

## Supporting information

### Alumina Grafted SBA-15 Sustainable Bifunctional Catalysts for Direct Cross-Coupling of Benzylic Alcohols to Diarylmethanes

C. Rajendran<sup>a</sup>, G. Satishkumar<sup>a\*</sup>, Charlotte Lang<sup>b</sup> and Eric M. Gaigneaux<sup>b</sup>

<sup>a</sup>*Advanced Materials and Catalysis Lab, Department of Chemistry,  
School of Advanced Sciences, Vellore Institute of Technology, Vellore-632014, Tamilnadu, India*  
<sup>b</sup>*Institute of Condensed Matter and Nanosciences, UCLouvain, 1348 Louvain-la-Neuve,  
Belgium.*

## **Table of Contents**

1. Materials and Methods	S3
2. Characterisation results	S6
3. General procedure for 4wt%AlSBA-15 catalysed construction of diarylmethane using 2-naphthylmethanol and phenylboronic acid	S10
4. Spent catalyst characterization results	S11
5. H <sup>1</sup> and C <sup>13</sup> NMR, GC-MS data of the obtained products	S12
6. References	S19
7. H <sup>1</sup> and C <sup>13</sup> NMR Spectra of all compounds	S20

## **1. Materials and Methods**

### **1.1 Materials**

All chemicals were purchased from commercial suppliers and used without further purification. TEOS, HCl, Sodium aluminate ( $\text{NaAlO}_2$ ) and  $\text{P}_{123}$  were purchased from Sigma Aldrich. Aluminium sec-tri butoxide ( $\text{Al}(\text{O}-\text{sec-Bu})_3$ ), Cerium nitrate ( $\text{Ce}(\text{NO}_3)_3 \cdot 6\text{H}_2\text{O}$ ), and di-chloroethane (DCE) bought from Alfa Aesar. Thin layer chromatography (TLC) plates (60 F254, 0.2 mm) and all other reagent grade chemicals, solvents were used as received from Avra synthesis Pvt. Ltd.

### **1.2. Characterization Methods**

Powder X-ray diffraction (XRD) patterns were obtained with a D8 Advance (BRUKER), diffractometer with a Cu K alpha ( $\lambda = 1.5418 \text{ \AA}$ ) radiation source 2.2 kW and a Lynx Eye detector (Silicon strip detector technology). XRD scans were recorded between  $2\theta = 0.75\text{--}5^\circ$  with a step size of 0.05 and step time of 1s for low angle and between  $2\theta = 10\text{--}90^\circ$  for wide angle analysis. The Scherrer equation is applied to determine crystal size  $h = K\lambda/[(B^2 - \beta^2)^{0.5} \cos(2\theta/2)]$ .  $\text{N}_2$  adsorption-desorption isotherms were performed at liquid nitrogen temperature (-196°C) using an Autosorb iQ and ASiQWin Version 5 model 6 analysers from Quantachrome. Before each measurement, samples were outgassed 5h at 300°C under Helium. Surface area, pore volume, and pore size distribution were obtained from  $\text{N}_2$  adsorption-desorption isotherms by using the conventional BET and BJH methods. High Resolution Transmission electron microscopy (HR-TEM) images were taken using a Model FEI-TECNAI G2-20 TWIN 200kV instrument, filament LaB6. The temperature-programmed desorption of ammonia ( $\text{NH}_3\text{-TPD}$ ) was conducted on Autosorb iQ and ASiQWin Version 5 model 6 analysers from Quantachrome. Typically, 0.100 g

of the dried sample was placed within a linear quartz tube (i.d. 12 mm) and pretreated in a helium flow at 400 °C for 1 h. Then, the sample was cooled down to 100°C and 20%NH<sub>3</sub>/He mixture gas was allowed to flow for 0.5 h. The sample was flushed with helium for 1 h to remove the physisorbed ammonia. NH<sub>3</sub>-TPD profile was recorded from 100 °C to 450 °C with a ramping rate of 10 °C min<sup>-1</sup>. CO<sub>2</sub>-TPD analysis was carried out using BELCAT II equipment. 50-100mg of the dried sample was placed within a linear quartz tube (i.d. 12 mm) and pretreatment heated up to 450°C at 10°C min<sup>-1</sup>, then held at maximum temperature for 30 min in flowing He (50 ml/min). Analysis conditions are then 10% CO<sub>2</sub> + He gas mixture flow for 1h at 50°C, then the physisorbed probe molecule was removed by flowing He (50 mL.min<sup>-1</sup>) for 1 h at 50°C, finally. CO<sub>2</sub>-TPD profile was recorded from 50 to 600°C at a heating rate of 10°C.min<sup>-1</sup> under the flow of helium to desorb the chemisorbed probe molecules. Pyridine chemisorption measurements were performed using a high vacuum setup and Fourier transform infrared (FT-IR) spectra were recorded in transmission mode using an IFS55 Equinox spectrometer (Bruker) equipped with a DTGS detector. 100 scans per spectrum were recorded between 400 and 4000 cm<sup>-1</sup> with a resolution of 4 cm<sup>-1</sup>. The pure catalyst powder was pressed into wafers (around 25mg) which were first degassed in the IR cell under vacuum (about 10<sup>-7</sup> Torr) at 250 °C for 2h to remove water and adsorbed species. The pyridine adsorption took place at room temperature for 10 minutes. The samples were then evacuated for 1h from room temperature to 200 °C (each 100 °C) to progressively desorb the pyridine. The IR spectra were measured after each desorption temperature and the IR spectra recorded after desorption at 100 °C were used for the calculation of the distribution between Brønsted and Lewis acid sites. The distribution between Lewis and Brønsted acid sites was calculated thanks to the integrated absorbance of bands due to pyridine absorption on Brønsted

(around 1550 cm<sup>-1</sup>) and Lewis (around 1450 cm<sup>-1</sup>) acid sites. The number of acid sites was calculated using equation 1 [1]:

$$n_T = \frac{A_L C_d}{\varepsilon_L m} + \frac{A_B C_d}{\varepsilon_B m} \quad \text{Equation 1}$$

Where  $n_T$  is the total number of pyridine adsorbed at each temperature, in  $\mu\text{mol.g}^{-1}$

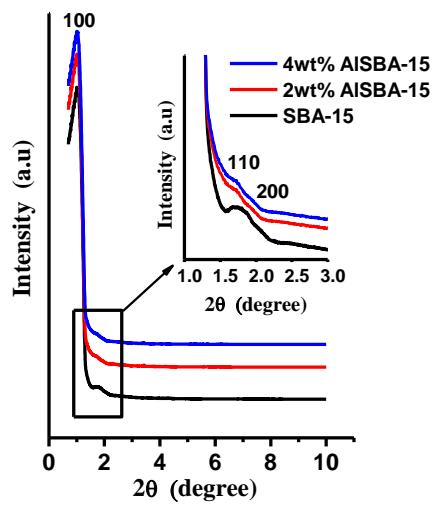
$A_L$  and  $A_B$  are the integrated absorbances of FTIR bands relative to Lewis or Brønsted sites respectively (cm<sup>-1</sup>)

$C_d$  is the cross sectional area of the wafer in cm<sup>2</sup>

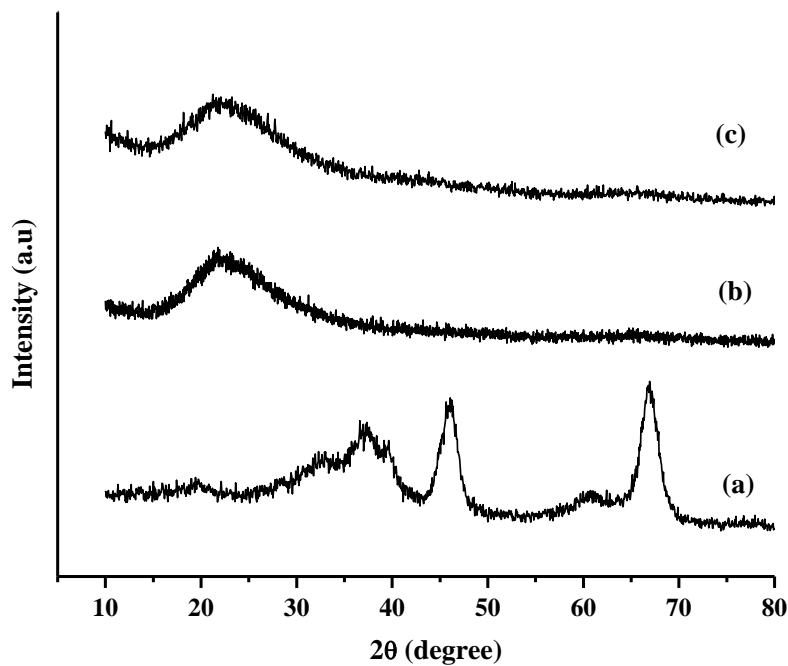
$\varepsilon_L = 1.44$  and  $\varepsilon_B = 0.59$  are the molar extinction coefficients in  $\text{cm}.\mu\text{mol}^{-1}$ , respectively for Lewis or Brønsted bands [1] and m is the mass of the water in g.

Magic angle spinning (MAS) <sup>27</sup>Al NMR spectra were recorded on ECX 400 - Jeol 400 MHz High-Resolution Multinuclear FT-NMR Spectrometer. NMR spectra were recorded on Bruker FT-NMR-400 MHz instrument for <sup>1</sup>H and <sup>13</sup>C NMR, respectively. All <sup>1</sup>H NMR spectra were reported in ppm unit (parts per million) and were measured relative to residual chloroform signal (7.26 ppm or 7.18 ppm) in the deuterated solvent. <sup>13</sup>C NMR spectra were reported in ppm relative to deuterated chloroform (77.23 ppm). The SEM analysis was carried out in SEM-Carl Zeiss with energy dispersive spectrometer (EDS) detector. XPS has been recorded using PHI 5000 Versa Probe II, FEI Inc. The products obtained has been confirmed through GC-MS Perkin Elmer, Clarus 680 model. The product yield was determined using Gas Chromatograph (Shimadzu 2014).

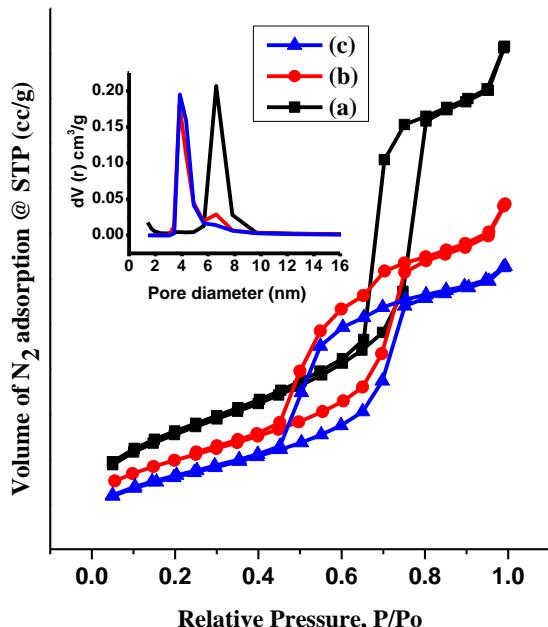
## 2. Characterisation results



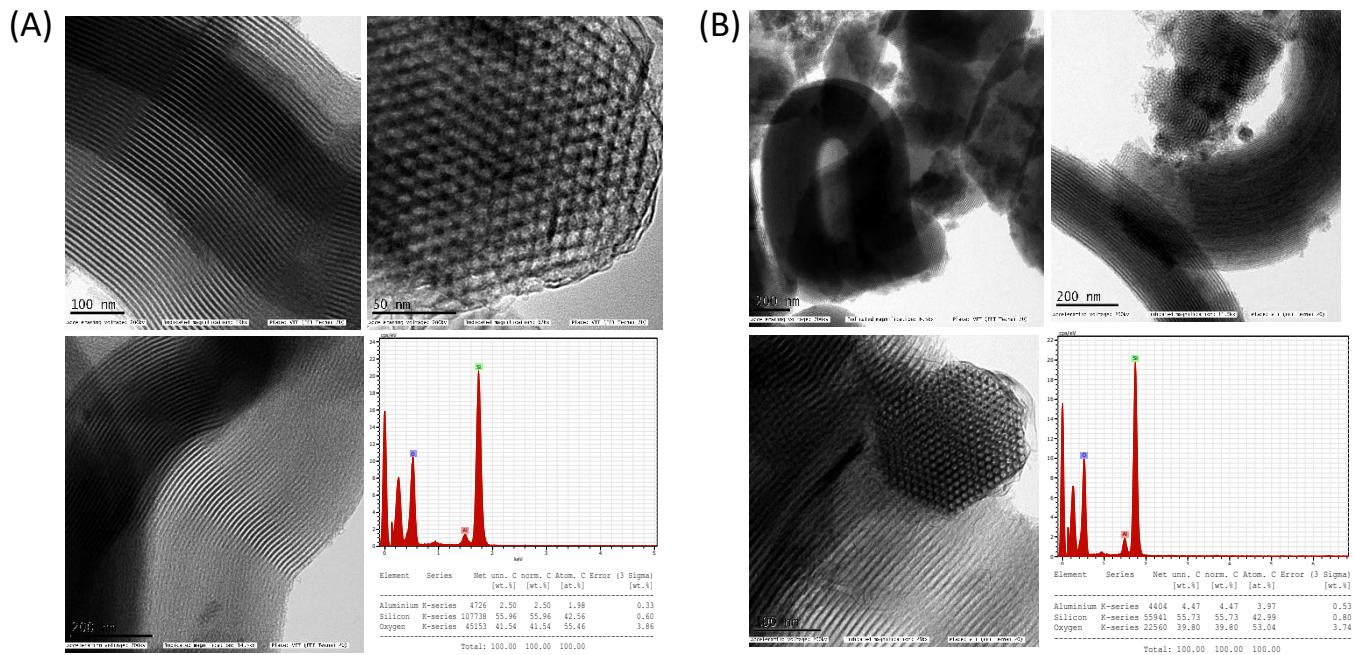
**Fig. S1.** Low angle XRD pattern of synthesized catalysts and support.



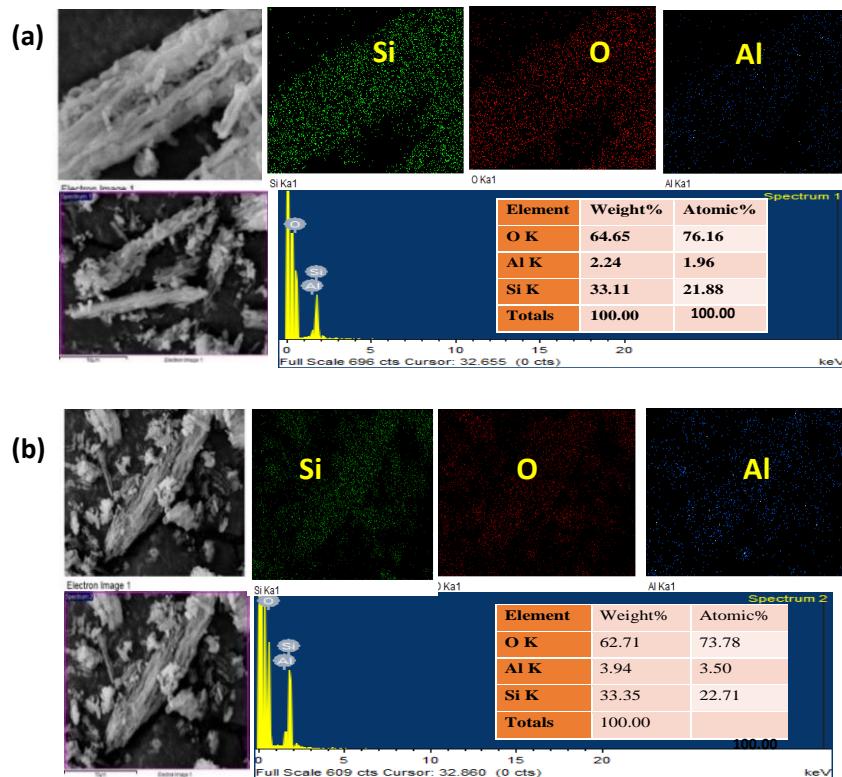
**Fig. S2.** Wide angle XRD pattern of synthesized catalysts (a)  $\gamma$ -Al<sub>2</sub>O<sub>3</sub>; (b) 2wt% AlSBA-15; (c) 4wt% AlSBA-15.



