Acceptor-induced cooperative supramolecular co-assembly with

emissive charge-transfer for supramolecular encryption

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1. Materials and methods

9,10-diiodoanthracene,^{S1} compounds 1,^{S2} and 4 ^{S3} (see Scheme S1 and S2) were synthesized according to the previously reported literatures. Other reagents and solvents used in the experiments were purchased from the commercial sources without further purification.

¹H NMR and ¹³C NMR spectra were obtained from Bruker Avance 400 instruments. Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry mass (MALDI-TOF-MS) measurements were recorded on a Bruker Autoflex Speed spectrometer with DCTB as the matrix. UV–Vis spectra were performed on a Shimadzu UV-2550 spectrometer. Fluorescence spectra were recorded on a Hitachi F-4600 FL spectrophotometer. The absolute fluorescence quantum yield (Φ_F) and time-resolved fluorescence lifetime experiments were recorded on an Edinburgh FLS980 transient steady-state fluorescence spectrometer. Φ_F was measured using a calibrated integrating sphere. Fourier transform infrared (FTIR) spectra were performed on a Nicolet iS50 infrared spectrometer. Transmission electron microscope (TEM) images were recorded on a JEM-2100 electron microscope. Dynamic light scattering (DLS) experiments were conducted on a Brookhaven BI-9000AT instrument. Solid-state ¹³C NMR spectra were recorded on a JEOL JNM-ECZ400R/S1 instrument.

Theoretical calculations: all of the optimized structures were performed on G09 software packages.^{S4} All of the elements were described by the B3LYP/6-31G(d) computational method. There are no imagery frequencies for the optimized geometries.

Mathematical fitting of the supramolecular co-assembly processes: the co-assembly of **G1/TCNB** is described using the nucleation-elongation cooperative model, which was developed by Meijer, *et al.*^{S5} This model is used to describe the co-assembly of **G1/TCNB** which exhibits a non-sigmoidal melting curve as shown in the temperaturedependent UV–Vis experiments. In terms of **G2/TCNB**, a sigmoidal curve is obtained by plotting the fraction of aggregated species (α_{agg}) against temperature, revealing the involvement of isodesmic co-assembly mechanism.^{S6} To acquire detailed thermodynamic parameters for the co-assembly processes of **G1/TCNB** and **G2/TCNB**, the normalized UV–Vis melting curves obtained by plotting α_{agg} ($\lambda = 568$ nm for **G1/TCNB**, and 489 nm for **G2/TCNB**) against temperature are fitted with the respective mathematical model. In addition, the data of solvent-dependent UV–Vis absorption spectra for **G1/TCNB** can be fitted with the solvent-dependent equilibrium model reported by the previous literature.^{S7}

Inkjet printing experiments: the printing experiments were performed on a commercially available inkjet printer (HP Deskjet 2131) with the customized HP803 black inkjet cartridge. The original black inks from the cartridge were removed, and the cartridge was washed with ethanol, water and dried with N_2 blowing. The inks of **G1** (0.5 mL, 1.0 mM) and **TCNB** (0.5 mL, 1.0 mM) were then loaded separately into the clean black cartridge to perform the printing experiments.





Figure S1. (a) UV–Vis and (b) fluorescence spectra of G1 (2.0×10^{-5} M) in CHCl₃ and MCH. $\lambda_{ex} = 453$ nm. (c) Temperature-dependent UV–Vis spectra of G1 (2.0×10^{-5} M) in MCH. (d) Concentration-dependent UV–Vis spectra of G1 in MCH (from 1.0×10^{-3} M to 5.0×10^{-5} M, 298 K). When switching the solvent from polar CHCl₃ to nonpolar MCH, the maximum absorption band of G1 is blue-shifted, whilst the band remains nearly unaltered in the temperature- and concentration- dependent UV–Vis spectra experiments. Such phenomena suggest a weak aggregation for G1 in the current situations.



