Supporting Information

For

N-Heterocyclic Silylene/Germylene Ligands in Au(I) Catalysis

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S1. Experimental Details

All experiments were carried out under an atmosphere of dry argon or in vaccuo using standard Schlenk technique and in a dinitrogen filled MBRAUN MB 150-G1 glovebox. The solvents used were purified by MBRAUN solvent purification system MB SPS-800. The starting materials **1** and **4** were prepared as reported in the literature. All other chemicals purchased from Aldrich were used without further purification. ¹H, ¹³C, ³¹P, ¹⁹F and ²⁹Si NMR spectra were recorded in Bruker 400 MHz spectrometer, using CDCl₃ as solvent with external standards SiMe₄ (¹H, ¹³C and ²⁹Si), 85% H₃PO₄ (³¹P) and CHF₃ (¹⁹F). Concentrated solution of the samples in CDCl₃ were sealed off in a NMR tube for measurement. Mass spectra were recorded using AB Sciex, 4800 plus MALDI TOF/TOF and HRMS ESI-TOF mass analyzer.

Synthesis of 2:

AgSbF₆ (0.171g, 0.5 mmol) was dissolved in DCM and added to the solution of **1** (0.325g, 0.5 mmol) in benzene. Immediately AgCl was precipitated out. After overnight stirring, AgCl was separated out from the reaction mixture by filtration and reduced the volume to 15 mL and kept it at 0°C. Colorless hexagonal shaped crystals suitable for X-ray analysis was observed after one day. Yield: 0.354g (75%). Mp: 145°C (decomposed). ¹H NMR (400.31 MHz, CDCl₃): δ 0.28 (*s*, 9H, Si*Me*₃), 0.43 (*s*, 9H, Si*Me*₃), 1.18 (*s*, 18H, C*Me*₃), 7.23-7.30 (*m*, 1H, Ph), 7.35-7.44 (*m*, 2H, Ph), 7.48 (*s*, 6H, C₆H₆) 7.52-7.69 (*m*, 2H, Ph) ppm. ¹³C NMR {¹H} (100.67 MHz, CDCl₃): δ 3.61 (Si*Me*₃), 5.38 (Si*Me*₃), 30.65 (C*Me*₃), 54.40 (CMe₃), 126.06, 126.24, 127.20, 127.39, 127.99, 128.68, 130.59 (Ph-C), 172.45 (NCN) ppm. ²⁹Si{¹H} NMR (79.495 MHz, CDCl₃, 298 K): δ 9.45 (*Si*Me₃), 8.39 (*Si*Me₃), -6.44 (*Si*N(SiMe₃)₂) ppm. MALDI: *m*/*z* [C₂₇H₄₇N₃Si₃Au]⁺: 616.05 [M-C₆H₆]⁺. Anal Calcd: C, 35.56; H, 5.33; N, 4.44. Found: C, 35.23; H, 5.15; N, 4.57.

Synthesis of 3:

AgSbF₆ (0.171g, 0.5 mmol) was dissolved in DCM and added to the solution of 1 (0.350g, 0.5 mmol) in *m*-xylene. Immediately AgCl was precipitated out. After overnight stirring, AgCl was separated out from the reaction mixture by filtration and reduced the volume to 15 mL and kept it at 0°C. Colorless block shaped crystals suitable for X-ray analysis was observed after one day. Yield: 0.329g (68%). Mp: 148°C (decomposed). ¹H NMR (400.31 MHz, CDCl₃): δ 0.25 (*s*, 9H, Si*Me*₃), 0.35 (*s*, 9H, Si*Me*₃), 1.14 (*s*, 18H, C*Me*₃), 2.40 (*s*, 6H, C₆H₄(C*H*₃)₂, 7.21-7.30 (*m*, 4H,

Ph), 7.35-7.45 (*m*, 3H, Ph), 7.47-7.54 (*s*, 1H, C₆*H*₆) 7.57-7.64 (*m*, 1H, Ph) ppm. ¹³C{¹H} NMR (100.67 MHz, CDCl₃): δ 3.39 (Si*Me*₃), 4.93 (Si*Me*₃), 20.47 (C₆H₄(CH₃)₂), 30.40 (C*Me*₃), 54.24 (CMe₃), 122.14, 125.89, 125.99, 127.08, 127.52, 128.12, 130.12, 130.40, (Ph-C), 172.38 (NCN) ppm. ²⁹Si{¹H} NMR (79.495 MHz, CDCl₃, 298 K): δ 9.34 (*Si*Me₃), 8.47 (*Si*Me₃), -8.85 (*Si*N(SiMe₃)₂). MALDI: *m/z* [C₂₇H₄₇N₃Si₃Au]⁺: 616.05 [M-C₆H₄(CH₃)₂]⁺. Anal Calcd: C, 37.00; H, 5.59; N, 4.32. Found: C, 36.73; H, 5.42; N, 4.40.

Synthesis of 4: 30 mL toluene was added into the mixture of 0.232 g (0.5 mmol) of germylene [{PhC(N*t*Bu)₂}Ge{N(SiMe₃)₂}] and 0.147 g (0.5 mmol) of AuCl.SMe₂ in a 100 mL schlenk flask. After overnight stirring at room temperature, the reaction mixture was dried completely and filtered in DCM through frit and crystalized in DCM/pentane mixture. Colorless crystals suitable for X-ray analysis was observed at 0°c. Yield: 0.290g (83%). Mp: >240°C. ¹H NMR (400.31 MHz, CDCl₃): δ 0.34 (*s*, 9H, Si*Me*₃), 0.53 (*s*, 9H, Si*Me*₃), 1.20 (*s*, 18H, C*Me*₃), 7.35-7.37 (*d*, J= 7.1 Hz, 2H, Ph), 7.44-7.55 (*m*, 3H, Ph) ppm. ¹³C NMR{¹H} (100.67 MHz, CDCl₃): δ 3.65 (Si*Me*₃), 5.14 (Si*Me*₃), 30.82 (C*Me*₃), 54.17 (CMe₃), 126.60, 126.88, 127.17, 127.23, 129.57, 130.79 (Ph-C), 168.95 (NCN) ppm. MALDI: *m*/*z* [C₂₁H₄₁N₃Si₂GeAuClNa]⁺: 720.08 [M+Na]⁺. Anal Calcd: C, 36.20; H, 5.93; N, 6.03. Found: C, 36.25; H, 5.95; N, 6.07.

Synthesis of 5: AgSbF₆ (0.171g, 0.5 mmol) was dissolved in DCM and added to the solution of 4 (0.348g, 0.5 mmol) in benzene. Immediately AgCl was precipitated out. After overnight stirring, AgCl was separated out from the reaction mixture by filtration and reduced the volume to 15 mL and kept it at 0°C. Colorless block shaped crystals suitable for X-ray analysis was observed after one day. Yield: 0.340g (69%). Mp: 210°C (decomposed). ¹H NMR (400.31 MHz, CDCl₃): δ 0.39 (*s*, 9H, Si*M*e₃), 0.56 (*s*, 9H, Si*M*e₃), 1.23 (*s*, 18H, C*M*e₃), 7.18-7.24 (*m*, 1H, Ph), 7.36-7.41 (*m*, 5H, Ph), 7.45-7.62 (*m*, 5H, Ph) ppm. ¹³C NMR {¹H} (100.67 MHz, CDCl₃): δ 4.69 (Si*M*e₃), 6.38 (Si*M*e₃), 32.00 (C*M*e₃), 55.36 (CMe₃), 127.02, 128.03, 128.41, 128.70, 128.77, 131.30, (Ph-C), 170.74 (NCN) ppm. MALDI: *m*/*z* [C₂₈H₅₀N₃Si₂GeAu]⁺: 661.97 [M-C₆H₆]⁺. Anal Calcd: C, 33.96; H, 5.09; N, 4.24. Found: C, 33.88; H, 5.15; N, 4.21.