**Fig. S3.** N<sub>2</sub> adsorption and desorption isotherm of synthesised catalysts and support  
(a) SBA-15, (b) 2wt% AlSBA-15 and (c) 4wt% AlSBA-15.



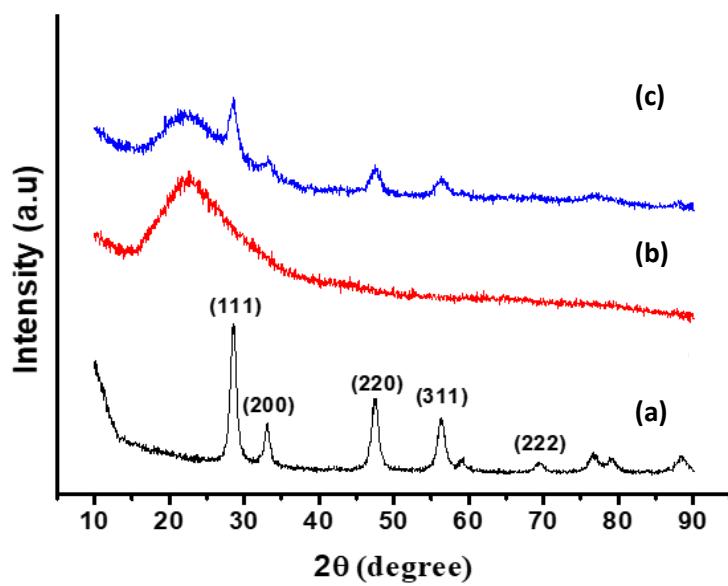
**Fig. S4.** HR-TEM images of (A) 2wt% AlSBA-15 and (B) 4wt% AlSBA-15



**Fig. S5.** SEM- EDAX elemental mapping of synthesised catalysts (a) 2wt% AlSBA-15  
(b) 4wt% AlSBA-15.

**Table S1:** Acidity Measurements of 2, 4wt% AlSBA-15 and  $\gamma$ -Al<sub>2</sub>O<sub>3</sub>

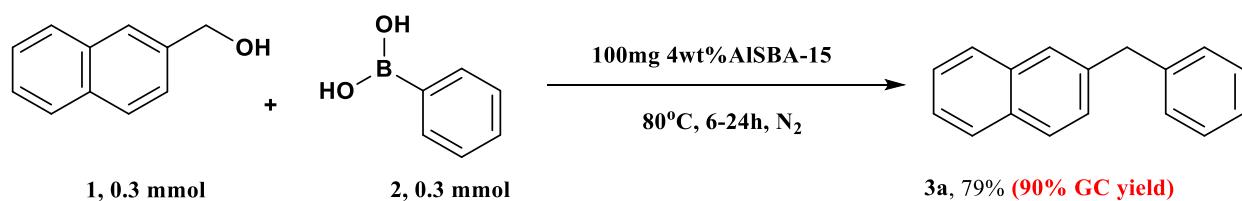
S. No	Catalyst	Temperature T (°C)	Amount of desorbed NH <sub>3</sub> (mmol g <sup>-1</sup> )	Temperature T(°C)	Amount of desorbed NH <sub>3</sub> (mmol g <sup>-1</sup> )
1	$\gamma$ -Al <sub>2</sub> O <sub>3</sub>	250	0.170	417	0.015
2	2wt% AlSBA-15	228	0.438	419	0.066
3	4wt% AlSBA-15	252	1.327	447	0.296

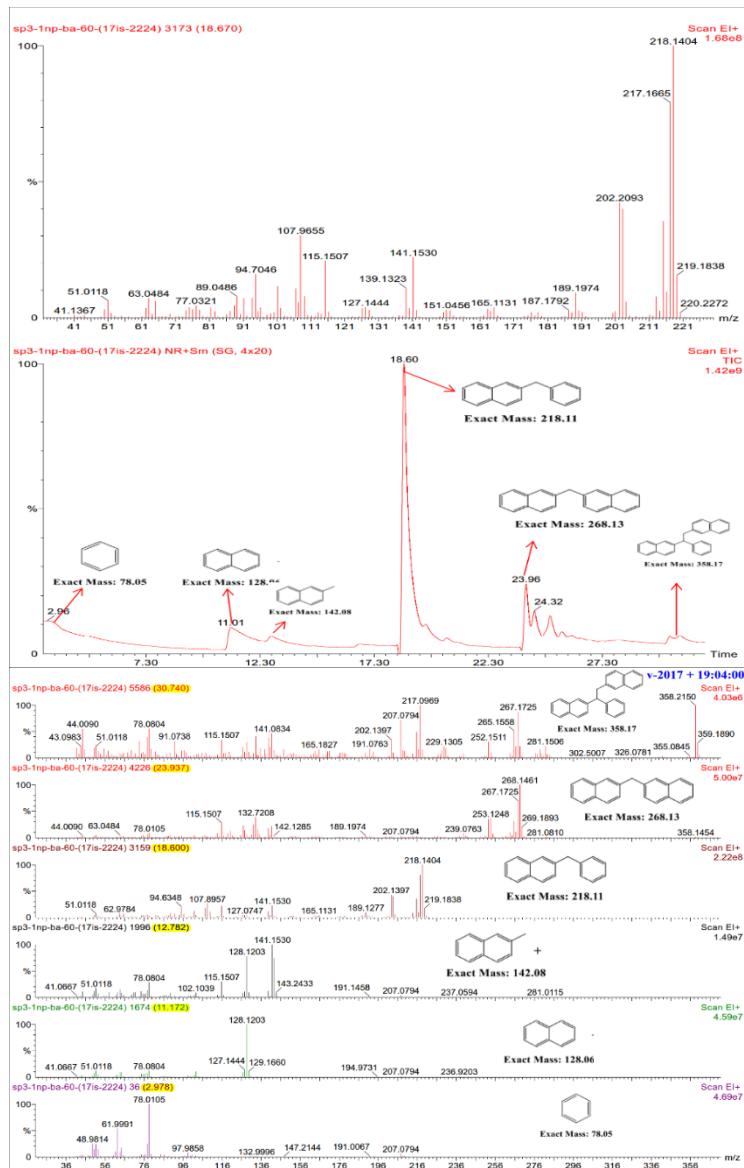


**Fig. S6.** Wide angel XRD pattern of synthesised catalysts and support. (a)  $\text{CeO}_2$  (**JCPDS Card No. 65-5923**), (b) IE4wt%AlSBA-15M, (c)  $\text{CeO}_2$ /IE4wt%AlSBA-15M.

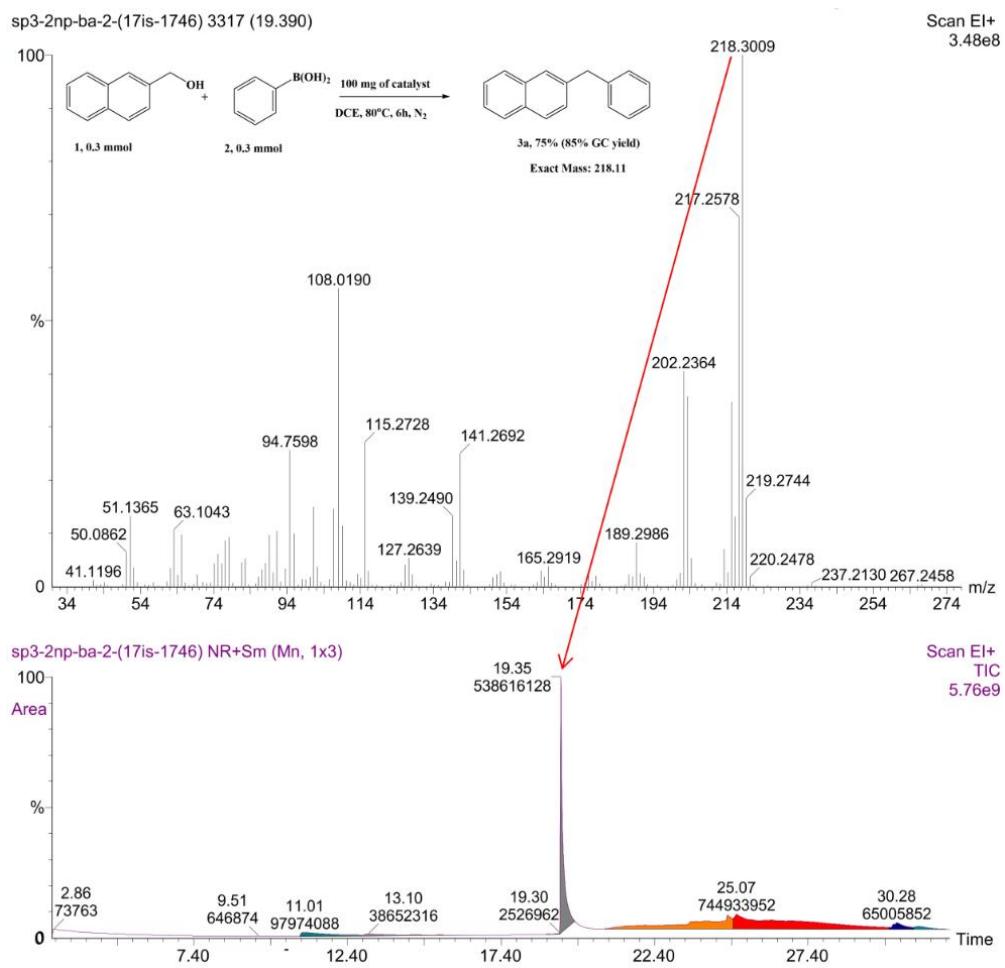
### **3. General procedure for 4wt%AlSBA-15 catalysed construction of diarylmethane using 2-naphthylmethanol and phenylboronic acid.**

The reaction was carried out by addition of 100mg AlSBA-15 catalyst to the mixture of 1 equiv. of 2-naphthalenemethanol (0.3 mmol), 1 equiv. of phenylboronic acid (0.3 mmol), in 2-3ml of dichloroethane solvent. The reaction mixture was stirred at 80°C for 6 h in N<sub>2</sub> atmosphere. After cooling, the reaction mixture was stirred with 10ml of ethyl acetate and the catalyst was separated from the reaction mixture by filtration. The filtrate was further concentrated to remove the solvent and used for GC-MS analysis. For column chromatography, 1.5 g of silica gel (100-200 mesh) was added to the residue and purified by eluting with hexane to get the desired product white solid (79%).



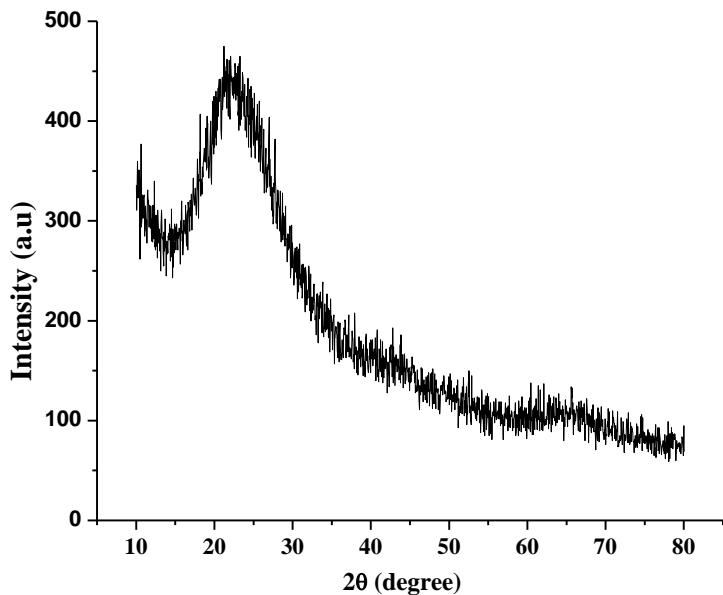


**Figure S7** GC-MS result of 4wt% AlSBA-15 catalyzed 2-naphthalenemethanol with phenylboronic acid at 60 °C (67% yield)

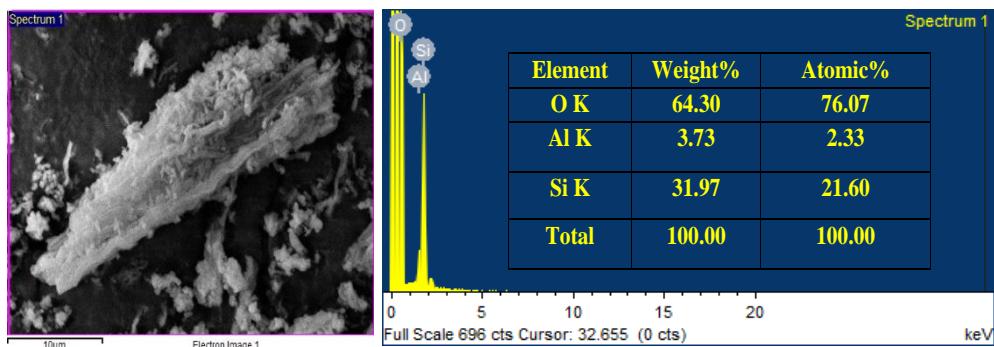


**Figure S8** GC-MS result of 4wt%AlSBA-15 catalyzed 2-naphthalenemethanol with phenylboronic acid at 80 °C (85% yield)

#### 4. Spent catalyst characterization results

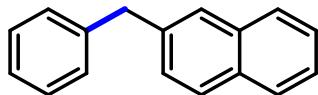


**Fig. S9.** Wide angle XRD pattern of spent catalyst 4wt% AlSBA-15.



**Fig. S10.** SEM-EDAX pattern of spent catalyst 4wt% AlSBA-15

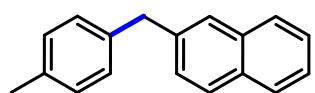
## 5. $^1\text{H}$ & $^{13}\text{C}$ NMR and GC-MS data of the obtained products



2-(4-bromobenzyl)naphthalene <sup>[2]</sup> (**3a**)

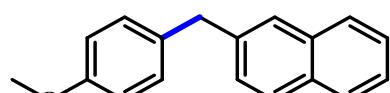
Followed the general procedure **1**, white solid (79% isolated yield).

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.77 – 7.74 (m, 3H), 7.62 (s, 1H), 7.46–7.39 (m, 2H), 7.32–7.27 (m, 3H), 7.23 – 7.20 (m, 3H), 4.13 (s, 2H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  140.00, 138.62, 133.63, 132.10, 129.09, 128.51, 128.09, 127.66, 127.64, 127.57, 127.11, 126.17, 125.99, 125.36, 42.13. GC-MS: Calc. for  $\text{C}_{17}\text{H}_{14}$  Exact Mass: 218.1096, Found Mass: 218.0726.



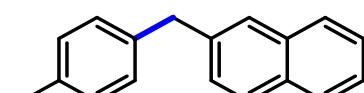
2-(4-methylbenzyl)naphthalene <sup>[2]</sup> (**3b**)

Followed the general procedure **1**, white solid (58% isolated yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.78 – 7.72 (m, 3H), 7.61 (s, 1H), 7.43 – 7.39 (m, 2H), 7.31 (d,  $J$  = 1.6 Hz, 1H), 7.10 – 7.09 (m, 4H), 4.09 (s, 2H), 2.3 (s, 3H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  138.93, 137.96, 135.67, 133.66, 132.11, 129.22, 128.94, 128.07, 127.67, 127.64, 127.58, 127.03, 125.96, 125.31, 41.72, 21.05. GC-MS: Calc. for  $\text{C}_{18}\text{H}_{16}$  Exact Mass: 232.1252 Found Mass: 232.0254.



