

Electronic Supplementary Information

Pandora's DNA Origami Box for the Site-specific Facet Protection of Gold Nanoparticles: A Building Block for Molecular Plasmonics

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Preparation of AuNS/AuNC and coating with ssDNA

Tween20-stabilized AuNS and AuNC were synthesized according to a reported method.² The synthesized and CTAB-stabilized particles were centrifuged and the precipitate was subsequently resuspended two times in 1 % sodium dodecyl sulfate (SDS) followed by three washing steps with 1 % Tween20. After the last centrifugation step, the pellets were collected and the concentration of AuNP was determined by UV-Vis extinction spectroscopy according to a reported method.³ Coating of the synthesized AuNS and AuNC with ssDNA was carried out following a published protocol.⁴

Agarose gel electrophoresis (AGE)

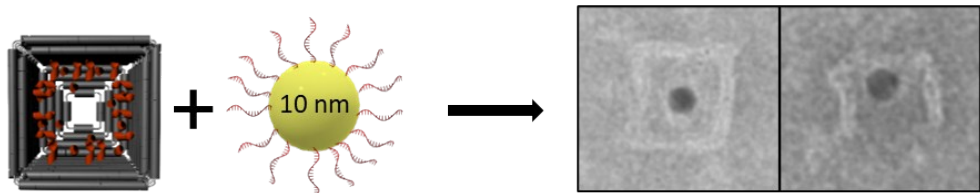
Purification of AuNP/origami box conjugates was achieved by using AGE in a 1 % agarose gel (containing 0.2 % Tween20) with 0.5xTBEMg as running buffer with 0.2 % Tween20 for 2 h at 4°C at 80 V. Isolation of origami band containing AuNP/origami conjugates was achieved by using spin column extraction (Freeze `n Squeeze Quantum prep, BioRad) at 10°C for 10 min and 250 g.

TEM Analysis

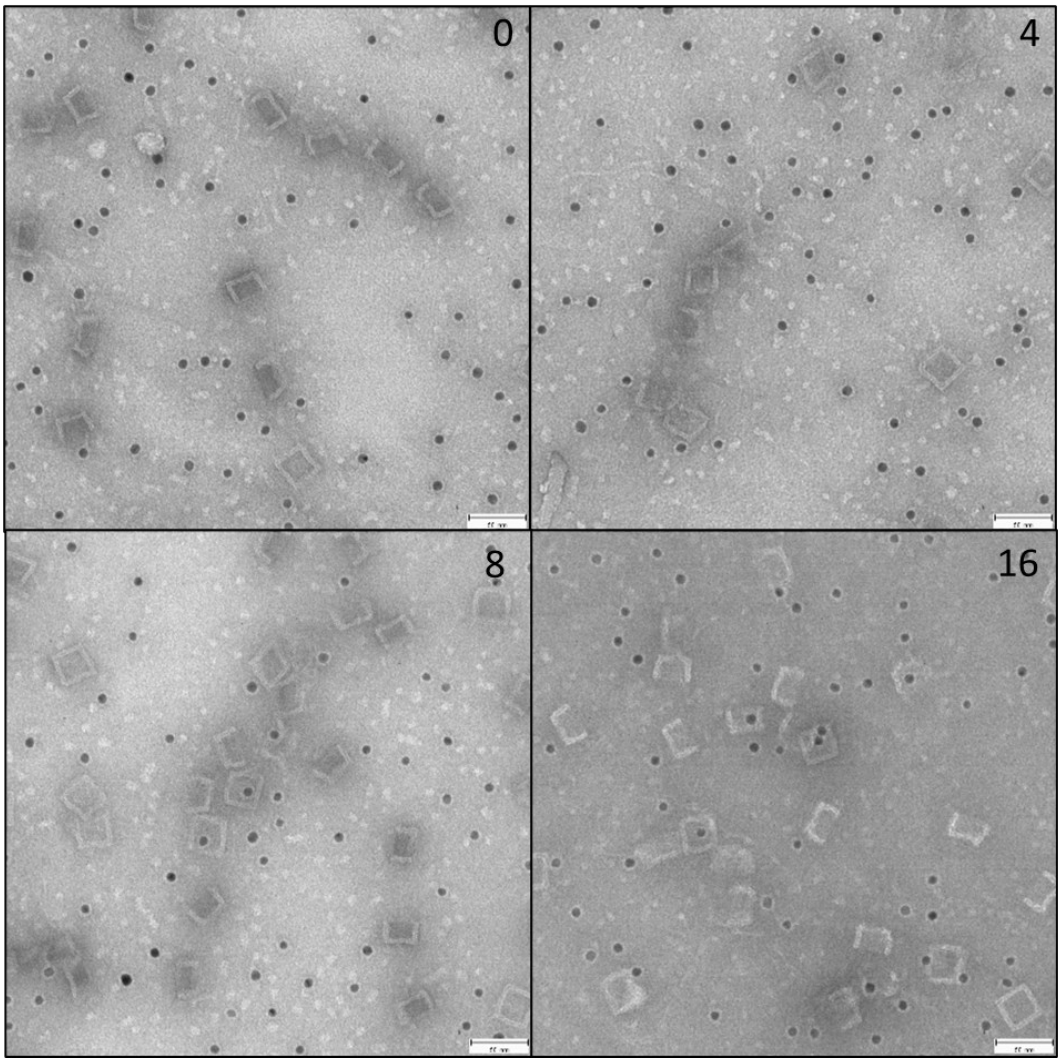
For the TEM analysis 400 mesh carbon-coated copper grids (Quantifoil) were glow charged and samples were drop-casted onto the grid. After incubation (2 min) the origamis were stained with 1 % uranyl formate before drying. The TEM images were obtained using a JEOL JEM 1400 Plus and a ZEISS EM910.