Synthesis of 6: $AgSbF_6$ (0.171g, 0.5 mmol) was dissolved in DCM and added to the solution of 4 (0.348g, 0.5 mmol) in toluene. Immediately AgCl was precipitated out. After overnight stirring, AgCl was separated out from the reaction mixture by filtration and reduced the volume to 15 mL and kept it at 0°C. Colorless block shaped crystals suitable for X-ray analysis was

observed after one day. Yield: 0.360g (72%). Mp: 212°C (decomposed). ¹H NMR (400.31 MHz, CDCl₃): δ 0.29 (*s*, 9H, Si*Me*₃), 0.46 (*s*, 9H, Si*Me*₃), 1.24 (*s*, 18H, C*Me*₃), 2.70 (*s*, 3H, C*H*_{3,toluene}), 7.04-7.10 (*m*, 1H, Ph), 7.15-7.18 (*m*, 2H, Ph), 7.28-7.34 (*m*, 1H, Ph), 7.36-7.62 (*m*, 6H, Ph) ppm. ¹³C NMR {¹H} (100.67 MHz, CDCl₃): δ 4.77 (Si*Me*₃), 6.36 (Si*Me*₃), 21.64 (*C*H_{3,toluene}), 32.09 (*CMe*₃), 55.64 (*C*Me₃), 125.49, 127.76, 128.06, 128.30, 128.41, 129.23, 129.39, 129.92, 131.01, 131.42 (Ph-*C*), 138.06 (N*C*N) ppm. MALDI: *m*/*z* [C₂₉H₅₂N₃Si₂GeAu]⁺: 662.25 [M-C₆H₅CH₃]⁺. Anal Calcd: C, 34.68; H, 5.22; N, 4.18. Found: C, 34.65; H, 5.25; N, 4.11.

Synthesis of 8:

AgSbF₆ (0.171g, 0.5 mmol) was dissolved in DCM and added to the solution of **7** (0.310g, 0.5 mmol) in benzene. Immediately AgCl was precipitaed out. After overnight stirring, AgCl was separated out from the reaction mixture by filtration and reduced the volume to 10 mL and kept it at 0°C. Colorless block shaped crystals suitable for X-ray analysis was observed after one day. Yield: 0.330g (71%). Mp: 230°C (decomposed). ¹H NMR (400.31 MHz, CDCl₃): δ 0.76 (d, 12H, *J*= 6.9 Hz, CH(*CH*₃)₂), 0.96 (d, 12H, *J*= 6.9 Hz, CH(*CH*₃)₂), 2.16-2.23 (m, 4H, *CH*(CH₃)₂), 6.97 (m, 1H, Ph), 7.04 (m, 4H, *J*= 7.8 Hz Ph), 7.22-7.28 (m, 4H, Ph), 7.39 (t, 2H, *J*= 7.8 Hz, C₆H₆) ppm. ¹³C NMR {¹H} (100.67 MHz, CDCl₃): δ 24.05, 24.25, 24.56, 28.63, 124.35, 125.08, 130.78, 133.95, 145.08, 184.36 ppm. MALDI: *m*/*z* [C₃₃H₄₀N₂Au]⁺: 585.31 [M-C₆H₆]⁺. Anal Calcd: C, 44.66; H, 4.96; N, 3.06. Found: C, 44.52; H, 4.51; N, 3.07.

Synthesis of 9: 30 mL toluene was added into 0.420 g of silylene [{PhC(N*t*Bu)₂}Si{N(SiMe₃)₂}] (1 mmol) in a 100 mL schlenk flask and CoCp(CO)₂ (0.133 mL, 1mmol) was added into the reaction mixture. After overnight stirring at room temperature, the reaction mixture was dried completely and 15 mL pentane was added into the reaction mixture and kept it at -30°C. Red colored crystals suitable for X-ray analysis was observed after two days. Yield: 0.390g (66%). Mp: 190°C. ¹H NMR (400.31 MHz, CDCl₃): δ 0.46 (*s*, 9H, Si*Me*₃), 0.76 (*s*, 9H, Si*Me*₃), 1.25 (*s*, 18H, C*Me*₃), 4.87 (*s*, 5H, *Cp*), 6.91-6.97 (*m*, 3H, Ph), 7.06 (*br*, 1H, Ph), 7.30 (*br*, 1H, Ph) ppm. ²⁹Si{¹H} NMR (79.495 MHz, CDCl₃, 298 K): δ 0.84 (*Si*Me₃), 4.90 (*Si*Me₃), 50.64 (*Si*N(SiMe₃)₂) ppm. MALDI: *m/z* [C₂₈H₄₉N₃Si₃OCo]⁺: 587.20 [M+H]⁺, 508.21 [M-Cp+2H]⁺. IR *v*_{CO} (solid): 1873 cm⁻¹. Anal Calcd: C, 57.30; H, 8.42; N, 7.16. Found: C, 57.38; H, 8.45; N, 7.21.

Synthesis of 10: 20 mL hexane was added into 0.646 g (1 mmol) of tris(2,4-di-tertbutylphenyl)phosphite in a 100 mL schlenk flask and $CoCp(CO)_2$ (0.133 mL, 1 mmol) was added into the reaction mixture and was refluxed for 5 days. After filtration, kept the reaction mixture at -30°C. Red colored crystals suitable for X-ray analysis was observed after two days. Yield: 0.570g (70%). Mp: 174-176°C. ¹H NMR (400.31 MHz, CDCl₃): δ 1.30 (*s*, 36H, C*Me*₃), 1.53 (*s*, 36H, C*Me*₃), 4.64 (*s*, 5H, *Cp*), 7.05 (*d*, *J*= 8.6 Hz, 3H, Ph), 7.36 (*s*, 3H, Ph), 7.68 (*d*, *J*= 8.6 Hz, 3H, Ph) ppm. ³¹P{¹H} NMR (161.976 MHz, CDCl3, 298K): δ 155.41 ppm. MALDI: *m/z* [C₄₉H₇₁PO4Co]⁺: 834.12 [M-2H]⁺, 755.18 [M-Cp-H]⁺. IR *v*_{CO} (solid): 1956 cm⁻¹. Anal Calcd: C, 72.30; H, 8.79. Found: C, 72.28; H, 8.81.

General procedure for the glycosylation:



Silylene-Au complex (2) (0.014 g, 0.0145 mmol) was added to a solution of donor 12a (0.100 g, 0.145 mmol) and acceptor 13a (0.073 g, 0.145 mmol) in anhydrous CH_2Cl_2 (4mL) containing 4Å MS powder (0.200 g) and stirred at 25 °C for 15 min, concentrated *in vacuo*, and resulting residue was purified by column chromatography (*n*-hexane/EtOAc) to afford disaccharide 14a (0.113 g, 75%) as a thick syrup.

Similar procedure was adopted for the synthesis of **14b**, **14c**, **14d**, **14e**, **14f**, **18** and **20** in case of other Au-complexes (**5** and **8**) also.

Preparation of compound 19: To a solution of compound **18** (0.350 g, 0.241 mmol, 1 eq), methanol-CH₂Cl₂ (1:1) (4 mL), a 1 M solution of NaOMe in MeOH (1 mL), was added and stirred for 1 h. The NaOMe was quenched by Amberlite IR-120, filtered, and concentrated in vacuo and the resulting residue was purified by column chromatography (*n*-hexane/EtOAc) to afford trisaccharide di-ol **19** (0.275 g, 92%) as a thick syrup

Compound Characterization Data

Compound (12a): Mishra, B.; Neralkar, M.; Hotha, S. Angew. Chem., Int. Ed. 2016, 55, 7786-7791.

1-*O*-(((1-ethynylcyclohexyl)oxy)carbonyl)-2-*O*-benzoyl-3,4,6-tri-*O*-benzyl α/β -Dmannopyranoside [α:β (1.0:3.0)] (**12b**): Syrup; [α]²⁵_D (CHCl₃, *c* 1.0): +13.8; IR (cm⁻¹, CHCl₃): 3250, 2935, 1730, 1625, 1455, 1260, 1130, 1080, 915, 705; ¹H NMR (400.31 MHz, CDCl₃): δ 1.19 – 1.27 (m, 2H), 1.40 – 1.45 (m, 4H), 1.50 – 1.60 (m, 6H), 1.67 – 1.73 (m, 2H), 1.78 – 1.86 (m, 2H), 1.88 – 2.01 (m, 2H), 2.03 – 2.15 (m, 2H), 2.38 (s, 1H), 2.41 (s, 1H), 3.67 – 3.73 (m, 2H), 3.76 – 3.91 (m, 7H), 4.03 – 4.25 (m, 1H), 4.50 – 4.67 (m, 8H), 4.73 – 5.35 (m, 5H), 5.38 – 6.34 (m, 3H), 7.12 – 7.19 (m, 14H), 7.27 – 7.34 (m, 16H), 7.38 – 7.43 (m, 4H), 7.52 – 7.57 (m, 2H), 7.99 – 8.01 (m, 4H); $^{13}C{^{1}H}$ NMR (100.67 MHz, CDCl₃): δ 22.3, 22.4, 22.5, 22.5, 24.9, 25.0, 36.4, 36.7, 36.7, 36.8, 68.1, 68.2, 72.5, 72.7, 73.4, 73.6, 73.6, 75.0, 75.1, 75.2, 75.2, 75.3, 75.6, 76.4, 77.2, 77.4, 78.2, 78.4, 79.8, 79.8, 82.5, 82.5, 93.5, 95.5, 127.7 – 128.0 (12C), 128.1 (6C), 128.4 – 128.5 (16C), 129.5, 129.7, 130.0 (4C), 133.3 (2C), 137.6, 137.9 (2C), 138.0 (2C), 138.1, 151.0, 151.3, 165.1, 165.5; HRMS (ESI-MS): m/z [C₄₃H₄₄O₉Na]⁺: 727.2874 [M+Na]⁺.