Figure S2. (a) UV–Vis and fluorescence spectra of **G0** (2.0×10^{-5} M) in CHCl₃ and MCH. λ_{ex} = 446 nm. (b) Temperature-dependent UV–Vis spectra of **G0** (2.0×10^{-5} M) in MCH.



Figure S3. (a) UV–Vis and fluorescence spectra of G2 (2.0×10^{-5} M) in CHCl₃ and MCH. λ_{ex} = 442 nm. (b) Temperature-dependent UV–Vis spectra of G2 (2.0×10^{-5} M) in MCH.



Figure S4. (a) UV–Vis absorbance changes upon gradual addition of TCNB into G1 (2.0×10^{-5} M) in MCH. (b) The intensity changes at λ =500 nm. (c)–(d) Job's plot between G1 and TCNB ([G1] + [TCNB] = 2.0×10^{-5} M), by plotting the intensity changes of UV–Vis absorbance band at 500 nm against the mole fraction of G1. Upon progressive addition of TCNB into the MCH solution of G1, the absorption band centred at 453 nm gradually decreases, whilst a broad and relatively lower-energy band locating between 500 and 625 nm emerges. The red-shifted band reaches to the maximum when adding equivalent TCNB, which suggests a 1 : 1 binding stoichiometry between G1 and TCNB. Moreover, the Job's plot experiments also demonstrate the 1 : 1 stoichiometry of the complex G1/TCNB.



Figure S5. UV–Vis and fluorescence spectra of (a) **G2** $(2.0 \times 10^{-5} \text{ M})$ and **G2/TCNB** $(2.0 \times 10^{-5} \text{ M})$ for each component) in MCH, and (b) **G0** $(2.0 \times 10^{-5} \text{ M})$ and **G0/TCNB** $(2.0 \times 10^{-5} \text{ M})$ for each component) in MCH. For the second-generation dendrimer **G2**, a new shoulder band between 477 and 525 nm emerges after mixing equivalent **G2** and **TCNB** together, which arises from the charge transfer (CT) interaction from donor **G2** to acceptor **TCNB**. In stark contrast, the CT process cannot occur between the zeroth-generation dendrimer **G0** and **TCNB**, as evidenced by the almost no changes of UV–Vis and fluorescence signals in MCH.



Figure S6. a) Molecular structures of **G1** and **TCNB**. b) Concentration-dependent ¹H NMR spectra (400 MHz, 298 K) of **G1/TCNB** in CDCl₃: (i) 1.0 mM, (ii) 2.0 mM, (iii) 4.0 mM, (iv) 6.0 mM, (v) 8.0 mM, and (vi) 10.0 mM. c) Photographs of i) **G1**, ii) **TCNB**, iii) **G1/TCNB** in concentrated CDCl₃ solution (10.0 mM for each component).



Figure S7. Solid-state ¹³C NMR of **G1**, **G1/TCNB**, and **TCNB**. Solid-state ¹³C NMR measurements are performed to determine the change of C atom chemical circumstances. The chemical shift of **G1** at around 110–120 ppm shifts downfield, and that of **TCNB** at 122 ppm shifts to 115 ppm, which indicate the reduction of π -electron density on **G1** and the increase of electron density of benzene ring on **TCNB**.



Figure S8. Molecular orbital diagrams of (a) **G1**, (b) **G1/TCNB**, and (c) **TCNB** *via* TD-DFT computation. The highest occupied molecular orbital (HOMO) of **G1/TCNB** is nearly completely localized on the electron-donor **G1**, whilst the lowest unoccupied molecular orbital (LUMO) is distributed on the acceptor **TCNB**, which indicates the electron transition from HOMO of donor to LUMO of acceptor.



Figure S9. (a) Temperature-dependent UV–Vis spectra of G1/TCNB (2.0×10^{-5} M) in MCH. Arrows indicate the spectral variations upon increasing temperature. (b) α_{agg} of G1/TCNB (on the basis of CT band at 568 nm) *versus* temperature in MCH. The obtained data feature a non-sigmoidal curve, denoting the adoption of nucleation–elongation cooperative mechanism. The solid lines denote the mathematical fitting of the curve according to the Meijer–Schenning–Van-der-Schoot model.



