

Supporting Information

Amphiphilic Dendrimers Control Protein Binding and Corona Formation on Liposome Nanocarriers

Jessica Wagner,^{†ab} Marcel Dillenburger,^{†a} Johanna Simon,^{ac} Jennifer Oberländer,^{ac} Katharina Landfester,^a Volker Mailänder,^{ac} David Y.W. Ng,^a Klaus Müllen,^{a*} Tanja Weil ^{a*}

^a Max Planck Institute for Polymer Research, Ackermannweg 10, 55128 Mainz (Germany)

^b Graduate School Materials Science in Mainz, Staudingerweg 9, 55128 Mainz (Germany)

^c Department of Dermatology, University Medical Center of the Johannes Gutenberg-University Mainz, Langenbeckstr. 1, 55131 Mainz (Germany)

† J.W. and M.D. contributed equally.

Table of Contents

1 Synthesis of dendrimers with different surface patterns	3
1.1 Materials and Instruments	3
1.2 Synthesis of a first-generation negatively charged amphiphilic polyphenylene dendrimer – (PS) ₄	5
1.3 Synthesis of a first-generation negatively charged polyphenylene dendrimer – S ₈	8
1.4 Synthesis of a first-generation positively charged amphiphilic polyphenylene dendrimer –(PN) ₄	12
1.5 Synthesis of a second-generation positively charged amphiphilic polyphenylene dendrimer – (PN) ₈	17
2 Protein corona on dendrimer-coated liposomes and polystyrene nanoparticles	20
2.1 Material and methods	20
2.2 Supplementary figures: Coating of liposomes and polystyrene nanoparticles and their respective protein corona	23
References	34

1 Synthesis of dendrimers with different surface patterns

1.1 Materials and Instruments

Chemicals and Solvents. All chemicals were obtained from commercial suppliers such as Sigma Aldrich, TCI, Acros Organics, Fisher Scientific, etc., and were utilized without further purification. The solvents (HPLC grade) were purchased from Fisher Scientific or Acros Organics. Chloroform (Rotisolv®, HPLC grade) for the *recycling*-gel permeation chromatography (GPC) system was purchased from Carl Roth. The deuterated solvents were purchased from Sigma Aldrich. Argon (Westfalen AG) was used for reactions under inert gas.

Silica Gel Chromatography. For thin-layer chromatography (TLC) Alugram Sil G/UV254 plates from Macherey-Nagel were used and the detection of substances was performed under UV light at 254 nm or 366 nm. Macherey-Nagel silica gel with particle size of 0.04–0.063 mm or 0.063–0.2 mm was used for preparative silica gel chromatography.

Gel Permeation Chromatography (GPC). Dendrimers with peripheral neopentyl-protected sulfonic acid groups were purified on a *recycling*-GPC purification system from Shimadzu. Dendrimers with deprotected peripheral sulfonic acid groups were purified by size exclusion chromatography using Sephadex® LH-20 in DMF.

Nuclear Magnetic Resonance Spectroscopy (NMR). All ^1H NMR and ^{13}C NMR spectra were recorded on a Bruker Avance III 300 MHz, Avance III 500 MHz, Avance III 700 MHz or Bruker Avance III 850 MHz spectrometer in deuterated solvents such as CD_2Cl_2 , $\text{THF}-d_8$ and $\text{DMSO}-d_6$ at 298 K. ^{13}C NMR were recorded in j-modulated spin-echo (JMOD) mode. Spectra were analyzed by applying the software MestReNova.

Matrix-assisted Laser Desorption/Ionization–Time of Flight (MALDI-TOF) Mass Spectrometry. MALDI-TOF mass spectra were measured on a Waters MALDI Synapt G2-SI or Bruker rapifleX MALDI-TOF/TOF. Precursors and uncharged dendrimers were dissolved in THF and measured by applying *trans*-2-[3-(4-*tert*-butylphenyl)-2-methyl-2-propenylidene]-malononitrile (DCTB) or dithranol as matrix. Deprotected sulfonic acid based dendrimers were solvated in DMF and diluted in a saturated α -cyano-4-hydroxycinnamic acid (CHCA) solution in water:acetonitrile (1:1) + 0.1% TFA. All dendrimers with peripheral pyridine groups were measured in tetrahydrofuran (THF) with tetracyanoquinodimethane (TCNQ) as matrix. Processing of data was performed in mMass.

Electrospray Ionization (ESI) Spectrometry. ESI spectra were measured on a Waters Synapt G2-SI. The samples were dissolved in methanol.

Atmospheric Pressure Chemical Ionization Mass Spectrometry (APCI-MS). APCI mass spectra of building blocks and intermediates were measured on an Advion expression-L Compact Mass Spectrometer (CMS) (Advion Inc. 61 Brown Rd, Suite 100, Ithaca, NY 14850, USA) by either applying the sample (solid) as an atmospheric solid analysis probe (ASAP) or measuring directly from TLC plates by an automated TLC plate reader (Plate express).

Field Desorption Mass Spectrometry (FD-MS). FD mass spectra of intermediates were measured on a VG Instruments ZAB 2-SE-FPD using an 8 kV accelerating voltage.

Nomenclature for Polyphenylene Dendrimers.

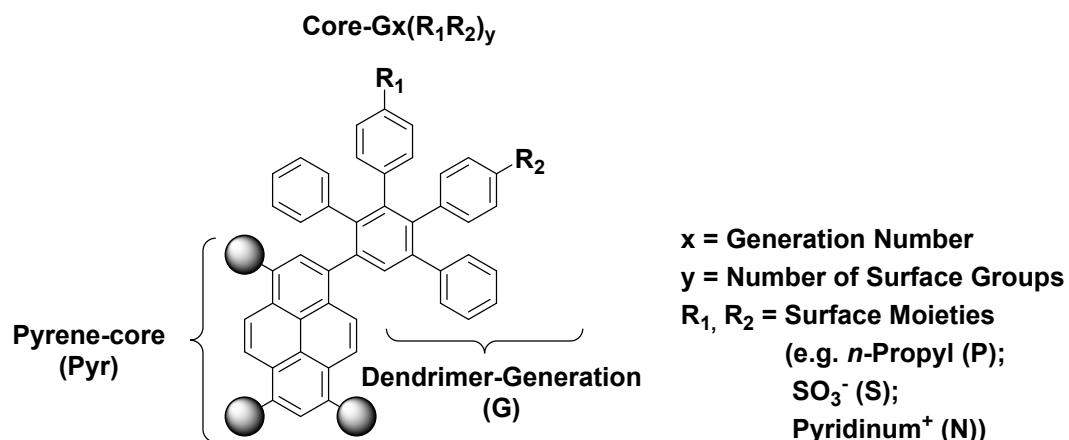


Figure S1. Systematic designation of polyphenylene dendrimers (PPDs)

1.2 Synthesis of a first-generation negatively charged amphiphilic polyphenylene dendrimer – (PS)₄

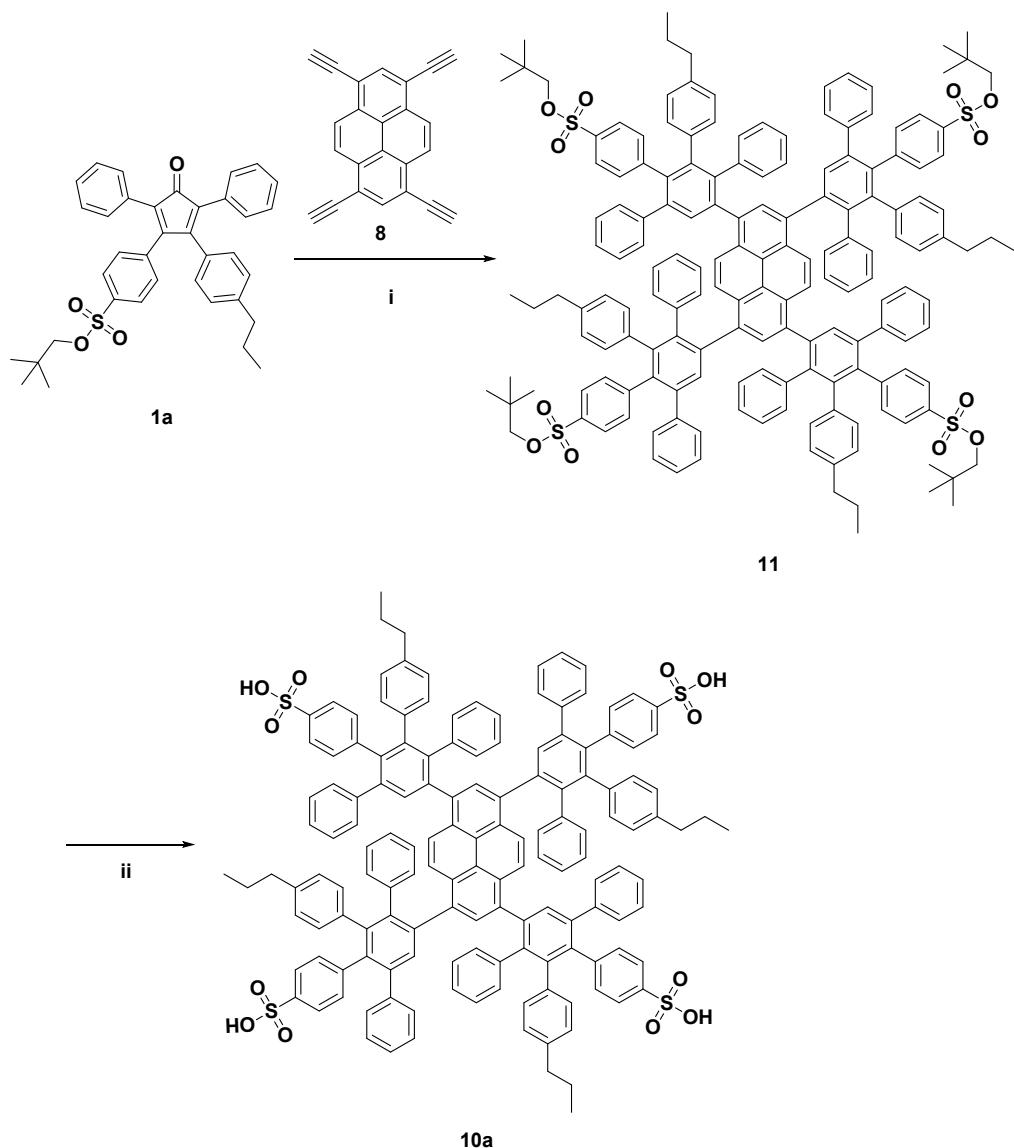


Figure S2. Synthesis of first-generation amphiphilic dendrimer **10a** with alternating sulfonic acid and *n*-propyl groups according to Stangenberg *et al.*¹ i) *o*-xylene, 145 °C, 48 h, 45% yield; ii) DMF, 180 °C, 48 h, 85% yield.

Neopentyl 4-(3-oxo-2,4-diphenyl-5-(4-propylphenyl)cyclopenta-1,4-dien-1-yl)benzenesulfonate (1a)

Surface building block **1a** was synthesized according to the literature in a four-step synthesis.²

³ All spectral data were in agreement with the literature.^{2,3}

¹H NMR (300 MHz, CD₂Cl₂): δ(ppm) = 7.70 (d, J = 8.5 Hz, 2H), 7.31–7.22 (m, 8H), 7.20–7.12 (m, 4H), 7.01 (d, J = 8.3 Hz, 2H), 6.83 (d, J = 8.2 Hz, 2H), 3.64 (s, 2H), 2.54 (t, J = 7.8, 7.2 Hz, 2H), 1.61 (dt, J = 13.7, 7.4 Hz, 2H), 0.92 (t, J = 7.3 Hz, 3H), 0.87 (s, 9H).

¹³C NMR (75 MHz, CD₂Cl₂): δ(ppm) = 200.22, 155.08, 152.76, 144.50, 139.78, 135.83, 131.36, 130.72, 130.66, 130.32, 129.56, 128.84, 128.74, 128.56, 128.09, 127.61, 125.62, 80.56, 38.26, 32.05, 26.27, 24.77, 14.02.

FD-MS: *m/z* calcd. for C₃₇H₃₆O₄S: 576.7, found 576.8 [M]⁺.

1,3,6,8-Tetraethynylpyrene (8)

Pyrene core **8** was synthesized in three steps according to the literature.⁴

Pyr-G1-(P_{Spn})₄ 11

The synthesis of neopentyl-protected dendrimer **11** was modified from the previously reported method.¹ 1,3,6,8-Tetraethynylpyrene (**8**) (10 mg, 33.5 μmol) and building block **1a** (116 mg, 201 μmol, 6 equiv) were dissolved in 3 mL *o*-xylene. The mixture was degassed with argon and stirred at 145 °C for 48 h in a sealed microwave tube. The product was precipitated in methanol, filtered off and purified by silica gel chromatography using a mixture of cyclohexane and THF (3:1). The product was further purified by *recycling*-GPC in chloroform to obtain dendrimer **11** as a yellow solid (38 mg, 45%).

¹H NMR (700 MHz, CD₂Cl₂): δ (ppm) = 7.98–7.73 (m, 5H), 7.60–7.31 (m, 11H), 7.26–6.23 (m, 66H), 3.54–3.30 (m, 8H), 2.50–2.27 (m, 8H), 1.52–1.38 (m, 8H), 0.88–0.66 (m, 48H).

¹³C NMR (176 MHz, CD₂Cl₂): δ(ppm) = 147.44, 147.16, 142.17, 141.36, 141.06, 140.65, 140.53, 140.10, 139.77, 138.10, 137.26, 137.01, 136.65, 136.32, 132.99, 132.81, 131.67, 130.37, 130.28, 128.20, 128.15, 127.92, 127.79, 127.45, 126.99, 126.72, 80.04, 37.68, 26.10, 24.67, 13.49.

MALDI-TOF: *m/z* calcd. for C₁₆₈H₁₅₄O₁₂S₄ 2491.03, found 2492.98 [M+H]⁺.

Pyr-G1-(PS)₄ 10a

Negatively charged amphiphilic dendrimer **10a** was synthesized according to the literature¹ with a modified purification protocol. Briefly, neopentyl-protected dendrimer **11** (12.0 mg, 4.81 µmol) was dissolved in 2 mL dry DMF, the resulting solution was degassed with argon and stirred at 180 °C for 48 h in a sealed microwave tube. Subsequently, DMF was removed *in vacuo*. The residue was dissolved in methanol and precipitated in diethyl ether. The resulting product was further purified by gel permeation chromatography (GPC) applying Sephadex LH-20 in DMF to obtain Pyr-G1-(PS)₄ **10a** as a light yellow solid (9 mg, 85%).

¹H NMR (700 MHz, DMSO): δ (ppm) = 8.02–6.20 (m, 82H), 2.44–2.28 (m, 8H), 1.52–1.31 (m, 8H), 0.79–0.58 (m, 12H).

¹³C NMR (176 MHz, DMSO): δ (ppm) = 145.51, 145.41, 141.11, 141.02, 139.64, 139.27, 137.01, 130.93, 130.36, 129.61, 129.54, 127.74, 127.70, 127.46, 127.43, 127.12, 126.72, 125.09, 124.18, 123.89, 36.54, 23.50, 13.12.

MALDI-TOF: *m/z* calcd. for C₁₄₈H₁₁₄O₁₂S₄ 2210.72, found 2210.73 [M]^{•+}.

