Electronic Supplementary Information

Catalytic O₂ activation with synthetic models of a-ketoglutarate dependent oxygenases based in triazacyclononane ligands

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1. Experimental Section

Materials and methods. Reagents and solvents used were of commercially available reagent quality unless otherwise stated. Preparation and handling of air-sensitive materials were performed under an inert atmosphere in a glove box. Acetonitrile and ¹⁸O₂ (99%) were purchased from Sigma-Aldrich and acetone from Scharlau. H₂¹⁸O (95% ¹⁸O-enriched) was received from ICON Isotopes. 1,4,7-tris(*p*-toluenesulfonyl)-1,4,7-triazacyclononane (Ts₃tacn) was obtained from Catexel.

Instrumentation

Infrared spectra were collected on a Nicolet Avatar 360 FTIR spectrometer. UV-Visible spectroscopy was performed in a 1 cm quartz cell using an Agilent Technology 8453 UV-Vis spectrophotometer equipped with a diode-array detector. NMR spectra were recorded on a Bruker DPX 400 MHz spectrometer in CDCl₃ or CD₃CN solvent using standard conditions and were referenced to the residual proton signal of the solvent. Elemental analysis was performed on a 4.1 Vario EL 3-elemental analyzer from Elementar. The ESI-MS experiments were performed with a Bruker esquire 6000 LC/MS chromatograph, using acetonitrile as a mobile phase. The product analyses after catalysis experiments were carried out on an Agilent Technology 7820A gas chromatograph with 16-sample automatic liquid sampler, flame ionization detector and EzChrom Elite Compact software. GC-MS analyses were performed on an Agilent Technology 7890A GC system equipped with 5975C inert XL EI/CI MSD with Triple-Axis Detector. The products were identified by comparison of their GC retention times and GC/MS with those of authentic compounds.

Synthesis and characterization of ligands

1,4,7-Triazacyclononane hexahydrochloride (H $_3$ **tacn·6HCl)** was prepared following a similar procedure to that described in the literature.¹

In a 250 mL round bottomed flask with a stirrer, Ts₃tacn (25 g, 0.041 mol) was dissolved in concentrated H₂SO₄ (50 mL). The resulting solution was heated at 170 °C for 30 min. After this period the solution was poured into cold ethanol (180 mL) which led to the formation of a white-brown solid. After dropwise addition of diethylether (270 mL) the resulting mixture was stored in the fridge overnight. The resulting brown solid was filtered off and dissolved in water (75 mL) and refluxed at 120°C for 30 min. The resulting solution was filtered through Celite[®] to remove impurities. Water evaporation from the filtrates afforded a brown oil, to which HCl conc. (18 mL) was added together with ethanol (75 mL). This caused the immediate precipitation of a yellowish solid, which was filtered and washed with ethanol affording 12.5 g of H₃tacn·6HCl. Yield: 86%. ¹H NMR (D₂O, 300 MHz, 25°C) δ , ppm: 3.49 (s, 12H). ¹³C NMR (D₂O, 400 MHz, 25 °C) δ , ppm: 51. Anal. Calcd. for C₆H₁₅N₃: C, 20.71; H, 6.08; N, 12.08. Found: C, 22.86; H, 5.82; N, 11.98.

1,4,7-tris(isopropyl)-1,4,7-triazacyclononane (*i***P** $_3$ **tacn)** was prepared following a similar procedure to that described in the literature.² In a 100 mL round bottomed flask H₃tacn·6HCl (2.35 g, 6.75 mmol), isopropylbromide (1.99 mL, 0.0212 mol) and K₂CO₃ (23.5 g, 0.170 mol) were dissolved in 50 mL of acetonitrile. The mixture was refluxed for 24 h at 90 °C. The resulting solution was filtered, and the acetonitrile from the filtrates was removed under reduced pressure. Afterwards NaOH 1 M (30 mL) was added to the resulting solid and the mixture was extracted with CH₂Cl₂ (3 x 20 mL). The combined organic phases were dried over MgSO₄, filtered and the solvent was removed under reduced pressure. 1.23 g of a yellowish oil corresponding to *i*Pr₃tacn were obtained. Yield = 71 %. ¹H

NMR (CDCl₃, 400 MHz, 25°C) δ, ppm: 0.94 (d, 18H), 2.61 (s, 12H), 2.85 (hept, 3H). ¹³C NMR (CDCl₃, 100 MHz, 25°C) δ, ppm: 18.49, 52.97, 54.56. ESI-MS (m/z): 256.2 [M+H]⁺..