Figure S10. (a) Solvent-dependent UV–Vis spectra of **G1/TCNB** (2.0×10^{-5} M). Arrows indicate the spectral changes upon increasing CHCl₃ volume fraction (*f*) in MCH. (b) α_{agg} as a function of *f* monitored at 500 nm. The red line denotes the mathematical fitting of the curve according to the solvent-dependent equilibrium model. The cooperativity parameter (σ) is determined to be 1.9×10^{-3} , demonstrating the involvement of a cooperative mechanism for the co-assembly process of **G1/TCNB**.



Figure S11. (a) Temperature-dependent UV–Vis spectra of **G2/TCNB** (2.0×10^{-5} M) in MCH. Arrows indicate the spectral variations upon increasing temperature. (b) α_{agg} of **G2/TCNB** (on the basis of CT band at 489 nm) *versus* temperature. The red line represents the non-linear fitting of the data according to the isodesmic mathematical model. c) Number-averaged degree of polymerization (DP_N) of **G2/TCNB** at different temperatures. d) Van't Hoff plot for the temperature dependence of the aggregation constant (K) of **G2/TCNB** in MCH. The sigmoidal melting curve obtained from the temperature-dependent UV–Vis spectra of **G2/TCNB** can be nicely fitted by the isodesmic model, which indicates the adoption of the isodesmic mechanism of the co-assembly process. According to the Van't Hoff plot, ΔG^0 of **G2/TCNB** was calculated to be –22.8 kJ mol⁻¹ at 298 K, which is lower than that of **G1/TCNB** (–36.0 kJ mol⁻¹).



Figure S12. Optimized geometries of structures a) G1 and b) G2, on the basis of DFT calculations at the level of B3LYP/6-31G(d).



Figure S13. (a) Photographs of the MCH solution of G1 (2.0×10^{-5} M) and G1/TCNB (2.0×10^{-5} M) irradiating with a laser pen. (b) DLS measurements of G1 and G1/TCNB in MCH. TEM micrographs of (c) G1 and (d)–(e) G1/TCNB on the copper grids. No aggregation can be detected for G1 in MCH, which is manifested by the results of no Tyndall effect, small DLS hydrodynamic diameter, and no morphology in TEM. The results are coincident with that of UV–Vis and fluorescence spectra measurements. When adding equivalent TCNB into the MCH solution of G1, the co-assembly G1/TCNB is prone to assemble into large-sized nanosheets, with the averaged hydrodynamic diameter of 750 nm.



Figure S14. (a) TEM micrograph of G2. (b) DLS measurements of G2 $(2.0 \times 10^{-5} \text{ M})$ and G2/TCNB $(2.0 \times 10^{-5} \text{ M})$ in MCH. (c)–(d) TEM micrographs of G2/TCNB. No microstructures can be detected for G2. In comparison, G2/TCNB is prone to assemble into nanoparticles with 265 nm in diameter.



Figure S15. Fluorescence decay profiles of (a) **G1** film ($\lambda_{ex} = 454$ nm, $\lambda_{em} = 552$ nm) and (b) **G1/TCNB** film ($\lambda_{ex} = 500$ nm, $\lambda_{em} = 619$ nm).



Figure S16. FTIR spectra of G1, TCNB and G1/TCNB. Depending on the FTIR measurements, the bands at 3114 and 3047 cm⁻¹ (C–H str) of TCNB slightly shift to the lower frequency of 3102 and 3041 cm⁻¹ in G1/TCNB, respectively, which are ascribed to the CT interaction from G1 to TCNB.



Figure S17. (a) Fluorescence spectra and (b) wavelength changes for the film of G1/TCNB as a function of CH_2Cl_2 vapor/dry cycles. Upon successively fuming with CH_2Cl_2 vapor and drying at room temperature, the emission signals of G1/TCNB can be reversibly switched for multiple cycles.