2-(4-methoxybenzyl)naphthalene <sup>[2]</sup> (**3c**)

Followed the general procedure **1**, Colorless oil (67% isolated yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.72 – 7.65 (m, 3H), 7.53 (s, 1H), 7.36 – 7.33 (m, 2H), 7.22 (d,  $J$  = 7.6 Hz, 1H), 7.06 (d,  $J$  = 8.48 Hz, 2H), 6.75 (d,  $J$  = 8.52 Hz, 2H), 4.00 (s, 2H), 3.70 (s, 3H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  158.05, 139.09, 133.64, 133.12, 132.08, 129.98, 128.06, 127.63, 127.61, 127.56, 126.93, 125.97, 125.31, 113.94, 55.28, 41.23. GC-MS: Calc. for  $\text{C}_{18}\text{H}_{16}\text{O}$  Exact Mass: 248.1201 Found Mass: 248.0522.

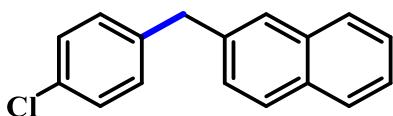


2-(4-fluorobenzyl)naphthalene <sup>[2]</sup> (**3d**)

Followed the general procedure **1**, white solid (70% isolated yield).

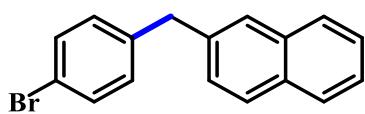
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.78 – 7.75 (m, 3H), 7.60 (s, 1H), 7.43 – 7.42 (m, 2H), 7.29 (t,  $J$  = 8.4 Hz, 1H), 7.18 (t,  $J$  = 8.0 Hz, 1H), 6.97 (t,  $J$  = 8.4 Hz, 2H), 4.10 (s, 2H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.83 (d,  $J_{\text{C}-\text{F}} = 252$  Hz), 138.43, 136.66 (d,  $J_{\text{C}-\text{F}} = 3.2$  Hz), 133.60, 132.12, 130.44

(d,  $J_{C3-F} = 7.6$  Hz), 128.21, 127.66, 127.56, 127.00, 126.67, 126.21, 125.21, 115.41(d,  $J_{C2-F} = 21.2$  Hz), 41.26. GC-MS: Calc. for C<sub>17</sub>H<sub>13</sub>F Exact Mass: 236.1001 Found Mass: 236.1233.



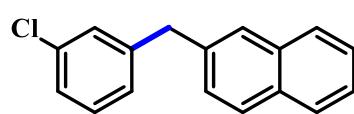
2-(4-chlorobenzyl)naphthalene <sup>[2]</sup> (**3e**)

Followed the general procedure **1**, white solid (70% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.80 – 7.75 (m, 3H), 7.60 (s, 1H), 7.45 – 7.42 (m, 2H), 7.22 – 7.19 (m, 3H), 7.09 (d,  $J = 8.32$  Hz, 2H), 4.04 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 139.45, 138.03, 133.58, 132.14, 131.99, 130.37, 128.62, 128.26, 127.66, 127.56, 127.43, 127.13, 126.13, 125.53, 41.42. GC-MS: Calc. for C<sub>17</sub>H<sub>13</sub>Cl Exact Mass: 252.0706, Found Mass: 252.0514.



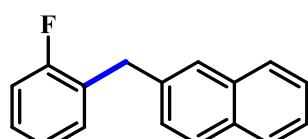
2-(4-bromobenzyl)naphthalene <sup>[3]</sup> (**3f**)

Followed the general procedure **1**, white solid (65% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.80 – 7.75 (m, 3H), 7.60 (s, 1H), 7.45 – 7.39 (m, 4H), 7.28 (s, 1H), 7.10 (d,  $J = 8$  Hz, 2H), 4.08 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 139.97, 139.31, 137.92, 133.58, 132.14, 131.57, 130.78, 128.26, 127.66, 127.55, 127.42, 127.14, 126.13, 125.54, 120.05, 41.48. GC-MS: Calc. for C<sub>17</sub>H<sub>13</sub>Br Exact Mass: 296.0201 Found Mass: 296.0501.



2-(3-chlorobenzyl)naphthalene <sup>[2]</sup> (**3g**)

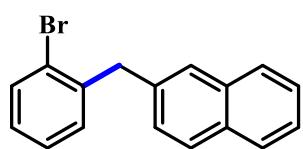
Followed the general procedure **1**, white solid (65% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.75 – 7.54 (m, 3H), 7.40 (s, 1H), 7.39 – 7.37 (m, 2H), 7.22 – 7.19 (m, 3H), 7.09 (d,  $J = 8$  Hz, 2H), 4.04 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 139.42, 138.0, 133.55, 132.11, 131.96, 130.34, 128.58, 128.22, 127.63, 127.52, 127.40, 127.10, 126.09, 125.49, 41.38. GC-MS: Calc. for C<sub>17</sub>H<sub>13</sub>Cl Exact Mass: 252.0706, Found Mass: 252.1511.



2-(2-fluorobenzyl)naphthalene <sup>[2]</sup> (**3h**)

Followed the general procedure **1**, white solid (63% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.80 – 7.75 (m, 3H), 7.65 (s, 1H), 7.44 – 7.41 (m, 2H), 7.37 (d,  $J = 7.32$  Hz, 1H), 7.19 – 7.16 (m, 2H), 7.07 – 6.97 (m, 2H), 4.16 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 162.03 (d,  $J_{C1-F} = 244$  Hz) 159.51, 137.34, 133.60, 132.14, 131.14 (d,  $J_{C4-F} = 5$  Hz), 128.20 (d,  $J_{C3-F} = 8$  Hz), 128.07, 127.99, 127.63 (d,  $J_{C2-F} = 6$  Hz), 127.42, 126.09

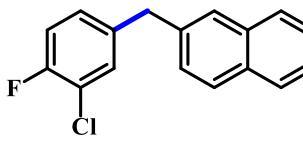
(d,  $J_{C6-F} = 7$  Hz), 125.43, 124.09, 115.47 (d,  $J_{C6-F} = 22$  Hz), 34.94 (d,  $J_{C7-F} = 3$  Hz). GC-MS: Calc. for C<sub>17</sub>H<sub>13</sub>F Exact Mass: 236.1001 Found Mass: 236.0925.



2-(2-bromobenzyl)naphthalene <sup>[4]</sup> (**3i**)

Followed the general procedure **1**, colorless oil (60% isolated yield).

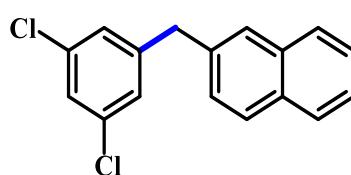
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.81 – 7.75 (m, 3H), 7.59 (d,  $J = 8.88$  Hz, 2H), 7.45 – 7.42 (m, 2H), 7.34 (d,  $J = 8.36$  Hz, 1H), 7.21 (d,  $J = 7.48$  Hz, 1H), 7.16 – 7.10 (m, 2H), 4.27 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 140.27, 137.02, 133.59, 132.91, 131.19, 128.11, 127.99, 127.64, 127.61, 127.56, 127.51, 127.37, 126.02, 125.46, 124.97, 41.91. GC-MS: Calc. for C<sub>17</sub>H<sub>13</sub>Br Exact Mass: 296.0201 Found Mass: 296.0363.



2-(3-chloro-4-fluorobenzyl) naphthalene (**3j**)

Followed the general procedure **1**, colorless oil (60% isolated yield).

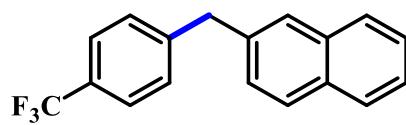
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.81 – 7.76 (m, 3H), 7.60 (s, 1H), 7.49 – 7.42 (m, 2H), 7.27 – 7.23 (m, 2H), 7.08 – 7.02 (m, 2H), 4.07 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 157.98 (d,  $J_{Cl-F} = 245.5$  Hz), 138.04 (d,  $J_{C4-F} = 4.05$  Hz), 137.53, 133.57, 132.21, 130.92, 128.60 (d,  $J_{C3-F} = 7.07$  Hz), 128.40, 127.69 (d,  $J_{C2-Cl-F} = 11.09$  Hz), 127.30, 127.20, 126.23, 125.66, 120.91, 120.73, 116.59 (d,  $J_{C2-F} = 20.9$  Hz), 41.04. GC-MS: Calc. for C<sub>17</sub>H<sub>12</sub>ClF Exact Mass: 270.0612, Found Mass: 270.0588.



2-(3,5-dichlorobenzyl)naphthalene (**3k**)

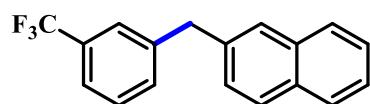
Followed the general procedure **1**, white solid (55% isolated yield).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) : δ (ppm) 7.81 – 7.76 (m, 3H), 7.60 (s, 1H), 7.47 – 7.44 (m, 2H), 7.25 (d,  $J = 8.44$  Hz, 1H), 7.20 (s, 1H), 7.09 (d,  $J = 1.56$  Hz, 2H), 4.05 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 144.40, 136.76, 134.96, 133.58, 132.29, 128.54, 127.73, 127.63, 127.50, 127.41, 127.28, 126.52, 126.31, 125.79, 41.5. GC-MS: Calc. for C<sub>17</sub>H<sub>12</sub>Cl<sub>2</sub> Exact Mass: 286.0316, Found Mass: 286.0924.



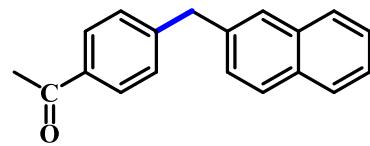
2-(4-(trifluoromethyl)benzyl)naphthalene<sup>[2]</sup> (**3l**)

Followed the general procedure **1**, white solid (60% isolated yield).<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.72 – 7.53 (m, 3H), 7.46 (s, 1H), 7.45 (d, *J* = 8 Hz, 2H), 7.39 – 7.36 (m, 2H), 7.34 (d, *J* = 7.46 Hz, 2H), 7.23 (d, *J* = 8 Hz, 1H), 4.09 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 145.11, 137.46, 133.61, 132.22, 128.73, 128.40, 127.71, 127.59, 127.41, 127.32, 126.24, 125.68, 125.52, 125.49, 125.45, 122.99, 41.89. GC-MS: Calc. for C<sub>18</sub>H<sub>13</sub>F<sub>3</sub> Exact Mass: 286.0969, Found Mass: 286.0929



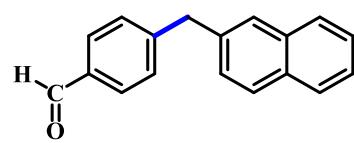
2-(3-(trifluoromethyl)benzyl)naphthalene (**3m**)

Followed the general procedure **1**, white solid (60% isolated yield).<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.82 – 7.77 (m, 3H), 7.63 (s, 1H), 7.54 (d, *J* = 8 Hz, 2H), 7.47 – 7.44 (m, 2H), 7.33 – 7.27 (m, 3H), 4.19 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 140.58, 132.42, 131.96, 131.64, 131.32, 131.00, 130.53, 129.55, 128.94, 128.39, 128.09, 127.68, 127.59, 127.36, 126.19, 125.63, 125.38, 124.77, 124.01, 123.15, 122.67, 41.86. GC-MS: Calc. for C<sub>18</sub>H<sub>13</sub>F<sub>3</sub> Exact Mass: 286.0969, Found Mass: 286.0950.



1-(4-(naphthalen-2-ylmethyl)phenyl)ethan-1-one<sup>[2]</sup> (**3n**)

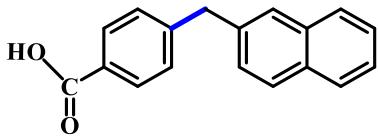
Followed the general procedure **1**, white solid (60% isolated yield).<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.83 – 7.76 (m, 4H), 7.71 (d, *J* = 8.4 Hz, 1H), 7.39 – 7.33 (m, 3H), 7.23 – 7.17 (m, 3H), 4.41 (s, 2H), 2.47 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 196.75, 145.44, 134.53, 134.25, 132.97, 130.94, 127.82, 127.75, 127.59, 126.51, 126.50, 125.12, 124.68, 124.52, 123.02, 38.10, 26.51. GC-MS: Calc. for C<sub>19</sub>H<sub>16</sub>O Exact Mass: 260.1201, Found Mass: 260.0920.



4-(naphthalen-2-ylmethyl)benzaldehyde<sup>[2]</sup> (**3o**)

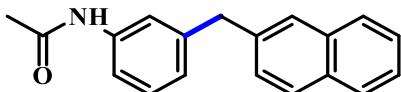
Followed the general procedure **1**, white solid (60% isolated yield).<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.96 (s, 1H), 7.80 – 7.76 (m, 5H), 7.62 (s, 1H), 7.46 – 7.43 (m, 2H), 7.37 (d, *J* = 7.92 Hz, 2H), 7.27 (d, *J* = 8.4 Hz, 1H), 4.19 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 192.02, 148.31, 137.25, 133.61, 132.22, 130.11, 129.70,

128.44, 127.72, 127.59, 127.41, 127.39, 126.27, 125.71, 42.27. GC-MS: Calc. for C<sub>18</sub>H<sub>14</sub>O Exact Mass: 246.1045, Found Mass: 246.0955.



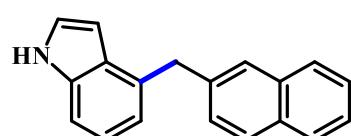
4-(naphthalen-2-ylmethyl)benzoic acid <sup>[5]</sup> (**3p**)

Followed the general procedure **1**, white solid (55% isolated yield). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ 12.81 (s, 1H), 8.01 (d, J = 9.28 Hz, 1H), 7.93 (d, J = 5.36 Hz, 1H), 7.85 (m, 3H), 7.49 (m, 3H), 7.44 (d, J = 6.8 Hz, 1H), 7.35 – 7.33 (d, J = 7.96 Hz, 2H), 4.51 (s, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>): δ 167.63, 146.60, 136.55, 134.06, 131.89, 129.93, 129.10, 129.08, 129.04, 127.94, 127.61, 126.60, 126.18, 126.16, 124.64, 38.51. GC-MS: Calc. for C<sub>18</sub>H<sub>14</sub>O<sub>2</sub> Exact Mass: 262.0994 Found Mass: 262.1025.



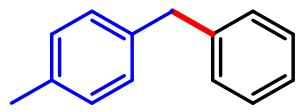
N-(3-(naphthalen-2-ylmethyl)phenyl)acetamide (**3q**)

Followed the general procedure **1**, white solid (45% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.72 (s, 1H), 8.05 – 7.98 (m, 1H), 7.87 – 7.77 (m, 2H), 7.75 – 7.68 (d, J = 7.7 Hz, 1H), 7.61 – 7.55 (d, J = 1.3 Hz, 1H), 7.51 – 7.41 (m, 3H), 7.38 – 7.26 (m, 2H), 7.07 – 7.00 (d, J = 8.0 Hz, 1H), 4.19 (s, 2H), 2.18 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 170.02, 138.70, 135.70, 133.59, 133.37, 132.78, 128.56, 127.71, 126.43, 126.32, 125.94, 125.39, 125.07, 122.91, 43.88, 23.30. GC-MS: Calc. for C<sub>19</sub>H<sub>17</sub>NO Exact Mass: 275.1310, Found Mass: 275.1315.



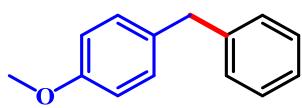
4-(naphthalen-2-ylmethyl)-1H-indole (**3r**)

Followed the general procedure **1**, white solid (50% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.89 (s, 1H), 7.73 – 7.62 (m, 3H), 7.58 (d, J = 7.8 Hz, 1H), 7.47 (d, J = 7.8 Hz, 1H), 7.37 – 7.33 (m, 3H), 7.14 – 7.12 (m, 2H), 7.06 – 6.97 (m, 2H), 6.85 (s, 1H), 6.48 (s, 1H), 4.20 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 136.92, 135.82, 133.16, 132.82, 131.98, 128.44, 127.92, 127.84, 127.61, 127.38, 126.63, 126.14, 126.13, 123.94, 121.61, 121.32, 111.95, 101.29, 38.04. GC-MS: Calc. for C<sub>19</sub>H<sub>15</sub>N Exact Mass: 257.1204 Found Mass: 257.1656.



