Supporting Information for:

A Bench-Stable Copper Photocatalyst for the Rapid Hydrophosphination of Activated and Unactivated Alkenes

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General Information

All manipulations were performed under a nitrogen atmosphere with dry, oxygen-free solvents using an M. Braun glovebox or standard Schlenk techniques unless otherwise noted. $CDCl_3$ was purchased and then degassed and dried over calcium hydride and then distilled and stored over 3 and 4 Å molecular sieves. Diphenylphosphine was synthesized according to literature procedures and stored under an inert atmosphere of N₂ prior to use.¹ Compounds **10**² was synthesized according to literature procedures. All other reagents were acquired from commercial sources and dried by conventional means, as necessary. ESI-mass spectra were collected on an AB-Sciex 4000 QTrap Hybrid Triple Quadrupole/Linear Ion trap mass spectrometer. ¹H, ²H, ¹³C, ³¹P, ³¹P{¹H} and ³¹P HMBC NMR spectra were recorded at 25 °C with a Bruker AXR 500 MHz or Varian 500 MHz spectrometer.

Safety note: Phosphines are toxic, and many primary phosphines are volatile. Redox reactions or metal catalysts have the potential for rapid hydrogen evolution from phosphine substrates. Appropriate precautions for toxic gases and hydrogen are necessary in this work.

Resonances in ¹H NMR spectra are referenced to the residual solvent resonance (CDCl₃ = δ 7.28) or tetramethylsilane (TMS, δ 0.0). Reported ³¹P{¹H} NMR resonances are relative to external 85% H₃PO₄. Spectral data for hydrophosphination products is consistent with literature reports.^{1, 3-10} Previously unreported products were purified by silica gel column chromatography silica gel column chromatography with a 3:1 hexanes: DCM liquid phase in an M. Braun glovebox.

 $CDCl_3$ is not visible in many of the spectra due to overlap with aromatic resonances of starting materials and products. The signal for acetylacetone is present in all catalytic reactions with **1** and is denoted by (\dagger) in the first catalytic hydrophosphination spectra of styrene and phenylphosphine in this document and then is not labeled in subsequent spectra. P-H bonds of some secondary phosphines are not visible in the crude spectra due to interaction with copper, but peaks become visible with the removal of copper by silica gel chromatography. Starting phosphines PhPH₂ and Ph₂PH signals are at approximately - 122 and - 40 ppm in the ³¹P{¹H} NMR spectra and are not labeled.

Single addition hydrophosphination product	*
Double addition hydrophosphination product	**
Ph ₂ P-PPh ₂	†
PhHP-PHPh	††
Acetylacetone	ŧ
Internal standard of 1,3,5-trimethoxybenzene, ferrocene or tetramethylsilane	TMB, Fc, TMS (not labeled in all spectra; exists at $\delta 0$)
External standard of PPh ₃	ES or E.S. PPh ₃

Table S1. Symbols for products in spectra:

Aromatic peaks and most overlapping peaks are not indicated

General Procedure for catalytic experiments

In an N₂ filled dry box, 0.76 mmol of phosphine (or 0.38 mmol where applicable) and 0.38 mmol of unsaturated substrate (and internal or external standard where applicable) were measured and mixed in 0.6 mL CDCl₃. This solution was either pipetted into a shell vial containing 0.019 mmol of catalyst in 100 µL of CDCl₃ or catalyst was added via stock solution. The resulting solution was quickly transferred to either an aluminum foil wrapped [-Young type polytetrafluoroethylene-valved NMR tube or an NMR tube with a disposable NMR cap which was subsequently wrapped with parafilm. Neat reactions were added directly to a J-Young type NMR tube. For reactions containing an external standard, a sealed capillary containing 5 mg of PPh₃ was added to the NMR tube before capping. Upon removal from dry box, the outside of the tube was washed with a 50:50 water: bleach to remove trace amounts of malodorous phosphine on the outside of the tube. An initial ¹H and ³¹P{¹H} NMR spectra was then obtained as soon as possible. After the initial NMR spectra, reactions were removed from the aluminum foil and placed in a photoreactor containing a Rexim G23 UV-A (9W) lamp (unless otherwise noted) at ambient temperature and shielded from external light. The temperature of the 360 nm photoreactor was measured to be 25-30 °C, depending on how long it had been in use. No efforts to control the temperature between this range were undertaken. Periodic ¹H and ³¹P{¹H} NMR spectra were collected. Conversions were determined by integration of ¹H and ³¹P{¹H} NMR spectra to starting materials and in many cases (see note below) confirmed by internal standard of either 1,3,5-trimethoxy benzene,¹¹ tetramethyl silane, or an external standard of PPh₃.

Note on the determination of conversions

Initial tests of reactivity were performed on all substrates, and conversions were determined by integration of product signals to starting material in the ¹H and ³¹P{¹H} NMR spectra. There was good agreement between ¹H and ³¹P{¹H} NMR spectra, and this method has precedent in catalytic hydrophosphination. ^{5, 6, 12, 13, 14, 15, 16, 17, 18}. To ensure the accuracy of this method, reactions from each class of substrates (styrenes, unactivated alkenes, alkynes, primary phosphines, secondary phosphines, etc.) were replicated with an internal standard of 1,3,5-trimethoxybenzene,¹⁹ or TMS for ¹H NMR spectra or an external standard (sealed capillary) of PPh₃ For ³¹P{¹H} NMR spectra.

For reactions between phenylphosphine and styrene, integration of product signals relative to a 1,3,5-trimethoxybenzene internal standard resulted in the same yield as integrating the starting materials to the products. Integration to an internal standard of TMS gave conversion *greater than 100%*. Thus, we had more confidence in the more conservative estimate of conversion obtained by our initial method.

Integration of product signals relative to a 1,3,5-trimethoxybenzene internal standard resulted in comparable conversions for the products of hydrophosphination of phenylphosphine with *p*-methylstyrene, and 1-hexene. At that point we were satisfied that our initial method of determining yield was satisfactory for primary phosphines and alkenes.

For reactions between diphenylphosphine and *p*-tertbutyl styrene there was good agreement our initial trial and a trial with 1,3,5-trimethoxybenzene. However, there was some discrepancy between the two methods in reaction between styrene and diphenylphosphine. There is some precedent for inconsistent behavior of styrene in hydrophosphination reactions.³ Conversions determination by integration relative to the 1,3,5-trimethoxybenzene internal standard results in a *higher* and *less* accurate yield than by relative integration. It appears that the internal standard interacts with copper and the two ¹H NMR signals of 1,3,5-trimethoxybenzene did not integrate in the correct 3:1 ratio and exhibited splitting (see figure 1 below), and internal standard protons exhibited splitting based on interaction with phosphorus in a ³¹P HMBC spectrum. This is surprising and not yet fully understood.