Ligand *i*Bu₃tacn was prepared following a modified literature protocol.³

N,N',N"-tri(isopropylcarbonyl)-1,4,7-triazacyclononane ((*i***PrCO)₃tacn). In a 500 mL two-necked round-bottomed flask H₃tacn·6HCl (3.23 g, 9.28 mmol) was dissolved in acetonitrile (250 mL). To this solution 18 equiv of triethylamine were added (23 mL, 167 mmol) and the reaction mixture was stirred at room temperature for 15 min. Then 3 equiv of isobutyryl chloride (3 mL, 27.8 mmol) were added dropwise.** The resulting solution was stirred overnight at 70 °C. After addition of water (100 mL) the acetonitrile solvent was removed under reduced pressure. The resulting aqueous mixture was extracted with CH_2Cl_2 (3 x 20 mL) and the combined organic phases were dried over MgSO₄. Filtration and solvent evaporation under reduced pressure afforded 2.81 g of (*i*PrCO)₃tacn as a white solid. Yield: 89 %. ¹H NMR (CDCl₃, 400 MHz, 25 °C) δ , ppm: 1.1 (d, 18H), 2.71 (hept, 3H), 3.39 (m, 6H), 3.69 (m, 6H). ¹³C NMR (CDCl₃, 100 MHz, 25 °C) δ , ppm: 19.69, 30.66, 48.44, 51.50, 178.49. ESI-MS (m/z): 362.24 [M+Na]⁺, 701.49 [2M+Na]⁺.

1,4,7-tris(isobutyl)-1,4,7-triazacyclononane (iBu₃tacn). In a 250 mL round-bottomed schlenck flask, (iPrCO)₃tacn (2.81 g, 8.3 mmol) was dissolved in CH₂Cl₂ (50 mL) and the solution was cooled down in an ice bath. Under a N₂ atmosphere 6 equiv LiAlH₄ (49 mL of a 1 M solution in diethylether, 49 mmol) were added dropwise and the resulting solution was stirred overnight at room temperature. After this period, ethyl acetate (50 mL) was slowly added at 0 °C under N₂. Evaporation of the solvents under reduced pressure afforded a white solid, which was stirred in 100 mL of CH₂Cl₂ for 2 h. The solid was filtered off and washed with CH₂Cl₂ (2 x 20 mL). The organic filtrates were extracted with NaOH 1 M (100 mL) and the aqueous phase was further extracted with CH₂Cl₂ (2 x 20 mL). The combined organic fractions were dried over MgSO₄ and filtered. Solvent evaporation under reduced pressure afforded 1.94 g of *i*Bu₃tacn as a white solid. Yield: 79%. ¹H NMR (CDCl₃, 400 MHz, 25 °C) δ , ppm: 0.9 (d, 18H), 1.65 (hept, 3H), 2.20 (d, 6H), 2.7 (s, 12H). ¹³C NMR (CDCl₃, 100 MHz, 25 °C) δ , ppm: 1.16, 21.24, 27.05, 29.85, 56.60, 68.06. ESI-MS (m/z): 298.32 [M+H]⁺.

Sodium benzoylformate (NaBF). In a 25 mL round-bottomed flask NaOH (262 mg, 6.55 mmol) was dissolved in methanol (5 mL) and to this solution benzoylformic acid (990 mg, 6.55 mmol) was added directly as a solid. The resulting mixture was stirred at room temperature for 2h. After this time the solvent was evaporated and the resulting white solid was dried in the vacuum line at 40 °C for 2 h. 1.07 g of NaBF were obtained as a white solid. Yield: 95%. ¹H NMR (D₂O, 400 MHz, 25°C) \square , ppm: 7.63 (t, 2H), 7.78 (t, 1H), 7.97 (d, 2H). ESI-MS (m/z): 148.7 [M-Na]⁺. Anal. Calcd. for C₈H₅NaO₃: C, 55.83; H, 2.93. Found: C, 55.37; H, 2.56.

