

Electronic Supplementary Information (ESI)

Helical nanofibers of *N*-(perfluorooctanoyl)cysteine ethyl ester in its coordination polymers of Ag⁺

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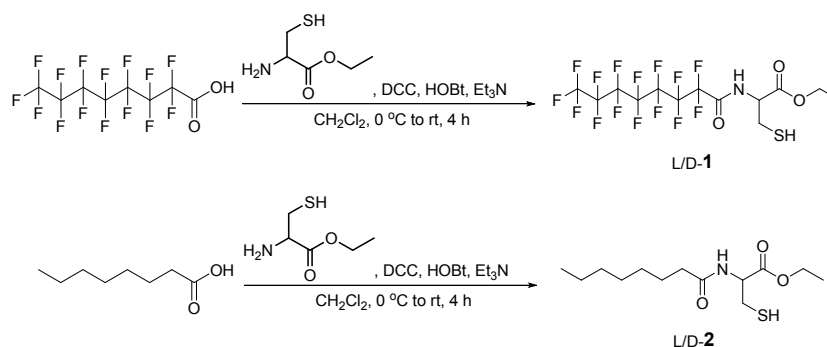
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S1. Experimental

S1.1 General

All chemicals and solvents purchased from Energy Chemical, except otherwise described, were used without further purification. ^1H NMR and ^{13}C NMR spectra were recorded using AVANCE III HD 500MHz in CDCl_3 with TMS. ^{19}F NMR spectra were recorded using Ascend 600 MHz in CDCl_3 with TMS. Absorption spectra were recorded on a Thermo Evolution 300 spectrophotometer. CD spectrometry was conducted using JASCO J-810 Circular Dichroism Chiroptical Spectrometer. Steady-state fluorescence measurements were conducted using a HITACHI F-4500 spectrofluorometer. HRMS (high-resolution MS) was recorded on Bruker Impact II.

S1.2 Synthesis



Scheme S1 Syntheses of L-1/D-1 and L-2/D-2.

L-1/D-1: L/D-cysteine ethyl ester (2 mmol), 1-hydroxybenzotriazole (2 mmol) and triethylamine (2 mmol) in CH_2Cl_2 (10 mL) were added pentadecafluorooctanoic acid (2 mmol). The mixture was stirred at 0 °C for 20 min, then dicyclohexylcarbodiimide (2 mmol) was added and stirred at room temperature for 4 h. After filtration and solvent evaporation, the crude product was purified by silica gel column chromatography, and white powder of L-1/D-1 was obtained (0.46 g, 42% yield).

L-1/D-2: L/D-cysteine ethyl ester (2 mmol), 1-hydroxybenzotriazole (2 mmol) and triethylamine (2 mmol) in CH_2Cl_2 (10 mL) were added octanoic acid (2 mmol). The mixture was stirred at 0 °C for 20 min, then dicyclohexylcarbodiimide (2 mmol) was added and stirred at room temperature for 4 h. After filtration and solvent evaporation, the crude product was purified by silica gel column chromatography, and light yellow powder of L-2/D-2 was obtained (0.21g, 60% yield).

L-1: ^1H NMR (500 MHz, CDCl_3) δ /ppm 7.28 (d, $J = 6.3$ Hz, 1H), 4.89–4.86 (m, 1H), 4.33–4.29 (m, 2H), 3.12–3.07 (m, 2H), 1.35 (t, $J = 9.0$ Hz 1H), 1.33 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ /ppm 168.43, 157.25, 117.12, 110.82, 110.65, 110.46, 110.19, 108.87, 108.37, 62.82, 54.09, 26.00, 14.12; **HRMS** (ESI^+) $[\text{M}+\text{Na}]^+$: calcd for $\text{C}_{13}\text{H}_{10}\text{F}_{15}\text{NO}_3\text{SNa}$ 568.0040, found 568.0038.

D-1: ^1H NMR (500 MHz, CDCl_3) δ /ppm 7.29 (d, $J = 5.6$ Hz, 1H), 4.89–4.86 (m, 1H), 4.36–4.26 (m, 2H), 3.15–3.05 (m, 2H), 1.34 (t, $J = 9.0$ Hz, 1H), 1.33 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ /ppm 168.42, 157.27, 117.12, 110.82, 110.65, 110.47, 110.20, 108.87, 108.38, 62.80, 54.08, 25.99, 14.11; **HRMS** (ESI^+) $[\text{M}+\text{Na}]^+$: calcd for $\text{C}_{13}\text{H}_{10}\text{F}_{15}\text{NO}_3\text{SNa}$ 568.0040, found 568.0043.

L-2: ^1H NMR (500 MHz, CDCl_3) : δ /ppm 6.34 (d, $J = 6.5$ Hz, 1H), 4.88–4.85 (m, 1H), 4.30–4.20 (m, 2H), 3.03–3.00 (m, 2H), 2.25 (t, $J = 7.7$ Hz, 2H), 1.67–1.61 (m, 2H), 1.32–1.27 (m, 12H), 0.87 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ /ppm 172.91, 170.20, 61.98, 53.38, 36.56, 31.65, 29.18, 28.96, 26.93, 25.59, 22.58, 14.19, 14.02; **HRMS** (ESI^+) $[\text{M}+\text{Na}]^+$: calcd for $\text{C}_{13}\text{H}_{25}\text{NO}_3\text{SNa}$ 298.1453, found 298.1448.

D-2: ^1H NMR (500 MHz, CDCl_3) δ /ppm 6.34 (d, $J = 6.6$ Hz, 1H), 4.88–4.85 (m, 1H), 4.28–4.21 (m, 2H), 3.03–3.00 (m, 2H), 2.26 (t, $J = 7.5$ Hz, 2H), 1.68–1.62 (m, 2H), 1.32–1.27 (m, 12H), 0.87 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ /ppm 172.89, 170.20, 61.98, 53.37, 36.57, 31.66, 29.18, 28.96, 26.93, 25.59, 22.58, 14.19, 14.02; **HRMS** (ESI^+) $[\text{M}+\text{Na}]^+$: calcd for $\text{C}_{13}\text{H}_{25}\text{NO}_3\text{SNa}$ 298.1453, found 298.1448.

S1.3 References

1. T. Terauchi, S. Machida and S. Komba, *Tetrahedron Lett.*, 2010, **51**, 1497.
2. M. Brindisi, S. Butini, S. Franceschini, S. Brogi, F. Trotta, S. Ros, A. Cagnotto, M. Salmona, A. Casagni, M. Andreassi, S. Saponara, B. Gorelli, P. Weikop, J. D. Mikkelsen, J. Scheel-Kruger, K. Sandager-Nielsen, E. Novellino, G. Campiani and S. Gemma, *J. Med. Chem.*, 2014, **57**, 9578.

S2. Additional data

S2.1 Experimental data

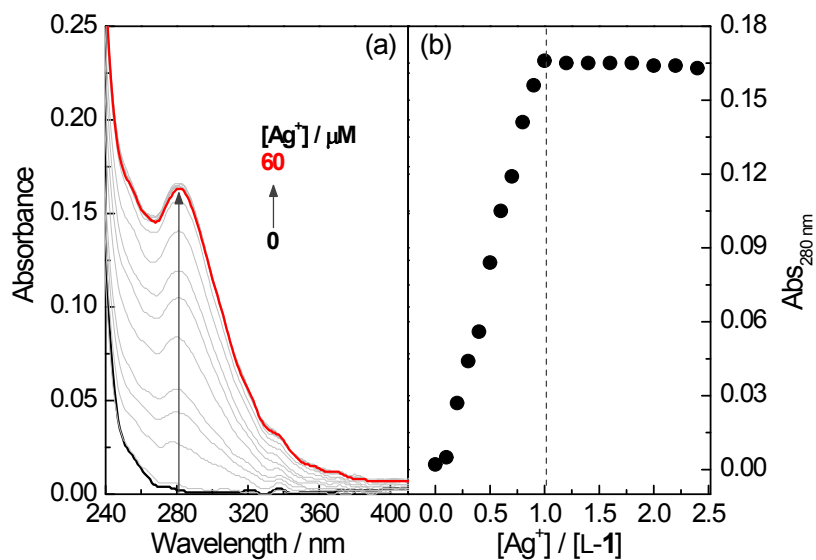


Fig. S1 (a) Absorption spectra of L-1 in EtOH in the presence of Ag⁺ and (b) plots of absorbance at 280 nm against concentration of Ag⁺. [L-1] = 25 μ M.

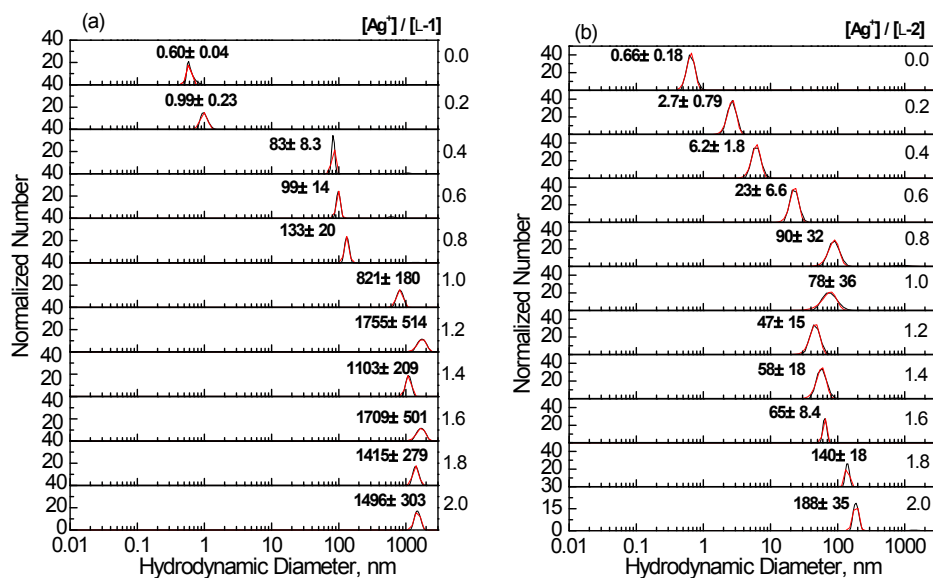


Fig. S2 Hydrodynamic diameters of L-1 (a) and L-2 (b) in EtOH in the presence of Ag⁺ of increasing concentration. [L-1] = [L-2] = 25 μ M.

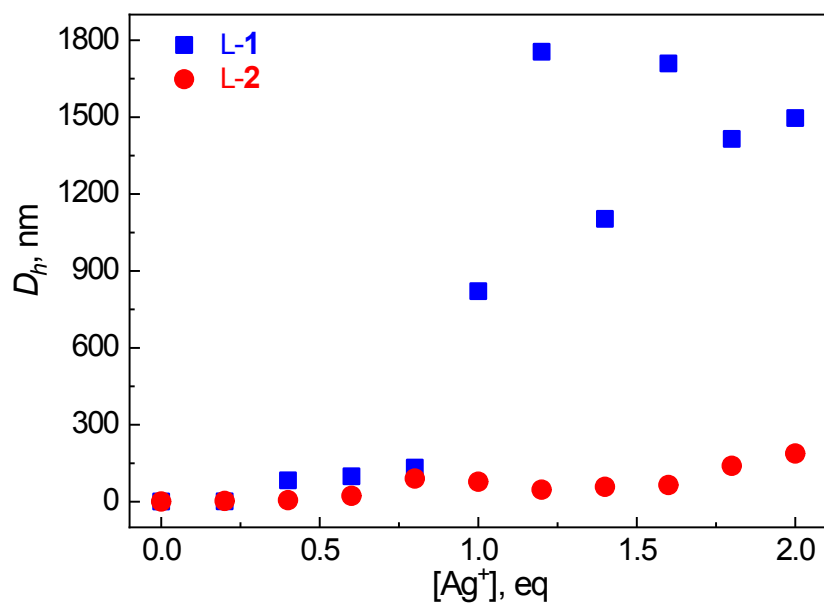


Fig. S3 Hydrodynamic diameters of L-1 (a) and L-2 (b) in EtOH in the presence of Ag⁺ of increasing concentration. [L-1] = [L-2] = 25 μ M.