References

1. Y. Ke, S. M. Douglas, M. Liu, J. Sharma, A. Cheng, A. Leung, Y. Liu, W. M. Shih and H. Yan, *J. Am. Chem. Soc.*, 2009, 15903–15908.
2. J. E. Park, Y. Lee and J. M. Nam, *Nano Lett.*, 2018, **18**, 6475–6482.
3. T. Hendel, M. Wuithschick, F. Kettemann, A. Birnbaum, K. Rademann and J. Polte, *Anal. Chem.*, 2014, **86**, 11115–11124.
4. S. Xu, H. Yuan, A. Xu, J. Wang and L. Wu, *Langmuir*, 2011, **27**, 13629–13634.



Binding via Hybridization of ssDNA coated AuNP (10 nm) “gold standard”					
No. of protruding strands inside Pandora	0	4	8	16	28
Binging efficiency	0.5%	2%	9%	14%	62%



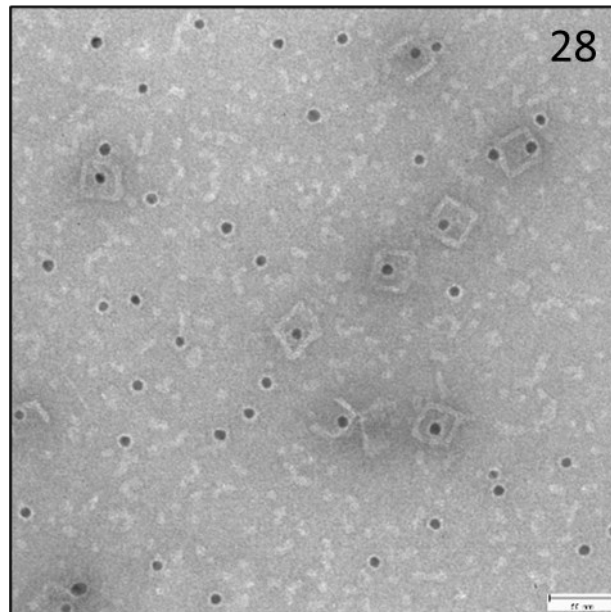


Fig. S 1 Top: **Incorporation of AuNS by hybridization** and representative TEM images. Middle: Binding yields of incorporation by using different numbers of protruding arms inside the cavity. Hybridization was tested in 0.5xTEMg with 0.15 % Tween20 and a 2.5-fold excess of AuNS. Bottom: TEM images (uranyl formate stained) were statistically analyzed by counting of origami structures ($n \geq 130$).