Compound (12c): Mishra, B.; Neralkar, M.; Hotha, S. Angew. Chem., Int. Ed. 2016, 55, 7786-7791.

1-*O*-(((1-ethynylcyclohexyl)oxy)carbonyl)-2,3,5-tri-*O*-benzyl α-D-arabinoside (**12d**): Syrup; [α]²⁵_D (CHCl₃, *c* 1.0): +33.2; IR (cm⁻¹, CHCl₃): 3020, 1746, 1518, 1215, 1049, 740, 669; ¹H NMR (400.31 MHz, CDCl₃): δ 1.39 (m, 1H), 1.54 – 1.65 (m, 1H), 1.65 – 1.79 (m, 4H), 1.92 – 2.00 (m, 2H), 2.22 – 2.36 (m, 2H), 2.71 (s, 1H), 3.69 (d, J = 4.9 Hz, 2H), 4.08 (dd, J = 5.8, 2.0 Hz, 1H), 4.23 (d, J = 2.8 Hz, 1H), 4.47 (q, J = 5.0 Hz, 1H), 4.53 – 4.60 (m, 3H), 4.61 (d, J = 2.1 Hz, 2H), 4.70 (d, J = 11.9 Hz, 1H), 6.25 (s, 1H), 7.29 – 7.43 (m, 15H); ¹³C{¹H} NMR (100.67 MHz, CDCl₃): δ 22.6, 22.6, 25.0, 36.7, 37.0, 69.4, 72.1, 72.1, 73.5, 75.2, 78.0, 82.9, 83.4, 83.4, 87.0, 103.2, 127.7, 127.8, 127.8, 127.8, 127.8, 127.8, 128.1, 128.1, 128.1, 128.4, 128.4, 128.4, 128.4, 128.5, 128.5, 137.2, 137.7, 138.0, 151.4; HRMS (ESI-MS): *m/z* [C₃₅H₃₈O₇Na]⁺: 593.2518 [M+Na]⁺.

Compound (12e): Sureshkumar, G.; Hotha, S. *Glycoconjugate. Journal*, 2012, *29*, 221-230.
Compound (12f): Thadke, S. A.; Mishra, B.; Hotha, S. *Org. Lett.* 2013, *15*, 2466-2469.
Compound (13a): Tiwari, V.; Badavath, V. N.; Singh, A. K.; Kandasamy, J. *Tetrahedron Lett.* 2018, *59*, 2511–2514.

Methyl 2,3,4-tri-*O*-benzoyl-6-*O*-(2,3,4,6-tetra-*O*-benzyl α/β-D-mannopyranosyl) α-Dglucopyranoside [α:β (3.0:1.0)] (**14a**): Syrup; $[α]^{25}_D$ (CHCl₃, *c* 1.0): +44.7; IR (cm⁻¹, CHCl₃): 3021, 2931, 1730, 1503, 1455, 1271, 1215, 1101, 919, 748, 699; ¹H NMR (400.31 MHz, CDCl₃): δ 3.37 (s, 3H), 3.43 (s, 3H), 3.48 – 3.57 (m, 2H), 3.59 – 3.74 (m, 7H), 3.80 – 4.02 (m, 5H), 4.03 – 4.26 (m, 3H), 4.29 – 4.42 (m, 3H), 4.42 – 4.67 (m, 9H), 4.68 – 5.09 (m, 7H), 5.14 – 5.37 (m, 4H), 5.48 – 5.66 (m, 2H), 6.00 – 6.25 (m, 2H), 7.14 – 7.20 (m, 4H), 7.20 – 7.41 (m, 50H), 7.41 – 7.56 (m, 4H), 7.83 – 8.01 (m, 12H); ¹³C{¹H} NMR (100.67 MHz, CDCl₃): δ 55.5, 55.6, 66.3, 68.2, 68.7, 69.0, 69.1, 69.5, 69.6, 69.9, 70.7, 70.7, 71.5, 72.1, 72.2, 72.2, 72.3, 72.7, 73.3, 73.5, 74.0, 74.2, 74.8, 74.9, 74.9, 75.1, 75.2, 76.1, 80.0, 82.2, 97.0, 97.1, 98.3, 102.4, 127.5 – 128.2 (24C), 128.3 – 128.5 (28C), 129.0, 129.2, 129.2, 129.2, 129.4, 129.4, 129.8 – 130.0 (12C), 133.2, 133.2, 133.4, 133.5, 133.5, 133.6, 138.3, 138.4, 138.5, 138.6, 138.7, 138.8, 139.0, 165.2, 165.2, 165.5, 165.9, 165.9, 166.0; HRMS (ESI-MS): *m/z* [C₆₂H₆₀O₁₄Na]⁺: 1051.3875 [M+Na]⁺.

Methyl 2,3,4-tri-*O*-benzoyl-6-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl α -D-mannopyranosyl) α -D-glucopyranoside (**14b**): Mp: 64°C; [α]²⁵_D (CHCl₃, *c* 1.0): +42.3; IR (cm⁻¹, CHCl₃): 3021, 1727, 1453, 1267, 1103, 1099, 1028, 745, 669; ¹H NMR (400.31 MHz, CDCl₃): δ 3.45 (s, 3H), 3.67 (dd, J = 11.0, 2.8 Hz, 1H), 3.74 (dd, J = 10.8, 2.9 Hz, 1H), 3.83 (dd, J = 10.8, 3.7 Hz, 1H), 3.87 (dd, J = 8.7 Hz, 1H), 4.01 (dd, J = 11.0, 5.3 Hz, 1H), 4.09 – 4.19 (m, 2H), 4.31 (ddd, J = 9.8, 4.9, 3.0 Hz, 1H), 4.48 (d, J = 12.0 Hz, 1H), 4.56 (dd, J = 15.4, 11.2 Hz, 2H), 4.70 (d, J = 12.0 Hz, 1H), 4.75 (d, J = 11.4 Hz, 1H), 4.94 (d, J = 10.9 Hz, 1H), 5.07 (d, J = 1.4 Hz, 1H), 5.28 (d, J = 3.6 Hz, 1H), 5.37 (dd, J = 10.2, 3.6 Hz, 1H), 5.57 – 5.72 (m, 2H), 6.23 (t, J = 9.9 Hz, 1H), 7.22 – 7.43 (m, 23H), 7.43 – 7.59 (m, 4H), 7.94 (m, 2H), 7.99 – 8.07 (m, 4H), 8.12 (m, 2H); ¹³C{¹H} NMR (100.67 MHz, CDCl₃): δ 55.7, 66.5, 68.3, 69.0, 69.1, 69.8, 70.7, 71.7, 71.9, 72.3, 73.5, 74.3, 75.3, 78.1, 97.1, 98.1, 127.6, 127.6, 127.6, 127.7, 127.7, 128.1, 128.1, 128.3, 128.3, 128.4, 128.4, 128.4, 128.4, 128.4, 128.5, 128.5, 128.5, 128.5, 128.6, 128.6, 128.6, 128.6, 129.1, 129.2, 129.4, 129.8, 129.8, 130.0, 130.1, 130.1, 130.1, 130.1, 130.1, 133.3, 133.3, 133.5, 133.5, 138.2, 138.6, 138.7, 165.4, 165.7, 166.0, 166.0; HRMS (ESI-MS): *m/z* [C₆₂H₅₈O₁₅Na]⁺: 1065.3690 [M+Na]⁺.