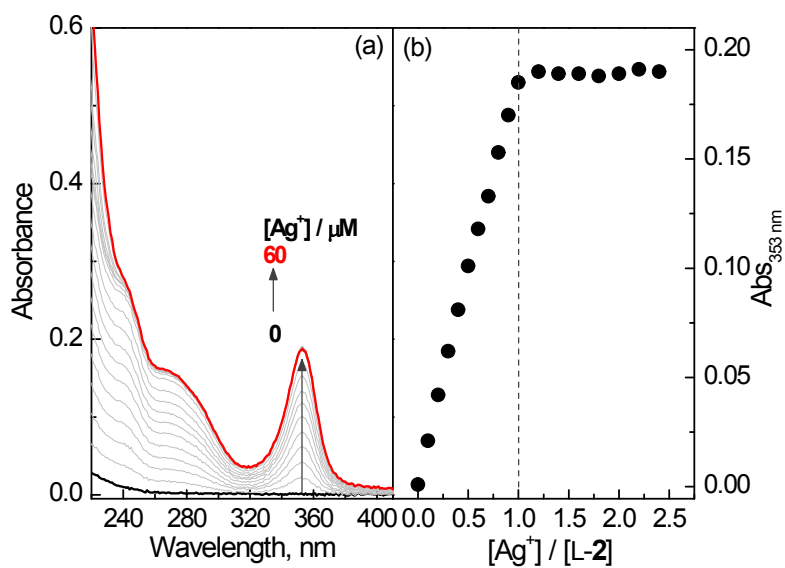


Fig. S4 (a) Absorption spectra of L-2 in EtOH in the presence of Ag⁺ and (b) plots of absorbance at 353 nm against concentration of Ag⁺. [L-2] = 25 μ M.

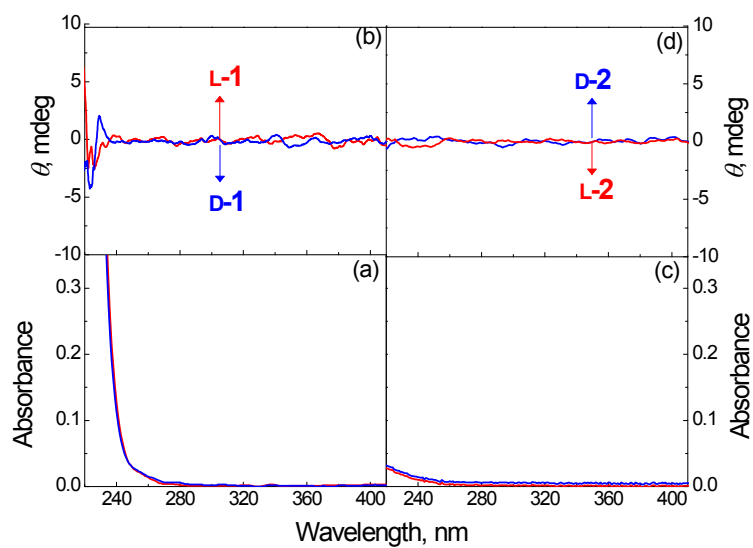


Fig. S5 Absorption and CD spectra of L-1/D-1 (a, b) and L-2/D-2 (c, d) in EtOH. $[L-1] = [D-1] = [L-2] = [D-2] = 25 \mu\text{M}$.

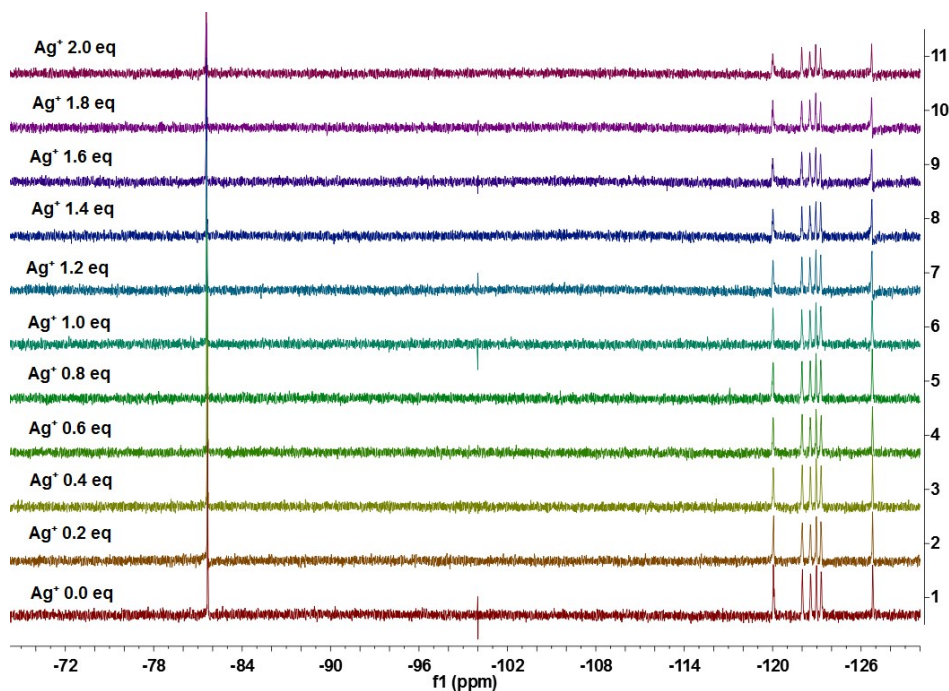


Fig. S6 ^{19}F NMR titrations of L-1 by Ag^+ in EtOD. $[L-1] = 50 \mu\text{M}$.

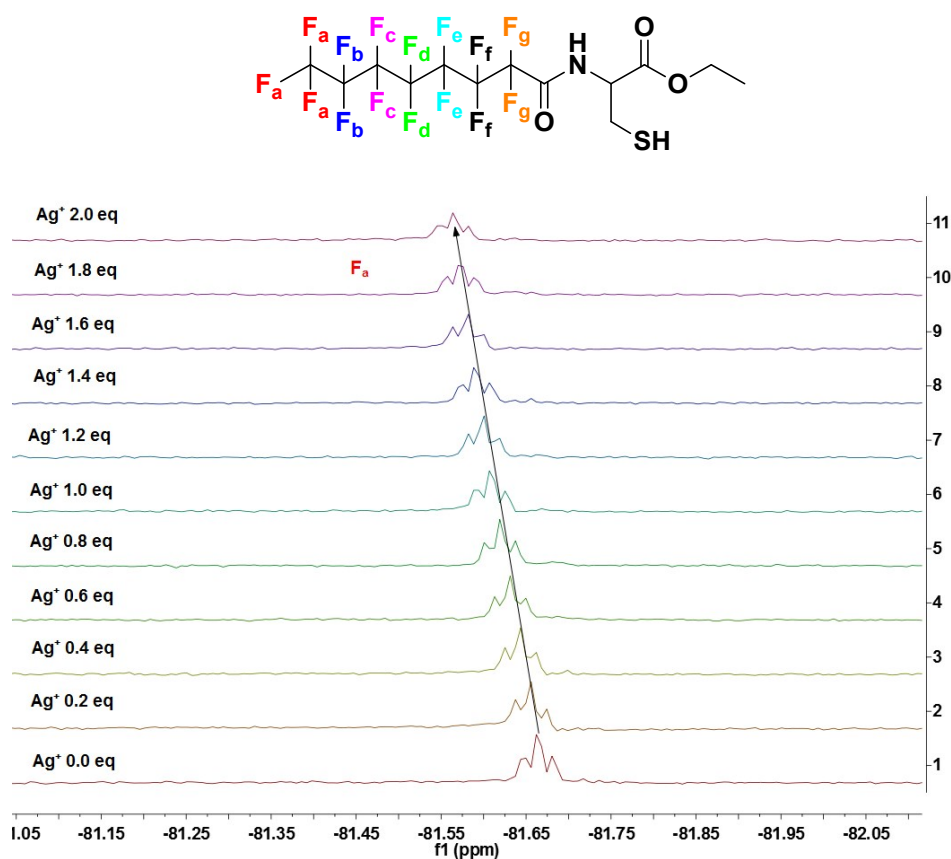


Fig. S7 Partial ¹⁹F NMR spectra of F_a of L-1 in the presence of Ag⁺ in EtOD. [L-1] = 50 μM.

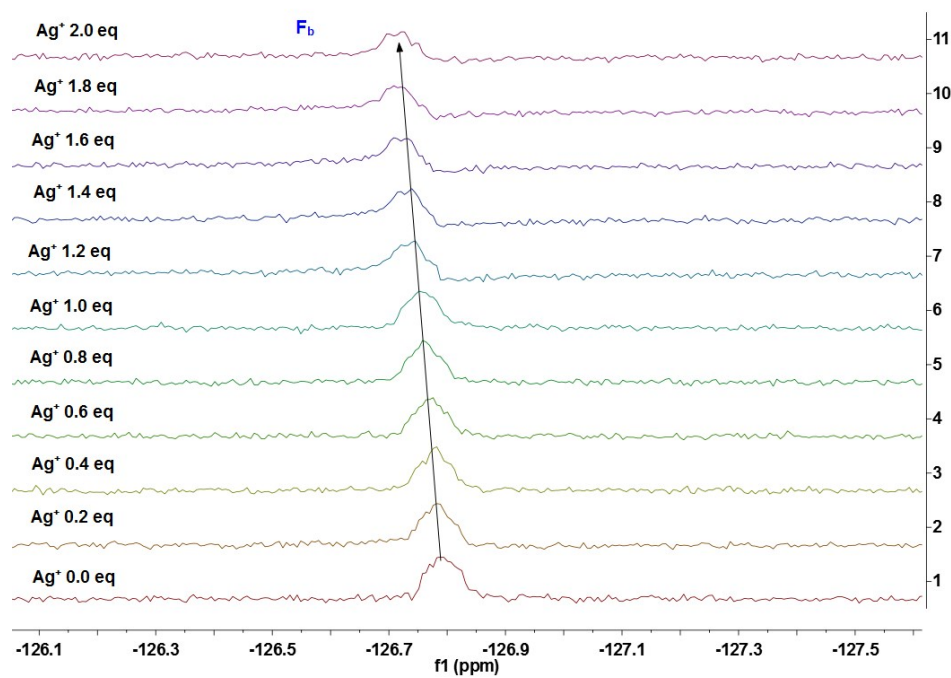


Fig. S8 Partial ¹⁹F NMR spectra of F_b of L-1 in the presence of Ag⁺ in EtOD. [L-1] = 50 μM.

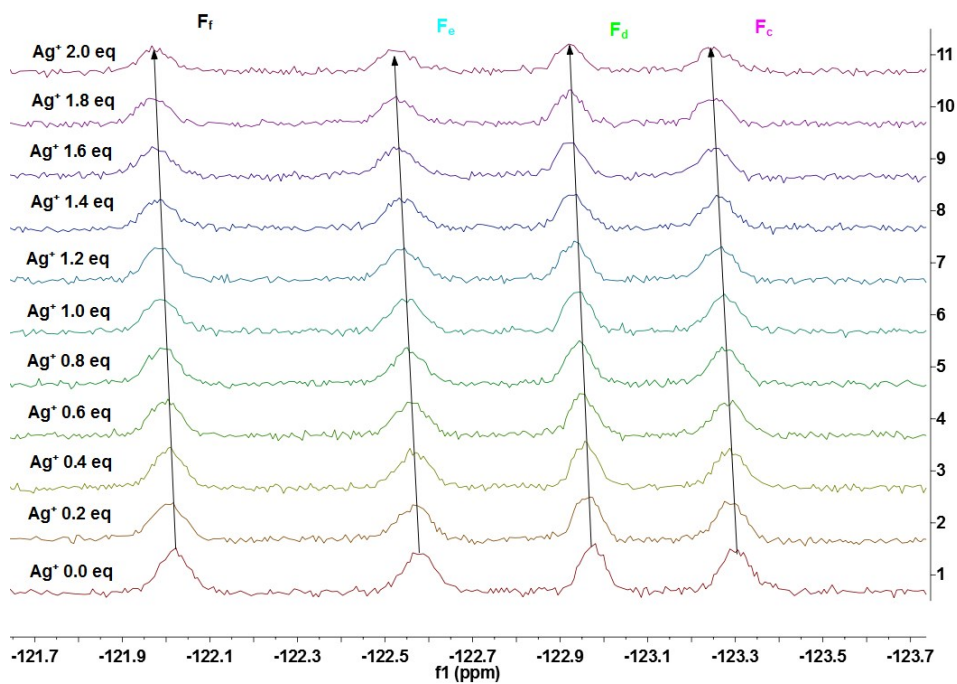


Fig. S9 Partial ^{19}F NMR spectra of F_c , F_d , F_e and F_f of L-1 in the presence of Ag^+ in EtOD. $[\text{L-1}] = 50 \mu\text{M}$.