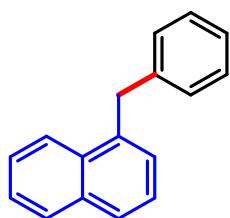
1-benzyl-4-methylbenzene<sup>[6]</sup> (**4a**)

Followed the general procedure **1**, white solid (65% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.28 – 7.24 (m, 2H), 7.22 – 7.16 (m, 3H), 7.10 – 7.05 (m, 4H), 3.93 (s, 2H), 2.33 – 2.30 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 141.46, 138.12, 135.58, 129.19, 128.91, 128.85, 128.47, 126.02, 41.55, 21.05. GC-MS: Calc. For C<sub>14</sub>H<sub>14</sub> Exact Mass: 182.1096 Found Mass: 182.0126.



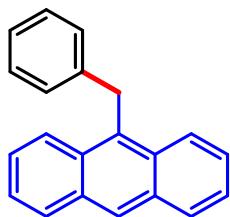
1-benzyl-4-methoxybenzene<sup>[6]</sup> (**4b**)

Followed the general procedure **1**, colorless oil (60% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.25 – 7.20 (m, 6H), 7.05 (s, 1H), 6.86 (s, 2H), 3.97 (s, 2H), 3.81 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 157.33, 141.02, 130.31, 129.66, 128.97, 128.25, 127.40, 125.77, 120.46, 110.39, 55.35, 35.85. GC-MS: Calc. for C<sub>14</sub>H<sub>14</sub>O Exact Mass: 198.1045, Found Mass: 198.0984.



2-(benzyl)naphthalene<sup>[2]</sup> (**4c**)

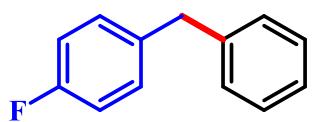
Followed the general procedure **1**, white solid (55% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.99 (t, *J* = 6.4 Hz, 1H), 7.86 – 7.80 (t, *J* = 6.4 Hz, 1H), 7.77 (d, *J* = 8.2 Hz, 1H), 7.46 – 7.40 (m, 3H), 7.29 – 7.24 (m, 3H), 7.20 - 7.18 (m, 3H) 4.45 (s, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 140.65, 136.64, 133.95, 132.15, 128.76, 128.68, 128.47, 127.35, 127.17, 126.07, 125.99, 125.57, 124.29, 39.06. GC-MS: Calc. for C<sub>17</sub>H<sub>14</sub> Exact Mass: 218.1096, Found Mass: 218.1096.



9-benzylnanthracene<sup>[7]</sup> (**4d**)

Followed the general procedure **1**, white solid (65% isolated yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.37 (s, 1H), 8.16 – 8.13 (m, 2H), 7.98 – 7.95 (m, 2H), 7.40 – 7.37 (m, 4H), 7.12 – 7.04 (m, 5H), 4.95 (s, 2H). <sup>13</sup>C NMR (100 MHz,

$\text{CDCl}_3$ )  $\delta$  140.84, 131.67, 130.55, 129.13, 128.47, 128.13, 126.57, 125.96, 125.90, 124.93, 124.81, 33.55. GC-MS: Calc. for  $\text{C}_{22}\text{H}_{17}\text{ClO}$  Exact Mass: 268.1252, Found Mass: 268.1240.



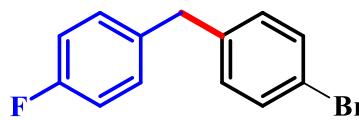
1-benzyl-4-fluorobenzene <sup>[8]</sup> (**4e**)

Followed the general procedure **1**, white solid (70% isolated yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.23 – 7.17 (m, 2H), 7.14 – 7.04 (m, 5H), 6.89 (t,  $J = 8.1$  Hz, 2H), 3.87 (s, 2H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.64 (d,  $J_{\text{C}1-\text{F}} = 242.3$  Hz), 140.95, 136.79 (d,  $J_{\text{C}4-\text{F}} = 3.1$  Hz), 130.33, 128.84 (d,  $J_{\text{C}3-\text{F}} = 29.5$  Hz), 126.21, 115.32 (d,  $J_{\text{C}2-\text{F}} = 20.9$  Hz), 41.09. GC-MS: CALC. For  $\text{C}_{13}\text{H}_{11}\text{F}$  Exact Mass: 186.0845 Found Mass: 186.0226.



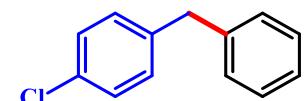
1-chloro-4-(4-fluorobenzyl)benzene <sup>[9]</sup> (**4f**)

Followed the general procedure **1**, Colorless oil (56% isolated yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.18 – 7.03 (m, 2H), 7.02 – 6.91 (m, 4H), 6.89 – 6.87 (d,  $J = 8.3$  Hz, 2H), 3.83 (s, 2H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.75 (d,  $J_{\text{C}1-\text{F}} = 243$  Hz), 139.41, 136.24 (d,  $J_{\text{C}4-\text{F}} = 3$  Hz), 132.06, 130.30 (d,  $J_{\text{C}3-\text{F}} = 8$  Hz), 128.66, 115.47 (d,  $J_{\text{C}2-\text{F}} = 21$  Hz), 40.40. GC-MS: Calc. For  $\text{C}_{13}\text{H}_{10}\text{ClF}$  Exact Mass: 220.0455 Found Mass: 220.0485.



1-bromo-4-(4-fluorobenzyl)benzene <sup>[10]</sup> (**4g**)

Followed the general procedure **1**, Colorless oil (62% isolated yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.30 (d,  $J = 8.4$  Hz, 2H), 7.01 – 6.98 (m, 2H), 6.92 (d,  $J = 8$  Hz, 2H), 6.85 (t,  $J = 8.1$  Hz, 2H), 3.78 (s, 2H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  162.77 (d,  $J_{\text{C}1-\text{F}} = 243$  Hz), 139.95, 136.15 (d,  $J_{\text{C}4-\text{F}} = 3$  Hz), 131.63, 130.58, 130.31 (d,  $J_{\text{C}3-\text{F}} = 7.67$  Hz), 120.12, 115.49 (d,  $J_{\text{C}2-\text{F}} = 21.1$  Hz), 40.47. GC-MS: Calc. For  $\text{C}_{13}\text{H}_{10}\text{BrF}$ : 263.9950, Found Mass: 263.9825.



1-benzyl-4-chlorobenzene <sup>[6]</sup> (**4h**)

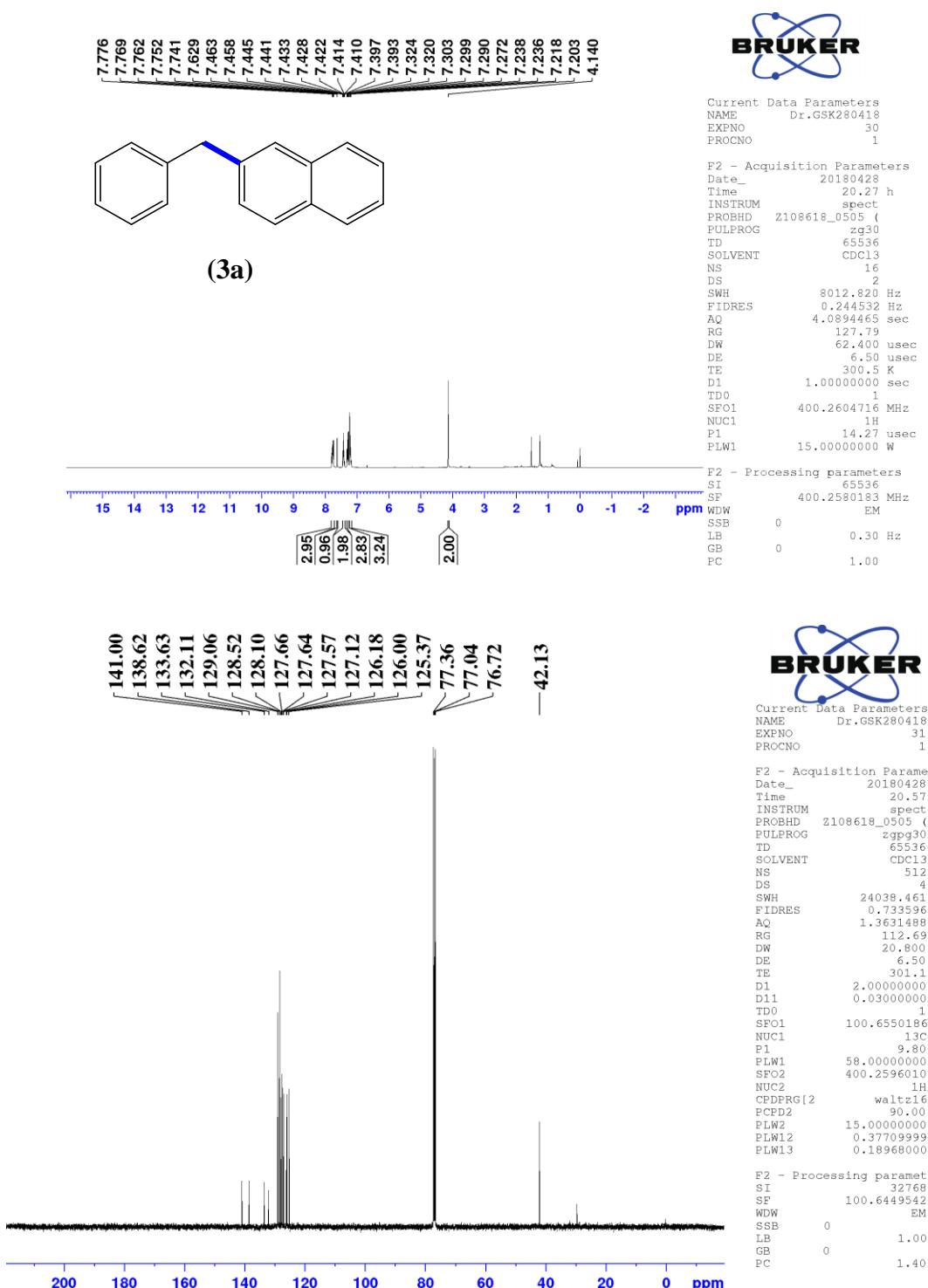
Followed the general procedure **1**, white solid (55% isolated yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.30 – 7.23 (m, 4H), 7.20 (s, 1H), 7.15 (d,  $J = 7.44$  Hz, 2H), 7.10 (d,  $J = 8.2$  Hz, 2H), 3.94 (s, 2H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  140.57, 139.60, 131.90, 130.27,

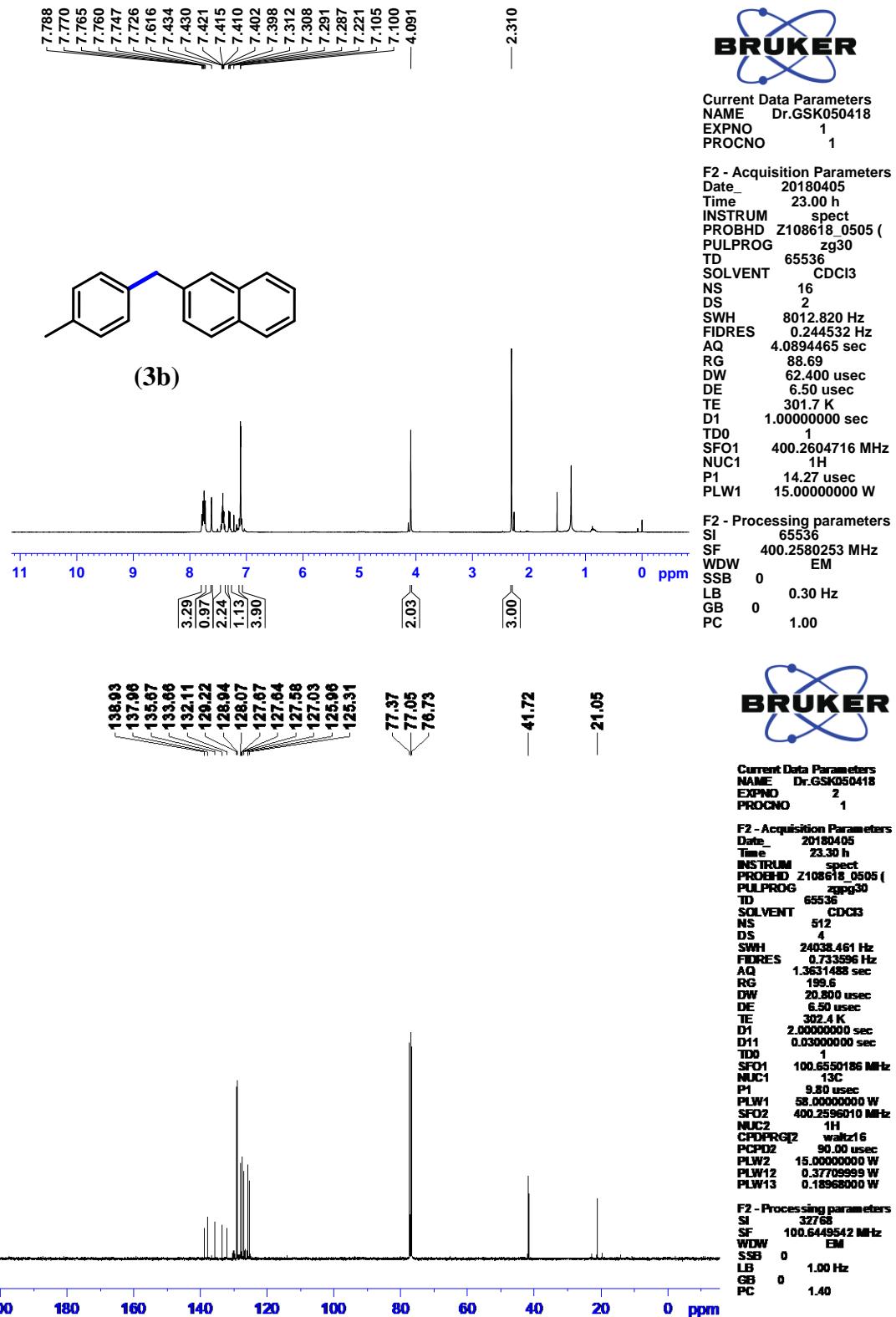
128.87, 128.57, 126.30, 41.25. GC-MS: Calc. For C<sub>13</sub>H<sub>11</sub>Cl Exact Mass: 202.0549, Found Mass: 202.0858