Methyl 2,3,4-tri-*O*-benzoyl-6-*O*-(2,3,4,6-tetra-*O*-benzoyl α -D-mannopyranosyl) α -D-glucopyranoside (**14c**): Mp: 72°C; $[\alpha]^{25}_{D}$ (CHCl₃, *c* 1.0): +2.9; IR (cm⁻¹, CHCl₃): 3045, 2910, 1720, 1625, 1228, 1035, 970; ¹H NMR (400.31 MHz, CDCl₃): δ 3.61 (s, 3H), 3.78 (d, *J* = 10.7 Hz, 1H), 4.09 (dd, *J* = 10.7, 6.1 Hz, 1H), 4.34 – 4.40 (m, 2H), 4.51 – 4.55 (m, 1H), 4.62 (dd, *J* = 11.9,1.7 Hz, 1H), 5.15 (s, 1H), 5.26 (dd, *J* = 10.2, 3.7 Hz, 1H), 5.33 (d, *J* = 3.6 Hz, 1H), 5.58 (td, *J* = 9.9, 1.6 Hz, 1H), 5.76 (dd, *J* = 3.1, 1.7 Hz, 1H), 5.98 (dd, *J* = 10.1, 3.2 Hz, 1H), 6.08 (t, *J* =

10.1 Hz, 1H), 6.21 (t, J = 9.9 Hz, 1H), 7.26 – 7.31 (m, 3H), 7.34 (m, 3H), 7.39 – 7.43 (m, 6H), 7.45 – 7.49 (m, 5H), 7.52 – 7.55 (m, 2H), 7.58 – 7.63 (m, 2H), 7.84 – 7.87 (m, 2H), 7.92 – 7.94 (m, 2H), 7.97 – 8.04 (m, 6H), 8.06 – 8.11 (m, 4H); ¹³C{¹H} NMR (100.67 MHz, CDCl₃): δ 55.8, 62.9, 66.6, 67.1, 68.5, 69.1, 69.6, 70.2, 70.5, 70.7, 72.3, 97.1, 97.6, 128.4 (4C), 128.5 (2C), 128.6 (6C), 128.7(2C), 128.9, 129.2, 129.3 (3C), 129.4, 129.5, 129.9 (6C), 130.0 (6C), 130.1 (2C), 133.1, 133.2, 133.2, 133.4, 133.5, 133.6, 133.6, 165.5, 165.5, 165.5, 165.7, 165.9, 165.9, 166.2; HRMS (ESI-MS): m/z [C₆₂H₅₂O₁₈Na]⁺: 1107.3046 [M+Na]⁺.

Methyl 2,3,4-tri-O-benzoyl-6-O-(2,3,5-tri-O-benzyl α/β -D-arabinofuranosyl) α-Dglucopyranoside $[\alpha:\beta (1.4:1.0)]$ (14d): $[\alpha]^{25}_{D}$ (CHCl₃, c 0.5): +74.8; IR (cm⁻¹, CHCl₃): 2920, 1724, 1452, 1258, 1102, 1024, 801, 704; ¹H NMR (400.31 MHz, CDCl₃): δ 3.36 (s, 3H), 3.44 (s, 3H), 3.50 - 3.52 (m, 4H), 3.61 (dd, J = 11.6, 5.8 Hz, 1H), 3.66 (dd, J = 11.1, 3.1 Hz, 1H), 3.85 - 10.03.99 (m, 3H), 3.99 - 4.12 (m, 4H), 4.18 - 4.23 (m, 3H), 4.41 - 4.51 (m, 7H), 4.52 - 4.61 (m, 7H)3H), 4.65 (d, J = 11.8 Hz, 1H), 4.82 (d, J = 11.7 Hz, 1H), 5.05 -5.07 (m, 2H), 5.18 -5.27 (m, 4H), 5.54 (dd, J = 10.2, 9.4 Hz, 1H), 5.63 (t, J = 9.8 Hz, 1H), 6.12 (td, J = 9.6, 1.0 Hz, 2H), 7.24 -7.54 (m, 48H), 7.80 - 8.02 (m, 12H); ${}^{13}C{}^{1}H$ NMR (100.67 MHz, CDCl₃); δ 55.5, 55.6, 65.6, 66.0, 68.5, 69.0, 69.2, 69.5, 69.6, 70.6, 70.7, 72.0, 72.0, 72.1, 72.1, 72.2, 72.3, 72.4, 73.3, 73.3, 80.4, 80.5, 83.2, 83.3, 84.2, 88.4, 96.8, 96.9, 101.3, 106.1, 127.5 - 127.9 (15C), 128.2 - 128.3 (15C), 128.4 (12C), 129.0, 129.1 (2C), 129.3 (2C), 129.4, 129.7 (4C), 129.8 (2C), 129.9 (6C), 133.0, 133.0, 133.2, 133.3, 133.3, 133.4, 137.6, 138.0, 138.0, 138.1, 138.1, 138.2, 165.1, 165.3, 165.8, 165.8, 165.8, 165.9; HRMS (ESI-MS): *m/z* [C₅₄H₅₂O₁₃Na]⁺: 947.3989 [M+Na]⁺.

Methyl 2,3,4-tri-*O*-benzoyl-6-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl α -D-mannopyranosyl) α -D-glucopyranoside (14e): See (14b).

Methyl 2,3,4-tri-*O*-benzoyl-6-*O*-(2-*O*-benzoyl-3,5-tri-*O*-benzyl β-D-ribofuranosyl) α-D-glucopyranoside (**14f**): Mp: 92°C; $[α]^{25}_D$ (CHCl₃, *c* 1.0): +56.4; IR (cm⁻¹, CHCl₃): 3020, 1729, 1514, 1272, 1215, 1105, 742, 669; ¹H NMR (400.31 MHz, CDCl₃): δ 3.33 (s, 3H), 3.44 (dd, J = 10.4, 6.1 Hz, 1H), 3.51 – 3.65 (m, 2H), 3.85 (d, J = 11.3 Hz, 1H), 4.07 – 4.16 (m, 2H), 4.19 – 4.25 (m, 1H), 4.27 (d, J = 11.5 Hz, 1H), 4.40 – 4.41(m, 2H), 4.48 (d, J = 11.4 Hz, 1H), 5.07 – 5.15 (m, 2H), 5.14 – 5.22 (m, 1H), 5.37 – 5.50 (m, 2H), 6.05 (t, J = 9.8 Hz, 1H), 7.05 – 7.46 (m, 22H), 7.76 – 7.80 (m, 2H), 7.81 – 7.85 (m, 2H), 7.87 – 7.91 (m, 2H), 7.95 – 7.99 (m, 2H); ¹³C{¹H} NMR (100.67 MHz, CDCl₃): δ 55.6, 66.6, 68.9, 69.8, 70.6, 71.2, 72.2, 73.0, 73.3, 74.3, 78.0, 80.8, 96.9, 105.9, 127.6, 127.7, 127.7, 127.8, 128.0, 128.0, 128.3, 128.3, 128.3, 128.3, 128.3,

128.4, 128.4, 128.4, 128.4, 128.5, 128.5, 128.5, 128.5, 129.1, 129.1, 129.3, 129.7, 129.7, 129.7, 129.9, 129.9, 129.9, 129.9, 130.0, 130.0, 133.1, 133.3, 133.4, 133.4, 137.6, 138.2, 165.3, 165.5, 165.9, 165.9; HRMS (ESI-MS): *m/z* [C₅₄H₅₀O₁₄Na]⁺: 945.3099 [M+Na]⁺.

Methyl 2,4-di-*O*-benzyl- α -D-mannopyranoside (17): Syrup; $[\alpha]^{25}_{D}$ (CHCl₃, *c* 0.5): +14.0; IR (cm⁻¹, CHCl₃): 3568, 3017, 2925, 910, 738, 564; ¹H NMR (400.31 MHz, CDCl₃): δ 2.48 (s, 2H), 3.30 (s, 3H), 3.58 (ddd, *J* = 9.8, 4.4, 2.8 Hz, 1H), 3.68 (t, *J* = 9.5 Hz, 1H), 3.71 (dd, *J* = 3.7, 1.6 Hz, 1H), 3.77 (dd, *J* = 11.9, 4.4 Hz, 1H), 3.84 (dd, *J* = 11.8, 2.9 Hz, 1H), 3.98 (dd, *J* = 9.2, 3.7 Hz, 1H), 4.59 (d, *J* = 11.8 Hz, 1H), 4.65 (d, *J* = 11.1 Hz, 1H), 4.70 (d, *J* = 11.8 Hz, 1H), 4.73 (d, *J* = 1.6 Hz, 1H), 4.88 (d, *J* = 11.2 Hz, 1H), 7.29 – 7.36 (m, 10H); ¹³C{¹H} NMR (100.67 MHz, CDCl₃): δ 54.9, 62.3, 71.3, 71.8, 73.2, 74.9, 76.4, 78.4, 98.3, 127.9, 128.0, 128.0, 128.1, 128.1, 128.2, 128.5, 128.5, 128.7, 128.7, 137.7, 138.4; HRMS (ESI-MS): *m*/*z* [C₂₁H₂₆O₆Na]⁺: 397.1635 [M+Na]⁺.