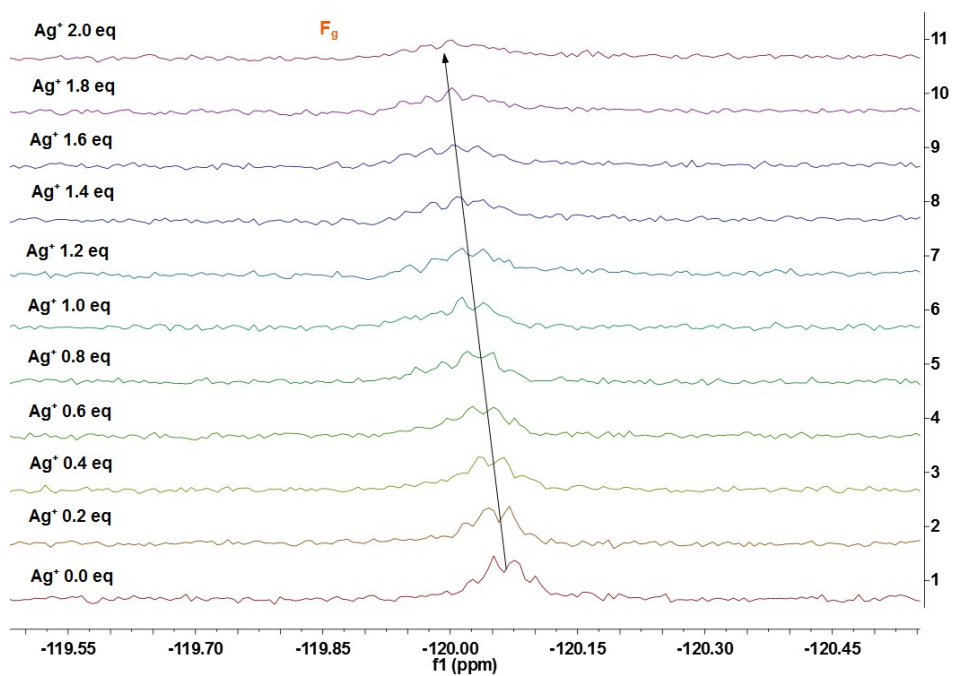


Fig. S10 Partial ^{19}F NMR spectra of F_g of L-1 in the presence of Ag^+ in EtOD. $[\text{L-1}] = 50 \mu\text{M}$.

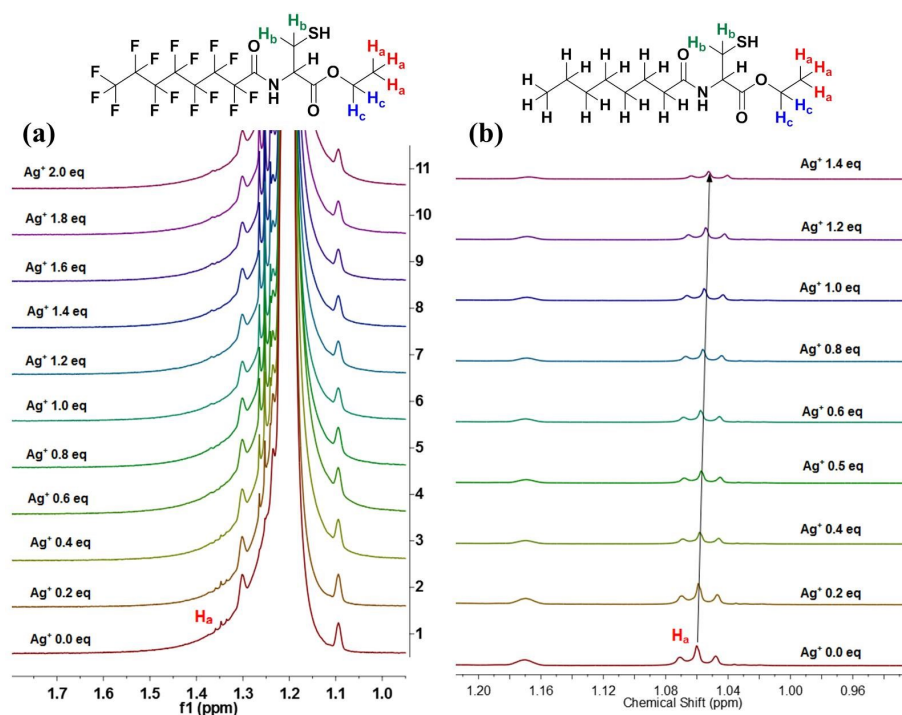


Fig. S11 Partial ^1H NMR spectra of H_a of L-1 (a) and L-2 (b) in the presence of Ag^+ in EtOD. $[\text{L-1}] = [\text{L-2}] = 50 \mu\text{M}$.

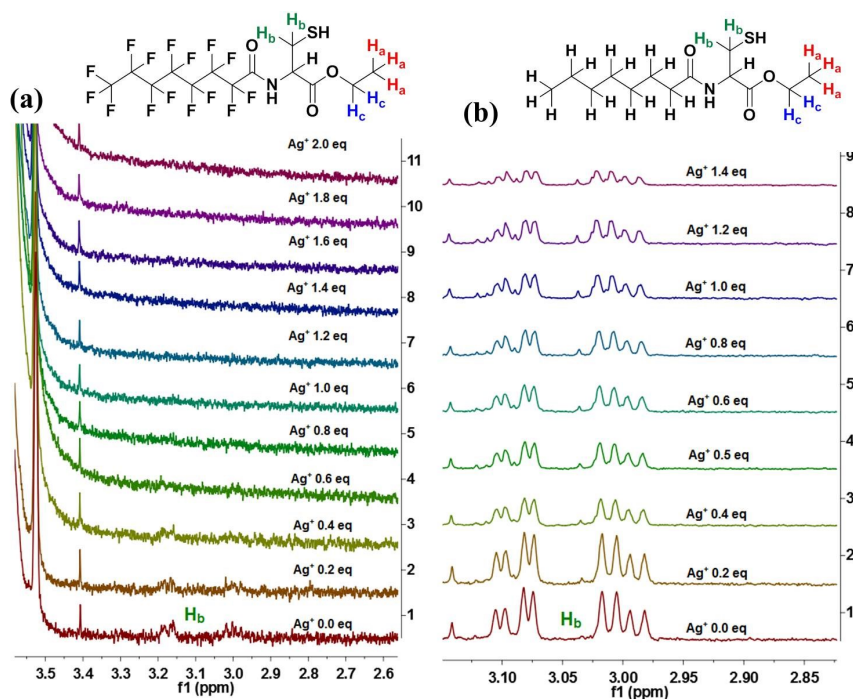


Fig. S12 Partial ^1H NMR spectra of H_b of L-1 (a) and L-2 (b) in the presence of Ag^+ in EtOD. $[\text{L-1}] = [\text{L-2}] = 50 \mu\text{M}$.

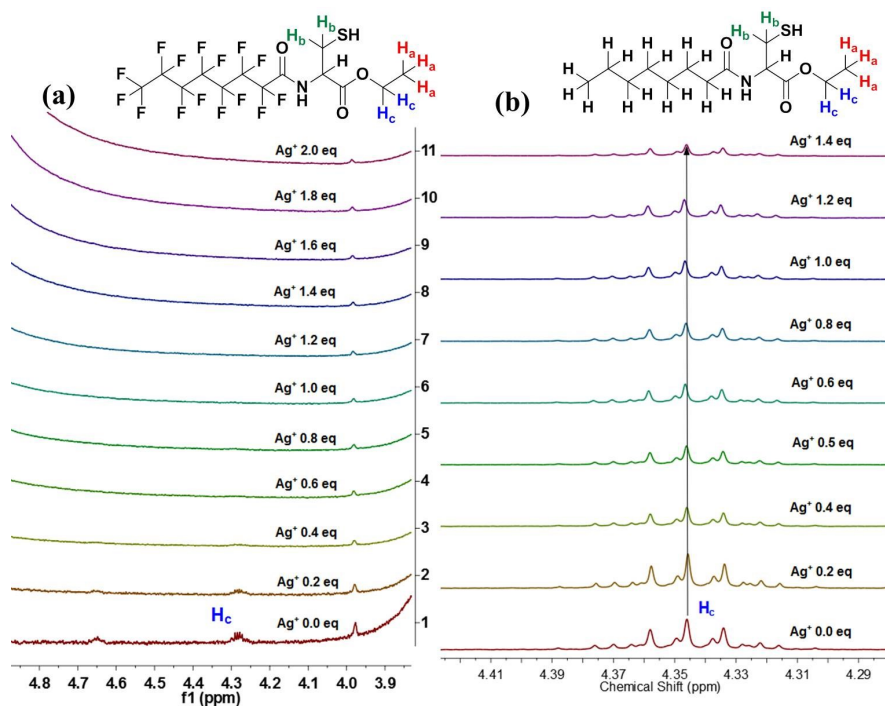


Fig. S13 Partial ^1H NMR spectra of H_c of L-1 (a) and L-2 (b) in the presence of Ag^+ in EtOD. $[\text{L-1}] = [\text{L-2}] = 50 \mu\text{M}$.

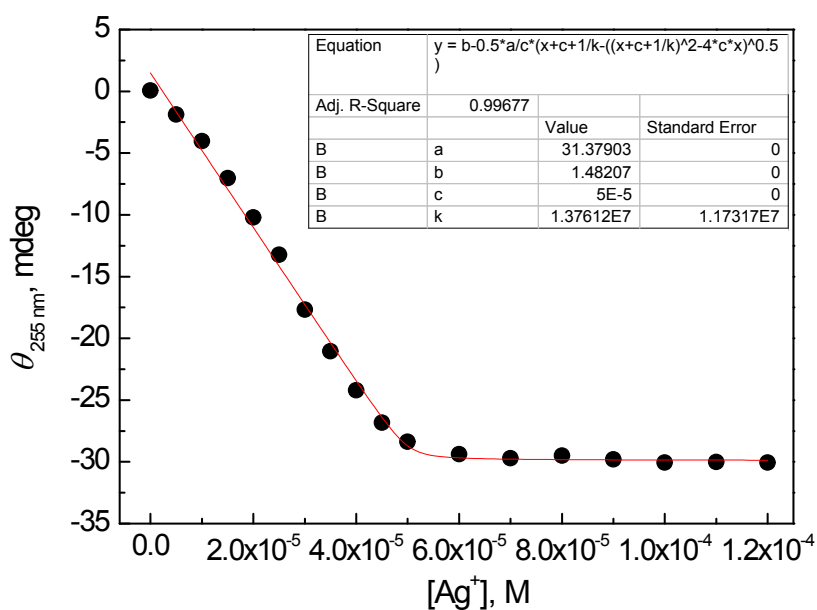


Fig. S14 Data fitting of CD signal at 255 nm of Ag^+ -L-1 in EtOH as a function of concentration of Ag^+ assuming a 1:1 stoichiometry. $[\text{L-1}] = 50 \mu\text{M}$, $[\text{Ag}^+] = 0 - 120 \mu\text{M}$.

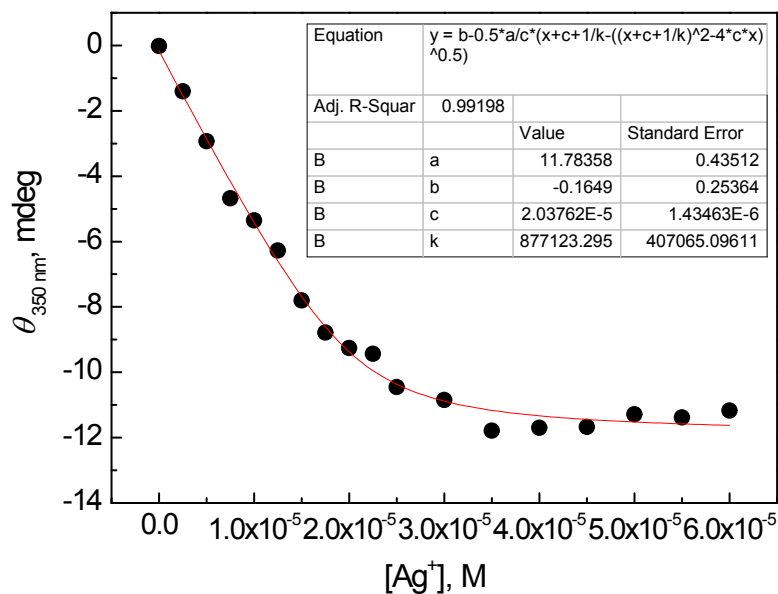


Fig. S15 Data fitting of CD signal at 350 nm of Ag^+ -L-2 in EtOH as a function of concentration of Ag^+ assuming a 1:1 stoichiometry. $[\text{L-2}] = 25 \mu\text{M}$, $[\text{Ag}^+] = 0 - 60 \mu\text{M}$.

