

Electronic Supplementary Information

Oxidative Addition of Bi-C Bonds to Pt(0): Reaction of Pt(PEt₃)₃ with 12-Phenyl- and 12-Chloro-5,6,7,12-tetrahydrodibenzo[*c,f*][1,5]azabismocines

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1. General.

All manipulations of air-sensitive materials were carried out under a nitrogen atmosphere using standard Schlenk techniques or in a glovebox. Anhydrous toluene, hexane and THF were purchased from Kanto Chemicals and degassed before use. C₆D₆, THF-*d*₈, and CD₂Cl₂ were dried over molecular sieves and degassed. NMR spectra were recorded on Jeol LA500, JNM-ECX400P spectrometers, or Bruker AVANCE III HD 600 spectrometer with a CryoProbe. Chemical shifts are reported in δ (ppm) and are referenced to the (residual) solvent signals for ¹H and ¹³C^{S1} or to the external standard (85% H₃PO₄ (0 ppm)) for ³¹P. Coupling constants were reported in Hertz. 5,6,7,12-Tetrahydrodibenz[*c,f*][1,5]azabismocines **1**,^{S2} **2a**,^{S2} and **2b**^{S3} and Pt(PEt₃)₄^{S4} were prepared according to literature procedures.

Safety Caution: Pt(PEt₃)₄ and PEt₃ are pyrophoric and should be handled with great care.

2. Reaction of 12-phenyl-5,6,7,12-tetrahydrodibenzo[*c,f*][1,5]azabismocine **1** with Pt(PEt₃)₃.

Monitoring of the reaction of **1 and Pt(PEt₃)₃ by NMR.** Pt(PEt₃)₄ (35.6 mg, 0.066 mmol) was placed in a Schlenk tube and heated at 60 °C for 10 min under vacuum to form Pt(PEt₃)₃ as an orange oil. Compound **1** (43.8 mg, 0.066 mmol) and C₆D₆ (0.8 mL) were added to the Schlenk tube to form an orange solution. Ca. 0.5 mL of the solution was transferred to an NMR tube and the tube was sealed with a PTFE valve. The reaction progress at room temperature was periodically monitored by NMR. After 91 h, volatiles were removed under vacuum and the residue was dissolved again in C₆D₆. The reaction was further monitored by NMR.

The progress of the reaction was shown in Fig. S1 (³¹P{¹H} NMR) and Fig. S2 (¹H NMR).

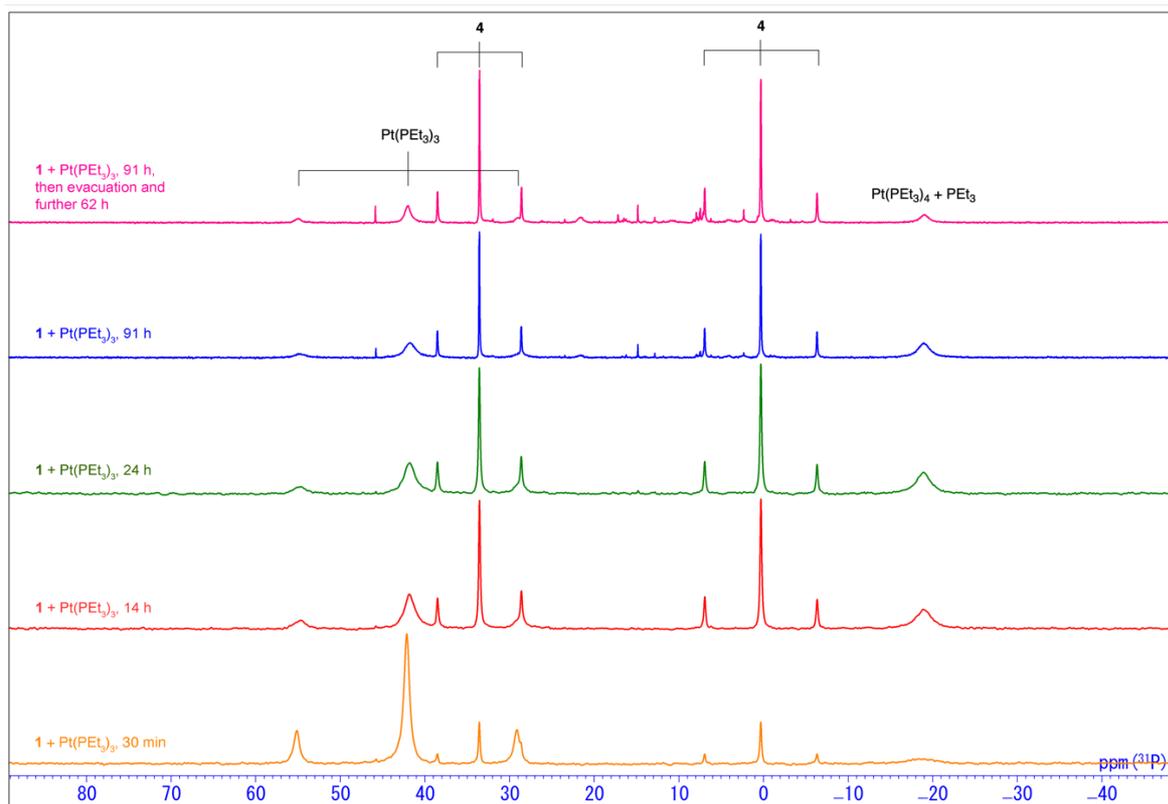


Fig. S1 $^{31}\text{P}\{^1\text{H}\}$ NMR spectra (161 MHz, C_6D_6) of the reaction mixture of **1** and $\text{Pt}(\text{PEt}_3)_3$ after 30 min (orange), 14 h (red), 24 h (green), 91 h (blue) and 91 h + evacuation + 62 h (pink).

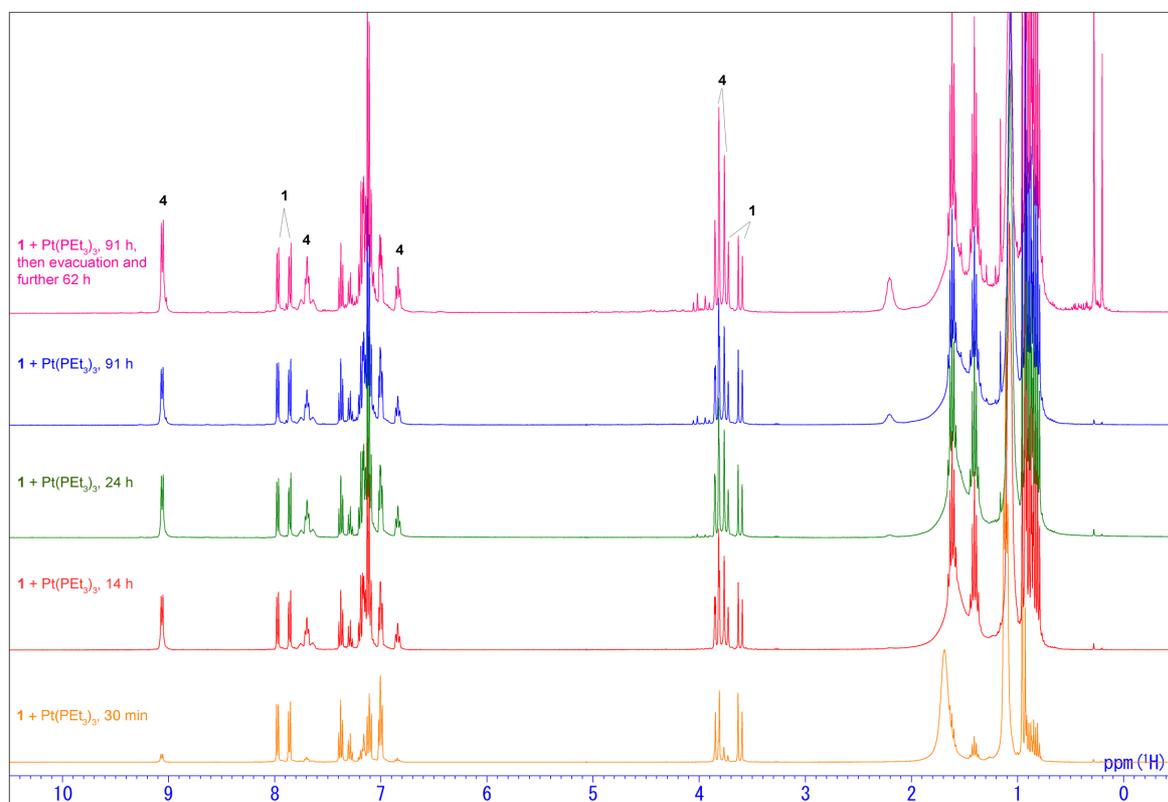
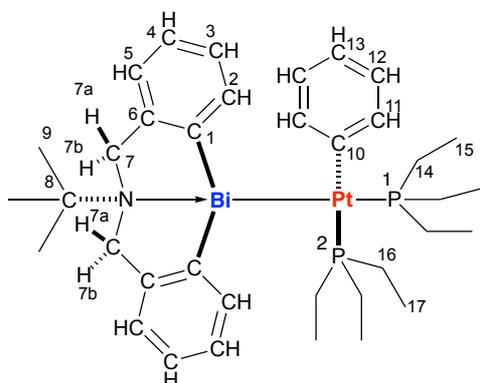


Fig. S2 ^1H NMR spectra (400 MHz, C_6D_6) of the reaction mixture of **1** and $\text{Pt}(\text{PEt}_3)_3$ after 30 min (orange), 14 h (red), 24 h (green), 91 h (blue) and 91 h + evacuation + 62 h (pink).

Synthesis of complex 4. A mixture of Pt(PET₃)₃ (0.378 mmol, prepared from 203 mg of Pt(PET₃)₄ by heating at 60 °C under vacuum) and **1** (254 mg, 0.380 mmol) in benzene (5 mL) was stirred at room temperature for 43 h. The color of the mixture changed from light orange to brown. A small aliquot of the mixture was taken for ¹H NMR analysis, which showed the molar ratio of **1**:**4** = ca 43:57. After the volatiles were removed under vacuum, the residue was again dissolved in benzene (6 mL) and stirred at room temperature for further 24 h. After the volatiles were removed under vacuum, the residue was dissolved in hexane (10 mL) and the hexane solution was kept at –32 °C for 4 days, resulting the formation of orange crystals. The supernatant was removed and the crystals were washed with cold hexane (cooled at –32 °C, 3 × 0.7 mL) and dried under vacuum to give **4** (151 mg, 41% yield). The stability of **4** in deuterated solvents was checked and found to be C₆D₆ > THF-*d*₈ (more than 2/3 of **4** changed to unknown species after 24 h) >> CD₂Cl₂ (immediate (partial) change to unknown species). However, signal separation was better in THF-*d*₈ than in C₆D₆. Therefore, full NMR characterization was done in THF-*d*₈. ¹H NMR (499 MHz, C₆D₆, Fig. S11) δ 0.78-0.93 (m, 18H), 0.92 (s, 9H), 1.40 (quint, *J* = 7.6 Hz, 6H), 1.61 (quint, *J* = 7.5 Hz, 6H), 3.76 (d, *J* = 15.1 Hz, 2H), 3.84 (d, *J* = 15.1 Hz, 2H), 6.86 (dt, *J* = 1.1, 7.3 Hz, 1H), 7.11-7.24 (m, 8H), 7.72 (dt, *J* = 1.1, 5.9 Hz, ³*J*_{Pt-H} = 45 Hz, 2H), 9.09 (d, *J* = 6.6 Hz, 2H). ¹³C NMR (126 MHz, C₆D₆) δ 8.46 (*J*_{Pt-C} = 12 Hz), 9.83 (*J*_{Pt-C} = 21 Hz), 15.5-16.1 (m), 20.6-21.3 (m), 28.04, 56.54, 57.28, 121.51, 126.27, 128.35, 128.76 (br, *J*_{Pt-C} = ca. 60 Hz), 139.50 (d, *J*_{P-C} = 2 Hz, *J*_{Pt-C} = ca. 28 Hz), 143.96 (*J*_{Pt-C} = ca. 33 Hz), 149.36, 150.27 (br). ³¹P NMR (202 MHz, C₆D₆) δ –0.22 (s, ¹*J*_{Pt-P} = 2149 Hz), 32.98 (s, ¹*J*_{Pt-P} = 1605 Hz).



Atomic numbering of complex **4** for the NMR signal assignment

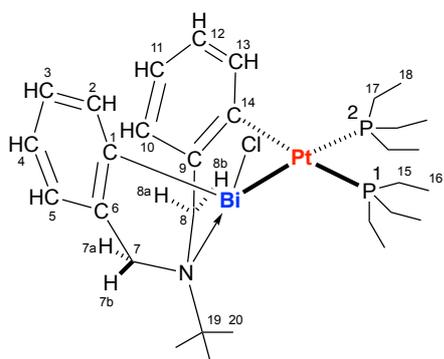
¹H NMR (600 MHz, THF-*d*₈, Fig. S13) δ 0.86-0.96 (m, 9H, H-17), 0.96 (s, 9H, H-9), 1.09-1.16 (m, 9H, H-15), 1.64 (quint, *J* = 7.6 Hz, 6H, H-14), 1.72 (quint, *J* = 7.5 Hz, 6H, H-16), 3.61 (d, *J* = 15.0 Hz, 2H, H-7a), 3.80 (d, *J* = 15.0 Hz, 2H, H-7b), 6.55 (t, *J* = 7.1 Hz, 1H, H-13), 6.81-6.88 (m, 4H, H-3 & H-12), 6.96-7.02 (m, 4H, H-4 & H-5), 7.37 (t with Pt satellites, *J* = 6.1 Hz, ³*J*_{Pt-H} = 44 Hz, 2H, H-11), 8.69 (d, *J* = 6.9 Hz, 2H, H-2). ¹³C {¹H} NMR (151 MHz, THF-*d*₈, Figs. S14-S18) δ 8.6 (s with Pt satellites, ³*J*_{Pt-C} = 12 Hz, C-15), 9.9 (s with Pt satellites, ³*J*_{Pt-C} = 21 Hz, C-17), 15.8-16.2 (m, C-14), 20.9-21.5 (m, C-16), 28.1 (s, C-9), 56.7 (s, C-8), 57.3 (s, C-7), 121.1 (s, C-13), 126.1 (s, C-4), 128.0 (s, C-5), 128.1 (s, C-3), 128.4 (dd with Pt satellites, ⁴*J*_{P-C} = 2, 6 Hz, ³*J*_{Pt-C} = 58 Hz, C-12), 139.7 (dd with Pt satellites, ³*J*_{P-C} = 1, 3 Hz, ²*J*_{Pt-C} = 28 Hz, C-11), 143.9 (s with Pt satellites, ³*J*_{Pt-C} = 33 Hz, C-2), 149.2 (s, C-6), 149.9 (br, C-1), 162.3 (d with Pt satellites, ²*J*_{P-C} = 101 Hz, ¹*J*_{Pt-C} = 721 Hz, C-10). ³¹P {¹H} NMR (243 MHz, THF-*d*₈, Fig. S19) δ –0.9 (s with Pt satellites, ¹*J*_{Pt-P} = 2147 Hz, P-2), 32.6 (s with

Pt satellites, $^1J_{\text{Pt-P}} = 1650$ Hz, P-1).

Anal. Calcd for $\text{C}_{36}\text{H}_{56}\text{BiNP}_2\text{Pt}$: C, 44.63%; H, 5.83%; N, 1.45%. Found: C, 44.95%; H, 5.78%; N, 1.38%.

3. Reaction of 12-chloro-5,6,7,12-tetrahydrodibenzo[*c,f*][1,5]azabismocine **2a** or **2b** with $\text{Pt}(\text{PEt}_3)_3$.

Synthesis of complex 5a. A mixture of $\text{Pt}(\text{PEt}_3)_3$ (0.30 mmol, prepared from 200 mg of $\text{Pt}(\text{PEt}_3)_4$ by heating at 60 °C under vacuum) and **2a** (149 mg, 0.30 mmol) in THF (20 mL) was stirred for 30 h at room temperature and then 60 h at 40 °C. After the volatiles were removed under vacuum, the residue was washed with hexane (2×20 mL) and then with hexane/THF (1/2, 15 mL). The residual yellow powder was dried under vacuum to give **5a** (144 mg, 52%). Because NMR signal separation was better in CD_2Cl_2 than in C_6D_6 , full NMR characterization was done in CD_2Cl_2 . ^1H NMR (499 MHz, C_6D_6 , Fig. S25) δ 0.63 (dt, $J_{\text{P-H}} = 15$ Hz, $J_{\text{H-H}} = 7.6$ Hz, 9H), 1.07-1.30 (m, 6H), 1.08 (dt, $J_{\text{P-H}} = 15$ Hz, $J_{\text{H-H}} = 7.6$ Hz, 9H), 1.18 (s, 9H), 1.98 (quasi sept, $J = 7.3$ Hz, 3H), 2.27 (quasi sept, $J = 7.6$ Hz, 3H), 3.40 (d, $J = 9.8$ Hz, 2H), 3.73 (d, $J = 15.8$ Hz, 2H), 4.09 (d, $J = 15.6$ Hz, 2H), 4.63 (d, $J = 9.9$ Hz, 2H), 6.39 (t, $J = 7.3$ Hz, 1H), 6.72 (t, $J = 7.3$ Hz, 1H), 6.85-6.94 (m, 2H), 6.98-7.28 (m, 3H), 8.99 (d, $J = 7.5$ Hz, 1H). ^{31}P NMR (202 MHz, C_6D_6) δ 0.54 (d, $^2J_{\text{P-P}} = 11$, $^1J_{\text{Pt-P}} = 2063$ Hz), 58.08 (d, $^2J_{\text{P-P}} = 11$, $^1J_{\text{Pt-P}} = 2036$ Hz).



