

Supporting Information

One-pot, Chemoselective Synthesis of Secondary Amines from Arylnitriles Using PdPt–Fe₃O₄ Nanoparticle Catalyst

Jin Hee Cho,[†] Sangmoon Byun,[‡] Ahra Cho,[†] and B. Moon Kim^{*†}

[†]Department of Chemistry, College of Natural Sciences, Seoul National University,
1 Gwanak-ro, Gwanak-gu, Seoul 08826, Republic of Korea

[‡]The Research Institute of Basic Sciences, Seoul National University, 1 Gwanak-ro,
Gwanak-gu, Seoul, 08826, Republic of Korea

Corresponding author's email address: kimbm@snu.ac.kr

Table of Contents

	page
1. Materials/Instrumentation.....	S3
2. Catalyst Characterization	
- Figure S1. HR-TEM analysis.....	S5
- Figure S2. SEM analysis.....	S6
- Figure S3. SEM-EDS analysis.....	S7
- Figure S4. HADDF & BF Cs-STEM analysis.....	S8
- Figure S5. STEM-EDS analysis of PdPt-Fe ₃ O ₄	S9
- Figure S6. The particle distribution of PdPt-Fe ₃ O ₄	S10
- Figure S7. ICP-AES data	S11
- Figure S8. XPS data.....	S12
- Figure S9. EDS line scan profile of a single bimetallic PdPt nanoparticle.....	S13
3. Supplementary figures and tables	
- Table S1. Optimization of condition of the hydrogenation of benzonitrile.....	S14
- Table S2. Comparison of the catalytic activity with other catalyst (symmetrical amine).....	S14
- Table S3. Comparison of yields with different nitrile and nitro compound stoichiometries....	S15
- Figure S10. SEM image of PdPt NPs with different supports.....	S16
- Figure S11. ICP-AES data of PdPt NPs with different supports.....	S16
- Figure S12. The particle size distribution with different supports.....	S17
- Figure S13. Kinetic data.....	S18
- Figure S14. SEM image of Pd _x Pd _y -Fe ₃ O ₄ NPs.....	S19
- Table S4. Comparison catalytic activity of Pd _x Pd _y -Fe ₃ O ₄ NPs.....	S19
- Table S5. Additive Screening.....	S20
- Table S6. Effect of HFIP as a co-solvent.....	S20
- Table S7. Comparison of yields with different nitrile and amine compound stoichiometries...	S21
- Table S8. Comparison of the catalytic activity with other catalyst (unsymmetrical amine).....	S21
- Figure S15. Recyclability of PdPt-Fe ₃ O ₄ NPs.....	S22
- Figure S16. HR-TEM image of used NPs.....	S23
4. NMR spectra.....	S24
5. References.....	S48

1. Materials/Instrumentation

ESCA (Electron Spectroscopy for Chemical Analysis)

1. Model: SIGMA PROBE (ThermoVG, U.K.)
2. Condition

X-ray source		Monochromatic <i>Al-K</i> (15 kV, 100W, 400 micrometer)
Wide scan	pass energy	50 eV
	step size	1.0 eV
Narrow scan	pass energy	20 eV
	step size	0.1 eV
Flood gun		off
Ion etching gun		off

3. Vacuum: 7×10^{-9} mB (x-ray on, flood gun off)
4. Calibration: C 1s (284.5 eV)
5. Program: Avantage (ThermoVG)

Transmission Electron Microscope II (ccd camera type)

1. Model: JEM-2100
2. Accelerating Voltage: 80 to 200 kV
3. Gatan Digital Camera (ORIUS-SC600)
4. Resolution: Point image: 0.23 nm
Lattice image: 0.14 nm
5. MAG: x50 ~ x1 500000
6. Camera Length: SA DIFF Mode: 80 ~ 2 000 mm

Cs-TEM (Cs corrected TEM with Cold FEG)

1. Model: JEM-ARM200F (Cold Field Emission Type)
2. Specifications:
 - a. HT: 60, 80, 120, 200 kV
 - b. Magnification: 50 to 2,000,000 X (TEM), 200 to 1,500,000 X (STEM)
 - c. Resolution
 - TEM mode: Lattice 0.07 nm/ Point 0.11 nm
 - STEM mode: 0.136 nm
 - d. Sample tilting
 - X / Y: $\pm 25^\circ$ / $\pm 25^\circ$
3. Analysis functions:
 - a. CCD Camera: UltraScan 1000XP (2,048 x 2,048 pixel)
 - b. EDS: SDD Type (Active area 100 mm²/ Solid angle 0.7 str)

Cs-STEM (Cs corrected STEM with Cold FEG)

1. Model: JEM-ARM200F (Cold Field Emission Type, JEOL)
2. Specifications
 - a. HT: 60, 80, 120, 200 kV
 - b. Magnification: 50 to 2,000,000 X (TEM), 200 to 1,500,000 X (STEM)
 - c. Resolution
 - STEM mode: HAADF 0.1 nm/ BF 0.136 nm
 - TEM mode: Point 0.23 nm
 - d. Sample tilting
 - X / Y: $\pm 35^\circ$ / $\pm 30^\circ$
3. Analysis functions
 - a. CCD Camera: UltraScan 1000XP (2,048 x 2,048 pixel)
 - b. EDS: SDD Type (Active area 100 mm²/ Solid angle 0.9 str)
 - c. EELS: Model 965 GIF Quantum ER

Fe-SEM (Field Emission SEM)

1. Model: JSM-7600F (Fe-SEM)
2. Specification
 - Resolution:
 - 1) 1.0 nm at 15 kV
 - 2) 1.5 nm at 1 kV
 - Voltage range: 0.1 ~ 30 kV
 - Maximum image size: 5,120 x 3,840 pixels
 - Probe current: 1pA ~ 200 nA
 - Tilt: -5 ~ 70°
 - Rotation: 360°
 - Working Distance: 1.5 ~ 25 mm

PdPt-Fe₃O₄ samples were analyzed on Cs-STEM (Cs corrected STEM with Cold FEG), Cs-TEM (Cs corrected TEM with Cold FEG) and High resolution-Transmission Electron Microscope (ccd camera type) installed at the National Center for Inter-university Research Facilities (NCIRF) at Seoul National University.

SEM images of PdPt-Fe₃O₄ were obtained with a JSM-7600F at a voltage of 15 kV installed at Seoul National University Research Institute of Advanced Materials.

2. Catalyst characterization

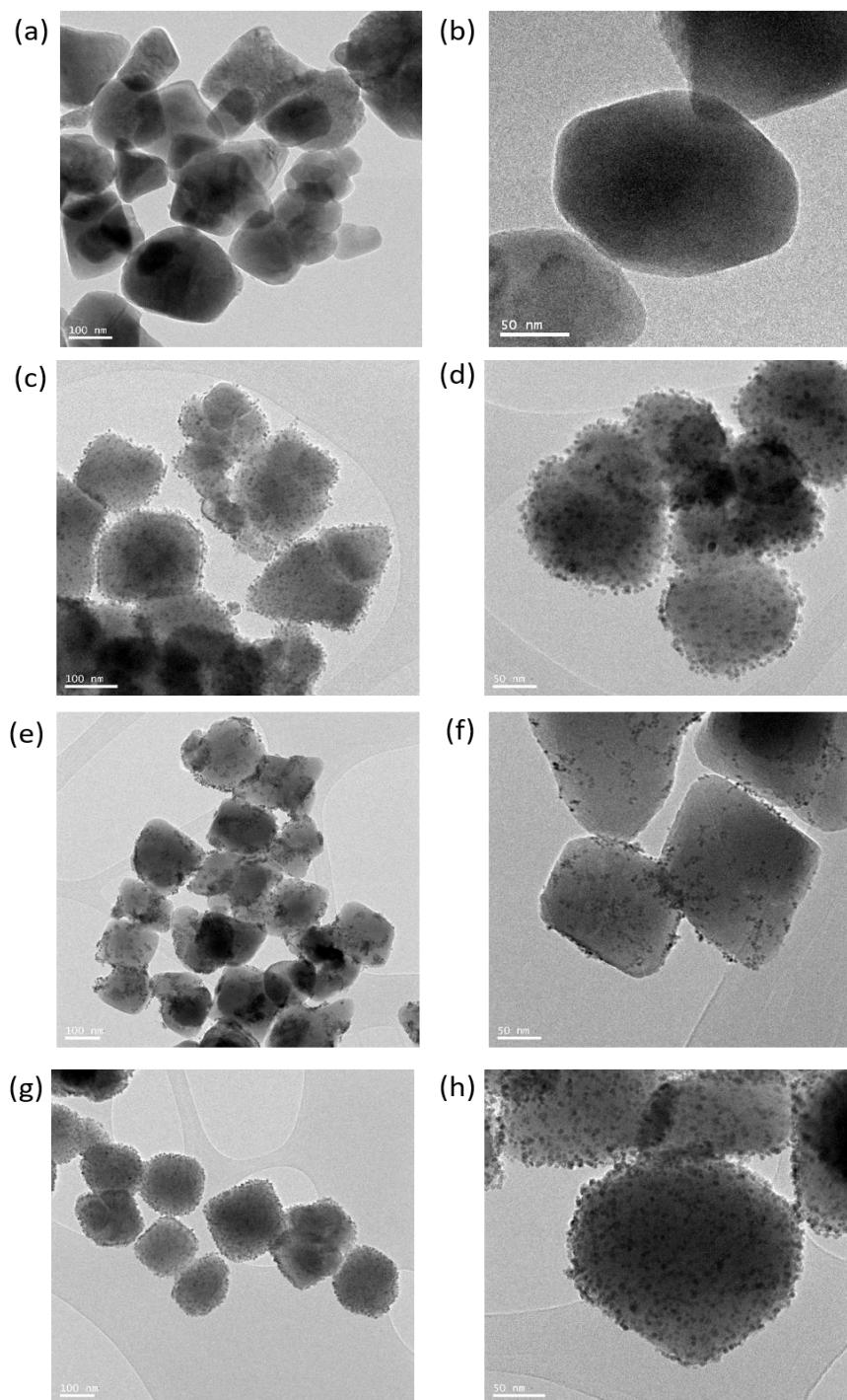


Figure S1. HR-TEM images of NPs. (a) and (b) Fe_3O_4 NPs; (c) and (d) Pd– Fe_3O_4 NPs; (e) and (f) Pt– Fe_3O_4 NPs; (g) and (h) PdPt– Fe_3O_4 NPs.

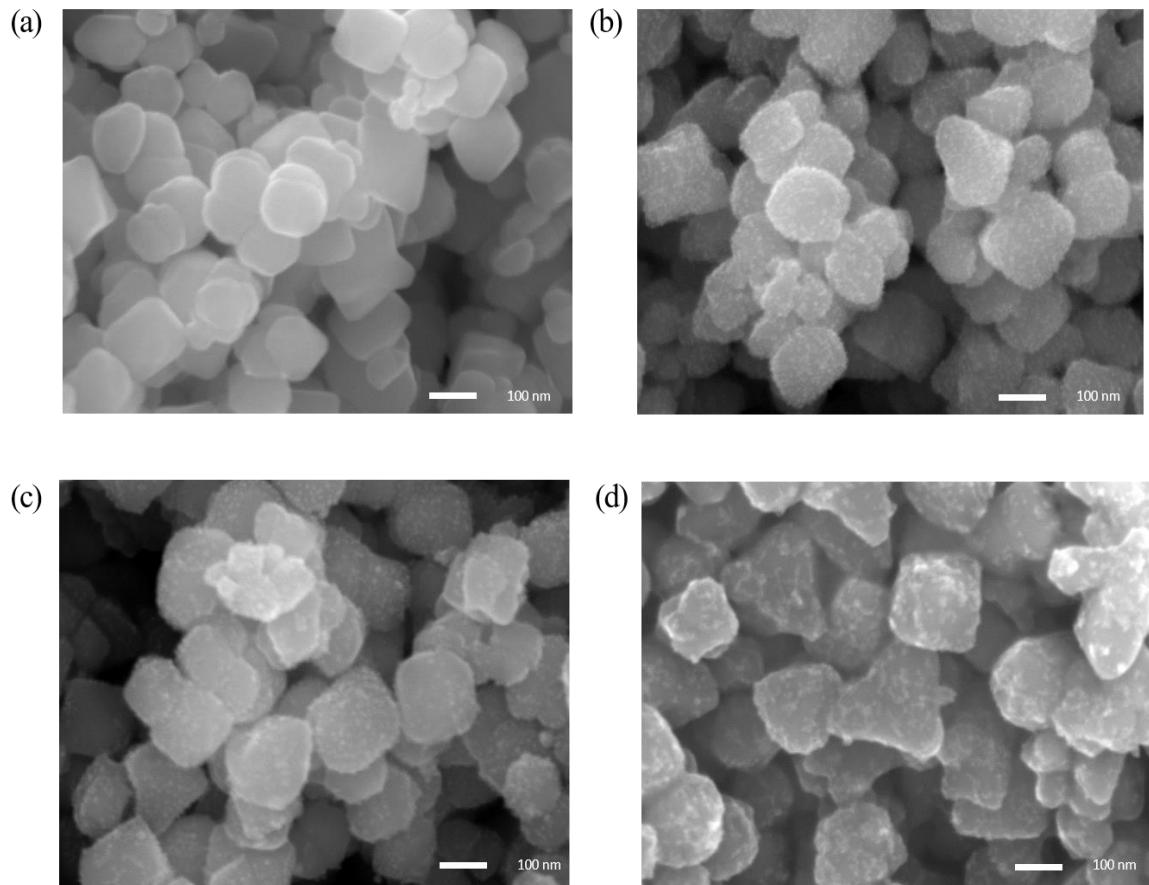


Figure S2. SEM image of (a) Fe₃O₄ NPs; (b) PdPt–Fe₃O₄ NPs; (c) Pd–Fe₃O₄ NPs; (d) Pt–Fe₃O₄ NPs.

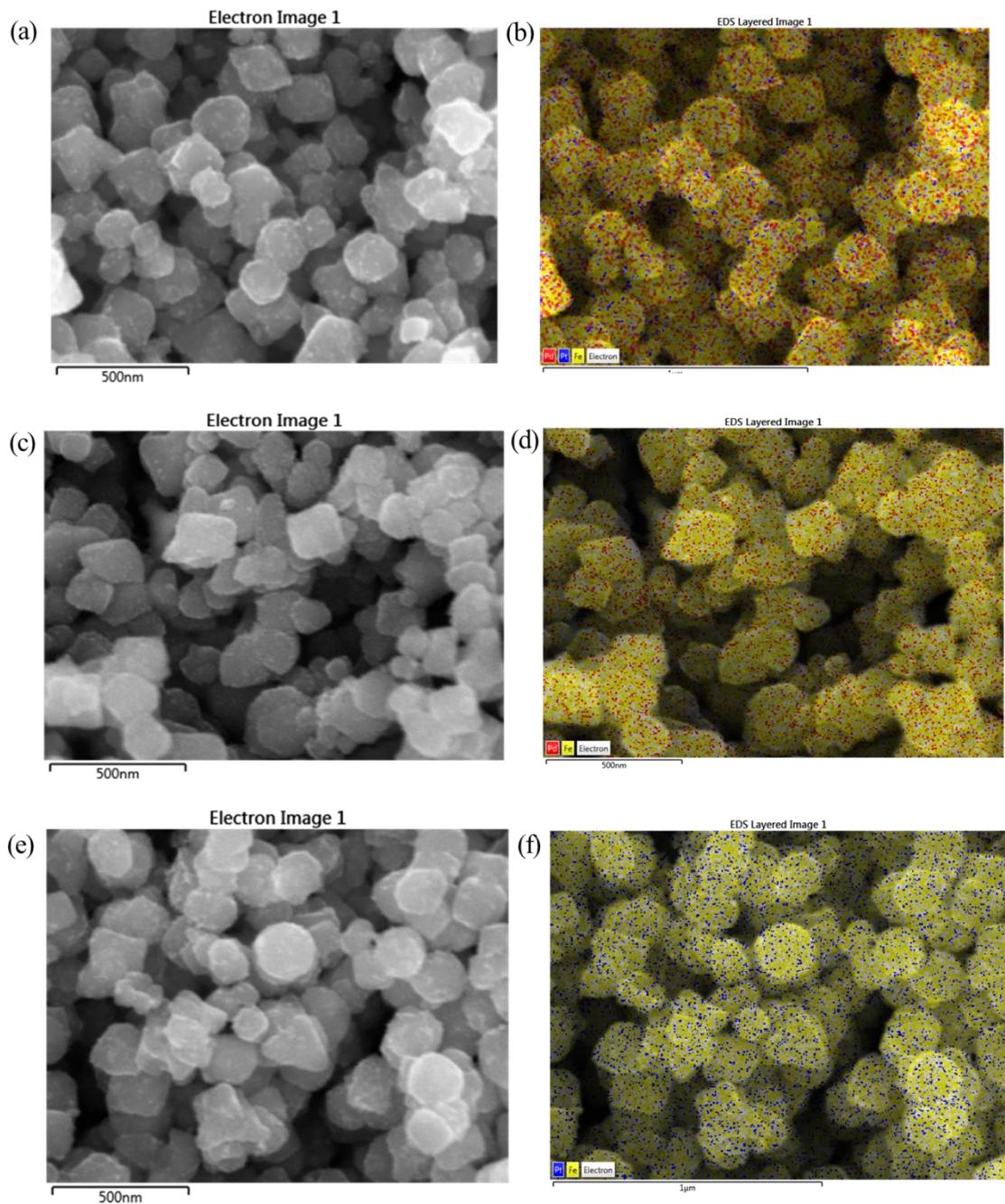


Figure S3. (a),(b) SEM-EDS images of PdPt- Fe_3O_4 NPs; (c),(d) SEM-EDS images of Pd- Fe_3O_4 NPs; (e),(f) SEM-EDS images of Pt- Fe_3O_4 NPs.

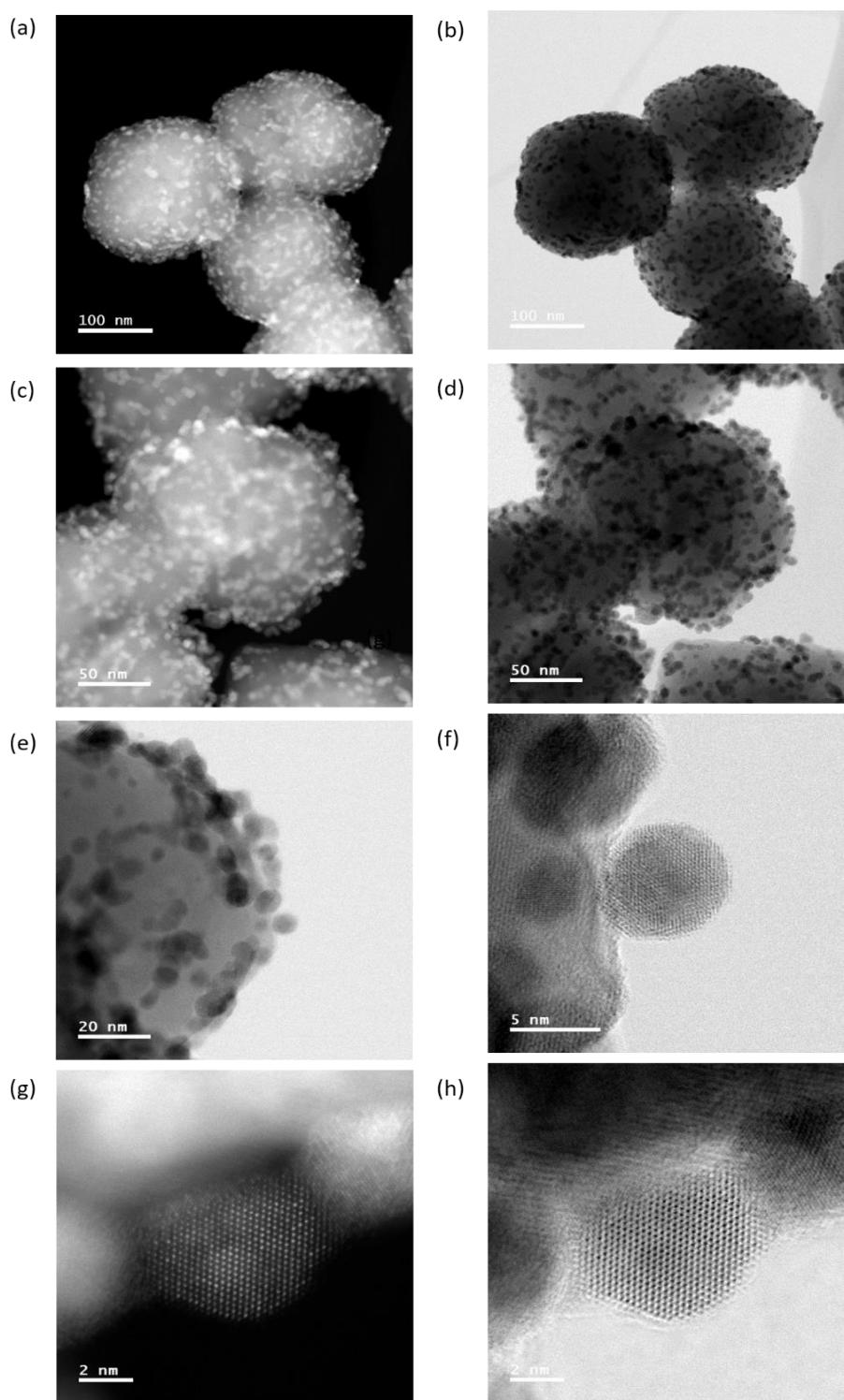


Figure S4. (a), (c), and (g) HADDF-STEM images of PdPt–Fe₃O₄ NPs; (b), (d), (e), (f), and (h) BF-STEM images of PdPt–Fe₃O₄ NPs.