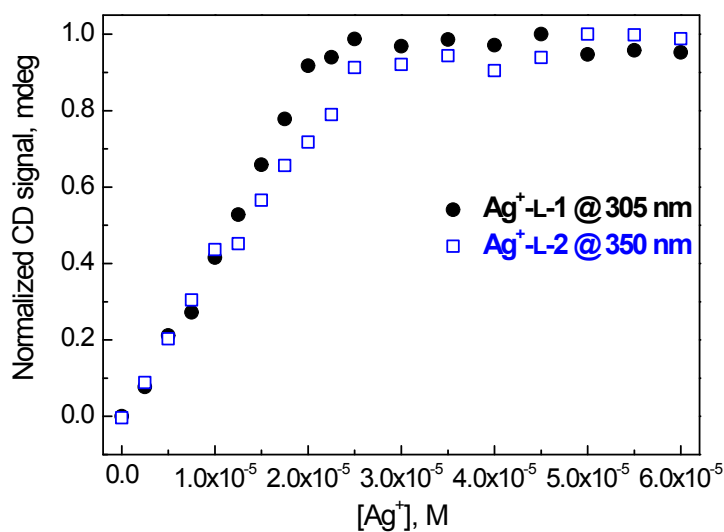


Fig. S16 Normalized CD signals at 305 nm of Ag^+ -L-1 and 350 nm of Ag^+ -L-2 in EtOH with increasing concentration of Ag^+ . $[\text{L-1}] = [\text{L-2}] = 25 \mu\text{M}$, $[\text{Ag}^+] = 0 - 60 \mu\text{M}$.

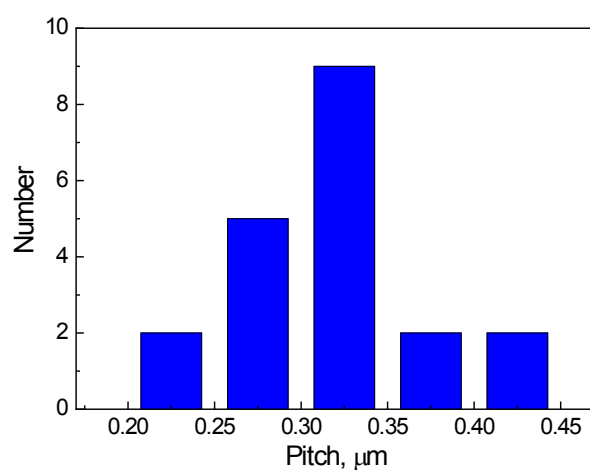


Fig. S17 Histogram of the helical pitch of structure observed in the SEM images from the Ag^+ -L-1 complex in EtOH. $[\text{L-1}] = [\text{Ag}^+] = 25 \mu\text{M}$.

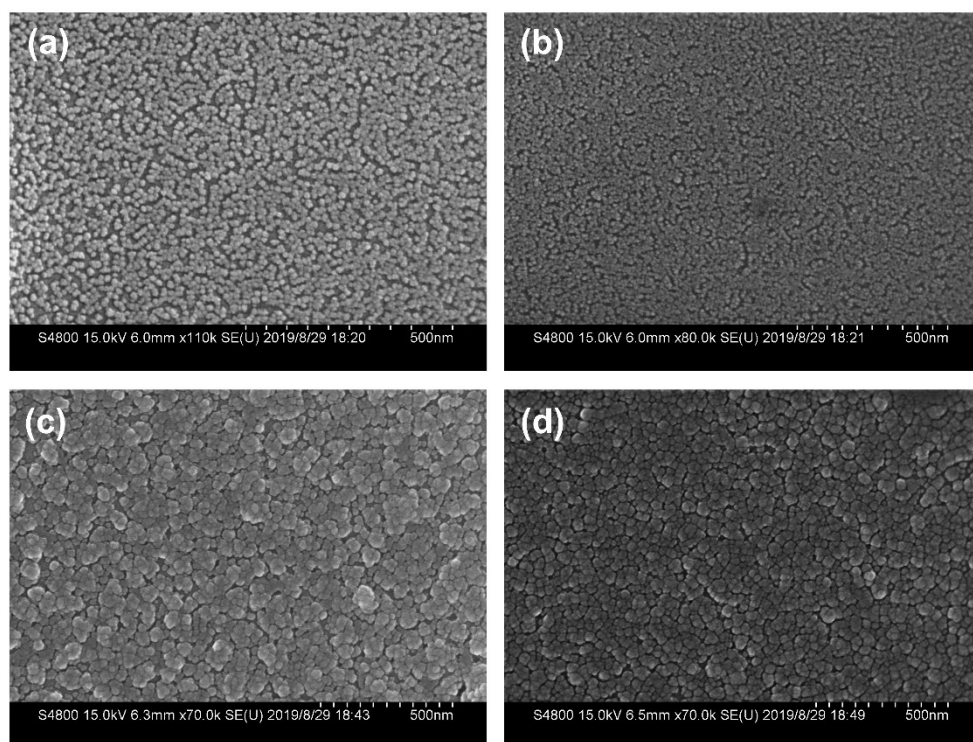


Fig. S18 SEM images of the structures of L-1 (a), D-1 (b) and L-2 (c), D-2 (d) formed in EtOH. $[\text{L-1}] = [\text{D-1}] = [\text{L-2}] = [\text{D-2}] = 25 \mu\text{M}$.

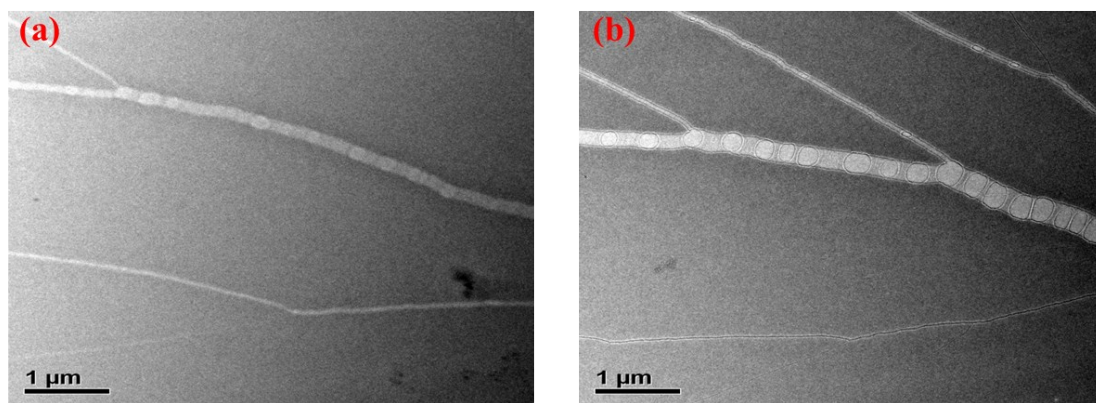


Fig. S19 TEM images of supramolecular structures formed in EtOH solutions of Ag^+ -L-1 (a) and Ag^+ -D-1 (b).

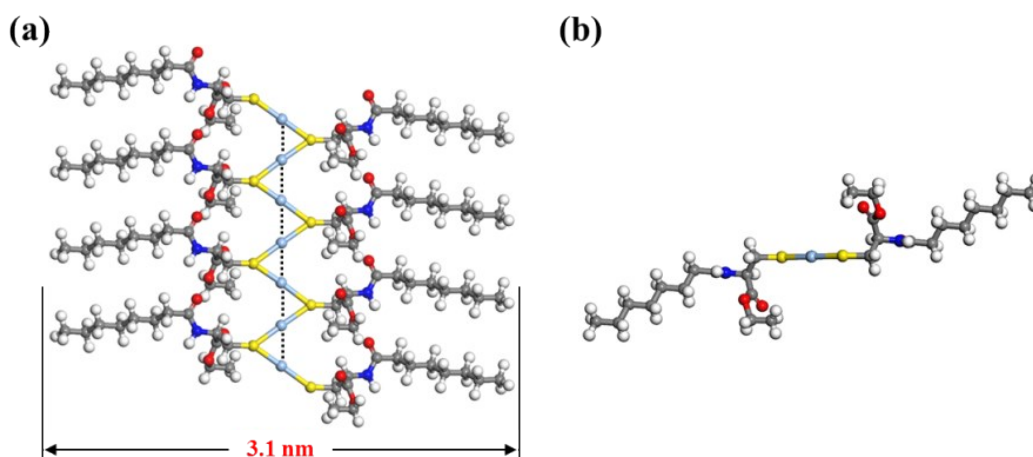


Fig. S20 Structural models of Ag^+ -2 polymeric chain from the front view (a) and the side view (b) using Materials Studio modeling. Dashed black lines highlight the $\text{Ag}^+\cdots\text{Ag}^+$ interactions.

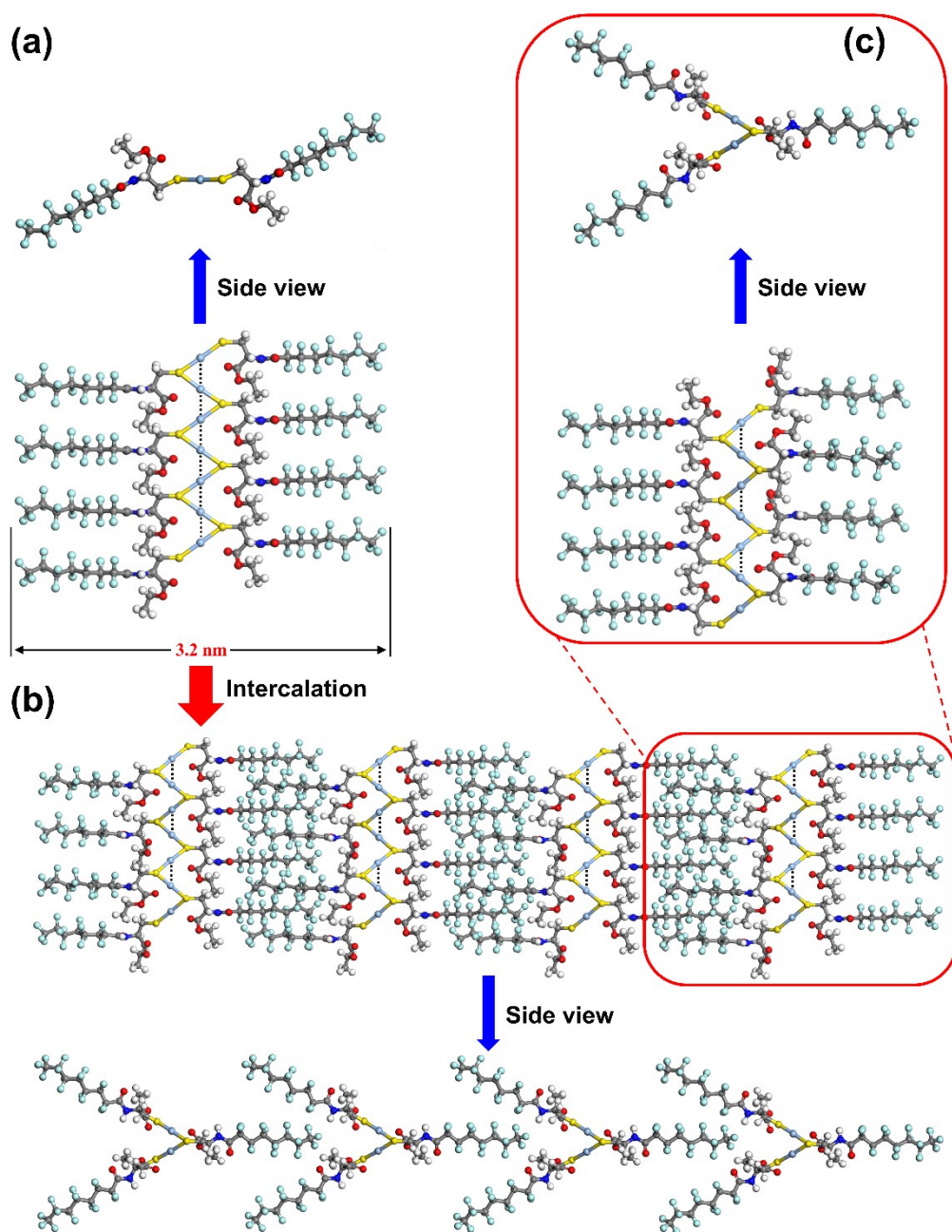


Fig. S21 (a) Structural models of single Ag^+ -1 polymeric chain. (b) Structural models of the inter-chain intercalation between Ag^+ -1 polymeric chains. (c) Single Ag^+ -1 polymeric chain extracted from the intercalation. Dashed black lines highlight the $\text{Ag}^+\cdots\text{Ag}^+$ interactions. The structural models are shown by using Materials Studio modeling. The extended $\text{Ag}^+\cdots\text{Ag}^+$ interactions in single polymeric chain (a) turn to the binuclear $\text{Ag}^+\cdots\text{Ag}^+$ interactions due to inter-chain intercalation (c).

