SUPPLEMENTARY INFORMATION

Candida albicans Steryl 6-O-Acyl-α-D-Mannosides Agonize Signalling through Mincle

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Figure S1. Signalling of aCM and aEM through mutants of hMincle.

Signalling of synthetic α CM and α EM through human Mincle R183V and hMincle MD chimera (residues 195–202 in hMincle is replaced by the corresponding region of Dectin-2 (residues 192–199)).¹ NFAT-GFP 2B4 reporter cells expressing either human Mincle (R183V)/FcR γ , human Mincle-MD chimera/FcR γ or FcR γ alone were tested for their reactivity to plate-bound trehalose dibehenate (TDB) and steryl mannosides. Quantities denote the amount in nmol; assays were performed in duplicate; the mean values and standard deviations are shown.



Figure S2. Signalling of aCAMs and aEAMs through mutants of hMincle.

Signalling of synthetic α CAMs and α EAMs through human Mincle R183V and hMincle MD chimera (residues 195–202 in hMincle is replaced by the corresponding region of Dectin-2 (residues 192–199)).¹ NFAT-GFP 2B4 reporter cells expressing either human Mincle (R183V)/FcR γ , human Mincle-MD chimera/FcR γ or FcR γ alone were tested for their reactivity to plate-bound trehalose dibehenate (TDB) and steryl mannosides. Quantities denote the amount in nmol; assays were performed in duplicate; the mean values and standard deviations are shown.

Experimental

General

Pyridine was distilled over KOH before use. Dichloromethane and THF were dried over alumina according to the method of Pangborn *et al.*² Reactions were monitored using thin layer chromatography, performed with silica gel 60 F254. Detection was effected by charring in a mixture of 5% sulfuric acid in methanol, 10% phosphomolybdic acid in EtOH, and/or visualizing with UV light. Flash chromatography was performed using silica gel 60according to the method of Still *et al.*³ NMR experiments were conducted on 400 MHz instruments, with chemical shifts referenced relative to residual protiated solvent, and are in ppm. $^{1}H^{-1}H$ COSY spectra were used to confirm proton assignments. Mass spectra were acquired using an ESI quadrupole orbitrap.

Cholesteryl 2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranoside (1)

A solution of cholesterol (0.516 g, 1.34 mmol) and 2,3,4,6-tetra-O-benzoyl-mannopyranosyl trichloroacetimidate⁴ (0.90 g, 1.21 mmol) in dry dioxane (134 mL) was stirred with activated 3 Å molecular sieves at r.t for 30 min. The mixture was then cooled to 0 °C and TfOH (0.036 g, 21 µL, 0.242 mmol) was added. The reaction mixture was stirred at 0 °C for 30 min. The reaction was quenched with NEt₃ (0.245 g, 2.42 mmol, 0.337 mL) at 0 °C, then was diluted with diethyl ether and the solid was removed by filtration through a pad of Celite and washed with diethyl ether. The combined filtrates were washed with aq. NaHCO₃, aq. brine and dried (MgSO₄). The solvent was removed under reduced pressure to give a crude residue that was purified by flash chromatography (Et₂O:pet. spirit, 5:95) to afford a white solid that was recrystallised from ethanol to afford the pure α-anomer 1 as a white solid (0.514 g, 44%). ¹H NMR (400 Hz, CDCl₃) δ 8.06-8.08 (4 H, t, J = 7.5Hz), 7.97-7.99 (2 H, d, J = 8.3 Hz), 7.83-7.85 (2 H, d, J = 8.2 Hz), 7.49-7.62 (3 H, m), 7.35-7.45 (7 H, m), 7.25-7.29 (2 H, t, J = 7.7 Hz), 6.02-6.07 (1 H, t, J = 9.9 Hz, H4), 5.93-5.96 (1 H, dd, J = 10.1, 3.2, H3), 5.64-5.65 (1 H, m, H2), 5.27 (1 H, m, CH=), 5.25 (1 H, d, J = 1.5 Hz, H1), 4.65-4.68 (1 H, dd, J = 11.6, 1.5 Hz, H6a), 4.53-4.57 (1 H, m, H5), 4.47-4.51 (1 H, dd, J = 11.7, 5.4 Hz)H6b), 3.59-3.66 (1 H, m, cholesteryl-H3), 2.46-2.48 (2 H, m), 1.82-2.04 (5 H, m), 0.83-1.60 (21 H, m, cholesteryl-CH), 1.03 (3 H, s), 0.92-0.93 (3 H, d, J = 6.4 Hz), 0.87 (3 H, d, J = 6.6 Hz), 0.86 (3 H, d, J = 6.6 Hz), 0.69 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 166.0, 165.4, 165.3, 140.0, 133.3, 133.0, 132.8, 129.7, 129.6, 129.6, 129.3, 129.0, 128.8, 128.4, 128.3, 128.2, 128.1, 122.1, 95.8, 78.4, 71.0, 70.0, 68.8, 67.1, 63.1, 56.6, 56.0, 49.9, 42.2, 39.9, 39.6, 39.4, 36.8, 36.5, 36.0, 35.6, 31.8, 31.7, 28.1, 27.9, 27.7, 24.2, 23.7, 22.7, 22.4, 20.9, 19.3, 19.2, 18.6, 11.7. HRMS (ESI⁺) m/z calcd. for [C₆₁H₇₂O₁₀+H]⁺: 965.5198, obsd: 965.5199.