1.3 Synthesis of a first-generation negatively charged polyphenylene dendrimer – **S₈**

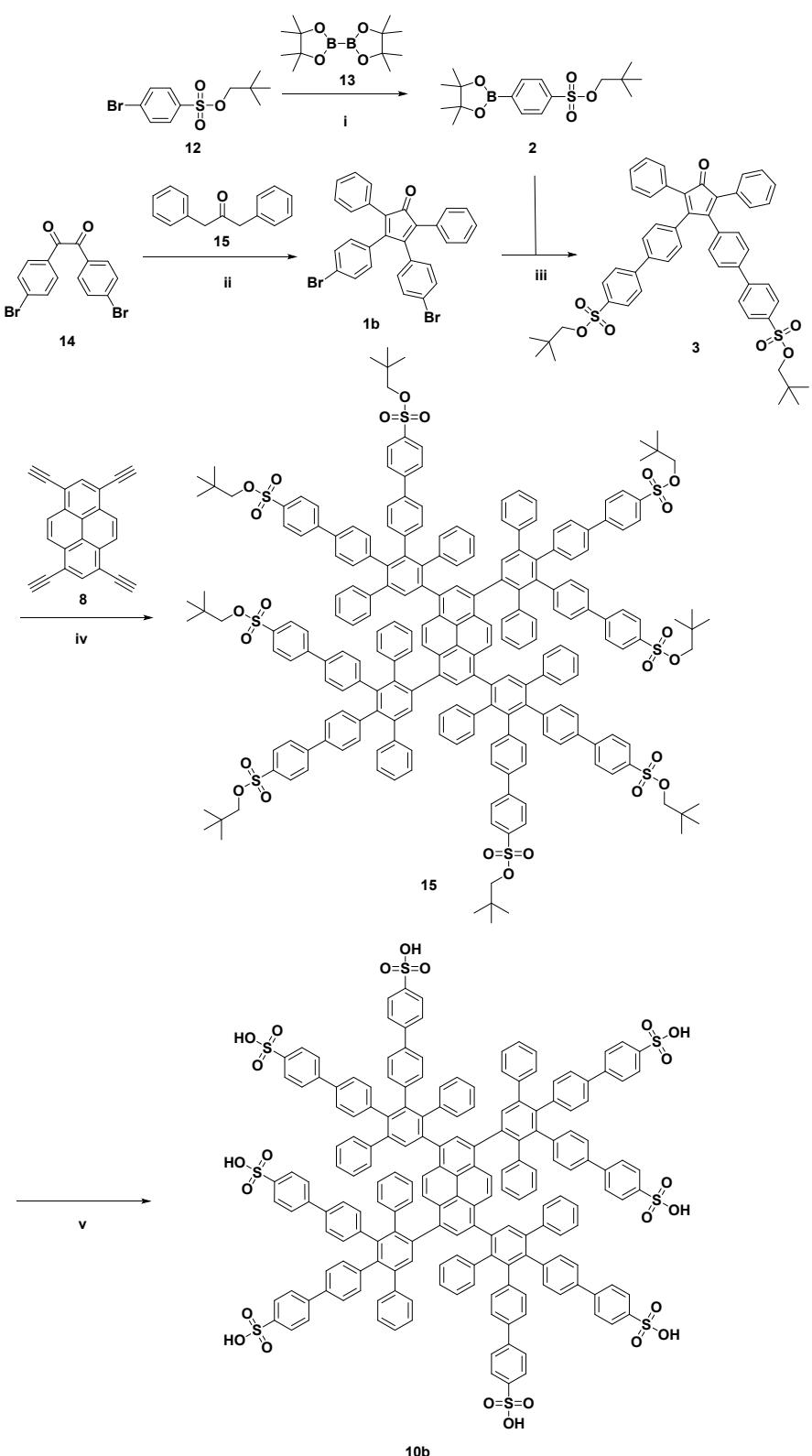


Figure S3. Synthesis of first-generation negatively charged dendrimer **10b**. i) KOAc, Pd(dppf)Cl₂, 1,4-dioxane, 90 °C, 6 h, 99% yield; ii) TBAH, ethanol, 85 °C, 0.5 h, 85% yield; iii) Pd(PPh₃)₄, K₂CO₃ (aq), 1,4-dioxane, 90 °C, 15 h, 34% yield; iv) o-xylene, 145 °C, 36 h, 27% yield; v) DMF, 180 °C, 48 h, 88% yield.

Neopentyl 4-bromobenzenesulfonate (12**)**

Compound **17** was synthesized according to the literature.^{2, 3} All spectral data were in agreement with the literature.²

¹H NMR (300 MHz, CD₂Cl₂): δ(ppm) = 7.79–7.70 (m, 4H), 3.67 (s, 2H), 0.89 (s, 9H).

¹³C NMR (75 MHz, CD₂Cl₂): δ(ppm) = 135.64, 133.12, 129.95, 129.27, 80.63, 32.08, 26.22.

FD-MS: *m/z* calcd. for C₁₁H₁₅BrO₃S: 307.2, found 308.7 [M+H]⁺.

Neopentyl 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzenesulfonate (2**)**

In a dry Schlenk flask neopentyl 4-bromobenzenesulfonate (**12**) (1.00 g, 3.26 mmol), bis(pinacolato)diboron (**13**) (1.24 g, 4.88 mmol, 1.5 equiv) and potassium acetate (703 mg, 7.16 mmol, 2.2 equiv) were dissolved in 15 mL dry 1,4-dioxane. After degassing with argon [1,1'-bis(diphenylphosphino)ferrocene]dichloropalladium(II) (Pd(dppf)Cl₂) (53.2 mg, 65.1 μmol, 0.02 equiv) was added under argon atmosphere. The reaction mixture was stirred at 90 °C for 6 h under argon atmosphere. Subsequently, ethyl acetate was added and the organic layer was washed with water. The organic layer was dried over magnesium sulphate. After removing the solvent *in vacuo* the crude product was purified by silica gel column chromatography using a mixture of cyclohexane and ethyl acetate (10:1) to obtain compound **2** as a colourless solid (1.14 g, 99%).

¹H NMR (300 MHz, CD₂Cl₂): δ (ppm) = 7.95 (d, *J* = 8.1 Hz, 2H), 7.86 (d, *J* = 8.1 Hz, 2H), 3.65 (s, 2H), 1.35 (s, 12H), 0.88 (s, 9H).

¹³C NMR (75 MHz, CD₂Cl₂): δ (ppm) = 138.47, 135.62, 127.23, 122.32, 84.95, 83.63, 79.89, 31.89, 26.50, 25.26, 25.06.

APCI-MS: *m/z* calcd. for C₁₇H₂₇BO₅S: 354.2, found 371.7 [M+NH₄]⁺.

3,4-Bis(4-bromophenyl)-2,5-diphenylcyclopenta-2,4-dien-1-one (1b**)**

1,2-Bis(4-bromophenyl)ethane-1,2-dione (**14**) (1.5 g, 4.08 mmol) and 1,3-diphenylacetone (**15**) (857 mg, 4.08 mmol) were dissolved in 150 mL ethanol and heated up to 85 °C. Subsequently, 1 M methanolic tetrabutylammonium hydroxide (TBAH) solution (1.06g, 4.08 mL, 4.08 mmol) was added. After stirring at 85 °C for 0.5 h, the reaction mixture was diluted in ethyl acetate. The organic layer was washed with water, dried over sodium sulphate and concentrated *in vacuo*. The product was precipitated in an ethanol water mixture to afford building block **1b** as a dark red solid (1.89 g, 85%).

¹H NMR (300 MHz, CD₂Cl₂): δ (ppm) = 7.41–7.32 (m, 4H), 7.32–7.25 (m, 6H), 7.24–7.15 (m, 4H), 6.90–6.77 (m, 4H).

¹³C NMR (75 MHz, CD₂Cl₂): δ (ppm) = 153.28, 132.24, 131.81, 131.38, 130.85, 130.49, 128.54, 128.18, 126.38, 123.35.

APCI-MS: *m/z* calcd. for C₂₉H₁₈Br₂O 539.97, found 540.7 [M+H]⁺.

Dineopentyl 4',4'''-(4-oxo-3,5-diphenylcyclopenta-2,5-diene-1,2-diyl)bis-[1,1'-biphenyl]-4-sulfonate) (3)

Building block **1b** (700mg, 1.29 mmol) and neopentyl 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzenesulfonate (**2**) were dissolved in 40 mL 1,4-dioxane. Then, an aqueous 2 M potassium carbonate solution (2.68 g, 9.68 mL, 19.4 mmol) was added and the mixture was degassed with argon. Tetrakis(triphenylphosphine)palladium(0) (Pd(PPh₃)₄) was added and the reaction mixture was stirred at 90 °C overnight under argon atmosphere. The crude mixture was diluted with dichloromethane and the organic layer was washed with water and dried over sodium sulfate. After removing the solvent *in vacuo* the crude product was purified by silica gel column chromatography using a mixture of cyclohexane and THF (5:1) to obtain building block **3** as a red solid (340 mg, 34%).

¹H NMR (300 MHz, CD₂Cl₂): δ (ppm) = 7.85 (dd, J = 48.4, 8.3 Hz, 8H), 7.52 (d, J = 8.1 Hz, 4H), 7.33–7.22 (m, 10H), 7.12 (d, J = 8.2 Hz, 4H), 0.89 (s, 18H).

¹³C NMR (75 MHz, CD₂Cl₂): δ = 200.21, 153.98, 145.80, 139.36, 135.30, 133.95, 131.15, 130.58, 128.84, 128.52, 128.11, 127.98, 127.30, 126.54, 80.23, 31.92, 26.10.

MALDI-TOF: *m/z* calcd. for C₅₁H₄₈O₇S₂ 3531.24, found 836.39 [M]^{•+}.

Pyr-G1-(Spen)₈ (**15**)

1,3,6,8-Tetraethynylpyrene (**8**) (6 mg, 20.1 μ mol) and building block **3** (101 mg, 121 μ mol, 6 equiv) were dissolved in 3 mL o-xylene, degassed with argon and stirred at 145 °C for 36 h in a sealed microwave tube. The solvent was removed *in vacuo* and the crude product was purified by silica gel chromatography using a mixture of cyclohexane and THF (2:1). The product was further purified by *recycling*-GPC in chloroform to obtain dendrimer **15** as a yellow solid (19 mg, 27%).

¹H NMR (700 MHz, CD₂Cl₂): δ (ppm) = 8.03 – 7.80 (m, 21H), 7.71 – 7.48 (m, 19H), 7.38 – 6.38 (m, 75H), 3.71 – 3.57 (m, 16H), 0.93 – 0.81 (m, 72H).

¹³C NMR (176 MHz, CD₂Cl₂): δ (ppm) = 146.12, 141.80, 141.54, 141.24, 140.17, 138.98, 136.90, 136.54, 136.19, 134.78, 132.82, 130.42, 130.33, 128.72, 128.68, 128.12, 128.07, 127.70, 127.61, 126.88, 126.28, 125.94, 80.12, 31.89, 26.09.

MALDI-TOF: *m/z* calcd. for C₂₂₄H₂₀₂O₂₄S₈ 3531.24, found 3532.72 [M+H]⁺.

Pyr-G1-S₈ (**10b**)

Pyr-G1-(Spen)₈ **15** (7.00 mg, 1.98 μ mol) was dissolved in 2 mL dry DMF, the resulting solution was degassed with argon and stirred at 180 °C in a sealed microwave tube for 48 h. The solvent was evaporated *in vacuo* and the product was precipitated in diethyl ether. The product was further purified by gel permeation chromatography (Sephadex LH-20) in DMF. Dendrimer **10b** was obtained as a slight yellow solid (5.2 mg, 88%).

¹H NMR (700 MHz, DMSO): δ (ppm) = 7.98–7.86 (m, 2H), 7.85–7.75 (m, 2H), 7.65–6.31 (m, 110H).

¹³C NMR (176 MHz, DMSO): δ (ppm) = 147.06, 146.43, 141.53, 140.92, 139.74, 139.06, 138.45, 136.59, 136.14, 131.79, 129.62, 127.63, 126.61, 125.99, 125.94, 125.43, 125.35, 125.01, 124.96, 124.51.

MALDI-TOF: *m/z* calcd. for C₁₈₄H₁₂₂O₂₄S₈ 2970.61, found 2971.59 [M+H]⁺, 2994.56 [M+Na]⁺, 3009.56 [M+K]⁺.

1.4 Synthesis of a first-generation positively charged amphiphilic polyphenylene dendrimer –(PN)₄

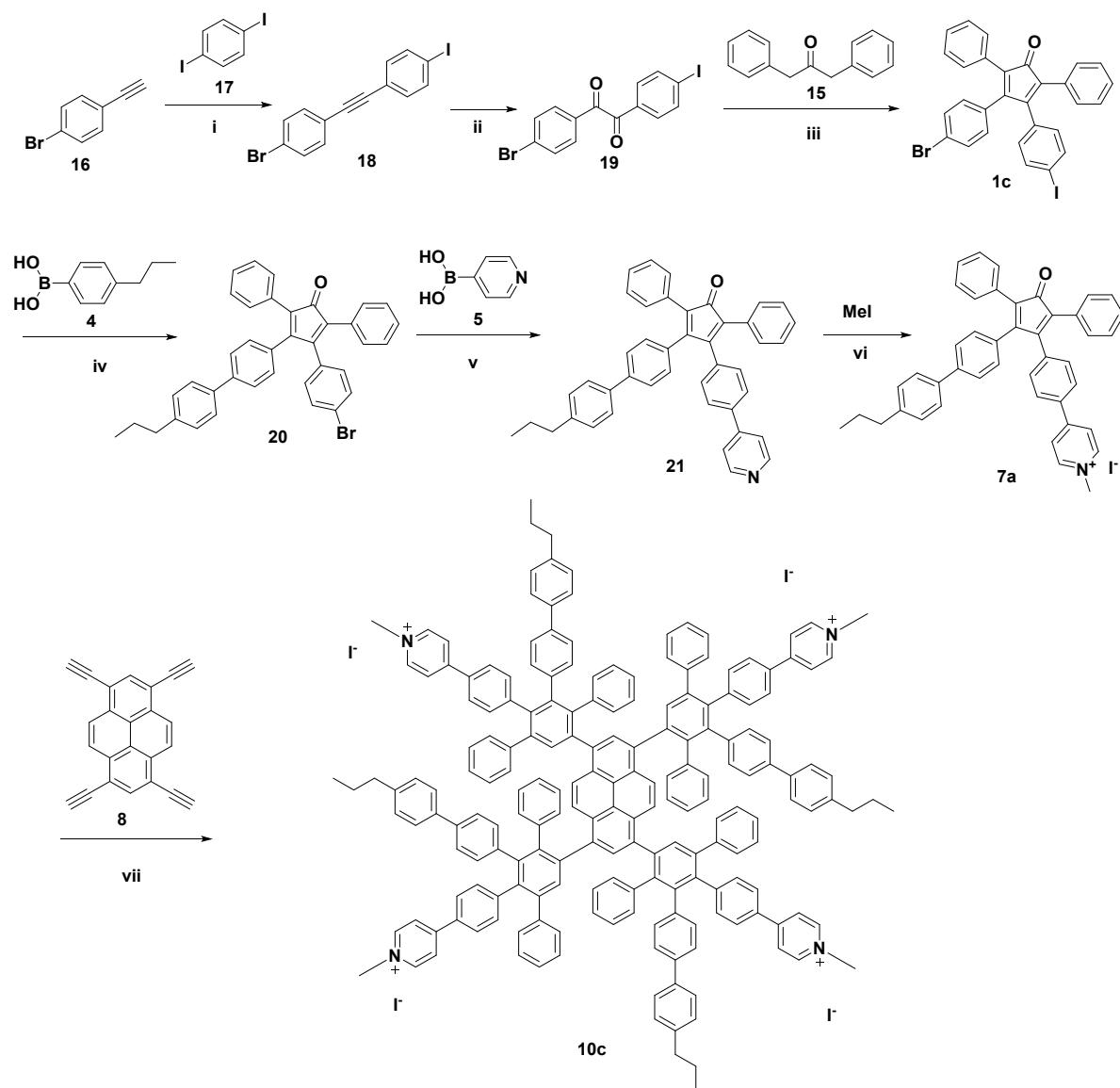


Figure S4. Synthesis of positively charged first-generation dendrimer **10c** with alternating pyridinium and *n*-propyl groups. i) Cul, PPh₃, PdCl₂(Ph₃P)₂, 1,4-dioxane, 40 °C, 15 h, 60% yield; ii) I₂, [Ru(p-cymene)Cl₂]₂, *tert*-butyl hydroperoxide (70% in H₂O), 1,4-dioxane, 50 °C, 15 h, 86% yield; iii) TBAH, ethanol, 85 °C, 20 min, 60% yield; iv) Pd(PPh₃)₄, K₂CO₃ (aq), 1,4-dioxane, 40 °C, 15 h; v) Pd(PPh₃)₄, K₂CO₃ (aq), 1,4-dioxane, 80 °C, 15 h, 60 % yield; vi) methanol, RT, 48 h, 70% yield; vii) DMSO, 140 °C, 3 d, 47% yield.

1-Bromo-4-((4-iodophenyl)ethynyl)benzene (18**)**

4-Bromophenylacetylene (2.0 g, 11.05 mmol) (**16**), 1,4-diiodobenzene (**17**) (7.29 g, 22.10 mmol, 2 equiv), copper(I) iodide (210 mg, 1.1 mmol, 0.1 equiv) and triphenylphosphine (579 mg, 2.21 mmol, 0.2 equiv) were dissolved in 200 mL 1,4-dioxane in a 500 mL flask. To this mixture 50 mL triethylamine was added and the solution was degassed with argon. Bis(triphenylphosphine)palladium(II) dichloride ($\text{PdCl}_2(\text{Ph}_3\text{P})_2$) (338 mg, 0.552 mmol) was added and the solution was stirred at 40 °C under argon atmosphere overnight. Then, the solution was filtered and the solvent was removed *in vacuo* followed by an extraction with dichloromethane. The organic layer was dried over magnesium sulphate and purified by silica gel flash chromatography using hexane as eluent to obtain compound **18** as a bright yellow solid (2.54 g, 60 %).

^1H NMR (300 MHz, THF- d_8): δ (ppm) = 7.75 (d, J = 7.9 Hz, 2 H), 7.55 (d, J = 8.0 Hz, 2 H), 7.42 (d, J = 8.1 Hz, 2 H), 7.27 (d, J = 8.0 Hz, 2 H).

^{13}C NMR (75 MHz, THF- d_8): δ (ppm) = 138.44, 134.62, 133.71, 133.62, 132.64, 132.43, 123.31, 123.17, 122.81, 95.06, 90.08.

APCI-MS: m/z calcd. for $\text{C}_{14}\text{H}_8\text{BrI}$: 381.9, found 383.9 [$\text{M}+2\text{H}]^+$.

1-(4-Bromophenyl)-2-(4-iodophenyl)ethane-1,2-dione (19**)**

Tolane **18** (1.0 g, 2.61 mmol), iodine (132 mg, 0.522 mmol, 0.2 equiv) and dichloro(*p*-cymene)ruthenium(II) dimer (16 mg, 0.026 mmol, 0.01 equiv) were added to a 50 mL flask and dissolved in as little 1,4-dioxane as possible. 1.5 mL (10.44 mmol, 4 equiv) of a 70 % *tert*-butyl hydroperoxide solution was slowly added to the solution. The mixture was stirred at 50 °C overnight. After evaporation of the solvent and purification via silica gel flash column chromatography using hexane as eluent followed by THF benzil **19** was obtained as a yellow solid (931 mg, 86 %).