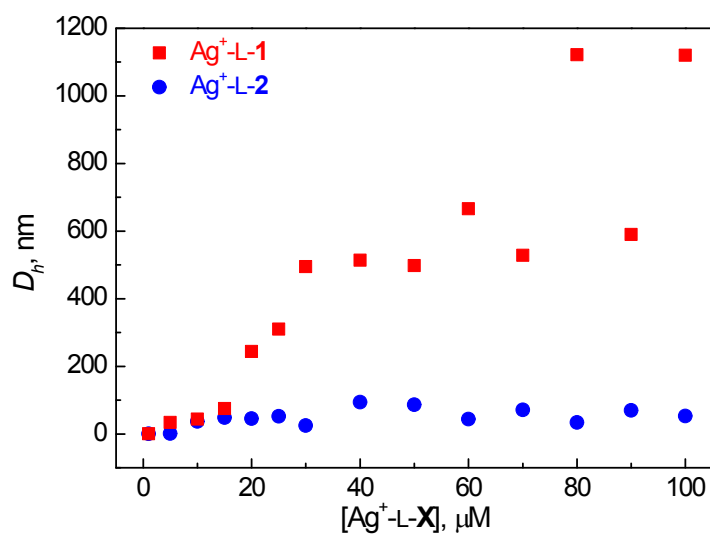


Fig. S22 Hydrodynamic diameters of Ag⁺-L-1 and Ag⁺-L-2 in EtOH in the presence of Ag⁺ of increasing concentration. [L-1] = [L-2] = [Ag⁺].

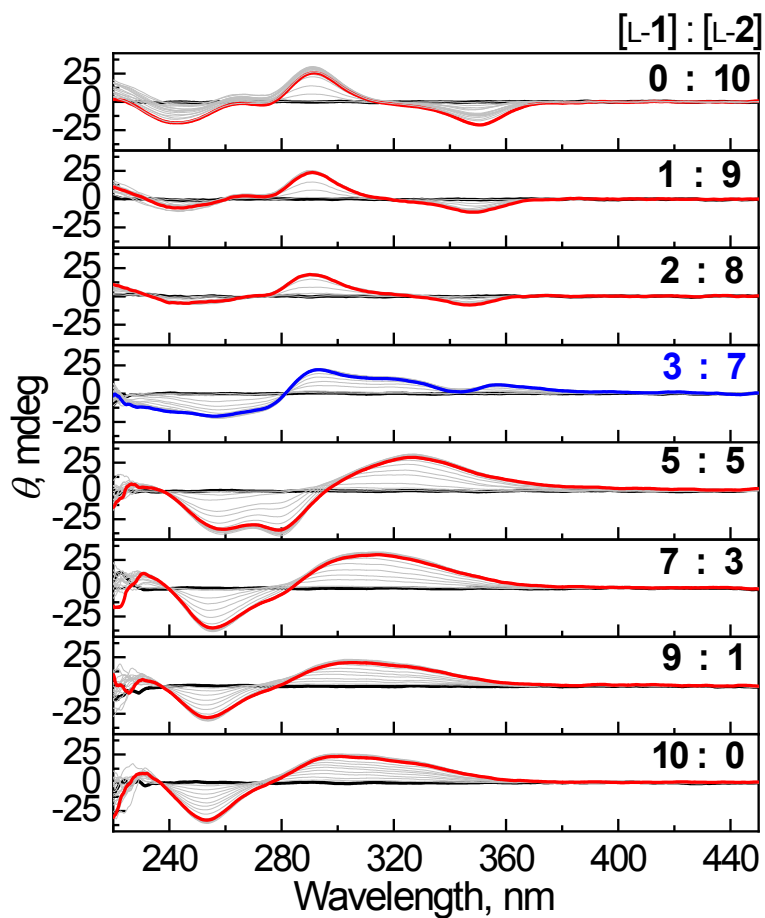


Fig. S23 CD spectra of L-1 and L-2 of varying molar ratio in EtOH in the presence of Ag⁺ of increasing concentration. [L-1] + [L-2] = 25 μM, [Ag⁺] = 0 - 60 μM.

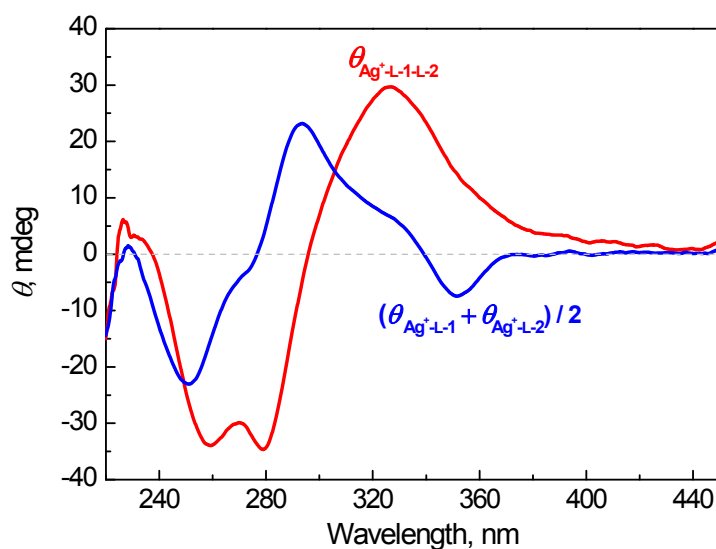


Fig. S24 Calculated (blue line) and experimental (red line) CD spectra of $\text{Ag}^+\text{-L-1-L-2}$ (molar ratio of L-1 to L-2 is 5:5) in EtOH. Calculated line is from individual CD spectra of $\text{Ag}^+\text{-L-1}$ and $\text{Ag}^+\text{-L-2}$ according to $\theta_{\text{calcd.}} = (\theta_{\text{Ag}^+\text{-L-1}} + \theta_{\text{Ag}^+\text{-L-2}})/2$. $[\text{L-1}] + [\text{L-2}] = [\text{Ag}^+] = 25 \mu\text{M}$.

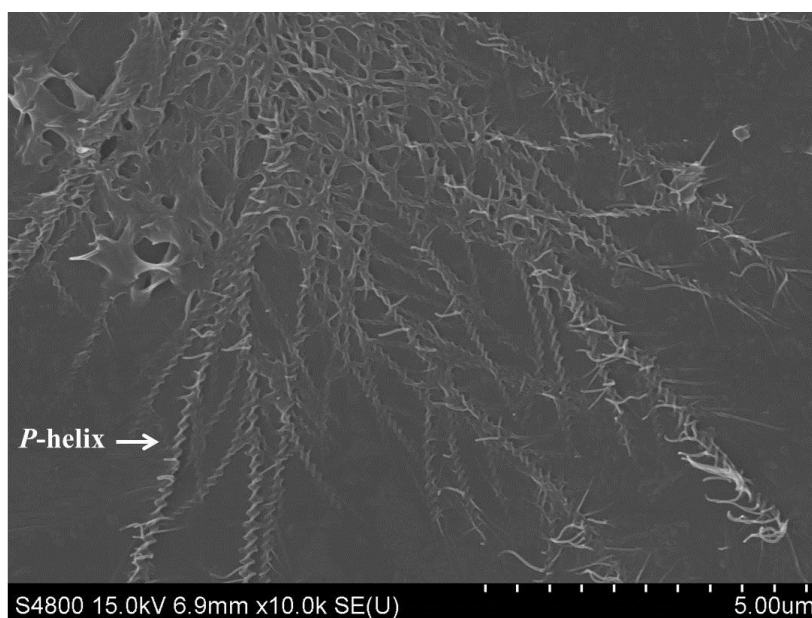


Fig. S25 SEM image of the mixed ligands of L-1 and L-2 of molar ratio 5:5 in EtOH in the presence of Ag^+ . $[\text{L-1}] + [\text{L-2}] = [\text{Ag}^+] = 25 \mu\text{M}$.

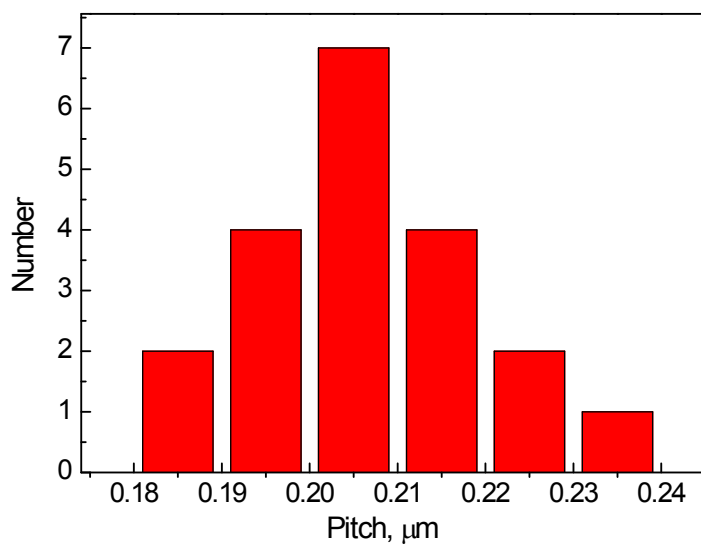


Fig. S26 Helical pitch of structures observed from Ag^+ -L-1-L-2 at L-1 to L-2 molar ratio of 5:5 in EtOH. $[\text{L-1}] + [\text{L-2}] = [\text{Ag}^+] = 25 \mu\text{M}$.

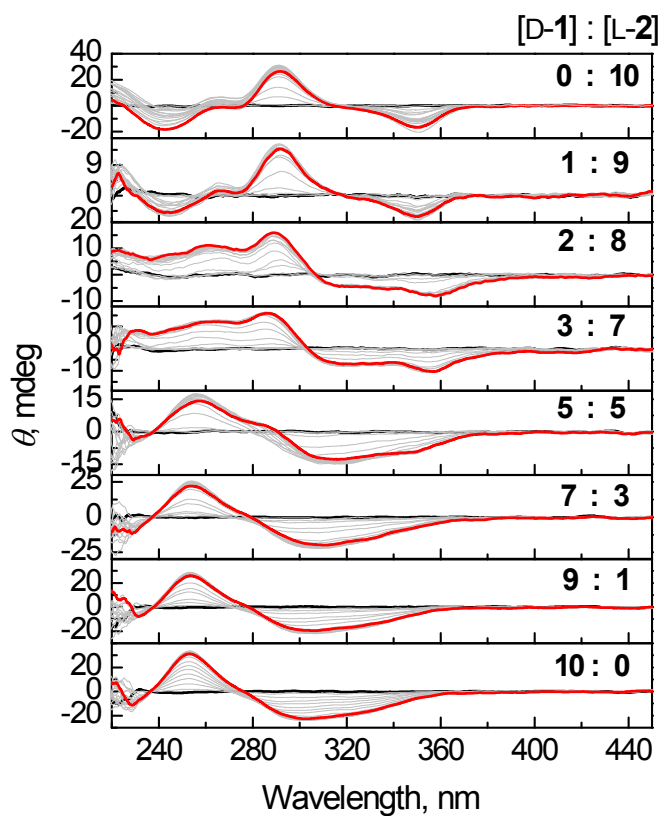


Fig. S27 CD spectra of the mixed ligands of D-1 and L-2 of different molar ratio in EtOH in the presence of Ag^+ . $[\text{D-1}] + [\text{L-2}] = 25 \mu\text{M}$, $[\text{Ag}^+] = 0 - 60 \mu\text{M}$.

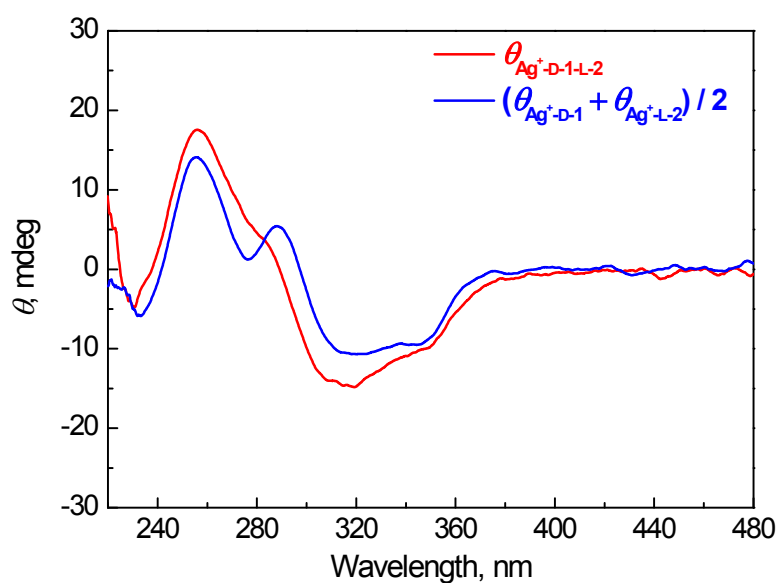


Fig. S28 Calculated (blue line) and experimental (red line) CD spectra of $\text{Ag}^+\text{-D-1-L-2}$ (molar ratio of **D-1** to **L-2** is 5:5) in EtOH. Calculated line is from the individual CD spectra of $\text{Ag}^+\text{-D-1}$ and $\text{Ag}^+\text{-L-2}$ according to $\theta_{\text{calcd.}} = (\theta_{\text{Ag}^+\text{-D-1}} + \theta_{\text{Ag}^+\text{-L-2}})/2$. $[\text{D-1}] + [\text{L-2}] = [\text{Ag}^+] = 25 \mu\text{M}$.