4. Supramolecular encryption applications of G1/TCNB

Figure S18. (a) Schematic illustration for **G1/TCNB** toward supramolecular encryption applications. (b) Digital photographs of the luminescent image at different stages. Scale bars: 5 mm. A cartoon pattern of "Bell Tower" is firstly printed on a non-fluorescent paper, which is in the encrypted state because it is invisible under both natural and UV light (Fig. S16b, i and ii). After the paper is printed with **G1** inks, the pre-printed pattern is still invisible under natural light, but appears with red emission color under UV light (Fig. S16b, iii and iv). Notably, the decrypted information could be authenticated by CH_2Cl_2 vapor, as evidenced by the emission color variation as well as the vanishment of the pattern (Fig. S16b, v). Subsequently, the red emission pattern could fully restore after the paper is drying. Besides, some other pattern can also be designed derived from **G1/TCNB** for supramolecular encryption applications (Fig. S16b, vi–x).



Figure S19. a) Schematic illustration of the sample preparation for quantitatively determining the molar stoichiometry of CH_2Cl_2 in the vapo-fluorochromic experiments. b) Fluorescence spectra change of the prepared G1/TCNB samples (28 mg of G1/TCNB CH_2Cl_2 solution)

versus the volatilization time of CH₂Cl₂ at room temperature ($\lambda_{ex} = 500 \text{ nm}$, $\lambda_{em} = 620 \text{ nm}$). The black, red and blue dots represent three repeated experiments. c) Fluorescence spectra and d) weight changes of the three sets of prepared G1/TCNB samples versus the volatilization time of CH₂Cl₂ at room temperature. An elaborate experiment is designed to quantitatively determine the equivalent ratio of CH_2Cl_2 to **G1/TCNB** in the encryption/decryption process. Specifically, a small amount of G1/TCNB CH₂Cl₂ solution (2.6 mg/mL) is firstly dropping onto a filter paper. Upon slowly volatilizing the CH₂Cl₂ vapor in air at room temperature, the emission color turns from green to red. Time-dependent fluorescence spectra are then employed to probe the emissive CT band variations of G1/TCNB versus the volatilization time of CH₂Cl₂. In Figure S19b, the emission signal at 620 nm gradually increases, then sharply reach to a plateau at around 170 s (the experiment is repeated three times), indicating the residual CH₂Cl₂ solvent at this time is the necessary minimum amount for the vapo-fluorochromic experiments. Simultaneously, the weight change of the prepared G1/TCNB sample versus the volatilization time of CH₂Cl₂ is proceeded under the same conditions (Figure S19d). At 170 s, the weight of the residual CH₂Cl₂ solvent is measured to be around 6.9 mg (Figure S19d, red dots), which is 126 times the weight of G1/TCNB. Additionally, two sets of similar experiments are performed, by separately dropping 15 mg and 36 mg of G1/TCNB CH₂Cl₂ solution (2.6 mg/mL) onto the filter papers (Figure S19c and S19d, black and blue dots). The equivalent ratio of CH₂Cl₂ to G1/TCNB is determined to be 115 and 133 times, respectively.



Figure S20. Photographs of the inkjet-printed fluorescent pattern on day 1 and after 6 months. The printed pattern derived from **G1/TCNB** features enough stability, as evidenced by unchanging of the emission color under the ambient conditions for at least 6 months.



Figure S21. Digital photographs of the cartoon pattern of "Bell Tower" firstly printed with TCNB as inks and then printed with G0 inks. Scale bars: 5 mm. As a control experiment, when G0 is utilized in place of G1 as fluorescent inks to decrypt the printed information, no distinguishable pattern emerges on the paper under both natural and UV light.

