Electronic Supplementary Information

Reversible hydrogen-bonded polymerization regulated by allosteric metal templation

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1. Materials, methods, and abbreviations

General

All chemicals, reagents and solvents were purchased from commercial suppliers and used, unless otherwise stated, without further purification. If needed, solvents were dried by literature known procedures. All yields were given as isolated yields. N1^[S1], 1^[S2], and 2^[S3] were prepared according to literature procedure.

NMR spectroscopy

The ¹H NMR and ¹³C NMR spectra were recorded with a Bruker AVANCE III (400 MHz or 600 MHz) spectrometer and calibrated against the residual proton signal or natural abundance carbon resonance of the used deuterated solvent from tetramethylsilane (TMS) as the internal standard. The chemical shifts δ are indicated in ppm and the coupling constants *J* in Hz. The multiplicities are given as s (singlet), d (doublet), dd (doublet of doublets), t (triplet), and m (multiplet).

Diffusion Ordered Spectroscopy (DOSY) were carried out at 298 K on a Bruker AVANCE III 600 spectrometer. The ledbpgp2s pulse sequence from Bruker Biospin was selected for the DOSY NMR by using gradients varied linearly from 5% up to 95% in 32 steps, with 16 scans per step. The diffusion time (Δ) was set at 20 ms, and the gradient length (δ) was set at 2 ms. Heptakis(2,3,6-tri-O-methyl)- β -cyclodextrin (peralkylated β -CD) was used as internal standard if necessary.

Mass spectrometry

Low-resolution electrospray ionization mass spectra (LR-ESI-MS) were obtained on LCMS2020. High-resolution electrospray ionization mass spectra (HR-ESI-MS) were recorded on an Agilent Technologies 6540 UHD Accurate-Mass. High-resolution mass spectra (HRMS) of **M3**+KPF₆ were measured on a Q-Exactive[™] HF/UltiMate[™] 3000 RSLCnano using a Nano roFlow meter with ProFlow technology in positive mode.

Melting point determination

The melting points were measured on a Haineng MP100 automatic melting point instrument.

Scanning electron microscopy (SEM)

SEM investigations were carried out on a ZEISS SUPRA 55 instrument.

Viscometry

Viscosity measurements were carried out with Ubbelohde micro viscometers (Shanghai Liangjing Glass Instrument Factory, 0.40 mm inner diameter) at 298 K in chloroform and acetonitrile.

X-ray crystal

Single crystals suitable for X-ray diffraction were selected and mounted in inert oil in cold gas stream and their X-ray diffraction intensity data was collected on a Rigaku XtaLAB Synergy diffractometer equipped with a Hypix6000HE detector, using Mo K α radiation ($\lambda = 0.7107$ Å).

Abbreviations

UPy = 2-ureido-4[1H]-pyrimidinone B18C6 = benzo-18-crown-6 DMSO = dimethyl sulfoxide M = mol/L br = broad Ar = aromatic group

2. Concentration-dependent ¹H NMR



Fig. S1. ¹H NMR spectra (300 MHz, CDCl₃/CD₃CN = 1:1, v/v, 298 K) of **M1** at different monomer concentrations. The asterisk symbols indicate solvent peaks.



Fig. S2. ¹H NMR spectra (300 MHz, $CDCl_3/CD_3CN = 1:1$, v/v, 298 K) of **M1** with equal molar of KPF₆ at different monomer concentrations. The asterisk symbols indicate solvent peaks.



Fig. S3. Concentration of cyclic monomers in CDCl₃/CD₃CN=1/1 (v/v) solutions versus increasing concentration of monomer **M1**, calculated from the ¹H NMR spectra. H₃ in **M1** was used for calculation.

3. DOSY experiments



Fig. S4. Two DOSY spectra (600 MHz, CDCl₃/CD₃CN = 1:1, v/v, 298 K) of (a) $M2 \supset K^+$ complex (16 mM) with the addition of permethylated β -CD as the internal standard and (b) $M1 \supset K^+$ complex (8 mM) with the addition of $N1^{[S1]}$ as the internal standard. Chemical structures of (c) permethylated β -CD and (d) a highly stable cyclic monomer of N1. The red peaks stand for proton signals from permethylated β -CD and the blue peaks belong to M2. The magenta peaks stand for peaks belonging to M1 and the green peaks represent peaks from N1. Asterisk symbols represent solvent peaks.