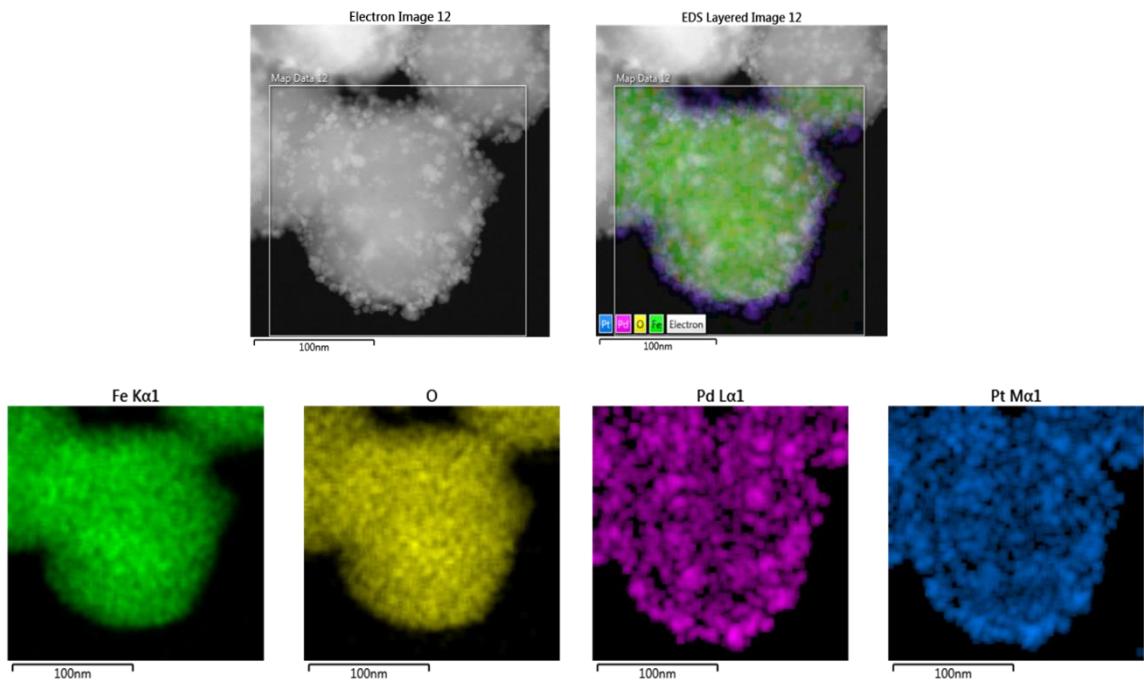


Figure S5. STEM-EDS images of PdPt–Fe₃O₄ NPs.

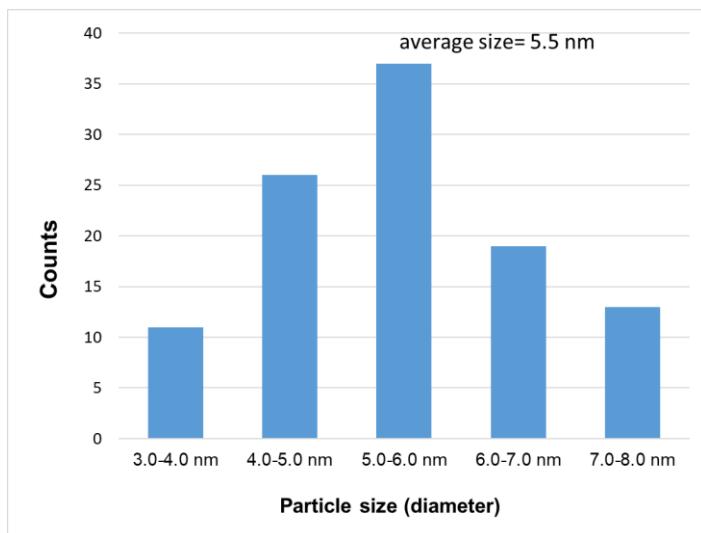
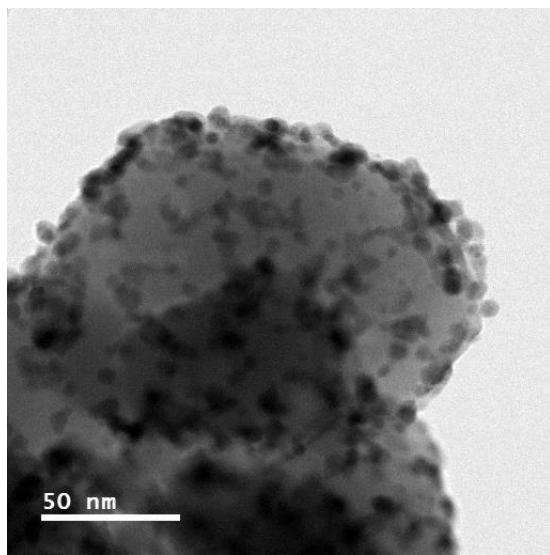


Figure S6. The particle size distribution of PdPt- Fe_3O_4 NPs.

ICP-AES Data		
Catalyst (NPs)	Pd (wt%)	Pt (wt%)
Pd–Fe ₃ O ₄	7.47	-
Pt–Fe ₃ O ₄	-	10.49
Fresh PdPt–Fe ₃ O ₄	3.94	6.06
PdPt–Fe ₃ O ₄ after 1 cycle reaction	3.87	5.91
PdPt–Fe ₃ O ₄ after 4 cycle reaction	3.86	5.91

Figure S7. ICP-AES data of PdPt–Fe₃O₄ NPs, Pd–Fe₃O₄ and Pt–Fe₃O₄ NPs.

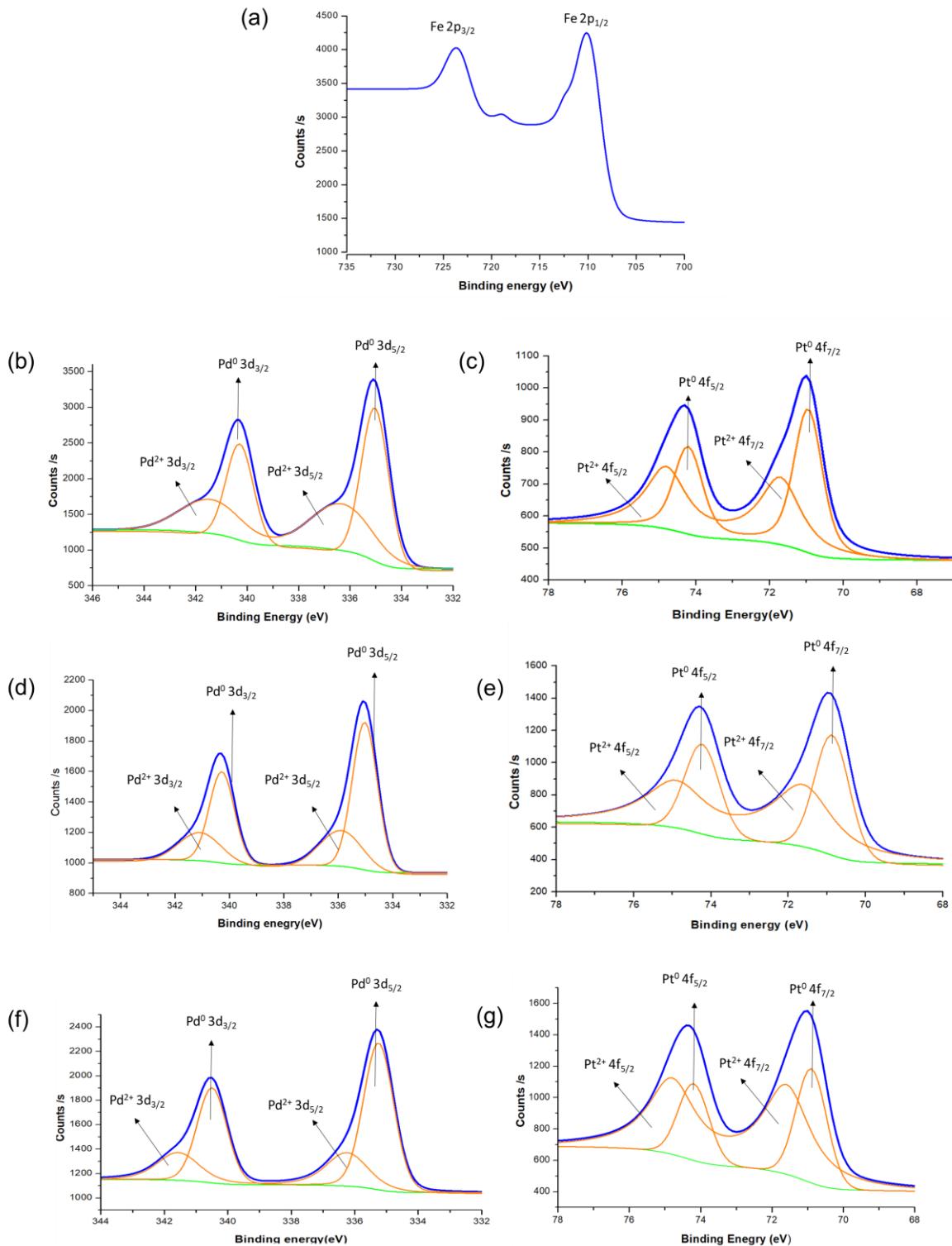


Figure S8. XPS data. (a) Fe 2p spectra of PdPt-Fe₃O₄; (b) Pd 3d spectra of Pd-Fe₃O₄; (c) Pt 4f spectra of Pt-Fe₃O₄; (d) Pd 3d spectra of fresh PdPt-Fe₃O₄; (e) Pt 4f spectra of fresh PdPt-Fe₃O₄; (f) Pd 3d spectra of PdPt-Fe₃O₄ after 1 cycle of the catalytic reaction; (g) Pt 3d spectra of PdPt-Fe₃O₄ after 1 cycle of the catalytic reaction

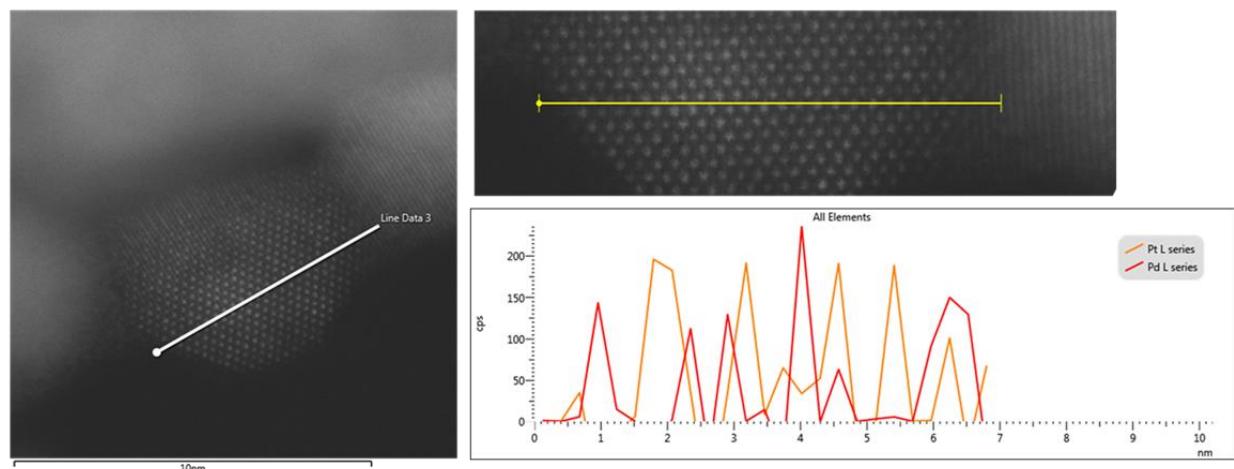


Figure S9. EDS line scan profile of a single bimetallic PdPt nanoparticle

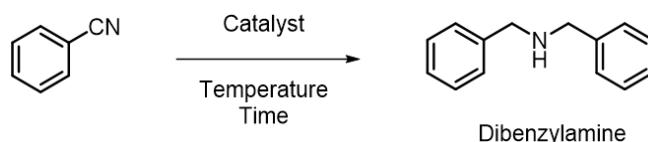
3. Supplementary information of tables and figures.

Table S1. Optimization of the reaction condition of the hydrogenation of benzonitrile^a

Entry	Catalyst	Reductant	Solvent	Conversion (%)	Selectivity ^b (%)			
					a	b	c	d
1	4 mol% Pd–Fe ₃ O ₄	NaBH ₄	MeOH	60	0	0	60	0
2	4 mol% Pt–Fe ₃ O ₄	NaBH ₄	MeOH	54	0	54	0	0
3	2 mol% PdPt–Fe ₃ O ₄	NH ₃ BH ₃	MeOH	38	0	37	1	0
4	2 mol% PdPt–Fe ₃ O ₄	NaBH ₄	MeOH	100	1	1	98	0
5	2 mol% PdPt–Fe ₃ O ₄	NaBH ₄	EtOH	81	75	5	1	0
6	2 mol% PdPt–Fe ₃ O ₄	NaBH ₄	i-PrOH	NR	-	-	-	-
7	2 mol% PdPt–Fe ₃ O ₄	NaBH ₄	Toluene	NR	-	-	-	-
8	2 mol% PdPt–Fe ₃ O ₄	NaBH ₄	H ₂ O	NR	-	-	-	-
9	2 mol% PdPt–Fe ₃ O ₄	H ₂	MeOH	NR	-	-	-	-
10	2 mol% PdPt–Fe ₃ O ₄	PhMe ₂ SiH	MeOH	NR	-	-	-	-

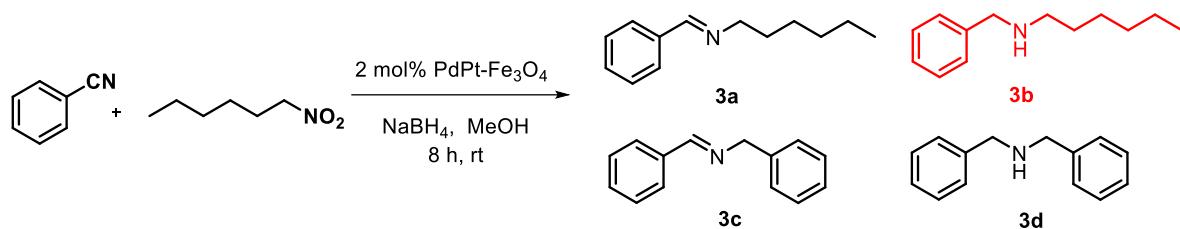
^a Reaction condition: 0.3 mmol benzonitrile, 0.6 mmol NaBH₄, 1.8 mL MeOH, 8 h, room temperature. ^b GC yield.

Table S2. Comparison of the catalytic activity of PdPt–Fe₃O₄ with other reported heterogeneous catalytic system for formation of dibenzylamine from benzonitrile



Entry	Catalyst	Reductant	Temperature	Time	Yield (%)
1 ^[1]	Rh/AlO ₃	H ₂ (1 bar)	RT	20 h	85
2 ^[2]	Pt nanowire	H ₂ (4 bar)	80 °C	24 h	99
3 ^[3]	Pd–Fe _{0.25} Cu _{0.25} /Fe ₃ O ₄	NH ₃ BH ₃	40 °C	1.5 h	98
4 ^[4]	Pd@mpg-C ₃ N ₄	H ₂ (10 bar)	RT	6 h	89
5 ^[5]	PtMo WNWs	H ₂ (1 bar)	40 °C	24 h	91
6 ^[6]	5NiO–Pd/SiC	H ₂ (1 bar)	30 °C	5 h	98
7	PdPt–Fe ₃ O ₄	NaBH ₄	RT	8 h	98

Table S3. Comparison of yields with different nitrile and nitro compound stoichiometries^a



Entry	benzonitrile (equivalent)	1-nitrohexane (equivalent)	Yield ^b			
			3a (%)	3b (%)	3c (%)	3d (%)
1	1.5	1	0	86	0	64
2	1.2	1	0	73	5	42
3	1	1	4	62	1	27
4	1	1.2	8	72	0	20
5	1	1.5	45	50	1	4

^a Reaction condition: 2.0 mol% PdPt-Fe₃O₄, 0.3 mmol scale, 0.6 mmol NaBH₄, 1.8 mL MeOH, 8 h, room temperature. ^b GC yield.

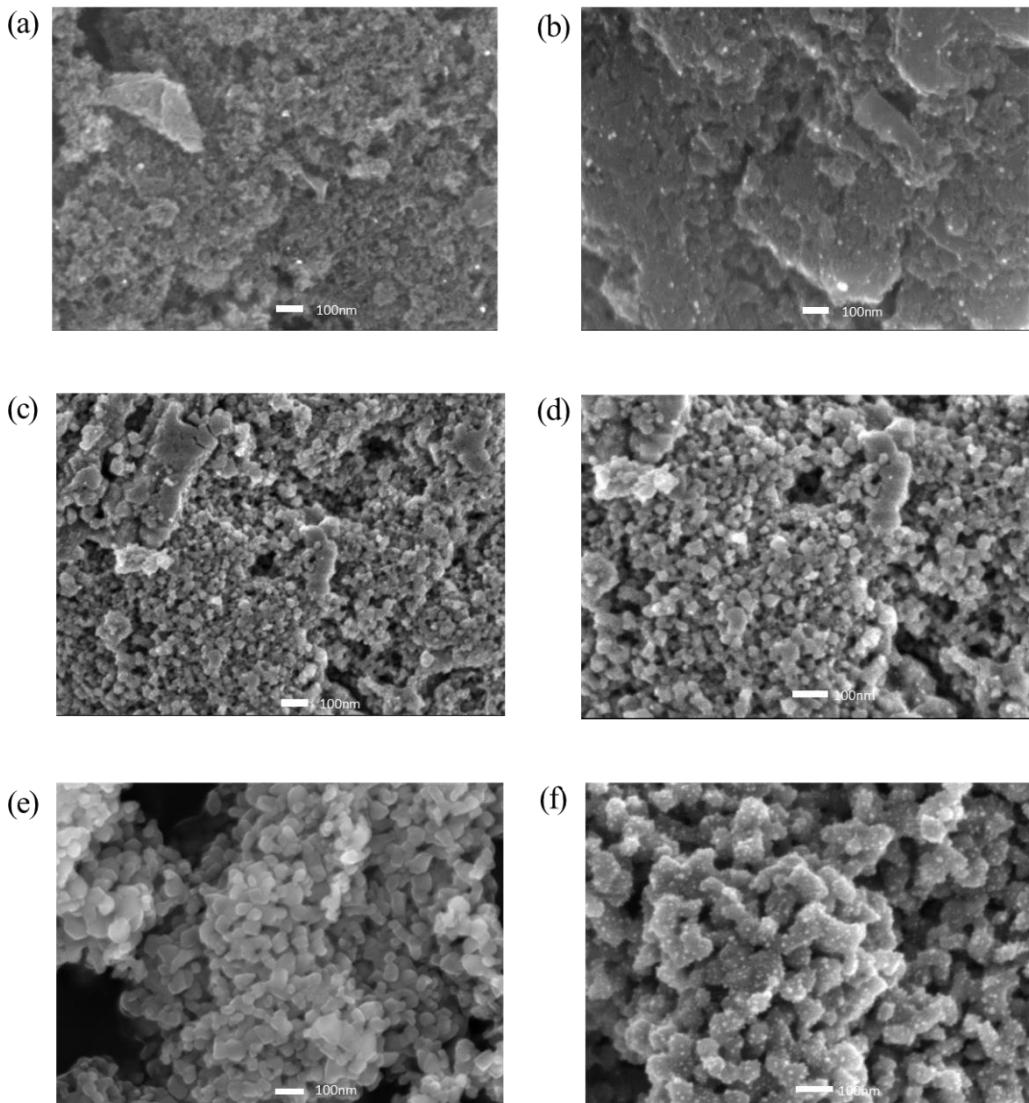


Figure S10. SEM images (a) C NPs; (b) PdPt–C NPs; (c) CeO₂ NPs; (d) PdPt–CeO₂ NPs; (e)TiO₂ NPs; (f) PdPt–TiO₂ NPs.

Catalyst	Pd (wt%)	Pt (wt%)	Pd : Pt
PdPt-C	0.94	0.84	2.05 : 1
PdPt-CeO ₂	2.88	4.78	1.13 : 1
PdPt-TiO ₂	1.04	1.58	1.21 : 1

Figure S11. ICP-AES data of PdPt NPs with different supports.