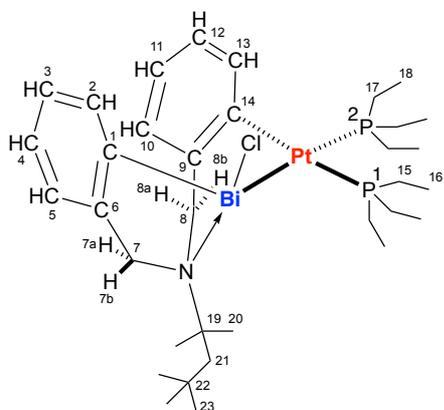
Atomic numbering of complex **5a** for the NMR signal assignment

^1H NMR (600 MHz, CD_2Cl_2 , Fig. S26) δ 0.92-0.98 (m, 9H, H-18), 1.20-1.27 (m, 9H, H-16), 1.46 (s, 9H, H-20), 1.43-1.52 (m, 3H, H-17), 1.57-1.65 (m, 3H, H-17), 2.12 (quasi septet, $J = 7.4$ Hz, 3H, H-15), 2.34 (quasi septet, $J = 7.6$ Hz, 3H, H-15), 3.61 (d, $J = 10.2$ Hz, 1H, H-8a), 3.98 (d, $J = 15.9$ Hz, 1H, H-7a), 4.31 (d, $J = 15.9$ Hz, 1H, H-7b), 4.69 (d, $J = 10.2$ Hz, 1H, H-8b), 6.08 (tt, $J = 1.4, 7.3$ Hz, 1H, H-11), 6.43 (t, $J = 7.3$ Hz, 1H, H-12), 6.71 (t, $J = 7.2$ Hz, 1H, H-3), 6.81 (t, $J = 7.5$ Hz, 1H, H-5), 6.83 (t, $J = 7.5$ Hz, 1H, H-4), 6.87 (td, $J = 1.7, 7.4$ Hz, 1H, H-10), 6.90-6.94 (m, 1H, H-13), 7.91 (d, $J = 7.4$ Hz, 1H, H-2). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_2Cl_2 , Figs. S27-S31) δ 8.5 (d with Pt satellites, $J_{\text{P-C}} = 1$ Hz, $^3J_{\text{Pt-C}} = 15$ Hz, C-18), 9.2 (s with Pt satellites, $^3J_{\text{Pt-C}} = 14$ Hz, C-16), 16.2-16.7 (m, C-17), 18.8-19.2 (m, C-15), 29.8 (br, C-20), 59.7 (s, C-7), 60.5 (s, C-19), 65.8 (s with Pt satellites, $^3J_{\text{Pt-C}} = 82$ Hz, C-8), 121.3 (d, $^5J_{\text{P-C}} = 3$ Hz, C-11), 124.3 (s, C-4), 125.59 (s, C-5), 125.66 (dd with Pt satellites, $^4J_{\text{P-C}} = 2, 5$ Hz, $^3J_{\text{Pt-C}} = 49$ Hz, C-12), 126.9 (s, C-3), 132.3 (dd with Pt satellites, $^4J_{\text{P-C}} = 2, 6$ Hz, $^3J_{\text{Pt-C}} = 41$ Hz, C-10), 139.0 (d, $^3J_{\text{P-C}} = 5$ Hz, C-13), 139.5 (s, C-2), 143.2 (t with Pt satellites, $^3J_{\text{P-C}} = 2$

Hz, $^2J_{\text{Pt-C}} = 26$ Hz, C-9), 151.3 (s, C-6), 161.6 (dd with Pt satellites, $^2J_{\text{P-C}} = 9$, 93 Hz, $^1J_{\text{Pt-C}} = 693$ Hz, C-14), 169.2 (br, C-1). $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CD_2Cl_2 , Fig. S32) δ 0.3 (d with Pt satellites, $^2J_{\text{P-P}} = 12$ Hz, $^1J_{\text{Pt-P}} = 2029$ Hz, P-1), 62.2 (d with Pt satellites, $^2J_{\text{P-P}} = 12$ Hz, $^1J_{\text{Pt-P}} = 2154$ Hz, P-2).

Anal. Calcd for $\text{C}_{30}\text{H}_{51}\text{BiClNP}_2\text{Pt}$: C, 38.86%; H, 5.54%; N, 1.51%. Found: C, 39.06%; H, 5.46%; N, 1.42%.

Synthesis of complex 5b. A mixture of $\text{Pt}(\text{PEt}_3)_3$ (0.33 mmol, prepared from 222 mg of $\text{Pt}(\text{PEt}_3)_4$ by heating at 60 °C under vacuum) and **2b** (183 mg, 0.33 mmol) in C_6H_6 (11 mL) was stirred for 92 h at room temperature and then for 23 h at 40 °C. After the volatiles were removed under vacuum, the residue was extracted with hexane (10 mL) and then with hexane/ C_6H_6 (3/1, 8 mL). Cooling the hexane/ C_6H_6 extract at -35 °C afforded **5b** as a yellow powder, which was separated and dried under vacuum (107 mg, 33%).



Atomic numbering of complex **5b** for the NMR signal assignment

^1H NMR (600 MHz, CD_2Cl_2) δ 0.92-0.98 (m, 9H, H-18), 1.07 (s, 9H, H-23), 1.21-1.28 (m, 9H, H-16), 1.43-1.73 (m, 13H, H-17, H-20 & one of H-21), 1.85 (d, $J = 13.9$ Hz, 1H, one of H-21), 2.12 (quasi septet, $J = 7.4$ Hz, 3H, H-15), 2.34 (quasi septet, $J = 7.6$ Hz, 3H, H-15), 3.66 (br s, 1H, H-8a), 4.00 (d, $J = 16.0$ Hz, 1H, H-7a), 4.30 (d, $J = 16.0$ Hz, 1H, H-7b), 4.68 (d, $J = 9.7$ Hz, 1H, H-8b), 6.08 (t, $J = 7.1$ Hz, 1H, H-11), 6.43 (t, $J = 7.3$ Hz, 1H, H-12), 6.71 (dt, $J = 1.0$, 7.2 Hz, 1H, H-3), 6.80-6.86 (m, 2H, H-4 & H-5), 6.87-6.93 (m, 2H, H-10 & H-13), 7.93 (d, $J = 6.5$ Hz, 1H, H-2). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_2Cl_2) δ 8.5 (d with Pt satellites, $J_{\text{P-C}} = 1$ Hz, $^3J_{\text{Pt-C}} = 15$ Hz, C-18), 9.3 (s, C-16), 16.2-16.7 (m, C-17), 18.8-19.2 (m, C-15), 33.2 (br), 32.1 (s, C-23), 36.4 (br), 44.2 (br), 58.9 (s, C-7), 64.5 (s with Pt satellites, $^3J_{\text{Pt-C}} = 81$ Hz, C-8), 64.7 (s, C-19), 121.3 (d, $^5J_{\text{P-C}} = 3$ Hz, C-11), 124.3 (s, C-4), 125.7 (dd with Pt satellites, $^4J_{\text{P-C}} = 1$, 5 Hz, $^3J_{\text{Pt-C}} = \text{ca. } 50$ Hz, C-12), 125.9 (s, C-5), 126.9 (s, C-3), 132.4 (br s with Pt satellites, $^3J_{\text{Pt-C}} = \text{ca. } 39$ Hz, C-10), 139.0 (d, $^3J_{\text{P-C}} = 5$ Hz, C-13), 139.5 (s, C-2), 143.6 (br, C-9), 151.9 (s, C-6), 161.5 (d with Pt satellites, $^2J_{\text{P-C}} = \text{ca. } 94$ Hz, $^1J_{\text{Pt-C}} = \text{ca. } 673$ Hz, C-14), 169.5 (br, C-1). $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CD_2Cl_2) δ 0.3 (br s with Pt satellites, $^1J_{\text{Pt-P}} = 2052$ Hz, P-1), 63.5 (d with Pt satellites, $^2J_{\text{P-P}} = 10$ Hz, $^1J_{\text{Pt-P}} = 2152$ Hz, P-2). Anal. Calcd for $\text{C}_{34}\text{H}_{59}\text{BiClNP}_2\text{Pt}$: C, 41.53%; H, 6.05%; N, 1.42%. Found: C, 41.64%; H, 6.07%; N, 1.53%.

Monitoring of the reaction of 2b and Pt(PEt₃)₃ by NMR. Pt(PEt₃)₄ (26.5 mg, 0.048 mmol) was placed in a Schlenk tube and heated at 60 °C for 10 min under vacuum to form Pt(PEt₃)₃ as an orange oil. Compound **2b** (32.1 mg, 0.048 mmol) and C₆D₆ (1.2 mL) were added to the Schlenk tube to form an almost homogeneous orange mixture. The mixture was transferred to an NMR tube and the tube was sealed with a PTFE valve. The reaction progress at room temperature was periodically monitored by NMR. The progress of the reaction was shown in Figs. S3 and S7 (³¹P{¹H} NMR) and Figs. S4-S6 (¹H NMR).

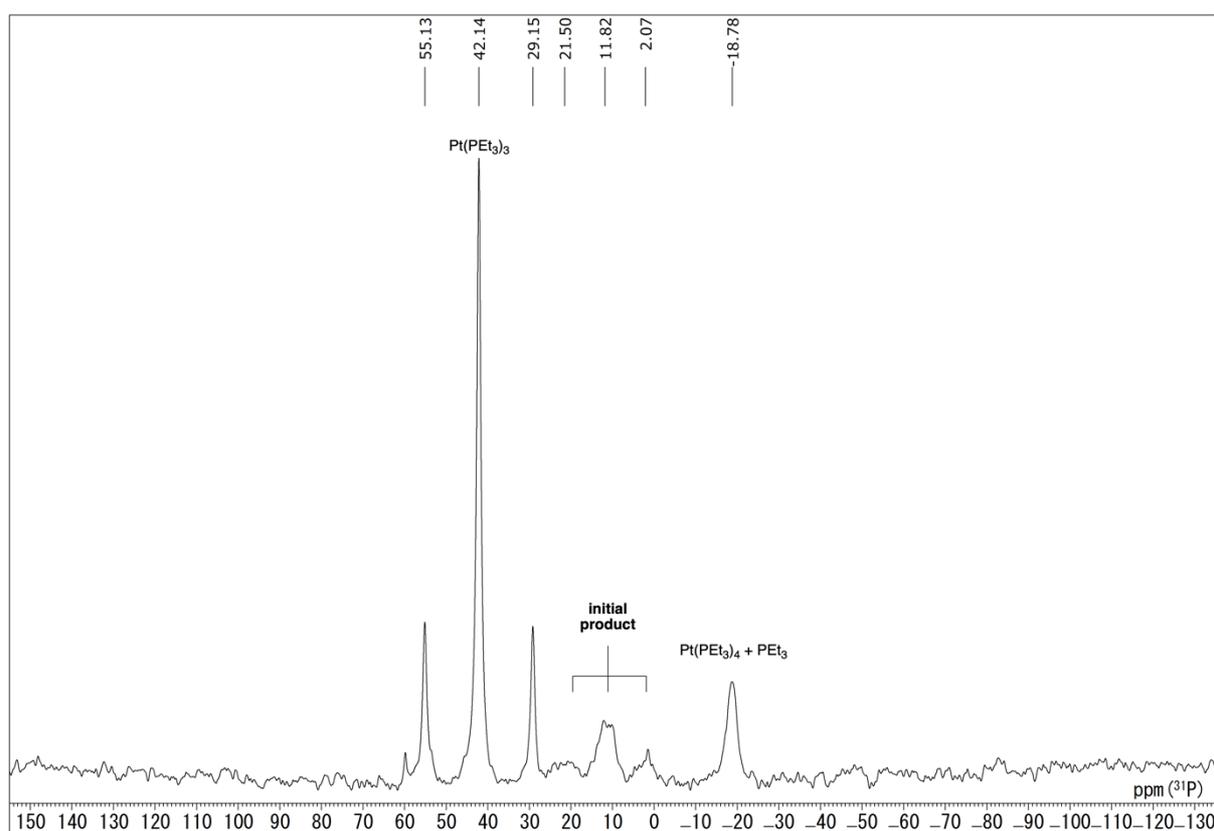


Fig. S3 ³¹P{¹H} NMR spectrum (161 MHz, C₆D₆) of the reaction mixture of **2b** and Pt(PEt₃)₃ in C₆D₆ after 30 min

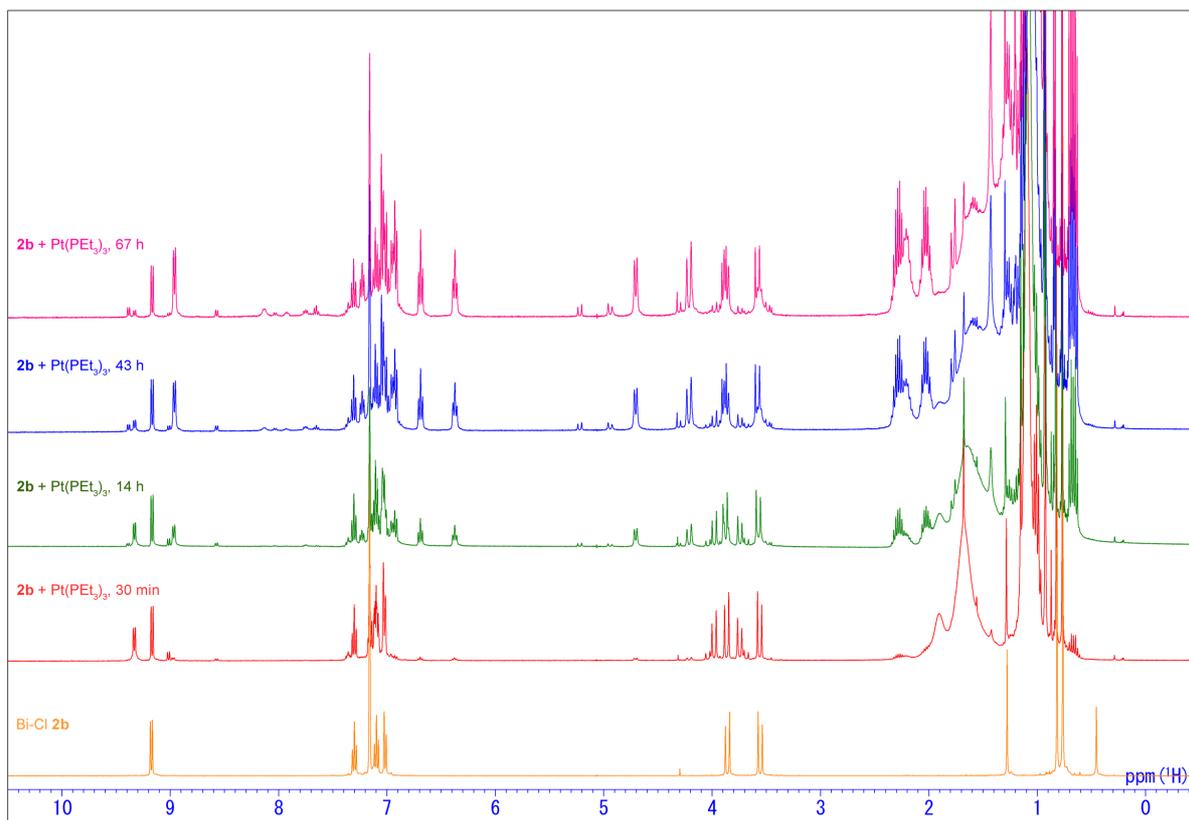


Fig. S4 ¹H NMR spectra (400 MHz, C₆D₆) of bismocine chloride **2b** (orange) and the reaction mixture of **2b** and Pt(PEt₃)₃ after 30 min (red), 14 h (green), 43 h (blue) and 67 h (pink).