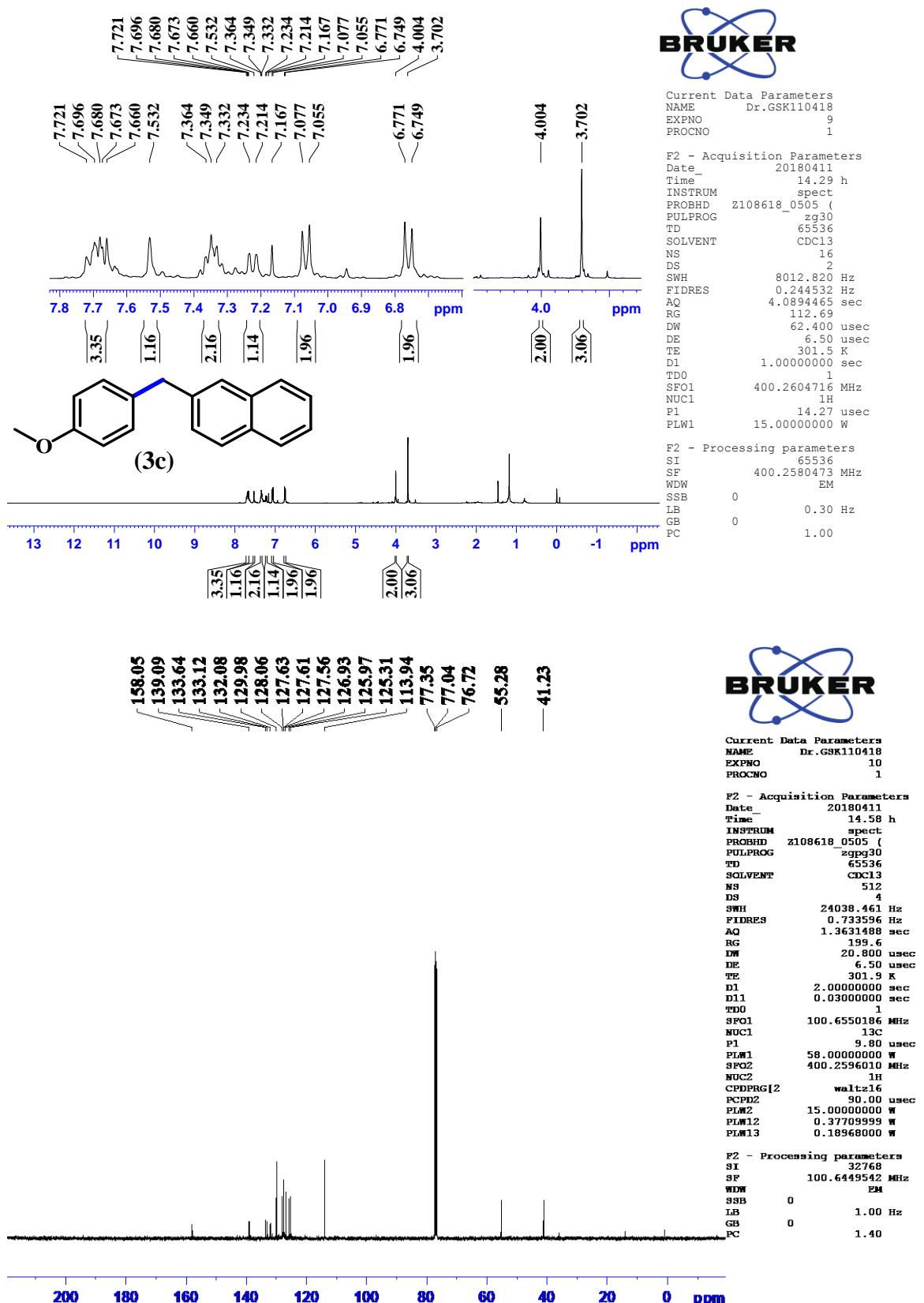
## 6. Reference

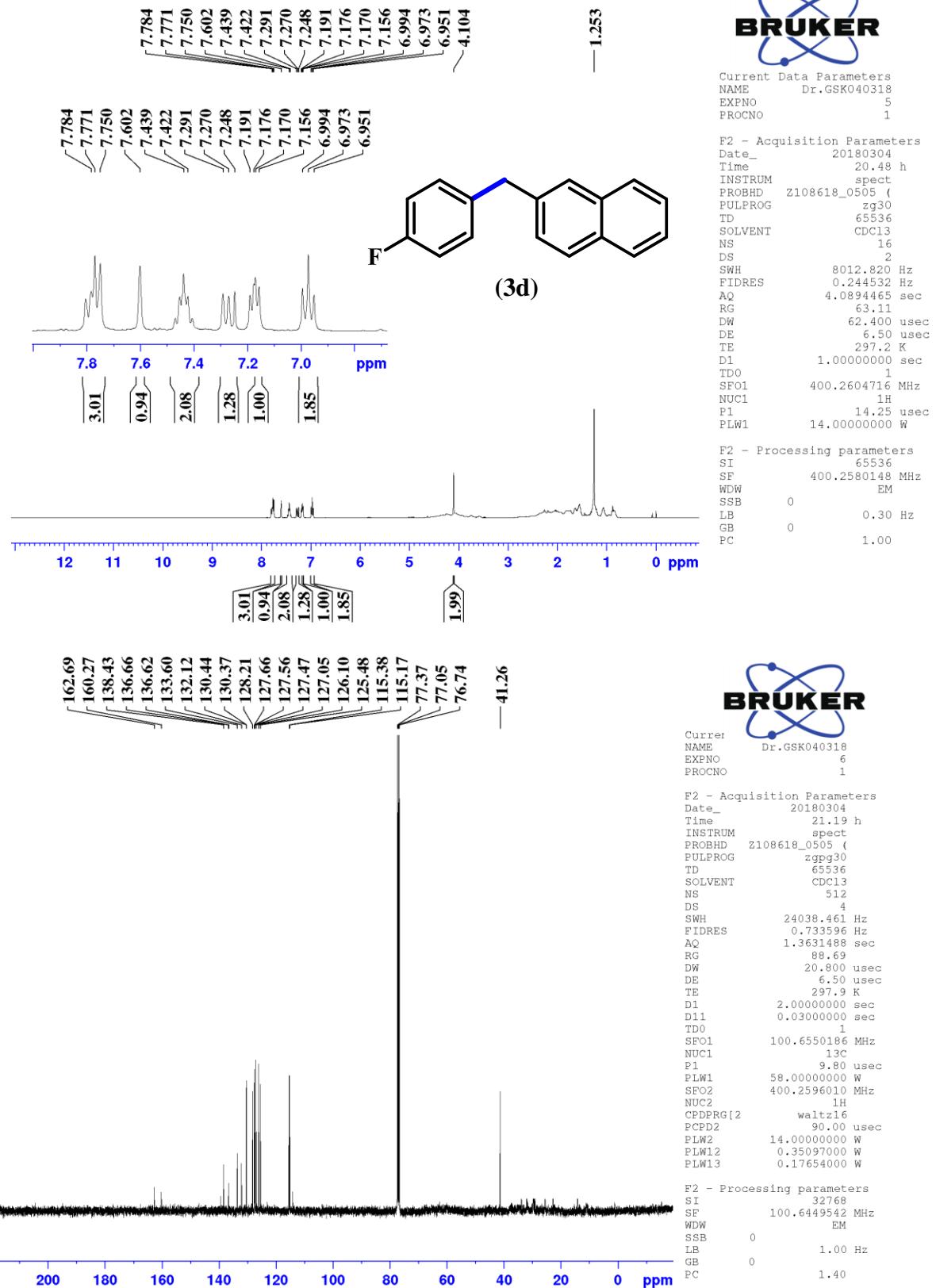
1. I. S. Pieta, M. Ishaq, R. P. K. Wells, J. A. Anderson, *Applied Catalysis A: General*, **2010**, *390*, 127-134.
2. Z. C. Cao, D.G. Yu, R.Y. Zhu, J. B. Wei, Z. J. Shi, *Chem. Commun.*, **2015**, *51*, 2683-2686.
3. X. Mo, J. Yakiwchuk, J. Dansereau, J. A. McCubbin, D. G. Hall, *J. Am. Chem. Soc.*, **2015**, *137* (30), 9694-9703.
4. C. Ramesh Kumar, K. T. V. Rao, P. S. Sai Prasad, N. Lingaiah, *J. Mol., Catal., A: Chemical* **2011**, *337* (1), 17-24.
5. M. Hasegawa, K. Takenouchi, K. Takahashi, T. Takeuchi, K. Komoriya, Y. Uejima, T. Kamimura, *J. Med., chemistry* **1997**, *40* (4), 395-407.
6. M. McLaughlin, *Org. Lett.*, **2005**, *7*, 4875-4878.
7. D. G. Yu, X. Wang, R. Y. Zhu, S. Luo, X. B. Zhang, B. Q. Wang, Z. J. Shi, *J. Am. Chem. Soc.*, **2012**, *134*, 14638-14641.
8. T. Suga, Y. Ukaji, *Org. Lett.*, **2018**, *20* (24), 7846-7850.
9. J. R. Schmink, N. E. Leadbeater, *Org. Lett.*, **2009**, *11* (12), 2575-2578.
10. L. Bering, K. Jeyakumar, A. P. Antonchick, *Org. Lett.*, **2018**, *20* (13), 3911-3914.

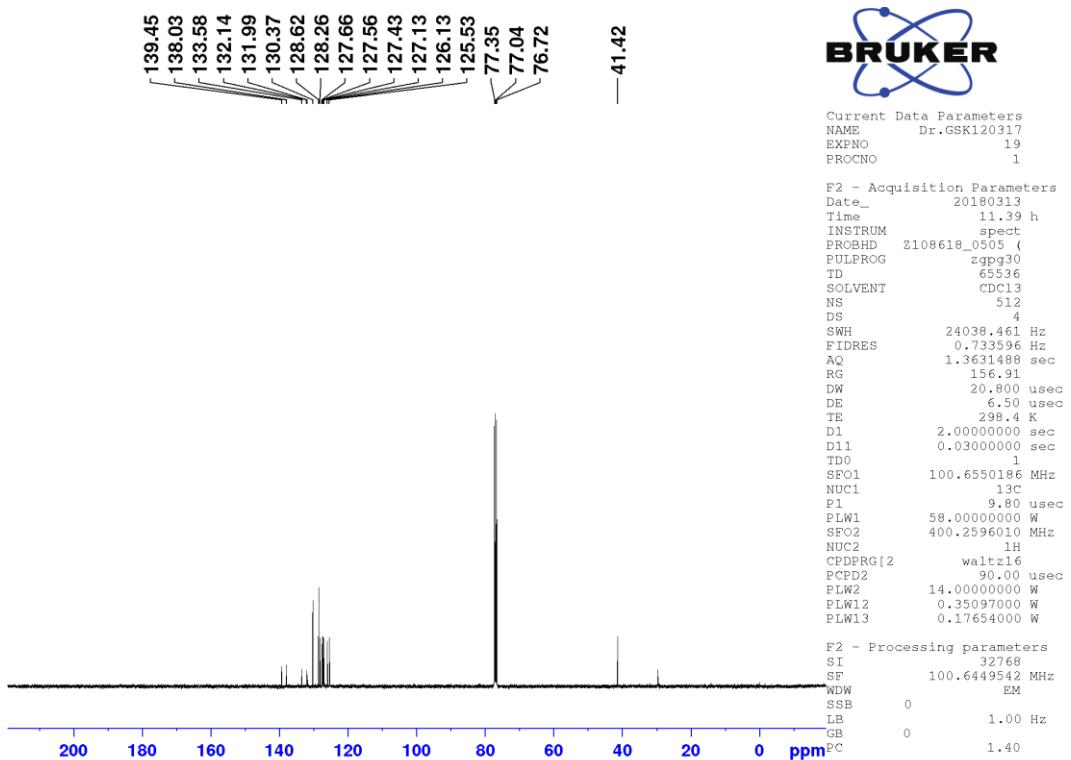
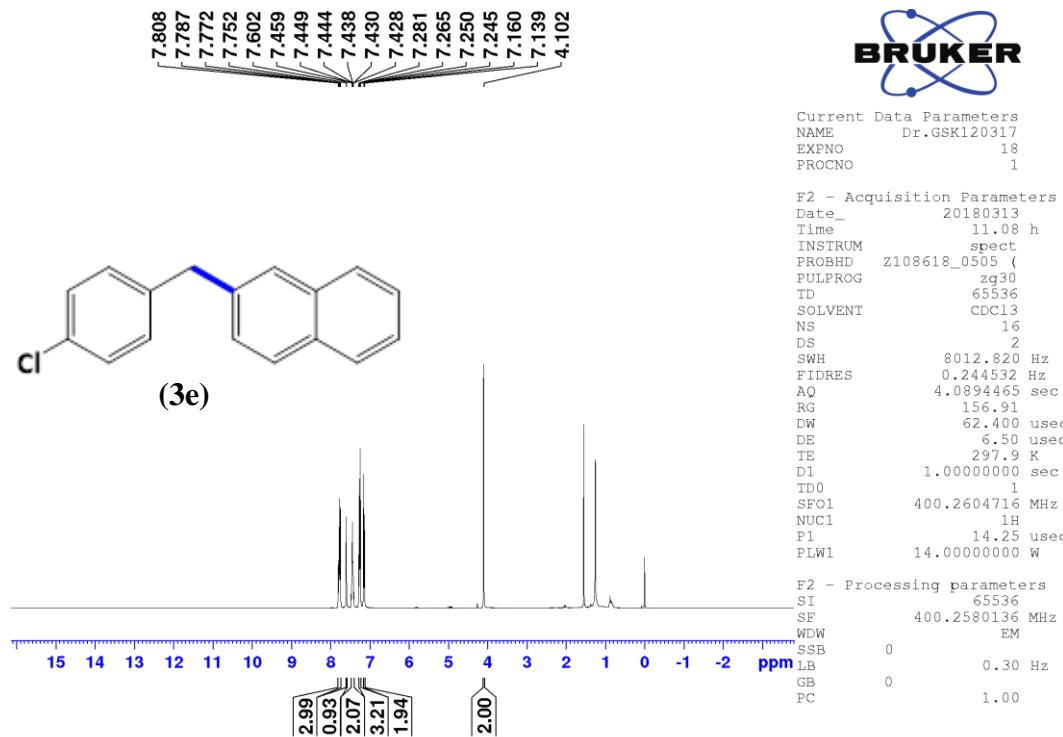
## 7. H<sup>1</sup> and C<sup>13</sup> NMR Spectra of all compounds