Cholesteryl α-D-mannopyranoside (3)

NaOMe (25% w/w in MeOH) was added dropwise into a solution of protected cholesteryl α -D-mannopyranoside **1** (0.514 g, 0.532 mmol) in dry MeOH:THF (3 mL: 2 mL) to achieve pH = 10. The solution was stirred at r.t for 1 h then was quenched with Dowex resin (H⁺ form), filtered and concentrated to afford a white solid. This solid was recrystallised from EtOAc containing a few drops of methanol to afford cholesteryl mannoside **3** (0.178 g, 61%). ¹H NMR (400 Hz, DMSO-d₆) δ 5.29-5.30 (1 H, d, *J* = 4.5 Hz, CH=), 4.75 (1 H, d, *J* = 1.5 Hz, H1), 4.67-4.68 (1 H, d, *J* = 4.8 Hz, OH-4), 4.63-4.65 (1 H, d, *J* = 4.4 Hz, OH-2), 4.51-4.52 (1 H, d, *J* = 5.9 Hz, OH-3), 4.39-4.42 (1 H, t, *J* = 5.8 Hz, OH-6), 3.61-3.66 (1 H, dd, *J* = 10.6, 6.1 Hz, H5), 3.54 (1 H, m, H2), 3.29-3.45 (5 H, m, H3,4,6a,6b,cholesteryl-H3), 2.31-2.35 (1 H, dd, *J* = 12.7, 4.3 Hz), 2.16-2.22 (1 H, t, *J* = 11.7 Hz), 1.76-1.97 (5 H, m), 0.99-1.55 (21 H, m, cholesteryl-CH), 0.95 (3 H, s), 0.89-0.90 (3 H, d, *J* = 6.4 Hz), 0.85-0.86 (3 H, d, *J* = 6.6 Hz), 0.83-0.84 (3 H, d, *J* = 6.6 Hz), 0.65 (3 H, s). ¹³C NMR (100 Hz, DMSO-d₆) δ 140.6, 121.1, 97.9, 75.2, 74.1, 70.9, 70.8, 67.1, 61.4, 56.2, 55.6, 49.5, 41.9, 36.6, 36.2, 35.7, 35.2, 31.4, 31.3, 27.8, 27.4, 23.9, 23.2, 22.7, 22.4, 20.6, 19.1, 18.6, 11.7. HRMS (ESI⁺) *m*/z calcd. for [C₃₃H₅₆O₆+H]⁺: 549.4150.

Cholesteryl 2,3,4-tri-O-trimethylsilyl-α-D-mannopyranoside (5)

Trimethylsilyl chloride (0.176 g, 1.62 mmol, 0.206 mL) and NEt₃ (0.170 mg, 1.68 mmol, 0.235 mL) were added to a stirred mixture of cholesteryl α-D-mannopyranoside **3** (0.178 g, 0.324 mmol) in CH₂Cl₂ (3 mL). The reaction was stirred overnight and then diluted with CH₂Cl₂ and aq. NaHCO₃. The combined organic layers were separated and washed with sat aq. NaHCO₃, water, aq. Brine, dried (Na₂SO₄) and concentrated. The residue was dissolved in CHCl₃ (3 mL) and NH₄OAc (55 mg, 0.71 mmol) was added. The reaction was allowed to stir for 48 h then concentrated and purified by flash chromatography (pre-treated with 4% NEt₃/pet. spirits, EtOAc:pet spirits 5:95) to afford 5 as a clear glass (248 mg, 76%). ¹H NMR (400 Hz, CDCl₃) δ 5.32-5.33 (1 H, dd, J = 3.4, 1.8 Hz, CH=), 4.73 (1 H, d, J = 1.9 Hz, H1), 3.84-3.88 (1 H, t, J = 9.1 Hz, H4), 3.76-3.78 (1 H, dd, J = 9.0, 2.5 Hz, H3, 3.73 - 3.76 (1 H, dd, J = 11.4, 2.7 Hz, H6a), 3.69 - 3.70 (1 H, m, H2), 3.66 - 3.68(1 H, m, H6b), 3.61-3.64 (1 H, m, H5), 2.28-2.31 (2 H, m), 1.94-2.01 (3 H, m), 1.83-1.85 (3 H, m), 0.86-1.60 (22 H, m, cholesteryl-CH), 1.00 (3 H, s), 0.90-0.91 (3 H, d, J = 6.2 Hz), 0.86-0.87 (3 H, d, J = 6.6 Hz), 0.85 (3 H, d, J = 6.6 Hz), 0.67 (3 H, s), 0.16 (9 H, s, 0.14 (9 H, s), 0.12 (9H, s). ¹³C NMR (100 Hz, CDCl₃) δ 140.8, 122.0, 99.1, 77.5, 74.4, 74.1, 72.6, 68.4, 62.4, 56.9, 56.3, 50.3, 42.5, 40.2, 39.9, 39.7, 37.2, 36.9, 36.3, 35.9, 32.1, 28.4, 28.2, 27.9, 24.4, 24.0, 23.0, 22.7, 21.2, 19.5, 18.9, 12.0, 0.83, 0.79, 0.52.

Ergosteryl 2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranoside (2)