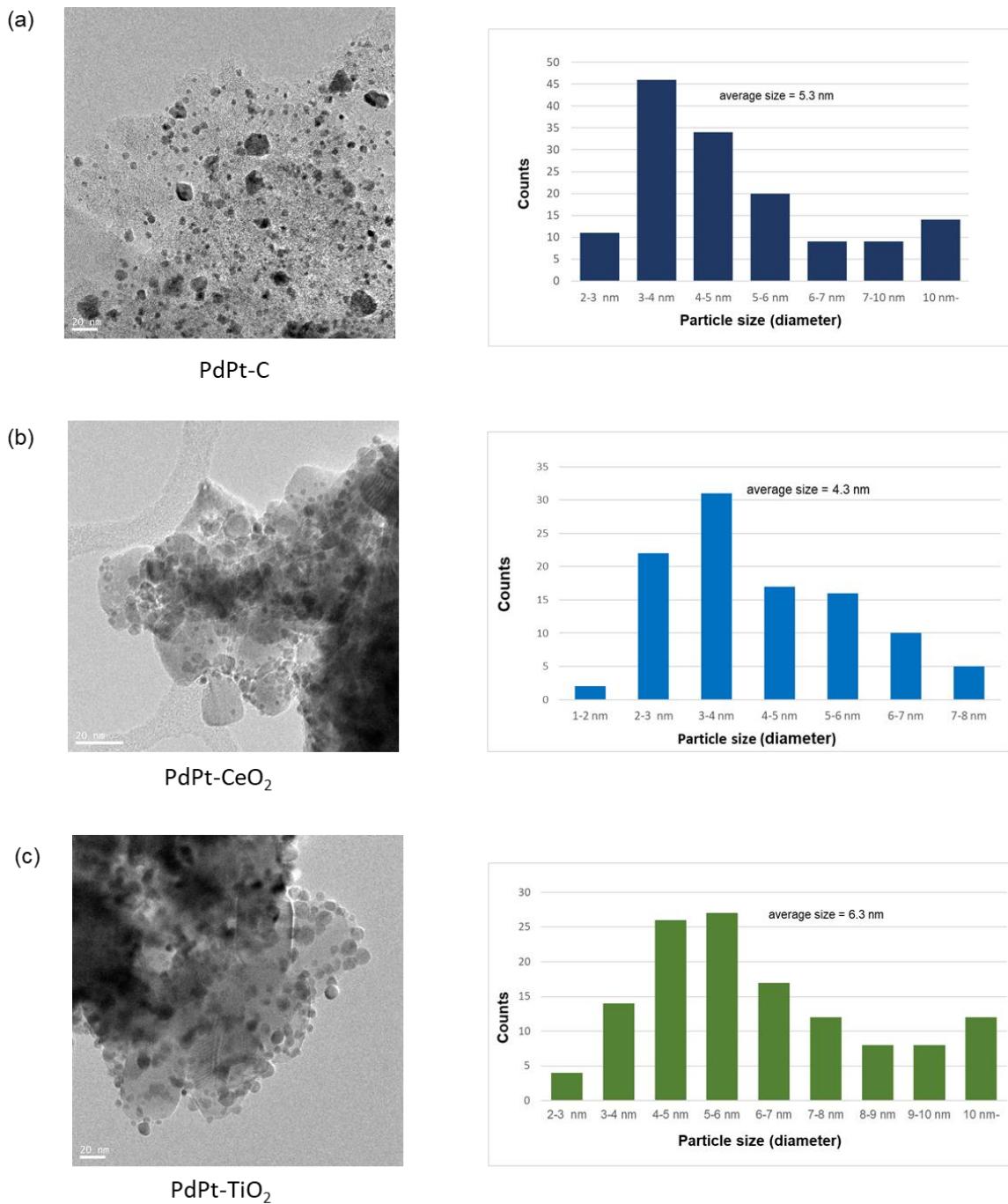
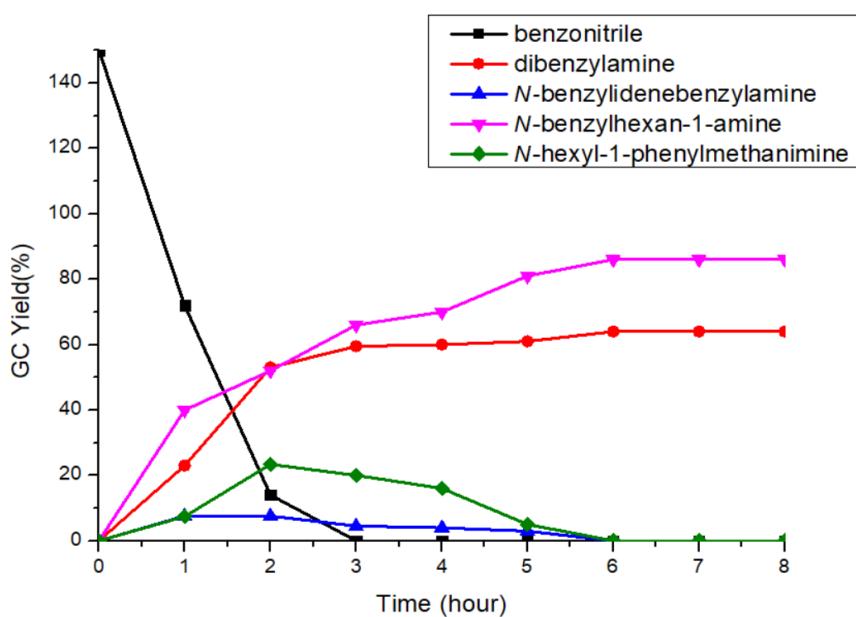


Figure S12. The particle size distribution. (a) PdPt-C NPs; (b)PdPt-CeO₂ NPs; (c) PdPt-TiO₂ NPs.



Reaction condition: 2.0 mol% catalyst, 0.45 mmol of benzonitrile, 0.30 mmol of 1-nitrohexane, 0.60 mmol NaBH₄, 1.8 mL MeOH, 8 h, room temperature.

Figure S13. Conversion versus time plot for *N*-benzylhexan-1-amine formation from the reductive amination of benzonitrile with 1-nitrohexane using PdPt–Fe₃O₄ NPs catalyst.

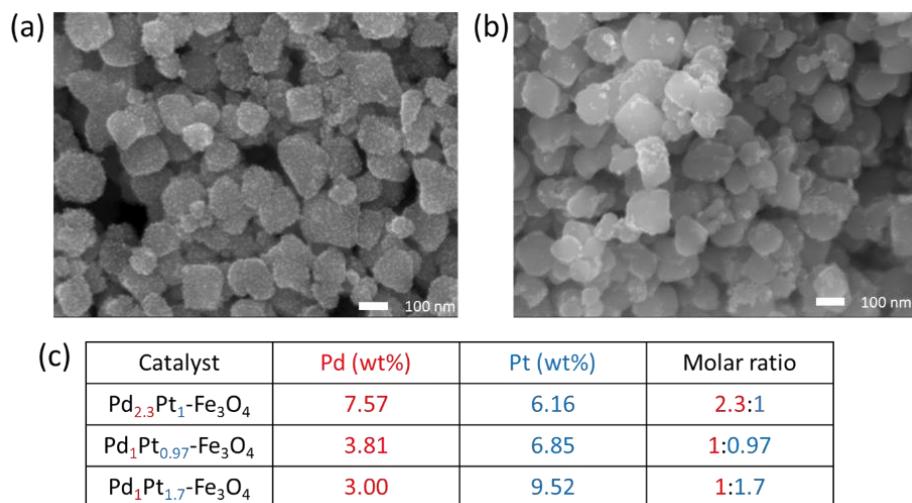
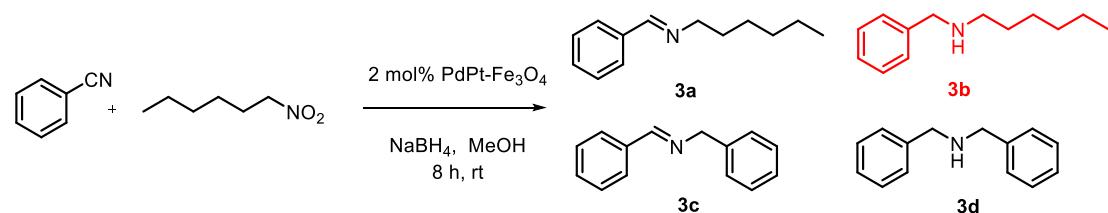


Figure S14. (a) SEM image of $\text{Pd}_{2.3}\text{Pd}_1\text{-Fe}_3\text{O}_4$ NPs; (b) SEM image of $\text{Pd}_1\text{Pd}_{1.7}\text{-Fe}_3\text{O}_4$ NPs; (c) ICP-AES data of $\text{Pd}_x\text{Pd}_y\text{-Fe}_3\text{O}_4$ NPs.

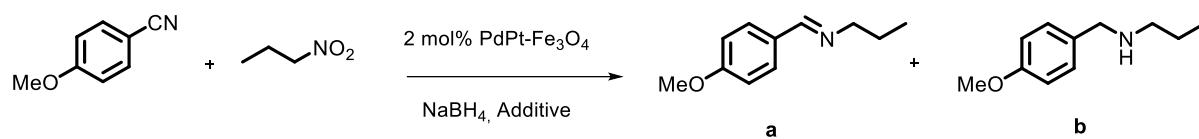
Table S4. Comparison catalytic activity of $\text{Pd}_x\text{Pd}_y\text{-Fe}_3\text{O}_4$ NPs for the unsymmetrical amine under the optimized condition^a



Catalyst	Benzonitrile (Conversion; 150%)	Yield (%)	Selectivity ^b				TON (Turnover number)
			3a (%)	3b (%)	3c (%)	3d (%)	
$\text{Pd-Fe}_3\text{O}_4$	40	27	0	27	0	10	13
$\text{Pd}_{2.3}\text{Pd}_1\text{-Fe}_3\text{O}_4$	150	78	1	78	1	70	39
$\text{Pd}_1\text{Pd}_{0.97}\text{-Fe}_3\text{O}_4$	150	86	1	86	1	62	43
$\text{Pd}_1\text{Pd}_{1.7}\text{-Fe}_3\text{O}_4$	68	0	55	0	13	0	-
$\text{Pt-Fe}_3\text{O}_4$	25	0	23	0	1	0	-

^a Reaction condition: 2.0 mol% catalyst (total metal contents is 4 mol%), 0.45 mmol benzonitrile, 0.30 mmol 1-nitrohexane, 0.60 mmol NaBH_4 , 1.8 mL MeOH, 8 h, room temperature. ^b GC yield

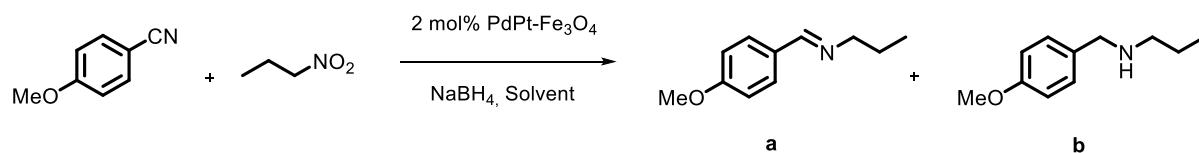
Table S5. Additive screenings toward the reductive amination of 4-methoxybenzonitrile with 1-nitropropane for unsymmetrical secondary amine^a



Entry	Additive (1eq)	Selectivity ^b	
		a (%)	b (%)
1	none	60	0
2	Acetic acid	37	0
3	NH ₄ OAc	51	0
4	NH ₄ Cl	65	0
5	TFE (2,2,2-Trifluoroethanol)	77	0
6	HFIP	61	2
7	ZnCl ₂	-	-

^a Reaction condition: 2.0 mol% PdPt–Fe₃O₄, 0.45 mmol 4-methoxybenzonitrile, 0.30 mmol 1-nitropropane, 0.90 mmol NaBH₄, 0.30 mmol additive, 1.8 mL MeOH, 24 h, room temperature. ^b GC yield.

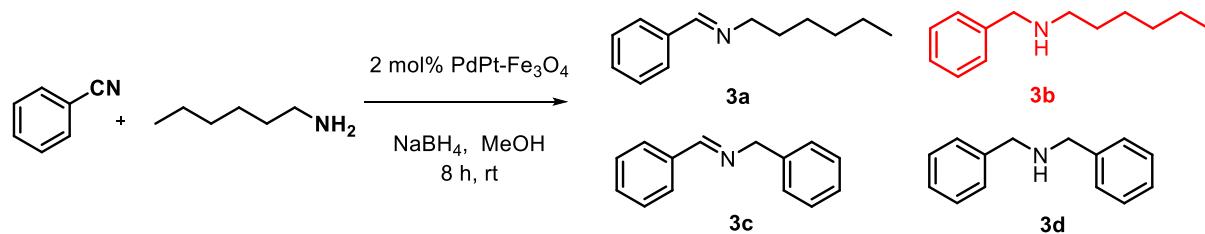
Table S6. Effect of HFIP co-solvent toward the reductive amination of 4-methoxybenzonitrile 1-nitropropane for unsymmetrical secondary amine^a



Entry	Solvent	Yield ^b	
		a (%)	b (%)
1	MeOH only	60	0
2	1 : 1	0	91
3	2 : 1	0	92
4	4 : 1	25	53
5	5 : 1	22	35
6	10 : 1	34	0
7	HFIP only	0	56

^a Reaction condition: 2.0 mol% PdPt–Fe₃O₄, 0.45 mmol 4-methoxybenzonitrile, 0.30 mmol 1-nitropropane, 0.90 mmol NaBH₄, 1.8 mL of solvent, 24 h, room temperature. ^b GC yield.

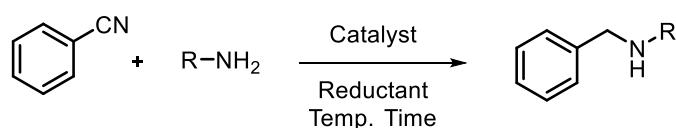
Table S7. Comparison of yields with different nitrile and amine compound stoichiometries^a



Entry	benzonitrile (equivalent)	1-hexylamine (equivalent)	Yield ^b			
			3a (%)	3b (%)	3c (%)	3d (%)
1	1.5	1	0	78	0	70
2	1.2	1	0	61	0	56
3	1	1	0	65	0	35
4	1	1.2	0	82	0	18
5	1	1.5	0	90	0	10

^a Reaction condition: 2.0 mol% PdPt-Fe₃O₄, 0.3 mmol scale, 0.6 mmol NaBH₄, 1.8 mL MeOH, 8 h, room temperature. ^b GC yield.

Table S8. Comparison of the synthesis of unsymmetrical secondary amines from aryl nitriles using the PdPt-Fe₃O₄ catalyst with those using other heterogeneous catalytic systems.



Entry	R-	Catalyst	Reductant	Temp.	Time	Yield
1 ^[7]	Phenyl	Pd/C	H ₂ (balloon)	rt	48 h	-
2 ^[2]	Pentyl	Pt nanowire	1 bar of H ₂ pressure (10 h) and 4 bars of H ₂ pressure (10 h)	80 °C	20 h	93%
3 ^[8]	Hexyl	Pt/C	6 bar of H ₂	105 °C	Flow rate of liquid (0.1 mL min ⁻¹)	91%
4	Hexyl	PdPt-Fe ₃ O ₄	NaBH ₄	rt	8 h	90% ^a (80% ^b)

^a GC yield. ^b Isolated yield.

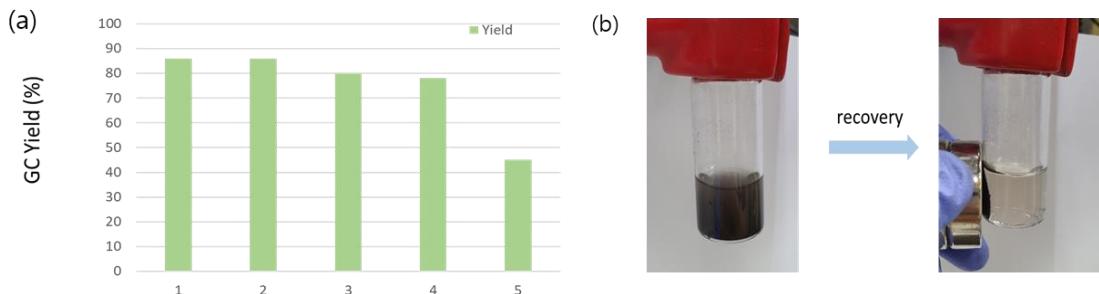
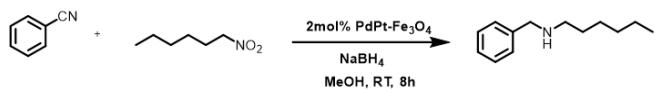


Figure S15. (a) Recyclability of PdPt–Fe₃O₄ for reductive amination of benzonitrile with 1-nitrohexane; (b) Recovery of catalyst using an external magnet.*

* Especially, when the fifth reaction was carried out, unlike the previous results, a mixture of *N*-benzylhexan-1-amine and *N*-hexyl-1-phenylmethanimine was produced. This indicates that as the reaction was repeated, the catalytic activity towards the imine reduction decreased significantly. We compared the HR-TEM images of both fresh and recovered catalysts (Figure S16). The HR-TEM image of the catalyst after 4th run showed considerable agglomeration of the PdPt nanoparticles compared to that of the fresh catalyst. To investigate the oxidation state change, we compared XPS data before and after the reaction (Figure S8) and found no change in the oxidation state. Also, ICP-AES confirmed that the Pd and Pt contents of the PdPt–Fe₃O₄ NPs from the 4th recycle had decreased only by 0.08 wt% and 0.15 wt%, respectively (Figure S7). Since the detachment of PdPt NPs was negligible, it can be confirmed that the catalyst activity was reduced due to the agglomeration of the PdPt nanoparticles, not from the catalyst detachment.

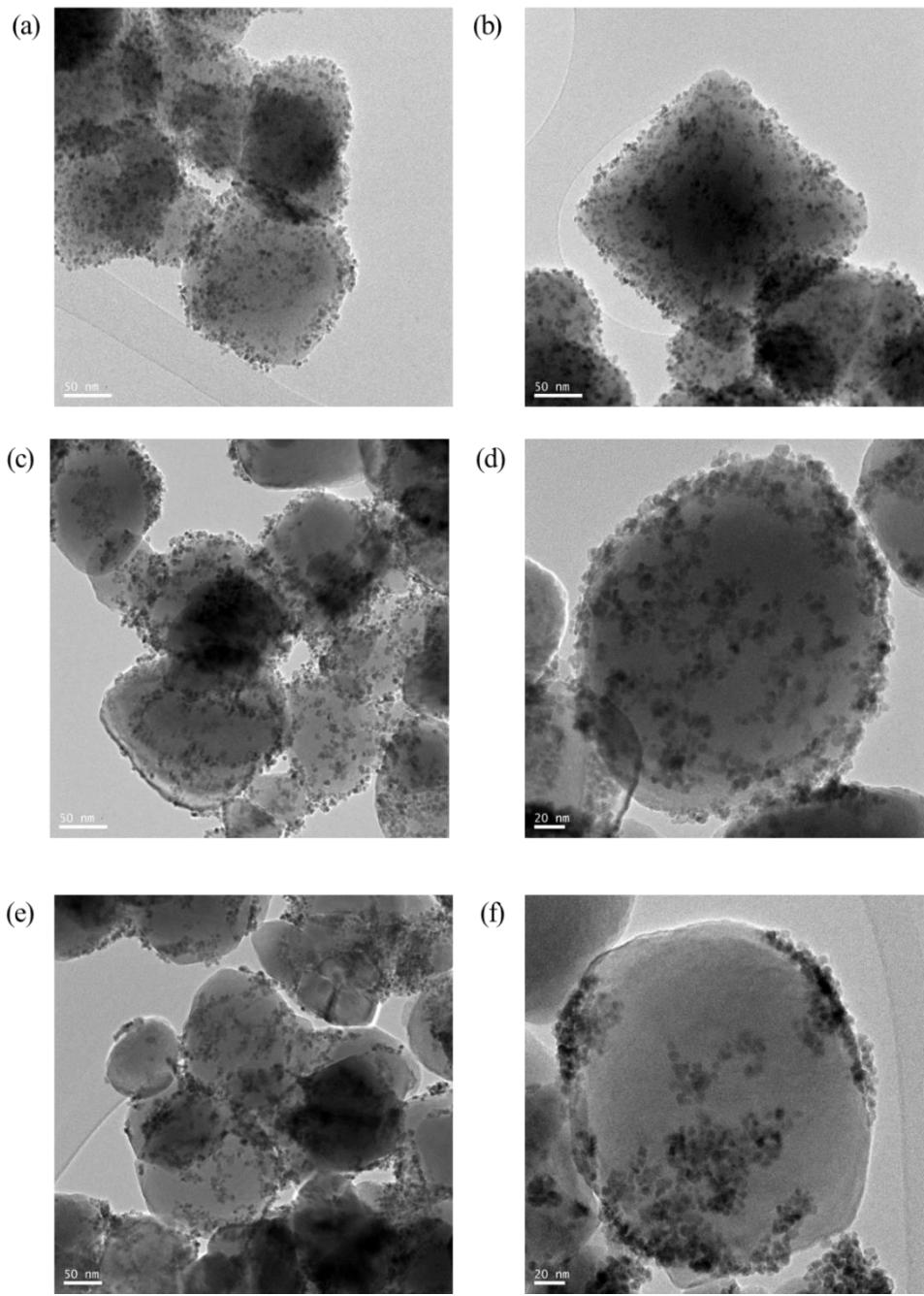
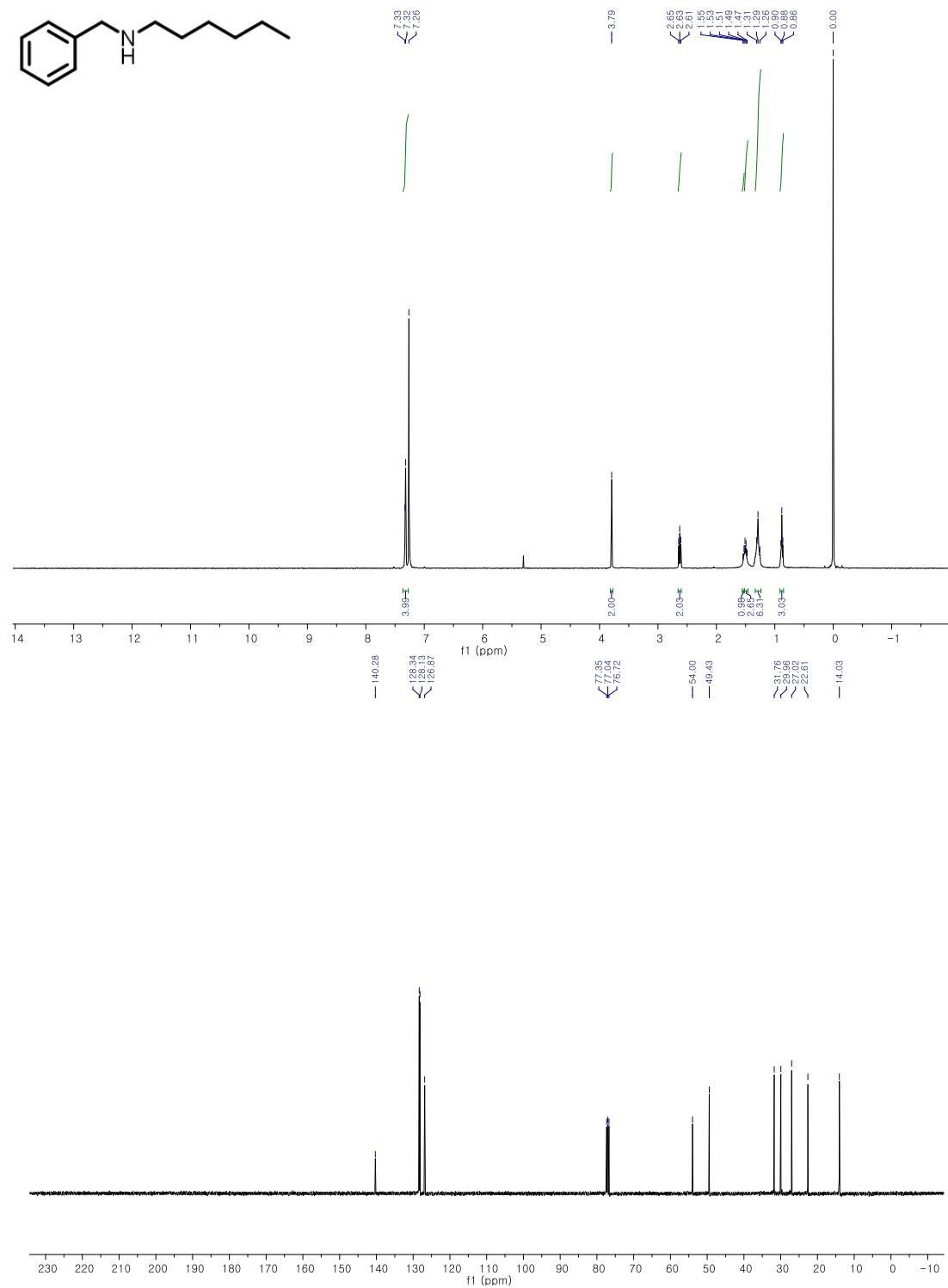


Figure S16. HR-TEM images of NPs. (a) and (b) fresh PdPt–Fe₃O₄ NPs; (c) and (d) PdPt–Fe₃O₄ NPs after 1 catalytic cycle; (e) and (f) PdPt–Fe₃O₄ NPs after 4 catalytic cycle.