Synthesis and characterization of complexes

[Fe(BF)(CF₃SO₃)(*i***Pr₃tacn)] (1BF).** Inside the glove box *i*Pr₃tacn (212 mg, 0.83 mmol) was dissolved in CH₃CN (250 µL). In a separate vial an equimolar amount of [Fe(CF₃SO₃)₂(CH₃CN)₂] (362 mg, 0.83 mmol) was solubilized in CH₃CN (500 µL). The colorless iron salt solution was poured onto the ligand solution which caused the immediate color change to pale yellow. After stirring for 5 min sodium benzoylformate (143 mg, 0.83 mmol) was added directly as a solid. The solution turned deep purple immediately. The reaction was stirred for 1 h at room temperature and filtered through a Celite[®] plug. Slow diethyl ether over the resulting solution afforded 387 mg of nice square purple crystals corresponding to **1BF**. Yield: 76%. ¹H NMR (CD₃CN, 400 MHz, 25 °C) δ , ppm: -14.55 (s, 6H), 11.07 (s, *m*-2H), 13.08 (s, *p*-1H), 23.95 (s, *o*-2H); 46.85 (s, 1H); 132.08 (s, 4H). ESI-MS (CH₃CN): m/z = 256.2757

 $[iPr_3tacn]^+$, 460.2260 $[Fe(BF)(iPr_3TACN)]^+$. UV-Vis (1.3 mM, CH₃CN, 25 °C) λ , nm: 370 (390 M⁻¹cm⁻¹), 540 (390 M⁻¹cm⁻¹), 568 (390 M⁻¹cm⁻¹). μ_{eff} (CD₃CN, 25°C): 5.25 μ_B .

[Fe(Bz)(*i***Pr₃tacn)(CH₃CN)](CF₃SO₃) (1Bz).** In a vial inside the glovebox *i*Pr₃tacn (50 mg, 0.19 mmol) was dissolved in acetonitrile (500 μL). An equimolar amount of $[Fe(OTf)_2(CH_3CN)_2]$ (84.7 mg, 0.19 mmol) was solubilized in acetonitrile (500 μL) in a separate vial and added dropwise onto the *i*Pr₃tacn solution, affording a pale yellow mixture. After stirring for 5 min, sodium benzoate (28 mg, 0.19 mmol) was added as a solid. The resulting solution was filtered through a Celite[®] plug. 113 mg of pale yellow crystals were isolated from diethyl ether diffusion over the solution of **1Bz**. Yield: 92%. ¹H NMR (CD₃CN, 400 MHz, 25°C) δ, ppm: -20.32 (s, 6H), 11.45 (s, *m*-2H), 18.08 (s, *p*-1H), 29.34 (s, *o*-2H); 44.65 (s, 1H); 130.49 (s, 4H). ESI-MS (CH₃CN): m/z = 256.2753 [*i*Pr₃tacn]⁺, 432.2322 [Fe(BZ)(*i*Pr₃tacn)]⁺. UV-Vis (1.3 mM, CH₃CN, 25°C) λ, nm: 324 (330 M⁻¹cm⁻¹), 378 (200 M⁻¹cm⁻¹). μ_{eff} (CD₃CN, 25°C): 5.03 μ_B.

[Fe₂(BF)₂(μ -Cl)(*i*Bu₃tacn)₂](ClO₄) (2BF-ClO₄). In the glove box, *i*Bu₃tacn (34 mg, 0.11 mmol) was dissolved in CH₃CN (500 μ L) and under stirring FeCl₂ (14 mg, 0.11 mmol) suspended in CH₃CN (1 mL) was added dropwise. To the resulting pale-yellow solution 2 equiv AgClO₄ (25 mg, 0.22 mmol) were added directly as a solid, causing the immediate precipitation of a white solid corresponding to AgCl. After stirring for 10 min, 1 equiv sodium benzoylformate (19 mg, 0.11 mmol) was added directly as a solid and the reaction was stirred for 1 h. Then the mixture was filtered using an Acrodisc obtaining a dark green solution. Slow diffusion of diethyl ether over this solution afforded 89 mg of orange crystals suitable for X-ray diffraction. Yield: 74%. ESI-MS (m/z): 1039.5204 [M-ClO₄]⁺.