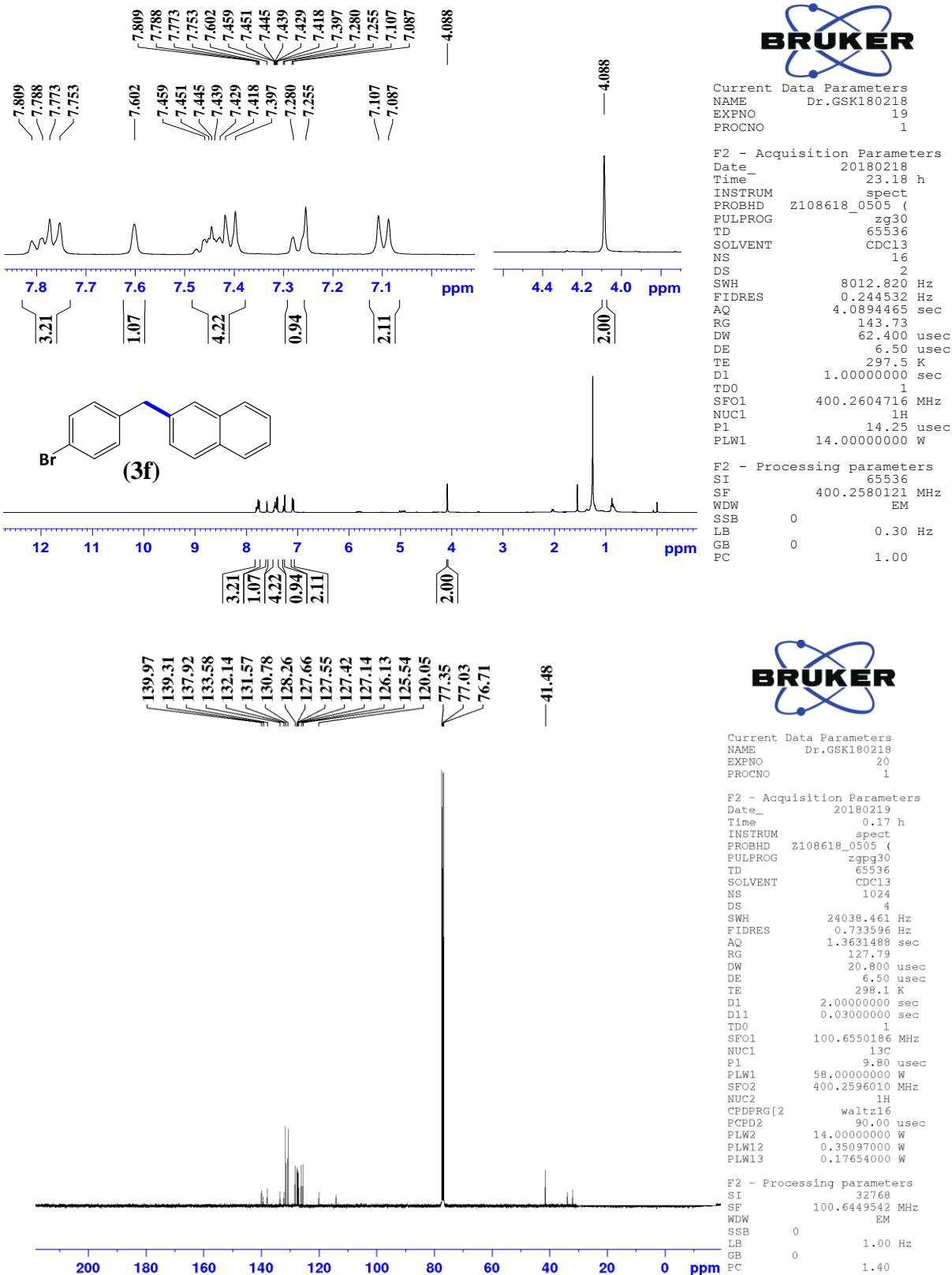


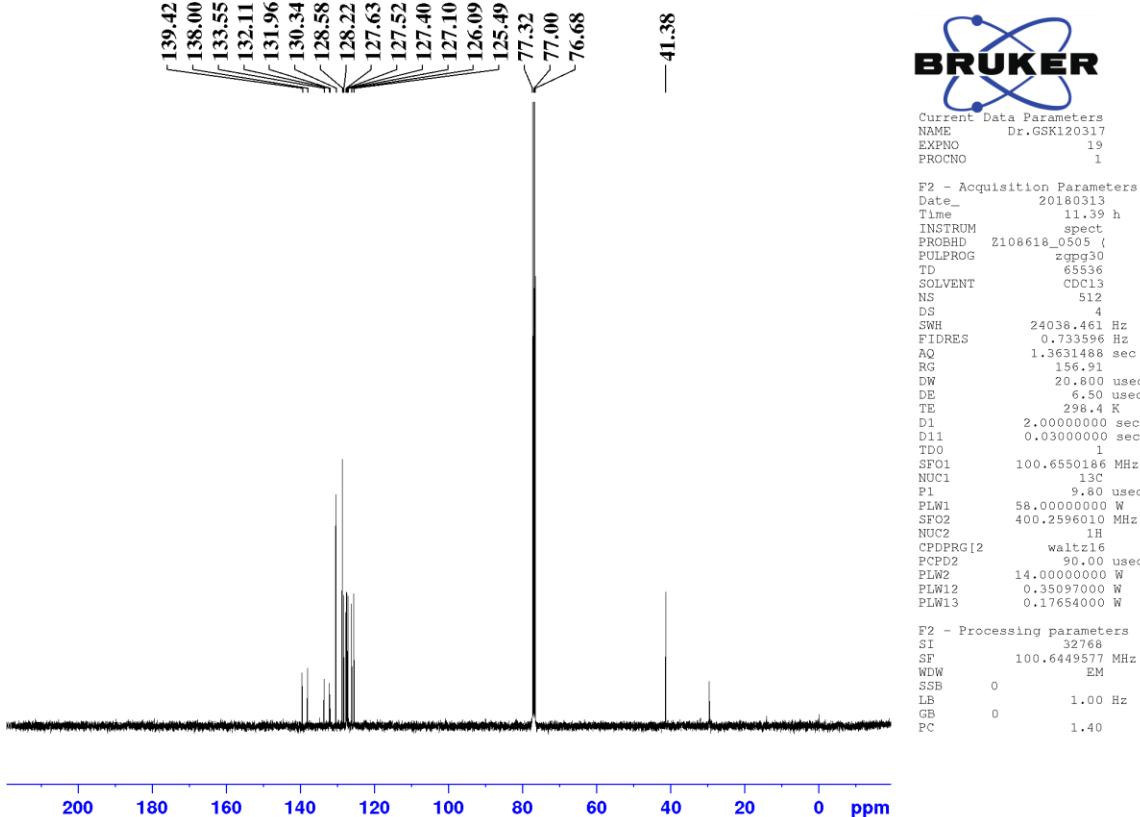
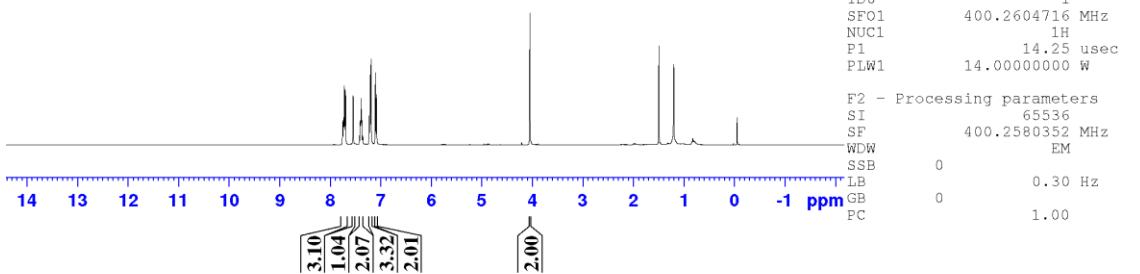
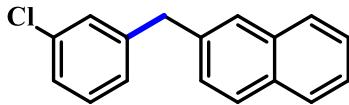












200 180 160 140 120 100 80 60 40 20 0 ppm



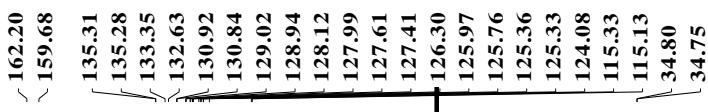
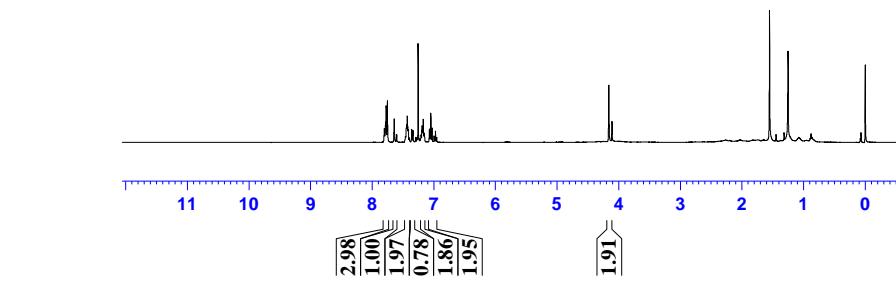
(3h)



Current Data Parameters  
NAME Dr.GSK291118  
EXPNO 13  
PROCNO 1

F2 - Acquisition Parameters  
Date 20181130  
Time 9.14 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 16  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 175.97  
DW 62.400 usec  
DE 6.50 usec  
TE 297.7 K  
D1 1.0000000 sec  
TDO 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 14.00 usec  
PLW1 16.0000000 W

F2 - Processing parameters  
SI 65536  
SF 400.2580105 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

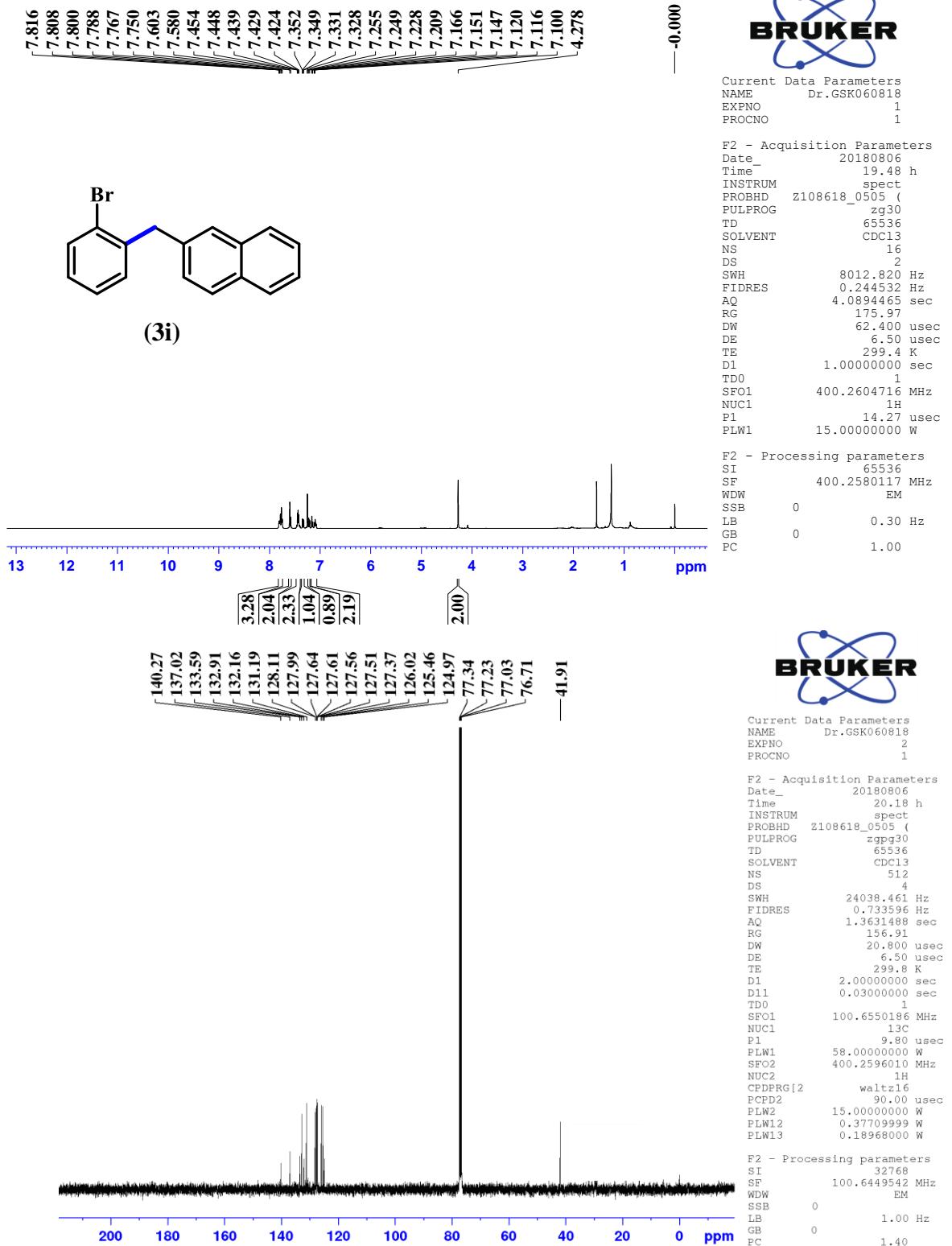


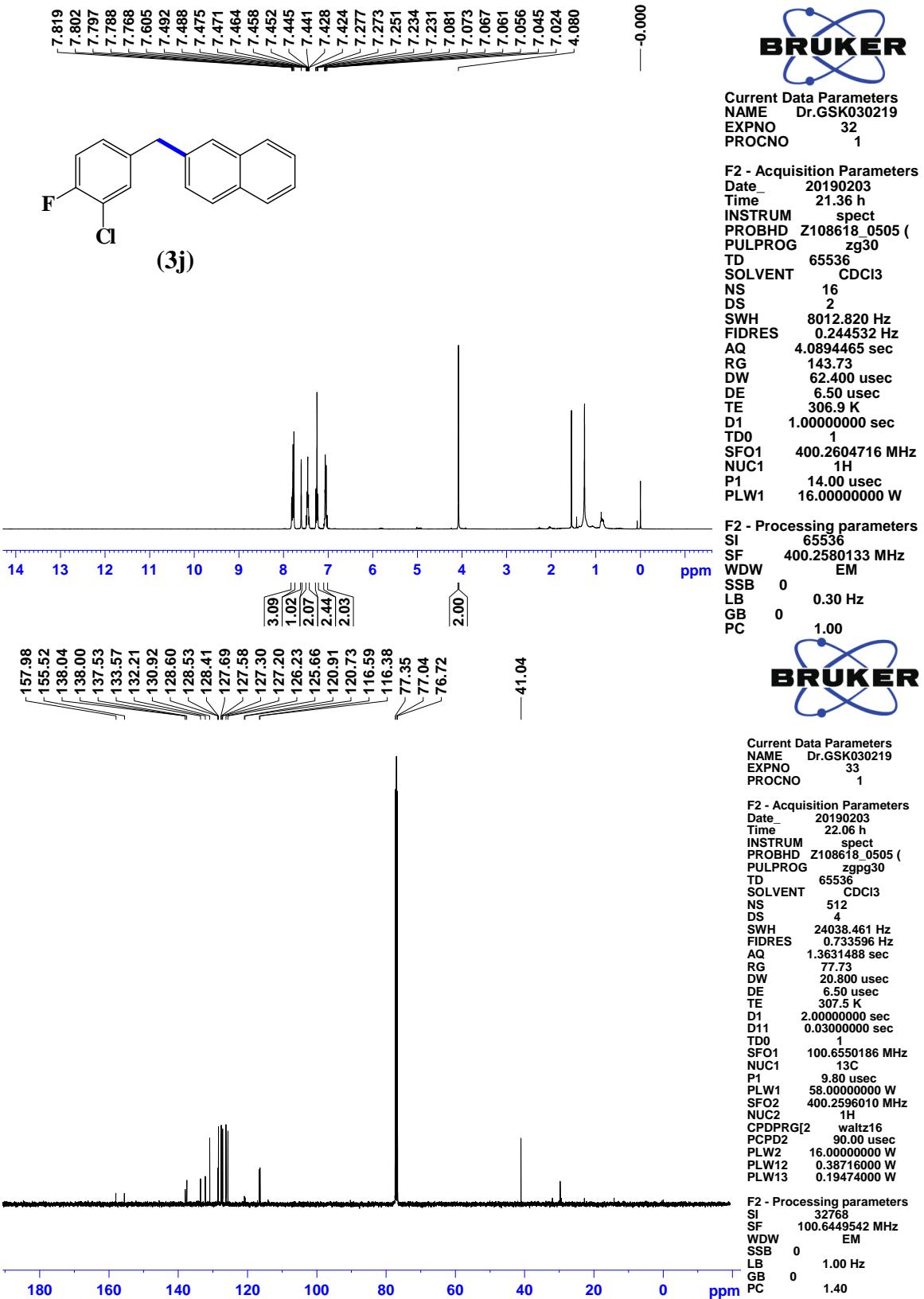
Current Data Parameters  
NAME Dr.GSK291118  
EXPNO 14  
PROCNO 1