4. NMR spectra

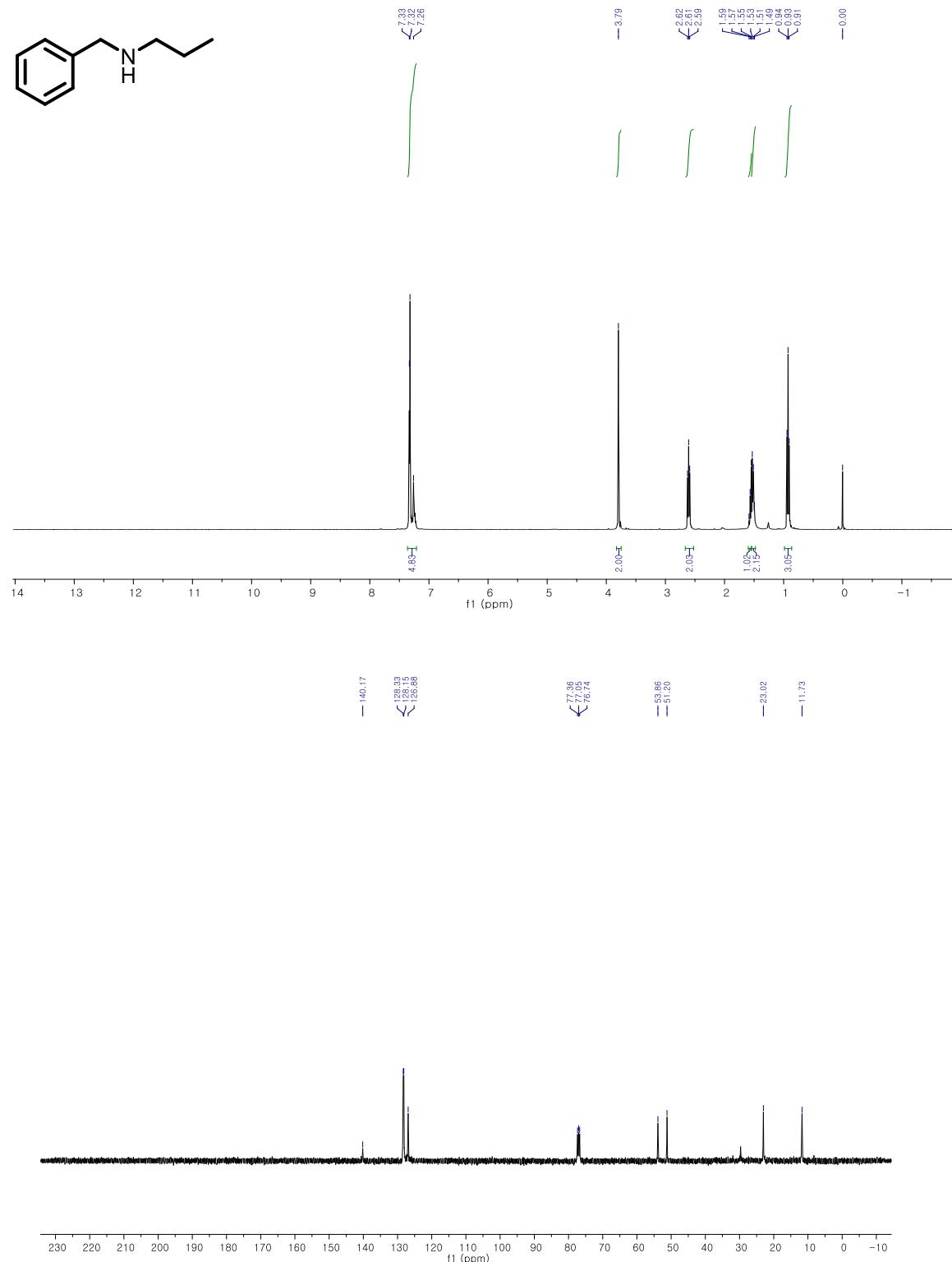
NMR S1, 3a, *N*-benzylhexan-1-amine^[9]

80% (153 mg, Yellow oil); ¹H NMR (400 MHz, CDCl₃) δ 7.32 (d, *J* = 4.4 Hz, 5H), 3.79 (s, 2H), 2.66 – 2.60 (m, 2H), 1.55 (br, 1H) 1.53 – 1.48 (m, 2H), 1.34 – 1.24 (m, 6H), 0.88 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (CDCl₃, 101 MHz) δ: 140.3, 128.3, 128.1, 126.8, 54.0, 49.4, 31.7, 29.9, 27.0, 22.6, 14.0



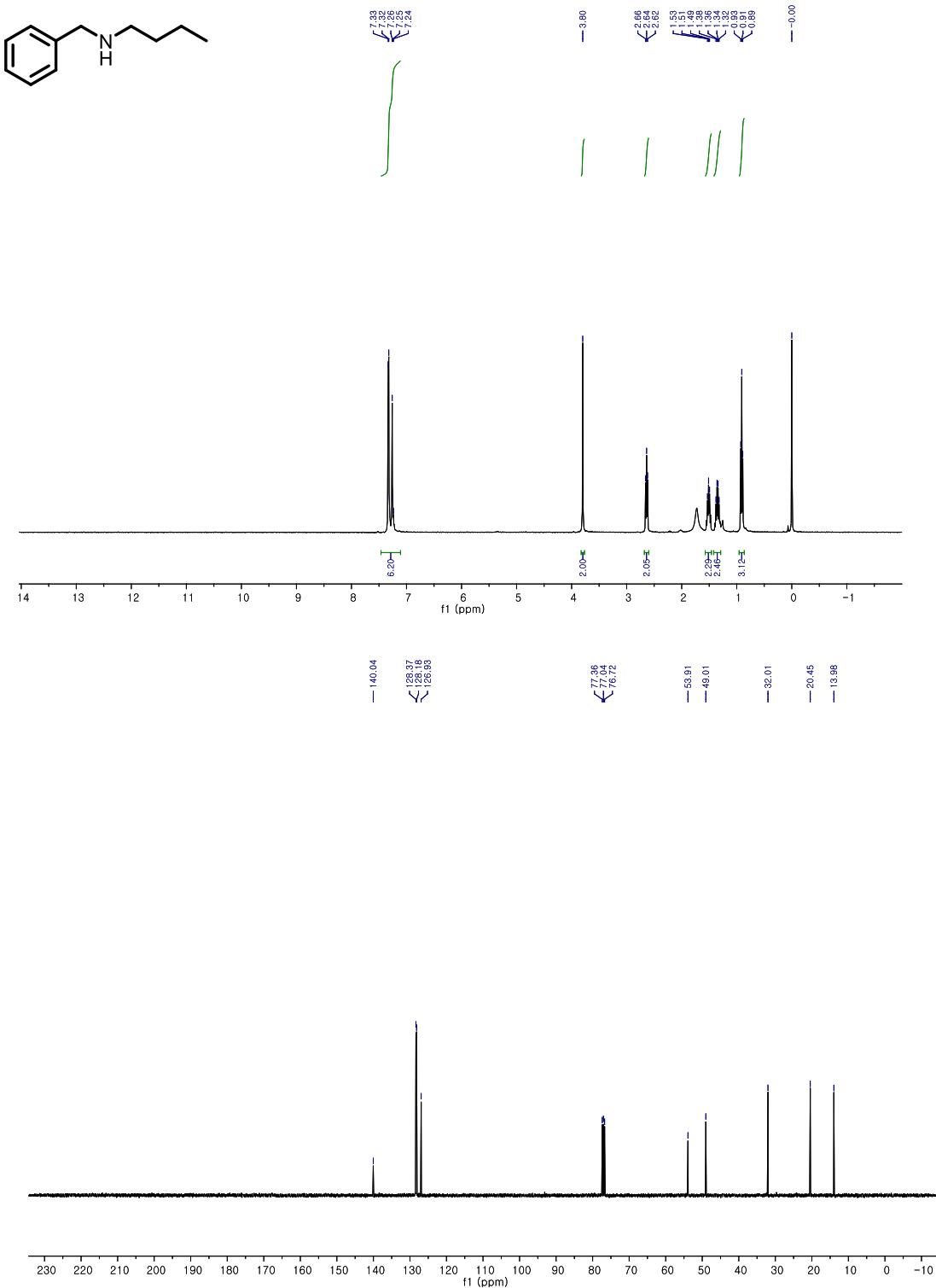
NMR S2, 3g, N-benzylpropan-1-amine^[10]

75% (111 mg, Colorless oil); ¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.21 (m, 5H), 3.79 (s, 2H), 2.61 (t, J = 7.2 Hz, 2H), 1.59 – 1.54 (br, 1H), 1.54 – 1.48 (m, 2H), 0.92 (t, J = 7.4 Hz, 3H). ¹³C NMR (CDCl₃, 101 MHz) δ: 140.2, 128.3, 128.2, 127.0, 53.9, 51.2, 23.0, 11.7



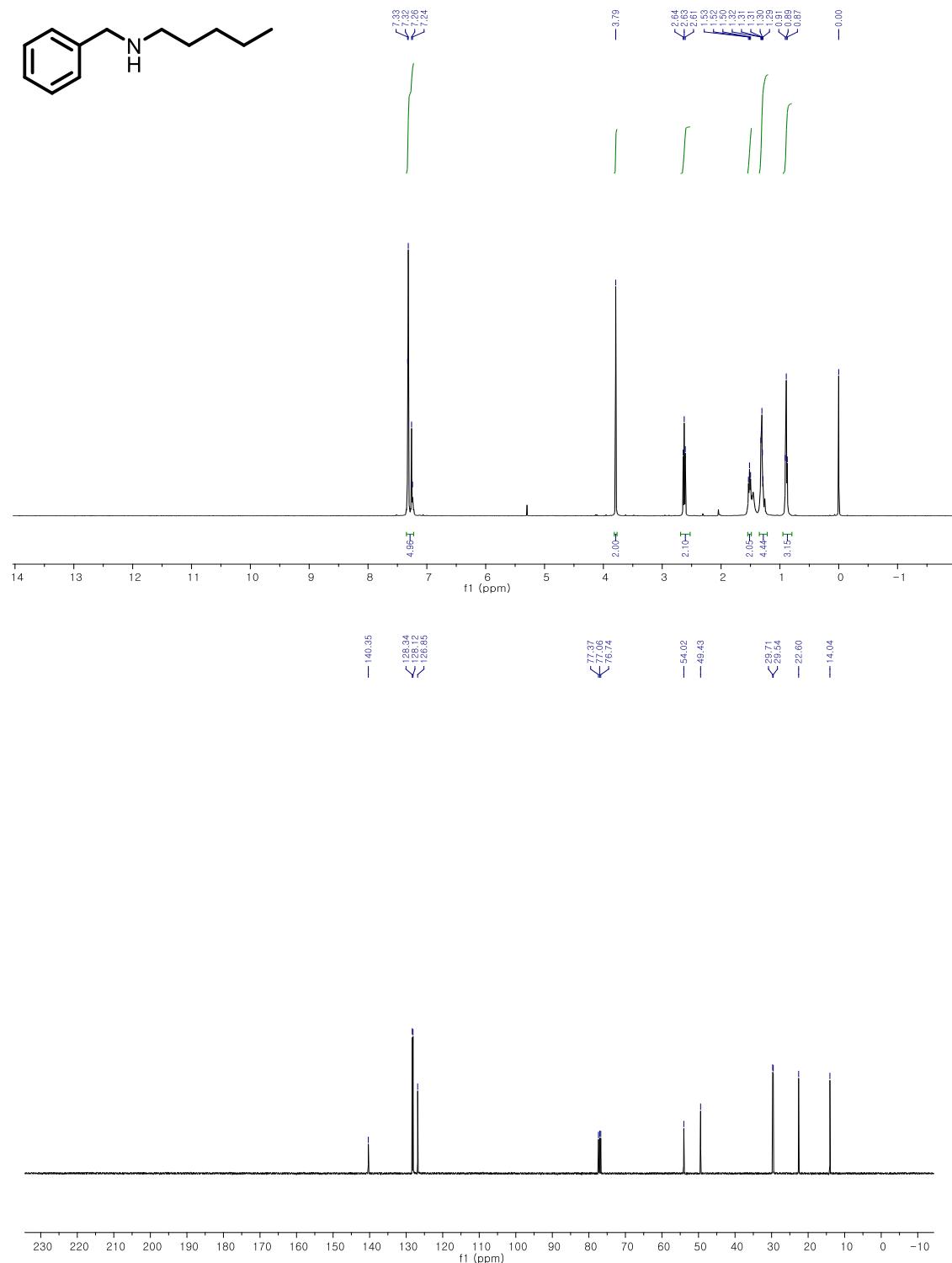
NMR S3, 3f, N-butylbenzylamine^[11]

76% (124mg, Yellow oil); ¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.21 (m, 5H), 3.80 (s, 2H), 2.68 – 2.60 (m, 2H), 1.53-1.48 (m, 2H), 1.38-1.32 (m, 2H), 0.91 (t, J = 7.3 Hz, 3H). ¹³C NMR (CDCl₃, 101 MHz) δ: 140.0, 128.4, 128.2, 126.9, 53.9, 49.0, 32.0, 20.5, 14.0.



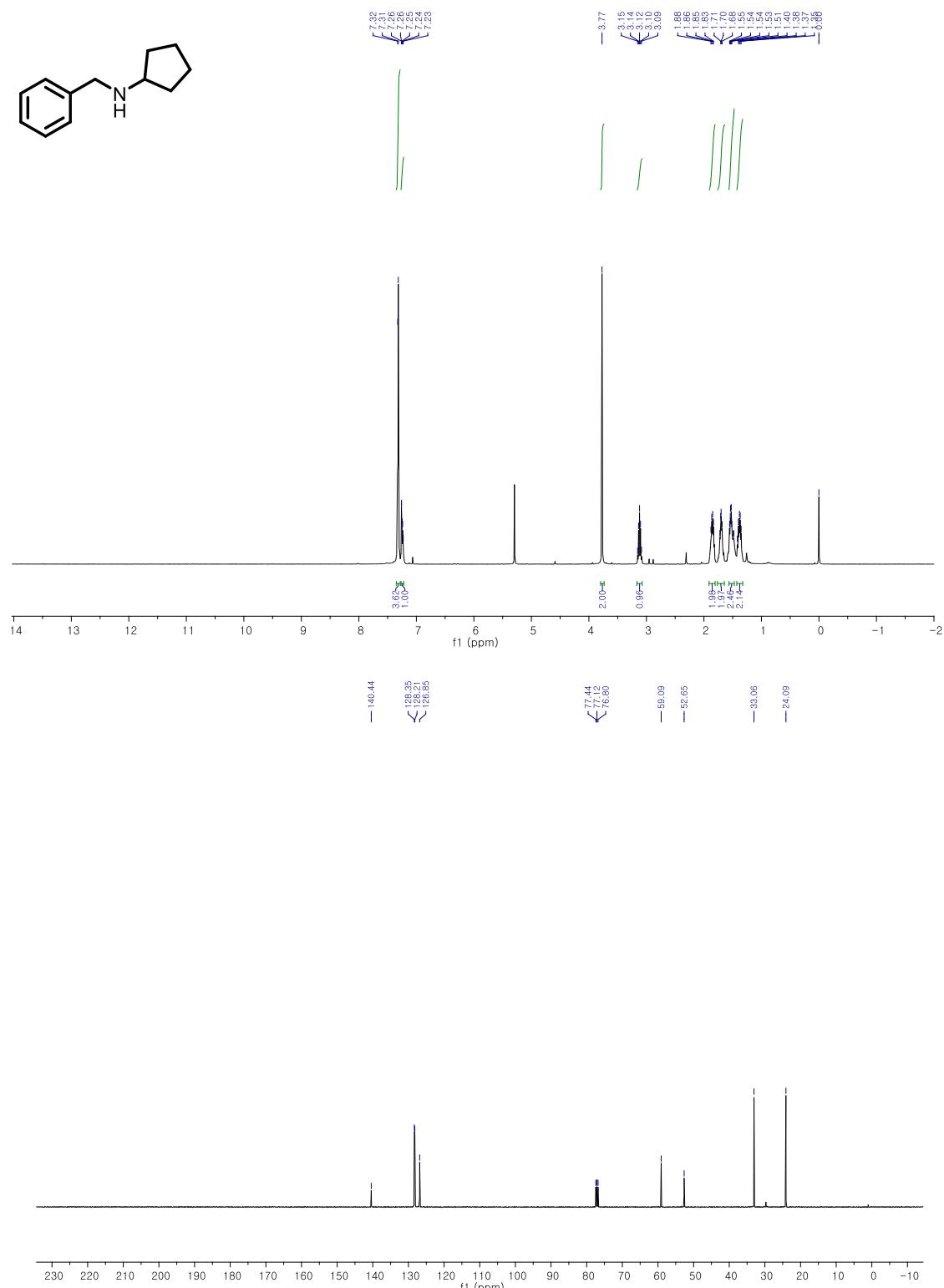
NMR S4, 3e, N-benzylpentan-1-amine^[12]

86% (152mg, Colorless oil) ; ¹H NMR (400 MHz, CDCl₃) δ 7.32–7.23 (m, 5H), 3.79 (s, 2H), 2.63 (t, *J* = 7.3 Hz, 2H), 1.55 – 1.48 (m, 2H), 1.38 – 1.26 (m, 4H), 0.89 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.4, 128.3, 128.1, 126.9, 54.0, 49.4, 29.8, 29.5, 22.6, 14.0.



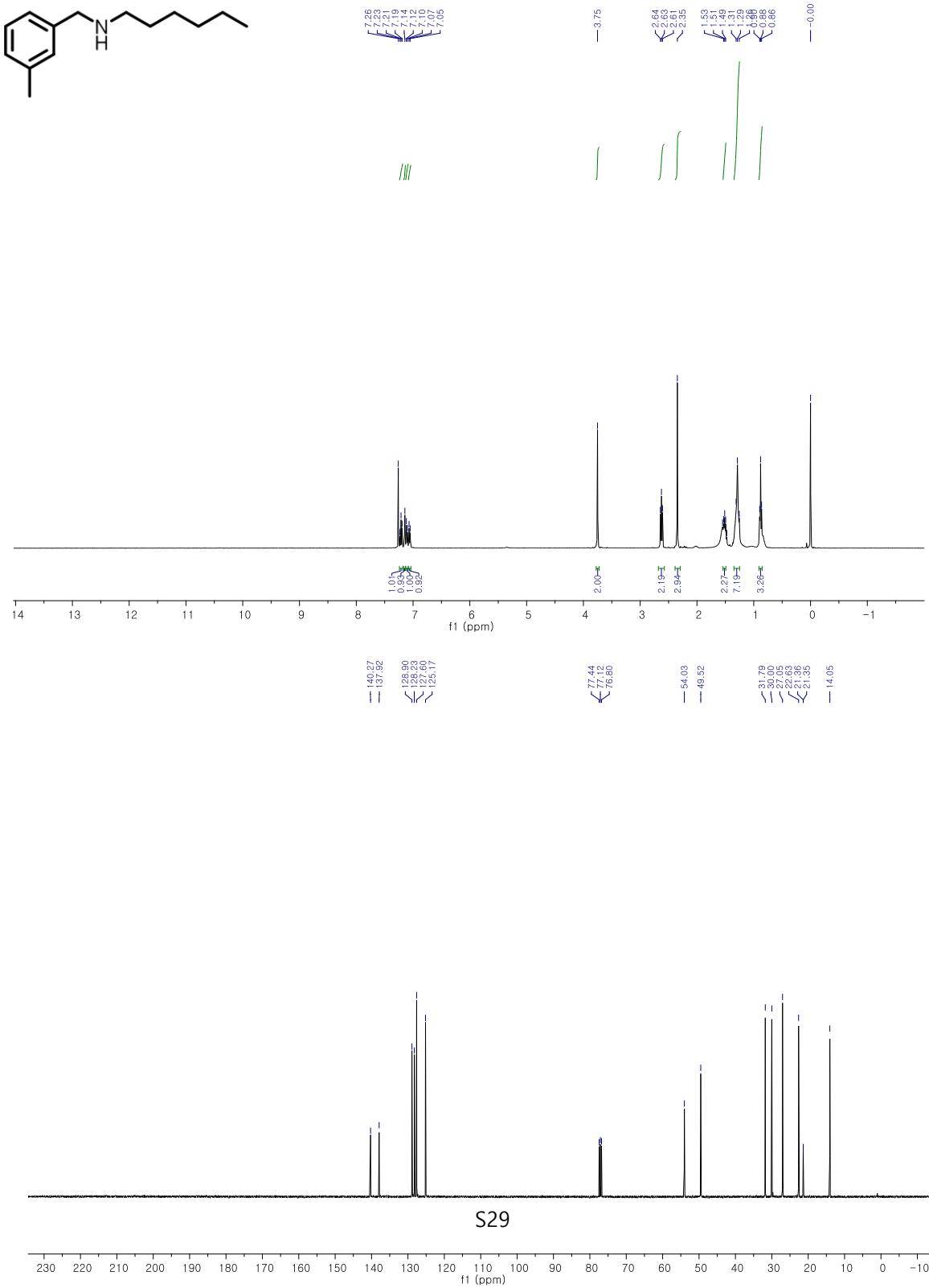
NMR S5, 3h, N-Benzyl-cyclopentyl-amine^[13]

80% (140 mg, Yellow oil); ¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.23 (m, 5H), 3.78 (s, 2H), 3.16 – 3.07 (m, 1H), 1.91 – 1.81 (m, 2H), 1.74 – 1.66 (m, 2H), 1.57 – 1.50 (m, 2H), 1.45 – 1.35 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 140.4, 128.4, 128.2, 126.9, 59.1, 52.7, 33.1, 24.1.