Methyl 2,4-di-*O*-benzyl-3,6-di-*O*-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl)- α -D-mannopyranoside (**18**): syrup; $[\alpha]^{25}_{D}$ (CHCl₃, *c* 0.5): +12.4; IR (cm⁻¹, CHCl₃): 3265, 3025, 2955, 1725, 917, 730, 571; ¹H NMR (400.31 MHz, CDCl₃): δ 3.22 (s, 3H), 3.65 – 3.71 (m, 3H), 3.73 – 3.85 (m, 5H), 3.87 – 3.96 (m, 2H), 4.02 – 4.10 (m, 4H), 4.13 – 4.19 (m, 2H), 4.47 (s, 1H), 4.49 – 4.50 (m, 3H), 4.51 – 4.56 (m, 3H), 4.65 – 4.68 (m, 4H), 4.71 – 4.76 (m, 3H), 4.80 (d, *J* = 11.0 Hz, 1H), 4.87 – 4.92 (m, 2H), 5.10 (d, *J* = 1.8 Hz, 1H), 5.33 (d, *J* = 1.7 Hz, 1H), 5.73 (t, *J* = 2.9 Hz, 1H), 5.77 (t, *J* = 2.6 Hz, 1H), 7.16 – 7.40 (m, 44H), 7.52 – 7.56 (m, 2H), 8.02 – 8.04 (m, 2H), 8.07 – 8.09 (m, 2H); ¹³C {¹H} NMR (101.67 MHz, CDCl₃): δ 54.9, 66.6, 68.8, 68.8, 69.1, 69.2, 69.5, 71.1, 71.2, 71.7, 71.9, 72.5, 72.5, 73.5, 73.6, 74.3, 74.5, 75.1, 75.1, 75.3, 77.6, 77.6, 78.3, 78.7, 98.3, 98.3, 99.8, 127.5, 127.6 (5C), 127.7 (2C), 127.8 (4C), 127.9 (4C), 128.0 (4C), 128.1 (4C), 128.3 (2C), 128.4 (10C), 128.5 (6C), 128.6 (2C), 130.0, 130.1 (5C), 133.2, 133.2, 138.0, 138.0, 138.0, 138.3, 138.6, 138.6, 138.7, 138.8, 165.6, 165.6; HRMS (ESI-MS): *m/z* [C₈₉H₉₀O₁₈Na]⁺: 1469.6021 [M+Na]⁺.

Methyl 2,4-di-*O*-benzyl-3,6-di-*O*-(3,4,6-tri-*O*-benzyl-α-D-mannopyranosyl)-α-Dmannopyranoside (**19**): syrup; $[\alpha]^{25}_{D}$ (CHCl₃, *c* 1.0): +46.1; IR (cm⁻¹, CHCl₃): 3554, 3275, 3037, 2963, 925, 716, 552; ¹H NMR (400.31 MHz, CDCl₃): δ 2.37 (s, 2H), 3.23 (s, 3H), 3.58 – 3.73 (m, 6H), 3.75 – 3.93 (m, 8H), 3.93 – 4.03 (m, 2H), 4.08 – 4.14 (m, 2H), 4.44 – 4.57 (m, 7H), 4.57 – 4.62 (m, 5H), 4.62 – 4.70 (m, 3H), 4.83 (t, J = 10.7 Hz, 2H), 5.07 (d, J = 1.8 Hz, 1H), 5.21 (d, J = 1.7 Hz, 1H), 7.12 – 7.34 (m, 40H); ${}^{13}C{}^{1}H$ NMR (101.67 MHz, CDCl₃): δ 54.9, 66.2, 68.2, 68.8, 68.9, 69.5, 71.3, 71.6, 71.6, 72.1, 72.2, 72.4, 73.5, 73.7, 74.4, 74.6, 75.0, 75.1, 75.1, 75.2, 77.9, 79.1, 79.7, 80.2, 98.3, 99.8, 101.6, 127.6, 127.7 (8C), 127.8 (2C), 127.9 (5C), 128.0 (6C), 128.1 (2C), 128.4 (8C), 128.5 (2C), 128.6 (6C), 138.0, 138.0, 138.2, 138.4, 138.4, 138.4, 138.6, 138.7; HRMS (ESI-MS): m/z [C₇₅H₈₂O₁₆Na]⁺: 1261.5495 [M+Na]⁺.

Methyl 2,4-di-O-benzyl-3,6-di-O-(2-O-(2,3,4,6-tetra-O-benzoyl α-Dmannopyranosyl) -3,4,6-tri-O-benzyl α -D-mannopyranosyl)- α -D-mannopyranoside (20): syrup; $[\alpha]^{25}_{D}$ (CHCl₃, c 0.5): +8.4; IR (cm⁻¹, CHCl₃): 3272, 3045, 2964, 1720, 1130, 945, 730, 583; ¹H NMR (400.31 MHz, CDCl₃): δ 3.25 (s, 3H), 3.55 – 3.75 (m, 6H), 3.78 – 3.86 (m, 2H), 3.86 – 3.95 (m, 3H), 3.99 – 4.04 (m, 5H), 4.11 - 4.18 (m, 2H), 4.29 (dt, J = 12.4, 2.5 Hz, 1H), 4.36 - 4.46 (m, 3H), 4.47 - 4.60 (m, 9H), 4.61 - 4.63 (m, 3H), 4.65 - 4.68 (m, 3H), 4.70 - 4.75 (m, 1H), 4.80 (s, 1H), 4.92 (t, J = 10.5Hz, 2H), 5.04 (d, J = 2.5 Hz, 1H), 5.13 (d, J = 2.5 Hz, 1H), 5.21 (d, J = 2.5 Hz, 1H), 5.34 (d, J = 2 2.5 Hz, 1H), 5.86 (d, J = 2.8 Hz, 1H), 5.89 – 5.98 (m, 3H), 6.13 (td, J = 10.1, 2.0 Hz, 1H), 6.19 (td, J = 10.1, 2.0 Hz, 1H), 6.88 - 7.06 (m, 5H), 7.06 - 7.23 (m, 15H), 7.28 - 7.33 (m, 26H), 7.35-7.46 (m, 14H), 7.51 - 7.64 (m, 4H), 7.83 - 7.91 (m, 8H), 8.04 - 8.06 (m, 4H), 8.12 - 8.14 (m, 4H); ¹³C{¹H} NMR (101.67 MHz, CDCl₃): δ 54.9, 62.7, 62.7, 66.4, 66.7, 66.9, 69.2, 69.2, 69.4, 69.6, 70.3, 70.3, 70.5, 70.6, 71.7, 71.9, 71.9, 72.2, 72.6, 72.7, 73.2, 73.4, 74.7, 74.8, 74.9, 74.9, 75.1, 75.4, 77.1, 77.4, 77.9, 78.7, 79.4, 79.7, 98.3, 99.5, 99.5, 99.8, 101.1, 127.1 (2C), 127.4 (2C), 127.5 – 127.7 (14C), 127.9 (2C), 128.0 (4C), 128.3 (4C), 128.4 (12C), 128.5 (12C), 128.7 (4C), 129.1, 129.2, 129.4, 129.5, 129.7, 129.7, 129.9 - 130.0 (16C), 130.1, 130.2, 133.0, 133.1, 133.1, 133.1, 133.4, 133.4, 133.4, 133.4, 138.2, 138.3, 138.3, 138.3, 138.5, 138.5, 138.7, 138.8, 165.2, 165.2, 165.4, 165.5, 165.5, 165.6, 166.2, 166.3; HRMS (ESI-MS): m/z [C₁₄₃H₁₃₄O₃₄Na]⁺: 2418.8691 [M+Na]+.

Entry	Catalyst	Mol	Yield
	·	%	(%)
1	1	10	-
2	TfOH	10	10
3	$AgSbF_6$	10	-
4	$[Et_3O]+[SbCl_6]-$	10	20
5	PPh ₃ AuCl/AgSbF ₆	10	75
6	$1/\text{AgSbF}_6$ (in situ)	10	42
	(DCM)		
7	$1/AgSbF_6$ (in situ)	10	55
	(DCM+benzene)		
8	$[IPr_2Au]^+[SbF_6]^-$	10	-

Table S1.1 Attempts of disaccharide synthesis using carbonate donors with different catalysts.^[a]

(a)Reagents and conditions: 12c (1 equiv.), 13a (1 equiv.), Catalyst, DCM, room temperature, 12 h

Table S1.2 Attempts of disaccharide synthesis using orthoester donors with different catalysts.^[b]

Entry	Catalyst	Mol %	Yield (%)
1	AgSbF ₆	10	30% (14f), 10% (16)
2	PPh ₃ AuCl/AgSbF ₆	10	65% (14f), 30% (16)
3	AuBr ₃	10	55% (14f), 15% (16)
4	1/AgSbF ₆ (in situ) (DCM)	10	76% (14f), 20% (16)
5	1/AgSbF ₆ (in situ) (DCM+benzene)	10	90% (14f), 2% (16)
6	$[IPr_2Au]^+[SbF_6]^-$	10	-

^[b]Reagents and conditions: **12f** (1 equiv.), **13a** (1 equiv.), Catalyst, DCM, room temperature, 12 h.