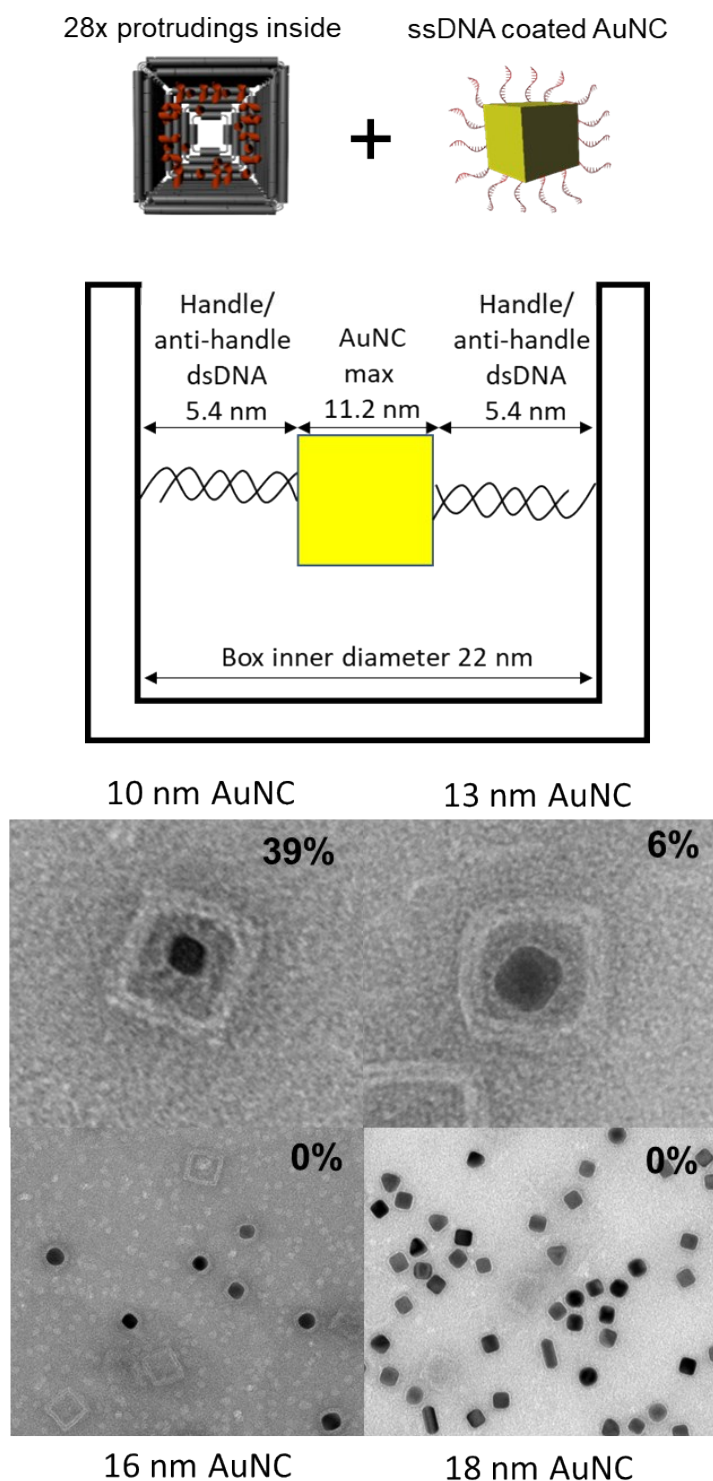
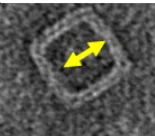
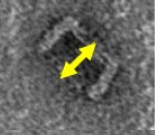
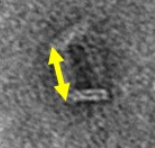


Fig. S 2 Top: Theoretical and experimentally determined maximum AuNC size. Top: Hybridization scheme of ssDNA-coated AuNC with maximum dimensions of AuNC fitting in the cavity (dimensions derived by design structure). Bottom: Experimentally determined yield for the incorporation of AuNC with different edge lengths (10-18 nm) into DNA origami boxes by hybridization. The unsuccessful incorporation of the larger 16 nm and 18 nm AuNC is attributed to steric hindrance and electrostatic repulsion.