5. Synthetic routes to dendrimers Gn (n=0-2)



Scheme S1. Synthetic routes to the dendrimers **G0** and **G1.** i) Pd(PPh₃)₂Cl₂, CuI, TEA; ii) K₂CO₃, DMF; iii) trimethylsilylacetylene, Pd(PPh₃)₂Cl₂, CuI, TEA; iv) tetrabutylammonium fluoride, THF; v) Pd(PPh₃)₂Cl₂, CuI, TEA.



Scheme S2. Synthetic routes to the dendrimer G2. i) K₂CO₃, DMF; ii) LiAlH₄, THF; iii) SOCl₂, DCM; iv) 5-bromoresorcinol, CsCO₃, MeCN; v) 2-methyl-3-butyn-2-ol, Pd(PPh₃)₂Cl₂, CuI, TEA; vi) KOH, toluene; vii) Pd(PPh₃)₂Cl₂, CuI, TEA.

5.1 Synthesis of dendrimer G0

9,10-diiodoanthracene (120 mg, 0.28 mmol), compound 1 (200 mg, 0.70 mmol), $Pd(PPh_3)Cl_2$ (19.6 mg, 0.08 mmol) and CuI (5.33 mg, 0.03 mmol) were stirred in TEA (20 mL) and stirred under nitrogen atmosphere. After stirring at 75 °C for 12 hours, the reaction mixture was evaporated to remove the solvent, and the residue was extracted with H_2O/CH_2Cl_2 for three times. The combined organic extracts were dried over

anhydrous Na₂SO₄, and the solvent was removed with a rotary evaporator. The residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 5 : 1 *v/v* as the eluent) to afford **G0** as an orange solid (172 mg, 82 %). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.69 (dd, *J* = 6.6, 3.3 Hz, 4H), 7.70 (d, *J* = 8.8 Hz, 4H), 7.63 (dd, *J* = 6.7, 3.2 Hz, 4H), 6.97 (d, *J* = 8.8 Hz, 4H), 4.03 (t, *J* = 6.6 Hz, 4H), 1.82 (dd, *J* = 14.7, 6.8 Hz, 4H), 1.51–1.45 (m, 4H), 1.28 (s, 32H), 0.89 (t, *J* = 6.8 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 159.80, 133.18, 132.15, 127.39, 126.58, 118.63, 115.60, 114.97, 102.68, 85.45, 68.38, 31.94, 29.67, 29.65, 29.61, 29.59, 29.41, 29.34, 29.30, 26.09, 22.68, 14.02. MALDI-TOF-MS *m/z*: [M + H]⁺, C₅₄H₆₇O₂, calculated 747.5141; found 747.5199.



Figure S22. ¹H NMR spectrum (400 MHz, CDCl₃, 298K) of G0.



Figure S23. ¹³C NMR spectrum (101 MHz, CDCl₃, 323 K) of G0.



Figure S24. MALDI-TOF-MS spectrum of G0.

5.2 Synthesis of compound 2

Compound **3** (217 mg, 0.30 mmol), Pd(PPh₃)₂Cl₂ (21.0 mg, 0.03 mmol) and CuI (5.07 mg, 0.03 mmol) were mixed in 20 mL of TEA and stirred under nitrogen atmosphere. Trimethylsilylacetylene (147 mg, 1.50 mmol) was added dropwise to the reaction mixture over 30 minutes. After stirring at 55 °C for 12 hours, the reaction mixture was evaporated to remove the solvent, and the residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 10 : 1 ν/ν as the eluent) to afford the product as an oil liquid (203 mg, 93%). It was then treated with tetrabutylammonium S17

fluoride (219 mg, 0.84 mmol) in THF (20 mL) at 25 °C for 6 hours. After the deprotection reaction, the solvent was removed *in vacuo*, and the residue was extracted with H₂O/CH₂Cl₂. The combined organic extracts were dried over anhydrous Na₂SO₄ and evaporated with a rotary evaporator. The residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 50: 1 *v*/*v* as the eluent) to afford compound **2** as a white solid (175 mg, 86 %). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.31 (d, *J* = 7.7 Hz, 4H), 6.90 (d, *J* = 7.5 Hz, 4H), 6.72 (s, 2H), 6.60 (s, 1H), 4.93 (s, 4H), 3.95 (t, *J* = 6.4 Hz, 4H), 3.03 (s, 1H), 1.78 (dd, *J* = 13.8, 6.9 Hz, 4H), 1.44 (d, *J* = 7.5 Hz, 4H), 1.26 (s, 32H), 0.88 (t, *J* = 6.3 Hz, 6H).