A solution of ergosterol (0.456 g, 1.15 mmol) and 2,3,4,6-tetra-O-benzoyl-mannopyranosyltrichloroacetimidate⁴ (0.741 g, 1.00 mmol) in dry CH₂Cl₂ (100 mL) was stirred with activated 3 Å molecular sieves at r.t for 30 min. The mixture was cooled to 0 °C then TBDMSOTf (0.053 g, 0.200 mmol, 46 µL) was added. The mixture was stirred at 0 °C for 30 min then was guenched with NEt₃ (0.202 g, 2.00 mmol, 0.278 mL) at 0 °C. The mixture was diluted with diethyl ether and the solids were removed by filtration through a pad of Celite and washed with diethyl ether. The combined filtrates were washed with aq. NaHCO₃, aq. brine and dried (MgSO₄). The solvent was removed under reduced pressure to give a crude residue, which was purified by flash chromatography (ether:pet. spirit, 5:95) to afford a white solid. The solid was recrystallised from ethanol to afford compound **2** as a white solid (0.537 g, 61%). ¹H NMR (400 Hz, CDCl₃) δ 8.06-8.08 (4 H, d, J = 7.1Hz), 7.97-7.99 (2 H, d, J = 8.3 Hz), 7.83-7.85 (2 H, d, J = 8.3 Hz), 7.29-2.62 (12 H, m), 6.03-6.07 (1 H, t, J = 9.8 Hz, H4), 5.93-5.96 (1 H, dd, J = 10.0, 2.6 Hz, H3), 5.66 (1 H, m, H2), 5.49-5.50 (1 H, m, H2), 5.H, dd, J = 5.3, 2.0 Hz, ring-B CH=), 5.36-5.37 (1 H, m, ring-B CH=), 5.38 (1 H, d, J = 1.7 Hz, H1), 5.19-5.35 (2 H, t, J = 7.1 Hz, C22/23 CH=CH), 4.64-4.66 (1 H, d, J = 10.9 Hz, H6a), 4.49-4.57 (2 H, m, H5,6b), 3.71-3.78 (1 H, ddd, J = 15.4, 11.0, 4.4 Hz, ergosteryl-H3), 1.20-2.10 (20 H, m, ergosteryl-CH), 1.04-1.05 (3 H, d, J = 6.6 Hz), 0.97 (3 H, s), 0.92-0.93 (3 H, d, J = 6.8 Hz), 0.83-0.85 (6 H, t, J = 7.1 Hz), 0.65 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 166.4, 165.7, 165.7, 141.6, 139.2, 135.7, 133.6, 133.3, 133.2, 133.1, 130.0, 130.0, 129.9, 128.7, 128.6, 128.5, 120.2, 116.5, 96.0, 71.3, 70.3, 69.2, 67.3, 63.4, 55.9, 54.8, 46.3, 43.0, 40.6, 39.2, 38.7, 38.2, 37.4, 33.3, 28.5, 28.3, 23.2, 21.3, 21.2, 20.1, 19.8, 17.8, 16.4, 12.2. HRMS (ESI⁺) m/z calcd. for $[C_{62}H_{70}O_{10}+Na]^+$: 997.4861, obsd: 997.4856.

Ergosteryl α-D-mannopyranoside (4)

NaOMe (25% w/w in MeOH) was added dropwise into a solution of protected ergosteryl α -D-mannopyranoside **2** (0.537 g, 0.550 mmol) in dried MeOH:THF (33 mL: 22 mL) to achieve pH = 10. The reaction was stirred at r.t for 1 h then was quenched with Dowex resin (H⁺ form), filtered and concentrated to afford a white solid. This solid was recrystallised from EtOAc containing a few drops of methanol to afford compound **4** (0.200 g, 65%). ¹H NMR (400 Hz, DMSO-d₆) δ 5.50-5.51 (1 H, d, *J* = 4.2 Hz, ring-B CH=), 5.34 (1 H, m, ring-B CH=), 5.15-5.27 (2 H, qd, *J* = 15.4, 7.4 Hz, C22/23 CH=CH), 4.79 (1 H, s, H1), 4.67-4.69 (1 H, d, *J* = 4.9 Hz, OH-H4), 4.65-4.66 (1 H, d, *J* = 4.4 Hz, OH-H2), 4.52-4.53 (1 H, d, *J* = 5.9 Hz, OH-H3), 4.41-4.44 (1 H, t, *J* = 5.8 Hz, OH-H6), 3.62-3.67 (1 H, dd, *J* = 10.4, 6.0 Hz, H5), 3.55 (1 H, s, H2), 3.37-3.52 (5 H, m, H3,4,6a,6b,ergosteryl-H3), 2.21-2.33 (1 H, t, *J* = 12.9 Hz), 1.21-2.02 (22 H, m, ergosteryl-CH), 1.00-1.02 (3 H, d, *J* = 6.6 Hz), 0.87-0.90 (3 H, d, *J* = 6.8 Hz), 0.87 (3 H, s), 0.79-0.82 (3 H, t, *J* = 6.1 Hz), 0.59 (3 H, s). ¹³C NMR (100 Hz, DMSO-d₆) δ 140.9, 140.2, 135.8, 131.9, 119.6, 116.5,

98.3, 74.6, 74.2, 71.3, 71.2, 67.6, 61.8, 55.5, 54.3, 46.0, 42.8, 42.5, 38.8, 38.5, 37.9, 37.1, 32.9, 28.4, 28.1, 23.0, 21.4, 21.0, 20.2, 19.9, 17.8, 16.4, 12.3. HRMS (ESI⁺) *m/z* calcd. for [C₃₄H₅₄O₆+H]⁺: 581.3813, obsd: 581.3810.

Ergosteryl 2,3,4-tri-*O*-trimethylsilyl-α-D-mannopyranoside (6)

Trimethylsilyl chloride (0.176 g, 1.62 mmol, 0.206 mL) and NEt₃ (0.170 mg, 1.68 mmol, 0.235 mL) were added to a stirred mixture of ergosteryl α -D-mannopyranoside **4** (0.181 g, 0.324 mmol) in CH₂Cl₂ (6 mL). The reaction was stirred overnight and then diluted with CH₂Cl₂ and aq. NaHCO₃. The combined organic layers were separated and washed with sat aq. NaHCO₃, water, aq. brine and dried (Na₂SO₄) and concentrated. The residue was dissolved in CHCl₃ (6 mL) and NH₄OAc (55 mg, 0.71 mmol) was added. The reaction was allowed to stir for 48 h then concentrated and purified by flash chromatography (pre-treated with 4% NEt₃/pet. spirits, EtOAc:pet spirits 5:95) to afford **6** as a clear glass (130 mg, 52%). ¹H NMR (400 Hz, CDCl₃) δ 5.53-5.54 (1 H, d, *J* = 4.4 Hz, ring-B CH=), 5.37 (1 H, s, ring-B CH=), 5.13-5.25 (2 H, m, C22/23 CH=CH), 4.76 (1 H, s, H1), 3.52-3.89 (9 H, m), 2.46-2.51 (1 H, dd, *J* = 14.0, 3.1 Hz), 2.30-2.37 (1 H, m), 1.25-1.89 (17 H, m, ergosteryl-CH), 1.02-1.04 (3 H, d, *J* = 6.7 Hz), 0.94 (3 H, s), 0.91-0.92 (3 H, d, *J* = 6.7 Hz), 0.81-0.84 (6 H, t, *J* = 6.3 Hz), 0.62 (3 H, s), 0.16 (9 H, s), 0.15 (9 H, s), 0.13 (9 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 141.0, 139.6, 135.4, 131.8, 119.5, 116.2, 98.7, 75.1, 74.0, 73.9, 72.3, 68.0, 62.0, 58.6, 55.6, 54.5, 46.0, 45.5, 42.7, 40.3, 39.0, 38.4, 38.0, 37.1, 33.0, 28.1, 28.2, 27.9, 22.9, 21.0, 19.8, 19.5, 17.5, 16.2, 11.9, 10.4, 0.55, 0.52, 0.24.