Reactivity tests. Inside the glovebox, complexes **1BF**, **1Bz** or **2BF-ClO**₄ (6.56 µmol) were dissolved in anhydrous solvent (5 mL acetonitrile or acetone) and 100 equiv methyl *p*-tolyl sulfide (656 µmol) were added to the mixture. The vial was capped with a rubber cap and taken out of the glovebox. A balloon filled with O₂ was gently bubbled through the solution. Different treatments were followed depending on the final compound to be analyzed. For *sulfoxide quantification*, the reaction was stopped after 4 hours and a known amount of biphenyl was added as an internal standard; the solution was filtered through a silica plug and washed with ethyl acetate. The resulting solution was analyzed by GC. For *benzoic acid* quantification, after 4 h of reaction half equivalent of 1,3,5trimethoxybenzene (TMB) was added as an internal standard, the solution was evaporated under vacuum and then HCl 3 M (10 mL) was added. The mixture was extracted 3 times with diethyl ether and the combined organic phases were washed twice with brine and finally dried over MgSO₄. After filtration, the resulting solution was concentrated and analyzed by ¹H NMR. Quantification of benzoic acid was carried out by comparison of the integrals of the aromatic protons of the TMB and the α -protons of the benzoic acid.

Isotope labeling experiments. Following the procedure described above but using ¹⁸O₂ instead of ¹⁶O₂, the incorporation of ¹⁸O into sulfoxide and benzoic acid products was determined by GC-MS and ESI-MS in negative mode, respectively. ¹⁸O-labeled water exchange experiments were carried out following the procedure described above but adding different amounts of H₂¹⁸O (10-80 equiv with respect to the Fe complex) prior to oxygenation. Analysis of the amount of ¹⁸O incorporated into the sulfoxide products was performed by GC-MS.

Catalytic tests. Inside the glovebox **1BF** (6.56 μ mol) was dissolved in acetone (5 mL). Then the appropriate amounts of methyl *p*-tolyl sulfide (20 – 100 equiv with respect to **1BF**) and HBF (3 – 100 equiv with respect to **1BF**) were added to the reaction mixture. The vial was capped with a rubber cap and taken outside of the glovebox. A balloon filled with O₂ gas was gently bubbled to the solution for 5 min and an O₂ atmosphere was kept for 4 hours under stirring. After this period, a known amount of biphenyl was added to the solution as an internal standard. The resulting mixture was filtered through a silica plug that was afterwards washed with ethyl acetate. The resulting solution was analyzed by GC and the amount of sulfoxide produced was determined.

Hammett plot analysis. Inside the glovebox **1BF** (6.56 μ mol) was dissolved in acetone (5 mL) and different amounts of pairs of p-X-thioanisoles were mixed until a total amount of 200 equiv of substrate were added. The vial was capped with a rubber cap and taken outside of the glovebox. A balloon filled with O₂ gas was gently bubbled to the solution for 5 min and an O₂ atmosphere was kept for 2 hours under stirring. After this period, biphenyl was added as an internal standard and the resulting solution was analyzed by GC.

2. Compounds characterization



Figure S1. ¹H NMR spectrum of H₃tacn·6HCl in D₂O at 25°C. *acetone



Figure S2. ^{13}C NMR spectrum of H3tacn·6HCl in D2O at 25°C







Figure S8. ¹³C NMR spectrum of (*i*PrCO)₃tacn in CDCl₃ at 25°C



Figure S10. ¹³C NMR spectrum of *I*Bu₃tacn in CDCl₃ at 25°C.





Figure S13. ¹H NMR spectrum of **2BF-ClO**₄ in CD₃CN at 25°C



Figure S14. ESI-MS spectrum of tacn 6HCl.







Figure S16. ESI-MS spectrum of of *i*Pr₃tacn.







Figure S18. ESI-MS spectrum of of *i*Bu₃tacn.



Figure S19. ESI-MS spectrum of **1BF** in CH₃CN. Inset: Isotopic distribution comparison between experimental and simulated data.



Figure S20. ESI-MS spectrum of **1Bz** in CH₃CN. Inset: Isotopic distribution comparison between experimental and simulated data.