^1H NMR (300 MHz, THF- d_8): δ (ppm) = 7.97 (d, J = 8.5 Hz, 2H), 7.88 (d, J = 8.6 Hz, 2H), 7.75 (d, J = 8.5 Hz, 2H), 7.70 (d, J = 8.5 Hz, 2H).

^{13}C NMR (75 MHz, THF- d_8): δ (ppm) = 193.57, 193.21, 139.43, 138.19, 133.33, 132.99, 132.27, 131.86, 130.91, 128.89, 125.95, 121.07, 104.30.

3-(4-Bromophenyl)-4-(4-iodophenyl)-2,5-diphenylcyclopenta-2,4-dien-1-one (1c**)**

Benzil **19** (500 mg, 1.2 mmol) and 1,3-diphenylacetone **3** (279 mg, 1.33 mmol, 1.1 equiv) were dissolved in 15 mL ethanol in a 50 mL flask. The solution was heated under reflux and 0.24 mL tetrabutylammonium hydroxide (1M in THF) were added. After 20 min the reaction was stopped by cooling in an ice bath. After cooling at –20 °C for a few hours the product was filtered off. Building block **1c** was received as a purple solid (424 mg, 60 %).

¹H NMR (300 MHz, CD₂Cl₂): δ (ppm) = 7.56 (d, J = 8.3 Hz, 2 H), 7.35 (d, J = 8.5 Hz, 2 H), 7.31–7.24 (m, 6 H), 7.24–7.17 (m, 4 H), 6.82 (d, J = 8.5 Hz, 2 H), 6.68 (d, J = 8.5 Hz, 2 H).

¹³C NMR (75 MHz, CD₂Cl₂): δ (ppm) = 200.17, 153.53, 153.42, 137.91, 132.92, 132.38, 131.96, 131.59, 131.53, 131.02, 131.01, 130.64, 128.70, 128.33, 126.57, 126.50, 123.50, 95.41.

APCI-MS: *m/z* calcd. for C₂₉H₁₈BrIO: 588.0, found 589.1 [M+H]⁺.

3-(4-Bromophenyl)-2,5-diphenyl-4-(4'-propyl-[1,1'-biphenyl]-4-yl)cyclopenta-2,4-dien-1-one (20**)**

Building block **1c** (230 mg, 0.390 mmol) and 4-propylbenzeneboronic acid (**4**) (83 mg, 0.507 mmol, 1.3 equiv) were added to a 50 mL flask and were dissolved in 25 mL 1,4-dioxane. To this solution 0.97 mL of a 2 M potassium carbonate solution were added and degassed with argon. Then, Pd(PPh₃)₄ (27 mg, 0.023 mmol, 0.06 equiv) was added and the reaction mixture was stirred at 40 °C overnight. The reaction mixture was filtered and extracted with DCM followed by a filtration over a silica gel flash column using a mixture of THF and hexane (1:19). Product **20** was used without further purification.

APCI-MS: *m/z* calcd. for C₃₈H₂₉BrO: 580.1, found 581.5 [M+H]⁺.

2,5-Diphenyl-3-(4'-propyl-[1,1'-biphenyl]-4-yl)-4-(4-(pyridin-4-yl)phenyl)cyclopenta-2,4-dien-1-one (21**)**

Building block **20** (380 mg, 0.653 mmol) and 4-pyridineboronic acid (**5**) (321 mg, 2.61 mmol, 4 equiv) were dissolved in 25 mL 1,4-dioxane in a 50 mL flask. 3.27 mL of a 2M potassium carbonate solution were added and the mixture was degassed. After adding Pd(PPh₃)₄ (45 mg, 0.038 mmol, 0.06 equiv) the mixture was stirred at 80 °C for two days under argon atmosphere. The solvent was removed followed by an extraction with DCM. The purification was performed by silica gel flash chromatography using a mixture of THF and hexane (1:2). Building block **21** was received as a violet solid (236 mg, 60%).

¹H NMR (300 MHz, CD₂Cl₂): δ (ppm) = 8.64–8.57 (m, 2 H), 7.58–7.43 (m, 8 H), 7.32–7.20 (m, 12 H), 7.12 (d, J = 8.4 Hz, 2 H), 7.03 (d, J = 8.4 Hz, 2 H), 2.66–2.55 (m, 2 H), 1.70–1.56 (m, 2 H), 0.94 (t, J = 7.3 Hz, 3 H).

¹³C NMR (75 MHz, CD₂Cl₂): δ (ppm) = 200.53, 154.81, 154.22, 152.06, 150.89, 150.83, 148.64, 147.60, 143.18, 141.65, 138.71, 138.51, 137.80, 136.47, 134.64, 132.17, 131.58, 131.44, 130.83, 130.78, 130.50, 129.66, 129.60, 129.58, 129.17, 128.87, 128.68, 128.65, 128.21, 128.12, 127.53, 127.15, 127.03, 126.80, 126.70, 126.13, 125.99, 122.09, 121.78, 38.16, 25.15, 14.17.

APCI-MS: *m/z* calcd. for C₄₃H₃₃NO: 579.3, found 580.4 [M+H]⁺.

1-Methyl-4-(4-(3-oxo-2,4-diphenyl-5-(4'-propyl-[1,1'-biphenyl]-4-yl)cyclopenta-1,4-dien-1-yl)phenyl)pyridin-1-i um (7a)

Building block **21** (220 mg, 0.38 mmol) was dissolved in 1 mL methyl iodide in a 10 mL Schlenk tube followed by the addition of 1 mL methanol. After two days, the product was precipitated by the addition of diethyl ether. After filtration, compound **7a** was obtained as a red-brown solid (192 mg, 70%).

¹H NMR (300 MHz, DMSO-*d*₆): δ (ppm) = 8.96 (d, J = 6.6 Hz, 2 H), 8.46 (d, J = 7.1 Hz, 2 H), 8.00 (d, J = 8.5 Hz, 2 H), 7.59 (d, J = 2.7 Hz, 2 H), 7.56 (d, J = 2.5 Hz, 2 H), 7.35–7.16 (m, 14 H), 7.08 (d, J = 8.4 Hz, 2 H), 4.29 (s, 3 H), 2.56 (t, J = 7.5 Hz, 2 H), 1.58 (h, J = 7.4 Hz, 2 H), 0.87 (t, J = 7.3 Hz, 3 H).

¹³C NMR (75 MHz, DMSO-*d*₆): δ (ppm) = 199.29, 154.27, 153.12, 153.03, 145.53, 141.99, 139.93, 136.65, 136.13, 133.09, 131.26, 130.43, 130.31, 130.20, 129.86, 129.59, 128.16, 128.11, 127.79, 127.60, 126.29, 126.11, 125.82, 124.93, 123.88, 47.04, 36.74, 23.91, 13.52.

ESI-MS: *m/z* calcd. for C₄₄H₃₆NO⁺: 594.28, found 592.22 [M+H]⁺.

Pyr-G1(PN)₄ (**10c**)

Building block **7a** (100 mg, 0.17 mmol) and 1,3,6,8-tetraethynylpyrene (**8**) (6.9 mg, 0.023 mmol, 0.14 equiv) were dissolved in 5 mL DMSO in a 10 mL Schlenk tube. After degassing with argon, the mixture was heated up to 140 °C for three days. After the addition of diethyl ether product **10c** precipitated and was filtered off. Purification was performed applying dialyses (1000 MWCO) in methanol. Dendrimer **10c** was received as yellow crystals (33.3 mg, 47 %).

¹H NMR (300 MHz, DMSO-*d*₆): δ (ppm) = 8.90 (d, J = 6.7 Hz, 8 H), 8.47–6.04 (m, 106 H), 4.25 (s, 12 H), 1.65–1.42 (m, 8 H), 0.86 (t, J = 6.5 Hz, 12 H).

¹³C-NMR (126 MHz, DMSO-*d*₆): δ (ppm) = 153.09, 145.46, 141.13, 140.68, 136.32, 132.60, 131.58, 130.31, 129.66, 128.88, 127.88, 126.60, 125.91, 124.83, 123.33, 46.88, 36.71, 24.64, 13.53.

ESI-MS: *m/z* calcd. for (C₁₉₆H₁₅₄N₄)⁴⁺: 641.31, found 641.14; *m/z* calcd. for (C₁₉₆H₁₅₄N₄)³⁺: 897.38, found 897.31; *m/z* calcd. for (C₁₉₆H₁₅₄N₄)²⁺: 1409.52, found 1409.44.

1.5 Synthesis of a second-generation positively charged amphiphilic polyphenylene dendrimer – (PN)₈

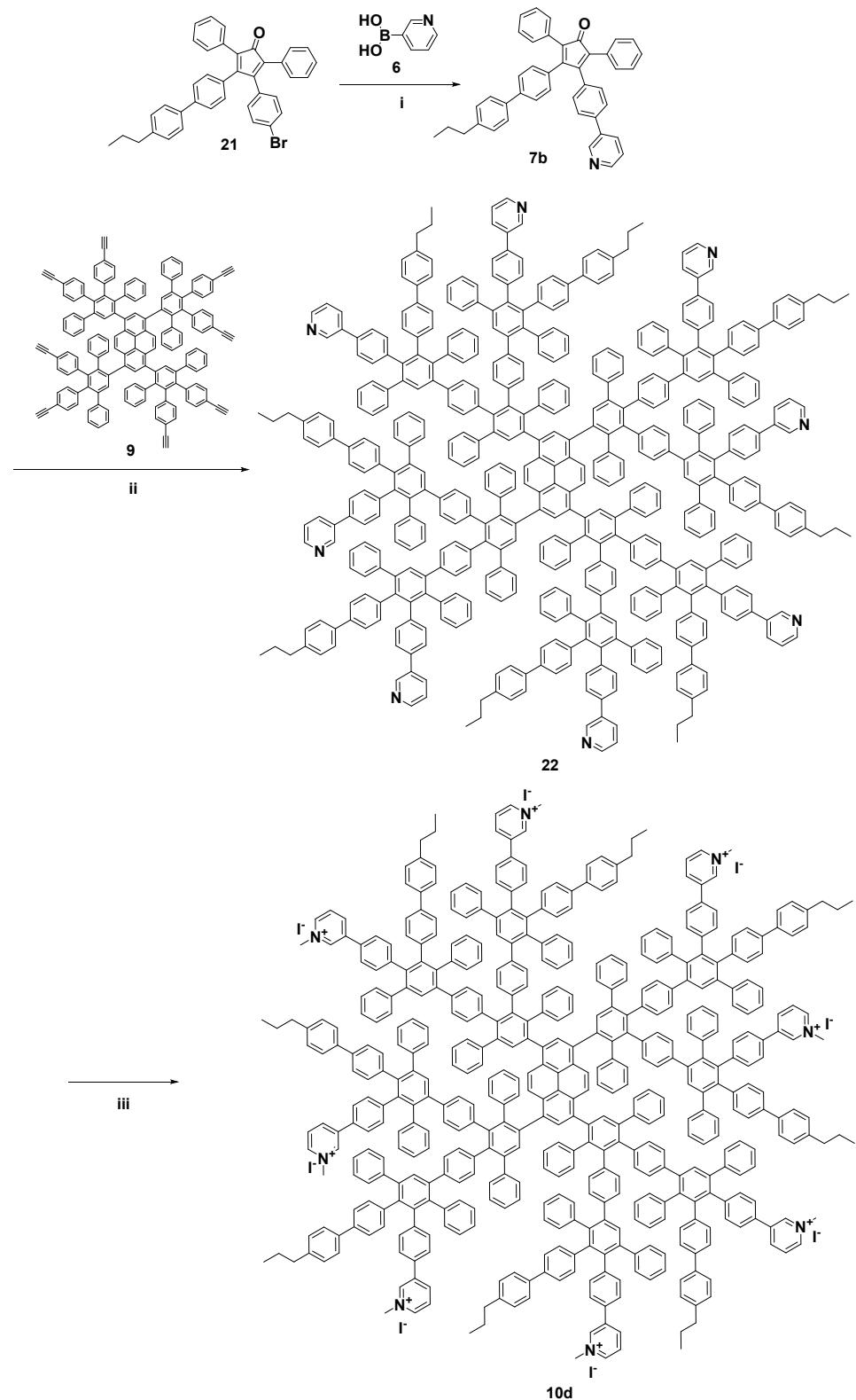


Figure S5. Synthesis of second-generation dendrimer **15d** with alternating pyridinium and *n*-propyl groups. i) $\text{Pd}(\text{PPh}_3)_4$, K_2CO_3 (aq), 1,4-dioxane, 80°C , 15 h, 50 % yield; ii) *o*-xylene, 160°C , 3 d, 64% yield; iii) methanol, RT, 15 h, 92% yield.

2,5-Diphenyl-3-(4'-propyl-[1,1'-biphenyl]-4-yl)-4-(4-(pyridin-3-yl)phenyl)cyclopenta-2,4-dien-1-one (7b**)**

Compound **21** (380 mg, 0.653 mmol) and 4-pyridineboronic acid (**6**) (321 mg, 2.61 mmol, 4 equiv) were dissolved in 25 mL 1,4-dioxane in a 50 mL flask. 3.27 mL of a 2M potassium carbonate solution were added and the mixture was degassed with argon. After adding Pd(PPh₃)₄ (45 mg, 0.038 mmol, 0.06 equiv) the mixture was stirred at 80 °C for two days under argon atmosphere. The solvent was removed *in vacuo* followed by an extraction with dichloromethane (DCM). The purification was performed by silica gel flash chromatography using a mixture of THF and hexane (1:2). Building block **7b** was received as a violet solid (112 mg, 50%).

¹H NMR (300 MHz, CD₂Cl₂): δ (ppm) = 8.82 (d, J = 2.4 Hz, 1 H), 8.55 (dd, J = 4.9, 1.6 Hz, 1 H), 7.88 (dt, J = 8.1, 2.1 Hz, 1 H), 7.55–7.43 (m, 5 H), 7.27 (d, J = 9.3 Hz, 14 H), 7.11 (d, J = 8.2 Hz, 2 H), 7.04 (d, J = 8.3 Hz, 2 H), 2.61 (t, J = 7.6 Hz, 2 H), 1.74–1.54 (m, 2 H), 0.94 (t, J = 7.3 Hz, 3 H).

¹³C NMR (75 MHz, CD₂Cl₂): δ (ppm) = 200.61, 154.85, 154.44, 149.31, 148.66, 143.16, 141.61, 138.41, 137.82, 136.12, 134.58, 133.56, 132.22, 131.62, 131.52, 130.79, 130.76, 130.50, 129.55, 128.63, 128.61, 128.14, 128.07, 127.14, 127.06, 126.76, 126.53, 126.13, 125.97, 124.08, 38.14, 25.12, 14.12.

APCI-MS: *m/z* calcd. for C₄₃H₃₃NO: 579.3, found 580.4 [M+H]⁺.

Pyr-G1-(ethynyl)₈ **9**

First generation dendrimer **9** was synthesized according to the literature.¹

¹H NMR (300 MHz, THF-*d*₈): δ (ppm) = 8.05–7.31 (m, 8 H), 7.29–7.02 (m, 28 H), 7.02–6.71 (m, 34 H), 6.71–6.43 (m, 12 H), 3.47–3.32 (m, 8 H).

¹³C NMR (126 MHz, THF-*d*₈): δ (ppm) = 142.00, 141.69, 132.76, 131.84, 131.49, 131.42, 130.98, 130.85, 128.69, 128.61, 121.22, 120.89, 78.90.

MALDI-TOF: *m/z* calcd. for C₁₅₂H₉₀: = 1914.70, found 1914.62 [M]⁺⁺.

Pyr-G2-(PN)₈ uncharged **22**

First generation dendrimer core **9** (50 mg, 0.0261 mmol) and building block **7b** (242 mg, 0.418 mmol, 16 equiv) were dissolved in 1 mL *o*-xylene in a 10 mL Schlenk tube. After degassing, the reaction mixture was heated up to 160 °C for three days. Subsequently, dendrimer **22** was precipitated in methanol and purified by GPC in THF. Dendrimer **22** was obtained as a yellow solid (106 mg, 64%).

¹H NMR (500 MHz, THF-*d*₈): δ (ppm) = 8.67 (d, *J* = 12.4 Hz, 8H), 8.40 (s, 8H), 7.80–7.72 (m, 8H), 7.56–6.30 (m, 274H), 1.60 (h, *J* = 6.1, 5.7 Hz, 16H), 0.90 (t, *J* = 7.3 Hz, 24H).

¹³C NMR (126 MHz, THF-*d*₈): δ (ppm) = 149.21, 148.92, 142.99, 142.59, 142.41, 142.35, 142.33, 142.17, 142.10, 141.75, 141.66, 141.39, 141.25, 141.17, 140.37, 140.27, 139.97, 139.81, 139.74, 139.42, 139.05, 138.89, 138.85, 138.77, 136.66, 136.63, 135.99, 135.71, 134.19, 134.16, 133.43, 133.09, 130.88, 129.61, 129.59, 128.58, 127.95, 127.26, 127.24, 126.68, 126.27, 125.99, 125.72, 124.10, 68.07, 67.92, 67.75, 67.57, 38.53, 25.94, 25.79, 25.64, 25.48, 14.15.

