

-Supporting Information-

Probing batch and continuous flow reactions in dry solvents: *Granulicella tundricula* Hydroxynitrile lyase (GtHNL)

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A. Gene and amino-acid sequences

GtHNL wild type – Gene sequence

5'CCATGGAGATTAAACGTGTTGGTCTCAGGCTCTGGTAAAGGTCCGGCTGATT
GGTCACTGGTACTGTTCGTATCGATCCGCTGTTCAAGGCTCCGGATCCGGCATTA
GTAGCTGGTCTGTTACCTTGAAACCGGGTGCTCGTACTGCTGGCATACTCA
TCCGTTAGGTCAAGACTCTGATTGTAAGTGCCTGGTTGTGGTGGCTCAGCGTGAA
GGTGGTGCTGTTGAAGAAATTCCGGGTGATGTTGTATGGTCTCTCCAGGTG
AAAAACACTGGCATGGTGCACCAACTACCGCTATGACCCACCTGGCTATCCA
GGAACGTCTGGATGGTAAAGCTGTTGATTGGATGGAACACGTTACTGATGAACAG
TACCGTCGTTAAGCTT -3'

GtHNL wild type – Aminoacid sequence

M E I K R V G S Q A S G K G P A D W F T G T V R I D P L F Q A P D P A L V A G A S
V T F E P G A R T A W H T H P L G Q T L I V T A G C G W A Q R E G G A V E E I H P
G D V V W F S P G E K H W H G A A P T T A M T H L A I Q E R L D G K A V D W M
E H V T D E Q Y R R A

GtHNL-A40H/V42T/Q110H

5'CCATGGAGATTAAACGTGTTGGTCTCAGGCTCTGGTAAAGGTCCGGCTGATT
GGTCACTGGTACTGTTCGTATCGATCCGCTGTTCAAGGCTCCGGATCCGGCATTA
GTAGCTGGTCACTCTACTACCTTGAAACCGGGTGCTCGTACTGCTGGCATACTCA
TCCGTTAGGTCAAGACTCTGATTGTAAGTGCCTGGTTGTGGTGGCTCAGCGTGAA
GGTGGTGCTGTTGAAGAAATTCCGGGTGATGTTGTATGGTCTCTCCAGGTG
AAAAACACTGGCATGGTGCACCAACTACCGCTATGACCCACCTGGCTATCCA
CGAACGTCTGGATGGTAAAGCTGTTGATTGGATGGAACACGTTACTGATGAACAG
TACCGTCGTTAAGCTT -3'

GtHNL-A40H/V42T/Q110H – Aminoacid sequence

M E I K R V G S Q A S G K G P A D W F T G T V R I D P L F Q A P D P A L V A G H S
T F E P G A R T A W H T H P L G Q T L I V T A G C G W A Q R E G G A V E E I H P
G D V V W F S P G E K H W H G A A P T T A M T H L A I H E R L D G K A V D W M E
H V T D E Q Y R R A

Aminoacids in red show the mutations at positions 40, 42 and 110.

B. SDS-PAGE purified *GtHNL-A40H/V42T/Q110H*

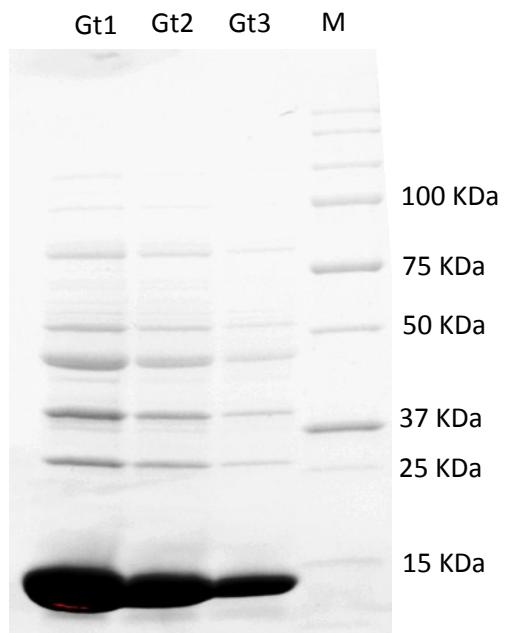


Figure S1. SDS PAGE of purified GtHNLA40H/V42T/Q110H. Sample M is the marker and G1 to Gt3 are different dilutions of purified GtHNLA40H/V42T/Q110H