By comparing the diffusion coefficients of a smaller methyl-substituted analogue $(M2 \supset K^+)_n$ (when n = 1, MW = 626 Da, 1.15×10^{-9} m² s⁻¹) and permethylated β -CD (6.76×10^{-10} m² s⁻¹, Figure S4a), as well as $(M1 \supset K^+)_n$ (7.94×10^{-10} m² s⁻¹, at 8 mM) and a highly stable UPy-based cyclic monomer N1^[S1] (1.02×10^{-9} m² s⁻¹, MW = 804 Da, Figure S4b), it can be concluded that K⁺ templates the formation of a dimeric species ($M1 \supset K^+$)₂ (MW = 1588 Da).

Moreover, by employing the Stokes-Einstein equation ($R_h = k_B T/(6\pi\eta D)$), the diffusion coefficients that were experimentally obtained could be converted to an approximate hydrodynamic radius (or Stokes radius).^[S4] k_B is the Boltzmann's constant and T is the temperature. The viscosity coefficient η of a mixed solution of CHCl₃ and CH₃CN (v/v = 1/1) is obtained from literature report ($\eta = 0.22$ cP).^[S5] Taking a solution of **M2** (8 mM, CHCl₃/ CH₃CN, v/v = 1/1) for example, R_h can be calculated by the following equation:

$$R_h = \frac{k_B T}{6\pi\eta D} = \frac{1.38 \times 10^{-23} \times 298}{6 \times 3.14 \times 0.22 \times 10^{-3} \times 1.15 \times 10^{-9}} = 0.86 \, nm$$

We have obtained a crystal structure of $M3 \supset K^+$ (vide infra). Inspired by this structure, we prepared a molecular model of $(M2 \supset K^+)_2$ (Figure S4). According to Stokes–Einstein equation assuming a spherical molecule, we measured the longest distance (diameter) in $M2 \supset K^+$ dimer by simplifying the supramolecular dimer to a geometric sphere and a spherical radius of 0.85 nm (Figure S5, $r = 16.963 \text{ Å} \div 2 = 0.85 \text{ nm}$) was shown, which was in good agreement with the experimental value.



Fig. S5. A molecular model of M2



Fig. S6. DOSY spectrum (600 MHz, CDCl₃, 298 K) of M1 in 4 mM.

By using the Stokes-Einstein equation (η of CHCl₃ is 0.54 cP), the size of cyclic monomer was estimated as follows:

$$R_h = \frac{k_B T}{6\pi\eta D} = \frac{1.38 \times 10^{-23} \times 298}{6 \times 3.14 \times 0.54 \times 10^{-3} \times 1.15 \times 10^{-9}} = 0.46 \ nm$$

which shows that the hydrodynamic radius of cyclic monomer was nearly half of the cyclic dimer (0.86 nm).



Fig. S7. DOSY spectrum (600 MHz, CDCl₃, 298 K) of M1 in 128 mM.

Furthermore, the size of supramolecular polymer at 128 mM ($D = 7.58 \times 10^{-11} \text{ m}^2 \text{ s}^{-1}$) was estimated as follows:

$$R_h = \frac{k_B T}{6\pi\eta D} = \frac{1.38 \times 10^{-23} \times 298}{6 \times 3.14 \times 0.22 \times 10^{-3} \times 7.58 \times 10^{-11}} = 5.3 \text{ nm}$$

4. Mass spectrometry of M1+KPF₆



Fig. S8. HR-MS (ESI, positive mode, $CHCl_3/CH_3CN = 1:5$, v/v) of $M3 + KPF_6$

The formation of dimer was also confirmed by high-resolution ESI-MS, with the signals being clearly observed at m/z = 793.3996, which correspond to $[2M1 + 2K]^{2+}$ (calculated as 793.4009). And m/z = 1731.7651, which correspond to $[2M1 + KPF_6 + K]^+$ (calculated as 1731.7666). Other peaks: m/z calcd for $[M1 + H]^+ = 755.4456$, found = 755.4437; $[2M1 + K]^+ = 1547.8387$, found = 1547.8357.