	Measured [nm]	Calculated [nm]	
Inner diameter:	25.4 ± 1.8	21.3	
Height:	26.1 ± 2.5	25.4	
Outer diameter:	34.5 ± 2.9	31.7	

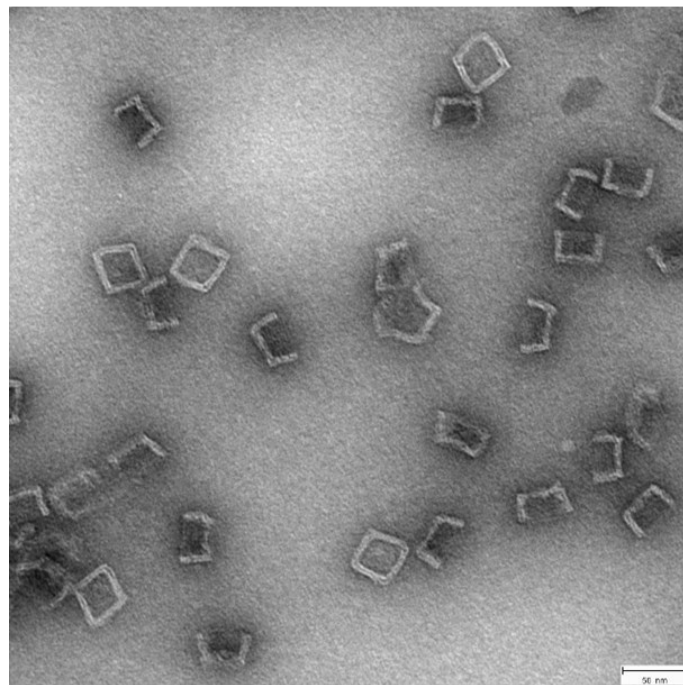


Fig. S 3 DNA origami box dimensions. Top: Theoretical values are derived by considering previously reported distance measurements.¹ Bottom: Measured values are derived from statistical analysis of DNA origami box TEM images (n=100).

