

Electronic Supplementary Information for

A selenium-coordinated palladium(II) *trans*-dichloride molecular rotor as a catalyst for site-selective annulation of 2-arylimidazo[1,2-*a*]pyridines

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EXPERIMENTAL SECTION

General. The ligand and complex were synthesized using standard Schlenk techniques. The catalysis reactions were carried out in pressure tube under open air conditions. HPLC grade DMF, CH₂Cl₂, EtOH, EtOAc, hexane, CH₃CN, DMAc, DMSO and xylene were used directly as received. Salicylaldehyde (Sigma Aldrich), benzyl bromide (Spectrochem), K₂CO₃ (Spectrochem), selenium powder (Sigma Aldrich), NaBH₄ (Spectrochem), Na₂SO₄ (Spectrochem), CDCl₃ (Sigma Aldrich), and PdCl₂ (Alfa Aesar, 99.9%) were used as purchased. All other reagents for catalysis reaction were purchased from local commercial sources and used as it is. NMR spectrum were recorded on a Bruker NMRs 400 MHz instruments at ambient probe temperatures and referenced as follows (δ , ppm): ¹H, residual internal CHCl₃ (7.26); ¹³C{¹H}, internal CDCl₃ (77.00). IR spectrums of new ligand and complex were taken on a Nicolet Protége 460 FT-IR spectrometer on KBr pellets. Melting points of new complexes were recorded in an open capillary. HRMS measurements were carried out by electrospray ionization (ESI) method on an Agilent Q-TOF LCMS spectrometer.

Synthesis of bis(2-(benzyloxy)benzyl)selane (2). A three necked round bottom flask was charged with selenium powder (4.00 mmol, 0.316 g) in 40 mL ethanol, and fitted with condenser under N₂ atmosphere. The mixture was refluxed and solid NaBH₄ (0.310 g, 8.2 mmol) was added in small portions until the mixture turned into colorless solution. A solution of 1-(benzyloxy)-2-(chloromethyl)benzene (**1**, 0.465 g, 2.00 mmol) in C₂H₅OH (5 mL) was added drop wise with stirring. After 12 h mixture was cooled to room temperature, solvent was reduced (~2 mL) by using rotary evaporated. The residue was placed at the top of a silica column (3 × 14 cm), which was eluted with hexanes (150 mL) and then hexanes/EtOAc (90:10 v/v). The solvent was evaporated from the product containing fractions (assayed by TLC) by rotary evaporation to give **2** as a yellow oil (0.768 g, 1.622 mmol, 81%). Anal. Calcd for C₂₈H₂₆O₂Se (473.47): C, 71.03; H, 5.54. Found: C, 70.92; H, 5.47.

NMR (CDCl₃, δ /ppm): ¹H (400 MHz) 7.47-7.45 (m, 2H), 7.38-7.31 (m, 3H), 7.19-7.15 (m, 2H), 6.91-6.83 (m, 2H), 5.01 (s, 2H, OCH₂), 3.87 (s, 2H, SeCH₂); ¹³C{¹H} (100 MHz) 156.2 (s),

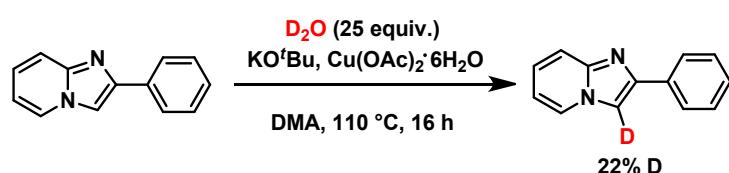
137.2 (s), 130.2 (s), 128.9 (s), 128.5 (s), 127.8 (s), 127.7 (s), 127.2 (s), 120.6 (s), 111.9 (s), 69.9 (s, OCH₂), 22.3 (s, SeCH₂). **IR** (cm⁻¹, powder film): 3216 (w), 3050 (w), 2875 (w), 1597 (s), 1489 (s), 1450 (s), 1242 (s), 694 (s). HRMS (ESI) calcd for C₂₈H₂₇O₂Se [M + H]⁺: 475.1176, found 475.1129.

Palladium complex of bis(2-(benzyloxy)benzyl)selane (3). A round bottom flask was charged with **2** (0.473 g, 1.00 mmol), PdCl₂(CH₃CN)₂ (0.130 g, 0.50 mmol), acetonitrile (15 mL), and fitted with a condenser. The mixture was stirred (2 h) while refluxing. The mixture was then cooled, passed through a short pad of silica gel (18 cm) and washed with CH₃CN (25 mL). The solvent was removed by rotary evaporation and residue was washed with cold *n*-pentane to give **3** as a yellow solid (0.417 g, 0.371 mmol, 74%). m.p: 296-298 °C. Anal. Calcd for C₅₆H₅₂Cl₂O₄PdSe₂ (1124.25): C, 59.83; H, 4.66. Found: C, 59.91; H, 4.58.

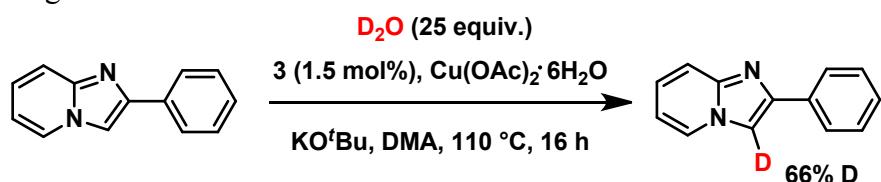
NMR (CDCl₃, δ/ppm): ¹H (400 MHz) 7.44-7.39 (m, 3H), 7.36-7.28 (m, 3H), 7.24-7.18 (m, 1H), 6.86-6.81 (m, 2H), 5.00 (s, 2H, OCH₂), 4.56 (d, 2H, *J* = 11.2 Hz, SeCH₂), 4.14 (d, 2H, *J* = 11.2 Hz, SeCH₂); ¹³C{¹H} (100 MHz) 156.6 (s), 136.8 (s), 131.7 (s), 129.2 (s), 128.5 (s), 127.6 (s), 127.1 (s), 124.0 (s), 120.7 (s), 111.8 (s), 69.8 (s, OCH₂), 30.6 (s, SeCH₂). **IR** (cm⁻¹, powder film): 3192 (m), 2914 (w), 2211 (m), 1645 (s), 1527 (s), 1210 (s), 1032 (s), 755 (s). HRMS (ESI) calcd for C₂₈H₂₇O₂Se [M - PdCl₂ + H]⁺: 475.1176, found 475.1118.

Procedure for Oxidative Annulation Reaction: A pressure tube was charged with 2-aryl imidazo[1,2-*a*]pyridine (0.250 mmol), 1,2-diarylethyne (1.1 equiv.), oxidant (2 equiv.), base (2 equiv.), and **3** (1.5 mol%) in 3 mL solvent. After that, the reaction mixture was refluxed at 110 °C for 16 h. The progress of reaction was monitored by TLC and reaction was quenched with aq. NH₄Cl. The reaction mixture was extracted with ethyl acetate (15 mL × 3). The crude product was purified by chromatography on silica-gel to afford annulated products **6a-6n** and **7o**.

Deuterium exchange experiment:



A 10 mL RBF was charged with 2-phenylimidazo[1,2-*a*]pyridine (1 equiv.), D₂O (25 equiv.), KO'Bu (2 equiv.), Cu(OAc)₂.6H₂O (2 equiv.), and DMA (3ml). The mixture was stirred at 110 °C under air for 16 h. The reaction mixture cooled to room temperature, quenched with water (5 mL) and diluted with EtOAc (15 mL). The layers were separated, and the aqueous layer was extracted with 3 x 10 mL of EtOAc. The organic layer is dried over Na₂SO₄, filtered, and concentrated on reduced pressure. The residue was purified by column chromatography on silica gel, eluting with 25-30% EtOAc /hexane. The isolated product was analyzed with ¹H NMR spectrometer and the result shown in figure s11.



A 10 mL RBF was charged with 2-arylimidazo[1,2-*a*]pyridine (1 equiv.), D₂O (25 equiv.), KO'Bu (2 equiv.), Cu(OAc)₂.6H₂O (2 equiv.), **3** (1.5 mol %), and DMA (3ml). The mixture was stirred at 110 °C under air for 16 h. The reaction mixture cooled to room temperature, quenched with water (5 mL) and diluted with EtOAc (15 mL). The layers were separated, and the aqueous layer was extracted with 3 x 10 mL of EtOAc. The organic layer is dried over Na₂SO₄, filtered, and concentrated on reduced pressure. The residue was purified by column chromatography on silica gel, eluting with 25-30% EtOAc/hexane. The isolated product was analyzed with ¹H NMR spectrometer and the result shown in figure s12.

Crystallography

A. A CH₃CN solution of **3** (5 mL) was slowly concentrated over a period of 14 days to afford suitable colorless plate like crystals. The data was collected and reported in Table s1. Cell parameters were obtained from 60 data frames using a 1° scan and refined with 11864 reflections. APEX2 was used to collect the integrated intensity information of each reflection by reducing the data frames.^{s1} Lorentz and polarization corrections were applied. The absorption correction program SADABS^{s2} was employed to correct the data for absorption effects. The space group was determined from systematic reflection conditions and statistical tests. The structure was refined

(weighted least squares refinement on F^2) to convergence.^{s3} Olex2^{s4} was employed for the final data presentation and structure plots. Non-hydrogen atoms were refined with anisotropic thermal parameters. Hydrogen atoms were fixed in idealized positions using a riding model. The absence of additional symmetry or voids was confirmed using PLATON (ADDSYM).^{s5} Molecule possess an inversion symmetry and symmetry generated atoms are not labeled in the Figure 1.

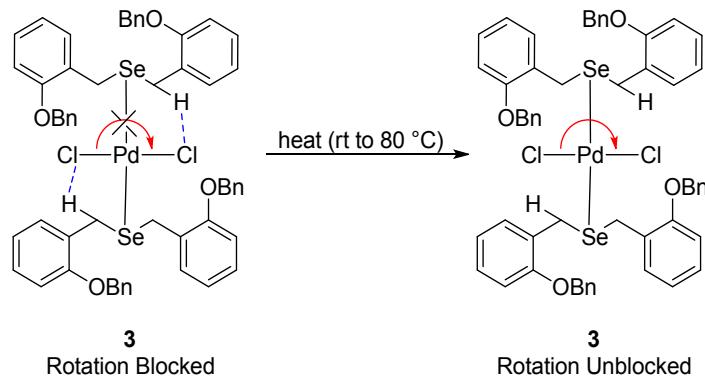
Table s1. Summary of crystallographic data.

3	
empirical formula	C ₅₆ H ₅₂ Cl ₂ O ₄ PdSe ₂
formula weight	1124.19
temperature [K]	110.0
diffractometer	Bruker Apex 2
wavelength [Å]	0.71073
crystal system	Triclinic
space group	P-1
unit cell dimensions:	
<i>a</i> [Å]	8.2319(4)
<i>b</i> [Å]	10.9599(5)
<i>c</i> [Å]	14.1131(8)
α [°]	101.73(2)
β [°]	100.17(2)
γ [°]	97.03(2)
<i>V</i> [Å ³]	1210.44(11)
<i>Z</i>	1
ρ_{calc} [Mg/m ³]	1.542
μ [mm ⁻¹]	2.044
F(000)	568
crystal size [mm ³]	0.10 × 0.08 × 0.01
Θ limit [°]	2.548 to 25.000
index range (<i>h</i> , <i>k</i> , <i>l</i>)	-9, 9, -13, 12, -16, 16
reflections collected	11864
independent reflections	4210
<i>R</i> (int)	0.0273
completeness to θ	98.8
max. and min. transmission	0.3334 and 0.2686
data/restraints/parameters	4210/0/295
goodness-of-fit on F^2	1.060
<i>R</i> indices (final) [$I > 2\sigma(I)$]	
<i>R</i> ₁	0.0253
<i>wR</i> ₁	0.0491
<i>R</i> indices (all data)	
<i>R</i> ₁	0.0352
<i>wR</i> ₂	0.0547
largest diff. peak and hole [eÅ ⁻³]	0.471 and -0.363

Table s2. Key crystallographic distances [Å] and angles [°].

3	
Pd-Se1	2.4391(3)
Pd-Cl1	2.2869(6)
Se-C1	1.986(3)
Se-C15	1.984(2)
Cl-Pd-Se	87.85(17) 92.15(17)
Cl-Pd-Cl	180.0
Se-Pd-Se	180.0

The proposed dynamic process in 3 and calculation of barriers from experiment:



Scheme 1: The proposed dynamic process in 3.

$$\Delta G^\ddagger = \Delta H^\ddagger - T\Delta S^\ddagger$$

$$\Delta E^\ddagger \approx \Delta H^\ddagger - P\Delta V^\ddagger$$

$$P\Delta V^\ddagger \approx 0 \text{ and } \Delta E^\ddagger \approx \Delta H^\ddagger$$

$T\Delta S^\ddagger \approx 0$ Since the two hybrids stay exactly *trans* and the change in the number of degrees of freedom is approximately zero; therefore

$$\Delta G^\ddagger = \Delta E^\ddagger$$

Minimum rate (k_{min}) at 298.15 K is 26 s^{-1} .

$$k_{min} \leq \frac{k_B T}{h} e^{-\Delta E^\ddagger / RT} = k$$

$$26 \leq 2.08366179 * 10^{10} * 298.15 * e^{-\Delta E^\ddagger / (1.9872 * 298.15)}$$

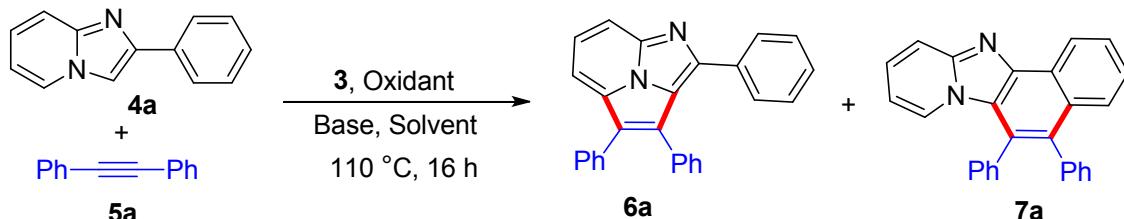
$$\ln(26) \leq \ln(2.08366179 * 10^{10} * 298.15) - \frac{\Delta E^\ddagger}{(1.9872 * 298.15)}$$

$$3.25809653802 \leq 29.4575744674 - \frac{\Delta E^\ddagger}{(592.48368)}$$

$$(29.4575744674 - 3.25809653802) * (592.48368) \leq \Delta E^\ddagger$$

$$\Delta E^\ddagger \leq 15.52 \text{ kcal/mol}$$

Table s3: Optimization of reaction conditions for cycloaromatization of 2-arylimidazo[1,2-*a*]pyridines (**4a**) with diphenylacetylene (**5a**)^a.



Entry no.	Catalyst (mol%)	Oxidant	Base	Solvent	Yield% (6a) ^b	Yield% (7a) ^b
1	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	—	DMAc	31	nd
2	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	K ₂ CO ₃	DMAc	58	nd
3	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMAc	67	nd
4	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	K ₃ PO ₄	DMAc	22	32
5	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	Na ₂ CO ₃	DMAc	11	nd
6	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	DIPEA	DMAc	03	nd
7	3 (1 mol%)	AgNO ₃	KO'Bu	DMAc	22	nd
8	3 (1 mol%)	AgOAc	KO'Bu	DMAc	52	nd
9	3 (1 mol%)	Ag ₂ CO ₃	KO'Bu	DMAc	26	nd
10	3 (1 mol%)	IBD	KO'Bu	DMAc	21	27
11	3 (1 mol%)	TBHP	KO'Bu	DMAc	nr	nr
12	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMF	36	23
13	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMSO	54	19
14	3 (1 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	Xylene	08	nd
15	3 (1.5 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMAc	74	nd
16	3 (2.0 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMAc	63	13
17 ^c	3 (1.5 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMAc	52	nd
18 ^d	3 (1.5 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMAc	55	nd
19	3 (1 mol%)	—	KO'Bu	DMAc	nd	nd
20	—	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMAc	nd	nd
21 ^e	3 (1.5 mol%)	Cu(OAc) ₂ .H ₂ O	KO'Bu	DMAc	13	nd

^aReaction conditions unless specified otherwise: **4a** (0.250 mmol), **5a** (1.1 equiv), oxidant (2 equiv), base (2 equiv), solvent (3 mL). nd = not determined, nr = no reaction. ^bIsolated yield. ^cat 90 °C. ^dat 10 h. ^eCu(OAc)₂.H₂O (10 mol%).

Computational Studies:

Initial geometry was taken from crystal structure **3** and optimized using QUICKSTEP module^{s6} of CP2k program package.^{s7} We used Perdew, Burke, and Ernzerhof (PBE) exchange–correlation functional (PBE XC functionals) and Goedecker-Teter-Hutter (GTH) pseudopotential^{s8} with MOLOPT type DZVP basis set^{s9} with an energy cut-off of 280 Ry. One of Cl-atom of optimized geometry **3** was further substituted as expected intermediate **A** to optimize *cis*- and *trans*- isomers. A visual inspection/representation was obtained using visual molecular dynamics program (VMD 1.9.2)^{s10} and the distances between particular atom/ring and centre of mass of aromatic ring was drawn using Tcl scripting language embedded in VMD.

