

## Supporting Information

### Immobilization of Arrestin-3 on different biosensor platforms for evaluating GPCR binding

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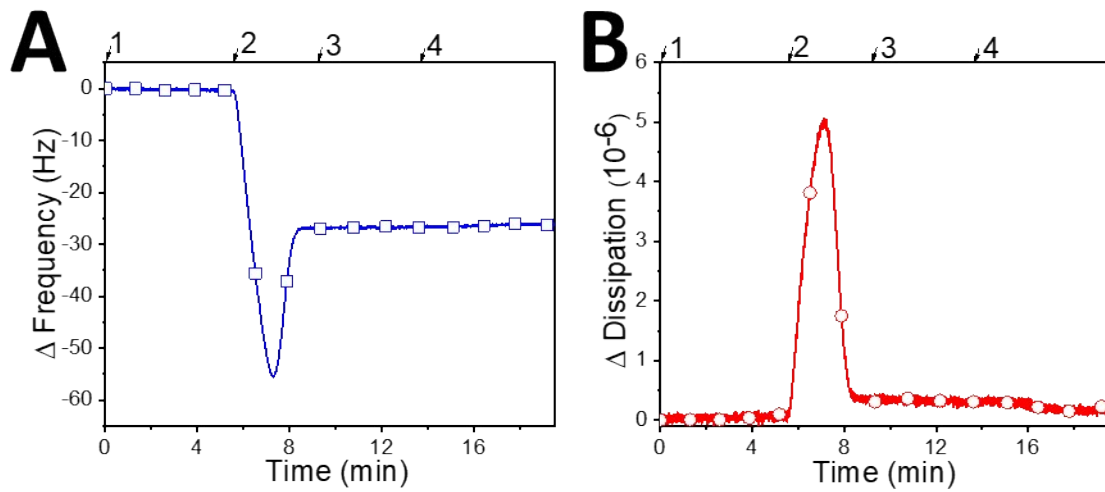
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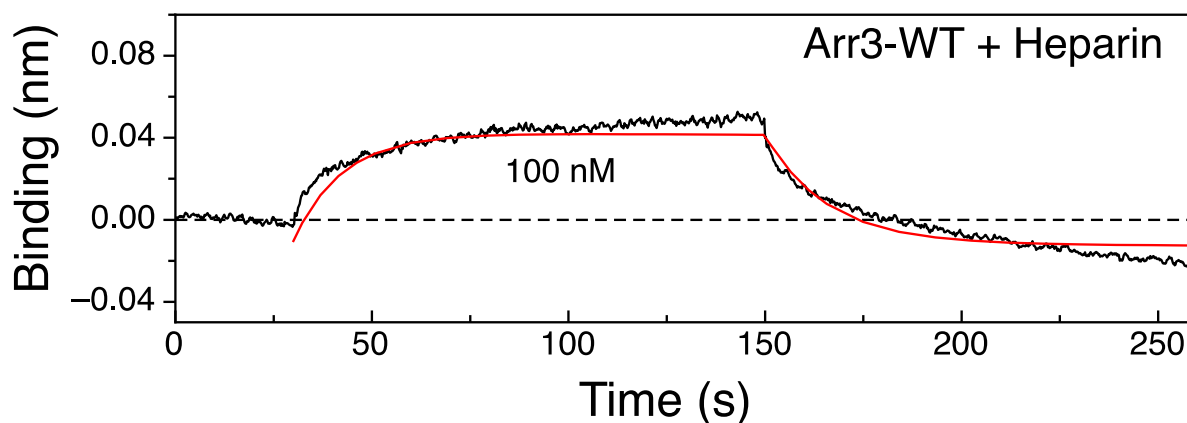
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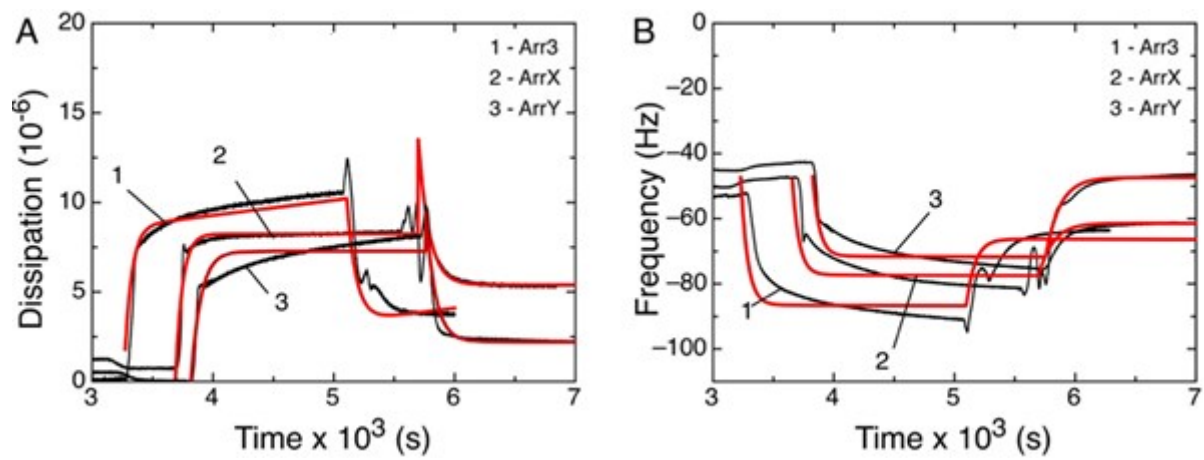
**Figure S1.** Changes in frequency (**A**) and in dissipation (**B**) were recorded as function of time upon formation of supported lipid bilayers on QCM-D sensors via vesicle fusion. The numbers at the top denote the injection of (1) buffer (20 mM HEPES, 150 mM NaCl at pH 7.4) containing 5 mM  $\text{MgCl}_2$ , (2) vesicles made of POPC:DGS-NTA ( $\text{Ni}^{2+}$ ) (95:5), (3) buffer, and (4) buffer without  $\text{MgCl}_2$ .