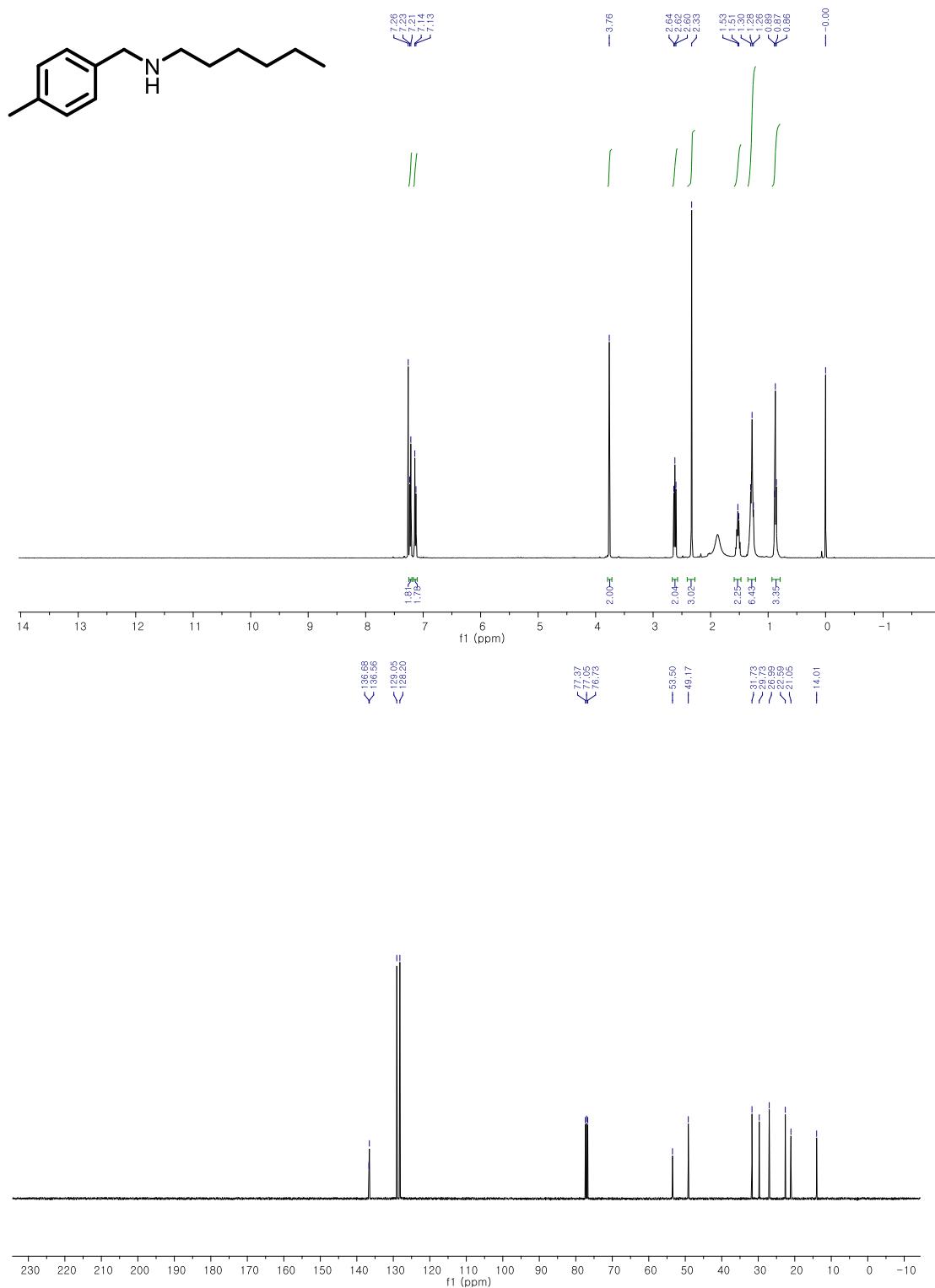
NMR S6, 3i, N-(3-methylbenzyl)hexan-1-amine

84% (172 mg, Yellow oil); ^1H NMR (400 MHz, CDCl_3) δ 7.21 (t, $J = 7.5$ Hz, 1H), 7.14 (s, 1H), 7.11 (d, $J = 7.6$ Hz, 1H), 7.06 (d, $J = 7.4$ Hz, 1H), 3.75 (s, 2H), 2.68 – 2.58 (m, 2H), 2.35 (s, 3H), 1.54 – 1.48 (m, 2H), 1.28 (t, $J = 11.0$ Hz, 6H), 0.88 (t, $J = 6.7$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 140.3, 137.9, 128.9, 128.2, 127.6, 125.1, 54.0, 49.5, 31.8, 30.0, 27.0, 22.6, 21.4, 21.3, 14.0. HR-MS (EI): calcd for $\text{C}_{14}\text{H}_{23}\text{N}$ 205.1890; found: 233.1831.



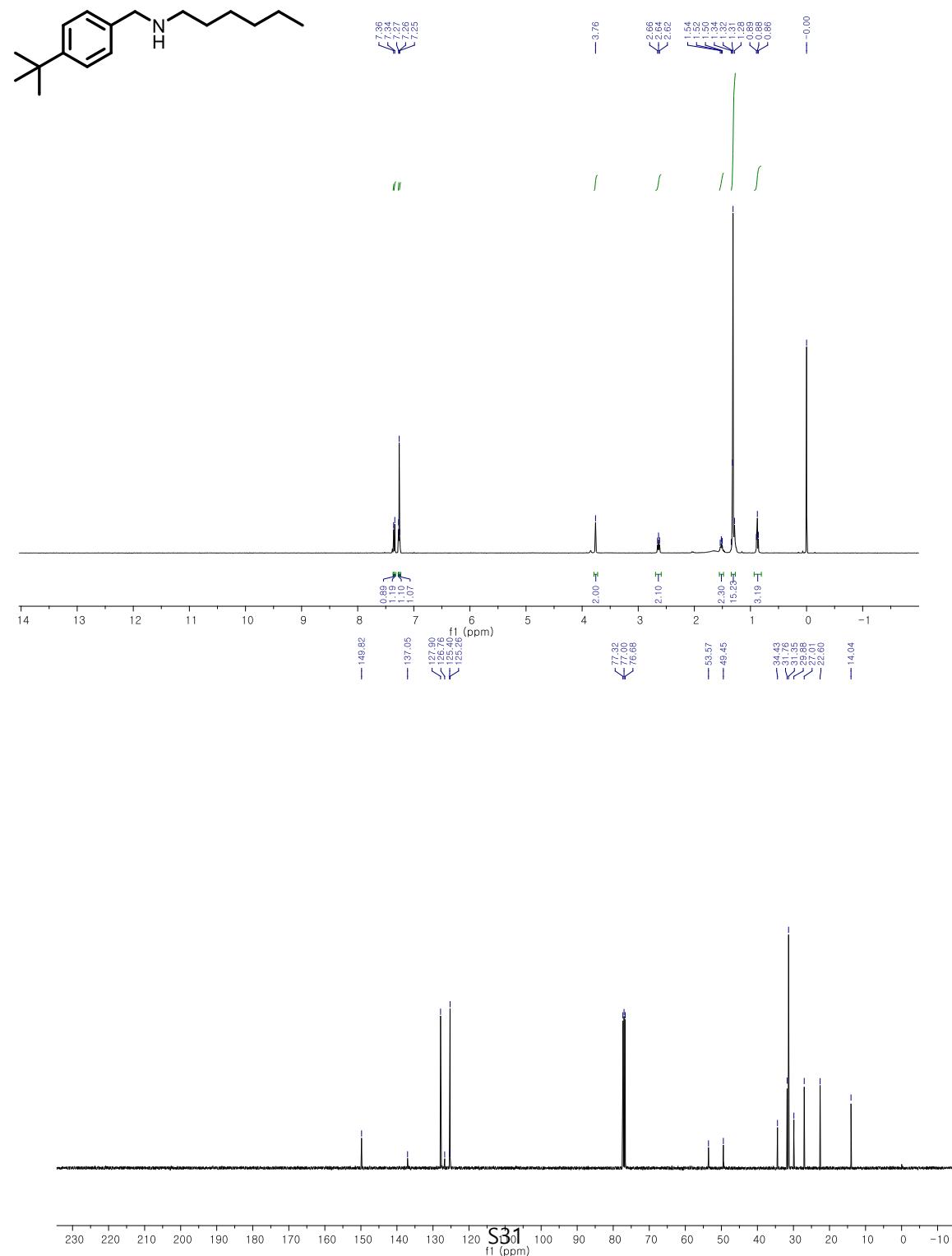
NMR S7, 3j, N-(4-methylbenzyl)hexan-1-amine^[14]

80% (164 mg, Colorless oil); ¹H-NMR (400 MHz, CDCl₃) 1H NMR (400 MHz, CDCl₃) δ 7.23-21 (d, 2H), 7.14-13 (d, 2 H), 3.76 (s, 2 H), 2.62 (t, 2H), 2.33(s, 3H), 1.53-1.51(m, 2H), 1.30-1.26 (m, 6H), 0.87 (t, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 136.7, 136.6, 129.0, 128.2, 53.5, 49.2, 31.7, 29.7, 27.0, 22.6, 21.0, 14.0.



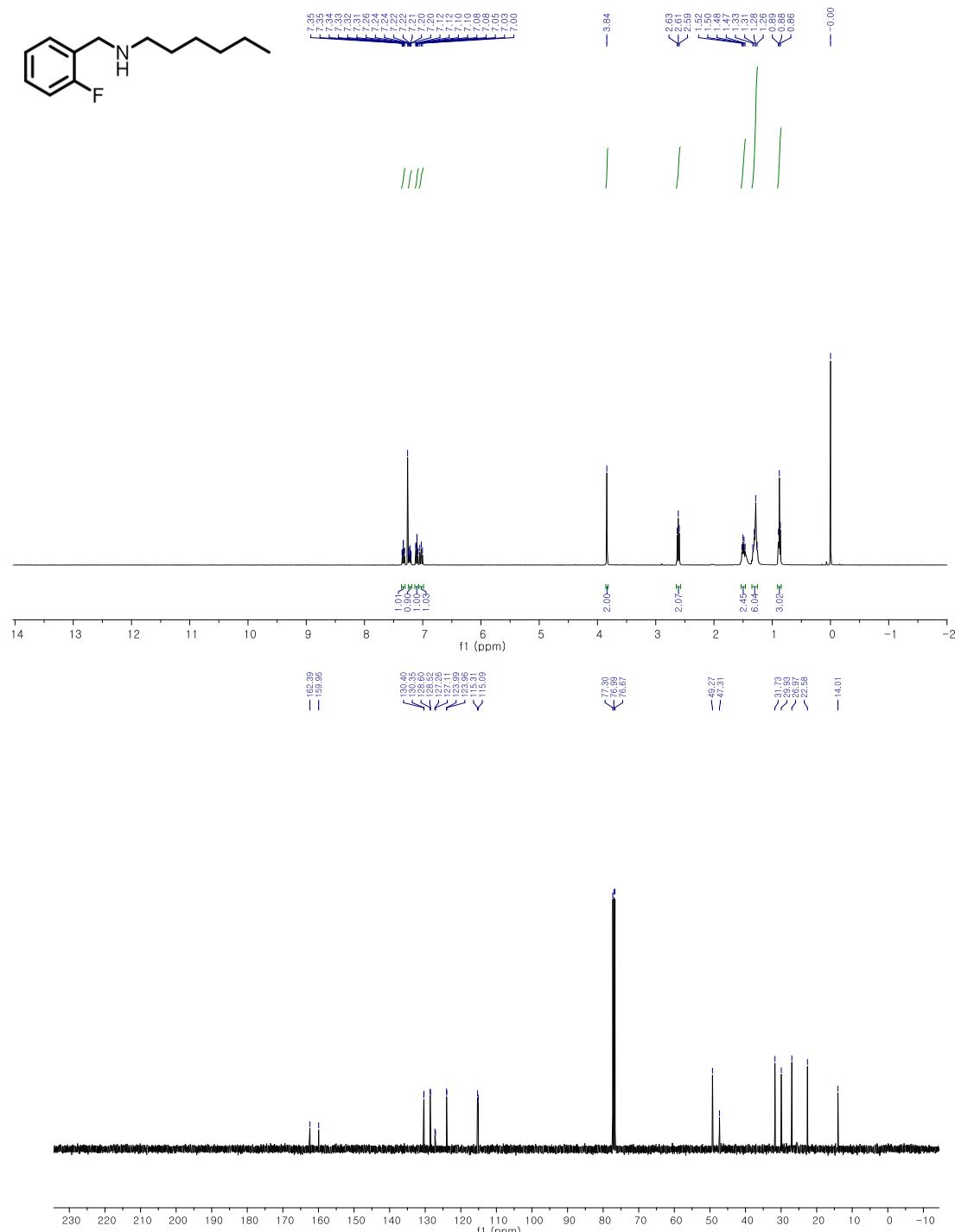
NMR S8, 3k, N-(4-(tert-butyl)benzyl)hexan-1-amine

60% (148mg, Colorless oil); ^1H NMR (400 MHz, CDCl_3) δ 7.36 (s, 1H), 7.34 (s, 1H), 7.27 (s, 1H), 7.25 (s, 1H), 3.76 (s, 2H), 2.64 (t, J = 7.3 Hz, 2H), 1.55 – 1.48 (m, 2H), 1.33 – 1.25 (m, 15H), 0.88 (t, J = 6.8 Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 149.8, 137.1, 127.9, 126.8, 125.4, 125.2, 53.6, 49.5, 34.4, 31.8, 31.3, 29.9, 27.0, 22.6, 14.0. HR-MS (EI): calcd for $\text{C}_{17}\text{H}_{29}\text{N}$ 247.2300; found: 247.2301.



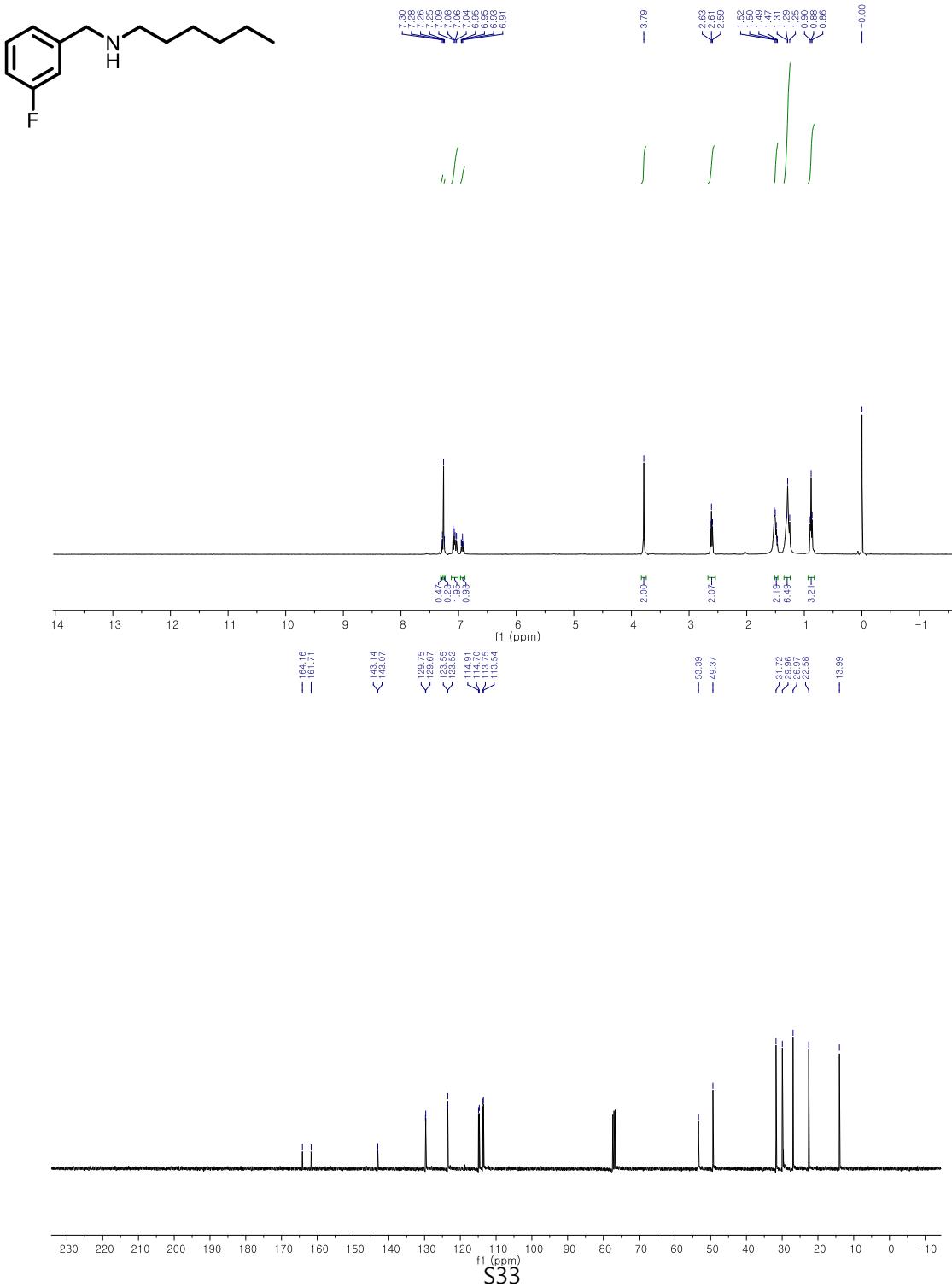
NMR S9, 3I, N-(2-fluorobenzyl)hexan-1-amine

62% (129.7 mg, Colorless oil); ^1H NMR (400 MHz, CDCl_3) δ 7.33 (td, $J = 7.6, 1.6$ Hz, 1H), 7.25 – 7.19 (m, 1H), 7.10 (td, $J = 7.5, 1.0$ Hz, 1H), 7.07 – 6.99 (m, 1H), 3.84 (s, 2H), 2.61 (t, $J = 7.2$ Hz, 2H), 1.52–1.47 (m, 2H), 1.33–1.26 (m, 6H). 0.88 (t, $J = 6.8$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 161.18 (d, $J = 245.2$ Hz), 130.37 (d, $J = 4.9$ Hz), 128.56 (d, $J = 8.2$ Hz), 127.19 (d, $J = 15.2$ Hz), 123.98 (d, $J = 3.5$ Hz), 115.20 (d, $J = 21.9$ Hz), 49.27, 47.31, 31.73, 29.93, 26.97, 22.58, 14.01. HR-MS (EI): calcd for $\text{C}_{13}\text{H}_{20}\text{FN}$ 209.1580; found: 209.1579.