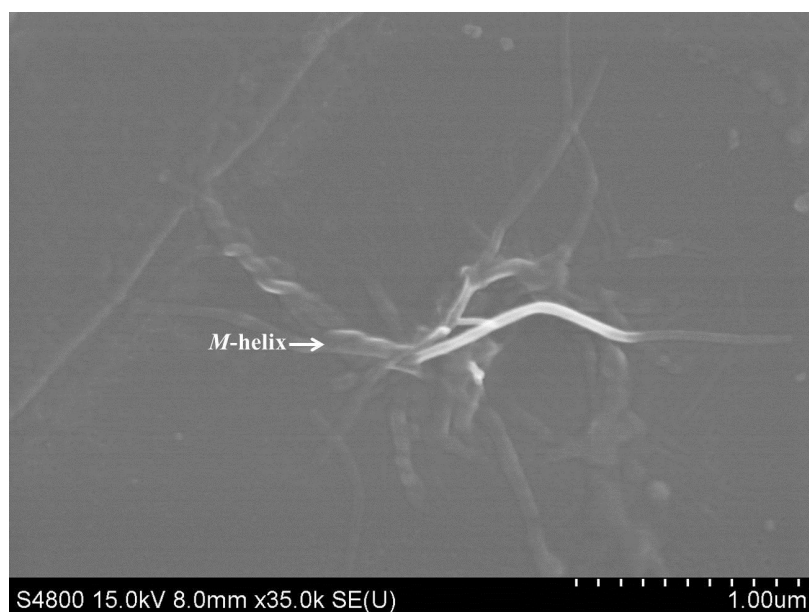


Fig. S29 SEM image of the mixed ligands of **D-1** and **L-2** at molar ratio of 5:5 in EtOH in the presence of Ag^+ . $[\text{D-1}] + [\text{L-2}] = [\text{Ag}^+] = 25 \mu\text{M}$.

S2.2 ^1H NMR, ^{13}C NMR, and ^{19}F NMR spectra of 1 and 2

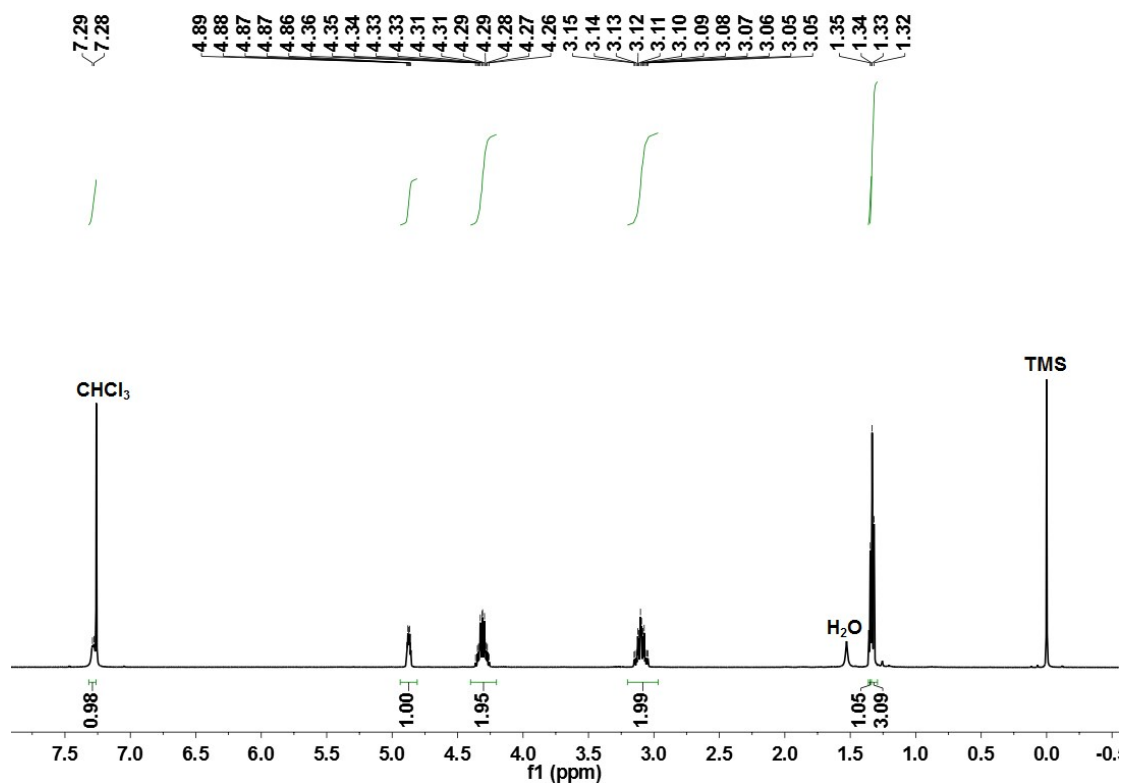


Fig. S30 ^1H NMR of L-1 (500 MHz, CDCl_3)

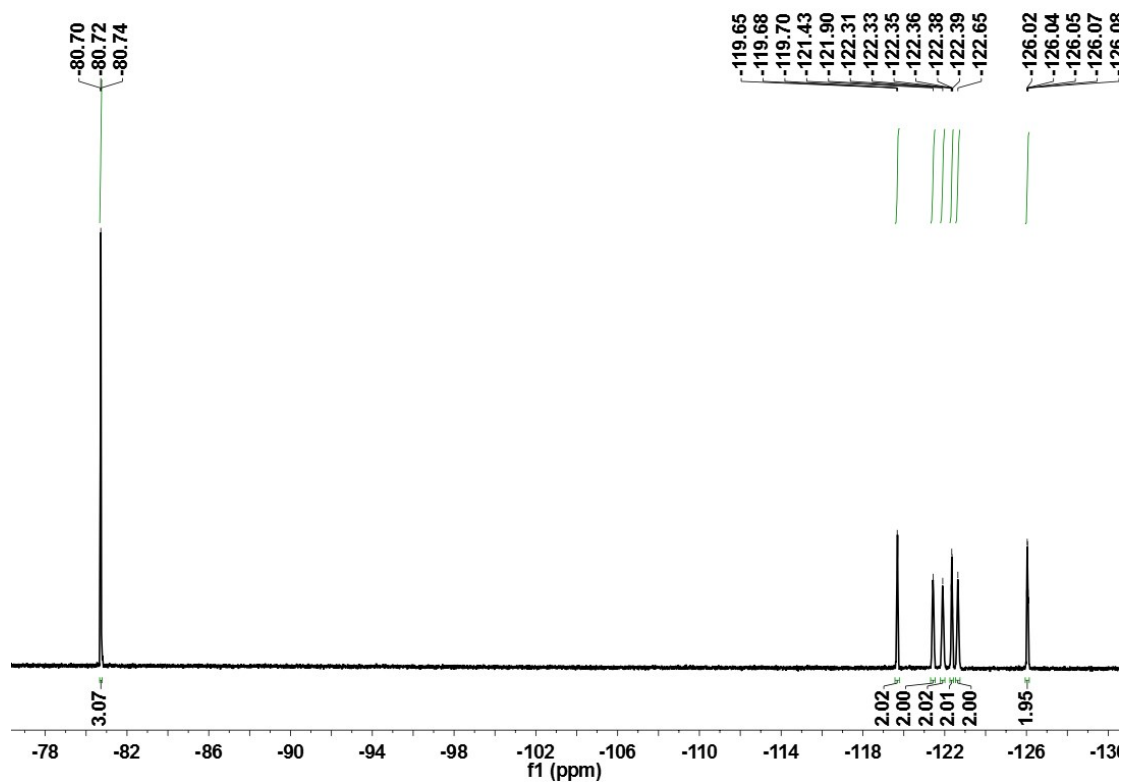


Fig. S31 ^{19}F NMR of L-1 (600 MHz, CDCl_3)

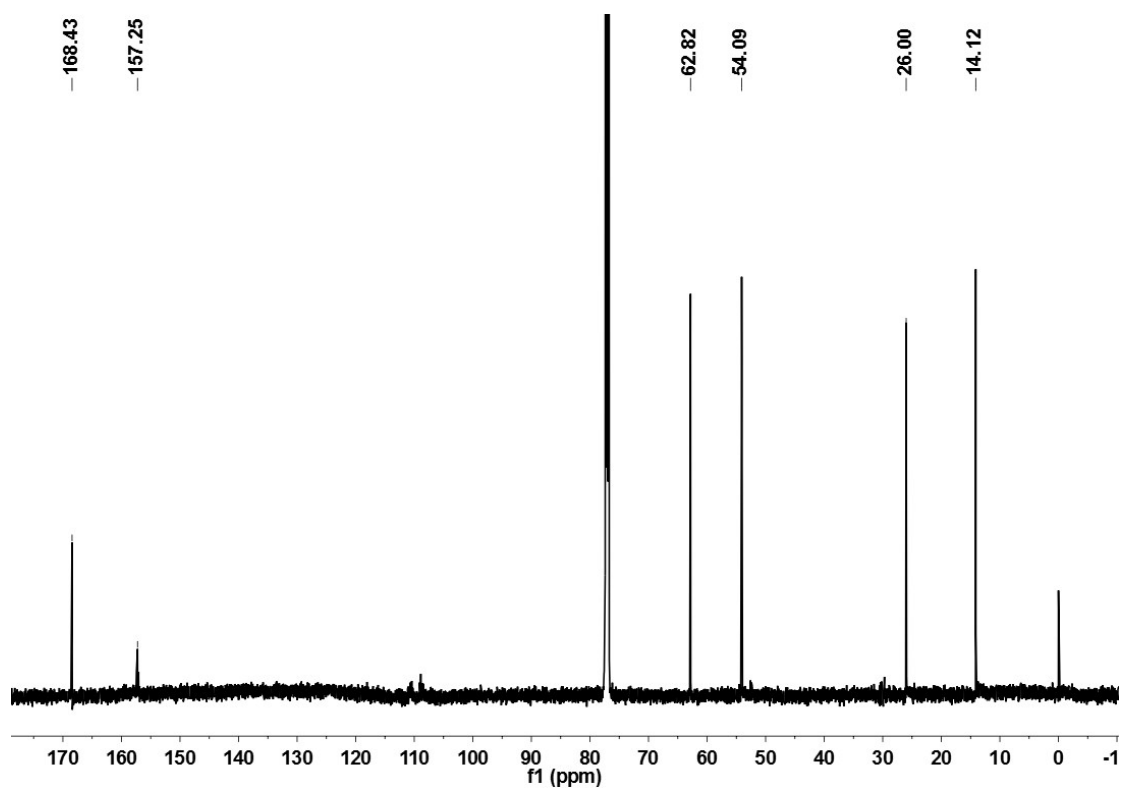


Fig. S32 ^{13}C NMR of L-1 (126 MHz, CDCl_3)