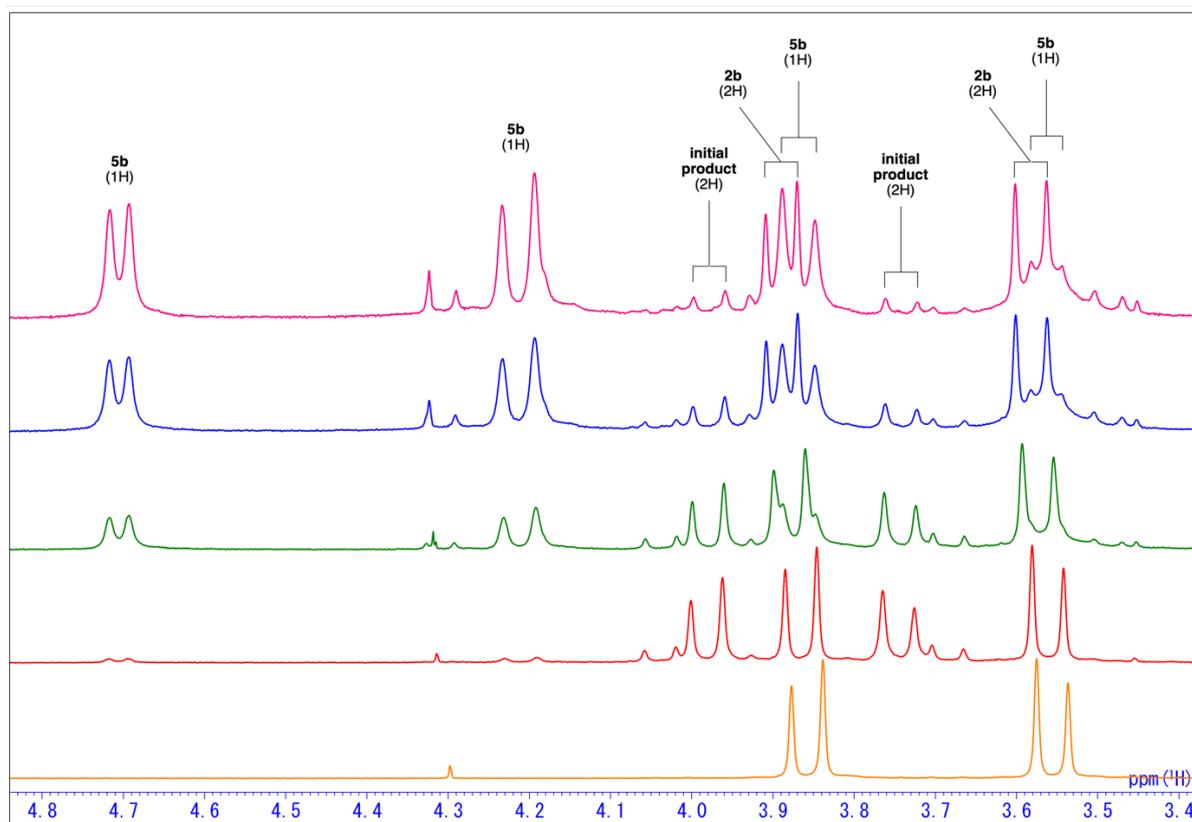


Fig. S5 Expansion (3.4 - 4.8 ppm region) of Fig. S4

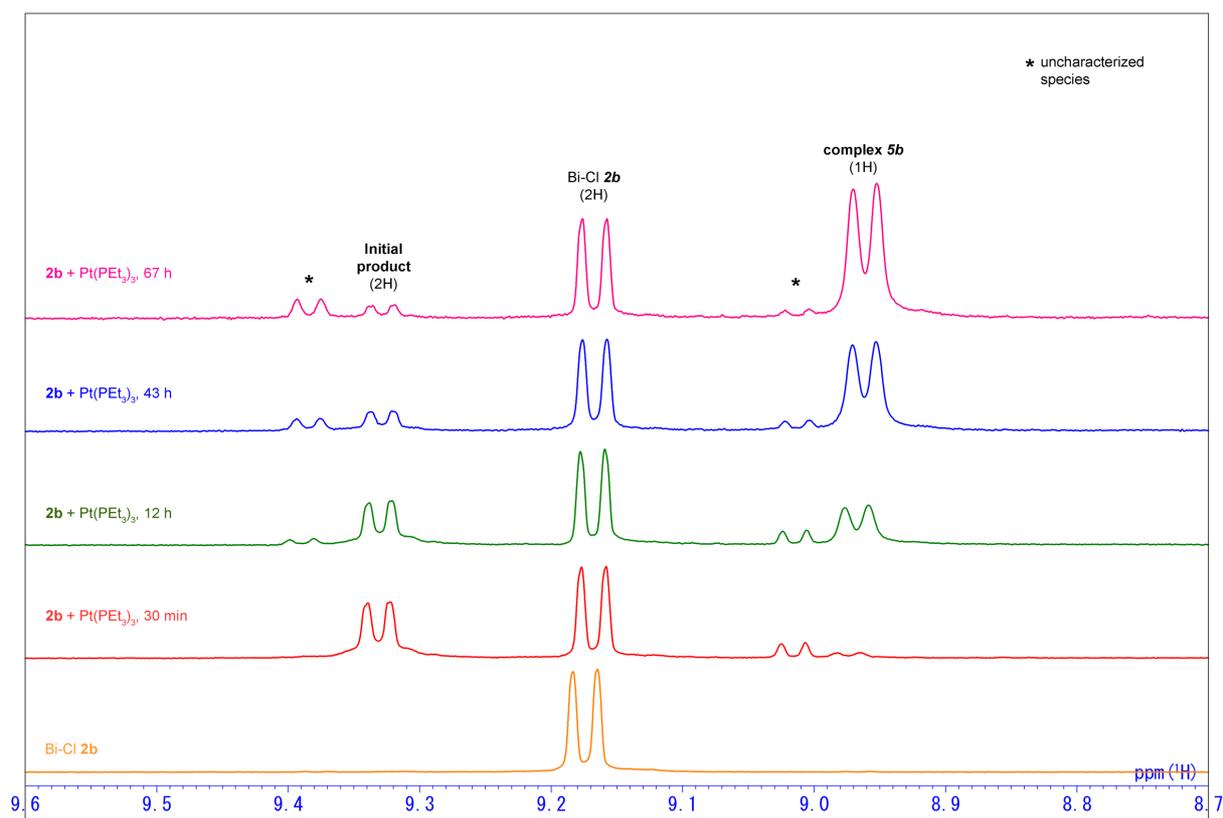


Fig. S6 Expansion (8.7 - 9.6 ppm region) of Fig. S4

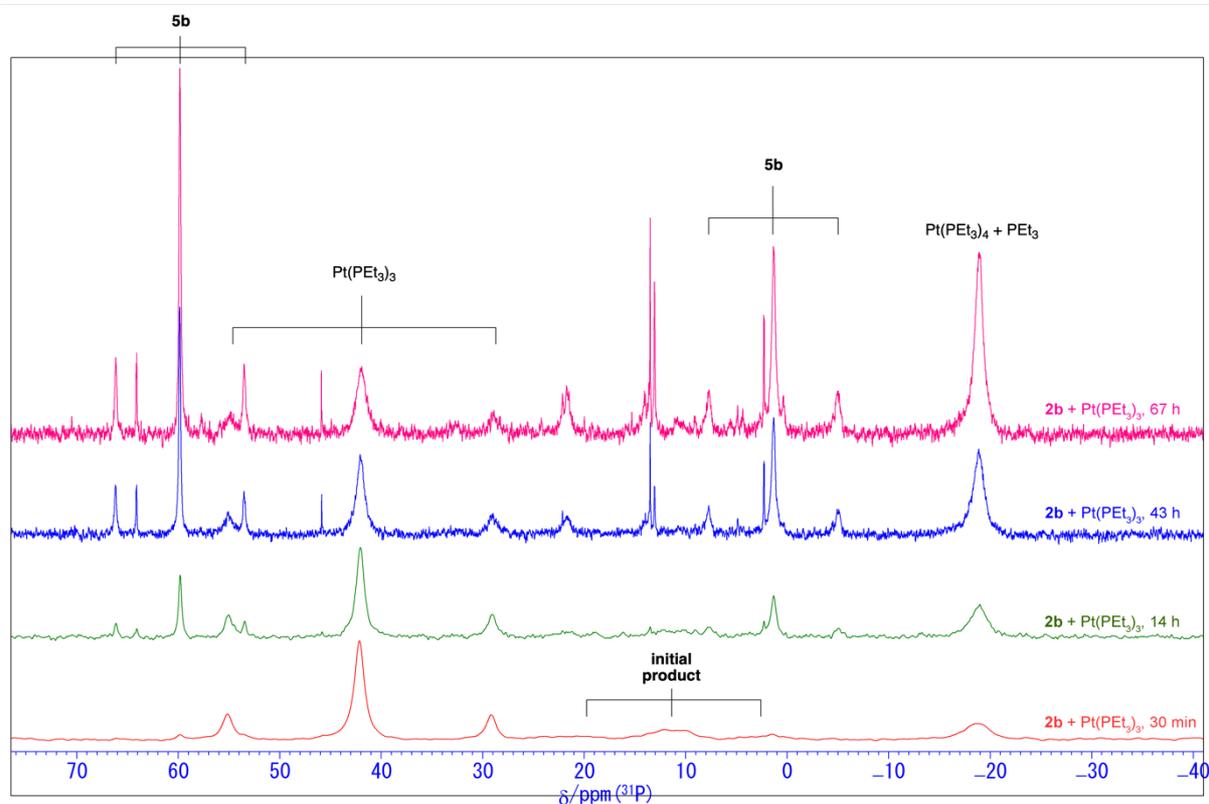
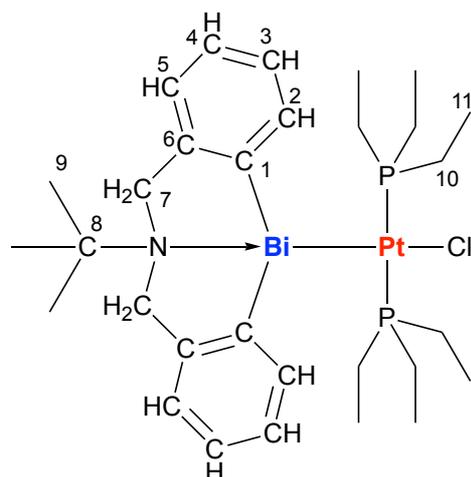


Fig. S7 $^{13}\text{P}\{^1\text{H}\}$ NMR spectra (161 MHz, C_6D_6) of the reaction mixture of **2b** and $\text{Pt}(\text{PEt}_3)_3$ after 30 min (red), 14 h (green), 43 h (blue) and 67 h (pink).

4. Synthesis of complex **6a**

Synthesis of complex **6a.** To a Schlenk tube containing $\text{Pt}(\text{PEt}_3)_3$ (0.24 mmol, prepared from 159 mg of $\text{Pt}(\text{PEt}_3)_4$ by heating at 60 °C under vacuum) was added a THF (12 mL) solution **2a** (90 mg, 0.18 mmol). The clear light orange solution was stirred for 30 min at room temperature. Then the volatiles were removed under vacuum to leave brown-yellow residue. The residue was extracted with hexane (1×10 mL, 3×5 mL). Each extract was kept in a freezer at -35 °C. From the first extract, **6a** was obtained as yellow crystals (42 mg, 25% yield). ^1H NMR (600 MHz, C_6D_6 , Fig. S38) δ 0.93-1.06 (m, 27H, H-9 & H-11), 1.89 (very broad s, 12H, H-10), 3.69 (d, $J = 15.4$ Hz, 2H, H-7), 3.85 (d, $J = 15.4$ Hz, 2H, H-7), 6.98 (dd, $J = 1.0, 7.3$ Hz, 2H, H-5), 7.10 (dt, $J = 1.5, 7.2$ Hz, 2H, H-3), 7.15 (dt, $J = 1.6, 7.2$ Hz, 2H, H-4), 9.34 (dd, $J = 1.5, 7.0$ Hz, 2H, H-2). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, C_6D_6 , Figs. S39-S41) δ 9.3 (br s, C-11), 17.0 (br s, C10), 27.9 (s, C-9), 57.1 (s, C-8), 57.3 (s, C-7), 127.25 (C-5), 127.33 (C-4), 128.8 (C-3), 142.4 (br s, C-1), 145.8 (s with Pt satellites, $^3J_{\text{Pt-C}} = 46$ Hz, C-2), 148.6 (C-6). ^{31}P NMR (243 MHz, C_6D_6 , Fig. S42) δ 0-21 (very broad multiplet). Anal. Calcd for $\text{C}_{30}\text{H}_{51}\text{BiClINP}_2\text{Pt}$: C, 38.86%; H, 5.54%; N, 1.51%. Found: C, 38.73%; H, 5.59%; N, 1.51%.



Atomic numbering of complex **6a** for the NMR signal assignment

5. Monitoring the change of complex **6a** in C_6D_6 or $\text{THF-}d_8$ by ^1H NMR

Complex **6a** (ca. 2 mg) was dissolved in C_6D_6 or $\text{THF-}d_8$ (0.5 mL). ^1H NMR spectra of the solution were measured periodically during 4 days (Fig. S8 (C_6D_6) and Fig. S9 ($\text{THF-}d_8$)). Another sample containing **6a** (ca. 2 mg) and PEt_3 (20 wt% toluene solution, 2.5 μL , ca. 2 equiv) was also prepared and monitored by ^1H NMR (Fig. S10).

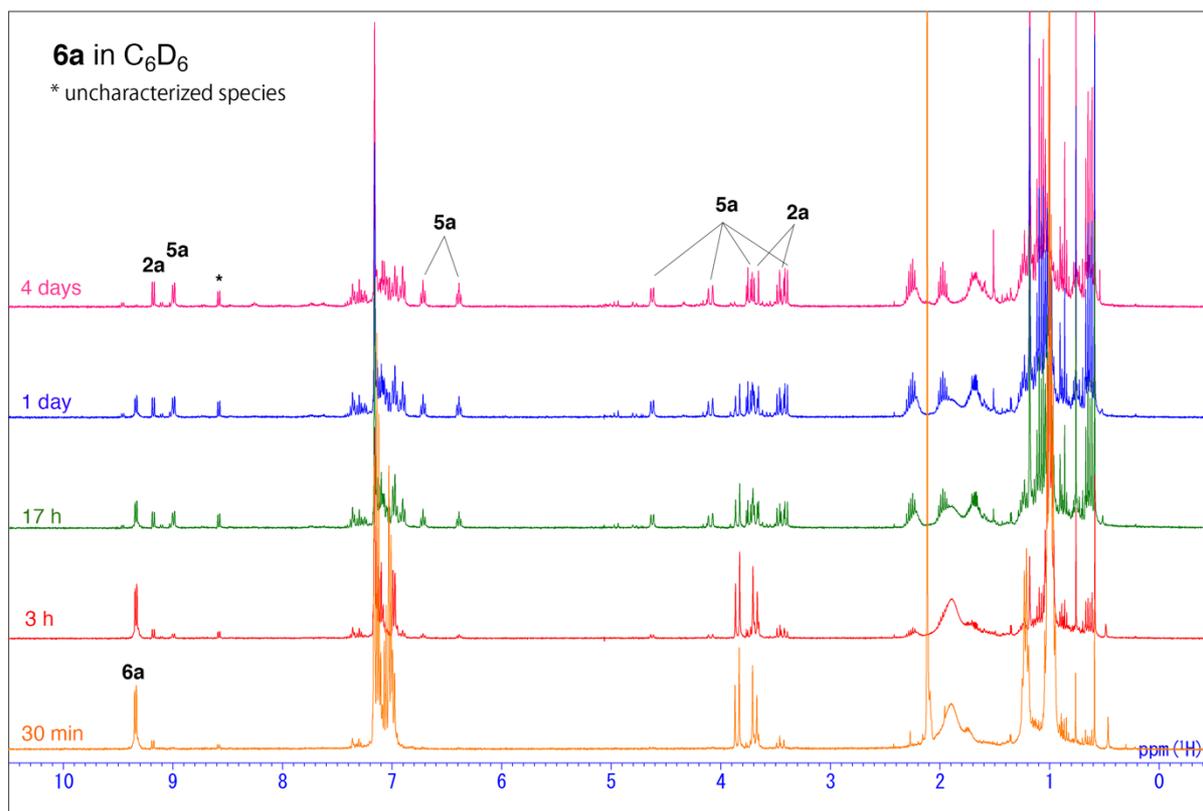


Fig. S8 Change of ¹H NMR spectra (400 MHz) of the C₆D₆ solution of **6a** after 30 min (orange), 3 h (red), 17 h (green), 1 day (blue) and 4 days (pink).

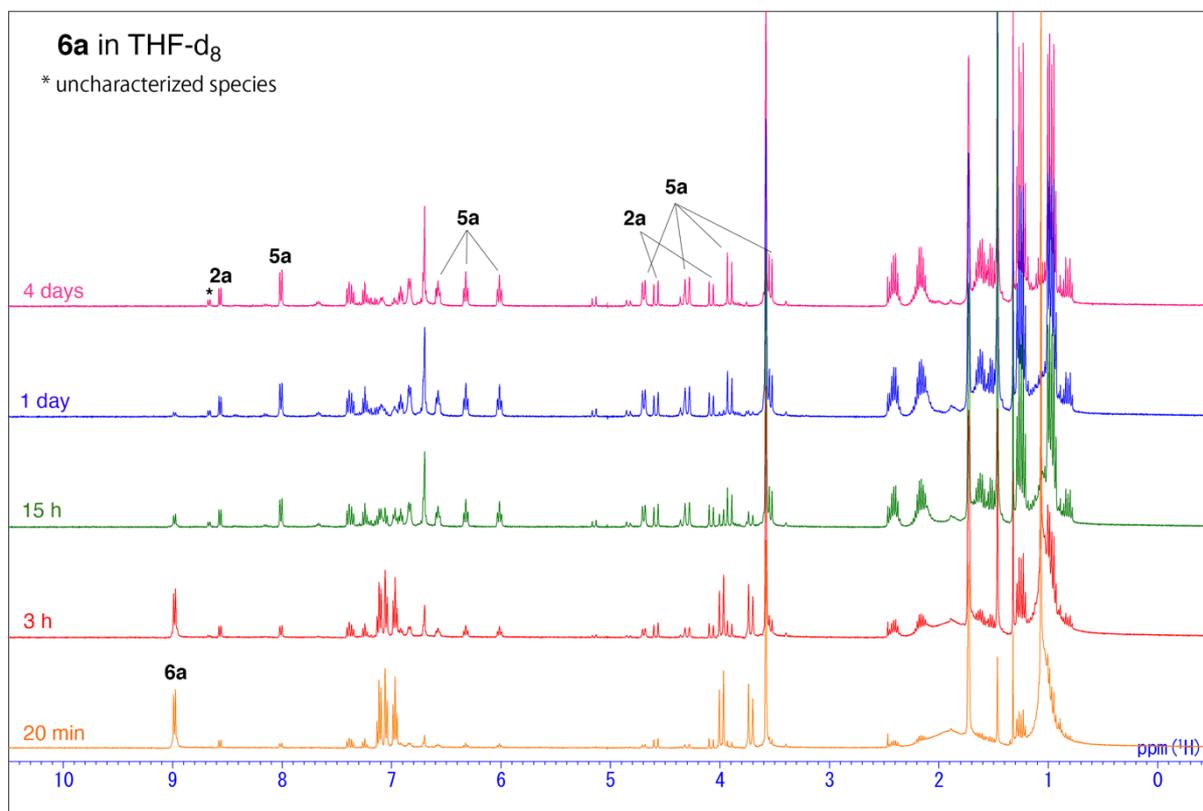


Fig. S9 Change of ¹H NMR spectra (400 MHz) of the THF-*d*₈ solution of **6a** after 20 min (orange), 3 h (red), 15 h (green), 1 day (blue) and 4 days (pink).