General Procedure for Acylation of cholesteryl and ergosteryl 2,3,4-tri-*O*-trimethylsilyl-α-Dmannosides

Acyl chloride (2 equiv) was added a solution of DMAP (1 equiv) and steryl 2,3,4-tri-*O*-trimethylsilylmannoside **5** or **6** (1 equiv) in pyridine/CH₂Cl₂ (3:97, 10 mL/mmol) at 0 °C. The solution was stirred at r.t for 2 h then MeOH (1 mL/mmol) was added and the mixture was stirred for 0.5 h. The solvent was evaporated and the residue was redissolved in CHCl₃ (10 mL/mmol) and Dowex 50WX8-200 (H⁺ form, approx. 100 mg) was added. After stirring for 1 h, the mixture was filtered and the filtrate concentrated. Flash chromatography (EtOAc + 1% AcOH) afforded the product as a white solid.

Cholesteryl 6-*O*-dodecanoyl-α-D-mannopyranoside (7)

The General Protocol for Acylation between **5** (30 mg, 0.039 mmol) and lauroyl chloride (17.1 mg, 0.078 mmol, 18 μ L) after flash chromatography afforded **7** as white solid (20.2 mg, 71%). ¹H NMR (400 Hz, CDCl₃) δ 5.33-5.35 (1 H, d, *J* = 4.3 Hz, CH=), 5.01 (1 H, s, H1), 4.55-4.59 (1 H, dd, *J* =

12.3, 3.9 Hz, H6a), 4.20-4.23 (1 H, d, J = 11.9 Hz, H6b), 3.91 (1 H, s, H2), 3.81-3.86 (2 H, dd, J = 19.4, 7.9 Hz, H3,5), 3.53-3.58 (1 H, t, J = 9.6 Hz, H4), 3.48-3.50 (1 H, m, cholesteryl-H3), 2.36-2.40 (2 H, t, J = 7.6 Hz), 2.29-2.31 (2 H, d, J = 7.8 Hz), 1.80 - 2.02 (6 H, m), 1.43-2.15 (12 H, m, cholesteryl-CH), 1.26 (18 H, m, lipid), 1.10 (8 H, m), 1.00 (3 H, s), 0.90-0.92 (3 H, d, J = 6.5 Hz), 0.87-0.89 (3 H, t, J = 6.7 Hz, ω -CH₃), 0.85-0.87 (3 H, d, 6.6 Hz), 0.86-0.87 (3 H, d, J = 6.6 Hz), 0.67 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 175.0, 140.4, 121.8, 97.6, 77.0, 71.2, 70.8, 70.3, 67.8, 63.2, 56.6, 56.0, 50.0, 42.2, 39.8, 39.6, 39.4, 36.8, 36.5, 36.0, 35.6, 34.10, 31.8, 31.8, 29.5, 29.4, 29.2, 29.1, 29.1, 28.1, 27.9, 27.5, 24.8, 24.2, 23.7, 22.7, 22.5, 22.4, 20.9, 19.2, 18.6, 14.0, 11.7. HRMS (ESI⁺) *m/z* calcd. for [C₄₅H₇₈O₇+Na]⁺: 753.5640, obsd: 753.5645.

Cholesteryl 6-O-tetradecanoyl-a-d-mannopyranoside (8)

The General Protocol for Acylation between **5** (30 mg, 0.039 mmol) and myristoyl chloride (19.3 mg, 0.078 mmol, 21.2 µL) after flash chromatography afforded **8** as white solid (22.4 mg, 76%). ¹H NMR (400 Hz, CDCl₃) δ 5.33-5.34 (1 H, d, J = 4.3 Hz, CH=), 4.99 (1 H, s, H1), 4.50-4.53 (1 H, dd, J = 11.8, 4.5 Hz, H6a), 4.25-4.27 (1 H, d, J = 11.7 Hz, H6b), 3.91 (1 H, s, H2), 3.82-3.85 (1 H, t, J = 8.4 Hz, H3), 3.47-3.49 (1 H, m, cholesteryl-H3), 3.33 (1 H, s, OH), 3.05 (1 H, s, OH), 2.70 (1 H, s, OH), 2.35-2.37 (2 H, t, J = 7.6 Hz), 2.30-2.31 (2 H, d, J = 7.0 Hz), 1.95-2.02 (2 H, m), 1.84-1.86 (2 H, m), 1.30-1.70 (18 H, m), 1.25 (22 H, m, lipid), 1.09-1.15 (6 H, m), 1.00 (3 H, s), 0.91-0.02 (3 H, d, J = 6.4 Hz), 0.87-0.89 (3 H, t, J = 6.4 Hz, ω -CH₃), 0.85-0.87 (6 H, J = 5.9 Hz), 0.67 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 174.8, 140.4, 121.8, 97.6, 77.0, 71.2, 70.9, 70.3, 67.8, 63.3, 56.6, 56.0, 50.0, 42.2, 39.8, 39.6, 39.4, 36.9, 36.5, 36.9, 36.5, 36.0, 35.7, 34.1, 31.8, 31.7, 29.6, 29.5, 29.4, 29.23, 29.19, 29.12, 28.1, 27.9, 27.5, 24.8, 24.1, 23.7, 22.7, 22.5, 22.4, 20.9, 19.2, 18.6, 14.0 11.7. HRMS (ESI⁺) *m/z* calcd. for [C₄₇H₈₂O₇+Na]⁺: 781.5953, obsd: 781.5950. The ¹H NMR spectrum was consistent with that reported.⁵