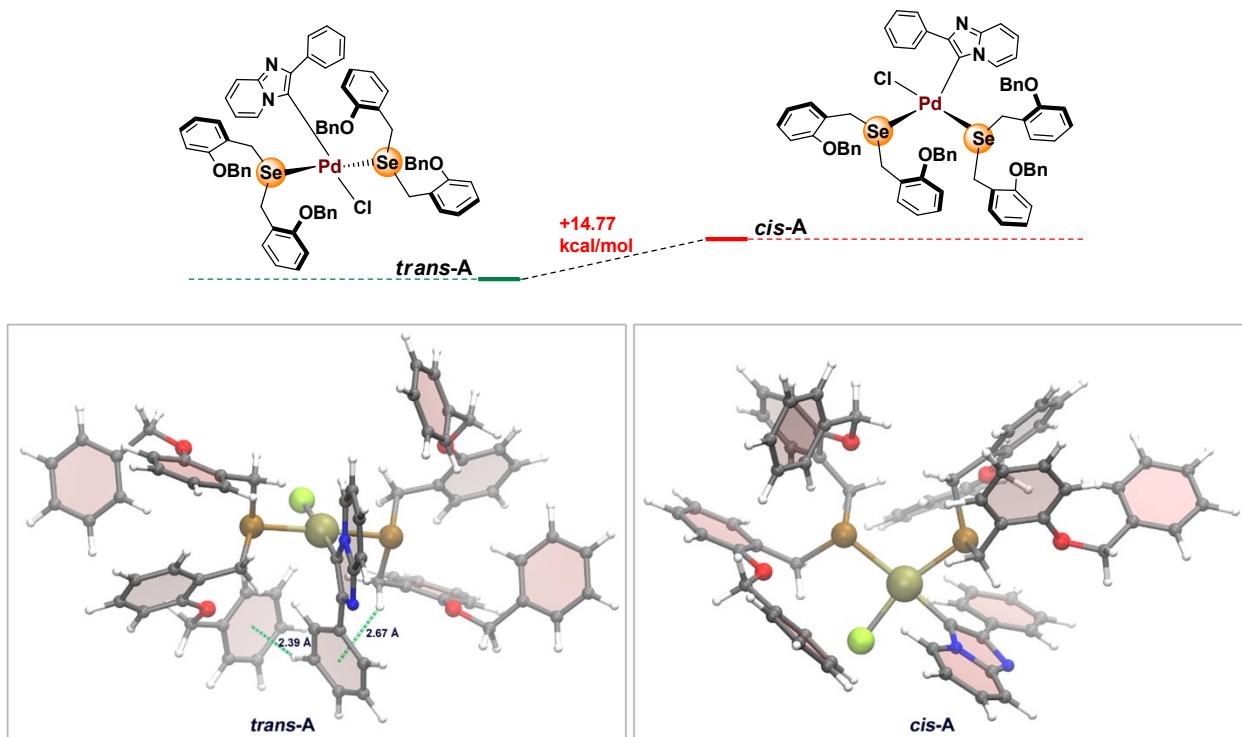


Figure s1: Geometry optimized structure of *trans*-A ($\text{CH}-\pi$ distances 2.39 Å and 2.67 Å) and *cis*-A isomer. [Color scheme: Pd-Tan (CPK), Se-Ochre (CPK), Cl-Lime (CPK), C-Gray, H-White, N-Blue (Licorice); aromatic rings-Glass1 (Paperchain)]

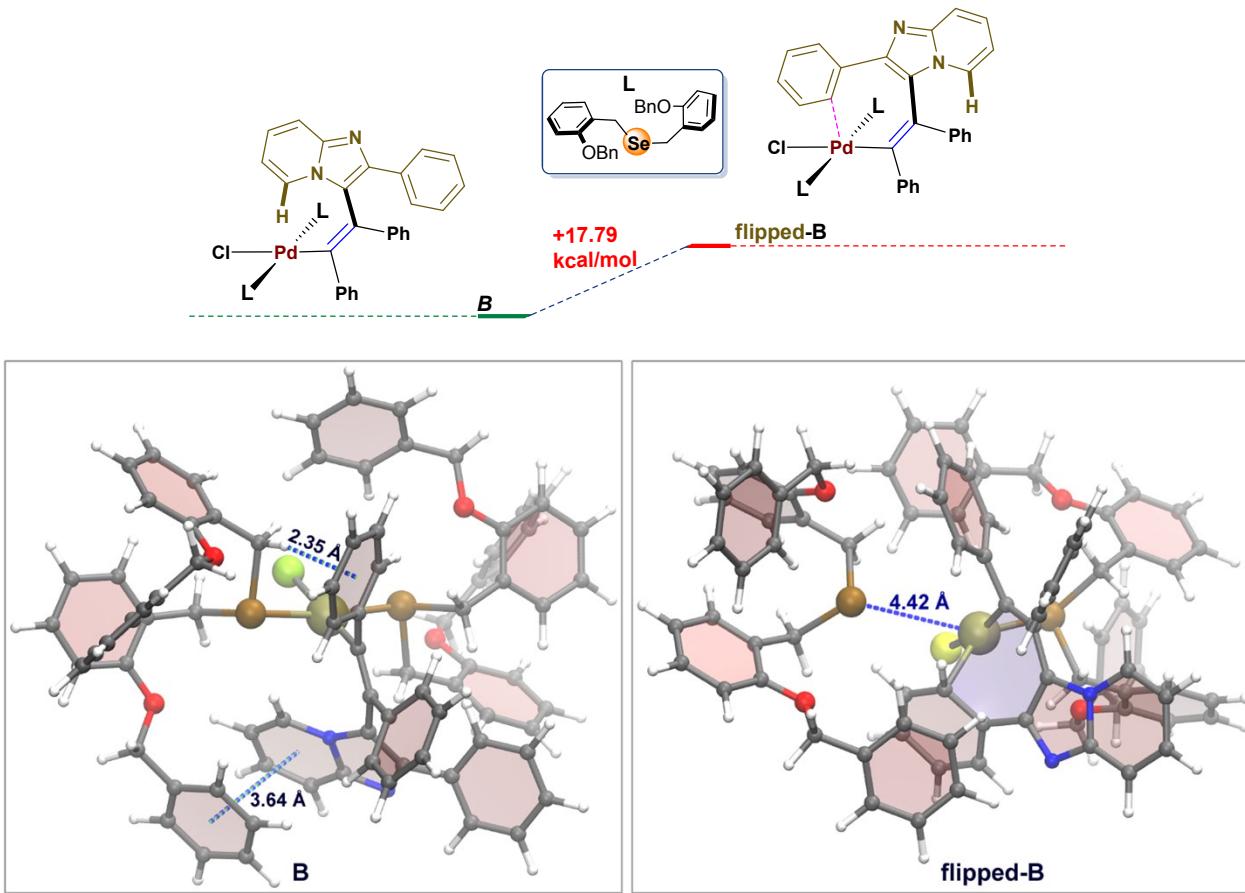
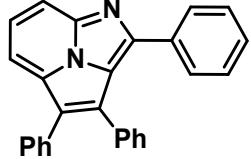


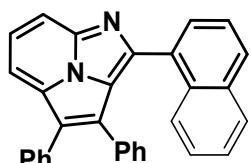
Figure s2: Geometry optimized structure of **B** ($\text{CH}-\pi$ distance 2.35 \AA and $\pi-\pi$ distance 3.64 \AA) and flipped-2-arylimidazo[1,2-*a*]pyridines of **B** (flipped-**B**). [Color scheme: Pd-Tan (CPK), Se-Ochre (CPK), Cl-Lime (CPK), C-Gray, H-White, N-Blue (Licorice); aromatic rings- Glass1 (Paperchain)]

In case of optimized structure of configuration **B**, $\text{CH}-\pi$ interactions^{s11} at distance 2.35 \AA and $\pi-\pi$ interactions at distance 3.64 \AA are observed. Whereas, flipped-2-arylimidazo[1,2-*a*]pyridines of **B** optimization leads to 7 membered ring formation with breaking of Pd-Se bond having a distance of 4.42 \AA . $\text{CH}-\pi$ interactions and $\pi-\pi$ interactions are not seen in between 2-arylimidazo[1,2-*a*]pyridines and **L** in **flipped-B** configuration of the complex and less favored with relatively higher energy i.e. 17.79 kcal/mol compared to configuration **B**.

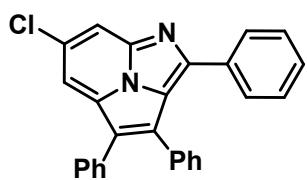
Spectroscopic data of products:



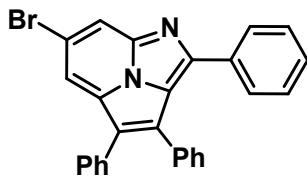
2,3,4-Triphenylimidazo[5,1,2-cd]indolizine (6a):^{s12} Yellow solid, m.p: 208-210 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.09 – 8.02 (m, 1H), 8.01 – 7.95 (m, 2H), 7.83 (d, *J* = 7.3 Hz, 2H), 7.47 (d, *J* = 7.3 Hz, 2H), 7.45 – 7.39 (m, 4H), 7.39 – 7.31 (m, 5H), 7.29 (d, *J* = 7.7 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 149.7 (s), 139.1 (s), 133.6 (s), 133.6 (s), 132.6 (s), 132.1 (s), 132.0 (s), 131.0 (s), 130.2 (s), 129.8 (s), 129.8 (s), 128.7 (s), 128.6 (s), 128.5 (s), 128.4 (s), 127.8 (s), 127.3 (s), 124.0 (s), 113.2 (s), 111.3 (s). HRMS (ESI) calcd for C₂₇H₁₉N₂ [M + H]⁺: 371.1548, found 371.1594.



2-(Naphthalen-1-yl)-3,4-diphenylimidazo[5,1,2-cd]indolizine (6b):^{s12} Yellow solid, m.p: 223-225 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.20 (dd, *J* = 8.6, 1.7 Hz, 1H), 8.15 (s, 1H), 8.11 (dd, *J* = 6.6, 1.9 Hz, 1H), 8.06 – 8.00 (m, 2H), 7.86 – 7.81 (m, 2H), 7.56 – 7.47 (m, 6H), 7.47 – 7.38 (m, 6H), 7.38 – 7.32 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 151.0 (s), 140.2 (s), 134.4 (s), 133.9 (s), 133.9 (s), 133.1 (s), 131.6 (s), 131.1 (s), 130.9 (s), 130.2 (s), 130.0 (s), 128.7 (s), 128.7 (s), 128.7 (s), 128.3 (s), 128.0 (s), 127.7 (s), 127.5 (s), 127.1 (s), 127.1 (s), 126.8 (s), 126.4 (s), 126.2 (s), 124.7 (s), 112.9 (s), 111.4 (s). (one carbon is not visible due to overlapping peaks)

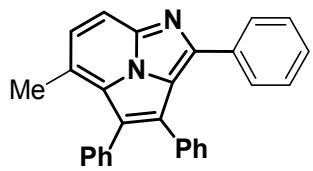


6-Chloro-2,3,4-triphenylimidazo[5,1,2-cd]indolizine (6c):^{s12} Brown solid, m.p: 222-224 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.18 (s, 1H), 7.94 (d, *J* = 7.3 Hz, 3H), 7.84 (s, 1H), 7.59 (d, *J* = 9.5 Hz, 2H), 7.45 (t, *J* = 7.6 Hz, 4H), 7.37 – 7.29 (m, 4H), 7.15 (d, *J* = 9.4 Hz, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 149.5 (s), 139.9 (s), 133.9 (s), 133.7 (s), 132.6 (s), 131.8 (s), 131.5 (s), 131.0 (s), 130.9 (s), 130.1 (s), 128.7 (s), 128.3 (s), 127.7 (s), 127.2 (s), 127.1 (s), 124.3 (s), 123.8 (s), 113.0 (s), 111.6 (s).

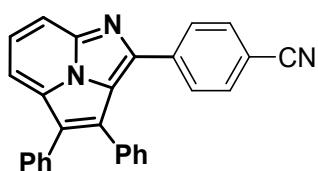


6-Bromo-2,3,4-triphenylimidazo[5,1,2-cd]indolizine (6d): Light brown solid, m.p: 235-237 °C; ¹H NMR (400 MHz, CDCl₃) 8.11 – 8.06 (m, 1H), 8.04 – 8.00 (m, 2H), 7.71 (d, *J* = 8.5 Hz, 2H), 7.53 – 7.47 (m, 3H), 7.47 – 7.38 (m, 8H), 7.38 – 7.32 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 149.82 (s), 140.25 (s), 134.21 (s), 134.03 (s), 132.90 (s), 132.11 (s), 131.86 (s), 131.80

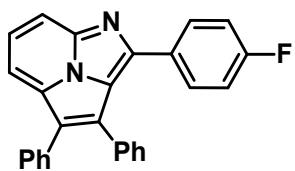
(s), 131.36 (s), 131.20 (s), 130.44 (s), 128.98 (s), 128.66 (s), 127.98 (s), 127.56 (s), 127.46 (s), 124.13 (s), 113.35 (s), 111.91 (s).



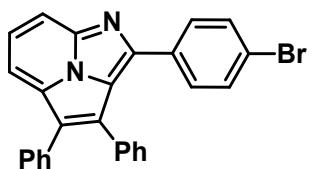
5-Methyl-2,3,4-triphenylimidazo[5,1,2-cd]indolizine (6e): Yellow solid, m.p: 184-186 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.92 – 7.76 (m, 4H), 7.51 – 7.26 (m, 13H), 2.85 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 150.8 (s), 139.9 (s), 138.6 (s), 134.1 (s), 134.1 (s), 133.7 (s), 131.8 (s), 131.3 (s), 131.0 (s), 130.2 (s), 129.6 (s), 129.3 (s), 128.6 (s), 128.5 (s), 128.3 (s), 127.0 (s), 126.7 (s), 123.9 (s), 113.8 (s), 111.9 (s), 22.9 (s).



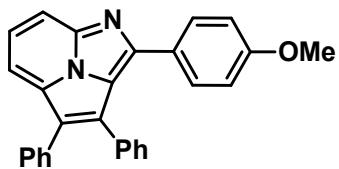
4-(3,4-Diphenylimidazo[5,1,2-cd]indolizin-2-yl)benzonitrile (6f): Orange solid, m.p: 238-240 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.14 – 8.09 (m, 1H), 8.04 (d, *J* = 4.3 Hz, 2H), 7.90 (d, *J* = 8.1 Hz, 2H), 7.55 (d, *J* = 8.1 Hz, 2H), 7.50 – 7.31 (m, 10H); ¹³C NMR (100 MHz, CDCl₃) δ 147.5 (s), 139.6 (s), 137.7 (s), 133.4 (s), 133.1 (s), 131.8 (s), 131.8 (s), 131.2 (s), 130.5 (s), 129.8 (s), 129.6 (s), 128.5 (s), 128.4 (s), 128.3 (s), 128.2 (s), 127.3 (s), 127.1 (s), 124.6 (s), 118.5 (s), 113.4 (s), 112.2 (s), 112.1 (s).



2-(4-Fluorophenyl)-3,4-diphenylimidazo[5,1,2-cd]indolizine (6g):^{s12} Yellow solid, m.p: 203-205 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.14 (d, *J* = 6.6 Hz, 1H), 8.08 – 8.00 (m, 2H), 7.84 (q, *J* = 8.6, 5.5 Hz, 2H), 7.51 – 7.33 (m, 10H), 7.00 (t, *J* = 8.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 163.8 (d, *J*_{C-F} = 258.8 Hz), 149.0 (s), 139.3 (s), 133.8 (s), 133.6 (d, *J*_{C-F} = 17.7 Hz), 131.9 (s), 131.8 (s), 131.7 (d, *J*_{C-F} = 8.4 Hz), 130.9 (s), 130.1 (s), 128.7 (s), 128.6 (s), 128.4 (s), 127.6 (s), 127.2 (s), 115.5 (d, *J*_{C-F} = 21.6 Hz), 113.1 (s), 111.2 (s).

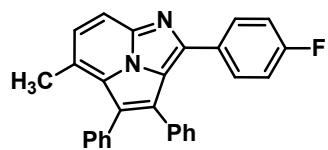


2-(4-Bromophenyl)-3,4-diphenylimidazo[5,1,2-cd]indolizine (6h): Yellow solid, m.p: 221-223 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.08 – 8.03 (m, 1H), 8.02 – 7.97 (m, 2H), 7.69 (d, *J* = 8.5 Hz, 2H), 7.50 – 7.29 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 149.5 (s), 140.0 (s), 133.9 (s), 133.7 (s), 132.6 (s), 131.8 (s), 131.6 (s), 131.0 (s), 130.9 (s), 130.1 (s), 128.7 (s), 128.4 (s), 127.7 (s), 127.2 (s), 127.2 (s), 123.8 (s), 113.0 (s), 111.6 (s).

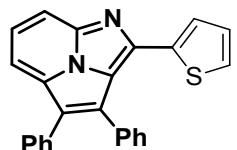


2-(4-Methoxyphenyl)-3,4-diphenylimidazo[5,1,2-cd]indolizine (6i):^{s12} Pale yellow solid, m.p: 198-200 °C; ¹H NMR (400 MHz,

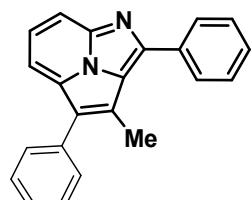
CDCl_3) δ 8.16 (d, $J = 7.4$ Hz, 1H), 8.09 – 7.97 (m, 2H), 7.84 (d, $J = 8.5$ Hz, 2H), 7.52 – 7.34 (m, 10H), 6.85 (d, $J = 8.4$ Hz, 2H), 3.86 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 160.8 (s), 147.6 (s), 140.2 (s), 134.2 (s), 134.1 (s), 131.5 (s), 131.4 (s), 131.1 (s), 131.0 (s), 130.2 (s), 128.6 (s), 128.5 (s), 128.1 (s), 126.9 (s), 126.8 (s), 126.3 (s), 113.8 (s), 112.4 (s), 110.6 (s), 55.3 (s).



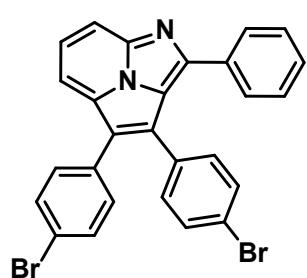
2-(4-Fluorophenyl)-5-methyl-3,4-diphenylimidazo[5,1,2-cd]indolizine (6j): Light brown solid, m.p: 184–186 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.98 (d, $J = 8.1$ Hz, 1H), 7.82 (dd, $J = 8.6, 5.5$ Hz, 2H), 7.74 (d, $J = 8.1$ Hz, 1H), 7.48 – 7.22 (m, 10H), 6.99 (t, $J = 8.7$ Hz, 2H), 2.56 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.6 (d, $J_{C-F} = 247.9$ Hz), 148.1 (s), 138.2 (s), 134.1 (s), 133.6 (s), 131.9 (s), 131.5 (d, $J_{C-F} = 7.4$ Hz), 131.4 (s), 131.1 (s), 130.0 (d, $J_{C-F} = 7.0$ Hz), 129.9 (s), 128.2 (s), 128.0 (s), 127.9 (s), 127.5 (s), 125.6 (s), 123.5 (s), 115.4 (d, $J_{C-F} = 21.5$ Hz), 111.0 (s), 17.8 (s).



3,4-Diphenyl-2-(thiophen-2-yl)imidazo[5,1,2-cd]indolizine (6k):^{s12} Brown solid, m.p: 209–211 °C; ^1H NMR (400 MHz, CDCl_3) δ 8.03 – 7.95 (m, 3H), 7.56 – 7.51 (m, 2H), 7.50 – 7.45 (m, 5H), 7.44 – 7.31 (m, 4H), 7.00 (dd, $J = 3.7, 0.9$ Hz, 1H), 6.96 – 6.92 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 144.9 (s), 140.0 (s), 137.7 (s), 134.1 (s), 133.8 (s), 131.5 (s), 131.1 (s), 130.8 (s), 130.0 (s), 129.1 (s), 128.7 (s), 128.6 (s), 128.5 (s), 128.4 (s), 128.0 (s), 127.5 (s), 127.2 (s), 127.0 (s), 123.4 (s), 112.8 (s), 111.0 (s).