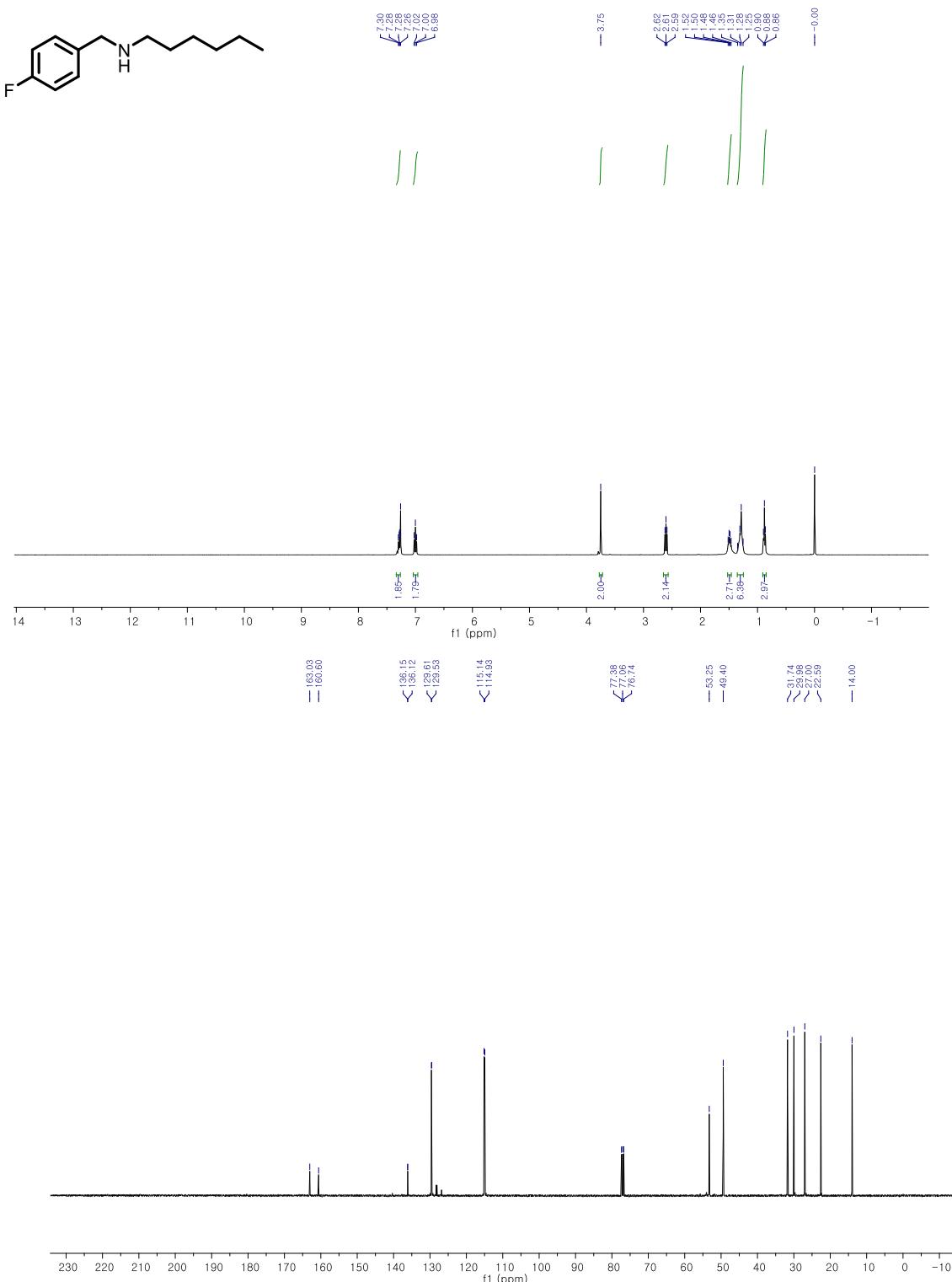
NMR S10, 3m, N-(3-fluorobenzyl)hexan-1-amine

60% (125 mg, Colorless oil); ^1H NMR (400 MHz, CDCl_3) δ 7.28 (t, $J = 10.8$ Hz, 1H), 7.07 (dd, $J = 14.8$, 8.6 Hz, 2H), 6.93 (td, $J = 8.3$, 2.0 Hz, 1H), 3.79 (s, 2H), 2.61 (t, $J = 7.2$ Hz, 2H), 1.57 – 1.42 (m, 2H), 1.37 – 1.24 (m, 6H), 0.88 (t, $J = 6.7$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 162.93 (d, $J = 245.6$ Hz), 143.10 (d, $J = 7.0$ Hz), 129.71 (d, $J = 8.2$ Hz), 123.53 (d, $J = 2.7$ Hz), 114.81 (d, $J = 21.2$ Hz), 113.54 (d, $J = 21.1$ Hz), 53.39, 49.37, 31.72, 29.96, 26.97, 22.58, 13.99. HR-MS (EI): calcd for $\text{C}_{13}\text{H}_{20}\text{FN}$ 209.1580; found: 209.1582.



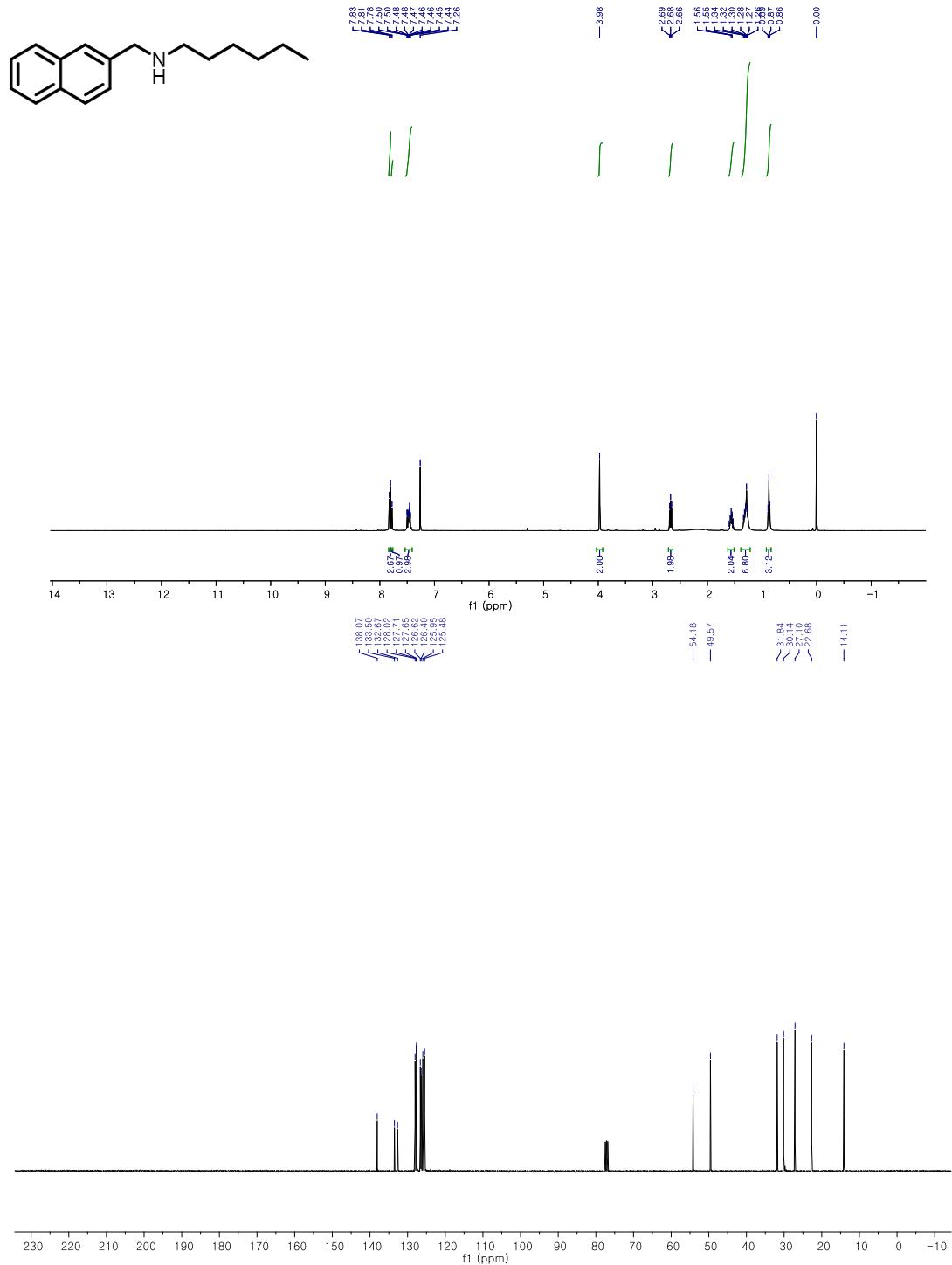
NMR S11, 3n, N-(4-fluorobenzyl)hexan-1-amine^[15]

81% (169mg, Colorless oil); ¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.26 (m, 2H), 7.09 – 6.85 (m, 2H), 3.75 (s, 2H), 2.60 (t, J = 7.2 Hz, 2H), 1.54 – 1.44 (m, 2H), 1.37 – 1.19 (m, 6H), 0.88 (t, J = 6.7 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 161.82 (d, J = 244.5 Hz), 136.14 (d, J = 3.1 Hz), 129.57 (d, J = 7.9 Hz), 115.04 (d, J = 21.2 Hz), 53.25, 49.40, 31.74, 29.98, 27.00, 22.59, 14.00.



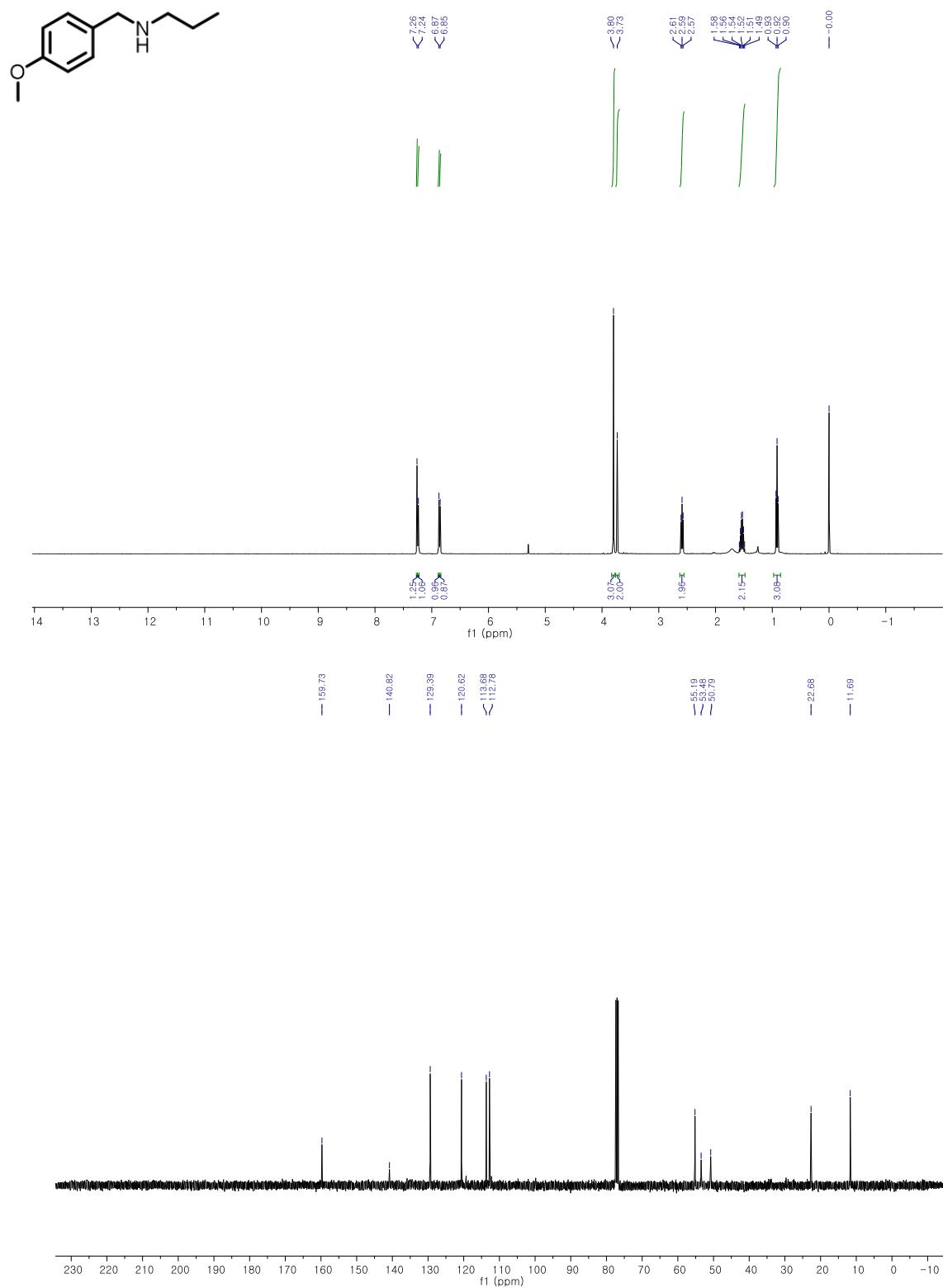
NMR S12, 3o, N-(naphthalen-2-ylmethyl)hexan-1-amine

87% (210mg, Yellow oil); ^1H NMR (400 MHz, CDCl_3) δ 7.82 (d, $J = 8.3$ Hz, 3H), 7.78 (s, 1H), 7.54 – 7.41 (m, 3H), 3.98 (s, 2H), 2.71 – 2.64 (m, 2H), 1.63 – 1.51 (m, 2H), 1.39 – 1.22 (m, 6H), 0.87 (t, $J = 6.8$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 138.07, 133.50, 132.67, 132.52, 129.71, 128.65, 128.62, 128.55, 128.48, 125.48, 54.18, 49.57, 31.84, 30.14, 27.10, 22.68, 14.11. HR-MS (EI): calcd for $\text{C}_{17}\text{H}_{23}\text{N}$ 241.1830; found: 241.1826.



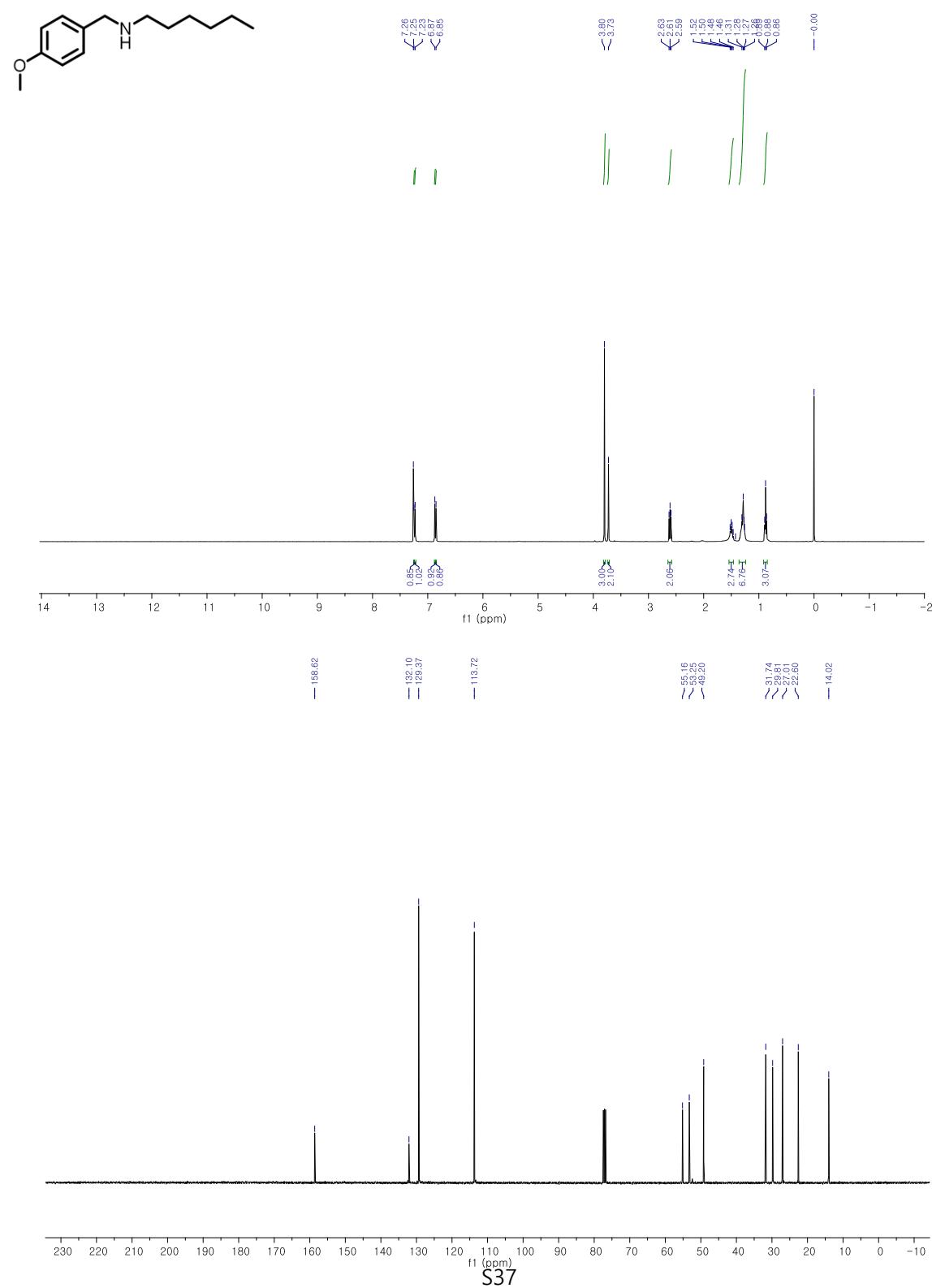
NMR S13, 3p, N-(4-methoxybenzyl)propan-1-amine^[16]

80% (143 mg Colorless oil) ; ¹H NMR (400 MHz, CDCl₃) δ 7.26 (s, 1H), 7.24 (s, 1H), 6.87 (s, 1H), 6.85 (s, 1H), 3.80 (s, 3H), 3.73 (s, 2H), 2.59 (t, J = 7.3 Hz, 2H), 1.59 – 1.48 (m, 2H), 0.92 (t, J = 7.4 Hz, 3H). ¹³C NMR(101 MHz, CDCl₃) δ 159.7, 140.8, 129.4, 120.6, 113.7, 112.8, 55.2, 53.5, 50.8, 22.7, 11.7.



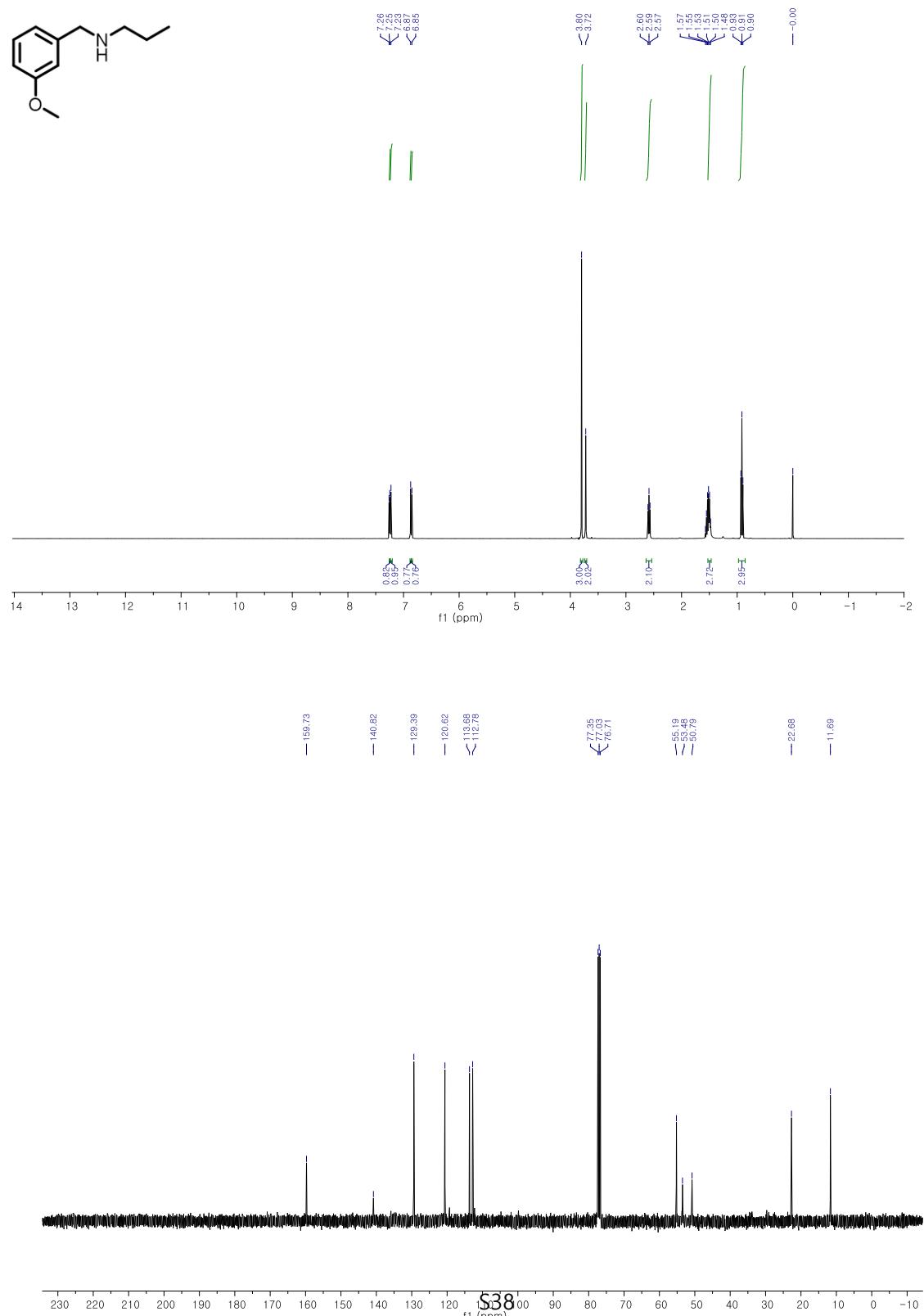
NMR S14, 3q, N-(4-methoxybenzyl)hexan-1-amine^[9]

71% (157 mg, Colorless oil); ¹H NMR (400 MHz, CDCl₃) δ 7.25 (s, 1H), 7.23 (s, 1H), 6.87 (s, 1H), 6.85 (s, 1H), 3.80 (s, 3H), 3.73 (s, 2H), 2.65 – 2.58 (m, 2H), 1.49 (dd, *J* = 14.7, 7.6 Hz, 2H), 1.28 (dd, *J* = 12.1, 6.5 Hz, 6H), 0.88 (t, *J* = 6.8 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 158.6, 132.1, 129.4, 113.7, 55.2, 53.3, 49.2, 31.7, 29.8, 27.0, 22.6, 14.0.