Conjugation of 19 nm AuNC with Pandora containing 28 thiols

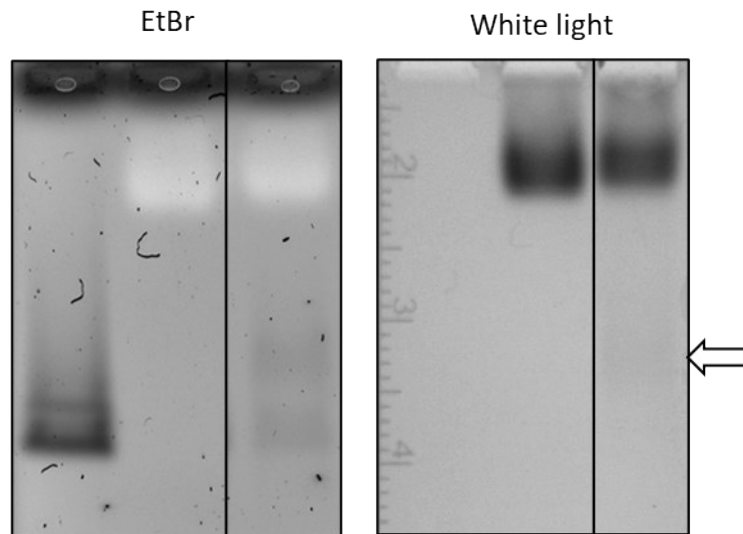


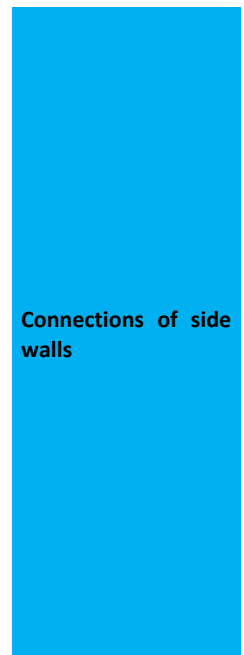
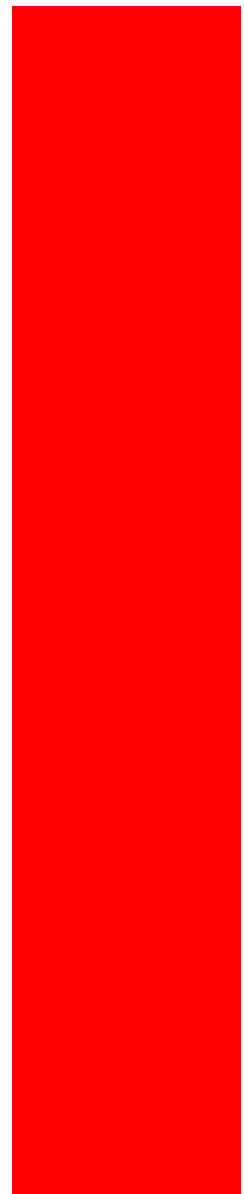
Fig. S 4 Purification of DNA origami AuNC conjugates by agarose gel electrophoresis. The analysis under fluorescence illumination allows to identify origami. Left: The origami structures stained with ethidium bromide (EtBr) appear dark in the fluorescence channel, while the AuNC (18 nm) appear bright due to their optical extinction. Right: The AuNC appear dark in the white-light image due to their pale reddish color. The electrophoretic mobility of the AuNC/DNA conjugates is higher than that of single or agglomerated AuNC which leads to a pale red conjugate band visible in the white-light channel indicated with an arrow.

Table S1 Staple sequences for the assembly of Pandora. Sequences include several modified staples, responsible for distinct features of the final construct. Staples are color-coded the same way as in the cadnano design.

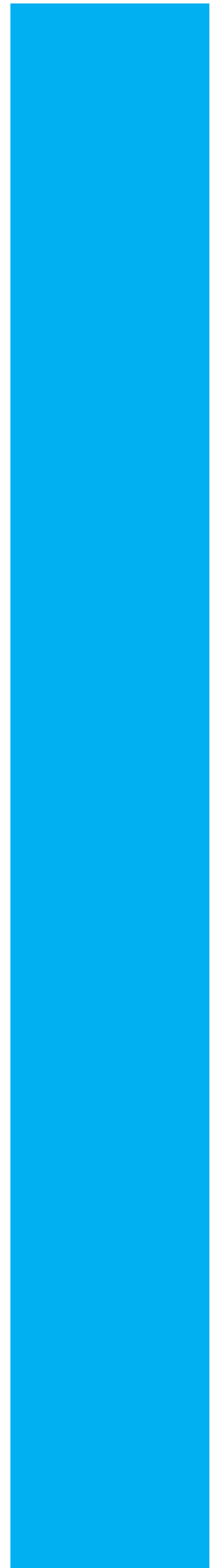
Sequence	Length	Description
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TTCGACTTG TAGAACGT CAGAAATAAATATA	31	
GAGAGGCGAGCTGATTGCCCTT CACCGC	28	
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ATTATCATAGCGCCAAAGACAAAAGGGC	28	
CCAACCTTTTAGAACAACTCAACATTAAATGTGAACCA	39	
CCTGTTTATCA	11	Core Staples, bottom plate
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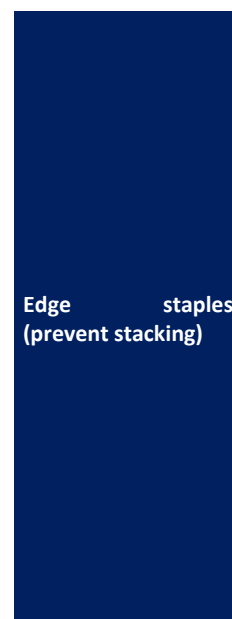
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Core staples, walls

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Edge staples
(prevent stacking)