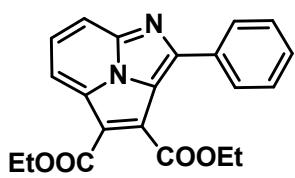


3-Methyl-2,4-diphenylimidazo[5,1,2-cd]indolizine (6l):^{s12} Yellow solid, m.p: 115–117 °C; ^1H NMR (400 MHz, CDCl_3) δ 8.33 – 8.28 (m, 2H), 7.98 (d, $J = 8.0$ Hz, 1H), 7.91 (t, $J = 7.6$ Hz, 1H), 7.84 (d, $J = 7.6$ Hz, 1H), 7.69 – 7.64 (m, 2H), 7.62 – 7.55 (m, 4H), 7.52 – 7.42 (m, 2H), 2.93 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.3 (s), 139.8 (s), 134.4 (s), 134.1 (s), 132.0 (s), 130.0 (s), 129.4 (s), 129.1 (s), 128.9 (s), 128.9 (s), 128.6 (s), 127.5 (s), 127.2 (s), 126.7 (s), 111.3 (s), 110.6 (s), 14.0 (s).



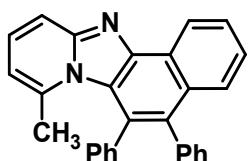
3,4-Bis(4-bromophenyl)-2-phenylimidazo[5,1,2-cd]indolizine (6m): Yellow solid, m.p: 200–202 °C; ^1H NMR (400 MHz, CDCl_3) δ 8.09 (d, $J = 7.7$ Hz, 1H), 8.03 – 7.92 (m, 2H), 7.83 (d, $J = 7.4$ Hz, 2H), 7.59 – 7.48 (m, 4H), 7.44 – 7.25 (m, 7H); ^{13}C NMR (100 MHz, CDCl_3) δ 151.5 (s), 140.1 (s), 133.4 (s), 132.6 (s), 132.57 (s), 132.1 (s), 131.9 (s), 131.7 (s).

(s), 131.3 (s), 130.1 (s), 129.7 (s), 129.67 (s), 128.5 (s), 127.3 (s), 126.0 (s), 123.8 (s), 122.8 (s), 121.6 (s), 112.7 (s), 111.8 (s).



Diethyl 2-phenylimidazo[5,1,2-cd]indolizine-3,4-dicarboxylate (6n):

Yellow solid, m.p: 119-121 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.42 – 8.35 (m, 1H), 8.26 (d, *J* = 7.8 Hz, 2H), 8.16 – 8.09 (m, 2H), 7.63 – 7.50 (m, 3H), 4.60 (q, *J* = 7.1 Hz, 2H), 4.53 (q, *J* = 7.1 Hz, 2H), 1.51 (t, *J* = 7.1 Hz, 3H), 1.45 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.6 (s), 163.3 (s), 154.6 (s), 140.9 (s), 132.9 (s), 130.8 (s), 130.1 (s), 129.4 (s), 129.1 (s), 129.1 (s), 116.4 (s), 116.2 (s), 112.7 (s), 62.7 (s), 61.1 (s), 14.4 (s), 14.1 (s).



8-Methyl-5,6-diphenylnaphtho[1',2':4,5]imidazo[1,2-a]pyridine (7o):^{s12}

Yellow solid, m.p: 271-273 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.10 (d, *J* = 8.0 Hz, 1H), 8.02 (d, *J* = 8.0 Hz, 2H), 7.77 (s, 3H), 7.64 – 7.51 (m, 3H), 7.47 (t, *J* = 7.7 Hz, 4H), 7.41 – 7.31 (m, 3H), 6.74 (d, *J* = 7.7 Hz, 1H), 2.67 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 139.6 (s), 137.9 (s), 134.0 (s), 131.6 (s), 131.6 (s), 131.0 (s), 130.2 (s), 129.6 (s), 129.1 (s), 129.1 (s), 128.6 (s), 128.5 (s), 128.1 (s), 127.0 (s), 126.9 (s), 123.3 (s), 120.8 (s), 114.8 (s), 112.6 (s), 112.2 (s), 111.0 (s), 29.7 (s).

Copies of spectral data

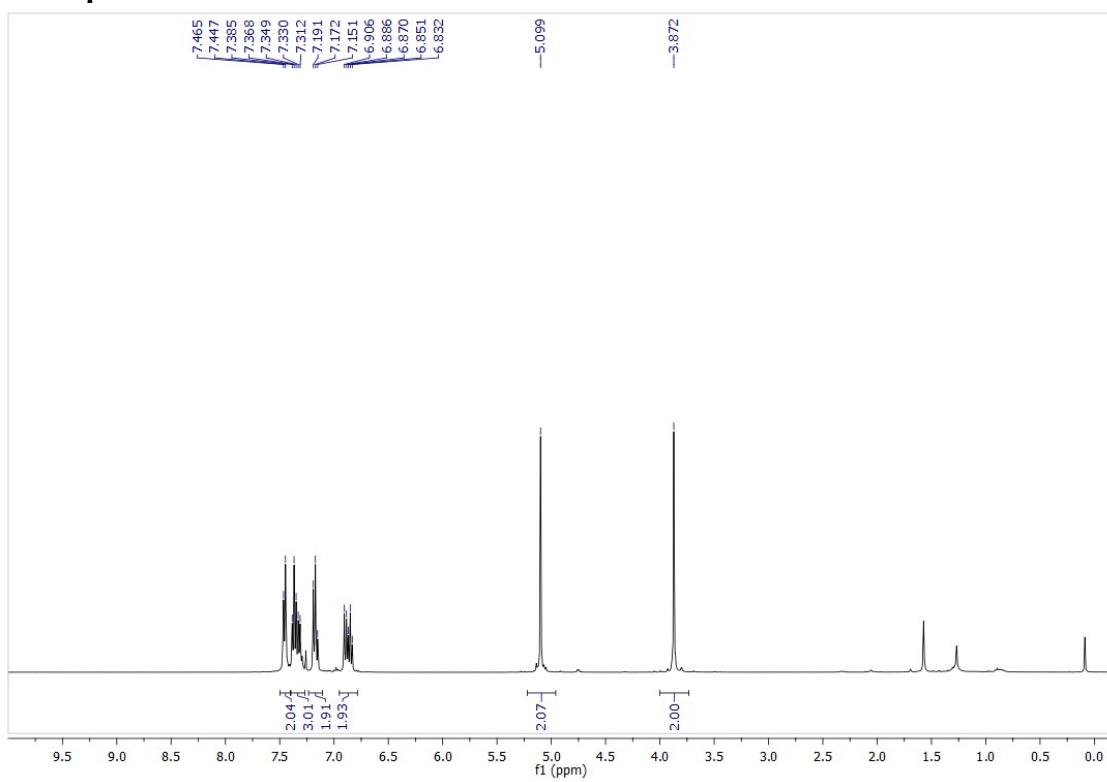


Figure s3. ^1H NMR spectrum of **2**.

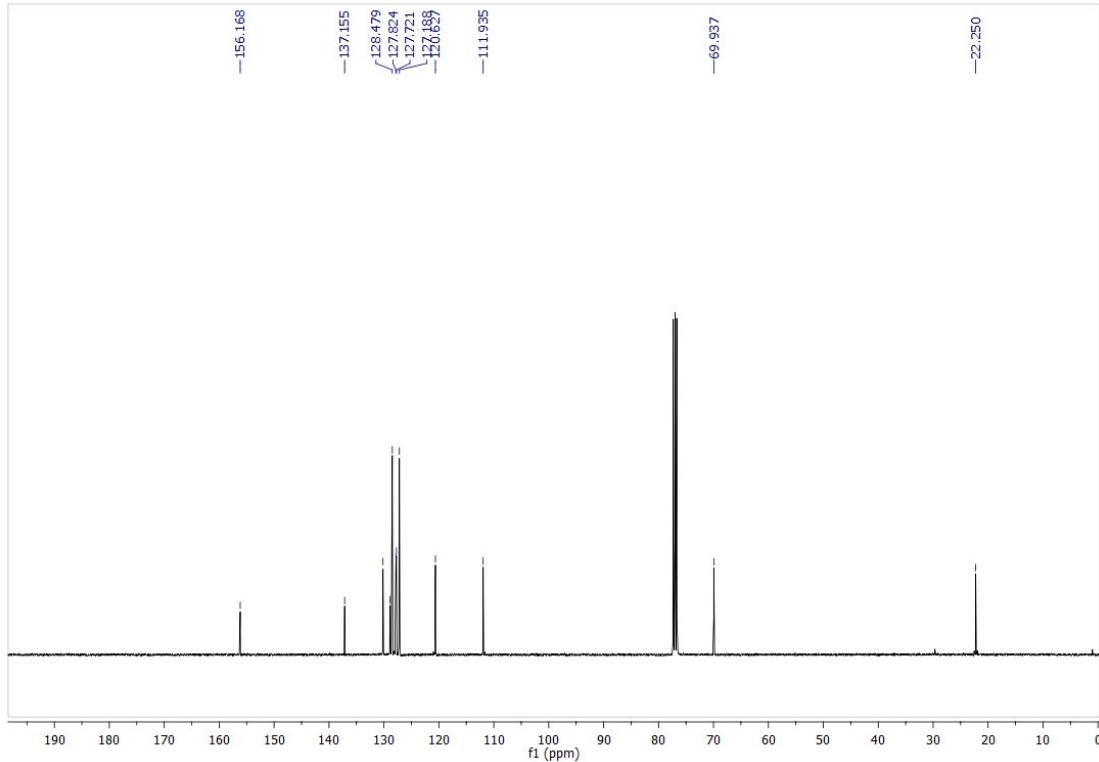


Figure s4. $^{13}\text{C}\{\text{H}\}$ NMR spectrum of **2**.

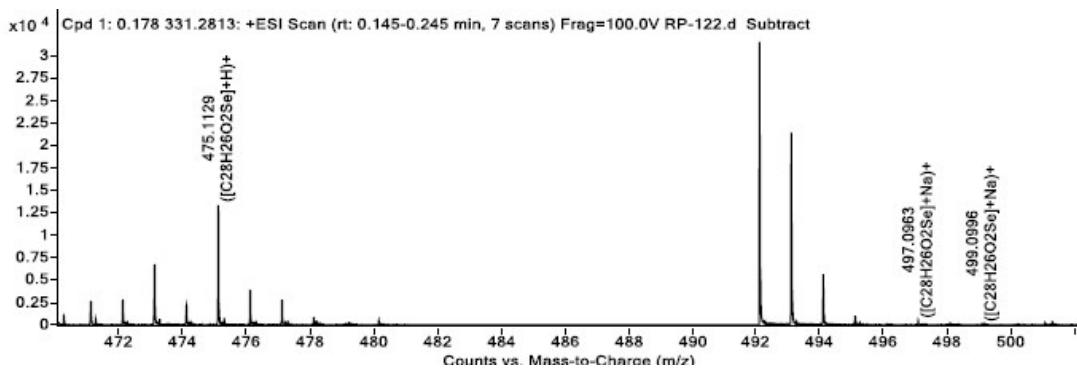


Figure s5. HRMS of **2**.

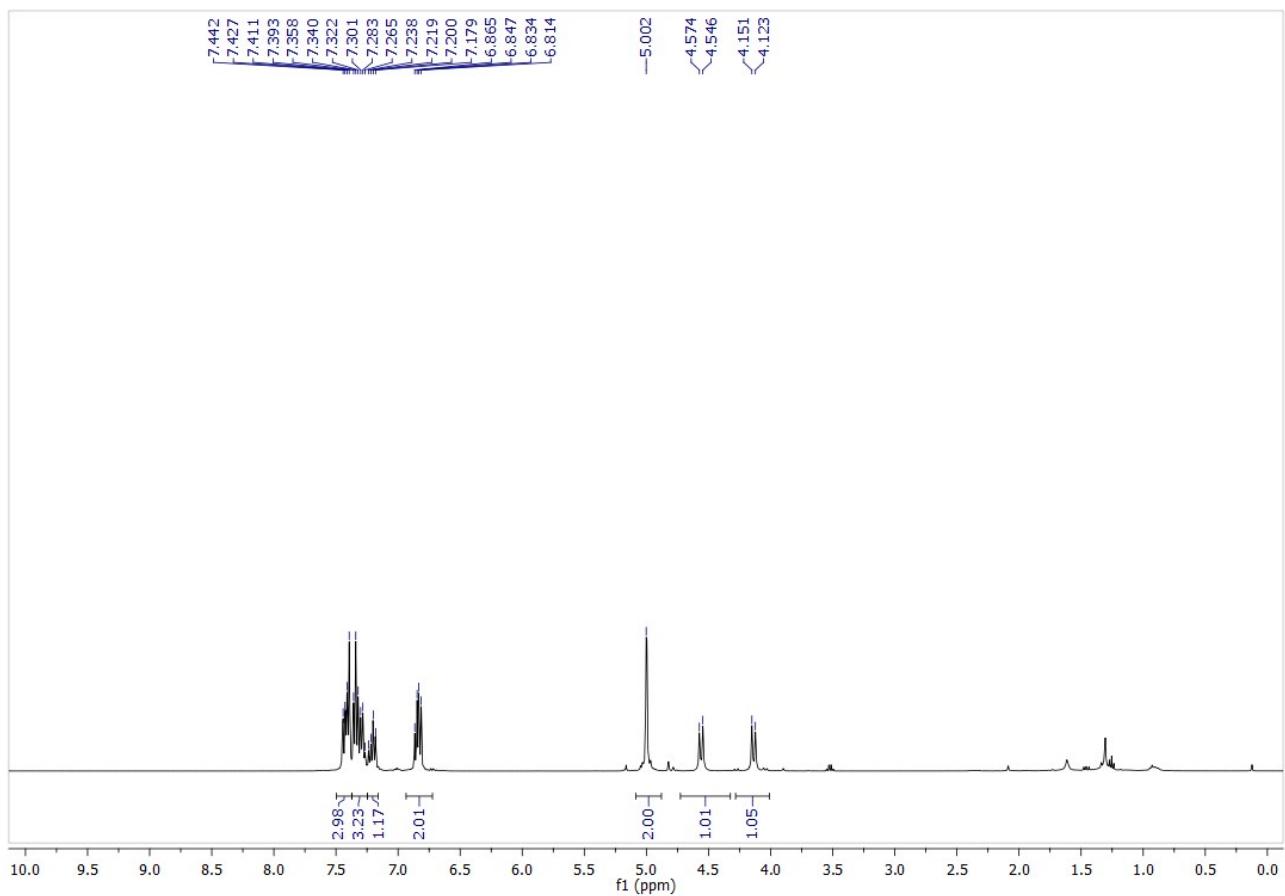


Figure s6. ^1H NMR spectrum of **3**.

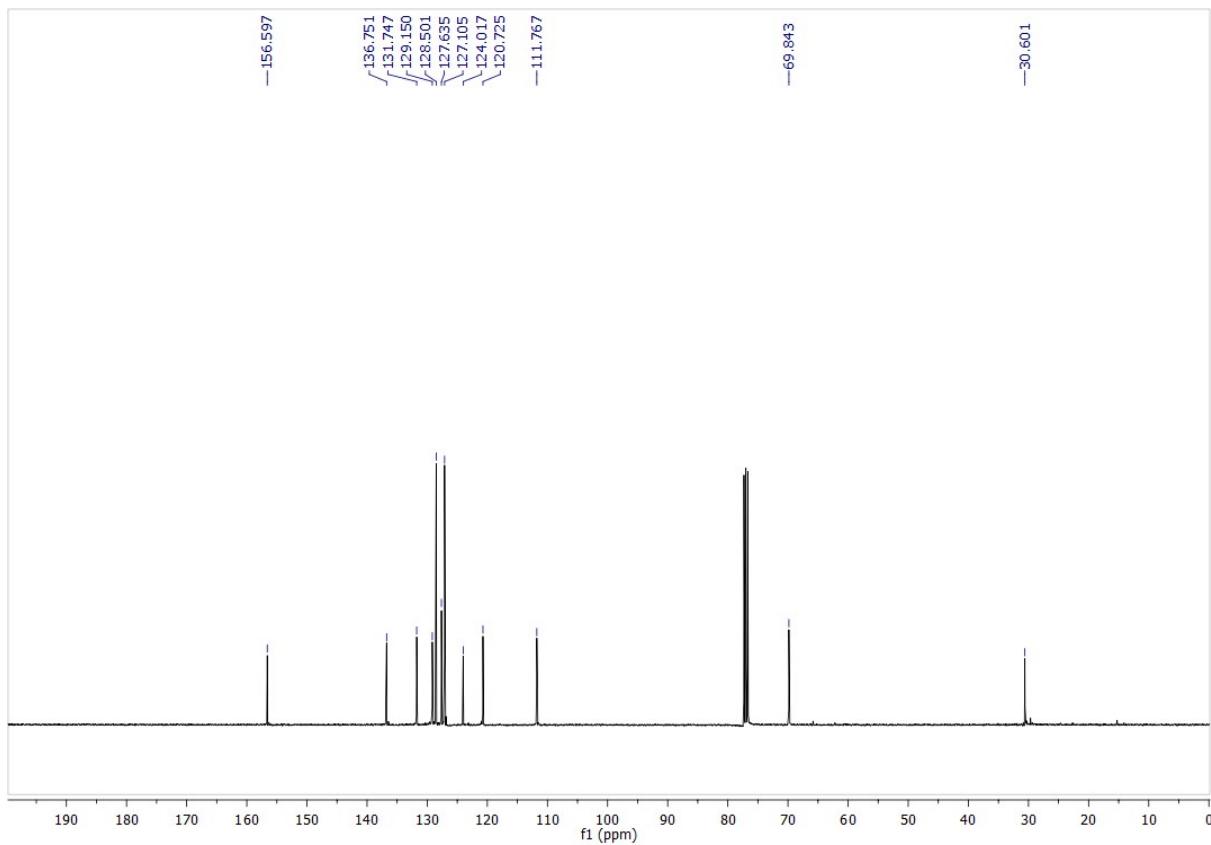


Figure s7. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **3**.

