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Electronic Supplementary Information

Strapped calix[4]pyrrole as a lithium salts selective receptor through separated ion-pair binding

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1. Materials and Measurements

All commercially available reagents were reagent grade and used without further purification. Tetrahydrofuran (THF) and CH₂Cl₂ were freshly distilled before each use. ¹H and ¹³C NMR spectra were recorded using a Bruker DPX 400 (400 MHz) spectrometer at 25 °C in MeOD, CDCl₃, CD₂Cl₂. MALDI-TOF-MS was performed on a Bruker Daltonics LRF20 with dithranol as the matrix. UV-Vis absorption spectra were recorded on a JASCO V-660 spectrometer. The SEM analysis of Li₂CO₃ was conducted by using a JEOL 7610F-PLUS instrument with energy-dispersive x-ray photoelectron spectroscopy (XPS) on a K-Alpha (Thermo Scientific, U.K.).

2. Synthesis

3: Chloroacetone (3.4 mL, 42 mmol) and sodium azide (5.4 g, 83 mmol) were added in acetone (100 mL) and the reaction mixture was stirred for 23 h at 25 °C. After the solvent was removed, the residue was extracted with CH₂Cl₂/H₂O, and the organic layer was concentrated. The residue was purified by column chromatography eluting with 16% ethylacetate/hexane to give **3** (4.2 g, 61%). ¹H NMR (400MHz, CDCl₃, 25 °C) δ (ppm): 3.95 (s, 2 H), 2.20 (s, 3 H).

4: Trifluoroacetic acid (1.5 mL) was added dropwise to **3** (2.0 g, 20 mmol) in pyrrole (50 mL, excess), and the reaction mixture was refluxed for 100 min. Triethylamine (1.5 mL) was added to a solution and the solution was concentrated under high vacuum to remove pyrrole. After extraction with CH_2Cl_2/H_2O and organic layer was washed with brine, the residue was

purified by column chromatography eluting with 40% CH₂Cl₂/Hexane to give **4** (3.4 g, 78%). ¹H NMR (400 MHz, CD₂Cl₂, 25 °C) δ (ppm): 8.15 (broad s, 2 H), 6.69 – 6.67 (m, 2 H), 6.15 – 6.13 (m, 2 H), 6.09 – 6.07 (m, 2 H), 3.73 (s, 2 H), 1.65 (s, 3 H).

5: NaH (540 mg, 22 mmol) was added to Schlenk flask, which was degassed under high vacuum. Anhydrous THF (20 mL) and triethyleneglycol (1.0 mL, 7.5 mmol) were added to the flask at 0 °C under N₂ and the reaction mixture was stirred. After 1 h, propargyl bromide (4.1 mL, 38 mmol) and KI (3 spoons with spatula) were added to the mixture solution, and stirred for 18 h at 25 °C. The reaction mixture was concentrated under high vacuum to remove propargyl bromide. After extraction with CH_2Cl_2/H_2O and organic layer was washed with brine, the residue was purified by column chromatography eluting with 40% ethylacetate/hexane to give **5** (1.6 g, 96%). ¹H NMR (400 MHz, CDCl₃, 25 °C) δ (ppm): 4.21 - 4.20 (d, J = 4 Hz, 4 H), 3.72 - 3.67 (m, 12 H), 2.43 - 2.42 (t, J = 4 Hz, 2 H).