Finally, ferrocene was used as a standard. There was still variability in the results, and ferrocene appeared to inhibit the catalysis.

Overall, we have sought the most accurate conversion data, which was not merely a function of a single integration standard. As needed, we have often reported lower conversion based on mitigating factors with various standards.



Figure S.1. Splitting of 1,3,5-trimethoxy benzene protons observed in ¹H NMR spectra of a catalytic reaction of styrene and phenylphosphine.



Figure S.2. Spectral distribution of Rexim G23 UV-A (9W) lamp as provided by manufacturer.

Photographs of the photoreactor



Figure S.3. Outside of the photoreactor containing a UV-A lamp



Figure S.4. Photograph of the inside of the photoreactor containing a UV-A lamp

Table S2.

S6

Ĺ	+ 2 PhPH ₂ <u>5 mol % of 1</u> <u>360 nm</u> <u>25 - 30 °C</u> <u>CDCl₃</u> <u>20 m</u>	Ph Ph H
Entry	Deviation from standard conditions	Conversion (%)
1 2	None 1 eg PhPH2	100 51
3	Control; without 1 , 360 nm	6
4	Ambient light, 60 °C	59
5	Visible Light LED	45
6	Ambient light	15
7	Dark	7
8	2 mol % of 1	100
9	1 mol % of 1	90
10	25 mol % of 1	40
11	Neat	100
12	Benzene-d ₆ solution	100

Standard condition: 0.38 mmol (39.5 mg) of styrene and 0.76 mmol phenylphosphine (83.5 mg) and 0.19 mmol (5 mg) of **1** in 600 μ L of CDCl₃. Changes in catalyst loading and phenylphosphine equivalents are with respect to styrene. Conversion to both secondary and tertiary phosphine determined by integration of ¹H and ³¹P{¹H} NMR spectra. The LED light bulb is a visible light 9 W, 800 Lumens 120 V, 60 Hz, 150 mA bulb





Note: The LED light bulb is a visible light 9 W, 800 Lumens 120 V, 60 Hz, 150 mA bulb

TABLE S3. $R = R' + PhR^{1}PH \xrightarrow{5 \text{ mol } \% \text{ of } 1}{25 - 30 \degree C} R' \xrightarrow{PPhR^{1}}{R'}$ $R' = R' + PhR^{1}PH \xrightarrow{25 - 30 \degree C}{CDCl_{3}} R'$				hR ¹		
entry	R	R'	RR'PH	Major Productª	Conversion % ^b	Z:E
1	Ph	Et	PhPH ₂	Et	80	3:1
2	Ph	Et	2 PhPH ₂	Ph PPhH	99	2.45:1
3	Et	Et	PhPH ₂	EtEt	48	1:1.7
4	Et	Et	2 PhPH ₂	PPhH	71	1:1.5
5	Ph	Ph	PhPH ₂	Ph	40	1.9:1
6	Ph	Ph	2 PhPH ₂	Ph PPhH	58	2.1:1
7	Ph	Н	2 PhPH ₂	P →	86 ^{c, d}	3:1e
8	Ph	Н	$0.5 PhPH_2$	Ph Ph Ph	88c, f	2.7:1e
9	Ph	Et	Ph ₂ PH	Ph PPh ₂	74	4.8:1
10	Ph	Н	Ph2PH		41	32:1
11	Ph	Н	2 Ph ₂ PH	Ph PPh ₂	77	37.5:1
12	Ph	Ph	Ph ₂ PH	Ph	12	1:0
13g	Ph	Ph	Ph ₂ PH		11	10:1
14	Ph	Ph	2 Ph ₂ PH	Pn PPn ₂	19 ^d (36 ^h)	35:1 ^h
Conditions: 0.38 mmol of alkyne and 0.38 or 0.76 mmol of PhPH ₂ or Ph ₂ PH and 0.019 mmol 1 [a] only the major stereoisomer is depicted [b] conversion to anti-markovnikov vinyl phosphine as determined by integration of ¹ H and ³¹ P{ ¹ H}MR spectra [c] double P-H bond activation product [d] 20 h [e] ratio of $Z = T$ product [f] 1 h [g] 5 mol % of PPh ₂ added [h] 72 h						
$\cup I L, L: E, L$	01 Δ ₁ Δ1Δ1 ₂ μισααίς [1] 1 11 [8] 5 11101 70 01 1 113 auαcu [11] 7 Δ 11					

S8 * = hydrophosphination product, ** = double addition hydrophosphination product, † or †† = Ph_2P-PPh_2 or PhPH-PhPH, \ddagger = acetylacetone, ES = PPh₃ external standard, TMB = 1,3,5-trimethoxy benzene, TMS = tetramethyl silane

Reactions of $\mathbf{1}$ with Ph₂PH

In an N₂ filled dry box, 0.38 mmol (70.7 mg) of diphenylphosphine, 0.019 mmol (5 mg) of **1**, 6.6 mg (0.039 mmol) of 1,3,5-trimethoxybenzene as an internal standard, and a closed capillary containing 5 mg of PPh₃ in 50 μ L of CDCl₃ as an external standard were measured and mixed in 0.8 mL in a J-Young type polytetrafluoroethylene-valved NMR tube wrapped in aluminum foil until an initial ¹H and ³¹P{¹H} NMR spectra was obtained. The reaction was then placed under 360 nm irradiation and monitored periodically by ¹H and ³¹P{¹H} NMR spectroscopy.



Figure S.6. Reaction of Cu(acac)₂ and 20 equiv. of Ph₂PH

Reaction of 1 with 20 equiv. of Ph₂PH - replication in toluene

In an N₂ filled dry box, 0.38 mmol (70.7 mg) of diphenylphosphine, 0.019 mmol (5 mg) of **1**, in 500 μ L of Toluene in 0.8 mL in a J-Young type polytetrafluoroethylene-valved NMR tube a ³¹P{¹H} NMR spectra was obtained.

These spectra provide evidence for a phosphido species of unknown speciation. A singlet integrating to 5% of the phosphorus in solution at -31.9 in addition to the free Ph₂PH. The signal remained a singlet in the coupled ³¹P NMR and is coupled only to aromatics in a ³¹P HMBC spectra. Our attempts at crystalizing a discreet intermediate from these solutions resulted only in powders that were insoluble in common organic solvents.