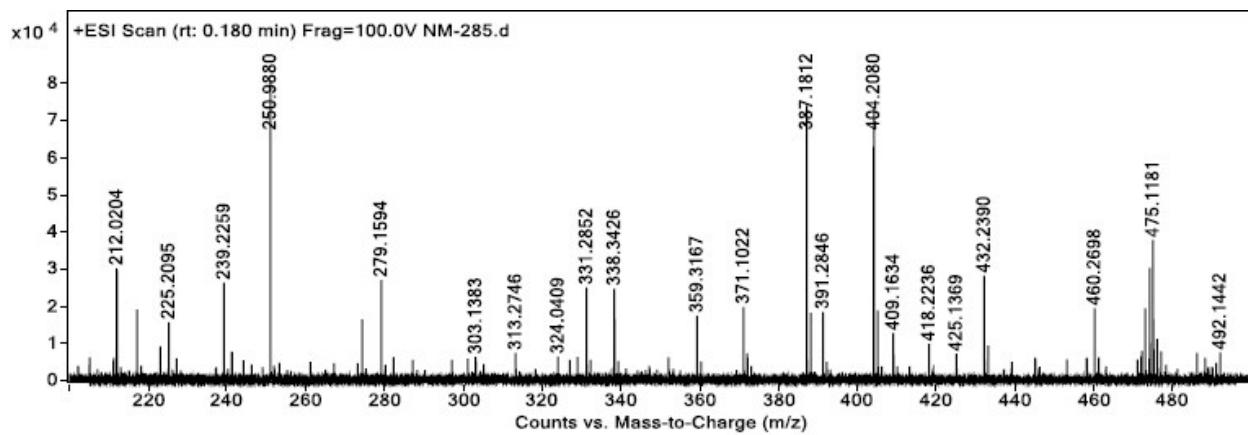


Figure s8. HRMS of **3**.

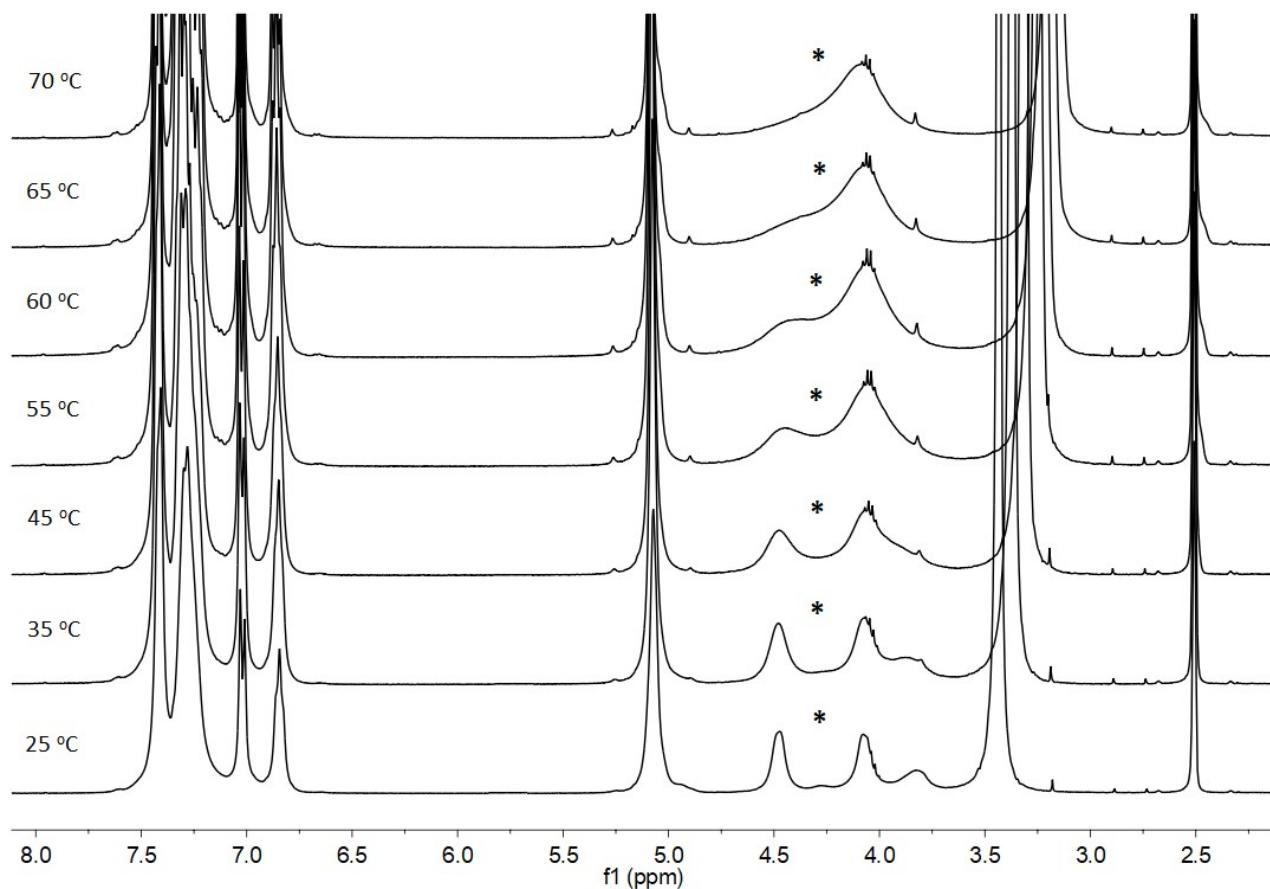


Figure s9. ^1H NMR spectra of **3** as a function of temperature (DMSO- d_6): Coalescing signals are denoted with a *.

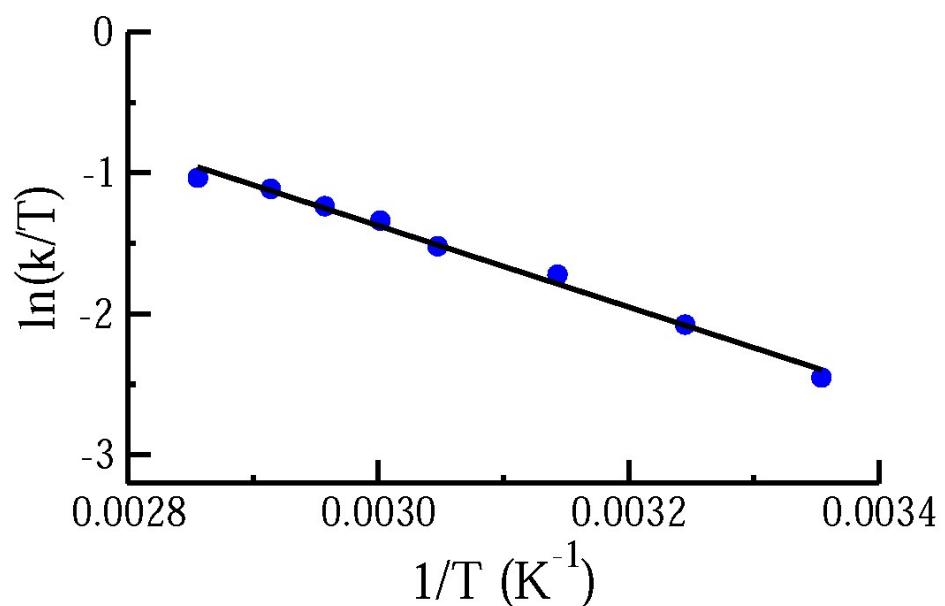


Figure s10. Eyring plot using rate constants derived from Figure 3 for the dynamic process of **3**.

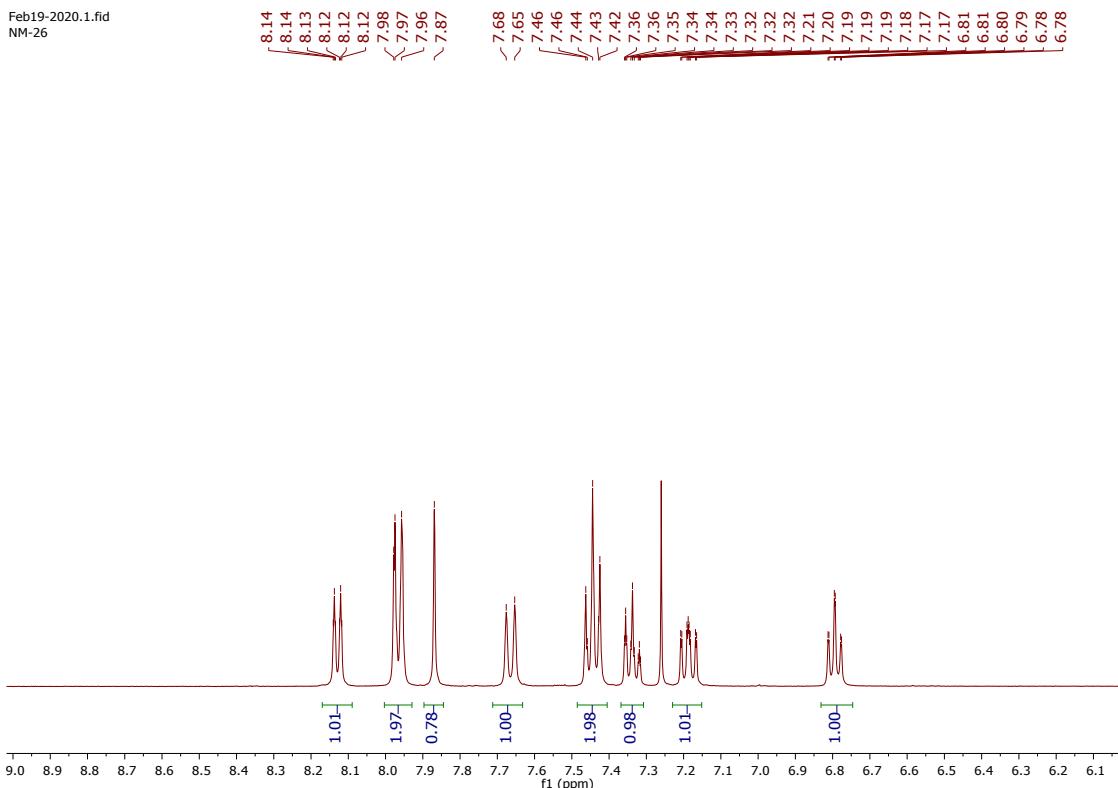


Figure s11. ^1H NMR spectrum of deuterium exchange experiment in absence of catalyst.

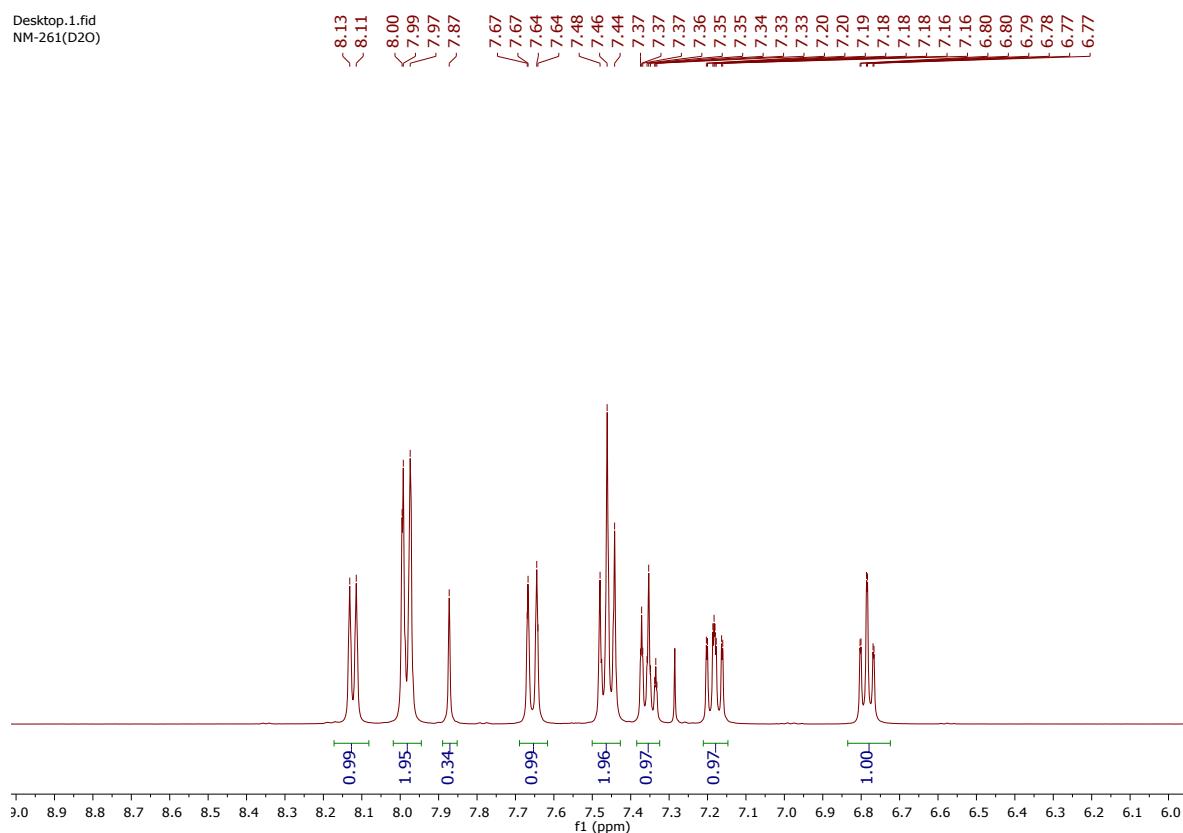


Figure s12. ^1H NMR spectrum of deuterium exchange experiment in presence of catalyst.

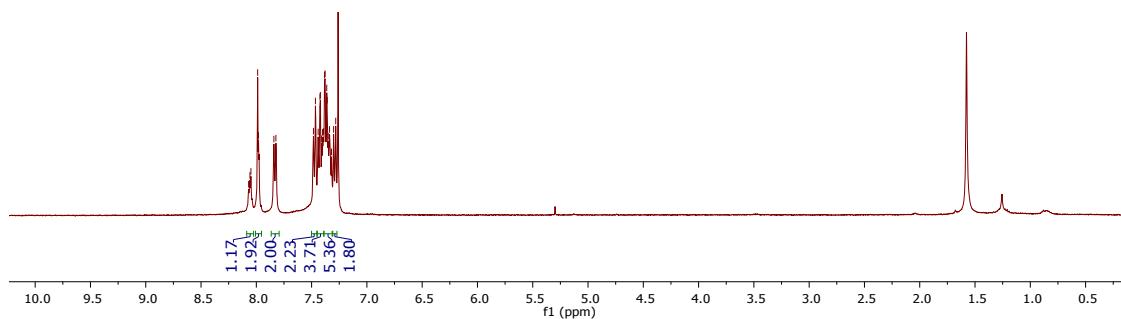
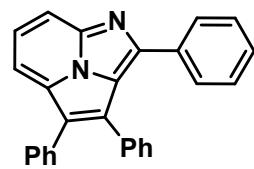


Figure s13. ^1H NMR spectrum of **6a**.

149.6708
139.1053
133.6482
133.5830
132.6226
132.0826
131.9523
130.9530
130.1609
129.8049
129.7973
128.7135
128.6054
128.4866
128.3609
127.7838
127.2791
123.9611
113.2087
~ 111.2607

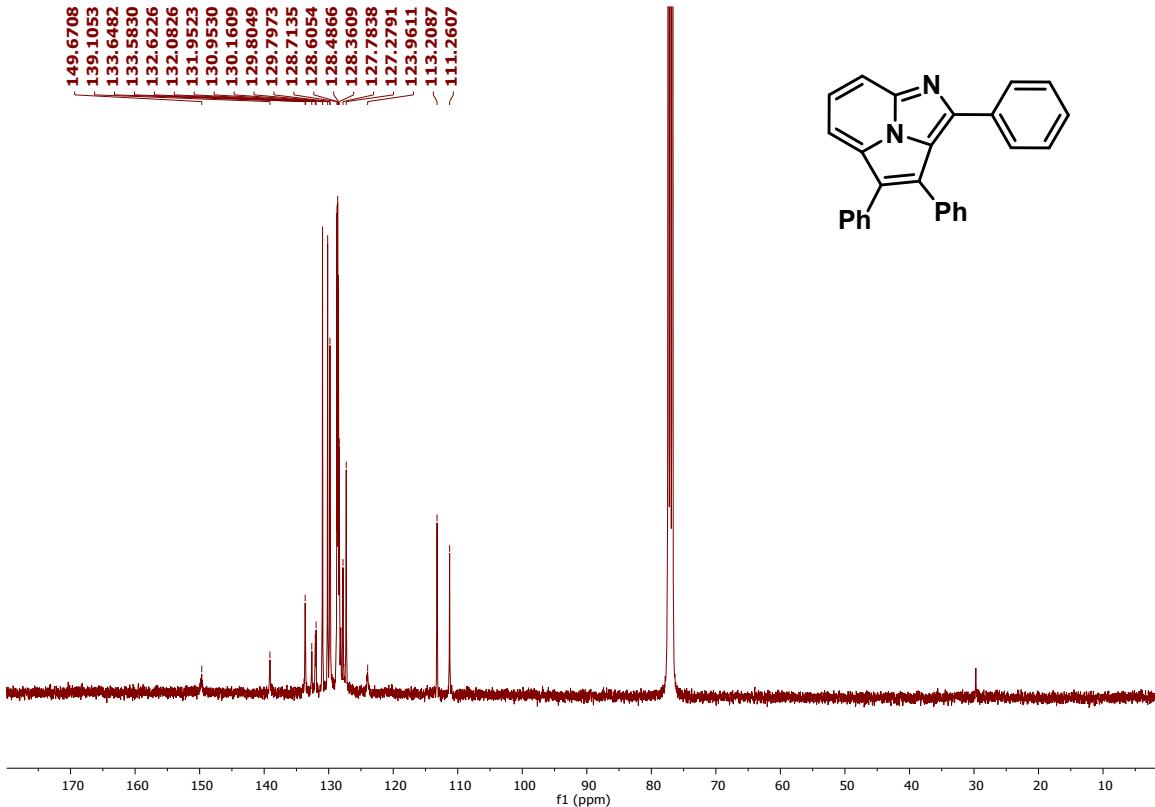
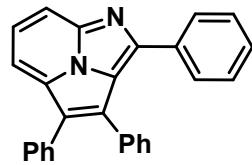


Figure s14. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6a**.