Figure S25. ¹H NMR spectrum (400 MHz, CDCl₃, 298 K) of compound 2.

5.3 Synthesis of dendrimer G1

9,10-diiodoanthracene (22.2 mg, 0.05 mmol), compound **2** (100 mg, 0.15 mmol), $Pd(PPh_3)Cl_2$ (14.0 mg, 0.02 mmol) and CuI (3.8 mg, 0.02 mmol) were mixed in TEA (20 mL) and stirred under nitrogen atmosphere. After stirring at 75 °C for 12 hours, the reaction mixture was evaporated to remove the solvent, and the residue was extracted with H_2O/CH_2Cl_2 for three times. The combined organic extracts were dried over

anhydrous Na₂SO₄, and the solvent was removed with a rotary evaporator. The residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 10 : 1 *v/v* as the eluent) to afford dendrimer **G1** as an orange solid (45 mg, 58 %). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.67 (dd, J = 6.6, 3.3 Hz, 4H), 7.64 (dd, J = 6.7, 3.2 Hz, 4H), 7.38 (d, J = 8.6 Hz, 8H), 7.01 (d, J = 2.2 Hz, 4H), 6.93 (d, J = 8.6 Hz, 8H), 6.69 (t, J = 2.2 Hz, 2H), 5.04 (s, 8H), 3.97 (t, J = 6.6 Hz, 8H), 1.82–1.76 (m, 8H), 1.47–1.42 (m, 8H), 1.29–1.24 (m, 64H), 0.89 (d, J = 6.6 Hz, 12H). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 159.92, 159.15, 132.10, 129.39, 128.30, 127.23, 126.85, 124.60, 118.37, 114.61, 110.65, 103.42, 102.45, 86.10, 70.14, 68.05, 31.91, 31.42, 30.16, 29.65, 29.63, 29.59, 29.58, 29.40, 29.34, 29.24, 26.03, 22.68, 14.13. MALDI-TOF-MS *m*/*z*: [M + H]⁺, C₁₀₆H₁₃₉O₈, calculated 1540.0470; found 1540.0391.



Figure S26. ¹H NMR spectrum (400 MHz, CDCl₃, 298 K) of dendrimer G1.



Figure S27. ¹³C NMR spectrum (101 MHz, CDCl₃, 298 K) of dendrimer G1.



Figure S28. MALDI-TOF-MS spectrum of dendrimer G1.

5.4 Synthesis of compound 8

Compound 4 (313 mg, 0.50 mmol), methyl 3,5-dihydroxybenzoate (38.0 mg, 0.23 mmol) and K₂CO₃ (300 mg, 2.17 mmol) were added into 30 mL of DMF. After stirred at 80 °C for 12 hours, the solvent was evaporated under reduced pressure and the residue was extracted with H₂O/CH₂Cl₂ for three times. The combined organic extracts were dried over anhydrous Na₂SO₄ and the solvent was removed with a rotary evaporator. The residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 20 : 1 v/v as the eluent) to afford compound **8** as a white solid (150 mg, 91 %). ¹H NMR S20

(400 MHz, CDCl₃) δ (ppm): 7.33 (d, *J* = 8.6 Hz, 4H), 7.28 (d, *J* = 2.3 Hz, 2H), 6.90 (d, *J* = 8.6 Hz, 4H), 6.77 (t, *J* = 2.3 Hz, 1H), 4.98 (s, 4H), 3.95 (t, *J* = 6.6 Hz, 4H), 3.90 (s, 3H), 1.77 (dd, *J* = 14.7, 6.8 Hz, 4H), 1.47–1.41 (m, 4H), 1.35–1.25 (m, 32H), 0.88 (t, *J* = 6.8 Hz, 6H).