Cholesteryl 6-*O*-hexadecanoyl-α-D-mannopyranoside (9)

The General Protocol for Acylation between **5** (30 mg, 0.039 mmol) and palmitoyl chloride (21.4 mg, 0.078 mmol, 23.7 µL) after flash chromatography afforded **9** as white solid (23.9 mg, 78%). ¹H NMR (400 Hz, CDCl₃) δ 5.33-5.34 (1 H, d, J = 3.2 Hz, CH=), 5.00 (1 H, s, H1), 4.54-4.55 (1 H, dd, J = 11.8, 4.3 Hz, H6a), 4.21-4.24 (1 H, d, J = 11.8 Hz, H6b), 3.91 (1 H, s, H2), 3.81-4.84 (2 H, m, H3,5), 3.55 (1 H, t, J = 9.6 Hz, H4), 3.47-3.49 (1 H, m, cholesteryl-H3), 2.35-2.39 (2 H, t, J = 7.4 Hz), 2.29-2.31 (2 H, d, J = 7.8 Hz), 1.84-1.89 (6 H, m), 1.04-1.62 (12 H, m, cholesteryl-CH), 1.25 (26 H, s, lipid), 1.11 (8 H, m), 0.99 (3 H, s), 0.90-0.92 (3 H, d, J = 5.6 Hz), 0.85-0.87 (6 H, d, J = 6.6 Hz), 0.87-0.89 (3 H, t, J = 6.7 Hz, ω -CH₃), 0.67 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 175.2,

140.7, 122.1, 97.9, 77.2, 71.5, 71.2, 70.6, 68.1, 63.6, 56.9, 56.3, 50.3, 42.5, 40.1, 39.9, 39.7, 37.2, 36.9, 36.4, 36.0, 34.4, 32.1, 32.0, 29.9, 29.8, 29.7, 29.6, 29.4, 29.4, 28.4, 28.2, 27.8, 25.2, 24.4, 24.0, 23.0, 22.9, 22.7, 21.2, 19.5, 18.9, 14.3, 12.0. HRMS (ESI⁺) *m/z* calcd. for [C₄₉H₈₆O₇+Na]⁺: 809.6266, obsd: 809.6258.

Cholesteryl 6-O-octadecanoyl-α-D-mannopyranoside (10)

The General Protocol for Acylation between **5** (30 mg, 0.039 mmol) and stearoyl chloride (23.6 mg, 0.078 mmol, 26.3 µL) after flash chromatography afforded **10** as white solid (24.8 mg, 78%). ¹H NMR (400 Hz, CDCl₃) δ 5.33-5.34 (1 H, d, J = 4.1 Hz, CH=), 5.01 (1 H, s, H1), 4.54-4.58 (1 H, dd, J = 12.2, 4,4 Hz, H6a), 4.22-4.24 (1 H, d, J = 11.4 Hz, H6b), 3.91 (1 H, s, H2), 3.81-3.88 (2 H, m, H3,5), 3.53-3.58 (1 H, t, J = 9.6 Hz, H4), 3.47-3.51 (1 H, m, cholesteryl-H3), 2.35-2.39 (2 H, t, J = 7.6 Hz), 2.29-2.31 (2 H, d, J = 7.7 Hz), 1.95-2.02 (2 H, m), 1.80-1.89 (3 H, m), 1.05-1.64 (13 H, m, cholesteryl-CH), 1.25 (30 H, lipid), 1.10 (8 H, m), 1.00 (3 H, s), 0.90-0.92 (3 H, d, J = 6.4 Hz), 0.87-0.89 (3 H, t, J = 6.8 Hz, ω -CH₃), 0.87 (3 H, d, J = 1.2 Hz), 0.85 (3 H, d, J = 1.2 Hz), 0.67 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 175.1, 140.5, 122.0, 97.7, 71.3, 71.0, 70.5, 67.9, 63.3, 56.73, 56.2, 50.1, 42.3, 39.6, 39.7, 39.5, 37.0, 36.7, 36.2, 35.8, 34.3, 31.9, 31.8, 29.7, 29.6, 29.5, 29.4, 29.3, 28.7, 28.2, 28.0, 27.6, 25.0, 24.3, 23.8, 22.8, 22.7, 22.5, 21.0, 19.3, 18.7, 14.1, 11.8. HRMS (ESI⁺) *m/z* calcd. for [C₅₁H₉₀O₇+Na]⁺: 837.6579, obsd: 837.6580.

Cholesteryl 6-*O*-oleyl-α-D-mannopyranoside (11)

The General Protocol for Acylation between **5** (30 mg, 0.039 mmol) and oleoyl chloride (23.5 mg, 0.078 mmol, 25.8 µL) after flash chromatography afforded **11** as white solid (27.2 mg, 86%). ¹H NMR (400 Hz, CDCl₃) δ 5.32-5.35 (3 H, m), 4.99 (1 H, s, H1), 4.51-4.55 (1 H, dd, J = 11.8, 4.1 Hz, H6a), 4.23-4.25 (1 H, d, J = 11.6 Hz, H6b), 3.91 (1 H, s, H2), 3.81-3.87 (2 H, m, H3,5), 3.56-3.59 (1 H, m), 3.46-3.49 (1 H, m, cholesteryl-H3), 2.34-2.38 (2 H, t, J = 7.6 Hz), 2.29-2.31 (2 H, d, J = 7.5 Hz), 2.01-2.00 (6 H, m), 1.29 (14 H, m, lipid), 1.26 (12 H, m, lipid), 1.11-1.86 (12 H, m, cholesteryl-CH), 0.99 (3 H, s), 1.11 (6 H, m), 0.90-0.92 (3 H, d, J = 6.5 Hz), 0.87-0.89 (3 H, t, J = 6.9 Hz, ω -CH₃), 0.85-0.89 (6 H, d, J = 6.6 Hz), 0.67 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 174.9, 140.4, 129.9, 129.5, 121.8, 97.6, 77.0, 71.2, 70.8, 70.3, 67.8, 63.2, 56.6, 56.0, 50.0, 42.2, 39.8, 39.6, 39.4, 36.8, 36.5, 36.0, 35.6, 34.1, 31.8, 31.7, 29.6, 29.4, 29.2, 29.0, 28.1, 27.9, 27.5, 27.1, 27.0, 24.8, 24.1, 23.7, 22.4, 20.9, 19.2, 18.6, 14.0, 11.7. HRMS (ESI⁺) *m/z* calcd. for [C₅₁H₈₈O₇+Na]⁺: 812.6530, obsd: 812.6538.