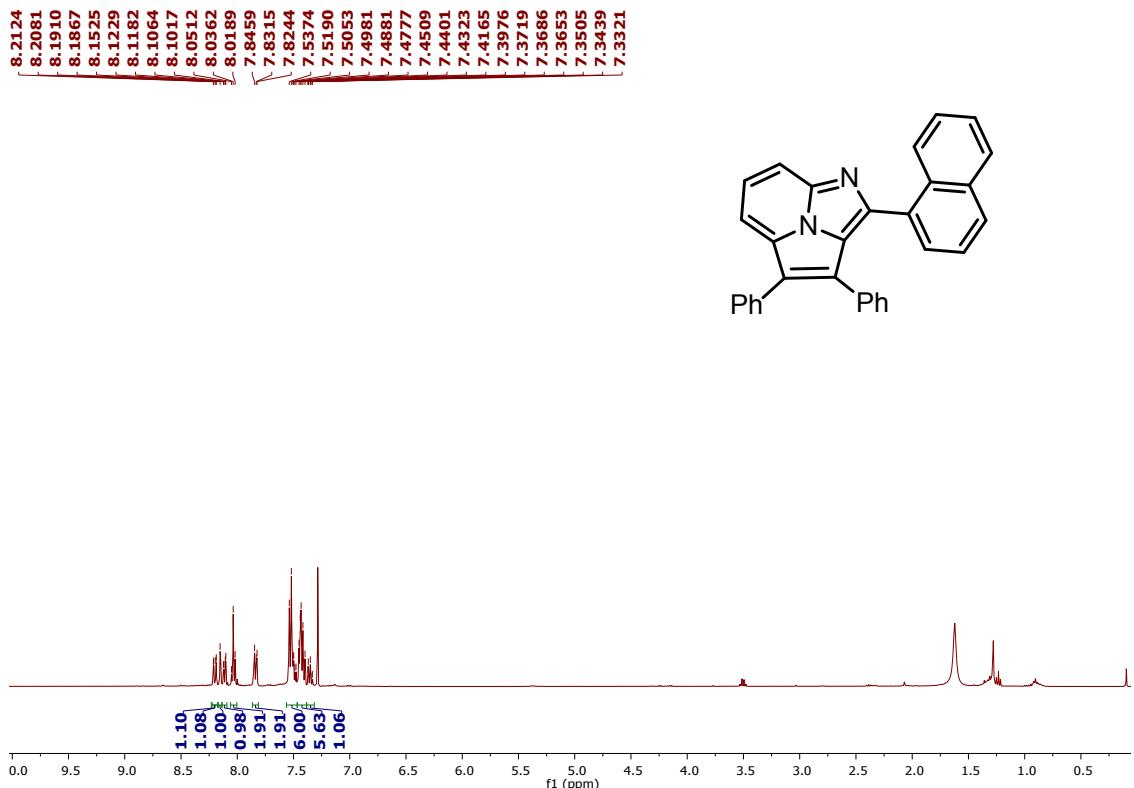


Figure s15. ^1H NMR spectrum of **6b**.

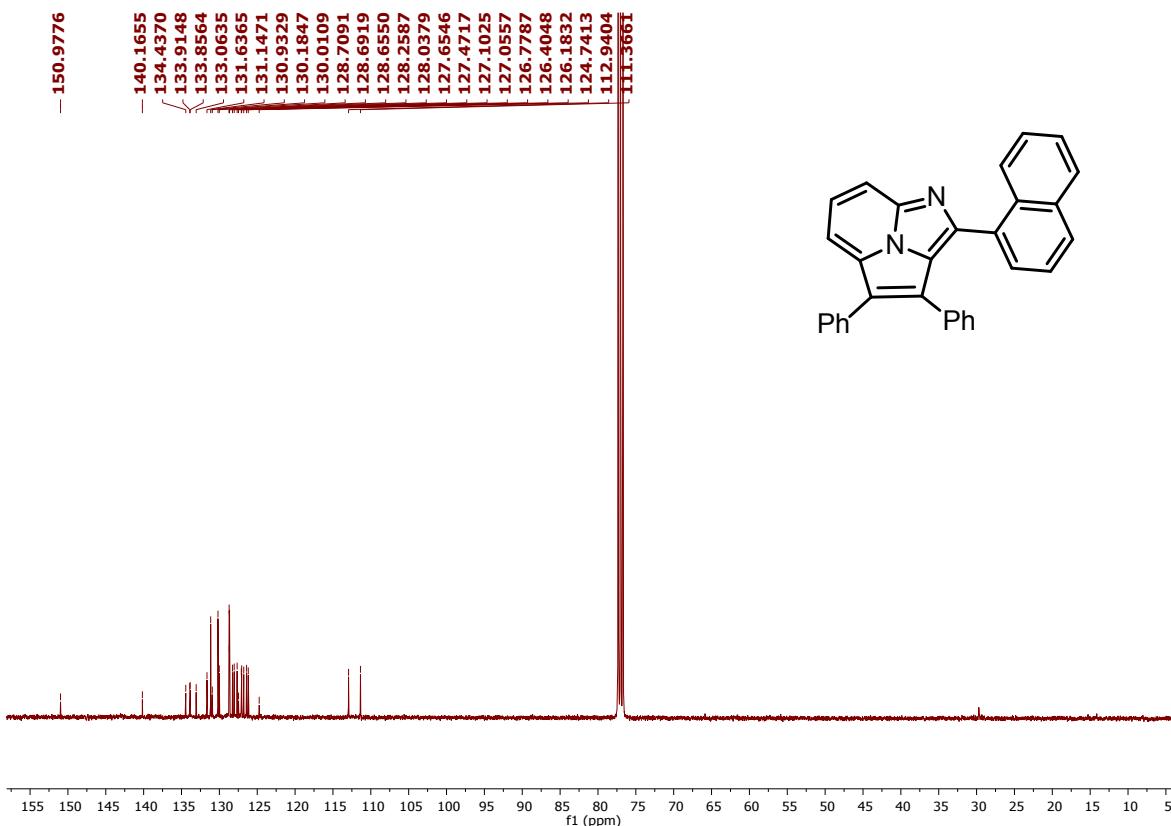


Figure s16. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6b**.

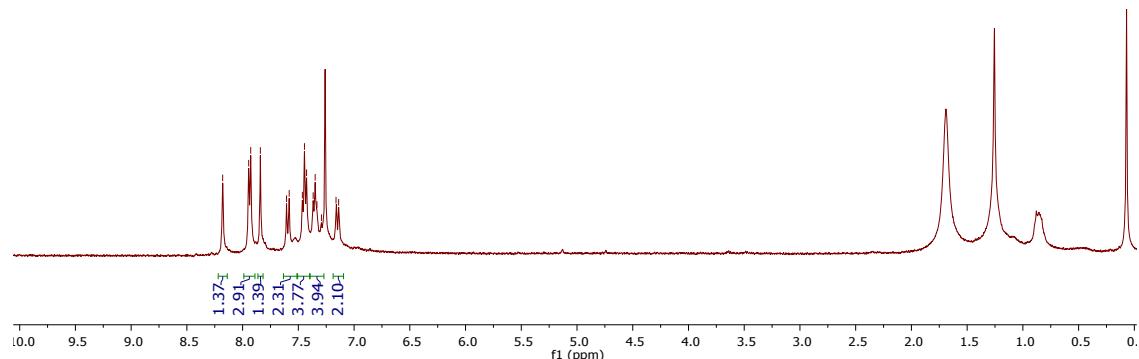
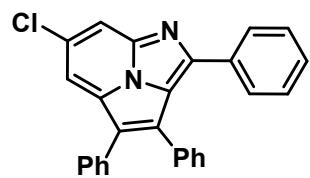


Figure s17. ¹H NMR spectrum of 6c.

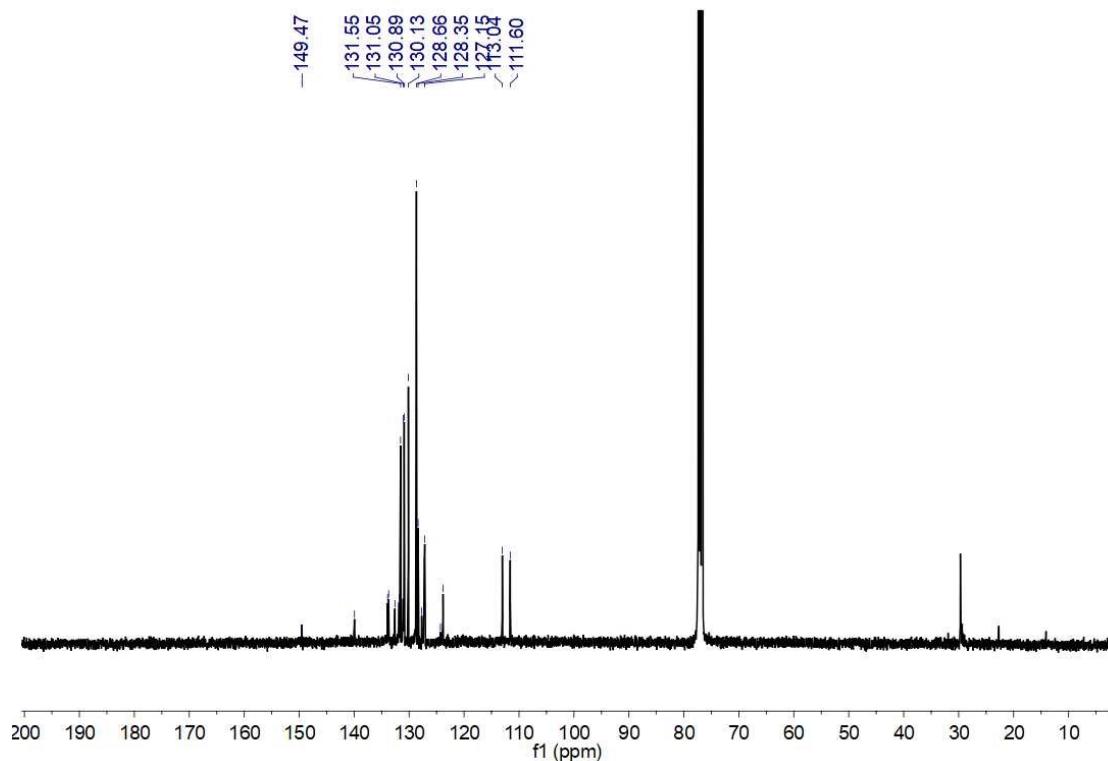


Figure s18. ¹³C{¹H} NMR spectrum of 6c.

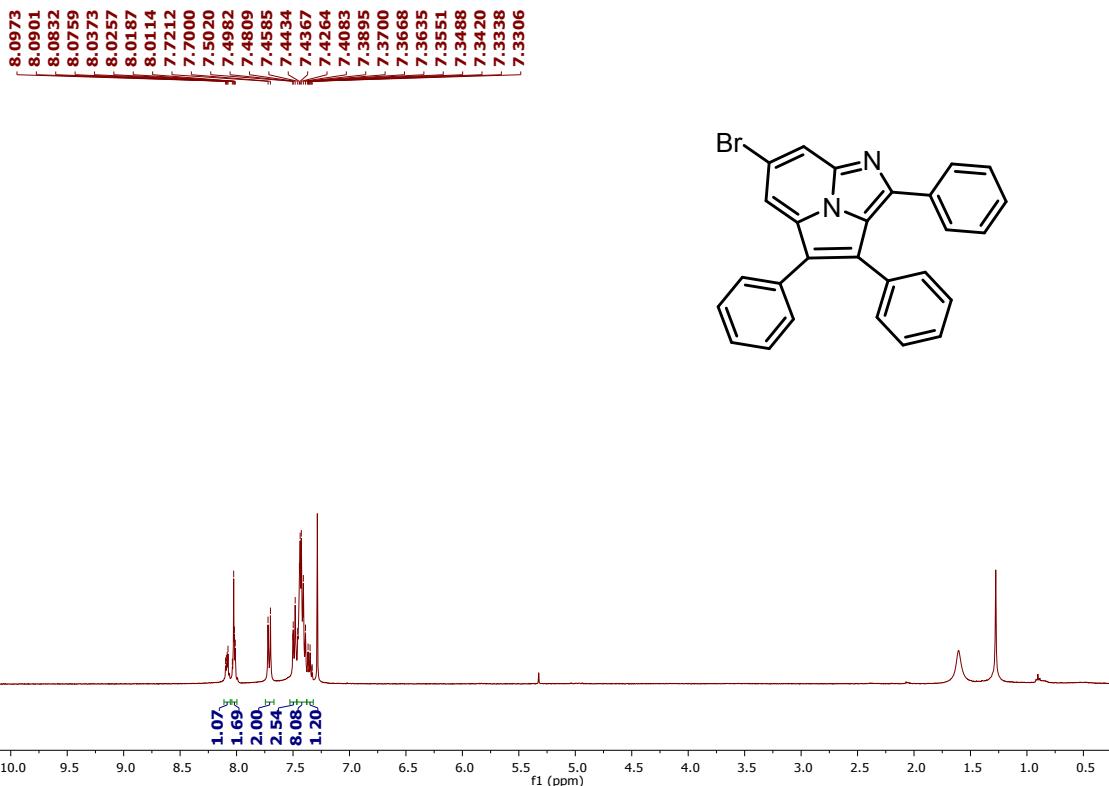


Figure s19. ^1H NMR spectrum of **6d**.

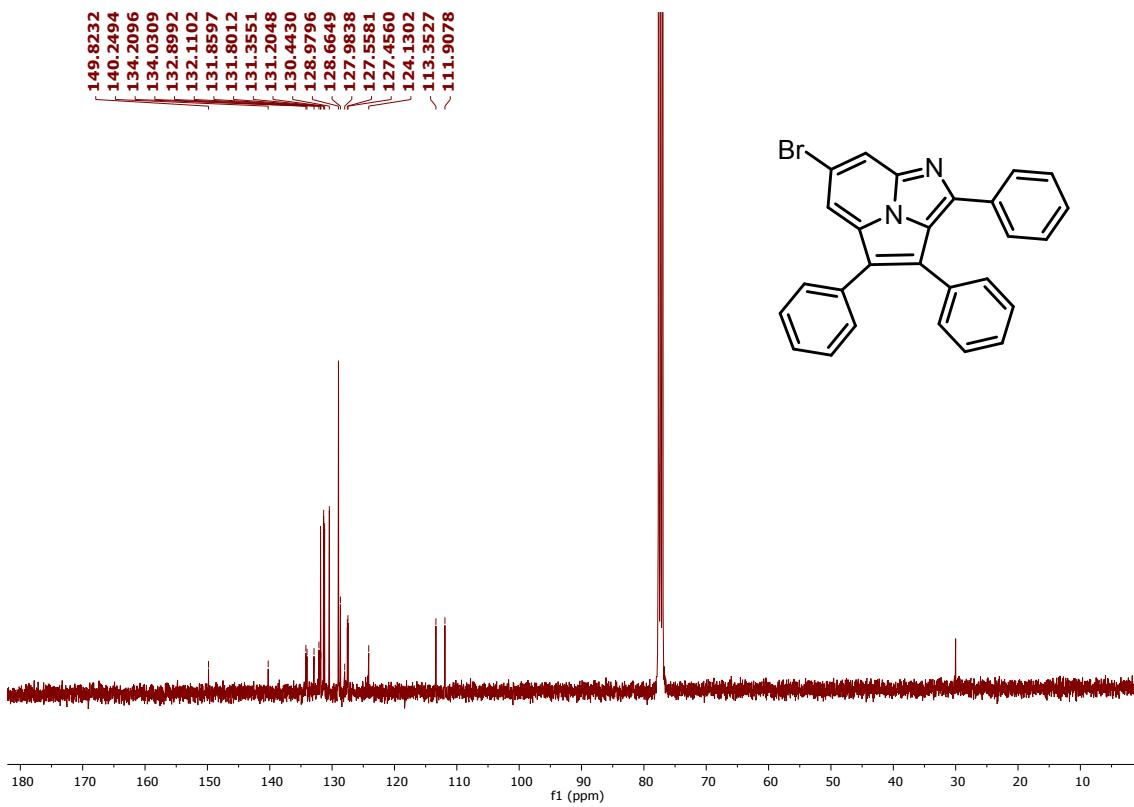


Figure s20. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6d**.

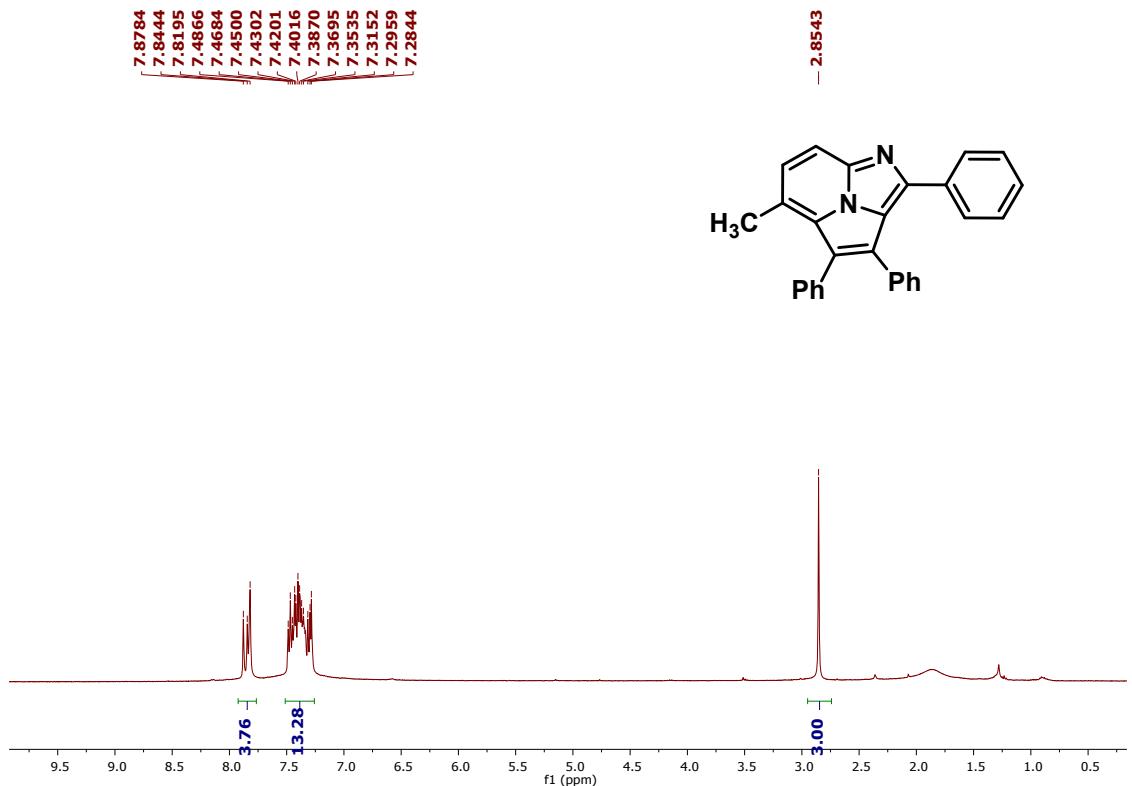


Figure s21. ^1H NMR spectrum of **6e**.