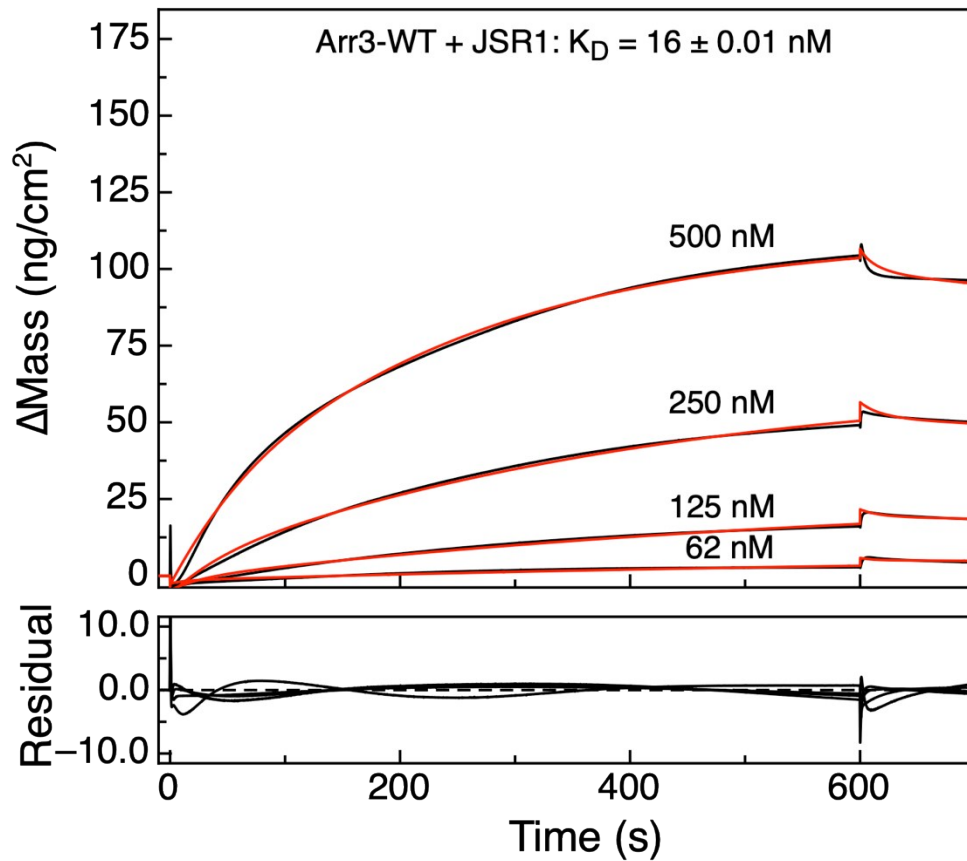
**Figure S2. Detection of heparin binding to Arr3-WT on the BLI sensors.** Representative heparin (100 nM) binding curve measured by BLI for (A) binding to Arr3-WT immobilized on the Ni<sup>2+</sup>-NTA tips. Number of replicates, n= 3 for each Arr3-WT.