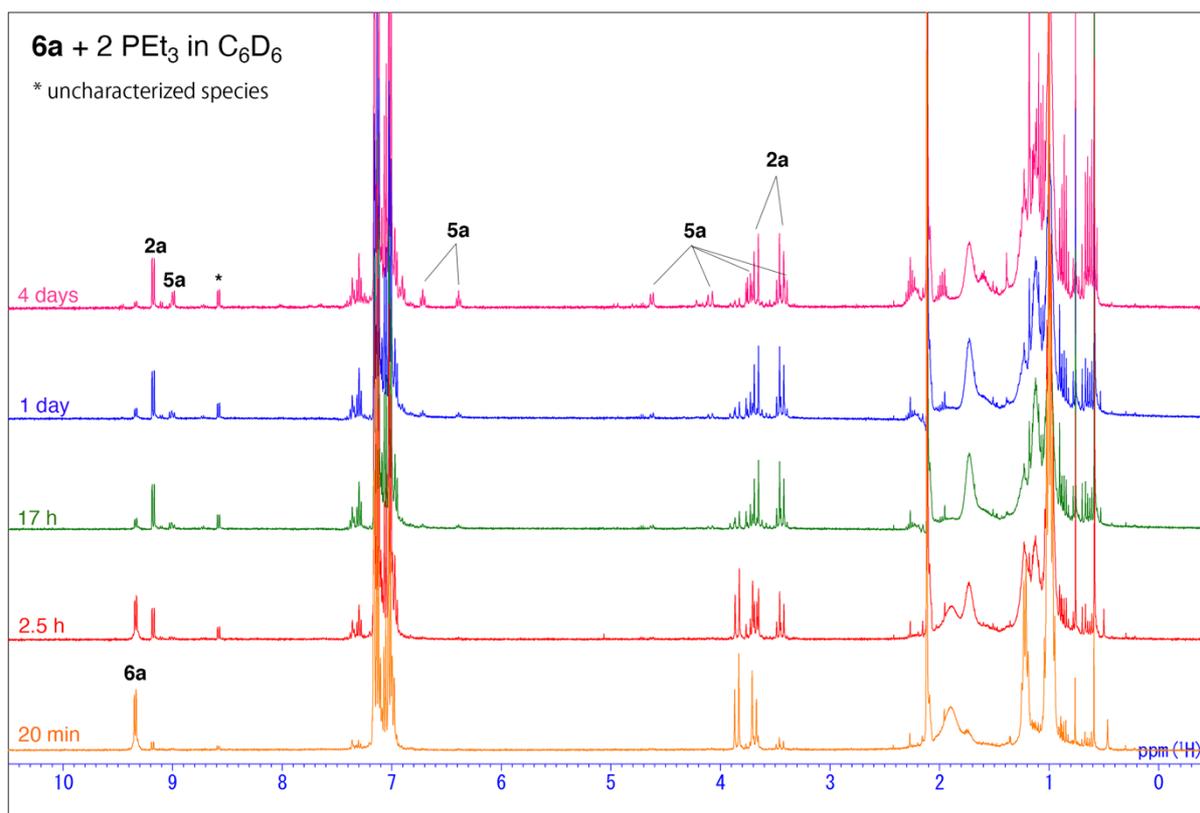


Fig. S10 Change of ¹H NMR spectra (400 MHz) of the C₆D₆ solution of **6a** and PEt₃ (ca. 2 equiv) after 20 min (orange), 2.5 h (red), 17 h (green), 1 day (blue) and 4 days (pink).

6. Single crystal X-ray structure analysis

Data collection of complexes **4** and **5a** was performed at $-120\text{ }^{\circ}\text{C}$ on a Bruker APEX II diffractometer using graphite monochromated Mo K α radiation. The determination of crystal class and unit cell parameters was carried out with the APEX2 program package.^{S5} The raw frame data were processed using SAINT^{S6} and SADABS^{S7} to yield the reduction data file. Structure solution and refinement were performed using CrystalStructure software package^{S8} with PATTY^{S9} and SHELXL^{S10} programs.

The crystals of **6a** was obtained directly from the hexane extract of the crude reaction mixture. Although the crystals were very small and their quality was not very good, recrystallization could not be attained due to the instability of **6a** in solution. Data collection of complex **6a** was performed at $-180\text{ }^{\circ}\text{C}$ on a Rigaku XtaLAB P200 diffractometer using multi-layer mirror optics monochromated Mo K α radiation. The determination of crystal class and unit cell parameters was carried out with the CrysAlisPro program package.^{S11} The raw frame data were processed using CrysAlisPro to yield the reduction data file. Structure solution and refinement were performed using Olex2 software package^{S12} with SHELXT and SHELXL programs.^{S10}

CCDC 1991384, 1991385 and 2043453 contain the supplementary crystallographic data for complexes **4**, **5a** and **6a**. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre at www.ccdc.cam.ac.uk/data_request/cif.

7. References:

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- S5 APEX2 Version 2009.9 ; Bruker AXS Inc., Madison, WI 2009.
- S6 SAINT Version 7.68A; Bruker AXS Inc., Madison, WI 2009.
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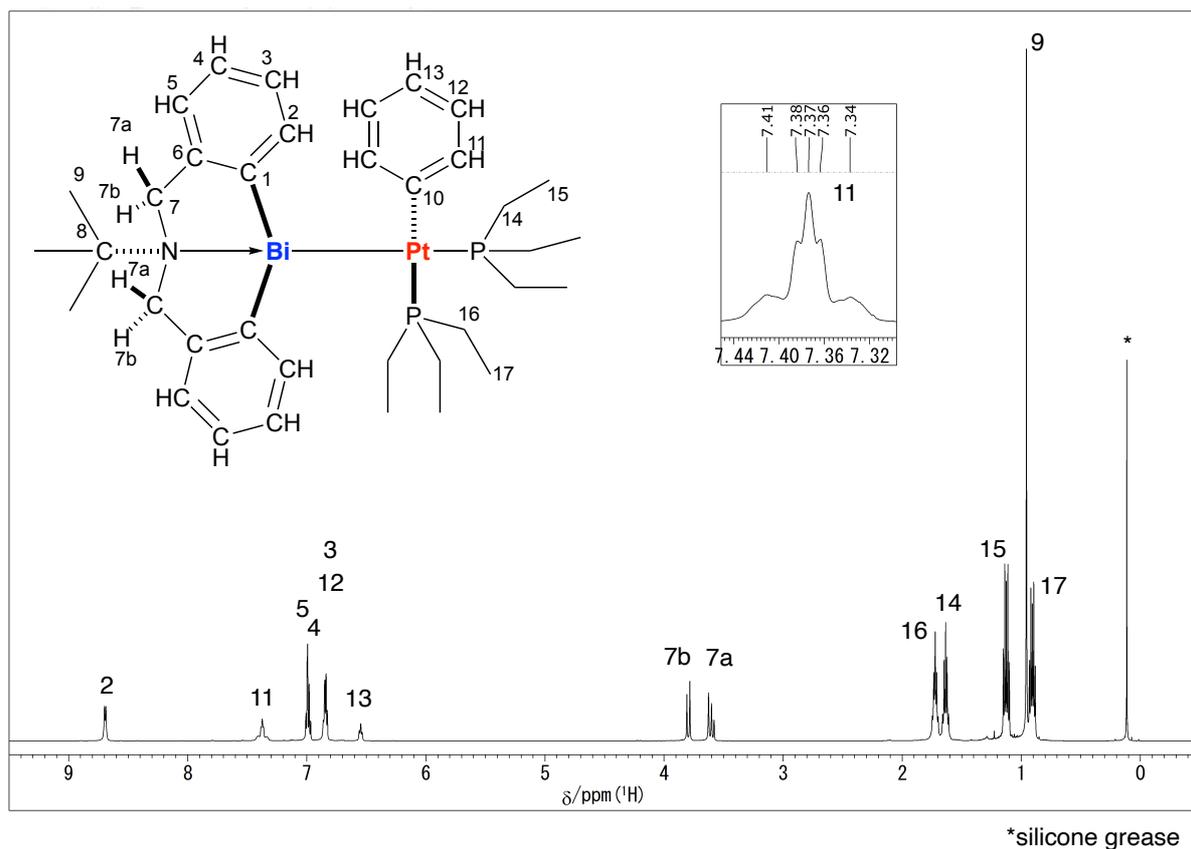


Fig. S13 ^1H NMR spectrum (600 MHz) of complex **4** in $\text{THF-}d_8$.

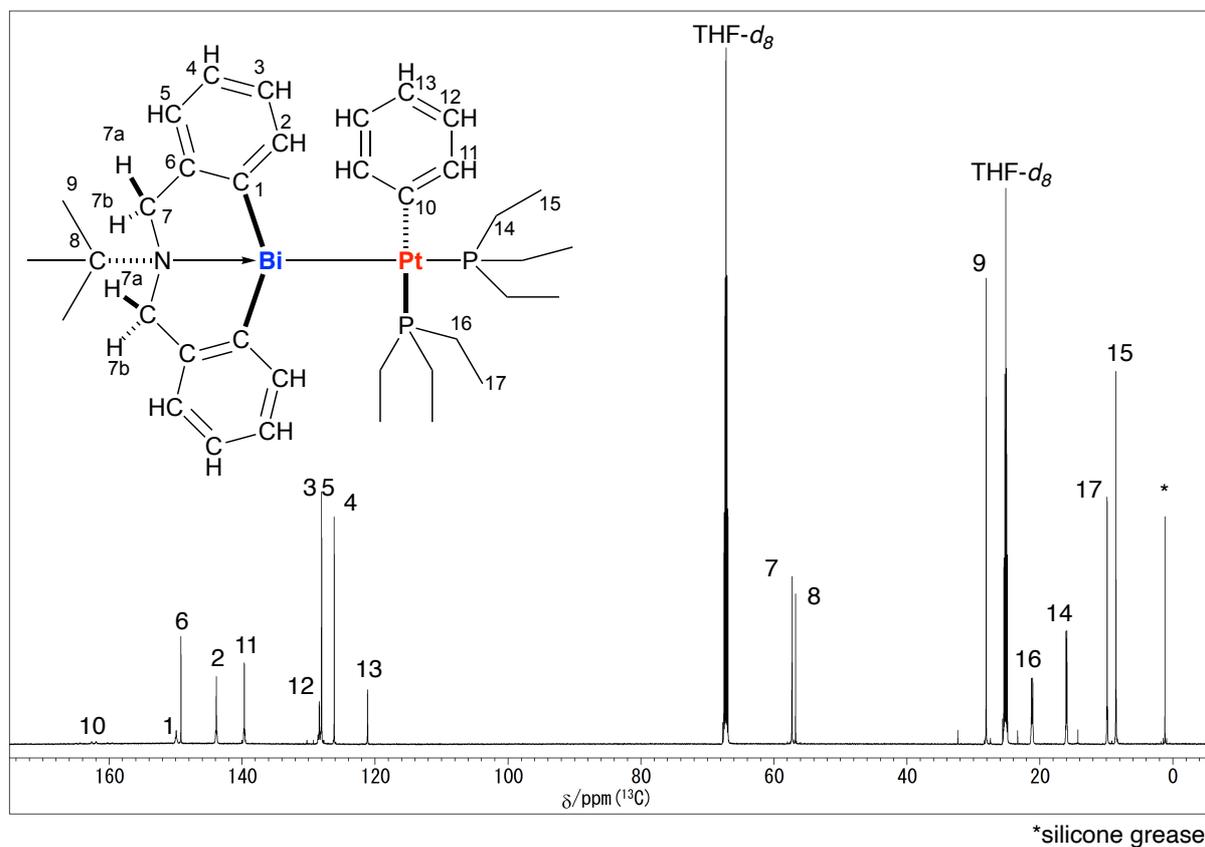


Fig. S14 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) of complex **4** in $\text{THF-}d_8$.

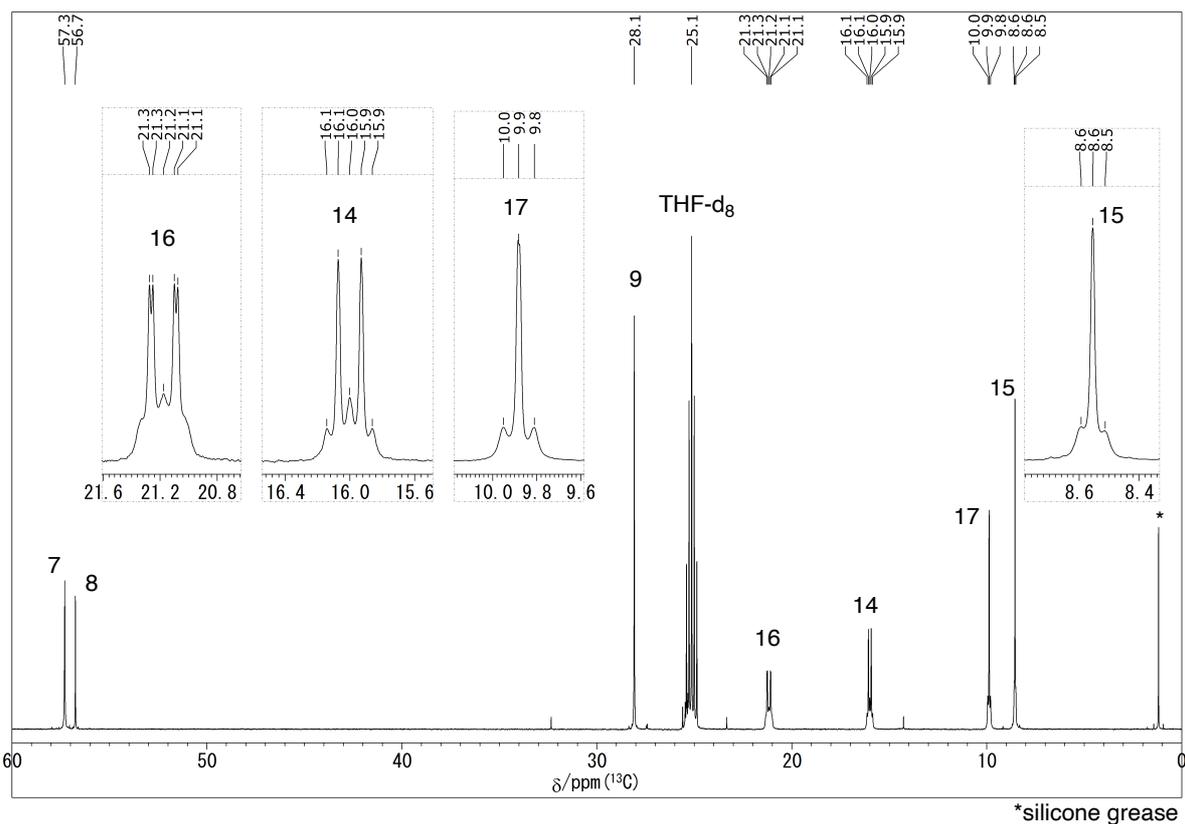


Fig. S15 A part (0–60 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) of complex **4** in $\text{THF-}d_8$.

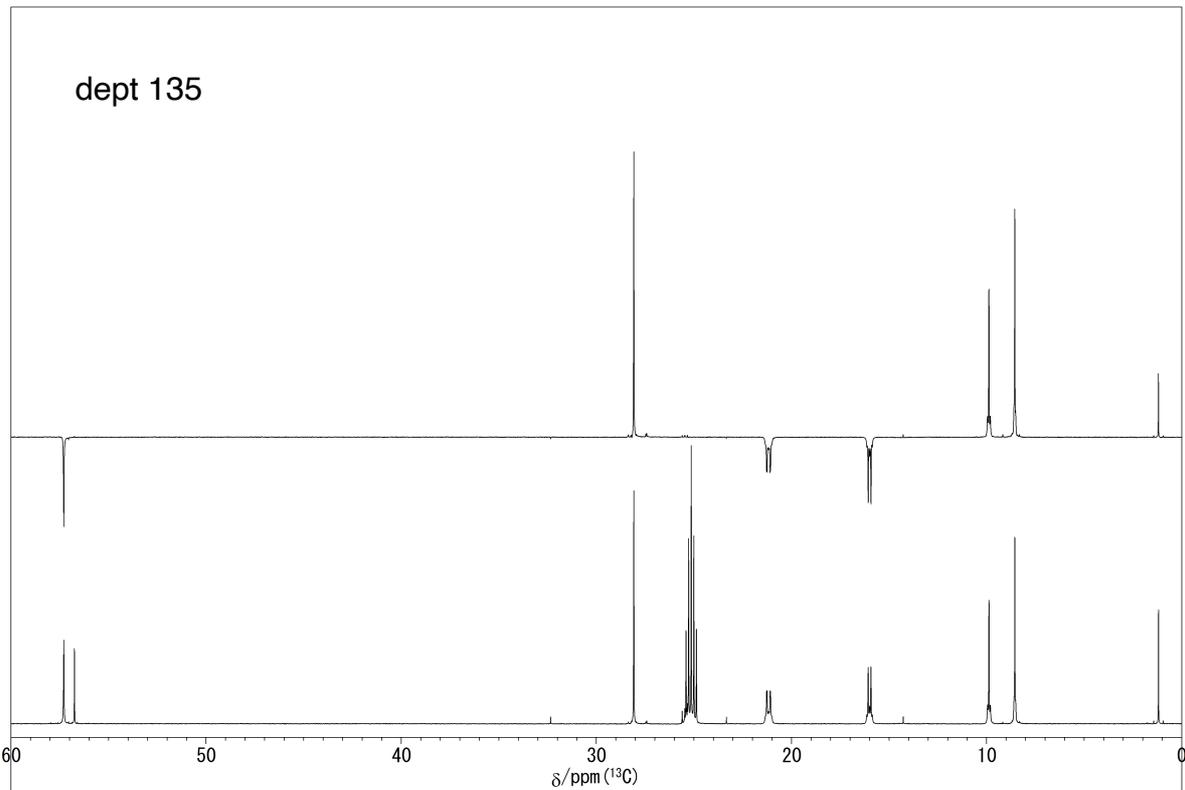


Fig. S16 A part (0–60 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) with dept 135 spectrum (upper) of complex **4** in $\text{THF-}d_8$.

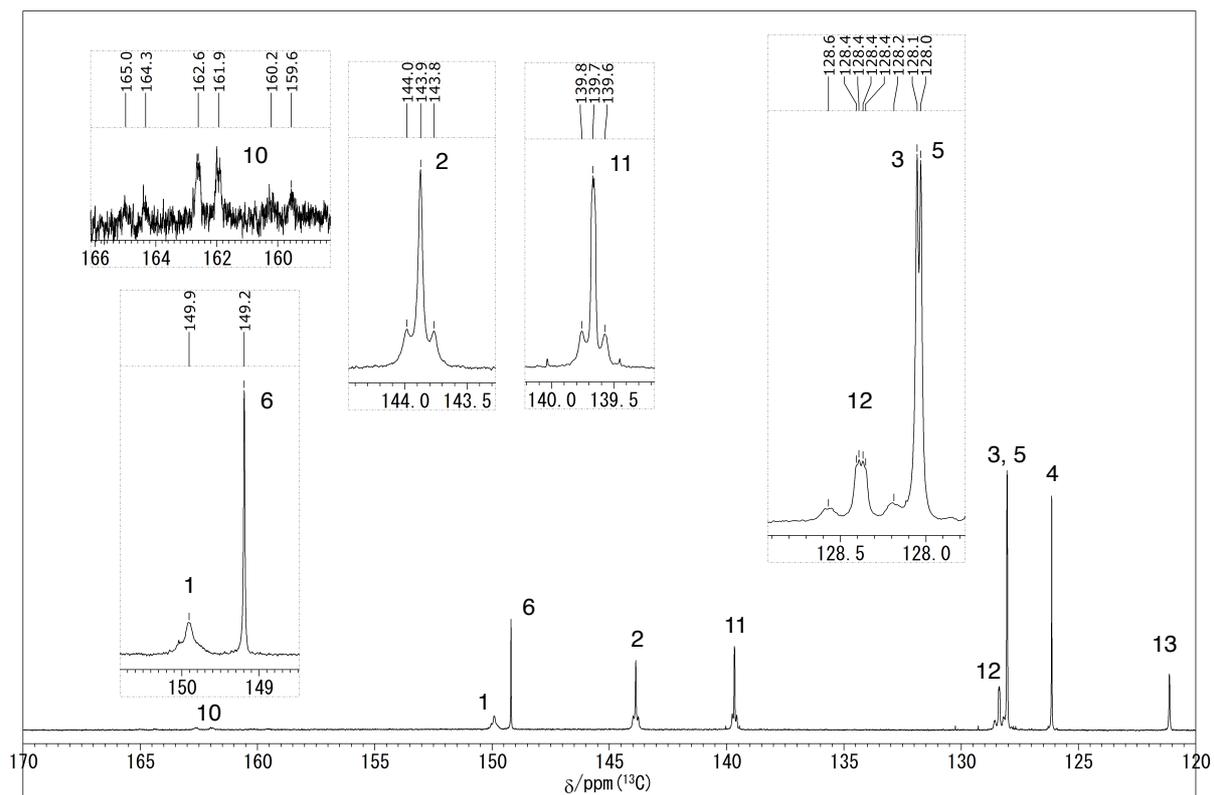


Fig. S17 A part (120–170 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) of complex **4** in $\text{THF-}d_8$.