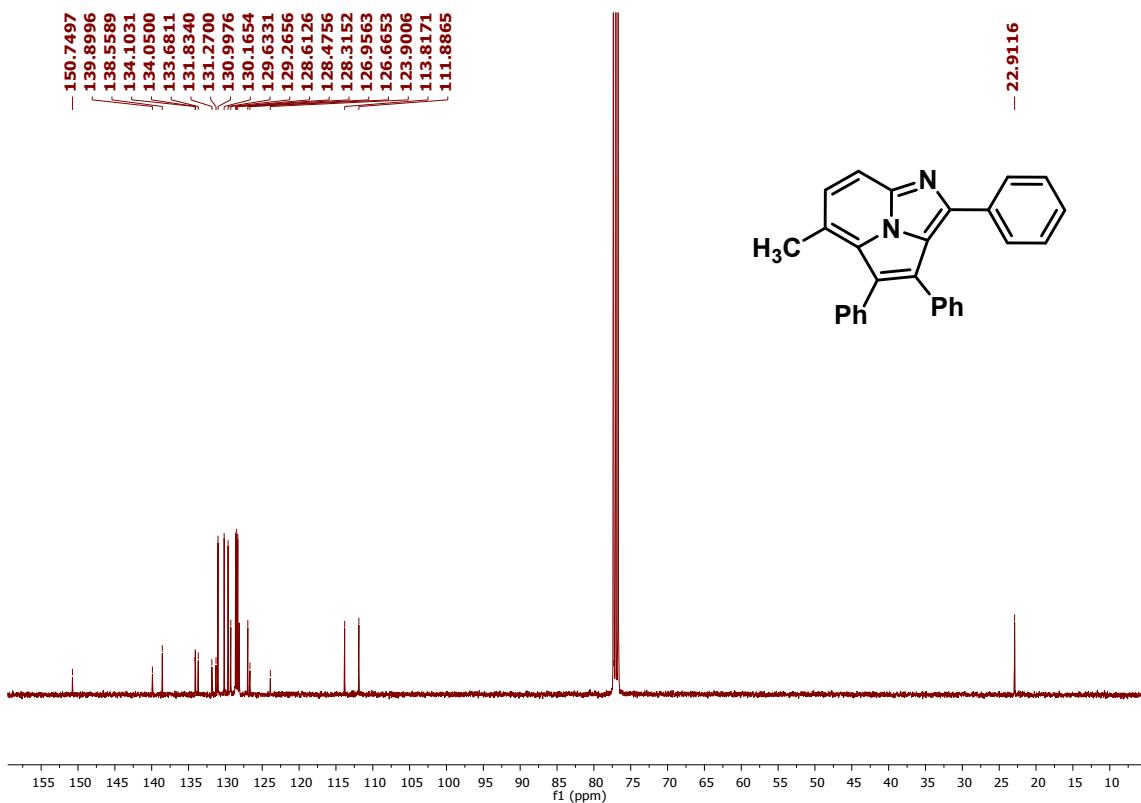


Figure s22. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6e**.

8.13
8.11
8.11
8.09
8.04
8.03
7.91
7.89
7.86
7.56
7.54
7.48
7.46
7.41
7.39
7.38
7.36
7.34
7.32
7.26

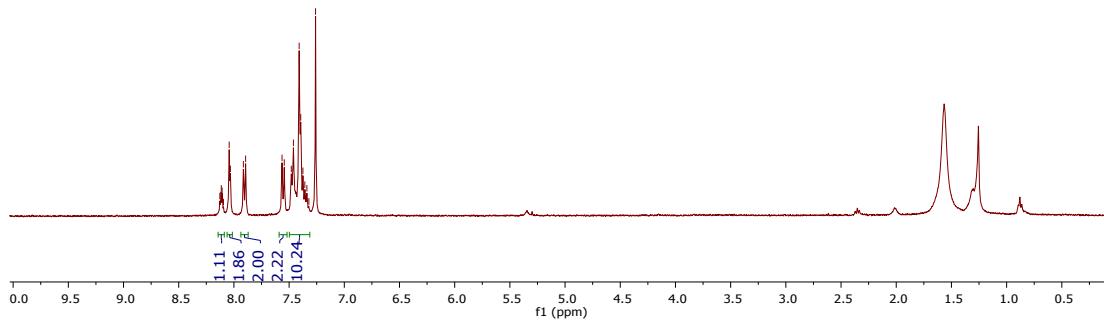
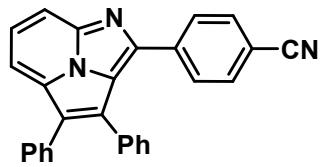
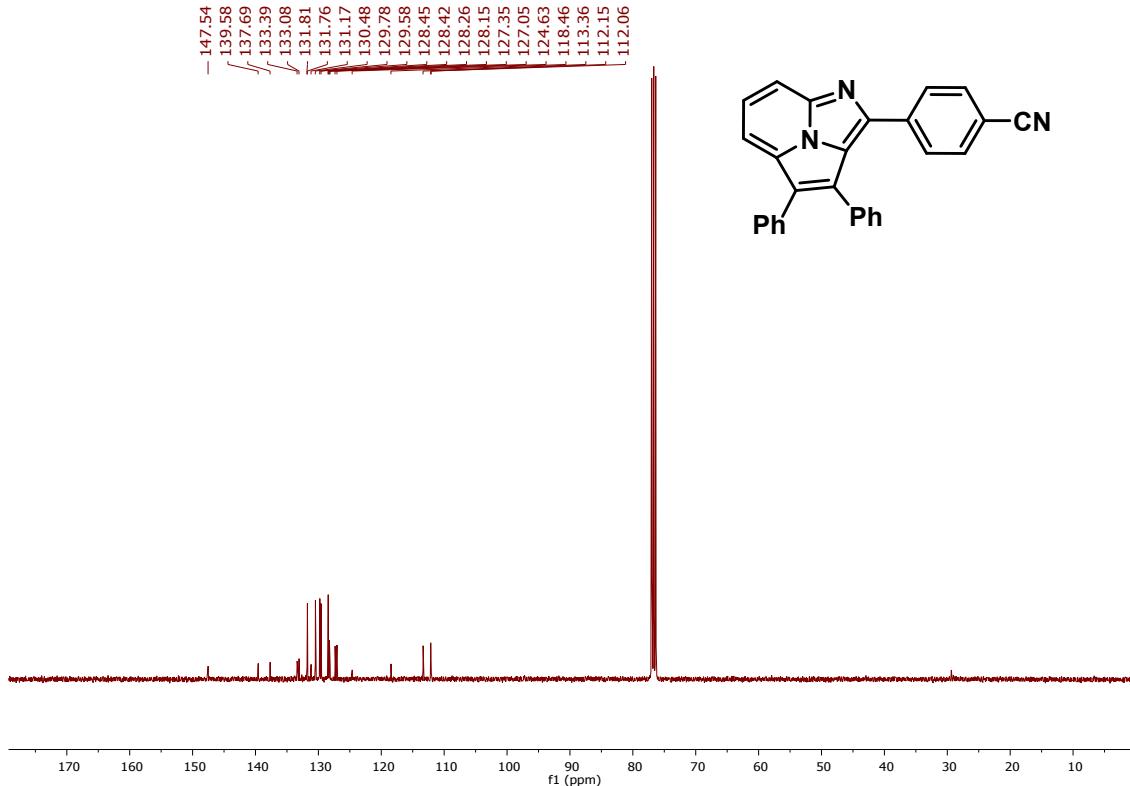
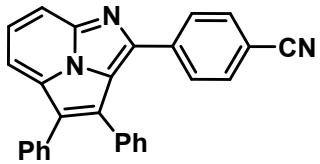


Figure s23. ^1H NMR spectrum of 6f.

147.54
139.58
137.69
133.39
133.08
131.81
131.76
131.17
130.48
129.78
129.58
128.45
128.42
128.26
128.15
127.35
127.05
124.63
118.46
113.36
112.15
112.06



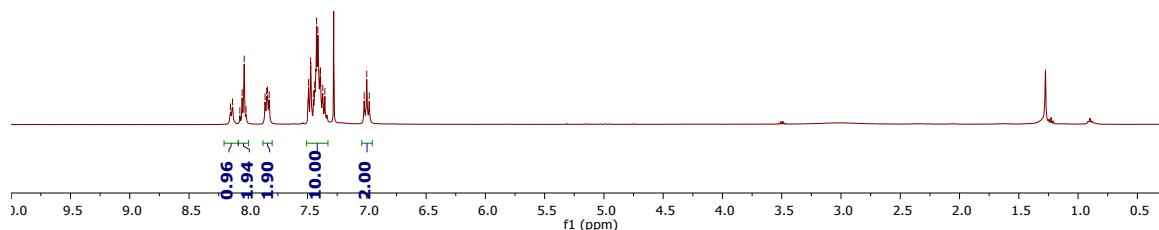
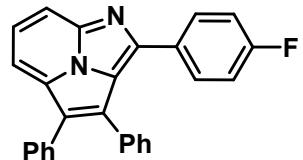


Figure s25. ^1H NMR spectrum of **6g**.

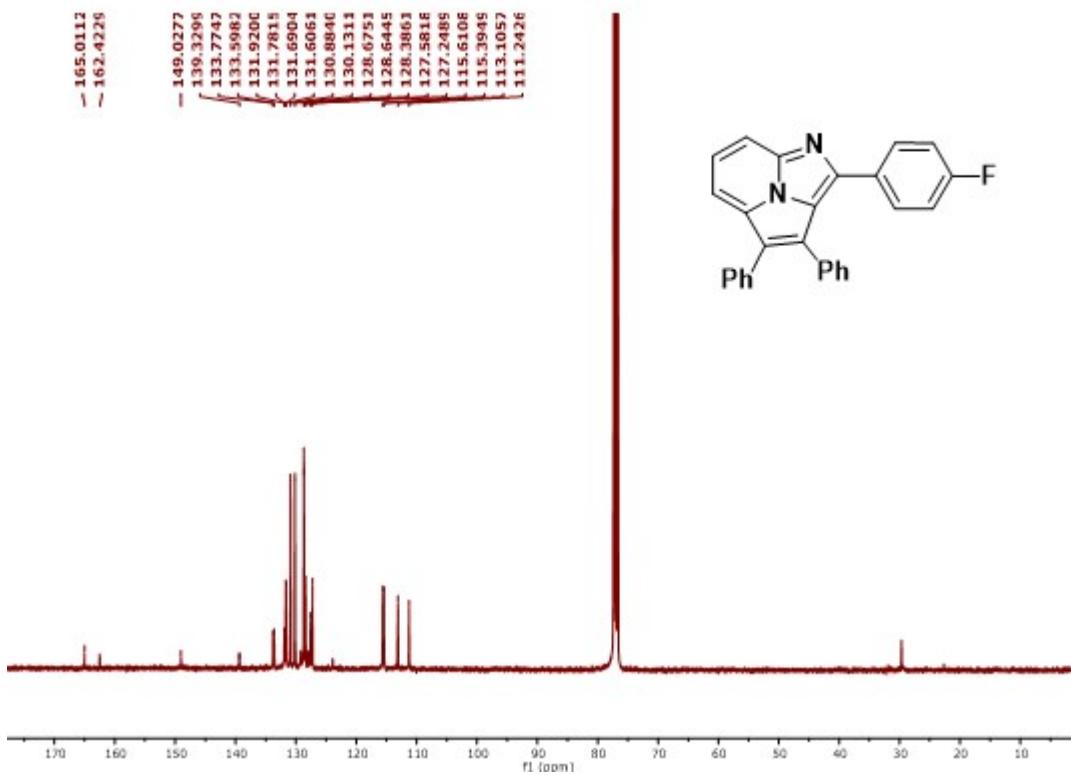
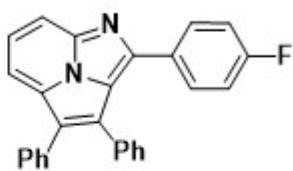


Figure s26. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6g**.

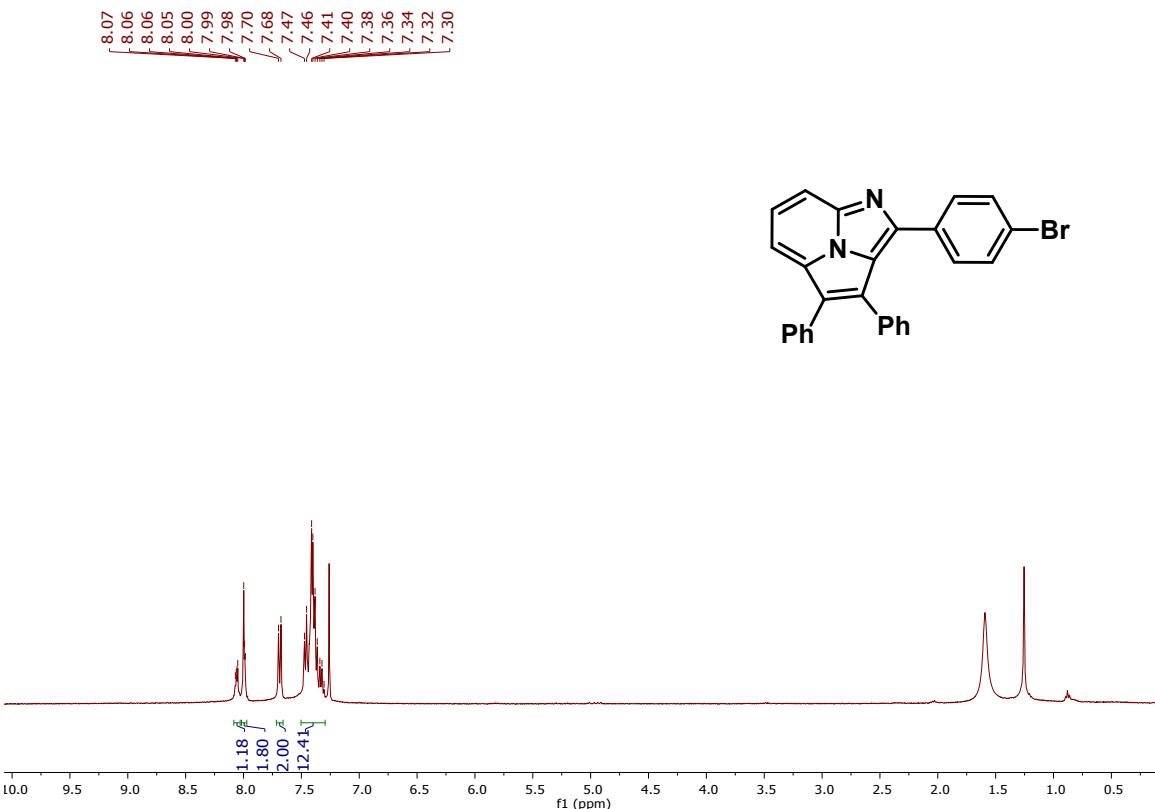


Figure s27. ¹H NMR spectrum of **6h**.

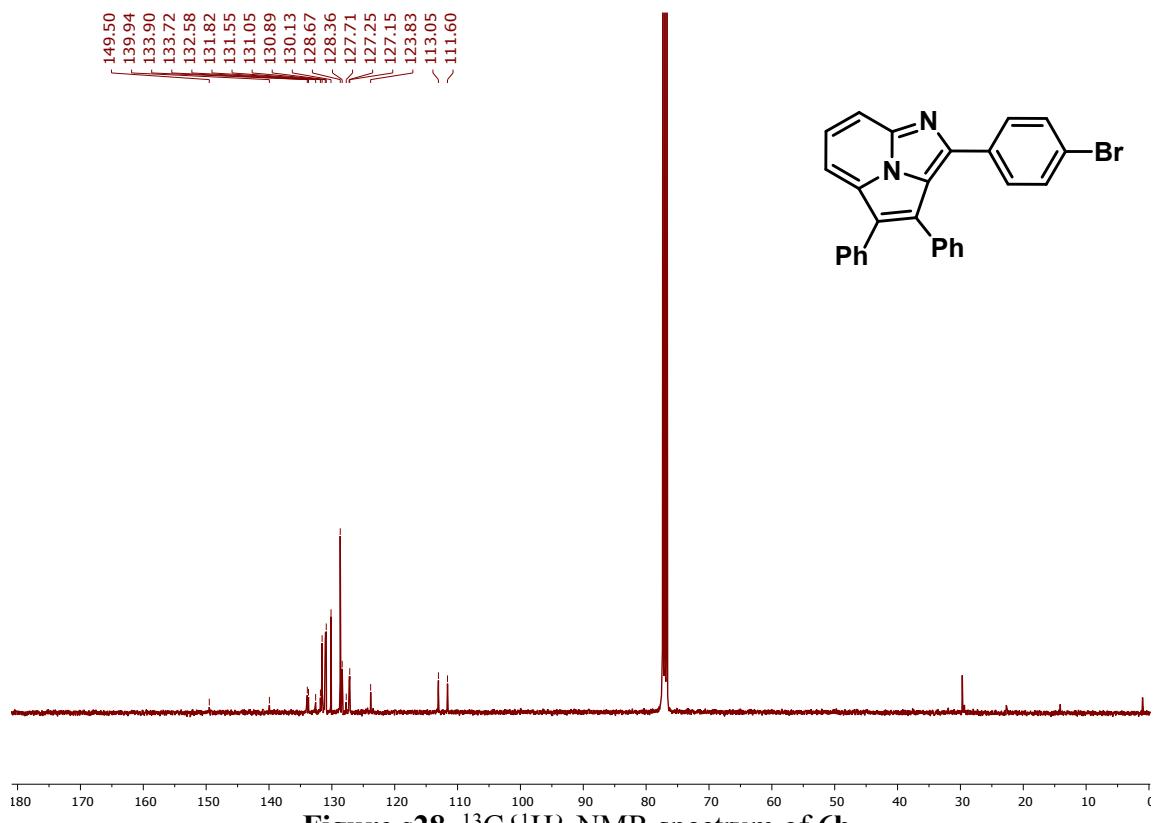


Figure s28. ¹³C{¹H} NMR spectrum of **6h**.