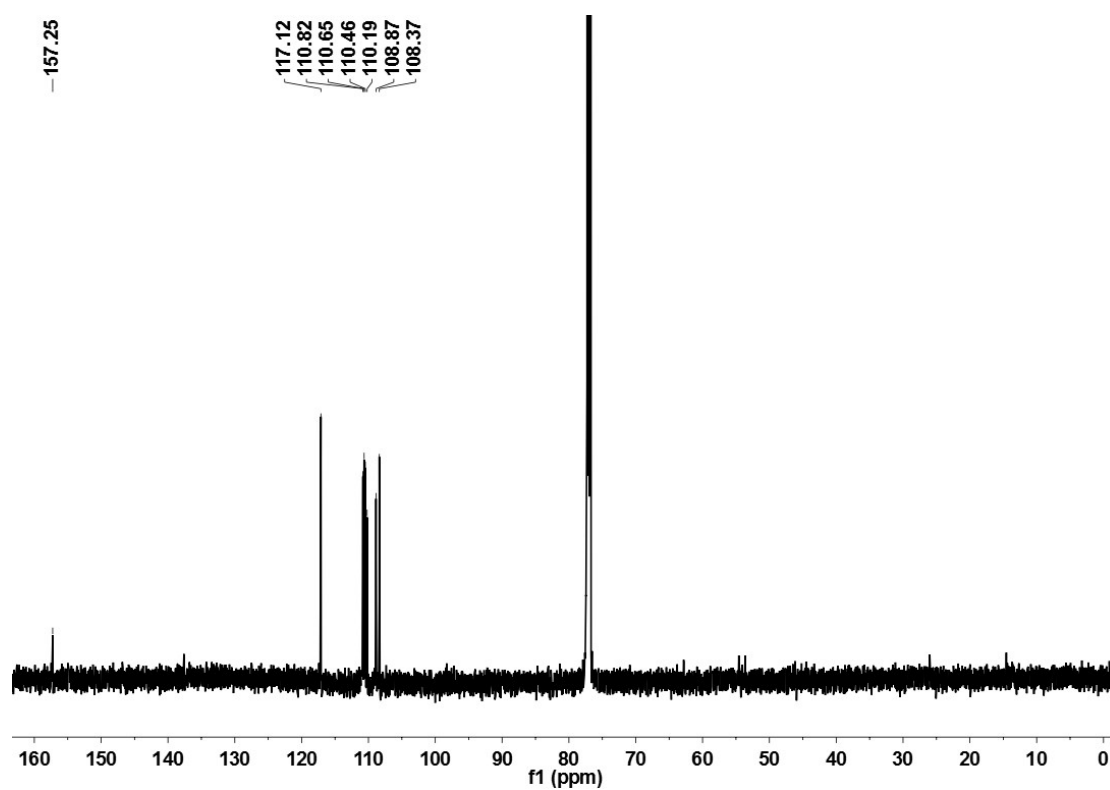


Fig. S33 ^{13}C NMR of L-1 (126 MHz, CDCl_3)

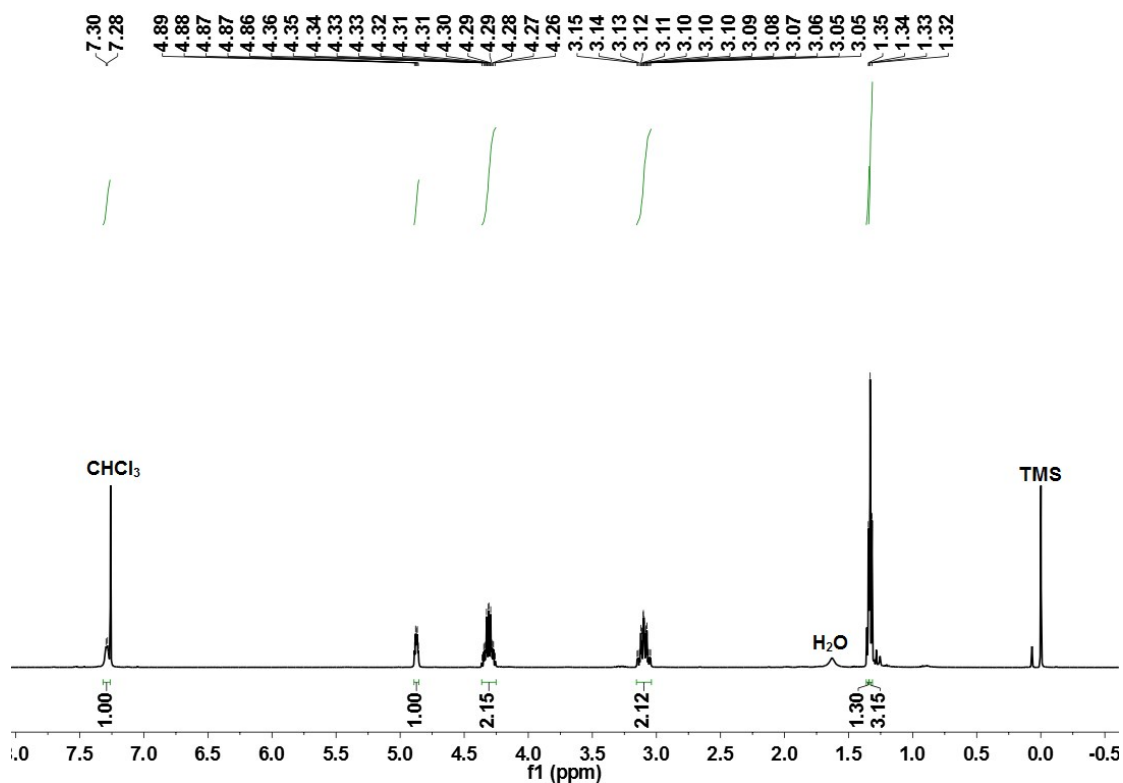


Fig. S34 ¹H NMR of D-1 (500 MHz, CDCl₃)

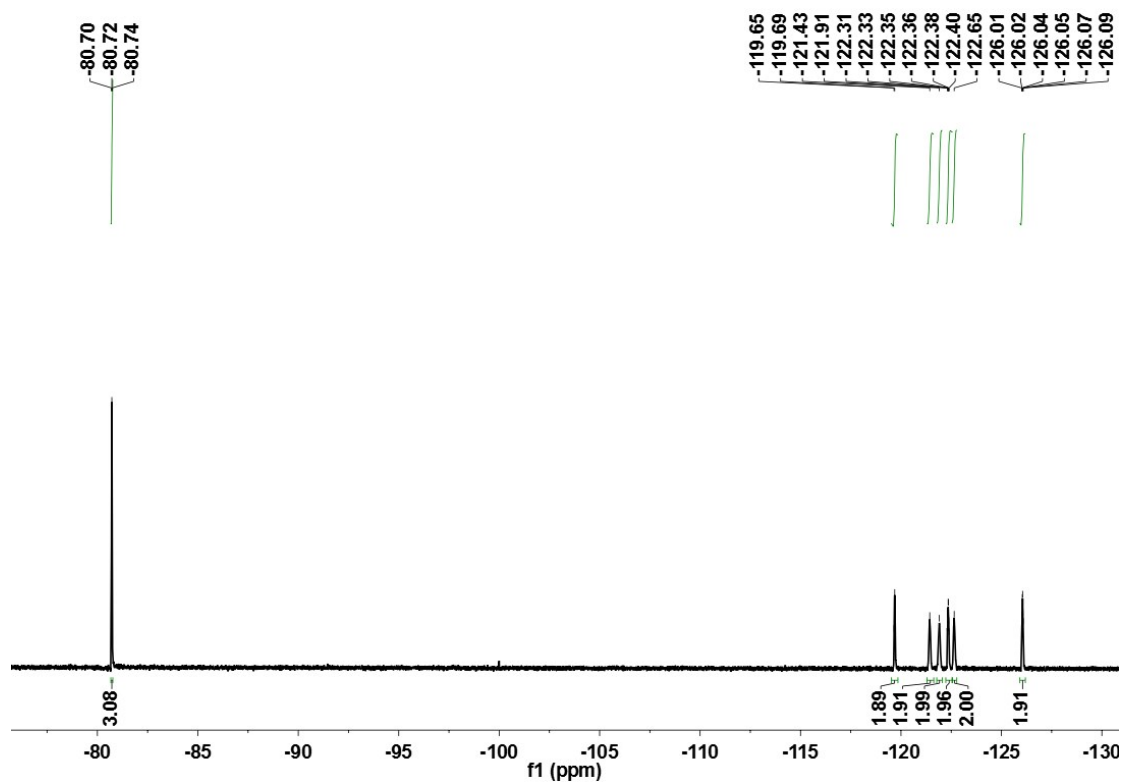


Fig. S35 ¹⁹F NMR of D-1 (600 MHz, CDCl₃)

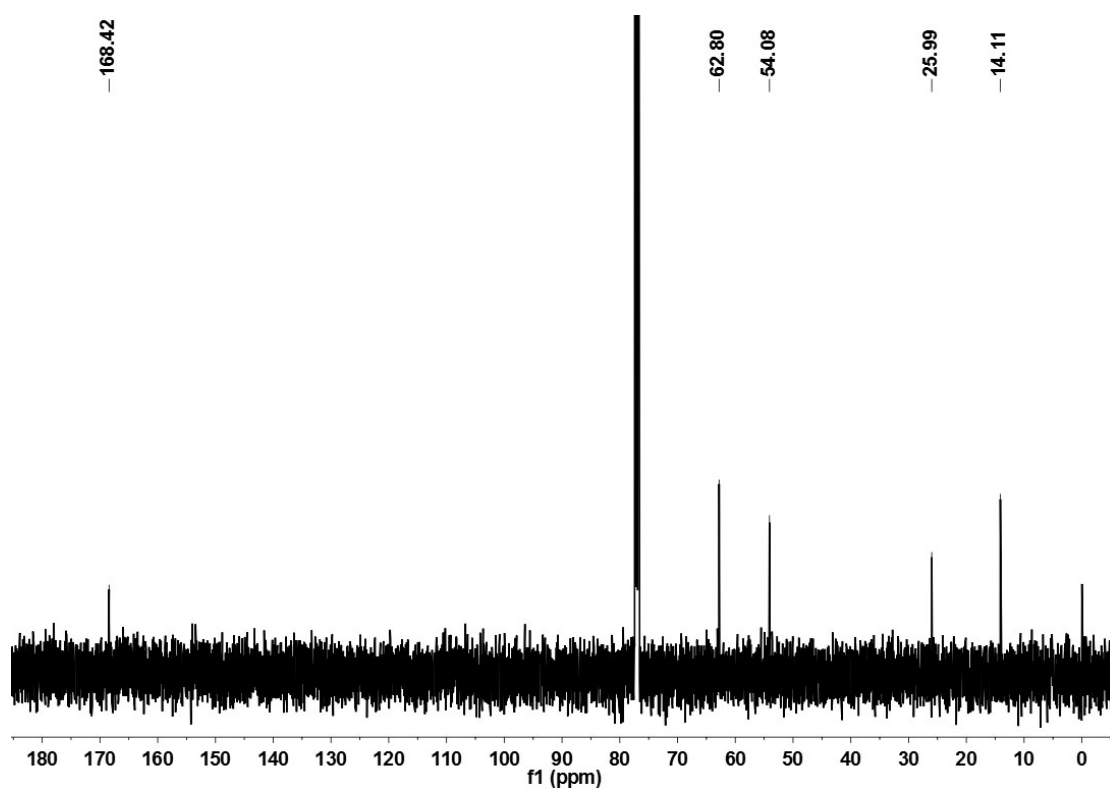


Fig. S36 ^{13}C NMR of D-1 (126 MHz, CDCl_3)

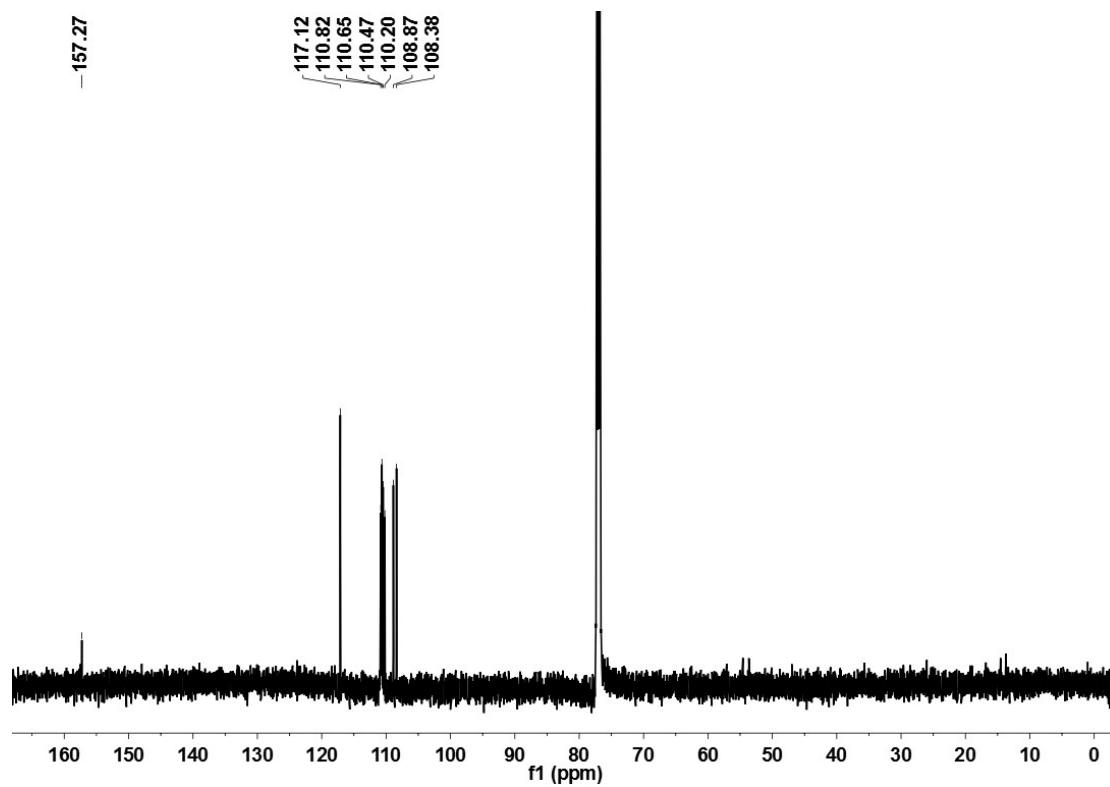


Fig. S37 ^{13}C NMR of D-1 (126 MHz, CDCl_3)