Figure S.8. ³¹P{¹H} NMR spectra (toluene, 202 MHz)

S10

* = hydrophosphination product, ** = double addition hydrophosphination product, † or †† = Ph_2P-PPh_2 or PhPH-PhPH, \ddagger = acetylacetone, ES = PPh₃ external standard, TMB = 1,3,5-trimethoxy benzene, TMS = tetramethyl silane Reaction of 1 with 20 equiv. of Ph₂PH - replication in toluene (continued)



Figure S.9. ³¹P HMBC NMR spectra (toluene d₈, 202 MHz)

Treatment of 1 with 1, 2, 4, 8, and 20 equiv. of Ph₂PH

In an N₂ filled dry box, 15 mg (0.057 mmol) of **1** was measured and mixed in \sim 0.6 mL CDCl₃ and transferred to a J-Young type polytetrafluoroethylene-valved NMR tube or an NMR tube. For the reaction with 20 equiv. of Ph₂PH only 5 mg of **1** was used. The corresponding equiv. of Ph₂PH were added by micropipette to the NMR tube and an initial ¹H and ³¹P{¹H} NMR spectra was obtained.

Caulton reported [(Ph₂PH)CuPPh₂]_n from the addition of multiple equivalents of Ph₂PH to Cu(O^tBu)₄.²⁰ Compound **1**, when treated with 2, 4, 8 and 20 equiv. of Ph₂PH in CDCl₃ solution, provided spectroscopic data consistent with observations made of [(Ph₂PH)CuPPh₂]_n. When compound **1** was treated with two equivalents of Ph₂PH a doublet with *J*_{PH} = 308 Hz was observed in the ¹H NMR spectra, which compares favorably to the literature report of 300 Hz reported by Caulton with Cu(O^tBu)₄. With both compounds, the doublet splitting decreases toward the value of free Ph₂PH (219 Hz) as additional Ph₂PH is present in solution. An additional similarity with the literature report was observed in the ³¹P{¹H} NMR spectra where the signal was shifted downfield when one equivalent of diphenylphosphine was added and the signal shifted back upfield toward the value of free Ph₂PH, coordinated Ph₂PH, and likely a phosphido species and is a weighted average of the shifts and coupling constants.



Figure S.10. Stacked ¹H NMR spectra (CDCl₃, 500 MHz)

S12 * = hydrophosphination product, ** = double addition hydrophosphination product, \dagger or $\dagger \dagger = Ph_2P-PPh_2$ or PhPH-PhPH, $\ddagger = acetylacetone$, ES = PPh₃ external standard, TMB = 1,3,5-trimethoxy benzene, TMS = tetramethyl silane

Treatment of 1 with 1, 2, 4, 8, and 20 equiv. of Ph₂PH (continued)



Figure S.11. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)

Hydrophosphination of styrene Eqn (1)

Prepared in accordance with the general procedure with two equiv. of phenylphosphine. In an N₂ filled dry box, 0.76 mmol of phenylphosphine (83.5 μ L, 83.5 mg) and 0.38 mmol of styrene (39.5 mg) were mixed in 0.5 mL ofCDCl₃. This solution was then pipetted into a 4 ml shell vial containing 5 mg (0.0019 mmol) **1** in 100 μ L CDCl₃. The resulting yellow solution was quickly transferred to an NMR tube, capped with a disposable NMR cap, parafilm was wrapped around the NMR cap, and the reaction was shielded from ambient light. After initial ¹H and ³¹P{¹H} NMR spectra were obtained, the reaction was irradiated with 360 nm light and monitored every 10 min. The yellow solution faded over time to a clear solution. After 10 min of irradiation both the ¹H and ³¹P{¹H} NMR spectra indicated 74% yield of secondary phosphine. After 20 min of irradiation, 100% of styrene was converted with 98:4:2 selectivity for the secondary hydrophosphination product compared to the PhHP-PHPh and tertiary products as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Together, these indicate a 96% conversion of styrene to secondary phosphine phenyl(2-phenylethyl)phosphine (PhPHCH₂CH₂Ph).

Two subsequent trials conducted with an internal standard of 1,3,5-trimethoxybenzene resulted in 95% conversion and 87% conversion to secondary phosphine product PhPHCH₂CH₂Ph.

³¹P{¹H} NMR signals for the products in the crude reaction mixtures are broad due to interaction with paramagnetic copper and sharpen upon removal of copper by filtering the crude residue through a plug of silica and evaporation of the volatile species by vacuum. Separation of secondary, tertiary and dehydrocoupled products can be achieved by silica gel column chromatography with a 3:1 hexanes: DCM liquid phase. 54 mg (0.25 mmol, 66% yield) of phenyl(2-phenylethyl)phosphine (PhPHCH₂CH₂Ph) was isolated. Spectra are consistent with previous reports.⁶



S14

* = hydrophosphination product, ** = double addition hydrophosphination product, \dagger or $\dagger \dagger = Ph_2P-PPh_2$ or PhPH-PhPH, $\dagger = acetylacetone$, ES = PPh₃ external standard, TMB = 1,3,5-trimethoxy benzene, TMS = tetramethyl silane



Hydrophosphination of styrene with two equiv. of phenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)



Hydrophosphination of styrene with two equiv. of phenylphosphine by 5 mol % of 1 under 360 nm irradiation (continued)

Figure S.14. ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz) at t = 20 min of 360 nm irradiation

Hydrophosphination of styrene with two equiv. of phenylphosphine by 5 mol % of 1 under 360 nm irradiation (Trial 2)

SD-4-15-IS



Figure S.15. ¹H NMR spectrum (CDCl₃, 500 MHz) at t = 20 min of 360 nm irradiation

Internal standard yield calculation: (1.99/(1.06/3)) * (0.0108 g/168.19 g/mol) / 0.38 mmol = 95% yield of secondary phosphine

Hydrophosphination of styrene with two equiv. of phenylphosphine with 5 mol % of **1** under 360 nm irradiation (continued)

SD-4-16-F7P



Figure S.16. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz) of three isolated fractions from trial 3. 0.54 grams (0.25 mmol) phenyl(2-phenylethyl)phosphine was recovered from fractions 5,6 and 7. Yield calculation: 35.5 mg + 15.3 mg + 3.2 mg = (0.054 g / 214.25 g/mol)/0.00038 mol = 66% yield



Hydrophosphination of styrene with one equiv. of diphenylphosphine by ${\bf 1}$ Eqn (2)

Prepared according to the general procedure with one equiv. of diphenylphosphine and 11.5 mg (0.068 mmol) of 1,3,5- trimethoxybenzene an internal standard. 100% conversion of styrene was measure after 5 h of 360 nm irradiation as measured by 1 H and $^{31}P{^{1}H}$ NMR spectroscopy. Spectra is in accordance with previous reports.²¹



Hydrophosphination of styrene with one equiv. of diphenyl phosphine by 5 mol % of **1** under 360 nm irradiation (continued)