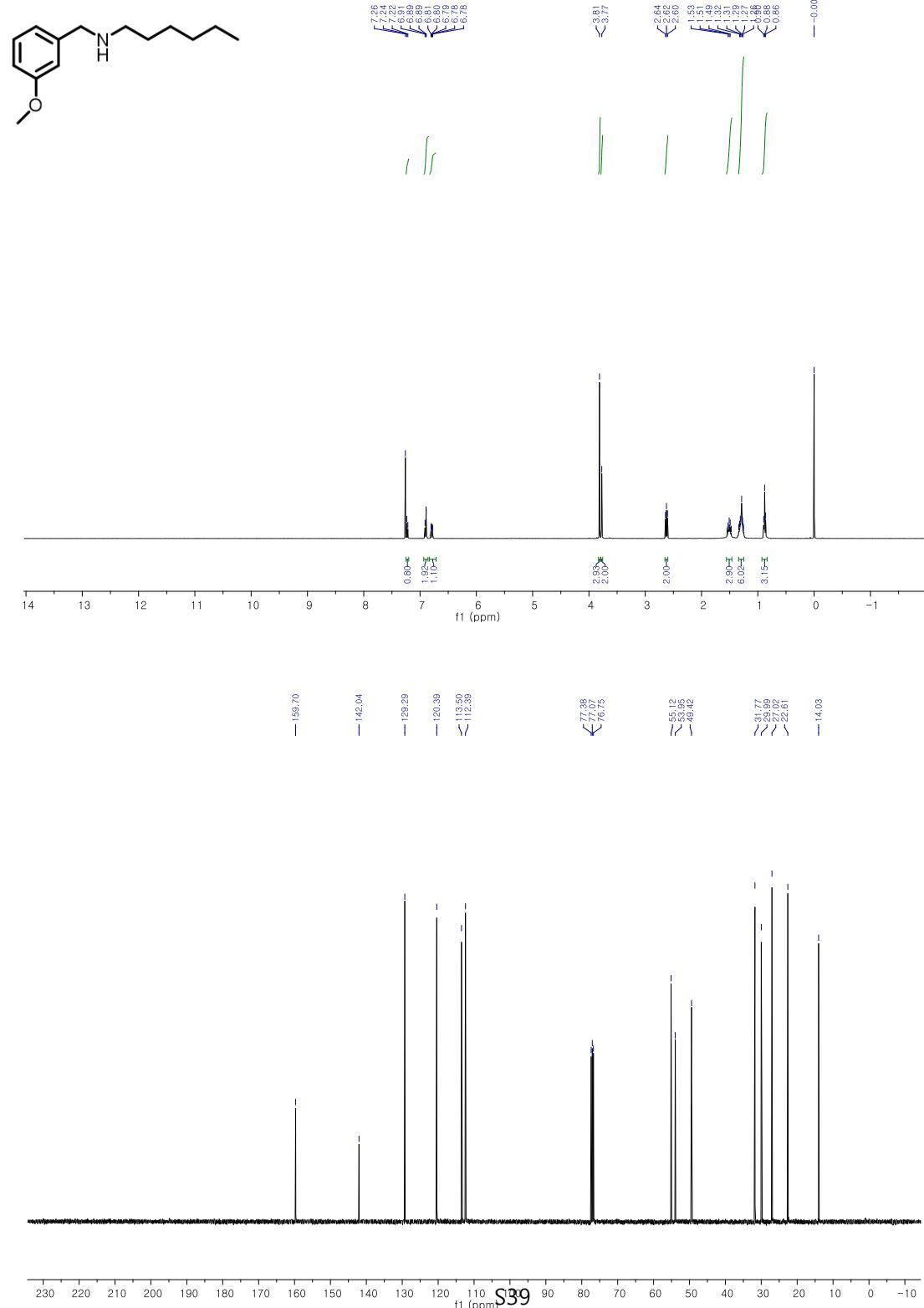
NMR S15, 3r, N-(3-methoxybenzyl)propan-1-amine^[10]

66% (118mg ,Colorless oil); ¹H NMR (400 MHz, CDCl₃) δ 7.25 (s, 1H), 7.23 (s, 1H), 6.87 (s, 1H), 6.85 (s, 1H), 3.80 (s, 3H), 3.72 (s, 2H), 2.59 (t, J = 7.2 Hz, 2H), 1.53 – 1.47 (m, 2H), 0.91 (t, J = 7.4 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 159.7, 140.8, 129.4, 120.62, 113.7, 112.8, 55.2, 53.5, 50.8, 22.7, 11.7.



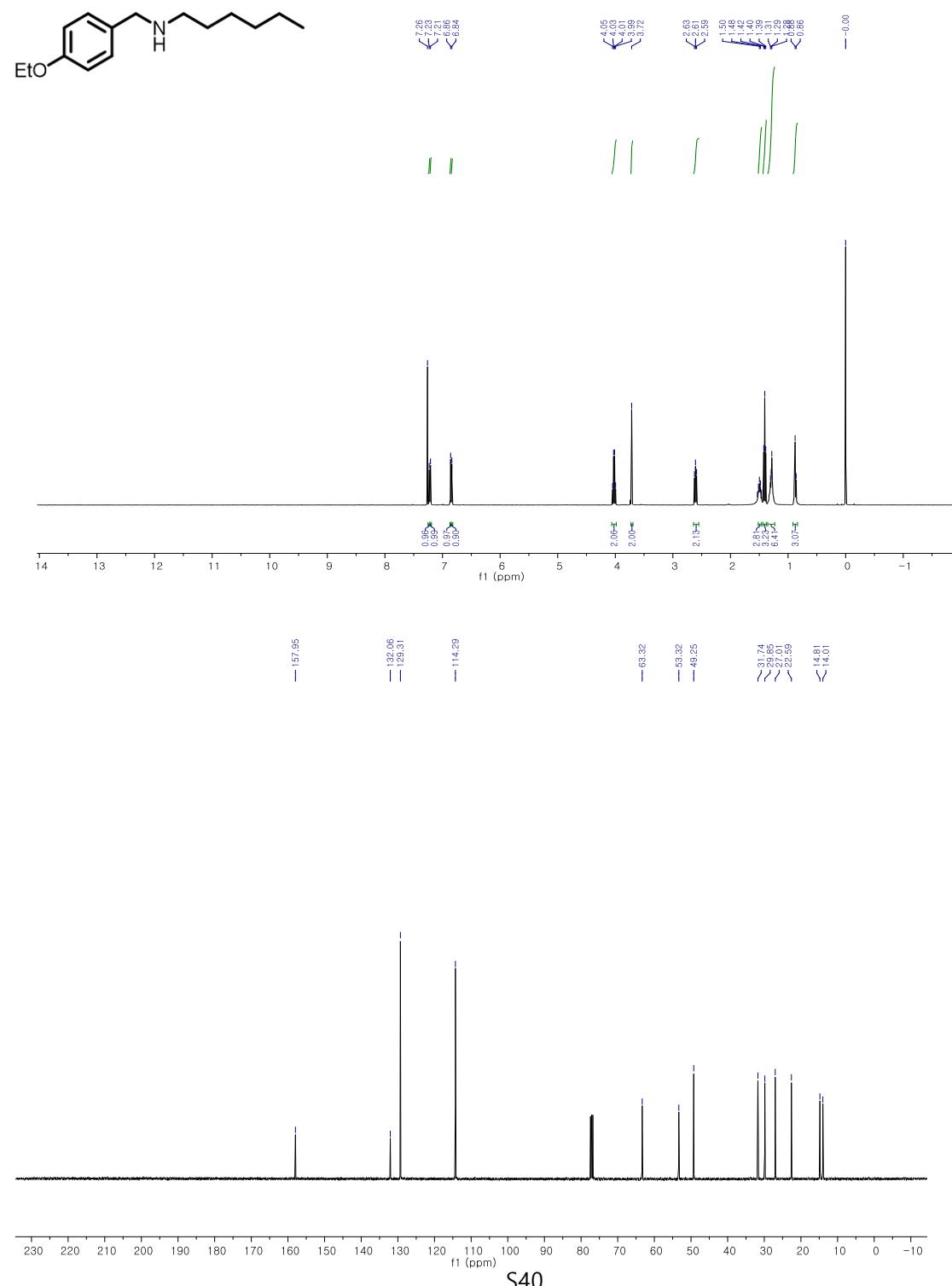
NMR S16, 3s, N-(3-methoxybenzyl)hexan-1-amine.

67% (148 mg, Yellow oil); ^1H NMR (400 MHz, CDCl_3) δ 7.23 (d, $J = 7.7$ Hz, 1H), 6.93 – 6.86 (m, 2H), 6.84 – 6.72 (m, 1H), 3.81 (s, 3H), 3.77 (s, 2H), 2.65 – 2.60 (m, 2H), 1.56 – 1.46 (m, 3H), 1.34 – 1.25 (m, 6H), 0.88 (t, $J = 6.8$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.70, 142.04, 129.29, 120.39, 113.50, 112.39, 55.12, 53.95, 49.42, 31.77, 29.99, 27.02, 22.61, 14.03. HR-MS (EI): calcd for $\text{C}_{14}\text{H}_{23}\text{NO}$ 221.1780; found: 221.1781.



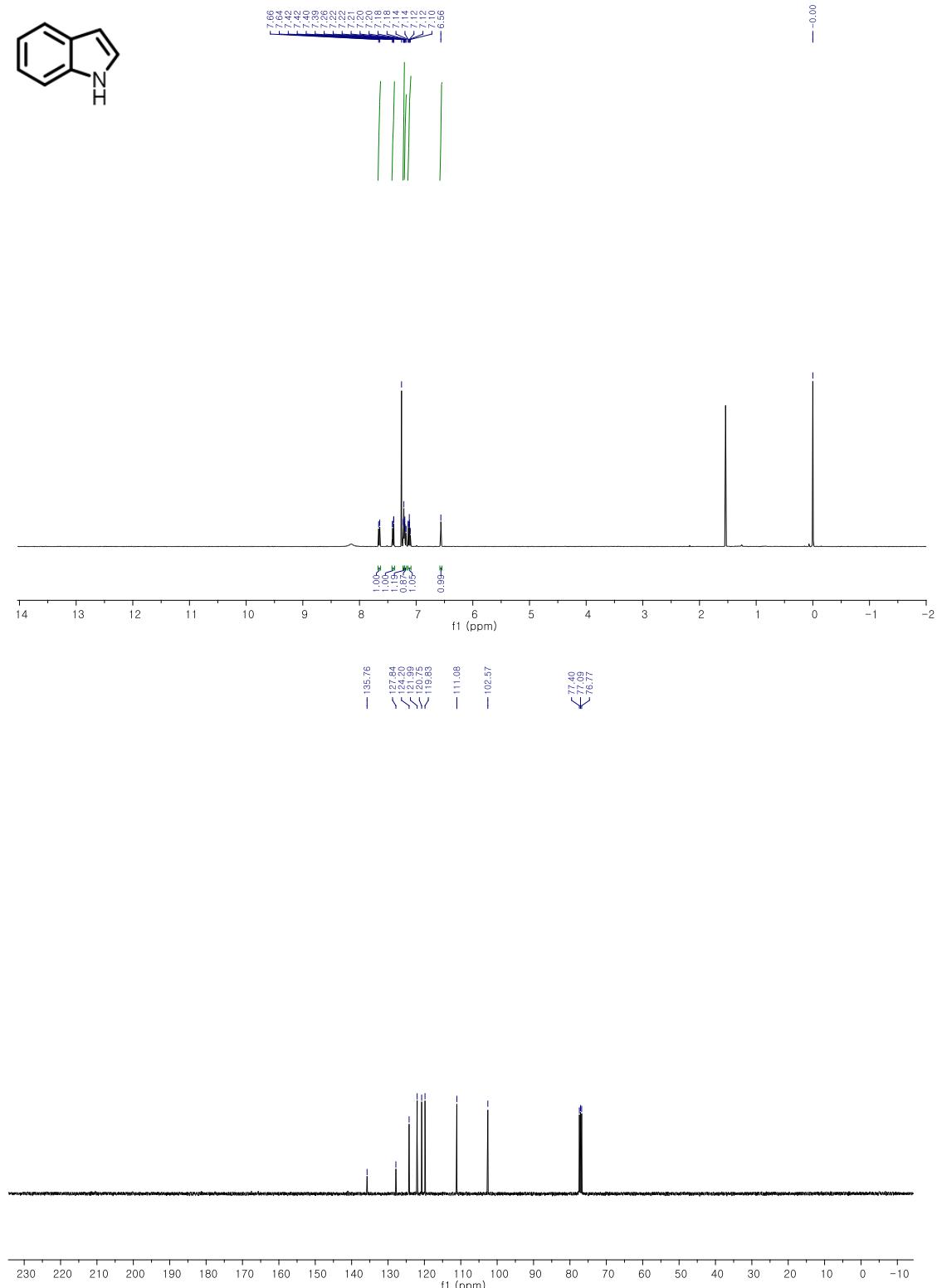
NMR S17, 3t, N-(4-ethoxybenzyl)hexan-1-amine.

73% (171 mg, Yellow oil); ^1H NMR (400 MHz, CDCl_3) δ 7.23 (s, 1H), 7.21 (s, 1H), 6.86 (s, 1H), 6.84 (s, 1H), 4.02 (q, $J = 7.0$ Hz, 2H), 3.72 (s, 2H), 2.64 – 2.55 (m, 2H), 1.53 – 1.45 (m, 2H), 1.40 (t, $J = 7.0$ Hz, 3H), 1.35 – 1.23 (m, 6H), 0.87 (d, $J = 7.1$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 157.95, 132.06, 129.31, 114.29, 63.32, 53.32, 49.25, 31.74, 29.85, 27.01, 22.59, 14.81, 14.01. HR-MS (EI): calcd for $\text{C}_{14}\text{H}_{23}\text{NO}$ 235.1936; found: 235.1931.



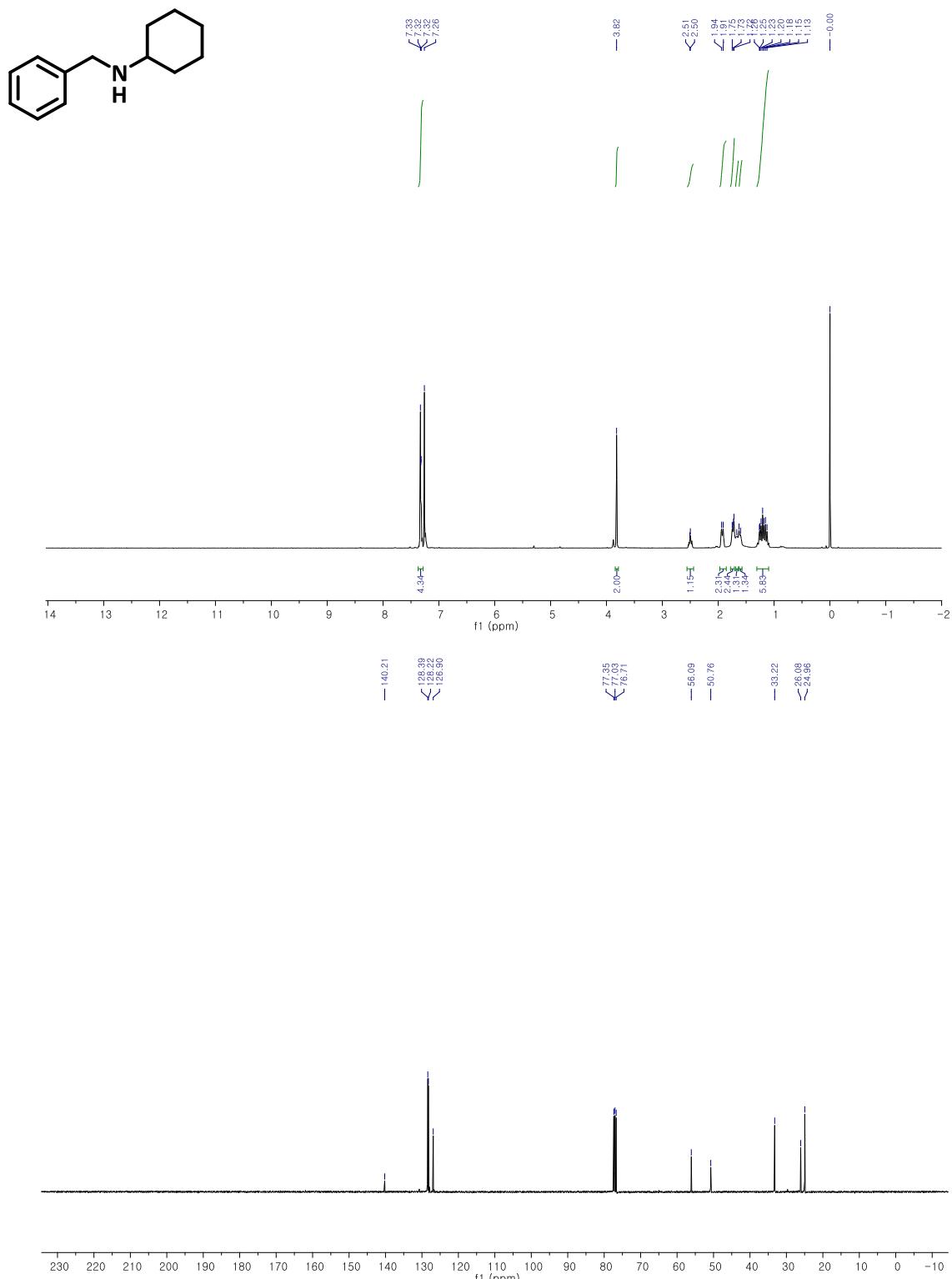
NMR S18, 3u, indole^[17]

93% (117mg, White solid); ^1H NMR (400 MHz, CDCl_3) δ 7.65 (d, $J = 7.9$ Hz, 1H), 7.41 (dd, $J = 8.1, 0.9$ Hz, 1H), 7.22 (t, $J = 2.8$ Hz, 1H), 7.19 (dd, $J = 8.1, 1.1$ Hz, 1H), 7.15 – 7.09 (m, 1H), 6.57 (s, 1H). ^{13}C NMR (101 MHz, CDCl_3) δ 135.8, 127.8, 124.2, 121.9, 120.8, 119.83, 111.0, 102.6.



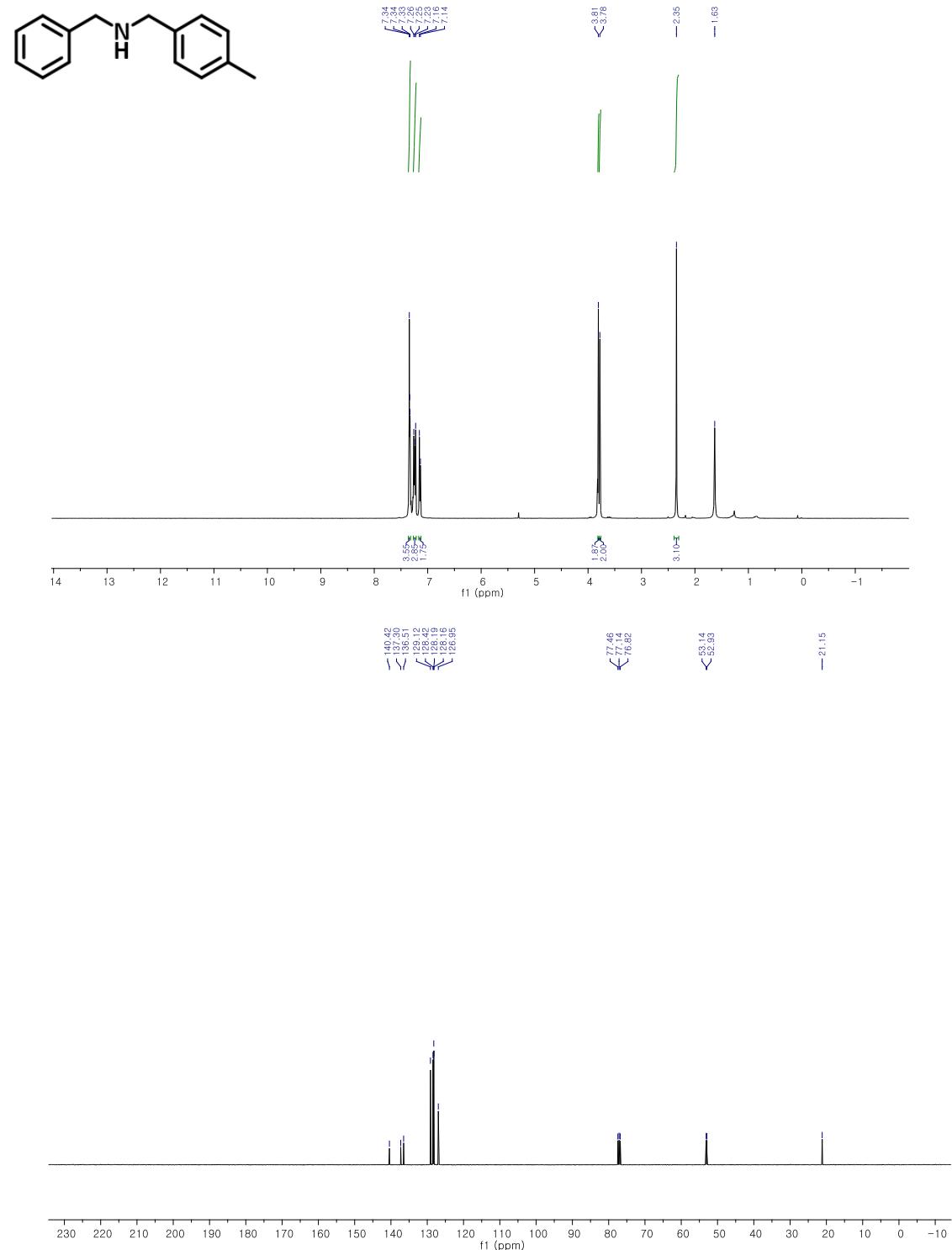
NMR S19, 5b, N-benzylcyclohexanamine^[18]

64% (60mg, Yellow oil); ¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.26 (m, 5H), 3.80 (s, 2H), 2.49 (d, *J* = 3.6 Hz, 1H), 1.91 (d, *J* = 11.4 Hz, 2H), 1.78 – 1.69 (m, 2H), 1.62–1.60 (br, 1H), 1.29 – 1.06 (m, 5H). ¹³C NMR (101 MHz, CDCl₃) δ 140.2, 128.4, 128.2, 126.9, 56.1, 50.8, 33.2, 26.1, 24.9.



