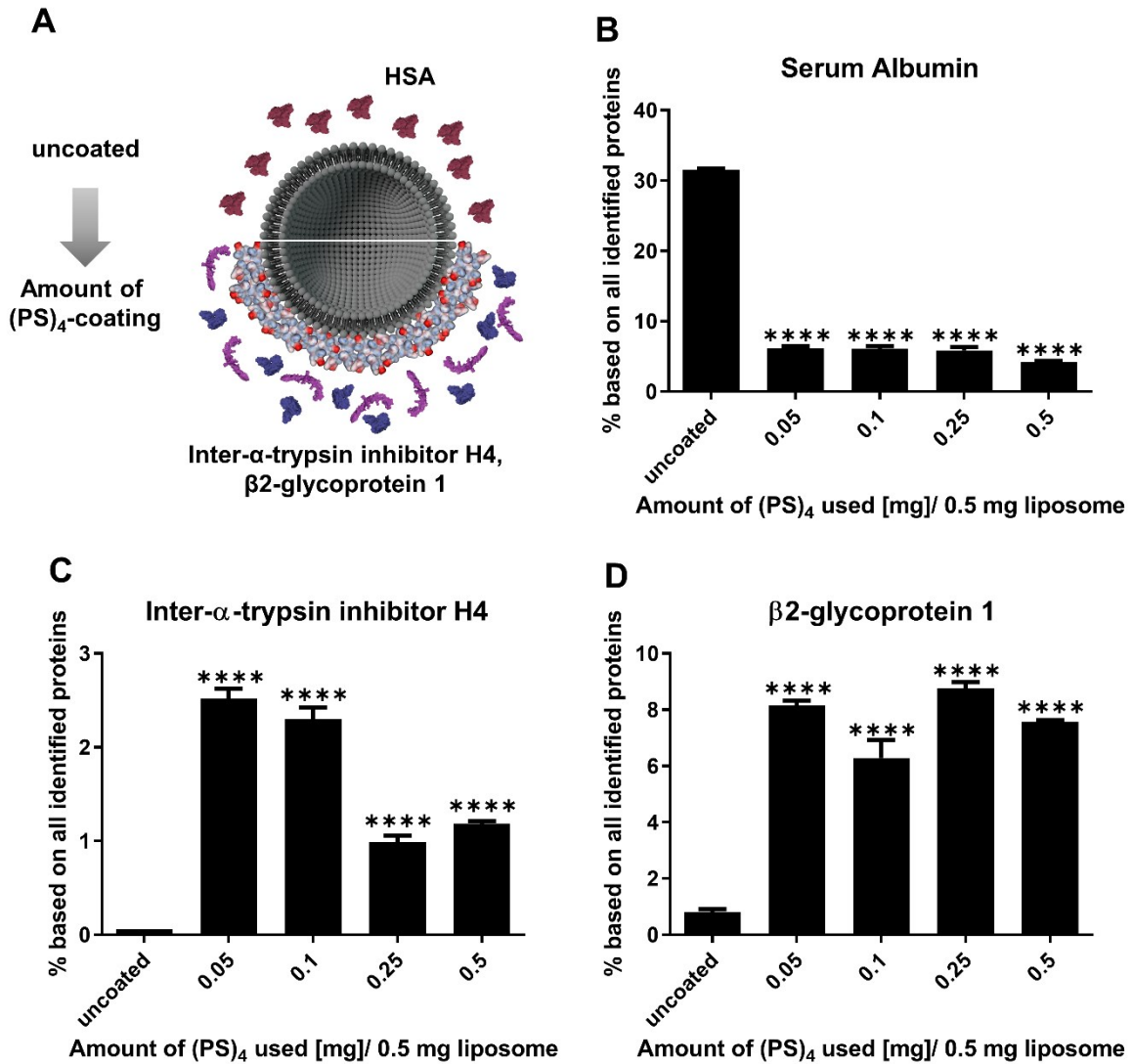
**Figure S11.** Comparison of adsorbed proteins between uncoated polystyrene nanoparticles (PS-NH<sub>2</sub>) and PS-dendrimers (PS-(PS)<sub>4</sub> and PS-S<sub>8</sub>). **(A)** Coating of PS-NH<sub>2</sub> lead to a different protein corona: Reduction of **(B)** serum albumin adsorption and enhancement of **(C)** fibrinogen, **(D)** vitronectin, **(E)** inter-α-trypsin inhibitor H4 and **(F)** β-2 glycoprotein 1 (n = minimum 2; one-way ANOVA, Tuckey's post-hoc multi-comparisons test; ns for  $p > 0.05$ , \* for  $p \leq 0.05$ , \*\* for  $p \leq 0.01$ , \*\*\* for  $p \leq 0.001$ , \*\*\*\* for  $p \leq 0.0001$ ).



**Figure S12.** Quantification of protein adsorption in human plasma by Pierce Assay for different (PS)<sub>4</sub> concentrations used for the coating to liposomes. The amount of proteins adsorbed increases with higher dendrimer concentrations. (n = 2; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; \* for  $p \leq 0.05$ , \*\*\*\* for  $p \leq 0.0001$ ).



**Figure S13.** Concentration dependency of dendrimer-coatings on protein adsorption in human plasma. **(A)** Comparison between uncoated liposomes and lipo-(PS)<sub>4</sub> at different dendrimer concentrations used for the liposome coating. The adsorption of the opsonins **(B)** Ig  $\gamma$ -2 and **(C)** complement C3 is significantly reduced even at low dendrimer concentrations, whereas the adsorption of **(D)** Clusterin, **(E)** Fibrinogen and **(F)** Vitronectin is significantly enhanced ( $n = 3$ ; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; \* for  $p \leq 0.05$ , \*\*\*\* for  $p \leq 0.0001$ ). A list of all identified proteins is provided in Table S3.



**Figure S14.** Concentration dependency of dendrimer-coatings specific for (PS)<sub>4</sub> on protein adsorption in human plasma. **(A)** Comparison between uncoated liposomes and lipo-(PS)<sub>4</sub> at different dendrimer concentrations used for the liposome coating. The adsorption of **(B)** serum albumin is significantly reduced even at low dendrimer concentrations, whereas the adsorption of **(C)** Inter- $\alpha$ -trypsin inhibitor H4 and **(D)**  $\beta$ 2-glycoprotein 1 is significantly enhanced ( $n = 3$ ; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; \* for  $p \leq 0.05$ , \*\*\*\* for  $p \leq 0.0001$ ). A list of all identified proteins is provided in Table S3.









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