C. Calculation of reaction volume in continuous flow

Interbead volume (Non porous glass beads)

A commonly used inter-beads volume for non-porous beads of similar or close diameters even for well packed columns is between 30% and 40% of the total volume. Since 150 mg of glass beads correspond to 0.5 mL bulk volume, the inter-beads volume is about:

$$0.5 \text{ mL} \times 35\% = 0.175 \text{ mL}$$

Pore volume (Celite R-633)

According to El-Sayed¹, Celite R-633 has a total pore volume of 1.46 mL g⁻¹.

$$0.150 \text{ g Celite R-633} \times 1.46 \text{ mL g}^{-1} = 0.219 \text{ mL}$$

$$\text{Reaction volume} = 0.219 \text{ mL} + 0.175 \text{ mL} = 0.394 \text{ mL}$$

D. Progress of the synthesis of (*R*)-mandelonitrile with non-immobilized *GtHNL-TV*

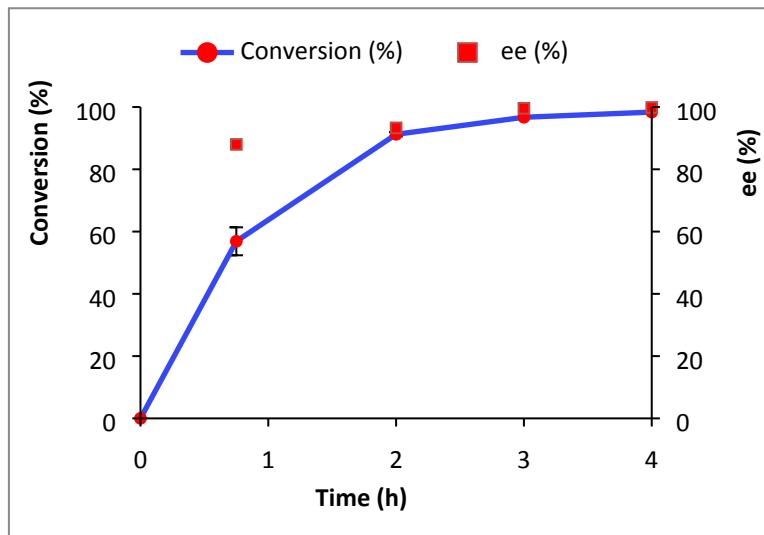


Figure S2: Kinetic trace for the synthesis of (*R*)-mandelonitrile with non-immobilized *GtHNL-TV*. Conditions: Ratio benzaldehyde : HCN in acetate buffered MTBE, pH 4, 1:4, *GtHNL-TV* (11 mg, 50 U). The reaction was stirred at 1000 rpm at 5 °C.