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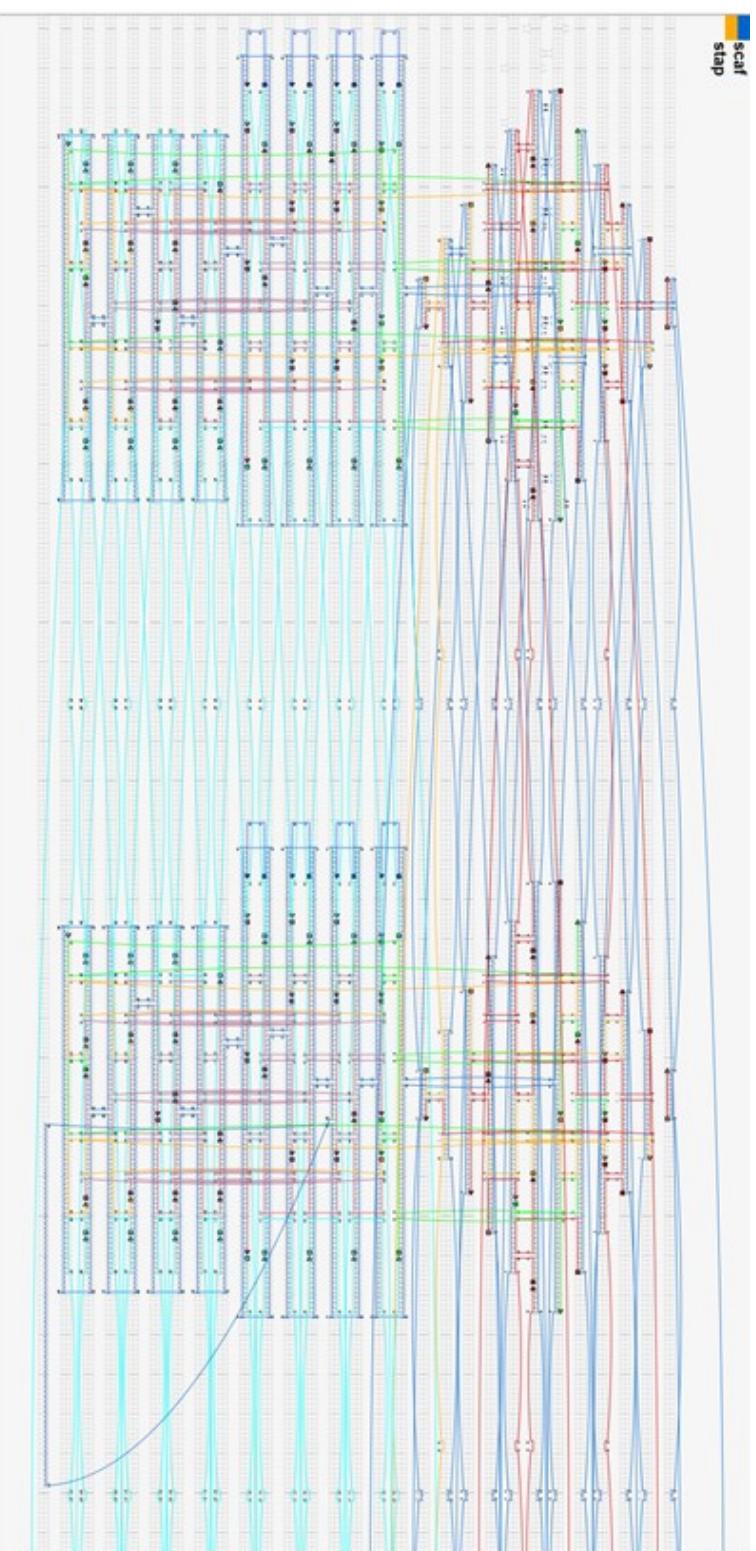
Fluorophore staple

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CCAAATCAAGCGGTCCATAATGGACCATCACGCAAATTAATTTTGATTGCCACTTTCCAC	60
TTTAATTTGCGCATCGTCAGGTTTATAACGGAACGTGCCGGTCTGATTGCCACTTTCCAC	60
GACCAGGGAACCAAGTAAGCAGATAGCCGAACAAAGTGATTGATTGCCACTTTCCAC	57

Thiol-GTGGAAAGTGGAATC	16
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Protruding staples
(These staples
protrude into
Pandora and can
bind thiolated-
oligonucleotides)

Thiol-staple (binds
the AuNC)