MALDI-TOF: *m/z* calcd. for C₄₈₈H₃₅₄N₈ 6324.79, found 6325.89 [M+H]⁺.

Pyr-G2-(PN)₈ **10d**

Dendrimer **22** (106 mg, 0.017 mmol) was dissolved in 1 mL methyl iodide in a 10 mL Schlenk tube followed by the addition of 1 mL methanol. After 24 h diethyl ether was added leading to precipitation of the product which was filtered off to obtain dendrimer **10d** as a yellow-brown solid (116 mg, 92 %).

¹H NMR (300 MHz, DMSO-*d*₆): δ (ppm) = 9.22 (s, 8 H), 8.81 (s, 8 H), 8.68 (s, 8 H), 8.03 (s, 8 H), 7.63–6.38 (m, 266 H), 4.27 (s, 24 H), 1.60–1.44 (m, 16 H), 0.83 (t, *J* = 7.0 Hz, 24 H).

¹³C NMR (126 MHz, DMSO-*d*₆): δ (ppm) = 153.21, 143.25, 140.89, 140.25, 137.99, 136.30, 132.30, 131.63, 130.83, 129.95, 129.48, 128.87, 127.78, 127.57, 125.88, 125.53, 124.62, 124.23, 47.95, 40.02, 39.86, 39.69, 39.52, 39.35, 39.19, 39.02, 36.69, 23.99, 13.52.

ESI-MS: *m/z* calcd. for (C₄₉₆H₃₇₈N₈)⁸⁺: 806.37, found 806.24; *m/z* calcd. for (C₄₉₆H₃₇₈N₈I)⁷⁺: 939.56, found 939.55; *m/z* calcd. for (C₄₉₆H₃₇₈N₈I₂)⁶⁺: 1117.30, found 1117.29; *m/z* calcd. for (C₄₉₆H₃₇₈N₈I₃)⁵⁺: 1366.14, found 1366.15; *m/z* calcd. for (C₄₉₆H₃₇₈N₈I₄)⁴⁺: 1739.40, found 1739.39.

2 Protein corona on dendrimer-coated liposomes and polystyrene nanoparticles

2.1 Material and methods

Liposome synthesis. Based on previous reports,⁵ amine functionalized liposomes were synthesized using 1,2-dioleoyl-sn-glycero-3-phosphoethanolamine (DOPE), L- α -phosphatidylcholine (egg PC) and cholesterol (Chol) with a molar ratio of egg PC:DOPE:Chol = 1:1:1 by film hydration followed by extrusion. Cholesterol and the lipids were dissolved in chloroform at a concentration of 10 mg mL⁻¹. All components (Egg PC = 835 μ L, DOPE = 767 μ L and Cholesterol = 398 μ L) were mixed with 2 mL of chloroform containing 1 vol% EtOH in a 50 mL round-bottomed flask. The mixture was dried using a rotary evaporator at room temperature (30 min, 450 mbar) and further at 42°C (30 min, 3 mbar). To remove organic solvent residuals the flask was placed into a vacuum oven for 1 hour. (Diameter \varnothing : 242 \pm 6 nm, ζ – Potential: - 52 \pm 6 mV)

Polystyrene nanoparticle synthesis. According to previous reports,⁶ amine functionalised polystyrene nanoparticles were prepared via direct miniemulsion. For colloidal stability, the ionic surfactant cetyl trimethyl ammonium chloride was used. To generate amino groups, 2-aminoethyl methacrylate hydrochloride (2 wt% to styrene) was copolymerized with styrene. Excessive surfactant was removed via dialysis. A detailed protocol is given in previous reports.⁷ (Diameter \varnothing : 98 \pm 10 nm, ζ – Potential: + 34 \pm 12 mV)

Coating of nanoparticles and liposomes with dendrimers. All dendrimers were dissolved in DMSO at a concentration of 10 mg mL⁻¹. PS-NH₂ nanoparticles (10 mg mL⁻¹, 100 μ L) and liposomes-NH₂ (3 mg mL⁻¹, 333 μ L) were mixed with the dendrimers (10 mg mL⁻¹, 100 μ L) and incubated at room temperature for 1 h. To remove unbound dendrimers, the liposome dispersion was diluted with 1 mL of PBS and the PS-nanoparticle dispersion with 1 mL of water. Afterwards, the mixture was centrifuged (20 000 g, 15 min, 4°C). Finally, polystyrene nanoparticles were resuspended in 100 μ L of water and liposomes were resuspended in 100 μ L of PBS.

Zeta Potential. Measurements were performed using a Nano Z Zetasizer (Malvern Instruments GmbH, Herrenberg, Germany). Uncoated and coated liposomes/PS-

nanoparticles (20 µL) were diluted with 1 mL of a 1 mM KCl solution and measured instantly at 25 °C. Each measurement was repeated in technical triplicate.

Human serum/plasma. Human blood serum and plasma was obtained from healthy donors from the Transfusion Center of the University Clinic of Mainz, Germany. For plasma preparation, citrate was used as an anticoagulant. Six serum batches and ten plasma batches were pooled together and stored at –20 °C. All experiments were performed in compliance with the relevant laws and institutional guidelines. The institutional ethics committee had approved the study (Landesärztekammer Rheinland-Pfalz, 837.439.12 (8540-F)). Written informed consent was obtained for any experimentation with human subjects.

Protein corona analysis. Uncoated and coated liposomes or polystyrene nanoparticles (10 mg mL⁻¹, 100 µL) were incubated with 1 mL of human serum or human plasma at 37 °C for 1 h. The protein corona coated liposomes and nanoparticles were centrifuged (20 000 g, 15 min, 4 °C) and washed with 1 mL PBS solution to remove unbound and loosely bound proteins. Overall, this step was repeated three times. The adsorbed corona proteins were detached from the surface by resuspending the liposome/nanoparticle pellet in 100 µL of an aqueous solution containing 2% SDS supplemented with 62.5 mM Tris-HCL (Sigma Aldrich, Germany). Afterwards, the dispersion was incubated for 5 min at 95 °C and centrifuged as described above. The supernatant contained the detached corona proteins.

Protein Assay. The protein concentration was determined by Pierce 660 nm Protein Assay (Thermo Fisher, Germany) according to manufacturer's instructions.

In solution digestion. Prior to LC-MS analysis, proteins were digested according to former protocols.^{7,8} Briefly, SDS was removed via Pierce detergent removal columns (Thermo Fisher, Germany) according to the manufacture's instruction. Afterwards, proteins were precipitated using ProteoExtract protein precipitation kit (Merck Millipore, Germany) overnight and the protein pellet was isolated via centrifugation (14 000 g, 10 min). Precipitated proteins were resuspended in RapiGest SF (Waters, USA) dissolved in 50 mM ammonium bicarbonate (Sigma Aldrich, Germany) and incubated at 80 °C for 15 min. Further, proteins were reduced with dithiothreitol at 56 °C for 45 min (5 mM, Sigma Aldrich, Germany) and alkylated with iodoacetoamide at room temperature in the dark for 1 h (15 mM, Sigma Aldrich, Germany). Digestion was carried out using a protein:trypsin ratio of 50:1 for 14–18 h at 37 °C and stopped by adding 2 µL hydrochloric acid (Sigma Aldrich, Germany). To remove degradation products of RapiGest SF, the peptide sample was centrifuged (14 000 g, 15 min, 4 °C).

Liquid chromatography-high resolution mass spectrometry (LC-MS). The peptide samples were spiked with 50 fmol of Hi3 EColi Standard (Waters, USA) for absolute protein quantification. Peptides were separated using a nanoACQUITY UPLC system equipped with a C18 nanoACQUITY Trap Column (5 µm, 180 µm x 20 mm, Waters, USA) and C18 analytic reversed phase column (1.7 µm, 75 µm x 150 mm, Waters, USA). The UPLC system was coupled to a Synapt G2-Si mass spectrometer (Waters, USA) and electrospray ionization (ESI) was carried in positive ion with a nanoLockSpray source. The high-resolution mass spectrometer was operated in resolution mode performing data-independent experiments (MS^E). Data was processed with MassLynx 4.1 and all proteins were identified with Progenesis QI. A reviewed human data base downloaded from Uniprot and the peptide sequence of Hi3 Ecoli standard (Chaperone protein CLpB, Water, USA) was added for absolute protein quantification. Based on the TOP3/Hi3 approach⁹ all proteins were quantified and the absolute amount in fmol is calculated.

Statistical Analysis. GraphPad Prism 8.4.2 Software was used for statistical analysis applying a one-way ANOVA followed by Tukey's post-hoc multiple comparisons test for all protein corona results in the main text and Figure S11 in order to compare the difference between both uncoated liposome/ PS nanoparticle and lipo-dendrimers/PS-dendrimers as well as between the different dendrimer-coatings. For zeta-potential and Pierce Assay and concentration dependency measurements a one-way ANOVA followed by Dunnett's post-hoc multiple comparisons test was used to compare the coatings with the uncoated liposomes/PS nanoparticles. Significance was considered at *p*-values such as not significant (ns) for *p* > 0.05, and significant * for *p* ≤ 0.05, ** for *p* ≤ 0.01, *** for *p* ≤ 0.001, **** for *p* ≤ 0.0001).

2.2 Supplementary figures: Coating of liposomes and polystyrene nanoparticles and their respective protein corona

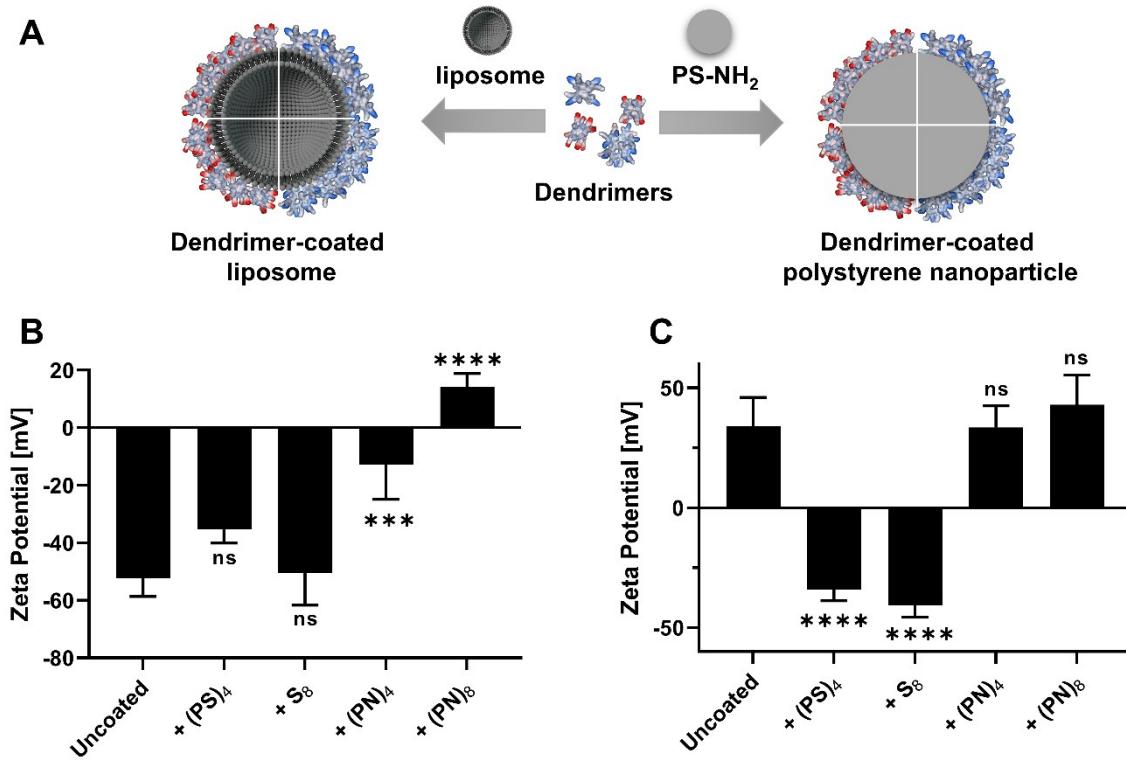


Figure S6. Verification of dendrimer-coating on liposomes and polystyrene nanoparticles ($PS-NH_2$). **(A)** Dendrimers were mixed with liposomes and ζ -potentials of **(B)** lipo-dendrimers and **(C)** PS-dendrimers were measured. $(PN)_4$ and S_8 did not show a clear change in ζ -potential values when mixed with liposomes indicating an insufficient coating. The same was observed for $(PN)_4$ and $(PN)_8$ when mixed with PS nanoparticles ($n = \text{minimum } 2$; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; ns for $p > 0.05$, *** for $p \leq 0.001$, **** for $p \leq 0.0001$).

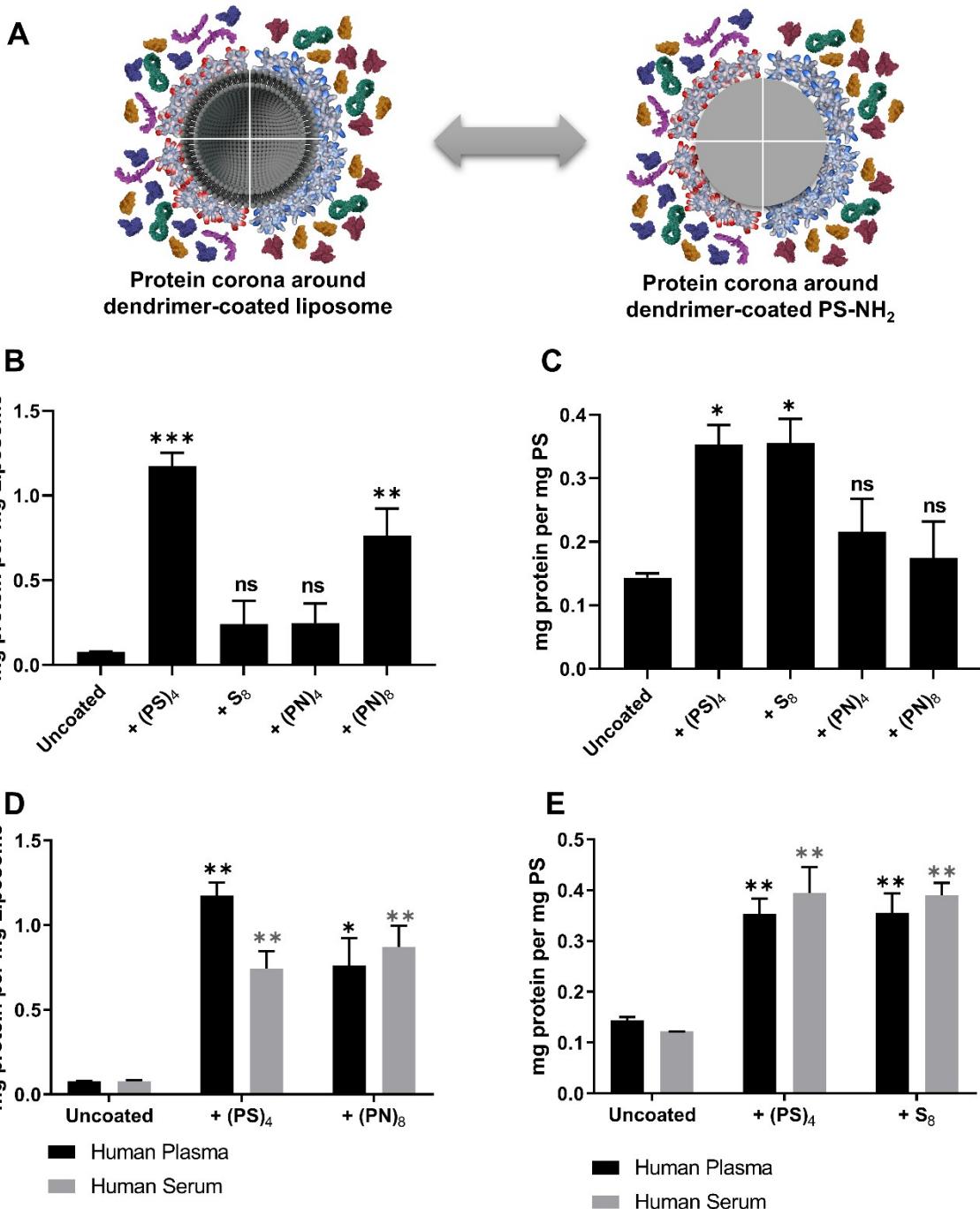


Figure S7. Verification of protein adsorption on lipo-dendrimers and PS-dendrimers: **(A)** Comparison of protein adsorption on lipo-dendrimers and PS-dendrimers. Quantification of protein adsorption by Pierce Assay for **(B)** lipo-dendrimers and **(C)** PS-dendrimers in blood plasma. Liposomes and PS nanoparticles with insufficient dendrimer binding did not show significant differences in protein quantities. Protein quantification by Pierce Assay for efficient dendrimer-coatings in human plasma and serum for **(D)** lipo-(PS)₄ and lipo-(PN)₈ as well as **(E)** PS-(PS)₄ and PS-S₈ ($n = \text{minimum } 2$; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; ns for $p > 0.05$, * for $p \leq 0.05$, ** for $p \leq 0.01$, *** for $p \leq 0.001$).