6: **4** (410 mg, 1.9 mmol) and **5** (196 mg, 0.86 mmol) were added in anhydrous THF (30 mL). CuSO₄:5H₂O (552 mg, 3.50 mmol) and sodium ascorbate (685 mg, 3.50 mmol) dissolved in H₂O were added, and the reaction mixture was refluxed for 12 h. After filtration, the solution was extracted with NaCl(aq)/CH₂Cl₂, and the organic layer was concentrated. The residue was purified by column chromatography eluting with 5% MeOH/Ethylacetate to give **6** (173 mg, 30%). ¹H NMR (400 MHz, CDCl₃, 25 °C) δ (ppm): 8.77 (broad s, 4 H), 6.63 (s, 4 H), 6.45 (s, 2 H), 6.09 (s, 4 H), 5.99 (s, 4 H), 4.82 (s, 4 H), 4.42 (s, 4 H), 3.52 – 3.48 (broad d, 12 H), 1.47 (s, 6 H); MALDI-TOF-MS m/z : cald. for C₃₄H₄₄N₁₀O₄: 656.78 [M+H]⁺; found : 657.524. 1: BF₃·OEt₂ (0.5 mL) was added dropwise to **6** (483 mg, 0.74 mmol) in acetone (200 mL, excess), and the reaction mixture was stirred for 2 days at room temperature. Triethylamine (0.5 mL) was added to a solution and the reaction mixture was extracted with CH₂Cl₂/H₂O, and the organic layer was concentrated. The residue was purified by column chromatography eluting with 5% MeOH/Ethylacetate to give **1** (91 mg, 17%). ¹H NMR (400 MHz, CD₂Cl₂, 25 °C) δ (ppm): 7.68 (broad s, 4 H), 7.01 (s, 2 H), 6.08 – 5.95 (m, 8 H), 4.87 (s, 4 H), 4.55 (s, 4 H), 3.61 – 3.59 (broad d, 12 H), 1.66 (s, 6 H), 1.54 (s, 6 H), 1.50 (s, 6 H); ¹³C NMR (100 MHz, CD₂Cl₂, 25 °C) δ (ppm): 144.73, 138.65, 132.85, 123.93, 105.41, 104.24, 70.50, 70.35, 70.09, 64.58, 40.93, 35.47, 31.12, 28.98, 26.84; MALDI-TOF-MS m/z : cald. for C₄₀H₅₂N₁₀O₄: 736.91 [M+H]⁺; found : 737.660.

3. The procedure for precipitation experiments

Using K_2CO_3 diluted in MeOH: Through liquid-liquid extraction, LiCl in aqueous phase were extracted to organic phase using 1 in a vial. To remove aqueous phase, the solution was filtered on hydrophobic membrane. The organic solution (CH₂Cl₂) became turbid upon addition of K₂CO₃ diluted in MeOH.

Using CO₂(g) bubbling: the excess of LiCl(s) was poured to the solution containing 5 mg/mL of 1 in CH₂Cl₂ and then the mixture was stirred for 1 h to extract LiCl. After the solution was filtered to remove salt, deionized water (1.0 mL) was added to the residue. Gaseous CO₂ bubbling was conducted at 0 °C.

4. X-ray crystallographic data of 1•LiCl complex

Data collection

The diffraction data from yellow crystals of 1•LiCl ($0.095 \times 0.054 \times 0.034$ mm3) mounted on a MiTeGen MicroMount© were collected at 100 on a ADSC Quantum 210 CCD diffractometer with synchrotron radiation (0.7000 Å) at Supramolecular Crystallography 2D, Pohang Accelerator Laboratory (PAL), Pohang, Korea. The ADSC Q210 ADX program¹ was used for data collection (detector distance is 63 mm, omega scan; $\Delta \omega = 1^{\circ}$, exposure time is 1 sec/frame for 1•LiCl and HKL3000sm (Ver. 703r)² was used for cell refinement, reduction and absorption correction. The crystal structures of 1•LiCl was solved by the direct method with SHELX-XT (Ver. 2014/5)³ and refined by full-matrix least-squares calculations with the SHELX-XL (Ver. 2016/4)⁴ program package.

Structure solution and refinement

The systematic absences in the diffraction data were uniquely consistent for the triclinic space group P-1 that yielded chemically reasonable and computationally stable results of refinement.⁴⁻⁵

A successful solution by the direct methods provided most non-hydrogen atoms from the Emap. The remaining non-hydrogen atoms were located in an alternating series of leastsquares cycles and difference Fourier maps. All non-hydrogen atoms were refined with anisotropic displacement coefficients. All hydrogen atoms were included in the structure factor calculation at idealized positions and were allowed to ride on the neighboring atoms with relative isotropic displacement coefficients. The final least-squares refinement of 597 parameters against 10770 data resulted in residuals R (based on F2 for I \geq 2 σ) and wR (based on F2 for all data) of 0.0526 and 0.1559, respectively. The final difference Fourier map was featureless.