E. Benzaldehyde and HCN kinetics

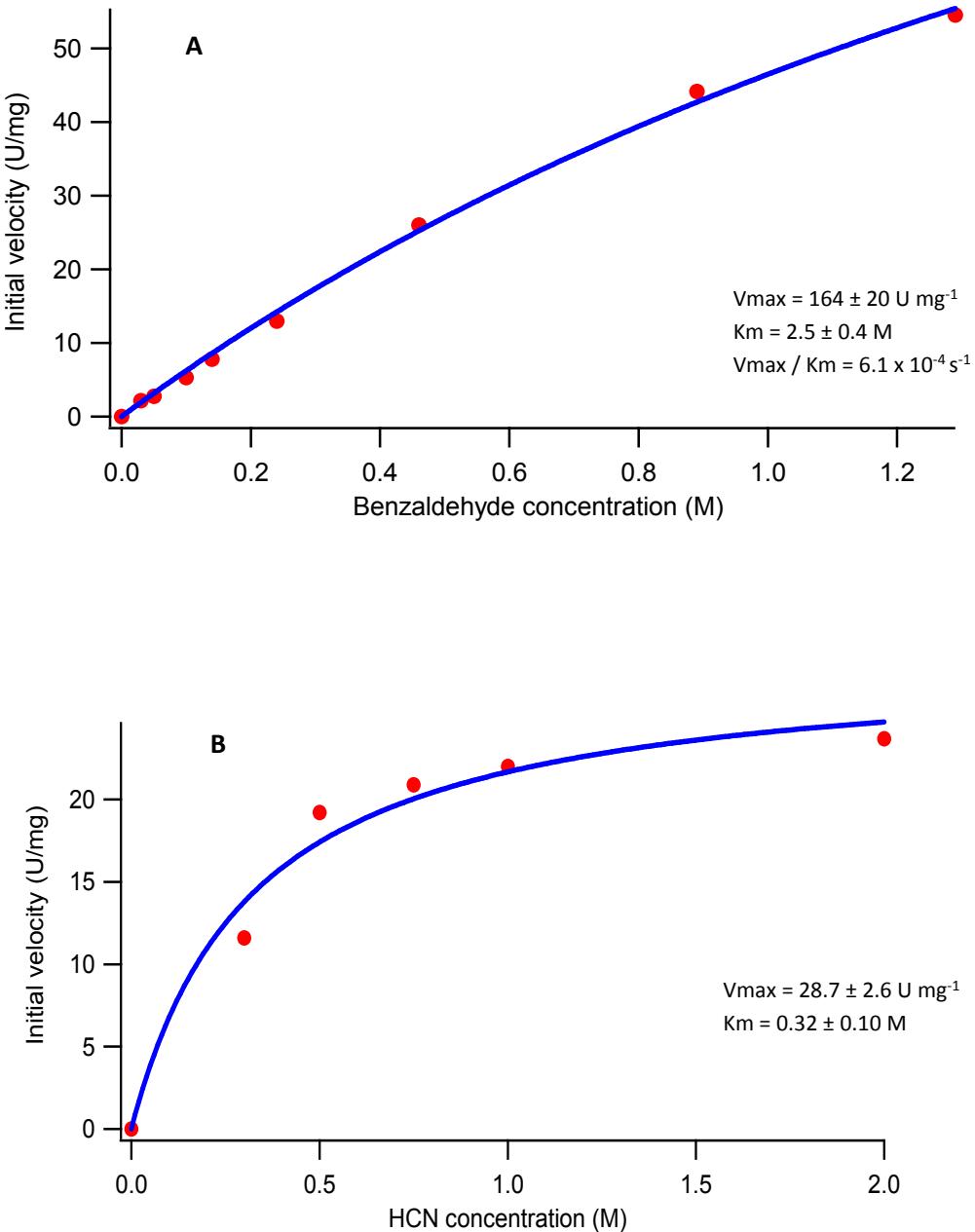


Figure S3. Kinetic traces for the synthesis of (*R*)-mandelonitrile at different benzaldehyde (S3-A) and HCN (S3-B) concentrations. Conditions: Ratio benzaldehyde : HCN in acetate buffered MTBE, pH 4, 1:4, 2 mL HCN in acetate buffered MTBE (1.5–2 M) pH 4, *GtHNL-TM* immobilized on 50 mg Celite R-633 (50 U). The reaction was stirred at 700 rpm, 5 °C. Single measurement points were fitted by using Michaelis-Menten equation. Standard deviation was calculated with Igor Pro 5.0.5 software