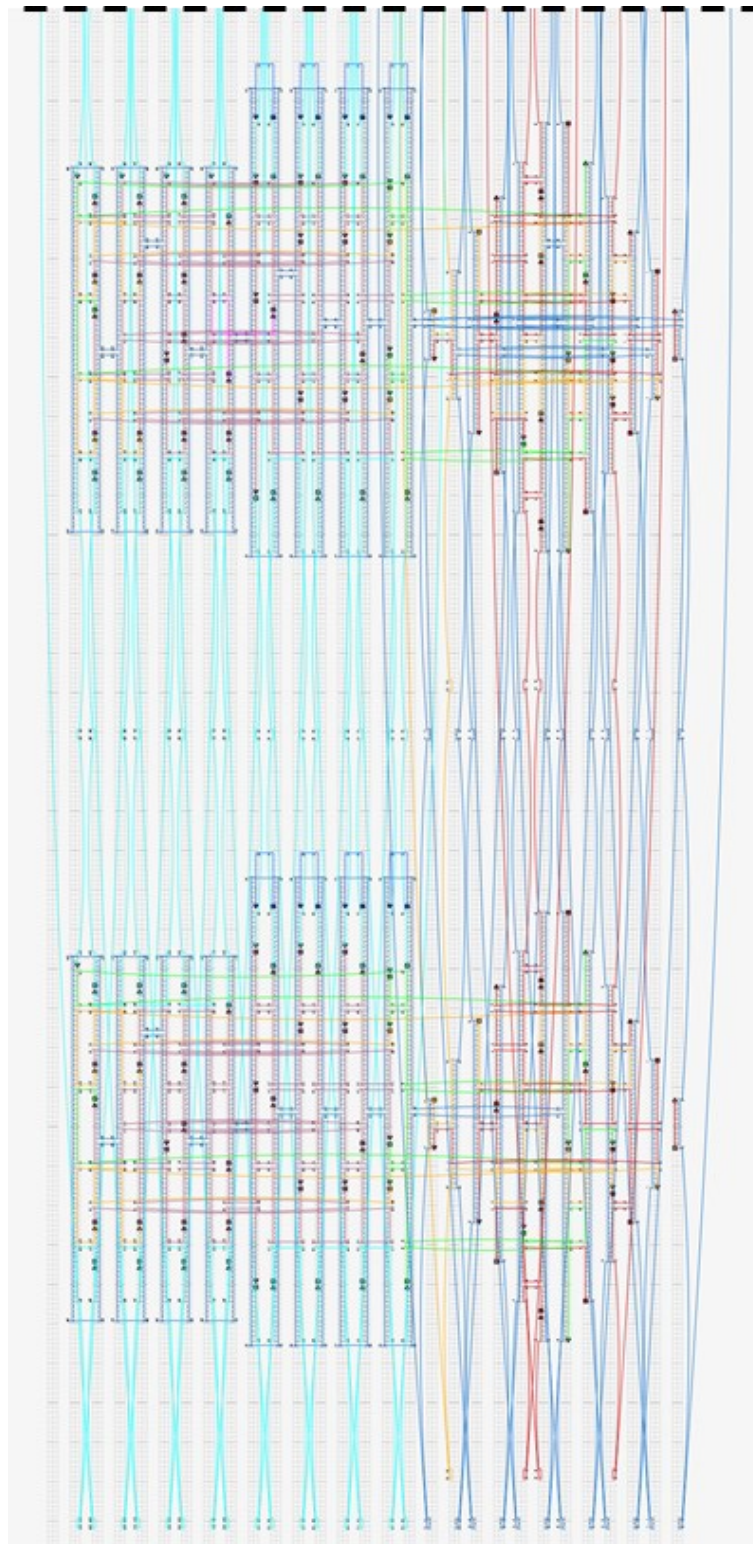


Fig. S 5 Cadnano design of Pandora. The structure consists of a bottom plate and four walls. Small single-stranded DNA segments are interspersed in between each face to release possible strain. Staples are color-coded (red: bottom plate core staples; green: Staples connecting bottom plate and walls; purple: wall core staples; cyan: Staples connecting walls; magenta: fluorophore-staple; dark blue: edge staples (prevent stacking); orange: protruding staples).