Summary

Crystal Data for C43H62.5CILiN11.5O7 (M =894.93 g/mol): triclinic, space group P-1 (no. 2), a = 9.7210(19) Å, b = 14.384(3) Å, c = 17.153(3) Å, $\alpha = 88.50(3)^{\circ}$, $\beta = 86.85(3)^{\circ}$, $\gamma = 77.08(3)^{\circ}$, V = 2334.0(9) Å3, Z = 2, T = 100 K, μ (synchrotron) = 0.138 mm-1, Dcalc = 1.273 g/cm3, 20744 reflections measured (3.672° $\leq 2\Theta \leq 55.996^{\circ}$), 10770 unique (Rint = 0.0198, Rsigma = 0.0310) which were used in all calculations. The final R1 was 0.0526 (I > 2σ (I)) and wR2 was 0.1559 (all data).

Table S1.	Crystal	data	and	structure	refineme	nt for	1•	Li	Cl
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Identification code	НССР3
Empirical formula	C43H62.5ClLiN11.5O7
Formula weight	894.93
Temperature/K	100
Crystal system	triclinic
Space group	P-1
a/Å	9.7210(19)
b/Å	14.384(3)
c/Å	17.153(3)
α/°	88.50(3)
β/°	86.85(3)
γ/°	77.08(3)
Volume/Å3	2334.0(9)
Ζ	2
pcalcg/cm3	1.273
μ/mm-1	0.138
F(000)	954.0
Crystal size/mm3	$0.095\times0.054\times0.034$
Radiation	synchrotron ($\lambda = 0.700$)
2O range for data collection/°	3.672 to 55.996
Index ranges	$\text{-13} \le h \le 13, \text{-19} \le k \le 19, \text{-20} \le l \le 21$
Reflections collected	20744
Independent reflections	10770 [Rint = 0.0198, Rsigma = 0.0310]
Data/restraints/parameters	10770/0/597
Goodness-of-fit on F2	1.059
Final R indexes [I>= 2σ (I)]	R1 = 0.0526, $wR2 = 0.1503$
Final R indexes [all data]	R1 = 0.0605, wR2 = 0.1559
Largest diff. peak/hole / e Å-3	0.62/-0.56



Figure S1. Partial ¹H NMR spectra of (a) 1 and 1 with excess of (b) LiF(s), (c) LiCl(s), (d) LiBr(s), (e) LiI(s), and (f) $LiClO_4(s)$ (400 MHz, $CD_2Cl_2:CD_3OD=9:1$, 297 K).



Figure S2. Partial ¹H NMR spectra to confirm complexation of **1**•LiCl in solution state. (a) only **1** (10 mM) and **1** with (b) excess of TBACl, (c) TBACl followed by excess of LiClO₄, (d) excess of LiCl. (all spectra were recorded in 10% CD₃OD/CD₂Cl₂, 297 K)



Figure S3. Partial ¹H NMR spectra to confirm complexation of 1•LiBr in solution state. (a) only 1 and 1 with (b) excess of TBABr, (c) TBABr followed by excess of LiClO₄, (d) excess of LiBr.(all spectra were recorded in 10% CD₃OD/CD₂Cl₂, 297 K)

Table S2. The binding constants of 1 with various guests in 10% MeOD-CD₂Cl₂.

К _а (М ⁻¹)	TBACI	TBABr	LiCIO ₄	LiCI	LiBr
1	377±1.07	61±1.04	4.4±1.13	60.5±1.02	19.5±1.01

 Table S3. The binding constant of other system in literatures.

Reference	Guest	Solvent	Binding constant
Ref. 7	LiCl	10% MeOD-CD ₂ Cl ₂	45±1
Ref. 8	LiCl	10% D ₂ O-THF-d ₈	(8.3±0.3)x10 ²



Figure S4. (a) stacked partial ¹H NMR spectra of **1**(10 mM) with LiCl in 10% CD₃OD/CD₂Cl₂, (b) chemical shift changes of **1**'s protons upon addition of LiCl, (c) binding constant of **1**• LiCl calculated by HypNMR2008.



Figure S5. (a) stacked partial ¹H NMR spectra of **1** (10 mM) with LiBr in 10% CD₃OD/CD₂Cl₂, (b) chemical shift changes of **1**'s protons upon addition of LiBr, (c) binding constant of **1**• LiBr calculated by HypNMR2008.



Figure S6. (a) stacked partial ¹H NMR spectra of **1** (10 mM) with LiClO_4 in 10% CD₃OD/CD₂Cl₂, (b) chemical shift changes of **1**'s protons upon addition of LiClO_4 , (c) binding constant of **1**•LiClO₄ calculated by HypNMR2008.