5. Crystallographic data for $M3 \supset K^+$



CCDC number	1962895
Empirical formula	$C_{64}H_{92}F_{12}K_2N_{16}O_{16}P_2$
Formula weight / g mol ⁻¹	1709.67
Temperature / K	159.99(10)
Crystal system	monoclinic
Space group	P2 ₁ /n
<i>a</i> / Å	14.5377(3)
b / Å	16.9875(3)
<i>c</i> / Å	31.4097(5)
$\alpha / ^{\circ}$	90
β / °	92.689(2)
γ∕°	90
Volume / Å ³	7748.4(2)
Ζ	4
$ ho_{ m calc}$ / g cm ⁻³	1.456
μ / mm ⁻¹	0.266
F/000	3537.0
Crystal size / mm ³	0.25 imes 0.2 imes 0.2
Index ranges	$-18 \le h \le 17, -22 \le k \le 22, -32 \le l \le 39$
Reflections collected	89281
Independent reflections	18990 [$R_{int} = 0.0292$, $R_{sigma} = 0.0285$]
Data/restraints/parameters	18990/36/1102
Goodness-of-fit on F^2	1.087
Final <i>R</i> indices [<i>I</i> > 2sigma(<i>I</i>)]	$R_1 = 0.0612, wR_2 = 0.1874$
<i>R</i> indices (all data)	$R_1 = 0.0887, wR_2 = 0.2077$
Largest diff. peak / hole / e·Å ⁻³	0.73/-0.64

Table S1 Crystal data and structure refinement parameter for M3

6. The complexation of K^+ with M4



Fig. S9. ¹H NMR spectra (300 MHz, $CDCl_3/CD_3CN = 1/1$, v/v, 298 K) of (a) M4 + KPF₆ and (b) M4. Complexed species are labeled in pink.

B18C6 (e) (d) (c) (b) (a) 15.5 14.5 13.5 11.5 8.5 7.5 6.5 5.5 12.5 10.5 9.5 δ/ppm

7. Topological transformation monitored by ¹H NMR

Fig. S10. Partial ¹H NMR spectra (300 MHz, $CDCl_3/CD_3CN = 1/1$, v/v, 298 K) of (a) **M1** (16 mM), (b) after addition of 1 equiv. KPF₆ (16 mM), (c) after addition of 1 equiv. B18C6 (16 mM), (d) after addition of 1 equiv. KPF₆ (16 mM), and (e) after addition of 1 equiv. B18C6 (16 mM). The asterisk symbols indicate solvent peaks and the red dot stand for another proton signals in B18C6.

Firstly, the reversible topological transformation was realized by benzo-18-crown-6 (B18C6). The process was monitored by ¹H NMR. A solution of **M1** (CDCl₃/CD₃CN = 1/1, v/v, 16 mM) at lower concentration was prepared, in which cyclic monomer and linear supramolecular polymer co-existed. Thus, two sets of peaks were appeared (Figure S8a). After the addition of 1 equiv. of KPF₆, there are only discrete cyclic dimers, resulting in one set of sharp peaks in ¹H NMR (Figure S8b). After the addition of 1 equiv. B18C6, these discrete cyclic dimers were transformed back into cyclic monomer and linear supramolecular polymer, resulting in the recovery of two sets of peaks in ¹H NMR (Figure S8c). This process can be repeated again (Figure S8d and 8e).



Fig. S11. ¹H NMR spectra (300 MHz, $CDCl_3/CD_3CN = 1:1$, v/v, 298 K) of (a) **M1** (128 mM), (b) after addition of 1 equiv. KPF₆, (c) after washing with water and supplementing with CD₃CN, (d) repeating the process of (b), and (e) repeating the process of (c).

Secondly, the reversible topological transformation was realized by water extraction. The process was also monitored by ¹H NMR. A solution of **M1** (0.5 mL, CDCl₃/CD₃CN = 1/1, v/v, 128 mM) at a relatively high concentration was prepared, in which linear supramolecular polymers were predominant. Thus, only one set of broad peaks was appeared (Figure S9a). After the addition of 1 equiv. of KPF₆, the linear supramolecular polymers were transformed into discrete cyclic dimers, resulting in one set of sharp peaks and changes of chemical shifts in ¹H NMR (Figure S9b). After washing with water (0.5 mL × 2), K⁺ ions were extracted into water phase and linear supramolecular polymers were recovered in organic solution, resulting in the recovery of one set of broad peaks in ¹H NMR (Figure S9c). Herein, CD₃CN was appropriately supplemented because part of the CD₃CN was also extracted into the aqueous phase. This reversible transformation processes can be repeated again (Figure S9d and 9e).