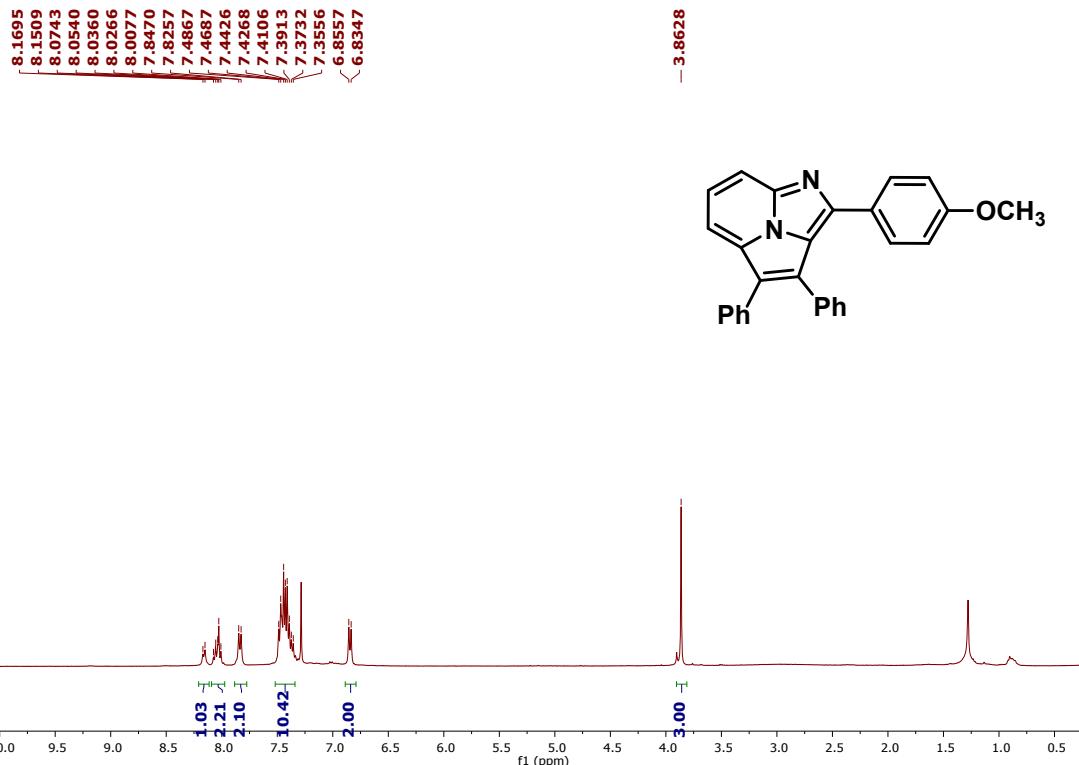


Figure s29. ^1H NMR spectrum of 6i.

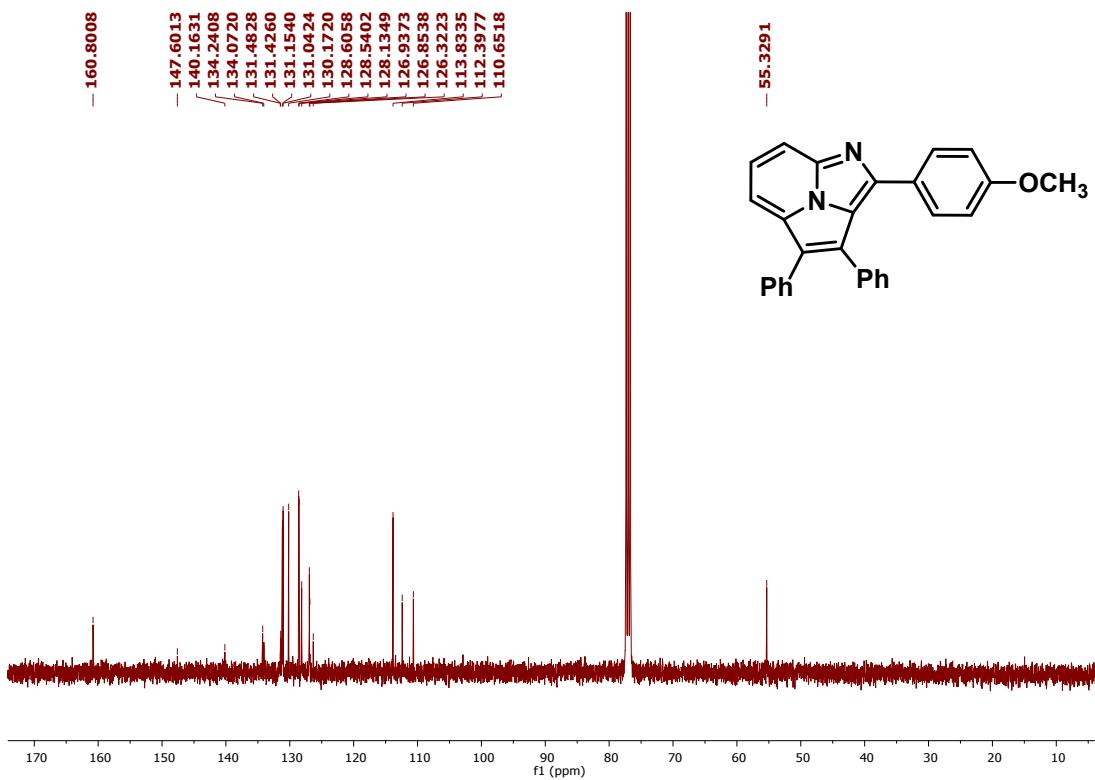
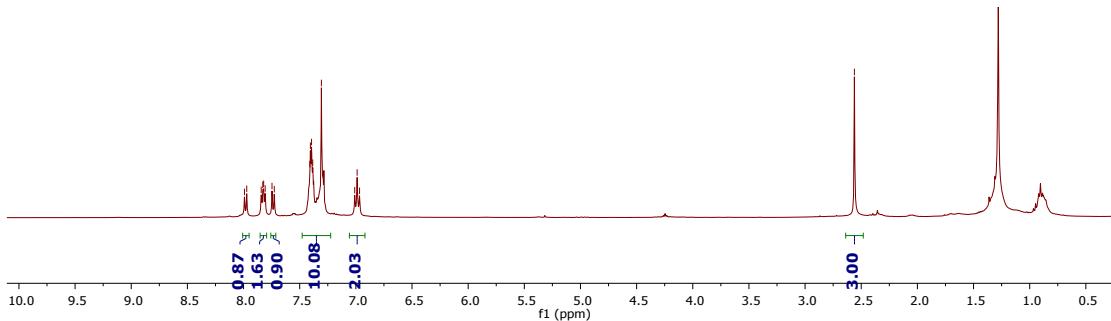
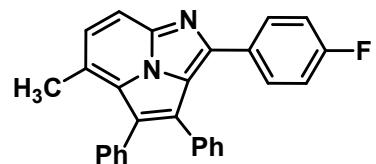


Figure s30. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of 6i.

7.9191
7.9716
7.8426
7.8288
7.8211
7.8073
7.7466
7.7263
7.4100
7.4049
7.3946
7.3839
7.3069
7.0103
6.9887
6.9670

- 2.5608



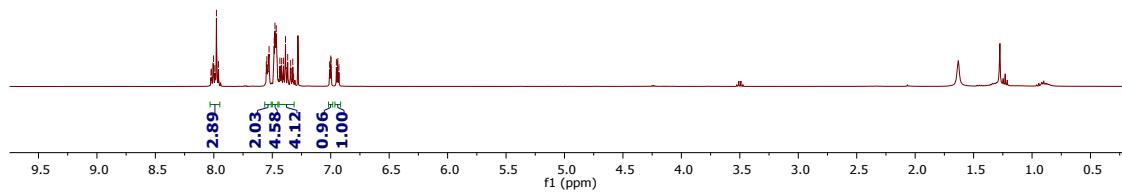
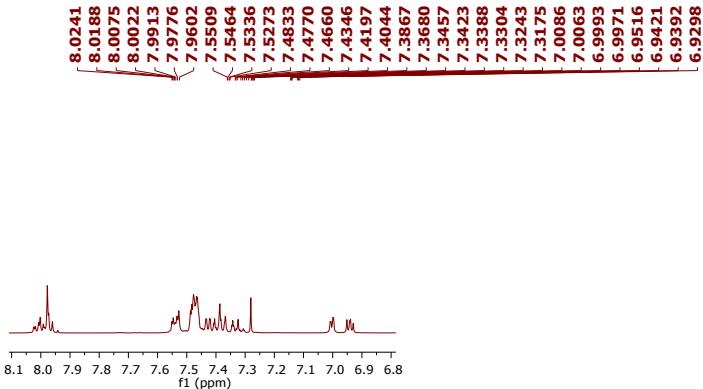


Figure s33. ^1H NMR spectrum of **6k**.

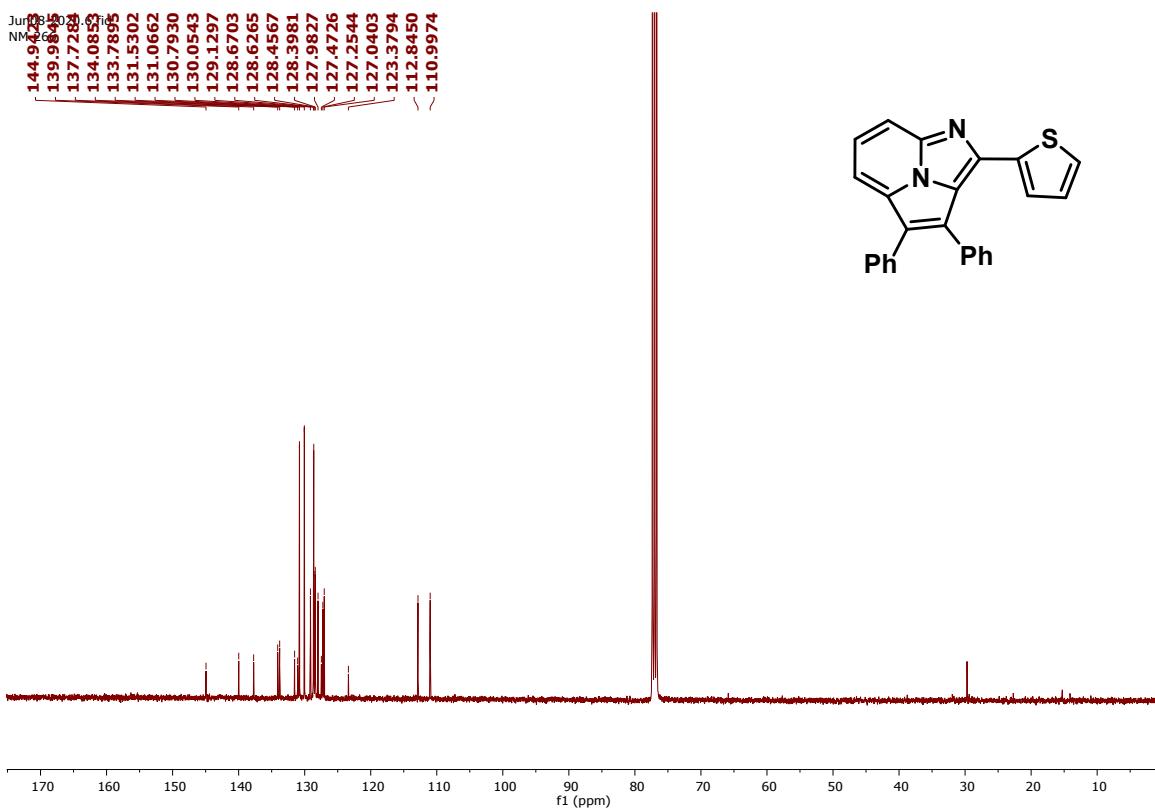


Figure s34. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6k**.



- 2.93

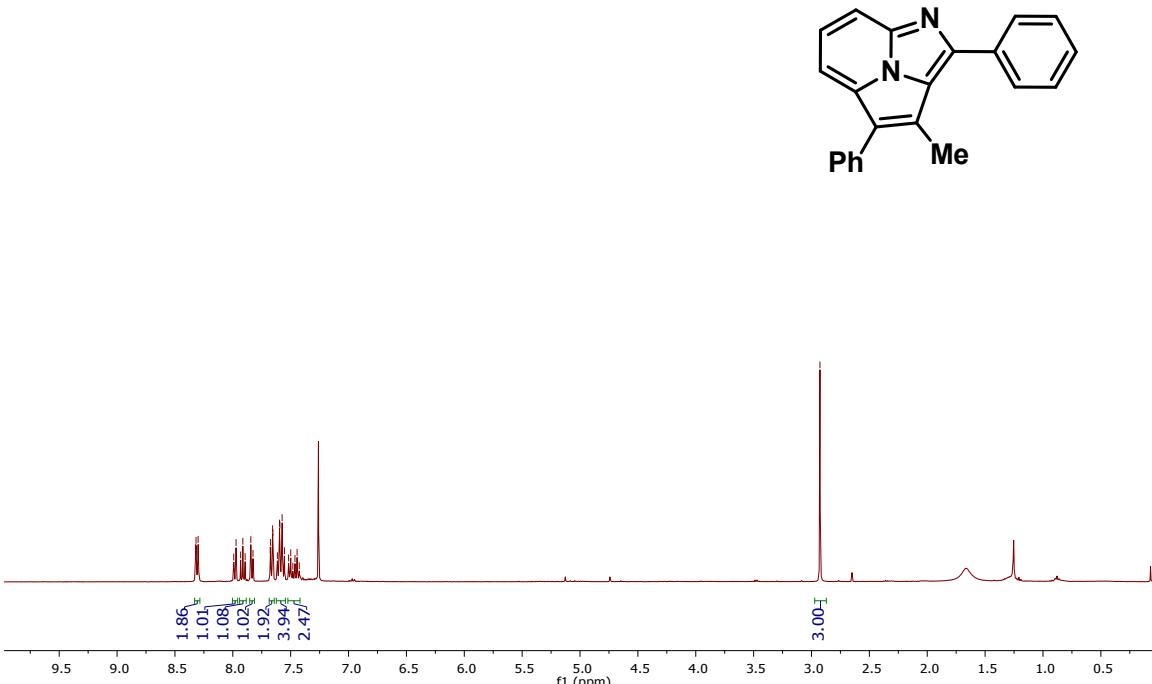


Figure s35. ^1H NMR spectrum of **6l**.



- 13.98

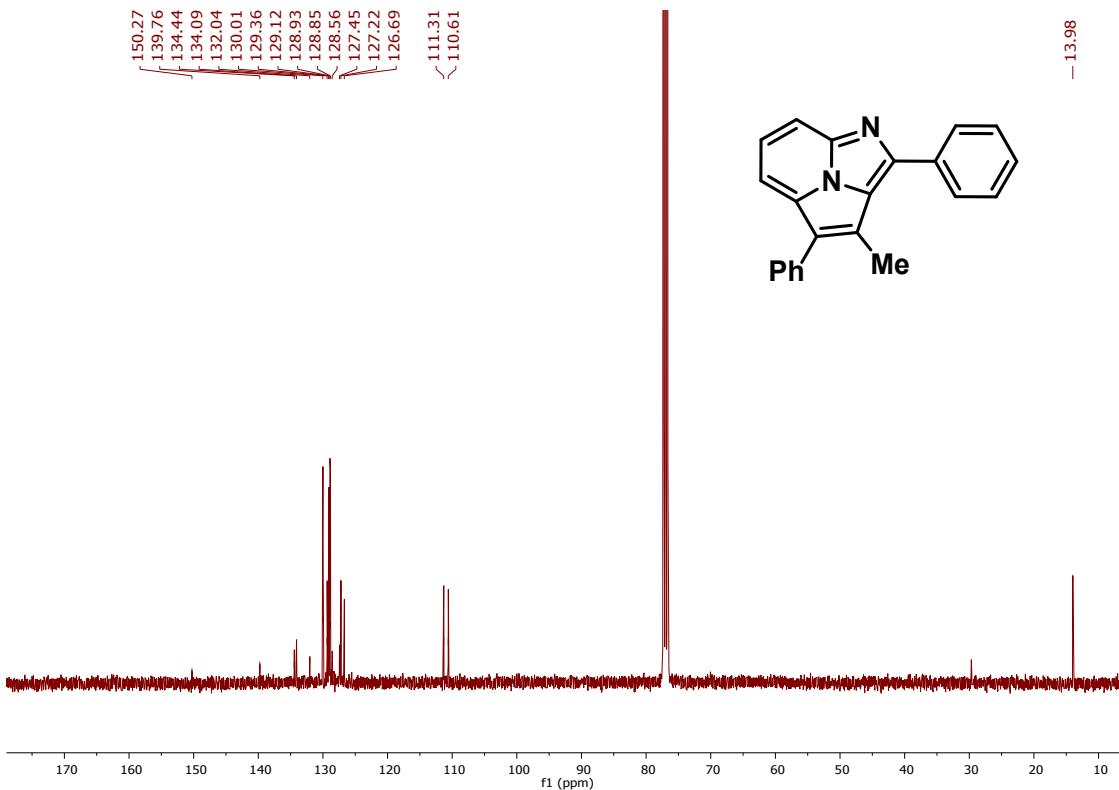
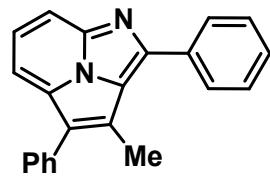


Figure s36. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6l**.

8.0954
8.0761
8.0233
8.0041
7.9847
7.9599
7.9409
7.8438
7.8253
7.5631
7.5422
7.5365
7.5153
7.4155
7.3982
7.3748
7.3561
7.3350
7.3138
7.3037
7.2835

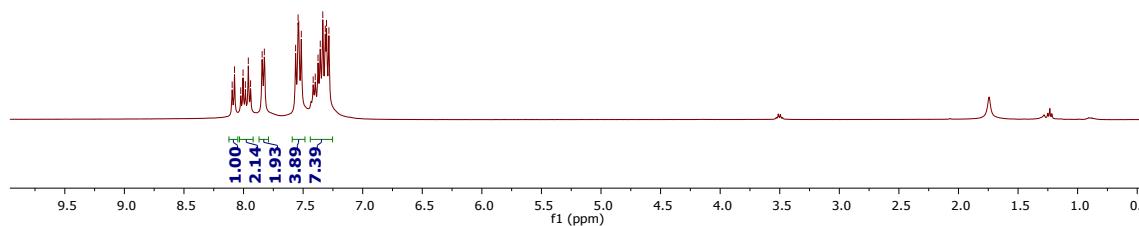
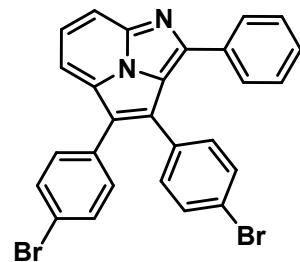


Figure s37. ^1H NMR spectrum of **6m**.