Ergosteryl 6-O-dodecanoyl-α-D-mannopyranoside (12)

The General Protocol for Acylation between **6** (23.5 mg, 0.030 mmol) and lauroyl chloride (13.1 mg, 0.060 mmol, 13.8 µL) after flash chromatography afforded **12** as white solid (16.2 mg, 73%). ¹H NMR (400 Hz, CDCl₃) δ 5.54-5.55 (1 H, d, J = 4.0 Hz, ring-B CH=), 5.36-5.37 (1 H, m, ring-B CH=), 5.13-5.25 (2 H, qd, J = 15.3, 7.3 Hz, C22/C23 CH=CH), 5.03 (1 H, s, H1), 4.52-4.56 (1 H, dd, J = 12.2, 4.6 Hz, H6a), 4.22-4.25 (1 H, d, J = 11.4 Hz, H6b), 3.92 (1 H, s, H2), 3.81-3.84 (2 H, m, H3,5), 3.57-3.59 (2 H, m, H4, ergosteryl-H3), 2.45-2.49 (1 H, dd, J = 14.5, 2.8 Hz), 2.34-2.38 (3 H, t, J = 7.7 Hz), 1.25 (18 H, m, lipid), 1.25-2.04 (18 H, m, ergsoteryl-CH), 1.02-1.04 (3 H, d, J = 6.6 Hz), 0.93 (3 H, s), 0.90-0.93 (3 H, d, J = 6.9 Hz), 0.86-0.89 (3 H, t, J = 6.8 Hz, ω -CH₃), 0.81-0.84 (6 H, t, J = 6.4 Hz), 0.62 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 175.2, 141.5, 139.6, 135.7, 132.1, 120.0, 116.4, 97.8, 75.7, 71.6, 71.2, 70.7, 68.0, 63.7, 55.9, 54.7, 46.3, 43.0, 42.9, 40.6, 39.2, 38.6, 38.2, 37.4, 34.5, 33.2, 32.1, 29.9, 29.8, 29.6, 29.5, 29.5, 28.5, 28.1, 25.2, 23.2, 22.9, 21.3, 21.2, 20.1, 19.8, 17.8, 16.3, 14.3, 12.2. HRMS (ESI⁺) *m*/*z* calcd. for [C₄₆H₇₆O₇+Na]⁺: 763.5483, obsd: 763.5480.

Ergosteryl 6-O-tetradecanoyl-α-D-mannopyranoside (13)

The General Protocol for Acylation between **6** (23.5 mg, 0.030 mmol) and myristoyl chloride (14.8 mg, 0.060 mmol, 16.3 µL) after flash chromatography afforded **13** as white solid (16.8 mg, 73%). ¹H NMR (400 Hz, CDCl₃) δ 5.55-5.56 (1 H, d, *J* = 3.8 Hz, ring B CH=), 5.37-5.38 (1 H, m, ring B CH=), 5.15-5.25 (2 H, m, C22/23 CH=CH), 5.04 (1 H, s, H1), 4.54-4.56 (1 H, d, *J* = 11.6 Hz, H6a), 4.23-4.25 (1 H, d, *J* = 11.9 Hz, H6b), 3.92 (1 H, s, H2), 3.81-3.87 (2 H, m, H3,5), 3.54-3.63 (2 H, m, H4, cholesteryl-H3), 3.56 (2 H, m), 2.46-2.50 (1 H, dd, *J* = 14.8, 2.4 Hz), 2.36-2.39 (3 H, m), 1.34-2.05 (16 H, m, ergosteryl-CH), 1.25 (22 H, m, lipid), 1.03-1.04 (3 H, d, *J* = 6.6 Hz), 0.94 (3 H, s), 0.91-0.92 (3 H, d, *J* = 6.8 Hz), 0.87-0.88 (3 H, t, *J* = 6.9 Hz, ω -CH₃), 0.82-0.85 (6 H, t, *J* = 7.2 Hz), 0.63 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 175.2, 141.5, 139.6, 135.7, 132.1, 120.0, 116.4, 97.8, 75.7, 71.5, 71.1, 70.7, 68.1, 63.5, 58.3, 55.9, 54.7, 46.3, 43.0, 40.6, 39.2, 38.6, 38.2, 37.4, 34.4, 33.2, 32.1, 29.9, 29.8, 29.7, 29.5, 29.4, 29.4, 28.4, 28.1, 25.2, 23.2, 22.9, 21.3, 21.2, 20.2, 20.1, 19.8, 17.8, 16.4, 14.3, 12.2, 8.3. HRMS (ESI⁺) *m/z* calcd. for [C₄₈H₈₀O₇+Na]⁺: 791.5796, obsd: 791.5794.