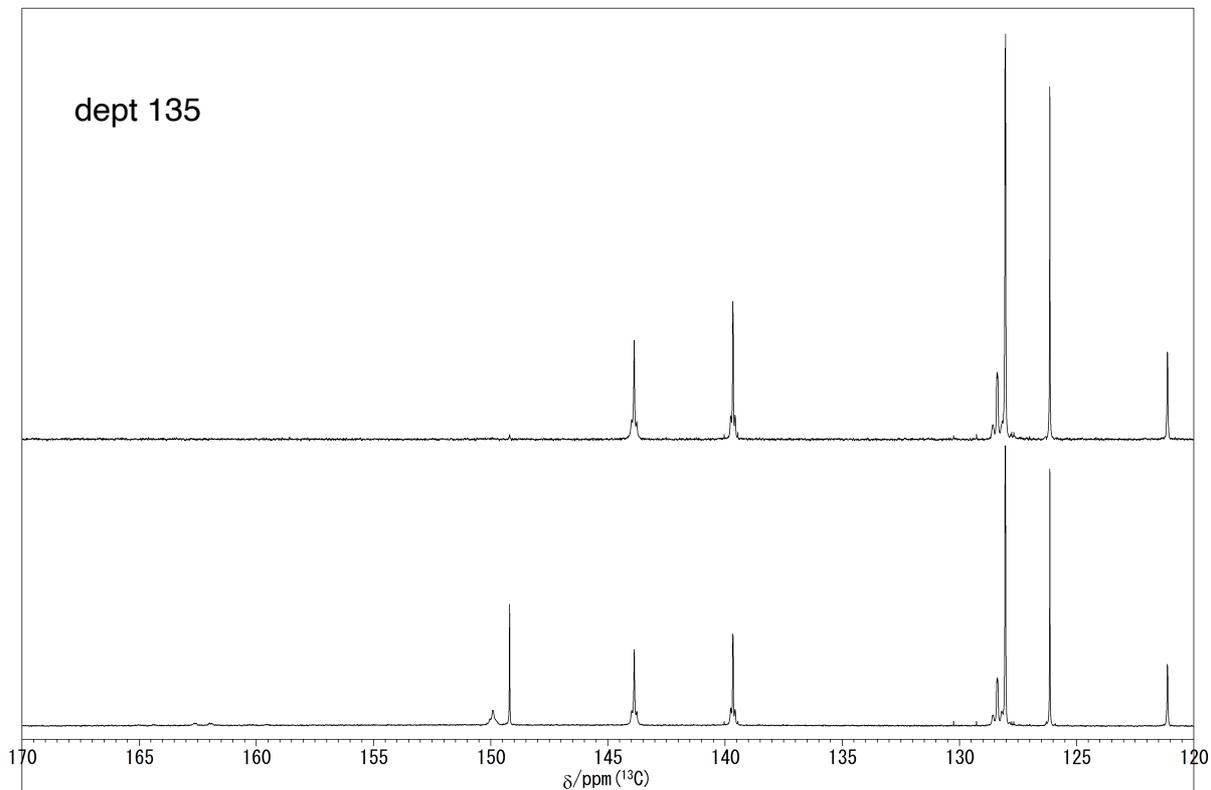


Fig. S18 A part (120–170 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) with dept 135 spectrum (upper) of complex **4** in $\text{THF-}d_8$.

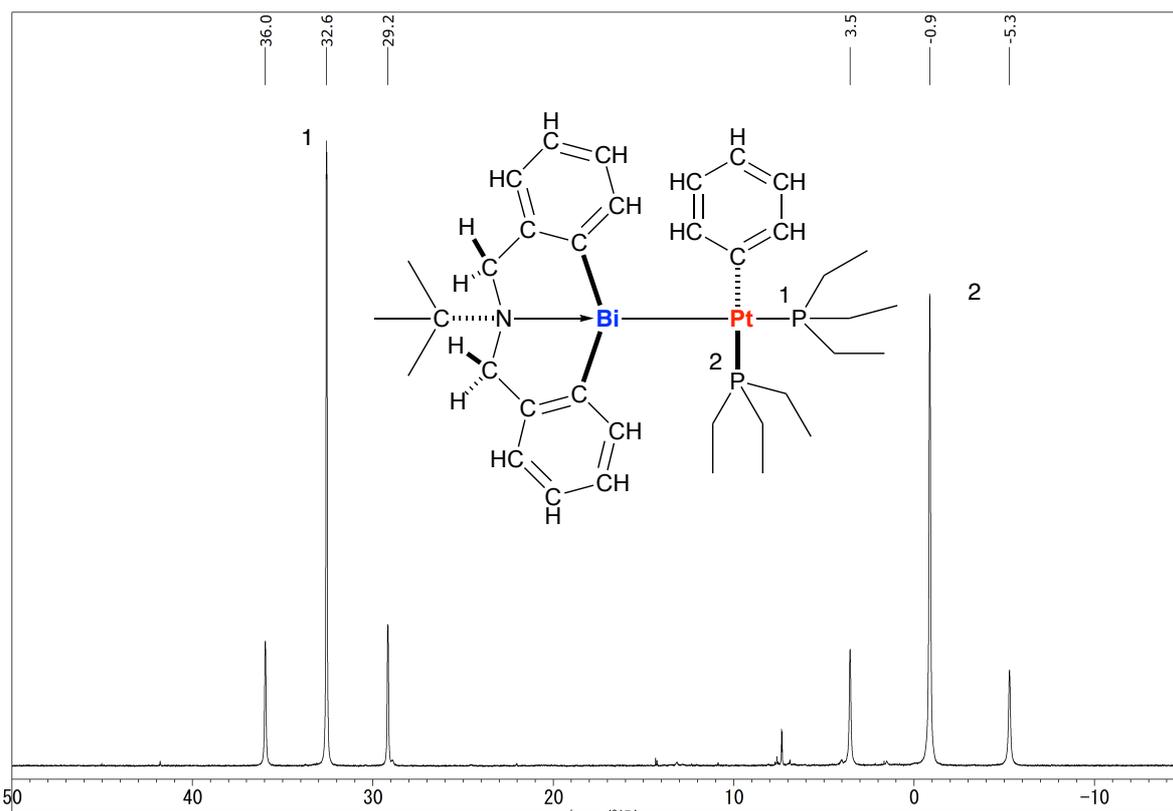


Fig. S19 $^{13}\text{P}\{^1\text{H}\}$ NMR spectrum (243 MHz) of complex 4 in THF- d_8 .

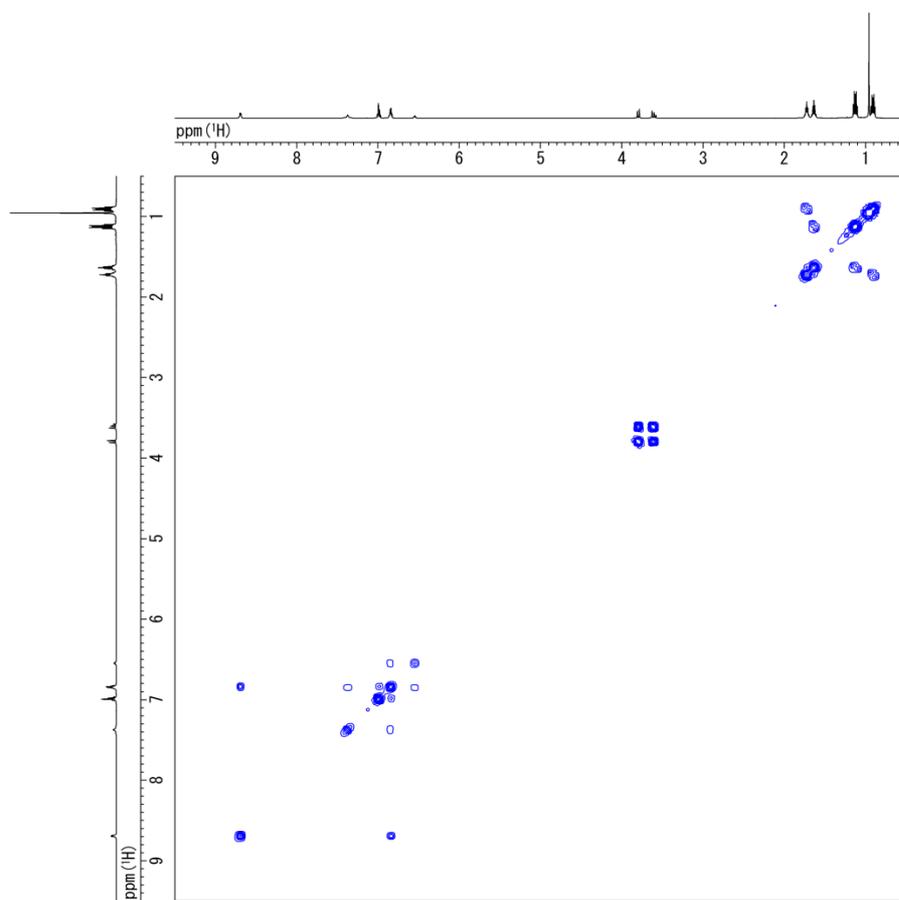


Fig. S20 $^1\text{H}-^1\text{H}$ COSY NMR spectrum (600 MHz) of complex 4 in THF- d_8 .

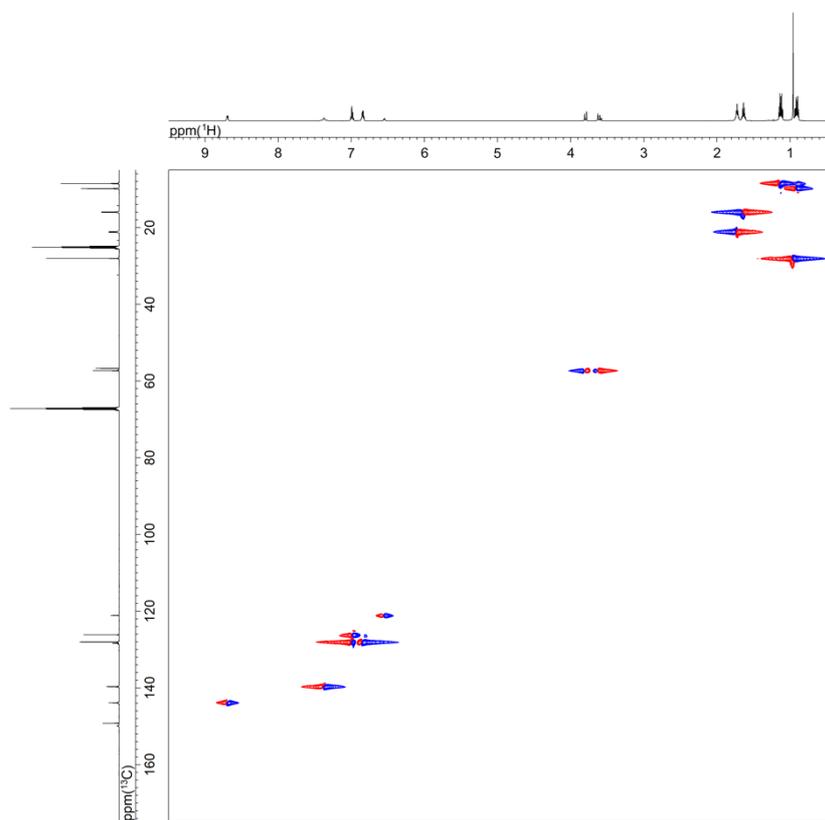


Fig. S21 ^1H - ^{13}C HSQC NMR spectrum (600 MHz for ^1H , 151 MHz for ^{13}C) of complex **4** in $\text{THF-}d_8$.

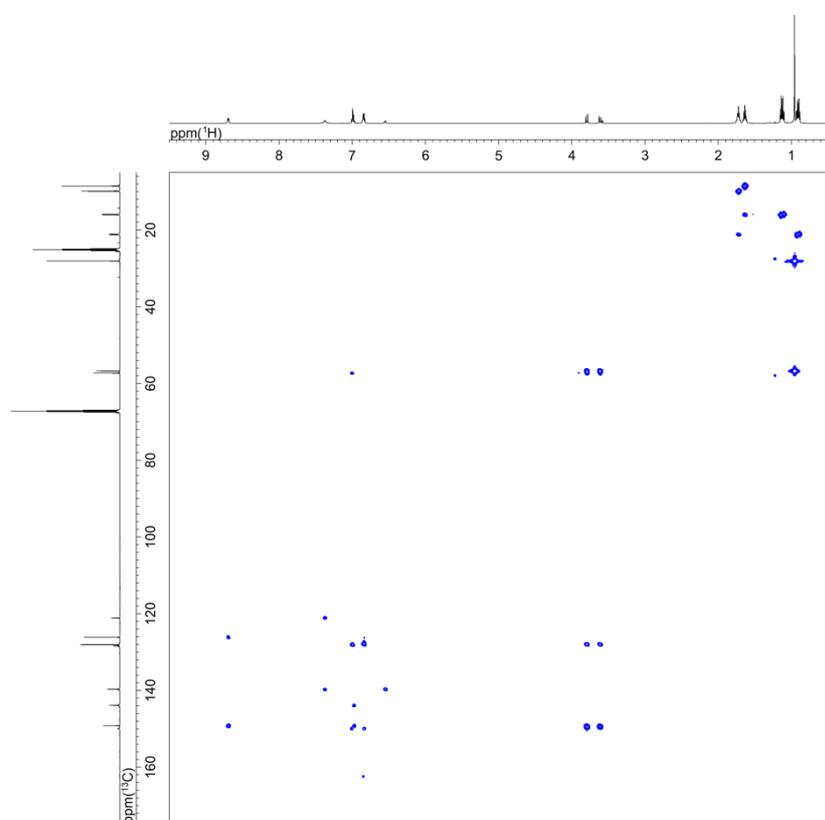


Fig. S22 ^1H - ^{13}C HMBC NMR spectrum (600 MHz for ^1H , 151 MHz for ^{13}C) of complex **4** in $\text{THF-}d_8$.

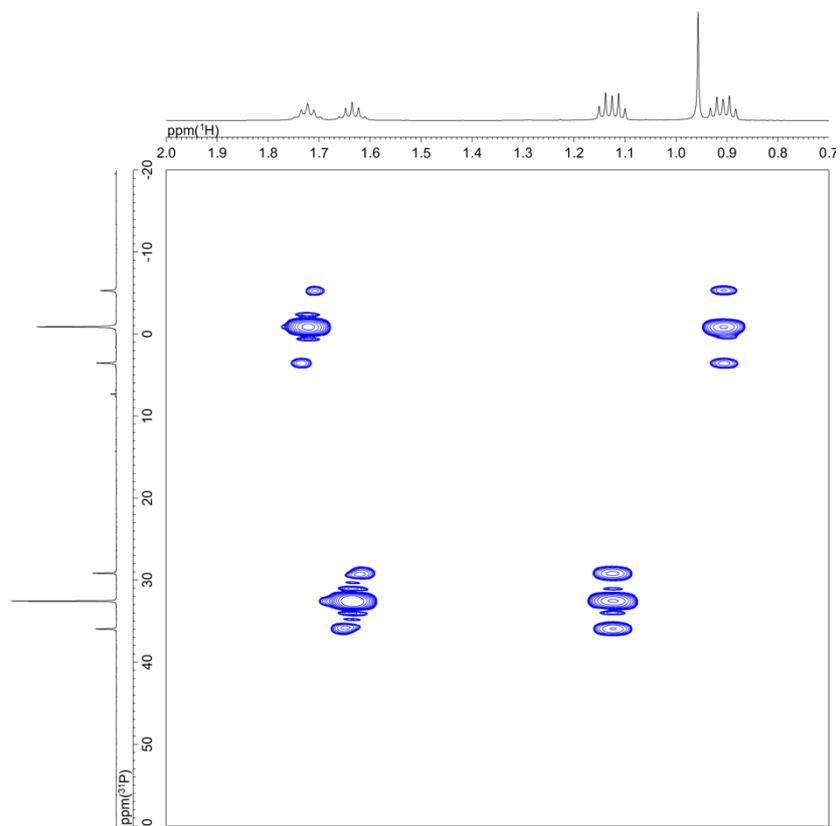


Fig. S23 ^1H - ^{31}P HMBC NMR spectrum (600 MHz for ^1H , 243 MHz for ^{31}P) of complex **4** in $\text{THF-}d_8$.