Figure S21. ESI-MS spectrum of **2BF-CIO**₄ in CH₃CN. Inset: Isotopic distribution comparison between experimental and simulated data.

3. Crystallographic data

1BF		1Bz		2BF-CIO ₄	
Fe1-N1	2.247(2)	Fe1-N1	2.203(2)	Fe1-N1	2.261(3)
Fe1-N2	2.248(2)	Fe1-N1	2.2127(18)	Fe1-N2	2.252(3)
Fe1-N3	2.239(2)	Fe1-N4	2.159(2)	Fe1-N3	2.212(3)
Fe1-01	2.0389(2)	Fe1-01	2.0868(18)	Fe1-01	2.105(2)
Fe1-O3	2.215(2)	Fe1-02	2.3366(19)	Fe1-O4	2.101(2)
Fe1-04	2.195(2)	Fe1-N3	2.203(2)	Fe1-Cl1	2.4513(10)
				Fe2-N4	2.220(3)
				Fe2-N5	2.220(3)
				Fe2-N6	2.243(3)
				Fe2-O2	2.112(2)
				Fe2-05	2.093(3)
				Fe2-Cl1	2.4696(10)

Table S1. Selected bond distances (Å) for 1BF, 1Bz and 2BF-ClO₄.

	1BF	1Bz	2BF-CIO ₄	
Formula	$C_{24}H_{38}F_{3}FeN_{3}O_{6}S$	$C_{27}H_{44}F_3FeN_5O_5S$	$C_{56}H_{98}C_{12}Fe_2N_6O_{11}$	
Molecular weight	609.48	663.58	1214.00	
Crystal system	Triclinic	Triclinic	Monoclinic	
Space group	P-1	P-1	P2(1)/n	
Wavelength (Å)	0.71073	0.71073	0.71073	
Crystal color	Purple	Colourless-yellow	Orange	
Т (К)	293(2)	180.(2)	100(2) K	
Size (mm)		0.1 x 0.1 x 0.1	0.25 x 0.14 x 0.14	
a (Å)	9.1103(9)	10.8333(8)	12.832(3)	
b (Å)	9.9440(10)	12.2736(8)	15.977(3)	
c (Å)	15.5023(16)	13.0120(9)	30.509(6)	
α (°)	77.338(2)	79.726(2)	90	
β (°)	89.422(2)	74.064(2)	98.773(4)	
γ (°)	88.622(2)	81.746(2)	90	
V (Å ³)	1369.8(2)	1628.8(2)	6182(2)	
hkl ranges	-12 ≤ h ≤ 12	-14 ≤ h ≤ 14	-16 ≤ h ≤ 16	
	-12 ≤ k ≤ 12	-16 ≤ k ≤ 16	-20 ≤ k ≤ 20	
	-8 ≤ ≤ 19	-17 ≤ 1 ≤ 17	-39 ≤ l ≤ 39	
ρ _{calc} (g cm⁻³)	1.478	1.353	1.304	
Ζ	2	2	4	
F(000)	640	700	2600	
μ (mm⁻¹)	0.691	0.586	0.616	
heta range (°)	1.346 to 28.297	3.35 to 28.24	1.64 to 27.50	
Absorption corr.		Multi-scan	Empirical	
T _{max} , T _{min}	0.989, 0.886	0.94, 0.53	1.0, 0.896233	
Refinement	Fsqd	Fsqd	Full-matrix least-squares on F ²	
Reflections	6023	7986	14176	
Data/restr./param.	6023 / 0 / 349	7986 / 0 / 445	14176 / 0 / 713	
Goodness-of-fit	1.008	1.094	1.110	
	2.000	0.0704		
к	0.0///	0.0704	0.0965	
K _w	0.1030	0.1385	0.1564	
Peak, hole (e A³)	0.479, -0.352	0.648, -0.576	1.063, -0.514	

Table S2. Selected crystallographic data for $\mathbf{1BF},\,\mathbf{1Bz}$ and $\mathbf{2BF}\text{-}\mathbf{ClO}_4$



Figure S22. Mercury diagram at 30% of probability level of **1BF**. Hydrogens and counteranion CF_3SO_3 were removed for clarity.



Figure S23. Mercury diagram at 30% of probability level of 1Bz. Hydrogens and counteranion ${\sf CF}_3{\sf SO}_3$ were removed for clarity.