Hydrophosphination of *p*-methylstyrene Eqn (1)

Prepared in accordance with the general procedure with two equiv. of phenylphosphine. After 25 min of 360 nm irradiation, 100% of *p*-methylstyrene was converted with 98:2:6 selectivity for the secondary hydrophosphination product compared to the tertiary product and dehydrocoupled product as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Together, these indicate a 96% conversion of styrene to secondary phosphine. Subsequent trials with an internal standard of 1,3,5-trimethoxybenzene added after 20 min confirm a greater than 90% conversion. Spectra are consistent with previous reports.⁶







Hydrophosphination of p-methylstyrene with two equiv. of phenylphosphine with 5 mol % of **1** under 360 nm irradiation (continued)

Figure S.20. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)



Hydrophosphination of *p*-methylstyrene with one equiv. of diphenylphosphine by $\mathbf{1}$ Eqn (2)

Prepared according to the general procedure with one equiv. of diphenylphosphine. 98% conversion of *p*-methylstyrene was observed after 5 h of 360 nm irradiation as measured by ¹H NMR and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.²¹





SD-3-225-5P	*/†	
t = 5 h		
SD-3-225-4P	1	
t = 4 h		
SD-3-225-3P		
t = 3 h		
SD-3-225-2P		
t = 1 h 45 m		
SD-3-225-1P		
t = 0		
130 110 90 80 70 60 50 40 30 20 31P muro \$ 22 Stacked 31P(1H) NMP spectra (CDCls 202 MHz)	10 0 -10 -30	-50 -70

Hydrophosphination of *p*-methylstyrene with one equiv. of diphenylphosphine and 5 mol % of **1** under 360 nm irradiation (continued)

Figure S.22. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)



Hydrophosphination of *p*-bromostyrene Eqn (1)

Prepared in accordance with the general procedure with two equiv. of phenylphosphine. After 20 min of 360 irradiation, 100% of styrene was converted with 97:3:2 selectivity for the secondary hydrophosphination product compared to the tertiary product and dehydrocoupled product as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Together, these indicate a 94% conversion of styrene to secondary phosphine. Spectra are consistent with previous reports.⁶



Hydrophosphination of p-bromostyrene with two equiv. of phenylphosphine with 5 mol % of **1** under 360 nm irradiation (continued)





Br

Hydrophosphination of p-bromostyrene with one equiv. of diphenylphosphine by **1** Eqn (2)

Prepared according to general procedure with one equiv. of diphenylphosphine. 88% conversion of *p*-bromostyrene was observed after 5 h and 98% after 8 h as measured by ¹H NMR and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.²¹







Hydrophosphination of p-bromostyrene with one equiv. of diphenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)

Figure S.26. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)



Hydrophosphination of 1-hexene Eqn (3), and Table 1, **3a**

Prepared by the general procedure. After 7 h of 360 nm irradiation, 72% conversion was measured by ¹H and ³¹P{¹H} NMR spectroscopy. After 20 h, 91% of 1-hexene was converted as measured by ¹H NMR and 78% was measured by ³¹P{¹H} NMR spectroscopy. Subsequent reactions measured to an internal standard yielded a similar result. Spectra are consistent with previous reports.⁶





S29

* = hydrophosphination product, ** = double addition hydrophosphination product, \dagger or $\dagger \dagger = Ph_2P-PPh_2$ or PhPH-PhPH, $\dagger = acetylacetone$, ES = PPh₃ external standard, TMB = 1,3,5-trimethoxy benzene, TMS = tetramethyl silane



Hydrophosphination of 1-hexene with two equiv. of phenylphosphine by 5 mol % of 1 under 360 nm irradiation (continued)



Hydrophosphination of 1-hexene with three equiv. of diphenylphosphine by ${\bf 1}$ – neat Table 1, ${\bf 3b}$

In an N₂ filled dry box, 2.28 mmol (424.2 mg, 396 μ L) of diphenylphosphine, 0.76 mmol of 1-hexene and 0.038 mmol (10 mg) of **1** were measured and mixed in a J-Young type polytetrafluoroethylene-valved NMR tube. Initial ¹³C and ³¹P{¹H} NMR spectra were then obtained. After the initial NMR spectra, the reaction was irradiated for 24 h under 360 nm irradiation. A second ¹³C and ³¹P{¹H} NMR spectra were then obtained. Then a ¹H NMR spectra was obtained after adding 200 μ L CDCl₃ mixed with 16.3 mg (0.097 mmol) 1,3,5-trimethoxybenzene as an internal standard. 100% conversion of 1-hexene occurred as measured by ¹H NMR spectroscopy. Ingratiation of the crude ¹H NMR spectra to the internal standard results in between 64 -74% conversion depending on peak integrated and agrees with integration of the ³¹P{¹H} NMR spectra. The catalyst was then removed by silica gel chromatography with 3:1 Hexanes DCM liquid phase in order to get cleaner integrations.

Note: This trial was scaled up in order to obtain spectra ³¹P NMR spectra of the reaction without addition of solvent and to allow for cleaner integrations after removal of catalyst. Comparable results were achieved under the standard conditions.



S31 * = hydrophosphination product, ** = double addition hydrophosphination product, \dagger or $\dagger \dagger = Ph_2P-PPh_2$ or PhPH-PhPH, $\dagger = acetylacetone$, ES = PPh₃ external standard, TMB = 1,3,5-trimethoxy benzene, TMS = tetramethyl silane

Hydrophosphination of 1-hexene with three equiv. of diphenylphosphine by **1** – neat (continued)



Figure S.30. ¹H NMR spectrum (CDCl₃, 500 MHz) at 24 h of 360 nm irradiation after addition of 1,3,5-trimethoxybenzene.



Hydrophosphination of 1-hexene with three equiv. of diphenylphosphine by **1** – neat (continued)

Figure S.31. ¹H NMR spectrum (CDCl₃, 500 MHz) of a fraction with 1 removed



Hydrophosphination of 1-hexene with three equiv. of diphenylphosphine by 1 – neat – (continued)

Figure S.32. ³¹P{¹H} NMR spectrum (CDCl₃, 202 MHz) of a fraction with 1 removed

Attempted hydrophosphination of 1-hexene with one equiv. of diphenylphosphine by 1 in CDCl₃

Prepared according to the general procedure with one equiv. of diphenylphosphine and 9.8 mg of 1,3,5trimethoxybenzene as an internal standard. 25% conversion of diphenylphosphine to a mixture of products was measured by ³¹P{¹H} NMR spectroscopy after 24 h of 360 nm irradiation. New alkyl peaks corresponding to 14% conversion were observed in the ¹H NMR spectra. However, we were unable to unambiguously characterize this as the anti-markovnikov hydrophosphination product due to overlapping peaks and thus is not reported.