**Table S1.** Heparin (100 nM) binding to Arr3-WT

Proteins	$K_D$ , nM	$k_a \times 10^5$ , M <sup>-1</sup> s <sup>-1</sup>	$k_d$ , s <sup>-1</sup>	Model for kinetic fit
Arr3-WT	400 ± 3	1.6 ± 0.4	0.064 ± 0.002	1:1 binding



**Figure S3.** The change in dissipation (A) and frequency (B) in the QCM-D measurements caused by the JSR1 (in DDM micelles) complex formation with arrestin-3 was fitted to the 1:1 binding model. Injected concentration of JSR1 was 1.59  $\mu\text{M}$ . Measurement was repeated 3 times for each arrestin ( $n = 3$ ). Obtained kinetic and equilibrium association and dissociation constants are summarized in **Table 6**.



**Figure S4. Analysis of JSR1 binding to Arr3-SLM.** (A) Representative kinetic fit to ‘two-state’ model for JSR1 binding to Arr3-WT immobilized on the Ni<sup>2+</sup>-NTA SLM of HPA sensor chip. Black curves are the experimental data whereas red curves are the fitted data.

**Table S2.** The equilibrium dissociation constants ( $K_D$ ) and kinetic association ( $k_{a1}$  and  $k_{a2}$ ) and dissociation ( $k_{d1}$  and  $k_{d2}$ ) constants obtained for JSR1 (activated) complexes with Arr3 using SPR. Constants were determined by the data fit to the ‘two-state’ model. They represent averages  $\pm$  standard deviations of three independent measurements.

Proteins	$K_D$ , nM*	$k_{a1} \times 10^5$ , $M^{-1}s^{-1}$	$k_{a2} \times 10^{-3}$ , $s^{-1}$	$k_{d1}$ , $s^{-1}$	$k_{d2}$ , $\times 10^{-3} s^{-1}$	Model for kinetic fit
Arr3-WT	137 $\pm$ 6.62	8980 $\pm$ 1200	7.86 $\pm$ 0.09	306 $\pm$ 53	5.29 $\pm$ 0.92	Two-state model
Arr3-X	18.0 $\pm$ 0.15	0.63 $\pm$ 0.006	5.94 $\pm$ 0.14	0.02 $\pm$ 0.002	3.89 $\pm$ 0.14	Two-state model
Arr3-Y	66.2 $\pm$ 20	38.7 $\pm$ 14	7.92 $\pm$ 1.01	0.93 $\pm$ 0.35	3.01 $\pm$ 0.32	Two-state model

\* For ‘two-state’ model apparent equilibrium dissociation constants was calculated as follows:

$$K_{Dapp} = \frac{k_{d1} \cdot k_{d2}}{k_{a1} \cdot (k_{a2} + k_{d2})}$$