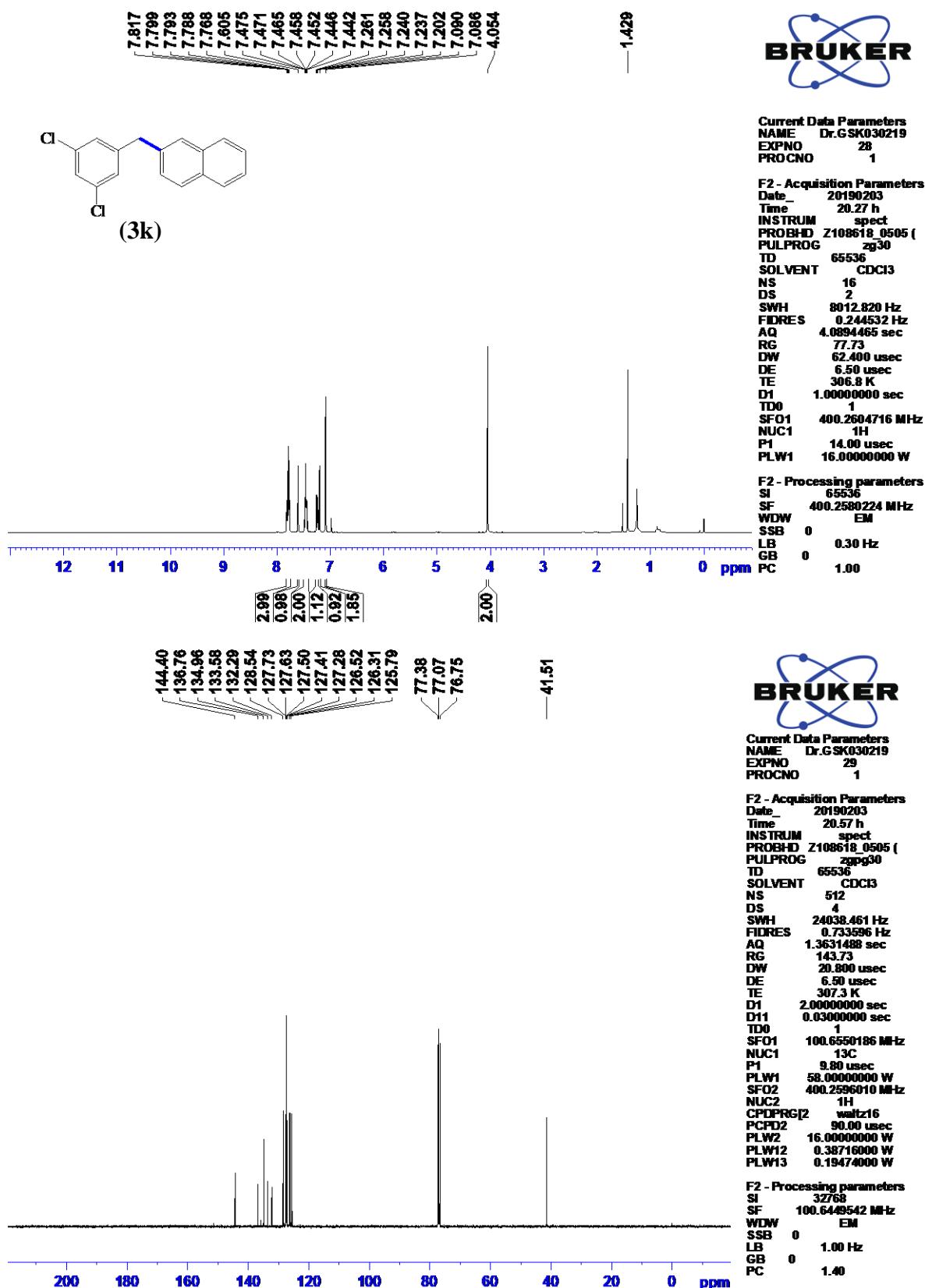
F2 - Acquisition Parameters  
Date 20181130  
Time 9.44 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 77.73  
DW 20.800 usec  
DE 6.50 usec  
TE 297.9 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TDO 1  
SFO1 100.6550186 MHz  
NUC1 13C  
P1 9.80 usec  
PLW1 58.00000000 W  
SFO2 400.2596010 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 90.00 usec  
PLW2 16.00000000 W  
PLW12 0.38716000 W  
PLW13 0.19474000 W

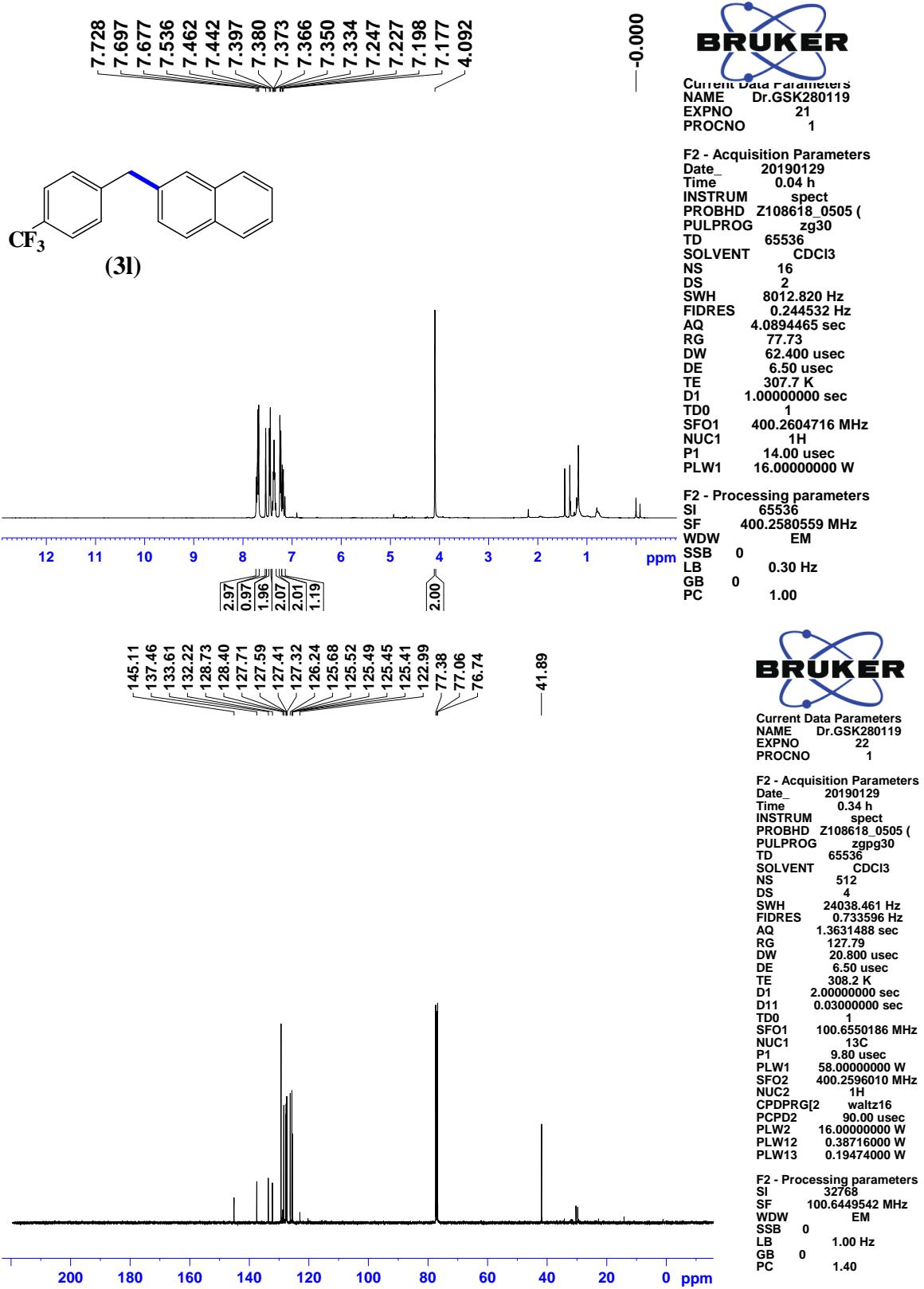
F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

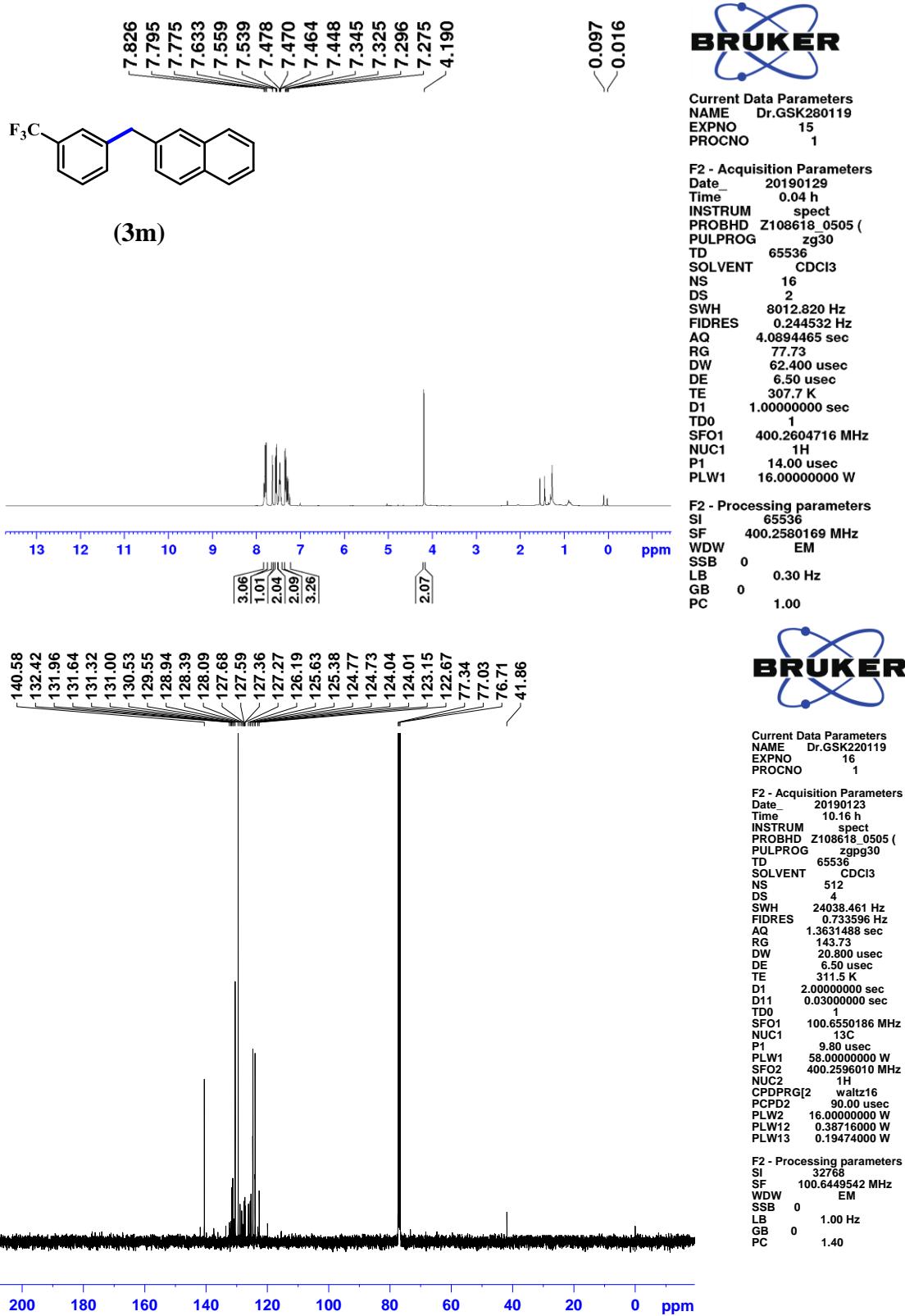


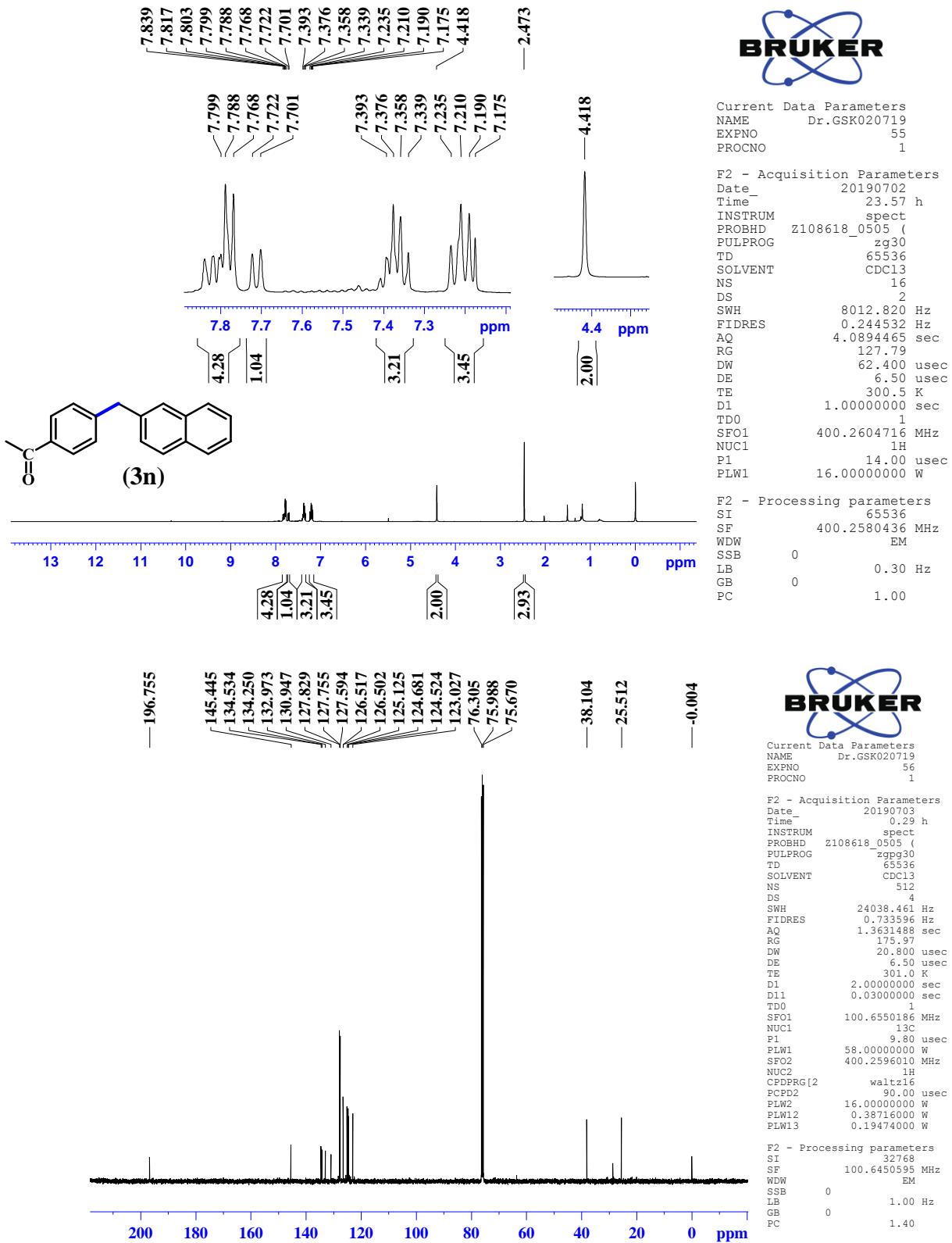














Current Data Parameters  
 NAME Dr.GSK100219  
 EXPNO 7  
 PROCNO 1

F2 - Acquisition Parameters  
 Date 20190211  
 Time 6.38 h  
 INSTRUM spect  
 PROBHD Z108618\_0505 (   
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 16  
 DS 2  
 SWH 8012.820 Hz  
 FIDRES 0.244532 Hz  
 AQ 4.0894465 sec  
 RG 77.73  
 DW 62.400 usec  
 DE 6.50 usec  
 TE 303.3 K  
 D1 1.0000000 sec  
 TD0 1  
 SFO1 400.2604716 MHz  
 NUC1 1H  
 P1 14.00 usec  
 PLW1 16.0000000 W