Figure S.33. Stacked ¹H NMR spectra (CDCl₃, 500 MHz)



Attempted hydrophosphination of 1-hexene with one equiv. of diphenylphosphine by $\mathbf{1}$ in CDCl₃ (continued)

Figure S.34. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)
Hydrophosphination of 1-heptene Table 1, **3c**

Prepared by the general procedure. After 16 h of 360 nm irradiation, 73% conversion was measured by 1 H and ${}^{31}P{}^{1}$ H NMR spectroscopy. Spectra are consistent with previous reports.⁶





Hydrophosphination of 1-heptene with two equiv. of phenylphosphine by 5 mol % of 1 under 360 nm irradiation (continued)



Hydrophosphination of 1-octene Table 1, **3d**

Prepared according to the general procedure with two equiv. of phenylphosphine. 88% conversion of 1–octene was observed after 16 h of 360 nm irradiation as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.⁶



S39 * = hydrophosphination product, ** = double addition hydrophosphination product, \dagger or $\dagger \dagger = Ph_2P-PPh_2$ or PhPH-PhPH, $\dagger = acetylacetone$, ES = PPh₃ external standard, TMB = 1,3,5-trimethoxy benzene, TMS = tetramethyl silane



Hydrophosphination of 1-octene with two equiv. of phenylphosphine by 5 mol % of 1 under 360 nm irradiation (continued)



Hydrophosphination of ethoxy ethene Table 1, **3e**

Prepared according to the general procedure with two equiv. of phenylphosphine. In an N₂ filled dry box, 0.76 mmol of phenylphosphine (83.5 μ L, 83.5 mg) and 0.38 mmol of ethoxy ethene (27.4 mg) were mixed in 0.5 mL CDCl₃. This solution was then pipetted into a 4 ml shell vial containing 5 mg (0.0019 mmol) **1** in 100 μ L CDCl₃. The resulting yellow solution was quickly transferred to an NMR tube, capped with a disposable NMR cap, and parafilm was wrapped around the NMR cap and shielded from ambient light. After initial ¹H and ³¹P{¹H} NMR spectra were obtained, the reaction was irradiated with 360 nm light and monitored periodically. The yellow solution faded over time to clear. After 8 h of 360 nm irradiation, 100% of ethoxy ethene was converted. Integration of the ¹H and ³¹P{¹H} NMR signals indicated an 84% conversion of ethoxy ethene to secondary phosphine. Replication of this reaction indicated that the reaction went to completion in 6 h. Spectra are consistent with previous literature reports.⁵





Hydrophosphination of ethoxy ethene with phenylphosphine with two equiv. of phenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)





Hydrophosphination of ethoxy ethene with three equiv. of diphenylphosphine by ${\bf 1}$ – neat Table 1, ${\bf 3f}$

Prepared according to the general procedure with the following modifications: 1) no solvent was used 2) three equiv. of diphenylphosphine was used 3) no initial ¹H and ³¹P{¹H} NMR spectra was obtained 4) After 24 h, 5 mg 1,3,5 trimethoxy-benzene was added and 0.5 ml of CDCl₃ was added to the NMR tube and ¹H and ³¹P{¹H} NMR spectra were obtained. 55% conversion to the hydrophosphination product was measured by integration to the internal standard and ¹H and ³¹P{¹H} NMR spectroscopy. Spectra is in accordance with the previous reports.²¹

SD-4-181-1H



Figure S.41. ¹H NMR spectrum (CDCl₃, 500 MHz) at 24 h of 360 nm irradiation

Hydrophosphination of ethoxy ethene with three equiv. of diphenylphosphine by 5 mol % of 1 – neat (continued)

SD-4-181-1P



Figure S.42. ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz) at 24 h of 360 nm irradiation

Hydrophosphination of allyl benzene Table 1, **3g**

Prepared by the general procedure with two equiv. of phenylphosphine. 74% conversion of allyl benzene was observed after 16 h of 360 nm irradiation as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.⁶





Ρh

Hydrophosphination of allyl benzene with two equiv. of phenyphosphine and 5 mol % of 1 under 360 nm irradiation

Hydrophosphination of allyl benzene with three equiv. of diphenylphosphine by ${\bf 1}$ – neat Table 1, ${\bf 3h}$

Prepared according to the general procedure with the following modifications: 1) no solvent was used 2) three equiv. of diphenylphosphine was used 3) no initial ¹H and ³¹P{¹H} NMR spectra was obtained 4) After 24 h, 5 mg 1,3,5 trimethoxy-benzene was added and 0.5 ml of CDCl₃ was added to the NMR tube and ¹H and ³¹P{¹H} NMR spectra were obtained. 70% conversion to the hydrophosphination product was measured by ³¹P{¹H} NMR spectroscopy. Spectra is in accordance with the previous reports.²²



Figure S.45. ³¹P NMR spectrum (CDCl₃, 202 MHz) at 24 h of 360 nm irradiation



Hydrophosphination of allyl benzene with three equiv. of diphenylphosphine by **1** (continued)

Figure S.46. ¹H NMR spectrum (CDCl₃, 500 MHz) of trial 1 at t = 24 h of 360 nm irradiation

Hydrophosphination of allyl chloride Table 1, **3i**

Prepared according to the general procedure with two equiv. of phenylphosphine and an internal standard of 10 mg of 1,3,5-trimethoxybenzene. 75% conversion of allyl chloride was measure after 18 h as measured by ¹H NMR spectroscopy. 63% conversion was measured by integration to an internal standard of 1,3,5-trimethoxybenzene and by ³¹P{¹H} NMR spectroscopy. ³¹P{¹H} Spectra are consistent with previous literature reports.²³

¹H spectra of this product is not previously reported. Purification of the crude product was achieved by flash column chromatography on silica gel with a 3:1 hexanes: DCM liquid phase but a small impurity persisted. The isolated yield of this product was 44% at 92% purity. The impurities were 4% of the double activation and 4% of an unknown product at -7 ppm in the ³¹P{¹H}NMR spectra.

¹H NMR (500 MHz, CDCl₃) δ 7.64 – 7.45 (m, 2H), 7.35 (m, 3H), 4.20 (d, *J* = 208.2 Hz, 1H), 3.72 – 3.33 (m, 2H), 2.20 – 1.75 (m, 4H)



Figure S.47. ¹H NMR spectrum (CDCl₃, 500 MHz)) at t = 18 h of 360 nm irradiation with catalyst removed by

Hydrophosphination of allyl chloride with two equiv. of phenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)







Hydrophosphination of allyl chloride with two equiv. of phenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)





Hydrophosphination of $\beta\text{-methyl}$ styrene Table 2, 4a

Prepared by the general procedure with two equiv. of phenylphosphine and a 50:50 mixture of E:Z isomers of β -methyl styrene. 100% conversion was observed at 20 h of 360 nm irradiation as measured by ¹H and ³¹P{¹H} NMR spectroscopy.