Figure S29. ¹H NMR spectrum (400 MHz, CDCl₃, 298 K) of compound 8.

5.5 Synthesis of compound 7

Compound **8** (287 mg, 0.40 mmol) was added into 30 mL of THF and stirred under nitrogen atmosphere at 0 °C. LiAlH4 (45.6 mg, 1.20 mmol) was then added dropwise to the reaction mixture over 30 minutes. After stirring at room temperature for 12 hours, the reaction mixture was quenched by methanol and the residue was extracted with H₂O/CH₂Cl₂ for three times. The combined organic extracts were dried over anhydrous Na₂SO₄, and the solvent was removed with a rotary evaporator. The residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 10 : 1 *v/v* as the eluent) to afford compound **7** as a white solid (245 mg, 89 %). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.33 (d, *J* = 8.6 Hz, 4H), 6.90 (d, *J* = 8.7 Hz, 4H), 6.61 (d, *J* = 2.2 Hz, 2H), 6.53 (t, *J* = 2.2 Hz, 1H), 4.95 (s, 4H), 4.63 (d, *J* = 4.4 Hz, 2H), 3.96 (t, *J* = 6.6 Hz, 4H), 1.77 (dd, *J* = 14.7, 6.8 Hz, 4H), 1.47–1.41 (m, 4H), 1.35–1.26 (m, 32H), 0.88 (d, *J* = 7.0 Hz, 6H).



5.6 Synthesis of compound 6

Compound 7 (200 mg, 0.29 mmol) was added into 30 mL of CH₂Cl₂ and stirred under nitrogen atmosphere at 0 °C. 1 mL of SOCl₂ was added dropwise to the reaction mixture over 10 minutes. After stirring at room temperature for 6 hours, the reaction mixture was quenched by H₂O and the residue was extracted with H₂O/CH₂Cl₂ for three times. The combined organic extracts were dried over anhydrous Na₂SO₄, and the solvent was removed with a rotary evaporator. The residue was then mixed with CsCO₃ (156 mg, 0.48 mmol) and 5-bromoresorcinol (22.7 mg, 0.12 mmol) in MeCN (20 mL) and stirred at 25 °C for 12 hours. After the reaction was complete, the solvent was removed in vacuo, and the residue was extracted with H₂O/CH₂Cl₂. The combined organic extracts were dried over anhydrous Na₂SO₄ and evaporated with a rotary evaporator. The residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 20 : 1 v/v as the eluent) to afford compound **6** as a white solid (138 mg, 75 %). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.32 (d, J = 8.6 Hz, 8H), 6.89 (d, J = 8.6Hz, 8H), 6.74 (d, *J* = 2.1 Hz, 2H), 6.63 (d, *J* = 2.1 Hz, 4H), 6.56 (d, *J* = 2.1 Hz, 2H), 6.51 (t, J = 2.1 Hz, 1H), 4.94 (d, J = 2.9 Hz, 12H), 3.95 (t, J = 6.6 Hz, 8H), 1.81–1.74 (m, 8H), 1.47-1.42 (m, 8H), 1.36-1.26 (m, 64H), 0.88 (t, J = 6.8 Hz, 12H).



Figure S31. ¹H NMR spectrum (400 MHz, CDCl₃, 298 K) of compound 6.