S2. Molecular Structure:



Figure S1. The molecular structure of **3** (ellipsoid are shown at the probability level of 50%). Hydrogen atoms are omitted for clarity.



Figure S2. The molecular structure of the deactivated catalyst of **8** (ellipsoids are shown at the probability level of 30%). Hydrogen atoms, *i*Pr groups and SbF₆ anion are omitted for clarity.



Figure S3. The molecular structure of the product obtained from the stoichiometric reaction of alkyne and catalyst **8** (ellipsoid are shown at the probability level of 50%). Hydrogen atoms are omitted for clarity.

S3. Crystal Data and Structure Refinements for 2, 3, 4, 8, 9 and 10.

X-ray Crystallography Details: Single crystals of suitable size, coated with paraffin oil was mounted for all the complexes. Crystal data for all the complexes were collected on a Bruker Smart Apex Duo diffractometer at 100 K using Mo K α radiation ($\lambda = 0.71073$ Å). Collected data were integrated by using SAINT and then absorption correction was done by multi-scan method using SADABS program. All the structures were solved by direct methods and refined by full-matrix least-squares methods against F² (SHELXL-2014/6). Crystallographic Information File (CIF) for the structures has been deposited to the Cambridge Crystallographic Data Centre as supplementary publication nos. 1819357 (2), 1819358 (3), 1990445 (4), 1954430 (8), 1990448 (9), 1990449 (10). These CIF can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

	2	3	4
Chemical formula	C ₂₇ H ₄₇ AuF ₆ N ₃ SbSi ₃	$C_{29}H_{51}AuF_6N_3SbSi_3$	C ₂₁ H ₃₉ AuClGeN ₃ Si ₂
Formula weight	930.66	958.71	694.74
Temperature	100(2)	100(2)	100(2)
Wavelength	0.71073	0.71073	0.71073
Crystal system	monoclinic	monoclinic	monoclinic
Space group	$P2_1/n$	$P2_1/c$	$P2_1/c$
Unit cell dimentions	<i>a</i> =11.831(7) Å	<i>a</i> =22.214(7) Å	<i>a</i> =8.9235(5)Å
	<i>b</i> =21.434(14) Å	<i>b</i> =18.276(6) Å	<i>b</i> =18.5223(13)Å
	<i>c</i> =14.563(10) Å	<i>c</i> =18.686(6) Å	<i>c</i> =17.8450(12) Å
	<i>α</i> = 90°	α=90°	<i>α</i> =90°
	$\beta = 98.575(18)^{\circ}$ $\beta = 93.575(18)^{\circ}$		β=101.332(2)°
	γ=90°	γ=90°	γ=90°
Volume	3652(4) Å ³	7576(4) Å ³	2892.0(3)Å ³
Ζ	4	8	4
Density (calculated)	1.693 g/cm ³	1.681 g/cm ³	1.596 g/cm ³

 Table S2. Crystal Data and Structure Refinement for 2, 3, 4, 8, 9 and 10.

Absorption coefficient	4.903 mm ⁻¹	4.729 mm ⁻¹	6.294 mm ⁻¹	
F(000)	1824	3776	1368	
Theta range for data2.28 to 25.25°		2.23 to 25.16°	2.20 to 25.25°	
collection				
Index ranges	-14<=h<=14	-26<=h<=26	-10<=h<=10	
	-25<=k<=25	-21<=k<=21	-22<=k<=22	
	-17<= <i>l</i> <=17	-22<=l<=22	-21<=l<=21	
Reflections collected	87805	104634	81573	
Independent reflections	6620 [R(int)=	13512 [R(int)=	5233 [R(int)=	
	0.1276]	0.2456]	0.0618]	
Coverage of	99.9%	99.4%	100%	
independent reflections				
Function minimized	$\Sigma w (Fo^2 - Fc^2)^2$	$\Sigma w (Fo^2 - Fc^2)^2$	$\Sigma w(Fo^2 - Fc^2)^2$	
Data/ restraints/ 6620/ 0/ 382		13512/ 108/ 849	5233/ 0/ 274	
parameters				
Goodness-of-fit on F2	1.015	1.078	1.119	
$\Delta \sigma \max$	0.001	0.001	0.002	
Final R indices	4974 data; [<i>I</i> >2σ(<i>I</i>)]	7556 data; $[I > 2\sigma(I)]$	4144 data; $[I > 2\sigma(I)]$	
	<i>R1</i> = 0.0363, <i>wR2</i> =	R1 = 0.0800, wR2 =	R1 = 0.0280, wR2 =	
0.0622		0.1492	0.0597	
all data, <i>R1</i> = 0.0649,		all data, $RI = 0.1724$,	all data, <i>R1</i> = 0.0457,	
<i>wR2</i> = 0.0708		wR2 = 0.1766	wR2 = 0.0668	
Largest diff. peak and 0.946 and -0.912 eÅ ⁻		2.265 and -1.323 eÅ ⁻	0.501 and -0.787 eÅ ⁻	
hole ³		3	3	
R. M. S deviation from 0.132 eÅ ⁻³		0.252 eÅ ⁻³	0.145 eÅ ⁻³	
mean				

	8	9	10
Chemical formula	$C_{33}H_{42}AuF_6N_2Sb$	C ₂₇ H ₄₆ CoN ₃ OSi ₃	C ₅₅ H ₇₅ CoO ₄ P
Formula weight	899.40	571.87	890.05

Temperature	100(2)	100(2)	100(2)	
Wavelength	0.71073	0.71073	0.71073	
Crystal system	monoclinic	triclinic	triclinic	
Space group	$P2_{1}/c$	P-1	P-1	
Unit cell dimentions	Unit cell dimentions a=13.4432(9) Å		<i>a</i> =12.132(3) Å	
	<i>b</i> =14.6762(9) Å	<i>b</i> =17.157(15) Å	<i>b</i> =13.703(5) Å	
	<i>c</i> =17.3789(8) Å	<i>c</i> =18.969(15)Å	<i>c</i> =17.467(6) Å	
	<i>α</i> = 90°	α=102.473(13)°	α=112.033(7)°	
	β=100.2060(10)°	β=91.56(2)°	β=104.987(7)°	
	γ=90°	<i>γ</i> =91.73(2)°	<i>γ</i> =92.737(8)°	
Volume	3374.5(3) Å ³	3092(5) Å ³	2565.8(14) Å ³	
Z	4	4	2	
Density (calculated) 1.770 g/cm ³		1.228 g/cm ³	1.152 g/cm ³	
Absorption coefficient 5.201 mm ⁻¹		0.695 mm ⁻¹	0.408 mm ⁻¹	
F(000)	F(000) 1752		958	
Theta range for data	2.25 to 25.25°	2.20 to 25.25	2.19 to 25.25	
collection				
Index ranges	-16<=h<=16	-11<=h<=11	-12<=h<=13	
	-17<=k<=17	-20<=k<=20	-16<=k<=16	
	-20<=l<=20	-22<=l<=22	-20<=l<=20	
Reflections collected	108354	85325	19606	
Independent reflections	6111 [R(int)=	11186 [R(int)=	8546 [R(int)=	
	0.0432]	0.1194]	0.1786]	
Coverage of	99.9%	99.8%	91.9%	
independent reflections				
Function minimized $\Sigma w(Fo^2 - Fc^2)^2$		$\Sigma w (Fo^2 - Fc^2)^2$	$\Sigma w (Fo^2 - Fc^2)^2$	
Data/ restraints/ 6111/ 0/ 396		11186/0/655 8546/0/552		
parameters				
Goodness-of-fit on F2	1.037	1.696 1.050		
$\Delta \sigma \max$	0.002	0.042	0.000	

Final R indices	5485 data; $[I > 2\sigma(I)]$	8239 data; [<i>I</i> >2σ(<i>I</i>)]	3619 data; $[I > 2\sigma(I)]$
	R1 = 0.0201, wR2 =	<i>R1</i> = 0.1474, <i>wR2</i> =	<i>R1</i> = 0.1149, <i>wR2</i> =
	0.0401	0.3961	0.2600
	all data, $RI = 0.0251$, all data, $RI = 0.1738$,		all data, <i>R1</i> = 0.2596,
	<i>wR2</i> = 0.0420	<i>wR2</i> = 0.4128	wR2 = 0.3260
Largest diff. peak and 2.132 and -1.629 eÅ ⁻		7.657 and -1.648	0.618 and -0.793
hole	3	eÅ-3	eÅ-3
R. M. S deviation from 0.082 eÅ ⁻³		0.257 eÅ ⁻³	0.112 eÅ ⁻³
mean			

S4. Deduction of hapticities in 2, 3 and 8

The assignment of hapticity number for the complexes having low hapticities (η^1 - η^3), has always been a complicated task as the difference between M-C bond distances is very less. Therefore, we used a method proposed by Alvarez and co-workers to deduce the hapticity of the metal-arene complexes given in *Organometallics*, 2014, **33**, 6660-6668.