Ergosteryl 6-O-hexadecanoyl-α-D-mannopyranoside (14)

The General Protocol for Acylation between **6** (23.5 mg, 0.030 mmol) and palmitoyl chloride (16.5 mg, 0.060 mmol, 18.2 μ L) after flash chromatography afforded **14** as white solid (21.5 mg, 90%). ¹H NMR (400 Hz, CDCl₃) δ 5.56 (1 H, d, *J* = 3.8 Hz, ring B CH=), 5.38-5.39 (1 H, m, ring B CH=), 5.21-5.26 (2 H, qd, *J* = 15.3, 7.5 Hz, C22/23 CH=CH), 5.05 (1 H, s, H1), 4.52-4.55 (1 H, dd, *J* = 12.0, 4.6 Hz, H6a), 4.26-4.28 (1 H, d, *J* = 11.2 Hz, H6b), 3.93 (1 H, s, H2), 3.83-3.85 (2 H, m, S11

H3,5), 3.56-3.62 (4 H, m), 2.48-2.51 (1 H, dd, J = 14.9, 2.3 Hz), 2.36-2.38 (4 H, m), 1.42-2.06 (15 H, m, ergosteryl- CH), 1.26 (26 H, m, lipid), 1.04-1.05 (3 H, d, J = 6.6 Hz), 0.95 (3 H, s), 0.92-0.93 (3 H, d, J = 6.8 Hz), 0.88-0.90 (3 H, t, J = 6.9 Hz, ω -CH₃), 0.83-0.86 (6 H, t, J = 7.2 Hz), 0.64 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 175.1, 141.5, 139.6, 135.7, 132.7, 119.9, 116.4, 97.8, 75.7, 71.6, 71.1, 70.7, 68.1, 63.6, 58.6, 55.9, 54.7, 46.3, 43.0, 40.6, 39.2, 38.6, 38.2, 37.4, 34.4, 33.2, 32.1, 30.0, 29.8, 29.7, 29.5, 29.4, 29.3, 28.4, 28.1, 25.2, 23.2, 23.0, 21.2, 21.2, 20.1, 19.8, 17.8, 16.4, 14.3, 12.2, 8.4. HRMS (ESI⁺) *m/z* calcd. for [C₅₀H₈₄O₇+Na]⁺: 819.6109, obsd: 819.6113.

Ergosteryl 6-O-octadecanoyl-α-D-mannopyranoside (15)

The General Protocol for Acylation between **6** (23.5 mg, 0.030 mmol) and stearoyl chloride (18.2 mg, 0.060 mmol, 20.3 µL) after flash chromatography afforded **15** as white solid (18.5 mg, 75%). ¹H NMR (400 Hz, CDCl₃) δ 5.54-5.55 (1 H, d, J = 3.8 Hz, ring B CH=), 5.36-5.37 (1 H, m, ring B CH=), 5.14-5.25 (2 H, qd, J = 15.3, 7.5 Hz, C22/23 CH=CH), 5.02 (1 H, s, H1), 4.47-4.50 (1 H, dd, J = 11.9, 5.1 Hz, H6a), 4.27-4.30 (1 H, d, J = 11.4 Hz, H6b), 3.91 (1 H, m, H2), 3.84 (2 H, m, H3,5), 3.57-3.60 (2 H, m), 2.46-2.50 (1 H, dd, J = 14.7, 2.3 Hz), 2.34-2.37 (3 H, t, J = 7.7 Hz), 1.25-2.07 (18 H, m, ergosteryl-CH), 1.25 (30 H, m, lipid) 1.03-1.04 (3 H, d, J = 6.6 Hz), 0.93 (3 H, s), 0.91-0.92 (3 H, d, J = 7.0 Hz), 0.86-0.89 (3 H, t, J = 6.9 Hz, ω -CH₃), 0.82-0.85 (6 H, t, J = 7.1 Hz), 0.62 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 175.0, 141.4, 139.5, 135.7, 132.1, 120.0, 116.4, 97.9, 75.8, 71.6, 71.2, 70.7, 68.1, 63.7, 55.9, 54.7, 46.3, 43.0, 42.9, 40.6, 39.2, 38.6, 38.2, 37.4, 34.5, 33.2, 21.1, 29.88, 29.82, 29.79, 29.51, 29.49, 29.40, 28.4, 28.1, 23.1, 22.9, 21.3, 21.2, 20.1, 19.8, 17.8, 16.3, 14.3, 12.2. HRMS (ESI⁺) *m*/*z* calcd. for [C₅₂H₈₈O₇+Na]⁺: 847.6422, obsd: 847.6424.

Ergosteryl 6-*O*-oleyl-α-D-mannopyranoside (16)

The General Protocol for Acylation between **6** (23.5 mg, 0.030 mmol) and oleoyl chloride (18.1 mg, 0.060 mmol, 19.8 μ L) after flash chromatography afforded **16** as white solid (20.5 mg, 82%). ¹H NMR (400 Hz, CDCl₃) δ 5.54-5.56 (1 H, dd, J = 5.2, 1.7 Hz, ring B CH=), 5.37-5.38 (1 H, m, ring B CH=), 5.33-5.35 (dt, 2 H, J = 5.6, 4.3 Hz), 5.15-5.25 (dq, 2 H, J = 15.3, 7.5 Hz, C22/23 CH=CH), 5.03 (1 H, s, H1), 4.53-4.56 (1 H, dd, J = 12.2, 4.5 Hz, H6a), 4.23-4.25 (1 H, d, J = 10.9 Hz, H6b), 3.92 (1 H, m, H2), 3.81-3.87 (2 H, m, H3,5), 3.55-3.62 (2 H, m), 2.46-2.50 (1 H, dd, J = 14.7, 2.4 Hz), 2.35-2.38 (3 H, m), 1.61-2.08 (18 H, m, ergosteryl-CH), 1.31 (14 H, m, lipid), 1.27 (12 H, m, lipid), 1.03-1.04 (3 H, d, J = 6.6 Hz), 0.93 (3 H, s), 0.91-0.92 (3 H, d, J = 6.8 Hz), 0.86-0.89 (3 H, t, J = 6.9 Hz), 0.82-0.85 (6 H, t, J = 7.2 Hz), 0.63 (3 H, s). ¹³C NMR (100 Hz, CDCl₃) δ 175.2, 141.5, 139.6, 135.7, 132.2, 130.2, 129.9, 120.0, 116.4, 97.8, 75.8, 71.6, 71.1, 70.7, 68.1, 63.6, 55.9, 54.7, 46.3, 43.0, 40.6, 39.2, 38.6, 38.2, 37.4, 34.4, 33.2, 32.1, 29.93, 29.88, 29.69, 29.49, S12

29.48, 29.35, 29.31, 28.4, 28.1, 27.39, 27.37, 25.1, 23.2, 22.8, 21.3, 21.2, 20.1, 19.8, 17.8, 16.4, 14.3, 12.2. HRMS (ESI⁺) *m/z* calcd. for [C₅₂H₈₆O₇+Na]⁺: 845.6266, obsd: 845.6270.