2BF·CIO₄

Figure S24. Mercury diagram at 30% of probability level of **2BF·ClO**₄. Hydrogens and counteranion ClO₄ were removed for clarity.

4. O₂ reactivity



Figure S25. UV-Vis spectra of the reaction of 1BF with O₂ ([Fe] = 1.3 mM) in CH₃CN at 25°C



Figure S26. Time trace of the decay of the band at 550nm together with the quantification over time of the production of HBz and methyl *p*-tolyl sulfoxide for the reaction of **1BF** and O_2 in CH₃CN at 25°C. $k_{obs} = (6.3 \pm 0.1) \times 10^{-5} \text{ s}^{-1} (\tau = 183 \text{ min})$



Figure S27. UV-vis spectra of the reaction of **2BF-ClO**₄ with O₂ ([Fe] = 1.3 mM) in CH₃CN at 25° C.



Figure S28. Time trace of the decay of the band at 600 nm from the reaction of **2BF-ClO**₄ with O₂ in CH₃CN at 25°C. k_{obs} = (5.9 ± 0.1) x 10⁻³ s⁻¹ (τ = 1.9 min)

5. Mechanistic studies



Figure S29. Hammett plot for the oxygenation of **1BF** in the presence of p-X-thioanisoles obtained in competition experiments by analysis of the final product mixtures.



Figure S30. Incorporation of ¹⁸O into the sulfoxide product obtained in the oxygenation of **1BF** in the presence of 100 equiv *p*-Cl-thioanisole or *p*-CH₃-thioanisole and variable amounts of $H_2^{18}O$ (from 0 to 80 equiv).



Figure S31. GC-MS spectrum of methyl *p*-tolyl sulfoxide formed from the oxidation of methyl *p*-tolyl sulfide in the reaction of **1BF** with ${}^{16}O_2$.



Figure S32. GC-MS spectrum of methyl *p*-tolyl sulfoxide formed from the oxidation of methyl *p*-tolyl sulfide in the reaction of **1BF** with ¹⁸O₂, exhibiting 90% ¹⁸O-incorporation.



Figure S33. ESI-MS spectrum of benzoic acid formed after the oxidation of methyl p-tolyl sulfide in the reaction of **1BF** with ${}^{16}O_2$.



Figure S34. ESI-MS spectrum of benzoic acid formed after the oxidation of methyl *p*-tolyl sulfide in the reaction of **1BF** with ¹⁸O₂, exhibiting 90% ¹⁸O-incorporation.

6. Catalytic studies



Figure S35. Formation of methyl *p*-tolyl sulfoxide over time in the oxygenation of methyl *p*-tolyl sulfide catalyzed by **1BF**.



protonated form of *i*Pr₃tacn.

Table S3. Screening of catalytic conditions



(20 - 100 equiv)

entry	HBF (equiv)	sulfide (equiv)	sulfoxide (TN)	Observations/hypothesis
1	20	20	8	Reference reaction
				Addition of reducing agents aiming at restoring deactivated ferric species
2	10	20	8	1 equiv. ascorbic acid (0.5 equiv added after 2h and 4h).
3	20	20	3	0.5 equiv. DDT added after 2 h
				Addition of sulfoxide to test possible product inhibition
4	10	20	7	Reaction run in the presence of 10 equiv thioanisole sulfoxide
				Addition of Fe(II) aiming at restoring activity by binding to the demetallated ligand
5	10	20	7	0.5 equiv $[Fe(CF_3SO_3)_2(CH_3CN)_2]$ and 0.5 equiv NaBF added after 2h
6	20	20	9	0.5 equiv $[Fe(CF_3SO_3)_2(CH_3CN)_2]$ and 0.5 equiv NaBF added after 2h
7	10	20	7	0.5 equiv [Fe(CF ₃ SO ₃) ₂ (CH ₃ CN) ₂] added after 2h
8	20	20	7	0.5 equiv $[Fe(CF_3SO_3)_2(CH_3CN)_2]$ added after 2h
				Addition of HBF using syringe pump
9	20	20	5	HBF added by syringe pump during 2h
10	15	20	6	HBF added by syringe pump during 4h

7. References

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