Table S3. Deduction of hapticities in 2, 3 and 8

Complex	$\mathbf{M}\text{-}\mathbf{C}^{\mathbf{a}}(\mathbf{d}_{1},\mathbf{d}_{2},$	Distance ratio ρ_1	Distance ratio ρ_2	η
	d ₃)			
2	2.37, 2.60, 2.77	1.09	1.16	1
3	2.31, 2.73, 2.78	1.18	1.20	1
8	2.29, 2.34, 2.88	1.02	1.25	2

^{*a*} M (=Au), $d_1 < d_2 < d_3$, ^{*b*} $\rho I = d_2/d_1$, ^{*c*} $\rho 2 = d_3/d_1$, if $\rho_1 \approx \rho_2 \gg 1$ then η^1 , if $\rho_2 > \rho_1 \approx 1$ then η^2 , and if $\rho_1 \approx \rho_2 \approx 1$ then η^3

S5. Computational Details.

Computational Methodology

Geometry optimizations were performed using GGA DFT functional BP86¹ with def2-SVP2² basis set using Gaussian 09³ software. Single point calculations were carried out using meta-GGA functional M06⁴ with def2-TZVPP² level of theory on the optimized geometries. Solvent effects were incorporated in single point calculations using PCM model⁵ by incorporating dichloromethane solvent (DCM). ΔE and $\Delta E^{\#}$ represent reaction enthalpy and enthalpy of activation respectively calculated by adding electronic energy at the M06/PCM/def2-TZVPP level of theory and zero point energy at the BP86/def2-SVP level of theory. ΔG and $\Delta G^{\#}$ represent the Gibbs free energy of reaction and Gibbs free energy of activation respectively, calculated by adding electronic energy at the M06/PCM/def2-TZVPP level of theory and the thermal correction to Gibbs free energy at 298.15 K and 1 atm at the BP86/def2-SVP level of theory. The population analysis was performed by natural bond orbital (NBO)⁶ method at M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory.

The model compound **A** for alkynyl glycosyl carbonate (glycosyl donor) was constructed by avoiding all –OR substitution on the carbohydrate ring with hydrogen, except at the nearest C atom of the anomeric carbon. At the nearest C of the anomeric carbon –O-CO-CH₃ group is retained to understand the anchimeric assistance of the protecting group on the oxocarbenium ion. Also cyclohexyl group in the leaving group is replaced by two methyl groups.



Scheme S1. Overall glycosidation reaction at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. ΔE and ΔG (in kcal/mol) represent reaction enthalpy and Gibbs free energy of reaction respectively at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory.



Scheme S2. Mechanism for glycosidation reaction by remote activation using **8** through intermediate **I11**. Energies are given in kcal/mol. ΔE and $\Delta E^{\#}$ (in kcal/mol) represent reaction enthalpy and enthalpy of activation respectively at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. ΔG and $\Delta G^{\#}$ (in kcal/mol) represent the Gibbs free energy of reaction and Gibbs free energy of activation respectively, at same level of theory at 298.15 K and 1 atm.



Scheme S3. Mechanism for glycosidation reaction by remote activation using 2 through intermediate I8'. Energies are given in kcal/mol. ΔE and $\Delta E^{\#}$ (in kcal/mol) represent reaction enthalpy and enthalpy of activation respectively at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. ΔG and $\Delta G^{\#}$ (in kcal/mol) represent the Gibbs free energy of reaction and Gibbs free energy of activation respectively, at same level of theory at 298.15 K and 1 atm.

*The transition state TS6' is obtained using additional keywords IOP(1/8=1) int = finegrid guess = mix



Scheme S4. Mechanism for glycosidation reaction by remote activation in presence of 8 with regeneration of catalyst. ΔE and $\Delta E^{\#}$ (in kcal/mol) represent reaction enthalpy and enthalpy of activation respectively at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. ΔG and $\Delta G^{\#}$ (in kcal/mol) represent the Gibbs free energy of reaction and Gibbs free energy of activation respectively, at same level of theory at 298.15 K and 1 atm.



Scheme S5. Mechanism for glycosidation reaction by remote activation in presence of 8 through the intermediate I13. ΔE and $\Delta E^{\#}$ (in kcal/mol) represent reaction enthalpy and enthalpy of activation respectively at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. ΔG and $\Delta G^{\#}$ (in kcal/mol) represent the Gibbs free energy of reaction and Gibbs free energy of activation respectively, at same level of theory at 298.15 K and 1 atm.

Discussion: We have carried out quantum mechanical calculations to explore the mechanism of model glycosidation reaction^{36,37} between alkynyl glycosyl carbonate A (glycosyl donor), and an acceptor B (ROH) catalyzed by N-heterocyclic carbene/silylene supported Au(I)-benzene complexes 8 and 2 at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory by considering DCM as solvent. The overall reaction is highly exothermic (Scheme S2, $\Delta E = -32.6$ kcal/mol) and exergonic ($\Delta G = -31.4$ kcal/mol). We have calculated different reaction pathways for glycosidation reaction (see SI for the details). Herein, we explain the most feasible mechanism of the glycosidation reaction catalyzed by N-heterocyclic carbene supported Au(I)-benzene complex 8 (Scheme 6).

Other plausible mechanisms are given in the Supporting Information. The optimized geometries (BP86/def2-SVP) and important geometrical parameters of reactants, products, intermediates, and transition states are given in supporting information (Figures S10 to S12).

The first step of the reaction involves the interaction of gold catalyst **8** with alkynyl glycosyl carbonate **A** to form a weakly bound complex **I1**, where the carbonyl oxygen of the carbonate group has a slight bonding interaction with hydrogen atoms of the benzene ring coordinated to Au (O···H distance = 2.487 Å). The reaction energy and Gibbs free energy for this step are found to be -3.6 kcal/mol and 8.1 kcal/mol, respectively. In the next step, the alkynyl group replaces the arene ring in **I1** to form Au-alkyne complex **I2**. The formation of **I2** is exothermic by 0.9 kcal/mol, and exergonic by 8.9 kcal/mol and corresponding energy barrier is low ($\Delta E^{\#} = 5.2$ kcal/mol and $\Delta G^{\#} = 9.3$ kcal/mol). The metal coordination stabilizes the π^* -MO (LUMO+1, E = -2.09 eV, Figure 4) centered on C=C bond in **I2** as compared to the corresponding MO on **A** (LUMO+1, E = -0.54 eV, Figure 4). This facilitates the intramolecular nucleophilic addition of the lone pair on carbonyl oxygen to π^* -MO of **I2** resulting the vinylgold complex **I3**. The reaction energy for the generation of **I3** is only slightly endothermic (0.2 kcal/mol) and exergonic (-0.4 kcal/mol) and the corresponding energy barrier is not very high ($\Delta E^{\#} = 10.6$ kcal/mol and $\Delta G^{\#} = 10.6$ kcal/mol).