5.7 Synthesis of compound 5

Compound 6 (200 mg, 0.13 mmol), Pd(PPh₃)₂Cl₂ (14.0 mg, 0.02 mmol) and CuI (3.38 mg, 0.03 mmol) were mixed in 20 mL of TEA and stirred under nitrogen atmosphere. 2-Methyl-3-butyn-2-ol (200 mg, 2.38 mmol) was added dropwise to the reaction mixture over 30 minutes. After stirring at 80 °C for 12 hours, the reaction mixture was evaporated to remove the solvent, and the residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 10 : 1 v/v as the eluent) to afford the product as an oil liquid (155 mg, 78%). It was then treated with KOH (16.0 mg, 0.30 mmol) in toluene (20 mL) at 85 °C for 6 hours. After the deprotection reaction, the solvent was removed, and the residue was extracted with H₂O/CH₂Cl₂. The combined organic extracts were dried over anhydrous Na₂SO₄ and evaporated with a rotary evaporator. The residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 20 : 1 v/v as the eluent) to afford compound 5 as a white solid (127 mg, 85 %). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.32 (d, J = 8.6 Hz, 8H), 6.89 (d, J = 8.6Hz, 8H), 6.72 (d, *J* = 2.2 Hz, 2H), 6.64 (d, *J* = 2.1 Hz, 4H), 6.60 (t, *J* = 2.2 Hz, 1H), 6.55 (t, J = 2.1 Hz, 2H), 4.95 (d, J = 5.0 Hz, 12H), 3.95 (t, J = 6.6 Hz, 8H), 3.04 (s, 1H), 1.78 (dd, J = 14.3, 7.3 Hz, 8H), 1.47–1.42 (m, 8H), 1.28 (d, J = 15.5 Hz, 64H), 0.90– S23



Figure S32. ¹H NMR spectrum (400 MHz, CDCl₃, 298 K) of compound 5.

5.8 Synthesis of dendrimer G2

9,10-diiodoanthracene (15.9 mg, 0.037 mmol), compound 5 (120 mg, 0.08 mmol), Pd(PPh₃)₂Cl₂ (9.80 mg, 0.04 mmol) and CuI (5.33 mg, 0.03 mmol) were stirred in TEA (20 mL) under nitrogen atmosphere. After stirring at 75 °C for 12 hours, the reaction mixture was evaporated to remove the solvent, and the residue was extracted with H₂O/CH₂Cl₂ for three times. The combined organic extracts were dried over anhydrous Na₂SO₄, and the solvent was removed with a rotary evaporator. The residue was purified by flash column chromatography (petroleum ether/CH₂Cl₂, 15:1 v/v as the eluent) to afford G2 as an orange solid (93.7 mg, 81 %). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.67 (dd, J = 6.7, 3.2 Hz, 4H), 7.63 (dd, J = 6.6, 3.2 Hz, 4H), 7.32 (d, J = 8.6Hz, 16H), 7.01 (d, *J* = 2.2 Hz, 4H), 6.89 (d, *J* = 8.7 Hz, 16H), 6.71 (d, *J* = 2.1 Hz, 8H), 6.68 (t, J = 2.1 Hz, 2H), 6.58 (t, J = 2.1 Hz, 4H), 5.06 (s, 8H), 4.96 (s, 16H), 3.93 (t, J = 6.6 Hz, 16H), 1.77 (dd, J = 14.2, 7.3 Hz, 16H), 1.59 (s, 16H), 1.42 (d, J = 7.5 Hz, 16H), 1.25 (s, 112H), 0.87 (t, J = 6.8 Hz, 24H). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 160.25, 159.79, 159.05, 138.84, 132.11, 129.31, 128.47, 127.21, 126.92, 124.68, 118.34, 114.54, 110.78, 106.29, 103.39, 102.38, 101.62, 86.23, 70.20, 69.94, 68.01, 31.90, S24

29.65, 29.62, 29.59, 29.57, 29.39, 29.34, 29.23, 26.02, 22.68, 14.12. MALDI-TOF-MS *m*/*z*: [M + H]⁺, C₂₁₀H₂₈₃O₂₀, calculated 3125.1128; found 3125.0681.



Figure S33. ¹H NMR spectrum (400 MHz, CDCl₃, 298 K) of dendrimer G2.



Figure S34. ¹³C NMR spectrum (101 MHz, CDCl₃, 298 K) of dendrimer G2.



Figure S35. MALDI-TOF-MS spectrum of dendrimer G2.

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