8. Synthetic procedures and characterizations



Scheme S1 Synthetic route of M1-M4

8.1 Synthesis of Compound M1

Imidazolide **2a** (0.94 g, 3.09 mmol) and **1a** (0.40 g, 1.41 mmol) were dissolved in dry CHCl₃ (30 mL) and this solution was stirred for 12 h under nitrogen at room temperature. To the reaction mixture CHCl₃ (20 mL) was added and the organic layer was washed with 1 M HCl (50 mL), saturated NaHCO₃ (50 mL), brine (50 mL), dried over anhydrous MgSO₄ and concentrated under reduced pressure. The resulting residue was subjected to column chromatography over silica gel (eluent: CHCl₃/MeOH = 100:1, v/v) to afford compound **M1** as a colorless viscous solid (0.90 g, 85%). ¹H NMR (300 MHz, CDCl₃): δ 13.16 (br s, 2H, N*H*), 11.98 (br s, 2H, N*H*), 10.34 (br s, 2H, N*H*), 6.86 (d, *J* = 18.6 Hz, 4H, Ar-*H*), 6.14-5.35 (m, 2H, alkylidene-*H*), 4.17 (m, 4H, OC*H*₂), 4.06-3.78 (m, 4H, OC*H*₂), 1.03-0.68 (m, 12H, C*H*₃) ppm. ¹³C NMR (75 MHz, DMSO-*d*₆): δ 170.7, 161.1, 154.4, 151.6, 148.4, 121.3, 114.4, 105.1, 69.4, 68.9, 68.3, 47.9, 39.2, 32.9, 29.1, 26.5, 22.2, 13.9, 11.8 ppm. ESI-MS (C₃₈H₅₈N₈O₈): *m/z* calcd for [M + H]⁺ = 755.45, found = 755.35 (81%); [M + Na]⁺ = 777.43, found = 777.30 (100%); HR-ESI-MS (C₃₈H₅₈N₈O₈): *m/z* calcd for [M + H]⁺ = 755.4450, found = 755.4441.



Fig. S13. ¹³C NMR (75 MHz, DMSO-*d*₆) spectrum for M1



Fig. S14. Electrospray ionization mass spectrum of M1

8.2 Synthesis of Compound M2

Imidazolide **2b** (0.61 g, 2.79 mmol) and compound **1a** (0.36 g, 1.27 mmol) were dissolved in dry CHCl₃ (30 mL) and this solution was stirred for 12 h under nitrogen at room temperature. To the reaction mixture CHCl₃ (20 mL) was added and the organic layer was washed with 1 M HCl (50 mL), saturated NaHCO₃ (50 mL), brine (50 mL), dried over anhydrous MgSO₄ and concentrated under reduced pressure. The resulting residue was subjected to column chromatography over silica gel (eluent: CHCl₃/MeOH = 100:1, v/v) to afford compound **M2** as a white solid (0.60 g, 81%). m.p.: 235-237 °C. ¹H NMR (300 MHz, DMSO-*d*₆): δ [ppm] = 11.50 (br s, 2H, N*H*), 7.05 (br s, 2H, N*H*), 7.01-6.92 (m, 2H, Ar-*H*), 6.90-6.83 (m, 2H, Ar-*H*), 5.75(s, 2H, alkylidene-*H*), 4.10-4.07 (m, 4H, OC*H*₂), 3.76-3.73 (m, 4H, OC*H*₂), 3.59-3.55 (t, *J* = 5.4 Hz, 4H, OC*H*₂), 3.35-3.32 (m, 4H, OC*H*₂), 2.07 (s, 6H, C*H*₃). ¹³C NMR (75 MHz, DMSO-*d*₆): δ [ppm] = 179.3, 161.8, 155.0, 151.5, 148.5, 121.4, 114.4, 104.7, 69.4, 69.0, 68.4, 39.2, 23.1. ESI-MS (C₂₆H₃₄N₈O₈): *m/z* calcd for [M + H]⁺ = 587.60, found = 587.09 (46%); [M + Na]⁺ = 609.59, found = 609.05 (100%); HR-ESI-MS (C₂₆H₃₄N₈O₈): *m/z* calcd for [M + H]⁺ = 587.2572, found = 587.2511; [M + Na]⁺ = 609.2392, found = 609.2391.