F. Substrate incubation for evaluation of background reaction during 8 hours

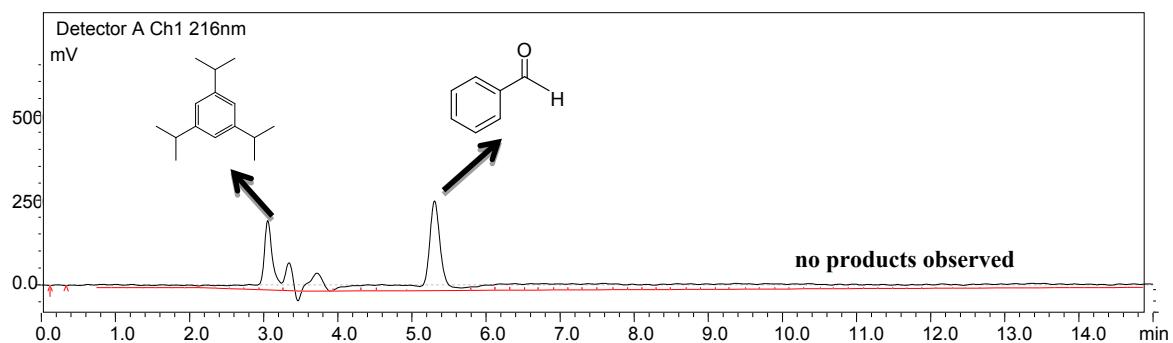


Figure S4. HPLC detection of benzaldehyde and 1,3,5 tri-isopropylbenzene during 8 hours of incubation:
Conditions: Ratio benzaldehyde : HCN in buffered MTBE, pH 4, 1:4, 100 μ L benzaldehyde (1 mmol), 2 mL HCN in acetate buffered MTBE, pH 4. The reaction was stirred at 1000 rpm at 5 °C.

G. Identification of substrates and products during the synthesis of (*R*)-mandelonitrile

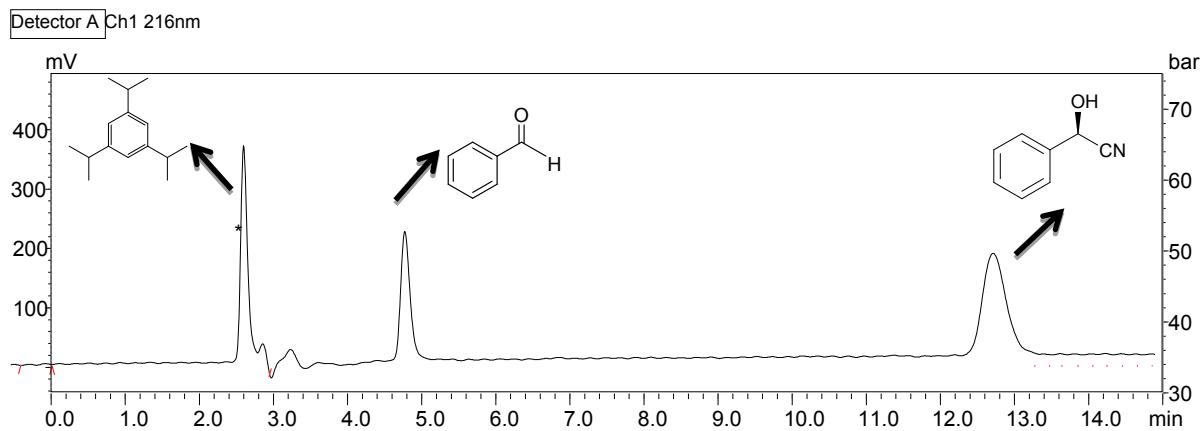


Figure S5. HPLC detection of benzaldehyde, 1,3,5 tri-isopropylbenzene and (*R*)-mandelonitrile. Conditions: Ratio benzaldehyde : HCN in buffered MTBE, pH 4, 1:4, a CFR with *GtHNL-TV* immobilised on 150 mg Celite R-633 (150 U). Reactions was performed at room temperature.