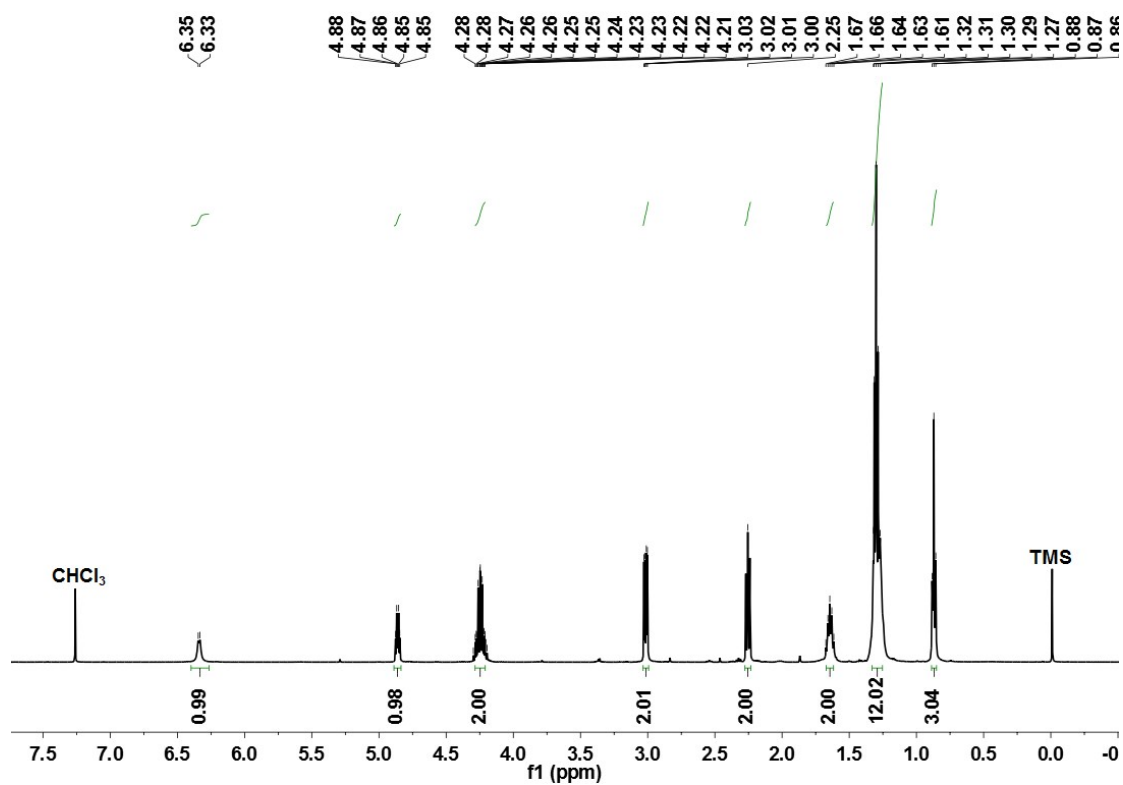


Fig. S38 ¹H NMR of L-2 (500 MHz, CDCl₃)

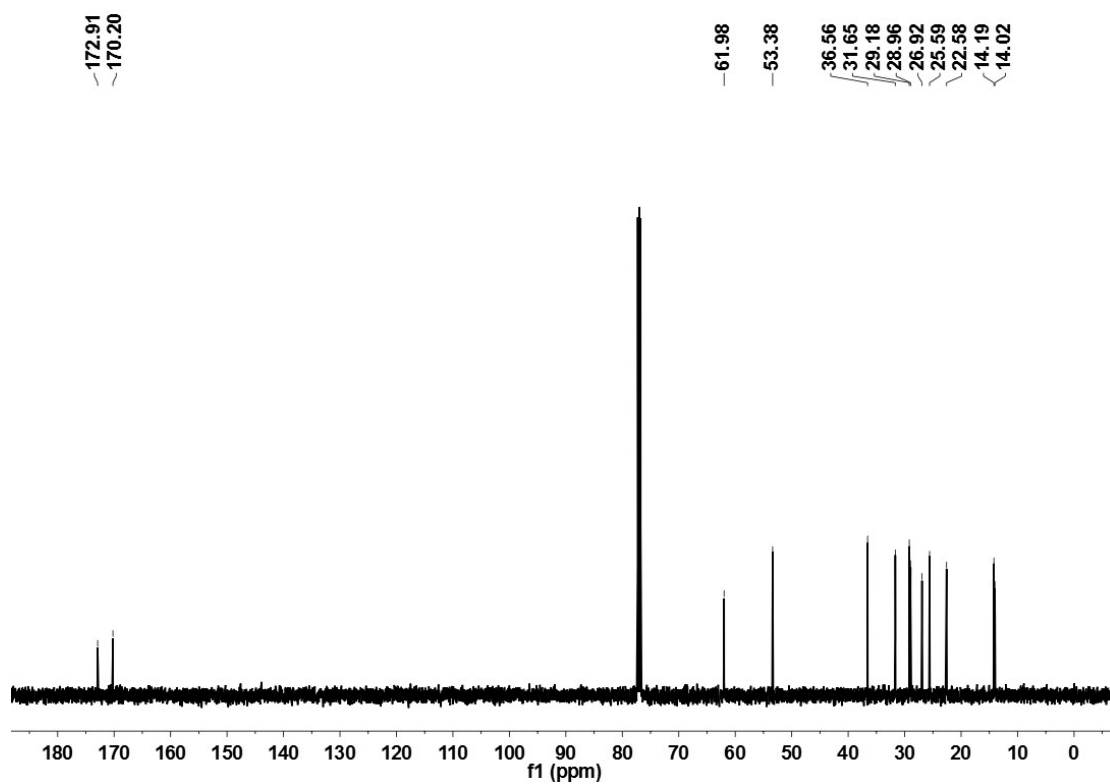


Fig. S39 ¹³C NMR of L-2 (126 MHz, CDCl₃)

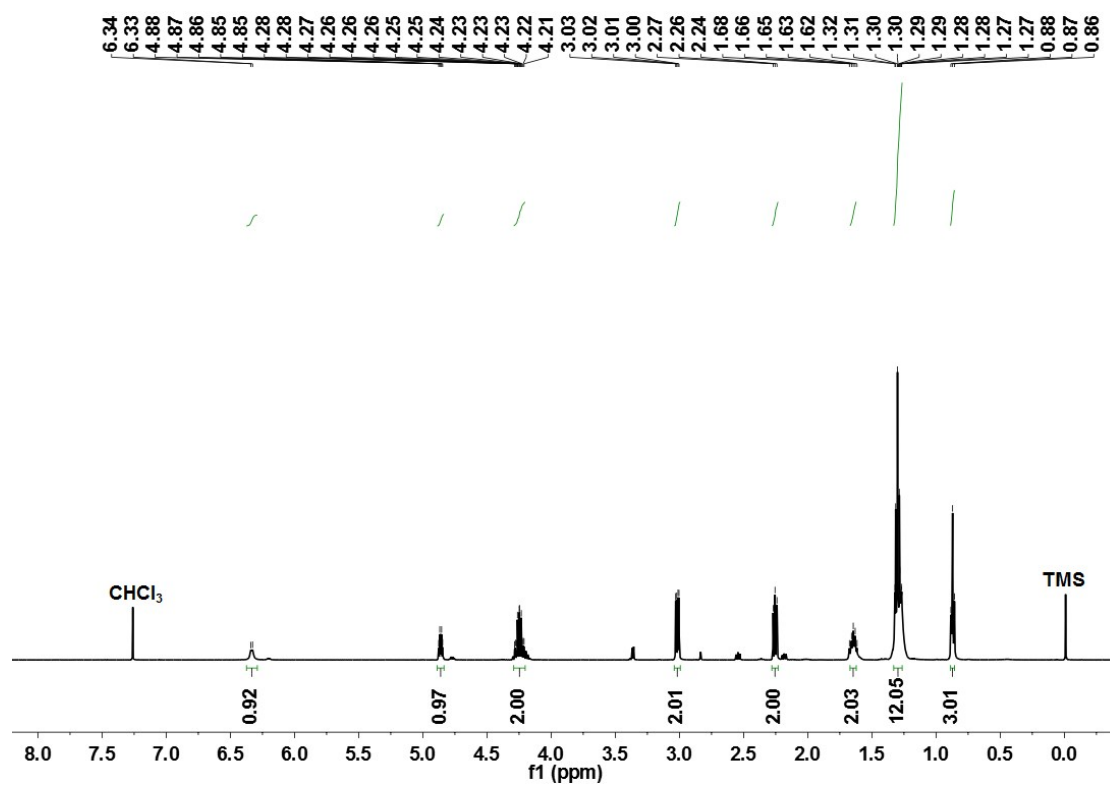


Fig. S40 ¹H NMR of D-2 (500 MHz, CDCl₃)

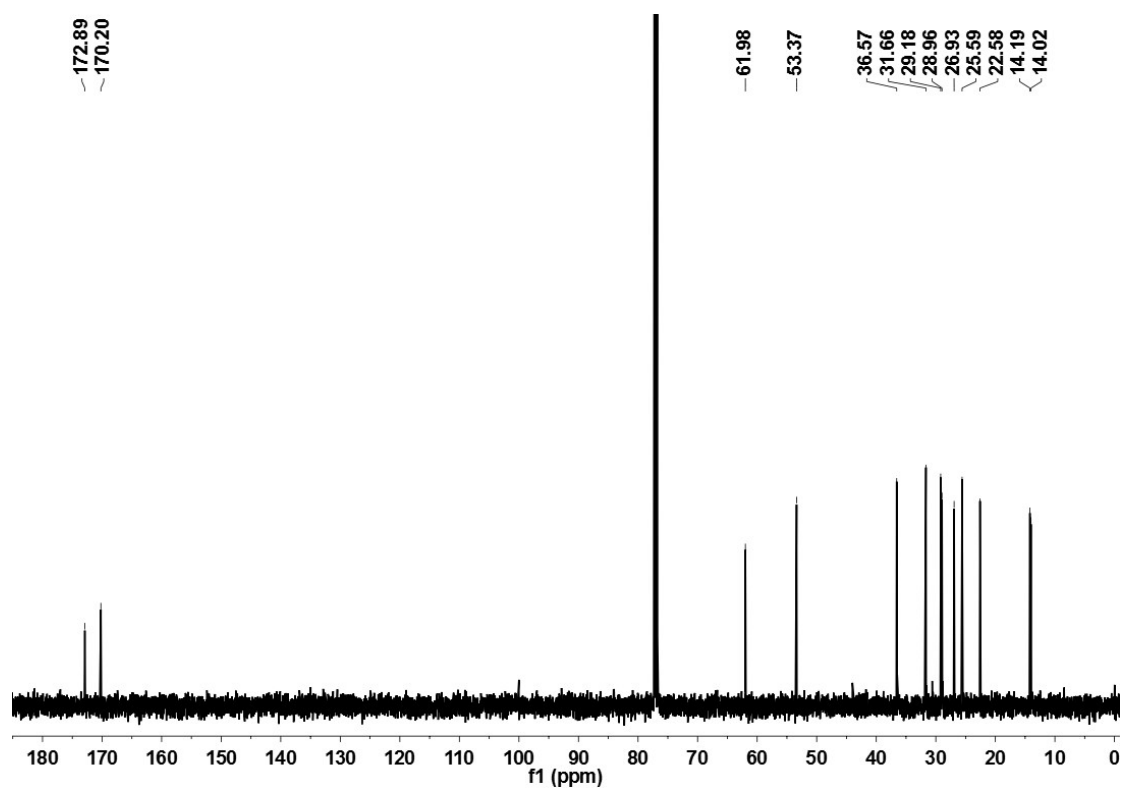


Fig. S41 ¹³C NMR of D-2 (126 MHz, CDCl₃)