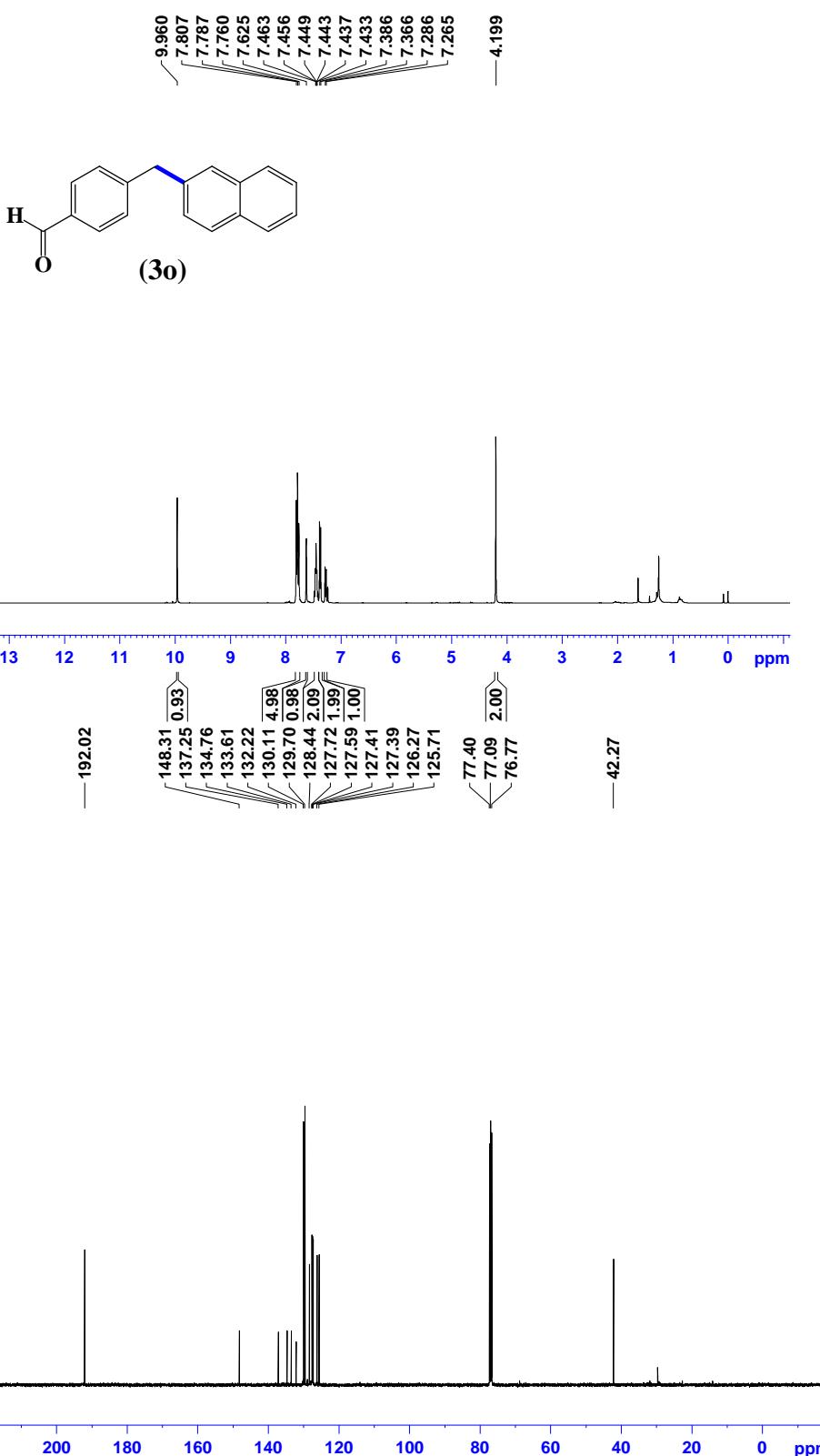
F2 - Processing parameters  
 SI 65536  
 SF 400.2580192 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

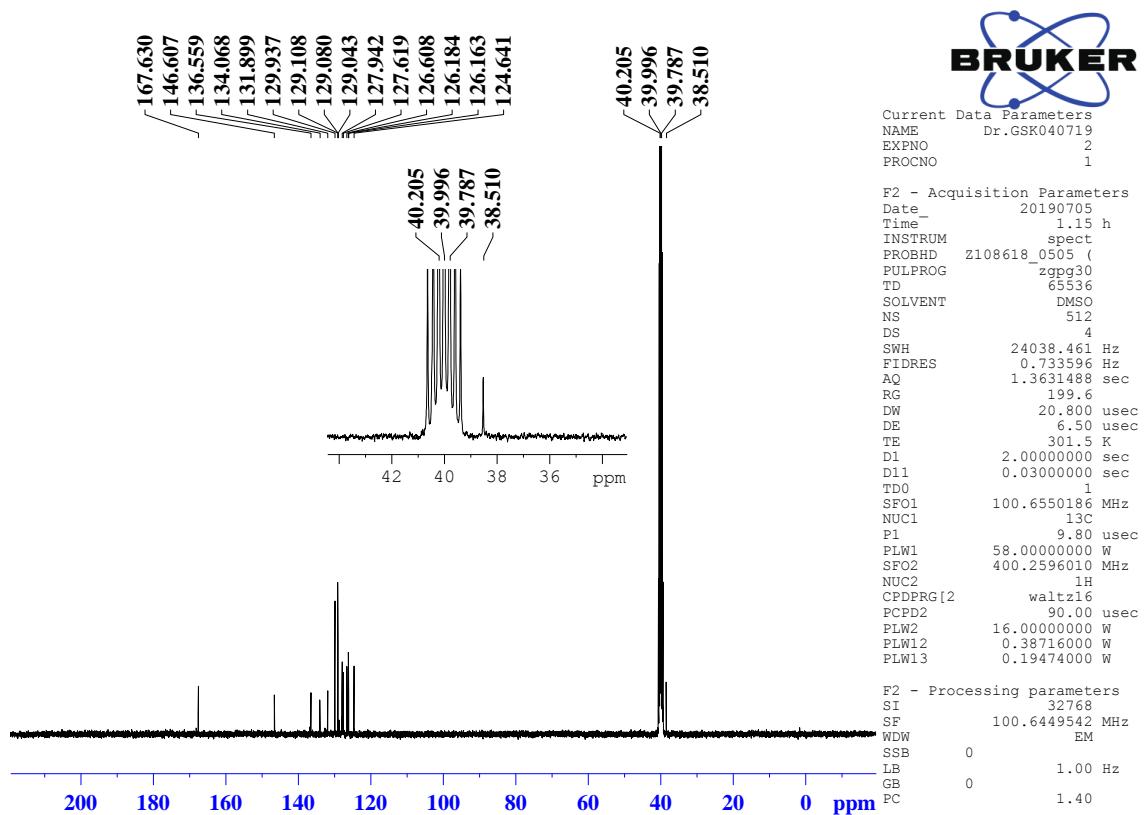
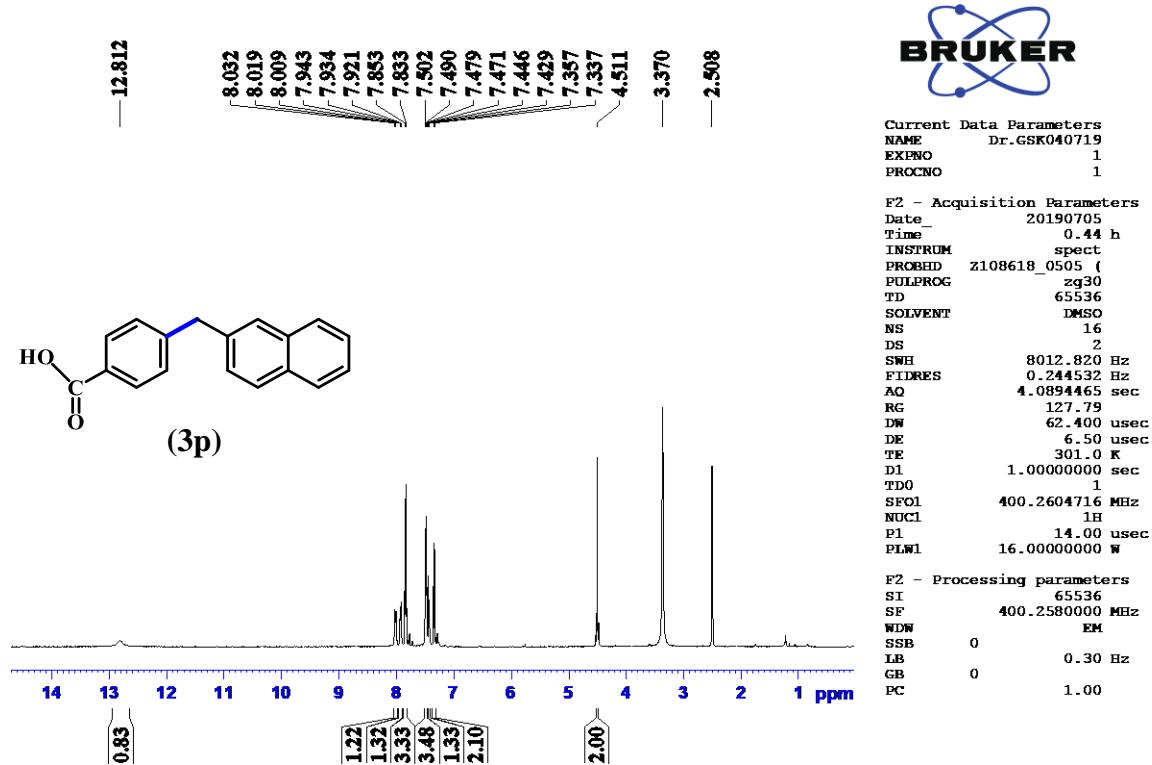


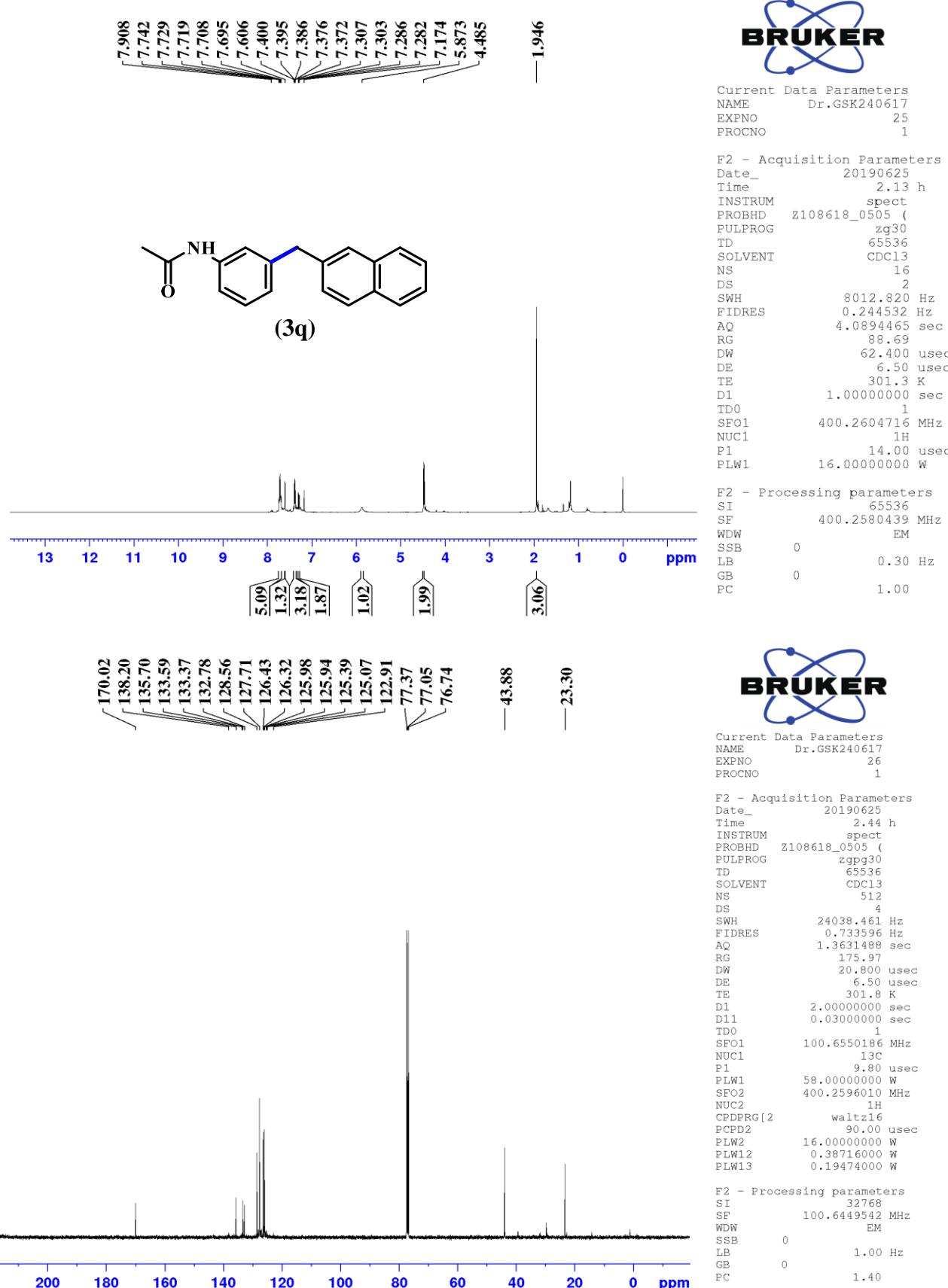
Current Data Parameters  
 NAME Dr.GSK100219  
 EXPNO 8  
 PROCNO 1

F2 - Acquisition Parameters  
 Date 20190211  
 Time 7.08 h  
 INSTRUM spect  
 PROBHD Z108618\_0505 (   
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 512  
 DS 4  
 SWH 24038.461 Hz  
 FIDRES 0.733596 Hz  
 AQ 1.3631488 sec  
 RG 112.69  
 DW 20.800 usec  
 DE 6.50 usec  
 TE 303.8 K  
 D1 2.0000000 sec  
 D11 0.0300000 sec  
 TD0 1  
 SFO1 100.6550186 MHz  
 NUC1 13C  
 P1 9.80 usec  
 PLW1 58.00000000 W  
 SFO2 400.2596010 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 90.00 usec  
 PLW2 16.00000000 W  
 PLW12 0.38716000 W  
 PLW13 0.19474000 W

F2 - Processing parameters  
 SI 32768  
 SF 100.6449542 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40







Current Data Parameters

NAME	Dr.GSK240617
EXPNO	25
PROCNO	1

F2 - Acquisition Parameters

Date_	20190625
Time	2.13 h
INSTRUM	spect
PROBHD	Z108618_0505 (
PULPROG	zg30
TD	65536
SOLVENT	CDCl <sub>3</sub>
NS	16
DS	2
SWH	8012.820 Hz
FIDRES	0.244532 Hz
AQ	4.0894465 sec
RG	88.69
DW	62.400 usec
DE	6.50 usec
TE	301.3 K
D1	1.00000000 sec
TD0	1
SFO1	400.2604716 MHz
NUC1	1H
P1	14.00 usec
PLW1	16.00000000 W

F2 - Processing parameters

SI	65536
SF	400.2580439 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	1.00



Current Data Parameters

NAME	Dr.GSK240617
EXPNO	26
PROCNO	1

F2 - Acquisition Parameters

Date_	20190625
Time	2.44 h
INSTRUM	spect
PROBHD	Z108618_0505 (
PULPROG	zg30
TD	65536
SOLVENT	CDCl <sub>3</sub>
NS	512
DS	4
SWH	24038.461 Hz
FIDRES	0.733596 Hz
AQ	1.3631488 sec
RG	175.97
DW	20.800 usec
DE	6.50 usec
TE	301.8 K
D1	2.00000000 sec
D11	0.03000000 sec
TD0	1
SFO1	100.6550186 MHz
NUC1	<sup>13</sup> C
P1	9.80 usec
PLW1	58.00000000 W
SFO2	400.2596010 MHz
NUC2	1H
CPDPG[2]	waltz16
PCPD2	90.00 usec
PLW2	16.00000000 W
PLW12	0.38716000 W
PLW13	0.19474000 W

F2 - Processing parameters

SI	32768
SF	100.6449542 MHz
WDW	EM
SSB	0
LB	1.00 Hz
GB	0
PC	1.40



(3r)



Current Data Parameters  
NAME Dr.GSK140818  
EXPNO 5  
PROCNO 1

F2 - Acquisition Parameters  
Date 20180817  
Time 14.43 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 16  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 156.91  
DW 62.400 usec  
DE 6.50 usec  
TE 296.9 K  
D1 1.0000000 sec  
TD0 1  
SFO1 400.2604716 MHz  
NUC1 1H  
P1 14.27 usec  
PLW1 15.0000000 W

F2 - Processing parameters  
SI 65536  
SF 400.2580428 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME Dr.GSK140818  
EXPNO 6  
PROCNO 1

F2 - Acquisition Parameters  
Date 20180817  
Time 15.13 h  
INSTRUM spect  
PROBHD Z108618\_0505 (   
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 512  
DS 4  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 143.73  
DW 20.800 usec  
DE 6.50 usec  
TE 297.6 K  
D1 2.0000000 sec  
D11 0.0300000 sec  
TD0 1  
SFO1 100.6550186 MHz  
NUC1 13C  
P1 9.60 usec  
PLW1 58.00000000 W  
SFO2 400.2596010 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 90.00 usec  
PLW2 15.00000000 W  
PLW12 0.37709999 W  
PLW13 0.18968000 W

F2 - Processing parameters  
SI 32768  
SF 100.6449542 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