H. Progress of the synthesis of (*R*)-mandelonitrile with immobilized *GtHNL-TV* in nylon and regular paper tea bag.

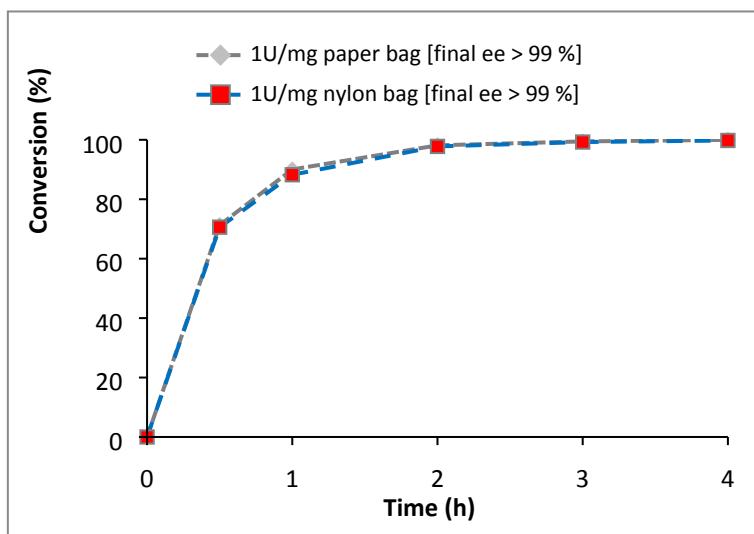


Figure S6: Synthesis of (*R*)-mandelonitrile with immobilized *GtHNL-TV* in nylon and regular paper tea bags. Paper tea bags are commonly nylon enforced. Conditions: Ratio benzaldehyde : HCN in acetate buffered MTBE, pH 4, 1:4, *GtHNL-TV* (11 mg, 50 U). The reaction was stirred at 1000 rpm at 5 °C.

I. Size of tea bags for BR and RBR reactions

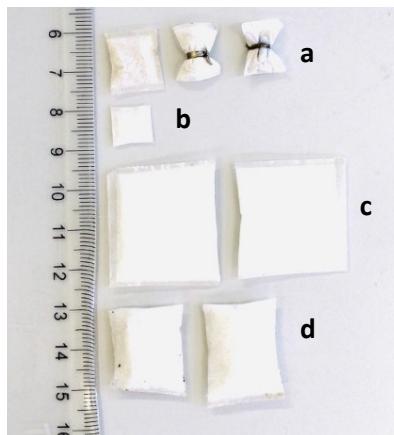


Figure S7: Tea bags with different sizes for tightly or loosely packed *GtHNL-TV* on Celite R-633. The ruler shows the size in cm. a) tightly packed Celite immobilised *GtHNL-TV* (50 mg) for BR, b) tightly packed Celite immobilised *GtHNL-TV* (18 mg) for BR, c) loosely packed Celite immobilised *GtHNL-TV* (773 mg) for RBR and d) tightly packed Celite immobilised *GtHNL-TV* (773 mg) for RBR.

Table S1: Size of tea bags for tightly or loosely packed *GtHNL-TV* on Celite R-633 for BR and RBR reactions.

Reactor	Type of packing	Size	Amount of enzyme -carrier
BR	Tight	1.3 x 1.6 cm	50 mg
BR	Tight	0.7 x 0.8	18 mg
RBR ¹	Loose	2.5 x 2.6 cm	773 mg
RBR ¹	Tight	1.8 x 2.2 cm	773 mg

¹ 2 bags with equal amount of *GtHNL-TV* on Celite.

SI References

- [1] A.H.M.M. El-Sayed, W.M. Mahmoud and R.W. Coughlin, *Biotechnol. Bioeng.*, **1990**, *36*, 83-91.