-151.4741
-140.1225
-133.4017
-132.6098
-132.5733
-132.0521
-131.9052
-131.6601
-131.2568
-130.1296
-129.6998
-129.6703
-128.5157
-127.2807
-125.9933
-123.8431
-122.7641
-121.5731
-112.7365
-111.7554

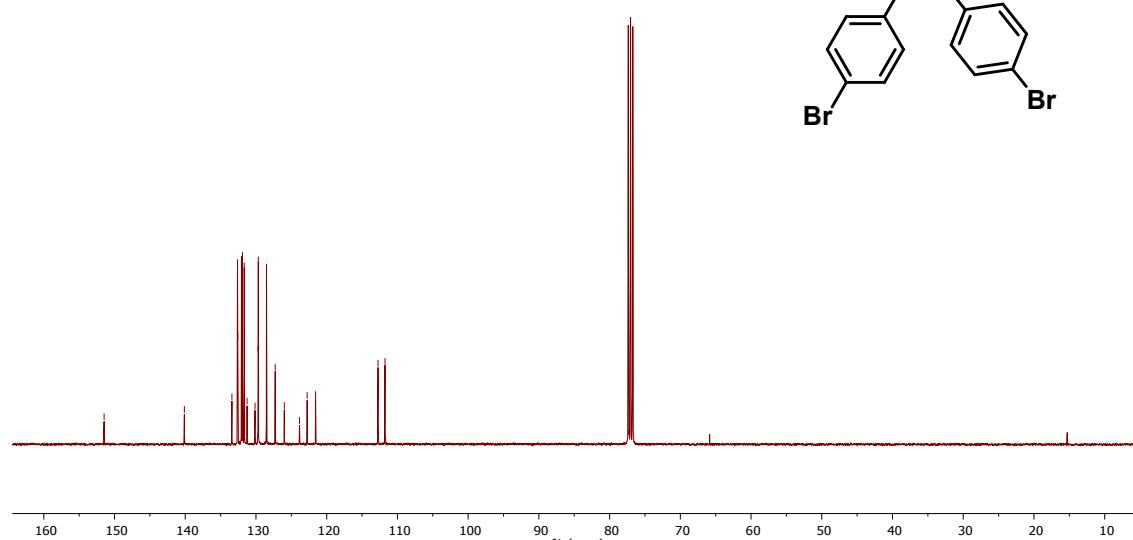
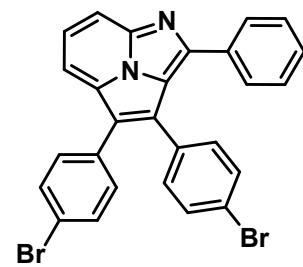


Figure s38. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6m**.

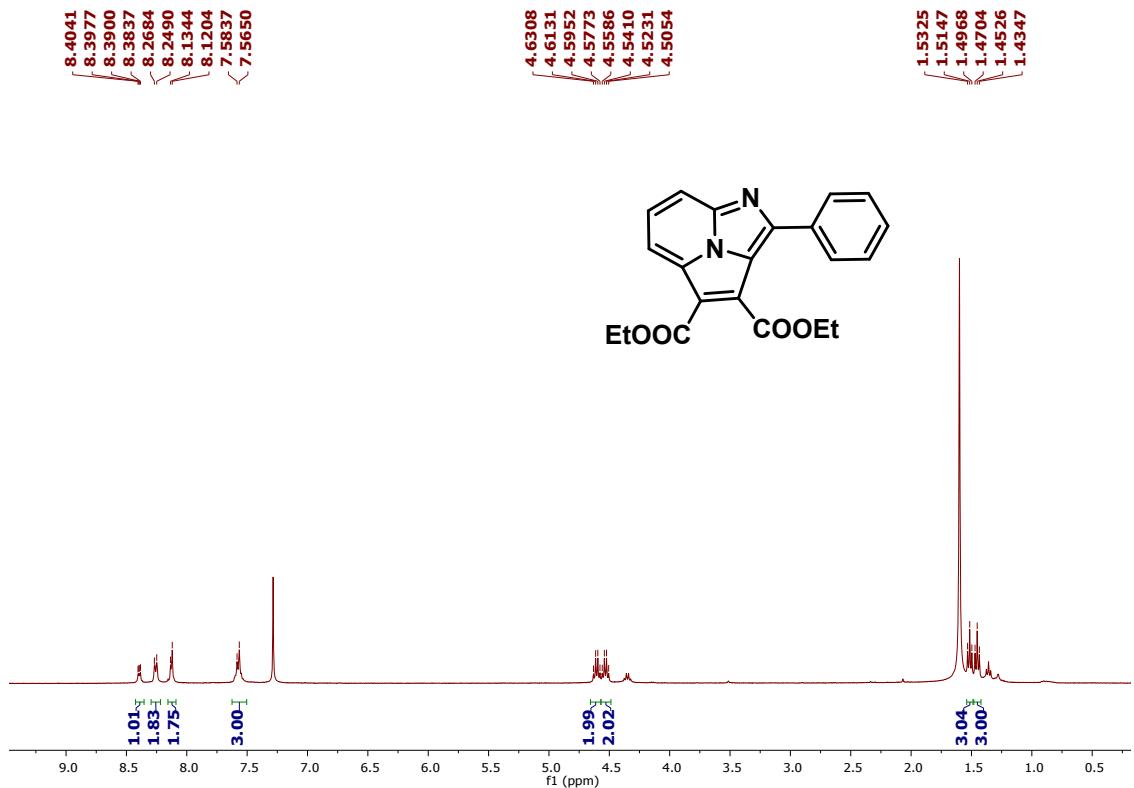


Figure s39. ¹H NMR spectrum of **6n**.

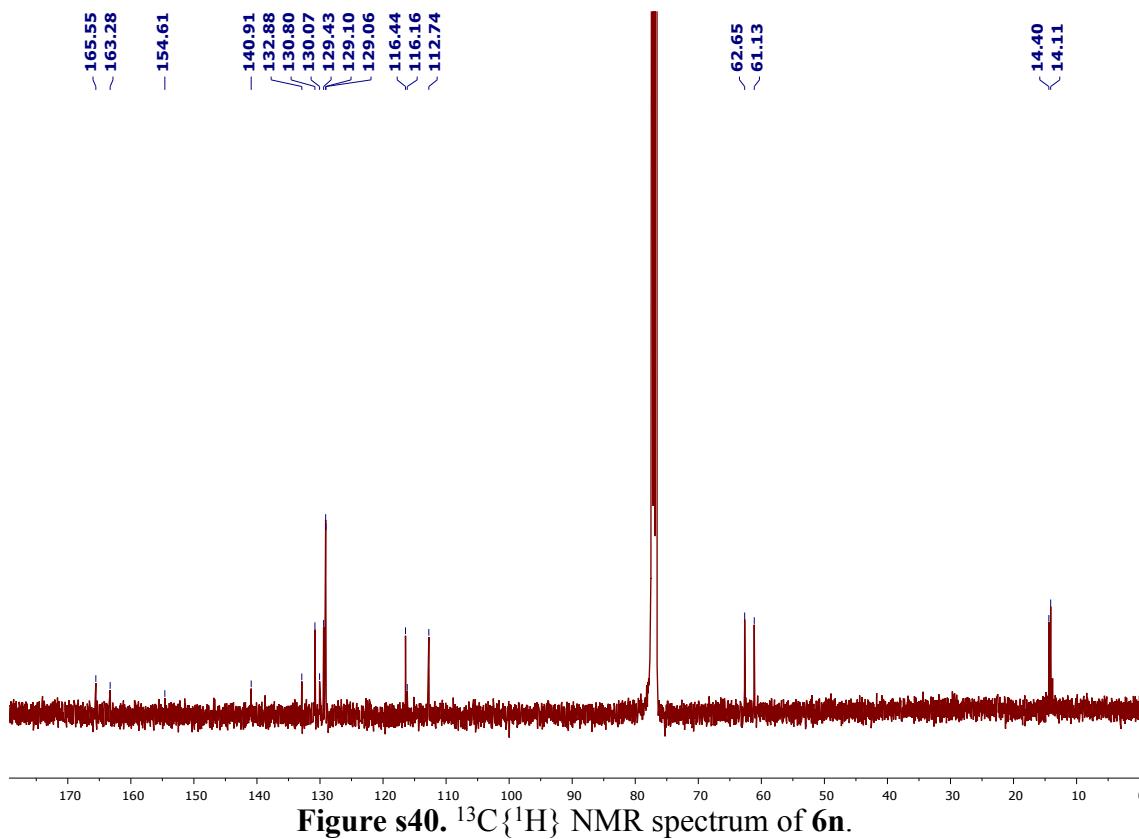


Figure s40. ¹³C{¹H} NMR spectrum of **6n**.

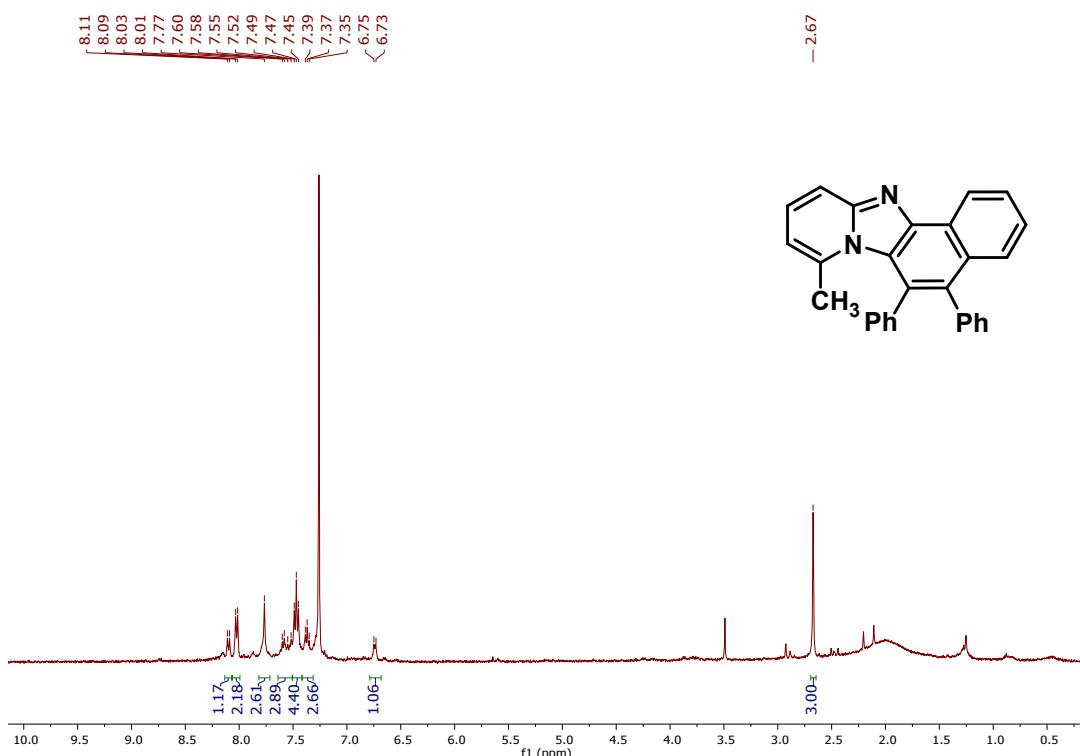


Figure s41. ^1H NMR spectrum of **70**.

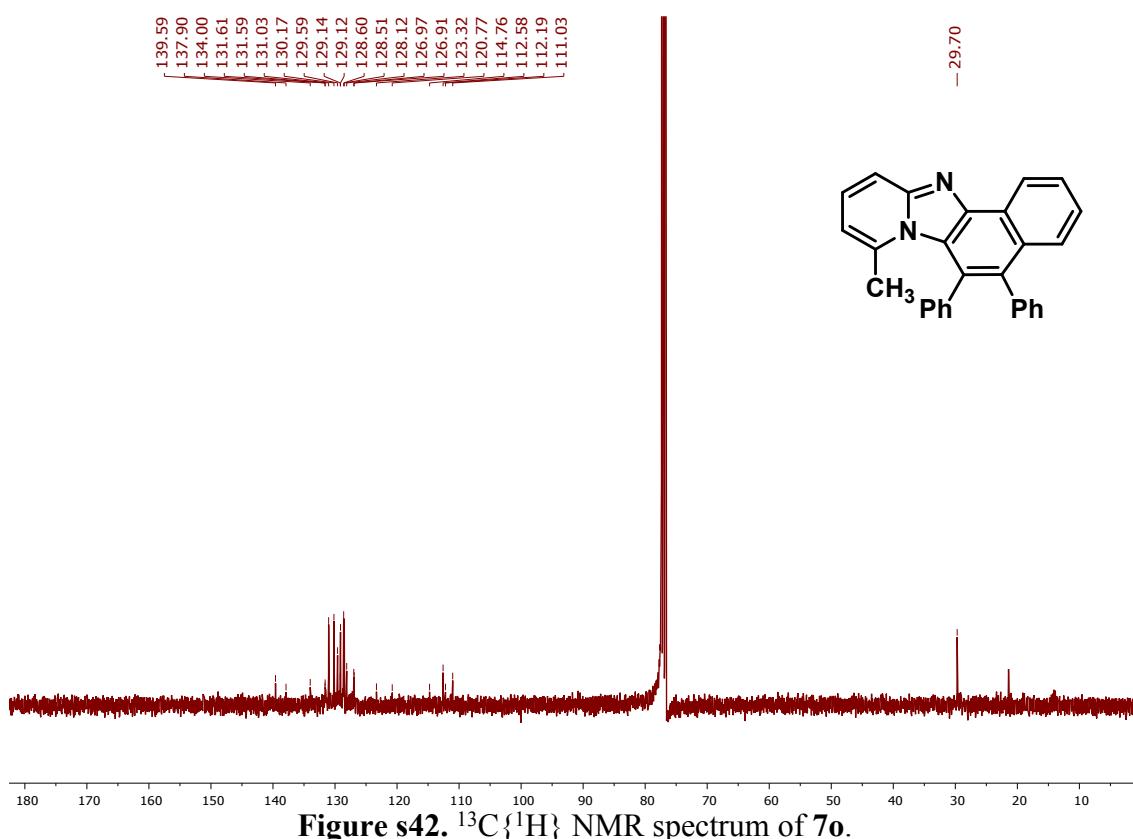


Figure s42. $^{13}\text{C}\{\text{H}\}$ NMR spectrum of **7o**.

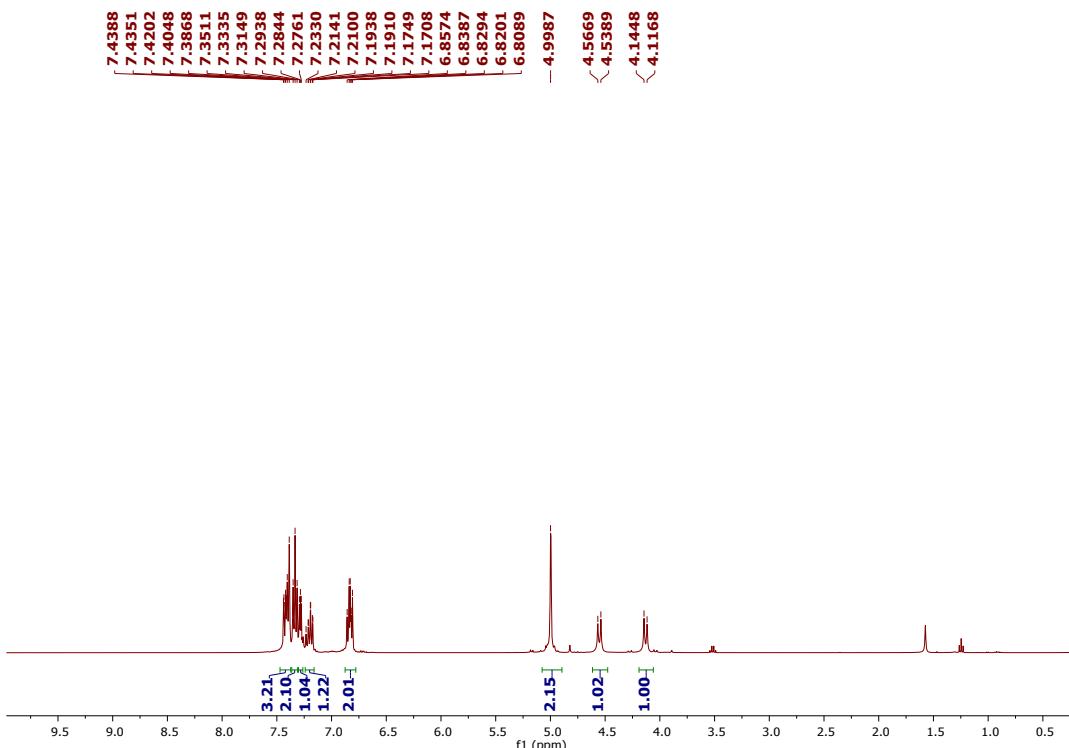


Figure s43. ^1H NMR spectrum of complex **3** after storing for three months at room temperature under open air conditions.

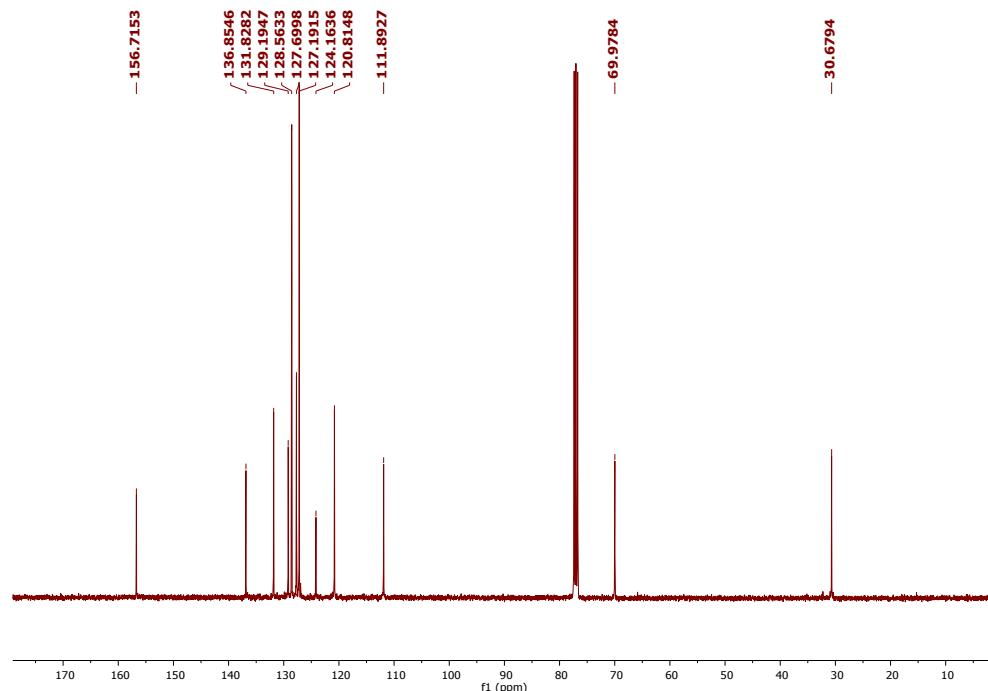


Figure s44. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of complex **3** after storing for three months at room temperature under open air conditions.

(Note: Peaks at 0.86 and 1.26 ppm in HNMRs and 29.76 in CNMRs are due to grease whereas peak at 1.56 in HNMRs is due to moisture of CDCl_3)

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