Fig. S16. ¹³C NMR (75 MHz, DMSO-*d*₆) spectrum for M2



Fig. S17. Electrospray ionization mass spectrum (CHCl₃/CH₃CN 1:5) of M2

8.3 Synthesis of Compound M3

Imidazolide **2c** (0.20 g, 0.78 mmol) and compound **1a** (0.10 g, 0.35 mmol) were dissolved in dry CHCl₃ (15 mL) and this solution was stirred for 12 h under nitrogen at room temperature. To the reaction mixture CHCl₃ (15 mL) was added and the organic layer was washed with 1 M HCl (30 mL), saturated NaHCO₃ (30 mL), brine (30 mL), dried over anhydrous MgSO₄ and concentrated under reduced pressure. The resulting residue was subjected to column chromatography over silica gel (eluent: CHCl₃/MeOH = 100:1, v/v) to afford compound **M3** as a white solid (0.18 g, 75%). m.p.: 148-150 °C. ¹H NMR (300 MHz, DMSO-*d*₆): δ [ppm] = 11.43 (br s, 2H, N*H*), 9.70 (br s, 2H, N*H*), 7.60 (br s, 2H, N*H*), 6.98-6.93 (m, 2H, Ar-*H*), 6.89-6.84 (m, 2H, Ar-*H*), 5.75(s, 2H, alkylidene-*H*), 4.13-4.07 (m, 8H, OC*H*₂), 3.76-3.73 (m, 4H, OC*H*₂), 3.57 (t, *J* = 5.3 Hz, 4H, OC*H*₂), 3.35-3.32 (m, 4H, OC*H*₂), 2.33 (t, *J* = 7.5 Hz, 4H, CH₂CH₂CH₂CH₃CH₃, 1.57-1.47 (m, 4H, CH₂CH₂CH₂CH₃), 1.33-1.20 (m, 4H, CH₂CH₂CH₂CH₃), 0.86 (t, *J* = 7.3 Hz, 6H, CH₂CH₂CH₂CH₃). ¹³C NMR (75 MHz, CDCl₃): δ [ppm] = 173.0, 157.0, 154.7, 152.3, 149.3, 121.6, 114.9, 105.7, 69.8, 69.6, 69.3, 39.8, 32.5, 29.0, 22.2, 13.8. ESI-MS (C₃₂H₄₆N₈O₈): *m/z* calcd for [M + H]⁺ = 671.76, found = 671.10 (36%); [M + Na]⁺ = 693.3331, found = 693.035.



Fig. S19. ¹³C NMR (75 MHz, CDCl₃) spectrum for M3



Fig. S20. HR-MS (ESI, positive mode, CHCl₃/CH₃CN = 1:5, v/v) of M3

8.4 Synthesis of Compound M4

Imidazolide **2a** (1.79 g, 5.9 mmol) and **1b** (1.00 g, 2.7 mmol) were dissolved in dry CHCl₃ (30 mL) and this solution was stirred for 12 h under nitrogen at room temperature. To the reaction mixture CHCl₃ (20 mL) was added and the organic layer was washed with 1 M HCl (50 mL), saturated NaHCO₃ (50 mL), brine (50 mL), dried over anhydrous MgSO₄ and concentrated under reduced pressure. The resulting residue was subjected to column chromatography over silica gel (eluent: CHCl₃/MeOH = 100:1, v/v) to afford compound **M4** as a colorless viscous solid (1.76 g, 77%). ¹H NMR (300 MHz, CDCl₃): δ 13.17 (br s, 2H, N*H*), 11.97 (br s, 2H, N*H*), 10.32 (br s, 2H, N*H*), 6.84 (d, *J* = 33.5 Hz, 4H, Ar-*H*), 5.79 (s, 2H, alkylidene-*H*), 4.21-3.98 (m, 4H, OC*H*₂), 3.92-3.80 (m, 4H, OC*H*₂), 3.77-3.61 (m, 12H, OC*H*₂), 3.56-3.42 (m, 4H, OC*H*₂), 2.35-2.20 (m, 2H, C*H*(CH₂)₂), 1.70-1.46 (m, 8H, C*H*₂), 1.33-1.25 (m, 8H, C*H*₂), 0.90-0.84 (m, 12H, C*H*₃) ppm. ¹³C NMR (75 MHz, CDCl₃): δ 173.0, 157.0, 155.4, 154.8, 149.0, 121.6, 115.0, 106.3, 70.8, 70.4, 69.8, 69.5, 68.8, 45.3, 39.5, 32.9, 29.3, 26.6, 22.5, 13.9, 11.7 ppm. ESI-MS (C4₂H₆6N₈O₁₀): *m/z* calcd for [M - H]⁺ = 841.48, found = 841.15; [M + Na]⁺ = 865.48, found = 865.20 (100%); HR-ESI-MS (C4₂H₆6N₈O₁₀): *m/z* calcd for [M - H]⁻ = 841.48, found = 841.15; [M + Na]⁺ = 843.4980, found = 843.4966.





Fig. S22. ¹³C NMR (75 MHz, CDCl₃) spectrum for M4





Line#:2 R.Time:2.200(Scan#:133) MassPeaks:456 Spectrum Mode:Averaged 2.167-2.233(131-135) Base Peak:865.20(282696) BG Mode:Calc Segment 1 - Event 1



Fig. S23. Electrospray ionization mass spectrum (CHCl₃/CH₃CN 1:5) of M4

9. References

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