Ph P.H Hydrophosphination of β -methyl styrene with two equiv. of phenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)



Figure S.51. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)



Hydrophosphination of $\alpha\text{-methyl}$ styrene with two equiv. of phenylphosphine by 1 Table 2, 4b

Prepared according to the general procedure with two equiv. of phenylphosphine. 92% conversion of α -methyl styrene was measure after 2 h and 15 min as measured by ¹H and ³¹P{¹H} NMR spectroscopy. 100% conversion was measured at 5 h of 360 nm irradiation. Spectra are consistent with previous reports.⁶



Figure S.52. Stacked ¹H NMR spectra (CDCl₃, 500 MHz). P-H protons are broadened into baseline in final spectra due to interaction with copper.





Hydrophosphination of α -methyl styrene with two equiv. of phenylphosphine with 5 mol % of **1** under 360 nm irradiation (continued)



Hydrophosphination of $\alpha\text{-methyl}$ styrene Table 2, $\boldsymbol{4c}$

Prepared by the general procedure with modified starting material amounts of 0.25 mmol (29.5 mg) α -methyl styrene, 0.375 mmol (70 mg) diphenylphosphine and 3.2 mg (0.0125 mmol, 5 mol %) **1** and 5 mg (0.03 mmol) of 1,3,5-trimethoxybenzene. 83% conversion was measured after 20 h of 360 nm irradiation as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.⁸

SD-4-146-2H

t = 20 h



Hydrophosphination of α -methyl styrene with 1.5 equiv. of diphenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)



t = 20 h



Ph

Hydrophosphination of 2,3-dimethyl,1-3-butadiene Table 2, **4d**

Prepared according to the general procedure with two equiv. of phenylphosphine. 100% conversion of 2,3dimethyl,1-3-butadiene was observed after 1 h 20 min of 360 nm irradiation as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.⁵





Hydrophosphination of 2,3-dimethyl,1-3-butadiene with two equiv. of phenylphosphine by 5 mol % of 1 under 360 nm irradiation (continued)

Hydrophosphination of 2,3-dimethyl,1-3-but adiene with one equiv. of diphenylphosphine by ${\bf 1}$ Table 2, ${\bf 4e}$

Prepared according to general procedure with one equiv. of diphenylphosphine. 100% conversion of 2,3dimethyl,1-3-butadiene was measured by ¹H and ³¹P{¹H} NMR spectroscopy after 8 h of 360 nm irradiation. Spectra is in accordance with previous reports.²¹



Ph P P Ph Hydrophosphination of 2,3-dimethyl,1-3-butadiene with one equiv. of diphenylphosphine and 5 mol % of 1 under 360 nm irradiation (continued)



Figure S.59. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)



Hydrophosphination of norbornene Table 2, **4f**

Prepared according to the general procedure with two equiv. of phenylphosphine. Greater than 99% of norbornene was converted after 50 min of 360 nm irradiation as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous literature reports.⁵







Hydrophosphination of norbornene with two equiv. of phenylphosphine with 5 mol % of **1** under 360 nm irradiation (continued)

Figure S.61. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)

 $\mathbf{P}_{\mathsf{H}}^{\mathsf{Ph}} = \mathbf{P}_{\mathsf{H}}^{\mathsf{Ph}}$

Hydrophosphination of acrylonitrile Table 2, **4g**

Prepared according to the general procedure with two equiv. of phenylphosphine and the modification that it was not irradiated because the initial ¹H and ³¹P{¹H} NMR spectra indicated 100% conversion of acrylonitrile. Spectra are consistent with previous reports.⁵



Figure S.62. ¹H NMR spectrum (CDCl₃, 500 MHz) at t = 25 min



Hydrophosphination of acrylonitrile with two equiv. of phenylphosphine by 5 mol % of **1** (continued)

Figure S.63. ³¹P{¹H} NMR spectrum (CDCl₃, 202 MHz) at t = 25 min

Hydrophosphination of acrylonitrile with one equiv. of diphenylphosphine by ${\bf 1}$ Table 2, ${\bf 4h}$

Prepared according to the general procedure with one equiv. of diphenylphosphine and the modification that it was not irradiated because the initial ¹H and ³¹P{¹H} NMR spectra indicated 98% conversion of acrylonitrile as measured by ¹H NMR and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.²¹

SD-4-117-1P

t = 20 m



Figure S.64. ³¹P{¹H} NMR spectrum (CDCl₃, 202 MHz) at t = 20 min

Hydrophosphination of methylacrylate Table 2, **4i**

Prepared according to the general procedure with two equiv. of phenylphosphine and the modification that it was not put under 360 nm irradiation because the reaction went to 99% completion by the initial ¹H and ³¹P{¹H} NMR spectra. Spectra are consistent with previous reports.⁵



Figure S.65. ¹H NMR spectrum (CDCl₃, 500 MHz) at t = 10 min



Hydrophosphination of methyl acrylate with two equiv. of phenylphosphine by 5 mol % of 1 (continued)

Figure S.66. ³¹P{¹H} NMR spectrum (CDCl₃, 202 MHz) at t = 10 m

Hydrophosphination of methylacrylate with one equiv. of diphenylphosphine by ${\bf 1}$ Table 2, ${\bf 4j}$

Prepared according to the general procedure with one equiv. of diphenylphosphine and the modification that it was not put under 360 nm irradiation because the reaction went to 99% completion by the initial 1 H and 31 P{ 1 H} NMR spectra. Spectra are consistent with previous reports. 21



Figure S.67. ³¹P{¹H} NMR spectrum (CDCl₃, 202 MHz) at t = 15 min

Hydrophosphination of 1-phenyl-1-butyne Eqn (4)

Prepared according to the general procedure with one equiv. of phenylphosphine and an external standard of PPh₃. 80% of 1-phenyl-1-butyne was converted after 24 h of 360 nm irradiation with 3:1 *Z:E* selectivity as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.⁴



Hydrophosphination of 1-phenyl-1-butyne with one equiv. of phenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)



Figure S.69. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz). ‡ Consitent with double hydrophosphination product.