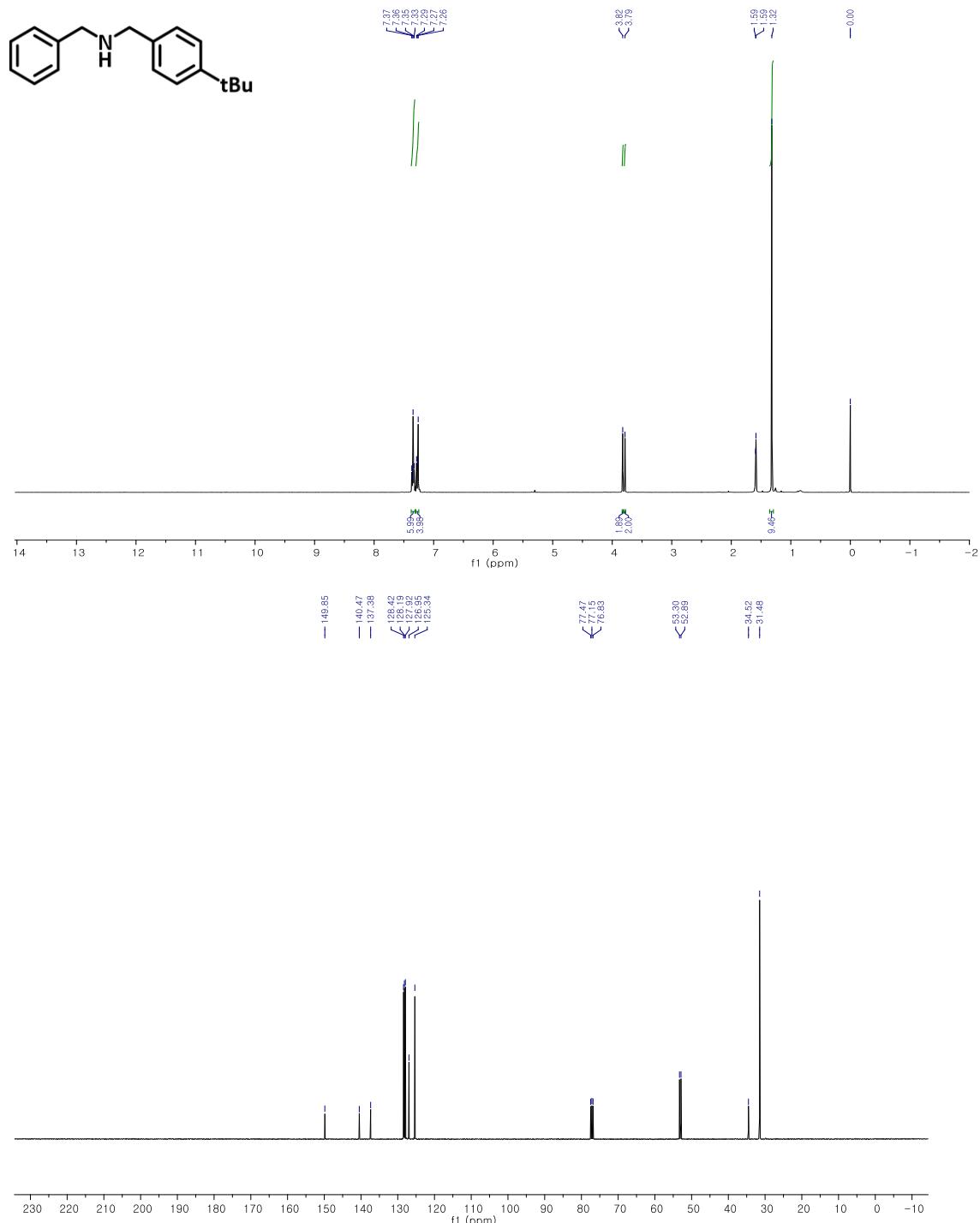
NMR S20, 5c, N-benzyl-4-methylbenzylamine^[19]

81% (80 mg, Yellow oil); ¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.31 (m, 4H), 7.25–7.20 (m, 3H), 7.12 (d, J = 7.9 Hz, 2H), 3.81 (s, 2H), 3.78 (s, 2H), 2.35 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.4, 137.3, 136.5, 129.1, 128.4, 128.2, 128.1, 126.9, 53.1, 52.9, 21.1.



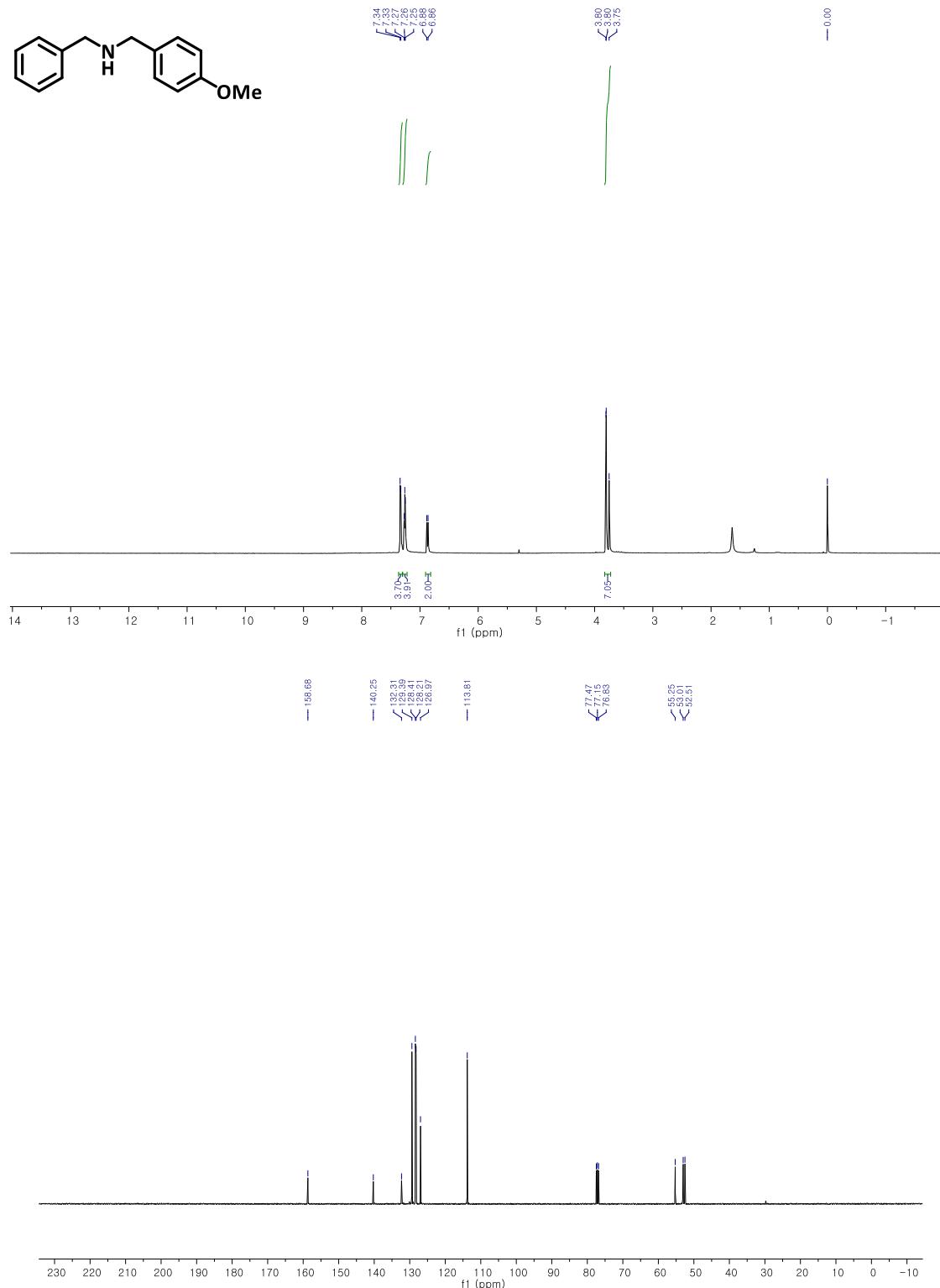
NMR S21, 5d, N-benzyl-1-(4-tert-butylbenzyl)amine^[20]

78% (99mg Yellow oil); ^1H NMR (400 MHz, CDCl_3) δ 7.37-7.33 (m, 5H), 7.29-7.16 (m, 4H), 3.82 (s, 2H), 3.79 (s, 2H), 1.32 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 149.9, 140.5, 137.4, 128.4, 128.2, 127.9, 127.9, 126.9, 125.3, 53.3, 52.9, 34.5, 31.5.



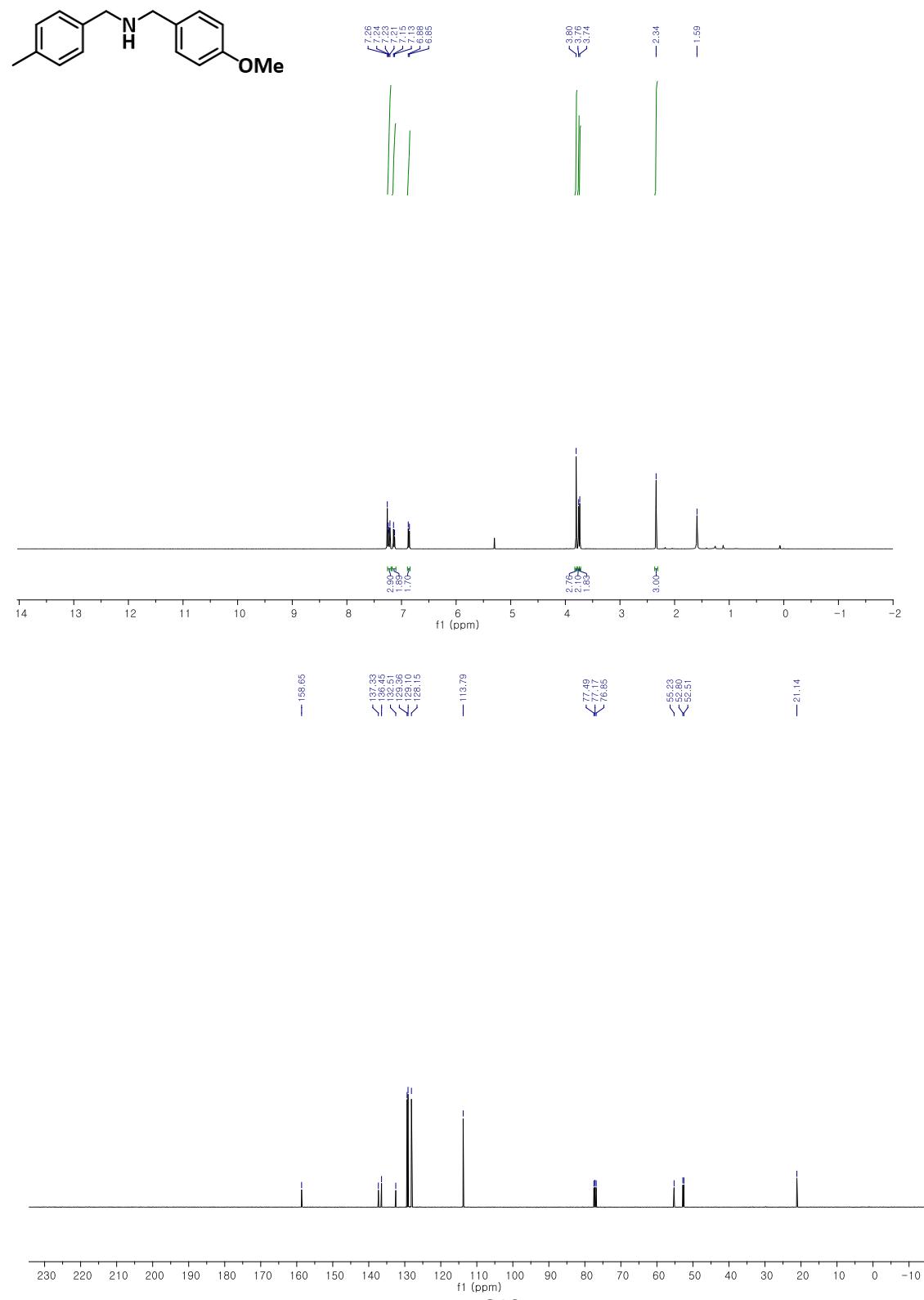
NMR S22, 5e, N-benzyl-1-(4-methoxyphenyl)methanamine^[12]

92% (104 mg, Yellow oil); ¹H NMR (400 MHz, CDCl₃) δ 7.34 - 7.22 (m, 7H), 6.87-6.86 (m, 2H), 3.83 – 3.72 (m, 7H). ¹³C NMR (101 MHz, CDCl₃) 158.7, 140.2 123.3 129.4 128.4 128.2, 126.9, 113.8, 55.3, 53.0, 52.5.



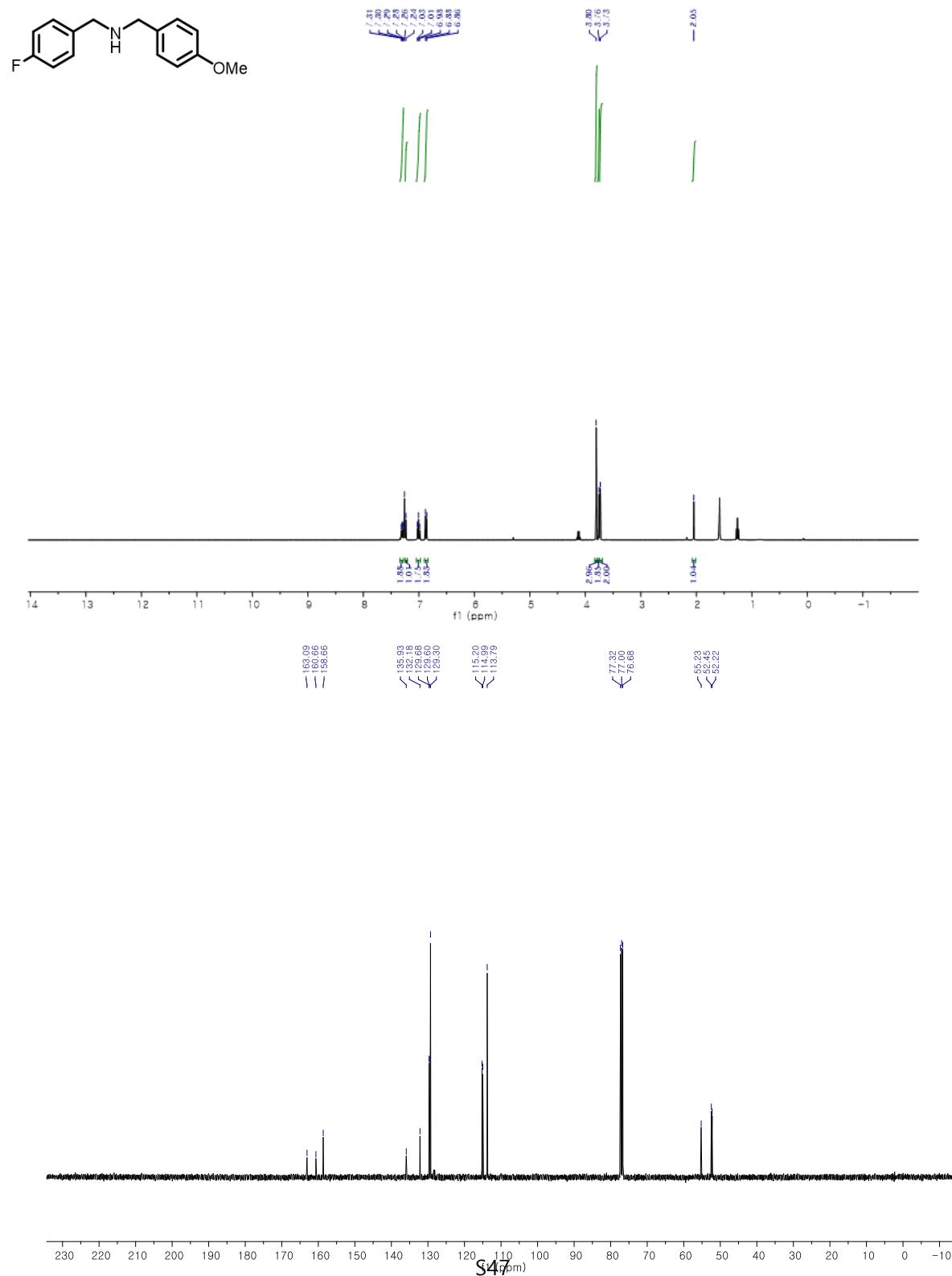
NMR S23, 5f, N-(4-methoxybenzyl)-1-(*p*-tolyl)methanamine^[14]

81% (104mg, Colorless oil); ¹H NMR (400 MHz, CDCl₃) δ 7.26 – 7.19 (m, 4H), 7.14 (d, *J* = 7.8 Hz, 2H), 6.87 (d, *J* = 8.6 Hz, 2H), 3.80 (s, 3H), 3.76 (s, 2H), 3.74 (s, 2H), 2.34 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 158.6, 137.3, 136.5, 132.5, 129.3, 129.1, 128.15, 113.8, 55.23, 52.80, 52.51, 21.1.



NMR S24, 5g, *N*-(4-fluorophenyl)methyl-1-(4-methoxyphenyl)methanamine^[21]

81% (99 mg, Yellow oil); ¹H NMR (400 MHz, CDCl₃) δ 7.30 (dd, *J* = 8.5, 5.6 Hz, 2H), 7.24 (m, 2H), 7.01 (t, *J* = 8.7 Hz, 2H), 6.87 (d, *J* = 8.6 Hz, 2H), 3.80 (s, 3H), 3.76 (s, 2H), 3.73 (s, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 161.88 (d, *J* = 244.4 Hz), 158.66, 135.93, 132.18, 129.64 (d, *J* = 7.9 Hz), 129.30, 115.10 (d, *J* = 21.2 Hz), 113.79, 55.23, 52.45, 52.22.



5. Reference

- [1] A. Galan, J. de Medoza, P. Prados, J. Rojo, A. M. Echavarren, *J. Org. Chem.* 1991, **56**, 452.
- [2] S. Lu, J. Wang, X. Cao, X. Li, H. Gu, *Chem. Commun.* 2014, **50**, 3512-3515.
- [3] L. Liu, Y. Liu, Y. Ai, J. Li, J. Zhou, Z. Fan, H. Bao, R. Jiang, Z. Hu, J. Wang, K. Jing, Y. Wang, Q. Liang, H. Sun, *iScience* 2018, **8**, 61-73.
- [4] Y. Li, Y. Gong, X. Xu, P. Zhang, H. Li, Y. Wang, *Catal. Commun.* 2012, **28**, 9-12.
- [5] S. Lu, P. Xu, X. Cao, H. Gu, *RSC Adv.* 2018, **8**, 8755-8760.
- [6] Z.-F. Jiao, J.-X. Zhao, X.-N. Guo, X.-L. Tong, B. Zhang, G.-Q. Jin, Y. Qin, X.-Y. Guo, *Catal. Sci. & Technol.* 2019, **9**, 2266-2272.
- [7] T. Ikawa, Y. Fujita, T. Mizusaki, S. Betsuin, H. Takamatsu, T. Maegawa, Y. Monguchi, H. Sajiki, *Org. Biomol. Chem.* 2012, **10**, 293-304
- [8] S. K. Sharma, J. Lynch, A. M. Sobolewska, P. Plucinski, R. J. Watson, J. M. J. Williams, *Catal. Sci. Technol.* 2013, **3**, 85-88.
- [9] Wetzel, S. Wockel, M. Schelwies, M. K. Brinks, F. Rominger, P. Hofmann and M. Limbach, *Org. Lett.* 2013, **15**, 266-269.
- [10] K. Orito, A. Horibata, T. Nakamura, H. Ushito, H. Nagasaki, M. Yuguchi, S. Yamashita and M. Tokuda, *J. Am. Chem. Soc.*, 2004, **126**, 14342-14343.
- [11] F. Mao, D. Sui, Z. Qi, H. Fan, R. Chen and J. Huang, *RSC Adv.* 2016, **6**, 94068-94073.
- [12] Z. Shao, S. Fu, M. Wei, S. Zhou and Q. Liu, *Angew. Chem. Int. Ed.* 2016, **55**, 14653-14657.
- [13] W. Zhang, X. Dong and W. Zhao, *Org. Lett.* 2011, **13**, 5386-5389.
- [14] H. Chung and Y. K. Chung, *J. Org. Chem.* 2018, **83**, 8533-8542.
- [15] P. T. K. Arachchige, H. Lee and C. S. Yi, *J.Org. Chem.* 2018, **83**, 4932-4947.
- [16] O. Saidi, A. J. Blacker, M. M. Farah, S. P. Marsden and J. M. Williams, *Chem. Commun.* 2010, **46**, 1541-1543.
- [17] Y. Motoyama, K. Kamo and H. Nagashima, *Org. Lett.* 2009, **11**, 1345-1348.
- [18] D. Wei, A. Bruneau-Voisine, D. A. Valyaev, N. Lugan and J. B. Sortais, *Chem. Commun.*, 2018, **54**, 4302-4305.
- [19] O. Y. Lee, K. L. Law and D. Yang, *Org. Lett.* 2009, **11**, 3302-3305.
- [20] S. Das, B. Join, K. Junge and M. Beller, *Chem. Commun.* 2012, **48**, 2683-2685.
- [21] C. J. Smith, C. D. Smith, N. Nikbin, S. V. Ley and I. R. Baxendale, *Org. Biomol. Chem.* 2011, **9**, 1927-1937.