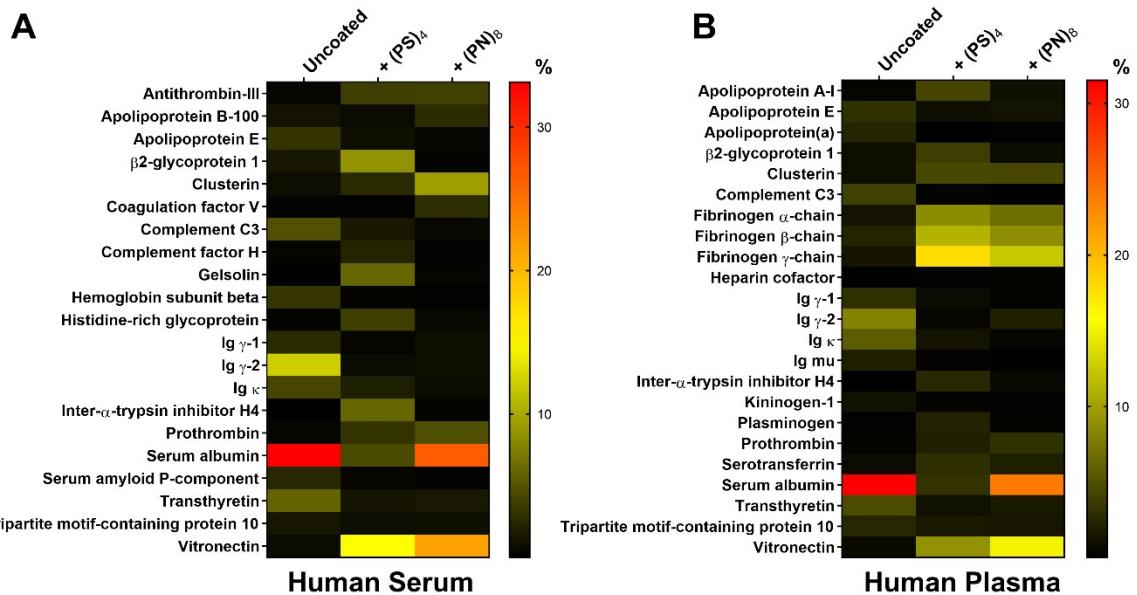


Figure S8. Heatmaps of adsorbed proteins to dendrimer-coated liposomes lipo-(PS)₄ and lipo-(PN)₈ in (A) blood serum and (B) blood plasma. A list of all identified proteins is provided in Table S1.

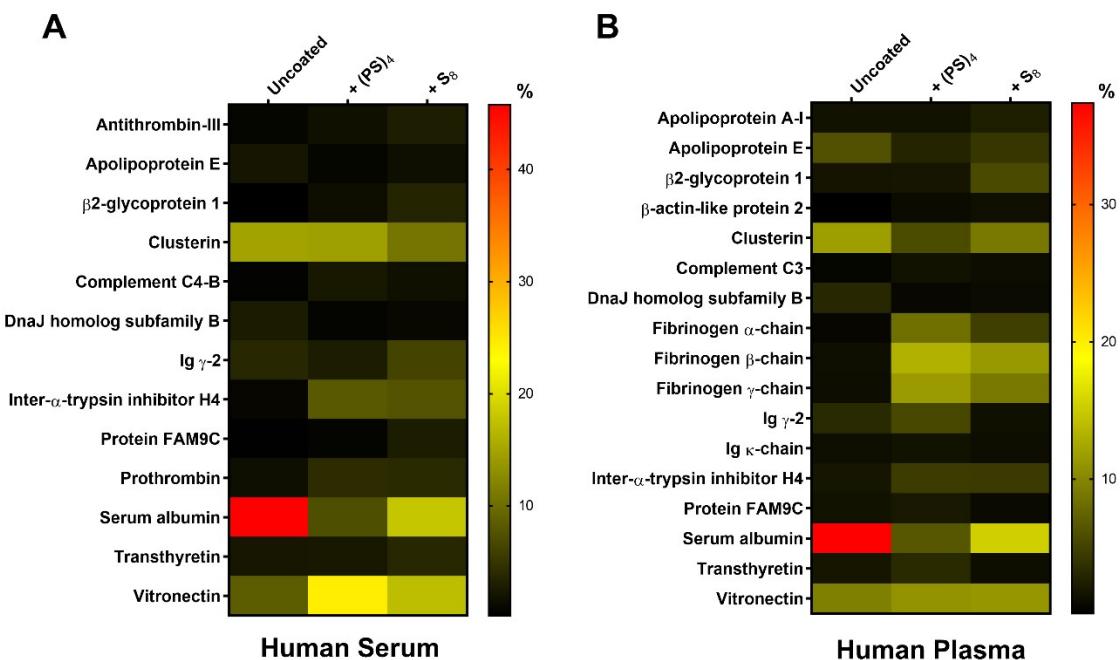


Figure S9. Heatmaps of adsorbed proteins to dendrimer-coated polystyrene nanoparticles PS-(PS)₄ and PS-S₈ in (A) blood serum and (B) blood plasma. A list of all identified proteins is provided in Table S2.

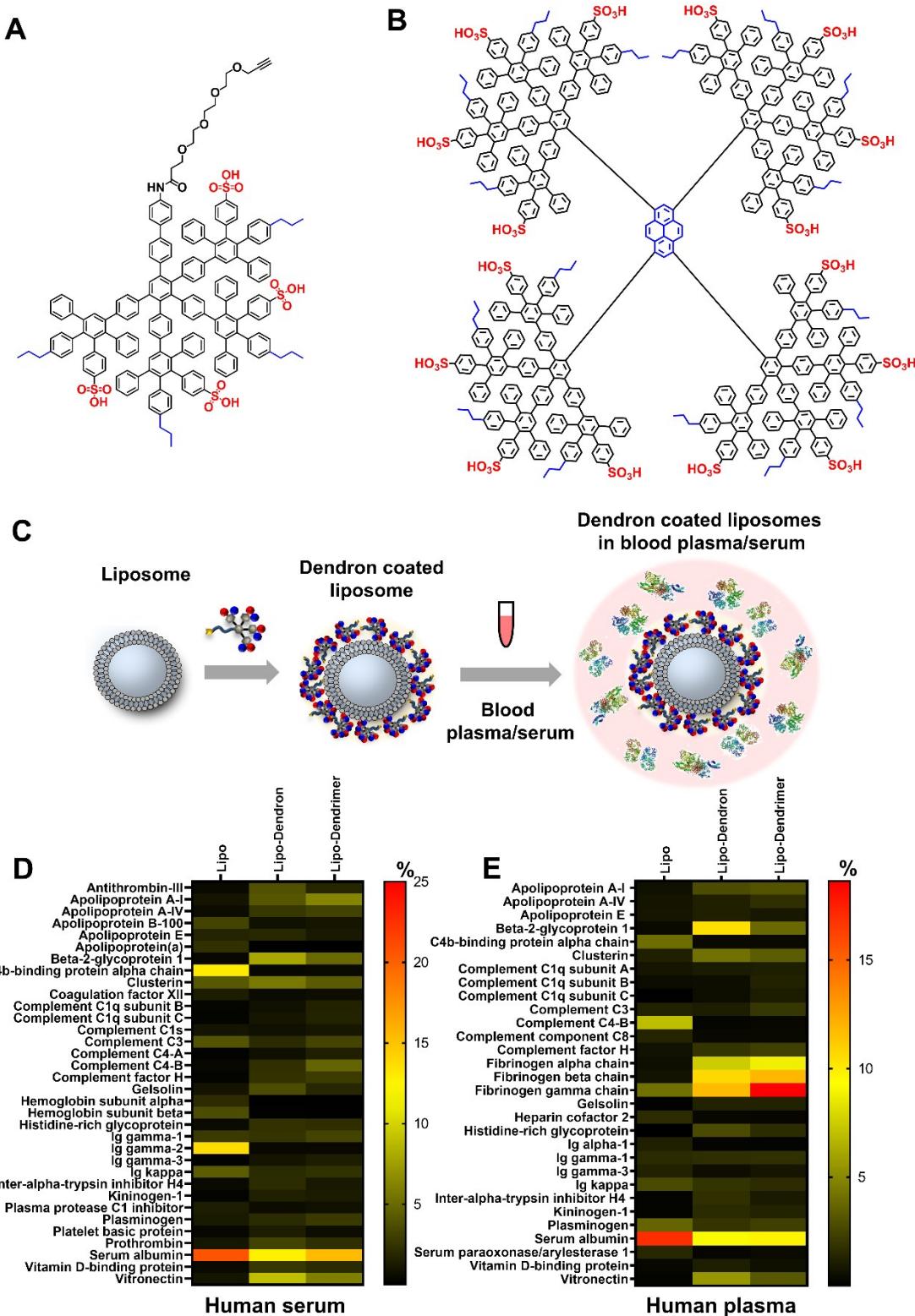


Figure S10. Protein corona on dendron- and dendrimer-coated liposomes. (A) Amphiphilic polyphenylene dendron³ and (B) amphiphilic polyphenylene dendrimer¹ with high density of surface pattern. (C) Coating of liposomes and addition of human serum and plasma. Heat map of adsorbed proteins on lipo-dendron and lipo-dendrimer in (D) human serum and (E) human plasma. (C-E). Reproduced from Wagner *et al.*³ (<https://doi.org/10.1002/anie.201913708>). Licensed under a Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).

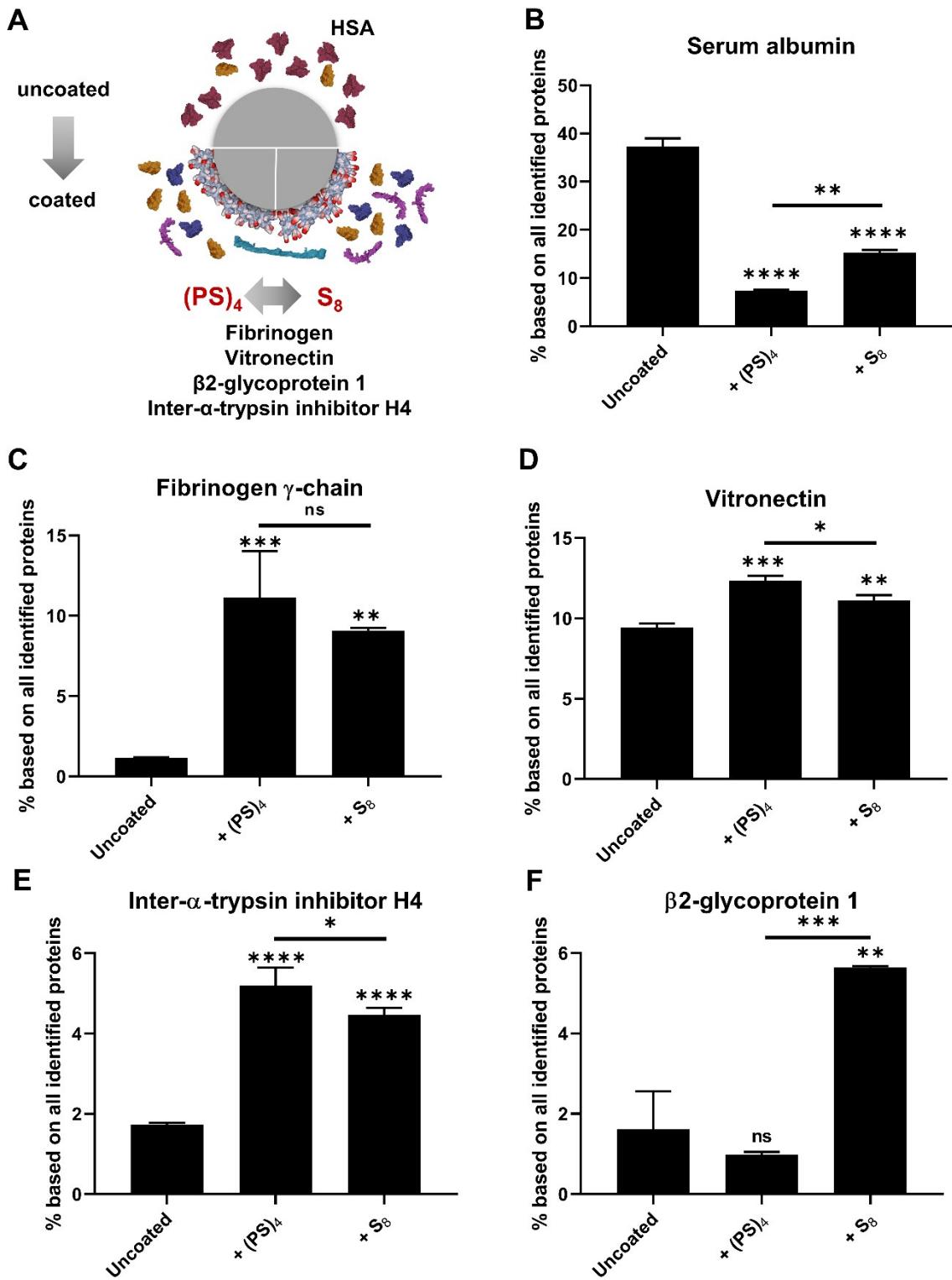


Figure S11. Comparison of adsorbed proteins between uncoated polystyrene nanoparticles ($PS-NH_2$) and PS-dendrimers ($PS-(PS)_4$ and $PS-S_8$). **(A)** Coating of $PS-NH_2$ lead to a different protein corona: Reduction of **(B)** serum albumin adsorption and enhancement of **(C)** fibrinogen, **(D)** vitronectin, **(E)** inter- α -trypsin inhibitor H4 and **(F)** $\beta 2$ -glycoprotein 1 ($n = \text{minimum } 2$; one-way ANOVA, Tukey's post-hoc multi-comparisons test; ns for $p > 0.05$, * for $p \leq 0.05$, ** for $p \leq 0.01$, *** for $p \leq 0.001$, **** for $p \leq 0.0001$).

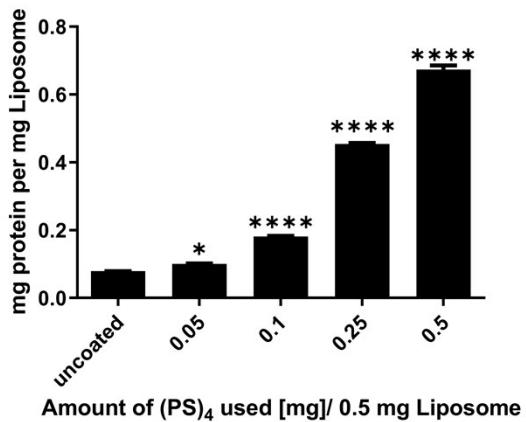


Figure S12. Quantification of protein adsorption in human plasma by Pierce Assay for different (PS)₄ concentrations used for the coating to liposomes. The amount of proteins adsorbed increases with higher dendrimer concentrations. (n = 2; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; * for $p \leq 0.05$, *** for $p \leq 0.0001$).

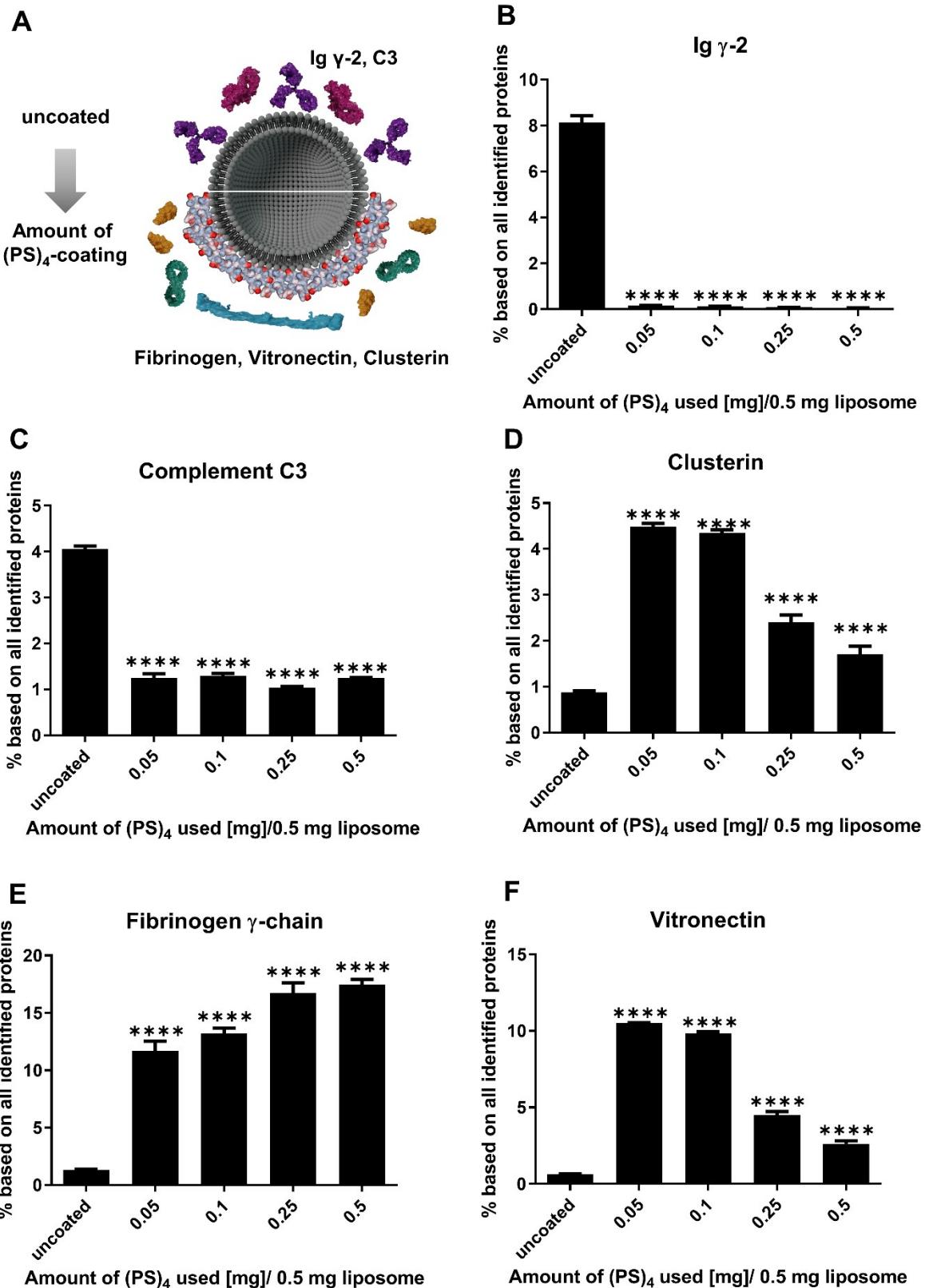


Figure S13. Concentration dependency of dendrimer-coatings on protein adsorption in human plasma. **(A)** Comparison between uncoated liposomes and lipo- $(PS)_4$ at different dendrimer concentrations used for the liposome coating. The adsorption of the opsonins **(B)** Ig γ -2 and **(C)** complement C3 is significantly reduced even at low dendrimer concentrations, whereas the adsorption of **(D)** Clusterin, **(E)** Fibrinogen and **(F)** Vitronectin is significantly enhanced ($n = 3$; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; * for $p \leq 0.05$, *** for $p \leq 0.0001$). A list of all identified proteins is provided in Table S3.