Hydrophosphination of 1-phenyl-1-butyne with one equiv. of diphenylphosphine by **1** Eqn (4) and Table S3

Prepared in accordance with the general procedure with one equiv. of diphenylphosphine. 74% conversion of 1phenyl-1-butyne with 1:1.1 *Z:E* selectivity was measured after 24 h of 360 nm irradiation. In Trial 2, 75% conversion was observed with 4.8:1 *Z:E* selectivity. The inconsistency is likely due to isomerization under UV light⁴ or on metal isomerization.²⁴




Hydrophosphination of 1-phenyl-1-butyne with one equiv. of diphenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)



Figure S.71. Stacked ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz)

Ft S→ Ph PPh₂

Hydrophosphination of phenylacetylene Eqn (4) and Table S3

Prepared in accordance with the general procedure with one equiv. of diphenylphosphine. 41% conversion of phenylacetylene was measured by ¹H and ³¹P{¹H} NMR spectroscopy after 24 h of 360 nm irradiation. Spectra is in accordance with previous reports.²¹





Hydrophosphination of phenylacetylene with one equiv. of diphenylphosphine by 5 mol % of **1** under 360 nm irradiation (continued)



Figure S.73. Stacked ¹H NMR spectra (CDCl₃, 500 MHz) at t = 24 h of 360 nm irradiation



Hydrophosphination of 3-hexyne Table S3

Prepared according to the general procedure with two equiv. of phenylphosphine and an external standard of PPh₃ added. 71% of 3-hexyne was converted after 24 h of 360 nm irradiation with 1:1.5 *Z:E* selectivity as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.⁴



Hydrophosphination of 3-hexyne with one equiv. of phenylphosphine by 5 mol % of 1 under 360 nm irradiation (continued)



Hydrophosphination of diphenylacetylene Table S3

Prepared according to general procedure with two equiv. of phenylphosphine and an external standard of PPh₃. 58% of diphenylacetylene was converted after 24 h of 360 nm irradiation with 2.1:1 *Z:E* selectivity as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Spectra are consistent with previous reports.²⁵



Hydrophosphination of phenylacetylene with two equiv. of phenylphosphine Table S3

Prepared by the general procedure with two equiv. of phenylphosphine and an external standard of PPh₃. 88% of phenylacetylene was converted to the double P-H activation product after 24 h of 360 nm irradiation with 2.7:1 Z,Z:E,Z selectivity as measured by ¹H and ³¹P{¹H} NMR spectroscopy. Z,Z is isomer is consistent with the literature.²⁶ The E:Z isomer has not been characterized by NMR spectroscopy for this compound but has been for several para substituted phenylacetylenes derivatives and our assignment.



Hydrophosphination of diphenylacetylene with two equiv. of diphenylphosphine by ${\bf 1}$ Table S3

Prepared in accordance with the general procedure with two equiv. of diphenylphosphine. 36% conversion of diphenylacetylene with 35:1 *Z:E* selectivity was measured by ¹H and ³¹P{¹H} NMR spectroscopy after 72 h 360 nm irradiation. Spectra are in accordance with previous reports.²¹



Ph

Characterization of new products: (E,Z)-diphenyl(1-phenylbut-1-en-2-yl)phosphine Hydrophosphination products of 1-phenyl-1-butyne and diphenylphosphine

Synthesized following the general procedure with one equiv. of diphenylphosphine and the modification that the reaction was scaled up and replicated with 0.99 mg (0.76 mmol) 1-phenyl-1-butyne, 140 mg (0.76 mmol) of diphenylphosphine, 10 mg **1** in 1 ml of CDCl₃. Purification of the crude product was achieved by flash column chromatography on silica gel with a 3:1 hexanes: DCM liquid phase. ³¹P{¹H} NMR spectroscopy indicated that the product was isolated in earlier fractions as a mixture of isomers and later fractions as the pure Z isomer. Over time, slow oxidation of the products occurred. The Z isomer peaks were determined by subtracting the signal for the previously characterized Z isomer phosphine oxide²⁷ from the ¹H, ¹³C, ³¹P, ³¹P{¹H} NMR spectra of later fractions. Assignments were verified by ³¹P HMBC. The E isomer peaks were determined by subtracting the Z isomer phosphine peaks from earlier fractions. The exact splitting patterns for the aromatic region of the products was hampered by overlap of peaks.



(Z)-diphenyl(1-phenylbut-1-en-2-yl)phosphine. ¹H NMR (500 MHz, CDCl₃) δ 7.45 – 7.29 (15H, Ar), 7.28 – 7.23 (m, 1H), 2.17 – 2.05 (qt, *J* = 7.3, 1.95 Hz, 2H), 0.99 (t, *J* = 7.3 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 140.98 (d, *J*_{C-P} = 29.2 Hz), 139.27 (d, *J*_{C-P} = 21.6 Hz), 137.67 (d, *J*_{C-P} = 7.4 Hz), 136.96 (d, *J*_{C-P} = 12.4 Hz), 133.32 (d, *J*_{C-P} = 18.7 Hz), 129.47 (d, *J*_{C-P} = 7.1 Hz), 128.32 (d, *J*_{C-P} = 6.2 Hz), 128.24 (s), 127.79 (s), 127.22 (s), 28.97 (d, *J*_{C-P} = 4.0 Hz), 13.54 (s). ³¹P{¹H} NMR (202 MHz, CDCl₃) δ -11.61 (s). MS calcd for C₂₂H₂₂P: 317.15 [M+H]+, found: 317.2.



(E)-diphenyl(1-phenylbut-1-en-2-yl)phosphine. ¹H NMR (500 MHz, CDCl₃) δ 7.53 – 7.24 (Ar), 6.42 (d, *J* = 10.3 Hz, 1H), 2.51 (dq, *J* = 14.9, 7.5 Hz, 2H), 1.14 (t, *J* = 7.5 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) unable to clearly assign peaks due to oxide impurity. ³¹P{¹H} NMR (202 MHz, CDCl₃) δ 1.11 (s). MS calcd for C₂₂H₂₂P: 317.15 [M+H]⁺, found: 317.2.



Figure S.79. ¹H NMR spectrum (CDCl₃, 500 MHz)



SD-4-77-Fraction8



Figure S.81. ³¹P{¹H} NMR spectra (hexanes/DCM 3:1, 202 MHz) of four isolated fractions



SD-4-77-F8P



Figure S.82. ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz) of 4 isolated fractions. Some oxidation of the products occurred.