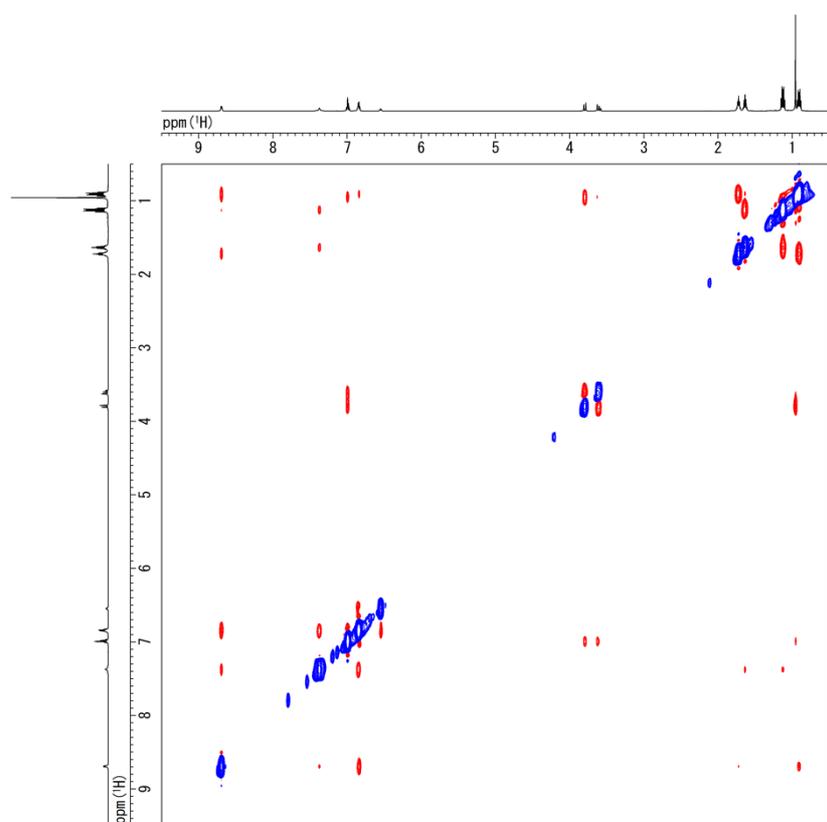


Fig. S24 NOESY NMR spectrum (600 MHz for ^1H) of complex **4** in $\text{THF-}d_8$.

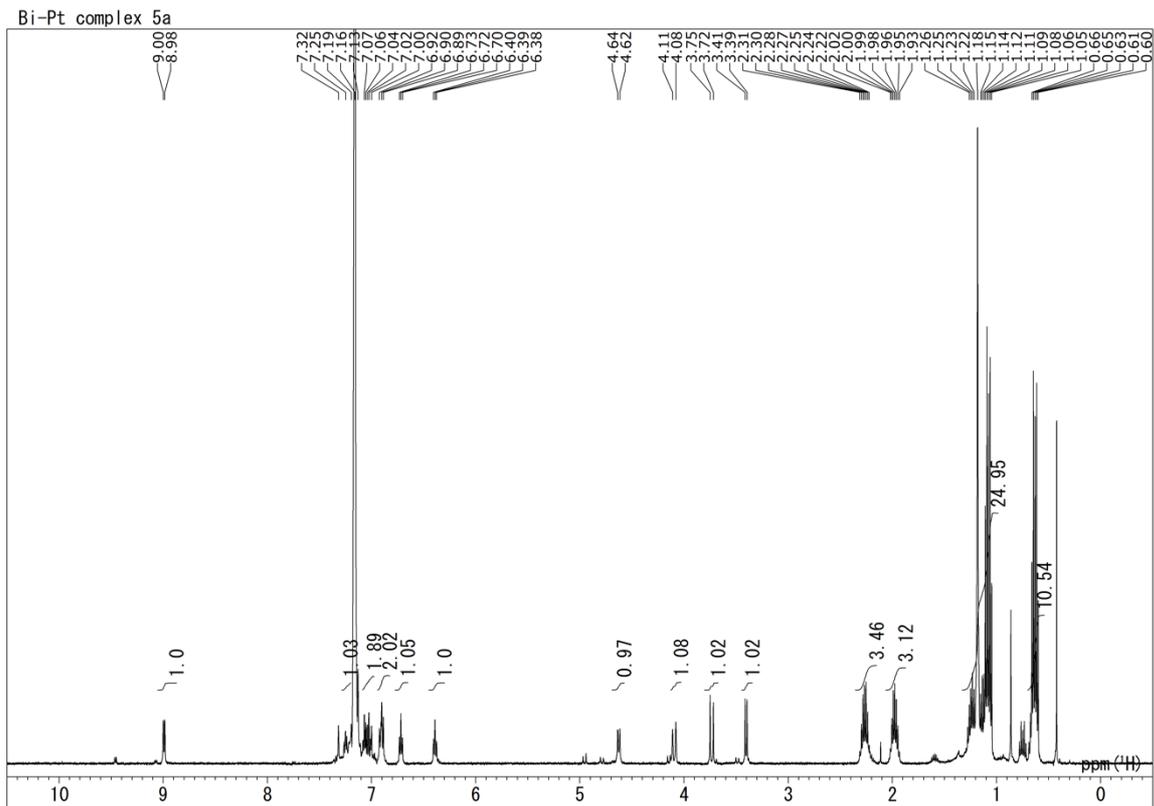


Fig. S25 ^1H NMR spectrum (499 MHz) of complex **5a** in C_6D_6 .

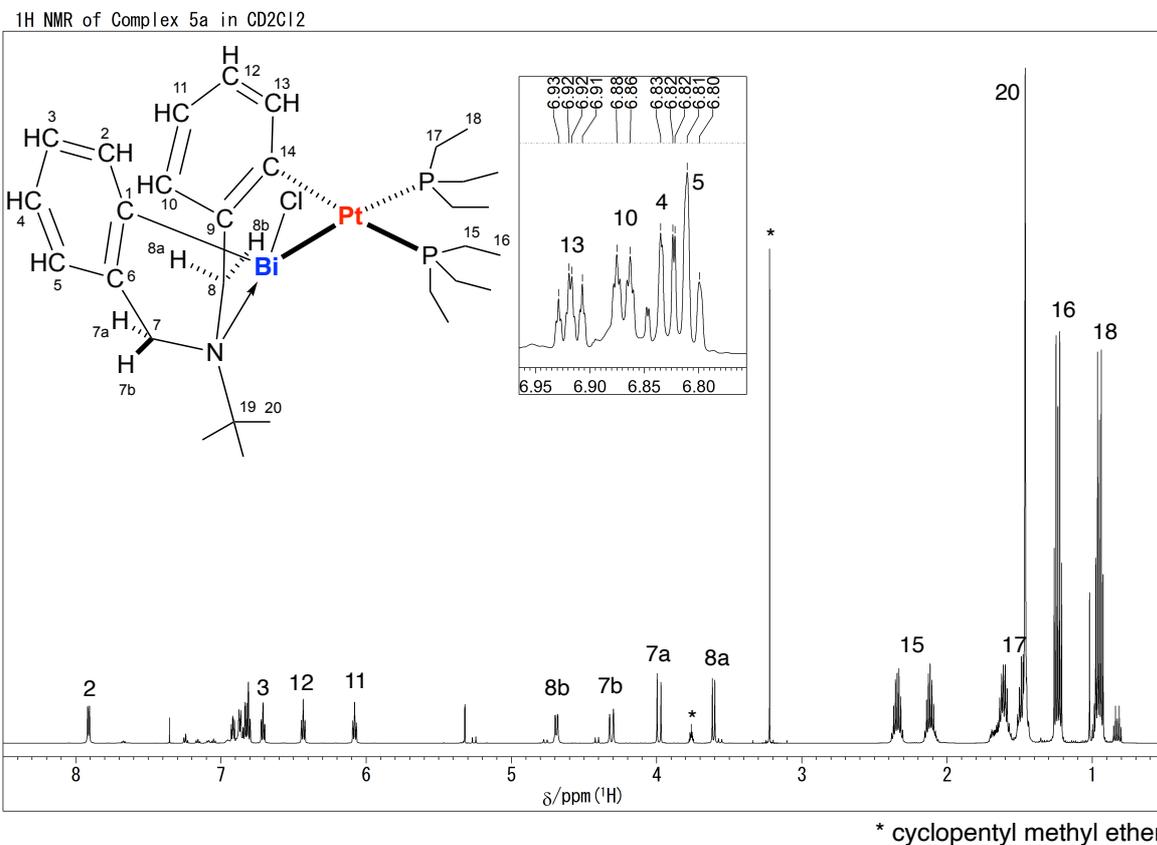


Fig. S26 ^1H NMR spectrum (600 MHz) of complex **5a** in CD_2Cl_2 .

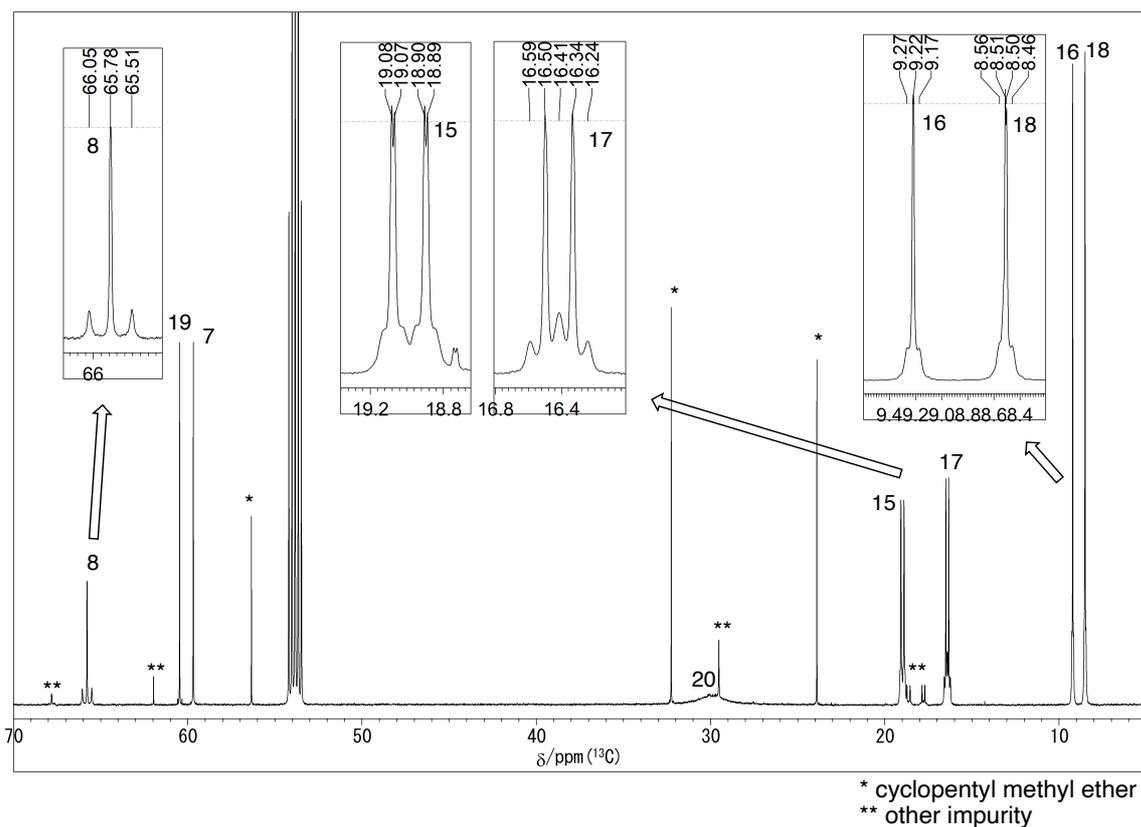


Fig. S29 A part (120–140 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) of complex **5a** in CD_2Cl_2 .

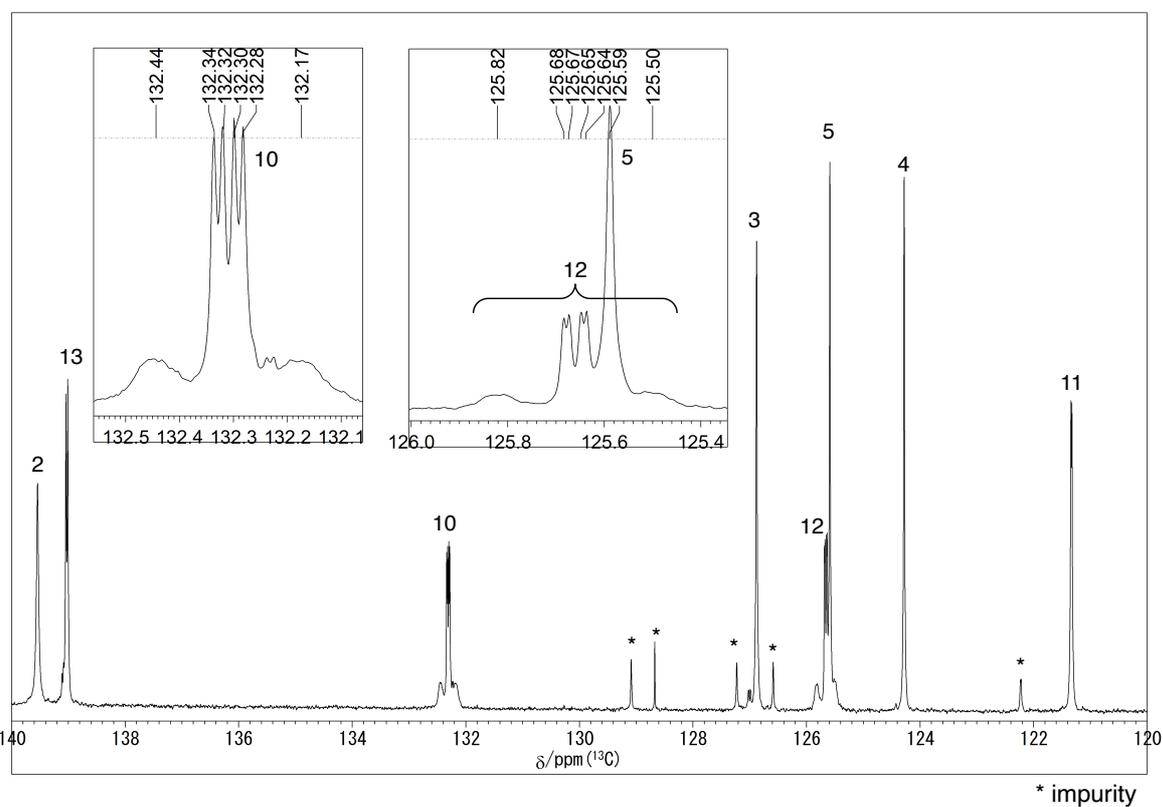


Fig. S30 A part (120–140 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) of complex **5a** in CD_2Cl_2 .

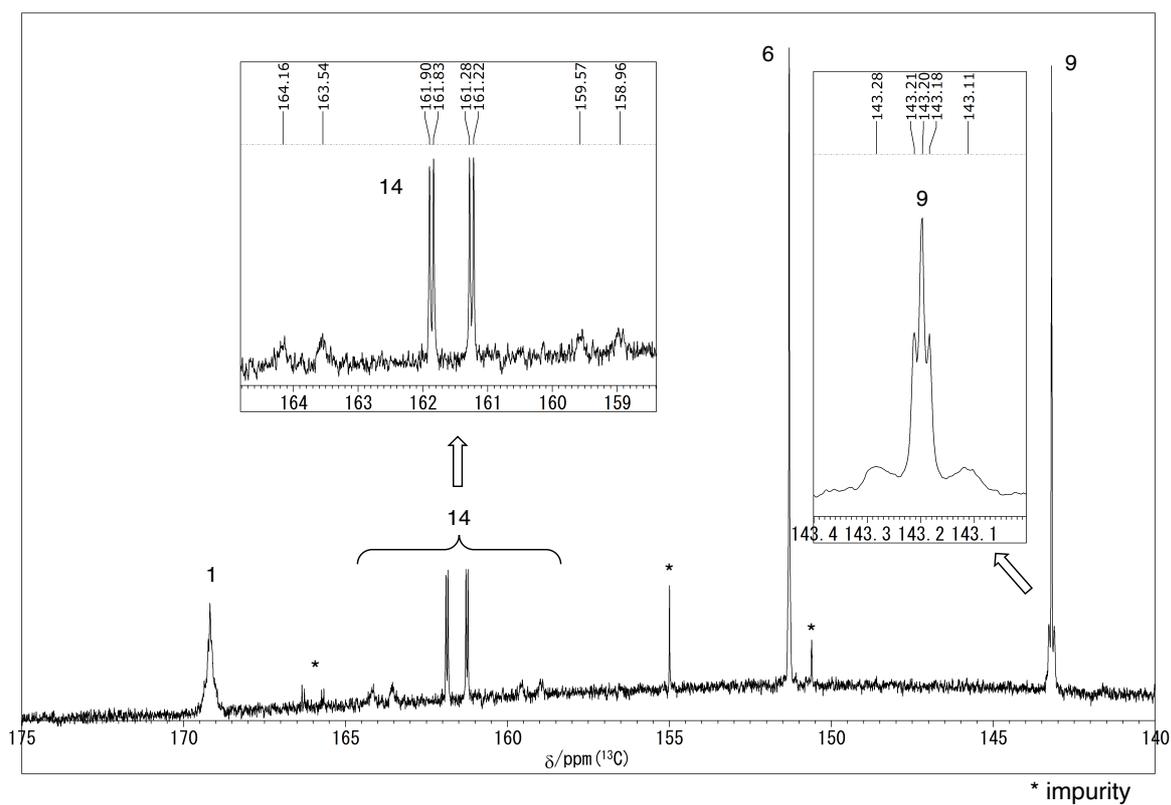


Fig. S31 A part (140–175 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) of complex **5a** in CD_2Cl_2 .