Mincle reporter cell signalling assays

2B4-NFAT-GFP reporter cells expressing mouse or human Mincle (or the R135L mutant;⁶ or the Mincle R183V and hMincle MD chimera (residues 195–202 in hMincle is replaced by the corresponding region of Dectin-2 (residues 192–199)¹), together with FcR γ , as well as an FcR γ only control, were prepared as described previously.^{7, 8} Each glycolipid was dissolved in chloroform methanol (2:1) at 5 or 10 mg mL⁻¹, then diluted in isopropanol, was added to 96-well plates at 20 μ L per well, followed by the evaporation of the solvent. Reporter cells were stimulated for 16–20 h, and activation of NFAT-GFP was monitored by flow cytometry.

Bone marrow-derived dendritic cell (BMDC) preparation

Mincle-deficient mice⁹ were backcrossed for at least fifteen generations with C57BL/6 mice. All mice were maintained in a filtered-air laminar-flow enclosure and given standard laboratory food and water ad libitum. All animal experiments were performed in accordance with Institute guidelines. Preparation of BMDCs followed previously reported approach.¹⁰ For preparing BMDCs, bone marrow cells were suspended in RPMI 1640 medium supplemented with 10% fetal calf serum and β -mercaptoethanol, plated in the presence of 20ng mL⁻¹ recombinant mouse granulocyte macrophage colony stimulating factor (BioLegend) and cultured for 7 days at 37 °C.

BMDC stimulation assay

To stimulate BMDCs, each glycolipid was dissolved in chloroform methanol (2:1) at 5 or 10 mg mL⁻¹, then diluted in isopropanol, was added to 96-well plates at 20 μ L per well, followed by the evaporation of the solvent. LPS was added at final concentration of 10 ng mL⁻¹. BMDCs were stimulated for 1 day, and the supernatant was harvested. Tumour necrosis factor (TNF) was measured by ELISA (BD Biosciences).

References

- 1. A. Furukawa, J. Kamishikiryo, D. Mori, K. Toyonaga, Y. Okabe, A. Toji, R. Kanda, Y. Miyake, T. Ose, S. Yamasaki and K. Maenaka, *Proc. Natl. Acad. Sci. USA*, 2013, **110**, 17438-17443.
- 2. A. B. Pangborn, M. A. Giardello, R. H. Grubbs, R. K. Rosen and F. J. Timmers, *Organometallics*, 1996, **15**, 1518-1520.
- 3. W. C. Still, M. Kahn and A. M. Mitra, J. Org. Chem., 1978, 43, 2923-2925.
- 4. F. Bien and T. Ziegler, *Tetrahedron: Asymmetry*, 1998, **9**, 781-790.

- 5. M. Shimamura, M. Yamamura, T. Nabeshima, N. Kitano, P. van den Elzen, H. Yesilkaya, P. Andrew and P. Illarionov, *Sci. Rep.*, 2017, 7, 9703.
- 6. R. Kiyotake, M. Oh-Hora, E. Ishikawa, T. Miyamoto, T. Ishibashi and S. Yamasaki, *J. Biol. Chem.*, 2015, **290**, 25322-25332.
- 7. E. Ishikawa, T. Ishikawa, Y. S. Morita, K. Toyonaga, H. Yamada, O. Takeuchi, T. Kinoshita, S. Akira, Y. Yoshikai and S. Yamasaki, *J. Exp. Med.*, 2009, **206**, 2879-2888.
- P. L. van der Peet, C. Gunawan, S. Torigoe, S. Yamasaki and S. J. Williams, *Chem. Commun.*, 2015, 51, 5100-5103.
- S. Yamasaki, M. Matsumoto, O. Takeuchi, T. Matsuzawa, E. Ishikawa, M. Sakuma, H. Tateno, J. Uno, J. Hirabayashi, Y. Mikami, K. Takeda, S. Akira and T. Saito, *Proc. Natl. Acad. Sci. USA*, 2009, 106, 1897-1902.
- 10. M. Nagata, Y. Izumi, E. Ishikawa, R. Kiyotake, R. Doi, S. Iwai, Z. Omahdi, T. Yamaji, T. Miyamoto, T. Bamba and S. Yamasaki, *Proc. Natl. Acad. Sci. USA*, 2017, **114**, E3285-e3294.

NMR Spectra

Cholesteryl 2,3,4,6-tetra-O-benzoyl- α -D-mannopyranoside (1) ¹H NMR





Cholesteryl α -D-mannopyranoside (3) ¹H NMR





5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0 0.8 0.6 0.4













Ergosteryl α-D-mannopyranoside (4) ¹H NMR















Cholesteryl 6-*O***-dodecanoyl**-α-**D**-mannopyranoside (7) ¹H NMR

















Cholesteryl 6-*O*-octadecanoyl-α-D-mannopyranoside (10) ¹H NMR





Ergosteryl 6-*O***-dodecanoyl-α-***D***-mannopyranoside (12)** ¹H NMR













Ergosteryl 6-*O*-octadecanoyl-α-D-mannopyranoside (15) ¹H NMR



Ergosteryl 6-*O*-oleyl-α-D-mannopyranoside (16) ¹H NMR



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