Characterization of new products: (1-phenylprop-2-yl)diphenylphosphine Hydrophosphination products of β -methyl styrene and phenylphosphine

Synthesized following the general procedure with two equiv. of phenylphosphine. Purification of the crude product was achieved in a dry box by flash column chromatography on silica gel with a 3:1 Hexanes: DCM liquid phase followed by evaporation of volatile materials by vacuum. Fraction 3 contained 25.3 mg (0.11 mmol, 29% yield) of the product in 98% purity by ³¹P{¹H} NMR spectroscopy. The 2% phosphorus impurity is likely the markovnikov addition product signal. Fractions 4,5, and 6 contained a total of 46.7 mg (za53.8%) of 100% pure product as determined by ³¹P NMR spectroscopy for a total isolated yield of 82% in 99% purity.

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(1-phenylprop-2-yl)diphenylphosphine: ¹H NMR (500 MHz, CDCl₃) δ 7.53 – 7.08 (10H, Ar), 4.11 (ddd, *J* = 211.2, 20.2, 4.9 Hz, 1H), 2.92 (dd, *J* = 12.8, 5.7 Hz, 1H), 2.63 – 2.46 (m, 1H), 2.37 – 2.22 (m, 1H), 1.13 – 0.96 (m, 3H). ³¹P{¹H} NMR (202 MHz, CDCl₃) δ -28.65 (s), -32.54 (s). ¹³C NMR (126 MHz, CDCl₃) δ 140.64 (s), 135.16 – 134.55 (m), 134.42 (d, *J* = 12.2 Hz), 129.03 (d, *J* = 11.9 Hz), 128.75 – 128.01 (m), 126.09 (s), 42.34 (d, *J* = 9.3 Hz), 41.76 (s), 31.61 (dd, *J* = 24.8, 9.3 Hz), 18.90 (s), 18.12 (d, *J* = 16.3 Hz).³¹P NMR (202 MHz, CDCl₃) δ -27.86 (d, *J* = 212.4 Hz), -31.77 (d, *J* = 211.9 Hz). MS calcd for C₁₅H₁₇P: 229.27 [M+H]+, found: 229.3



Figure S.83. ¹H NMR spectrum (CDCl₃, 500 MHz) of (1-phenylprop-2-yl)diphenylphosphine. Residual ammounts of residual solvent (DCM, hexanes) are visible.







Figure S.84. ³¹P NMR spectrum (CDCl₃, 202 MHz) of (1-phenylprop-2-yl)diphenylphosphine

SD-4-22-F6P



Figure S.85. ³¹P{¹H} NMR spectra (CDCl₃, 202 MHz) of two pure fractions of (1-phenylprop-2-yl)diphenylphosphine



References

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- 1. H. Gulyás, J. Benet-Buchholz, E. C. Escudero-Adan, Z. Freixa and P. W. N. M. van Leeuwen, *Chem. Eur. J.*, 2007, **13**, 3424.
- 2. I. Paterson, S. J. Fink, L. Y. W. Lee, S. J. Atkinson and S. B. Blakey, Org. Lett., 2013, 15, 3118.
- 3. B. J. Ackley, J. K. Pagano and R. Waterman, Chem. Commun., 2018, 54, 2774.
- 4. C. A. Bange and R. Waterman, ACS Catal., 2016, 6, 6413.
- 5. C. A. Bange, M. A. Conger, B. T. Novas, E. R. Young, M. D. Liptak and R. Waterman, ACS Catal., 2018, 8, 6230.
- 6. M. B. Ghebreab, C. A. Bange and R. Waterman, *J. Am. Chem. Soc.*, 2014, **136**, 9240.
- 7. M. D. Gordon and L. D. Quin, J. Am. Chem. Soc., 1976, **98**, 15.
- 8. A. K. King, A. Buchard, M. F. Mahon and R. L. Webster, Chem. Eur. J., 2015, 21, 15960.
- 9. R. Waterman, Organometallics, 2007, 26, 2492.
- 10. D. G. Yakhvarov, Y. S. Ganushevich and O. G. Sinyashin, Mendeleev Commun., 2007, 17, 197.
- 11. A. K. King, K. J. Gallagher, M. F. Mahon and R. L. Webster, Chem. Eur. J. , 2017, 23, 9039.
- 12. A. O. Tolpygin, A. V. Cherkasov, G. K. Fukin, T. A. Kovylina, K. A. Lyssenko and A. A. Trifonov, *Eur. J. Inorg. Chem.*, 2019, 2019, 4289.
- 13. N. A. Isley, R. T. Linstadt, E. D. Slack and B. H. Lipshutz, *Dalton Trans.*, 2014, 43, 13196.
- 14. M. P. Cibuzar, S. G. Dannenberg and R. Waterman, Isr. J. Chem., 2019, 60, 446.
- 15. I. V. Lapshin, O. S. Yurova, I. V. Basalov, V. Y. Rad'kov, E. I. Musina, A. V. Cherkasov, G. K. Fukin, A. A. Karasik and A. A. Trifonov, *Inorg. Chem.*, 2018, **57**, 2942.
- 16. W. S. Tay, X.-Y. Yang, Y. Li, S. A. Pullarkat and P.-H. Leung, *Dalton Trans.*, 2019, **48**, 4602.
- 17. R. G. Belli, K. M. E. Burton, S. A. Rufh, R. McDonald and L. Rosenberg, Organometallics, 2015, 34, 5637.
- 18. L. Routaboul, F. Toulgoat, J. Gatignol, J.-F. Lohier, B. Norah, O. Delacroix, C. Alayrac, M. Taillefer and A.-C. Gaumont, *Chem. Eur. J.*, 2013, **19**, 8760.
- 19. A. K. King, K. J. Gallagher, M. F. Mahon and R. L. Webster, *Chem. Eur. J.*, 2017, **23**, 9039.
- 20. T. H. Lemmen, G. V. Goeden, J. C. Huffman, R. L. Geerts and K. G. Caulton, Inorg. Chem., 1990, 29, 3680.
- 21. B. T. Novas, C. A. Bange and R. Waterman, Eur. J. Inorg. Chem., 2019, 2019, 1640.
- 22. G. Phillips, S. Hermans, J. R. Adams and B. F. G. Johnson, Inorganica Chim. Acta, 2003, 352, 110.
- 23. H. Schmidt, J. Prakt. Chem., 1997, 339, 482.
- 24. M. M. I. Basiouny, D. A. Dollard and J. A. R. Schmidt, ACS Catal., 2019, 9, 7143.
- 25. G. Zhao, F. Basuli, U. J. Kilgore, H. Fan, H. Aneetha, J. C. Huffman, G. Wu and D. J. Mindiola, *J. Am. Chem. Soc.*, 2006, **128**, 13575.
- 26. N. T. Coles, M. F. Mahon and R. L. Webster, Chem. Commun., 2018, 54, 10443.
- 27. H. Wang, Y. Li, Z. Tang, S. Wang, H. Zhang, H. Cong and A. Lei, ACS Catal, 2018, 8, 10599.