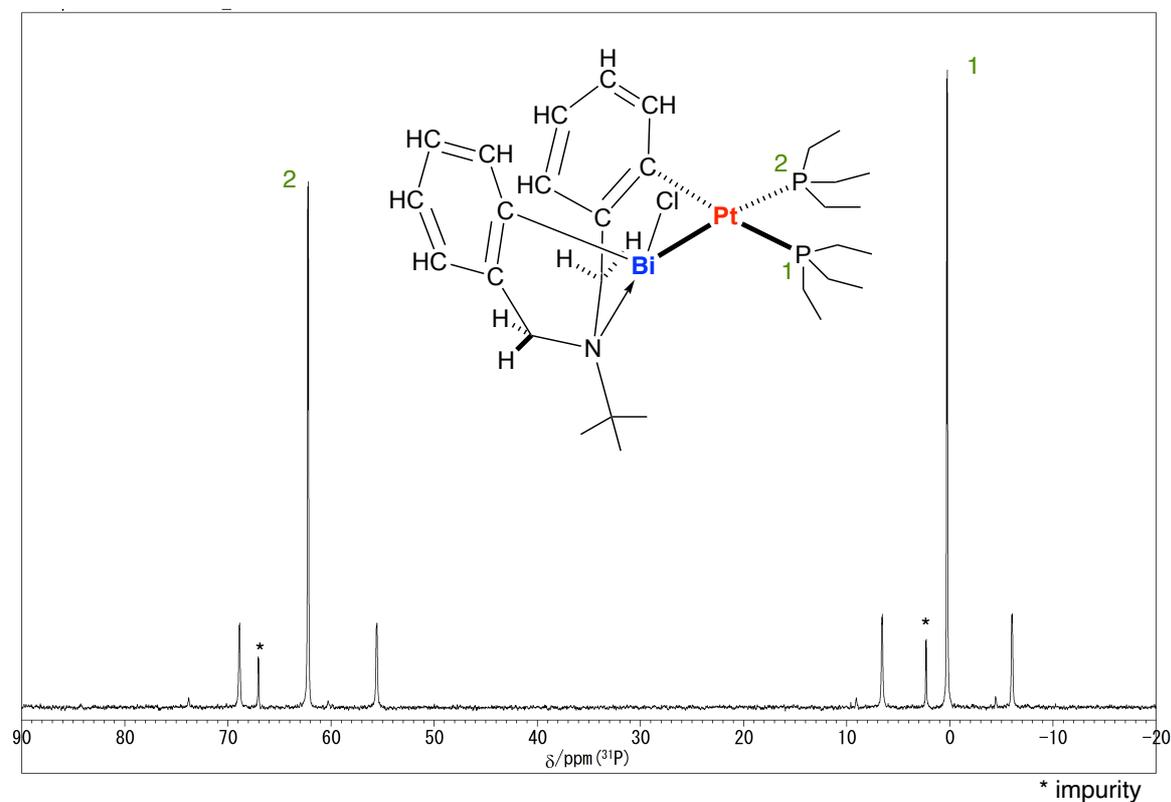


Fig. S32 $^{13}\text{P}\{^1\text{H}\}$ NMR spectrum (162 MHz) of complex **5a** in CD_2Cl_2 .

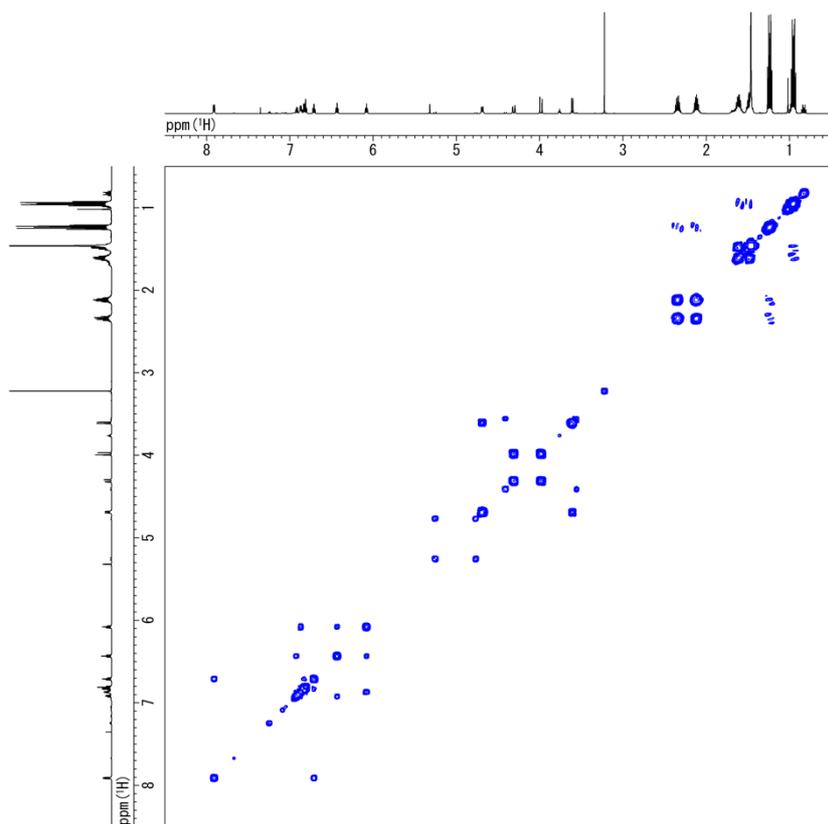


Fig. S33 ^1H - ^1H COSY NMR spectrum (600 MHz) of complex **5a** in CD_2Cl_2 .

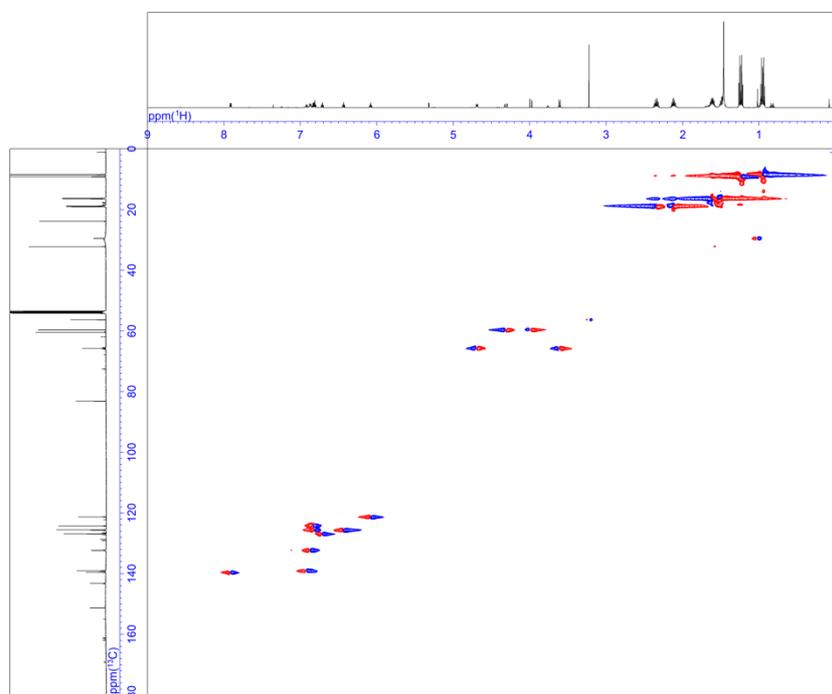


Fig. S34 ^1H - ^{13}C HSQC NMR spectrum (600 MHz for ^1H , 151 MHz for ^{13}C) of **5a** in CD_2Cl_2 .

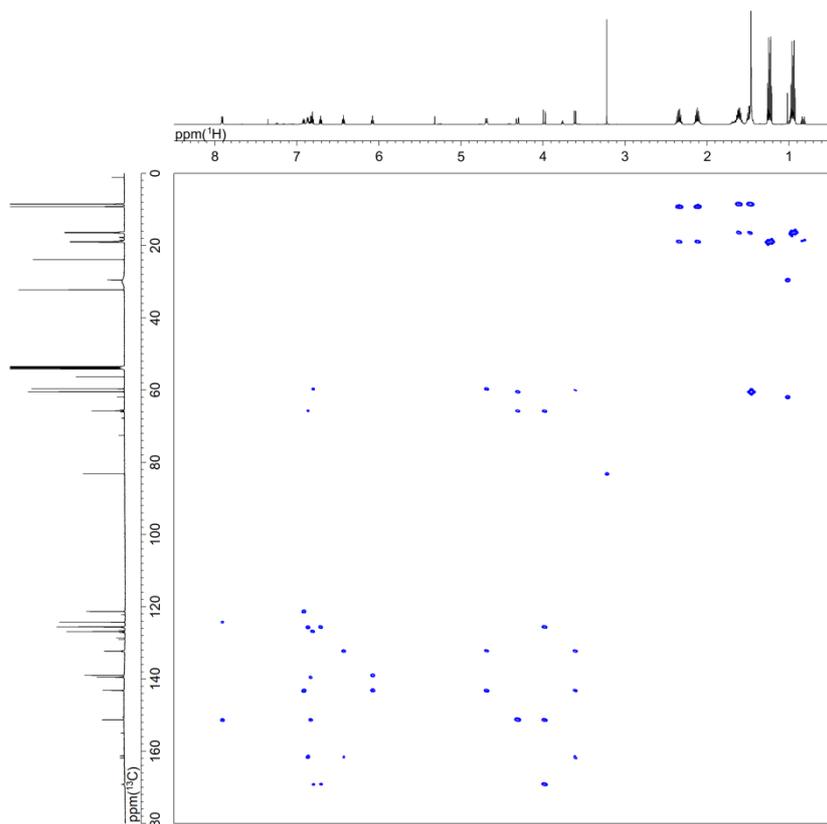


Fig. S35 ^1H - ^{13}C HMBC NMR spectrum (600 MHz for ^1H , 151 MHz for ^{13}C) of complex **5a** in CD_2Cl_2 .

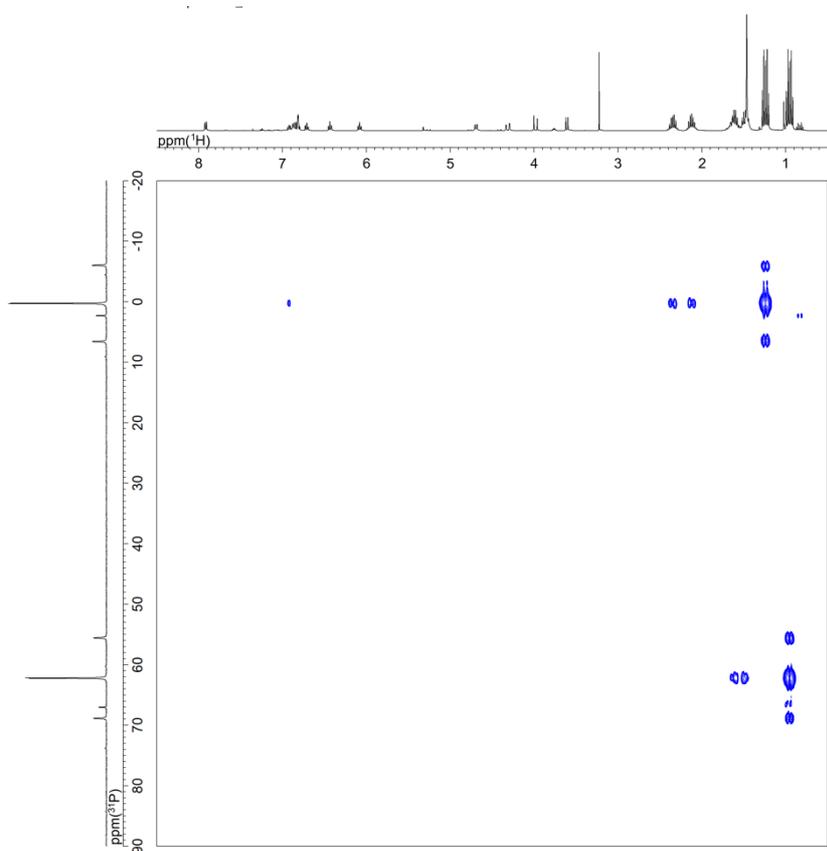


Fig. S36 ^1H - ^{31}P HMBC NMR spectrum (400 MHz for ^1H , 162 MHz for ^{31}P) of complex **5a** in CD_2Cl_2 .

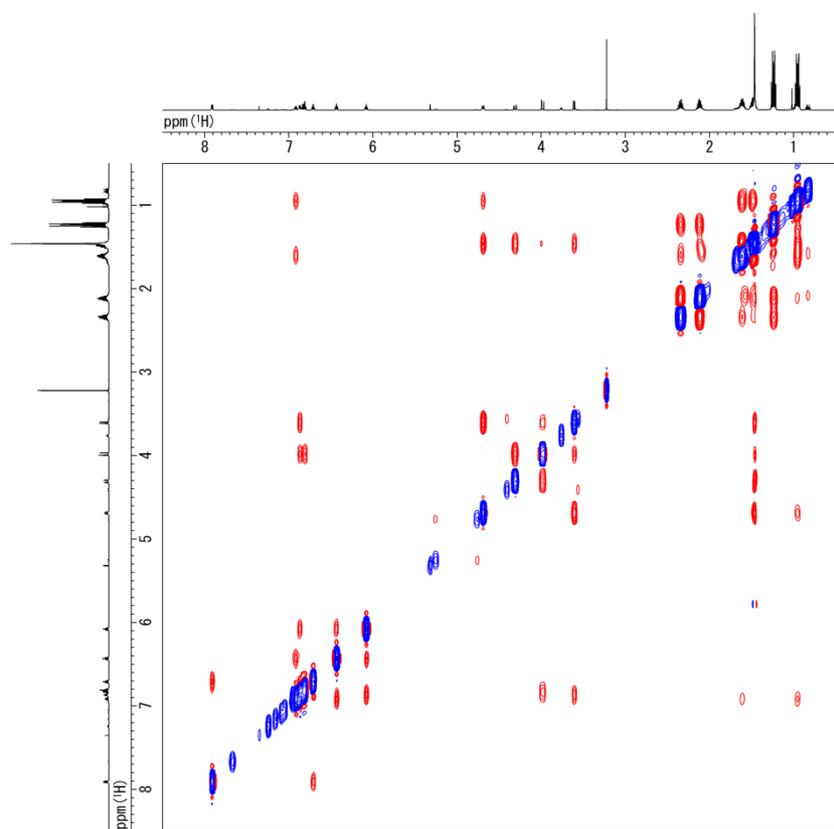


Fig. S37 NOESY NMR spectrum (600 MHz for ^1H) of complex **5a** in CD_2Cl_2 .

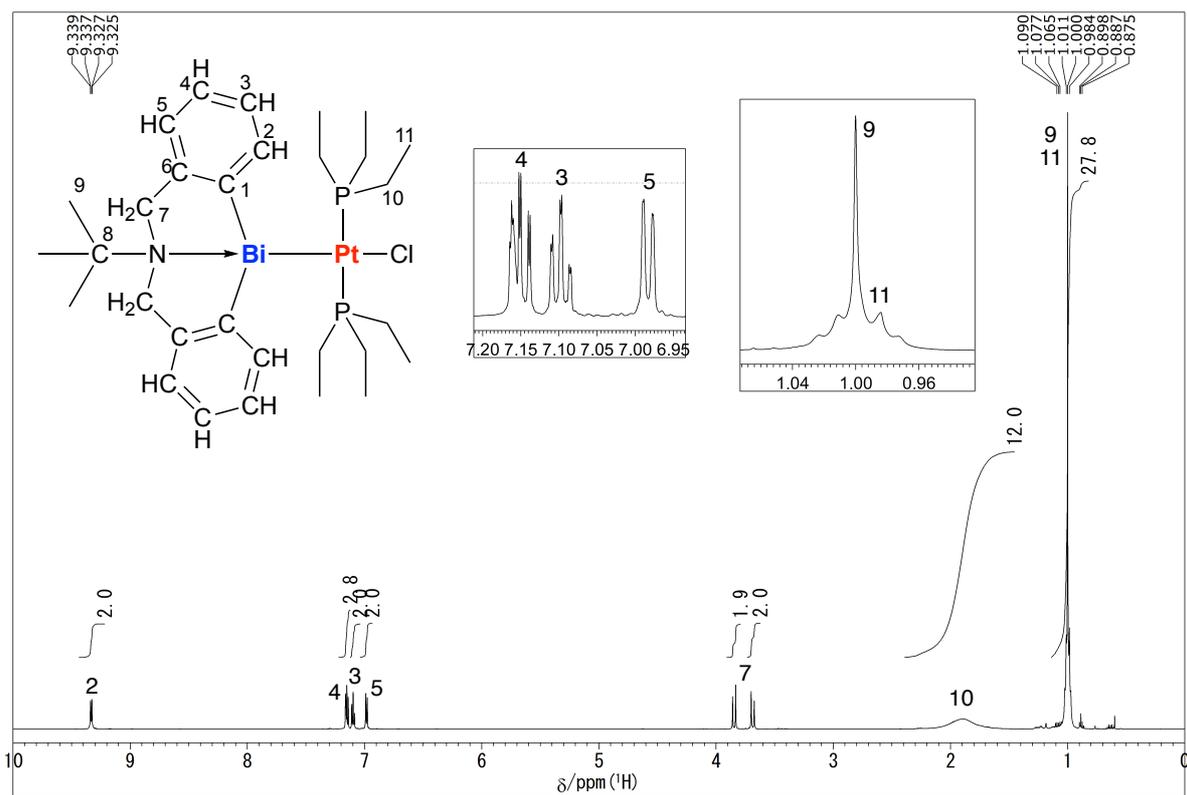


Fig. S38 ^1H NMR spectrum (600 MHz) of complex **6a** in C_6D_6 .

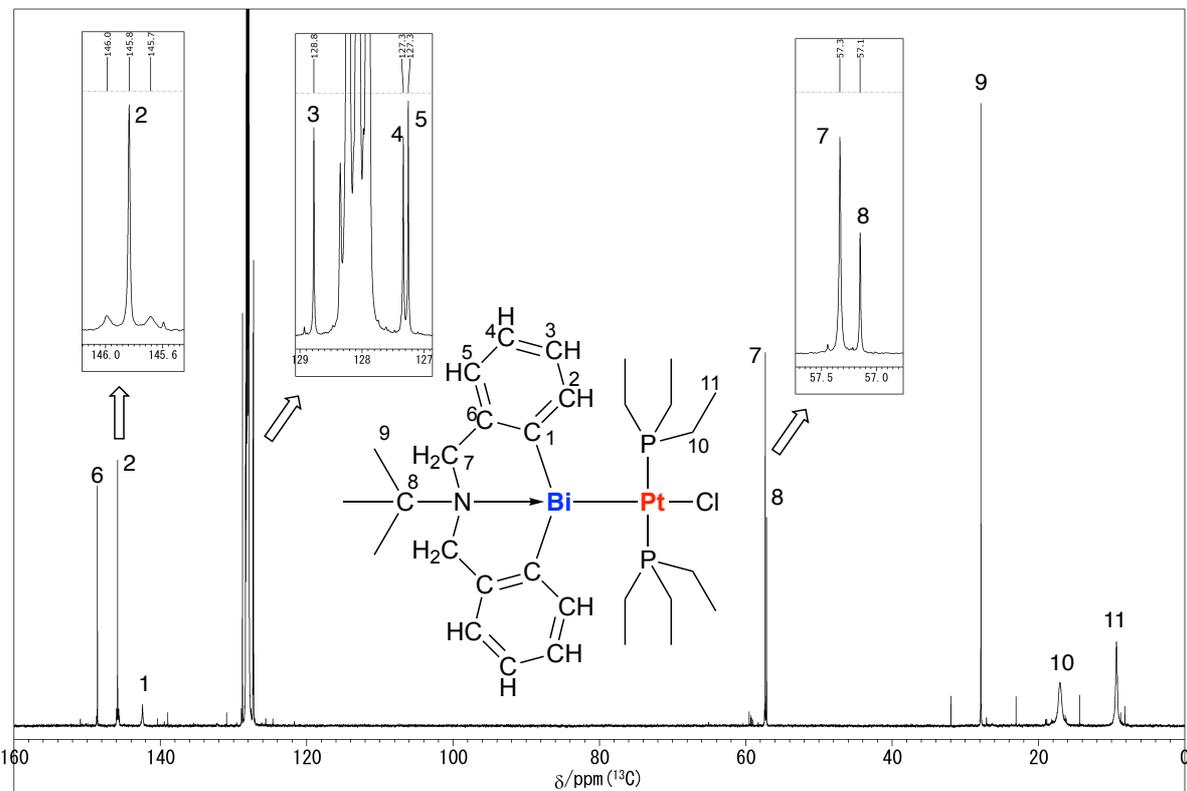


Fig. S39 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) of complex **6a** in C_6D_6 .