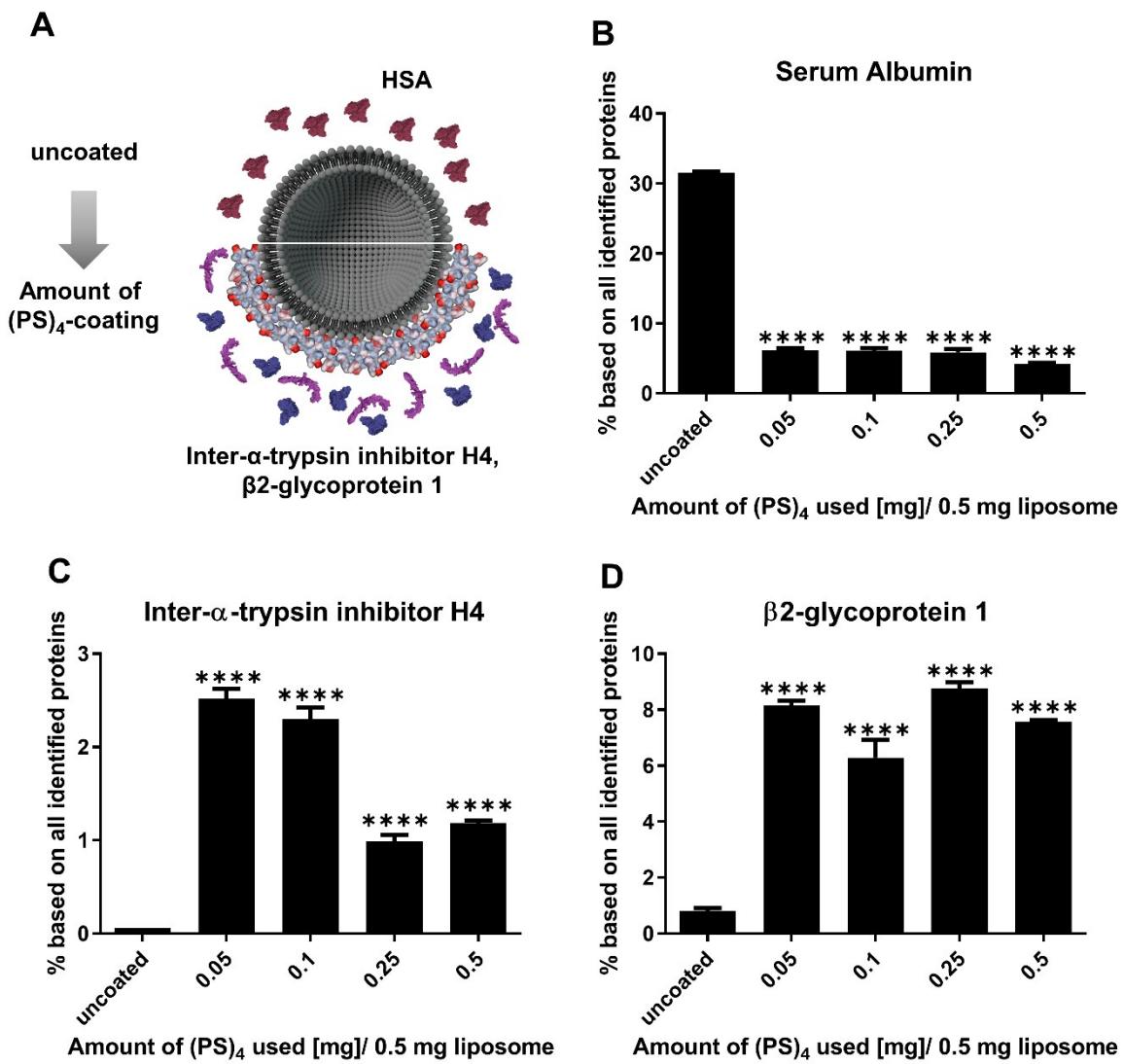


Figure S14. Concentration dependency of dendrimer-coatings specific for $(PS)_4$ on protein adsorption in human plasma. **(A)** Comparison between uncoated liposomes and lipo- $(PS)_4$ at different dendrimer concentrations used for the liposome coating. The adsorption of **(B)** serum albumin is significantly reduced even at low dendrimer concentrations, whereas the adsorption of **(C)** Inter- α -trypsin inhibitor H4 and **(D)** β 2-glycoprotein 1 is significantly enhanced ($n = 3$; one-way ANOVA, Dunnett's post-hoc multi-comparisons test; * for $p \leq 0.05$, **** for $p \leq 0.0001$). A list of all identified proteins is provided in Table S3.

Table S1. List of all identified corona proteins on lipo-dendrimers lipo-(PS)4 and lipo-(PN)8 in human serum and plasma.

Accession	Peptide count	Unique peptides	Description	Average in %								Standard deviation			
				Plasma		Unadjusted		Serum		Unadjusted		Plasma		Serum	
				(PS)4	(PN)8	(PS)4	(PN)8	(PS)4	(PN)8	(PS)4	(PN)8	(PS)4	(PN)8	(PS)4	(PN)8
62367_P02981	6	3	Actin, actinic smooth muscle OS-Homo sapiens GN- <i>GACTA2</i> PE=1 SV=1	0.43	0.00	0.01	0.00	0.13	0.07	0.28	0.00	0.01	0.00	0.05	0.25
62361_P05262	9	3	Actin, cytoplasmic 1 OS-Homo sapiens GN- <i>ACTB</i> PE=1 SV=1	0.56	0.08	0.12	0.06	0.48	0.32	0.01	0.02	0.00	0.06	0.36	0.11
P02763	3	3	Alpha-1-acid glycoprotein 1 OS-Homo sapiens GN- <i>ORM1</i> PE=1 SV=1	0.28	0.01	0.00	0.35	0.10	0.03	0.01	0.00	0.00	0.01	0.14	0.02
P01011	2	2	Alpha-1-antichymotrypsin OS-Homo sapiens GN- <i>SERPINAS1</i> PE=1 SV=2	0.04	0.00	0.01	0.03	0.07	0.01	0.00	0.01	0.00	0.08	0.01	
P01099	14	12	Alpha-1-antitrypsin OS-Homo sapiens GN- <i>SERPINAA1</i> PE=1 SV=3	0.67	0.11	0.16	0.54	0.24	0.27	0.04	0.01	0.00	0.01	0.17	0.16
P04217	4	4	Alpha-2-macroglobulin OS-Homo sapiens GN- <i>AMGB</i> PE=1 SV=1	0.03	0.20	0.00	0.02	0.06	0.37	0.02	0.01	0.00	0.01	0.08	0.51
P02997	10	10	Alpha-2-tryptinin OS-Homo sapiens GN- <i>TRYPTIN</i> PE=1 SV=3	0.09	0.26	0.14	0.06	0.25	0.37	0.01	0.00	0.01	0.24	0.34	
P02765	11	11	Alpha-2-HS glycoprotein OS-Homo sapiens GN- <i>AHSG</i> PE=1 SV=1	0.04	0.46	0.04	0.02	0.90	0.09	0.00	0.01	0.00	0.70	0.05	
P01023	11	9	Alpha-2-macroglobulin OS-Homo sapiens GN- <i>AMGB</i> PE=1 SV=3	0.21	0.02	0.01	0.11	0.04	0.06	0.00	0.00	0.00	0.04	0.03	
P01019	8	7	Angiotensinogen OS-Homo sapiens GN- <i>ANGTG</i> PE=1 SV=1	0.00	0.02	0.00	0.05	0.21	0.00	0.00	0.00	0.01	0.22	0.04	
P01917	4	2	Antithrombin III OS-Homo sapiens GN- <i>ANTX</i> PE=1 SV=1	0.22	0.00	0.01	0.10	0.10	0.02	0.00	0.00	0.00	0.00	0.1	
P02098	38	37	Antithrombin III OS-Homo sapiens GN- <i>AP0001</i> PE=1 SV=1	0.80	0.90	0.82	0.30	5.23	4.84	0.07	0.04	0.01	5.66	5.16	
P02647	32	32	Apolipoprotein A-I OS-Homo sapiens GN- <i>APOA1</i> PE=1 SV=1	0.42	4.65	1.02	0.21	1.16	2.05	0.02	0.12	0.02	0.00	0.94	1.75
P02652	8	8	Apolipoprotein A-II OS-Homo sapiens GN- <i>AP002</i> PE=1 SV=1	0.04	0.17	0.13	0.03	0.21	0.19	0.01	0.07	0.00	0.00	0.17	0.23
P06727	41	41	Apolipoprotein A-IV OS-Homo sapiens GN- <i>AP003</i> PE=1 SV=3	0.15	0.63	0.91	0.12	1.07	0.86	0.03	0.15	0.07	0.01	0.38	0.72
P04114	155	131	Apolipoprotein C-I OS-Homo sapiens GN- <i>AP004</i> PE=1 SV=2	1.43	0.14	0.25	1.12	0.73	6.38	0.02	0.01	0.00	0.03	0.38	
P02643	2	2	Apolipoprotein C-II OS-Homo sapiens GN- <i>AP005</i> PE=1 SV=1	0.06	0.26	0.15	0.05	1.14	0.05	0.01	0.01	0.00	1.59	1.53	
P05090	8	7	Apolipoprotein C-III OS-Homo sapiens GN- <i>AP006</i> PE=1 SV=1	0.49	0.13	0.16	0.05	0.43	0.53	0.02	0.00	0.00	0.03	0.17	
P02649	28	27	Apolipoprotein E OS-Homo sapiens GN- <i>APOE</i> PE=1 SV=1	2.96	0.84	1.08	3.01	0.88	0.41	0.05	0.04	0.00	0.32	0.06	
P08519	12	10	Arginin N-methyltransferase OS-Homo sapiens GN- <i>CNN1</i> PE=1 SV=1	2.40	0.11	0.08	0.28	0.49	0.08	0.04	0.01	0.00	0.44	0.07	
P04030	14	11	Band 3 protein transmembrane protein OS-Homo sapiens GN- <i>CBP3</i> PE=1 SV=3	1.63	0.02	0.00	1.11	0.06	0.20	0.01	0.01	0.00	0.16	0.35	
P02749	21	20	Beta-2-glycoprotein B OS-Homo sapiens GN- <i>PGPB</i> PE=1 SV=3	0.83	4.30	0.88	1.46	0.86	4.40	0.09	0.13	0.26	0.01	5.69	0.57
P04003	19	17	Carboxypeptidase N catalytic chain OS-Homo sapiens GN- <i>CNP1</i> PE=1 SV=1	0.10	0.02	0.00	0.07	0.09	0.02	0.00	0.00	0.00	0.13	0.03	
P07106	5	5	Centrosome-associated protein OS-Homo sapiens GN- <i>CNP530</i> PE=1 SV=1	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.03	
P12098	29	29	Chitinase OS-Homo sapiens GN- <i>CHIT1</i> PE=1 SV=1	0.88	4.64	4.37	0.72	1.14	4.28	0.03	0.13	0.19	0.03	1.67	6.58
P00740	20	20	Coagulation factor IX OS-Homo sapiens GN- <i>F9</i> PE=1 SV=2	0.07	0.92	0.68	0.01	0.31	1.80	0.00	0.02	0.01	0.00	0.28	2.92
P12259	42	34	Coagulation factor V OS-Homo sapiens GN- <i>F5</i> PE=1 SV=2	0.16	1.19	0.06	0.23	0.21	7.11	0.01	0.02	0.01	0.00	0.14	11.55
P08709	6	5	Coagulation factor VII OS-Homo sapiens GN- <i>F7</i> PE=1 SV=1	0.05	0.44	0.26	0.08	0.14	0.01	0.01	0.03	0.00	0.09	0.01	
P04293	5	4	Coagulation factor VIII OS-Homo sapiens GN- <i>F8</i> PE=1 SV=2	0.01	0.03	0.00	0.05	0.00	0.00	0.00	0.00	0.00	0.00	0.03	
P02974	17	16	Coagulation factor IX OS-Homo sapiens GN- <i>F11</i> PE=1 SV=1	0.03	0.26	0.31	0.02	0.34	0.19	0.01	0.04	0.00	0.00	0.14	0.06
P00748	12	11	Coagulation factor XII OS-Homo sapiens GN- <i>F12</i> PE=1 SV=3	0.02	0.03	0.02	0.02	0.71	0.20	0.00	0.00	0.00	0.01	0.59	0.14
P07366	12	12	Complement C1r subcomponent OS-Homo sapiens GN- <i>C1R</i> PE=1 SV=2	0.03	0.09	0.04	0.06	0.58	3.09	0.00	0.07	0.01	0.00	0.42	4.64
P05981	7	4	Complement C1s subcomponent OS-Homo sapiens GN- <i>C1S</i> PE=1 SV=1	0.00	0.09	0.07	0.00	0.13	3.29	0.00	0.02	0.01	0.00	0.10	5.55
P06685	7	6	Complement C2 OS-Homo sapiens GN- <i>C2</i> PE=1 SV=1	0.00	0.10	0.00	0.02	0.00	0.33	0.00	0.02	0.00	0.00	0.17	
P00514	106	95	Complement C3 OS-Homo sapiens GN- <i>C3</i> PE=1 SV=2	4.05	0.32	0.07	4.42	0.37	3.71	0.01	0.03	0.00	0.02	0.87	0.22
P05024	65	3	Complement C4 A OS-Homo sapiens GN- <i>C4A</i> PE=1 SV=2	1.25	0.06	0.12	0.88	0.22	0.23	0.00	0.01	0.00	0.05	0.17	0.21
P05025	66	3	Complement C4 B OS-Homo sapiens GN- <i>C4B</i> PE=1 SV=2	1.07	0.05	0.11	0.75	0.19	0.20	0.00	0.01	0.00	0.05	0.18	
P13671	17	13	Complement component C6 OS-Homo sapiens GN- <i>C6</i> PE=1 SV=3	0.06	0.01	0.00	0.05	0.18	0.00	0.00	0.00	0.00	0.00	0.03	
P01425	10	9	Complement component C7 OS-Homo sapiens GN- <i>C7</i> PE=1 SV=3	0.28	0.16	0.00	0.05	0.25	0.52	0.00	0.01	0.00	0.00	0.01	0.51
P07354	11	10	Complement component C8 OS-Homo sapiens GN- <i>C8B</i> PE=1 SV=3	0.03	0.15	0.03	0.05	0.04	0.09	0.00	0.01	0.00	0.00	0.18	0.06
P02751	30	24	Complement component C9 OS-Homo sapiens GN- <i>C9</i> PE=1 SV=2	0.81	0.05	0.02	0.76	0.27	0.41	0.01	0.00	0.00	0.00	0.15	0.27
P06396	44	42	Complement factor B OS-Homo sapiens GN- <i>CFB</i> PE=1 SV=2	0.04	0.07	0.02	0.06	1.18	0.16	0.00	0.00	0.00	0.00	1.15	0.12
P07476	5	5	Complement factor D OS-Homo sapiens GN- <i>CDFD</i> PE=1 SV=5	0.01	0.02	0.01	0.00	0.22	0.00	0.00	0.00	0.00	0.20	0.01	
P00601_CD2980	65	59	Complement factor H-related protein C OS-Homo sapiens GN- <i>CFHR3</i> PE=1 SV=1	0.23	0.01	0.00	0.00	0.33	0.42	0.01	0.01	0.00	0.00	0.37	0.17
P05151	15	13	Complement factor H-related protein F OS-Homo sapiens GN- <i>CFHR1</i> PE=1 SV=2	0.13	0.29	0.06	0.01	0.59	0.33	0.00	0.01	0.00	0.01	1.96	0.30
P36698	8	7	Complement factor H-related protein G OS-Homo sapiens GN- <i>CFHR2</i> PE=1 SV=2	0.04	0.02	0.06	0.03	0.05	0.02	0.00	0.00	0.00	0.04	0.02	
P66995	2	2	Complement factor H-related protein H OS-Homo sapiens GN- <i>CFHR3</i> PE=1 SV=2	0.15	0.06	0.03	0.01	0.53	0.24	0.01	0.01	0.00	0.15	0.05	
42_P0210_P06	4	4	Complement factor H-related protein I OS-Homo sapiens GN- <i>CFHR4</i> PE=1 SV=1	0.26	0.35	0.33	0.00	0.69	0.31	0.00	0.01	0.00	0.16	0.04	
P02679	40	40	Complement factor I OS-Homo sapiens GN- <i>CFI</i> PE=1 SV=2	1.32	39.69	0.95	0.36	0.50	0.69	0.00	0.03	0.00	0.08	0.20	
P02751	30	24	Complement factor J OS-Homo sapiens GN- <i>CFJ</i> PE=1 SV=4	0.15	0.06	0.04	0.01	0.23	0.63	0.01	0.00	0.00	0.01	0.07	0.99
P06396	44	42	Gelsolin OS-Homo sapiens GN- <i>GMA</i> PE=1 SV=1	0.13	0.91	0.20	0.00	5.21	0.46	0.00	0.00	0.00	0.00	4.47	0.13
P14136	6	3	Glial fibrillary acidic protein OS-Homo sapiens GN- <i>GIFAP</i> PE=1 SV=1	0.19	0.01	0.04	0.11	0.20	0.36	0.04	0.01	0.00	0.04	0.24	0.33
P22352	2	2	Glutathione peroxidase-binding protein 1 OS-Homo sapiens GN- <i>GIPBP1</i> PE=1 SV=2	0.00	0.01	0.00	0.00	0.20	0.03	0.00	0.00	0.00	0.33	0.06	
P00383	7	6	Insulin-like growth factor-binding protein 1 OS-Homo sapiens GN- <i>IGFBP1</i> PE=1 SV=2	0.05	0.02	0.00	0.00	0.28	0.00	0.00	0.00	0.00	0.30	0.01	
P13823	21	19	Insulin-like growth factor-binding protein 1 OS-Homo sapiens GN- <i>IGFBP1</i> PE=1 SV=1	0.21	0.39	0.27	0.07	0.93	0.00	0.00	0.01	0.00	0.75	0.34	
P01424	50	50	Ig gamma 1 chain C region OS-Homo sapiens GN- <i>IGHG2</i> PE=1 SV=2	2.92	0.78	0.27	0.50	0.10	0.00	0.01	0.04	0.00	0.29	0.57	
P01042	29	27	Ig gamma 1 chain C region OS-Homo sapiens GN- <i>IGHG3</i> PE=1 SV=2	8.13	0.47	1.79	12.17	0.46	0.69	0.31	0.02	0.13	0.21	0.42	
P18428	10	9	Ig kappa 2 chain C region OS-Homo sapiens GN- <i>IGHG4</i> PE=1 SV=1	0.30	0.18	0.01	0.01	0.22	0.09	0.04	0.00	0.01	0.05	0.06	
P07355	5	3	Ig kappa 2 chain C region OS-Homo sapiens GN- <i>IGHM</i> PE=1 SV=3	0.06	0.05	0.01	0.00	0.32	0.00	0.00	0.00	0.00	0.08	0.01	
P15284	17	15	Immunoglobulin heavy variable 3d 6S OS-Homo sapiens GN- <i>IGHV4 54</i> PE=1 SV=1	2.04	0.16	0.06	0.12	1.73	0.43	0.06	0.02	0.01	0.04	2.13	
P36955	14	13	Proteoglycan 4 OS-Homo sapiens GN- <i>SPN101</i> PE=1 SV=1	0.43	0.04	0.06	0.52	0.19	0.30	0.04	0.00	0.00	0.25	0.30	
P00734	46	43	Proteinase inhibitor 1 OS-Homo sapiens GN- <i>PRIN1</i> PE=1 SV=2	0.05	0.02	0.01	0.02	2.26	3.56	0.32	0.01	0.01	0.02	2.80	3.15
P04050	5														