Figure S7. DFT optimized structure of 1-LiCl complex, (a) water-separated ion pair (solvent bridged), (b) contact ion pair, and (c) separated ion pair.⁶



Figure S8. ¹H-NOESY spectrum of 1 (10 mM) in 10% MeOD-CD₂Cl₂.



Figure S9. ¹H-NOESY spectrum of 1·LiClO₄ (10 mM) in 10% MeOD-CD₂Cl₂.



Figure S10. ¹H-NOESY spectrum of 1·LiCl (2.5 mM) in 10% MeOD-CD₂Cl₂.



Figure S11. ¹H NMR spectra of (a) **1** (5 mM) and **1** after extraction with (b) 25% saturated LiCl (aq), (c) 50% saturated LiCl (aq). (d) 80% saturated LiCl (aq), (e) 95% saturated LiCl (aq), and (f) 100% saturated LiCl (aq). (all spectra were recorded CD₂Cl₂, 297 K).



Figure S12. ¹H NMR spectra of (a) **1** (5 mM) and **1** after extraction with excess of (b) NaCl(aq), (c) KCl(aq), (d) NaBr(aq), and (e) KBr(aq) (all spectra were recorded CD₂Cl₂, 297 K).



Figure S13. ¹H NMR spectra of (a) 1 (5 mM) and 1 after extraction with (b) saturated LiBr(aq), (c) excess LiBr(s), (d) excess LiBr(s), NaBr(s) and KBr(s), (e) excess of NaBr (s), and (f) excess of KBr (s) (400 MHz, CD_2Cl_2 , 297 K).



Figure S14. UV/Vis spectral titration for methyl orange (**MO**) as a pH indicator using 0.05 M HCl and its photographs: (a) and (e) only deionized water, the solution containing lithium carbonate extracted by (b) and (f) 20 mg of 1, (c) and (g) 40 mg of 1.



Figure S15. (a) The binding isotherm graph for acid-base titration and (b) the amount of lithium carbonate extracted by 1 was summarized on table.



Figure S16. $Li_2CO_3(s)$ was precipitated by addition of K_2CO_3 diluted in methanol into the solution containing **1**-LiCl complex; (a) before addition of K_2CO_3 and (b) after addition of K_2CO_3 .



Figure S17. The images of SEM/EDS mapping of oxygen and carbon for A) commercial lithium carbonate and B) lithium carbonate extracted by **1**.

¹H and ¹³C NMR spectra of each compounds



Figure S18. ¹H NMR spectrum of 3.



Figure S19. ¹H NMR spectrum of 4.



Figure S20. ¹H NMR spectrum of 5.



Figure S21. ¹H NMR spectrum of 6.



Figure S22. ¹H NMR spectrum of 1.



Figure S23. ¹³C NMR spectrum of 1.



Figure S24. MALDI-TOF-MS spectrum of 1.

References

(1) Arvai, A. J.; Nielsen, C. ADSC Quantum-210 ADX Program, Area Detector System Corporation: Poway, CA, USA, 1983.

(2) Otwinowski, Z.; Minor, W. Methods in Enzymology; Carter Jr., C. W. Jr.; Sweet, R. M., Eds.; Academic Press: New York, 1997, vol. 276, part A, pp. 307-326.

(3) Sheldrick, G. M. Acta Cryst. 2015, A71, 3-8.

(4) Sheldrick, G. M. Acta. Cryst. 2015, C71, 3-8.

(5) Dolomanov, O. V.; Bourhis, L. J.; Gildea, R. J.; Howard, J. A. K.; Puschmann, H. J. Appl. Cryst. 2009, 42, 339-341.

(6) Gaussian 09, Revision A.02, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, and D. J. Fox, Gaussian, Inc., Wallingford CT, 2009.

(7) Q. He, Z. Zhang, J. T. Brewster, V. M. Lynch, S. K. Kim and J. L. Sessler, *J. Am. Chem. Soc.*, 2016, 138, 9779-9782

(8) Q. He, N. J. Williams, J. H. Oh, V. M. Lynch, S. K. Kim, B. A. Moyer and J. L. Sessler, *Angew. Chem. Int. Ed.*, 2018, 57, 11924-11928