In the next step, the glycosidic bond undergoes cleavage to form the intermediate I4, which is also facilitated by the donation of lone pair of electrons on pyranose oxygen to σ^* - orbital glycosidic bond. The formation of I4 is slightly endergonic process (1.2 kcal/mol) and endothermic (2.5 kcal/mol), and the corresponding change in free energy barrier is 7.4 kcal/mol. The bound complex I4 is then separated into two intermediates viz. oxocarbenium ion I5 and the cyclic carbonate vinyl gold complex I6. The oxocarbenium ion I5 is now susceptible to interact with nucleophiles such as the aglycone (ROH) molecule (B) leading to new complex I7. The nucleophilic attack of alcohol oxygen atom O_{OH}, which is a part of aglycone moiety in I7 on to the anomeric carbon of the oxocarbenium ion, produces a new intermediate I8. The corresponding energy values are given in Scheme 6. Subsequently, I8 interacts with the vinyl gold complex I6 to from a weakly bound complex I9. The HOMO of I9 (Figure S9a) is \Box -MO of C=C bond with

comparatively higher coefficient on C_{AuL}. Note that the natural charge on C_{AuL} is -0.64 e, whereas that in the other carbon atom of C=C is slightly positive (0.20 e). Hence, this promotes the electrophilic addition of positively charged H_{SbF6-} (0.55 e) to electron rich C_{AuL} . This results in the cleavage of Au- C_{AuL} σ bond, leading to the formation of Aualkene π complex I10 and the disaccharide molecule C. The product formation step is highly exothermic as well as exergonic ($\Delta E = -30.4$ kcal/mol and $\Delta G = -41.3$ kcal/mol). The alkynyl group in alkynyl glycosyl carbonate A now competes with the alkene in I10 to coordinate with Au(I) and leads to the formation of I2, and thus the catalytic cycle continues. This step is exothermic by -0.2 kcal/mol, and endergonic by 3.2 kcal/mol and the corresponding kinetic energy barrier is $\Delta E^{\#} = 6.3$ kcal/mol, and $\Delta G^{\#} = 20.5$ kcal/mol. The reaction mechanism for the glycosidation reaction catalyzed by silvlene-supported gold-benzene complex 2 is given in Scheme S6. The extent of elongation of C≡C bond length in I2 (1.257 Å) is larger than that in I2' (1.244 Å). Hence, the C≡C in NHC supported Au(I)-alkyne complex (I1) is more activated by the donation of π electrons from an alkyne to Au as well as a greater back donation from Au \rightarrow alkyne in comparison with Au(I)-alkyne complex supported by silvlene (I2'). The activation of alkyne, and thereby formation of intermediate I3 is more favorable in the case of reaction catalyzed by 8. The barrier for the formation of similar intermediate I3', catalyzed by 2 is highly energy demanding. Both step-wise, as well as the concerted mechanism, indicate that the energy barrier for the product formation step is high in the case of reaction catalyzed by 2. The DFT studies on the glycosidation reaction between glycosyl carbonate A and an aglycone molecule **B** (ROH) catalyzed by N-heterocyclic carbene/silylene supported Au(1)-benzene complexes 8 and 2, indicate that the NHC supported complex 8 is better catalyst than 2.



Scheme S6: Feasible mechanism for glycosidation reaction by remote activation in the presence of **2**. ΔE and $\Delta E^{\#}$ (in kcal/mol) represent reaction enthalpy and enthalpy of activation respectively at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. ΔG and $\Delta G^{\#}$ (in kcal/mol) represent the Gibbs free energy of reaction and Gibbs free energy of activation respectively, at same level of theory at 298.15 K and 1 atm.



Figure S4. a) Experimental and b) optimized (BP86/def2-SVP) geometry of complexes 2 ($L^{1}L^{2}SiAuBz^{+}$, where $L^{1} = N(SiMe_{3})_{2}$, $L^{2} = (Ph)C(Nt-Bu)_{2}$ and $Bz = C_{6}H_{6}$), (c) Experimental and d) optimized (BP86/def2-SVP) geometry of 8 (NHCAu(Bz)⁺, NHC=N-heterocyclic carbene with (i-Pr)₂Ph substituent on each N.



Figure S5. Energy profile diagram for glycosidation reaction by remote activation in presence of **8** as per the mechanism given in Scheme 7. Energies are given in kcal/mol at M06/PCM/Def2-TZVPP//BP86/Def2-SVP level of theory. Activation energy for each step is also given above in kcal/mol.



Figure S6. Energy profile diagram for (Scheme S2) of glycosidation reaction by remote activation in presence of **8**. Energies are given in kcal/mol at M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory (DCM solvent). Activation energy for each step is given above in kcal/mol.



Figure S7. Energy profile diagram for glycosidation reaction by remote activation in presence of **2** as per the mechanism given in Scheme S6. Energies are given in kcal/mol at M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. Activation energy for each step is given above in kcal/mol.



Figure S8. Energy profile diagram for of glycosidation reaction by remote activation in presence of **2** as per the mechanism given in Scheme S3. Energies are given in kcal/mol at M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. Activation energy for each step is given above in kcal/mol.



Figure S9. Important molecular orbitals of at M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory of a) **I9** and b) **I11** c) **I6'** and d) **I8'**. Eigen values (eV) are given in parenthesis.



TS3

Figure S10. Optimized geometries and important geometrical parameters of reactants, intermediates and transition states at the BP86/def2-SVP level of theory. Distances are given in angstroms and angles in degrees.





Figure S11. Optimized geometries and important geometrical parameters of intermediates, transition states and products at the BP86/def2-SVP level of theory. Distances are given in angstroms and angles in degrees.



Figure S12. Optimized geometries and important geometrical parameters of intermediates, transition states and products at the BP86/def2-SVP level of theory. Distances are given in angstroms and angles in degrees.



gure S13. Optimized geometries and important geometrical parameters of intermediates and transition states at the BP86/def2-SVP level of theory. Distances are given in angstroms and angles in degrees.



Figure S14. Optimized geometries and important geometrical parameters of intermediates and transition states mentioned at the BP86/def2-SVP level of theory. Distances are given in angstroms and angles in degrees.


Figure 15: The low lying π^* -molecular orbitals centered on C=C in a) A and b) I2 at the M06/PCM/def2-TZVPP//BP86/def2-SVP level of theory. Eigen values (eV) are given in parenthesis.

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Fig S15. IR Spectrum of 9 in solid state



Fig S16. IR Spectrum of 10 in solid state



¹³C NMR Spectrum (100.67 MHz, CDCl₃) of compound 2





¹H NMR spectrum (400.31 MHz, CDCl₃) of compound **3**



²⁹Si NMR Spectrum (79.495 MHz, CDCl₃) of **2**





²⁹Si NMR Spectrum (79.495 MHz, CDCl₃) of **3**





¹³C NMR Spectrum (100.67 MHz, CDCl₃) of compound 4









¹³C NMR Spectrum (100.67 MHz, CDCl₃) of compound 6







¹H NMR spectrum (400.31 MHz, CDCl₃) of compound 9

²⁹Si NMR Spectrum (79.495 MHz, CDCl₃) of 9



³¹P NMR spectrum (161.976 MHz, CDCl₃) of **10**

5.0 4.5 f1 (ppm)

4.0 3.5

3.0

2.5

6.0

5.5

9.5

8.5

8.0

7.5

7.0 6.5

9.0

1.5

1.0 0.5

2.0





¹³C NMR Spectrum (100.67 MHz, CDCl₃) of compound **12b**



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of compound 12b





\ 127.8 127.8 127.8 127.8 128.4 83.4 83.4 22.6 73.5 72.1 72.1 103.2 - 87.0 37.0 36.7 25.0 69.4 138.0 137.7 137.2 151.4 180 C 170 30 90 f1 (ppm) 160 150 140 130 120 110 100 80 70 60 50 40 20 10

¹³C NMR Spectrum (100.67 MHz, CDCl₃) of Compound **12d**







¹³C NMR Spectrum (100.67 MHz, CDCl₃) of Compound 14a



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of Compound 14a





¹³C NMR Spectrum (100.67 MHz, CDCl₃) of Compound 14b



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of Compound 14b





DEPT NMR Spectrum (100.67 MHz, CDCl₃) of Compound 14c





¹H NMR Spectrum (400.31 MHz, CDCl₃) of Compound 14d

¹³C NMR Spectrum (100.67 MHz, CDCl₃) of Compound 14d



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of Compound 14d





¹H NMR Spectrum (400.31 MHz, CDCl₃) of Compound **14f**

^{13}C NMR Spectrum (100.67 MHz, CDCl_3) of Compound 14f



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of Compound 14f





¹³C NMR Spectrum (100.67 MHz, CDCl₃) of Compound **17**



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of Compound 17


¹H NMR Spectrum (400.31 MHz, CDCl₃) of compound **18**



 ^{13}C NMR Spectrum (100.67 MHz, CDCl₃) of compound 18



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of compound 18



¹H NMR Spectrum (400.31 MHz, CDCl₃) of compound **19**



¹³C NMR Spectrum (100.67 MHz, CDCl₃) of compound **19**



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of compound 19



¹H NMR Spectrum (400.31 MHz, CDCl₃) of compound **20**



¹³C NMR Spectrum (100.67 MHz, CDCl₃) of compound 20



DEPT NMR Spectrum (100.67 MHz, CDCl₃) of compound 20