Table S2. List of all identified corona proteins on PS nanoparticles coated with (PS)₄ and S₈.

Accession	Peptide count/unique peptide	Description	Average in %						Standard deviation					
			Plasma Uncoated			Serum			Plasma Uncoated			Serum		
			(PS)4	S8	Uncoated	(PS)4	S8	Uncoated	(PS)4	S8	Uncoated	(PS)4	S8	Uncoated
6/PA021/PS2671P	9	Actin, actin, smooth muscle OS-Homo sapiens GN-ACTA2 PE=1 SV=1	0.81	0.17	0.30	0.24	0.14	0.35	0.03	0.08	0.10	0.02	0.05	0.04
O709/PS2621OS052	13	Actin, cytoplasmic 1 OS-Homo sapiens GN-ACTB PE=1 SV=1	0.95	1.47	0.57	0.33	0.80	1.08	0.02	0.01	0.02	0.06	0.02	0.04
P02763/PS10652	4	Alpha-1-acid glycoprotein 1 OS-Homo sapiens GN-ORRM1 PE=1 SV=1	0.02	0.05	0.01	0.02	0.03	0.03	0.00	0.01	0.00	0.00	0.00	0.00
:Q57W2/PS0VUR7	3	Ankyrin repeat domain-containing protein 1 202A OS-Homo sapiens GN-ANKR2D0A2 PE=4 SV=1	0.08	0.08	0.05	0.06	0.08	0.09	0.00	0.01	0.00	0.00	0.01	0.07
P01008	32	Antithrombin-III OS-Homo sapiens GN-SERPINC1 PE=1 SV=1	0.02	0.05	0.02	0.70	1.58	2.72	0.00	0.02	0.00	0.07	0.03	1.54
P02647	25	Apolipoprotein A-I OS-Homo sapiens GN-APOA1 PE=1 SV=1	1.49	1.47	2.33	3.24	1.84	4.16	0.00	0.15	0.15	0.02	0.07	0.86
P02652	2	Apolipoprotein A-II OS-Homo sapiens GN-APOA2 PE=1 SV=1	0.16	0.02	0.09	0.04	0.03	0.03	0.00	0.03	0.04	0.00	0.01	0.01
P06727	28	Apolipoprotein A-V OS-Homo sapiens GN-APOA4 PE=1 SV=3	1.39	0.69	0.67	0.33	1.43	0.57	0.04	0.12	0.03	0.16	0.07	0.46
P04114	40	Apolipoprotein B-100 OS-Homo sapiens GN-APOB PE=1 SV=1	0.12	0.12	0.09	0.08	0.10	0.18	0.01	0.01	0.00	0.05	0.00	0.10
P02649	20	Apolipoprotein E OS-Homo sapiens GN-APOE PE=1 SV=1	6.19	3.34	4.11	2.05	0.71	1.52	0.39	0.28	0.06	0.07	0.01	0.31
Q9647	3	Arf-GAP with GTG-Pase, Arf-GAP with GTG-Pase containing protein 3 OS-Homo sapiens GN-AGAP3 PE=1 SV=2	0.14	1.20	0.10	0.13	0.13	0.18	0.00	0.00	0.00	0.00	0.00	0.1
Q9357	2	BAG family member 2 OS-Homo sapiens GN-BAG2 PE=1 SV=3	0.00	0.00	0.02	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
P02730	8	Band 3 anion transport protein OS-Homo sapiens GN-CLCA4 PE=1 SV=3	0.00	0.03	0.04	0.02	0.10	0.05	0.00	0.02	0.00	0.00	0.00	0.00
P02749	15	Beta-2-glycoprotein 1 OS-Homo sapiens GN-APOH PE=1 SV=3	1.61	0.98	0.64	0.21	1.41	3.49	0.95	0.07	0.04	0.00	0.03	0.42
Q56291	8	Beta-actin-like protein 2 OS-Homo sapiens GN-ACBL2 P1 PE=1 SV=2	0.22	1.04	1.34	0.07	0.06	0.04	0.05	0.01	0.04	0.06	0.02	0.03
Q9379	3	Biotin-acyl-CoA synthetase OS-Homo sapiens GN-CHBS PE=1 SV=1	0.03	0.09	0.05	0.02	0.31	0.07	0.00	0.02	0.00	0.00	0.02	0.00
P06723	2	Broad-complex transcriptional regulator 1 OS-Homo sapiens GN-BCOR1 P1 PE=1 SV=2	1.35	0.26	0.40	0.09	0.18	0.40	0.05	0.01	0.02	0.41	0.07	0.27
P04003	18	C4b-binding protein alpha chain OS-Homo sapiens GN-C4BPB1 PE=1 SV=2	0.38	0.52	0.30	0.29	0.08	0.09	0.10	0.02	0.01	0.09	0.03	0.02
Q9614	3	Carboxypeptidase B2 OS-Homo sapiens GN-CPBP2 PE=1 SV=2	0.00	0.00	0.00	0.02	0.11	0.05	0.00	0.00	0.00	0.01	0.00	0.02
P10909	28	Clustering OS-Homo sapiens GN-CLU PE=1 SV=1	11.74	5.90	9.09	14.63	14.37	10.69	0.23	0.26	0.09	0.84	0.18	2.05
P00740	7	Cogulation factor V OS-Homo sapiens GN-CAFV PE=1 SV=2	0.19	0.18	0.09	0.09	0.14	0.11	0.01	0.02	0.00	0.00	0.00	0.00
P12259	28	Cogulation factor X OS-Homo sapiens GN-CAFX PE=1 SV=2	0.15	0.30	0.33	0.34	0.16	0.34	0.01	0.01	0.01	0.05	0.00	0.13
P02051	10	Complement C1 component OS-Homo sapiens GN-CC1P1 PE=1 SV=1	0.14	0.34	0.16	0.13	0.19	0.17	0.00	0.01	0.00	0.01	0.00	0.12
P02747	4	Complement Clq subcomponent OS-Homo sapiens GN-CC1QC PE=1 SV=3	0.05	0.04	0.04	0.07	0.17	0.05	0.00	0.01	0.01	0.01	0.01	0.04
P00736	16	Complement C1s subcomponent OS-Homo sapiens GN-CC1R1 PE=1 SV=2	0.02	0.04	0.06	0.06	0.29	0.11	0.00	0.00	0.00	0.01	0.00	0.01
P09871	3	Complement C1s subcomponent OS-Homo sapiens GN-CC1R1 P1 PE=1 SV=1	0.05	0.14	0.13	0.07	0.17	0.08	0.00	0.00	0.01	0.01	0.00	0.05
P01022	107	Complement C3 OS-Homo sapiens GN-CC3 PE=1 SV=2	1.01	1.13	0.74	0.41	0.81	0.00	0.00	0.00	0.00	0.00	0.00	0.00
P00044	45	Complement C4-A OS-Homo sapiens GN-CC4A PE=1 SV=2	0.01	0.02	0.01	0.14	0.18	0.11	0.00	0.00	0.00	0.04	0.04	0.00
P00505	46	Complement C4-B OS-Homo sapiens GN-CC4B PE=1 SV=2	0.07	1.03	0.66	0.39	2.26	1.58	0.02	0.06	0.02	0.04	0.25	0.46
P13671	8	Complement component C6 OS-Homo sapiens GN-CC6 PE=1 SV=3	0.08	0.15	0.20	0.08	0.53	0.22	0.02	0.08	0.03	0.03	0.06	0.06
P10643	5	Complement component C7 OS-Homo sapiens GN-CC7 PE=1 SV=2	0.00	0.01	0.02	0.00	0.08	0.00	0.00	0.00	0.00	0.01	0.00	0.02
P02748	11	Complement component C8 OS-Homo sapiens GN-CC8 PE=1 SV=2	0.07	0.12	0.13	0.09	0.38	0.22	0.00	0.00	0.01	0.00	0.02	0.10
P00703	2	Complement factor B OS-Homo sapiens GN-CCB1 PE=1 SV=2	0.00	0.00	0.02	0.01	0.11	0.03	0.00	0.01	0.00	0.01	0.00	0.00
P08613/PS03985	47	Complement factor H OS-Homo sapiens GN-CHP1 PE=1 SV=4	0.06	0.35	0.13	0.10	0.52	0.19	0.01	0.05	0.00	0.01	0.02	0.12
Q03591/PS36980	11	Complement factor H-related protein 1 OS-Homo sapiens GN-CHRH1 PE=1 SV=2	0.01	0.28	0.73	0.06	0.78	0.58	0.00	0.04	0.07	0.03	0.03	0.03
Q98X6	13	Complement factor H-related protein 1 OS-Homo sapiens GN-CHRH1 PE=1 SV=1	0.13	0.11	0.15	0.06	0.29	0.19	0.01	0.00	0.00	0.02	0.00	0.05
O16555	3	Dihydroxyacetone-phosphate reductase OS-Homo sapiens GN-DHAR1 PE=1 SV=2	0.24	0.11	0.16	0.12	0.24	0.15	0.01	0.00	0.00	0.00	0.00	0.02
P02749	2	DnaJ homolog subfamily 8 member 1 OS-Homo sapiens GN-DNAJ8 P1 PE=1 SV=2	3.06	0.09	0.07	2.85	0.53	0.39	0.03	0.02	2.13	0.04	0.03	0.02
Q87502	2	Elongator complex alpha chain OS-Homo sapiens GN-CPFA1 PE=1 SV=2	0.37	0.46	0.20	0.32	0.36	0.49	0.01	0.02	0.01	0.01	0.01	0.42
P02671	43	Fibrinogen alpha chain OS-Homo sapiens GN-FGB PE=1 SV=2	0.72	7.17	4.80	0.22	0.16	0.21	0.04	0.77	0.15	0.13	0.03	0.15
P02675	48	Fibrinogen beta chain OS-Homo sapiens GN-FGBP PE=1 SV=2	1.25	15.17	12.27	0.08	0.10	0.07	0.09	0.21	0.13	0.00	0.01	0.48
P02679	37	Fibrinogen gamma chain OS-Homo sapiens GN-FGGP PE=1 SV=3	1.15	11.15	9.06	0.06	0.21	0.07	0.04	0.87	0.18	0.00	0.02	0.02
P02749	2	Genetic oligomeric glucose-regulated protein OS-Homo sapiens GN-GORP1 PE=1 SV=2	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
P02750	27	Genomic OS-Homo sapiens GN-GN P1 PE=1 SV=1	0.29	0.28	0.27	0.21	0.58	0.30	0.08	0.07	0.00	0.13	0.02	0.16
P14136	8	Gial fibrillary acidic protein OS-Homo sapiens GN-GFAP1 PE=1 SV=1	0.23	0.14	0.12	0.22	0.09	0.23	0.01	0.01	0.00	0.00	0.01	0.01
P22352	4	Glutathione peroxidase 1 OS-Homo sapiens GN-GPX1 PE=1 SV=2	0.01	0.04	0.07	0.02	0.18	0.05	0.00	0.01	0.00	0.00	0.00	0.01
P04046	4	Gluceraldehyde-3-phosphate dehydrogenase OS-Homo sapiens GN-GAPDH PE=1 SV=3	0.01	0.05	0.02	0.04	0.09	0.05	0.00	0.00	0.00	0.00	0.01	0.04
P00078	6	Haptoglobin-related protein OS-Homo sapiens GN-HPR1 PE=1 SV=2	0.05	0.04	0.02	0.05	0.04	0.03	0.00	0.00	0.00	0.00	0.00	0.02
P00079	4	Hemoglobin subunit alpha OS-Homo sapiens GN-HBBA1 PE=1 SV=2	0.00	0.01	0.01	0.13	0.15	0.06	0.00	0.00	0.00	0.00	0.00	0.00
P00070	5	Hemoglobin subunit beta OS-Homo sapiens GN-HBBP1 PE=1 SV=2	0.00	0.16	0.20	0.04	0.14	0.16	0.00	0.04	0.01	0.00	0.00	0.02
P00071	11	Heme oxygenase 1 chain C region OS-Homo sapiens GN-HO1 PE=1 SV=1	0.00	0.16	0.29	0.03	0.06	0.00	0.00	0.00	0.00	0.00	0.00	0.05
P01876	7	Hspalpha 1 chain C region OS-Homo sapiens GN-HSPH1 P1 PE=1 SV=2	0.16	0.18	0.14	0.13	0.18	0.07	0.01	0.02	0.01	0.00	0.00	0.05
P16424	59	Hspalpha 1 chain C region OS-Homo sapiens GN-HSPH1 PE=1 SV=1	0.00	0.16	0.25	0.09	0.35	0.33	0.01	0.03	0.04	0.02	0.04	0.08
P01875	15	Ig alpha 1 chain C region OS-Homo sapiens GN-IGHA1 PE=1 SV=2	0.49	0.77	0.48	0.94	1.50	0.84	0.02	0.03	0.01	0.06	0.01	0.67
P01859	11	Ig gamma 2 chain C region OS-Homo sapiens GN-IGHG2 P1 PE=1 SV=2	3.27	3.84	3.88	3.65	2.68	6.16	0.22	0.07	0.04	0.18	0.22	1.31
P01860	14	Ig gamma 3 chain C region OS-Homo sapiens GN-IGHG3 P1 PE=1 SV=2	0.00	0.05	0.06	0.05	0.13	0.05	0.00	0.00	0.00	0.00	0.00	0.03
P00334	7	Ig kappa chain C region OS-Homo sapiens GN-IGKC P1 PE=1 SV=1	1.29	1.32	1.14	1.05	1.20	1.20	0.04	0.08	0.06	0.24	0.06	0.05
P00374	7	Ig lambda 2 chain C region OS-Homo sapiens GN-IGLC2 P1 PE=1 SV=1	0.09	0.18	0.25	0.15	0.44	0.32	0.02	0.01	0.02	0.04	0.07	0.02
P03871	16	Ig mu chain C region OS-Homo sapiens GN-IGM1 PE=1 SV=3	0.37	0.29	0.22	0.49	0.56	0.30	0.01	0.01	0.01	0.03	0.03	0.10
P04220	11	Ig mu heavy chain disease protein OS-Homo sapiens PE-1 SV=1	0.04	0.06	0.09	0.06	0.02	0.05	0.00	0.00	0.00	0.00	0.00	0.01
P00602	3	Immunoglobulin E receptor OS-Homo sapiens GN-IEPE1 PE=1 SV=2	0.05	0.08	0.06	0.13	0.47	0.18	0.01	0.01	0.01	0.01	0.01	0.06
P05354	7	Immunoglobulin E receptor OS-Homo sapiens GN-IEPE1 P1 PE=1 SV=2	0.00	0.03	0.01	0.02	0.17	0.08	0.00	0.01	0.01	0.00	0.01	0.00
P00747	14	Inter-alpha-trypsin inhibitor heavy chain 4 OS-Homo sapiens GN-ITIH4 P1 PE=1 SV=4	1.73	5.20	4.46	0.77	8.28	7.51	0.04	0.44	0.18	0.08	0.54	1.54
P02770	4	Inter-alpha-trypsin inhibitor heavy chain 4 OS-Homo sapiens GN-ITIH4 PE=1 SV=3	0.22	0.16	0.22	0.42	1.61	0.77	0.01	0.00	0.01	0.06	0.17	0.01
P07737	2	Ion channel-interacting protein 2 OS-Homo sapiens GN-IPIP2 PE=1 SV=1	0.23	0.14	0.33	0.21	0.39	0.26	0.01	0.00	0.01	0.02	0.03	0.01
P12259</td														

Table S3. List of all identified corona proteins on lipo-(PS)₄ in human plasma applying different dendrimer amounts.