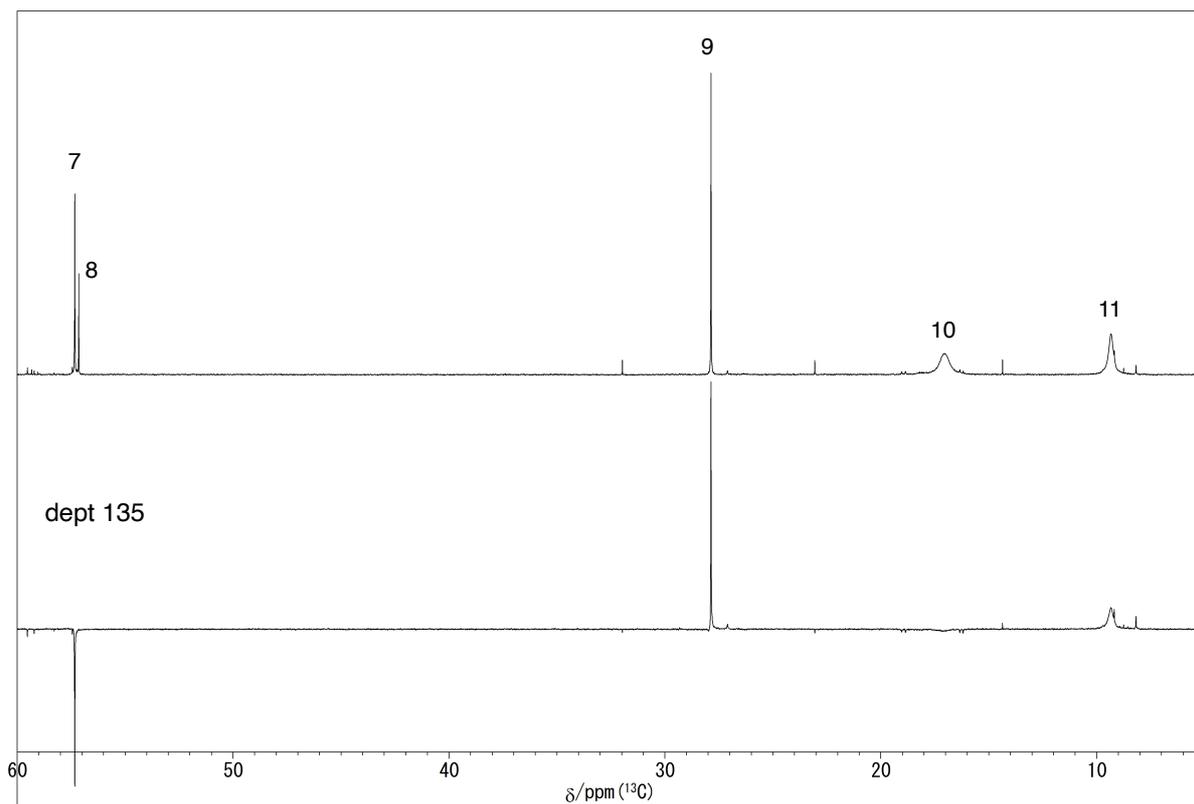


Fig. S40 A part (5–60 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) with dept 135 spectrum (lower) of complex **6a** in C_6D_6 .

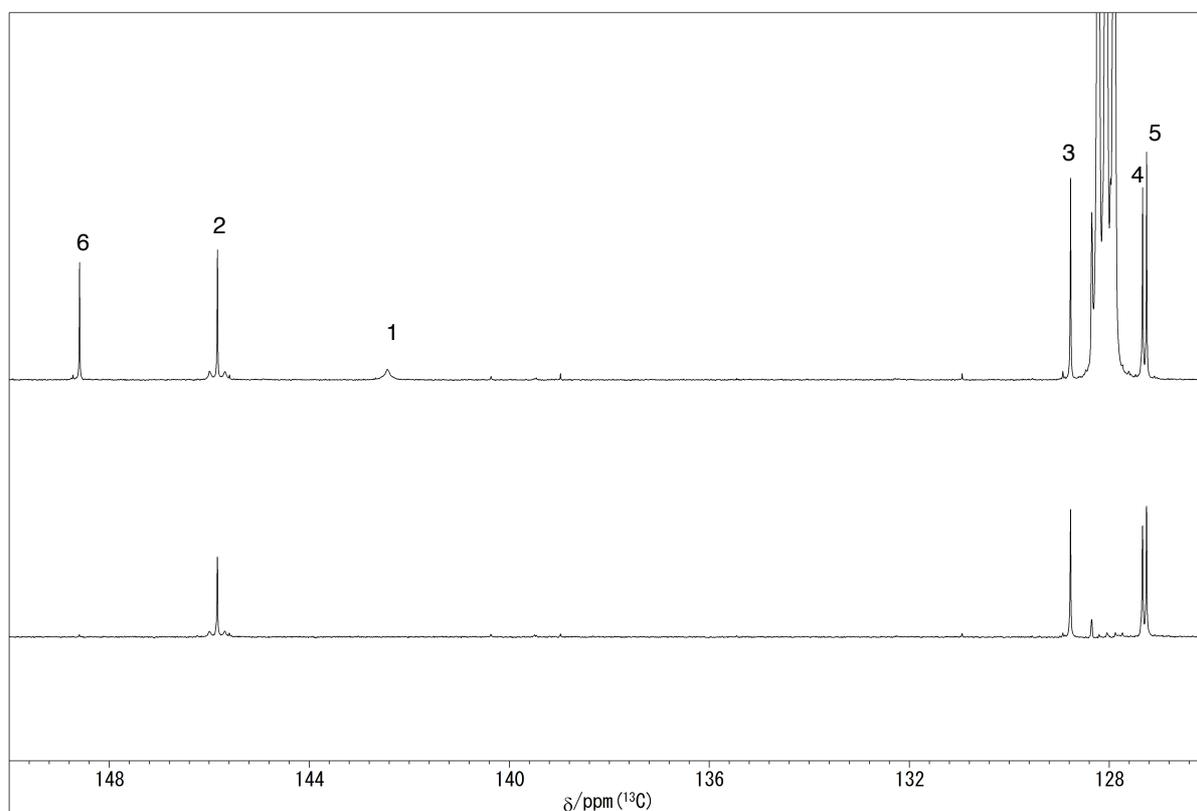


Fig. S41 A part (126–150 ppm region) of $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (151 MHz) with dept 135 spectrum (lower) of complex **6a** in C_6D_6 .

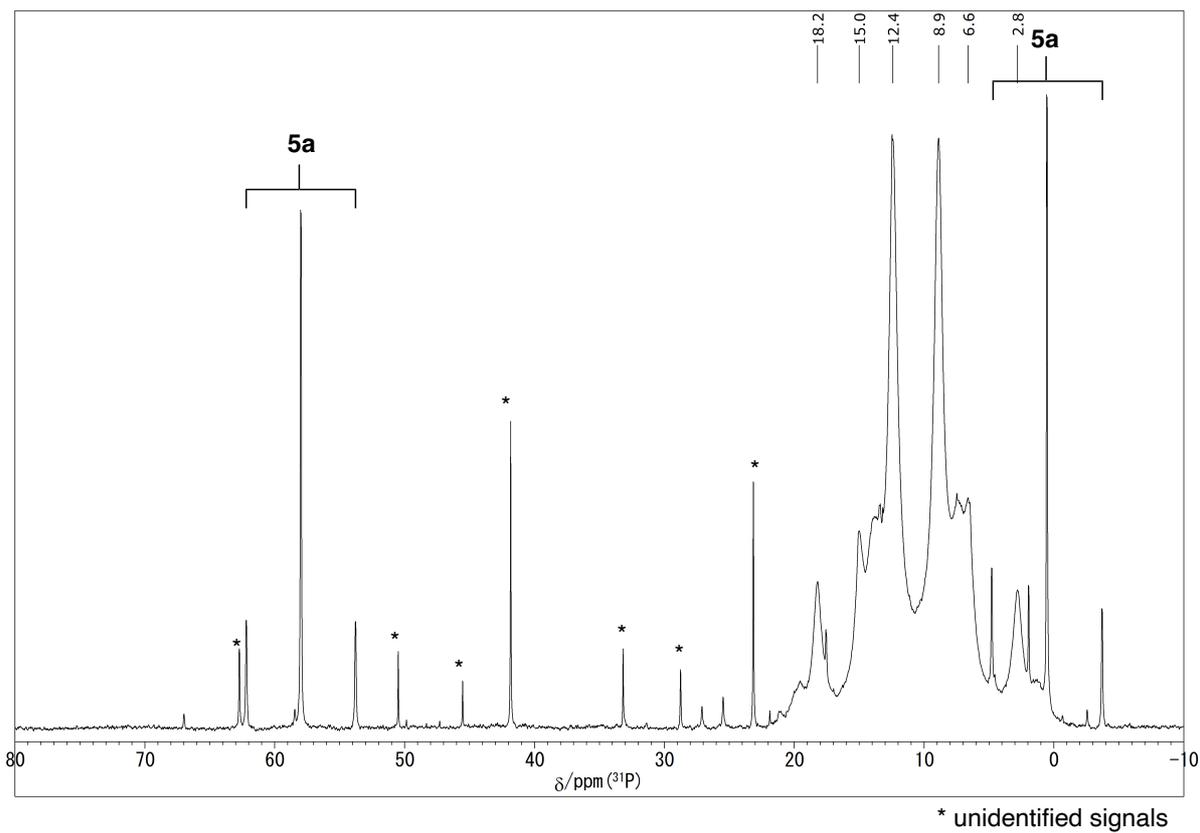


Fig. S42 $^{13}\text{P}\{^1\text{H}\}$ NMR spectrum (243 MHz) of complex **6a** in C_6D_6 . The spectrum was obtained 2 h after the sample preparation and contained weak signals of **5a** and unidentified species.

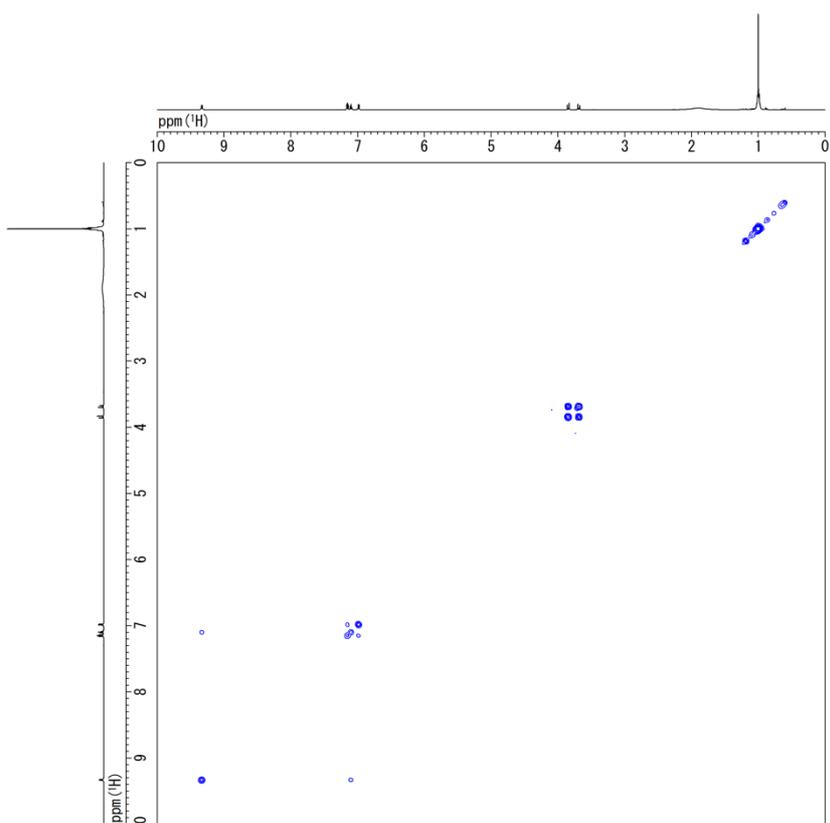


Fig. S43 $^1\text{H}\text{-}^1\text{H}$ COSY NMR spectrum (600 MHz) of complex **6a** in C_6D_6 .

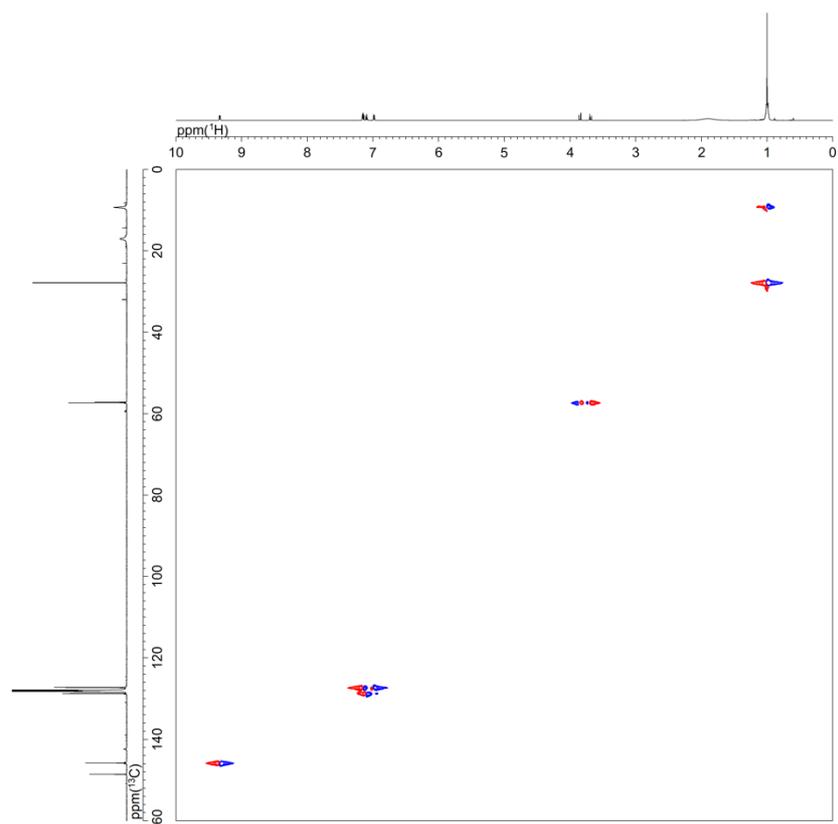


Fig. S44 ^1H - ^{13}C HSQC NMR spectrum (600 MHz for ^1H , 151 MHz for ^{13}C) of complex **6a** in C_6D_6 .

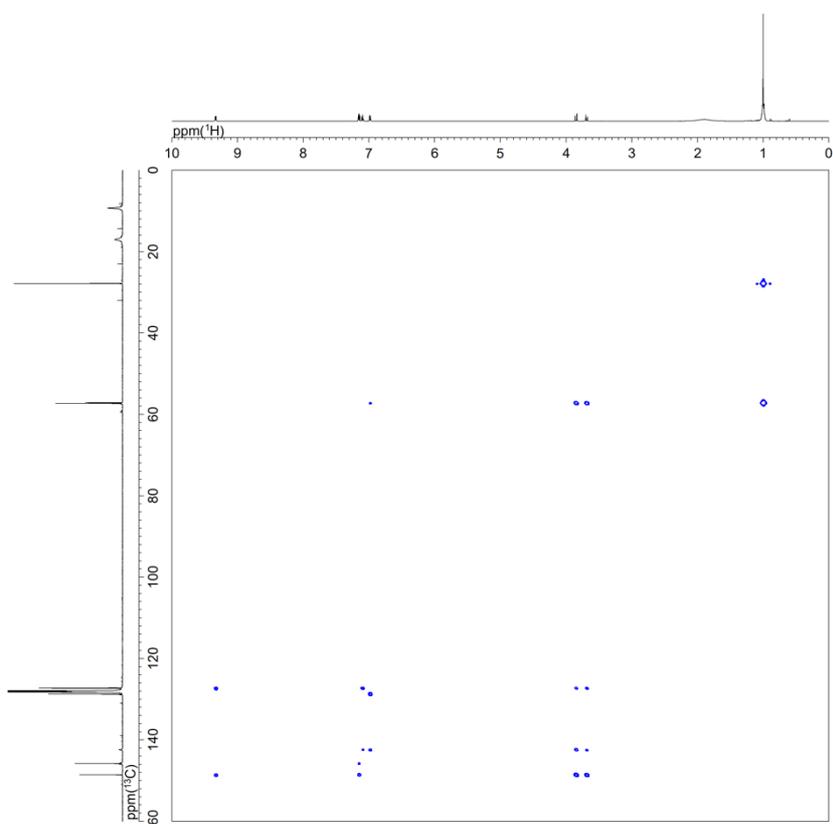


Fig. S45 ^1H - ^{13}C HMBC NMR spectrum (600 MHz for ^1H , 151 MHz for ^{13}C) of complex **6a** in C_6D_6 .