Accession	Peptide count	Unique peptides	Description	Average in %						Standard deviation						
				Amount Dendrimer [mg]			n=3			Amount Dendrimer [mg]			n=3			
				0.5	0.25	0.1	0.5	0.25	0.1	0.5	0.25	0.1	0.5	0.25	0.1	
P02790	8	8	Band 3 anion transport protein OS=Homo sapiens GN=SLC4A1 PE=1 SV=3	0.01680706	0.02324408	0.0550601	0.1223086	0.00149152	0.00123881	0.00380245	0.00497245					
Q93696	3	3	Geranylgeranyl transferase type-2 subunit alpha OS=Homo sapiens GN=RABGTTA PE=1 SV=2	0.71593041	0.11633758	0.11090014	0.08070386	0.02537714	0.00809266	0.01039344	0.00631582					
P04004	21	20	Vitronectin OS=Homo sapiens GN=VTN PE=1 SV=1	2.61623035	4.49555647	0.8432212	0.10503001	0.19632669	0.3339972	0.1113526	0.0280207					
P02671	55	51	Fibrinogen alpha chain OS=Homo sapiens GN=FGA PE=1 SV=2	12.6128433	11.04928687	7.42876573	6.3568448	0.1857229	0.16358085	0.01820355	0.10160731					
P08697	12	12	Alpha 2-antiplasmin OS=Homo sapiens GN=SERPINF2 PE=1 SV=3	1.46620385	1.55898519	0.60703084	0.53672887	0.04204691	0.04064677	0.02526979	0.02191196					
P01859	6	2	'lg gamma-2 chain C region OS=Homo sapiens GN=IGH2B PE=1 SV=2	0.0570516	0.07610793	0.12606853	0.1602325	0.00217081	0.0022423	0.00428111						
P22352	3	3	Glutathione peroxidase 3 OS=Homo sapiens GN=GPRX3 PE=1 SV=2	0.03743359	0.12336943	0.00373591	0.0221532	0.00275734	0.00414055							
P01023	18	17	Alpha 2-macroglobulin OS=Homo sapiens GN=A2M PE=1 SV=3	0.08735203	0.05376149	0.0982822	0.11998797	0.00049738	0.00189113	0.00436343	0.00638202					
P02751	60	52	Fibrinectin OS=Homo sapiens GN=FNI PE=1 SV=4	0.57868631	0.25293768	0.05059392	0.06630739	0.006630739	0.00673793	0.01738048						
Q98XR6	14	12	Complement factor H-related protein 5 OS=Homo sapiens GN=CFHR5 PE=1 SV=1	0.07299998	0.09718554	0.16110797	0.17446556	0.00630946	0.0354998	0.0055798	0.00840534					
P00738	16	9	Haptoglobin OS=Homo sapiens GN=HP PE=1 SV=1	0.16270924	0.22484115	0.26720294	0.26720294	0.00622559	0.0805067	0.01556167	0.00804511					
P02765	14	13	Alpha-3-HS-glycoprotein OS=Homo sapiens GN=AHSG PE=1 SV=1	2.41049562	1.47697963	1.60167713	1.06861451	0.05941193	0.13424621	0.06836366	0.02353633					
Q99969	3	3	Retinoic acid receptor responder protein 2 OS=Homo sapiens GN=RARE2 PE=1 SV=1	0.01025176	0.01581746	0.05181746	0.00012223	0.00060709	0.00152326	0.00517599						
P01019	8	8	Angiotensinogen OS=Homo sapiens GN=AGT PE=1 SV=1	0.20665355	0.16854789	0.1308342	0.1127002	0.0104055	0.00638945	0.00615027	0.00149482					
Q72501	2	2	Cytoplasmic polyadenylation element-binding protein 2 OS=Homo sapiens GN=CPEB2 PE=2 SV=3	0.03024327	0.05175592	0.08467073	0.08268388	0.00123022	0.00486943	0.0248735						
P03950	2	2	Angiogenin OS=Homo sapiens GN=ANG PE=1 SV=1	0.03457441	0.05053631	0.12948308	0.15297768	0.00353075	0.0353602	0.00213504	0.00645584					
P01834	5	5	Ig kappa Chain C region OS=Homo sapiens GN=IGKC PE=1 SV=1	0.81092834	0.1071255	0.17417849	0.16678958	0.00522793	0.10910821	0.04479627	0.04105914					
P02747	7	7	Complement C1q subcomponent C1 OS=Homo sapiens GN=C1QC PE=1 SV=3	0.37687082	0.22735333	0.119945107	0.19898215	0.00484563	0.19454451	0.00978533	0.00621545					
Q14624	52	50	Inter-alpha-trypsin inhibitor heavy chain H4 OS=Homo sapiens GN=ITIH4 PE=1 SV=4	1.18472059	0.22993392	0.21598874	0.02708899	0.0674759	0.1252438	0.10620918						
P02766	11	11	Transferrin OS=Homo sapiens GN=TRT PE=1 SV=1	0.1439157	0.25215136	0.3675698	0.38807946	0.00481693	0.17176171	0.00278445	0.01197111					
P04003	17	15	C4b protein chain OS=Homo sapiens GN=CBPA4 PE=1 SV=2	0.15859543	0.12517593	0.24679932	0.28744553	0.00471119	0.11503804	0.00838267	0.01827057					
P04196	20	19	Histidine-rich glycoprotein OS=Homo sapiens GN=Hrg PE=1 SV=1	1.5098255	2.08719334	2.4169188	2.35625971	0.0515938	0.19736139	0.05345164	0.0651156					
P19827	12	12	Inter-alpha-trypsin inhibitor heavy chain H1 OS=Homo sapiens GN=ITIH1 PE=1 SV=3	0.16303914	0.13708334	0.30321668	0.30346427	0.00387944	0.0803767	0.00432455	0.01333936					
Q03591	16	3	Complement factor H-related protein 3 OS=Homo sapiens GN=CFHR1 PE=1 SV=2	0.47170624	0.06416013	0.00039973	0.10078835	0.002589545	0.06323134	0.01735261	0.0598228					
P10909	26	26	Clusterin OS=Homo sapiens GN=CLU PE=1 SV=1	1.70210362	2.40842967	0.45258286	0.48551293	0.1719434	0.15604558	0.066049	0.0795851					
P06396	42	40	Gelsolin OS=Homo sapiens GN=GSM PE=1 SV=1	2.48904078	0.08576336	0.49717086	0.29573167	0.0895163	0.15705022	0.07228916	0.04721548					
P18428	13	13	Lipoprotein OS=Homo sapiens GN=LGP PE=1 SV=3	0.17653499	0.25170531	0.27685887	0.28517126	0.0285130	0.00847545	0.00831553						
P01876;P01877	8	8	lg alpha 1 chain C region OS=Homo sapiens GN=IGH1A PE=1 SV=2	0.13359181	0.13239058	0.17865486	0.16771346	0.01134741	0.01088698	0.00880662						
P00747	60	53	Plasminogen OS=Homo sapiens GN=PLG PE=1 SV=2	2.83835469	0.27704672	1.87983331	1.8198172	0.08305253	0.23170924	0.0464916	0.04196102					
P04756	4	3	Hepatocyte growth factor activator OS=Homo sapiens GN=HGFA PE=1 SV=1	0.05168923	0.07433913	0.13766185	0.16119391	0.01415156	0.05050551	0.00628391	0.00550703					
P01008	21	20	Antithrombin-III OS=Homo sapiens GN=SERPINC1 PE=1 SV=1	0.30242272	0.26526182	0.21636288	0.18974942	0.01467999	0.1246892	0.00793888	0.00695791					
P03952	21	20	Plasma kallikrein OS=Homo sapiens GN=KLK1B PE=1 SV=1	1.3806749	0.18585173	0.28932841	0.38325441	0.00535319	0.02069694	0.03866093	0.02648168					
P02774	39	36	Vitamin D-binding protein OS=Homo sapiens GN=GC PE=1 SV=1	1.7244466	1.65911354	1.29967688	0.99266404	0.04602358	0.3991976	0.11932767	0.0414988					
P15169	2	2	Carboxypeptidase N catalytic chain OS=Homo sapiens GN=CNP1 PE=1 SV=2	0.1819817	0.1775688	0.26782649	0.26782649	0.00336966	0.1512492	0.00409354	0.011673					
P02649	24	22	Apolipoprotein E OS=Homo sapiens GN=APE PE=1 SV=2	0.9238183	1.7168897	1.05656929	0.31591665	0.0440916	0.03981678	0.03404351						
P04220	11	2	lg mu heavy chain disease protein OS=Homo sapiens GN=COMP PE=1 SV=1	0.38937698	0.08793439	0.1223758	0.16771346	0.0085120	0.01024769	0.00880662						
P05452	7	6	Tetranectin OS=Homo sapiens GN=CLEC3B PE=1 SV=2	0.08870501	0.05586580	0.05023185	0.04093707	0.00205018	0.00132219	0.0358967						
P05154	13	13	Plasma serine protease inhibitor OS=Homo sapiens GN=SERPINAS PE=1 SV=3	0.41714988	0.18931502	0.13766185	0.16120391	0.01415156	0.05050551	0.00628391	0.00550703					
P01871	12	12	Serum alpha-1-proteinase inhibitor OS=Homo sapiens GN=IGH4 PE=1 SV=1	0.61074171	0.52635612	0.44995973	0.4319126	0.00872013	0.06000401	0.00406104	0.00914983					
P02675	58	53	Fibrinogen beta chain OS=Homo sapiens GN=FGB PE=1 SV=2	9.44596863	9.62658883	6.67545533	6.53161727	0.08206924	0.13453753	0.04585982	0.10025157					
P25858	7	7	Insulin-like growth-factor-binding protein complex alpha-lariat subunit OS=Homo sapiens GN=IGFALS PE=1 SV=1	0.07055625	0.05059392	0.05387387	0.05050882	0.00260214	0.03044018	0.00265697	0.00282788					
P49747	4	4	Cartilage oligomeric matrix protein OS=Homo sapiens GN=COMP PE=1 SV=2	0.4711	0.50474744	0.6905577	0.74800734	0.00426694	0.07047451	0.0485716						
P02776;P10720	3	3	Platelet factor 4 OS=Homo sapiens GN=Pf4 PE=1 SV=2	0.5950421	0.22776944	0.28817788	0.34631335	0.03197061	0.1852544	0.04040158	0.00633333					
P06727	31	31	Apolipoprotein IV-A OS=Homo sapiens GN=APOA4 PE=1 SV=3	0.84393857	1.22004209	0.10134462	0.2156368	0.05113985	0.11554096	0.07785851	0.01724384					
P07360	4	4	Complement component C8 OS=Homo sapiens GN=C8B PE=1 SV=3	0.03295717	0.03575861	0.05561421	0.0022776	0.00127755	0.00426623	0.022239						
P08575	9	9	lg gamma-1 chain C region OS=Homo sapiens GN=IGH4 PE=1 SV=1	0.32405394	0.4079448	0.43769948	0.41567138	0.01215993	0.1961741	0.01944907	0.01809518					
P13671	30	29	Complement component C6 OS=Homo sapiens GN=C6B PE=1 SV=3	0.47446176	0.15428497	0.49221261	0.53815045	0.01300008	0.01282204	0.00767384	0.01061598					
P15113	10	9	Serocollagenase enhancer 1 OS=Homo sapiens GN=PCOLCE PE=1 SV=2	1.08006848	0.58380694	0.60861961	0.64086642	0.01306745	0.02277837	0.00463202	0.00469559					
P07920	17	16	Hemopexin OS=Homo sapiens GN=HPX PE=1 SV=2	17.44946	16.703128	13.2144823	11.6938728	0.45366663	0.92647455	0.00372778	0.01755048					
P05155	16	16	Plasmin protease inhibitor OS=Homo sapiens GN=CD5L PE=1 SV=1	0.16703356	0.12390013	0.09978223	0.10481417	0.00303187	0.01442929	0.00054261	0.0022166					
P02652	6	6	Apolipoprotein A-II OS=Homo sapiens GN=APOA2 PE=1 SV=1	0.34234117	0.125251624	0.24063297	0.21269458	0.0113327	0.017813	0.01948378	0.01301247					
P29622	4	4	Kallistatin OS=Homo sapiens GN=APOLI PE=1 SV=5	0.08210583	0.04868062	0.06475611	0.04067371	0.00129711	0.00205082	0.01427367	0.01219571					
P10643	10	9	Protein C8 OS=Homo sapiens GN=C8B PE=1 SV=3	0.12368874	0.1173945	0.13533231	0.16401935	0.00484725	0.01150353	0.00568691	0.00570464					
P01861	9	4	lg gamma-4 chain C region OS=Homo sapiens GN=C8B PE=1 SV=1</													

References

- 1 R. Stangenberg, Y. Wu, J. Hedrich, D. Kurzbach, D. Wehner, G. Weidinger, S. L. Kuan, M. I. Jansen, F. Jelezko, H. J. Luhmann, D. Hinderberger, T. Weil and K. Müllen, *Adv. Healthcare Mater.*, 2015, **4**, 377.
- 2 R. Stangenberg, I. Saeed, S. L. Kuan, M. Baumgarten, T. Weil, M. Klapper and K. Müllen, *Macromol. Rapid Commun.*, 2014, **35**, 152.
- 3 J. Wagner, L. Li, J. Simon, L. Krutzke, K. Landfester, V. Mailänder, K. Müllen, D. Y. W. Ng, Y. Wu and T. Weil, *Angew. Chem. Int. Ed.*, 2020, **59**, 5712.
- 4 S. Bernhardt, M. Kastler, V. Enkelmann, M. Baumgarten and K. Müllen, *Chem. Eur. J.*, 2006, **12**, 6117.
- 5 M. Gai, J. Simon, I. Lieberwirth, V. Mailänder, S. Morsbach and K. Landfester, *Polym. Chem.*, 2020, **11**, 527.
- 6 S. Lerch, M. Dass, A. Musyanovych, K. Landfester and V. Mailänder, *Eur. J. Pharm. Biopharm.*, 2013, **84**, 265; D. Baumann, D. Hofmann, S. Nullmeier, P. Panther, C. Dietze, A. Musyanovych, S. Ritz, K. Landfester and V. Mailänder, *Nanomedicine*, 2013, **8**, 699.
- 7 J. Simon, T. Wolf, K. Klein, K. Landfester, F. R. Wurm and V. Mailänder, *Angew. Chem. Int. Ed.*, 2018, **57**, 5548.
- 8 S. Schöttler, G. Becker, S. Winzen, T. Steinbach, K. Mohr, K. Landfester, V. Mailänder and F. R. Wurm, *Nat. Nanotechnol.*, 2016, **11**, 372; M. Kokkinopoulou, J. Simon, K. Landfester, V. Mailänder and I. Lieberwirth, *Nanoscale*, 2017, **9**, 8858.
- 9 J. C. Silva, M. V. Gorenstein, G.-Z. Li, J. P. C. Vissers and S. J. Geromanos, *Mol. Cell. Proteomics*, 2006, **5**, 144.